

Supporting Information

One-step green synthesis of polypyrrole-Au nanocomposite and its application in myoglobin aptasensor

Chong Sun^a, Daoying Wang^{a,*}, Zhiming Geng^a, Ling Gao^b, Muhan Zhang^a, Huan Bian^a, Fang Liu^a,
Yongzhi Zhu^a, Haihong Wu^a, Weimin Xu^{a,*}

^a*Institute of Agricultural Products Processing, Jiangsu Academy of Agricultural Sciences, Nanjing 210014, China*

^b*Jiangsu Collaborative Innovation Center of Biomedical Functional Materials, College of Chemistry and Materials Science, Nanjing Normal University, Nanjing 210023, China*

*Corresponding author.

E-mail address: daoyingwang@yahoo.com (D. Wang); weiminxu2002@aliyun.com (W. Xu)

Tel.: +86 25 84390065, Fax: +86 25 84390065

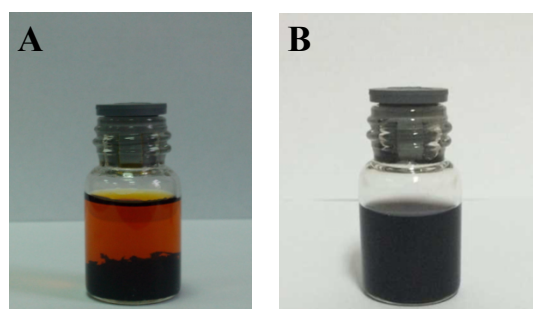


Fig. S1 Photograph of (A) PPy and (B) PPy-Au NC.

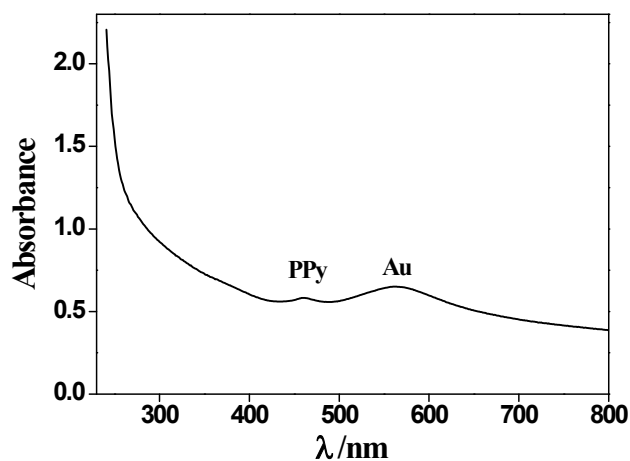


Fig. S2 UV-*vis* adsorption spectrum of PPy-Au NC.

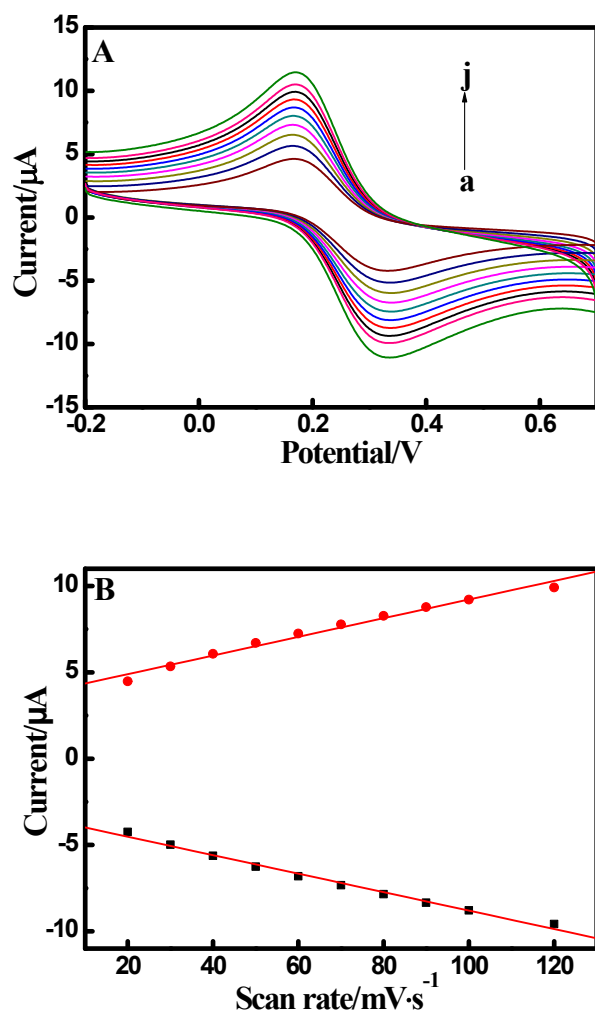


Fig. S3 (A) CVs of MBA/(PPy-Au)/APTES/GCE in 0.1 M PBS (pH=6.0) containing 10 mM $[\text{Fe}(\text{CN})_6]^{3-/4-}$ (1:1) solution and 0.1 M KCl at different scan rates. Scan rate (from a to j): 20, 30, 40, 50, 60, 70, 80, 90, 100, 120 $\text{mV}\cdot\text{s}^{-1}$. (B) Plots of anodic and cathodic peak current vs. scan rate.

