

Supplementary information for

**Protein-based Sensitive, Selective and Rapid Fluorescence Detection  
of Picric Acid in Aqueous Media**

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## Experimental section

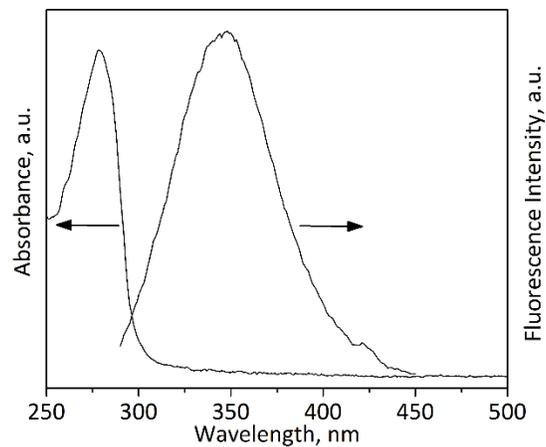
**Reagents.** Bovine serum albumin (BSA,  $\geq 98\%$ , agarose gel electrophoresis lyophilized) was purchased from Sigma Aldrich. Sodium phosphate buffer (0.01 M, pH=7.4) was used for preparation of BSA solution. Nitro based explosives such as picric acid (PA,  $\geq 99\%$ ), 2,4-dinitrotoluene (DNT, 97%), 1,3-dinitrobenzene (DNB,  $\geq 99\%$ ), 2,4-dinitrophenol (DNP,  $\geq 98\%$ ), 4-nitrophenol (4-NP, 99%), and 3-nitrophenol (3-NP, 99%) were purchased from Sigma Aldrich. 1,3,5-trinitroperhydro-1,3,5-triazine (RDX) (1000  $\mu\text{g/mL}$  in acetonitrile) was bought from Chem Service. 2,4,6-trinitrotoluene (TNT, 1000  $\mu\text{g/mL}$  in acetonitrile), 2,4,6-trinitrophenylmethylnitramine (Tetryl, 1000  $\mu\text{g/mL}$  in acetonitrile) was obtained from Ultra Scientific, while nitromethane (NM, 99+%) was bought from Acros Organics. 2-Nitrotoluene (2-NT,  $\geq 99.0\%$ ) was obtained from Fluka. Octogen (HMX, 1000  $\mu\text{g/mL}$  in acetonitrile: methanol) was purchased from SPEX CertiPrep. Pentaerythritol tetranitrate (PETN, 1000  $\mu\text{g/mL}$  in acetonitrile) was bought from Cerilliant.

**Equipment.** The absorption spectra were performed on a Cary 50 UV-vis spectrophotometer (Agilent Technologies). Fluorescence emission spectra were measured using a Varian Cary Eclipse fluorescence spectrometer (Agilent Technologies). Circular dichroism measurements were conducted on a JASCO (J-710) spectropolarimeter by fixing the concentrations of BSA while varying the analyte concentrations.

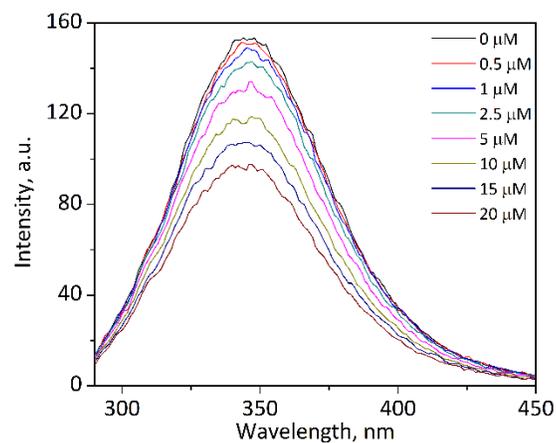
**Detection of nitro explosives in aqueous samples.** The sensing experiments were carried out by monitoring the fluorescence quenching behavior of BSA solution upon the addition of target explosives at room temperature through a fluorescence spectrometer, in which the excitation and emission slits width were adjusted according to the fluorescence intensities. Briefly, 3 mL of BSA solution was placed in a disposal cuvette and then the target solution was added successively. Each titration was repeated at least three times and consistent results are reported. The fluorescence emission experiments were excited at 279 nm, and the emission data were collected in the wavelength range of 290-450 nm. The quenching percentage (QP) was calculated using the equation as follows:

