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Catalytic Asymmetric Hydrogenation of Quinoline Carbocycles: Unusual Chemoselectivity in Hydrogenation of Quinolines

Ryoichi Kuwano,*,†,‡ Ryuhei Ikeda,† and Kazuki Hirasada†

[†]Department of Chemistry, Graduate School of Sciences, and International Research Center for Molecular Systems (IRCMS), Kyushu University, 6-10-1 Hakozaki, Higashi-ku, Fukuoka 812-8581, Japan

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[‡]JST ACT-C, 6-10-1 Hakozaki, Higashi-ku, Fukuoka 812-8581, Japan

General and Materials. All NMR spectra were measured with Bruker AVANCE 400 or AVANCE III HD 400 Nanobay (9.4 T magnet) spectrometer at ambient temperature. In ¹H NMR spectra, chemical shifts (ppm) referenced to internal tetramethylsilane (0.00 ppm in CDCl₃). In ¹³C NMR spectra, chemical shifts (ppm) referenced to the carbon signal of the deuterated solvents (77.0 ppm in CDCl₃). IR spectra and melting points were measured with JASCO FT/IR-4100 and Büchi Melting Point B-545, respectively. Elemental analyses and high-resolution mass spectra (FAB) were performed by Service Centre of Elementary Analysis of Organic Compounds and Network Joint Research Center for Materials and Devices (Institute for Materials Chemistry and Engineering, Kyushu University), respectively. Column chromatographies were performed with silica gel 60 (230–400 mesh, Merck).

Ethyl acetate (EtOAc) was dried with phosphorus pentoxide. 2-Propanol (i-PrOH), triethylamine (Et₃N), and 1,8-diazabicyclo[5,4,0]undec-7-ene (DBU) were dried with calcium hydride. These solvents and reagents were distilled under nitrogen atmosphere. Dry N,N-dimethylformamide (DMF, Aldrich) Toluene (Guaranteed Reagent) and tetrahydrofuran (THF) (HPLC grade, without inhibitor) were deoxidized by purging with nitrogen for 30 min and dried with an alumina and copper (GlassContour $Ru(\eta^3$ -methallyl)₂(cod),¹ (S,S)-(R,R)-PhTRAP column system Co.). $(2k)^{4}$ $\{\text{Ru}(p\text{-cvmene})[(S,S)\text{-}(R,R)\text{-PhTRAP}]\}\text{Cl}^3$ 8-phenylquinoline 8-(trifluoromethanesulfoxy)quinoline, and 8-phenyl-5,6,7,8-tetrahydroquinolin-8-ol were prepared 2-Phenylquinoline (2a), methyl quinoline-6-carboxylate (2b), according to literature procedures. 6-isopropylquinoline (2c), 3-methoxyquinoline (2d), 4-methoxyquinoline (2e), 5-methoxyquinoline (2f), 6-methoxyquinoline (2g), 7-methoxyquinoline (2h), 8-methoxyquinoline (2i), 8-methylquinoline (2o), 8-hydroxyquinoline, dry sodium hydride, (triisopropylsilyloxy)methyl chloride, 4-methoxyphenylboronic tetrakis(triphenylphosphine)palladium(0) acid, lithium chloride, $[Pd(PPh_3)_4],$ 4-(trifluoromethyl)phenylboronic acid, 2-methylphenylboronic acid, sodium carbonate (Na₂CO₃), palladium(II) acetate [Pd(OAc)₂], 1,1'-bis(diphenylphosphino)ferrocene (DPPF), 0.5 M solution of cyclohexylzinc bromide in THF, 1.0 M solution of tetrabutylammonium fluoride (TBAF) in THF, acetic acid (AcOH), and methanesulfonic acid (MsOH) were purchased and used without further purification.

Preparation of Substrates 2

8-[(Triisopropylsilyloxy)methoxy|quinoline (2j).

8-Hydroxyquinoline (726 mg, 5.0 mmol) was placed in a three-neck flask, which was equipped with a stirring bar, rubber septum, three-way stopcock, and dropping funnel having a rubber septum. In the dropping funnel, dry sodium hydride (138 mg, 5.8 mmol) and a stirring bar were placed. After the reaction vessel was evacuated and charged with nitrogen gas three times, dry DMF (12 mL \times 2) was added into the flask and dropping funnel. After the flask was immersed in an ice-bath, the suspension of NaH was added dropwise to the solution of 8-hydroxyquinoline at 0°C for 20 min. To the resulting

solution was added (triisopropylsilyloxy)methyl chloride (1.4 mL, d 0.96 g/mL, 6.0 mmol) with a syringe through the septum. The reaction mixture was stirred at 0°C for 5 min and at ambient temperature for 12 h. After water was added, the resulting mixture was extracted three times with Et₂O. The combined organic layer was dried over Na₂SO₄, and then evaporated under reduced pressure after filtration. The residue was purified with a flash column chromatography on silica gel (EtOAc/hexane = 1/3) to give 2j (1.40 g, 84%) as colorless oil: ¹H NMR (400 MHz, CDCl₃, TMS) δ 1.07 (d, J = 7.3 Hz, 18H), 1.20 (septet, J = 7.3 Hz, 3H), 5.75 (s, 2H), 7.38 (dd, J = 4.2, 8.3 Hz, 1H), 7.40–7.49 (m, 2H), 7.52 (dd, J = 1.7, 7.5 Hz, 1H), 8.10 (dd, J = 1.5, 8.1 Hz, 1H), 8.93 (dd, J = 1.7, 4.3 Hz, 1H); ¹³C {¹H} NMR (100 MHz, CDCl₃) δ 11.8, 17.7, 88.6, 112.3, 120.4, 121.3, 126.6, 129.3, 135.7, 140.3, 149.2, 153.6; IR (neat) 2946, 2867, 1496, 1467, 1378, 1250, 1158, 1064, 984, 789, 755 cm⁻¹; Anal. Calcd for C₁₉H₂₉NO₂Si: C, 68.83; H, 8.82; N, 4.22. Found: C, 68.70; H, 8.84; N, 4.09.

8-(4-Methoxyphenyl)quinoline [57479-35-3] (21).6

8-(Trifluoromethanesulfoxy)quinoline (1.39 g, 5.0 mmol), 4-methoxyphenylboronic acid (790 mg, 5.2 mmol), lithium chloride (636 mg, 15 mmol), and Pd(PPh₃)₄ (289 mg, 0.25 mmol) were placed in a 50 mL two-neck flask, which was equipped with a stirring bar, rubber septum, and Dimroth condenser having a three-way stopcock. After the reaction vessel was evacuated and charged with nitrogen gas three times, dry toluene (18 mL) and freshly prepared 1 M aqueous solution of Na₂CO₃ (3.2 mL, 3.2 mmol) were added into the flask, and then the resulting mixture was stirred under reflux for 60 h. After cooled to ambient temperature, the mixture was washed with water. The aqueous layer was extracted three times with EtOAc. The combined organic layer was dried over Na₂SO₄, and then evaporated under reduced pressure after filtration. The residue was purified with a flash column chromatography on silica gel (EtOAc/hexane = 1/20) to give **21** (892 mg, 76%) as a colorless solid: ¹H NMR (400 MHz, CDCl₃, TMS) δ 3.87 (s, 3H), 7.04 (d, J = 8.7 Hz, 2H), 7.39 (dd, J = 4.2, 8.3 Hz, 1H), 7.57 (t, J = 7.6 Hz, 1H), 7.66 (d, J = 8.7 Hz, 2H), 7.71 (dd, J = 1.4, 7.1 Hz, 1H), 7.78 (dd, J = 1.3, 8.1 Hz, 1H), 8.17 (dd, J = 1.7, 8.2 Hz, 1H), 8.95 (dd, J = 1.7, 4.2 Hz, 1H); ¹³C {¹H} NMR (100 MHz, CDCl₃) δ 55.3, 113.5, 120.9, 126.3, 127.0, 128.8, 129.9, 131.7, 131.9, 136.2, 140.5, 146.1, 150.1, 159.0.

8-[4-(Trifluoromethyl)phenyl]quinoline [1275591-20-2] (2m).

The procedure for preparing **21** was followed with use of 4-(trifluoromethyl)phenylboronic acid (985 mg, 5.2 mmol). The reaction mixture was stirred under reflux for 32 h. The crude product was purified with a flash column chromatography on silica gel (EtOAc/hexane = 1/20) to give **2m** (508 mg,

37%) as a colorless solid: ¹H NMR (400 MHz, CDCl₃, TMS) δ 7.42 (dd, J = 4.2, 8.3 Hz, 1H), 7.61 (t, J = 7.6 Hz, 1H), 7.70–7.77 (m, 3H), 7.81 (d, J = 8.2 Hz, 2H), 7.86 (dd, J = 1.2, 8.1 Hz, 1H), 8.21 (dd, J = 1.6, 8.3 Hz, 1H), 8.94 (dd, J = 1.7, 4.2 Hz, 1H); ¹³C {¹H} NMR (100 MHz, CDCl₃) δ 121.2, 124.4 (q, J = 272 Hz), 124.9 (q, J = 4 Hz), 126.3, 128.3, 128.7, 129.3 (q, J = 32 Hz), 130.4, 130.9, 136.3, 139.4, 143.2, 145.7, 150.5.

8-(2-Methylphenyl)quinoline [57479-11-5] (2n).8

The procedure for preparing **21** was followed with use of 2-methylphenylboronic acid (714 mg, 5.3 mmol). The reaction mixture was stirred under reflux for 22 h. The crude product was purified with a flash column chromatography on silica gel (EtOAc/hexane = 1/5) to give **2n** (972 mg, 89%) as a colorless solid: 1 H NMR (400 MHz, CDCl₃, TMS) δ 2.04 (s, 3H), 7.27–7.36 (m, 5H), 7.53–7.59 (m, 2H), 7.80 (dd, J = 3.0, 6.6 Hz, 1H), 8.14 (dd, J = 1.6, 8.2 Hz, 1H), 8.89 (dd, J = 1.7, 4.2 Hz, 1H); 13 C { 1 H} NMR (100 MHz, CDCl₃) δ 20.4, 120.8, 125.3, 126.0, 127.4, 127.6, 128.3, 129.7, 130.1, 130.3, 136.0, 136.8, 139.8, 141.4, 146.5, 150.3.

8-Cyclohexylquinoline [1539656-77-3] (2p).

In a nitrogen-filled drybox, Pd(OAc)₂ (112 mg, 0.50 mmol) and DPPF (306 mg, 0.55 mmol) were placed in a 5 mL screw-capped vial equipped with a stirring bar. After dry THF (5.0 mL) was added, the vial was sealed with a screw cap containing a PTFE/silicone septum and then removed from the drybox. The solution temperature 10 was stirred ambient for min. 8-(Trifluoromethanesulfoxy)quinoline (1.39 g, 5.0 mmol) was placed in a 100 mL two-neck flask, which was equipped with a stirring bar, rubber septum, and Dimroth condenser having a three-way stopcock. After the reaction vessel was evacuated and charged with nitrogen gas three times, the catalyst solution prepared above was transferred through a cannula into the flask, and a 0.5 M solution of cyclohexylzinc bromide in THF (55 mL, 28 mmol) was added with a syringe to the solution. The mixture was stirred under reflux for 60 h. After cooled to ambient temperature, the mixture was washed with water. The aqueous layer was extracted three times with EtOAc. The combined organic layer was dried over Na₂SO₄, and then evaporated under reduced pressure after filtration. The residue was purified with a flash column chromatography on silica gel (EtOAc/hexane = 1/20) to give 2p (837 mg, 79%) as pale yellow oil: ¹H NMR (400 MHz, CDCl₃, TMS) δ 1.26–1.39 (m, 1H), 1.45–1.79 (m, 4H), 1.78–1.92 (m, 3H), 1.97–2.05 (m, 2H), 4.04 (tt, J = 3.3, 11.8 Hz, 1H), 7.33 (dd, J = 4.1, 8.2 Hz, 1H), 7.48 (t, J = 7.6 Hz, 1H), 7.58 (d, J = 7.2 Hz, 1H), 7.61 (d, J = 8.0 Hz, 1H), 8.08 (dd, J = 1.6, 8.2 Hz, 1H), 8.93 (dd, J = 1.7,

4.1 Hz, 1H); 13 C { 1 H} NMR (100 MHz, CDCl₃) δ 26.5, 27.0, 34.1, 37.1, 120.6, 125.4, 125.7, 126.4, 128.3, 136.3, 146.1, 146.6, 149.1; IR (neat) 2925, 2849, 1496, 1447, 826, 795 cm $^{-1}$; Anal. Calcd for $C_{15}H_{17}N$: C, 85.26; H, 8.11; N, 6.63. Found: C, 85.32; H, 8.06; N, 6.55.

Catalytic Asymmetric Hydrogenation of Quinolines 2

Procedure A: General Procedure for the Optimization of the Hydrogenation of 2b (Table 1). $Ru(\eta^3$ -methallyl)₂(cod) (1.6 mg, 50 μ mol) and (S,S)-(R,R)-PhTRAP (1) (4.4 mg, 5.5 μ mol) were placed in a 2.0 mL Schlenk tube, which was equipped with a stirring bar, rubber septum and three-way stopcock. After the reaction vessel was evacuated and charged with nitrogen gas three times, a solvent (1.0 mL) and a base (50 μ mol, if liquid) were added into the Schlenk tube through the septum by using a syringe. catalyst solution was stirred for 10 min at ambient temperature. Methyl quinoline-6-carboxylate (2b) (93.6 mg, 0.50 mmol), a base (50 μ mol, if solid), and a stirring bar were placed in a 50 mL test tube, which was sealed with a rubber septum, and then the tube was evacuated and charged with nitrogen gas The catalyst solution was transferred through a cannula into the test tube. After the septum was removed, the test tube was quickly inserted into the nitrogen-purged stainless autoclave, and then the autoclave was sealed immediately. Hydrogen gas was introduced into the autoclave until the pressure gauge indicated over 5.0 MPa, and then the pressure was carefully released to 0.1 MPa. This procedure was repeated twice, and finally the inside of the autoclave was pressurized with hydrogen to The mixture was vigorously stirred at 80°C for 24 h. The autoclave was cooled to room temperature, and then excess hydrogen gas was released carefully. The reaction mixture was evaporated under reduced pressure. The residue was analyzed with ¹H NMR in order to determine its composition, and then purified with a flash column chromatography (EtOAc/hexane = 2/1) on silica gel to give the desired 5,6,7,8-tetrahydroquinoline-6-carboxylate **3b**. Results of the selected experiments are shown in Tables 1 and S-1.

Table S-1. Effect of ligand on the ruthenium-catalyzed hydrogenation of **2b**.

$$\begin{array}{c} \text{MeO}_2\text{C} \\ \text{N} \\ & \begin{array}{c} \text{Ru} \, (\eta^3\text{-methallyl})_2(\text{cod}) \, (1.0\%) \\ \text{ligand} \, (1.1\% \, \text{or} \, 2.2\%), \, \text{DBU} \, (10\%) \\ \hline \\ \text{H}_2 \, (5.0 \, \text{MPa}), \, \textit{i-PrOH}, \, 80^{\circ}\text{C}, \, 24 \, \text{h} \\ \\ \text{2b} \\ \end{array} \\ \begin{array}{c} \text{MeO}_2\text{C} \\ \text{N} \\ \end{array} \\ \begin{array}{c} \text{MeO}_2\text{C} \\ \text{N} \\ \end{array} \\ \begin{array}{c} \text{MeO}_2\text{C} \\ \text{N} \\ \end{array} \\ \begin{array}{c} \text{N} \\ \text{N} \\ \text{H} \\ \end{array}$$

entry	ligand	conv. (%) ^{b,c}	$3b:4b^b$
1	DPPE	>99	1:99
2	DPPP	>99	2:98
3	DPPB	>99	3:97
4	DPPPent	>99	5:95
5	DPPBz	>99	1:99
6	DPEphos	58	3:97
7	Xantphos	>99	13:87

8	DPPF	64	19:81
9^d	$P(c ext{-Hex})_3$	49	33:67
10^d	PPh_3	>99	34:66
11	Josiphos L1	>99	4:96
12	(R)- (S) -BPPFA	>99	4:96
13	(S,S)-DIOP	>99	3:97
14	(S,S)-Chiraphos	72	2:98
15	(R)-BINAP	68	4:96
16	(R,R)-Me-DuPHOS	>99	2:98
16 ^d	Monophos L2	>99	2:98
17^d	(R)-MeO-MOP	96	25:75 ^e
18	(S,S)- (R,R) -PhTRAP (1)	>99	85:15 ^f

^a Reactions were conducted on a 0.50 mmol scale in 1.0 mL of *i*-PrOH under 5.0 MPa of H₂ at 80°C for 24 h. The ratio of **2b**:Ru(η^3 -methallyl)₂(cod):ligand:DBU was 100:1.0:1.1:10 unless otherwise noted. ^b Determined by ¹H NMR analysis. ^c A small amount of isopropyl ester was formed. ^d The ratio of Ru(η^3 -methallyl)₂(cod):ligand was 1.0:2.2. ^e **3b** was obtained with 25% ee. ^f **3b** was obtained with 35% ee.

Procedure B: General Procedure for the Catalytic Asymmetric Hydrogenation of Quinolines 2 (Tables 2 and 3). Ru(η^3 -methallyl)₂(cod) (1.6 mg, 50 μ mol) and (S,S)-(R,R)-PhTRAP (1) (4.4 mg, 5.5 μ mol) were placed in a 2.0 mL Schlenk tube, which was equipped with a stirring bar, rubber septum and three-way stopcock. After the reaction vessel was evacuated and charged with nitrogen gas three times, dry *i*-PrOH or EtOAc (1.0 mL) was added into the Schlenk tube through the septum by using a syringe.

The catalyst solution was stirred for 10 min at ambient temperature. A quinoline 2 (0.25 mmol), K_2CO_3 (6.9 mg, 50 μ mol), and a stirring bar were placed in a 50 mL test tube, which was sealed with a rubber septum, and then the tube was evacuated and charged with nitrogen gas three times. The catalyst solution was transferred through a cannula into the test tube. After the septum was removed, the test tube was quickly inserted into the nitrogen-purged stainless autoclave, and then the autoclave was sealed immediately. Hydrogen gas was introduced into the autoclave until the pressure gauge indicated over 5.0 MPa, and then the pressure was carefully released to 0.1 MPa. This procedure was repeated twice, and finally the inside of the autoclave was pressurized with hydrogen to 5.0 MPa. The mixture was vigorously stirred for 24 h. The autoclave was cooled to room temperature, and then excess hydrogen gas was released carefully. The reaction mixture was evaporated under reduced pressure. The residue was purified with a flash column chromatography on silica gel to give the desired 5,6,7,8-tetrahydrouquinoline 3.

2-Phenyl-5,6,7,8-tetrahydroquinoline [1570-04-3] (3a) (eq 1).9

Procedure A was followed with use of Et₃N (7.0 μ L, d 0.726 g/mL, 50 μ mol) and EtOAc. 2-Phenylquinoline (**2a**) (102 mg, 0.50 mmol) was used in place of **2b**. The crude product was purified with a flash column chromatography on silica gel (EtOAc/hexane = 1/20) to give **3a** (101 mg, 97%) as colorless oil: ¹H NMR (400 MHz, CDCl₃, TMS) δ 1.80–1.88 (m, 2H), 1.89–1.97 (m, 2H), 2.80 (t, J = 6.3 Hz, 2H), 3.00 (t, J = 6.3 Hz, 2H), 7.34–7.47 (m, 5H), 7.94 (d, J = 7.8 Hz, 2H); ¹³C { ¹H} NMR (100 MHz, CDCl₃) δ 22.8, 23.2, 28.5, 32.8, 117.9, 126.8, 128.3, 128.6, 130.7, 137.4, 139.9, 154.6, 157.2.

(-)-Methyl 5,6,7,8-tetrahydroquinoline-6-carboxylate [76384-36-6] (3b) (Table 1, entry 6).¹⁰

Procedure A was followed with use of methyl quinoline-6-carboxylate (**2b**) (94.0mg, 0.50 mmol), Cs₂CO₃ (16.3 mg, 50 μ mol), and EtOAc. The crude product was purified with a flash column chromatography on silica gel (EtOAc/hexane = 1/3) to give **3b** (82.9 mg, 86%) as pale yellow oil: $[\alpha]_D^{26}$ = -18.7 (*c* 1.12, CHCl₃) (for 35% ee of **3b**); ¹H NMR (400 MHz, CDCl₃, TMS) δ 1.93–2.05 (m, 1H), 2.26–2.34 (m, 1H), 2.75–2.83 (m, 1H), 2.91–3.10 (m, 4H), 3.74 (s, 3H), 7.05 (dd, J = 4.7, 7.6 Hz, 1H), 7.39 (d, J = 7.6 Hz, 1H), 8.37 (d, J = 4.7 Hz, 1H); ¹³C {¹H} NMR (100 MHz, CDCl₃) δ 25.7, 30.8, 31.4, 39.2, 51.8, 121.1, 130.0, 136.7, 147.2, 155.9, 175.2.

The enantiomeric excess of **3b** was determined to be 31% ee by the HPLC analysis with Chiralcel OD-H (4.6 mm $\phi \times 250$ mm): 10 or 30% 2-propanol in hexane, 0.5 mL/min flow, at 35°C, UV 268 nm detection, (+) $t_1 = 9.5$ min, (-) $t_2 = 10.8$ min (30% 2-propanol in hexane).

(-)-6-Isopropyl-5,6,7,8-tetrahydroquinoline [76384-36-6] (3c) (Table 1, entry 12).¹¹

Procedure A was followed with use of DBU (7.5 μ L, d 1.018 g/mL, 50 μ mol) and i-PrOH. 6-Isopropylquinoline (2c) (85.8 mg, 0.50 mmol) was used in place of 2b. The reaction was conducted The crude product was purified with a flash column chromatography on silica gel (EtOAc/hexane = 1/10 to 1/3) to give 3c (29.0 mg, 33%) as colorless oil 6-isopropyl-1,2,3,4-tetrahydroquinoline (**4c**) (40.0 mg, 46%) as colorless oil. **3c**: $[\alpha]_D^{25} = -64.9$ (*c* 1.17, CHCl₃); ¹H NMR (400 MHz, CDCl₃, TMS) δ 0.976 (d, J = 6.7 Hz, 3H), 0.981 (d, J = 6.7 Hz, 3H), 1.44– 1.69 (m, 3H), 1.99–2.09 (m, 1H), 2.53 (dd, J = 10.3, 16.3 Hz, 1H), 2.78 (dd, J = 4.2, 16.3 Hz, 1H), 2.83– 2.95 (m, 1H), 2.98–3.07 (m, 1H), 7.01 (dd, J = 4.7, 7.6 Hz, 1H), 7.35 (d, J = 7.6 Hz, 1H), 8.34 (d, J = 4.7Hz, 1H); ¹³C {¹H} NMR (100 MHz, CDCl₃) δ 19.7, 19.8, 26.4, 32.0, 32.4, 32.7, 40.3, 120.8, 132.1, 136.9, 146.7, 157.4. **4c**: ¹H NMR (400 MHz, CDCl₃, TMS) δ 1.19 (d, J = 6.9 Hz, 6H), 1.90–1.97 (m, 2H), 2.68-2.81 (m, 3H), 3.27 (t, J = 5.5 Hz, 2H), 3.71 (br, 1H), 6.43 (d, J = 8.0 Hz, 1H), 6.80-6.86 (m, 2H); ¹³C {¹H} NMR (100 MHz, CDCl₃) δ 22.4, 24.3, 27.0, 33.2, 42.1, 114.4, 121.4, 124.6, 127.4, 137.6, 142.7; IR (neat) 3402, 2954, 2865, 1616, 1511, 1297, 813 cm⁻¹; Anal. Calcd for C₁₂H₁₇N: C, 82.23; H, 9.78; N, 7.99. Found: C, 82.11; H, 9.80; N, 7.89.

