

Supporting Information

A rhodamine hydrazide-based fluorescent probe for sensitive and selective detection of hypochlorous acid and its application in living cells

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1. UV-vis absorption spectra of probe **RGNH** with different levels of HOCl

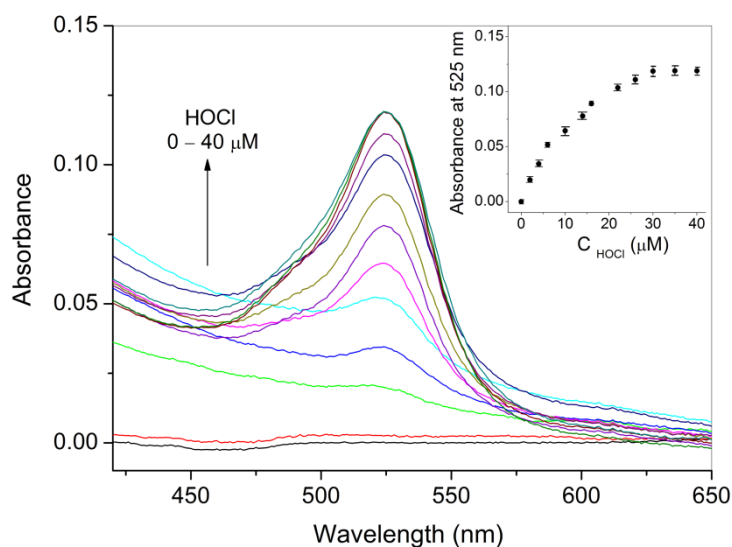


Fig. S1 UV-vis absorption spectra of probe **RGNH** (10 μM) with different levels of HOCl (bottom to top: 0, 2, 4, 6, 10, 14, 16, 22, 26, 30, 35, 40 μM) in PBS (10 mM, pH 7.4, 1% ethanol, v/v). Inset: Absorbance changes of probe **RGNH** (10 μM) at 525 nm as a function of HOCl concentration.

2. Effect of the ratio of ethanol to water on the fluorescence response of probe **RGNH** to HOCl

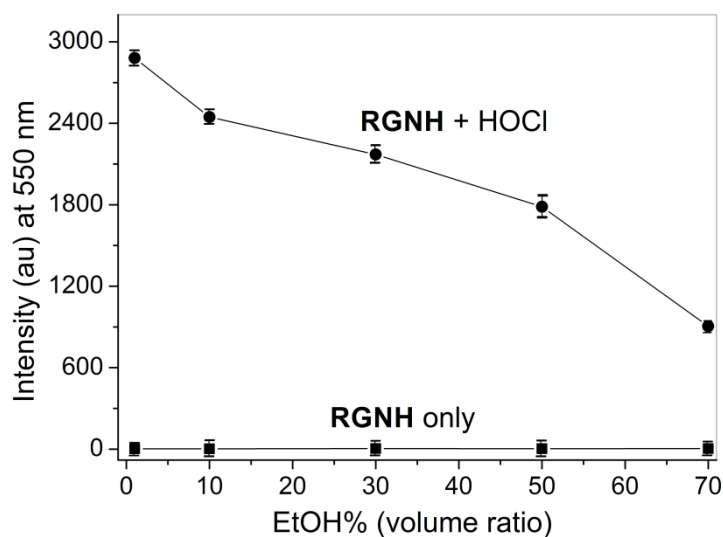


Fig. S2 Effect of the ratio of ethanol to water on the fluorescence intensity (550 nm) of probe **RGNH** (10 μM) in the absence and presence of HOCl (20 μM) in PBS (10 mM, pH 7.4). $\lambda_{\text{ex}} = 500 \text{ nm}$.

