



Fig. S1. Influences of different NaCl concentrentions in Buffer-3 containing 50 nM Hg<sup>2+</sup> on the performance of E-DNA sensor. All other conditions are identical. Data were evaluated by SWV.



Fig. S2. Voltammetric behavior of a clean Au electrode in 0.5 M H<sub>2</sub>SO<sub>4</sub>.

Table S1	Performance of	comparison betwee	n the proposal	strategy	and o	other
electroch	emical DNA de	tection assays for I	<b>Ig</b> <sup>2+</sup> .			

Method	Linear range	Detection	Analytical	Reference
		limit	technique	
DNA functionalized graphene	8-100 nM	5 nM	DPV	12
Hg <sup>2+</sup> catalyst HAuCl <sub>4</sub> /NH <sub>2</sub> OH	0.5-120 nM	0.06 nM	DPV	14
reaction				
Inhibition of activity urease enzyme	6-60 nM	56 nM	СТ	19
SRP enzymatic E-DNA sensor	0.5 nM-1 μM	0.3 nM	CV	20
DNA structure-switching	0.1 nM-5 µM	0.06nM	DPV	21
Hg <sup>2+</sup> -induced DNA hybridization	1nM -10 μM	0.6 nM	CV	22
DNA conformational switch and Exo		0.2 nM	DPV	23
III's activity				
DNA conformational switch	1 nM-2.0 µM	0.5 nM	ASV	38
T-Hg <sup>2+</sup> -T complex	0.1 nM-10	0.1 nM	EIS	39
	μΜ			
Our E-DNA senosr	0.01-500 nM	1 pM	SWV	

Abbreviations: SRP, streptavidin-horseradish peroxidase; E-DNA, electrochemical DNA; CV, cyclic voltammograms; EIS, electrochemical impedance spectroscopy; DPV, differential pulse voltammetry; SWV, square wave voltammetry; ASV, anodic stripping voltammetry; CT, chronoamperometry.



**Fig. S3** Fluorescence spectra ( $\lambda_{EX}$ =494nm;  $\lambda_{EM}$ =517nm) of Exo III incubation buffer at different incubation times (a-g: 0, 5, 10, 20, 30, 40 and 60 min) for detecting 50 nM Hg<sup>2+</sup>. 30 µL of buffer solution was diluted to 1 mL for the fluorescence test.