The enantiomeric excess of 3c was determined to be 62% ee by the HPLC analysis with Chiralcel OD-H (4.6 mm $\phi \times 250$ mm): 4% 2-propanol in hexane, 0.5 mL/min flow, at 35°C, UV 268 nm detection, (+) $t_1 = 15.2$ min, (-) $t_2 = 18.2$ min.

3-Methoxy-5,6,7,8-tetrahydroquinoline [405174-69-8] (3d) (Table 2, entry 1).10

Procedure B was followed with use of 3-methoxyquinoline (**2d**) (40.2 mg, 0.25 mmol). The reaction was conducted in *i*-PrOH at 80°C. The crude product was purified with a flash column chromatography on silica gel (EtOAc/hexane = 1/1) to give **3d** (37.4 mg, 91%) as pale yellow oil: ¹H NMR (400 MHz, CDCl₃, TMS) δ 1.75–1.83 (m, 2H), 1.83–1.91 (m, 2H), 2.75 (t, J = 6.2 Hz, 2H), 2.85 (t, J = 6.4 Hz, 2H), 3.81 (s, 3H), 6.88 (d, J = 2.6 Hz, 1H), 8.07 (d, J = 2.6 Hz, 1H); ¹³C {¹H} NMR (100 MHz, CDCl₃) δ 22.6, 23.3, 29.0, 31.6, 55.5, 121.1, 132.5, 134.5, 149.4, 153.7.

4-Methoxy-5,6,7,8-tetrahydroquinoline [860202-86-4] (3e) (Table 2, entry 2).¹²

Procedure B was followed with use of 4-methoxyquinoline (**2e**) (39.8mg, 0.25 mmol). The reaction was conducted in *i*-PrOH at 80°C. The crude product was purified with a flash column chromatography on silica gel (MeOH/EtOAc = 1/10) to give **3e** (40.4 mg, 99%) as pale yellow oil: ¹H NMR (400 MHz, CDCl₃, TMS) δ 1.73–1.88 (m, 4H), 2.62 (t, J = 6.2 Hz, 2H), 2.87 (t, J = 6.2 Hz, 2H), 3.83 (s, 3H), 6.57 (d, J = 5.6 Hz, 1H), 8.26 (d, J = 5.6 Hz, 1H); ¹³C {¹H} NMR (100 MHz, CDCl₃) δ 22.06, 22.13, 22.7, 32.4, 55.0, 103.0, 121.0, 147.8, 157.8, 163.4.

(+)-5-Methoxy-5,6,7,8-tetrahydroquinoline (3f) (Table 2, entry 3).

Procedure B was followed with use of 5-methoxyquinoline (**2f**) (40.0 mg, 0.25 mmol). The reaction was conducted in *i*-PrOH at 80°C. The crude product was purified with a flash column chromatography on silica gel (EtOAc/hexane = 1/1) to give **3f** (16.5 mg, 40%) as colorless oil: $[\alpha]_D^{25} = +1.8$ (*c* 1.11, CHCl₃); ¹H NMR (400 MHz, CDCl₃, TMS) δ 1.77–1.87 (m, 1H), 1.94–2.14 (m, 3H), 2.87 (dt, J = 17.8, 6.8 Hz, 1H), 2.99 (dt, J = 17.8, 6.2 Hz, 1H), 3.46 (s, 3H), 4.34 (t, J = 5.2 Hz, 1H), 7.11 (dd, J = 4.7, 7.6 Hz, 1H), 7.67 (d, J = 7.6 Hz, 1H), 8.44 (dd, J = 1.3, 4.7 Hz, 1H); ¹³C {¹H} NMR (100 MHz, CDCl₃) δ 18.4, 26.9, 32.1, 56.2, 76.5, 121.1, 132.2, 136.8, 148.6, 157.5; IR (neat) 2940, 1576, 1443, 1351, 1087, 803 cm⁻¹; HRMS (FAB) Calcd for C₁₀H₁₄NO: 164.1075. Found: m/z = 164.1055 ([M+H]⁺).

The enantiomeric excess of **3f** was determined to be 42% ee by the HPLC analysis with Chiralcel OD-H (4.6 mm $\phi \times 250$ mm): 10% 2-propanol in hexane, 0.5 mL/min flow, at 35°C, UV 268 nm detection, (+) $t_1 = 10.1$ min, (-) $t_2 = 13.7$ min.

(+)-6-Methoxy-5,6,7,8-tetrahydroquinoline [75414-06-1] (3g) (Table 2, entry 4).¹³

Procedure B was followed with use of 6-methoxyquinoline (**2g**) (39.7 mg, 0.25 mmol). The reaction was conducted in *i*-PrOH at 80°C. The crude product was purified with a flash column chromatography on silica gel (EtOAc/hexane = 1/1) to give **3g** (32.0 mg, 79%) as pale yellow oil: $[\alpha]_D^{25}$ = +6.3 (*c* 1.15, CHCl₃); ¹H NMR (400 MHz, CDCl₃, TMS) δ 1.96–2.06 (m, 1H), 2.08–2.17 (m, 1H), 2.83 (dd, J = 6.7, 16.7 Hz, 1H), 2.92 (dt, J = 17.6, 6.9 Hz, 1H), 3.01–3.14 (m, 2H), 3.43 (s, 3H), 3.69–3.75 (m, 1H), 7.04 (dd, J = 4.7, 7.6 Hz, 1H), 7.37 (d, J = 7.6 Hz, 1H), 8.37 (d, J = 4.7 Hz, 1H); ¹³C {¹H} NMR (100 MHz, CDCl₃) δ 27.0, 29.4, 34.4, 55.9, 74.6, 121.1, 129.4, 137.2, 147.1, 156.4.

The enantiomeric excess of 3g was determined to be 58% ee by the HPLC analysis with Chiralpak AS-H (4.6 mm $\phi \times 250$ mm): 4% 2-propanol in hexane, 0.5 mL/min flow, at 35°C, UV 268 nm detection, (-) $t_1 = 14.1$ min, (+) $t_2 = 18.4$ min.

(-)-7-Methoxy-5,6,7,8-tetrahydroquinoline (3h) (Table 2, entry 5).

Procedure B was followed with use of 7-methoxyquinoline (**2h**) (39.8 mg, 0.25 mmol). The reaction was conducted in *i*-PrOH at 80°C. The crude product was purified with a flash column chromatography on silica gel (EtOAc/hexane = 1/1) to give **3h** (34.1 mg, 84%) as pale yellow oil: $[\alpha]_D^{25} = -9.6$ (*c* 1.11, CHCl₃); ¹H NMR (400 MHz, CDCl₃, TMS) δ 1.89–1.98 (m, 1H), 1.99–2.07 (m, 1H), 2.74 (dt, J = 16.9, 6.6 Hz, 1H), 2.93 (dt, J = 16.9, 6.6 Hz, 1H), 3.00 (dd, J = 6.2, 17.3 Hz, 1H), 3.21 (dd, J = 4.8, 17.3 Hz, 1H), 3.43 (s, 3H), 3.76–3.82 (m, 1H), 7.04 (dd, J = 4.7. 7.7 Hz, 1H), 7.37 (d, J = 7.7 Hz, 1H), 8.37 (d, J = 4.7 Hz, 1H); ¹³C {¹H} NMR (100 MHz, CDCl₃) δ 25.1, 27.0, 38.0, 55.9, 75.1, 121.1, 131.3, 136.2, 147.2, 154.9; IR (neat) 2930, 1577, 1446, 1096, 790 cm⁻¹; HRMS (FAB) Calcd for C₁₀H₁₄NO: 164.1075. Found: m/z = 164.1102 ([M+H]⁺).

The enantiomeric excess of **3h** was determined to be 37% ee by the HPLC analysis with Chiralcel OD-H (4.6 mm $\phi \times 250$ mm): 10% or 20% 2-propanol in hexane, 0.5 mL/min flow, at 35°C, UV 268 nm detection, (–) $t_1 = 9.4$ min, (+) $t_2 = 11.3$ min (20% 2-propanol in hexane).

(+)-8-Methoxy-5,6,7,8-tetrahydroquinoline [75414-07-2] (3i) (Table 2, entry 9).¹³

Procedure B was followed with use of 8-methoxyquinoline (**2i**) (159 mg, 1.0 mmol). The reaction was conducted in EtOAc (1.0 mL) at 60°C. The crude product was purified with a flash column chromatography on silica gel (EtOAc/hexane = 1/1) to give **3i** (154 mg, 94%) as pale yellow oil: $[\alpha]_D^{24}$ = +37.6 (*c* 1.04, CHCl₃); ¹H NMR (400 MHz, CDCl₃, TMS) δ 1.72–1.90 (m, 2H), 1.95–2.07 (m, 1H), 2.20–2.29 (m, 1H), 2.72 (ddd, J = 5.8, 10.1, 17.0 Hz, 1H), 2.84 (dt, J = 17.0, 4.7 Hz, 1H), 3.54 (s, 3H), 4.36 (t, J = 3.8 Hz, 1H), 7.14 (dd, J = 4.7, 7.7 Hz, 1H), 7.42 (d, J = 7.7 Hz, 1H), 8.48 (d, J = 4.7 Hz, 1H); ¹³C { ¹H} NMR (100 MHz, CDCl₃) δ 17.5, 27.5, 28.4, 57.0, 77.6, 122.8, 132.9, 137.2, 147.2, 155.2.

The enantiomeric excess of **3i** was determined to be 82% ee by the HPLC analysis with Chiralcel OD-H (4.6 mm $\phi \times 250$ mm): 10% or 20% 2-propanol in hexane, 0.5 mL/min flow, at 35°C, UV 275 nm detection, (+) $t_1 = 8.7$ min, (-) $t_2 = 10.2$ min (20% 2-propanol in hexane).

(+)-8-[(Triisopropylsilyloxy)methoxy]-5,6,7,8-tetrahydroquinoline (3j) (Table 3, entry 1).

Procedure B was followed with use of 8-[(triisopropylsilyloxy)methoxy]quinoline (2j) (78.8 mg, 0.24 mmol). The reaction was conducted in EtOAc at 60°C. The crude product was purified with a flash

column chromatography on silica gel (EtOAc/hexane = 1/3) to give **3j** (77.2 mg, 97%) as colorless oil: $[\alpha]_D^{26} = +33.4$ (c 1.04, CHCl₃); ¹H NMR (400 MHz, CDCl₃, TMS) δ 1.02–1.18 (m, 21H), 1.73–1.82 (m, 1H), 1.89–1.98 (m, 1H), 2.01–2.14 (m, 1H), 2.29–2.38 (m, 1H), 2.71 (ddd, J = 5.9, 10.0, 16.9 Hz, 1H), 2.83 (dt, J = 16.9, 4.8 Hz, 1H), 4.76 (t, J = 4.0 Hz, 1H), 5.19 (d, J = 4.9 Hz, 1H), 5.34 (d, J = 4.9 Hz, 1H), 7.11 (dd, J = 4.7, 7.7 Hz, 1H), 7.39 (d, J = 7.7 Hz, 1H), 8.44 (d, J = 4.7 Hz, 1H); ¹³C {¹H} NMR (100 MHz, CDCl₃) δ 12.0, 17.85, 17.89, 28.3, 30.3, 74.8, 90.0, 122.6, 132.8, 137.0, 147.2, 155.9; IR (neat) 2944, 2867, 1500, 1470, 1376, 1318, 1251, 1156, 1084, 1061, 984 cm⁻¹; Anal. Calcd for C₁₉H₃₃NO₂Si: C, 68.01; H, 9.91; N, 4.17. Found: C, 67.99; H, 9.93; N, 4.05.

The enantiomeric excess of 3j was determined to be 81% ee by the HPLC analysis with Chiralcel OD-H (4.6 mm $\phi \times 250$ mm): 1% 2-propanol in hexane, 0.5 mL/min flow, at 35°C, UV 268 nm detection, (+) $t_1 = 9.3$ min, (-) $t_2 = 10.9$ min.

(+)-8-Phenyl-5,6,7,8-tetrahydroquinoline [56826-78-9] (3k) (Table 3, entry 2).¹³

Procedure B was followed with use of 8-phenylquinoline (**2k**) (52.3 mg, 0.25 mmol). The reaction was conducted in *i*-PrOH at 40°C for 48 h. The crude product was purified with a flash column chromatography on silica gel (EtOAc/hexane = 1/5) to give **3k** (46.5 mg, 87%) as colorless oil: $[\alpha]_D^{24}$ = +34.5 (*c* 1.00, CHCl₃); ¹H NMR (400 MHz, CDCl₃, TMS) δ 1.65–1.76 (m, 1H), 1.78–1.90 (m, 1H), 1.92–2.02 (m, 1H), 2.17–2.27 (m, 1H), 2.79 (dt, *J* = 17.0, 6.7 Hz, 1H), 2.88 (dt, *J* = 17.0, 6.1 Hz, 1H), 4.30 (t, *J* = 6.1 Hz, 1H), 6.98 (d, *J* = 7.2 Hz, 2H), 7.01 (dd, *J* = 4.8, 7.8 Hz, 1H), 7.14 (t, *J* = 7.3 Hz, 1H), 7.23 (t, *J* = 7.5 Hz, 2H), 7.40 (d, *J* = 7.8 Hz, 1H), 8.37 (d, *J* = 4.8 Hz, 1H); ¹³C {¹H} NMR (100 MHz, CDCl₃) δ 19.3, 28.8, 32.7, 47.5, 121.0, 125.6, 128.0, 128.4, 132.8, 136.6, 146.2, 147.3, 158.3.

The enantiomeric excess of **3k** was determined to be 73% ee by the HPLC analysis with Chiralcel OD-H (4.6 mm $\phi \times 250$ mm): 10% 2-propanol in hexane, 0.5 mL/min flow, at 35°C, UV 268 nm detection, (+) $t_1 = 9.8$ min, (-) $t_2 = 10.8$ min.

(+)-8-(4-Methoxyphenyl)-5,6,7,8-tetrahydroquinoline (31) (Table 3, entry 3).

Procedure B was followed with use of 8-(4-methoxyphenyl)quinoline (**2l**) (58.5 mg, 0.25 mmol). The reaction was conducted in *i*-PrOH at 60°C. The crude product was purified with a flash column chromatography on silica gel (EtOAc/hexane = 1/2) to give **3l** (56.1 mg, 94%) as colorless oil: $[\alpha]_D^{27}$ = +39.8 (*c* 1.02, CHCl₃); ¹H NMR (400 MHz, CDCl₃, TMS) δ 1.67–1.78 (m, 1H), 1.80–2.00 (m, 2H), 2.16–2.26 (m, 1H), 2.81 (dt, J = 16.8, 6.7 Hz, 1H), 2.89 (dt, J = 16.8, 6.1 Hz, 1H), 3.75 (s, 3H), 4.26 (t, J = 6.0 Hz, 1H), 6.80 (d, J = 8.6 Hz, 2H), 6.91 (d, J = 8.6 Hz, 2H), 7.04 (dd, J = 4.6, 7.6 Hz, 1H), 7.42 (d, J = 7.6

Hz, 1H), 8.39 (d, J = 4.6 Hz, 1H); ¹³C {¹H} NMR (100 MHz, CDCl₃) δ 19.5, 29.0, 32.9, 46.9, 55.1, 113.6, 121.1, 129.4, 132.9, 136.7, 138.6, 147.4, 157.6, 158.7; IR (neat) 2934, 1511, 1444, 1246, 1177, 1035, 827, 796 cm⁻¹; Anal. Calcd for C₁₆H₁₇NO: C, 80.30; H, 7.16; N, 5.85. Found: C, 80.16; H, 6.89; N, 5.86.

The enantiomeric excess of **31** was determined to be 72% ee by the HPLC analysis with Chiralcel OD-H (4.6 mm $\phi \times 250$ mm): 10% 2-propanol in hexane, 0.5 mL/min flow, at 35°C, UV 268 nm detection, (+) $t_1 = 13.7$ min, (-) $t_2 = 16.9$ min.

(+)-8-[4-(Trifluoromethyl)phenyl]-5,6,7,8-tetrahydroquinoline (3m) (Table 3, entry 4).

Procedure B was followed with use of 8-[4-(trifluoromethyl)phenyl]quinoline (**2m**) (68.3 mg, 0.25 mmol). The reaction was conducted in *i*-PrOH at 60°C. The crude product was purified with a flash column chromatography on silica gel (EtOAc/hexane = 1/3) to give **3m** (60.8 mg, 88%) as colorless oil: $[\alpha]_D^{27} = +18.7$ (c 1.01, CHCl₃); ¹H NMR (400 MHz, CDCl₃, TMS) δ 1.71–2.01 (m, 3H), 2.22–2.32 (m, 1H), 2.84 (dt, J = 17.0, 6.5 Hz, 1H), 2.92 (dt, J = 17.0, 6.5 Hz, 1H), 4.36 (t, J = 6.4 Hz, 1H), 7.08 (dd, J = 4.7, 7.7 Hz, 1H), 7.12 (d, J = 8.1 Hz, 2H), 7.46 (d, J = 7.7 Hz, 1H), 7.52 (d, J = 8.1 Hz, 2H), 8.38 (d, J = 4.7 Hz, 1H); ¹³C (¹H} NMR (100 MHz, CDCl₃) δ 19.7, 28.9, 32.9, 47.8, 121.6, 124.3 (q, J = 272 Hz), 125.2 (q, J = 4 Hz), 128.1 (q, J = 32 Hz), 128.9, 133.1, 137.0, 147.6, 150.5, 157.6; IR (neat) 2938, 1446, 1327, 1163, 1120, 1067, 833, 790 cm⁻¹; Anal. Calcd for C₁₆H₁₄F₃N: C, 69.30; H, 5.09; N, 5.05. Found: C, 69.32; H, 5.08; N, 5.09.

The enantiomeric excess of **3m** was determined to be 72% ee by the HPLC analysis with Chiralpak AS-H (4.6 mm $\phi \times 250$ mm): 10% 2-propanol in hexane, 0.5 mL/min flow, at 35°C, UV 268 nm detection, (–) $t_1 = 7.9$ min, (+) $t_2 = 8.7$ min.

(+)-8-(2-Methylphenyl)-5,6,7,8-tetrahydroquinoline (3n) (Table 3, entry 5).

Procedure B was followed with use of 8-(2-methylphenyl)quinoline (**2n**) (54.1 mg, 0.25 mmol). The reaction was conducted in *i*-PrOH at 60°C. The crude product was purified with a flash column chromatography on silica gel (EtOAc/hexane = 1/3) to give **3n** (30.8 mg, 56%) as a colorless solid: $[\alpha]_D^{25}$ = +39.0 (*c* 1.00, CHCl₃); ¹H NMR (400 MHz, CDCl₃, TMS) δ 1.66–1.77 (m, 1H), 1.81–1.94 (m, 2H), 2.13–2.22 (m, 1H), 2.39 (s, 3H), 2.80 (dt, J = 16.7, 6.8 Hz, 1H), 2.90 (dt, J = 16.7, 6.0 Hz, 1H), 4.50 (t, J = 6.1 Hz, 1H), 6.53 (d, J = 7.3 Hz, 1H), 6.97–7.08 (m, 3H), 7.15 (d, J = 7.3 Hz, 1H), 7.40 (d, J = 7.6 Hz, 1H), 8.37 (d, J = 4.7 Hz, 1H); ¹³C {¹H} NMR (100 MHz, CDCl₃) δ 19.4, 19.5, 29.0, 30.5, 44.2, 120.9, 125.4, 125.7, 128.9, 130.4, 133.0, 135.4, 136.5, 144.3, 147.4, 158.9; IR (neat) 3045, 2940, 2857, 1571,

1489, 1443, 807, 789, 757, 729 cm⁻¹; Anal. Calcd for $C_{16}H_{17}N$: C, 86.05; H, 7.67; N, 6.27. Found: C, 85.77; H, 7.69; N, 6.30.

The enantiomeric excess of **3n** was determined to be 41% ee by the HPLC analysis with Chiralcel OJ-H (4.6 mm $\phi \times 250$ mm): 10% 2-propanol in hexane, 0.5 mL/min flow, at 35°C, UV 268 nm detection, (-) $t_1 = 12.2$ min, (+) $t_2 = 13.3$ min.

(-)-8-Methyl-5,6,7,8-tetrahydroquinoline [52601-66-8] (30) (Table 3, entry 7).¹³

Procedure B was followed with use of 8-methylquinoline (**20**) (35.9 mg, 0.25 mmol). {Ru(p-cymene)[(S,S)-(R,R)-PhTRAP]}Cl (5.5 mg, 5.0 μ mol) and DBU (8.2 mg, 55 μ mol) were used in place of [Ru(η^3 -methallyl)₂(cod)]–(S,S)-(R,R)-PhTRAP and K₂CO₃, respectively. The reaction was conducted in i-PrOH at 60°C. The crude product was purified with a flash column chromatography on silica gel (EtOAc/hexane = 1/1) to give **30** (25.9 mg, 71%) as colorless oil: [α]_D²⁶ = -29.7 (c 1.05, CHCl₃); ¹H NMR (400 MHz, CDCl₃, TMS) δ 1.37 (d, J = 7.1 Hz, 3H), 1.60–1.80 (m, 2H), 1.83–1.94 (m, 1H), 1.98–2.08 (m, 1H), 2.69–2.84 (m, 2H), 3.00 (sextet, J = 6.6 Hz, 1H), 7.01 (dd, J = 4.7, 7.6 Hz, 1H), 7.33 (d, J = 7.6 Hz, 1H), 8.39 (d, J = 4.7 Hz, 1H); ¹³C {¹H} NMR (100 MHz, CDCl₃) δ 20.0, 21.3, 29.4, 31.2, 35.6, 120.8, 131.8, 136.6, 146.9, 161.3.

The enantiomeric excess of **30** was determined to be 79% ee by the HPLC analysis with Chiralcel OB-H (4.6 mm $\phi \times 250$ mm): 4% 2-propanol in hexane, 0.5 mL/min flow, at 35°C, UV 268 nm detection, (-) $t_1 = 9.2$ min, (+) $t_2 = 10.0$ min.

(-)-8-Cyclohexyl-5,6,7,8-tetrahydroquinoline [75414-11-8] (3p) (Table 3, entry 8). 13

Procedure B was followed with use of 8-cyclohexylquinoline (**2p**) (52.8 mg, 0.25 mmol). {Ru(p-cymene)[(S,S)-(R,R)-PhTRAP]}Cl (5.5 mg, 5.0 μ mol) and DBU (8.2 mg, 55 μ mol) were used in place of [Ru(η^3 -methallyl)₂(cod)]–(S,S)-(R,R)-PhTRAP and K₂CO₃, respectively. The reaction was conducted in i-PrOH at 60°C. The crude product was purified with a flash column chromatography on silica gel (EtOAc/hexane = 1/1) to give **3p** (46.4 mg, 86%) as colorless oil: [α]_D²⁶ = -75.7 (c 1.00, CHCl₃); ¹H NMR (400 MHz, CDCl₃, TMS) δ 0.97–1.43 (m, 6H), 1.56–1.80 (m, 6H), 1.86–1.97 (m, 2H), 2.30–2.41 (m, 1H), 2.63–2.78 (m, 2H), 2.79–2.88 (m, 1H), 6.98 (dd, J = 4.8, 7.6 Hz, 1H), 7.30 (d, J = 7.6 Hz, 1H), 8.41 (d, J = 4.8 Hz, 1H); ¹³C {¹H} NMR (100 MHz, CDCl₃) δ 21.6, 24.1, 26.70, 26.73, 27.0, 27.7, 29.5, 31.5, 41.3, 45.9, 120.3, 133.2, 136.3, 146.7, 159.8.