3. Kinetic behavior of the fluorescence intensity changes of probe **RGNH** with HOCl

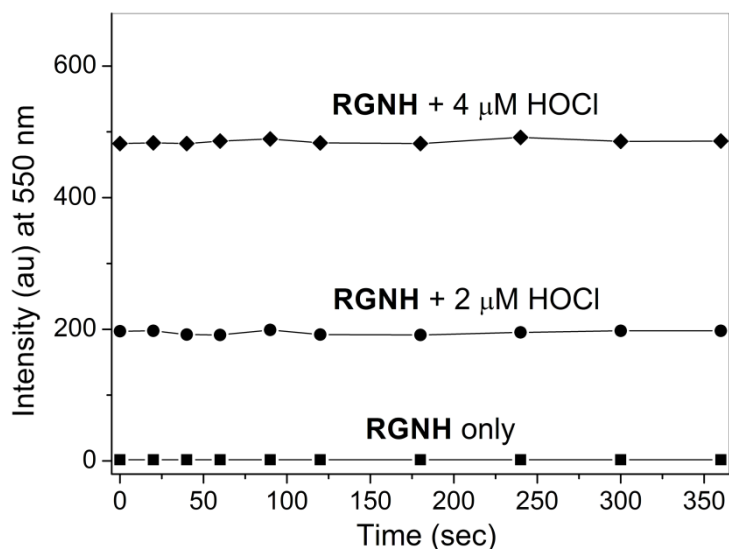


Fig. S3 Kinetic behavior of the fluorescence intensity changes (550 nm) of probe **RGNH** (10 μM) in the absence and presence of HOCl (2, or 4 μM) in PBS (10 mM, pH 7.4, 1% ethanol, v/v). $\lambda_{\text{ex}} = 500$ nm.

4. Fluorescence intensity changes of probe **RGNH** with HOCl and other ROS/RNS

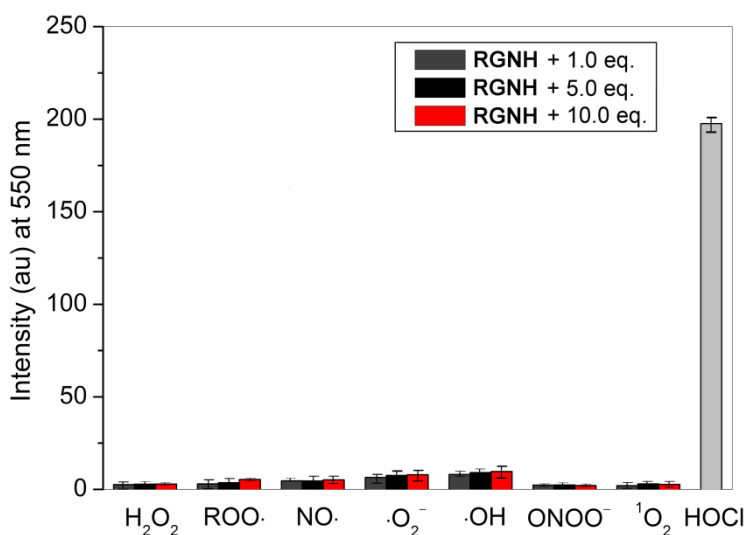


Fig. S4 Fluorescence intensity changes (550 nm) of probe **RGNH** (10 μM) upon addition of HOCl (2 μM) or other ROS/RNS (10, 50, 100 μM) in PBS (10 mM, pH 7.4, 1% ethanol, v/v). $\lambda_{\text{ex}} = 500$ nm.

5. Color changes of probe **RGNH** with HOCl and other ROS/RNS



RGNH only HOCl H₂O₂ ROO[·] NO[·] ·OH ·O₂⁻ ONOO⁻ ¹O₂⁻ HOCl + L-methionine

Fig. S5 Color changes of probe **RGNH** (10 μM) upon addition of HOCl (20 μM), other ROS/RNS (100 μM), or HOCl (20 μM) plus L-methionine (200 μM).

