Electronic Supplementary Information (ESI)

Fluorescence sensing ADP over ATP and PPi in 100% aqueous solution

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1. General Experimentals.

Starting materials were purchased from commercial suppliers and were used without further purification. 9,10-bisaminomethylanthracene (2)and 2-Bromomethyl-6-pivalamidopyridine (3) were prepared according the previously published methods (see below). All solvents were purified by the most used methods before use. Distilled water was used after passing through a Millipore-Q ultrapurification system. NMR spectra were measured on Varian Mercury 400 and 600 instruments. HR-MS data were obtained with an LC/Q-TOF MS spectrometer. UV-vis spectra and fluorescent spectra were recorded on an Agilent Cary 100 UV-vis spectrophotometer and an Agilent Cary Eclipse fluorescence spectrophotometer, respectively. Both UV-vis and fluorescence spectrophotometer are equipped with a temperature controller. Standard quartz cuvettes with a 10 mm lightpath are used for all UV-vis spectra and fluorescent spectra measurements. Cell imaging was performed in an inverted fluorescence microscopy with a 20× objective lens.

Fluorescent Studies of ZnL upon Addition of Analytes. For a typical optical measurements, 3.0 mL of ZnL (5 μ M) in a HEPES buffer (10 mM, pH 7.2) was placed in a quartz cell at 25 °C. The fluorescent spectra were then recorded immediately after addition of analytes.

The fluorescence quantum yields of **ZnL**, **ZnL**-2ADP and **ZnL**-2ATP were determined to be $\Phi = 0.02$, 0.99, and 0.50 in HEPES buffer (10 mM, pH 7.2) at 25°C, using quinine sulfate ($\Phi_f = 0.58$ in 1N H₂SO₄) as standard.

Cell Culture and Imaging. HeLa cells were cultured in Dulbecco's Modified Eagle's Medium (DMEM) supplemented with 10% FBS (fetal bovine serum), 100 mg/mL penicillin and 100 μ g/mL streptomycin in a 5% CO₂, water saturated incubator at 37 °C. Before cell imaging experiments, HeLa cells were seeded in 12-well culture plate for one night. For living cell imaging experiments, cells were incubated with 25 μ M of **ZnL** for 30 min at 37 °C and washed three times with prewarmed PBS buffer, and then imaged immediately. For apyrase treated experiments, HeLa cells were pretreated with 5 units apyrase for 30 min at 37 °C,

washed three times with prewarmed PBS buffer, and then incubated with 25 μ M of **ZnL** (or incubated with 125 μ M of ADP for 30 min prior to addition of probe **1** for a control experiment) for 30 min at 37 °C. Cell imaging was then carried out after washing cells with prewarmed PBS buffer.

2. Synthesis of ZnL



Synthesis of 1: To a solution of 2^{s1} (236 mg, 1.00 mmol) and 3^{s2} (1.14 g, 4.2 mmol) in anhydrous CH₃CN (30 mL) was added K₂CO₃ (621 mg, 4.50 mmol). The reaction mixture was then heated to reflux for 12 hours, and the solvent was evaporated under reduced pressure. To the residue was added dichloromethane (60 mL), and the organic phase was washed with water and brine followed by drying over Na₂SO₄. After removal of the solvent under reduced pressure, the crude product was purified by silica gel column (eluent: dichloromethane /methanol = 50:1 (v/v)) to give a yellow powder (800 mg, 80% yield). Mp 116-118 °C. ¹H NMR (400 MHz, CDCl₃) δ 8.41-8.39 (dd, *J* = 6.8, 3.2 Hz, 4H), 8.08 (s, 4H), 8.01 (d, *J* = 8.0 Hz, 4H), 7.50 (t, *J* = 8.0 Hz, 4H), 7.43 (dd, *J* = 6.8, 3.2 Hz, 4H), 6.98 (d, *J* = 7.6 Hz, 4H), 4.57 (s, 4H), 3.74 (s, 8H), 1.31 (s, 36H). ¹³C NMR (100 MHz, CDCl₃): δ 176.99, 157.59, 150.57, 138.35, 130.97, 130.31, 125.51, 124.83, 119.07, 111.82, 60.36, 51.16, 39.70, 27.42. IR (KBr) v_{max} (cm⁻¹): 3437, 2964, 1688, 1578, 1518, 1453, 1306, 1152. HR-MS Calc. for C₆₀H₇₃N₁₀O₄⁺ (M + H⁺) 997.5811, found 997.5791.





¹³C NMR spectrum of **1** in CDCl₃.



