## Synthesis of Bradyrhizose, a Unique Inositol-fused Monosaccharide Relevant to a Nod-factor Independent Nitrogen Fixation

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General remarks for the synthesis: All reactions were carried out under argon with regular solvents in glassware, unless otherwise noted. The chemicals were reagent grade as supplied. Analytical thin-layer chromatography was performed using silica gel 60 F254 glass plates. Compound spots were visualized by UV light (254 nm) and by heating with a solution with 10% H<sub>2</sub>SO<sub>4</sub> in ethanol. Flash column chromatography was performed on silica gel. NMR spectra were referenced using Me<sub>4</sub>Si (0 ppm), residual CHCl<sub>3</sub> (<sup>1</sup>H NMR  $\delta$  = 7.26 ppm, <sup>13</sup>C NMR  $\delta$  = 77.0 ppm). Peak and coupling constant assignments are based on <sup>1</sup>H NMR, COSY, HSQC, and NOESY. Splitting patterns were indicated as s (singlet), d (doublet), t (triplet), q (quartet), and brs (broad singlet) for <sup>1</sup>H NMR data. ESI-MS and MALDI-MS were run on an IonSpec Ultra instrument using HP5989A or VG Quattro MS. Optical rotations were measured using a Perkin-Elmer 241 polarimeter. [ $\alpha$ ]<sub>D</sub> values are given in 10<sup>-1</sup> deg cm<sup>2</sup> g<sup>-1</sup>.



To a mixture of tri-*O*-acetyl-D-glucal (10.0 g, 36.7 mmol), methyl acrylate (6.60 mL, 73.6 mmol), and Cu(OAc)<sub>2</sub> (6.68 g, 36.8 mmol) in DMA (50 mL) and AcOH (50 mL) was added Pd(OAc)<sub>2</sub> (4.14 g, 18.4 mmol) at 65 °C. After stirring for 2.5 h under O<sub>2</sub>, the mixture was filtered and diluted with ethyl acetate. The mixture was washed with water, saturated NaHCO<sub>3</sub>, and brine, respectively. The organic layer was dried, filtered, and then concentrated. The residue was purified by silica gel column chromatography (petroleum ether/ethyl acetate, 6:1 to 4:1 to 2:1) to provide glycal **2** (9.94 g, 76%) as a yellow syrup: <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  = 7.20 (d, *J* = 15.9 Hz, 1H), 6.98 (s, 1 H), 5.63 (d, *J* = 15.8 Hz, 1H), 5.58 (d, *J* = 2.2 Hz, 1H), 5.15 (t, *J* = 3.4 Hz, 1H), 4.52–4.47 (m, 1H), 4.46–4.42 (m, 1H), 4.18 (dd, *J* = 11.8, 4.1 Hz, 1H), 3.72 (s, 3H), 2.09 (s, 3H), 2.08 (s, 3H), 2.07



To a solution of glycal 2 (9.94 g, 27.9 mmol) in MeOH (50 mL) was added MeONa (430 mg, 7.96 mmol) at RT. After stirring for 4 h, the mixture was neutralized with  $H^+$  resins, and was then filtered and concentrated.

To a suspension of the residue above (3.31 g, 14.4 mmol) in anhydrous THF (150 mL) was added *m*-CPBA (5.97 g, 29.4 mmol) at RT. After 1.5 h, anhydrous MeOH (50 mL) was added, and the stirring continued overnight. The mixture was then concentrated.

To a solution of the residue above in anhydrous acetonitrile (100 mL) and 2,2-dimethoxypropane (9.0 mL, 73.2 mmol) was added *p*-TsOHH<sub>2</sub>O (100 mg, 0.53 mmol) at RT. After 40 min, another portion of *p*-TsOHH<sub>2</sub>O (60 mg, 0.35 mmol) was added and the stirring continued for 3.5 h. The reaction was quenched with TEA (3 mL). The mixture was concentrated and purified by silica gen column chromatography (petroleum ether/ethyl acetate, 1:1) to give ester **3** (4.18 g, 91%) as a colorless syrup:  $[\alpha]_D^{27} = 72.6$  (*c* = 0.9 in CHCl<sub>3</sub>); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta = 7.01$  (d, *J* = 15.7 Hz, 1H), 6.20 (d, *J* = 15.7 Hz, 1H), 4.38 (s, 1H), 3.94–3.88 (m, 2H), 3.87–3.84 (m, 2H), 3.75–3.68 (m, 4H), 3.31 (s, 3H), 3.05 (s, 1H), 3.02–2.95 (br s, 1H), 1.51 (s, 3H), 1.41 ppm (s, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta = 166.8$ , 149.2, 122.2, 103.1, 100.2, 76.6, 71.7, 71.2, 63.7, 62.2, 55.3, 51.8, 29.2, 19.2 ppm; HR-ESI calcd for C<sub>14</sub>H<sub>23</sub>O<sub>8</sub> [M + H]<sup>+</sup> 319.1387; found 319.1385.



