

Supporting Information

CFBSA: A Novel and Practical Chlorinating Reagent

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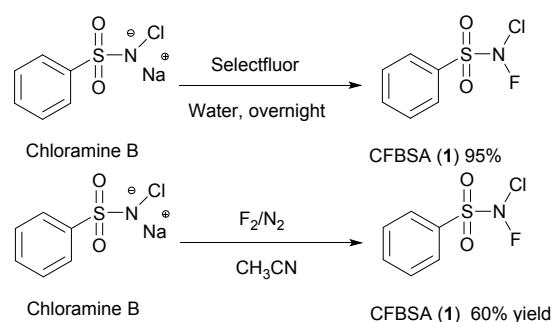
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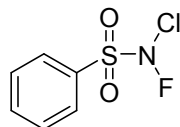
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1. General Information

All the starting chemicals were commercially available and used without further purification. Substrates were purchased from Energy chemical Co. Ltd. and Damas-beta Co. Ltd. Fluorinating reagents (Selectfluor) were purchased from Shanghai Science Bio-pharmaceutical Co. Ltd. Dry acetonitrile was obtained by refluxing with CaH₂. Flash column chromatography was performed using silica gel (300-400 mesh). All ¹H NMR spectra were recorded on a Bruker spectrometer at 400 MHz. The ¹⁹F NMR spectra were recorded on a Bruker spectrometer at 376 MHz. The ¹³C NMR spectra were recorded on a Bruker spectrometer at 100 MHz. Chemical shifts (δ value) were reported in ppm down field from internal tetramethylsilane (TMS). *J* values are reported in Hz. IR spectrum (Film) were recorded on a Nicolet 6700 spectrophotometer in the range of 400~4000 cm⁻¹. HRMS (EI) Ms Spectra were recorded on a Waters GCT Premier ms spectrometer with electron impact.



Preparation of *N*-chloro-*N*-fluorobenzenesulfonamide (CFBSA): (1) To a 250 mL round bottom flask equipped with a large magnetic stir bar, Chloroamine B (5.35 g, 0.025 mol, 1.0 equiv), Selectfluor (13.50 g, 0.038 mol, 1.5 equiv) were all dissolved in water (100 mL) at room temperature. The reaction was allowed to stir at room temperature overnight (~12 h). The mixture was extracted with dichloromethane (3 \times 30 mL), the organic layer was dried with sodium sulfate. Solvent was removed under reduced pressure and the crude product (yellow liquid) was further purified through flash column chromatography. (2) To a 250 mL round bottom flask equipped with a large magnetic stir bar, Chloroamine B (5.35 g, 0.025 mol, 1.0 equiv), 100 mL CH₃CN were added. 20% F₂ was bubbled. The reaction was monitored by TLC. After the disappearance of starting material, the solvent was removed by reduced pressure and the crude product was further purified through flash column chromatography.



***N*-chloro-*N*-fluorobenzenesulfonamide (CFBSA).** Yellow liquid; $R_f = 0.55$ (10% ethyl acetate/petroleum ether); ¹H NMR (400 MHz, CDCl₃): δ 8.07 (d, *J* = 7.8 Hz, 2H), 7.86 (t, *J* = 7.5 Hz, 1H), 7.69 (t, *J* = 7.9 Hz, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 136.78, 131.66, 129.70, 128.74. ¹⁹F NMR (376 MHz, CDCl₃): δ 17.72 (s, 1F). IR (neat) $\nu = 3071, 1582, 1450, 1390, 1190, 1085, 753, 682, 683, 615, 559$ cm⁻¹; HRMS-EI (*m/z*) *calcd.* for (C₆H₅O₂ClFNS): 208.9714, *found* 208.9716.

2. Typical procedures

Standard procedure for the chlorination of carbonyl compounds: To a mixture of carbonyl compound (0.25 mmol, 1.0 equiv) and potassium carbonate (0.38 mmol, 1.5 equiv) in acetonitrile (3.0 mL) were added chlorinating reagent (CFBSA, 0.30 mmol, 1.2 equiv) with stirring at room temperature. The reaction was monitored by thin layer chromatography until completion. After the complete conversion of starting material, the reaction mixture was concentrated and directly purified by column chromatography on silica gel.

NOTES: For substrates that showed a mixture of mono and dichlorinated products with 1.2 equiv CFBSA, addition of another 1.2 equiv CFBSA would promote the complete conversion of monochlorinated products into dichlorinated ones.

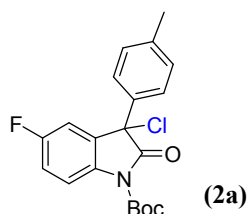
Standard procedure for the chlorination of heterocycles: To a solution of heterocycle (0.25 mmol, 1.0 equiv) in acetonitrile (3.0 mL) were added chlorinating reagent (CFBSA, 0.30 mmol, 1.2 equiv) with stirring at room temperature. The reaction was monitored by thin layer chromatography until completion. After the complete conversion of starting material, the reaction mixture was concentrated and directly purified by column chromatography on silica gel.

NOTES: For substrates that showed a mixture of mono and dichlorinated products with 1.2 equiv CFBSA, addition of another 1.2 equiv CFBSA would promote the complete conversion of monochlorinated products into dichlorinated ones.

Standard procedure for the chlorination of *N*-alkylaniline analogues: To a solution of *N*-alkylaniline (0.25 mmol, 1.0 equiv) in acetonitrile (3.0 mL) was added chlorinating reagent CFBSA (0.30 mmol, 1.2 equiv) with stirring at room temperature. The reaction was monitored by thin layer chromatography until completion. After the complete conversion of starting material, the reaction mixture was concentrated and directly purified by column chromatography on silica gel.

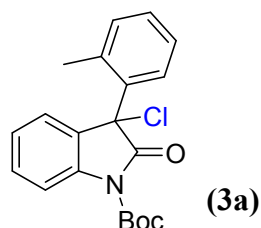
NOTES: For substrates that showed a mixture of mono and dichlorinated or trichlorinated products with 1.2 equiv CFBSA, addition of another 1.2 equiv or 2.4 equiv CFBSA would promote the complete conversion of monochlorinated products into dichlorinated or trichlorinated ones.

3. Analytical Data for the Products

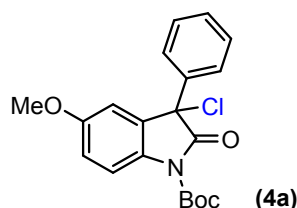


N-tert-butoxycarbonyl-3-chloro-3-(4-methylphenyl)-5-fluoro-2-oxindole.³ Procedure: Using 1.2 equiv CFBSA provided **2a** in 87% yield. White solid; $R_f = 0.61$ (10% ethyl acetate/petroleum ether); ^1H NMR (400 MHz, CDCl_3) δ 7.98 (dd, $J = 4.4, 8.4$ Hz, 1H), 7.37 (d, $J = 8.0$ Hz, 2H), 7.20-7.12 (m, 4H), 2.35 (s, 3H), 1.61 (s, 9H); ^{19}F NMR (376 MHz, CDCl_3) δ -115.95 (s, 1F); ^{13}C NMR (100 MHz, CDCl_3) δ 170.77, 160.26 (d, $J = 243.9$ Hz), 149.08, 139.67, 135.14 (d, $J = 2.5$ Hz), 132.92, 130.79 (d, $J = 8.4$ Hz), 129.52, 127.71, 117.63 (d, $J = 22.8$ Hz), 117.31 (d, $J = 7.7$ Hz), 113.39 (d, $J = 24.7$ Hz), 85.41, 66.24 (d, $J = 1.6$ Hz), 28.10, 21.23.

On this TLC plate, S: Starting material; R: Reaction mixture: the upper point is the product, the lower point is N-fluorobenzulfonamide; C: CFBSA



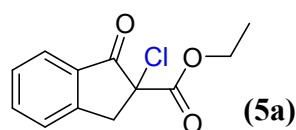
***N*-tert-butoxycarbonyl-3-chloro-3-(2-methylphenyl)-2-oxindole.**³ Procedure: Using 1.2 equiv CFBSA provided **3a** in 86% yield. White solid; $R_f = 0.40$ (10% ethyl acetate/petroleum ether); ^1H NMR (400 MHz, CDCl_3) δ 8.16 (d, $J = 8.0\text{ Hz}$, 1H), 7.94 (d, 8.0 Hz, 1H), 7.42-7.33 (m, 2H), 7.32-7.27 (m, 1H), 7.16-7.11 (m, 1H), 7.09 (d, $J = 7.2\text{ Hz}$, 1H), 7.04 (dd, $J = 1.2, 7.6\text{ Hz}$, 1H), 1.78 (s, 3H), 1.66 (s, 9H); ^{13}C NMR (100 MHz, CDCl_3) δ 171.11, 149.22, 135.17, 131.87, 130.66, 129.37, 129.36, 126.36, 125.58, 124.70, 115.61, 85.28, 69.16, 28.19, 20.17.



***N*-tert-butoxycarbonyl-3-chloro-3-phenyl-5-methoxy-2-oxindole.**⁴ Procedure: Using 1.2 equiv CFBSA provided **3a** in 79% yield. White solid; $R_f = 0.55$ (10% ethyl acetate/petroleum ether); ^1H NMR (400 MHz, CDCl_3) δ 7.91 (d, $J = 8.8\text{ Hz}$, 1H), 7.53-7.50 (m, 2H), 7.39-7.36 (m, 3H), 7.00-6.94 (m, 2H), 3.81 (s, 3H), 1.61 (s, 9H); ^{13}C NMR (100 MHz, CDCl_3) δ 171.14, 157.55, 149.18, 136.46, 132.52, 130.09, 129.30, 128.72, 127.98, 116.89, 116.51, 111.37, 85.08, 67.00, 55.89, 28.16.

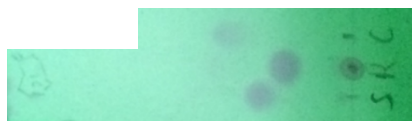


On this TLC plate, S: Starting material; R: Reaction mixture; C: CFBSA

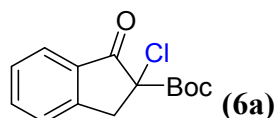


Ethyl 2-chloro-1-oxo-indan-2-carboxylate.⁵ Procedure: Using 1.2 equiv CFBSA provided **5a** in 89% yield. White solid; $R_f = 0.30$ (10% ethyl acetate/petroleum ether); ^1H NMR (400 MHz, CDCl_3) δ 7.86 (d, $J = 8.0\text{ Hz}$, 1H), 7.70 (t, $J = 7.6\text{ Hz}$, 1H), 7.49-7.44 (m, 2H), 4.27 (q, $J = 7.2,$

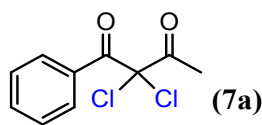
14.4 Hz, 2H), 4.09 (d, $J = 17.6$ Hz, 1H), 3.56 (d, $J = 17.6$ Hz, 1H), 1.26 (t, $J = 7.2$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 195.24, 167.23, 150.71, 136.54, 132.63, 128.71, 126.43, 126.08, 68.11, 63.54, 43.53, 14.08.



On this TLC plate, S: Starting material; R: Reaction mixture; C: CFBSA



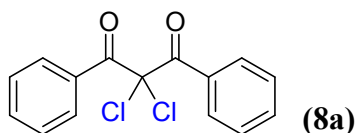
Tert-butyl-2-chloro-1-oxo-indan-2-carboxylate.⁶ Procedure: Using 1.2 equiv CFBSA provided **6a** in 83% yield. White solid; $R_f = 0.70$ (10% ethyl acetate/petroleum ether); ^1H NMR (400 MHz, CDCl_3) δ 7.84 (d, $J = 7.6$ Hz, 1H), 7.68 (t, $J = 7.6$ Hz, 1H), 7.48-7.43 (m, 2H), 4.00 (d, $J = 17.6$ Hz, 1H), 3.53 (d, $J = 17.6$ Hz, 1H), 1.42 (s, 9H); ^{13}C NMR (100 MHz, CDCl_3) δ 195.69, 165.98, 150.76, 136.30, 132.89, 128.56, 126.34, 125.89, 84.52, 68.88, 43.63, 27.80.



2,2-Dichloro-1-phenyl-1,3-butanedione.⁷ Procedure: Using 2.4 equiv CFBSA provided **7a** in 88% yield. White solid; $R_f = 0.76$ (10% ethyl acetate/petroleum ether); ^1H NMR (400 MHz, CDCl_3) δ 8.10-8.05 (m, 2H), 7.66-7.61 (m, 1H), 7.54-7.46 (m, 2H), 2.46 (s, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 192.31, 186.02, 134.60, 130.97, 130.68, 128.78, 86.57, 25.04.



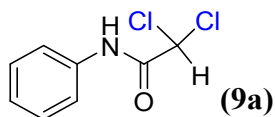
On this TLC plate, S: Starting material; R: Reaction mixture; C: CFBSA



1,3-Diphenyl-2,2-dichloro-1,3-propanedione.⁸ Procedure: Using 2.4 equiv CFBSA provided **8a** in 86% yield. Yellow solid; $R_f = 0.54$ (10 % ethyl acetate/petroleum ether); ^1H NMR (400 MHz, CDCl_3) δ 7.99-7.95 (m, 4H), 7.56-7.51 (m, 2H), 7.42-7.37 (m, 4H); ^{13}C NMR (100 MHz, CDCl_3) δ 185.48, 134.39, 131.58, 130.60, 128.82, 87.68.



On this TLC plate, S: Starting material; R: Reaction mixture; C: CFBSA

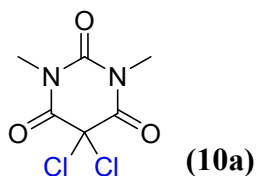


2,2-
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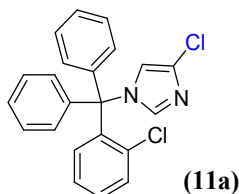
Dichloro-N-phenyl-acetamide.⁹ Procedure: Using 2.4 equiv CFBSA **9a** in 84% yield. White solid; $R_f = 0.31$ (10% ethyl acetate/petroleum ether); ^1H NMR (400 MHz, CDCl_3) δ 8.18 (s, br, 1H), 7.58-7.54 (m, 2H), 7.41-7.23-7.18 (m, 1H), 6.05 (s, 1H); ^{13}C NMR (100 MHz, CDCl_3) δ 136.34, 129.37, 125.84, 120.36, 66.99.



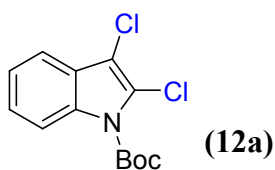
On this TLC plate, S: Starting material; R: Reaction mixture; C: CFBSA



5,5-Dichloro-1,3-dimethyl-2,4,6(1H,3H,5H)-pyrimidinetrione.¹⁰ Procedure: Using 2.4 equiv CFBSA provided **10a** in 79% yield. White solid; $R_f = 0.35$ (10% ethyl acetate/petroleum ether); ^1H NMR (400 MHz, CDCl_3) δ 3.42 (s, 6H); ^{13}C NMR (100 MHz, CDCl_3) δ 161.42, 148.86, 71.85, 30.57.

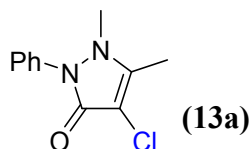


4-Chloro-1-((2-chlorophenyl)diphenylmethyl)-1H-imidazole.¹ Procedure: Using 1.2 equiv CFBSA provided **11a** in 77% yield. White solid; $R_f = 0.43$ (50% ethyl acetate/petroleum ether); ^1H NMR (400 MHz, CDCl_3) δ 7.44 (dd, $J = 1.2, 7.6$ Hz, 1H), 7.39-7.36 (m, 7H), 7.32 (s, 1H), 7.29 (dd, $J = 1.2, J = 7.6$ Hz, 1H), 7.19-7.16 (m, 4H), 6.94 (dd, $J = 1.6, 8.0$ Hz, 1H), 6.66 (s, 1H); ^{13}C NMR (100 MHz, CDCl_3) δ 140.16, 139.74, 137.57, 135.61, 132.44, 130.45, 130.20, 129.20, 128.53, 128.28, 127.25, 117.50, 76.00.



Tert-butyl-2,3-dichloro-1H-indole-1-carboxylate. Procedure: Using 2.4 equiv CFBSA provided **12a** in 75% yield. Colorless liquid; $R_f = 0.55$ (5% ethyl acetate/petroleum ether); ^1H NMR (400

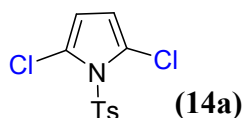
MHz, CDCl₃) δ 8.11(d, J = 8.0 Hz, 1H), 7.53-7.50 (m, 1H), 7.51 (dd, J = 1.2, 8.0 Hz, 1H), 7.38-7.27(m, 2H), 1.69(s, 9H); ¹³C NMR (100 MHz, CDCl₃) δ 148.63, 134.35, 126.00, 125.68, 123.79, 121.76, 117.92, 115.46, 112.18, 85.64, 28.29. IR (neat) ν = 3317, 3165, 1743, 1477, 1174, 797, 751, 685, 556 cm⁻¹; HRMS-EI (m/z) *calcd.* for (C₁₃H₁₃Cl₂NO₂) 289.0323; *found* 289.0324.



4-Chloro-1,2-dihydro-1,5-dimethyl-2-phenyl-3H-pyrazol-3-one.¹¹ Procedure: Using 1.2 equiv CFBSA provided **13a** in 88% yield. White solid; R_f = 0.42 (50% ethyl acetate/petroleum ether); ¹H NMR (400 MHz, CDCl₃) δ 7.46 (t, J = 7.6 Hz, 2H), 7.40 (d, J = 7.6 Hz, 2H), 7.31 (t, J = 7.6 Hz, 1H), 3.07 (s, 3H), 2.30(s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 161.57, 152.01, 134.78, 129.41, 127.22, 124.27, 103.51, 36.59, 11.35.



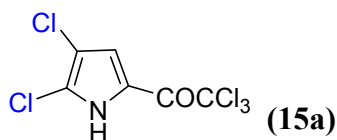
On this TLC plate, S: Starting material; R: Reaction mixture; the upper point is the product, the lower point is N-F bensulfonamide; C: CFBSA



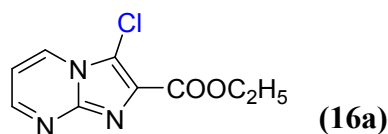
2,5-Dichloro-1-tosyl-1H-pyrrole.¹ Procedure: Using 2.4 equiv CFBSA provided **14a** in 86% yield. White solid; R_f = 0.66 (10% ethyl acetate/petroleum ether); ¹H NMR (400 MHz, CDCl₃) δ 7.89 (d, J = 8.0 Hz, 2H), 7.34 (d, J = 8.0 Hz, 2H), 6.10 (s, 2H), 2.44 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 146.04, 135.46, 130.16, 127.69, 117.67, 112.76, 21.86.



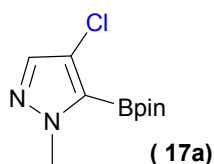
On this TLC plate, S: Starting material; R: Reaction mixture; C: CFBSA; M: Mixture of "S" and "R"



2,2,2-Trichloro-1-(4,5-dichloro-1H-pyrrol-2-yl)ethanone.¹² Procedure: Using 2.4equiv CFBSA provided **15a** in 78% yield. Yellow solid; R_f = 0.64 (10% ethyl acetate/petroleum ether); ¹H NMR (400 MHz, CDCl₃) δ 9.87 (s, 1H), 7.31 (d, J = 3.2 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 172.29, 123.58, 120.49, 120.40, 113.30, 94.05.



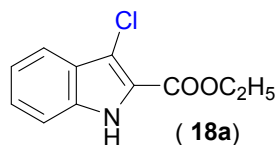
Ethyl 3-chloroimidazo[1,2-a]pyrimidine-2-carboxylate.¹ Procedure: Using 1.2 equiv CFBSA provided **16a** in 89% yield. Yellow solid; $R_f = 0.57$ (ethyl acetate); $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 8.70 (dd, $J = 2.0$ Hz, 4.0 Hz, 1H), 8.47 (dd, $J = 2.0$, 6.8 Hz, 1H), 7.10 (dd, $J = 4.0$, 6.8 Hz, 1H), 4.47 (q, $J = 6.8$, 12 Hz, 2H), 1.43 (t, $J = 7.2$ Hz, 3H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 161.84, 152.62, 145.77, 132.34, 131.29, 113.14, 110.61, 61.75, 14.40.