$$\text{QP \%} = (1 - I/I_0) \times 100\%$$

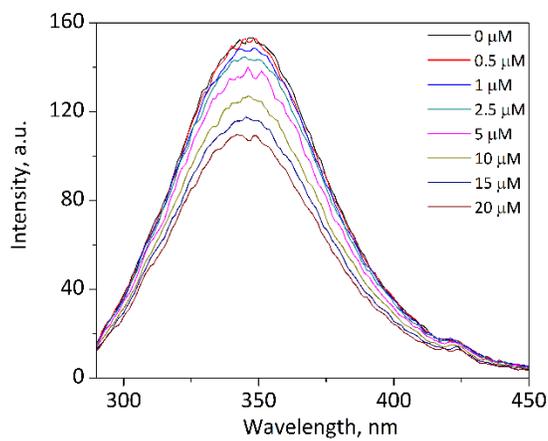
where  $I_0$  is the initial fluorescence intensity in the absence of analyte,  $I$  is the fluorescence intensity in the presence of corresponding analyte.



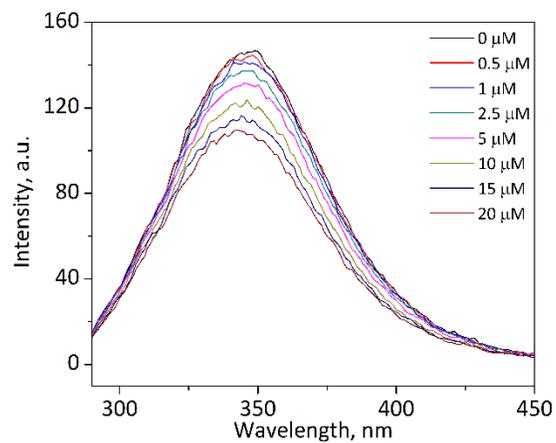
**Fig. S1** Absorption and emission spectra of BSA solution (emission spectrum upon excitation at 279 nm).



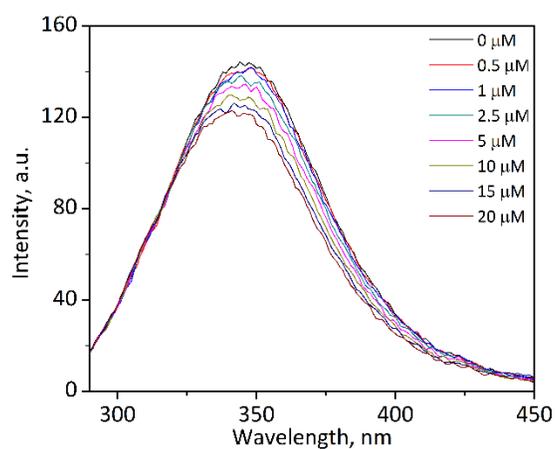
**Fig. S2** Fluorescence spectra of BSA upon addition of Tetryl in aqueous solution.



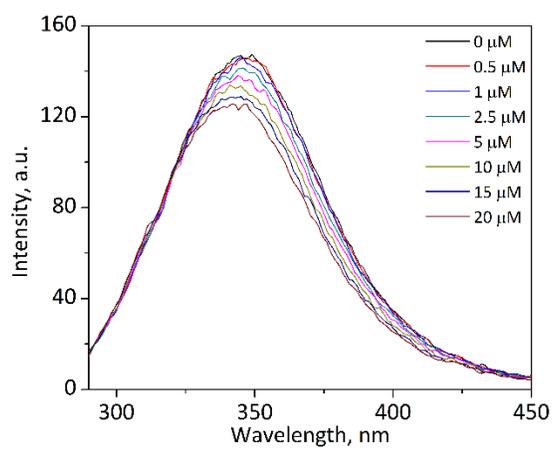
**Fig. S3** Fluorescence spectra of BSA upon addition of 2-NT in aqueous solution.