The enantiomeric excess of **3p** was determined to be 81% ee by the HPLC analysis with Chiralcel OD-H (4.6 mm $\phi \times 250$ mm): 4% 2-propanol in hexane, 0.5 mL/min flow, at 35°C, UV 268 nm detection, (–) $t_1 = 8.1$ min, (+) $t_2 = 9.0$ min.

Deprotection of 3j (eq 2).

(R)-5,6,7,8-Tetrahydroquinolin-8-ol [451466-81-2] (3q).9

Compound **3j** (81% ee, 62.0 mg, 0.18 mmol) was placed in a 2.0 mL Schlenk tube, which was equipped with a stirring bar, rubber septum and three-way stopcock. After the reaction vessel was evacuated and charged with nitrogen gas three times, a 1.0 M solution of TBAF in THF (0.36 mL, 0.36 mmol) was added into the tube through the septum by using a syringe. The solution was stirred at ambient temperature for 5 h. After the solution was acidified with AcOH and then diluted with water, the resulting mixture was extracted three times with CH_2Cl_2 . The combined organic layer was dried over Na_2SO_4 , and then evaporated under reduced pressure after filtration. The residue was purified with a flash column chromatography on silica gel (MeOH/EtOAc = 1/10) to give **3q** (24.7 mg, 90%) as a colorless solid: $[\alpha]_D^{24} = -52.0$ (c 1.00, $CHCl_3$); lit. $[\alpha]_D^{20} = -65$ (c 1.05, $CHCl_3$); $[\alpha]_D^{20} = -65$ (c 1.05, $[\alpha]_D^{20} = -65$) ($[\alpha]_D^{20} = -65$) (

The enantiomeric excess of 3q was determined to be 80% ee by the HPLC analysis with Chiralcel OD-H (4.6 mm $\phi \times 250$ mm): 10% 2-propanol in hexane, 0.5 mL/min flow, at 35°C, UV 268 nm detection, (R) $t_1 = 12.4$ min, (S) $t_2 = 15.2$ min.

Mechanistic Study on the Ruthenium-Catalyzed Hydrogenation of 2. Preparation of 8-phenyl-5,6-dihydroquinoline [343320-64-9] (5k).

MsOH (400 mg, 4.2 mmol) was placed in a 10 mL Schlenk tube, which was equipped with a stirring bar, rubber septum, and cold finger. Toluene (3.0 mL) and then 8-phenyl-5,6,7,8-tetrahydroquinolin-8-ol (225 mg, 1.0 mmol) were added into the tube. The solution was stirred under reflux for 12 h. After saturated Na₂CO₃ aq. was added, the mixture was extracted three times with EtOAc. The combined organic layer was washed with brine, dried over Na₂SO₄, and

then evaporated under reduced pressure after filtration. The residue was purified with a flash column chromatography on silica gel (EtOAc/hexane = 1/10) to give **5k** (202 mg, 97%) as a colorless solid: mp. 78.0–78.1°C; ¹H NMR (400 MHz, CDCl₃, TMS) δ 2.46 (dt, J = 4.7, 8.0 Hz, 2H), 2.90 (t, J = 8.0 Hz, 2H), 6.43 (t, J = 4.7 Hz, 1H), 7.05 (dd, J = 4.9, 7.5 Hz, 1H), 7.30 (t, J = 7.1 Hz, 1H), 7.37 (t, J = 7.3 Hz, 2H), 7.41–7.47 (m, 3H), 8.40 (dd, J = 1.7, 4.9 Hz, 1H); ¹³C {¹H} NMR (100 MHz, CDCl₃) δ 23.0, 27.8, 121.6, 127.2, 128.0, 128.8, 132.2, 134.7, 139.5, 140.9, 147.1, 153.6; IR (neat) 3051, 2936, 2888, 2829, 1560, 1492, 1431, 796, 754, 697 cm⁻¹; Anal. Calcd for C₁₆H₁₇N: C, 86.05; H, 7.67; N, 6.27. Found: C, 85.77; H, 7.69; N, 6.30.

Hydrogenation of 5k (eq 3).

The procedure B for the asymmetric hydrogenation was followed with use of 5k (51.6 mg, 0.25 mmol). The reaction was conducted in *i*-PrOH at 60°C for 1 h. The ¹H NMR analysis of the resulting mixture was indicated that 40% of 5k was remained. The crude product was purified with a flash column chromatography on silica gel (EtOAc/hexane = 1/5) to give 3k (32.2 mg, 62%) as colorless oil.

The enantiomeric excess of 3k was determined to be 11% ee (–) by the HPLC analysis with Chiralcel OD-H (4.6 mm $\phi \times 250$ mm): 10% 2-propanol in hexane, 0.5 mL/min flow, at 35°C, UV 268 nm detection.

Deuteration of 2i (eq 4).

The procedure B for the asymmetric hydrogenation was followed with use of 2i (40.0 mg, 0.25 mmol). The reaction was conducted in *i*-PrOH at 60°C under 1.0 MPa of D₂ for 24 h. The ¹H NMR analysis of the resulting mixture was indicated that it contained 2i in 61%, 3i-d in 15%, 3'-d in 6%, and 4i-d in 16%. The crude product was purified with a flash column chromatography on silica gel (EtOAc/hexane = 1/1) to give 3i-d (1.5 mg, 4%) as colorless oil. The ¹H NMR spectrum of 3i-d is given in Figure S-51. The methoxy group was used as the internal standard for calibrating the integration of each peak. In the recovered starting material, deuterium was incorporated at the 2-position in 57%.

The enantiomeric excess of **3i-d** was determined to be 79% ee (+) by the HPLC analysis with Chiralcel OD-H (4.6 mm $\phi \times 250$ mm): 10% 2-propanol in hexane, 0.5 mL/min flow, at 35°C, UV 268 nm detection.

As shown in Scheme S-1, the hydrogenation of **2i** was carried out under the above condition to compare the deuteration with the hydrogenation. The resulting mixture contains 87% ee of **3i** in 60% and **3'** in 40%. The substrate **2i** was completely consumed without formation of **4i**.

Scheme S-1. The hydrogenation of **2i** under 1.0 MPa of H₂.

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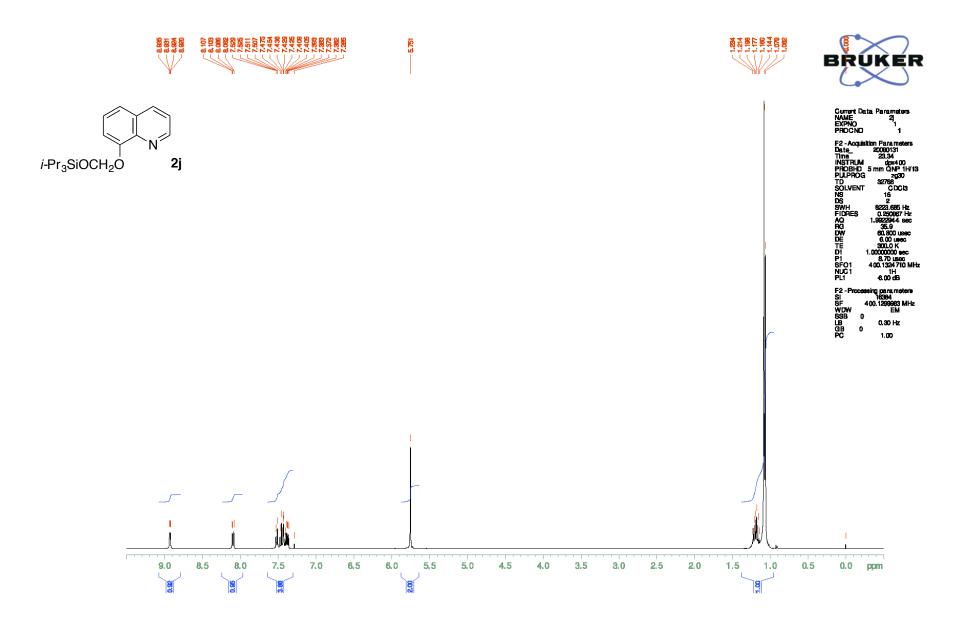


Figure S-1. ¹H NMR spectrum (CDCl₃) of 2j.

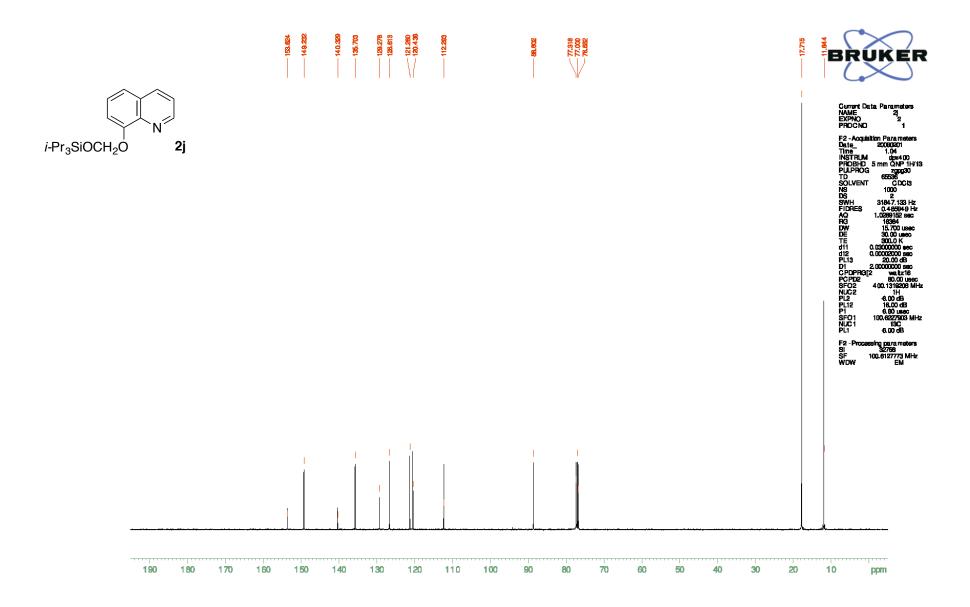


Figure S-2. ¹³C {¹H} NMR spectrum (CDCl₃) of 2j.

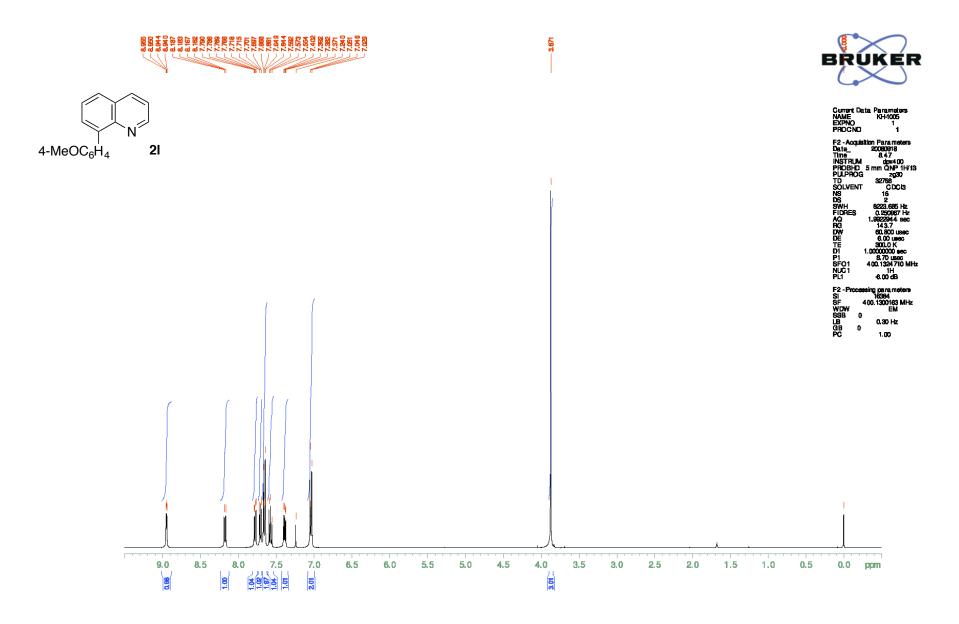


Figure S-3. ¹H NMR spectrum (CDCl₃) of 2l.

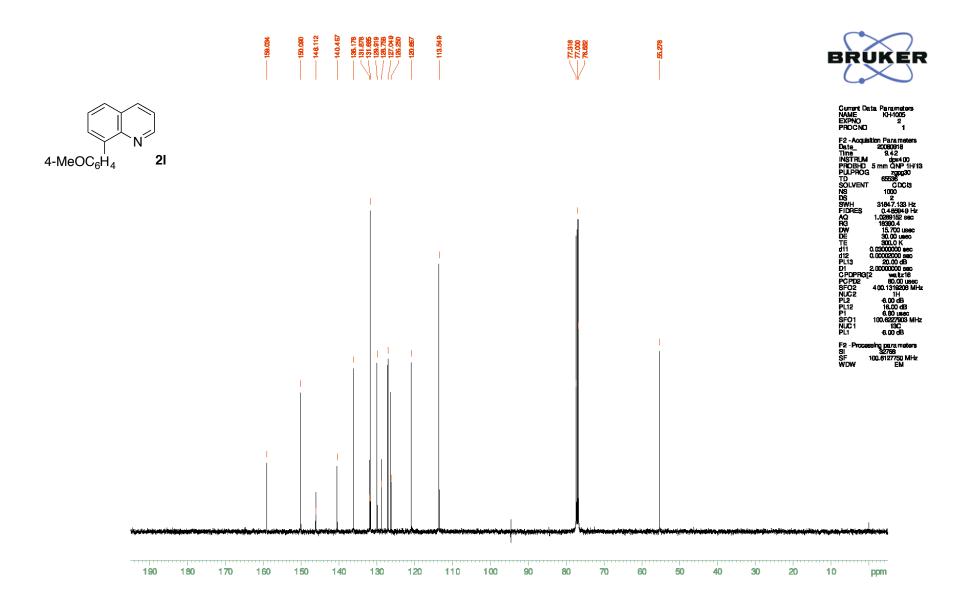


Figure S-4. ¹³C {¹H} NMR spectrum (CDCl₃) of 2l.

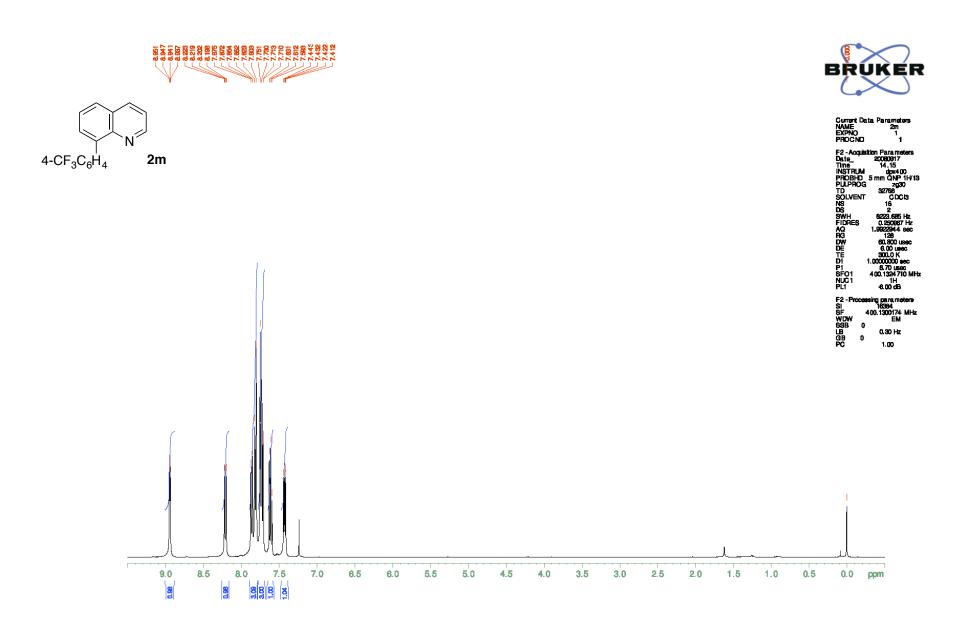


Figure S-5. ¹H NMR spectrum (CDCl₃) of **2m**.

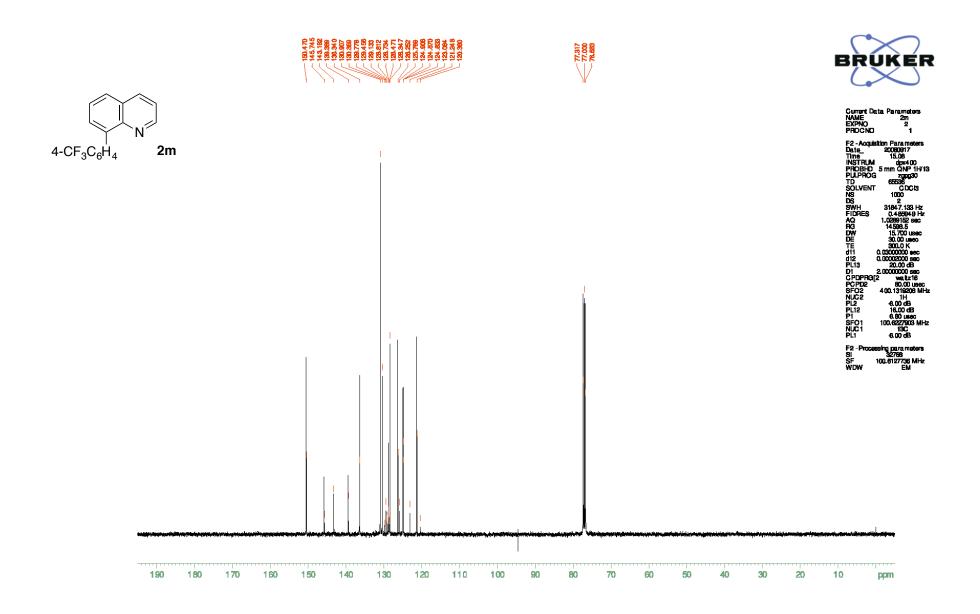


Figure S-6. 13 C $\{^{1}$ H $\}$ NMR spectrum (CDCl₃) of 2m.

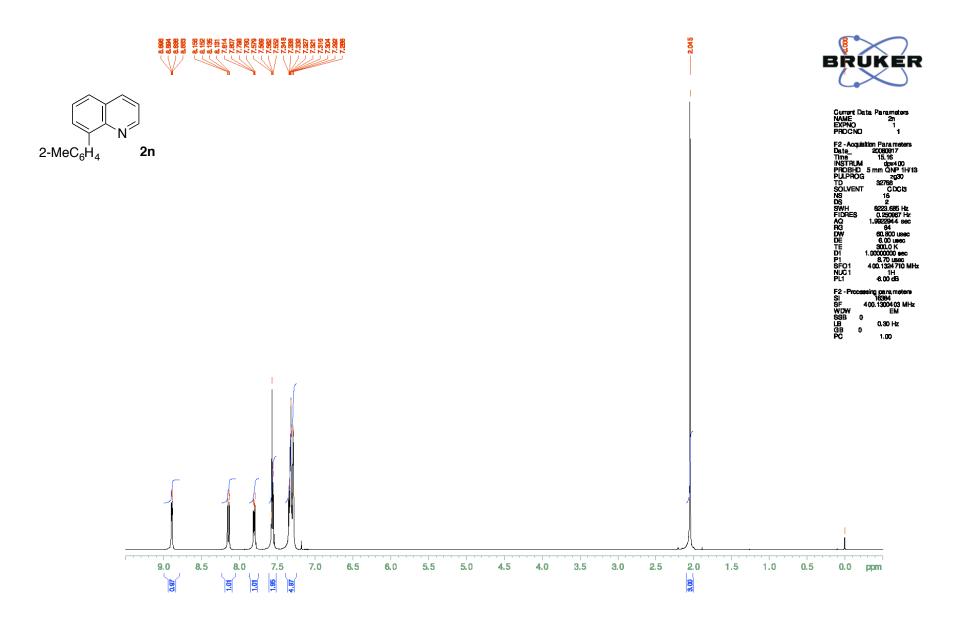


Figure S-7. ¹H NMR spectrum (CDCl₃) of 2n.

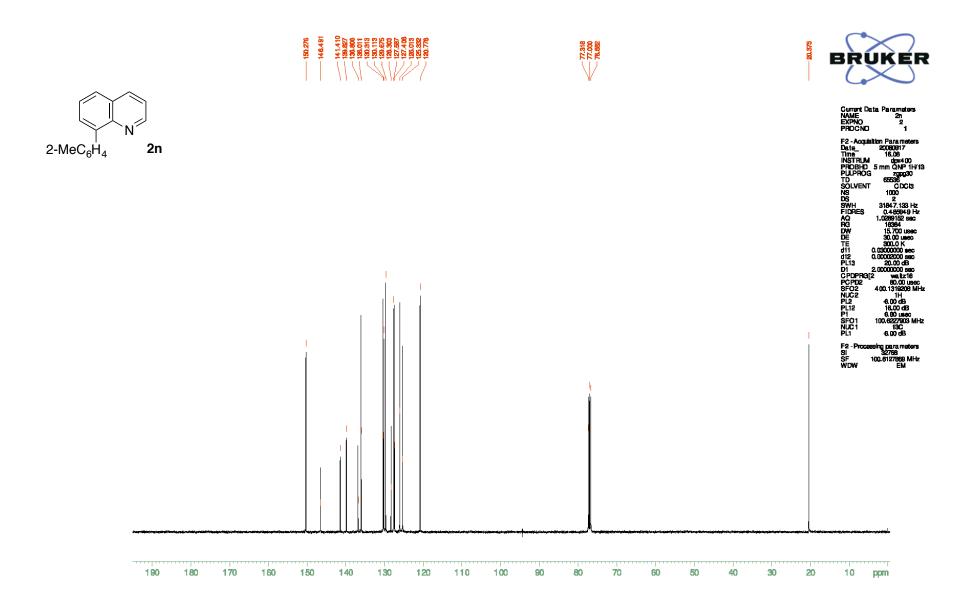


Figure S-8. 13 C $\{^{1}$ H $\}$ NMR spectrum (CDCl₃) of 2n.