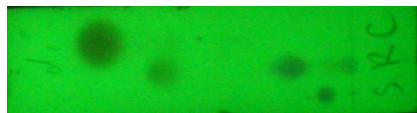
4-Chloro-1-methyl-5-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)-1H-pyrazole.¹ Procedure: Using 1.2 equiv CFBSA provided **17a** in 89% yield. Yellow solid; $R_f = 0.49$ (10% ethyl acetate/petroleum ether); $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.42 (s, 1H), 4.04 (s, 3H), 1.36 (s, 12H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 137.34, 120.01, 100.10, 84.39, 40.59, 24.91.



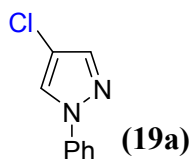
On this TLC plate, S: Starting material; R: Reaction mixture; C: CFBSA



Ethyl 3-chloro-1H-indole-2-carboxylate.¹ Procedure: Using 1.2 equiv CFBSA provided **18a** in 92% yield. White solid; $R_f = 0.67$ (16% ethyl acetate/petroleum ether); $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 9.20 (s, br, 1H), 7.74-7.71 (m, 1H), 7.42-7.35 (m, 2H), 7.25-7.20 (m, 1H), 4.48 (q, $J = 7.2$, 14.4 Hz, 2H), 1.46 (t, $J = 7.2$ Hz, 3H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 161.16, 134.86, 126.70, 126.36, 122.52, 121.41, 120.36, 112.59, 112.19, 61.58, 14.50.



On this TLC plate, S: Starting material; R: Reaction mixture; C: CFBSA

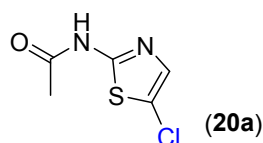


4-Chloro-1-phenyl-1H-pyrazole.¹ Procedure: Using 1.2 equiv CFBSA provided **19a** in 83% yield. White solid; $R_f = 0.40$ (16% ethyl acetate/petroleum ether); $^1\text{H NMR}$ (400 MHz, CDCl_3) δ

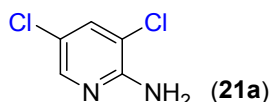
7.9 (s, 1H), 7.65 (s, 1H), 7.64 (d, $J = 8.0$ Hz, 3H), 7.46 (t, $J = 7.6$ Hz, 2H); ^{13}C NMR (100 MHz, CDCl_3) δ 139.85, 139.60, 129.68, 127.13, 124.95, 119.09, 112.51.



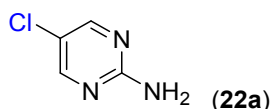
On this TLC plate, S: Starting material; R: Reaction mixture; C: CFBSA



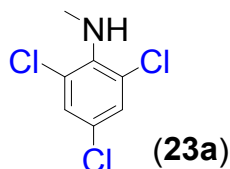
***N*-(5-chlorothiazol-2-yl)acetamide.**¹³ Procedure: Using 1.2 equiv CFBSA provided **20a** in 83% yield. White solid; $R_f = 0.38$ (50% ethyl acetate/petroleum ether); ^1H NMR (400 MHz, CDCl_3) δ 7.49 (s, 1H), 3.39 (br,s, 1H), 2.14 (s, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 168.94, 155.96, 135.52, 117.81, 22.23.



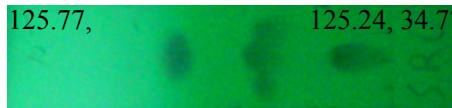
3,6-dichloropyridin-2-amine.¹⁴ Procedure: Using 2.4 equiv CFBSA provided **21a** in 84% yield. White solid; $R_f = 0.70$ (10% ethyl acetate/petroleum ether); ^1H NMR (400 MHz, CDCl_3) δ 7.93 (d, $J = 2.0$ Hz, 1H), 7.51 (d, $J = 2.0$ Hz, 1 H), 5.01 (s, 2H); ^{13}C NMR (100 MHz, CDCl_3) δ 153.35, 144.37, 136.75, 120.43, 115.30.



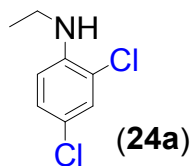
5-chloropyrimidin-2-amine.¹⁵ Procedure: Using 1.2 equiv CFBSA provided **22a** in 78% yield. White solid; $R_f = 0.70$ (50% ethyl acetate/petroleum ether); ^1H NMR (400 MHz, CDCl_3) δ 8.26 (s, 2 H), 6.90 (s, 2H); ^{13}C NMR (100 MHz, CDCl_3) δ 162.03, 156.09, 117.23.



2,4,6-Trichloro-*N*-methyl benzenamine.¹⁶ Procedure: Using 3.6 equiv CFBSA provided **23a** in 78% yield. Yellow oil; $R_f = 0.58$ (5% ethyl acetate/petroleum ether); ^1H NMR (400 MHz, CDCl_3) δ 7.22 (s, 2H), 3.89 (br s, 1H), 3.00 (s, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 142.95, 128.68, 125.77, 125.24, 34.77.



On this TLC plate, S: Starting material; R: Reaction mixture; C: CFBSA

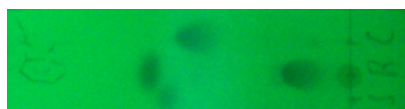


2,4-Dichloro-N-ethylbenzenamine.¹⁷ Procedure: Using 2.4 equiv CFBSA provided **24a** in 81% yield. Yellow oil; $R_f = 0.58$ (5% ethyl acetate/petroleum ether); $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.25 (d, $J = 2.4$ Hz, 1H), 7.10 (dd, $J = 2.4, 8.8$ Hz, 1H), 6.58 (d, $J = 8.8$ Hz, 1H), 4.35 (br s, 1H), 3.18 (q, $J = 7.2, 14.4$ Hz, 2H), 1.30 (t, $J = 7.2$ Hz, 3H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 142.95, 128.73, 127.83, 120.93, 119.24, 111.73, 38.43, 14.68.



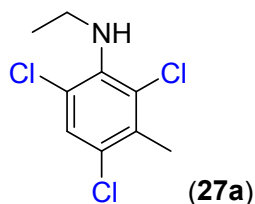
On this TLC plate, S: Starting material R: Reaction mixture C: CFBSA

2-Methyl-4,6-dichloro-N-methylbenzenamine.¹⁸ Procedure: Using 2.4 equiv CFBSA provided **25a** in 83% yield. Yellow oil; $R_f = 0.58$ (10% ethyl acetate/petroleum ether); $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.18 (d, $J = 2.4$ Hz, 1H), 7.01 (d, $J = 2.0$ Hz, 1H), 3.73 (s, br, 1H), 2.82 (s, 3H), 2.32 (s, 3H). $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 144.33, 132.21, 129.97, 126.70, 126.32, 126.17, 35.22, 19.29.



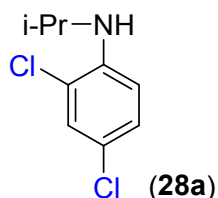
On this TLC plate, S: Starting material; R: Reaction mixture; C: CFBSA

3-Methyl-2,4,6-trichloro-N-methylbenzenamine. Procedure: Using 3.6 equiv CFBSA provided **26a** in 88% yield. Yellow oil. $R_f = 0.67$ (10% ethyl acetate/petroleum ether); $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.27 (s, 1H), 3.90 (s, br, 1H), 2.97 (s, 3H), 2.42 (s, 3H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 143.15, 133.94, 128.52, 127.36, 126.58, 123.26, 34.89, 18.23. IR (neat) $\nu = 3396, 2957, 1779, 1453, 1416, 1246, 1125, 818, 727, 647$ cm^{-1} . HRMS-EI (m/z) *calcd.* for ($\text{C}_8\text{H}_8\text{Cl}_3\text{N}$) 222.9722; *found* 222.9724.

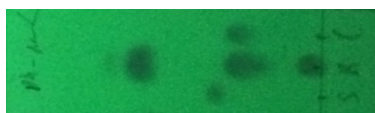


3-Methyl-2,4,6-trichloro-N-ethylbenzenamine. Procedure: Using 3.6 equiv CFBSA provided **27a** in 81% yield. Yellow oil; $R_f = 0.71$ (10 % ethyl acetate/petroleum ether); $^1\text{H NMR}$ (400 MHz,

CDCl₃) δ 7.28 (s, 1H), 3.74 (s, br, 1H), 3.32 (q, J = 7.2, 2H), 2.42 (s, 3H), 1.20 (t, J = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 141.89, 133.93, 128.45, 127.96, 126.83, 123.87, 42.64, 18.30, 16.11. IR (neat) ν = 3379, 2968, 1754, 1445, 1380, 1136, 798, 729, 574 cm⁻¹. HRMS-EI (m/z) *calcd.* for (C₉H₁₀Cl₃N) 236.9879; *found* 236.9875.

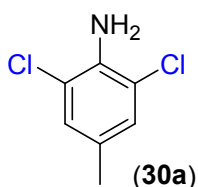


2,4,6-Trichloro-N-(1-methylethyl)benzenamine.¹⁸ Procedure: Using 3.6 equiv CFBSA provided **28a** in 83% yield. Yellow liquid; R_f = 0.64 (10% ethyl acetate/petroleum ether); ¹H NMR (400 MHz, CDCl₃) δ 7.23 (s, 1H), 7.08 (d, J = 8.8 Hz, 1H), 6.57 (d, J = 8.8 Hz, 1H), 4.21 (s, br, 1H), 3.65-3.57 (m, 1H), 1.24 (d, J = 6.4, 6H); ¹³C NMR (100 MHz, CDCl₃) δ 142.02, 128.93, 127.81, 120.74, 119.46, 112.44, 44.52, 22.85.

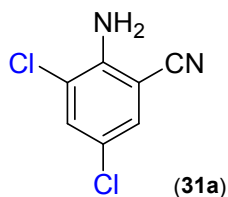


On this TLC plate, S: Starting material; R: Reaction mixture; C: CFBSA

2,4,6-trichloroaniline.¹⁹ Procedure: Using 3.6 equiv CFBSA provided **29a** in 70% yield. Yellow oil; R_f = 0.73 (10% ethyl acetate/petroleum ether); ¹H NMR (400 MHz, CDCl₃) δ 7.17(s, 2 H), 4.43(s, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 139.06, 127.62, 121.86, 119.72.

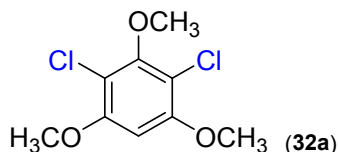


2,6-dichloro-4-methylaniline.²⁰ Procedure: Using 2.4 equiv CFBSA provided **30a** in 62% yield. Yellow solid; R_f = 0.79 (10% ethyl acetate/petroleum ether); ¹H NMR (400 MHz, CDCl₃) δ 7.00(s, 2H), 2.21(s, 2H), 1.25(s, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 137.69, 128.38, 119.59, 29.84, 20.18.

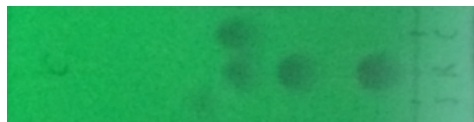


2-amino-3,5-dichlorobenzonitrile.²¹ Procedure: Using 2.4 equiv CFBSA provided **31a** in 60% yield. white solid; R_f = 0.30 (10% ethyl acetate/petroleum ether); ¹H NMR (400 MHz, CDCl₃) δ 7.44 (d, J = 2.4 Hz, 1H), 7.31 (d, J = 2.0 Hz, 1H), 4.85 (s, 2H); ¹³C NMR (100 MHz, CDCl₃) δ

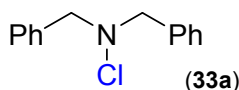
145.06, 133.17, 121.97, 120.29, 115.78, 97.64.



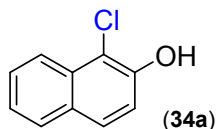
1,3,5-Trimethoxy-2,6-dichlorobenzene.²² Procedure: Using 3.6 equiv CFBSA provided **32a** in 90% yield. White solid; $R_f = 0.36$ (10% ethyl acetate/petroleum ether); $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 6.37 (s, 1H), 3.91 (s, 6H), 3.88 (s, 3H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 154.93, 153.99, 109.84, 93.31, 60.75, 56.66.



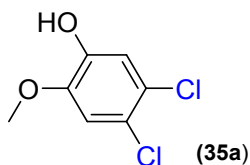
On this TLC plate, S: Starting material; R: Reaction mixture; C: CFBSA



N-chloro-N-(phenylmethyl)benzenemethanamine.²³ Procedure: Using 1.2 equiv CFBSA provided **33a** in 75% yield. White liquid; $R_f = 0.54$ (10% ethyl acetate/petroleum ether); $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.45-7.33 (m, 10H), 4.19 (s, 4H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 137.19, 129.21, 128.51, 128.00, 67.27.



3,5-Dichloro-2-methoxyphenol.²⁴ Procedure: Using 2.4 equiv CFBSA provided **34a** in 64% yield. Yellow liquid; $R_f = 0.2$ (10% ethyl acetate/petroleum ether); $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.00 (s, 1H), 6.90 (s, 1H), 5.60 (br, s, 1H), 3.88 (s, 3H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 145.89, 145.03, 124.30, 122.78, 116.15, 112.44, 56.50.



1-chloronaphthalen-2-ol.²⁵ Procedure: Using 1.2 equiv CFBSA provided **35a** in 77% yield. White solid; $R_f = 0.70$ (10% ethyl acetate/petroleum ether); $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 8.08 (d, $J = 8.0$ Hz, 1H), 7.45 (dt, $J = 1.2, 7.6$ Hz, 1H), 7.43 (d, $J = 10.0$ Hz, 1H), 7.33 (d, $J = 7.6$ Hz, 1H), 6.34 (d, $J = 10.0$ Hz, 1H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 186.00, 144.99, 140.77, 131.37, 130.77, 129.66, 129.365, 127.04, 122.72, 80.59.

Differential scanning calorimetry (DSC) trace

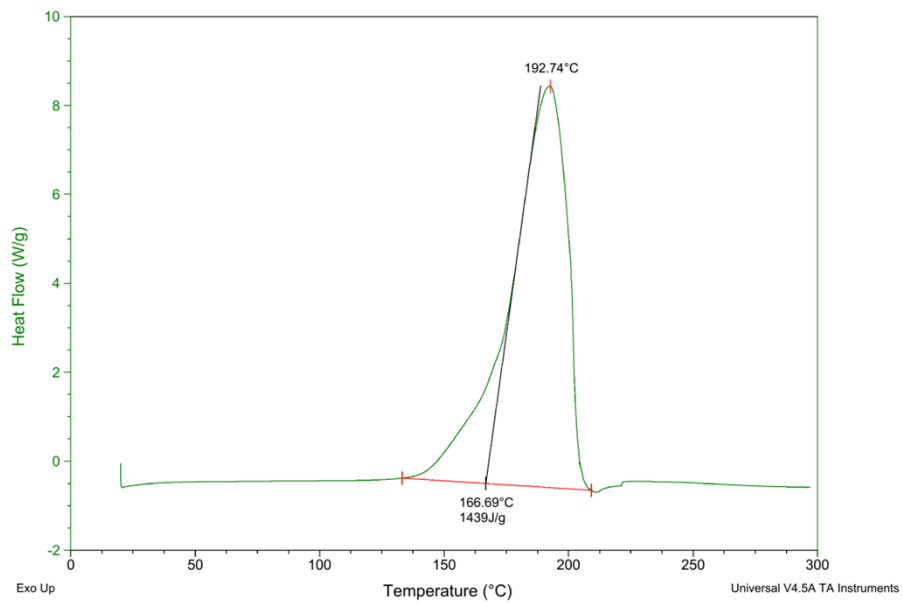
Table 1. DSC analysis of CFBSA

Thermal profile for CFBSA	
Onset Temperature:	130 °C
Exothermmaximum:	192 °C
ΔH : Enthalpy	1439J/g

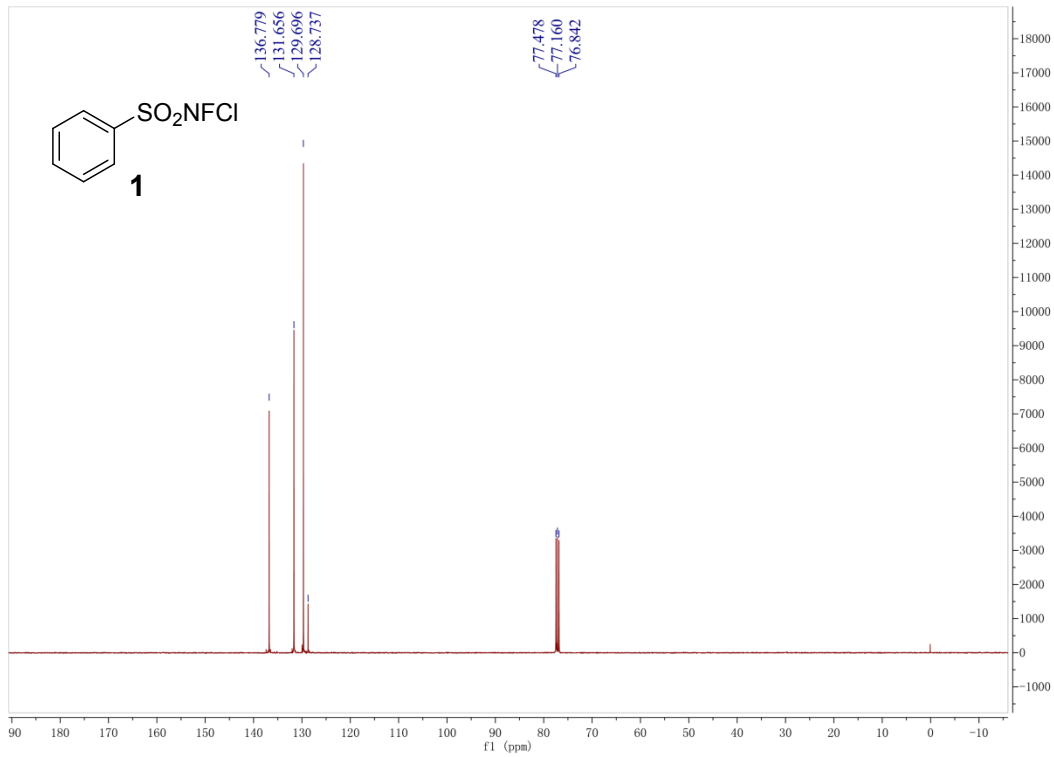
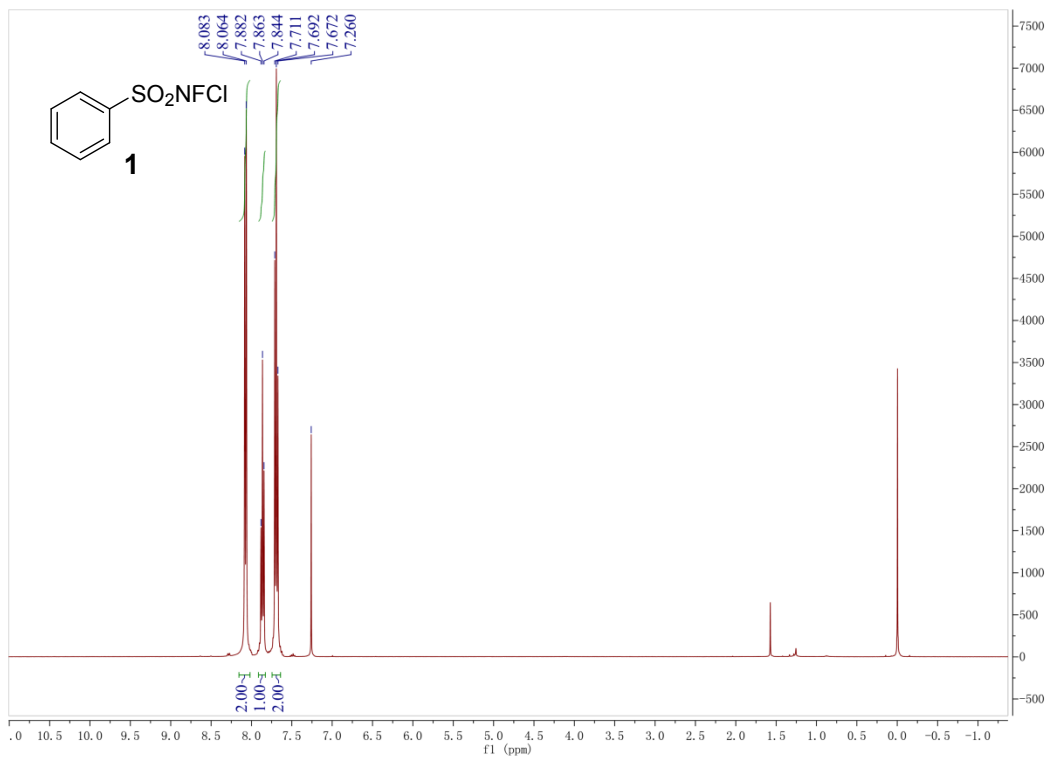
Sample: 2013546-LZ-01
Size: 0.9800 mg
Method: Liu,PR

