

1-Phenylethynylpyrene (PEPy) as a novel blue-emitting dye for qPCR

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Supporting Information

¹H NMR of 3

¹³C NMR of 3

¹H NMR of 5

¹³C NMR of 5

Table S1. Brightness of AMCA- or PEPy-modified oligonucleotides and their duplexes with fully matched and mismatched complementary DNA.

Table S2. Labeled oligonucleotides and TaqMan probes, used in the study. MS data

Table S3. Sequences of primers used in the study

Oligonucleotide 6A - HPLC traces and MS spectrum.

Oligonucleotide 6T - HPLC traces and MS spectrum

Oligonucleotide 6C - HPLC traces and MS spectrum

Oligonucleotide 6G - HPLC traces and MS spectrum

Oligonucleotide 7A - HPLC traces and MS spectrum

Oligonucleotide 7T - HPLC traces and MS spectrum

Oligonucleotide 7C - HPLC traces and MS spectrum

Oligonucleotide 7G - HPLC traces and MS spectrum

Oligonucleotide 9.1 - HPLC traces and MS spectrum

Oligonucleotide 9.2 - HPLC traces and MS spectrum

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Oligonucleotide PEPy-P4z - HPLC traces and MS spectrum

Oligonucleotide PEPy-P8z - HPLC traces and MS spectrum

Oligonucleotide PEPy-G1z - HPLC traces and MS spectrum

Oligonucleotide PEPy-G2z - HPLC traces and MS spectrum

Oligonucleotide PEPy-G3z - HPLC traces and MS spectrum

Oligonucleotide PEPy-G4z - HPLC traces and MS spectrum

Oligonucleotide AMCA-P4z - HPLC traces and MS spectrum

Oligonucleotide AMCA-P8z - HPLC traces and MS spectrum

Oligonucleotide AMCA-G1z - HPLC traces and MS spectrum

Oligonucleotide AMCA-G2z - HPLC traces and MS spectrum

Oligonucleotide AMCA-G3z - HPLC traces and MS spectrum

Oligonucleotide AMCA-G4z - HPLC traces and MS spectrum

Secondary structure of P4z probes optimized by Mfold. Raw and normalized data of qPCR

Secondary structure of P8z probes optimized by Mfold. Raw and normalized data of qPCR

Secondary structure of G1z probes optimized by Mfold. Raw and normalized data of qPCR

Secondary structure of G2z probes optimized by Mfold. Raw and normalized data of qPCR

Secondary structure of G3z probes optimized by Mfold. Raw and normalized data of qPCR

Secondary structure of G4z probes optimized by Mfold. Raw and normalized data of qPCR

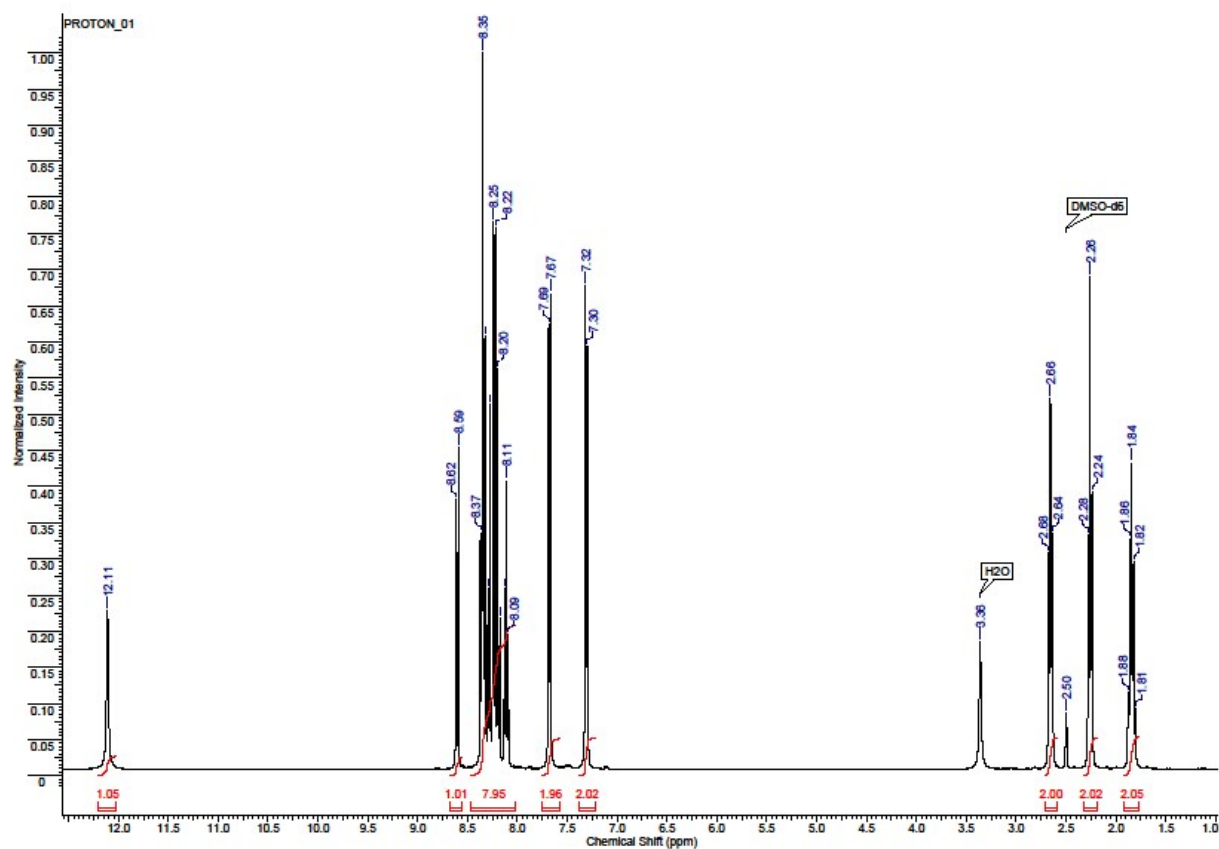
Melting curves for PEPy-labeled P8z, P4z, G1z, G2z, G3z, G4z probes.

qPCR data for cross-talk studies between “blue” and “green” channels.

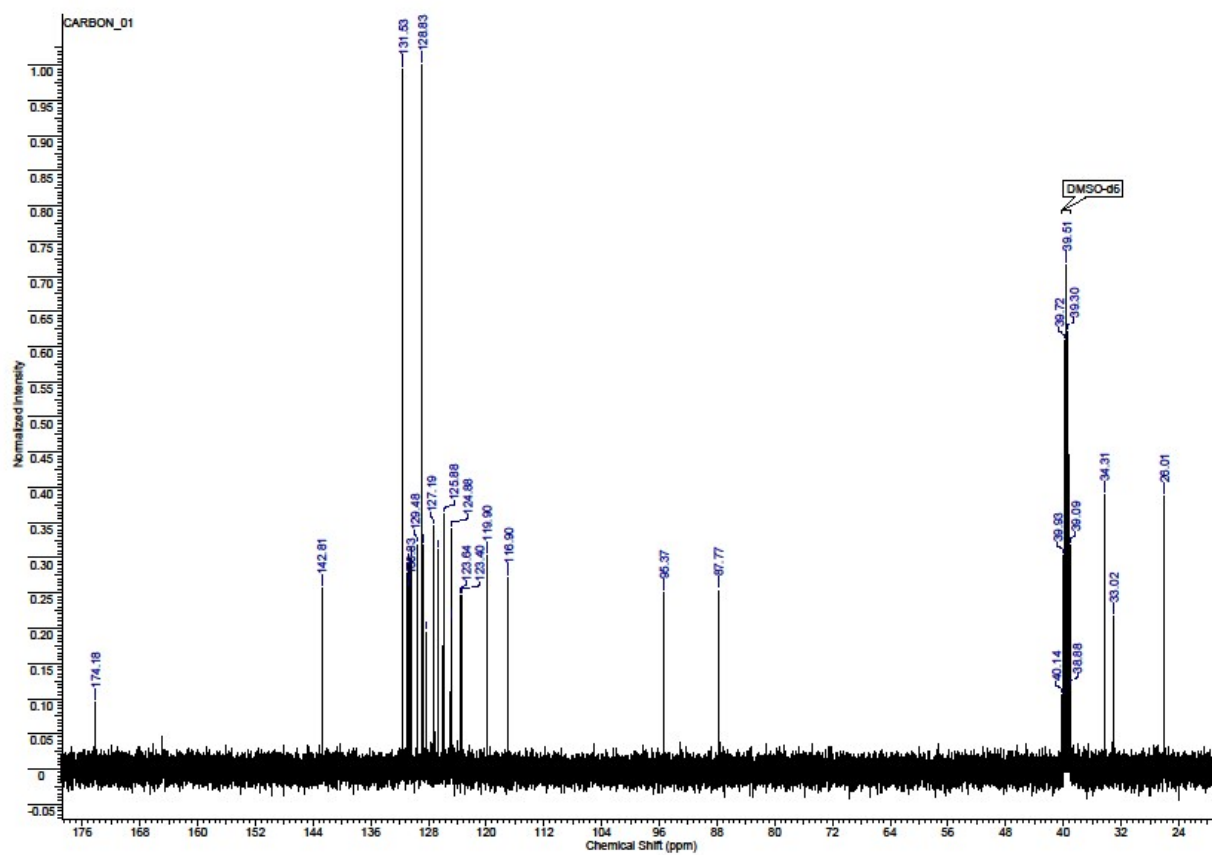
Table S4. qPCR data for AMCA, Alexa 350 and PEPy dual labeled probes.

Table S5. Normalized fluorescence data for AMCA, Alexa 350 and PEPy dual labeled probes in end-point qPCR.

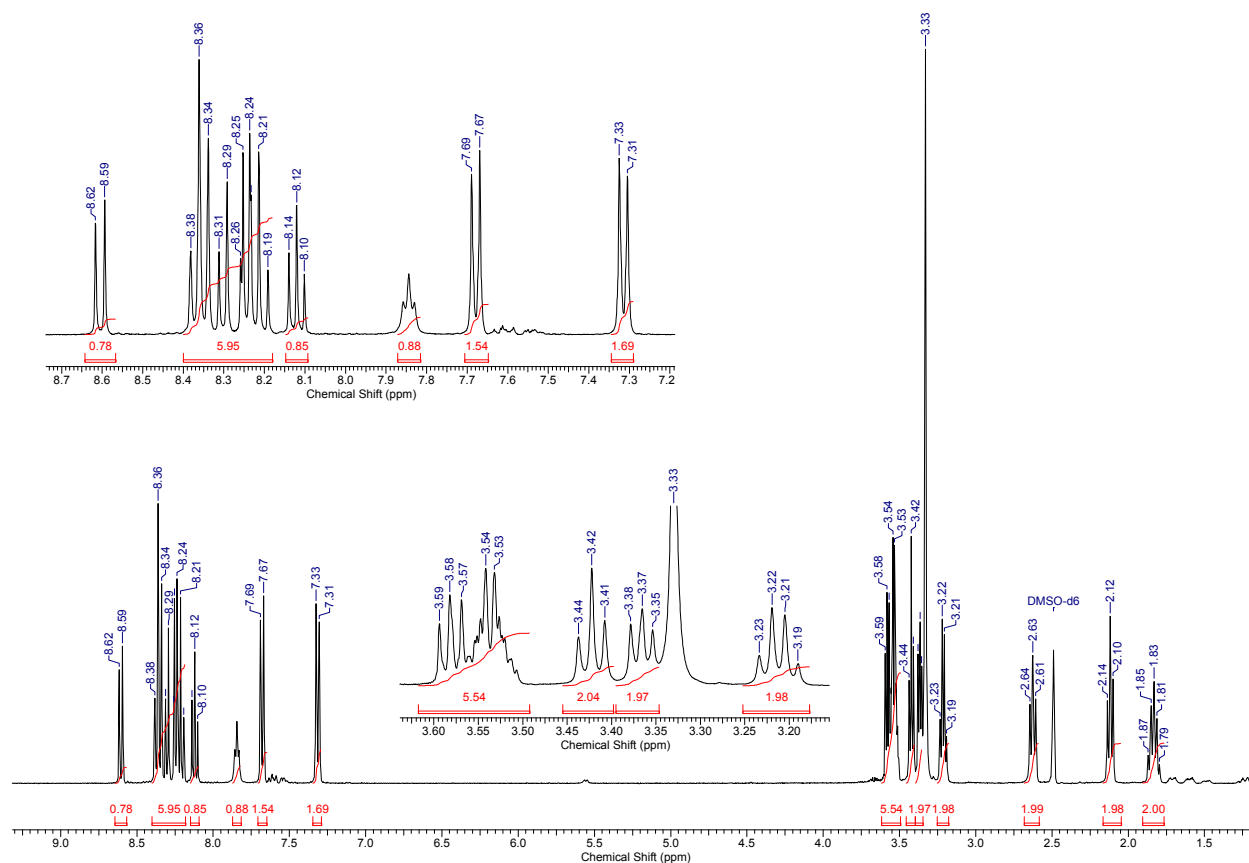
^1H NMR of 3



^{13}C NMR of 3



^1H NMR of **5**



^{13}C NMR of **5**

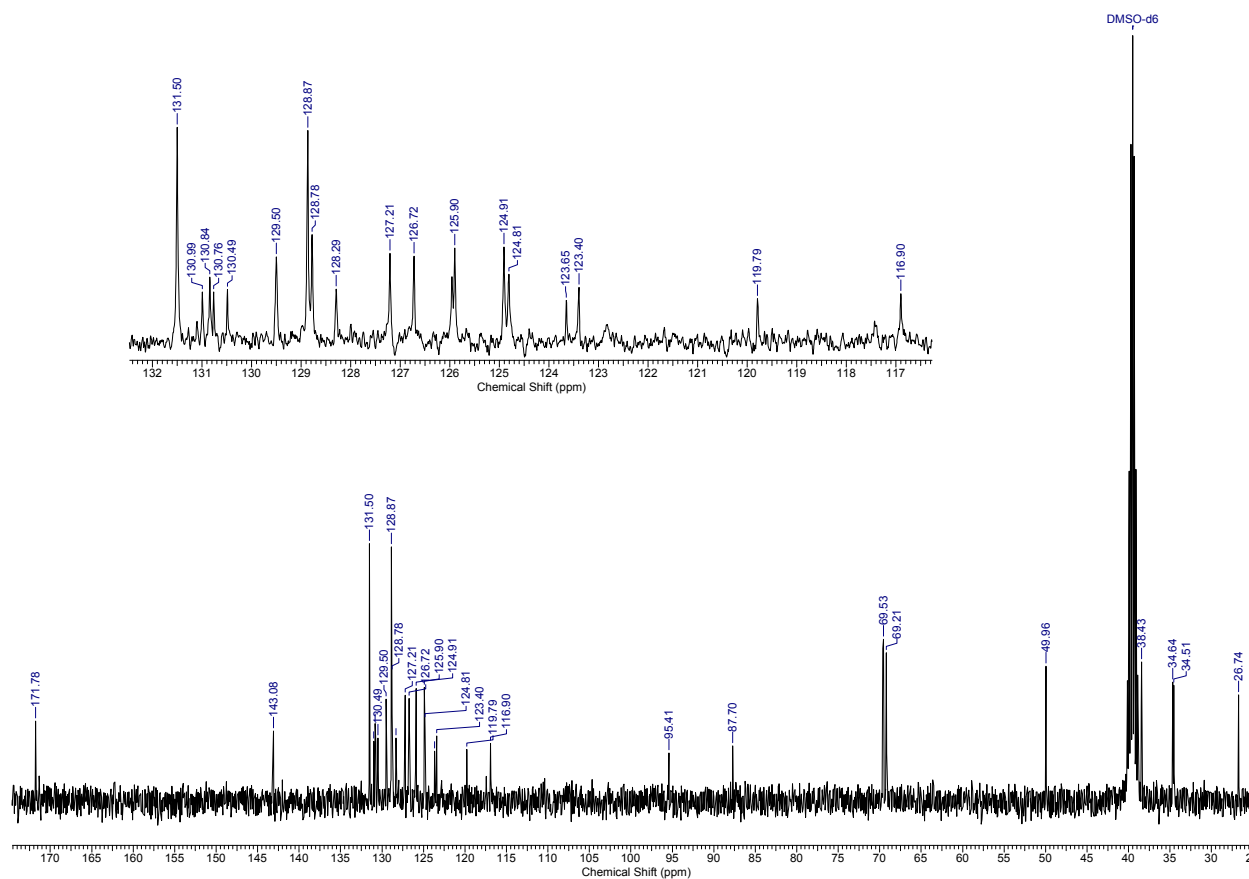


Table S1. Quantum yields (Φ_f) of fluorescence of AMCA- or PEPy-modified oligonucleotides and their duplexes with fully matched and mismatched complementary DNA.^a

Oligonucleotides 6 and 7 (5'→3') and their duplexes with targets 8	X = AMCA			X = PEPy		
	ON	Φ_f , %	Brightness, $L \cdot M^{-1} cm^{-1}$	ON	Φ_f , %	Brightness, $L \cdot M^{-1} cm^{-1}$
X-A ACCCAACTGAAGCAGCA	6A	58±4	11000±800	7A	63±4	28000±1800
X-T ACCCAACTGAAGCAGCA	6T	52±3	9900±700	7T	47±3	21000±1700
X-C ACCCAACTGAAGCAGCA	6C	55±3	10500±700	7C	51±3	23000±1600
X-G ACCCAACTGAAGCAGCA	6G	61±4	11600±800	7G	52±3	23000±1600
X-A ACCCAACTGAAGCAGCA ACAAACATA A TGGGTTGACTTCGTCGT	6A/8A	45±3	8600±600	7A/8A	53±3	23700±1600
X-T ACCCAACTGAAGCAGCA ACAAACATA A TGGGTTGACTTCGTCGT	6T/8A	48±3	9100±600	7T/8A	51±3	22800±1600
X-C ACCCAACTGAAGCAGCA ACAAACATA A TGGGTTGACTTCGTCGT	6C/8A	46±3	8700±600	7C/8A	40±3	17900±1200
X-G ACCCAACTGAAGCAGCA ACAAACATA A TGGGTTGACTTCGTCGT	6G/8A	46±3	8700±600	7G/8A	46±3	20600±1400
X-A ACCCAACTGAAGCAGCA ACAAACATA T TGGGTTGACTTCGTCGT	6A/8T	49±3	9300±600	7A/8T	59±3	26400±1800
X-T ACCCAACTGAAGCAGCA ACAAACATA T TGGGTTGACTTCGTCGT	6T/8T	52±3	9900±700	7T/8T	48±3	21500±1500
X-C ACCCAACTGAAGCAGCA ACAAACATA T TGGGTTGACTTCGTCGT	6C/8T	51±3	9700±700	7C/8T	40±3	17900±1200
X-G ACCCAACTGAAGCAGCA ACAAACATA T TGGGTTGACTTCGTCGT	6G/8T	50±3	9500±600	7G/8T	37±2	16500±1100
X-A ACCCAACTGAAGCAGCA ACAAACATA C TGGGTTGACTTCGTCGT	6A/8C	49±3	9300±600	7A/8C	53±3	23700±1600
X-T ACCCAACTGAAGCAGCA ACAAACATA C TGGGTTGACTTCGTCGT	6T/8C	50±3	9500±600	7T/8C	39±3	17400±1200
X-C ACCCAACTGAAGCAGCA ACAAACATA C TGGGTTGACTTCGTCGT	6C/8C	45±3	8600±600	7C/8C	50±3	22400±1500
X-G ACCCAACTGAAGCAGCA ACAAACATA C TGGGTTGACTTCGTCGT	6G/8C	43±3	8200±600	7G/8C	28±2	12500±900
X-A ACCCAACTGAAGCAGCA ACAAACATA G TGGGTTGACTTCGTCGT	6A/8G	48±3	9100±600	7A/8G	51±3	22800±1600
X-T ACCCAACTGAAGCAGCA ACAAACATA G TGGGTTGACTTCGTCGT	6T/8G	53±3	10100±700	7T/8G	36±2	16100±1100
X-C ACCCAACTGAAGCAGCA ACAAACATA G TGGGTTGACTTCGTCGT	6C/8G	51±3	9700±700	7C/8G	22±2	9800±700
X-G ACCCAACTGAAGCAGCA ACAAACATA A TGGGTTGACTTCGTCGT	6G/8G	49±3	9300±650	7G/8G	43±3	19200±1300

^a Mismatched nucleosides are highlighted in red. Measurements were taken for 3.0×10^{-7} M solutions in hybridization buffer (1x PBS, pH 7.0) at 20°C. Excitation wavelength 365 nm, fluorescence maxima are 442 nm (AMCA), 399 nm, 420 nm (PEPy); for the technique of Φ_f calculation see Experimental part.