To a solution of ester **3** (5.02 g, 15.8 mmol) in anhydrous DCM (90 mL) was added DIBAL-H (1 M in cyclohexane, 50.0 mL, 50.0 mmol) at -70 °C. After 40 min, the mixture was warmed to 0 °C, and H<sub>2</sub>O (2.0 mL), aqueous NaOH (15%, 2.0 mL), and H<sub>2</sub>O (5.0 mL) were added subsequently. The mixture was then moved to RT and stirred for 15 min. Anhydrous MgSO<sub>4</sub> was added, and the stirring was continued for another 15 min. The mixture was then filtered, eluted with DCM and MeOH (10:1), and then concentrated.

To a solution of the residue above and imidazole (1.62 g, 23.8 mmol) in anhydrous DCM (50 mL) was added TBDPSCl (5.0 mL, 19.5 mmol) at RT. The mixture was stirred for 40 min, and then quenched with saturated NaHCO<sub>3</sub>. The stirring was continued for another 5 min. The mixture was diluted with ethyl acetate, and washed with brine. The organic layer was dried, filtered, and

concentrated.

To a solution of the residue above and DMAP (150 mg, 1.23 mmol) in anhydrous DCM (50 mL) were added TEA (3.30 mL, 23.7 mmol) and acetic anhydride (1.80 mL, 19.1 mmol) at RT. After stirring for 2.5 h, the reaction was quenched with saturated NaHCO<sub>3</sub>. The mixture was diluted with ethyl acetate, and was then washed with brine. The organic layer was dried, filtered, and concentrated. The residue was purified by silica gel column chromatography (petroleum ether/ethyl acetate, 6:1 to 4:1) to give **4** (7.40 g, 82%) as a colorless syrup:  $[\alpha]_D^{27} = 39.3$  (*c* = 1.4 in CHCl<sub>3</sub>); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta = 7.67-7.65$  (m, 4H), 7.43–7.37 (m, 6H), 5.97–5.86 (m, 2H), 5.31 (d, *J* = 9.7 Hz, 1H), 4.33 (s, 1H), 4.20 (d, *J* = 4.4 Hz, 2H), 4.10 (t, *J* = 9.5 Hz, 1H), 3.90–3.80 (m, 3H), 3.33 (s, 3H), 2.03 (s, 3H), 1.48 (s, 3H), 1.38 (s, 3H), 1.06 ppm (s, 9H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>)  $\delta = 169.6$ , 135.510, 135.499, 133.62, 133.60, 130.7, 130.3, 129.7, 127.68, 127.67, 104.1, 99.9, 76.4, 72.3, 69.8, 64.4, 63.8, 62.4, 55.2, 29.2, 26.8, 20.8, 19.23, 19.20 ppm; HR-ESI calcd for C<sub>31</sub>H<sub>42</sub>O<sub>8</sub>SiNa [M + Na]<sup>+</sup> 593.2541; found 593.2551.



To a solution of **4** (10.5 g, 18.4 mmol) in MeOH (70 mL) was added Dowex 50WX2 (350 mg) at RT. After stirring for 3 h, the mixture was filtered and concentrated.

To a solution of the residue above, imidazole (1.89 g, 27.8 mmol), and PPh<sub>3</sub> (5.84 g, 22.3 mmol) in anhydrous THF (60 mL) was added I<sub>2</sub> (5.60 g, 22.1 mmol). The mixture was heated to 60 °C and stirred for 1 h. The mixture was diluted with ethyl acetate, washed with saturated Na<sub>2</sub>SO<sub>3</sub> and brine, respectively. The organic layer was dried, filtered, and concentrated. The residue was purified by silica gel column chromatography (petroleum ether/ethyl acetate, 2:1) to give iodide **5** (11.8 g, 100%) as a yellow syrup:  $[\alpha]_D^{26} = 60.7$  (c = 0.3 in CHCl<sub>3</sub>); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta = 7.66-7.64$  (m, 4H), 7.45–7.36 (m, 6H), 5.98 (dt, J = 15.6, 4.6 Hz, 1H), 5.81 (dt, J = 15.6, 1.6 Hz, 1H), 5.17 (d, J = 9.4 Hz, 1H), 4.37 (s, 1H), 4.21 (d, J = 4.0 Hz, 2H), 3.77 (td, J = 9.4, 6.2 Hz, 1H), 3.65 (dd, J = 10.6, 2.4 Hz, 1H), 3.54 (ddd, J = 9.3, 7.0, 2.3 Hz, 1H), 3.44–3.39 (m, 4H), 2.38 (d, J = 6.2 Hz, 1H), 2.18 (s, 1H), 2.08 (s, 3H), 1.05 ppm (s, 9H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta = 172.0$ , 135.5, 133.59, 133.56, 131.5, 129.7, 128.7, 127.7, 103.5, 75.9, 75.6, 71.5, 63.8, 55.7, 26.8, 20.9, 19.2, 7.0 ppm; HR-ESI calcd for C<sub>28</sub>H<sub>37</sub>O<sub>7</sub>ISiNa [M + Na]<sup>+</sup> 663.1245; found 663.1256.



To a solution of iodide 5 (653 mg, 1.02 mmol) in DCM (10 mL) was added Dess-Martin

periodinane (645 mg, 1.52 mmol) at RT. After stirring for 2 h, the reaction was quenched with saturated  $Na_2S_2O_3$  and saturated  $NaHCO_3$ . The stirring continued for another 3 h. The mixture was diluted with ethyl acetate, washed with brine. The organic layer was dried, filtered, and concentrated.