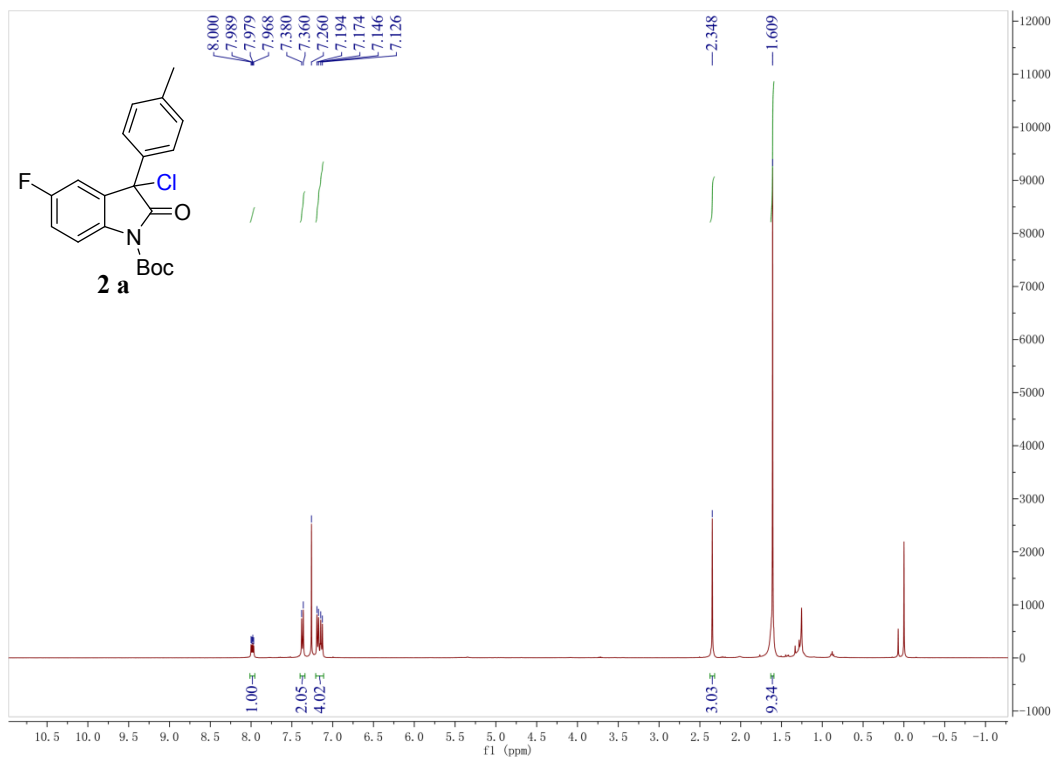
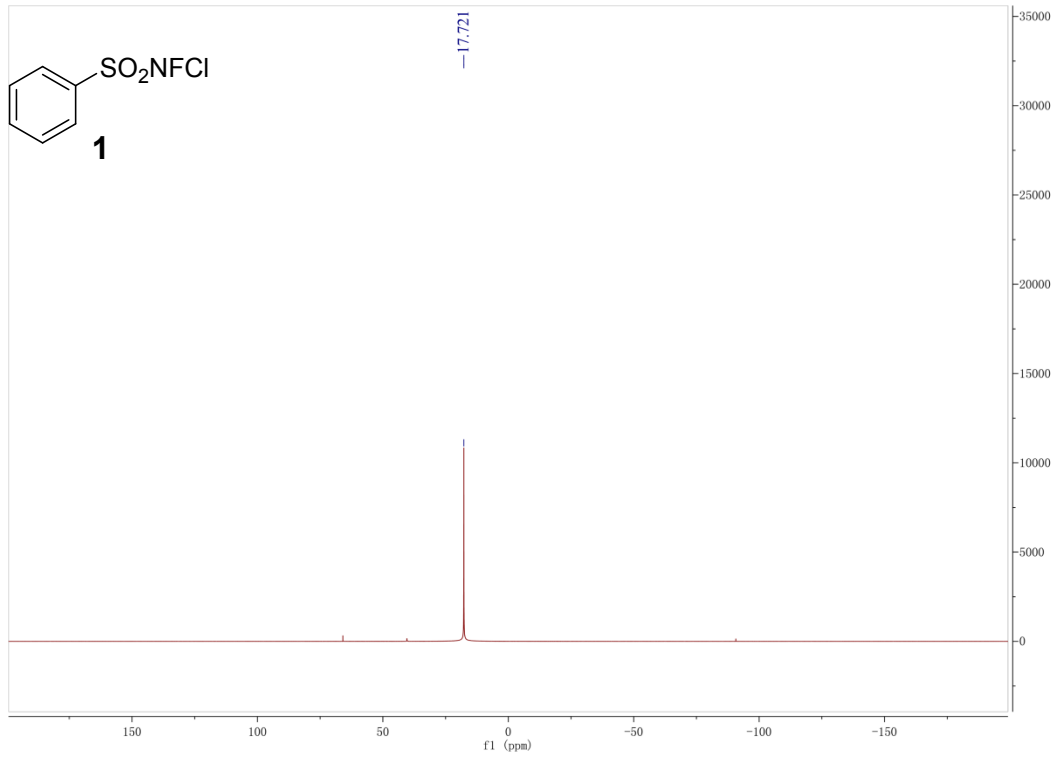
DSC

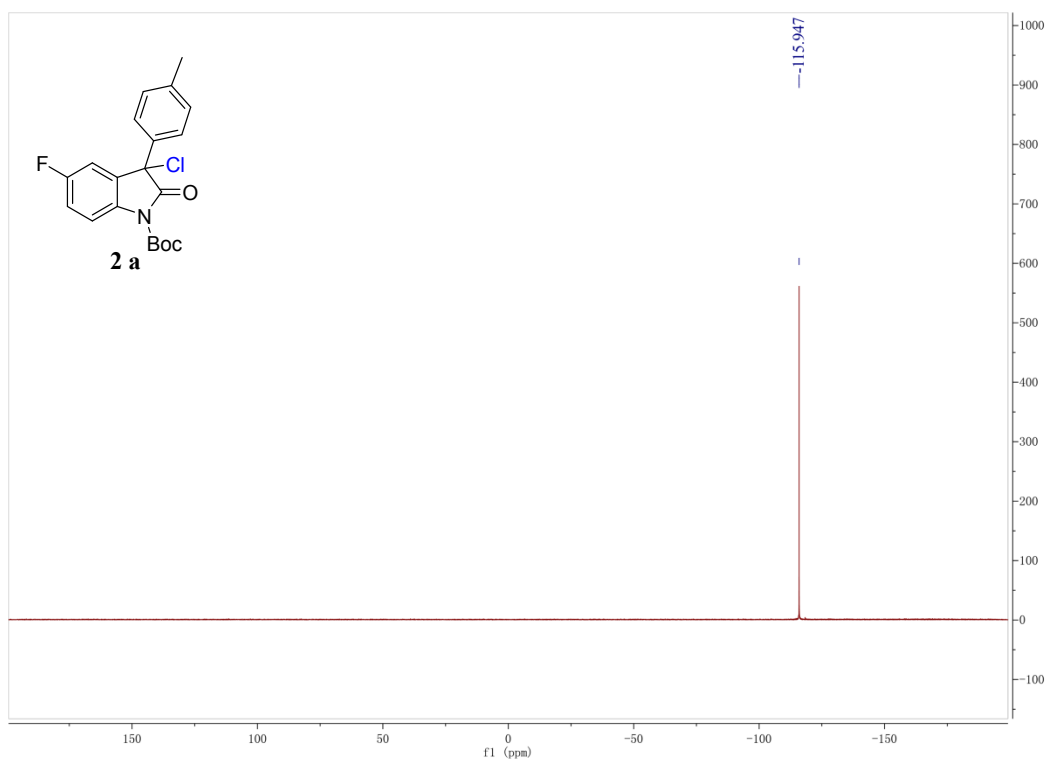
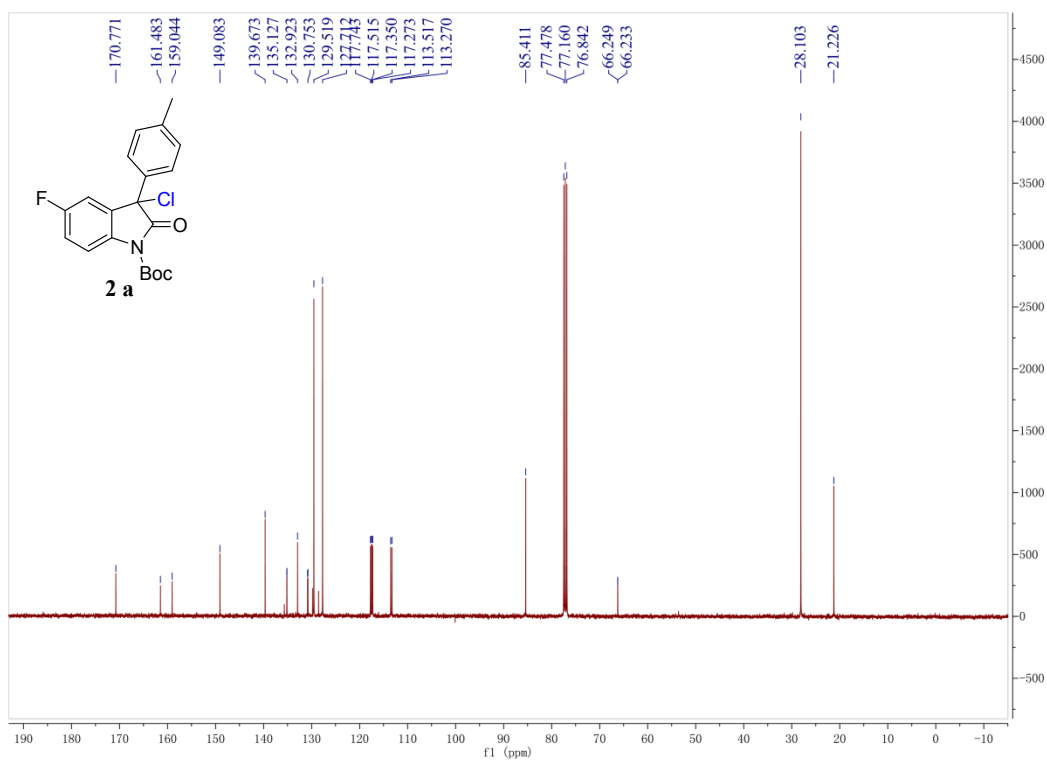
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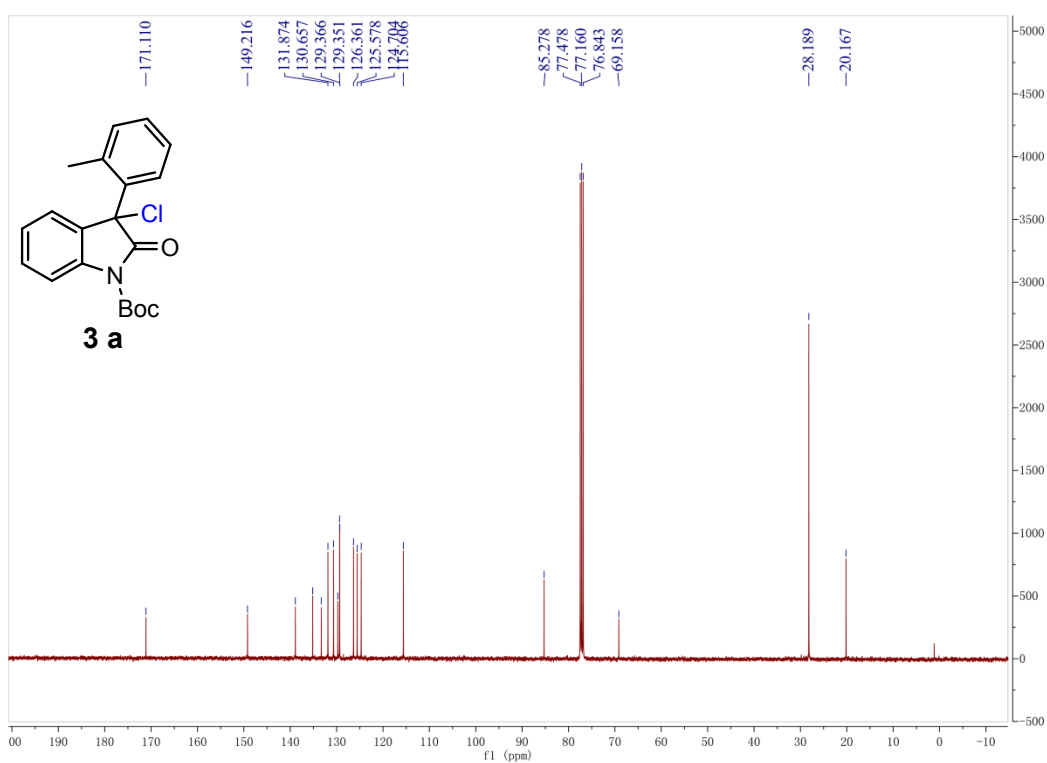
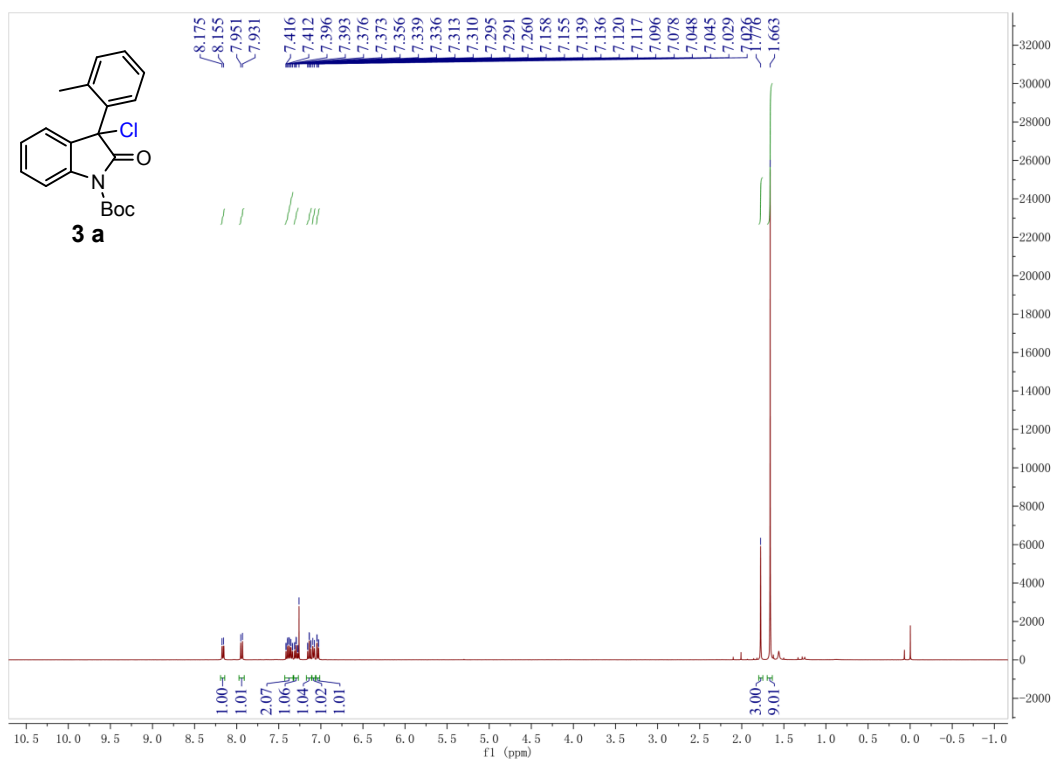


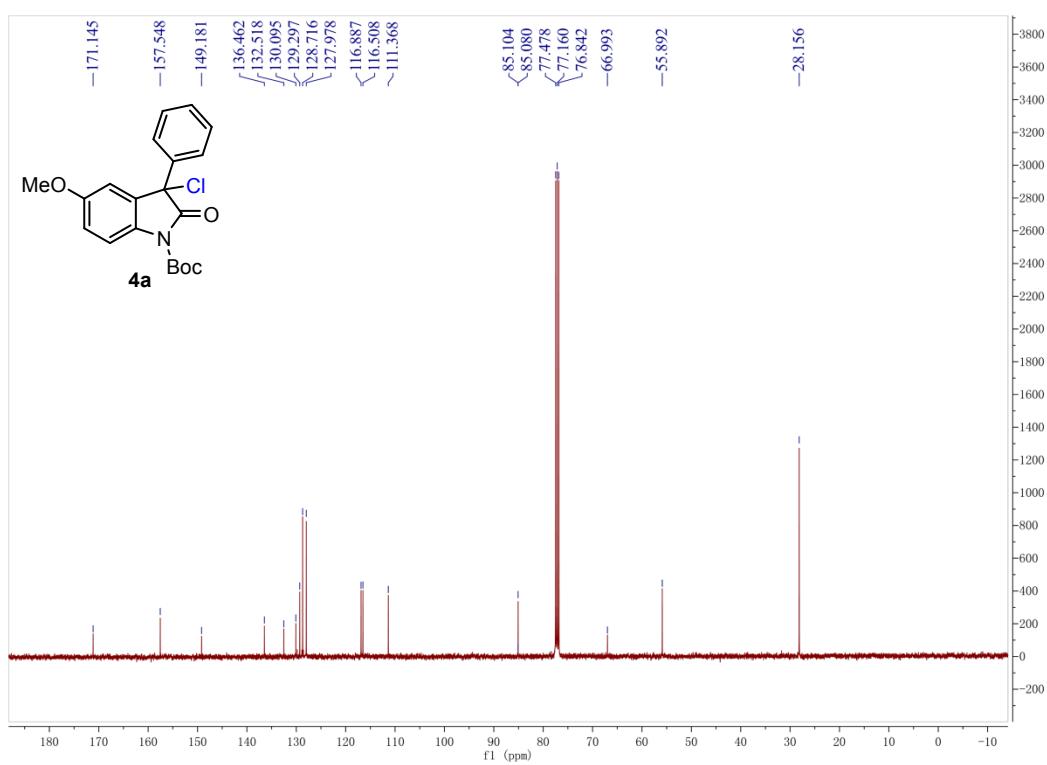
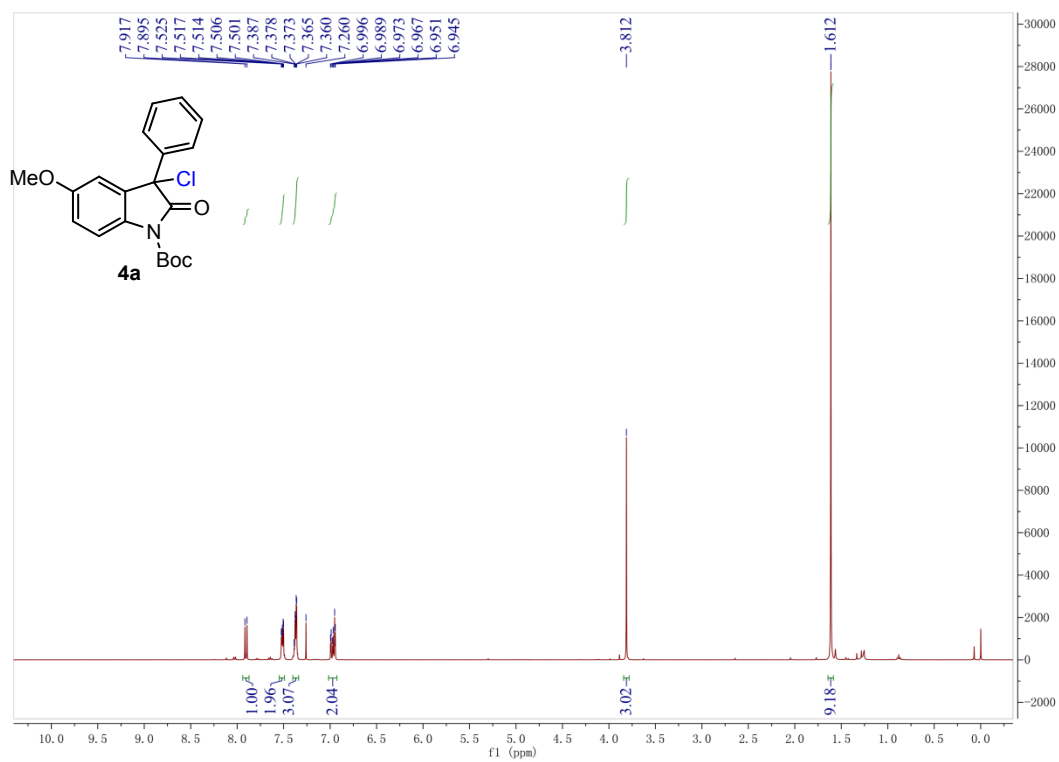
^1H , ^{13}C NMR and ^{19}F NMR spectra