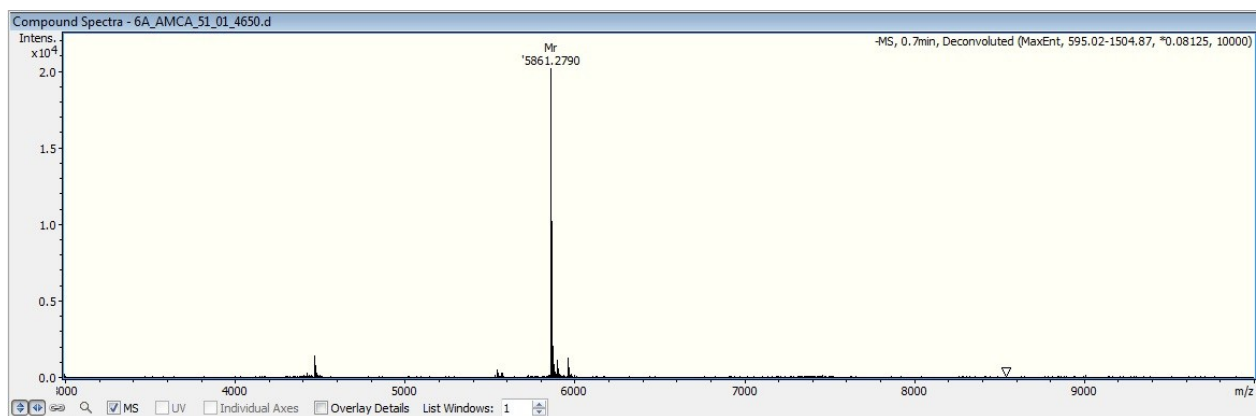
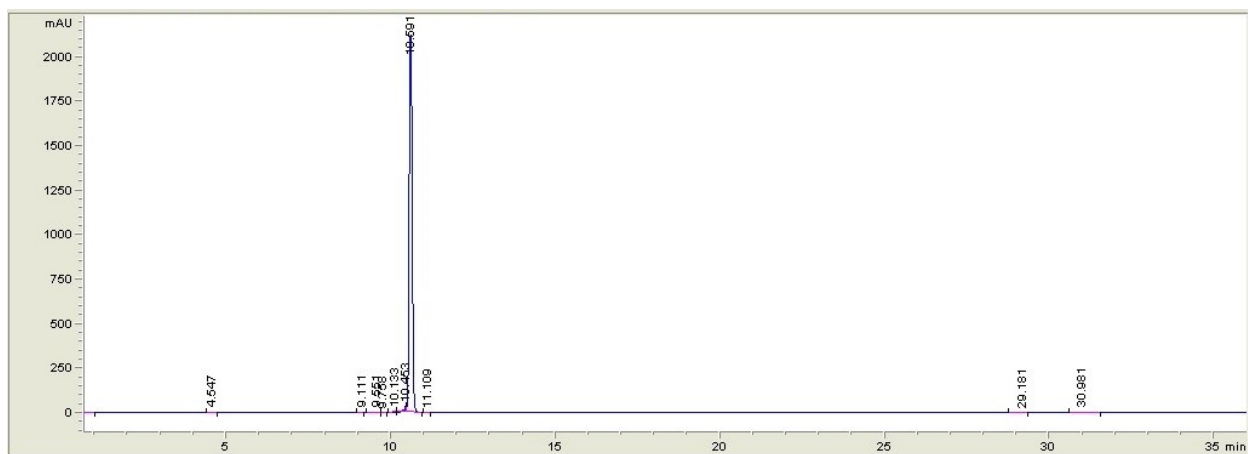
Table S2. Labeled oligonucleotides and Taqman probes, used in the study. MS data

N	Name	Oligonucleotide, 5'-3'	ESI-MS (calcd/ observed)
1	6A	AMCA-AACCCCAACTGAAGCAGCA	5867 / 5861
2	6T	AMCA-TACCCCAACTGAAGCAGCA	5858 / 5852
3	6C	AMCA-CACCCCAACTGAAGCAGCA	5843 / 5837
4	6G	AMCA-GACCCCAACTGAAGCAGCA	5883 / 5877
5	7A	PEPy-AACCCCAACTGAAGCAGCA	6286 / 6282
6	7T	PEPy-TACCCCAACTGAAGCAGCA	6277 / 6273
7	7C	PEPy-CACCCCAACTGAAGCAGCA	6262 / 6258
8	7G	PEPy-GACCCCAACTGAAGCAGCA	6302 / 6298
9	9.1	PEPy-AAACATATAATACGG	5408 / 5405
10	9.2	AMCA-AAACATATAATACGG	4990 / 4984
11	9.3	PEPy-TTTTTTTTTTAAACATATAATACGG	8450 / 8446
12	9.4	AMCA-TTTTTTTTTTAAACATATAATACGG	8032 / 8025
13	9D	CCGTATTATATGTTT-DABCYL	4978 / 4975
14	PEPy-P4z	PEPy-CAC TCT GAC TAC TAC CTT TAA ACA GAG CG-Dabcyl	10165 / 10159
15	PEPy-P8z	PEPy-GGA CTG CAG TCR TTG CTG TTG AAC- Dabcyl	8743 / 8738 8759 / 8754
16	PEPy-G1z	PEPy-CTG AGC TTT AGT YAA GGC AAA TAA TGC TCA G-Dabcyl	10902 / 10896 10917 / 10911
17	PEPy-G2z	PEPy-CAG CGT CTA GTG ATC CCG TTA TTG GC-Dabcyl	9312 / 9307
18	PEPy-G3z	PEPy-TAC CCA ACT GAA GCA GCA ACA GGG TA-Dabcyl	9341 / 9336
19	PEPy-G4z	PEPy-CTG AAC YTG TCG GCC ATC CTT TGG TT-Dabcyl	9263 / 9258 9278 / 9273
20	AMCA-P4z	AMCA-CAC TCT GAC TAC TAC CTT TAA ACA GAG CG-Dabcyl	9747 / 9738
21	AMCA-P8z	AMCA-GGA CTG CAG TCR TTG CTG TTG AAC-Dabcyl	8325 / 8317 8341 / 8333
22	AMCA-G1z	AMCA-CTG AGC TTT AGT YAA GGC AAA TAA TGC TCA G-Dabcyl	10484 / 10475 10499 / 10490
23	AMCA-G2z	AMCA-CAG CGT CTA GTG ATC CCG TTA TTG GC-Dabcyl	8894 / 8886
24	AMCA-G3z	AMCA-TAC CCA ACT GAA GCA GCA ACA GGG TA-Dabcyl	8923 / 8914
25	AMCA-G4z	AMCA-CTG AAC YTG TCG GCC ATC CTT TGG TT-Dabcyl	8845 / 8837 8860 / 8852

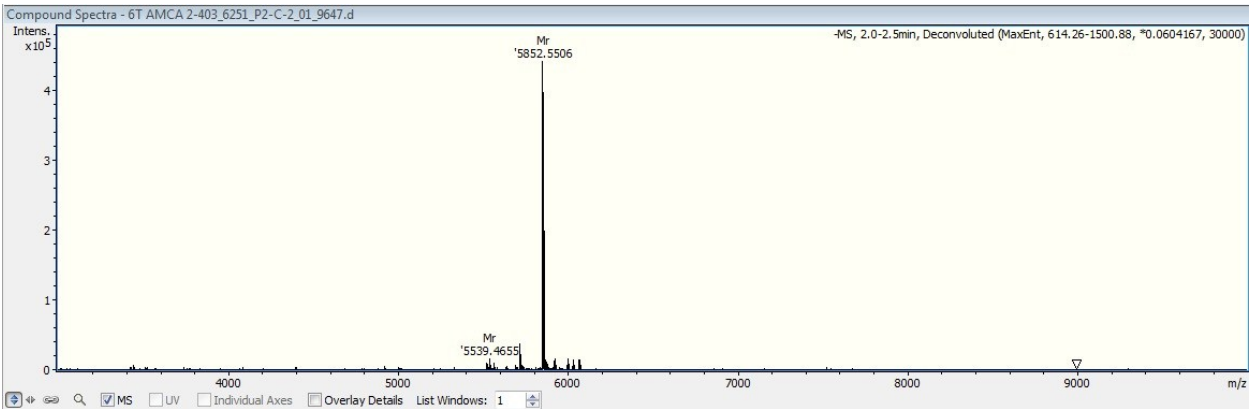
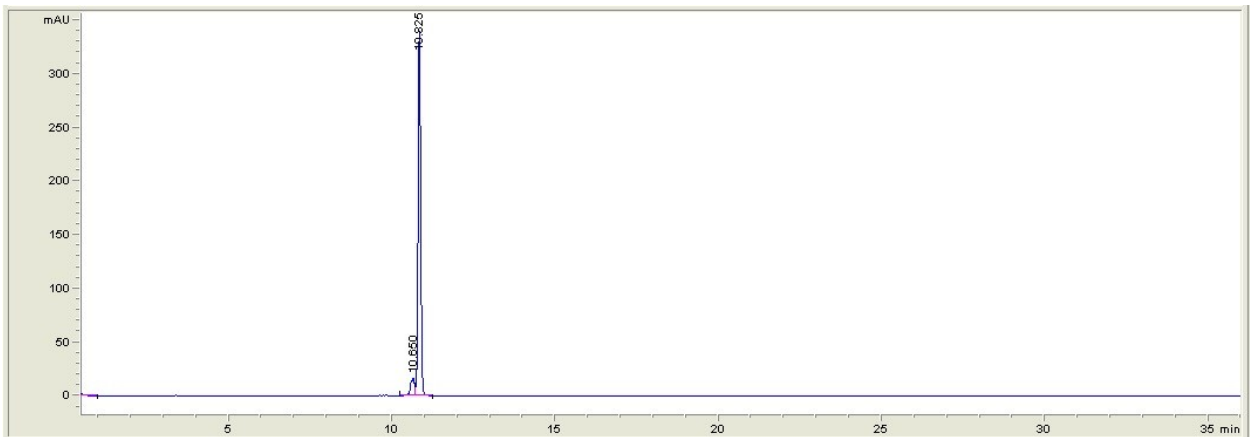
Table S3. Sequences of primers used in the study.

Primer		Target gene	Location (GenBank accession number)
VP4 for	TAAAATGGCTTCGCTCATTTATAGACA		
VP4 rev	GGTTATAAATGGTCCGAAATATCATATA		
VP7 for	GGCTTTAAAAGAGAGAATTTCCGTCTGG		
VP7 rev	TCAACTACAGTATAAAATACTTGCCACCA		
4P385	GAGAATAAGCAGTTTAAACGTAGA	P[4]	379-401 (M32559)
2T1	CTATTGTTAGAGGTTAGAGTC	P[4]	494-474 (M32559)
1G86	GATATCAATCATTTCTACTCAAC	G1	87-108 (M21843)
1G310	GATTTGAGTACTTGCTTCAGT	G1	327-307 (M21843)
9T3P	GTCCAGTTGCAGTGTAGC	G3	501-484 (AF161823)
3G276	CCTAACTTCGACTTTATGTT	G3	276-295 (AF161823)
9T2	GTT AGA AAT GAT TCT CCA CT	G2	281-262 (U73974)
2G170	CTC TGA TAT CAC CAT TTG TG	G2	173-192 (U73974)
9T4	GGG TCG ATG GAA AAT TCT	G4	423-440 (D86284)
4G319	CCA ACT CAA ATT AGT GAC AC	G4	707-726 (D86284)
8P214	CCT TAT CAR CCT ACT ACA TTT AC	P8	210-232 (M21014)
1-T1m	GTA TCT ACT GGA TTA ACG TGC	P8	336-356 (M21014)

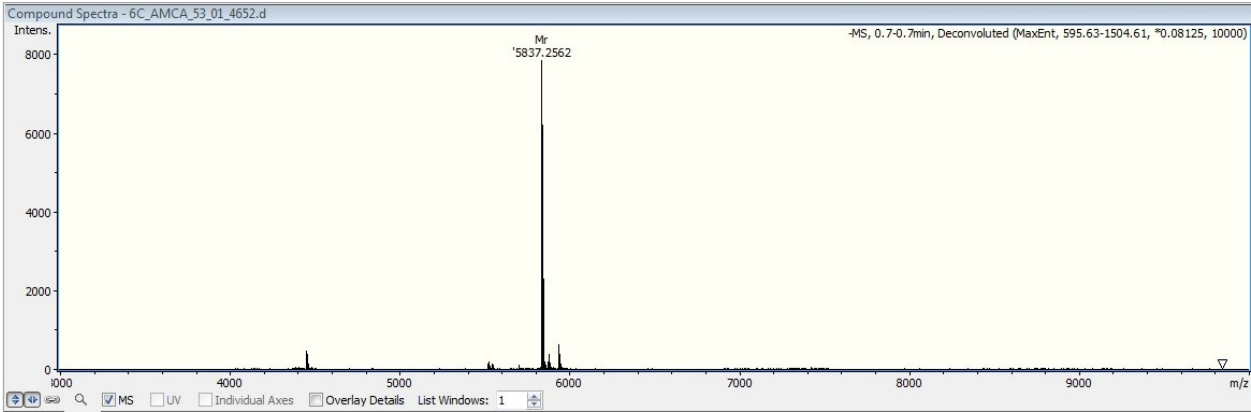
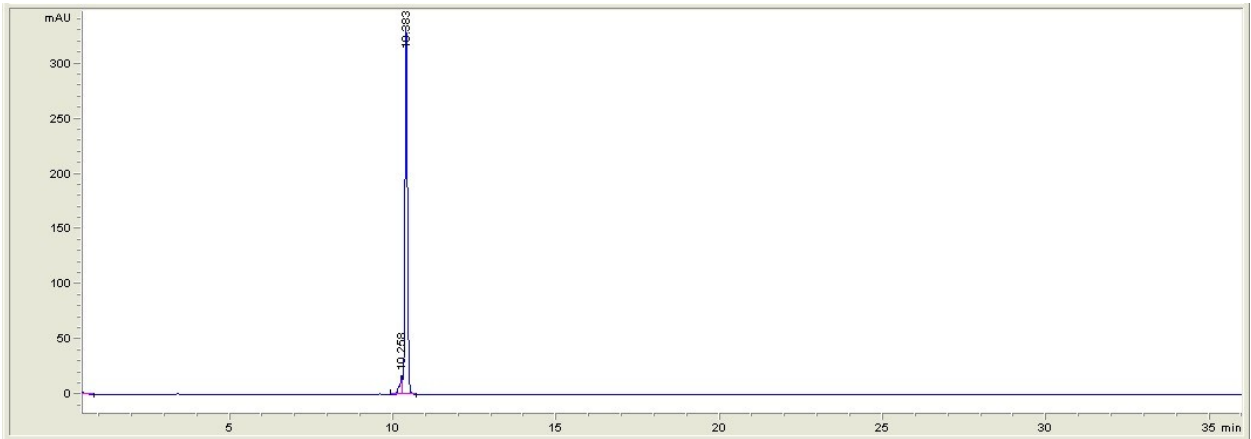
Oligonucleotide 6A - HPLC traces and MS spectrum.



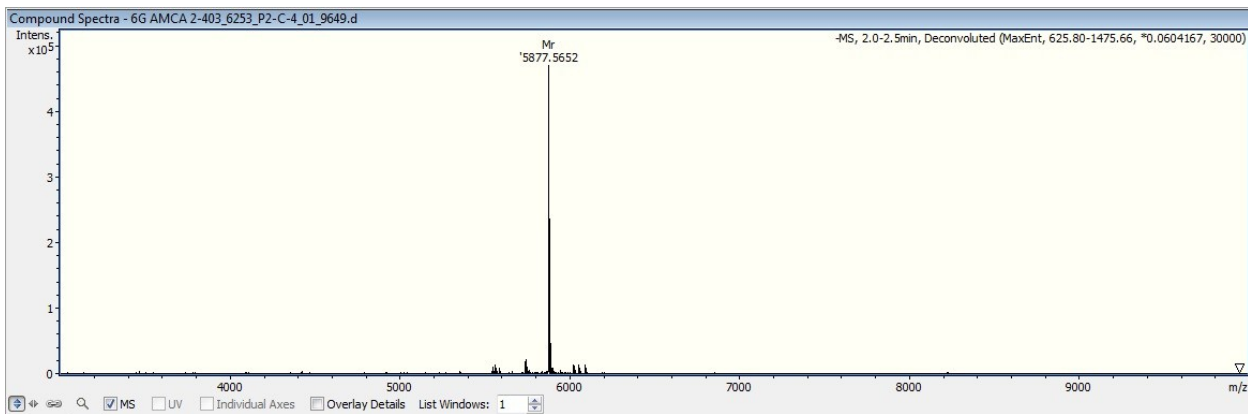
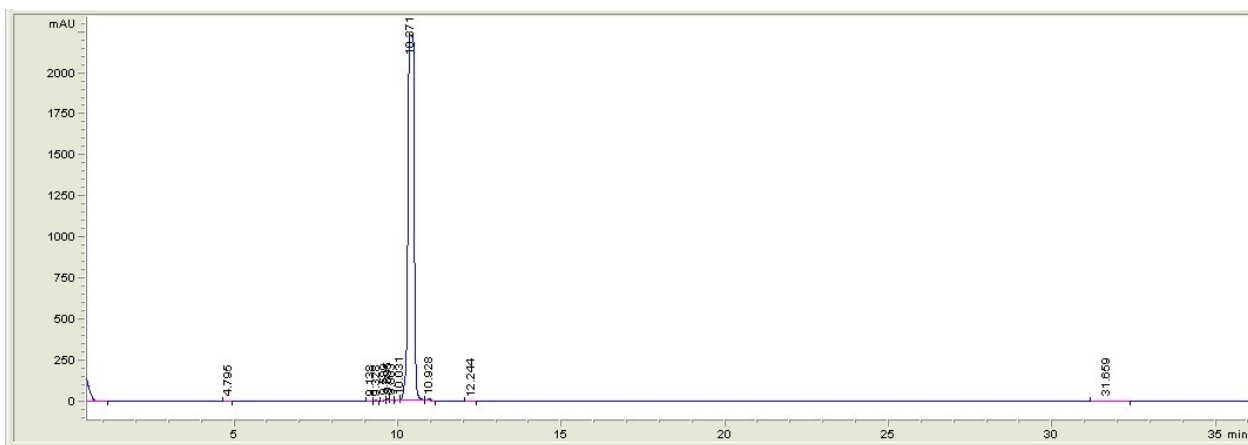
Oligonucleotide 6T - HPLC traces and MS spectrum



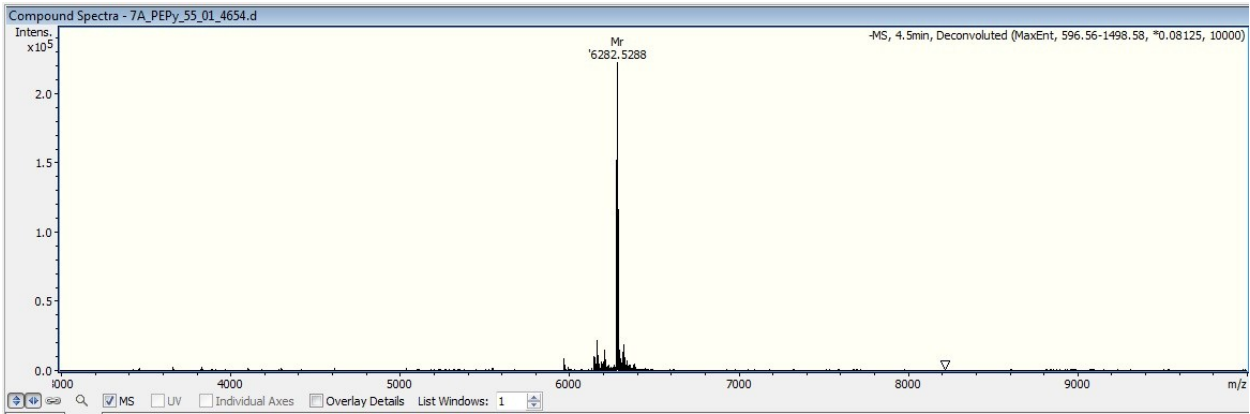
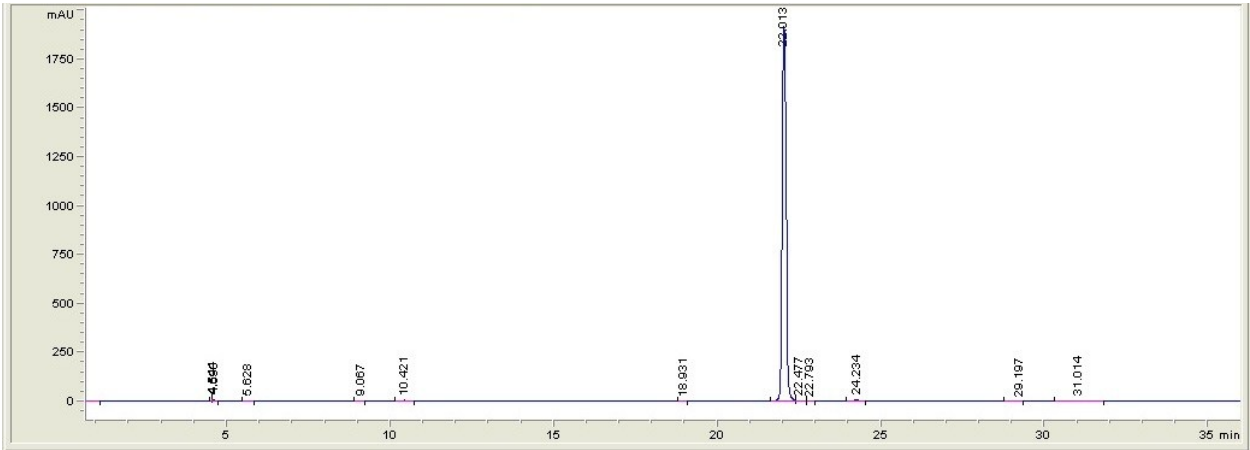
Oligonucleotide 6C - HPLC traces and MS spectrum