6. Analysis of the reaction product between probe **RGNH** and HOCl by high-resolution ESI-MS

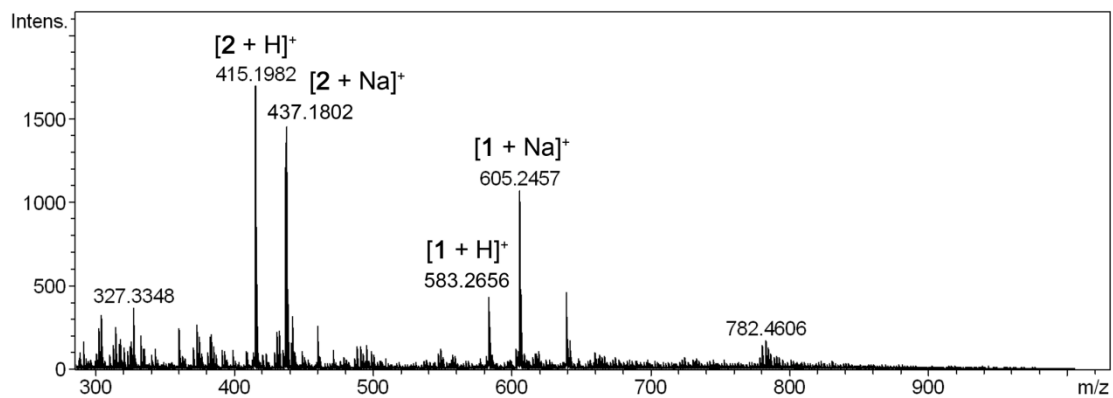
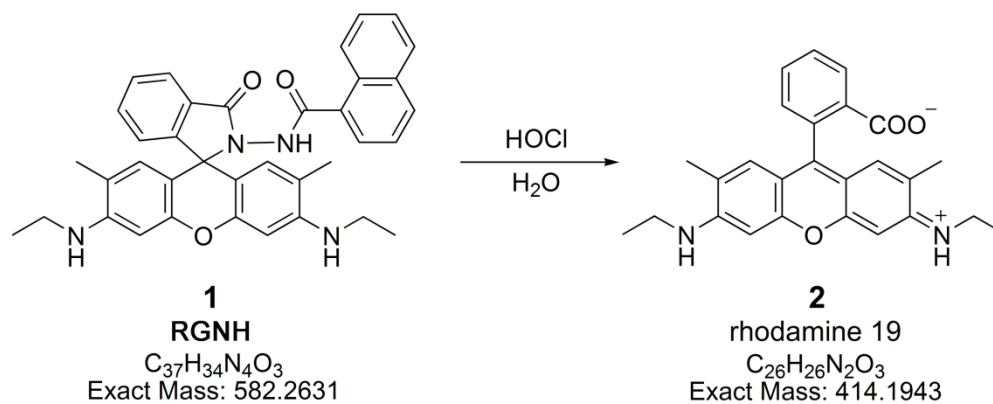


Fig. S6 High-resolution ESI-MS spectrum of probe **RGNH** in the presence of HOCl.

7. ESI-MS and ^1H NMR spectra of rhodamine 19, the product isolated from the reaction of probe **RGNH** with HOCl via column chromatography

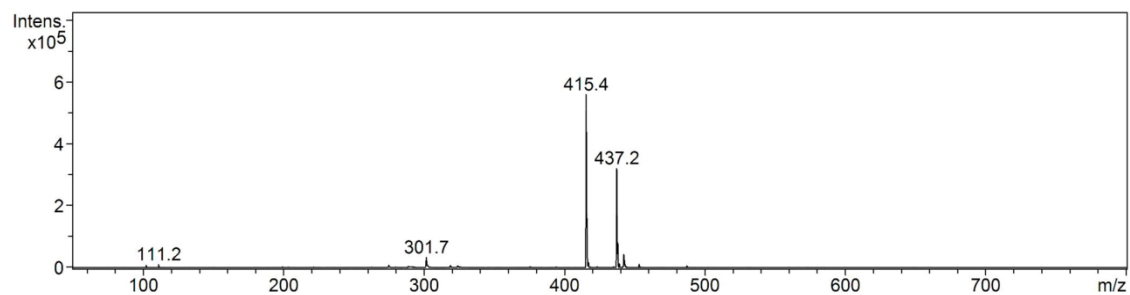
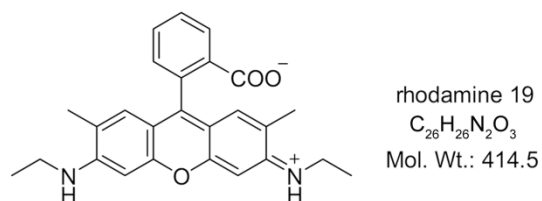


Fig. S7 ESI-MS spectrum of rhodamine 19.