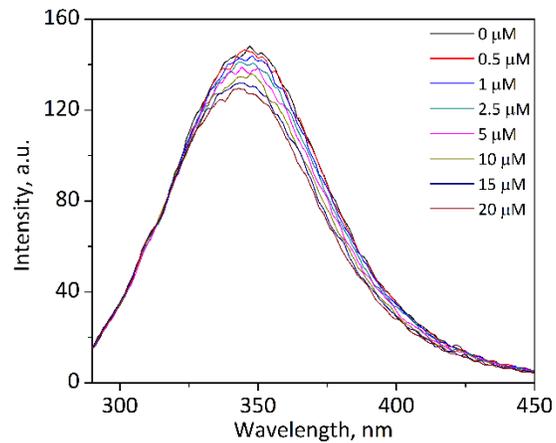
**Fig. S4** Fluorescence spectra of BSA upon addition of TNT in aqueous solution.



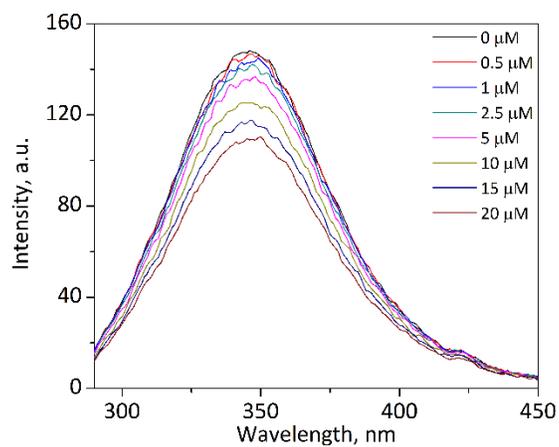
**Fig. S5** Fluorescence spectra of BSA upon addition HMX of in aqueous solution.



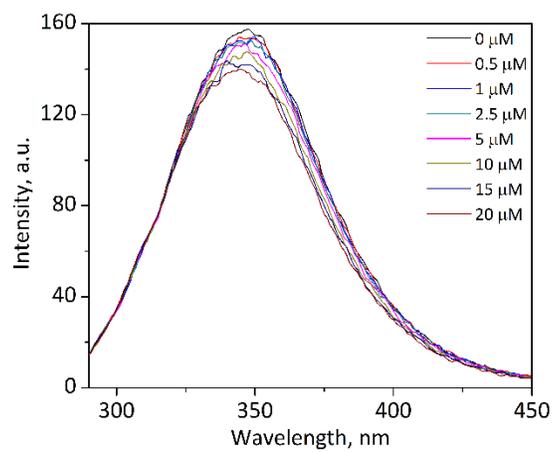
**Fig. S6** Fluorescence spectra of BSA upon addition of PETN in aqueous solution.



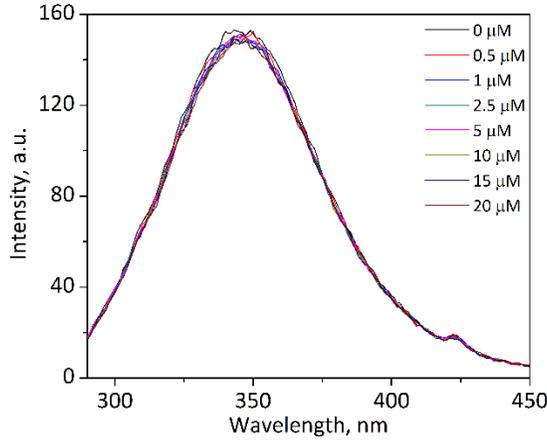
**Fig. S7** Fluorescence spectra of BSA upon addition RDX of in aqueous solution.



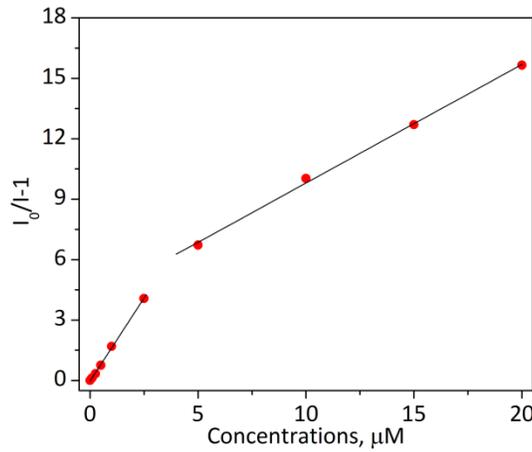
**Fig. S8** Fluorescence spectra of BSA upon addition of DNT in aqueous solution.