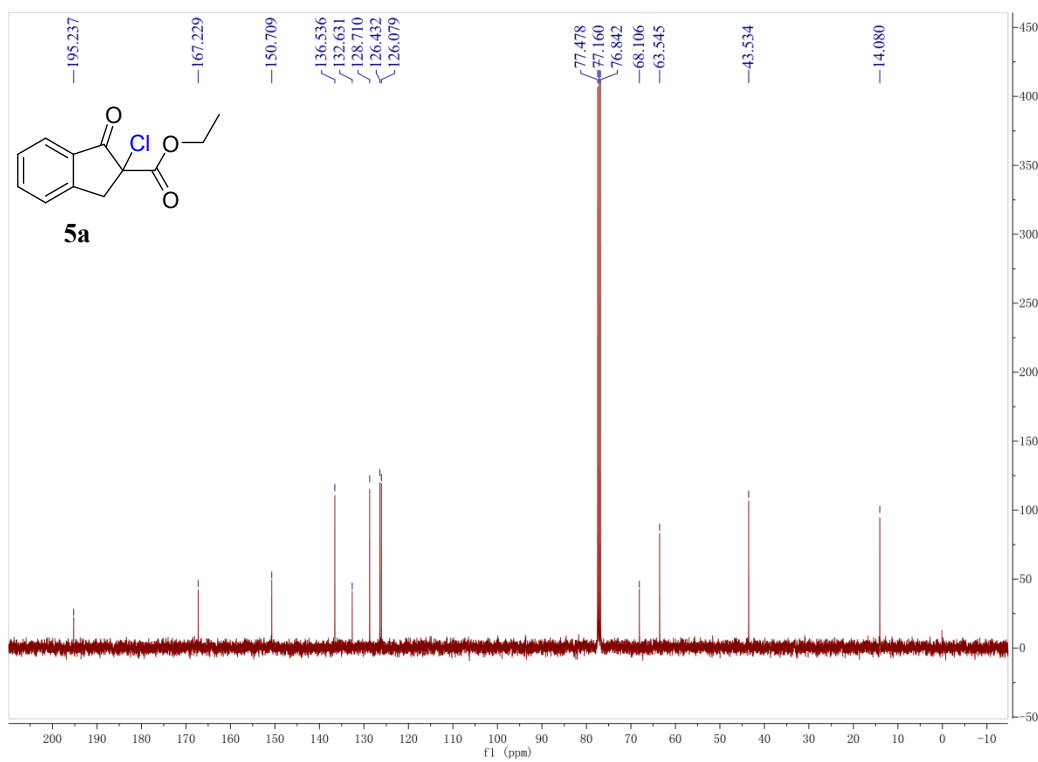
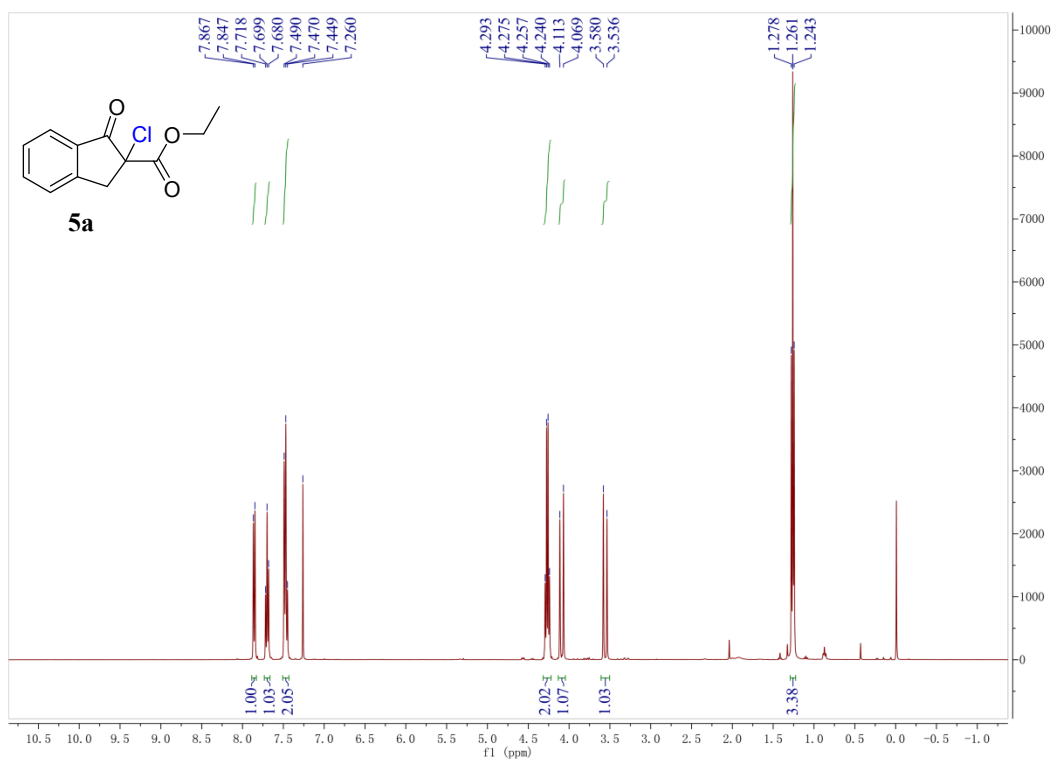


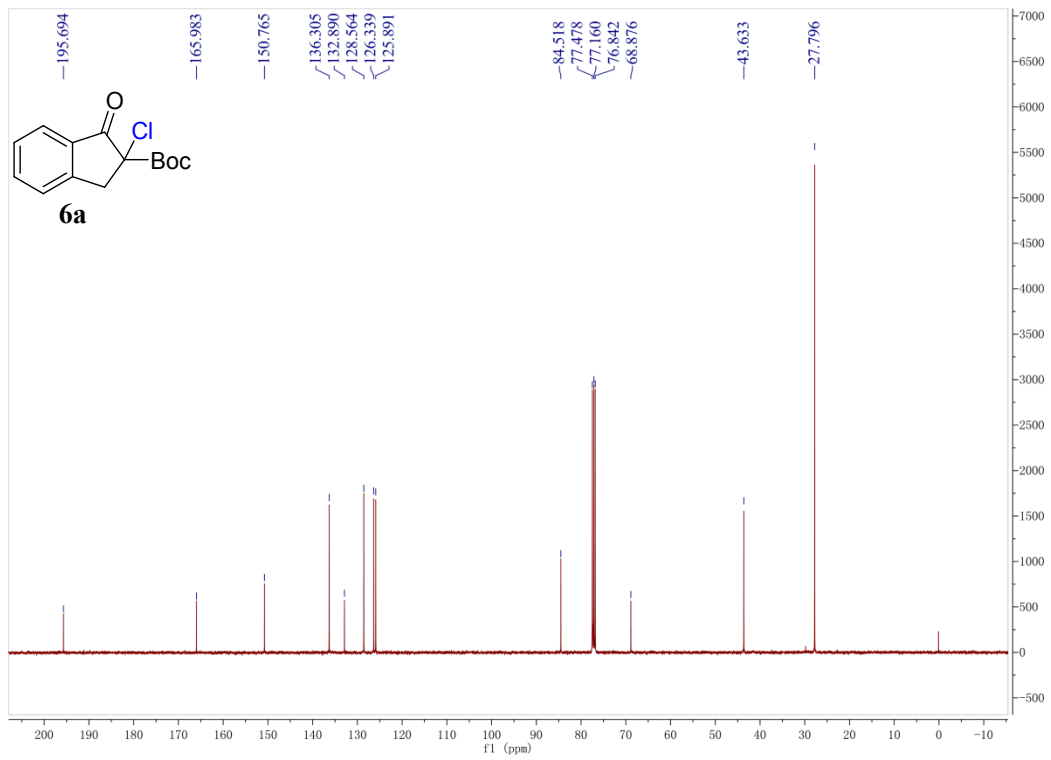
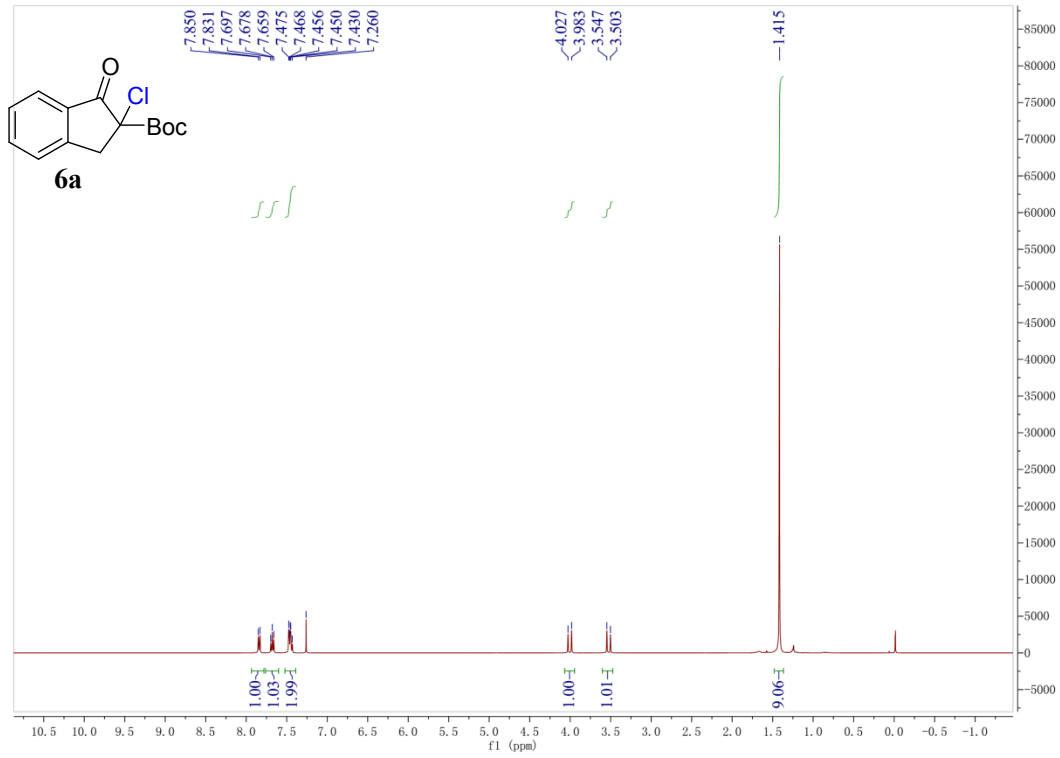


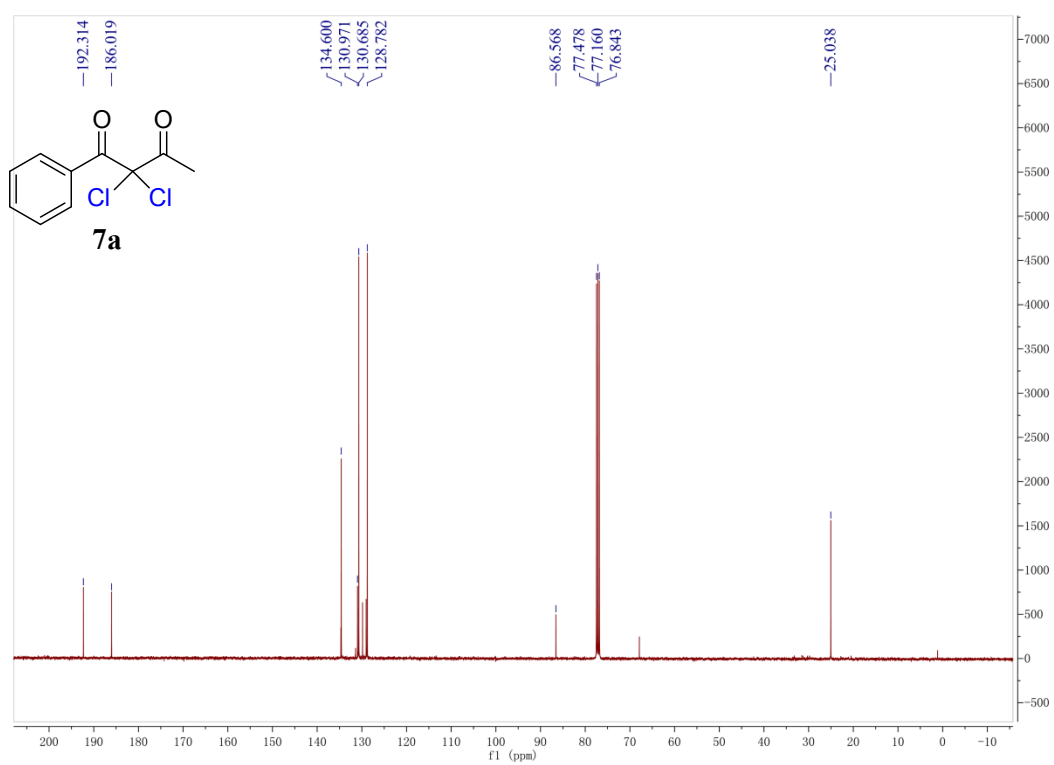
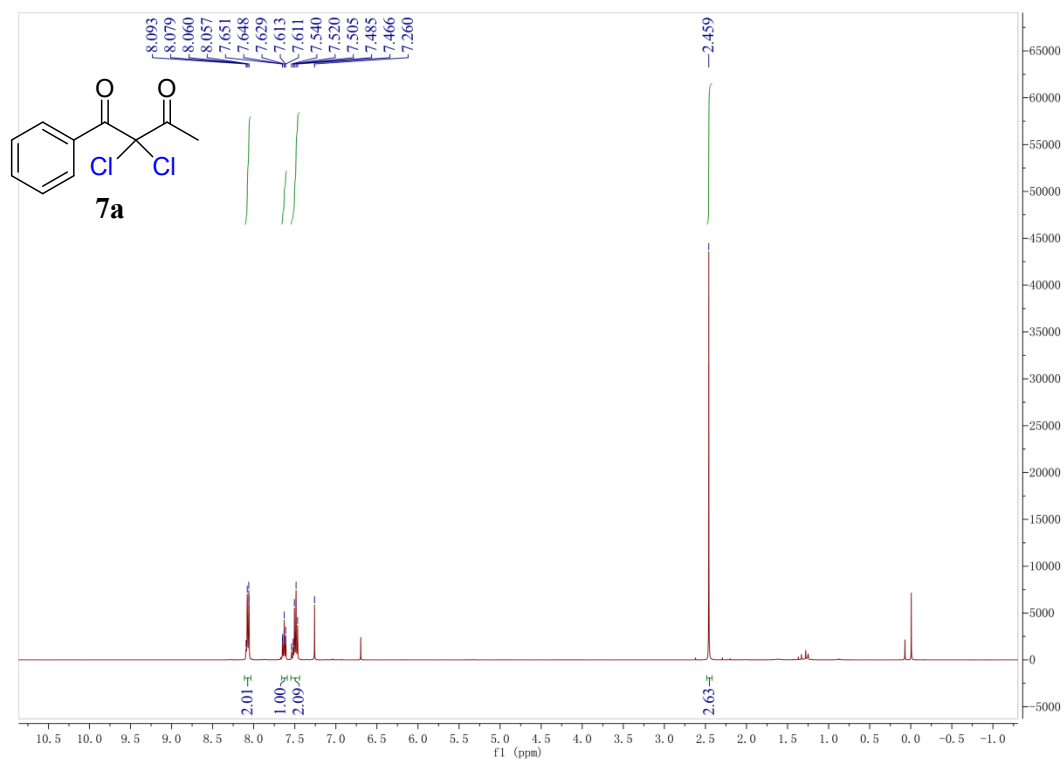


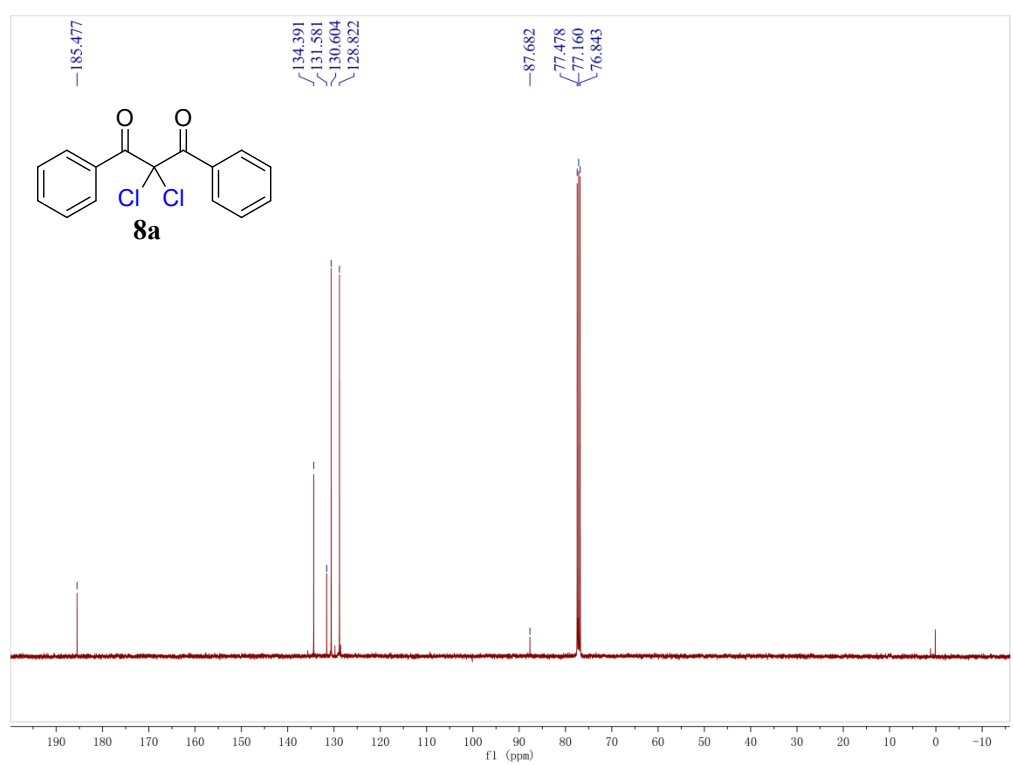
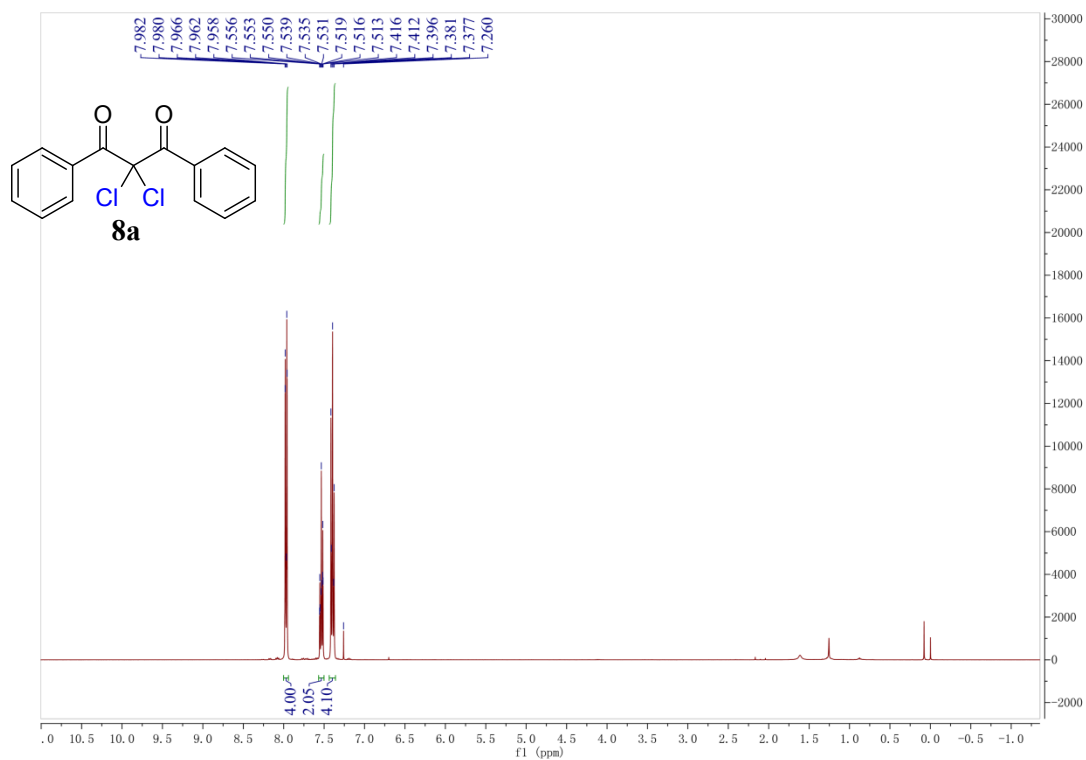