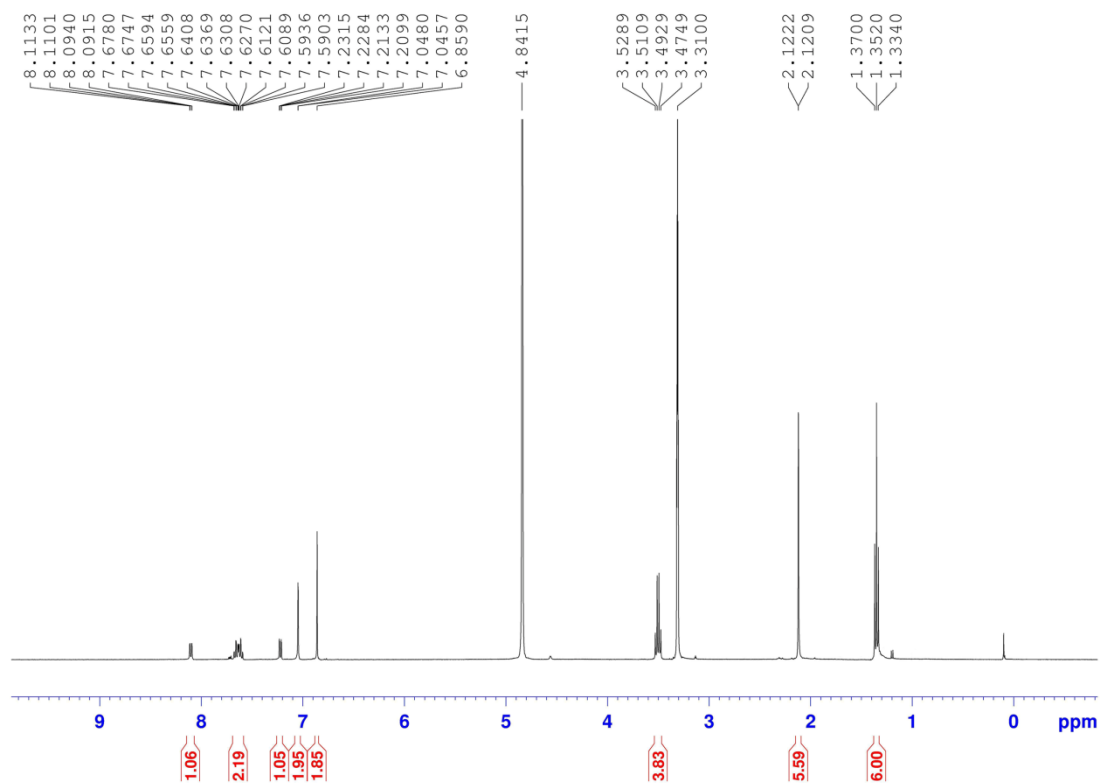


Fig. S8 ^1H NMR spectrum of rhodamine 19.

8. ESI-MS, high-resolution ESI-MS, ^1H NMR, and ^{13}C NMR spectra of probe **RGNH**

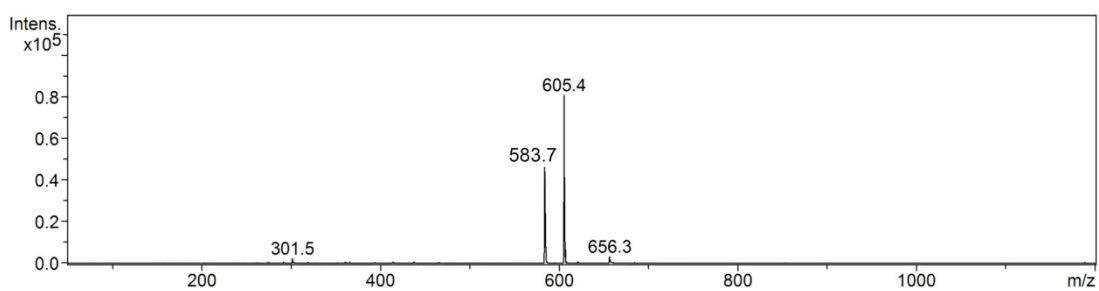
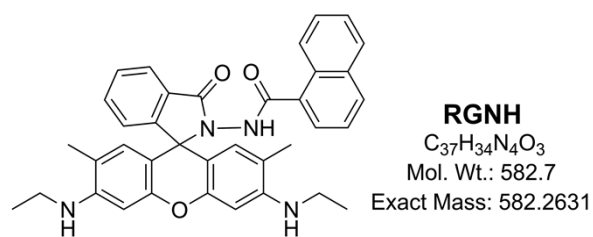


Fig. S9 ESI-MS spectrum of **RGNH**.