To a solution of the residue above in anhydrous THF (10 mL) was added TEA (0.14 mL, 1.0 mmol) at RT. After 2 h, TLC showed the reactant was consumed. The mixture was then cooled to -70 °C, and MeLi (1 M in E<sub>2</sub>O, 5.0 mL, 5.0 mmol) was added. After stirring for 1.5 h at -70 °C, the reaction was quenched with MeOH. The mixture was diluted with ethyl acetate, washed with water and brine, respectively. The organic layer was dried, filtered, and concentrated. The residue was purified by silica gel column chromatography (petroleum ether/ethyl acetate, 3:1) to provide triol **6** (278 mg, 56%) as a colorless syrup:  $[\alpha]_D^{26} = 40.8$  (c = 0.2 in CHCl<sub>3</sub>); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta = 7.69-7.66$  (m, 4H), 7.45–7.36 (m, 6H), 5.98–5.88 (m, 2H), 4.93 (s, 1H), 4.72 (d, J = 0.9 Hz, 1H), 4.45 (s, 1H), 4.29 (d, J = 3.3 Hz, 2H), 3.78 (s, 1H), 3.41 (s, 3H), 2.21 (s, 1H), 1.54 (s, 3H), 1.06 ppm (s, 9H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta = 160.8$ , 135.57, 135.56, 133.8, 133.6, 130.8, 129.7, 127.7, 104.4, 96.1, 75.6, 75.5, 73.4, 63.9, 55.6, 26.9, 23.7, 19.2 ppm; HR-MALDI calcd for C<sub>27</sub>H<sub>40</sub>O<sub>6</sub>NSi [M + NH<sub>4</sub>]<sup>+</sup> 502.2619; found 502.2611.



To a solution of triol **6** (3.00 g, 6.19 mmol) in anhydrous DMF (30 mL) was added NaH (2.20 g, 55.0 mmol) at 0  $^{\circ}$ C. After 25 min, TBAI (2.29 g, 6.19 mmol) and BnBr (6.80 mL, 57.2 mmol) were added, and the mixture was moved to RT. The mixture was stirred for another 40 min, quenched with MeOH, and diluted with ethyl acetate. The mixture was washed with water and brine, respectively, and was then concentrated.

To a solution of the residue above in THF (40 mL) was added TBAF (1 M in THF, 7.0 mL, 7.0 mmol) at RT. The mixture was stirred overnight and concentrated.

To a solution of the residue above, TEA (0.86 mL, 6.2 mmol), and DMAP (65 mg, 0.53 mmol) in anhydrous DCM (30 mL) was added acetic anhydride (0.58 mL, 6.2 mmol) at RT. After stirring for 1 h, the mixture was diluted with ethyl acetate, and was then washed with saturated NaHCO<sub>3</sub> and brine, respectively. The organic layer was dried, filtered, and concentrated. The residue was purified by silica gel column chromatography (petroleum ether/ethyl acetate, 10:1) to give **7** (2.47 g, 71%) as a yellow syrup:  $[\alpha]_D^{27} = 22.3$  (c = 1.0 in CHCl<sub>3</sub>); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta = 7.36-7.32$  (m, 15H), 5.92–5.91 (m, 2H), 4.87–4.75 (m, 5H), 4.71–4.52 (m, 6H), 4.10 (s, 1H), 3.45 (s, 3H), 1.99 (s, 3H), 1.68 ppm (s, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta = 170.7$ , 158.6, 139.5, 138.9, 138.8, 131.6, 128.3, 128.2, 127.4, 127.31, 127.26, 127.05, 127.97, 102.9, 96.7, 80.0, 79.9, 75.2, 66.3, 64.8, 64.7, 55.9, 23.7, 20.9 ppm; HR-ESI calcd for C<sub>34</sub>H<sub>38</sub>O<sub>7</sub>Na [M + Na]<sup>+</sup> 581.2510; found 581.2523.



To a solution of **7** (2.47 g, 4.42 mmol) in 1,4-dioxane (40 mL) and H<sub>2</sub>O (20 mL) were added Hg(OAc)<sub>2</sub> (3.00 g, 9.41 mmol) and AcOH (0.6 mL) at 60 °C. After 5 min, NaCl (0.26 g, 4.4 mmol) was added. After stirring for 1.5 h, the mixture was diluted with ethyl acetate. After washing with brine, the organic layer was dried, filtered, and concentrated. The residue was purified by silica gel column chromatography (petroleum ether/ethyl acetate, 4:1) to give ketone **8** (2.09 g, 87%) as a colorless syrup:  $[\alpha]_D^{27} = -21.4$  (c = 0.3 in CHCl<sub>3</sub>); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta = 7.45$  (d, J = 7.3 Hz, 2H), 7.34–7.23 (m, 13H), 6.00 (dt, J = 16.3, 5.5 Hz, 1H), 5.92 (d, J = 16.3 Hz, 1H), 5.05 (d, J = 11.6 Hz, 1H), 4.89 (d, J = 12.3 Hz, 1H), 4.72–4.65 (m, 3H), 4.61–4.57 (m, 3H), 4.39 (s, 1H), 4.08 (t, J = 3.4 Hz, 1H), 3.22 (dd, J = 15.0, 3.2 Hz, 1H), 2.52 (dd, J = 15.0, 3.8 Hz, 1H), 2.01 (s, 3H), 1.69 ppm (s, 3H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>)  $\delta = 209.4$ , 170.9, 139.4, 138.8, 138.7, 133.4, 129.3, 128.4, 128.32, 128.31, 127.8, 127.5, 127.44, 127.40, 127.3, 127.3, 86.8, 82.2, 81.2, 75.6, 72.5, 67.9, 66.6, 64.6, 42.1, 20.9, 19.1 ppm; HR-ESI calcd for C<sub>33</sub>H<sub>36</sub>O<sub>7</sub>Na [M + Na]<sup>+</sup> 567.2353; found 567.2342.