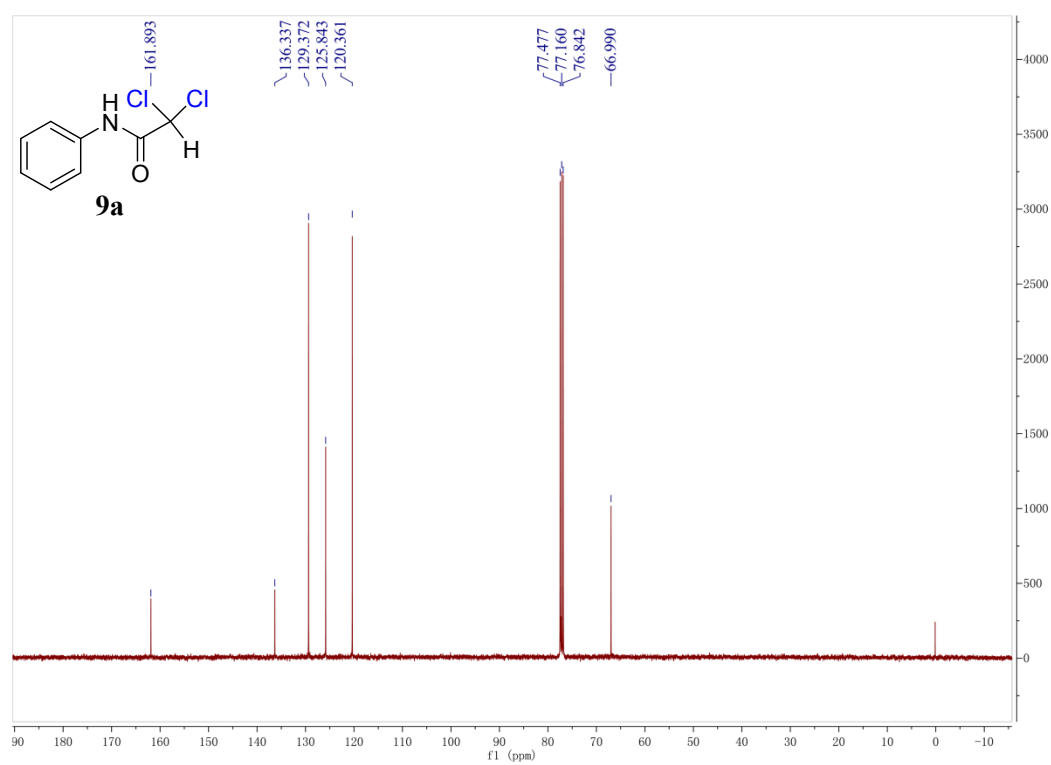
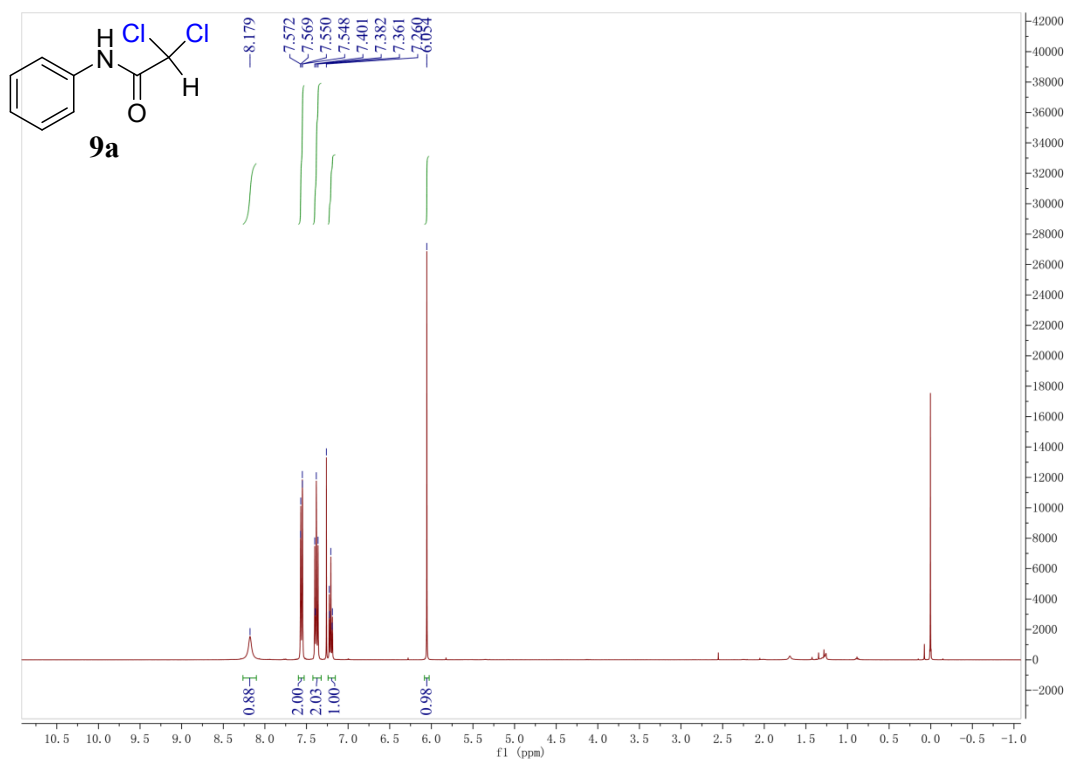


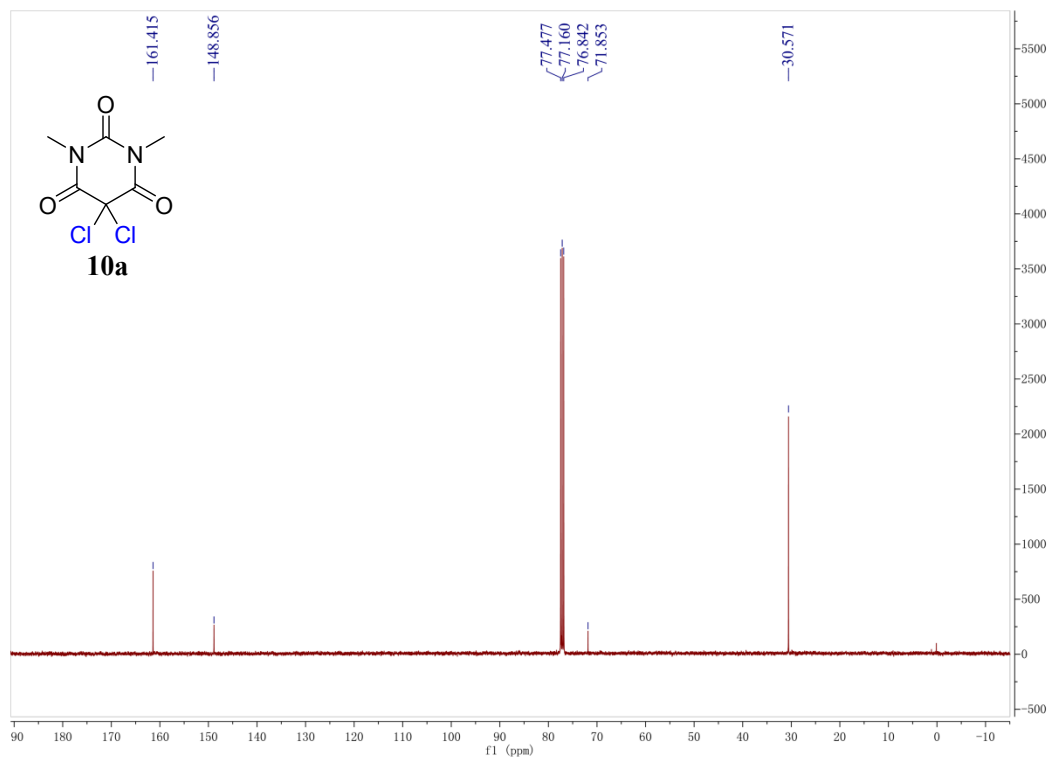
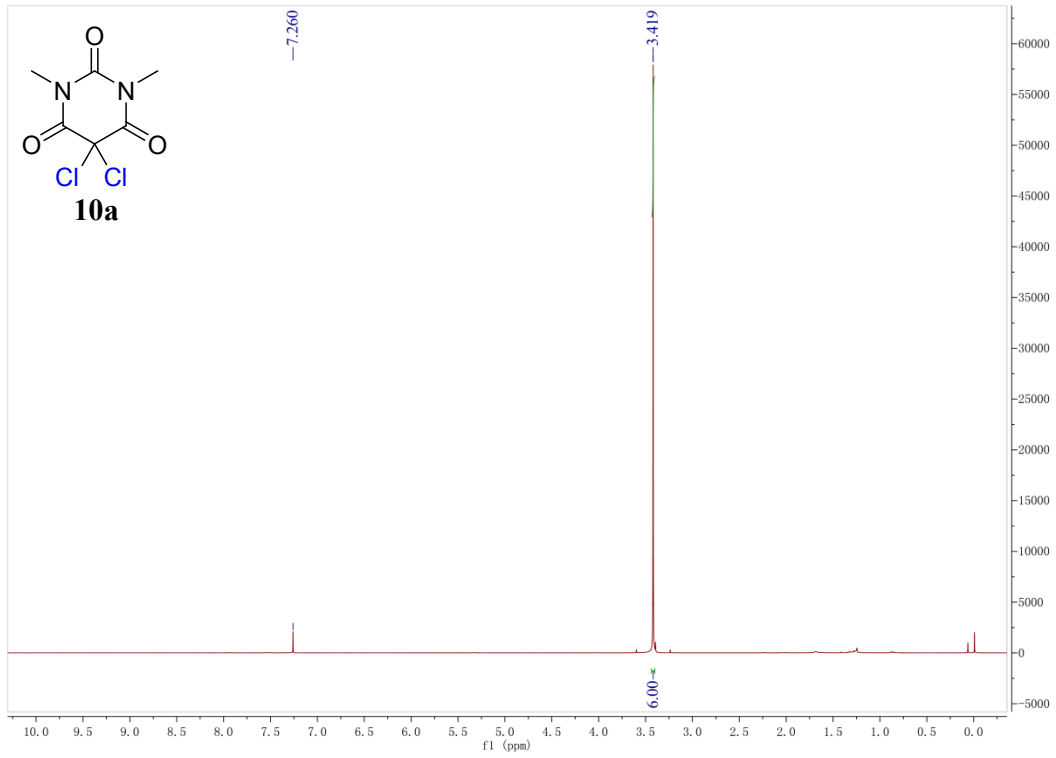


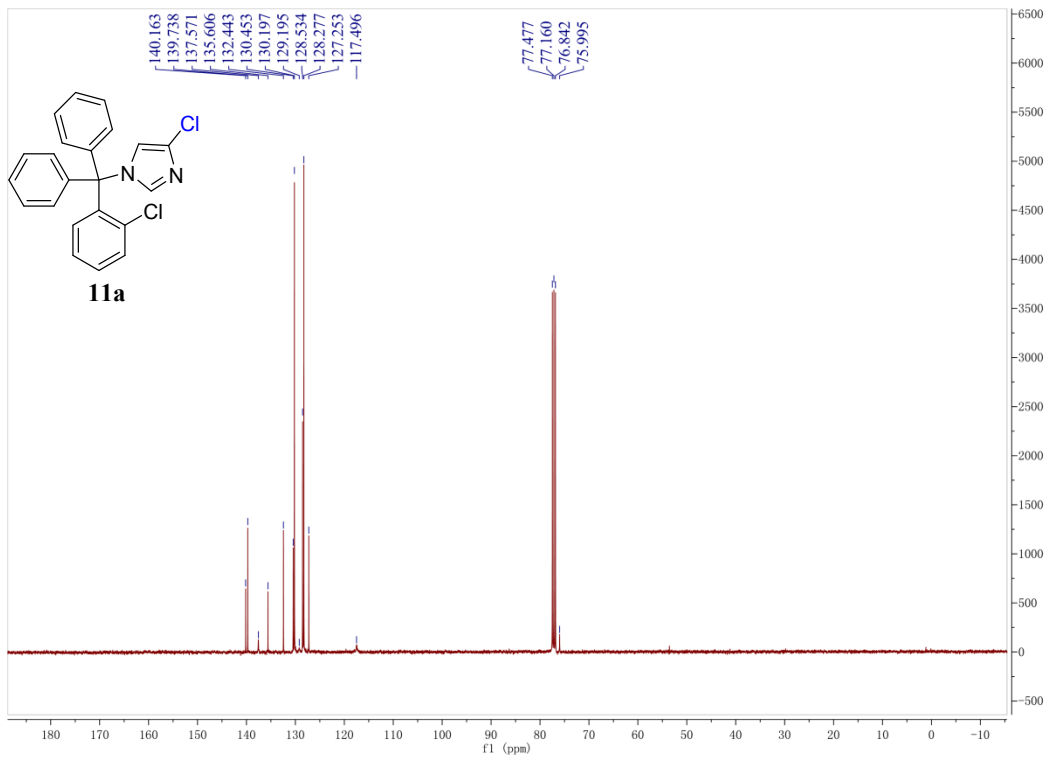
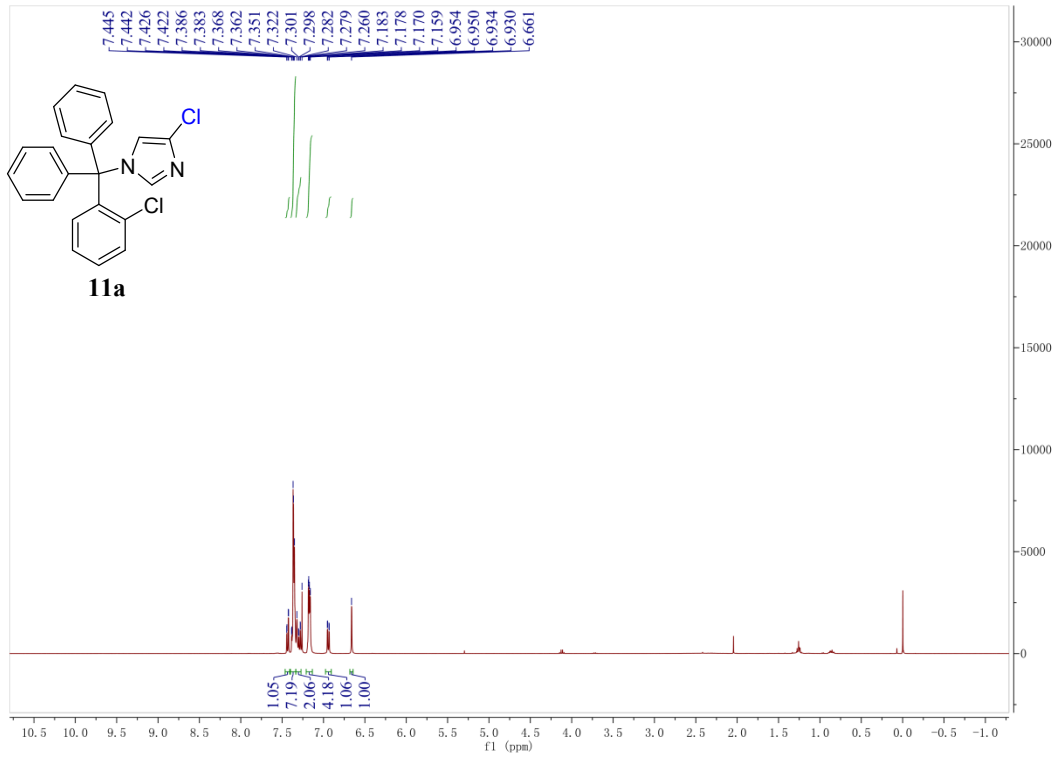


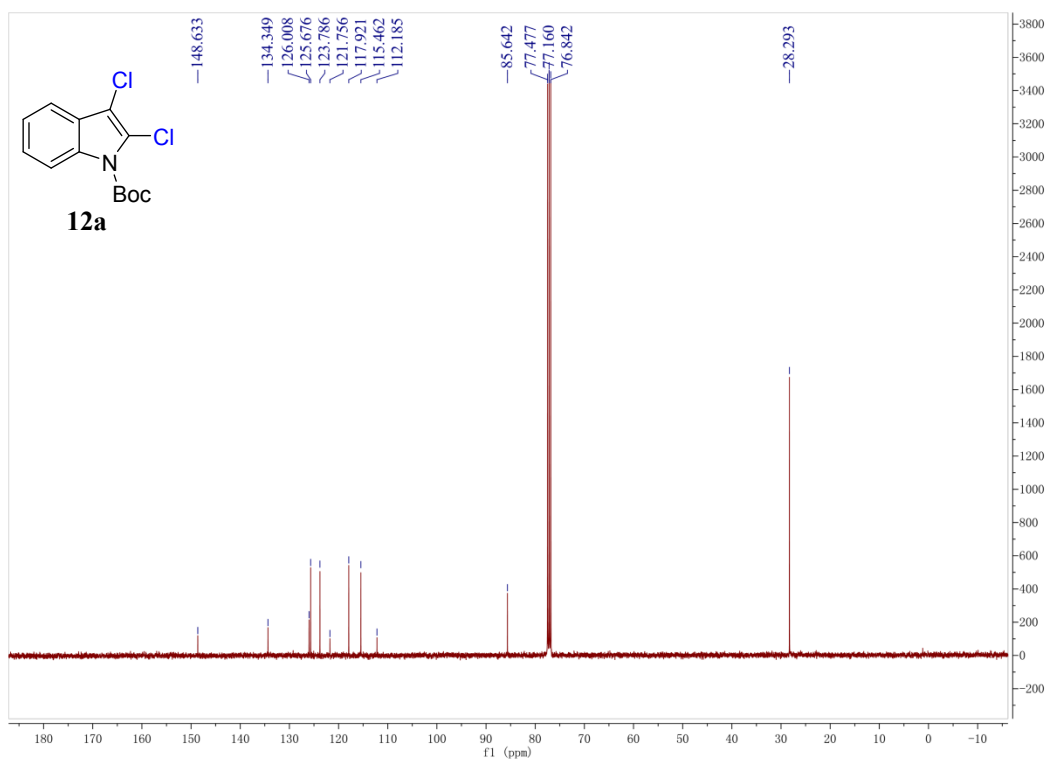
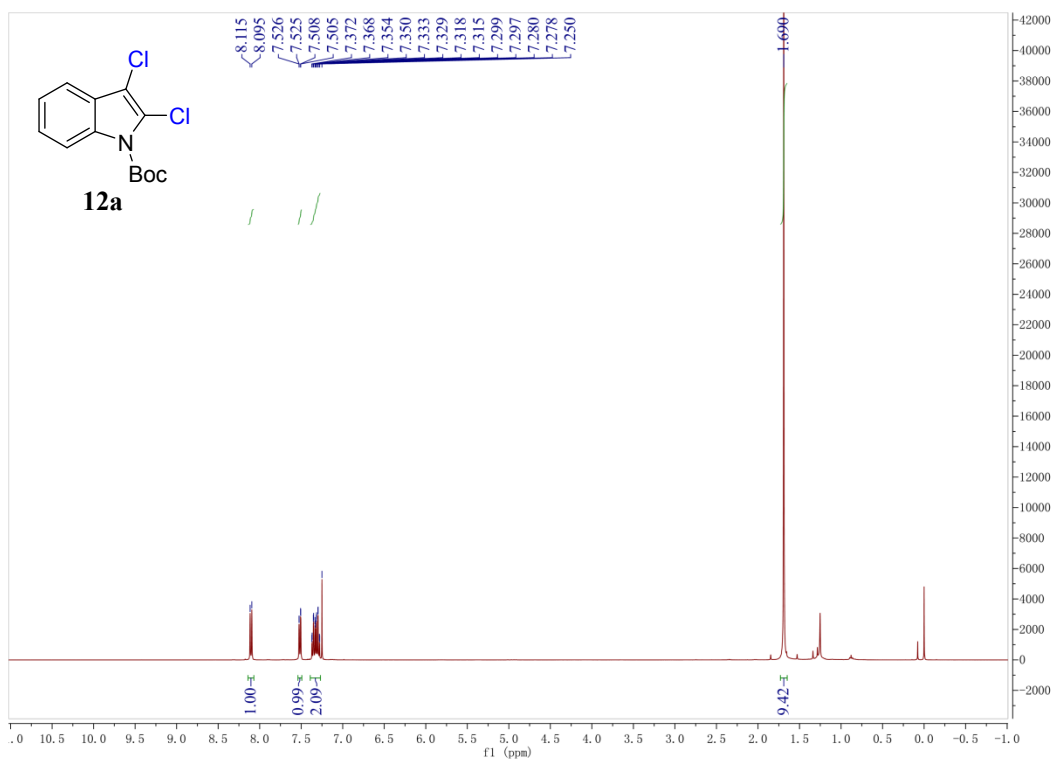