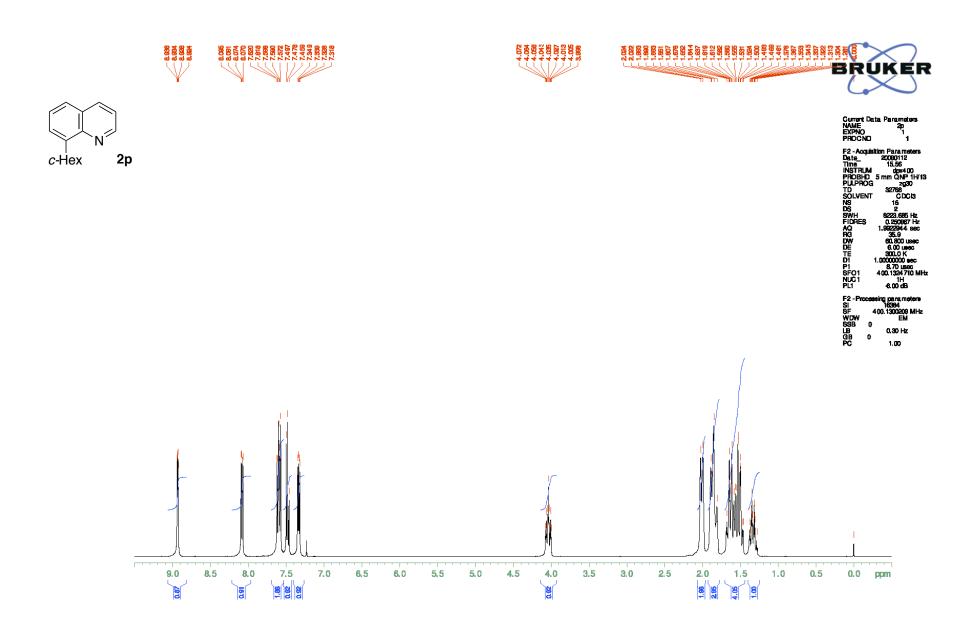


Figure S-9. ¹H NMR spectrum (CDCl₃) of **2p**.

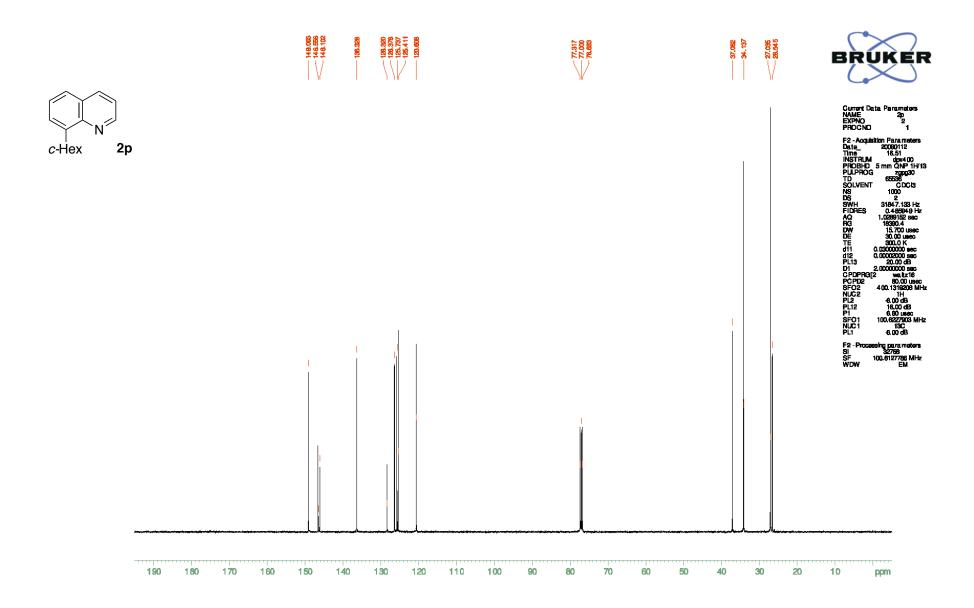


Figure S-10. 13 C $\{^{1}$ H $\}$ NMR spectrum (CDCl₃) of 2p.

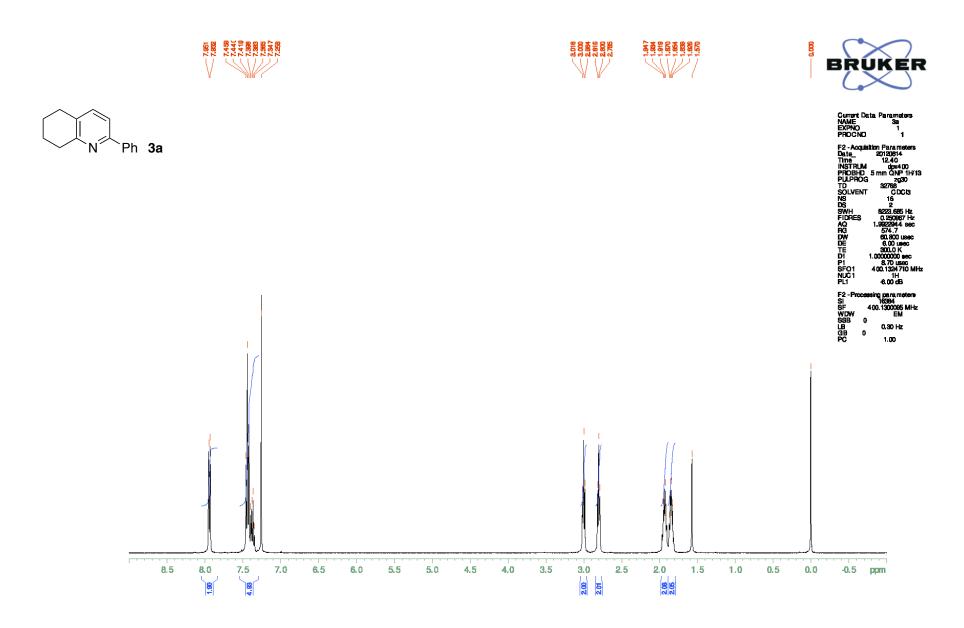


Figure S-11. ¹H NMR spectrum (CDCl₃) of 3a.

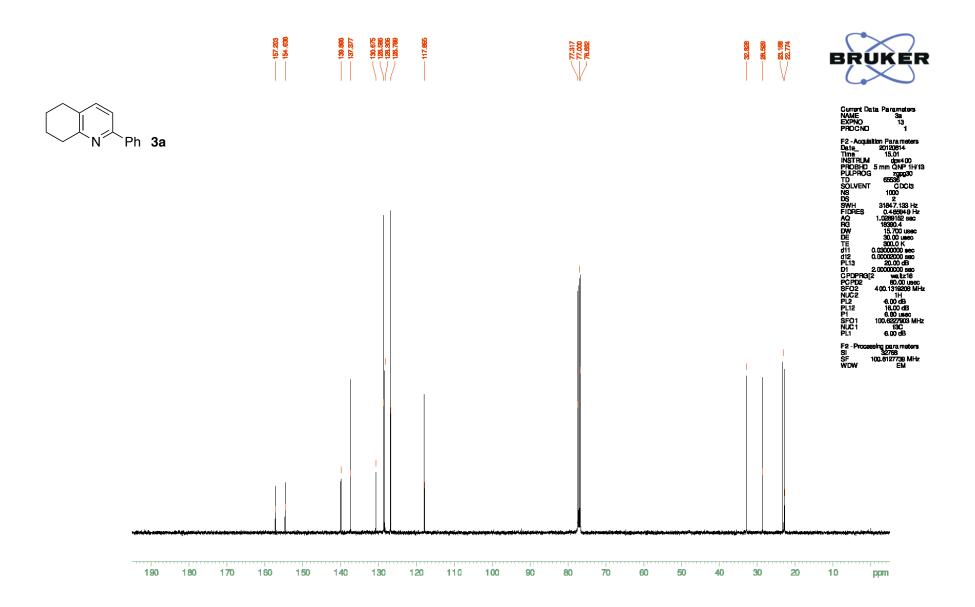


Figure S-12. 13 C $\{^{1}$ H $\}$ NMR spectrum (CDCl₃) of 3a.

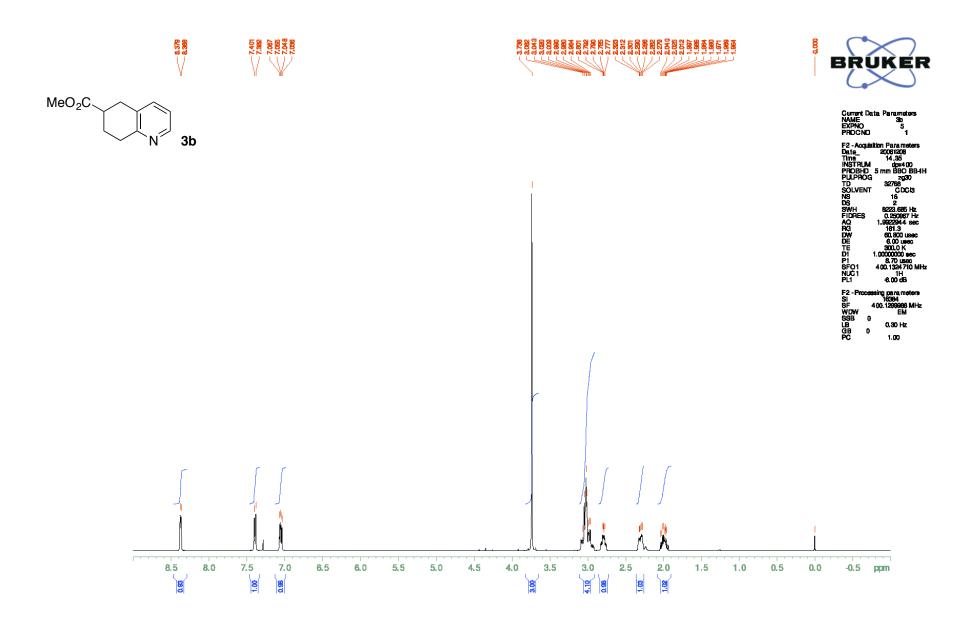


Figure S-13. ¹H NMR spectrum (CDCl₃) of **3b**.

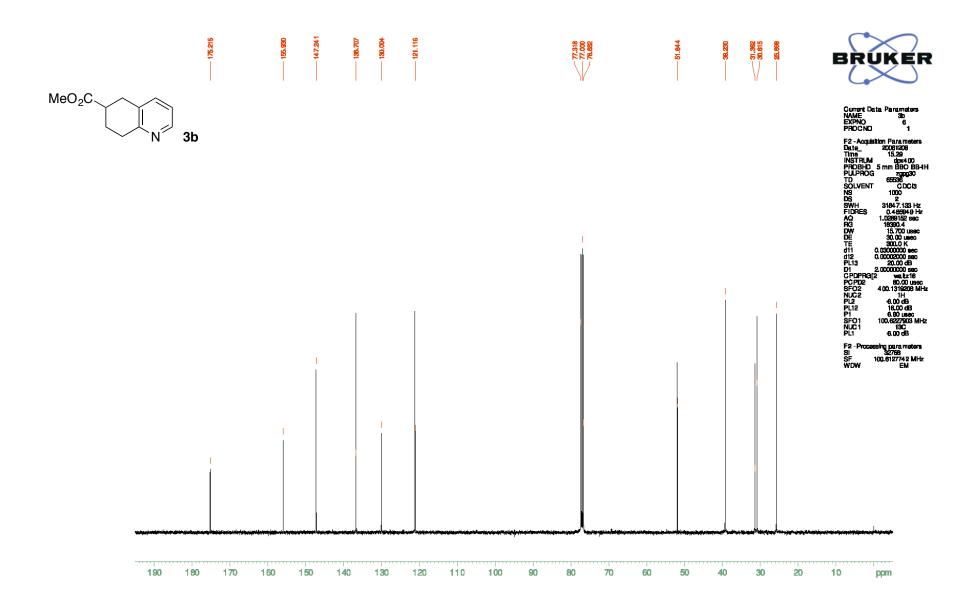


Figure S-14. 13 C $\{^{1}$ H $\}$ NMR spectrum (CDCl₃) of **3b**.

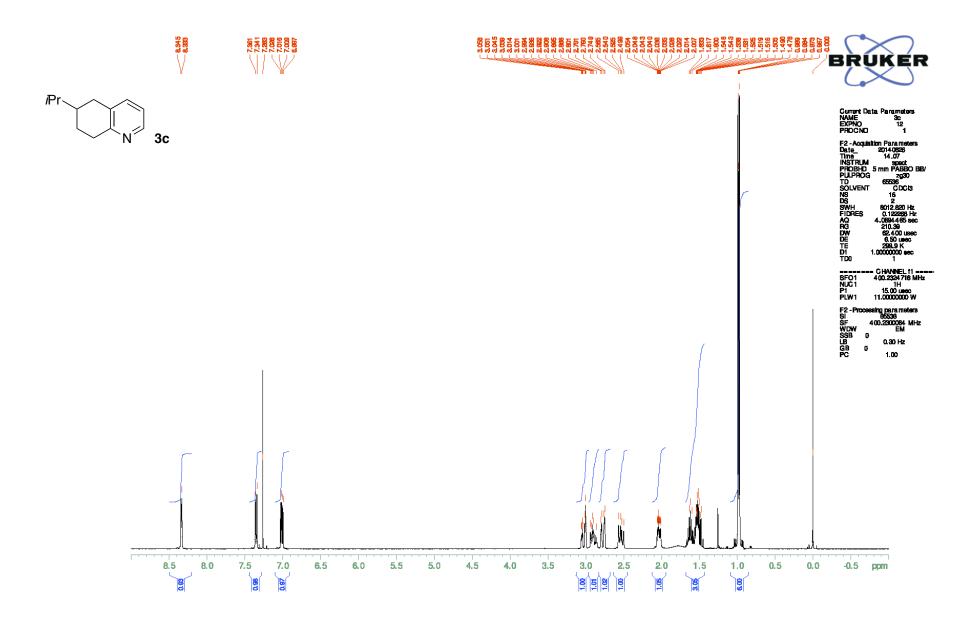


Figure S-15. ¹H NMR spectrum (CDCl₃) of 3c.

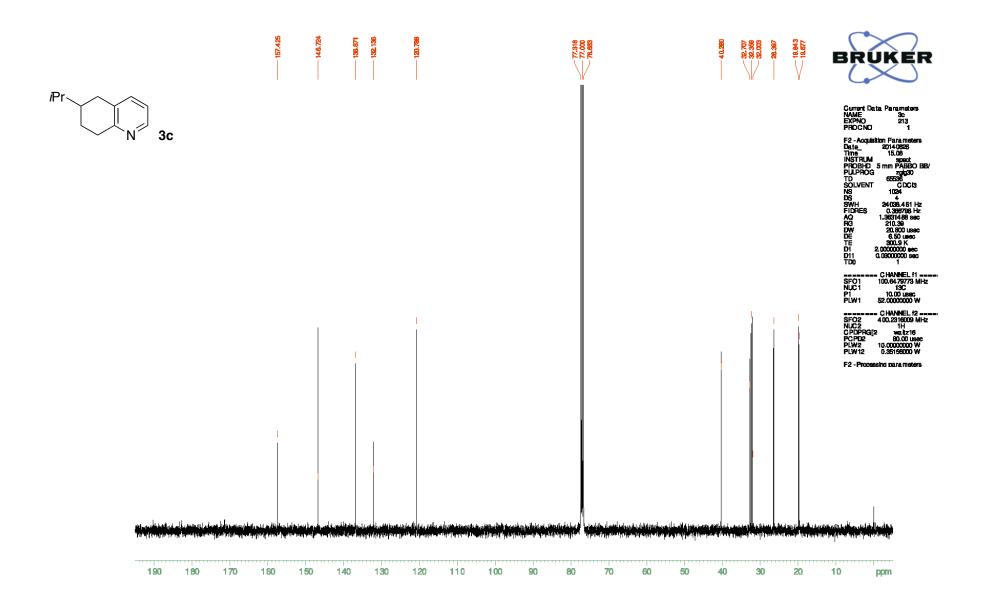


Figure S-16. 13 C $\{^{1}$ H $\}$ NMR spectrum (CDCl₃) of 3c.

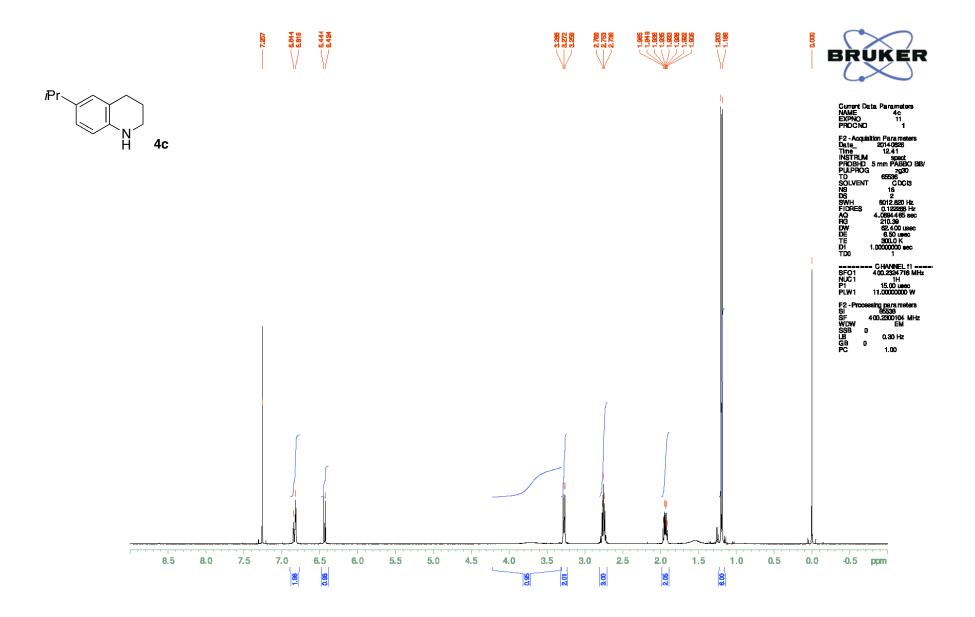


Figure S-17. ¹H NMR spectrum (CDCl₃) of 4c.

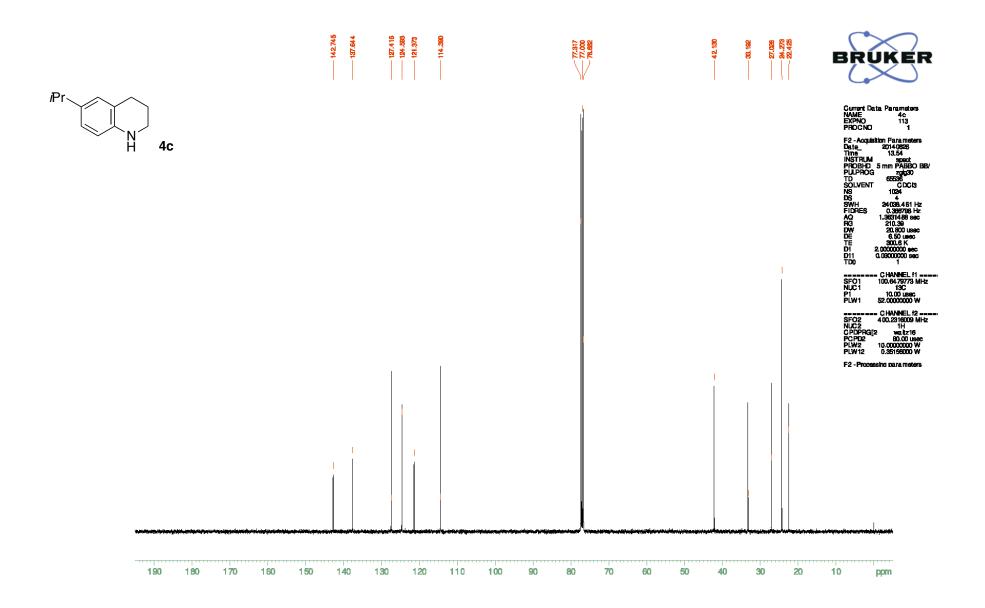


Figure S-18. 13 C $\{^{1}$ H $\}$ NMR spectrum (CDCl₃) of 4c.

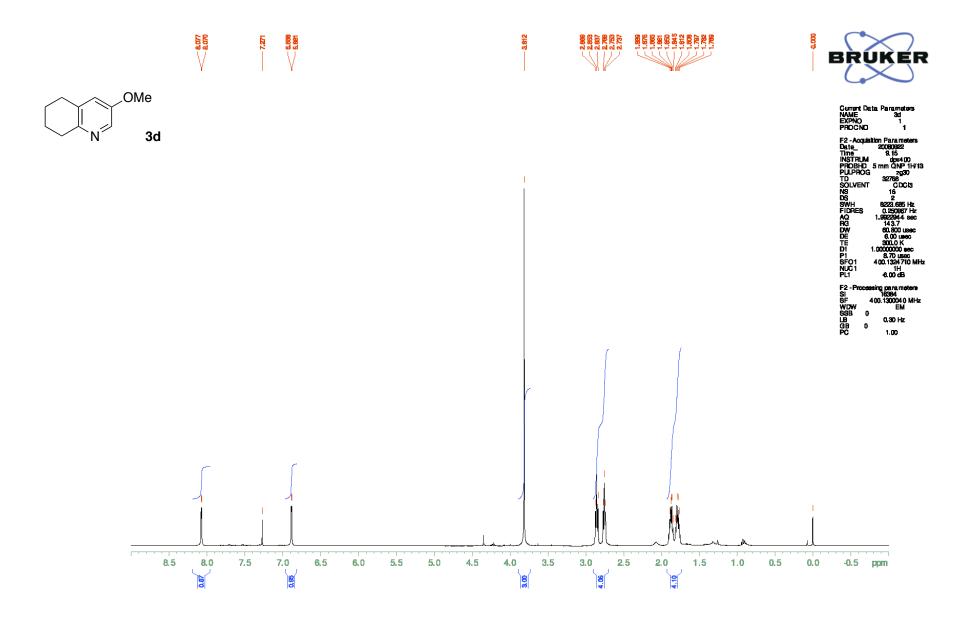


Figure S-19. ¹H NMR spectrum (CDCl₃) of 3d.

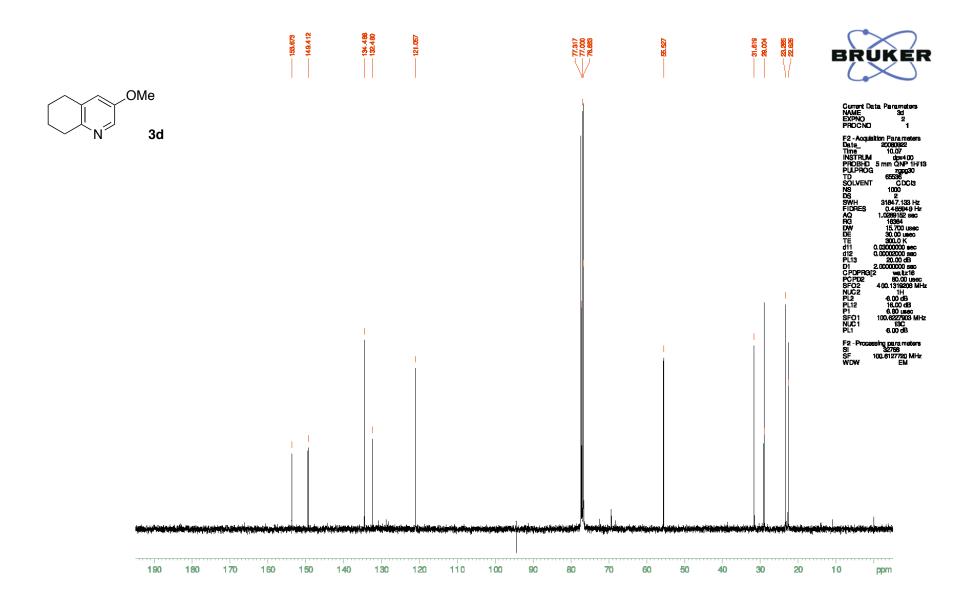


Figure S-20. 13 C $\{^{1}$ H $\}$ NMR spectrum (CDCl₃) of 3d.