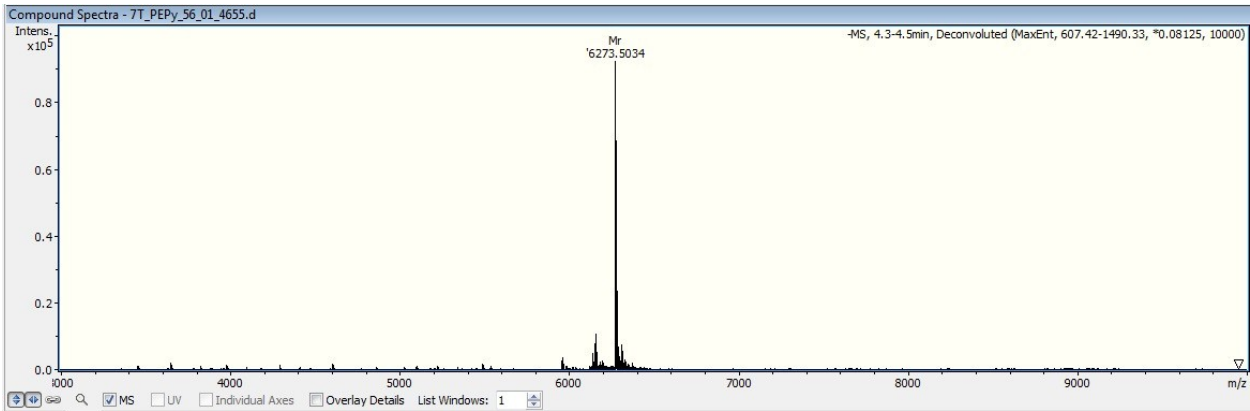
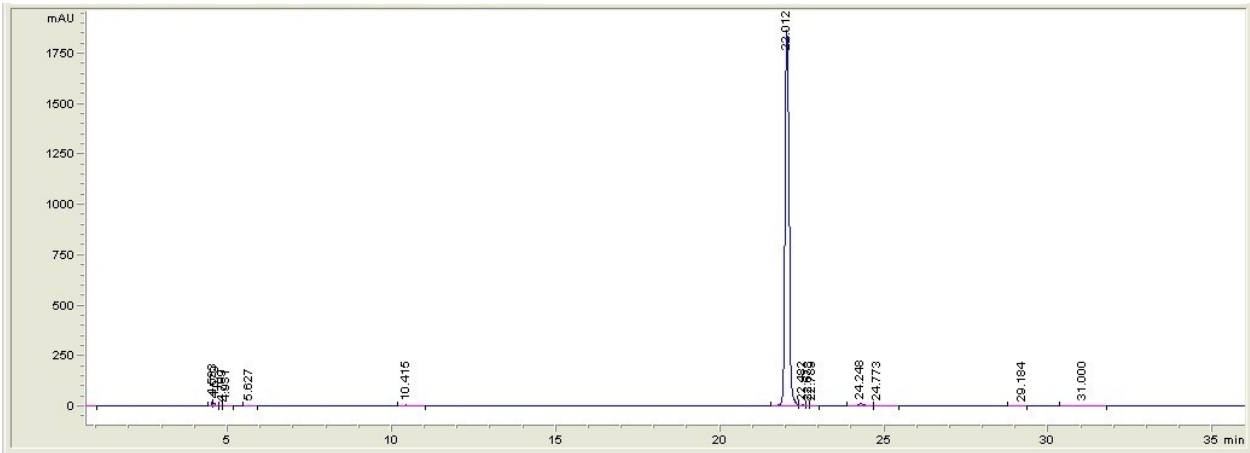
Oligonucleotide 6G - HPLC traces and MS spectrum



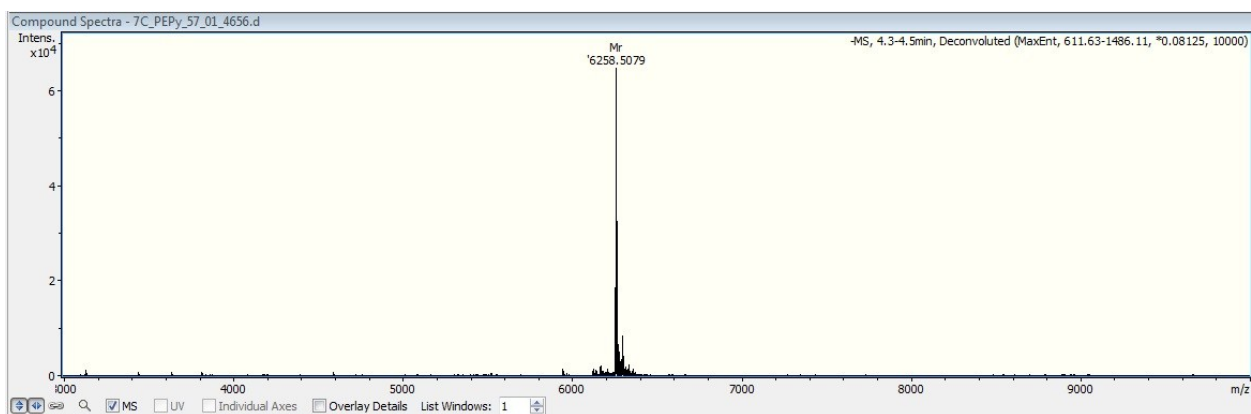
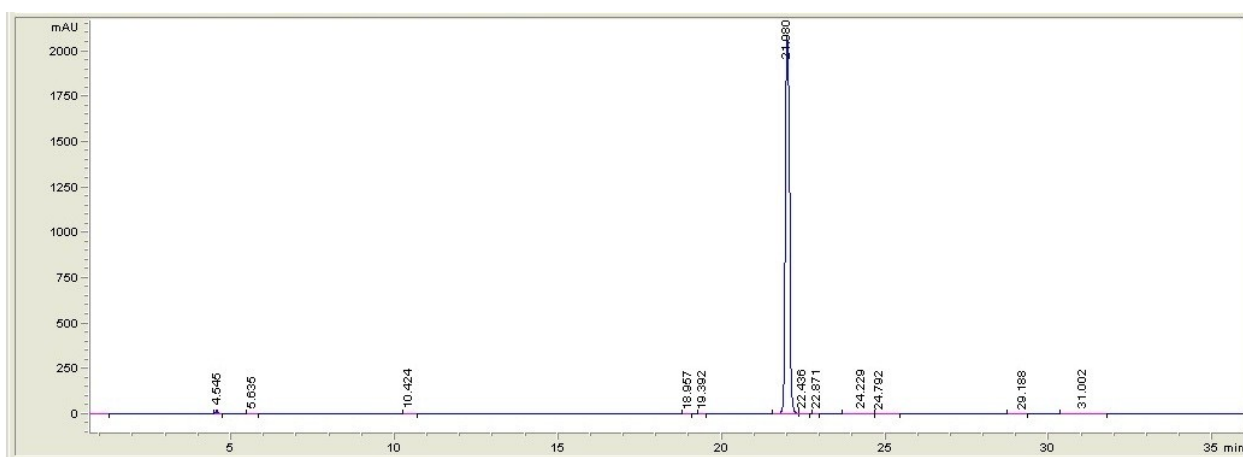
Oligonucleotide 7A - HPLC traces and MS spectrum



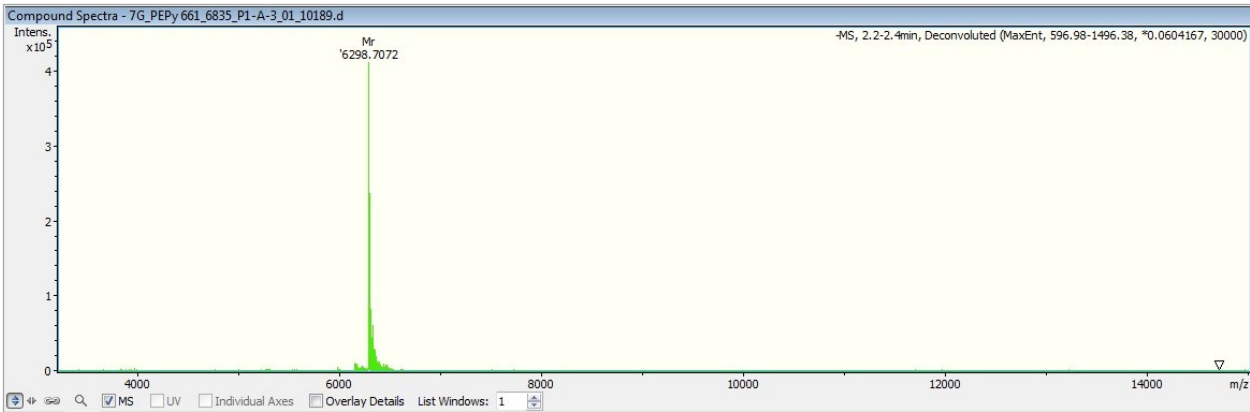
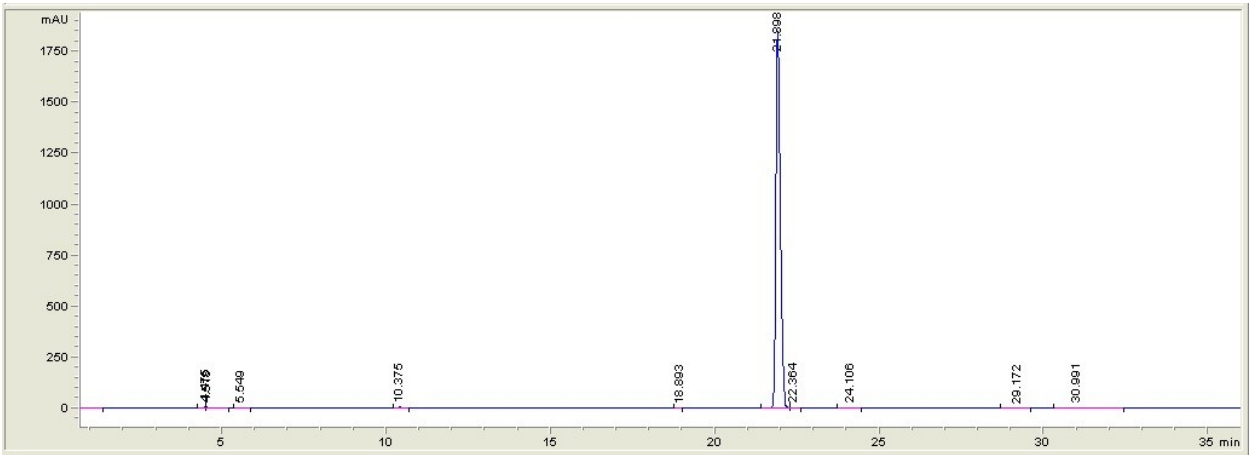
Oligonucleotide 7T - HPLC traces and MS spectrum



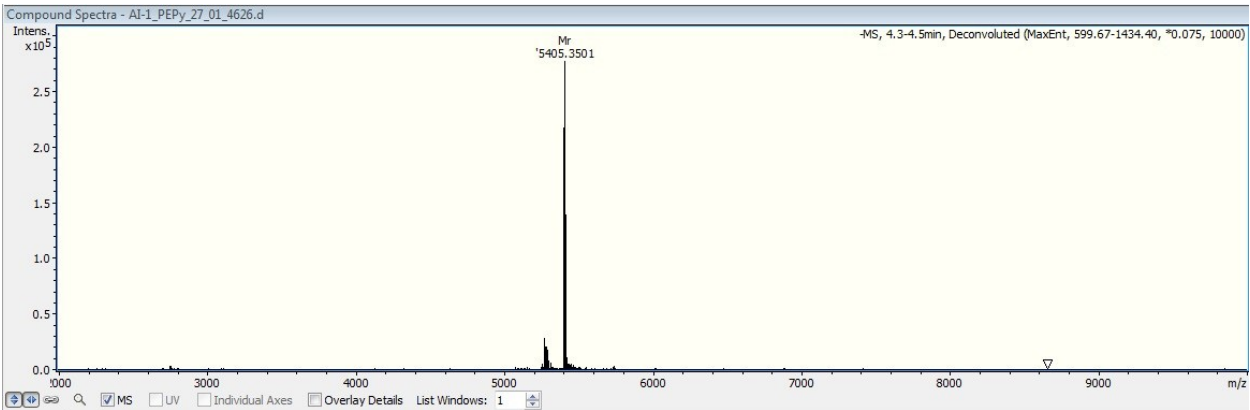
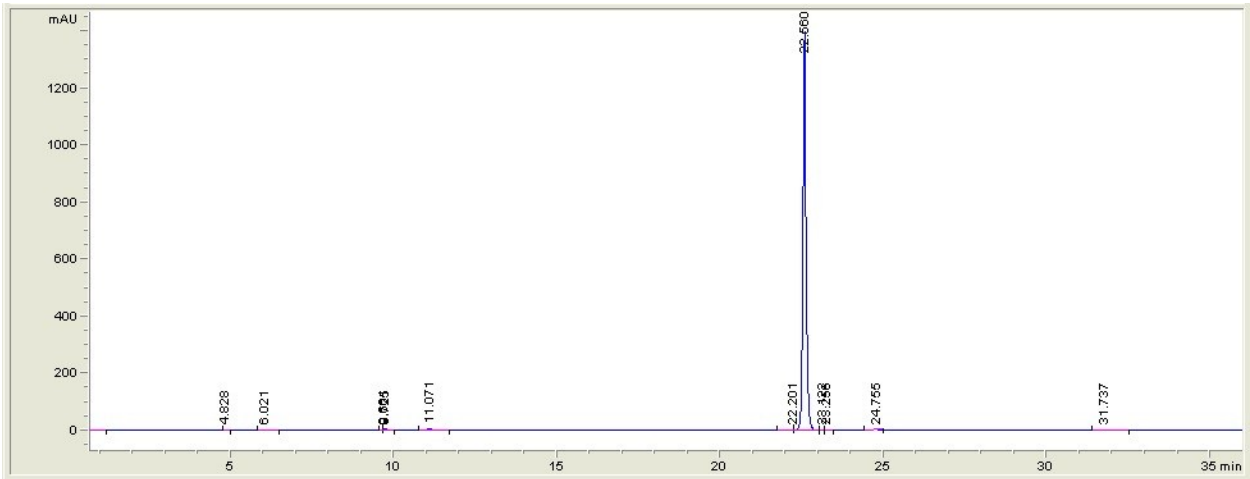
Oligonucleotide 7C - HPLC traces and MS spectrum



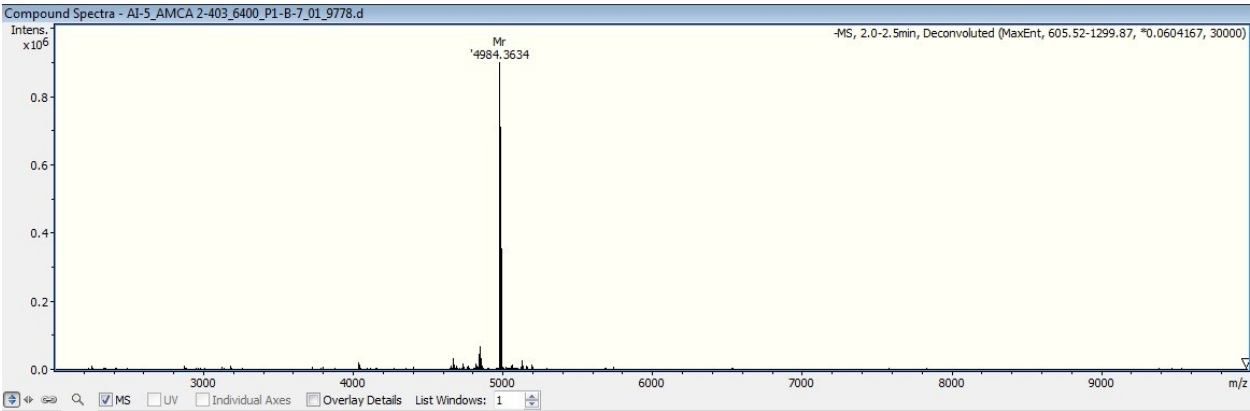
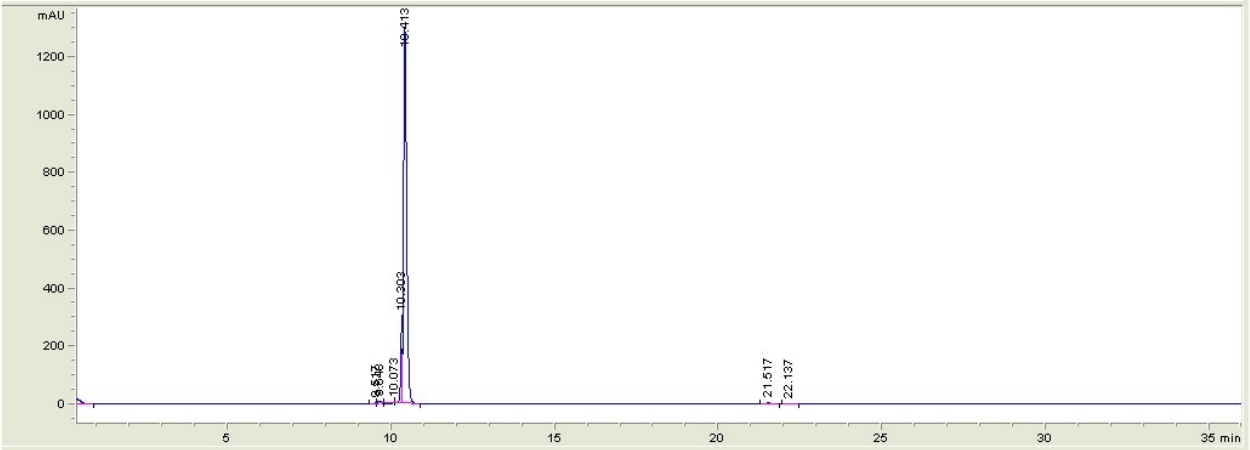
Oligonucleotide 7G - HPLC traces and MS spectrum



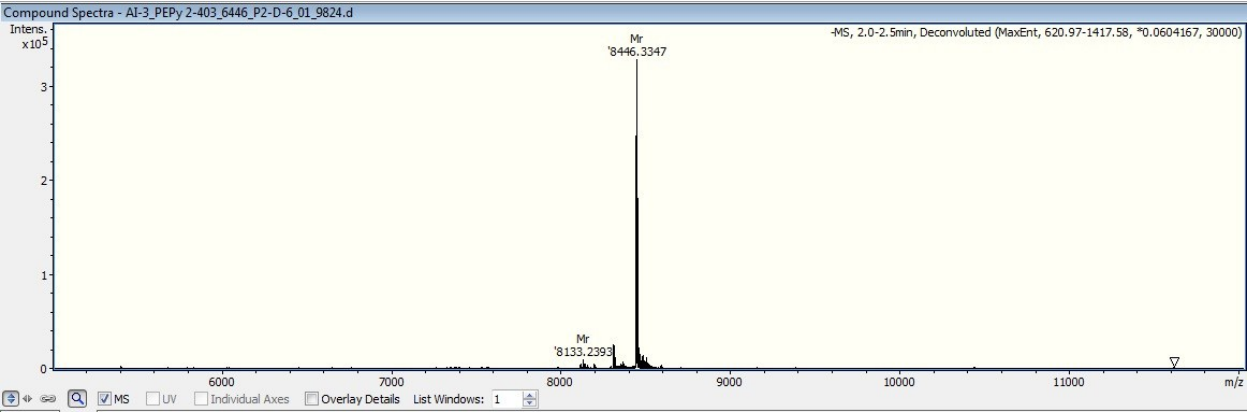
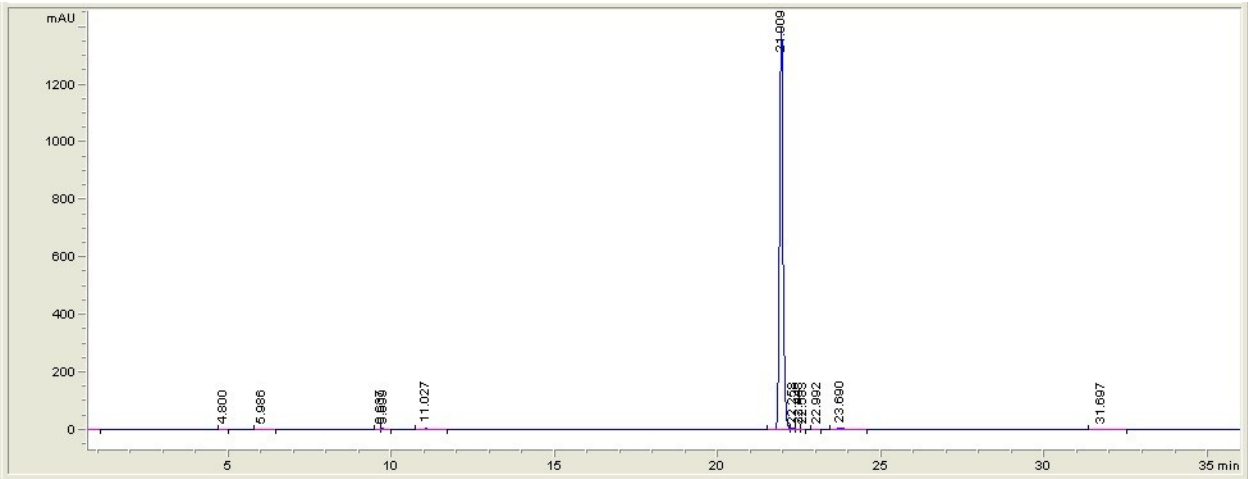
Oligonucleotide 9.1 - HPLC traces and MS spectrum