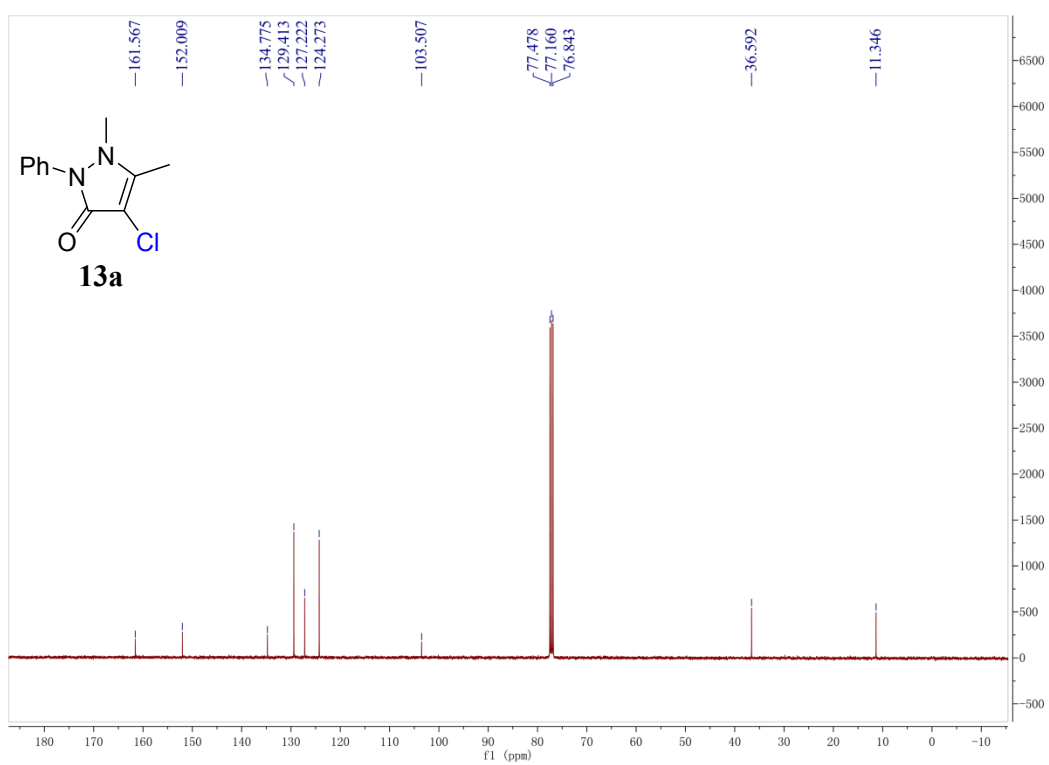
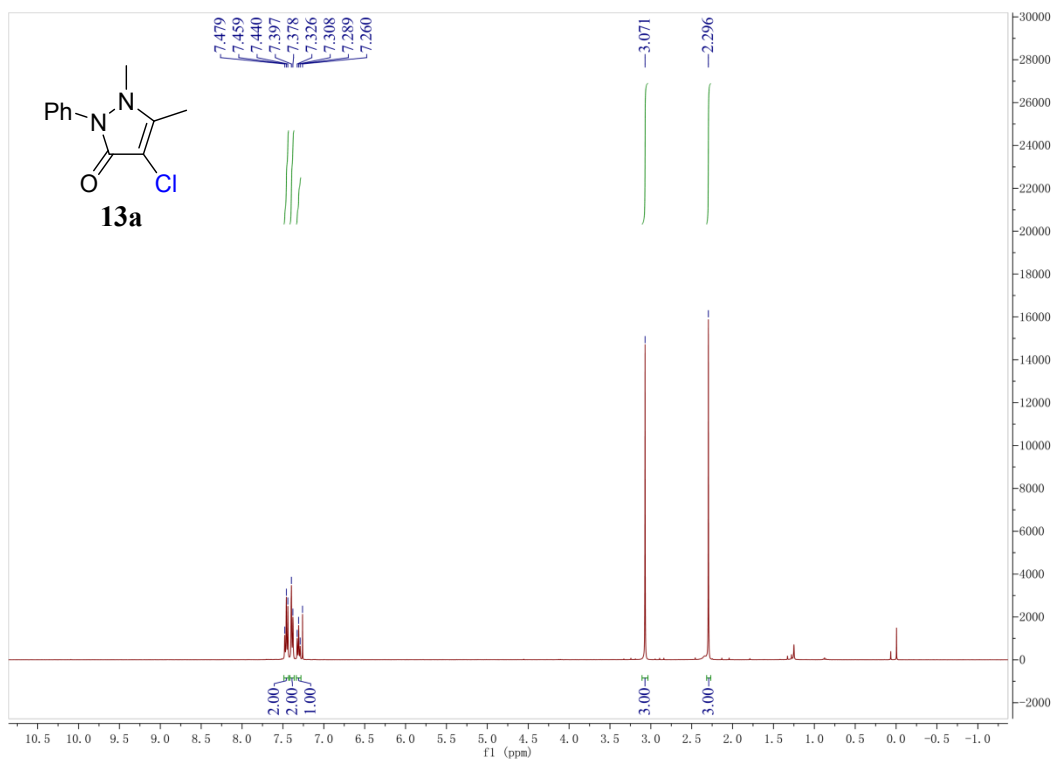


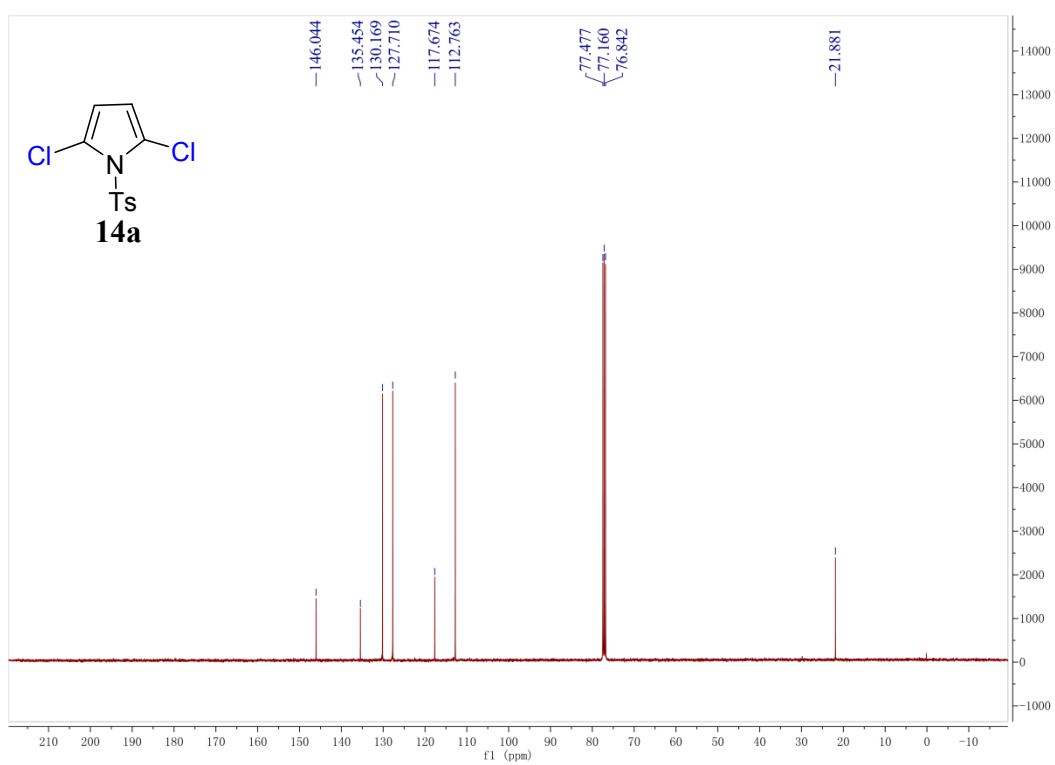
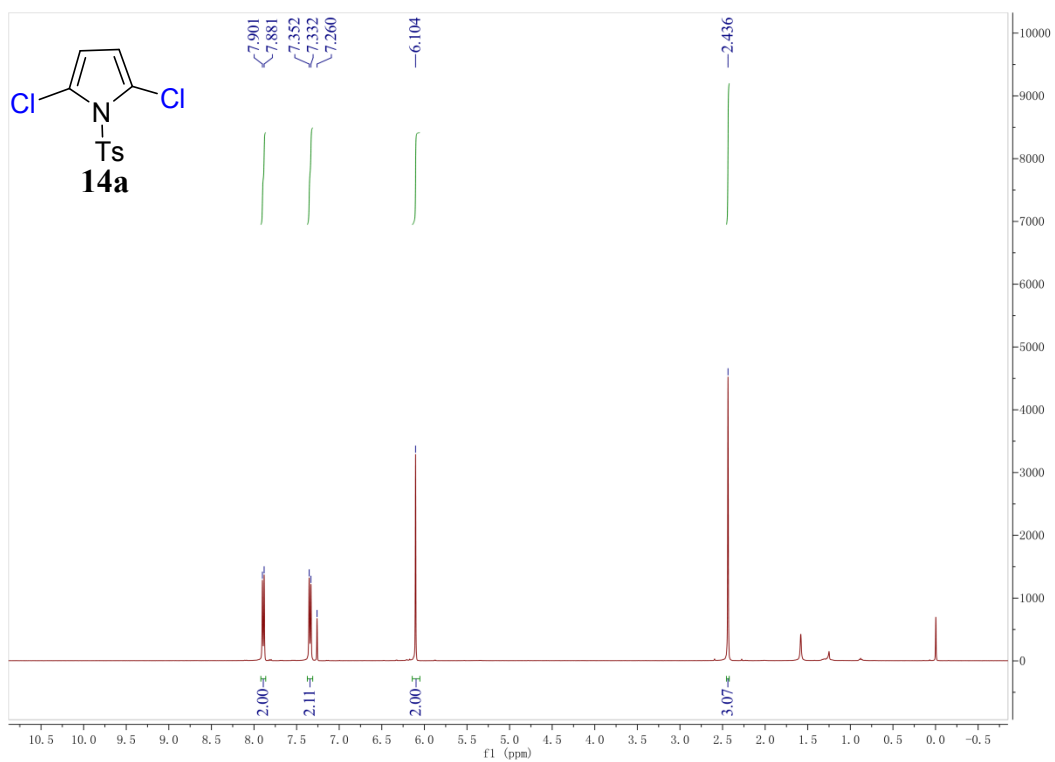


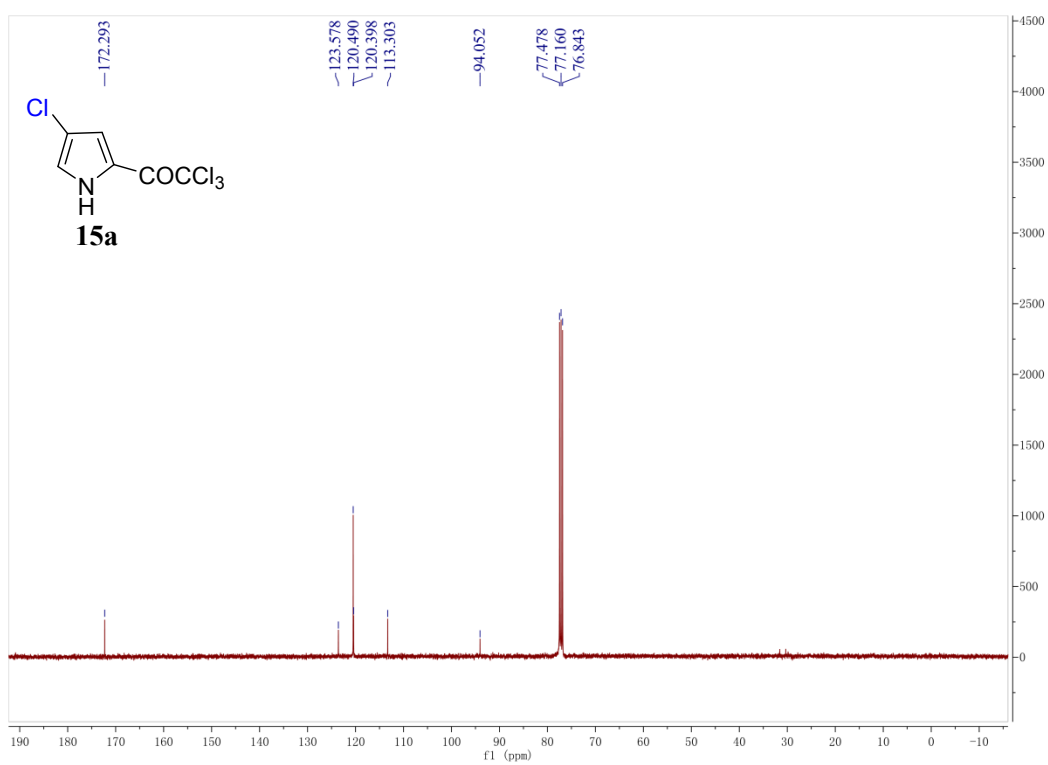
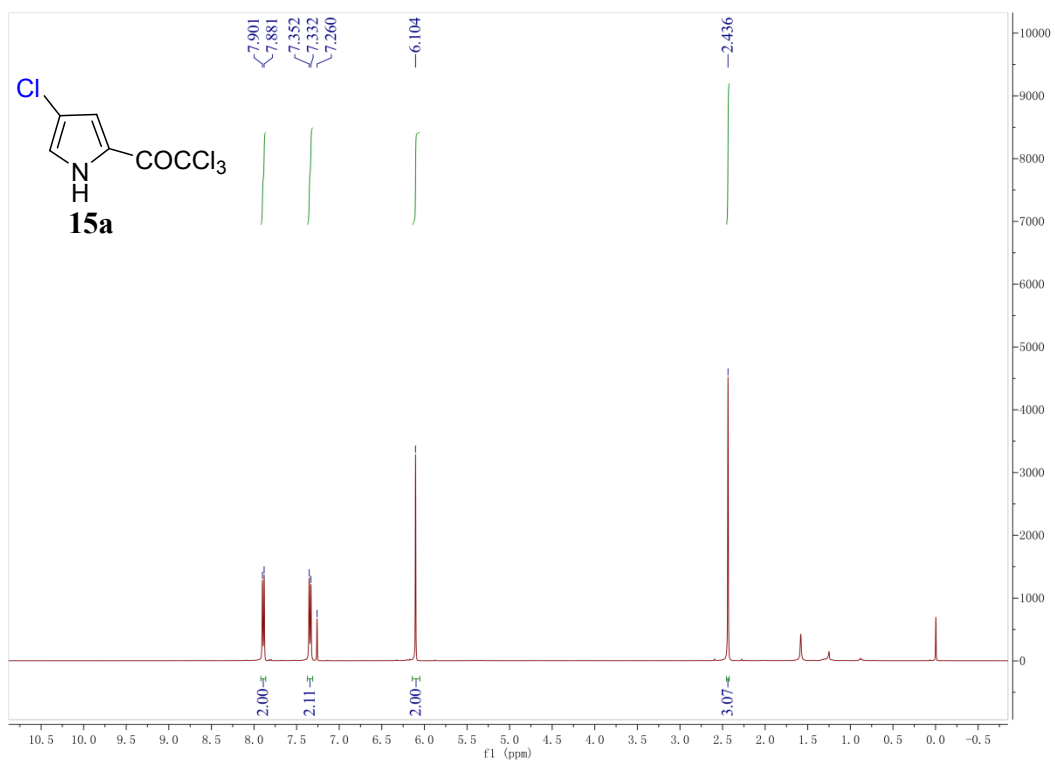


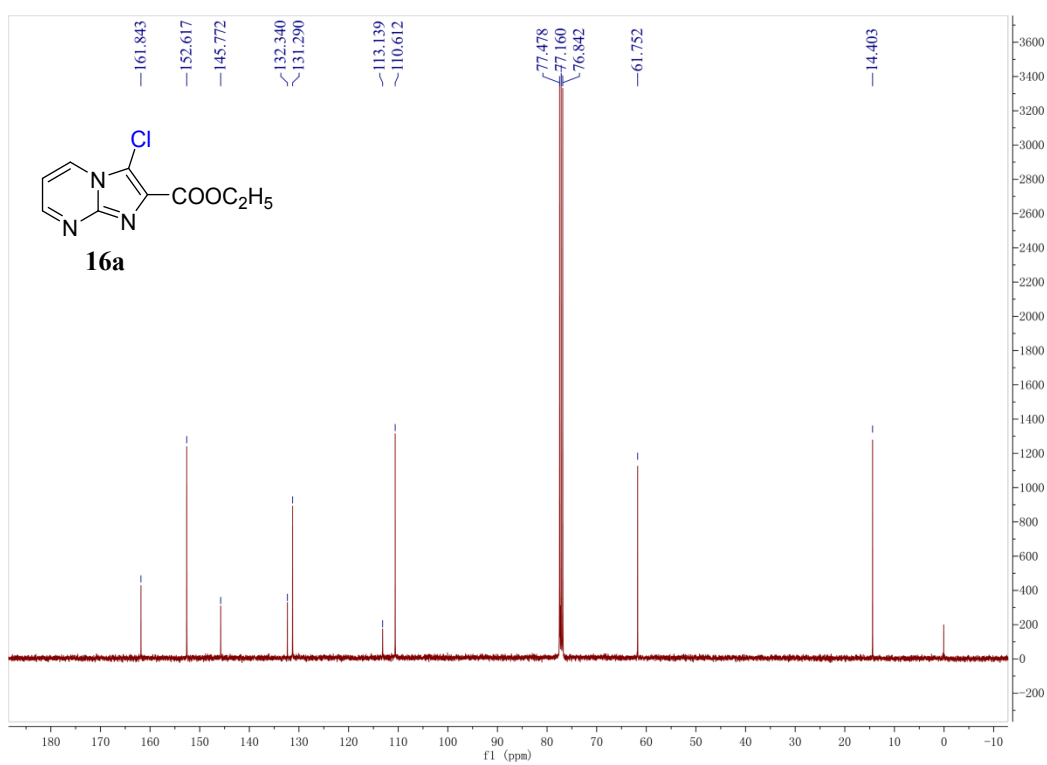
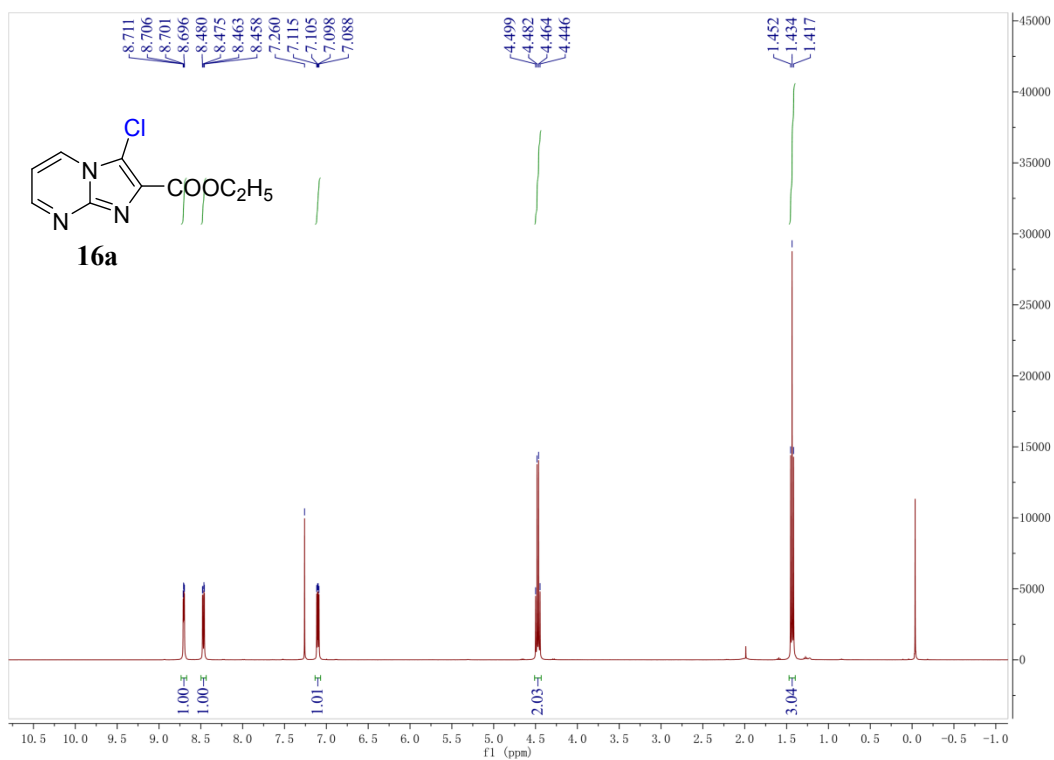