HR-MS spectrum of 1

Synthesis of L: To 600 mg of **1** was added 50 ml of 4 M KOH and CH₃CH₂OH 50 mL. The mixture was heated to reflux for 16 hours, and the CH₃CH₂OH was evaporated under reduced pressure. The aqueous solution was then extracted with ethyl acetate (4 × 30 mL), dried over Na₂SO₄, and concentrated in vacuo to yield a brown yellow solid, 330 mg (83%). Mp 254-256°C. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.52 (s, 4H), 7.48 (s, 4H), 7.30-7.31 (d, *J* = 4.0 Hz, 4H), 6.54-6.56 (d, *J* = 8.0 Hz, 4H), 6.26-6.28 (d, *J* = 8.0 Hz, 4H), 5.82 (s, 8H), 4.52 (s, 4H), 3.52 (s, 8H). ¹³C NMR (100 MHz, DMSO-*d*₆) δ 159.14, 157.40, 137.33, 130.73, 125.72, 125.16, 110.61, 106.05, 59.75, 50.09. IR (KBr) v_{max} (cm⁻¹): 3446, 3360, 3223, 2929, 1639, 1572, 1484, 1384, 1293, 1170, 1011, 833, 803, 764, 710; ESI-MS: m/z found 661.4 (M + H⁺); HR-MS Calc. for C₄₀H₄₁N₁₀⁺ (M + H⁺) 661.351, found 661.351. Elemental analysis calcd for C₄₀H₄₀N₁₀: C 72.70, H 6.10, N 21.20; found: C 72.71, H 6.037, N 21.15.

 $\begin{array}{c} -8.52 \\ -8.52 \\ 7.31 \\ 7.33 \\ 7.7.30 \\ 6.56 \\ 6.28 \\ -5.82 \\ -5.82 \\ -5.82 \\ -3.52 \\ -3.52 \\ -3.52 \\ -3.52 \\ -2.50 \\ -2$



¹³C NMR spectrum of **L** in DMSO- d_6 .



HR-MS spectrum of L

Synthesis of ZnL: To a solution of **L** (33 mg, 0.05 mmol) in 5 mL of MeOH, was added Zn(NO₃)₂·6H₂O (33 mg, 0.11 mmol), and the mixture was stirred for 1 h at rt. After concentrating under reduced pressure, the obtained solid was recrystallized from MeOH to afford **ZnL** (41mg, 81%). Mp >300 °C. ¹H NMR (400 MHz, DMSO): 8.08(s, 4H), 7.61(s, 4H), 7.54 (s, 4H), 6.93 (s, 8H), 6.72 (d, *J* = 7.8 Hz, 4H), 6.28 (s, 4H), 4.93 (s, 4H), 3.51-3.26 (m, 8H). ¹³C NMR (100 MHz, DMSO) δ 169.58, 161.69, 150.68, 142.06, 141.77, 136.35, 135.71, 122.00, 120.91, 66.88, 61.71. IR (KBr) v_{max} (cm⁻¹): 3446, 3360, 1639, 1572, 1484, 1384, 1293, 1170, 1011, 833, 803, 764. HR-MS Calc. for C₄₀H₄₀N₁₃O₉Zn₂⁺ [M-NO₃⁻]⁺ 974.1649, found 974.1953. Elemental analysis calcd for C₄₀H₄₀N₁₀Zn₂(NO₃)₄·H₂O: C 45.43, H 4.00, N 18.54; found: C 45.52, H 3.91, N 18.75.



¹³C NMR spectrum of **ZnL** in DMSO- d_6 .



HR-MS spectrum of ZnL

(S1) Gassensmith, J. J.; Arunkumar, E.; Barr, L.; Baumes, J. M.; DiVittorio, K. M.; Johnson, J. R.; Noll, B. C.; Smith, B. D. J. Am. Chem. Soc. 2007, 129, 15054–15059.

(S2) Livieri, M.; Mancin, F.; Tonellato, U.; Chin, J.; Cowley, A. Chem. Commun. 2008, 392–394.

3. Additional Spectra and Data



Figure S1. Fluorescence colour (under 365 nm light) changes of **ZnL** (5 μ M) in 10 mM aqueous HEPES buffer solution (pH = 7.2) with 50 μ M of different anions (from left to right: none, ATP, ADP, AMP, PPi, Citrate, PO₄³⁻, HPO₄²⁻, H₂PO₄⁻, F⁻, Cl⁻, Br⁻, I⁻, ClO₄⁻, CO₃²⁻, HCO₃⁻, NO₃⁻, NO₂⁻, AcO⁻, N₃⁻, SO₄²⁻, HSO₄⁻, SCN⁻, S₂O₇²⁻, S²⁻, S₂O₃²⁻).