**Fig. S9** Fluorescence spectra of BSA upon addition of DNB in aqueous solution.



**Fig. S10** Fluorescence spectra of BSA upon addition of NM in aqueous solution.



**Fig. S11** Linear fitting of Stern-Volmer plot for PA explosive detection using 0.5 μM BSA.

The Stern-Volmer plot for PA in 0.5 μM BSA solution was shown in Fig. S11, and the fitting parameters were obtained. The S-V plot is nearly linear at low concentrations and deviates downward from linearity in a higher concentration range. In the low concentration range, the  $K_{SV}$  value was calculated as  $1.65 \times 10^6 \text{ M}^{-1}$ , and the correlation coefficient  $R^2=0.999$ ; In the high concentration range, the  $K_{SV}$  value was calculated as  $5.90 \times 10^5 \text{ M}^{-1}$ , and the correlation coefficient  $R^2=0.997$ .

The efficiency of energy transfer between the donor and the acceptor,  $E$ , could be calculated by the following equation<sup>1-3</sup>:

$$E = 1 - \frac{F}{F_0} = \frac{R_0^6}{R_0^6 + r_0^6} \quad (1)$$

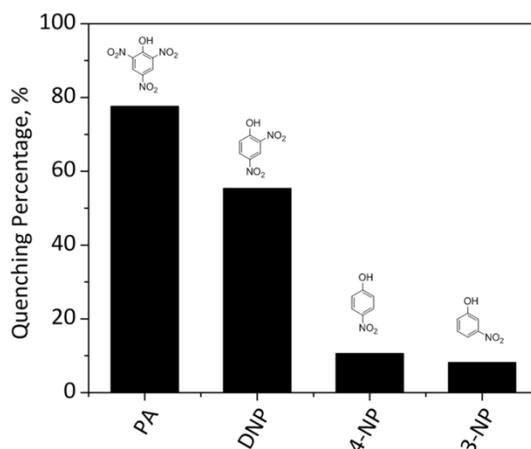
where  $r_0$  represents the distance between the donor and acceptor, and  $R_0$  is the critical distance at which energy transfer efficiency equals to 50%. The value of  $R_0$  is calculated using the equation:

$$R_0^6 = 8.79 \times 10^{-5} K^2 n^{-4} \Phi J(\lambda) \text{ (in } \text{\AA}^6 \text{)} \quad (2)$$

Where  $K^2$  is the orientation factor related to the geometry of the donor-acceptor dipole,  $n$  is the refractive index of the medium,  $\Phi$  is the fluorescence quantum yield of the donor, and  $J(\lambda)$  in  $M^{-1} \text{ cm}^{-1} \text{ nm}^4$ , expresses the degree of spectral overlap between the donor emission and the acceptor absorption,  $J(\lambda)$  is given by:

$$J(\lambda) = \frac{\int_0^\infty F(\lambda) \varepsilon(\lambda) \lambda^4 d\lambda}{\int_0^\infty F(\lambda) d\lambda} \quad (3)$$

Where  $F(\lambda)$  is the corrected fluorescence intensity of the donor in the wavelength  $\lambda$  to  $\lambda + \Delta\lambda$ ,  $\varepsilon(\lambda)$  is the extinction coefficient of the acceptor at  $\lambda$  in  $M \text{ cm}^{-1}$ . In the present case,  $K^2 = 2/3$ ,  $n = 1.36$ , and  $\Phi = 0.15$ . When  $C(\text{PA}) = C(\text{BSA}) = 0.50 \mu\text{M}$ ,  $E = 0.432$ , according to Eqs. (1)-(3), we could calculate that  $J(\lambda)$  for picric acid,  $J(\lambda) = 1.16 \times 10^{14} M^{-1} \text{ cm}^{-1} \text{ nm}^4$ ,  $R_0 = 2.58 \text{ nm}$ , and  $r_0 = 2.70 \text{ nm}$ . The degree of spectra overlap  $J(\lambda)$  for other analytes were also calculated and the results were shown in Table S4.