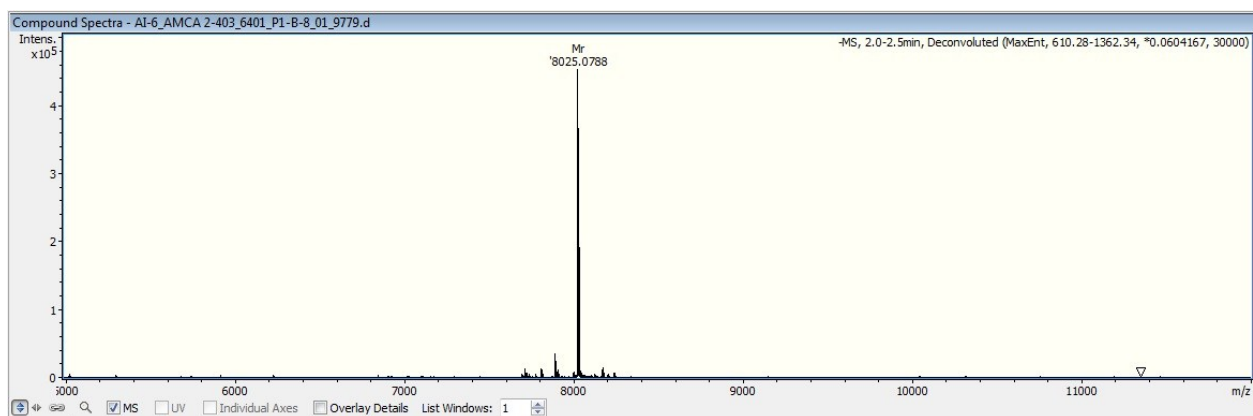
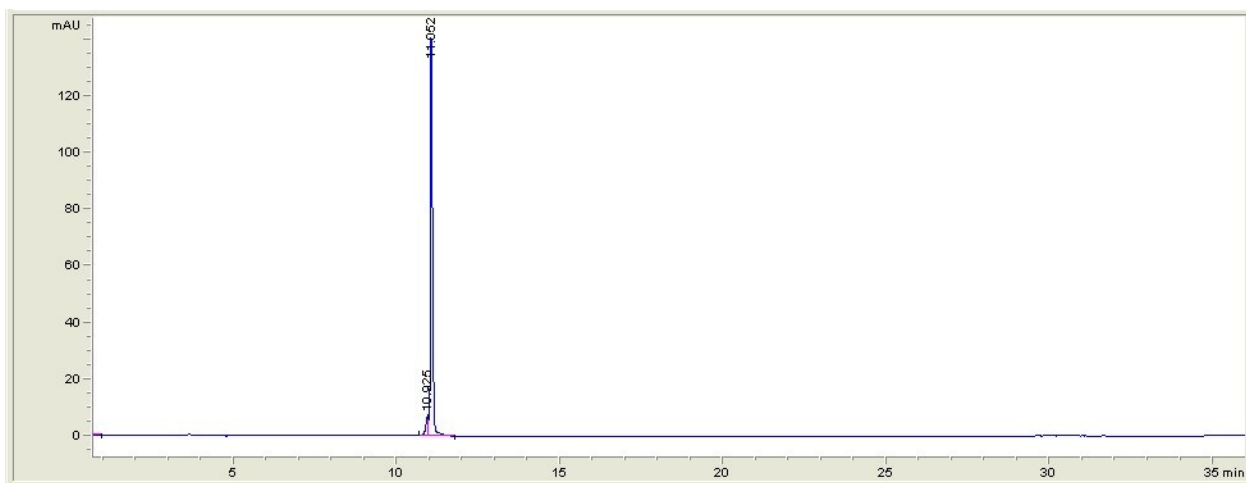
Oligonucleotide 9.2 - HPLC traces and MS spectrum



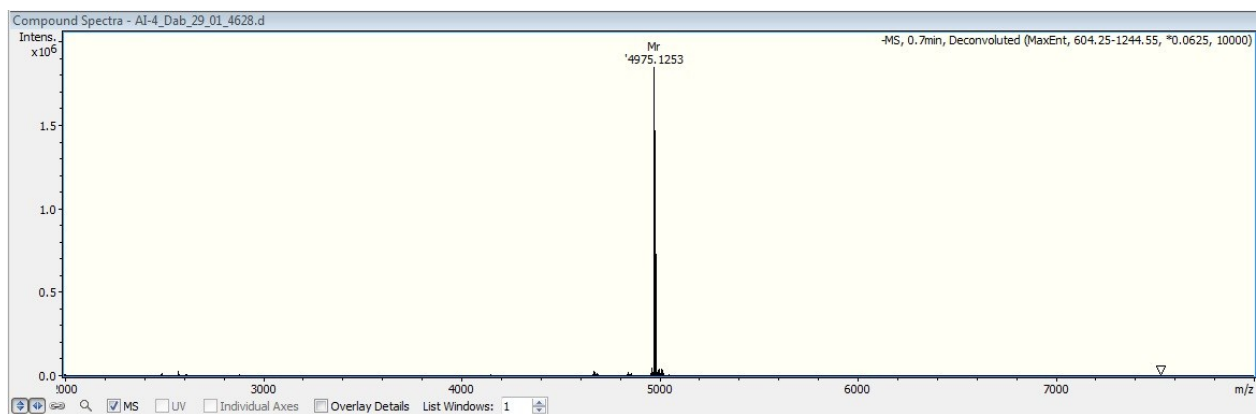
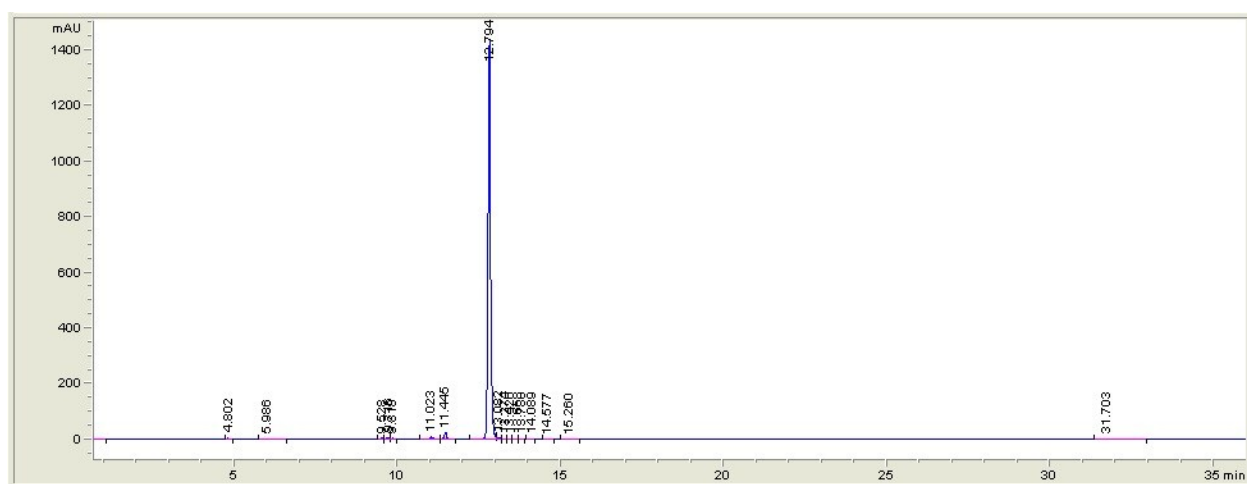
Oligonucleotide 9.3 - HPLC traces and MS spectrum



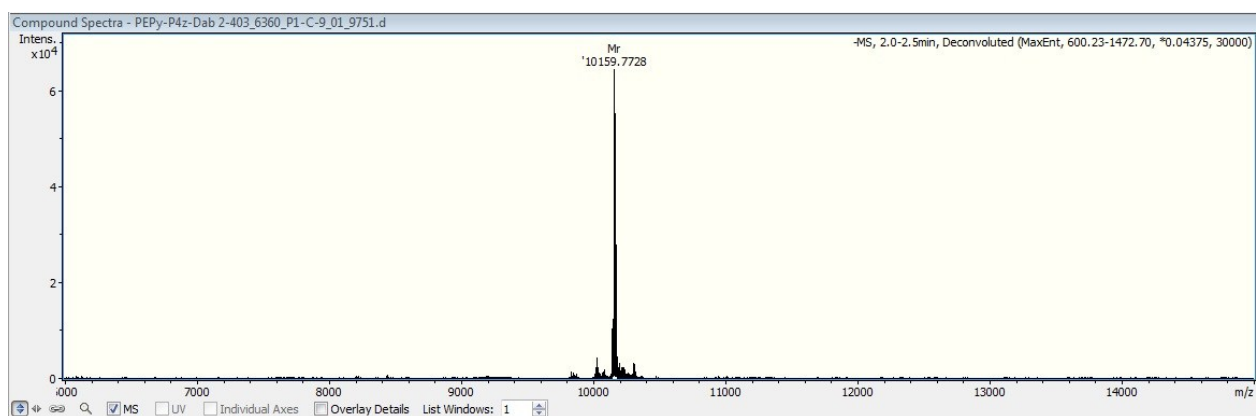
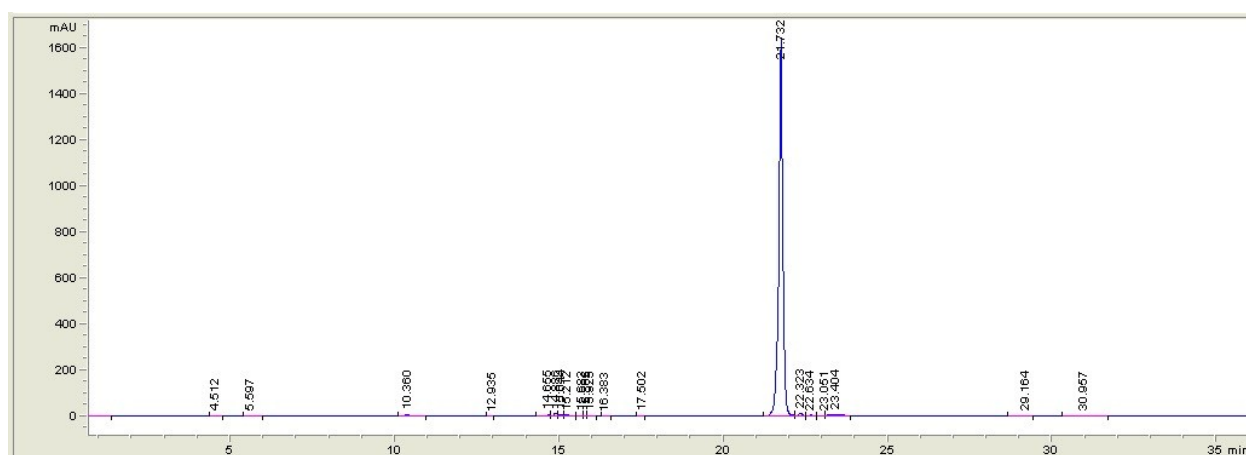
Oligonucleotide 9.4 - HPLC traces and MS spectrum



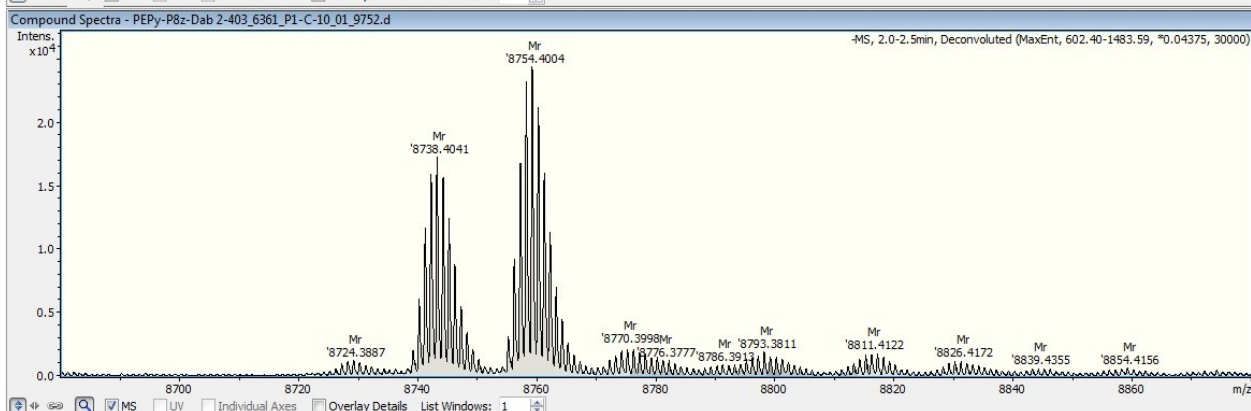
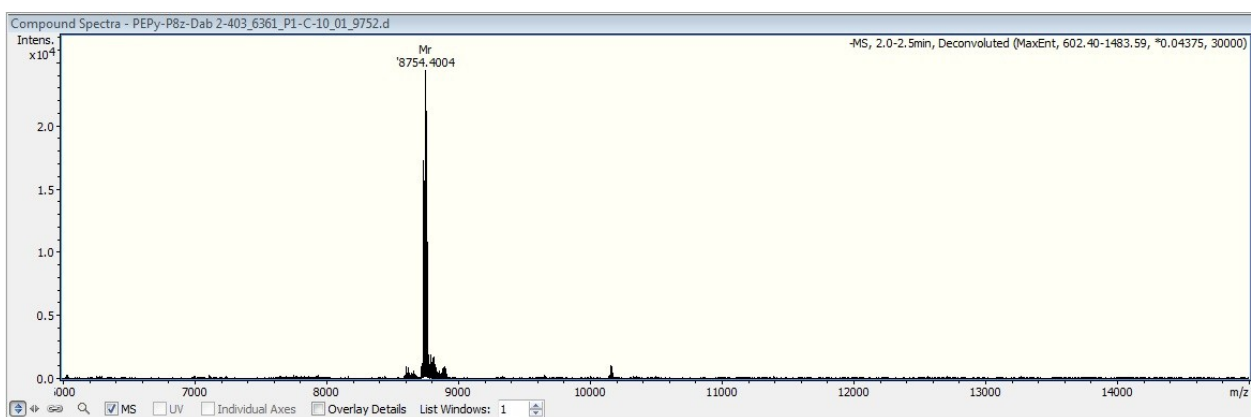
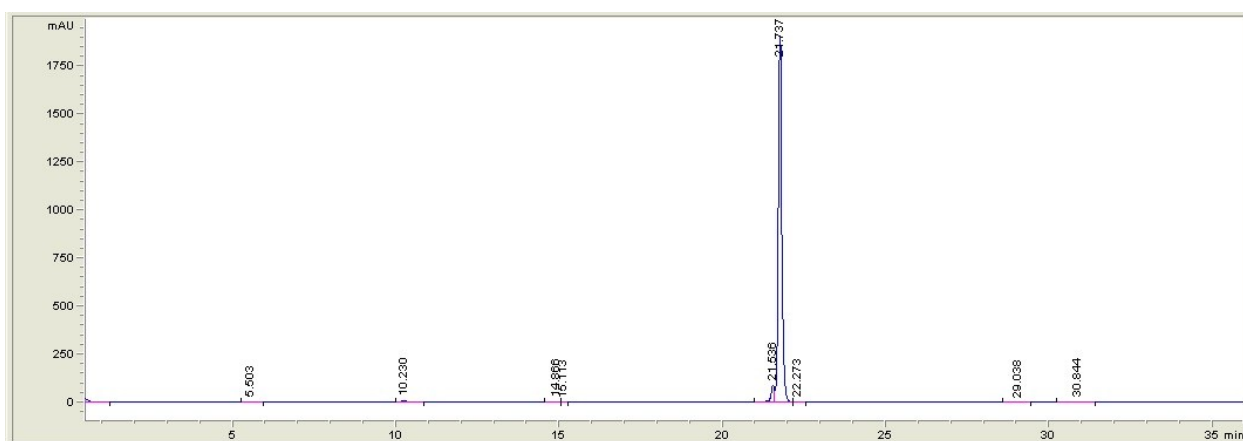
Oligonucleotide 9D - HPLC traces and MS spectrum



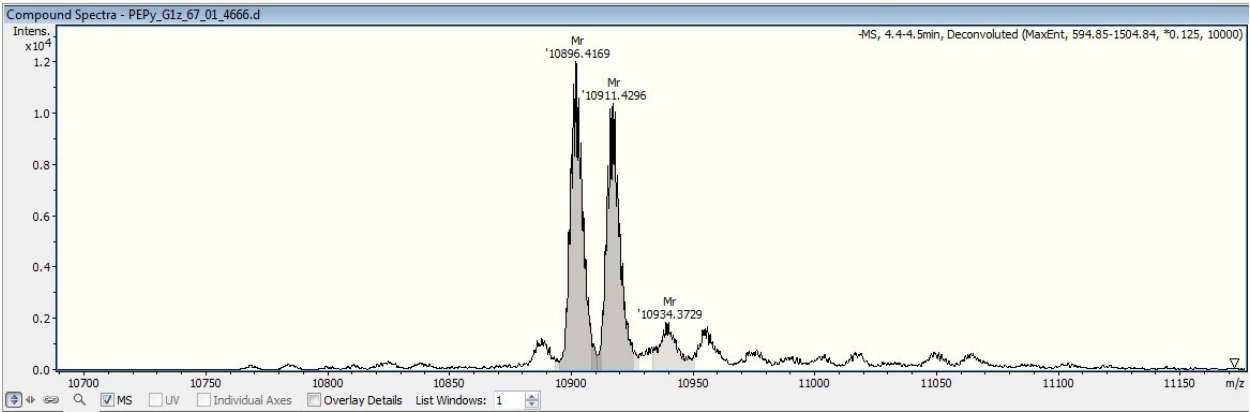
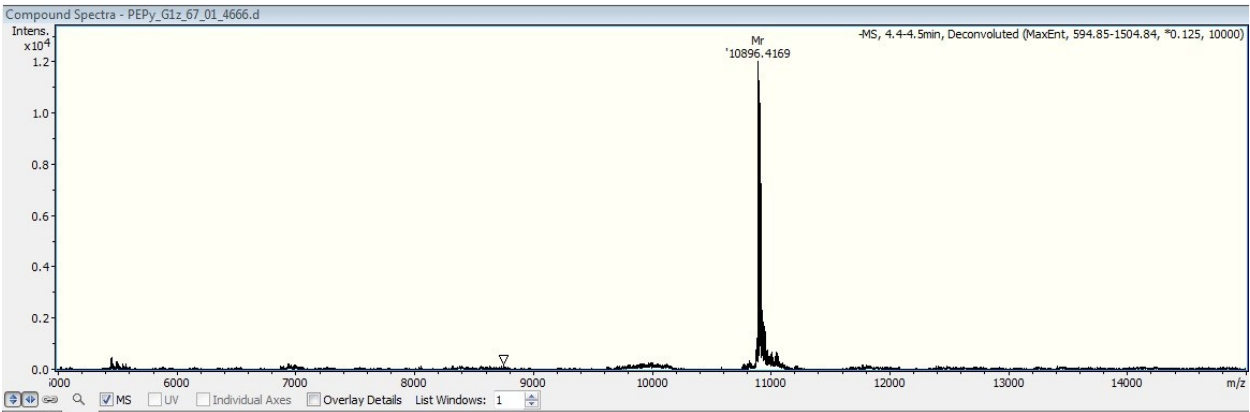
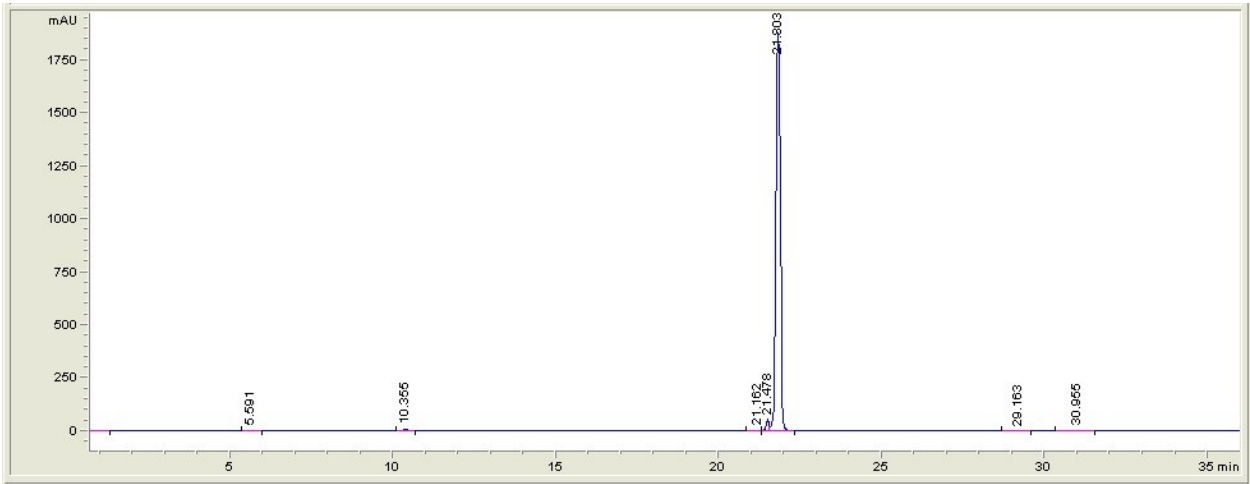
Oligonucleotide PEPy-P4z - HPLC traces and MS spectrum