Figure S2. The effect of pH on the fluorescence intensity changes of **ZnL** (5 μ M) in the absence and in the presence of ATP (50 μ M) or ADP (50 μ M) at 25°C. $\lambda_{ex} = 380$ nm, $d_{em} = d_{ex} = 5$ nm.



Figure S3. Left: Fluorescence emission spectra of **ZnL**(5 μ M) in the presence of various nucleotides (50 μ M); Right: A plot of the relative fluorescent intensity (I/I₀) of **ZnL** (5 μ M) at 416 nm against 50 μ M of different nucleotides (from left to right: none, ADP, GTP, GDP, GMP, UTP, UDP, UMP, CTP, CDP, CMP). $\lambda_{ex} = 380$ nm, $d_{em} = d_{ex} = 5$ nm.



Figure S4. Fluorescence colour (under a 365 nm light) changes of **ZnL** (5 μ M) in 10 mM aqueous HEPES buffer solution (pH = 7.2) with 50 μ M of different anions (from left to right: none, ATP, ADP, AMP, GTP, GDP, GMP, UTP, UDP, UMP, CTP, CDP, CMP).



Figure S5. Left: Fluorescent titration of complex **ZnL** (5 μ M) upon the addition of AMP (0-1250 μ M) in HEPES buffer (10 mM, pH = 7.2) solution at 25°C; Right: The saturation curve of fluorescent intensity changes at 430 nm. $\lambda_{ex} = 380$ nm, $d_{em} = d_{ex} = 5$ nm.



Figure S6. Left: Fluorescent titration of complex **ZnL** (5 μ M) upon the addition of CTP (0-50 μ M) in HEPES buffer (10 mM, pH = 7.2) solution at 25°C; Right: The saturation curve of fluorescent intensity changes at 430 nm. $\lambda_{ex} = 380$ nm, $d_{em} = d_{ex} = 5$ nm.



Figure S7. Left: Fluorescent titration of complex **ZnL** (5 μ M) upon the addition of CDP (0-50 μ M) in pH = 7.2, 10 mM HEPES buffer solution at 25°C; Right: The saturation curve of fluorescent intensity changes at 430 nm. $\lambda_{ex} = 380$ nm, $d_{em} = d_{ex} = 5$ nm.



Figure S8. Fluorescent titration of complex **ZnL** (5 μ M) upon the addition of CMP (0-150 μ M) in HEPES buffer (10 mM, pH = 7.2) solution at 25°C. λ_{ex} = 380 nm, d_{em} = d_{ex} = 5 nm.



Figure S9. Left: Fluorescent titration of complex **ZnL** (5 μ M) upon the addition of GTP (0-50 μ M) in HEPES buffer (10 mM, pH = 7.2) solution at 25°C; Right: The saturation curve of fluorescent intensity changes at 430 nm. $\lambda_{ex} = 380$ nm, $d_{em} = d_{ex} = 5$ nm.



Figure S10. Left: Fluorescent titration of complex **ZnL** (5 μ M) upon the addition of GDP (0-50 μ M) in HEPES buffer (10 mM, pH = 7.2) solution at 25°C; Right: The saturation curve of fluorescent intensity changes at 430 nm. $\lambda_{ex} = 380$ nm, $d_{em} = d_{ex} = 5$ nm.



Figure S11. Fluorescent titration of complex **ZnL** (5 μ M) upon the addition of GMP (0-250 μ M) in HEPES buffer (10 mM, pH = 7.2) solution at 25°C. λ_{ex} = 380 nm, d_{em} = d_{ex} = 5 nm.



Figure S12. Left: Fluorescent titration of complex **ZnL** (5 μ M) upon the addition of UTP (0-100 μ M) in HEPES buffer (10 mM, pH = 7.2) solution at 25°C; Right: The saturation curve of fluorescent intensity changes at 430 nm. $\lambda_{ex} = 380$ nm, $d_{em} = d_{ex} = 5$ nm.



Figure S13. Left: Fluorescent titration of complex **ZnL** (5 μ M) upon the addition of UDP (0-150 μ M) in HEPES buffer (10 mM, pH = 7.2) solution at 25°C; Right: The saturation curve of fluorescent intensity changes at 430 nm. $\lambda_{ex} = 380$ nm, $d_{em} = d_{ex} = 5$ nm.



Figure S14. Fluorescent titration of complex **ZnL** (5 μ M) upon the addition of UMP (0-250 μ M) in HEPES buffer (10 mM, pH = 7.2) solution at 25°C. λ_{ex} = 380 nm, d_{em} = d_{ex} = 5 nm.