To a mixture of  $Me_4NB(OAc)_3$  (5.04 g, 19.2 mmol) in MeCN (50 mL) was added AcOH (2.20 mL, 38.4 mmol) at 0 °C. After 30 min, ketone **8** (2.09 g, 3.84 mmol) in MeCN (20 mL) was added. The mixture was then moved to RT. After stirring for another 2 h, the reaction was quenched with potassium sodium tartrate, and the stirring continued for another 1 h. The mixture was diluted with ethyl acetate and washed with brine. The organic layer was dried, filtered, and concentrated. The residue was employed in the next step without further purification.

To a solution of the residue (2.10 g, 3.84 mmol) in anhydrous pyridine (8 mL) was added BzCl (0.60 mL, 5.17 mmol) dropwise at 0 °C. After stirring for 1 h, the reaction was quenched with H<sub>2</sub>O. The mixture was diluted with ethyl acetate and then washed with brine. The organic layer was dried, filtered, and concentrated. The residue was purified by silica gel column chromatography (petroleum ether/ethyl acetate, 4:1) to give alcohol **9** (1.93 g, 77%) as a colorless syrup:  $[\alpha]_D^{27} = 17.4$  (c = 0.9 in CHCl<sub>3</sub>); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta = 8.04$  (d, J = 7.3 Hz, 2H), 7.54 (t, J = 7.4 Hz, 1H), 7.41 (t, J = 7.7 Hz, 2H), 7.34–7.16 (m, 15H), 5.98 (dt, J = 16.2, 5.4 Hz, 1H), 5.91 (d, J = 16.4 Hz, 1H), 5.75 (dd, J = 10.1, 6.2 Hz, 1H), 5.03 (d, J = 11.6 Hz, 1H), 4.88 (d, J = 12.3 Hz, 1H), 4.73–4.66 (m, 3H), 4.60–4.58 (m, 3H), 4.16 (s, 1H), 3.95 (s, 1H), 2.36 (s, 1H), 2.24–2.17 (m, 2H), 1.99 (s, 3H), 1.82 ppm (s, 3H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>)  $\delta = 170.9$ , 165.6,

139.8, 139.22, 139.16, 134.3, 133.0, 130.5, 129.6, 128.6, 128.4, 128.29, 128.26, 128.19, 127.5, 127.3, 127.19, 127.18, 127.16, 127.1, 82.0, 76.5, 74.1, 67.5, 65.2, 64.7, 31.3, 20.9 ppm; HR-ESI calcd for  $C_{40}H_{42}O_8Na$  [M + Na]<sup>+</sup> 673.2772; found 673.2784.



To a solution of alcohol **9** (1.93 g, 2.96 mmol) in anhydrous DCM (10 mL) was added Dess-Martin periodinane (1.85 g, 4.36 mmol) at RT. The mixture was stirred for 1.5 h, and saturated NaHCO<sub>3</sub> (20 mL) and saturated Na<sub>2</sub>S<sub>2</sub>O<sub>3</sub> (20 mL) were added, and the stirring continued overnight. The mixture was diluted with ethyl acetate and then washed with brine. The organic layer was dried, filtered, and concentrated.

To a mixture of the residue above in DCM (2 mL) and MeOH (18 mL) was added NaBH<sub>4</sub> (0.504 g, 13.3 mmol) at 0 °C. After stirring for 0.5 h, the reaction was quenched with saturated NH<sub>4</sub>Cl, and the stirring continued for another 5 min. The mixture was diluted with ethyl acetate and then washed with brine. The organic layer was dried, filtered, and concentrated.