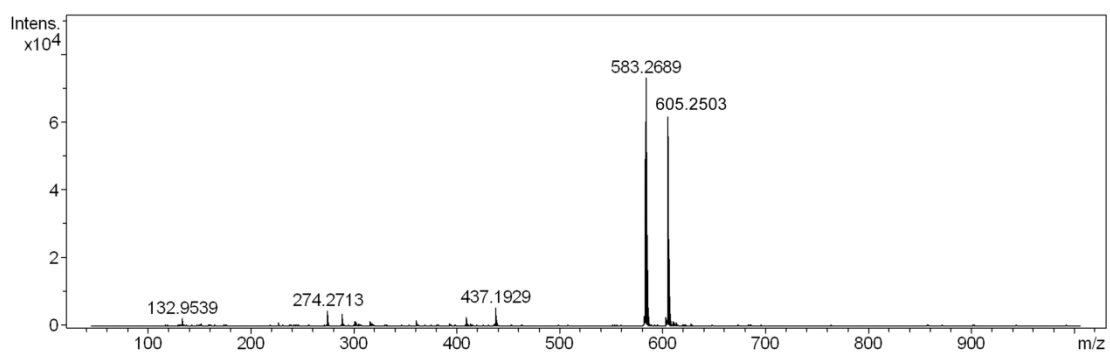


Fig. S10 High-resolution ESI-MS spectrum of **RGNH**.

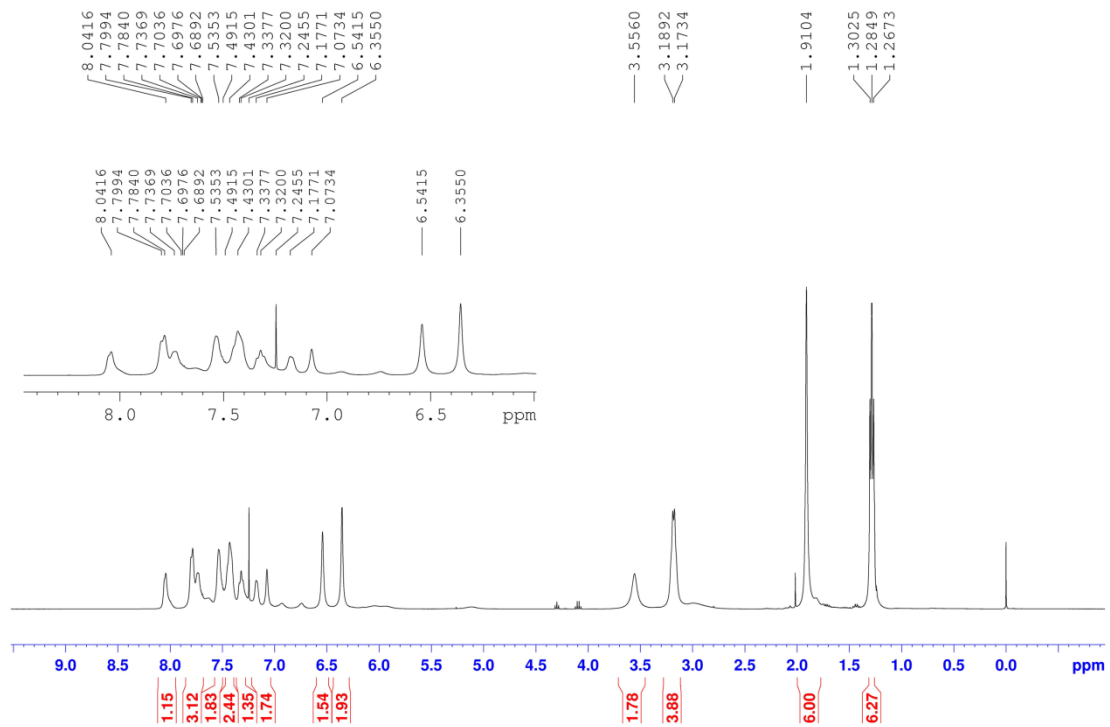


Fig. S11 ^1H NMR spectrum of RGNH.

