Core-shell nano-sized magnetic molecularly imprinted solid phase extractant coupled with HPLC for the selective isolation and determination of 17βestradiol in lake water sample

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Fig. S1. Effect of volume ratio of ethanol to HAc in elution solvent.



Fig. S2. Molecular structures of estrogens used in this study.



Fig. S3. Reusability of CS-Fe₃O₄@E2-MIPs and CS-Fe₃O₄@NIPs.

Table S1

Polymers	Batch	1	2	3	4	5	6	Average
CS-Fe ₃ O ₄ @E2-MIPs	$Q (\mathrm{mg \ g^{-1}})$	16.42	16.32	16.39	16.45	16.37	16.30	16.38
	RSD	3.7	8.0	6.7	5.7	5.8	8.9	6.5
CS-Fe ₃ O ₄ @NIPs	$Q (\mathrm{mg}~\mathrm{g}^{-1})$	4.382	4.362	4.322	4.362	4.380	4.342	4.358
	RSD	3.2	5.1	9.4	5.4	3.6	7.6	5.7

Reproducibility of CS-Fe₃O₄@E2-MIPs and CS-Fe₃O₄@NIPs.

Table S2

- Lake water	E2									
	1.0 ng mL ⁻¹		10.0 ng ml	L-1	100.0 ng mL ⁻¹					
	Recovery (%)	RSD (%)	Recovery (%)	RSD (%)	Recovery (%)	RSD (%)				
Intra-day	94.2	4.3	96.7	3.7	98.3	3.2				
Inter-day	95.1	5.8	96.2	4.2	97.9	3.9				

Recoveries of CS-Fe₃O₄@E2-MIPs binding E2 for spiked lake water samples (n=5).

 C_{18} -SPE method was used to pretreat the spiked lake water sample, and the chromatogram was compared with MIP-SPE method developed in this work as shown in Fig. S3. It is obvious that there is no peak corresponding to other impurities in the chromatogram of spiked lake water which was pretreated by MIP-SPE (Fig. S4A). However, after being treated by C_{18} -SPE, the peak height of E2 was lower and some other peaks of impurities appeared in the chromatogram (Fig. S4B). The results confirm that CS-Fe₃O₄@E2-MIPs has good adsorption capacity and selectivity to E2.



Fig. S4. Chromatograms of elution of absorbed CS-Fe₃O₄@E2-MIPs (A), and C₁₈-SPE (B).



Fig. S5. Chromatograms of lake water sample spiked with E2 at the concentration of 100.0 ng $mL^{-1}(A)$, elution of absorbed CS-Fe₃O₄@E2-MIPs (B), and E2 standard sample (C).