To a solution of the residue above and 2,6-lutidine (1.00 mL, 8.59 mmol) in anhydrous DCM (15 mL) was added TBSOTf (1.00 mL, 4.35 mmol) at 0 °C. The mixture was moved to RT and stirred for 0.5 h. The reaction was quenched with MeOH. The mixture was diluted with ethyl acetate and then washed with brine. The organic layer was dried, filtered, and concentrated. The residue was purified by silica gel column chromatography (petroleum ether/ethyl acetate, 15:1) to provide **10** (2.01 g, 89%) as a yellow syrup:  $[\alpha]_D^{27} = 9.7$  (c = 0.5 in CHCl<sub>3</sub>); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta = 8.07$  (dd, J = 8.4, 1.4 Hz, 2H), 7.59–7.56 (m, 1H), 7.47–7.45 (m, 2H), 7.35–7.16 (m, 15H), 5.91–5.82 (m, 2H), 5.31 (dd, J = 12.3, 4.6 Hz, 1H), 4.92 (d, J = 13.8 Hz, 1H), 4.89 (d, J = 11.4 Hz, 1H), 4.83 (d, J = 13.8 Hz, 1H), 4.67 (d, J = 11.2 Hz, 1H), 4.60 (d, J = 11.4 Hz, 1H), 4.55 (d, J = 11.2 Hz, 1H), 4.53–4.51 (m, 2H), 3.96 (dd, J = 11.8, 3.9 Hz, 1H), 3.53 (s, 1H), 2.27 (q, J = 12.1 Hz, 1H), 2.11 (dt, J = 12.1, 4.3 Hz, 1H), 1.92 (s, 3H), 1.83 (s, 3H), 0.91 (s, 9H), 0.13 (s, 3H), 0.12 ppm (s, 3H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  = 170.6, 165.6, 141.1, 139.4, 139.1, 133.9, 133.1, 130.4, 129.6, 128.5, 128.15, 128.14, 128.08, 127.3, 127.2, 127.09, 127.07, 127.0, 126.6, 126.1, 86.0, 82.9, 82.3, 76.1, 75.3, 72.6, 67.5, 65.6, 64.5, 33.5, 25.6, 20.7, 18.0, 12.9, -4.0, -5.0 ppm; assignments are labeled in the corresponding NOESY spectrum; HR-ESI calcd for  $C_{46}H_{56}O_8SiNa [M + Na]^+$  787.3637; found 787.3668.



A mixture of AD-mix α (3.22 g, 2.30 mmol), K<sub>2</sub>OsO<sub>4</sub>·H<sub>2</sub>O (18 mg, 0.049 mmol), MeSO<sub>2</sub>NH<sub>2</sub> (223

mg, 2.34 mmol),  $K_2S_2O_8$  (652 mg, 2.41 mmol), (DHQ)<sub>2</sub>PHAL (120 mg, 0.154 mmol) in *t*BuOH (4 mL) and H<sub>2</sub>O (4 mL) was stirred for 0.5 h, and **10** (1.19 g, 1.55 mmol) was then added. After stirring for 24 h, the reaction was quenched with saturated Na<sub>2</sub>S<sub>2</sub>O<sub>3</sub>. The mixture was diluted with ethyl acetate and then washed with brine. The organic layer was dried, filtered, and concentrated. The residue was purified by silica gel column chromatography (petroleum ether/ethyl acetate, 6:1) to give the corresponding diol (1.073 g).

To a solution of the diol (925 mg) above in anhydrous MeOH (20 mL) was added Mg(OMe)<sub>2</sub> (7% in MeOH, 4.5 mL, 2.97 mmol). The mixture was heated at 50 °C for 1 h, and was then neutralized with 5% HCl. The mixture was diluted with ethyl acetate and then washed with brine. The organic layer was dried, filtered, and concentrated. The residue was purified by silica gel column chromatography (petroleum ether/ethyl acetate, 3:1) to give triol **11** (784 mg, 76%) as a colorless syrup:  $[\alpha]_D^{27} = -8.2$  (c = 0.3 in CHCl<sub>3</sub>); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta = 8.04$  (d, J = 8.0 Hz, 2H), 7.57 (t, J = 7.4 Hz, 1H), 7.46–7.14 (m, 17H), 5.25 (dd, J = 12.2, 4.6 Hz, 1H), 5.08–5.04 (m, 2H), 4.96 (d, J = 11.4 Hz, 1H), 4.68–4.61 (m, 3H), 4.55 (d, J = 7.0 Hz, 1H), 4.51–4.48 (m, 2H), 3.73 (s, 1H), 3.71–3.68 (m, 1H), 3.66–3.63 (m, 1H), 3.54–3.49 (m, 1H), 2.86 (d, J = 8.4 Hz, 1H), 2.53 (d, J = 9.1 Hz, 1H), 2.29 (dd, J = 24.1, 12.0 Hz, 1H), 2.10–2.05 (m, 1H), 1.82 (s, 3H), 0.89 (s, 9H), 0.18 (s, 3H), 0.15 ppm (s, 3H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>)  $\delta = 165.5$ , 139.8, 138.7, 138.6, 133.3, 130.2, 129.6, 128.53, 128.52, 128.3, 128.1, 127.7, 127.2, 127.0, 83.1, 82.8, 80.4, 75.7, 75.2, 71.7, 71.5, 69.5, 66.5, 66.3, 65.6, 33.2, 25.8, 18.0, 12.5, -3.6, -4.5 ppm; HR-ESI calcd for C<sub>44</sub>H<sub>56</sub>O<sub>9</sub>SiNa [M + Na]<sup>+</sup> 779.3586; found 779.3568.