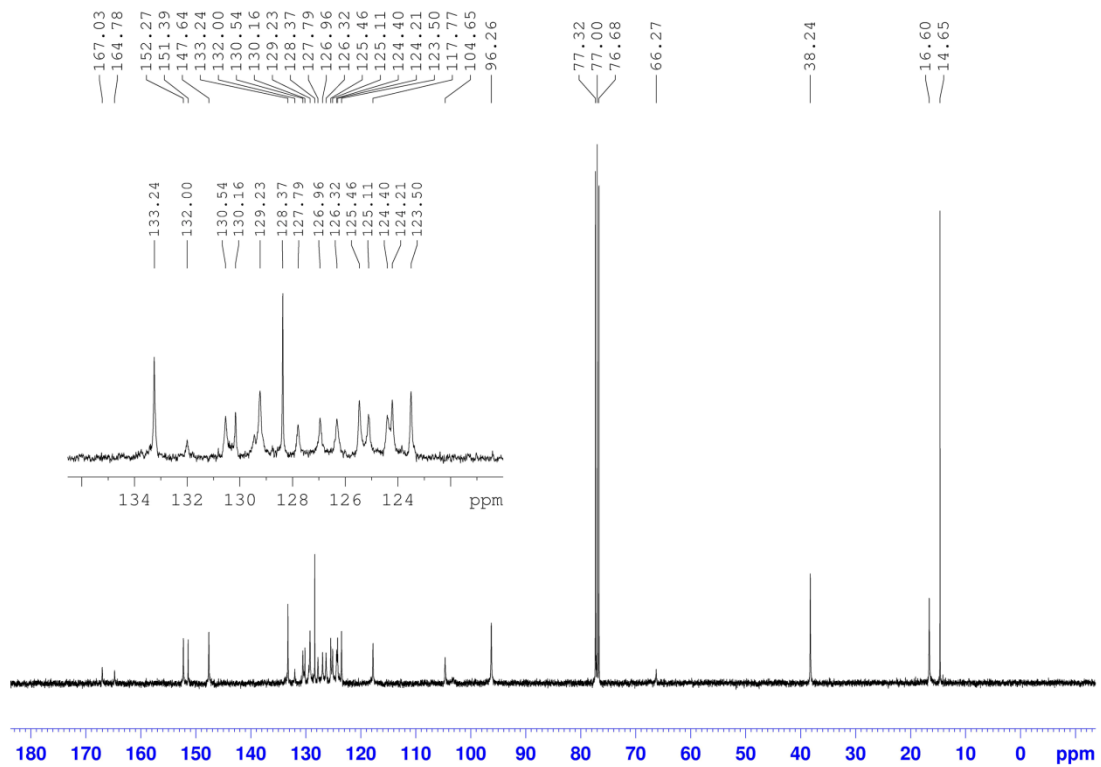
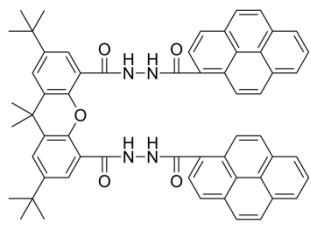
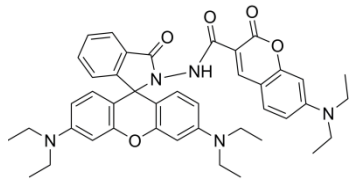
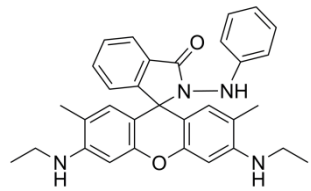
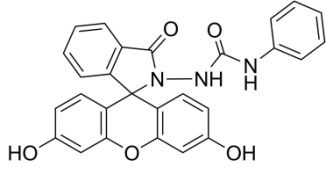
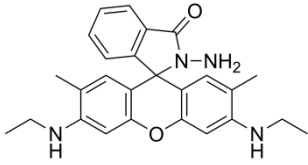
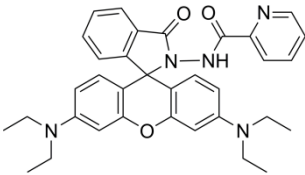
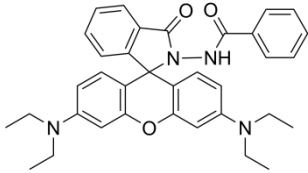
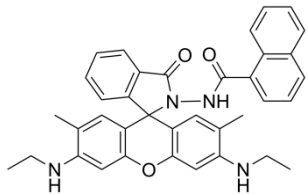