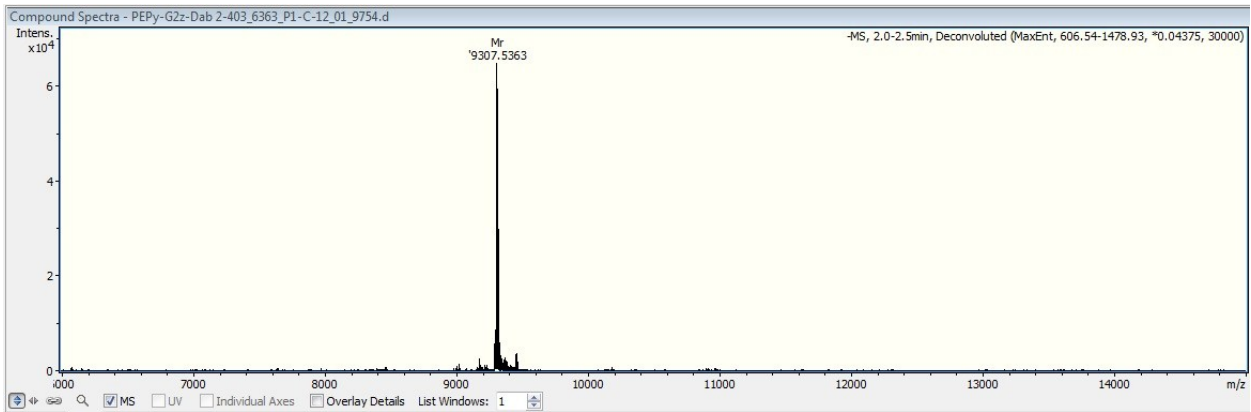
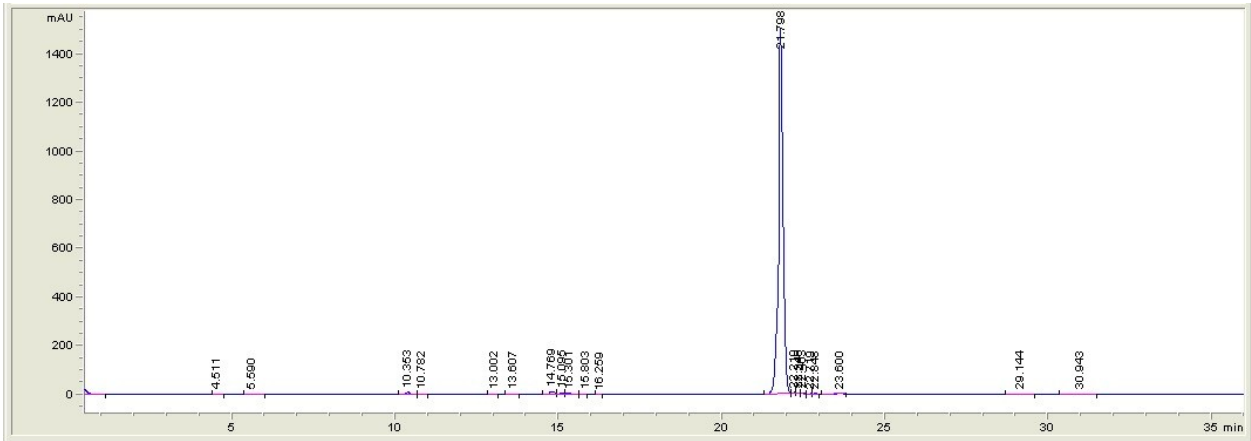
Oligonucleotide PEPy-P8z - HPLC traces and MS spectrum



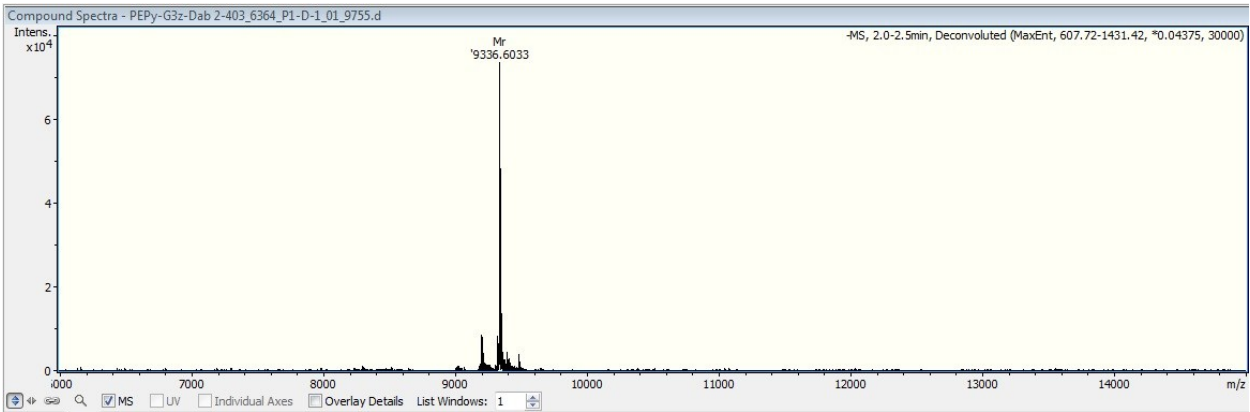
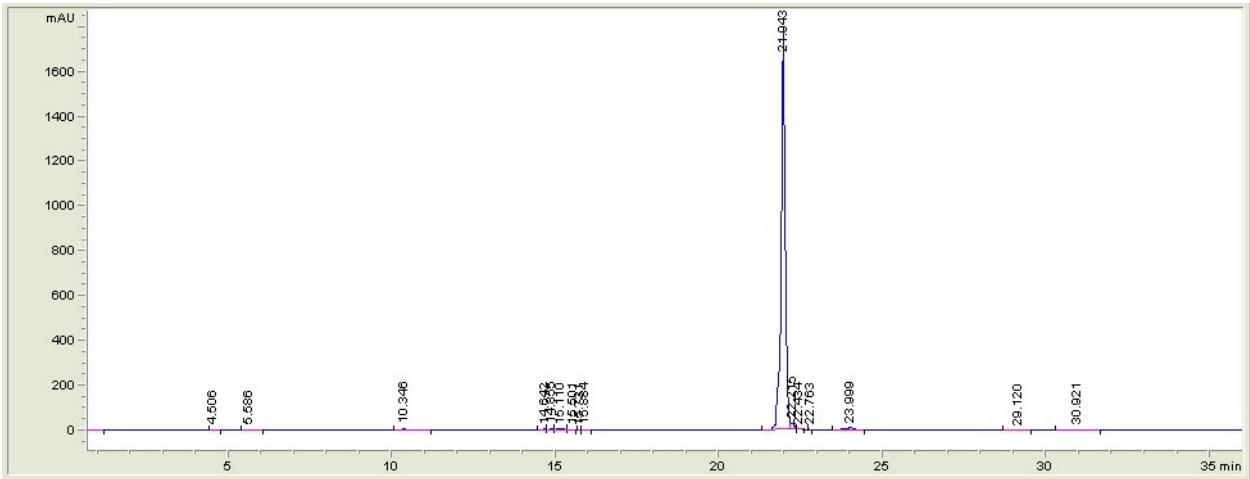
Oligonucleotide PEPy-G1z - HPLC traces and MS spectrum



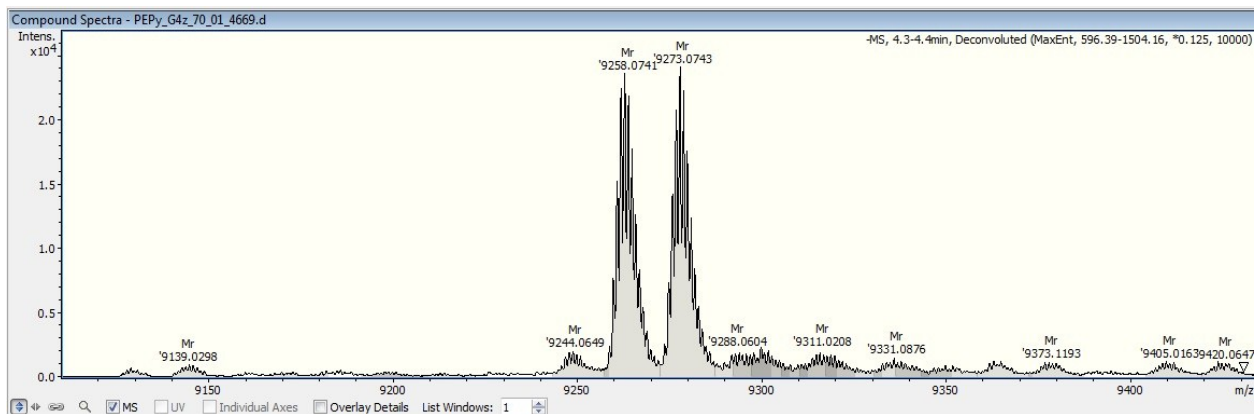
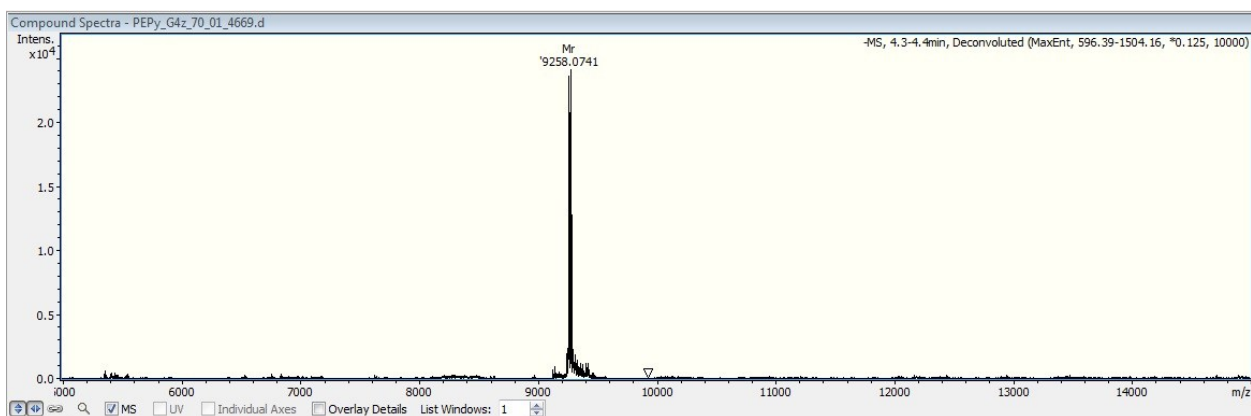
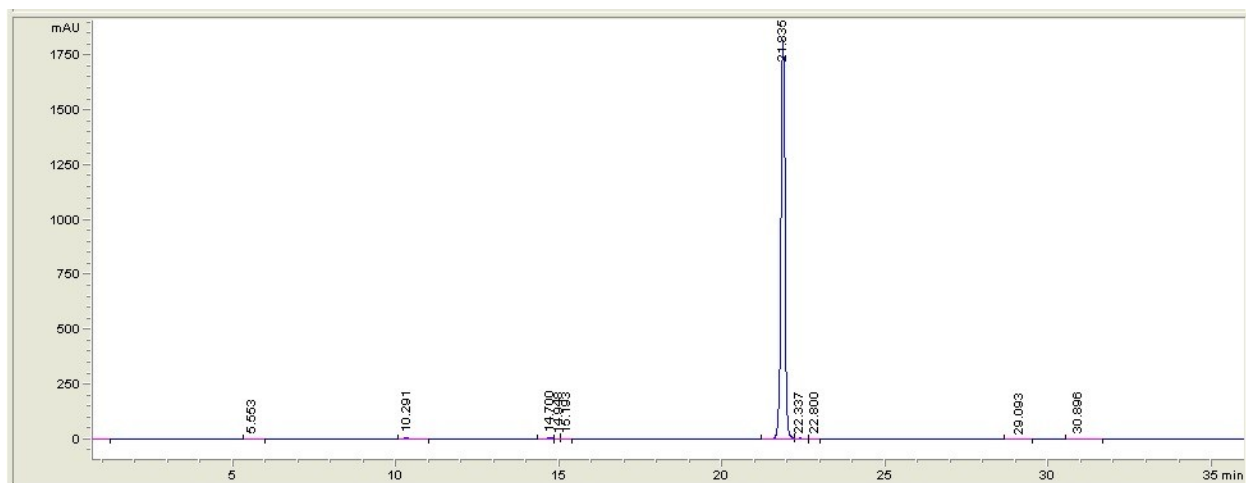
Oligonucleotide PEPy-G2z - HPLC traces and MS spectrum



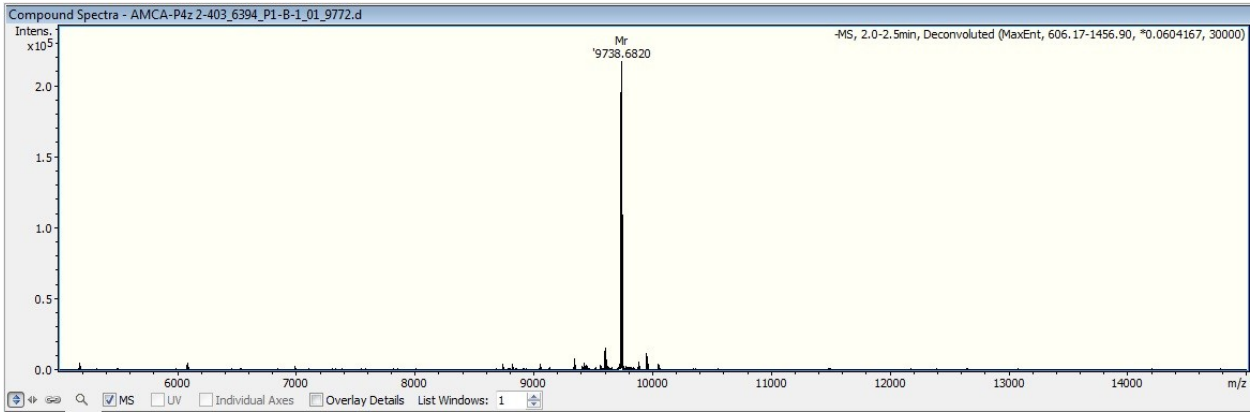
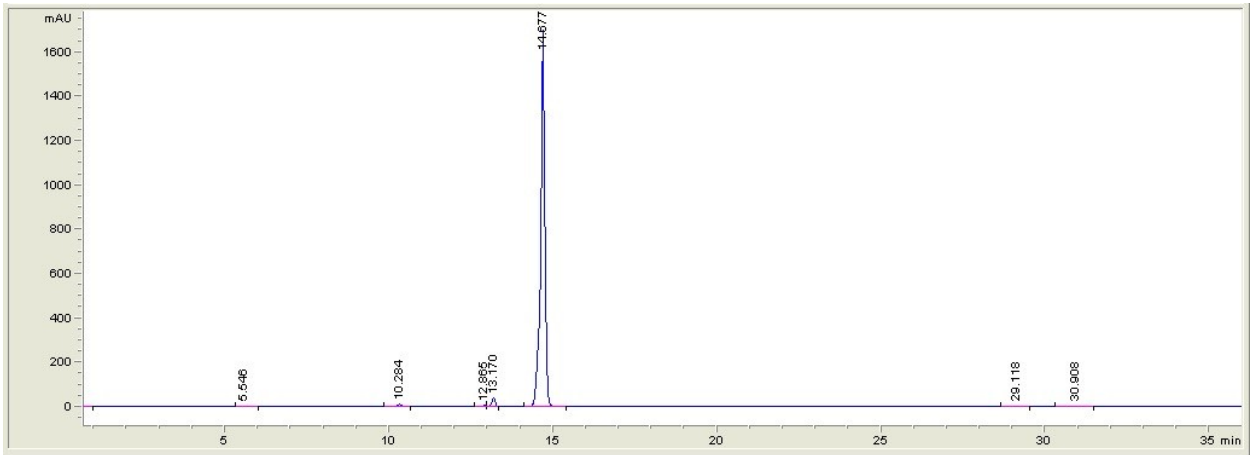
Oligonucleotide PEPy-G3z - HPLC traces and MS spectrum



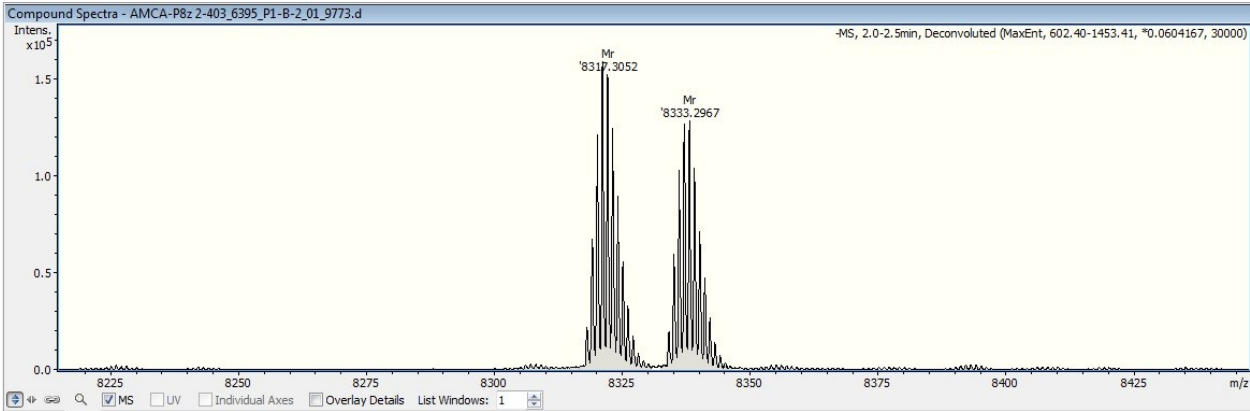
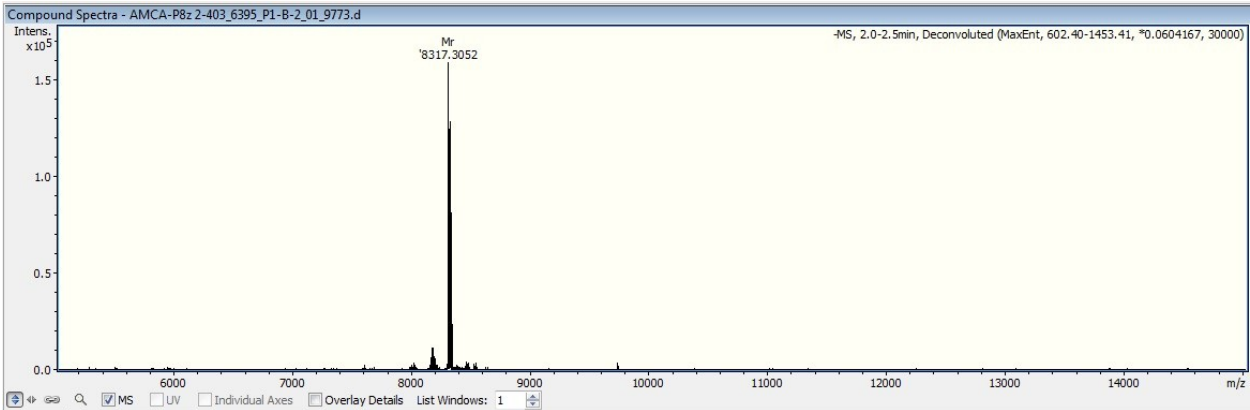
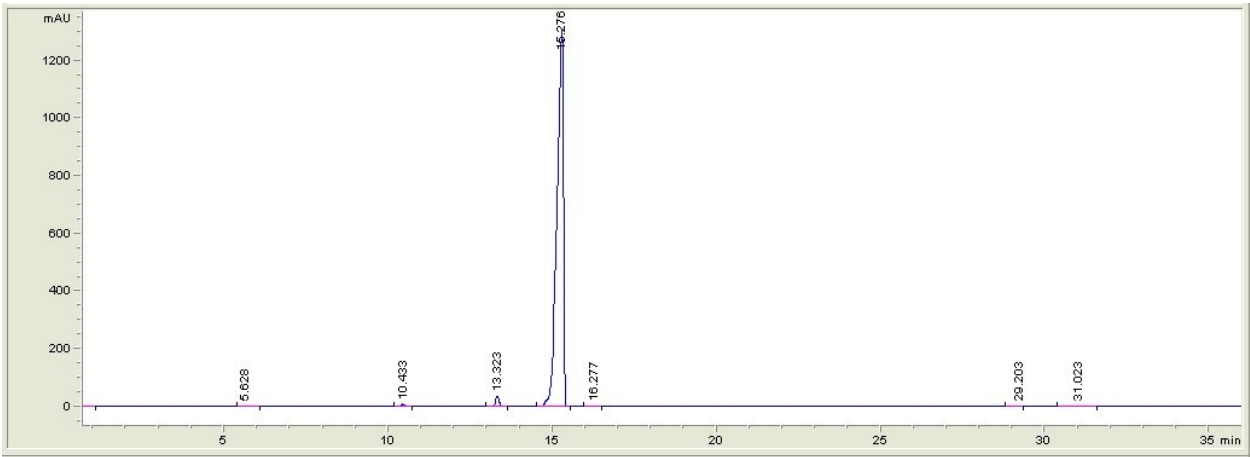
Oligonucleotide PEPy-G4z - HPLC traces and MS spectrum