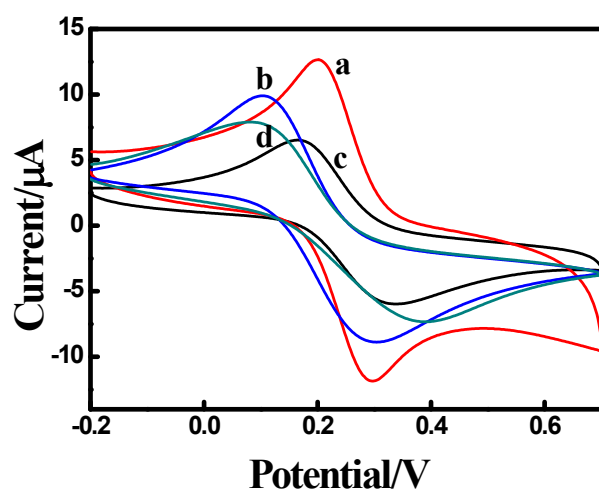


Fig. S4 (A) CVs of (a) MBA/(PPy-Au)/APTES/GCE, (b) MBA/Au/APTES/GCE, (c) Mb/MBA/(PPy-Au)/APTES/GCE and (d) Mb/MBA/Au/APTES/GCE in 0.1 M PBS (pH = 6.0) containing 10 mM $[\text{Fe}(\text{CN})_6]^{3-/4-}$ (1:1) solution and 0.1 M KCl.

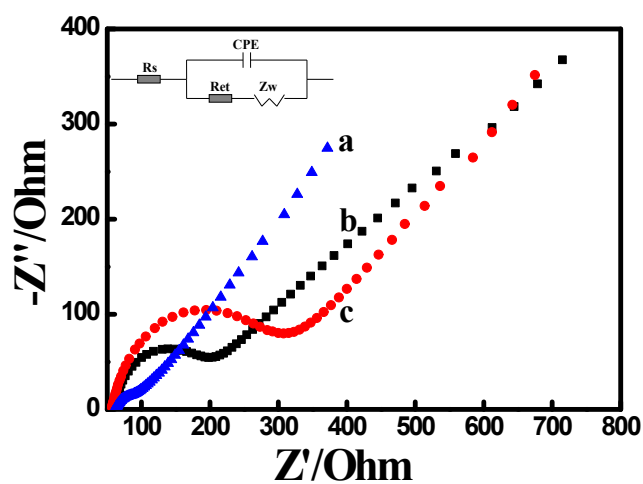


Fig. S5 Electrochemical impedance spectra of (a) (PPy-Au)/APTES/GCE, (b) MBA/(PPy-Au)/APTES/GCE and (c) Mb/MBA/(PPy-Au)/APTES/GCE recorded at the open circuit potential in 10 mM $[\text{Fe}(\text{CN})_6]^{3-/4-}$ (1:1) solution containing 0.1 M KCl. Inset is a schematic of the equivalent circuit.

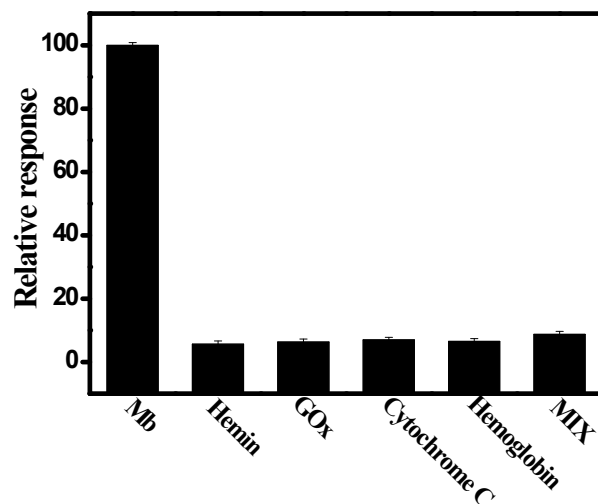


Fig. S6 Specificity of the assay for $0.05 \text{ g}\cdot\text{L}^{-1}$ Mb, four interfering substances and mixture of the four interferences, including hemin, GOx, cytochrome c and hemoglobin were $5 \text{ g}\cdot\text{L}^{-1}$.

Table S1. Comparison of analytical performance of aptasensor and other determination methods

Determination methods	Linear Range	Detection limit	R ²	Ref.
poly(HEMA-MATrp) nanofilm SPR sensor	0.1-1.0 $\mu\text{g}\cdot\text{mL}^{-1}$	87.6 $\text{ng}\cdot\text{mL}^{-1}$	0.98	45
MIP/Au-SPE biosensor	0.852-4.26 $\mu\text{g}\cdot\text{mL}^{-1}$	2.25 $\mu\text{g}\cdot\text{mL}^{-1}$	/	46
TiO ₂ nanotubes sensor	0.001-0.1 $\text{mg}\cdot\text{mL}^{-1}$	1 $\mu\text{g}\cdot\text{mL}^{-1}$	/	47
SPR immunosensor	100-1700 $\text{ng}\cdot\text{mL}^{-1}$	/	>0.98	48
MBA/(PPy-Au)/APTES/GCE	0.0001- 0.15 $\text{g}\cdot\text{L}^{-1}$	30.9 $\text{ng}\cdot\text{mL}^{-1}$	0.9931	This work

/ represents relevant data which were not provided in these references.

Table S2. Comparison of two methods obtained in practical samples.

Samples	muscle-1	muscle-2	muscle-3	muscle-4
Colorimetry ($\mu\text{mol}\cdot\text{g}^{-1}$)	1.59 \pm 0.18	1.40 \pm 0.25	0.25 \pm 0.31	2.12 \pm 0.41
Aptasensors ($\mu\text{mol}\cdot\text{g}^{-1}$)	1.48 \pm 0.02	1.05 \pm 0.08	0.17 \pm 0.05	1.75 \pm 0.03