To a solution of triol **11** (750 mg, 0.99 mmol) in anhydrous DCM (10 mL) were added trichloroisocyanuric acid (604 mg, 2.60 mmol) and TEMPO (1.8 mg, 0.012 mmol) at  $-10^{\circ}$ C. After stirring for 2 h, the reaction was quenched with saturated Na<sub>2</sub>S<sub>2</sub>O<sub>3</sub>. The mixture was diluted with ethyl acetate and then washed with brine. The organic layer was dried, filtered, and concentrated.

To a solution of the residue above in THF (10 mL) was added TBAF (1 M in THF, 1.50 mL, 1.50 mmol) at RT. After stirring for 1 h, the mixture was acidified by 5% HCl. The mixture was diluted with ethyl acetate and then washed with brine. The organic layer was dried, filtered, and concentrated. The residue was purified by silica gel column chromatography (petroleum ether/ethyl acetate, 4:1 to 2:1 to 1:1) to give **12** (378 mg, 60%) as a white foam: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  = 8.04–8.02 (m, 4H), 7.58–7.54 (m, 2H), 7.45–7.15(m, 34H), 5.53 (d, *J* = 12.1 Hz, 1H), 5.45 (d, *J* = 11.7 Hz, 1H), 5.33–5.26 (m, 3H), 5.21 (d, *J* = 12.1 Hz, 1H), 5.16 (d, *J* = 11.7 Hz, 1H), 5.07 (dd, *J* = 10.9, 4.4 Hz, 2H), 4.72–4.60 (m, 5H), 4.51–4.46 (m, 2H), 4.19–4.09 (m, 4H), 4.04–3.97 (m, 2H), 3.86–3.78 (m, 3H), 3.75 (s, 1H), 3.54 (s, 1H), 3.40 (dd, *J* = 11.6, 4.0 Hz, 1H), 3.09 (s, 1H), 2.57 (d, *J* = 6.5 Hz, 1H), 2.30–2.02 (m, 4H), 1.79 (s, 3H), 1.77 ppm (s, 3H); <sup>13</sup>C

NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  = 165.53, 165.48, 139.6, 139.4, 138.6, 138.5, 137.7, 137.5, 133.34, 133.31, 130.0, 129.9, 129.6, 128.76, 128.75, 128.53, 128.52, 128.32, 128.30, 128.28, 128.25, 128.23, 128.15, 128.09, 128.0, 127.8, 127.7, 127.5, 127.4, 127.1, 127.03, 127.00, 126.8, 97.6, 92.6, 88.7, 88.6, 82.5, 82.3, 79.7, 76.9, 76.6, 76.3, 76.2, 75.9, 75.71, 75.67, 73.4, 72.2, 69.7, 68.98, 68.95, 67.2, 65.7, 65.6, 29.5, 29.4, 12.34, 12.31 ppm; HR-ESI calcd for  $C_{38}H_{40}O_9Na [M + Na]^+$  663.2565; found 663.2533.



To a solution of 12 (33 mg, 0.061 mmol), TEA (0.10 mL, 0.72 mmol), and DMAP (2 mg, 0.016 mmol) in anhydrous DCM (2 mL) was added BzCl (50 µl, 0.43 mmol) at RT. The mixture was stirred for 2 h, and then another portion of DMAP (32 mg, 0.26 mmol) was added. The stirring continued overnight, and the reaction was quenched with saturated NaHCO<sub>3</sub>. The mixture was diluted with ethyl acetate, and then washed with brine. The organic layer was dried, filtered, and concentrated. The residue was purified by silica gel column chromatography (petroleum ether/ethyl acetate, 5:1) to provide S1 as a white foam (35 mg, 67%): <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  8.10 (d, J = 8.3 Hz, 2H), 8.04 (d, J = 8.3 Hz, 2H), 7.93 (d, J = 8.3 Hz, 2H), 7.67–7.17 (m, 24H), 6.68 (d, J = 3.9 Hz, 1H), 5.87 (dd, J = 10.3, 4.0 Hz, 1H), 5.68 (d, J = 11.5 Hz, 1H), 5.38 (dd, J = 12.2, 4.9 Hz, 1H), 5.34 (d, J = 11.5 Hz, 1H), 5.21 (d, J = 11.0 Hz, 1H), 4.73 (d, J = 11.0 Hz, 1H), 4.69 (d, J = 10.4 Hz, 1H), 4.65 (d, J = 10.4 Hz, 1H), 4.49 (d, J = 10.4 Hz, 1H), 4.33 (s, 1H), 4.07 (dd, J = 12.2, 3.7 Hz, 1H), 4.01 (s, 1H), 2.32 (q, J = 12.1 Hz, 1H), 2.14 (dd, J = 7.6, 4.4 Hz, 1H), 1.87 ppm (s, 4H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>) δ 165.9, 165.4, 164.7, 139.0, 138.3, 137.3, 133.7, 133.4, 133.2, 129.9, 129.9, 129.8, 129.7, 129.5, 129.4, 128.8, 128.7, 128.6, 128.4, 128.3, 128.2, 128.0, 127.8, 127.6, 127.3, 127.2, 90.9, 88.8, 82.5, 76.7, 76.3, 75.6, 75.3, 69.9, 69.8, 69.6, 65.9, 29.3, 12.2 ppm; assignments are labeled in the corresponding NOESY spectrum.