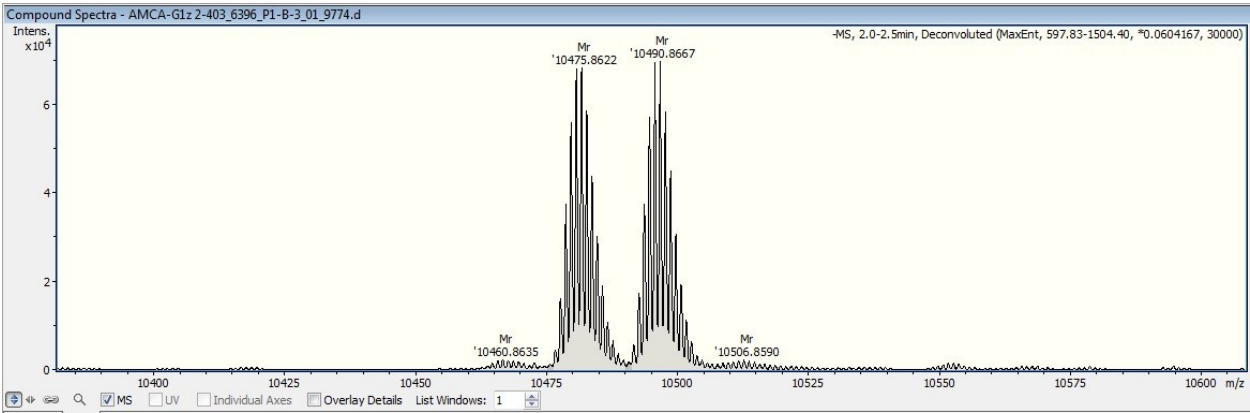
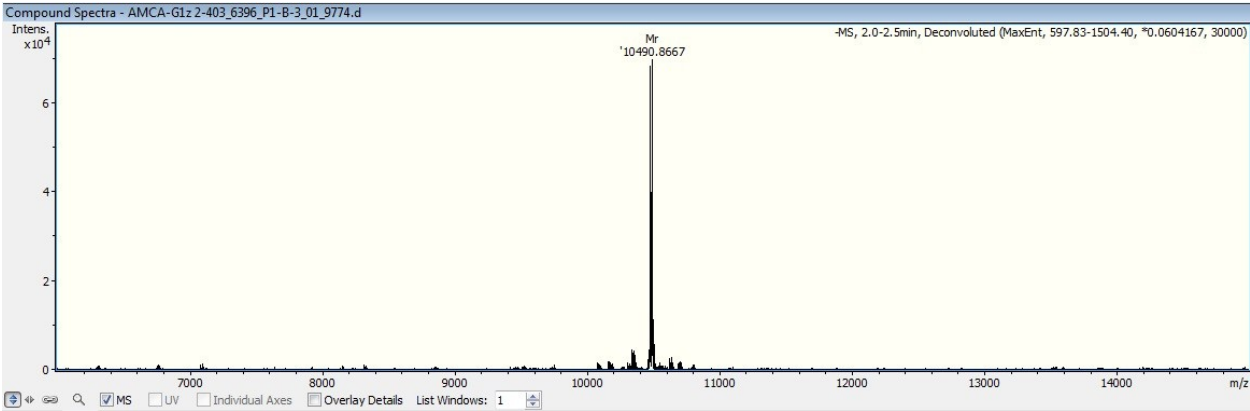
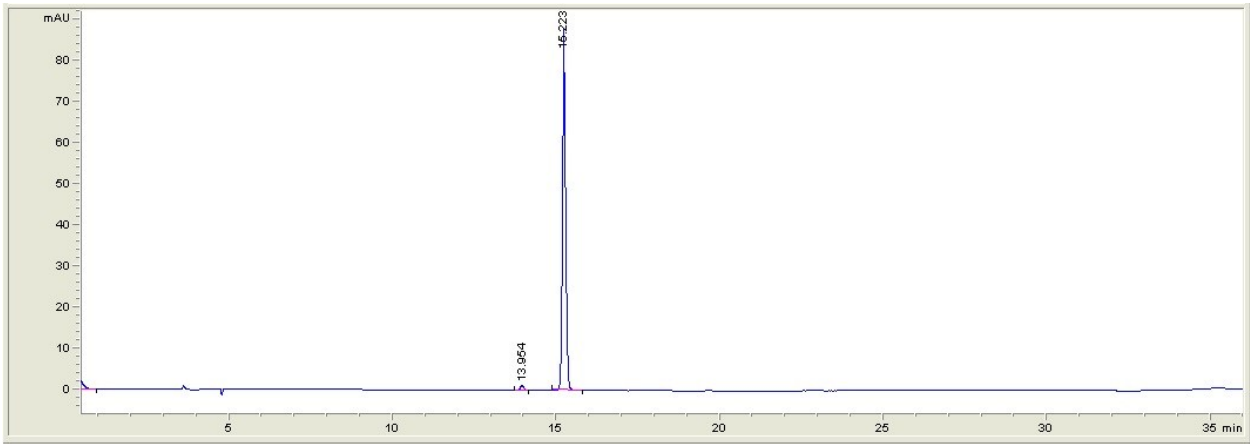
Oligonucleotide AMCA-P4z - HPLC traces and MS spectrum



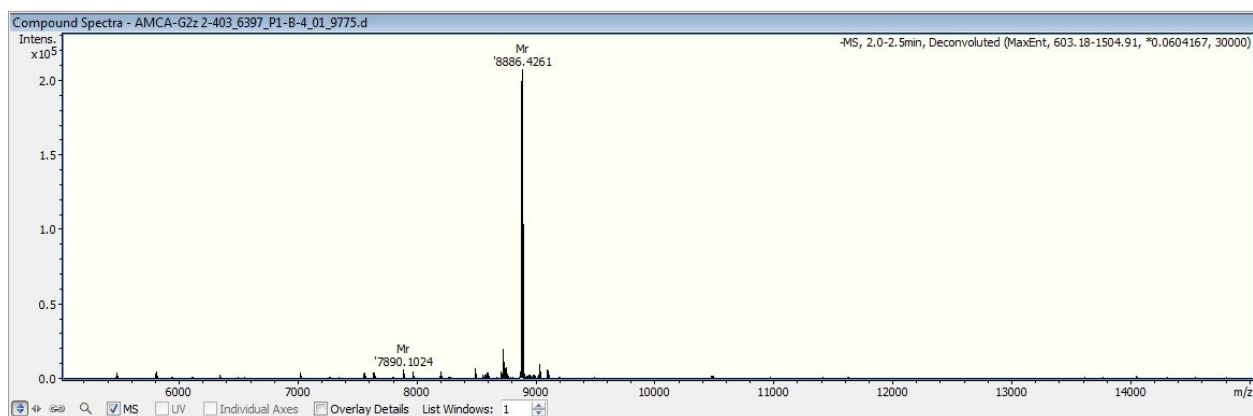
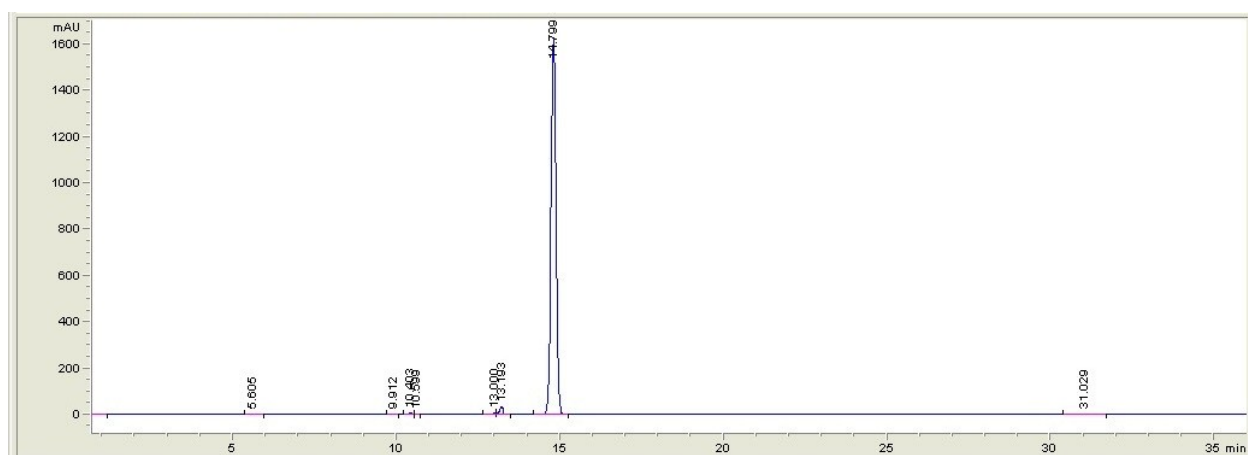
Oligonucleotide AMCA-P8z - HPLC traces and MS spectrum



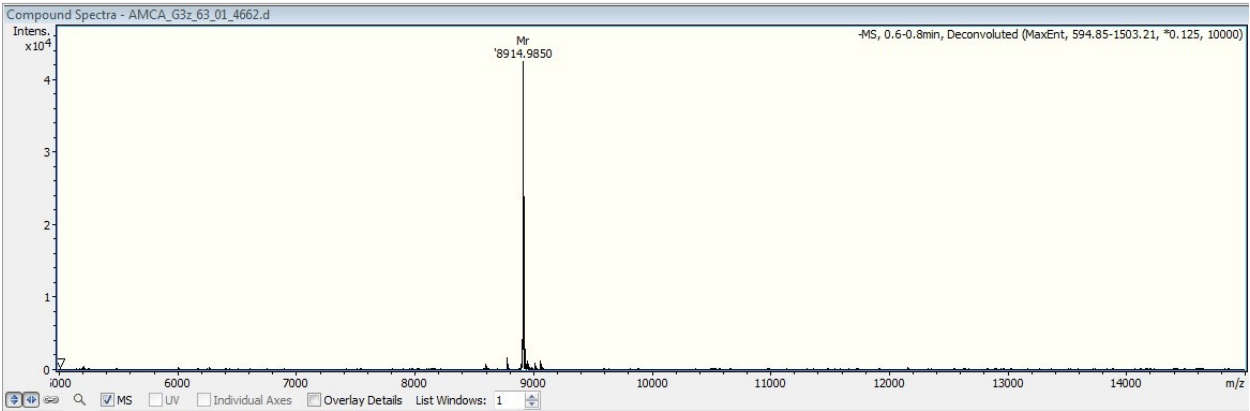
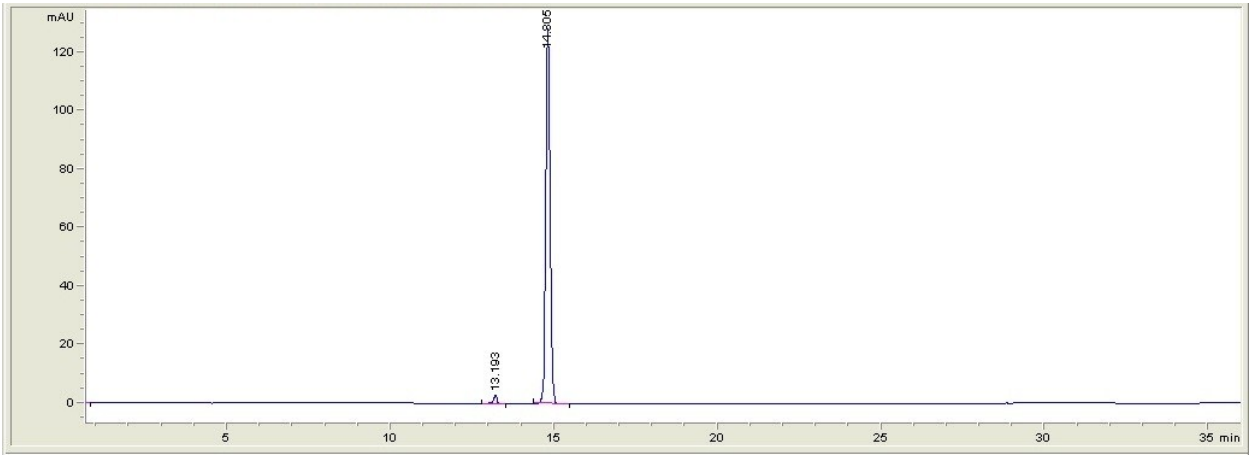
Oligonucleotide AMCA-G1z - HPLC traces and MS spectrum



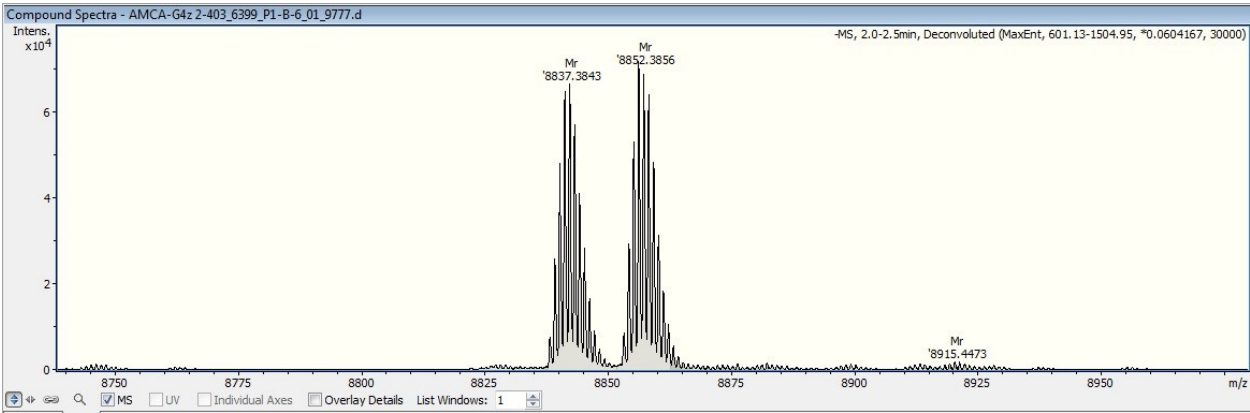
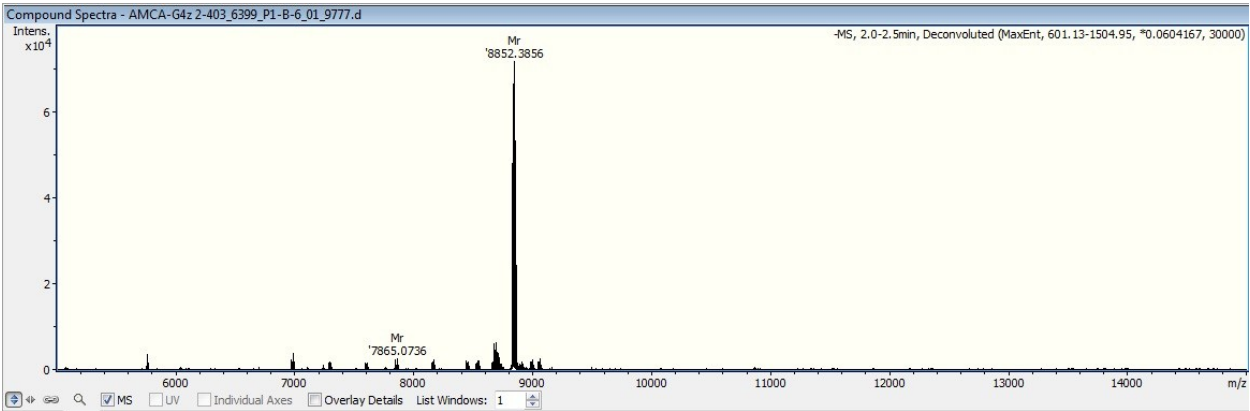
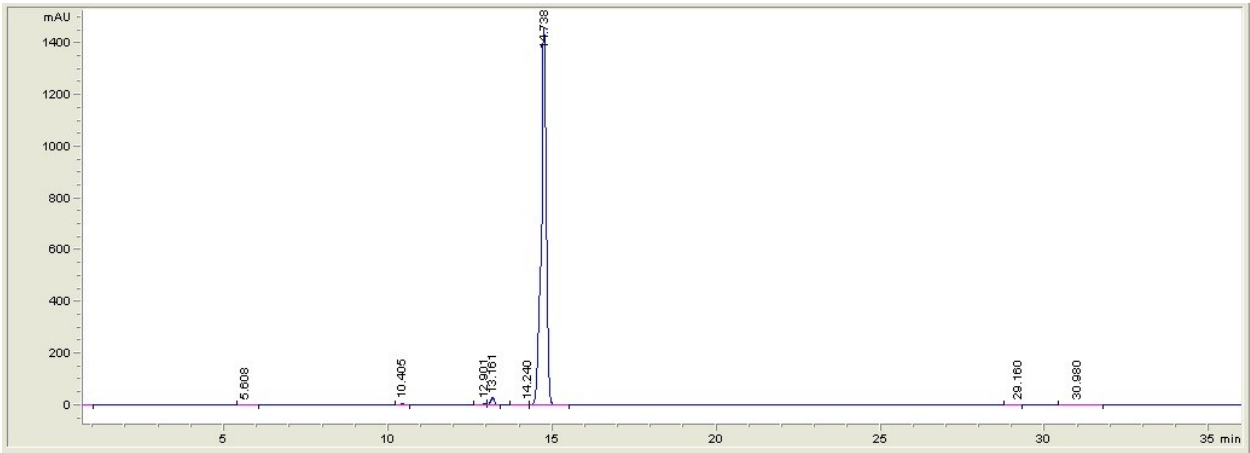
Oligonucleotide AMCA-G2z - HPLC traces and MS spectrum



Oligonucleotide AMCA-G3z - HPLC traces and MS spectrum

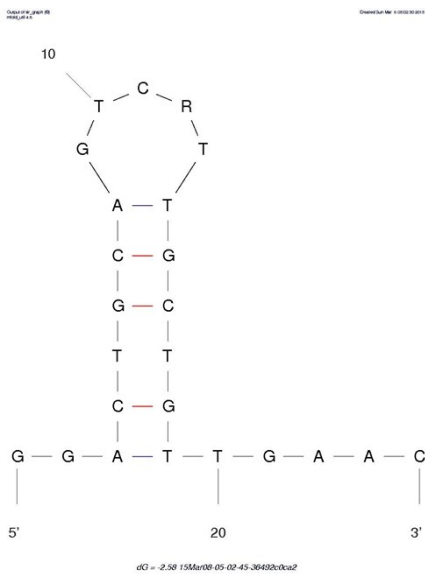


Oligonucleotide AMCA-G4z - HPLC traces and MS spectrum

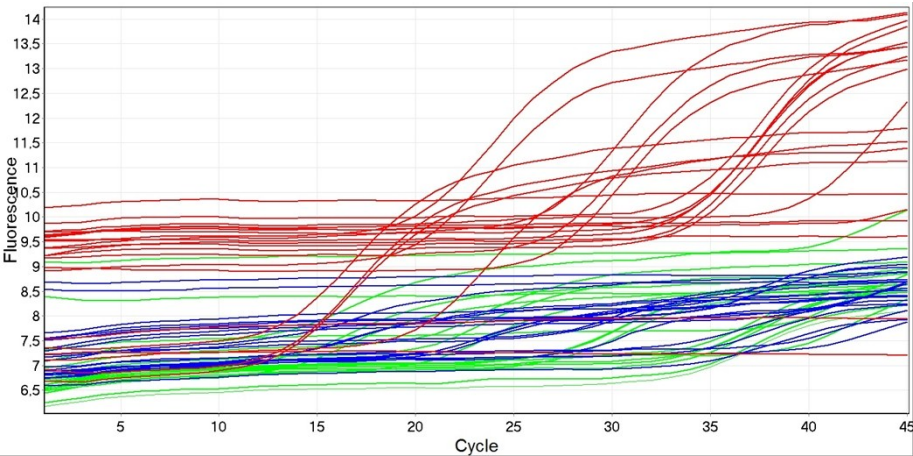


The diagram shows a circular plasmid DNA molecule with a linear double-stranded DNA fragment. The circular molecule is composed of 18 nucleotides: 10 on the left, 20 on the right, and 18 in the middle (5'-T-A-C-G-A-T-T-T-T-T-T-T-T-T-T-3'). The linear fragment is a double-stranded DNA molecule with a 5' end on the left and a 3' end on the right. The top strand of the linear fragment is 5'-C-A-C-G-C-G-3' and the bottom strand is 3'-G-C-G-C-G-C-5'. The linear fragment is connected to the circular molecule at the 5' end of the top strand (C) and the 3' end of the bottom strand (G).

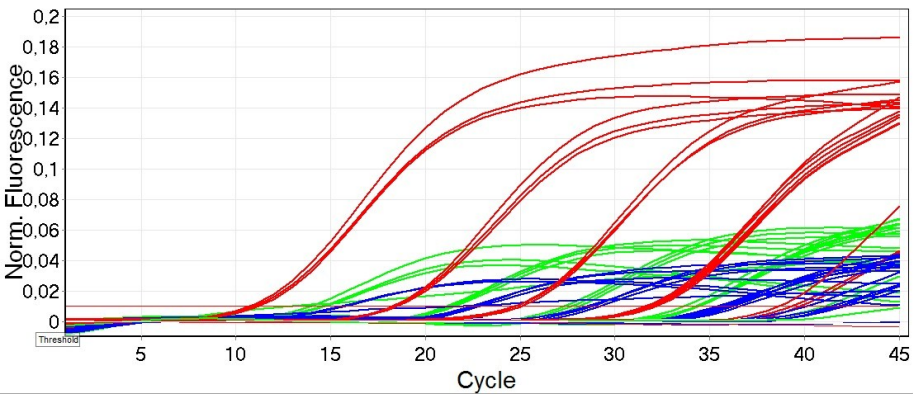
Secondary structure of P8z probes optimized by Mfold. Raw and normalized data of qPCR