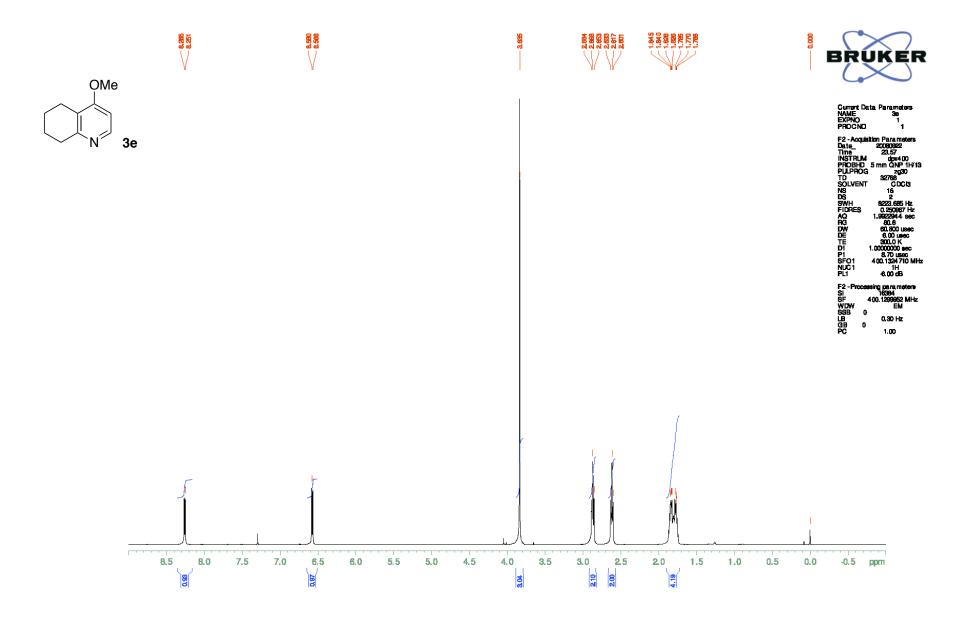


Figure S-21. ¹H NMR spectrum (CDCl₃) of **3e**.

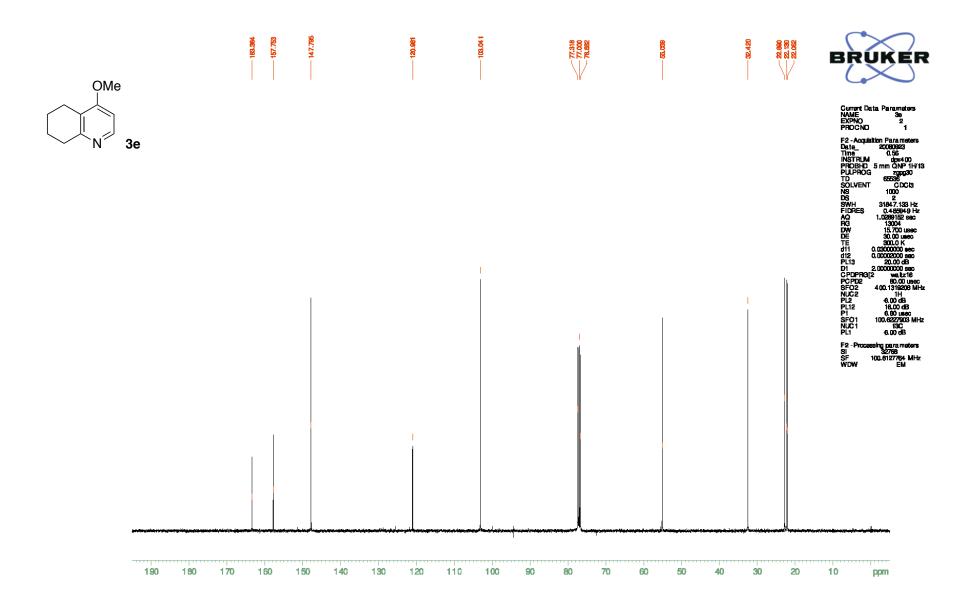


Figure S-22. 13 C $\{^{1}$ H $\}$ NMR spectrum (CDCl₃) of 3e.

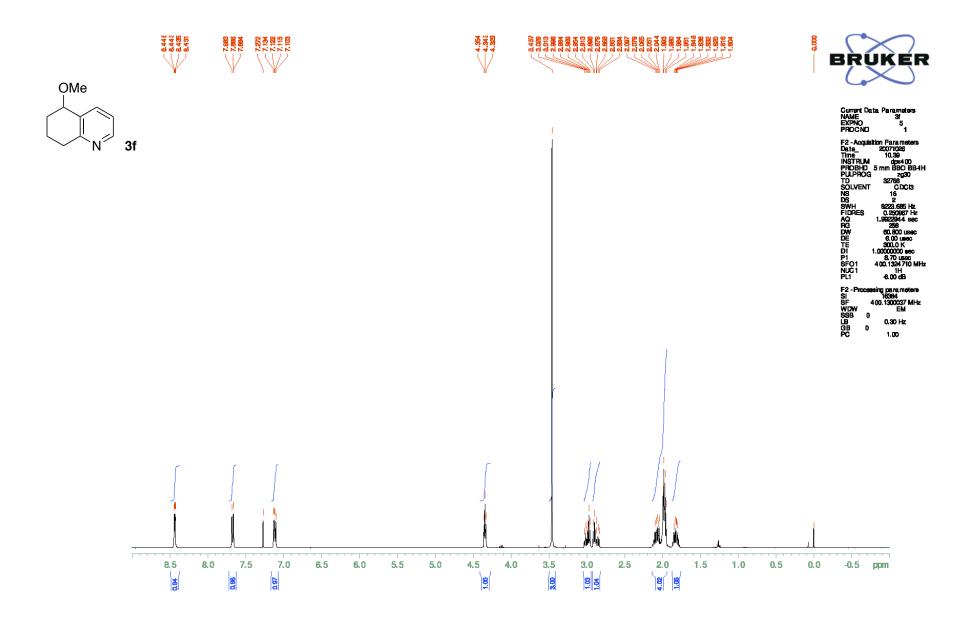


Figure S-23. ¹H NMR spectrum (CDCl₃) of 3f.

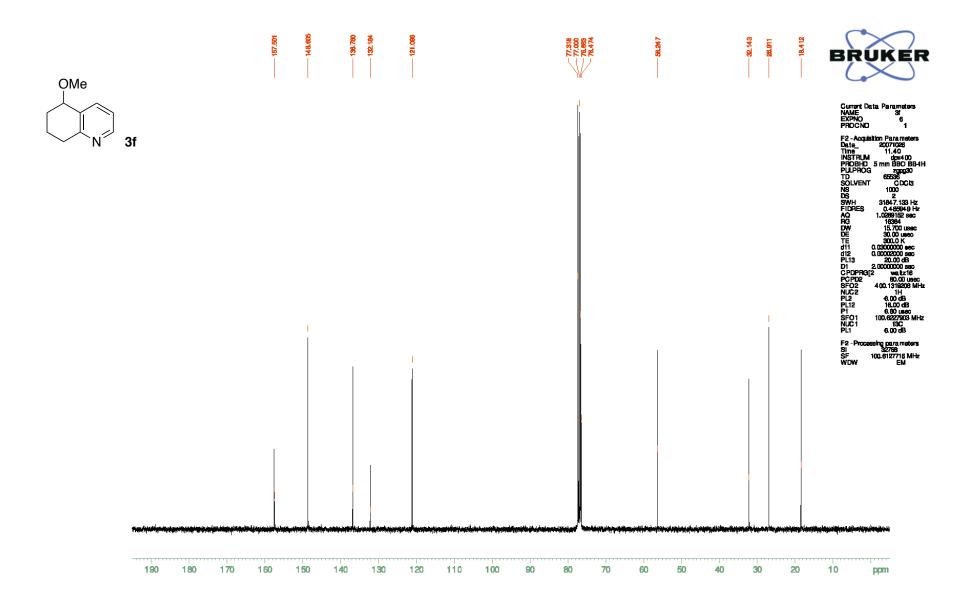


Figure S-24. 13 C $\{^{1}$ H $\}$ NMR spectrum (CDCl₃) of 3f.

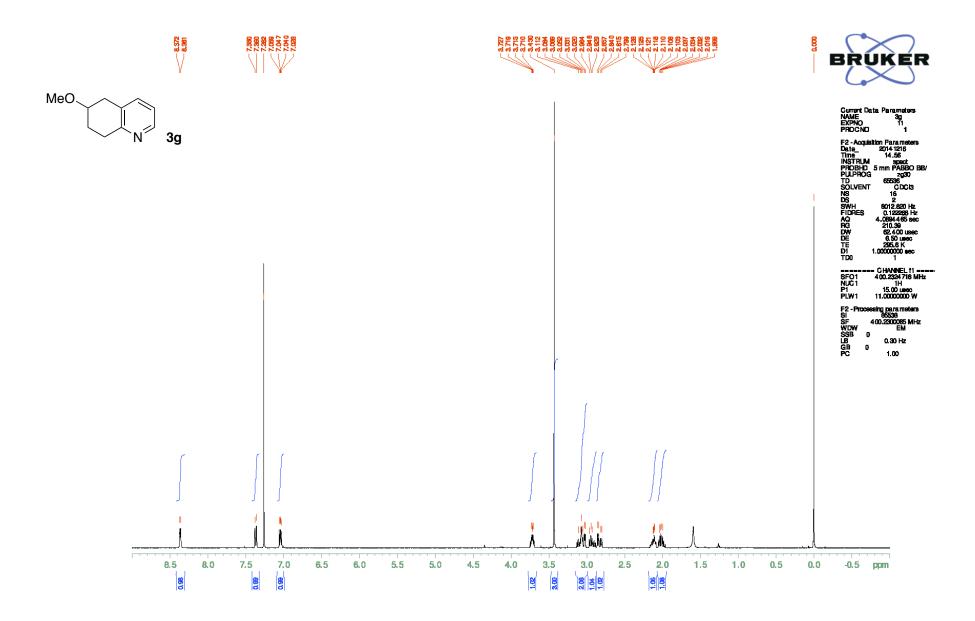


Figure S-25. ¹H NMR spectrum (CDCl₃) of 3g.

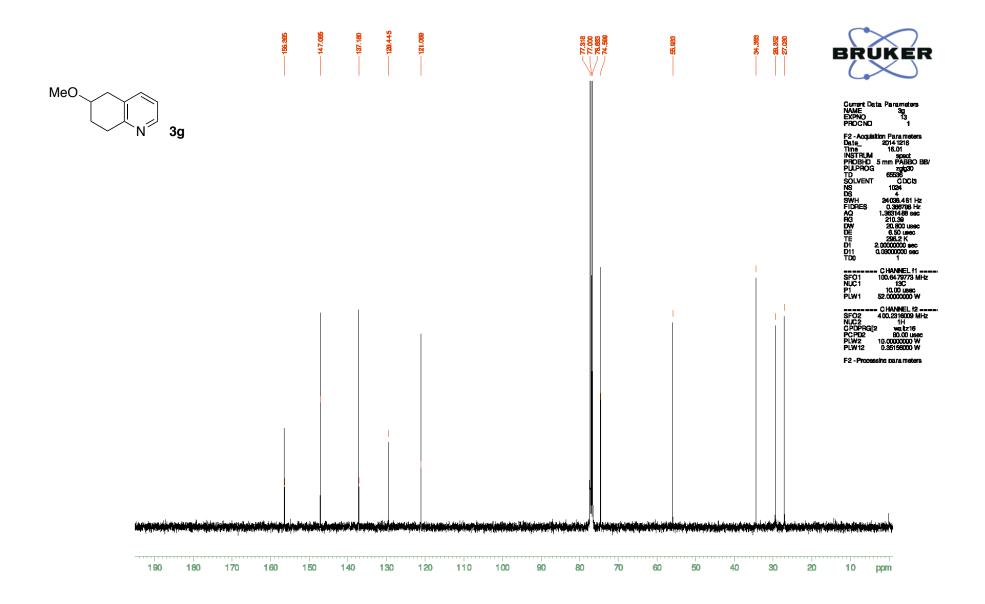


Figure S-26. 13 C $\{^{1}$ H $\}$ NMR spectrum (CDCl₃) of 3g.

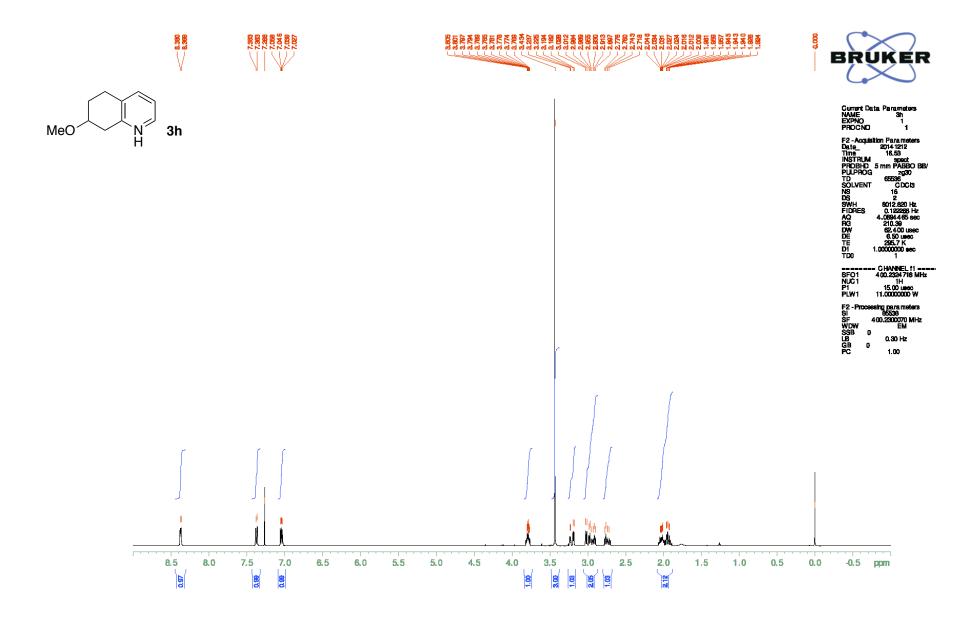


Figure S-27. ¹H NMR spectrum (CDCl₃) of 3h.

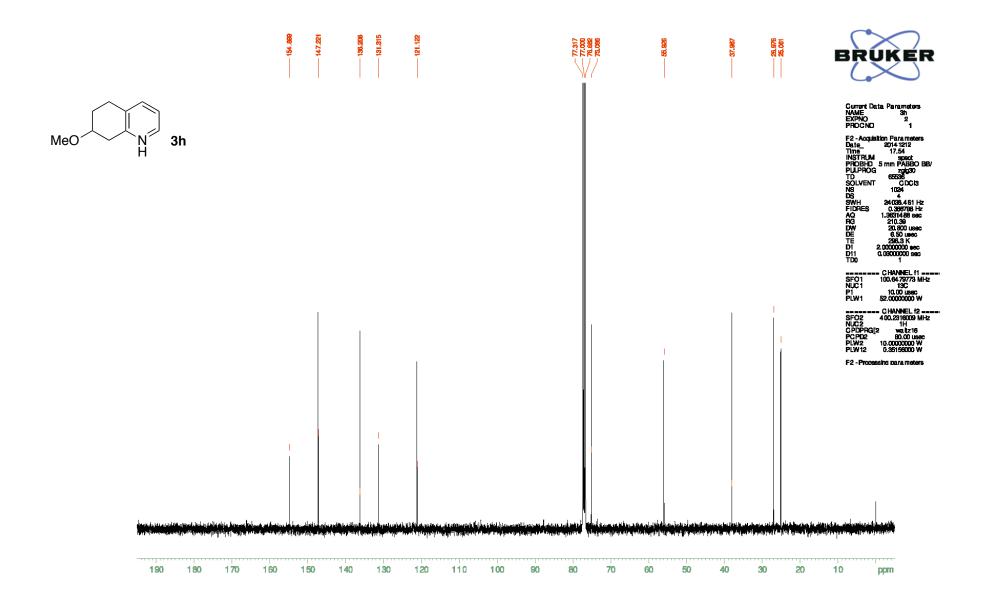


Figure S-28. 13 C $\{^{1}$ H $\}$ NMR spectrum (CDCl₃) of 3h.

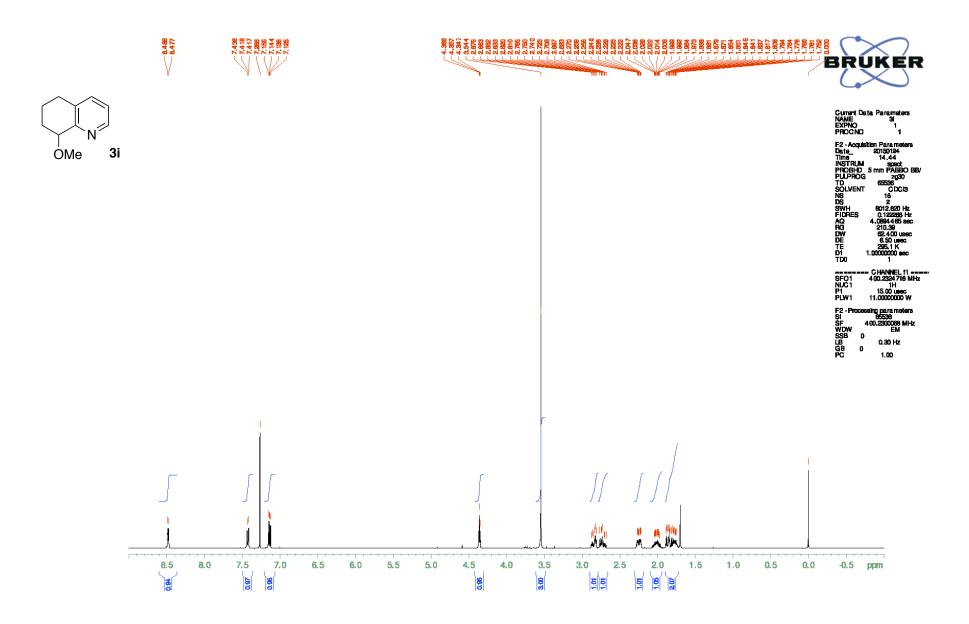


Figure S-29. ¹H NMR spectrum (CDCl₃) of 3i.

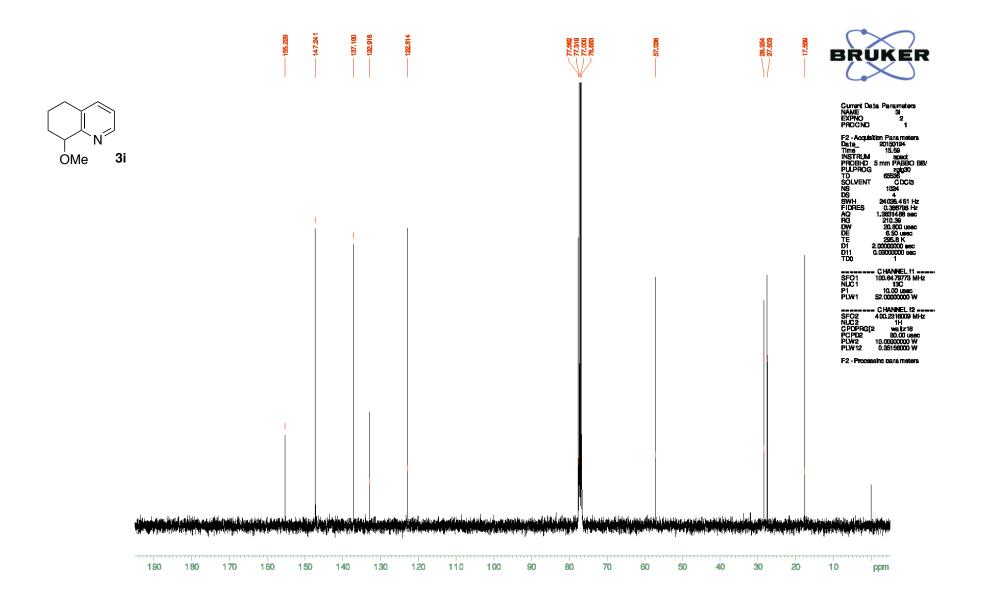


Figure S-30. ${}^{13}C \{{}^{1}H\}$ NMR spectrum (CDCl₃) of 3i.

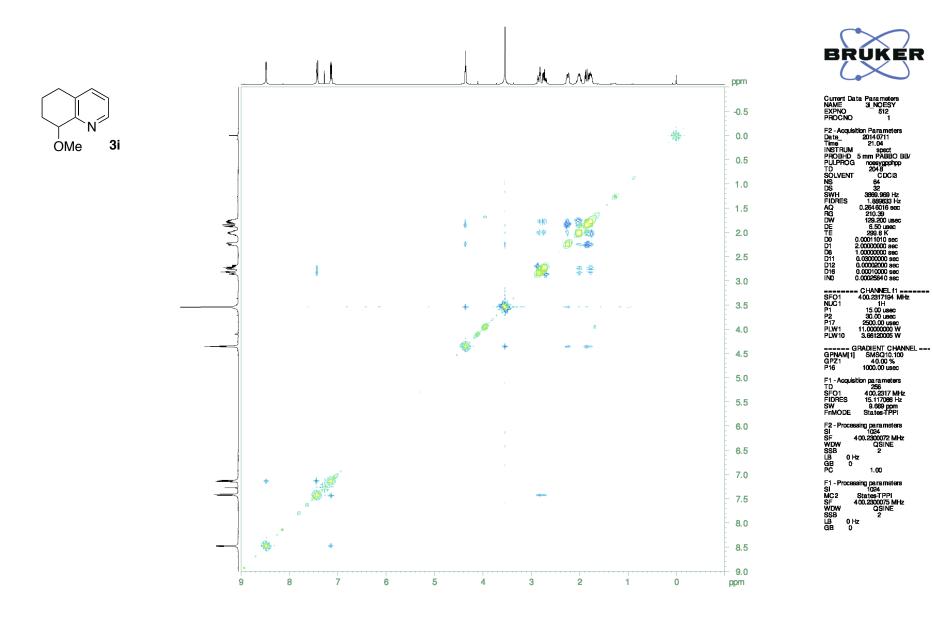
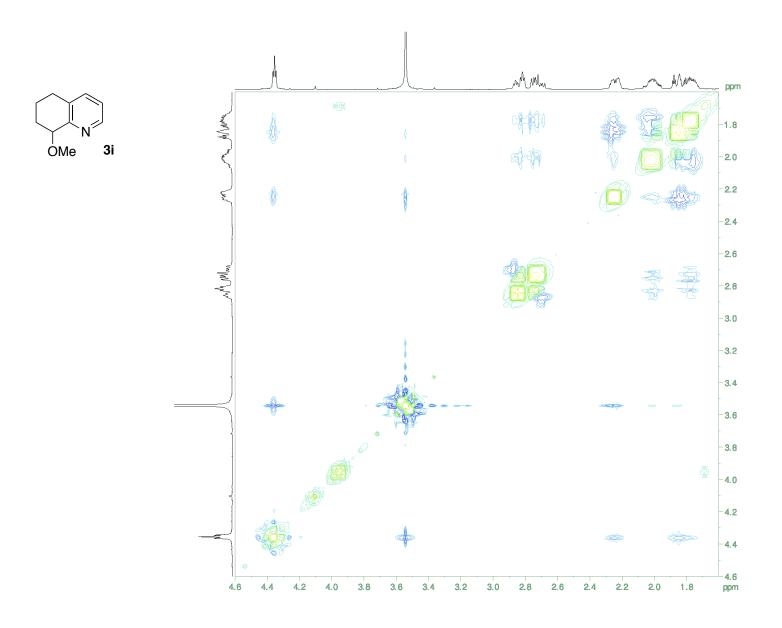


Figure S-31. NOESY spectrum (CDCl₃) of 3i.





