Supporting Information

Silver nanoparticles aggregates on metal fiber for solid phase microextraction-surface enhanced Raman spectroscopy detection of polycyclic aromatic hydrocarbons

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1. The SERS effect of oxidized AgNPs/Ag-Cu fiber.



Figure S1. The SERS effect of oxidized AgNPs/Ag-Cu fiber. The AgNPs/Ag-Cu fibers were stored in air for several days, and then immersed in 1.00×10^{-3} M propanethiol for 12 h. The SERS-active SPME fibers were immersed in 1.00×10^{-5} M fluoranthene and their SERS spectra were recorded. The intensity decreases with the increase of oxidation time. The SERS signals of fluoranthene disappear, when the AgNPs/Ag-Cu fibers are stored in air for 3 days.

2. Cyclic voltammetric of AgNPs/Ag-Cu fiber and PTH modified AgNPs/Ag-Cu fiber in





Figure S2. (A) Cyclic voltammetric of AgNPs/Ag-Cu fiber in 0.1 M KCl solution, (B) Cyclic voltammetric of PTH modified AgNPs/Ag-Cu fiber in 0.1 M KCl solution. The complete self-assembled monolayer over the entire surface of the nanoparticle was evaluated by cyclic voltammetric incorporating three electrodes: a working electrode, a reference electrode (Ag/AgCl) and an auxiliary electrode (Ti).¹⁻³

3. SEM image of commercial Ag-Cu fiber.



Figure S3. SEM image of commercial Ag-Cu fiber.

4. SEM images of the SERS-active fiber for sixteen cycles (A); twenty cycles (B); after



modification with PTH (C); Cross-sectional SEM image of the SERS-active fiber (D).

Figure S4. SEM image of silver nanoparticles on the silver-copper fiber with the assistant of Sn²⁺, for sixteen cycles (A), for twenty cycles (B); (C) SEM image of the PTH modified AgNPs/Ag-Cu fiber with sixteen cycles; (D) Cross-sectional SEM image of the SERS-active fiber .



5. The XPS spectra of the SERS-active fiber after modification with PTH.

Figure S5. The XPS spectra of the SERS-active fiber after modification with PTH. (A) Survey scan for the PTH modified silver nanoparticles, immobilized on silver-copper fiber; Ag 3d (B), S 2p (C) regions of the XPS spectrum of the PTH modified silver nanoparticles, immobilized on silver-copper fiber and (D) XPS spectra for the SPME fiber before propanethiol self-assembled (red) and after propanethiol self-assembled (black). All of the reported spectra have been normalized to the same counts value. Silver binding energy shifted to lower value by 0.2 eV might be attributing to the formation of

6. The XPS spectra of quantitative Ag3d.



Figure S6. The XPS spectra of quantitative Ag3d.

7. SERS spectra of PTH (black line), Raman spectrum of liquid PTH served as a reference (red line).



Figure S7. SERS spectra of PTH (black line), Raman spectrum of liquid PTH served as a

reference (red line). Compared with the normal Raman spectrum of liquid propanethiol,

both the number of Raman peaks and relative intensities had some differences.

Spectral data on the assignment of vibrational band of liquid propanethiol, SERS spectra of $Ag-S(CH_2)_2CH_3$ as shown in the following table.⁴⁻⁶

CH ₃ (CH ₂) ₂ SH, (pure liquid)	Assignment	Ag-S(CH ₂) ₂ CH ₃ , SERS(10 ⁻³ M),	Assignment	
369s	CCC bend,	703s	C-S stretch,	
	gauche		trans	
655vs	C-S stretch,	785vw	CH ₂ rock,	
	gauche		gauche	
710m	C-S stretch,	899w	CH ₂ rock,	
	trans		gauche	
735s	C-S stretch,	1030m	C-C stretch,	
	gauche		trans	
776m	CH ₂ rock,	1090m	C-C stretch,	
	gauche		gauche	
813w	CSH deformation,			
	trans			
1036vs	C-C stretch,			
	trans			

Band frequencies are given in cm⁻¹; vibration intensities are denoted as vs(very strong), s(strong), m(medium), w(weak), vw(very week); T and G represent rotational isomers of propanethiol in the trans and the gauche conformations with respect to the central C–C bond, respectively.

8. SERS spectra of fluoranthene on the SERS-active SPME fiber with different alkanethiol.



Figure S8. SERS spectra of fluoranthene $(1.00 \times 10^{-5} \text{ M})$ on the SERS-active SPME fiber with different alkanethiol.

9. Simulated (FDTD) spatial of the electric field of crossed silver nanoparticles.



Figure S9. Simulated (FDTD) spatial of the electric field of crossed silver nanoparticles.

10. The length of the three alkanethiols, which were optimized using Gaussian 03 program (B3LYP, 6-31G).



Figure S10. Structure optimization and length of alkanethiols.