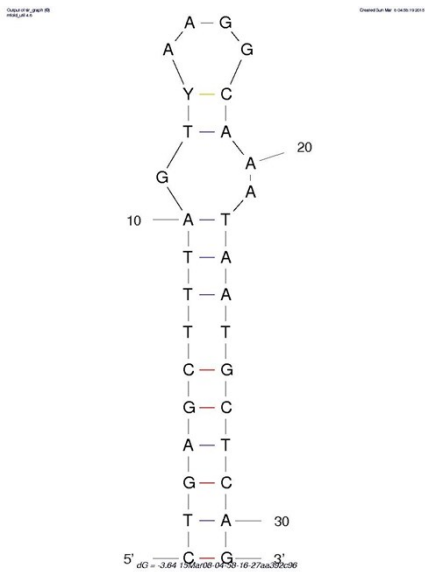
qPCR - raw data



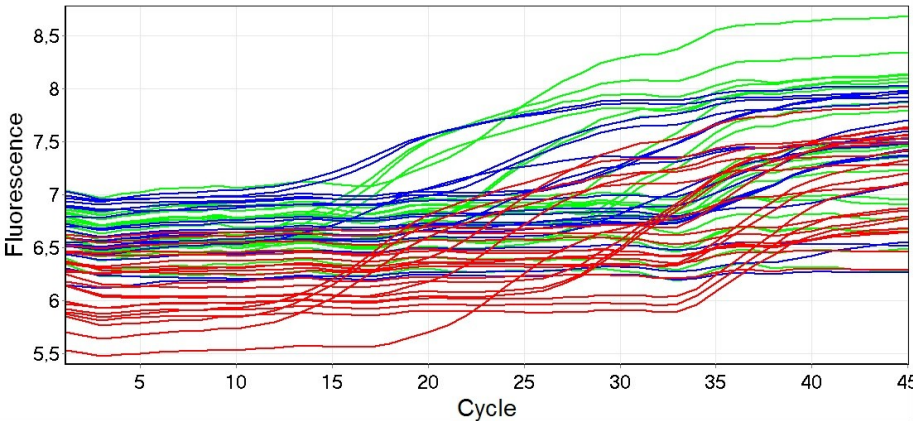
qPCR - normalized data



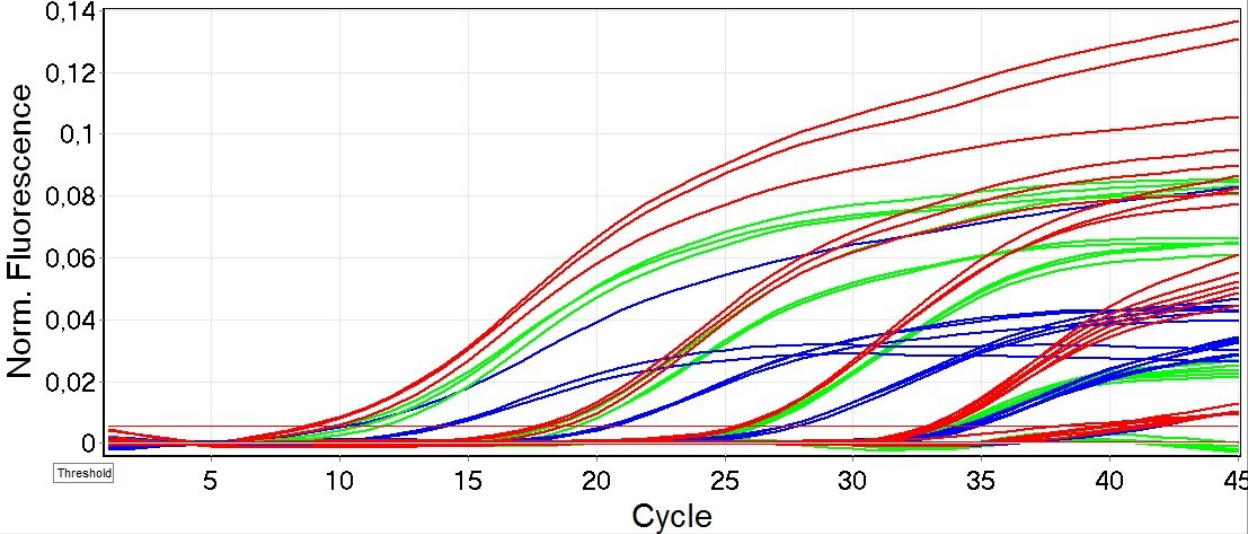
Secondary structure of G1z probes optimized by Mfold. Raw and normalized data of qPCR



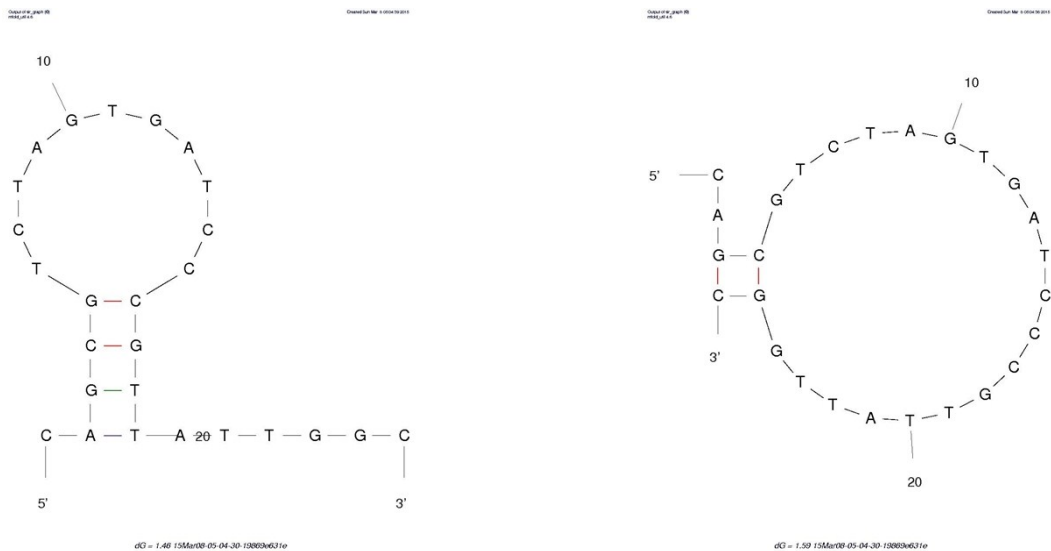
qPCR - raw data



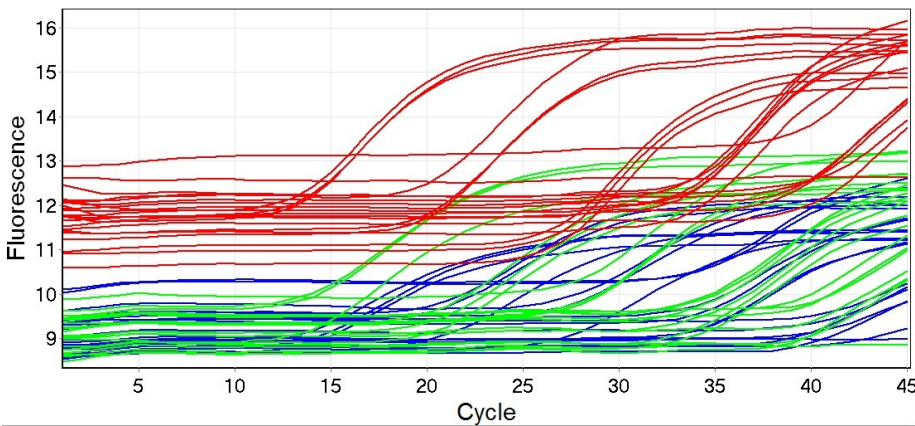
qPCR - normalized data



Secondary structure of G2z probes optimized by Mfold. Raw and normalized data of qPCR



qPCR - raw data



qPCR - normalized data

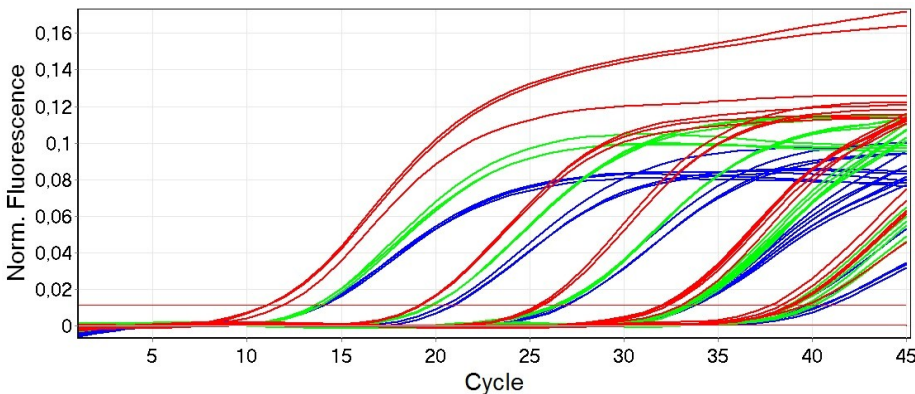


Diagram of a DNA hairpin loop structure. The top part is a circular loop with the sequence 5'-G-T-C-A-A-C-A-G-C-A-C-3'. The bottom part is a stem with the sequence 5'-C-G-C-G-C-A-T-A-3'. The stem is formed by base pairing between the loop's sequence and a complementary sequence. The stem sequence is 5'-C-G-C-G-C-A-T-A-3'.

3'pdr12747.ppt01-05
2024-04-16

5' — C —
|
T —
|
G —
|
A —
|
A —
|
C — Y — T —
| | |
G — C — G —
| | |
G —
|
C — C — A —
| | |
G — G — T —
|
T —
|
3' — T —

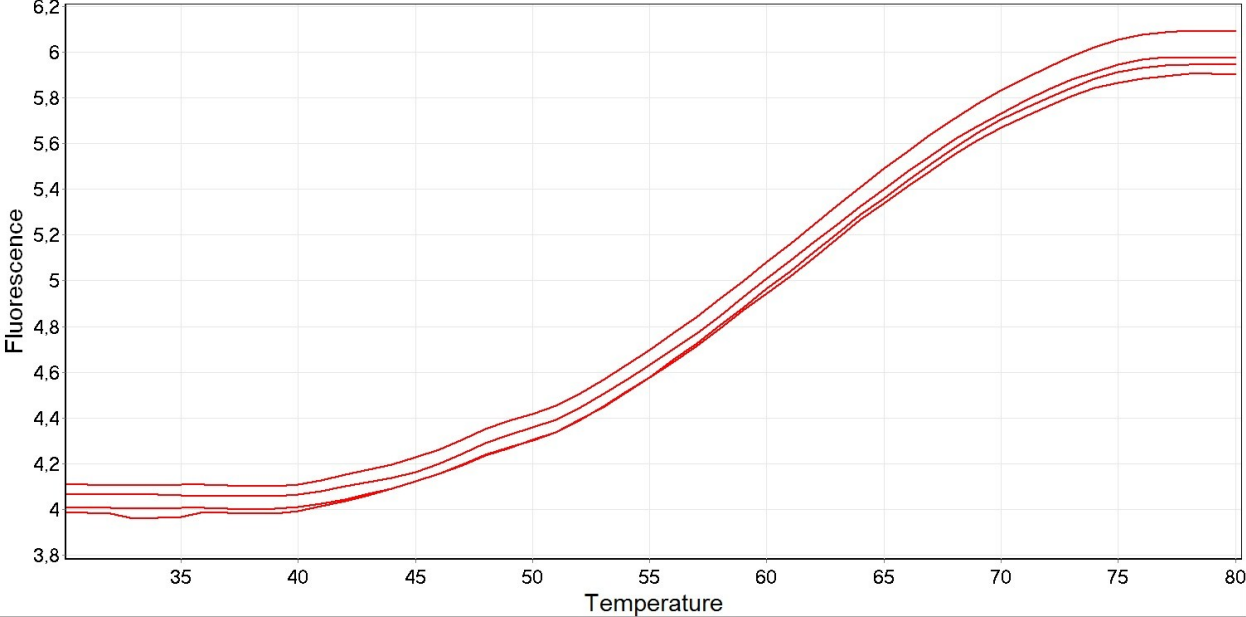
10 20

0.42 = -0.46 / 554609.05-09.40 / 1160316-56

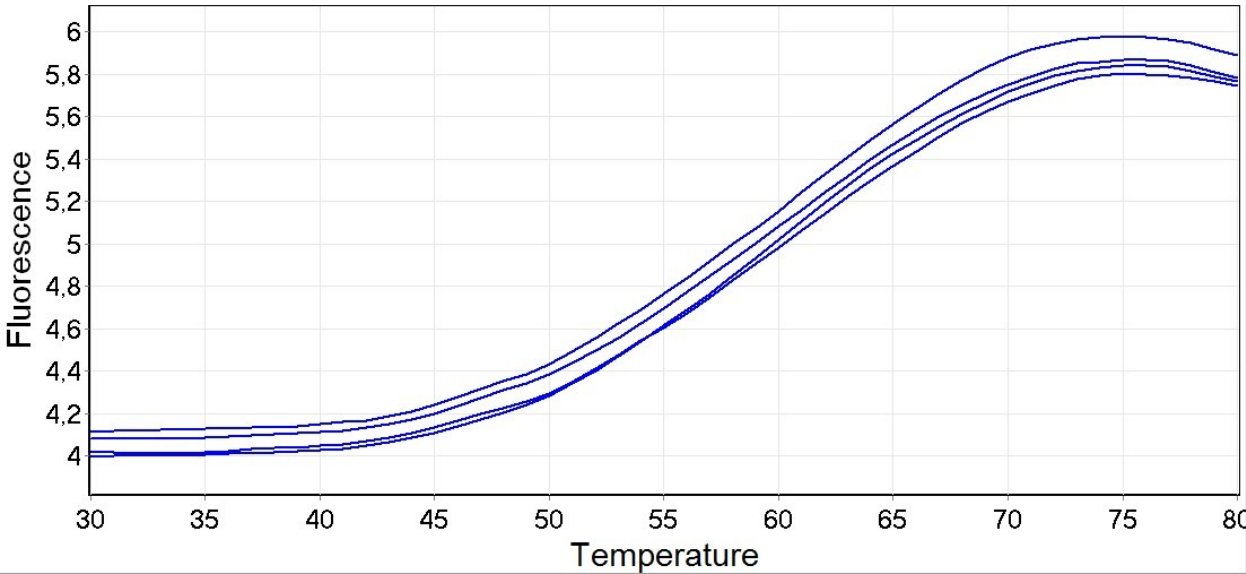
Melting curves for PEPy-labeled P8z, P4z, G1z, G2z, G3z, G4z probes.

