Electronic Supplementary Material

Carboxylic-group-functionalized single-walled carbon nanohorns as

peroxidase mimetics and their application to glucose detection

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Fig. S1 FTIR spectra of SWCNHs-COOH.



Fig. S2 Dependence of the SWCNHs-COOH peroxidase-like activity on A) pH, B) temperature, C) H_2O_2 concentrations, and D) SWCNHs-COOH concentrations. Experiments were carried out using 20 µg mL⁻¹ SWCNHs-COOH in 500 µL of 0.1 M phosphate buffer with 0.8 mM TMB as substrate. The concentration of H_2O_2 was 100 mM at pH 3.5 and 35 °C unless otherwise stated. The maximum point in each curve was set as 100%.

Catalyst	Substrate	K _m (mM)	V _{max} (10 ⁻⁸ M S ⁻¹)	Reference
SWCNHs-COOH	H ₂ O ₂	49.8	2.07	Present work
	ТМВ	0.506	2.28	
HRP	H ₂ O ₂	3.70	8.71	- 1
	ТМВ	0.434	10.0	
HCNTs	H ₂ O ₂	0.02	-	2
	ТМВ	41.42	-	
C-Dots	H ₂ O ₂	26.77	30.61	- 3
	ТМВ	0.039	3.61	
GO-COOH	H ₂ O ₂	3.99	3.85	- 4
	ТМВ	0.0237	3.45	
C ₆₀ [C(COOH) ₂] ₂	H ₂ O ₂	24.58	4.011	5
	ТМВ	0.2333	3.473	

Table S1 Comparison of the apparent Michaelis-Menten constant (K_m) and maximumreaction rate (V_{max}) between SWCNHs-COOH and other enzyme mimics



Fig. S3 Typical absorption spectra of the colorimetric method by using GOx and SWCNHs-COOH catalyzed color reaction in the absence (a) and the presence (b) of 2 mM glucose.



Fig. S4 Typical absorption spectra of the colorimetric method by using GOx and SWCNHs-COOH catalyzed color reaction in the absence of 0, 0.1, 0.3, 0.5, 0.7, 1.0, 1.5, 2.0 mM glucose. Inset: the linear calibration plot for glucose determination. The error bars represent the standard deviation of the three measurements.

Enzyme mimics	Linear range (µM)	Detection limit (µM)	Refs.
SWCNHs-COOH	100-2000	100	Present work
HCNTs	0.5-115	0.12	2
C-Dots	1-500	0.4	3
GO-COOH	1-20	1	4
C ₆₀ [C(COOH) ₂] ₂	1-40	0.5	5

Table S2. Comparison of colorimetric methods based on enzyme mimics for glucose detection.

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