**Fig. S12** Comparison of quenching percentage of BSA fluorescent sensors upon addition of various 2.5  $\mu\text{M}$  nitrophenols.

As shown in Fig. S12, the quenching efficiency follows the order  $\text{PA} > \text{DNP} > 4\text{-NP} > 3\text{-NP}$ , which is in complete agreement with the order of acidity of these analytes, where the  $\text{pK}_a$  values of the quenchers are 0.38 for PA, 4.89 for DNP, 7.15 for 4-NP, and 8.4 for 3-NP. In addition, the quenching efficiency of PA and DNP are much higher than their corresponding structure-similar nitro compounds (TNT, DNT and DNB). This feature when coupled with the magnitudes of the overall integrals may explain the molecular basis for the unprecedented selectivity for PA, as other nitro compounds do not have a hydroxyl group and so they cannot interact efficiently with amino groups of BSA and so result in a very low quenching efficiency. In addition, the interaction of the hydroxyl groups with the basic sites in BSA is expected to follow the order of their acidity.

**Table S1** Comparison of present results with previous fluorescent picric acid sensors

| Reference   | $K_{SV}$ ( $M^{-1}$ )                | Detection limit | Selectivity      |
|---|--------------------------------------|-----------------|------------------|
| <i>J. Mater. Chem.</i> , 2009, 19, 7347             | /                                    | 4.8 ppb         | High             |
| <i>Adv. Funct. Mater.</i> , 2009, 19, 905;          | $1.09 \times 10^5$                   | 0.2 ppm         | High             |
| <i>Macromol. Rapid Commun.</i> 2010, 31, 834        | $9.09 \times 10^3$                   | 1 ppm           | /                |
| <i>Chem. Commun.</i> , 2011, 47, 4505               | /                                    | 98.5 ppb        | High             |
| <i>Chem. Commun.</i> , 2011, 47, 10046              | 2100                                 | /               | Low              |
| <i>J. Photochem. Photobiol., A</i> 2011, 217, 356   | $5 \times 10^4$                      | 2.3 ppb         | Low              |
| <i>ACS Appl. Mater. Interfaces</i> 2011, 3, 1245    | $5.7 \times 10^3$                    | 70 ppb          | High             |
| <i>Inorg. Chem.</i> 2011, 50, 1506                  | $1 \times 10^5$                      | /               | Low              |
| <i>Inorg. Chem.</i> 2012, 51, 13072                 | /                                    | /               | Low              |
| <i>Chem. Commun.</i> , 2012, 48, 7167               | $2.5 \times 10^5$                    | 0.4 ppm         | Low              |
| <i>J. Mater. Chem.</i> , 2012, 22, 11574            | $3.04 \times 10^4$                   | 23 ppb          | Moderate         |
| <i>Anal. Chem.</i> , 2012, 84, 8415                 | /                                    | 32.3 ppb        | High             |
| <i>Chem. Commun.</i> , 2012, 48, 5007               | $9.9 \times 10^4$                    | 3-300 ppb       | Low              |
| <i>Angew. Chem. Int. Ed.</i> , 2013, 52, 2881       | $3.5 \times 10^4$                    | 0.916 ppm       | High             |
| <i>ACS Appl. Mater. Interfaces</i> , 2013, 5, 672   | $1.55 \times 10^4$                   | 80.2 ppb        | Moderate         |
| <i>ACS Appl. Mater. Interfaces</i> , 2013, 5, 5373. | $3.2 \times 10^6$                    | 0.9 ppb         | High             |
| <i>Chem. Commun.</i> , 2013, 49, 4764.              | $1.9 \times 10^4$                    | /               | High             |
| <i>J. Am. Chem. Soc.</i> , 2013, 135, 17310;        | $7.8 \times 10^4$                    | 1 ppm           | High             |
| <i>J. Org. Chem.</i> , 2013, 78, 1306               | $3.8 \times 10^4$                    | /               | High             |
| <i>Chem. Commun.</i> , 2014, 50, 6031               | $5.28 \times 10^4$                   | /               | Moderate         |
| <i>ACS Applied Mater. Inter.</i> 2014, 6, 10722     | $1.32 \times 10^5$                   | 125 ppb         | High             |
| <i>Asian J. Org. Chem.</i> 2014, 3, 805             | $3.57 \times 10^5$                   | 0.23 ppb        | High             |
| <b>Present Study</b>                                | <b><math>1.65 \times 10^6</math></b> | <b>3.9 ppb</b>  | <b>Very high</b> |