Figure S15. Fluorescent titration of complex **ZnL** (5 μ M) upon the addition of PPi (0-25 μ M) in HEPES buffer (10 mM, pH = 7.2) solution at 25°C. $\lambda_{ex} = 380$ nm, $d_{em} = d_{ex} = 5$ nm. This data suggests that PPi induced an anthracene-excimer ($\lambda_{em} \sim 480$ nm) formation of the **ZnL** solution, which is similar to the result of our recent report (see: Huang, F.; Feng, G. *RSC Adv.* **2014**, *4*, 484–487.). However, the fluorescence enhancements induced by PPi are far less than that of ADP.





Figure S16. Fluorescent titration of complex **ZnL** (5 μ M) upon the addition of citrate, PO₄³⁻, F⁻ and Cl⁻ (0-500 μ M) as representative anionic analytes in HEPES buffer (10 mM, pH = 7.2) solution at 25°C. $\lambda_{ex} = 380$ nm, $d_{em} = d_{ex} = 5$ nm.



Figure S17. Job's plot examined between (a) **ZnL** and ATP and (b) **ZnL** and ADP in HEPES buffer (10 mM, pH = 7.2) solution at 25°C. [ATP or ADP] + [**ZnL**] = 5 μ M, λ_{ex} = 380 nm, $d_{ex} = d_{em} = 5$ nm.



Figure S18. High resolution mass spectrum of a mixture of **ZnL** and ADP. The peak at m/z = 918.1172 corresponds to formula $C_{60}H_{76}N_{20}Na_4O_{26}P_4Zn_2^{2+}$ (calculated mass $1836.2353 = 2 \times 918.11765$), which can be assigned to **ZnL**⁴⁺+2ADP³⁺+4Na⁺⁺+6H₂O as shown in the mass spectrum.



Figure S19. High resolution mass spectrum of a mixture of **ZnL** and ATP. The peak at m/z = 1821.1535 corresponds to formula $C_{60}H_{68}N_{20}NaO_{26}P_6Zn_2^+$ (**ZnL**+2ATP+Na⁺), which was calculated as 1821.15145. The peak at m/z = 1887.09481 corresponds to formula $C_{60}H_{65}N_{20}Na_4O_{26}P_6Zn_2^+$ (**ZnL**+2ATP-3H⁺+4Na⁺), which was calculated as 1887.09728.



Figure S20. (a) ¹H NMR spectra change and (b) ³¹P NMR spectra change of **ZnL** upon addition of ADP in DMSO- d_6 at 25 °C. [**ZnL**] = 10 mM, [ADP] = 20 mM. Still, we cannot get very good NMR spectra with high resolution due to the reason of low solubility, but it is clear that **ZnL** binds ADP and a new complex was formed.

4. Quantum Mechanical Calculations

Quantum mechanical calculations based on the density functional theory (DFT) were investigated to identify the configuration of **ZnL-2ADP** and **ZnL-2ATP**. The calculations were performed by Gaussian09 suite. Both of the two structures were optimized at the DFT level with the B3LYP functional. The 6-31G(d) basis set was used for C, H, N, O, and P atoms, while the LANL2DZ basis set was used for Zn atom. The charge values were set to 0 for **ZnL-2ADP** and -2 for **ZnL-2ATP**. The optimized configurations of **ZnL-2ADP** and **ZnL-2ATP** are displayed below. For

ZnL-2ADP, the two Zn²⁺ ions link **L** and ADP together via five coordination sites, respectively. The Zn²⁺ forms coordination bonds with three nitrogen atoms (bond lengths are 2.201Å, 2.171Å, and 2.292Å) of **L** and two oxygen atoms (bond lengths are 2.028Å and 1.999Å) of ADP. Different from the configuration of **ZnL-2ADP**, the Zn²⁺ ion in **ZnL-2ATP** join L and ATP together via six coordination sites, including three bonds with nitrogen atoms (bond lengths are 2.312Å, 2.304Å, and 2.332Å) of **L** and other three with oxygen atoms (bond lengths are 2.065Å, 2.042Å, and 2.082Å) of ATP. The results suggest that both **ZnL-2ADP** and **ZnL-2ATP** configurations are stable by forming coordination bonds via Zn²⁺ ions. Furthermore, the adenine structure in ADP have π - π stacking interaction with anthracene of **L** in **ZnL-2ATP**.



Figure S21. DFT calculated bond lengths between Zn^{2+} and the coordinated three nitrogen atoms for ADP and ATP, respectively.



Figure S22. DFT calculated interactions between **ZnL** and ADP. Top: View with hydrogen atoms. Bottom: Hydrogen atoms are omitted for clarity.



Figure S23. DFT calculated interactions between **ZnL** and ATP. Top: View with hydrogen atoms. Bottom: Hydrogen atoms are omitted for clarity.