Current Data Parameters NAME 3i_NOESY EXPNO 512 PROCNO 1

PROCNO 1
F2 - Acquisition Parameters
Date 2014 0711
Time 21.04
INSTRUM spect
PROSHD 5 mm PABBO BB/
mossygophop
TID VENT
BS 8
BS 808 989 1½
FIDRES 1.88693 99 1½
FIDRES 20,2646016 sec
RG 210.39
DW 128.200 usec
DE 6.50 usec
TE 299 6 K
DD 0.00011010 sec
D12 0.00000000 sec
D10 0.00000000 sec

===== GRADIENT CHANNEL === GPNAM[1] SMSQ10.100 GPZ1 40.00 % P16 1000.00 usec

F1 - Acquisition parameters TD 256 SFO1 400.2317 MHz FIDRES 15.117098 Hz SW 9.669 ppm FnMODE States-TPPI

F2 - Processing parameters
SI 1024
SF 400.2300072 MHz
WDW 2SINE
SSB 2
LB 0 Hz
GB 0
PC 1.00

F1 - Processing parameters SI 1024
MC2 States-TPPI SF 400.2800075 MHz WDW QSINE SB 2 LB 0 Hz GB 0

Figure S-32. NOESY spectrum (CDCl₃) of 3i (δ 1.6–4.6 ppm).

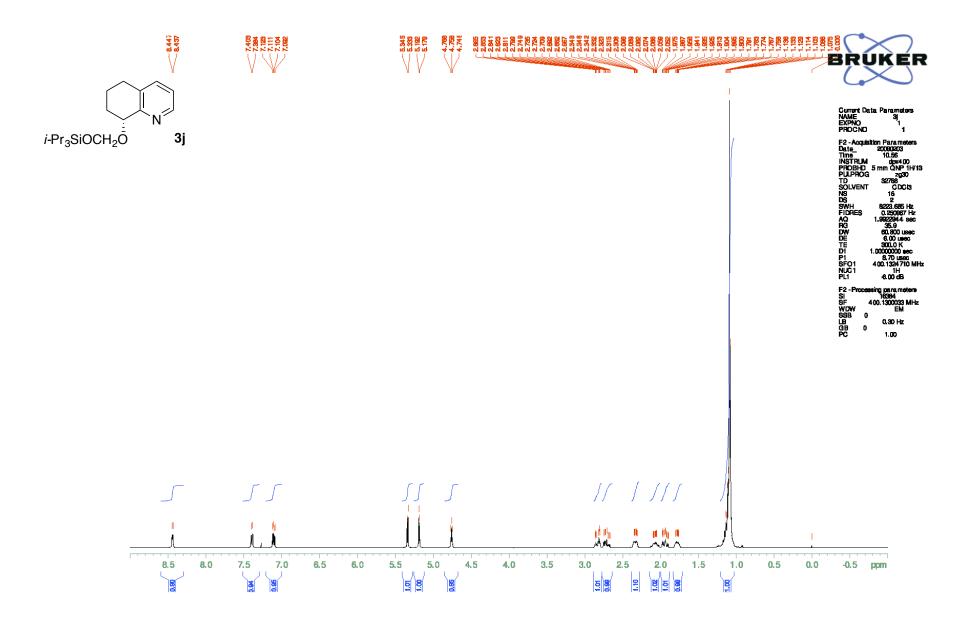


Figure S-33. ¹H NMR spectrum (CDCl₃) of 3j.

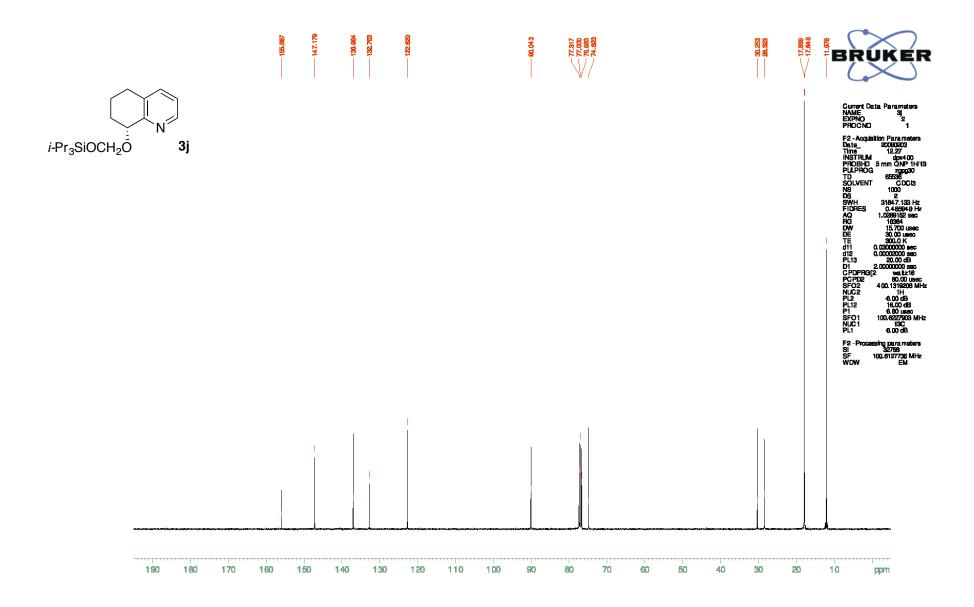


Figure S-34. 13 C $\{^{1}$ H $\}$ NMR spectrum (CDCl₃) of 3j.

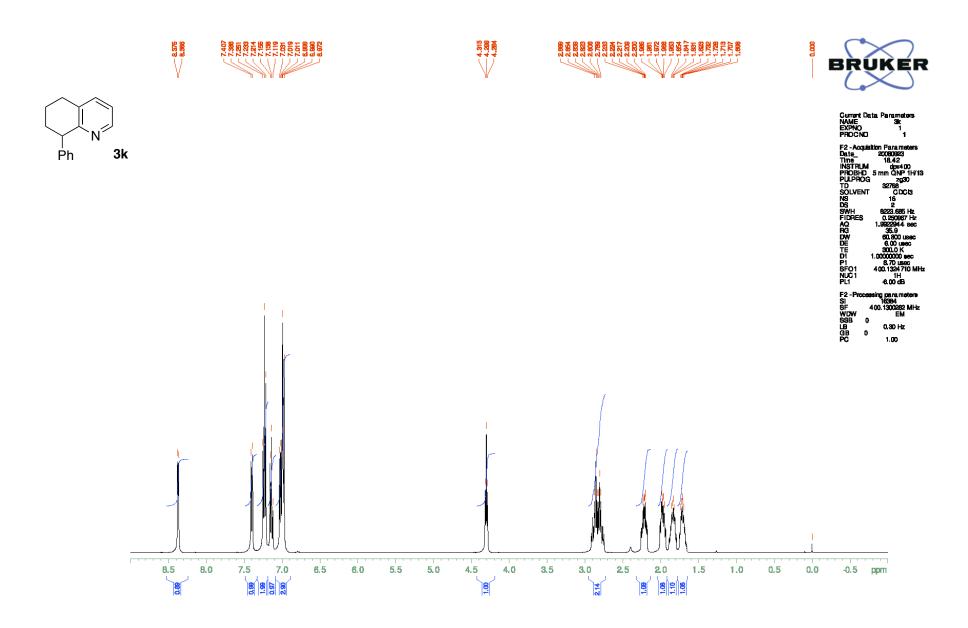


Figure S-35. ¹H NMR spectrum (CDCl₃) of 3k.

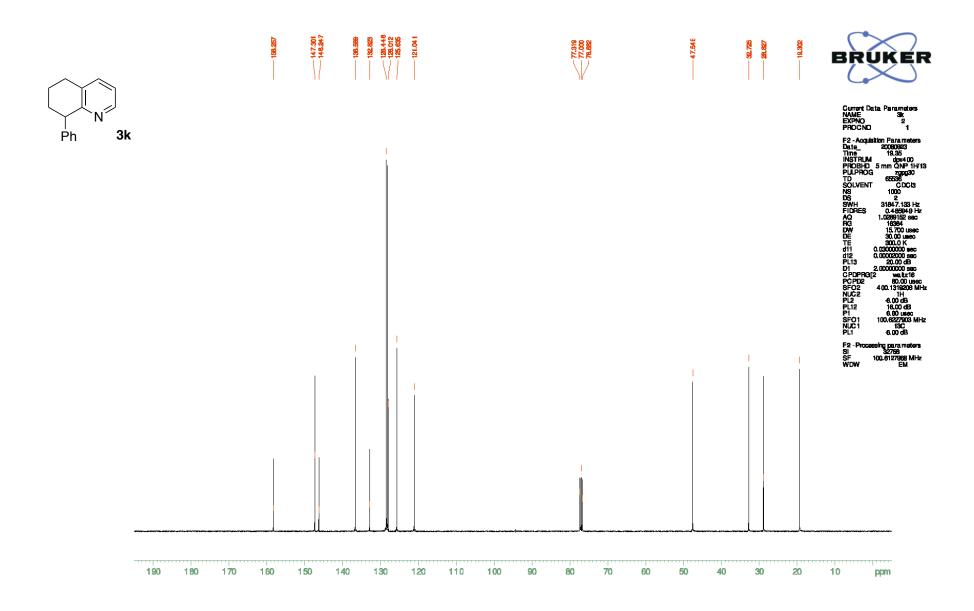


Figure S-36. 13 C $\{^{1}$ H $\}$ NMR spectrum (CDCl₃) of 3k.

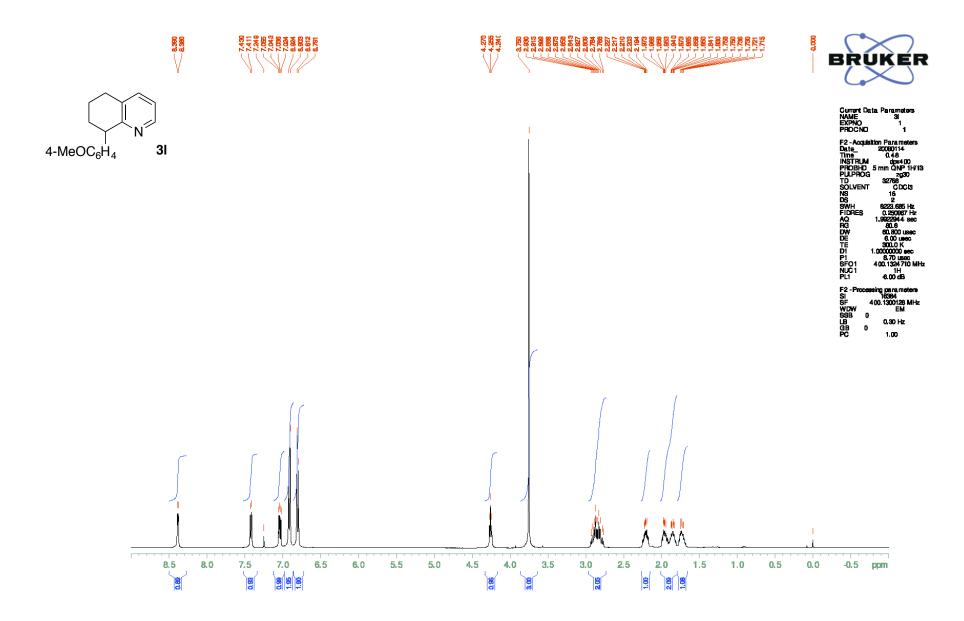


Figure S-37. ¹H NMR spectrum (CDCl₃) of 3l.

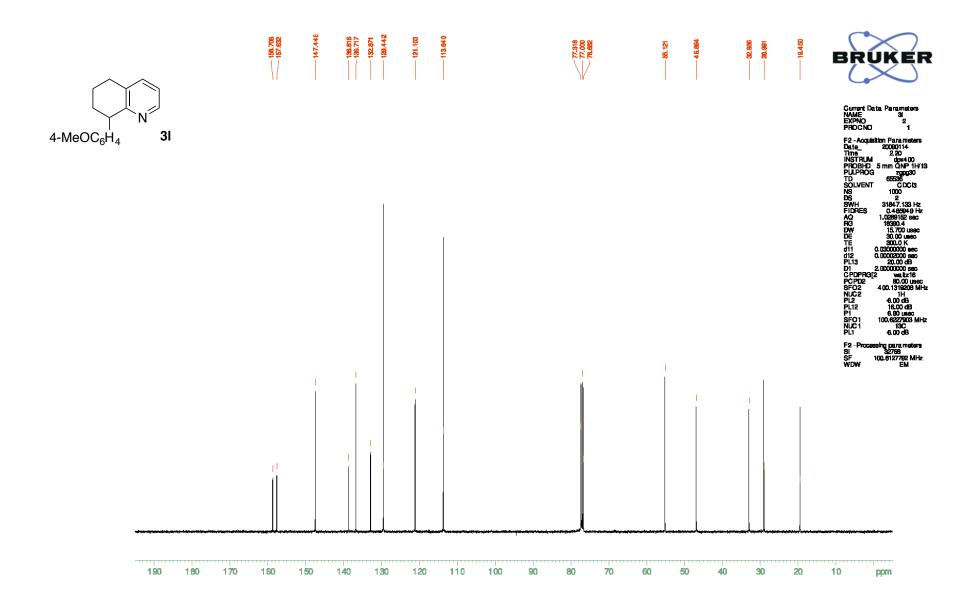


Figure S-38. 13 C $\{^{1}$ H $\}$ NMR spectrum (CDCl₃) of 3l.

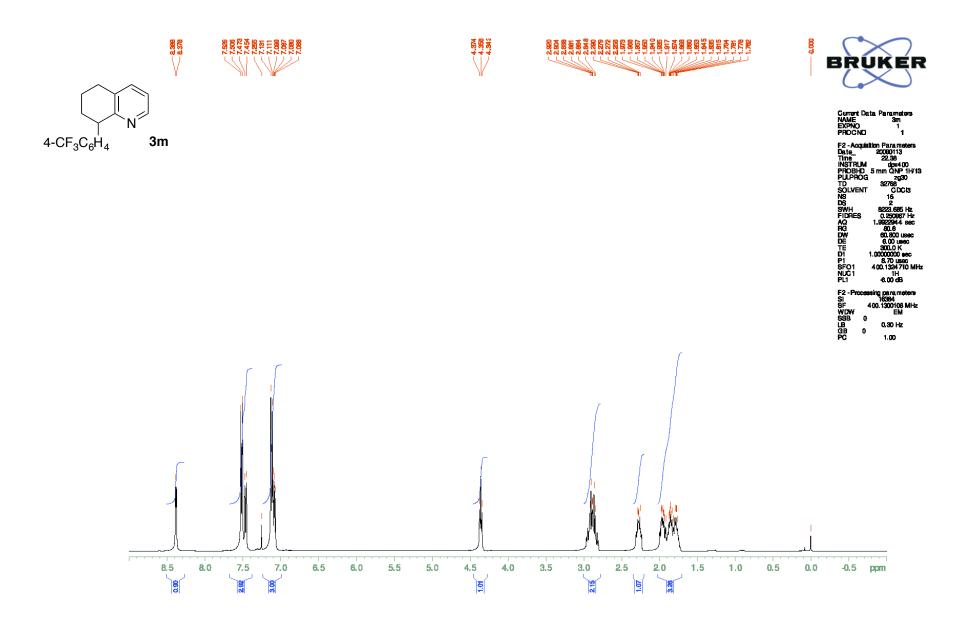


Figure S-39. ¹H NMR spectrum (CDCl₃) of 3m.

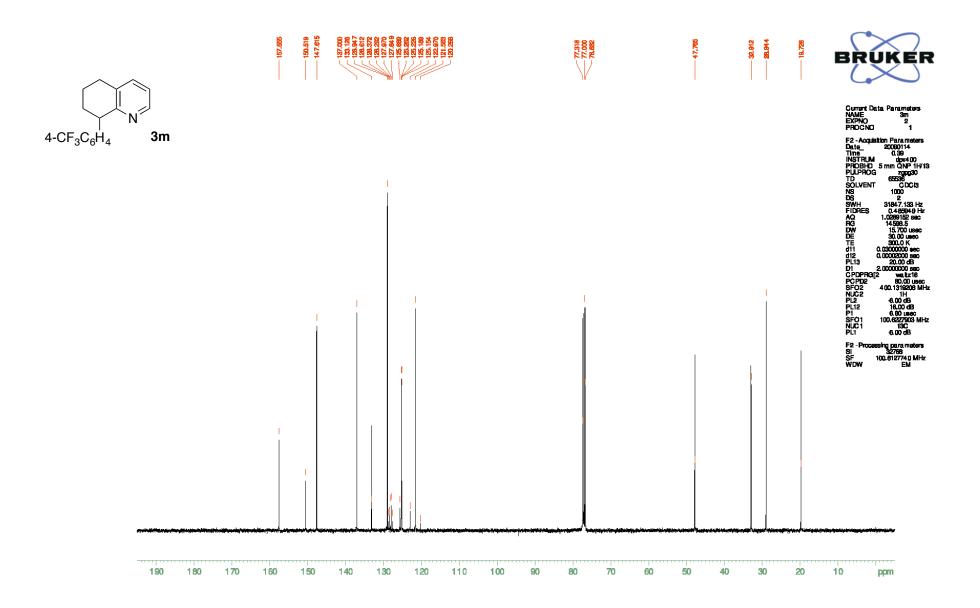


Figure S-40. 13 C $\{^{1}$ H $\}$ NMR spectrum (CDCl₃) of 3m.

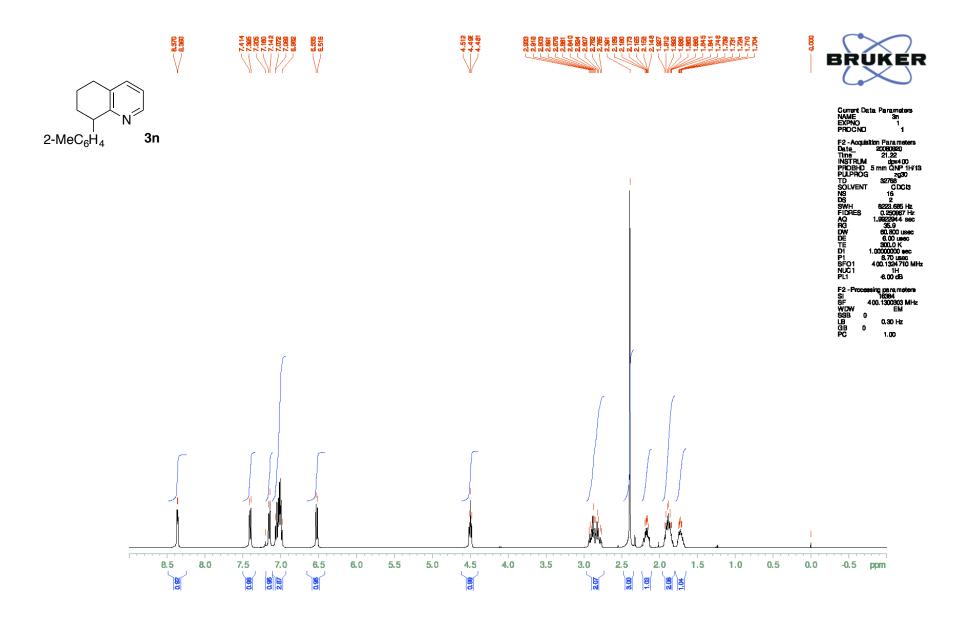


Figure S-41. ¹H NMR spectrum (CDCl₃) of 3n.

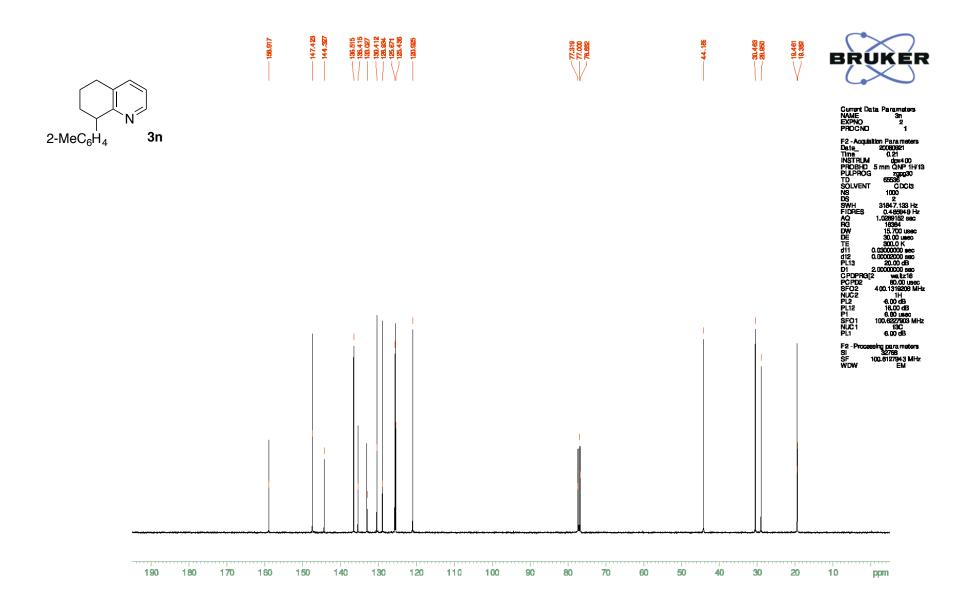


Figure S-42. ¹³C {¹H} NMR spectrum (CDCl₃) of 3n.

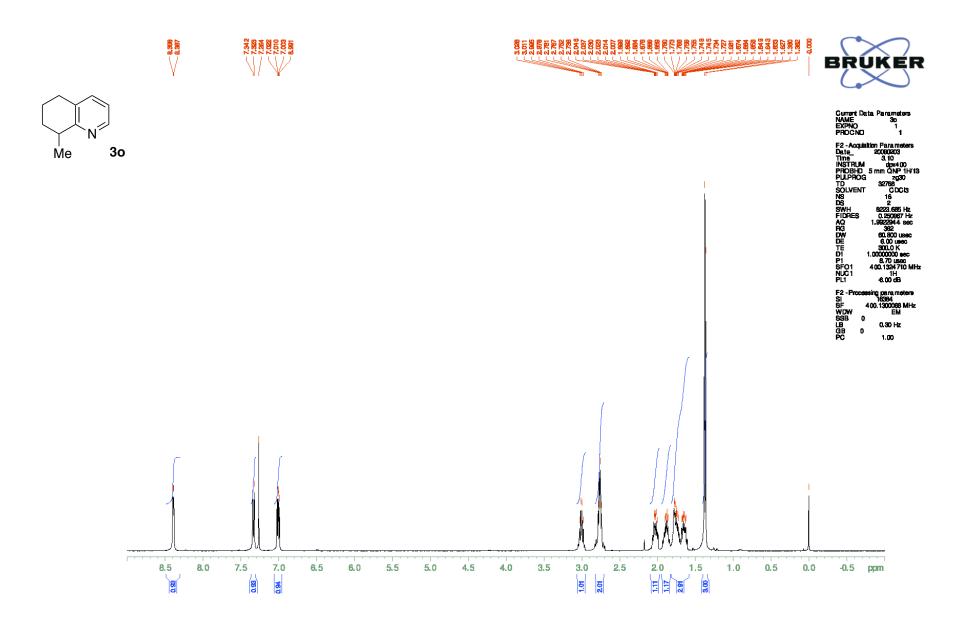


Figure S-43. ¹H NMR spectrum (CDCl₃) of 30.