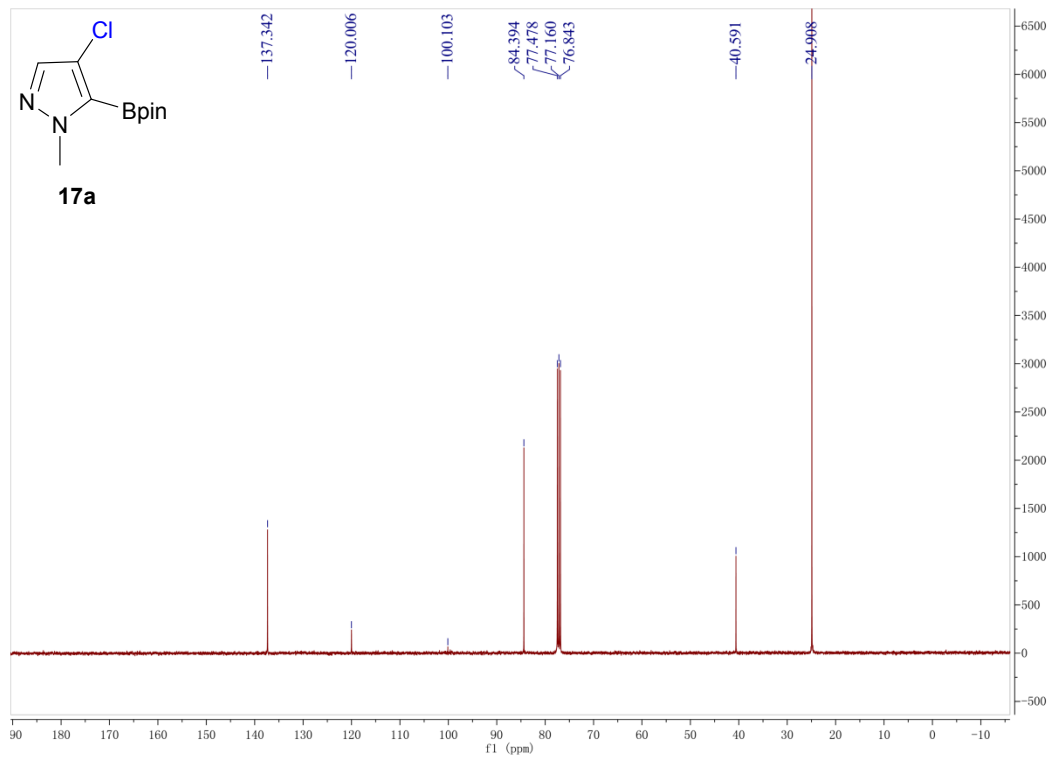
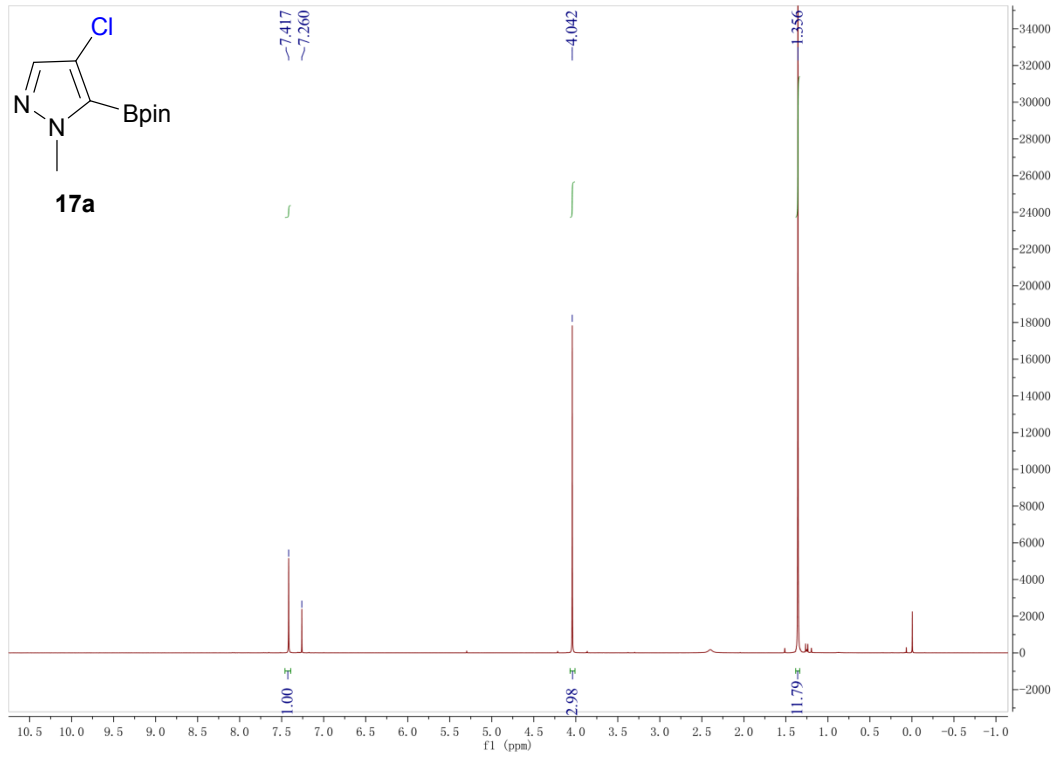


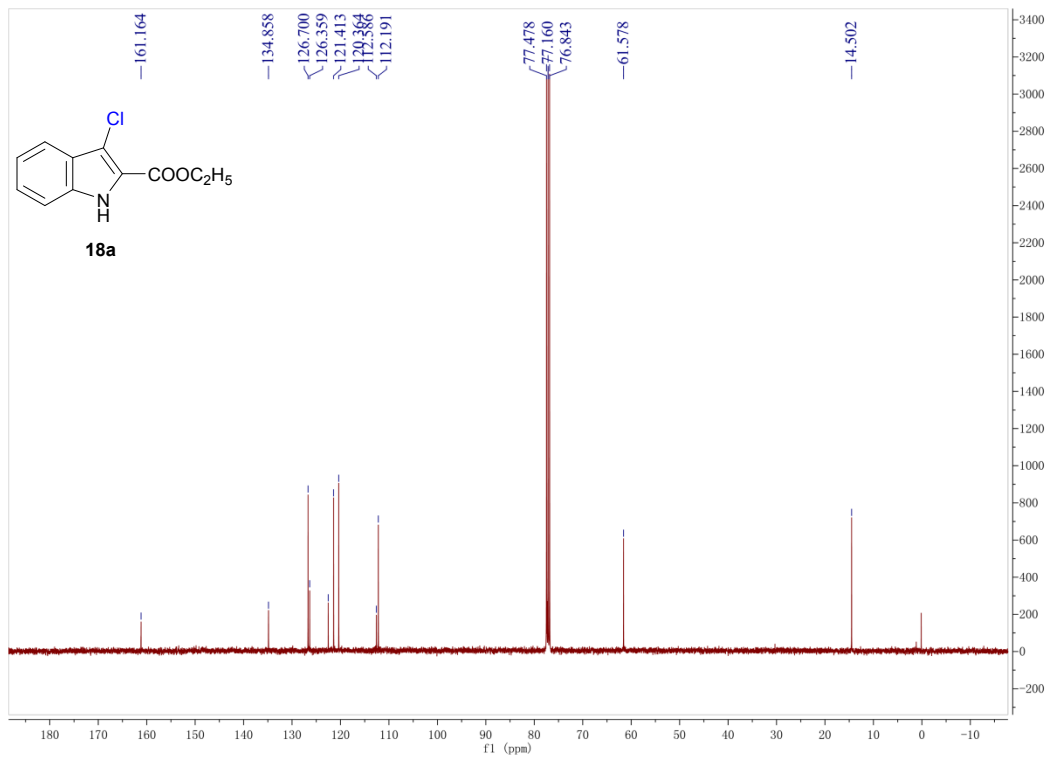
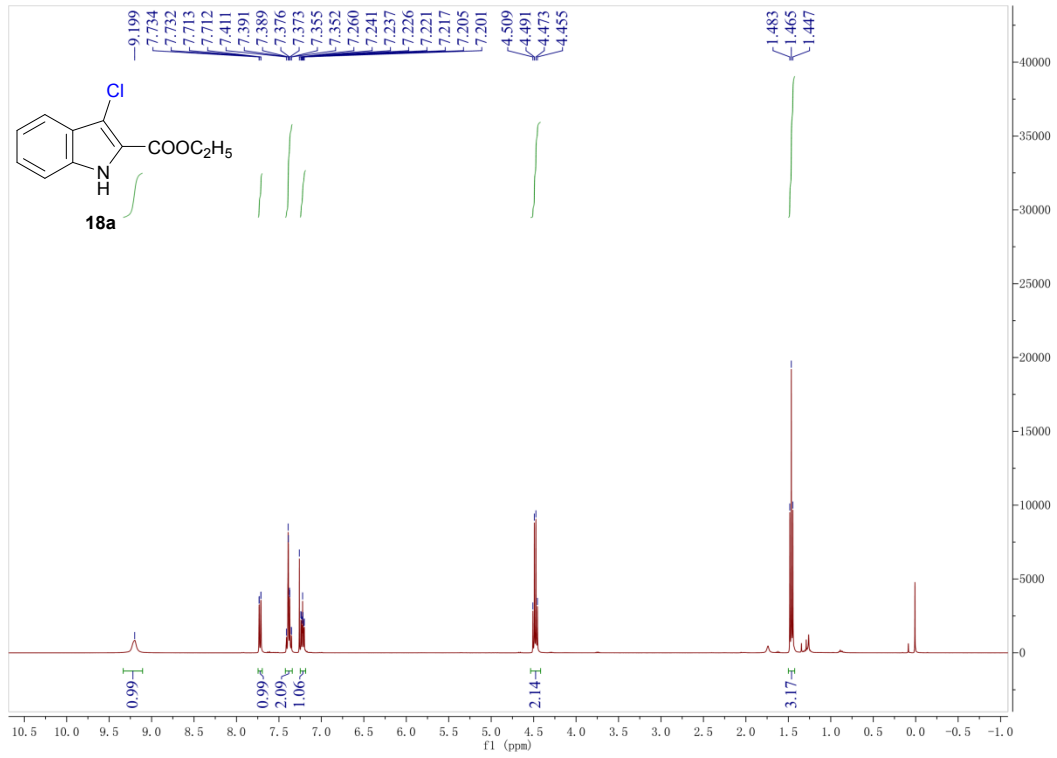


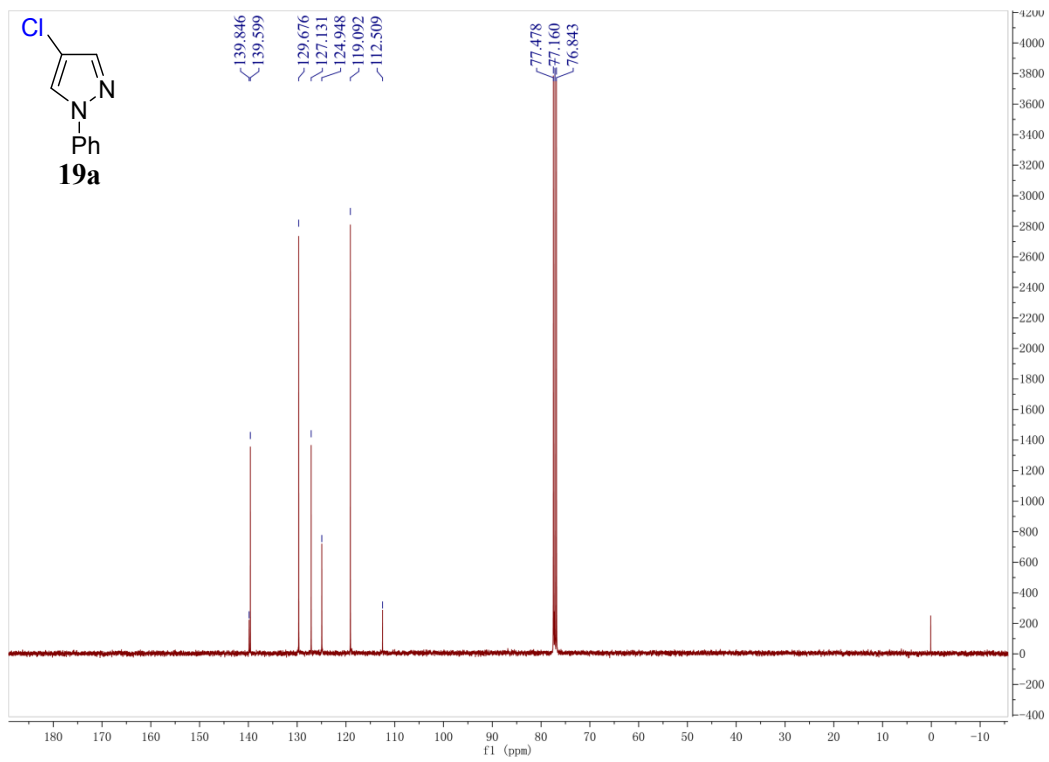
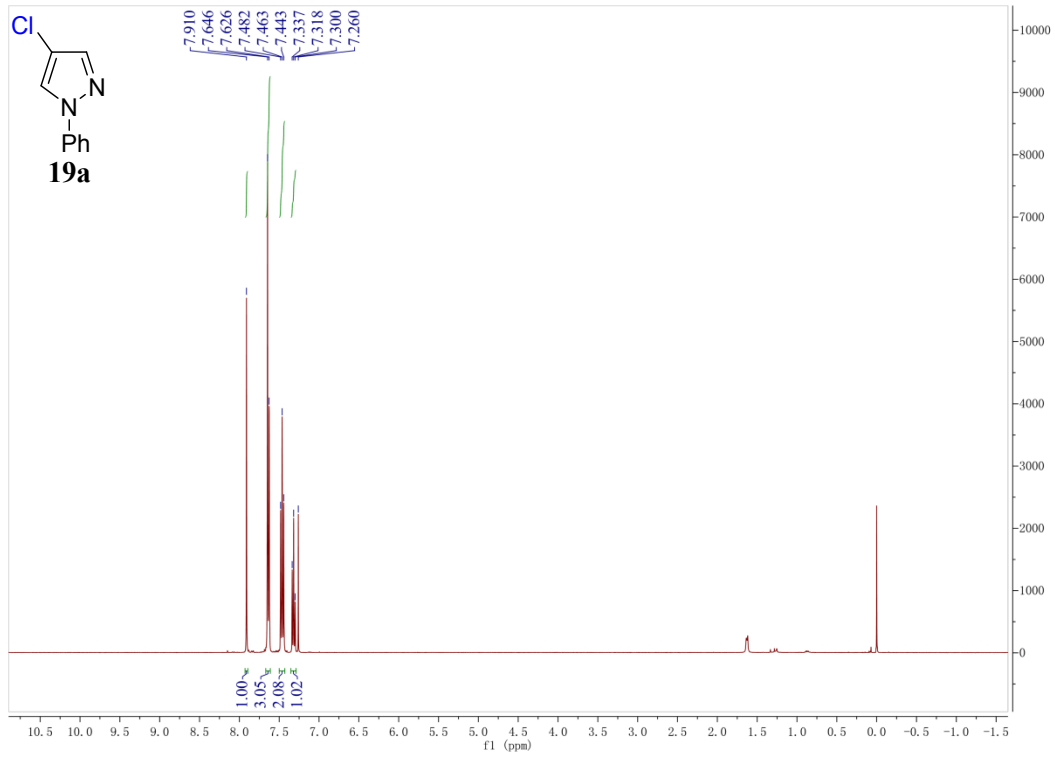


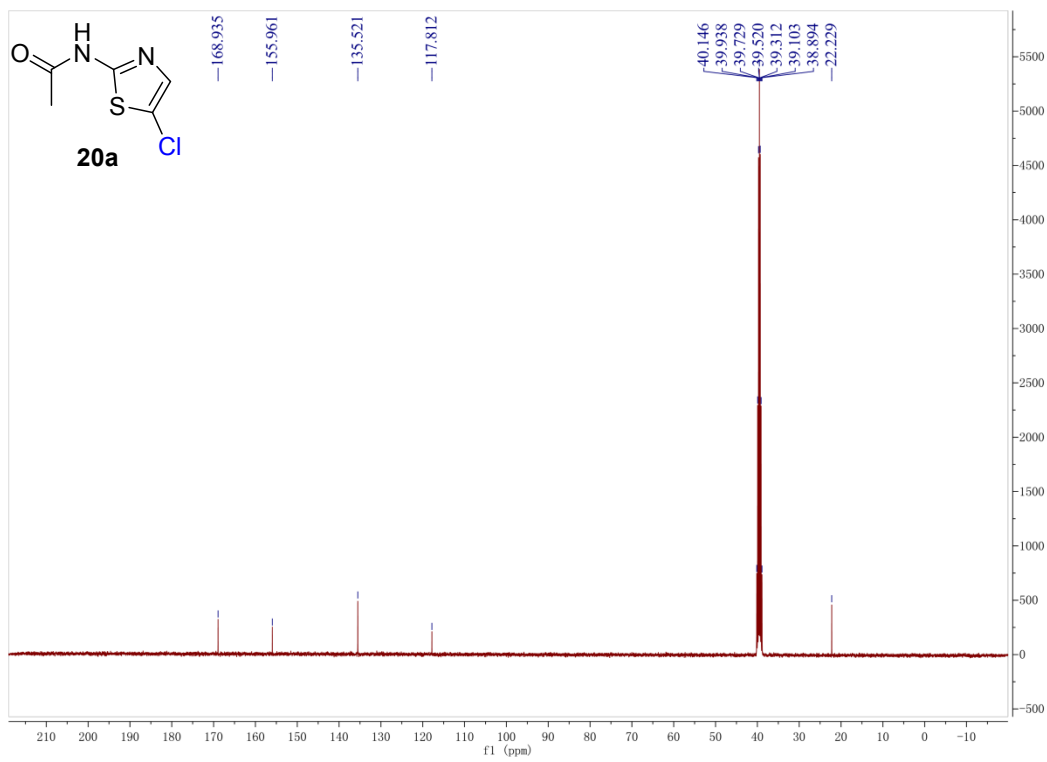
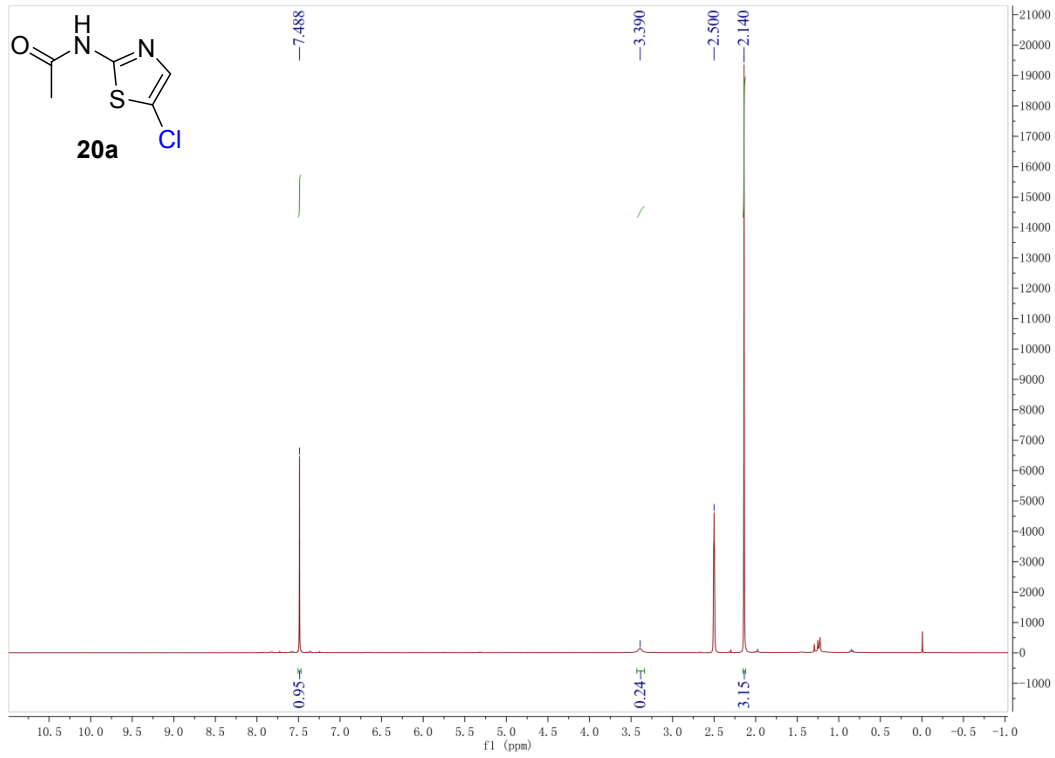