PEPy-P8z

Heating

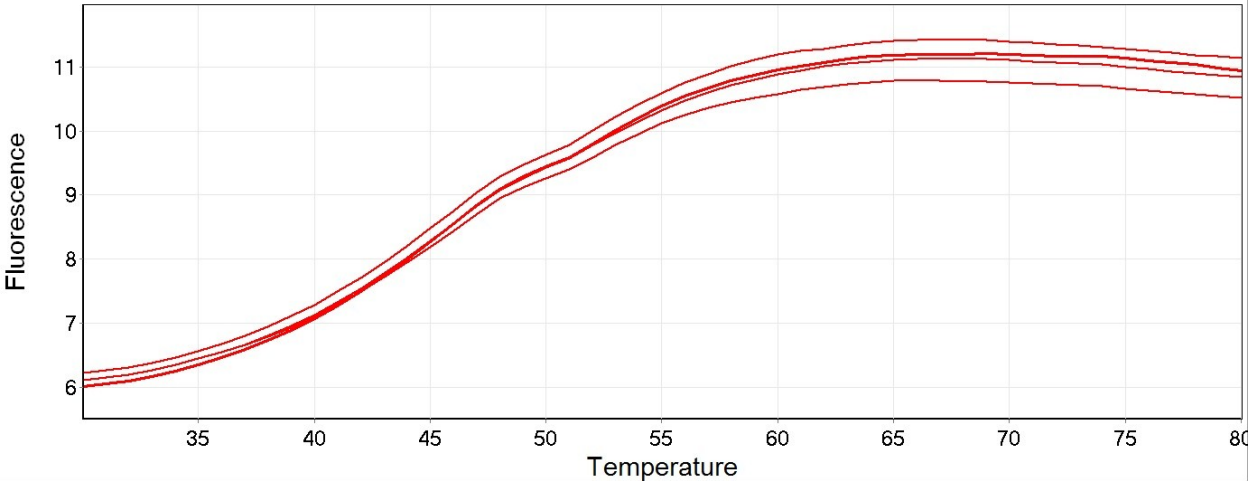


Cooling

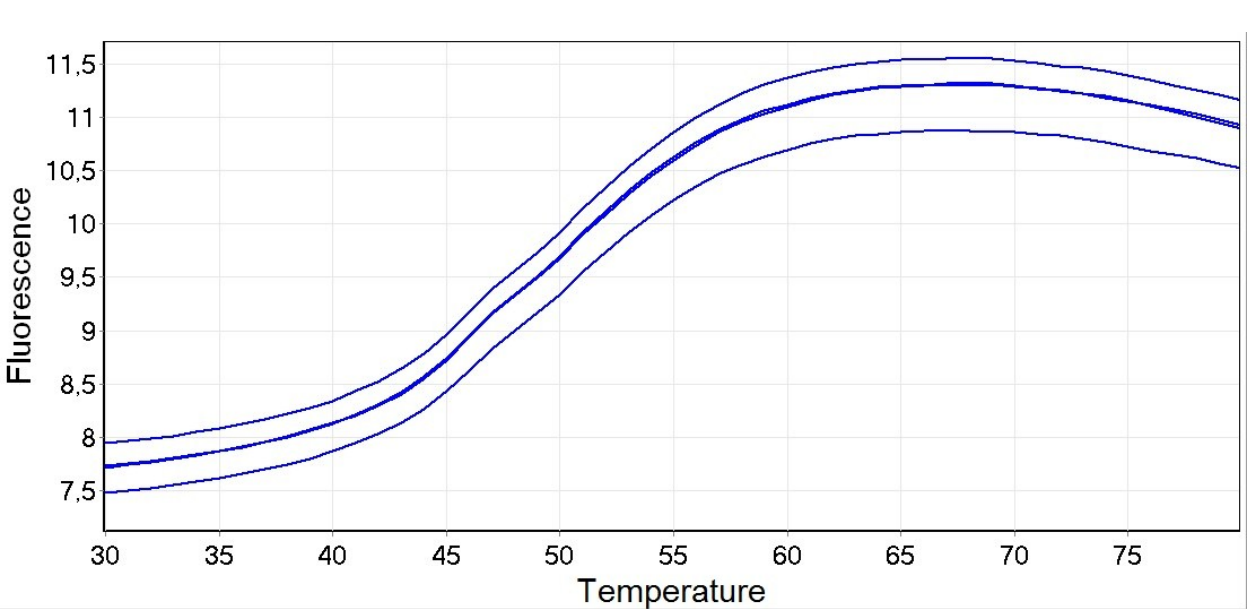


PEPy-P4z

Heating

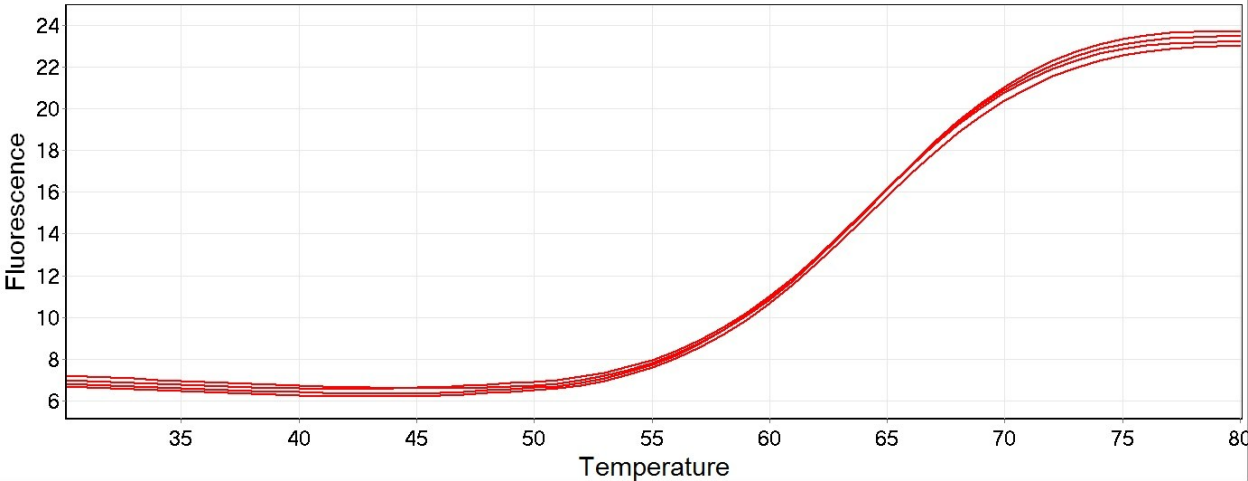


Cooling

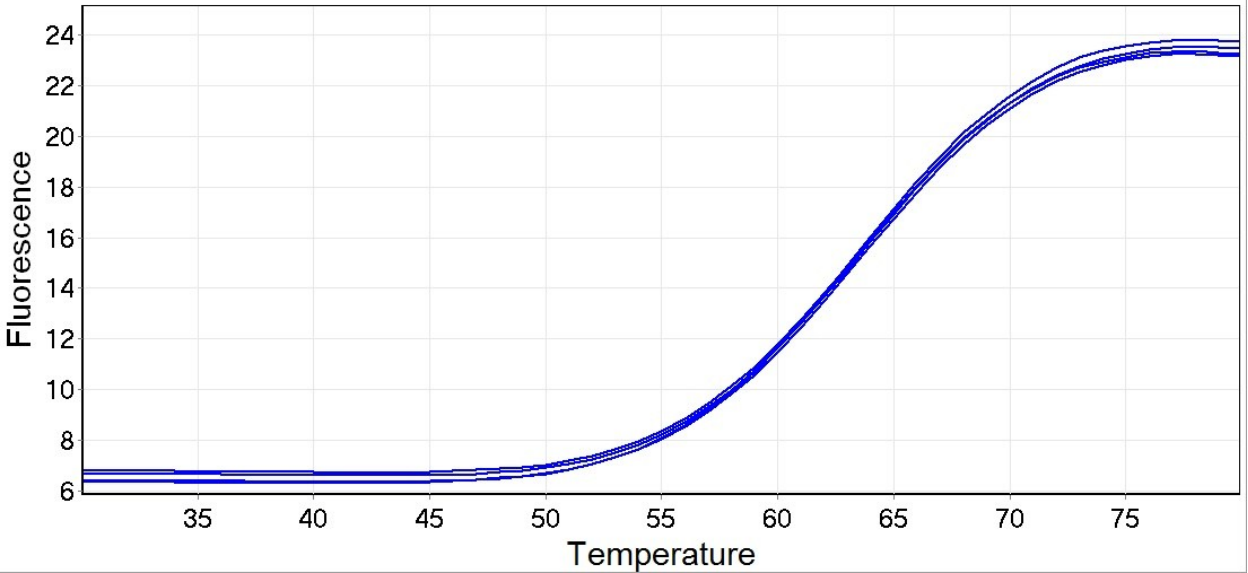


PEPy-G1z

Heating

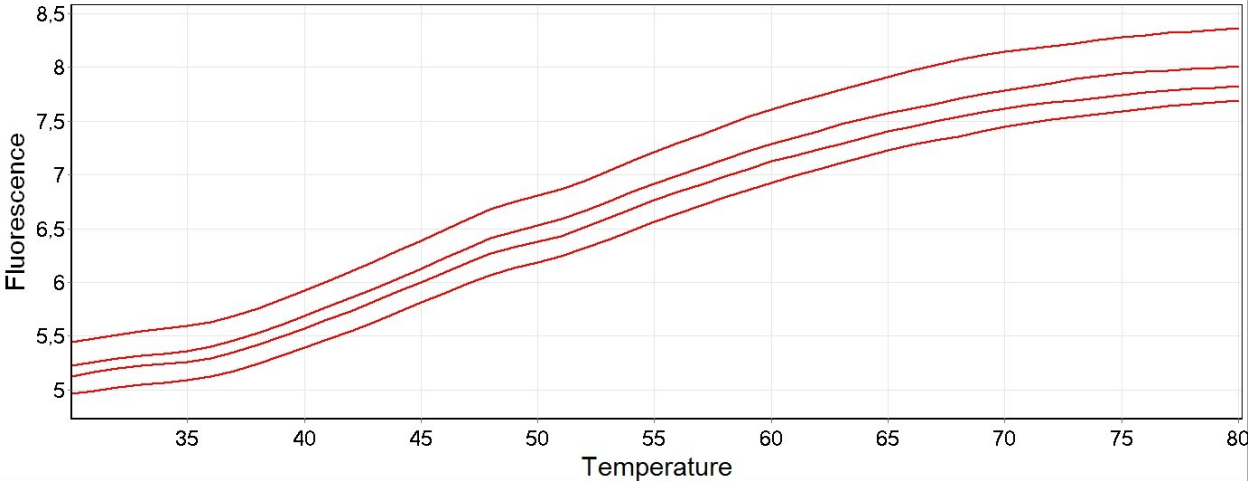


Cooling

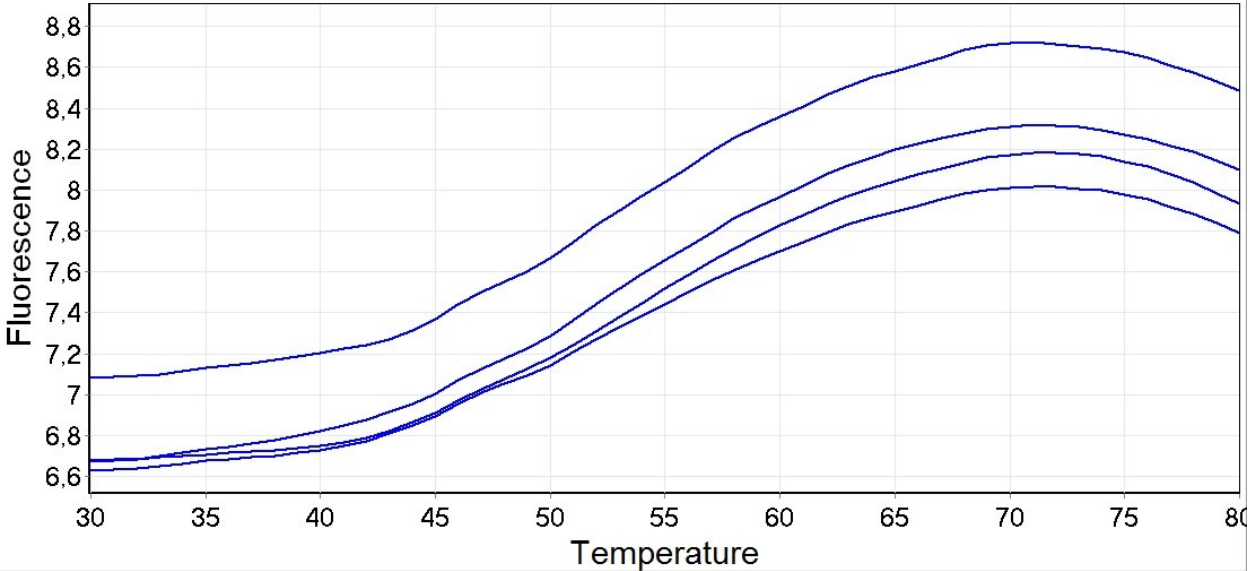


PEPy-G2z

Heating

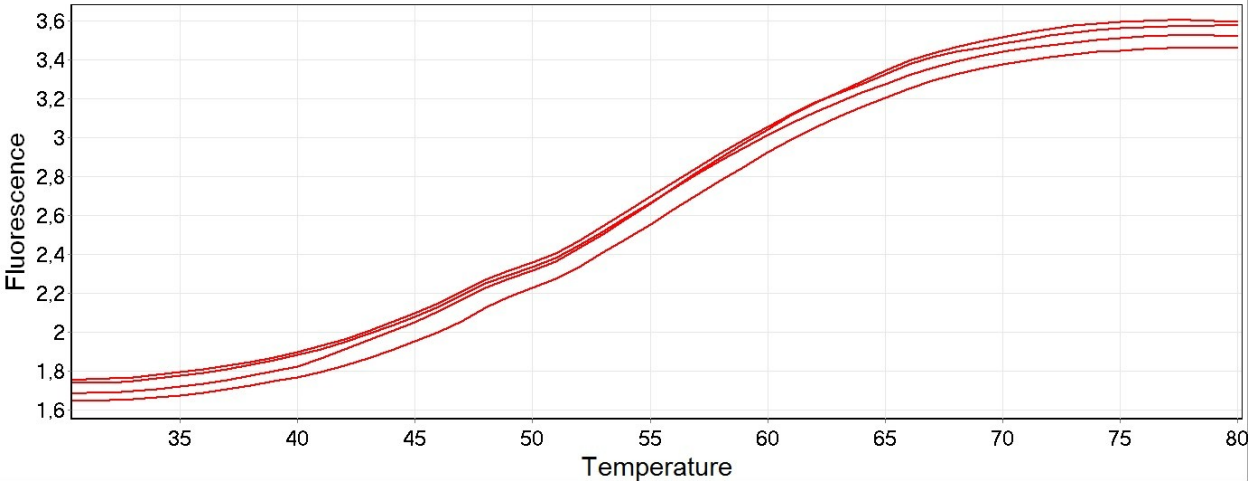


Cooling

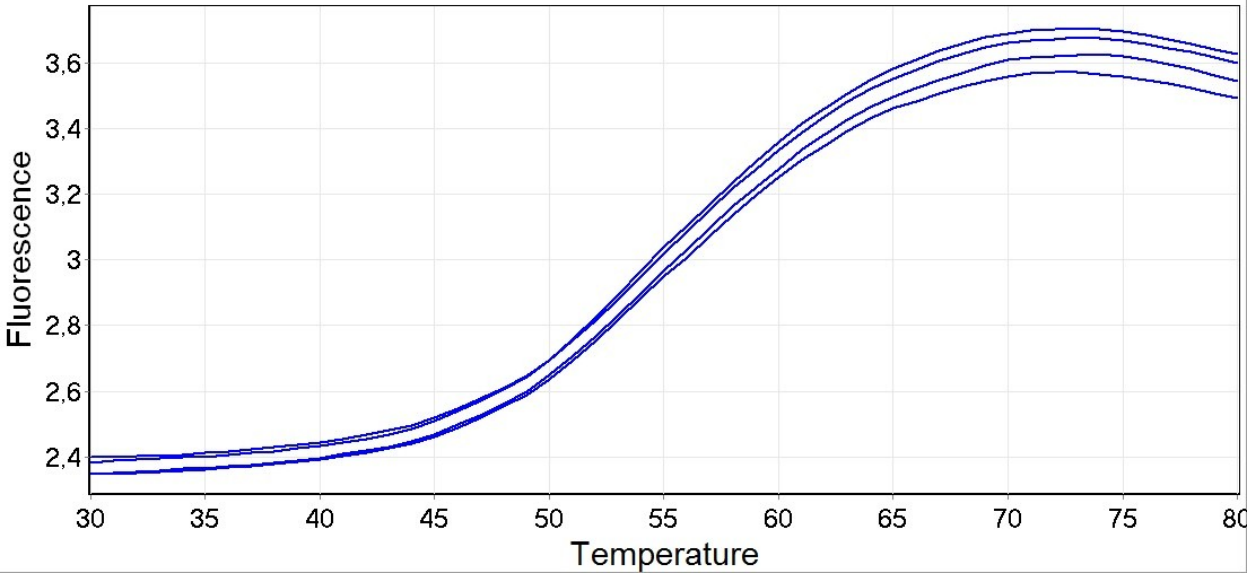


PEPy-G3z

Heating

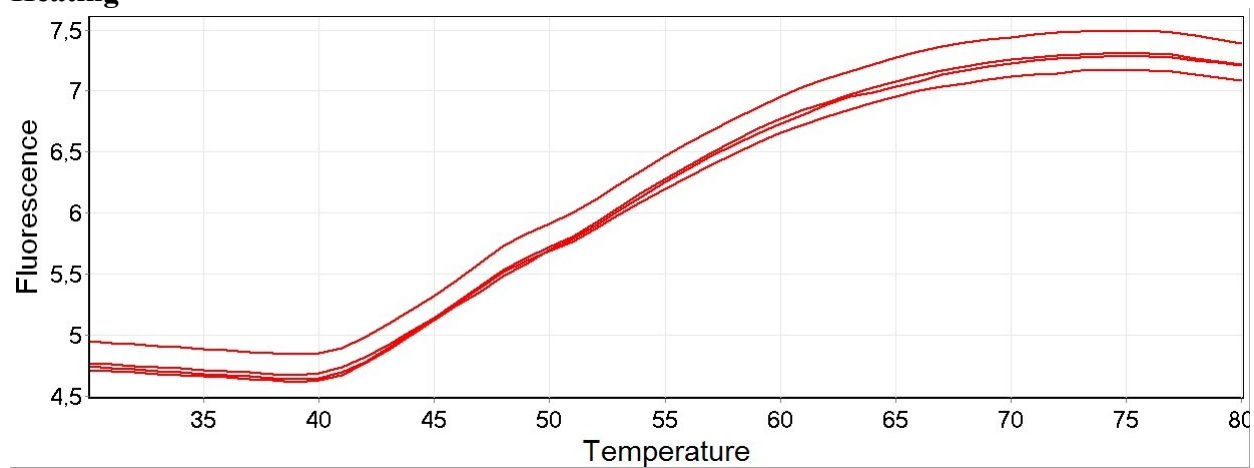


Cooling

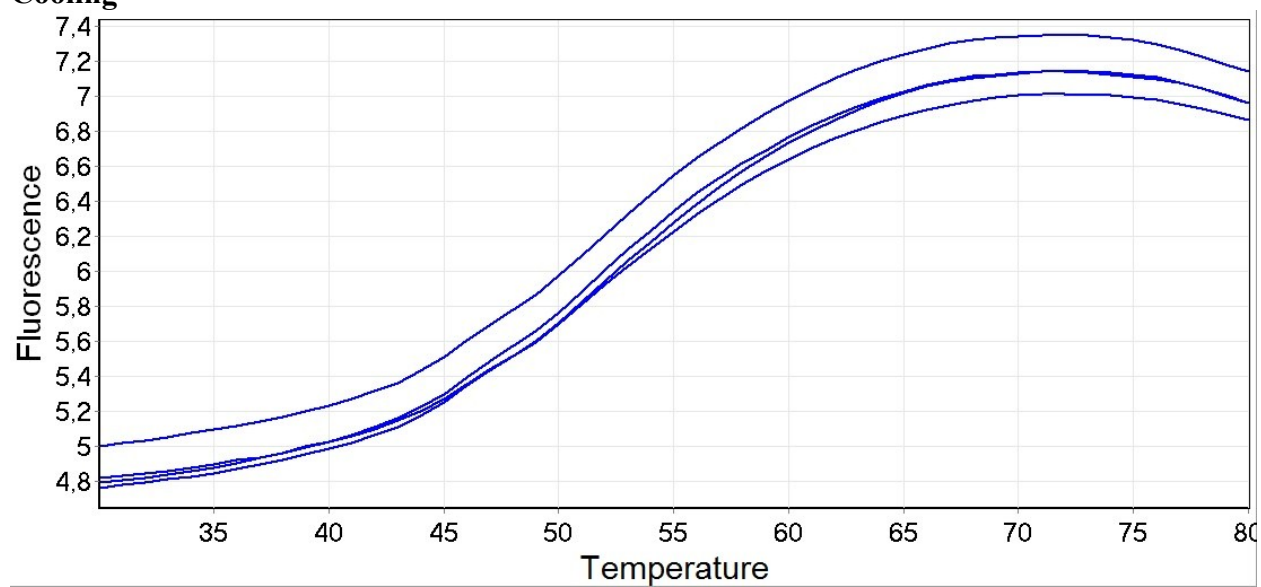


PEPy-G4z

Heating



Cooling



qPCR data for cross-talk studies between “blue” and “green” channels.

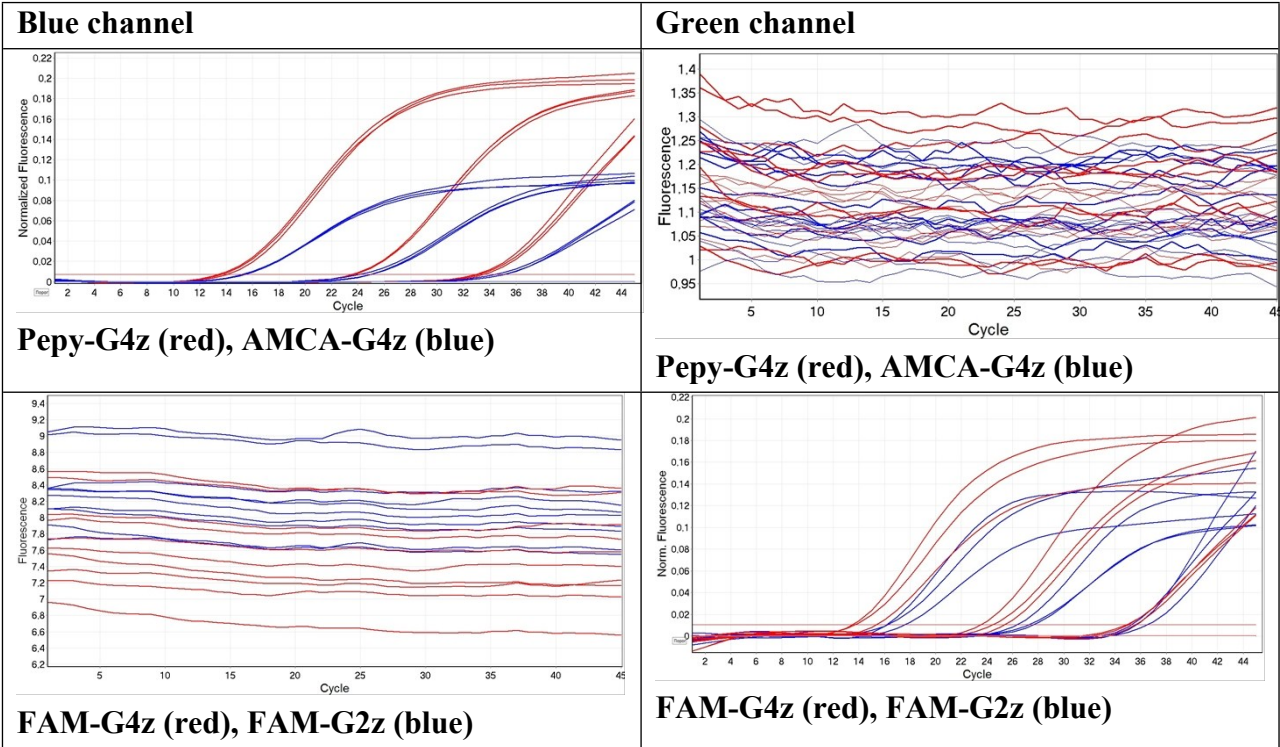


Table S4. qPCR data for AMCA, Alexa 350 and PEPy dual labeled probes.

Probe	Probe sequence, 5'-3'	qPCR data					
		Target DNA, copies/reaction	Cq (PEPy)	Cq (AMCA)	Cq (Alexa350)	ΔCq ($Ct_{AMCA} - Ct_{PEPy}$)	ΔCq ($Ct_{Alexa350} - Ct_{PEPy}$)
P4z	Dye -CAC TCT GAC TAC TAC CTT TAA ACA GAG CG- Dabcy1	$1.0 \cdot 10^{10}$	18.9±0.4	21.6±0.3	21.6±0.2	2.7±0.4	2.7±0.3
		$1.0 \cdot 10^8$	25.8±0.3	28.9±0.2	26.9±0.3	3.1±0.3	1.1±0.3
		$1.0 \cdot 10^6$	32.8±0.2	35.9±0.3	35.0±0.4	3.1±0.3	2.2±0.4
		$1.0 \cdot 10^4$	40.5±0.3	41.9±0.3	42.5±0.3	1.4±0.3	2.0±0.3
		$1.0 \cdot 10^2$	-	-	-	-	-
P8z	Dye -GGA CTG CAG TCR TTG CTG TTG AAC- Dabcy1	$1.0 \cdot 10^{10}$	11.0±0.4	15.7±0.4	14.4±0.4	4.7±0.3	3.4±0.4
		$1.0 \cdot 10^8$	18.4±0.3	22.6±0.3	21.3±0.3	4.2±0.3	2.9±0.3
		$1.0 \cdot 10^6$	24.8±0.4	28.9±0.2	27.7±0.4	4.1±0.4	2.9±0.4
		$1.0 \cdot 10^4$	31.9±0.3	36.0±0.3	34.2±0.3	4.1±0.3	2.3±0.3
		$1.0 \cdot 10^2$	38.7±0.4	41.5±0.3	40.1±0.3	3.8±0.4	1.4±0.4
G1z	Dye -CTG AGC TTT AGT YAA GGC AAA TAA TGC TCA G- Dabcy1	$1.0 \cdot 10^{10}$	10.6±0.3	14.5±0.3	12.0±0.3	3.9±0.3	1.4±0.3
		$1.0 \cdot 10^8$	19.2±0.2	21.9±0.3	19.9±0.3	2.7±0.3	0.7±0.3
		$1.0 \cdot 10^6$	26.7±0.3	28.9±0.3	27.1±0.3	2.2±0.3	0.4±0.3
		$1.0 \cdot 10^4$	34.1±0.3	35.9±0.3	35.0±0.3	1.8±0.3	0.9±0.3
		$1.0 \cdot 10^2$	43.3±0.3	44.4±0.3	>45	1.1±0.3	>1.7
G2z	Dye -CAG CGT CTA GTG ATC CCG TTA TTG GC- Dabcy1	$1.0 \cdot 10^{10}$	11.5±0.3	14.1±0.3	13.8±0.3	2.6±0.3	2.3±0.2
		$1.0 \cdot 10^8$	19.3±0.3	20.9±0.3	19.9±0.3	1.6±0.3	0.6±0.3
		$1.0 \cdot 10^6$	25.6±0.3	26.9±0.3	26.5±0.3	1.3±0.3	0.9±0.3
		$1.0 \cdot 10^4$	32.3±0.3	33.7±0.3	33.4±0.3	1.4±0.3	1.1±0.3
		$1.0 \cdot 10^2$	38.8±0.3	41.0±0.3	39.5±0.3	2.2±0.3	0.7±0.3
G3z	Dye -TAC CCA ACT GAA GCA GCA ACA GGG TA- Dabcy1	$1.0 \cdot 10^{10}$	10.5±0.3	11.4±0.3	11.1±0.3	0.9±0.3	0.8±0.3
		$1.0 \cdot 10^8$	17.1±0.3	19.2±0.3	17.9±0.3	0.8±0.3	0.8±0.3
		$1.0 \cdot 10^6$	24.0±0.3	25.9±0.3	24.6±0.3	1.9±0.3	0.6±0.3
		$1.0 \cdot 10^4$	30.9±0.3	32.9±0.3	31.4±0.3	0.5±0.3	0.5±0.3
		$1.0 \cdot 10^2$	36.8±0.3	40.2±0.3	39.5±0.3	3.4±0.3	2.7±0.3
G4z	Dye -CTG AAC YTG TCG GCC ATC CTT TGG TT- Dabcy1	$1.0 \cdot 10^{10}$	12.7±0.3	15.0±0.3	14.8±0.5	2.3±0.5	2.1±0.3
		$1.0 \cdot 10^8$	19.2±0.3	21.3±0.3	21.5±0.3	2.1±0.3	2.3±0.3
		$1.0 \cdot 10^6$	25.6±0.3	27.6±0.3	27.7±0.3	2.0±0.3	2.1±0.3
		$1.0 \cdot 10^4$	32.6±0.3	34.7±0.3	34.5±0.3	2.1±0.3	1.9±0.3
		$1.0 \cdot 10^2$	39.1±0.3	41.3±0.3	41.0±0.3	2.2±0.3	1.9±0.3

Table S5. Normalized fluorescence data for AMCA, Alexa 350 and PEPy dual labeled probes in end-point qPCR.

Probe	Probe sequence, 5'-3'	Fluorescence data	
		Dye	Increase of normalized fluorescence
P4z	Dye -CAC TCT GAC TAC TAC CTT TAA ACA GAG CG- Dabcy1	PEPy	0.14±0.01
		Alexa 350	0.06±0.01
		AMCA	0.05±0.01
P8z	Dye -GGA CTG CAG TCR TTG CTG TTG AAC- Dabcy1	PEPy	0.15±0.01
		Alexa 350	0.06±0.01
		AMCA	0.04±0.01
G1z	Dye -CTG AGC TTT AGT YAA GGC AAA TAA TGC TCA G- Dabcy1	PEPy	0.10±0.01
		Alexa 350	0.08±0.01
		AMCA	0.04±0.01
G2z	Dye -CAG CGT CTA GTG ATC CCG TTA TTG GC- Dabcy1	PEPy	0.13±0.01
		Alexa 350	0.12±0.01
		AMCA	0.09±0.01
G3z	Dye -TAC CCA ACT GAA GCA GCA ACA GGG TA- Dabcy1	PEPy	0.16±0.01
		Alexa 350	0.08±0.01
		AMCA	1.5±0.01
G4z	Dye -CTG AAC YTG TCG GCC ATC CTT TGG TT- Dabcy1	PEPy	19.5±0.01
		Alexa 350	24.5±0.01
		AMCA	1.3±0.01