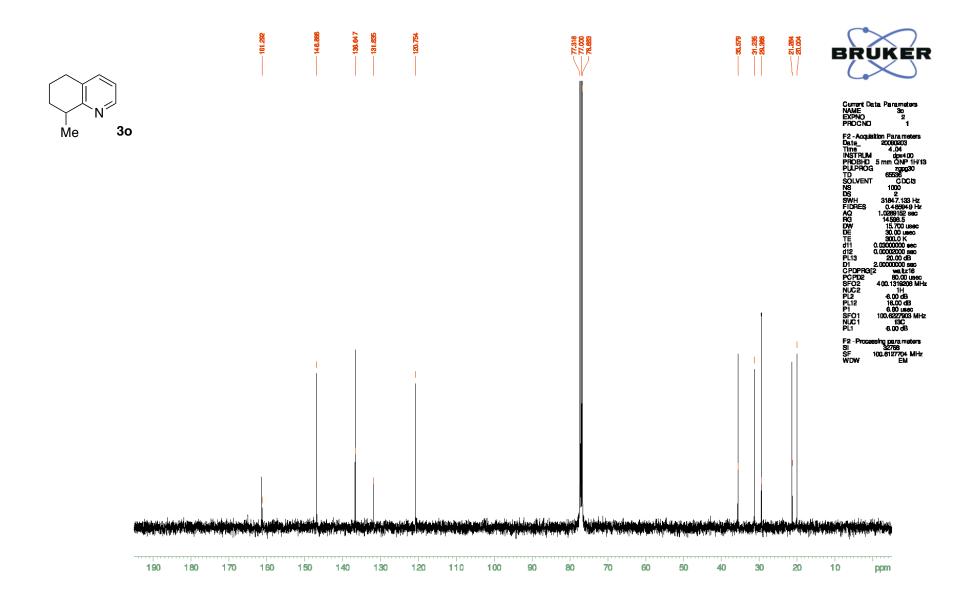


Figure S-44. 13 C $\{^{1}$ H $\}$ NMR spectrum (CDCl₃) of 30.

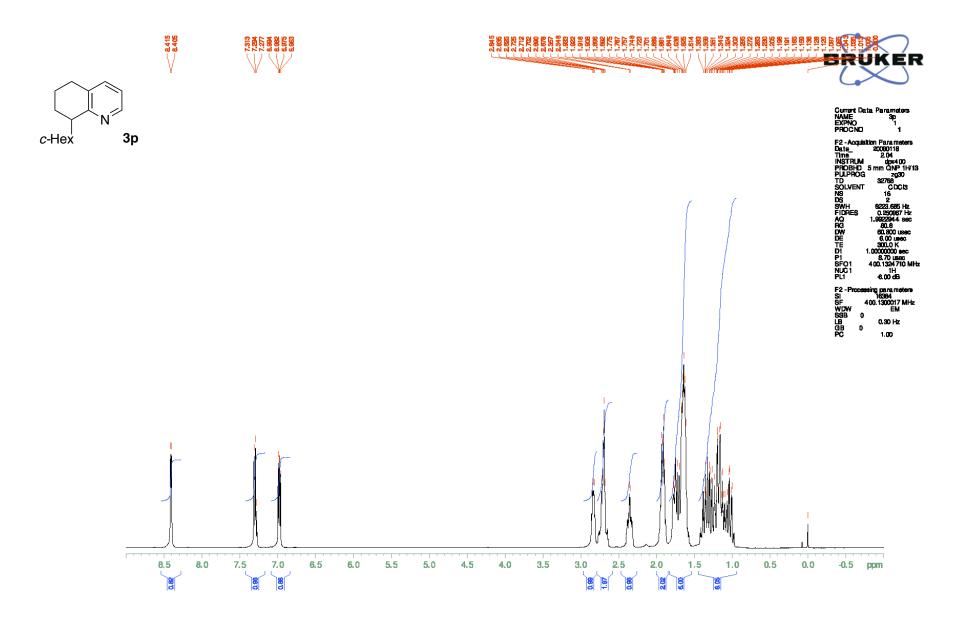


Figure S-45. ¹H NMR spectrum (CDCl₃) of **3p**.

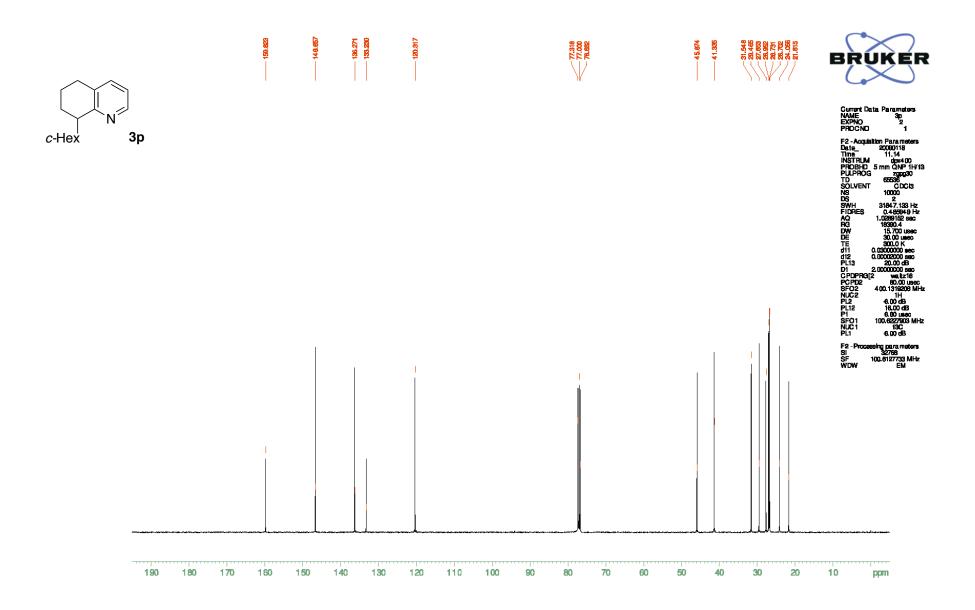


Figure S-46. 13 C $\{^{1}$ H $\}$ NMR spectrum (CDCl₃) of **3p**.

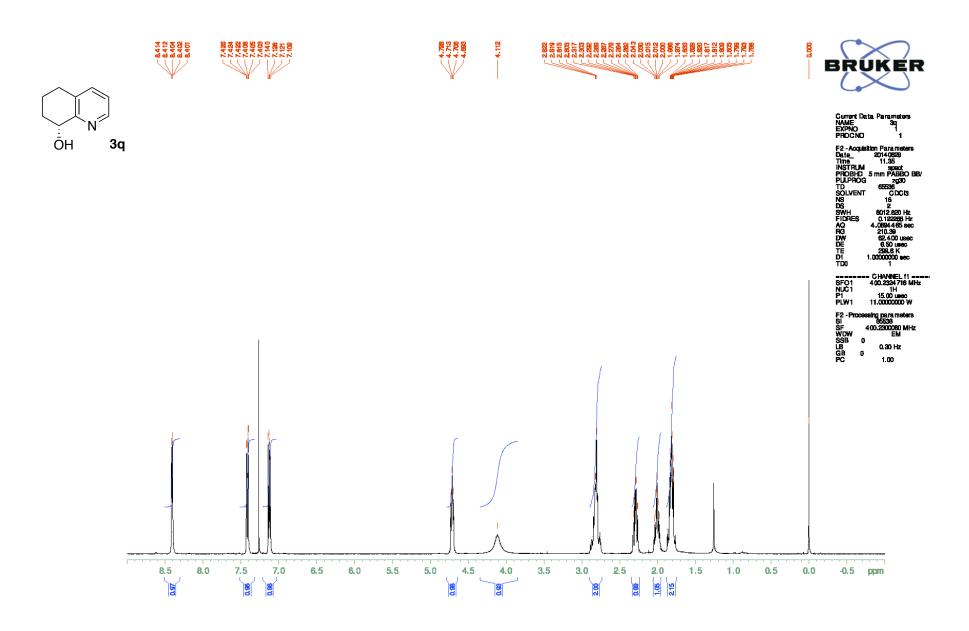


Figure S-47. ¹H NMR spectrum (CDCl₃) of 3q.

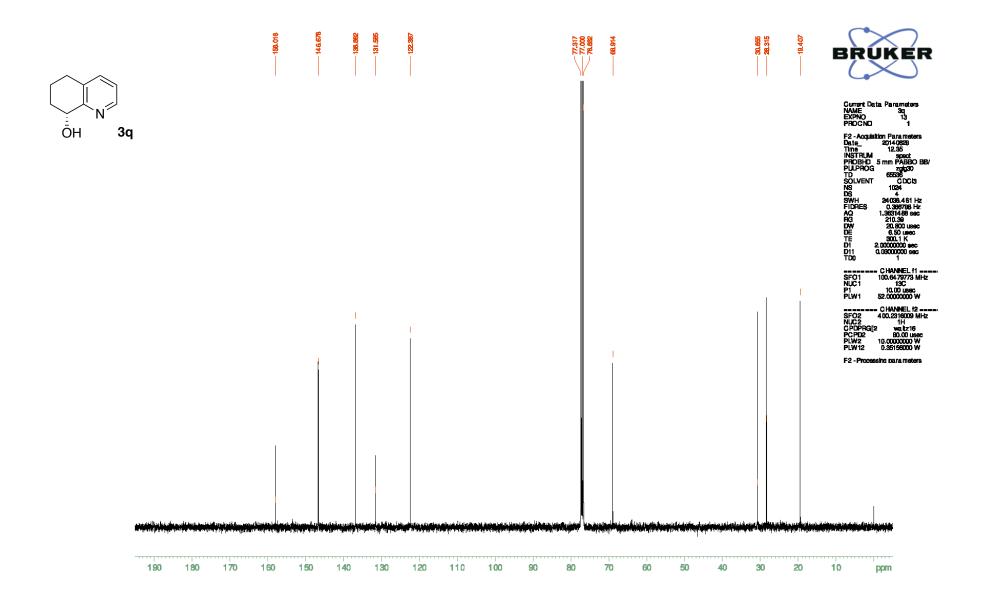


Figure S-48. 13 C $\{^{1}$ H $\}$ NMR spectrum (CDCl₃) of 3q.

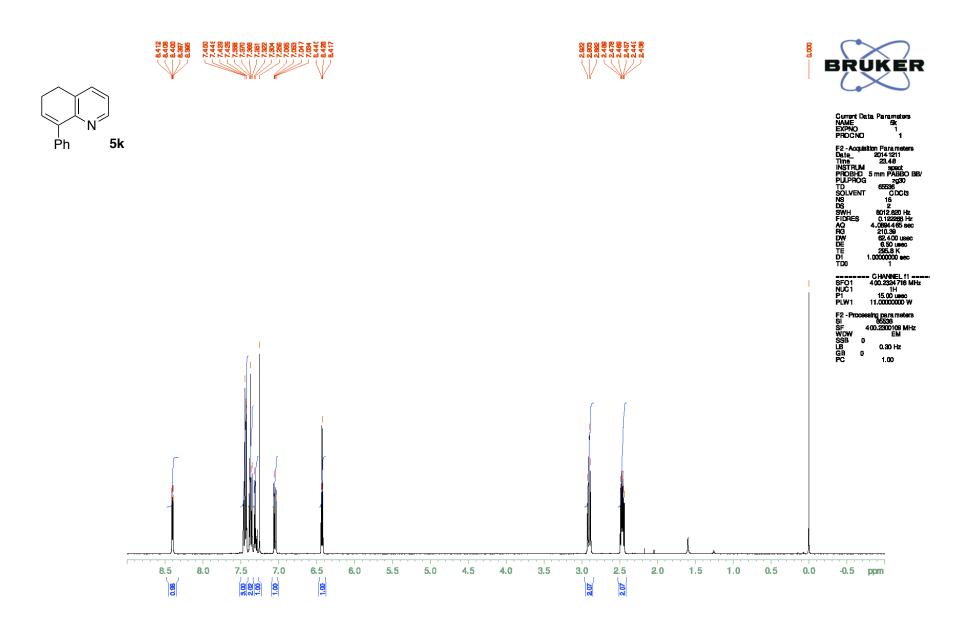


Figure S-49. ¹H NMR spectrum (CDCl₃) of **5k**.

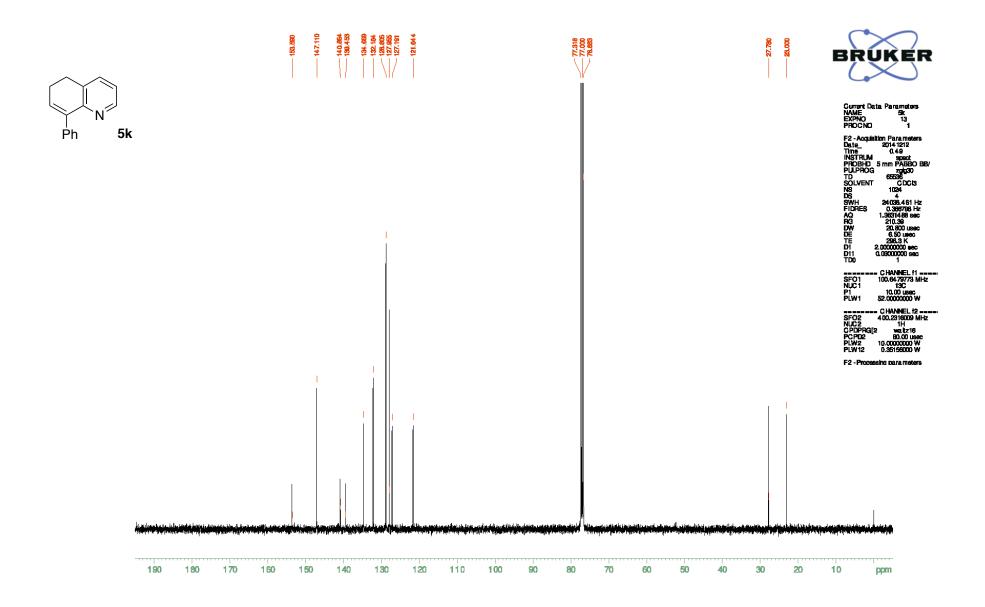


Figure S-50. 13 C $\{^{1}$ H $\}$ NMR spectrum (CDCl₃) of 5k.

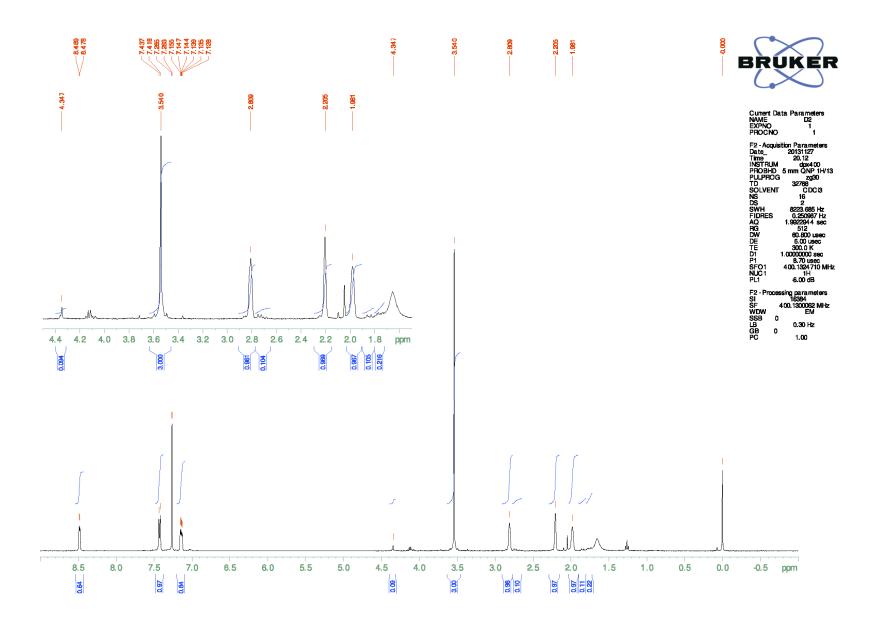


Figure S-51. ¹H NMR spectrum (CDCl₃) of 3i-d, which was obtained from the deuteration of 2i.

The charts of chiral HPLC analyses are given in the following pages. The translation of the Japanese words in column headings is given in figures below.

Type 1

Results								
計算結果	Name of peak	Integral	Integral (%)	Height	Calibration	S	ymmetry S	eparation
No. Rt(min)	ピーク名	面積	面積(%)	高さ	定量結果	NTP	対称性	分離度
1 9.51		49055, 200	50. 9759	3039		7810. 3	1. 314	2.777
2 10.76		47177. 000	49. 0241	2676		8384. 9	1. 276	
		96232, 200	100.0000	5715				

Type 2

3: 268 nm, 4 nm結果 Name of peak Pk # Results 名前	Retention time	Integral	Integral, %	Baseline code
	保持時間	面積	面積%	ヘ・ースラインコート・
1 2	15. 05	13076881	34. 594	BI
	17. 12	24723711	65. 406	BI
トータル Total		37800592	100.000	

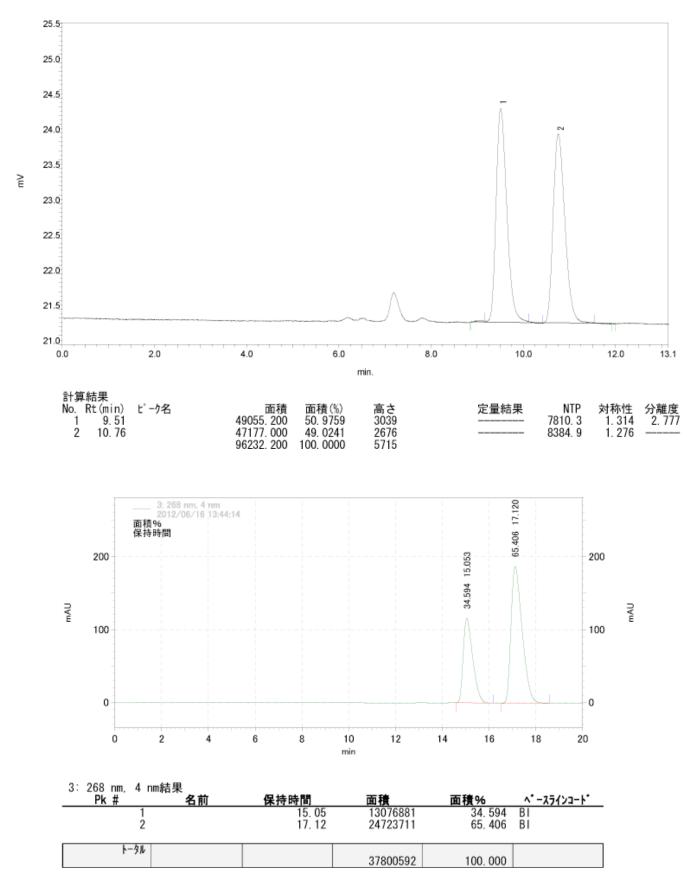


Figure S-52. The charts of chiral HPLC analyses of racemic **3b** (upper), and the hydrogenation product obtained from entry 6 of Table 1 (lower).

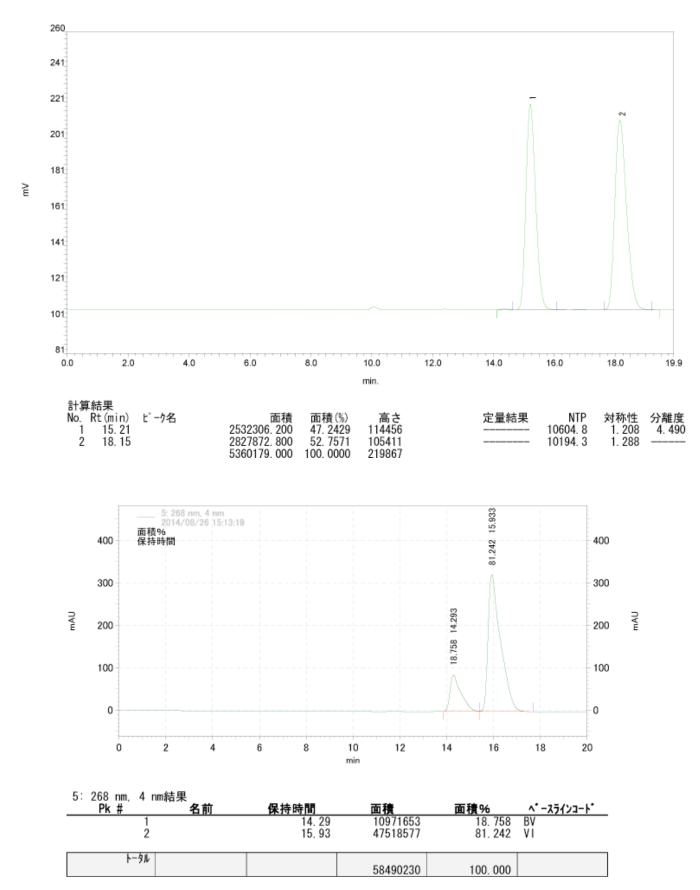


Figure S-53. The charts of chiral HPLC analyses of racemic **3c** (upper), and the hydrogenation product obtained from entry 12 of Table 1 (lower).

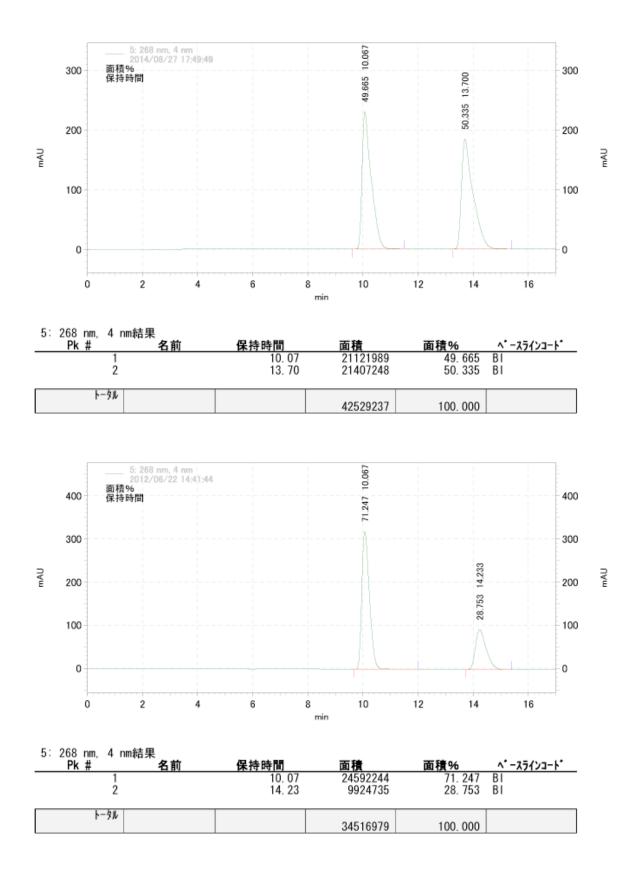


Figure S-54. The charts of chiral HPLC analyses of racemic **3f** (upper), and the hydrogenation product obtained from entry 3 of Table 2 (lower).