11. The enhancement factor of different alkanethiol.



Figure S11. The enhancement factor of different alkanethiol, which was normalized d=0. 12. SERS spectra of fluoranthene $(1.00 \times 10^{-5} \text{ M})$ on the SERS-active SPME fiber with different cycles.



Figure S12. SERS spectra of fluoranthene $(1.00 \times 10^{-5} \text{ M})$ on the SERS-active SPME fiber with different cycles.

13. The stability of the SERS-active SPME fiber. SERS spectra of 1.00×10^{-5} M fluoranthene were recorded from fibers stored over a number of days under ambient

conditions.



Figure S13. (A) Stability of the SERS-active SPME fiber. SERS spectra of 1.00×10^{-5} M fluoranthene were recorded from fibers stored over a number of days under ambient conditions, (B) The normalized Intensity at 1612 cm⁻¹ over 7 days.

14. The uniformity of the SERS-active SPME fiber probed with 1.00×10^{-5} M fluoranthene without normalization.



Figure S14. The uniformity of the SERS-active SPME fiber probed with 1.00×10^{-5} M fluoranthene without normalization.

15. The reproducibility of the SERS-active SPME fibers probed with 1.00×10^{-8} M fluoranthene.



Figure S15. The reproducibility of the SERS-active SPME fibers probed with 1.00×10^{-8}

M fluoranthene.

16. Structures of PAHs in this work.



Fluoranthene

Pyrene

Benzo(b)fluoranthene

Figure 16. Structures of PAHs in this work

17. The SERS spectra of benzo(b)fluoranthene



Figure S17. The SERS spectra of benzo(b)fluoranthene

18. Selection of the internal standard peak.

Raman peak position	1030 cm ⁻¹	1090 cm ⁻¹
Vibrational mode	ν(C-C) _T	v(C-C) _G
RSD	5.66%	7.51%

Table S1. Comparison of the relative standard deviation (RSD) with different internal standard. T and G represent rotational isomers of propanethiol in the trans and the gauche conformations with respect to the central C-C bond, respectively.²



19. The calculation of limit of detection (LOD).

LOD was determined as three times the standard deviation.

y=three times the standard deviation.

x = the limit of detection.

	standard	liner equation	detection limit
	deviation		
fluoranthene	0.00209	Logarithm <i>y</i> =-1.54622+0.20912	7.56×10 ⁻¹⁰
		logarithm x	

Table S2. The LOD of fluoranthene.

20. The calculation of EF (Enhancement factor) of AgNPs/Ag-Cu fiber.

The SERS enhancement factor for p-aminothiophenol was calculated by the following equation:

 $EF = (I_{SERS}/I_{Raman}) \cdot (N_{bulk}/N_{ads})$

$$N_{ads} = A/A_{sum} \cdot V_{ads} \cdot C_1$$

$$N_{bulk} = A \cdot H_{eff} \cdot C_{sol}$$

N_{ads} is the number of p-aminothiophenol molecules under laser radiation adsorbed on substrate;

A_{eff} is the effective area of spot size;

A_{sum} is the area of the fiber substrate, id=0.35mm, length=1cm;

 V_{ads} is the volume that is spotted onto the paper substrate, 10 μ L;

 C_1 is the concentration of the solution that is to be measured.

 N_{bulk} is the number of p-aminothiophenol molecules in the scattering volume in bulk solution;

 H_{eff} is the effective length of of the scattering volume and here was estimated as the depth of field, 2.2 mm;

C_{sol} is 0.1 M p-aminothiophenol power for the non-SERS measurement.

 $EF(fluoranthene) = (I_{SERS}/I_{Raman}) \cdot (A_{eff} \cdot H_{eff} \cdot C_{sol}) / (A_{eff} / A_{sum} \cdot V_{sum} \cdot C_1)$

 $= (I_{SERS}/I_{Raman}) \cdot (A_{sum} \cdot H_{eff} \cdot C_{sol}) / (V_{ads} \cdot C_1)$ $= 0.0024178 \cdot (I_{SERS} \cdot C_{sol}) / (I_{Raman} \cdot C_1)$ $= 1.59 \times 10^5$

References

1. K. Bandyopadhyay, M. Sastry, V. Paul, K. Vijayamohanan, *Langmuir* **1997**, 13, 866-869.

- 2. M. French, S. E. Creager, Langmuir 1998, 14, 2129-2133.
- 3. J. D Zhang, Q. J. Chi, J. Ulstrup, Langmuir 2006, 22, 6203-6213
- 4. R. E. Pennington, D. W. Scott, H. L. Finke, J. P. McCullough, J. F. Messerly, I. A.

Hossenlopp, G. Waddington, J. Am. Chem. Soc. 1956, 78, 3266-3272.

5. A. Kudelski, J. Raman Spectrosc. 2003, 34, 853-862.

6. N. D. Strekal, A. E. German, G. A. Gachko, S. A. Maskevich, Optics and Spectroscopy

2000, 89, 834-840.