To a solution of triol **11** (81 mg, 0.11 mmol) in THF (2 mL) was added TBAF (1 M in THF, 0.15 mL, 0.15 mmol) at RT. After stirring for 0.5 h, the mixture was diluted with ethyl acetate, and was then washed with 5% HCl and brine, respectively. The organic layer was dried, filtered, and concentrated. The residue was purified by silica gel column chromatography (petroleum ether/ethyl acetate, 1:1, then DCM/MeOH, 20:1) to give the corresponding tetraol (66 mg, 96%).

To a mixture of the tetraol (64 mg, 0.10 mmol) and trichloroisocyanuric acid (63 mg, 0.27 mmol) in anhydrous DCM (3 mL) was added TEMPO (0.2 mg, 1.28  $\mu$ mol) at 0 °C. After stirring for 1 h, the reaction was then quenched with saturated Na<sub>2</sub>S<sub>2</sub>O<sub>3</sub>. The mixture was diluted with

ethyl acetate and then washed with brine. The organic layer was dried, filtered, and concentrated.

To a solution of the residue above in anhydrous DCM (2 mL) was added DIBAL-H (1 M in cyclohexane, 0.50 mL, 0.50 mmol) at -70 °C. After stirring for 40 min, the reaction was quenched with H<sub>2</sub>O. The mixture was diluted with ethyl acetate. NaHCO<sub>3</sub> (0.7 g) and Na<sub>2</sub>SO<sub>4</sub> (0.7 g) were then added, and the stirring continued for another 2 h. The mixture was filtered (eluted with ethyl acetate) and concentrated. The residue was purified by silica gel column chromatography (petroleum ether/ethyl acetate, 1:2, then DCM/MeOH, 20:1) to give **13** (33 mg, 84%) as a white foam: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  = 7.35–7.21 (m, 24H), 5.48 (d, *J* = 12.1 Hz, 0.6H), 5.41 (d, *J* = 11.8 Hz, 1H), 5.29 (t, *J* = 2.8 Hz, 0.6H), 5.17 (d, *J* = 12.1 Hz, 0.6H), 5.12 (d, *J* = 11.8 Hz, 1H), 5.07–5.03 (m, 1.6H), 4.81–4.77 (m, 1.6H), 4.74–4.69 (m, 3H), 4.56 (t, *J* = 6.1 Hz, 1H), 4.15 (s, 1H), 4.13–4.07 (m, 2H), 4.01–3.95 (m, 0.6H), 3.88–3.73 (m, 4.6H), 3.62 (s, 1H), 3.46 (d, *J* = 2.4 Hz, 0.6H), 3.24 (dd, *J* = 12.0, 3.7 Hz, 1H), 3.05 (s, 1H), 2.58 (d, *J* = 6.4 Hz, 0.6H), 2.10 (p, *J* = 12.0 Hz, 1.6H), 1.92 (dt, *J* = 12.0, 4.3 Hz, 1H), 1.86–1.81 (m, 2H), 1.53 (s, 1.8H), 1.51 (s, 3H).



To a solution of **13** (50 mg, 0.093 mmol) in MeOH (3 mL) was added 10% Pd/C (202 mg, 0.19 mmol) at RT. The mixture was stirred for 2 days under H<sub>2</sub> atmosphere, and was then filtered and concentrated. The residue was purified by RP-18 column chromatography (H<sub>2</sub>O) to give bradyrhizose **1** (25 mg, 100%) as a white foam:  $[\alpha]_D^{25} = 6.5$  (c = 0.2 in H<sub>2</sub>O); HR-ESI calcd for C<sub>10</sub>H<sub>18</sub>O<sub>8</sub>Cl [M + Cl]<sup>-</sup> 301.0696; found 301.0694.

## General remarks for the NMR spectroscopy of bradyrhizose

All 1D and 2D <sup>1</sup>H-NMR spectra were recorded on a solution of 10 mg bradyrhizose (1) in 0.5 mL of D<sub>2</sub>O, TDE-d3, DMSO-d6 on Bruker 600 DRX equipped with a cryo probe. Spectra were calibrated with internal acetone [ $\delta_{H}$  2.225,  $\delta_{C}$  31.45]. 2D-DQF COSY spectra were acquired with 4096×1024 data points in both  $F_2$  and  $F_1$  dimensions. Quadrature indirect dimensions are achieved through States-TPPI method; spectra are processed applying a Qsine function to both dimensions and data matrix was zero-filled by factor of 2 before Fourier transformation. Coupling constants were determined on a first order basis from 2D phase sensitive DQF-COSY.<sup>[S2-S3]</sup> Nuclear Overhauser enhancement spectroscopy (NOESY) are measured using data sets ( $t_2 \times t_1$ ) of 4096 × 512 points, mixing times of 400-600 ms are used. Total correlation spectroscopy experiments (TOCSY) were performed with a spinlock time of 100 ms, using data sets ( $t_2 \times t_1$ ) of 4096 × 512 points. In homonuclear experiments the data matrix is zero-filled in the *F1* dimension to give a matrix of 4096 x 2048 points and is resolution enhanced in both dimensions by a 90° shifted Qsine function before Fourier transformation. Heteronuclear single quantum coherence (HSQC),

HSQC-NOESY (mixing time 500 ms), and heteronuclear multiple bond correlation (HMBC) experiments were measured in the <sup>1</sup>H-detected mode via single quantum coherence with proton decoupling in the <sup>13</sup>C domain, using data sets of 2048 x 256 points. Experiments were carried out in the phase-sensitive mode according to the method of States *et al.*<sup>[S4]</sup> HMBC experiment was optimized for 6 Hz coupling constant. In all heteronuclear experiments, the data matrix was extended to 2048 x 1024 points using forward linear prediction extrapolation.