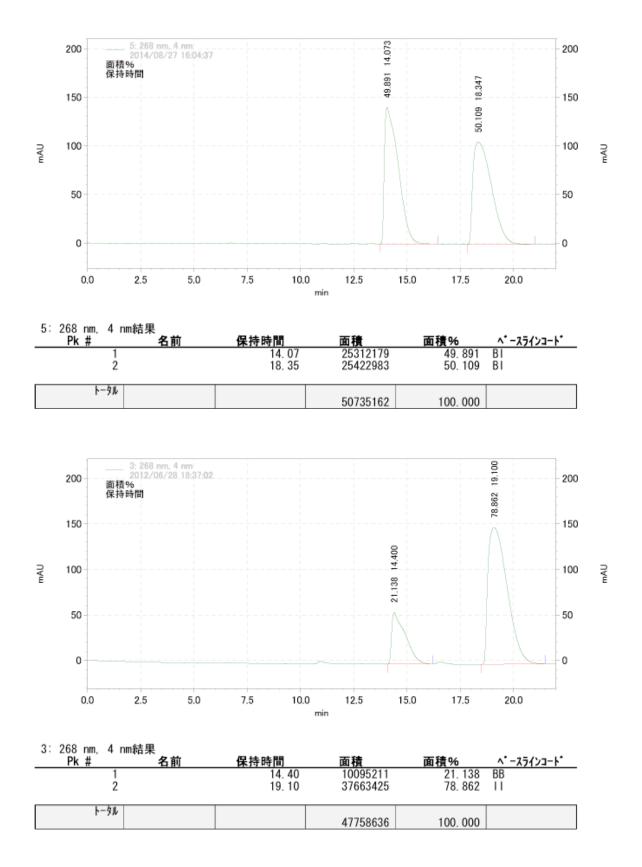


Figure S-55. The charts of chiral HPLC analyses of racemic **3g** (upper), and the hydrogenation product obtained from entry 4 of Table 2 (lower).

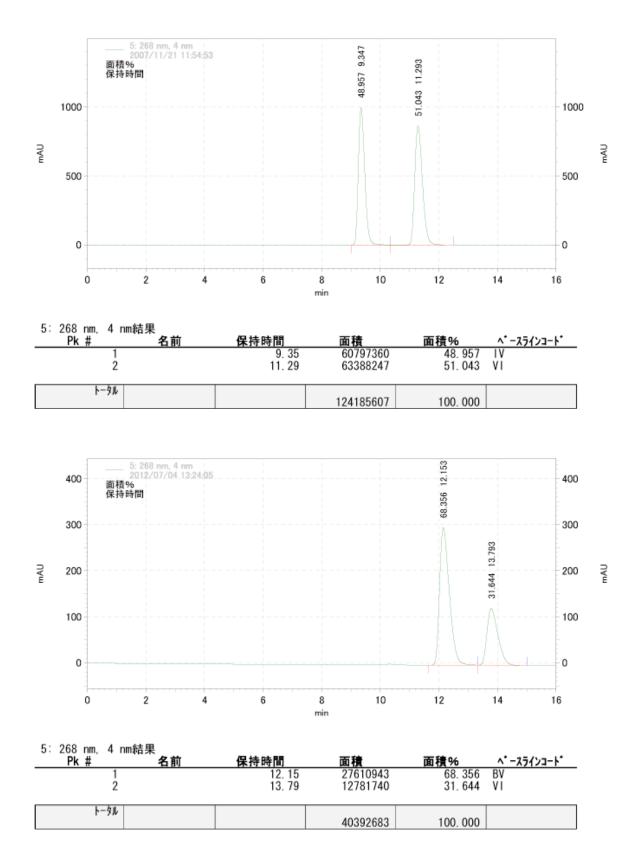


Figure S-56. The charts of chiral HPLC analyses of racemic **3h** (upper), and the hydrogenation product obtained from entry 5 of Table 2 (lower).

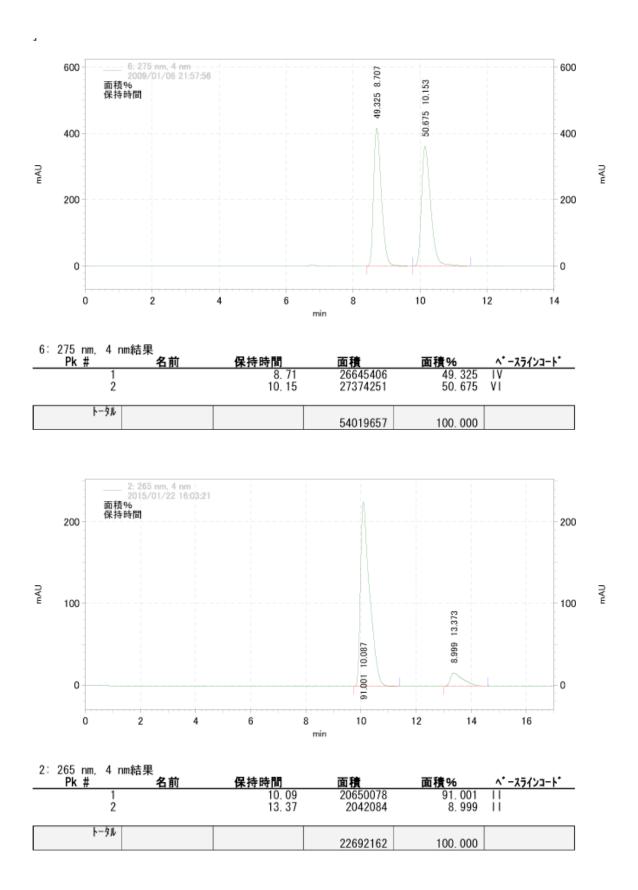


Figure S-57. The charts of chiral HPLC analyses of racemic **3i** (upper), and the hydrogenation product obtained from entry 8 of Table 2 (lower).

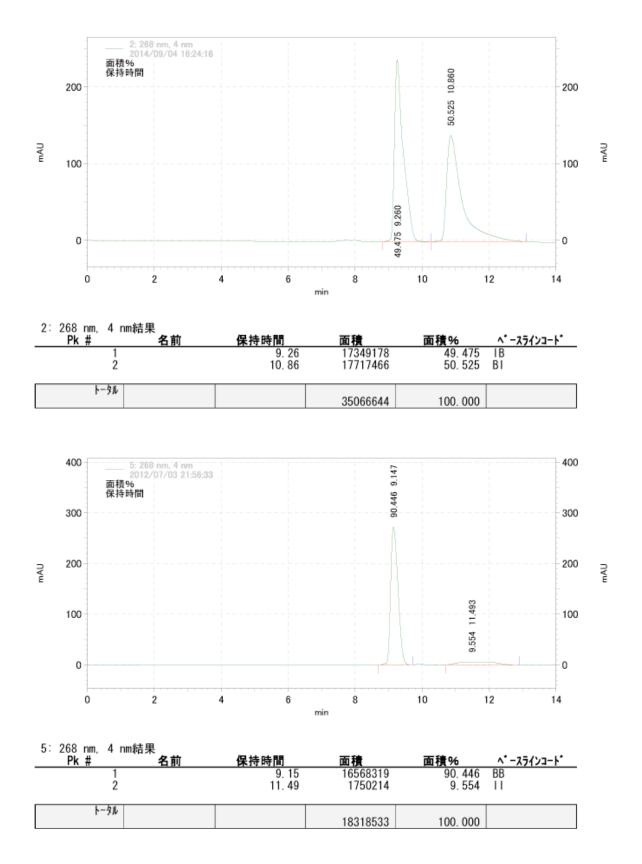


Figure S-58. The charts of chiral HPLC analyses of racemic **3j** (upper), and the hydrogenation product obtained from entry 1 of Table 3 (lower).

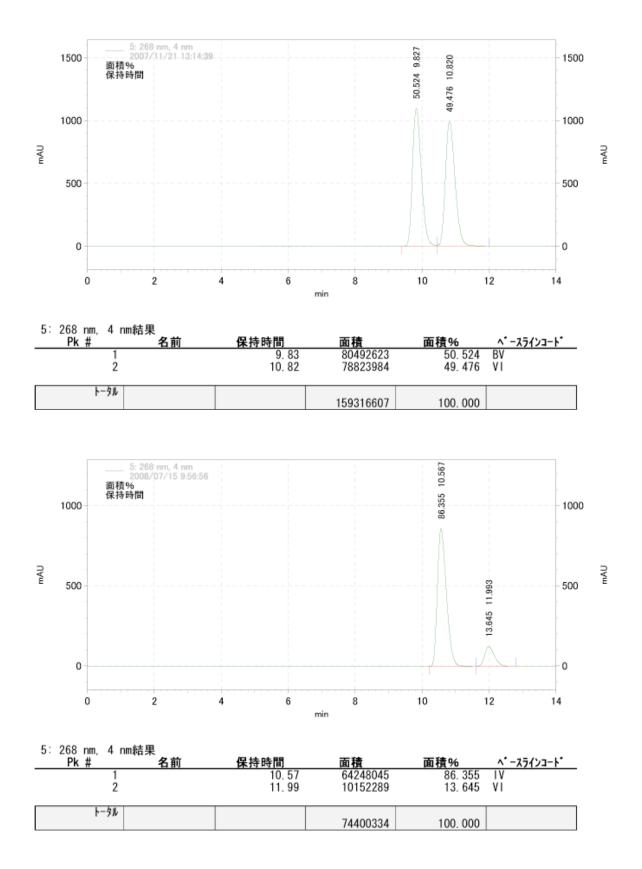


Figure S-59. The charts of chiral HPLC analyses of racemic **3k** (upper), and the hydrogenation product obtained from entry 2 of Table 3 (lower).

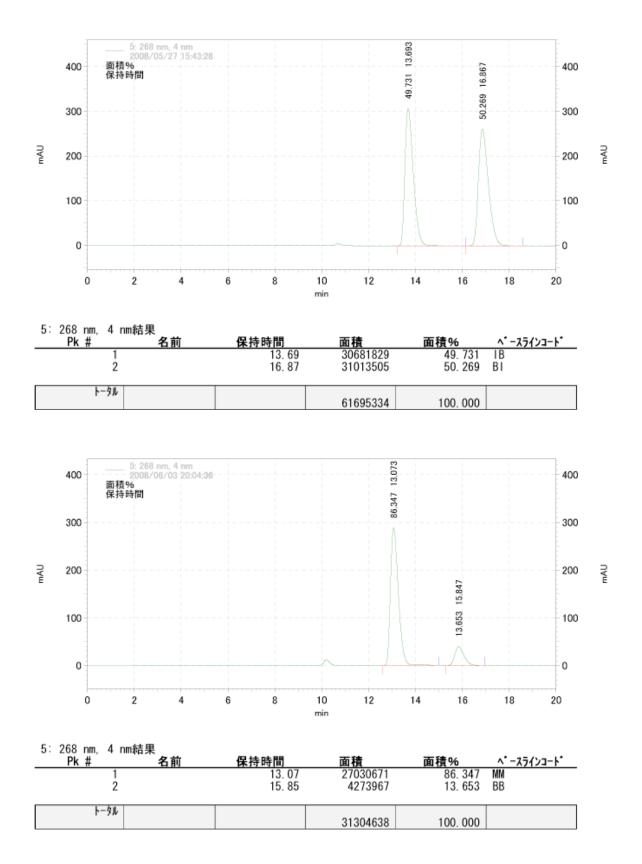


Figure S-60. The charts of chiral HPLC analyses of racemic **31** (upper), and the hydrogenation product obtained from entry 3 of Table 3 (lower).

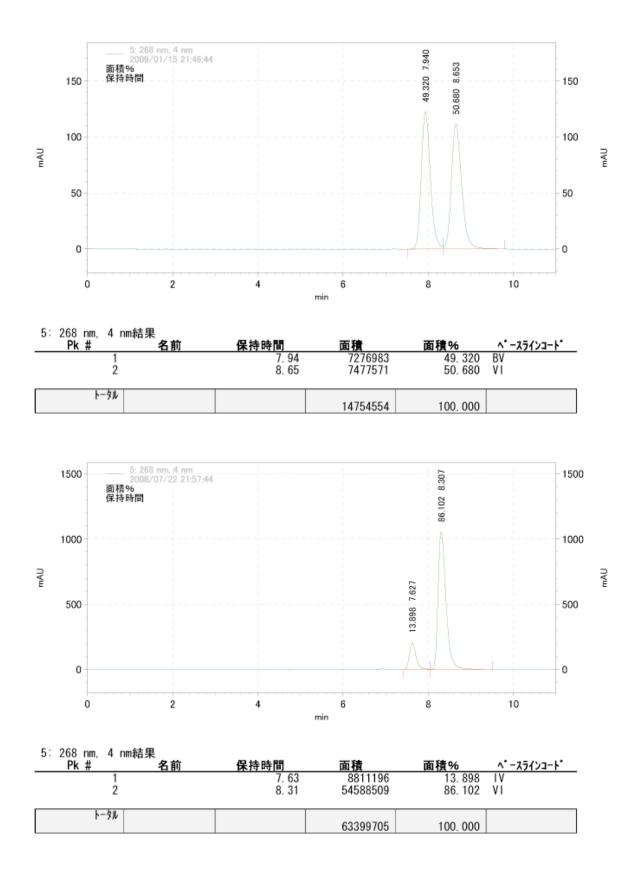


Figure S-61. The charts of chiral HPLC analyses of racemic **3m** (upper), and the hydrogenation product obtained from entry 4 of Table 3 (lower).

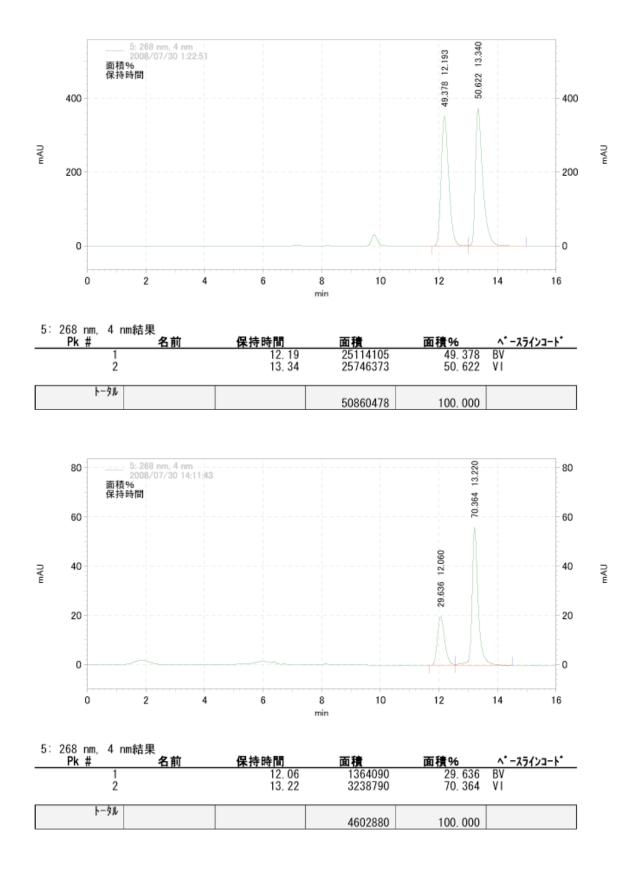


Figure S-62. The charts of chiral HPLC analyses of racemic **3n** (upper), and the hydrogenation product obtained from entry 5 of Table 3 (lower).

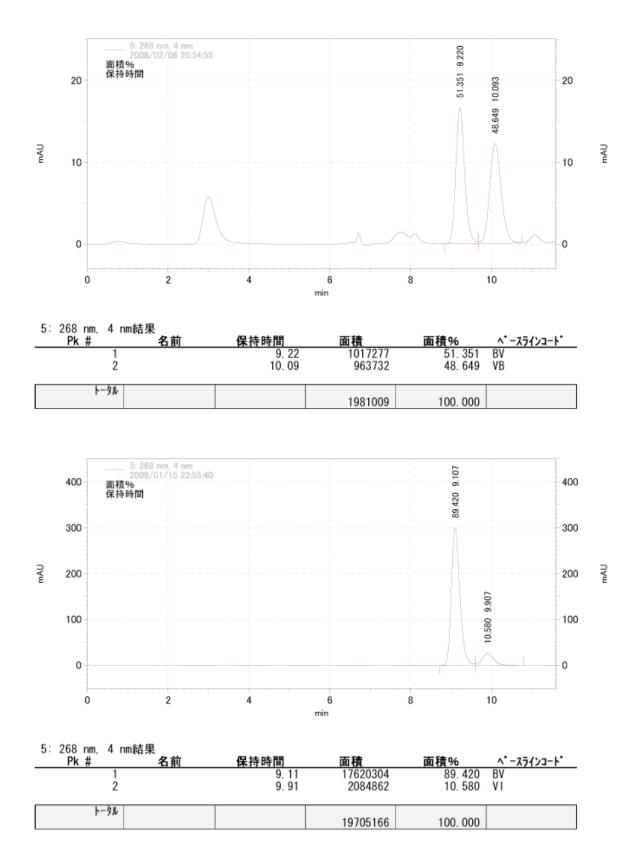


Figure S-63. The charts of chiral HPLC analyses of racemic **30** (upper), and the hydrogenation product obtained from entry 7 of Table 3 (lower).

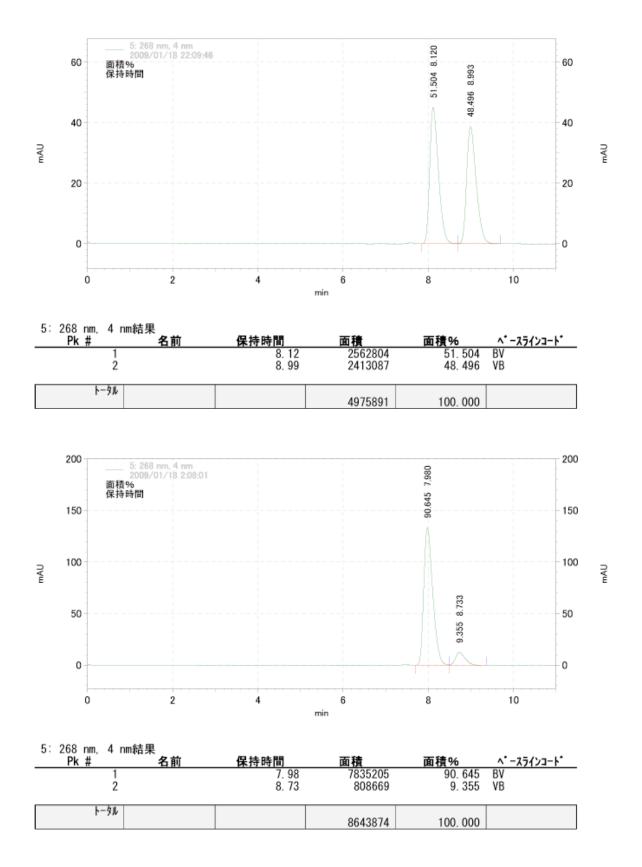


Figure S-64. The charts of chiral HPLC analyses of racemic **3p** (upper), and the hydrogenation product obtained from entry 8 of Table 3 (lower).

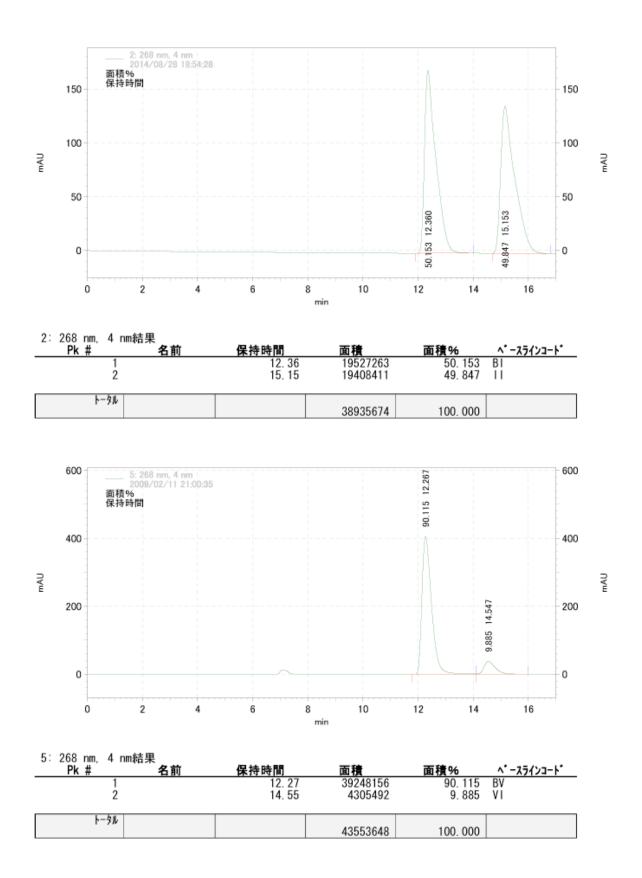


Figure S-65. The charts of chiral HPLC analyses of racemic **3q** (upper), and the product obtained from the deprotection of **3j** in eq 2 (lower).

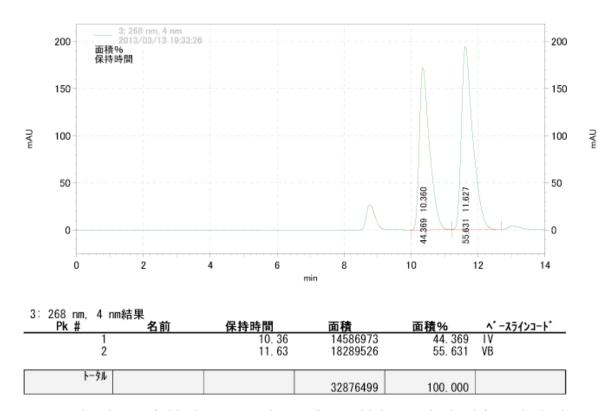


Figure S-66. The charts of chiral HPLC analyses of **3k**, which was obtained from the hydrogenation of **5k** in eq 3.

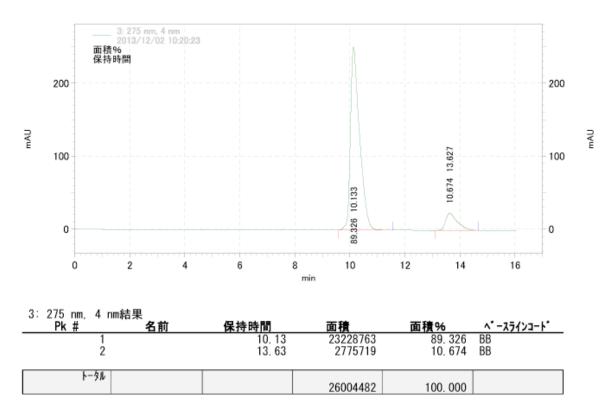


Figure S-67. The charts of chiral HPLC analyses of **3i-d**, which obtained from the deuteration of **2i** in eq 4.

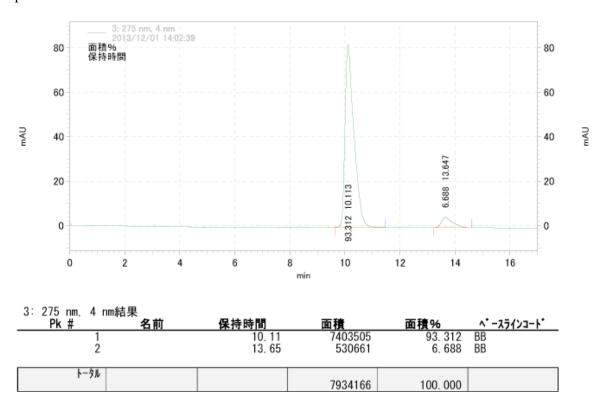


Figure S-68. The charts of chiral HPLC analyses of **3i**, which obtained from the hydrogenation of **2i** under 1.0 MPa of H₂.