**Table S2** Fluorescent quenching constant  $K_{SV}$  values for the various explosive analytes

| Analytes | $K_{SV}$ ( $M^{-1}$ ) |
|----------|-----------------------|
| PA       | $1.65 \times 10^6$    |
| Tetryl   | $2.85 \times 10^4$    |
| TNT      | $1.64 \times 10^4$    |
| DNT      | $1.77 \times 10^4$    |
| 2-NT     | $1.98 \times 10^4$    |
| DNB      | $5.39 \times 10^3$    |
| NM       | /                     |
| RDX      | $5.62 \times 10^3$    |
| PETN     | $7.92 \times 10^3$    |
| HMX      | $7.86 \times 10^3$    |

**Table S3** LUMO and HOMO energies calculated for the various explosive analytes (at the B3LYP/6-31G\* level of theory)<sup>4-6</sup>

| Analytes | LUMO (eV)    | HOMO (eV)    | Band gap (eV) |
|----------|--------------|--------------|---------------|
| PA       | -3.89(-4.32) | -8.22(-8.60) | 4.33(4.28)    |
| Tetryl   | -3.92        | -8.11        | 4.19          |
| TNT      | -3.48        | -8.44        | 4.96          |
| DNT      | -2.97        | -8.11        | 5.14          |
| 2-NT     | -2.75        | -7.56        | 4.81          |
| DNB      | -3.43        | -7.99        | 4.56          |
| NM       | -1.90        | -8.03        | 6.13          |
| RDX      | -2.53        | -8.25        | 5.71          |
| PETN     | -3.08        | -8.71        | 5.63          |
| HMX      | -2.72        | -8.30        | 5.58          |

**Table S4** Degree of spectra overlap integral  $J(\lambda)$  values for the various explosive analytes.

| Analytes | $J(\lambda)$ ( $M^{-1} \text{ cm}^{-1} \text{ nm}^4$ ) |
|----------|--|
| PA       | $1.16 \times 10^{14}$                                  |
| Tetryl   | $8.42 \times 10^{12}$                                  |
| TNT      | $7.10 \times 10^{12}$                                  |
| DNT      | $6.10 \times 10^{12}$                                  |
| 2-NT     | $1.19 \times 10^{13}$                                  |
| DNB      | $2.12 \times 10^{12}$                                  |
| NM       | $7.84 \times 10^{11}$                                  |
| RDX      | $7.72 \times 10^{11}$                                  |
| PETN     | $9.82 \times 10^{12}$                                  |
| HMX      | $1.08 \times 10^{12}$                                  |

## References

1. J. R. Lakowicz, *Principles of Fluorescence Spectroscopy*, Springer, Singapore, 2006.
2. N. Shahabadi and M. Maghsudi, *J. Mol. Struct.*, 2009, **929**, 193-199.
3. Y.-Z. Zhang, X. Xiang, P. Mei, J. Dai, L.-L. Zhang and Y. Liu, *Spectrochim. Acta, Part A*, 2009, **72**, 907-914.
4. J. C. Sanchez, A. G. DiPasquale, A. L. Rheingold and W. C. Trogler, *Chem. Mater.*, 2007, **19**, 6459-6470.
5. S. S. Nagarkar, B. Joarder, A. K. Chaudhari, S. Mukherjee and S. K. Ghosh, *Angew. Chem. Int. Ed.*, 2013, **52**, 2881-2885.
6. Z. Hu, B. J. Deibert and J. Li, *Chem. Soc. Rev.*, 2014, <http://dx.doi.org/10.1039/C1034CS00010B>.