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Figure SA. The isomeric equilibrium mixture of bradyrhizose as detected by NMR.



Figure SB: <sup>1</sup>H NMR spectrum of bradyrhizose in D<sub>2</sub>O





Figure SC: HSQC NMR spectrum of bradyrhizose in D<sub>2</sub>O. Cross peaks are labelled as indicated in Table S1

Figure SD: HSQC (blue) and HMBC (red) NMR spectra of bradyrhizose in D<sub>2</sub>O; key long range scalar correlations are shown. Cross peaks are labelled as indicated in Table S1



Figure SE: HSQC (blue) and HSQC-NOESY (green) NMR spectra of bradyrhizose in D<sub>2</sub>O; key NOE correlations are shown. Cross peaks are labelled as indicated in Table S1



93.15 <sup>1</sup> Ј <sub>С,Н</sub> = 163
' <i>J</i> <sub>C,H</sub> = 163
70.3
68.2
72
67.17
04.4
31.1
69.1 (64.7)
75.9
76.9
14.39

Table S1. Isomer distribution of reducing bradyrhizose. <sup>1</sup>H and <sup>13</sup>C chemical shifts (ppm) in D<sub>2</sub>O, coupling constants <sup>3</sup> $J_{H,H}$  and <sup>1</sup> $J_{C,H}$  (Hz).  $\beta$  and  $\alpha$  anomers, pyranose form (A-B), furanose forms (C-D) and alternative ring closure (E) (% is a rough estimation)

Figure SF1-2: <sup>1</sup>H NMR spectrum of bradyrhizose in DMSO and its zoom in the anomeric region. Signals are labelled as indicated in Table S2







Figure SG: HSQC NMR spectrum of bradyrhizose in DMSO. Cross peaks are labelled as indicated in Table S2

Table S2. Isomer distribution of reducing bradyrhizose. <sup>1</sup>H and <sup>13</sup>C chemical shifts (ppm) in DMSO, coupling constants <sup>3</sup> $J_{H,H}$  and <sup>1</sup> $J_{C,H}$  (*Hz*).  $\beta$  and  $\alpha$  anomers, pyranose form (A-B), furanose forms (C-D) and alternative ring closure (E) (% is a rough estimation)

	<sup>1</sup> H and <sup>13</sup> C β-anomer <mark>A residue</mark> (54.1%)		β-anomer ${}^{1}$ H and ${}^{13}$ C α-anomer (54.1%) B residue (22.6%)		<sup>1</sup> H and <sup>13</sup> C β-anomer <mark>C</mark> residue (8%)		<sup>1</sup> H and <sup>13</sup> C α-anomer D residue (3.5%)		<sup>1</sup> H and <sup>13</sup> C β-anomer E residue (15.7%)	
1	4.26 <sup>3</sup> J <sub>H,H</sub> = 7.4 <sup>1</sup> J <sub>C,H</sub> = 156	97.87	4.865	93.08	4.77 <sup>3</sup> J <sub>H,H</sub> = 3.64 <sup>1</sup> J <sub>C,H</sub> = 178	101.85	4.95	93.99	4.73 <sup>1</sup> J <sub>C,H</sub> = 160	93.6
2	2.21	72.87	3.504	69.6	3.88	82.5	4.00	76.53	3.42	71.78
3	3.37	79.19	3.605	75.4	4.07	76.9	4.404	75.58	3.66	68.3
4		72.23		73.06		87.15		86.38		73.5
5	3.19	70.59	3.646	65.6	3.22	66.0	3.24	65.8	3.53	64.9
6 <sub>ax</sub> 6 <sub>eq</sub>	1.71 1.55	32.0	1.45 1.67	32.3	1.68 1.45	37.3	1.39 1.68	37.2	1.58 1.58	3.59
7	3.32	72.6	3.32	73.0	3.43	71.88	3.53	71.58	3.21	72.8
8		76.8		75.5		76.9		76.9		75.6
9	3.29	79.7	3.23	77.5	3.32	72.48	3.32	72.77	3.24	77.47
10	1.09	16.1	1.11	16.5	1.00	15.9	0.97	15.7	1.10	16.29

Figure SH: <sup>1</sup>H NMR spectrum of bradyrhizose in TFE. Relative abundances: anomer A ( $\beta$ , 53.5%), anomer B ( $\alpha$ , 25.1%), anomer C ( $\beta$ , 5.6%), anomer D ( $\alpha$ , 5.0 %), anomer E ( $\beta$ , 10.7%)





























