Fig. S12 ^{13}C NMR spectrum of RGNH.

9. Comparison of the recently reported hydrazone-based fluorescent probes for HOCl or OCl⁻

Table S1. Comparison of the recently reported hydrazone-based fluorescent probes for HOCl or OCl⁻.

Probe	Response time	Detection limit	pH span of detection	Comments
 <p><i>Anal. Methods</i>, 2014, 6, 609</p>	30 min	0.35 μM	5–9	HEPES buffer (pH 7.4) containing 50% (v/v) ethanol I ₃₇₅ /I ₄₉₅ = 152 fold Ratiometric fluorescent probe
 <p><i>Anal. Chim. Acta</i>, 2013, 775, 100</p>	within 1 min	0.024 μM	7.5–11	PBS buffer (pH 8.5) containing 40% (v/v) DMF I ₅₇₈ /I ₅₀₁ = 3955 fold Ratiometric fluorescent probe
 <p><i>Anal. Methods</i>, 2012, 4, 616</p>	more than 60 min	5.0 nM	6–11.3	water Fluorescent turn-on response I ₅₄₈ = over 82 fold 10 equiv Cu ²⁺ and Hg ²⁺ induced around 10-fold enhancement at 548 nm
 <p><i>Anal. Methods</i>, 2012, 4, 4334</p>	within 2 min	0.33 μM	no data available	PBS buffer (pH 7.4) with less than 1% ethanol Fluorescent turn-on response I ₅₂₈ = 156 fold Hg ²⁺ induced 80-fold enhancement at 530 nm

Probe	Response time	Detection limit	pH span of detection	Comments
 <p><i>Talanta</i>, 2011, 85, 779</p>	10 min	0.06 μM	4.5–8	<p>HEPES buffer (pH 7.4) containing 1% (v/v) DMF</p> <p>Fluorescent turn-on response</p> <p>$I_{550} = 270$ fold</p> <p>Cu^{2+} induced 100-fold enhancement at 550 nm</p> <p>Proton (especially under acid conditions, pH < 6) induced great enhancement at 550 nm</p> <p>Imaging both exogenous and endogenous HOCl in living cells</p> <p>Low cytotoxicity when probe concentration below 100 μM</p>
 <p><i>Org. Biomol. Chem.</i>, 2011, 9, 432</p>	10 s	~ 1.0 nM	4–13	<p>$\text{Na}_2\text{B}_4\text{O}_7/\text{NaOH}$ buffer (pH 12.0) containing 70% (v/v) methanol</p> <p>Fluorescent turn-on response</p> <p>$I_{580} > 420$ fold</p>
 <p><i>Chem. Eur. J.</i>, 2008, 14, 4719</p>	30 min	27 nM	9.2–12.7	<p>$\text{Na}_2\text{B}_4\text{O}_7/\text{NaOH}$ buffer (pH 12.0) containing 30% (v/v) THF</p> <p>Fluorescent turn-on response centered at 578 nm</p>
 <p><i>This work</i>, RGNH</p>	completed immediately after the sample solutions were prepared	3.3 nM	5–14	<p>PBS buffer (pH 7.4) with 1% (v/v) ethanol</p> <p>Fluorescent turn-on response</p> <p>$I_{550} = 1890$ fold</p> <p>Imaging both exogenous and endogenous HOCl in living cells</p> <p>Flow cytometry analysis of endogenous HOCl in living cells</p> <p>Low cytotoxicity when probe concentration below 100 μM</p>