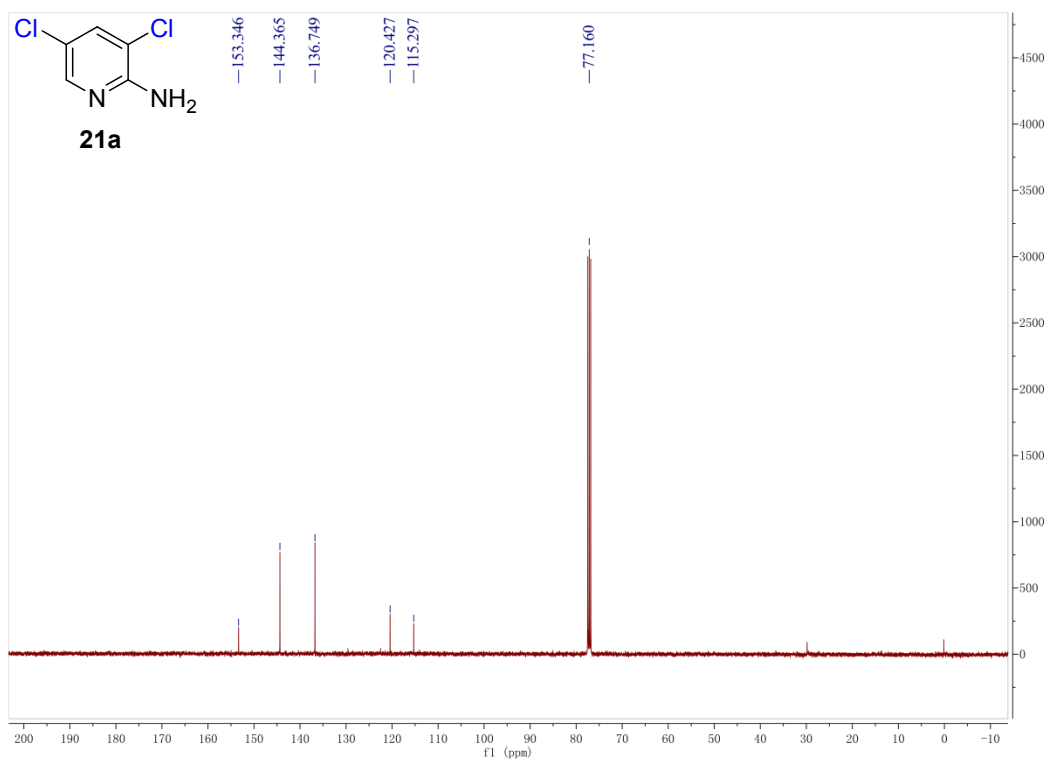
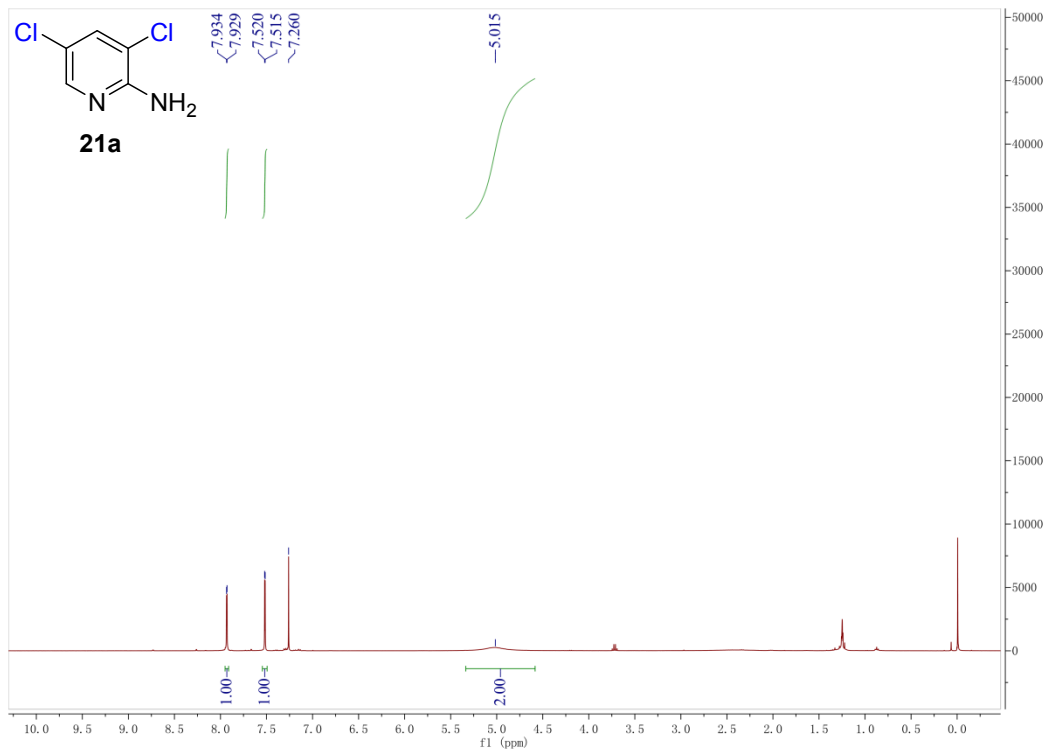


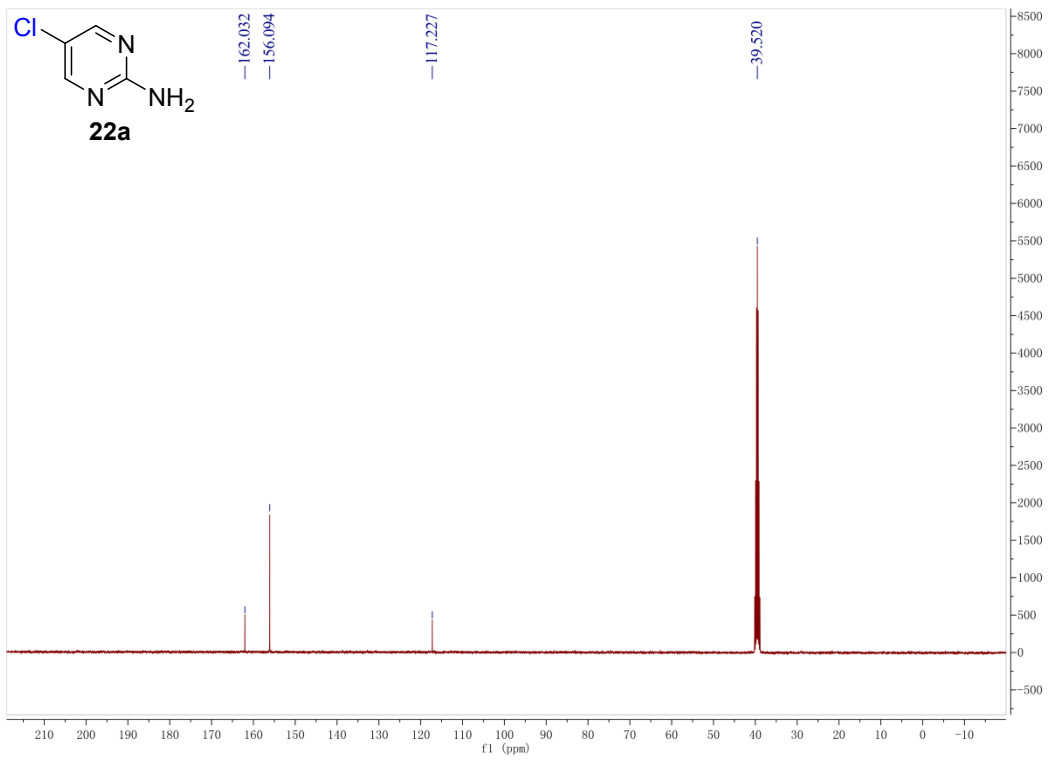
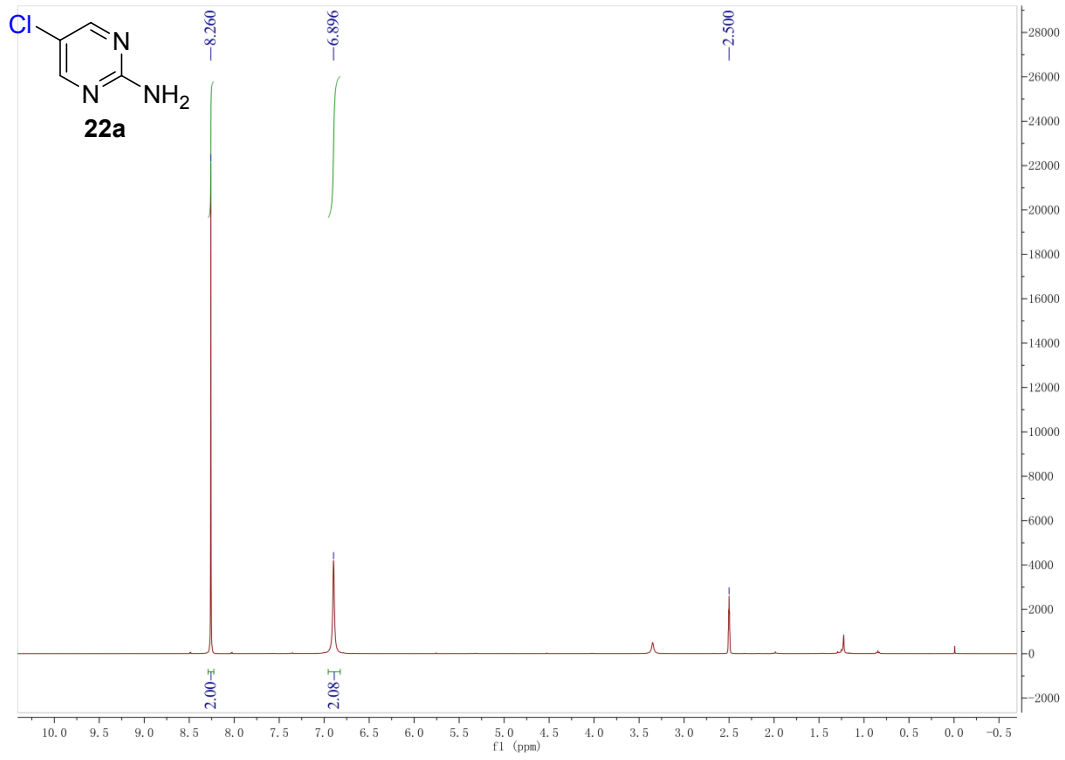


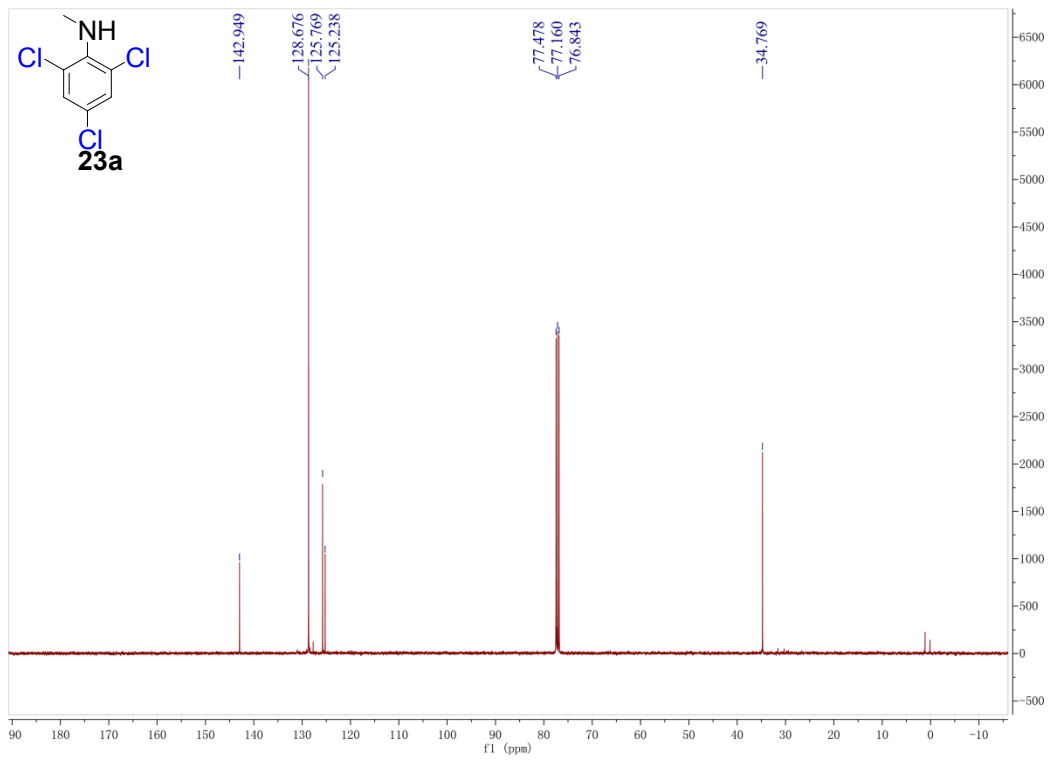
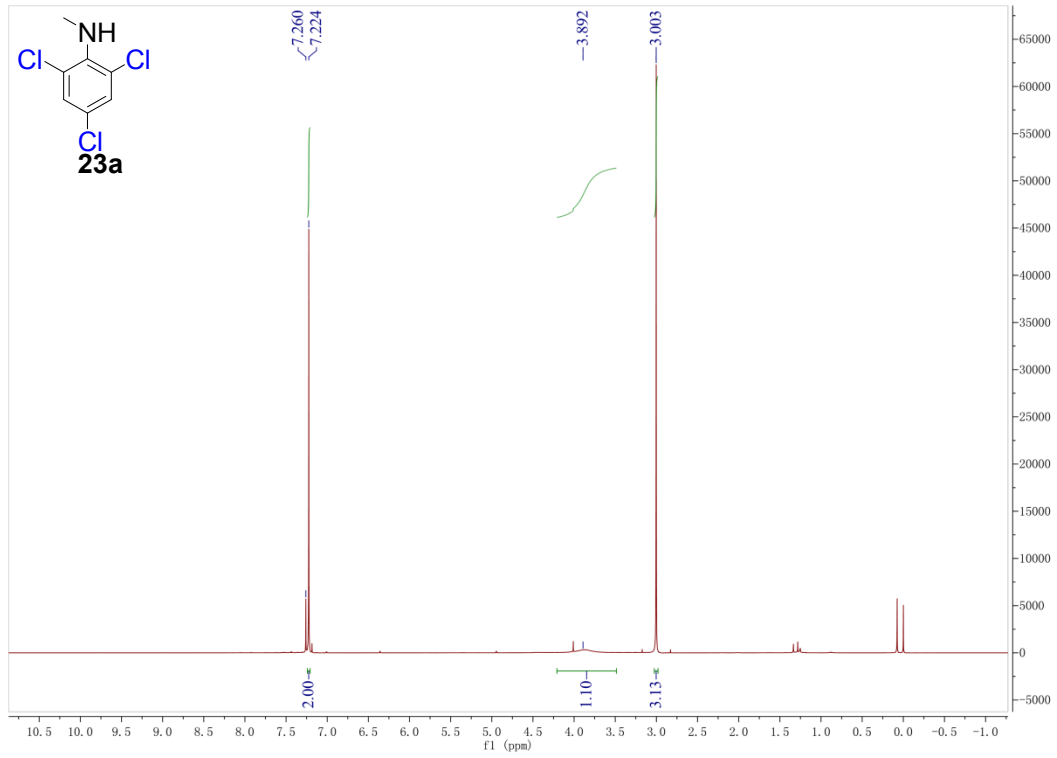


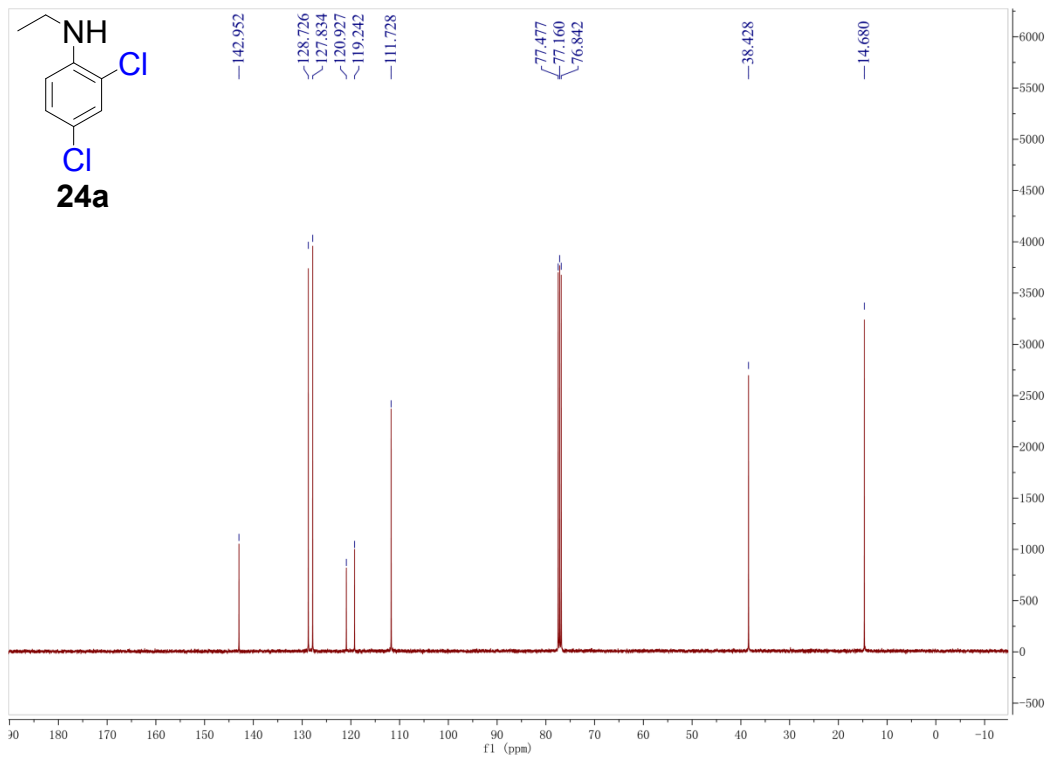
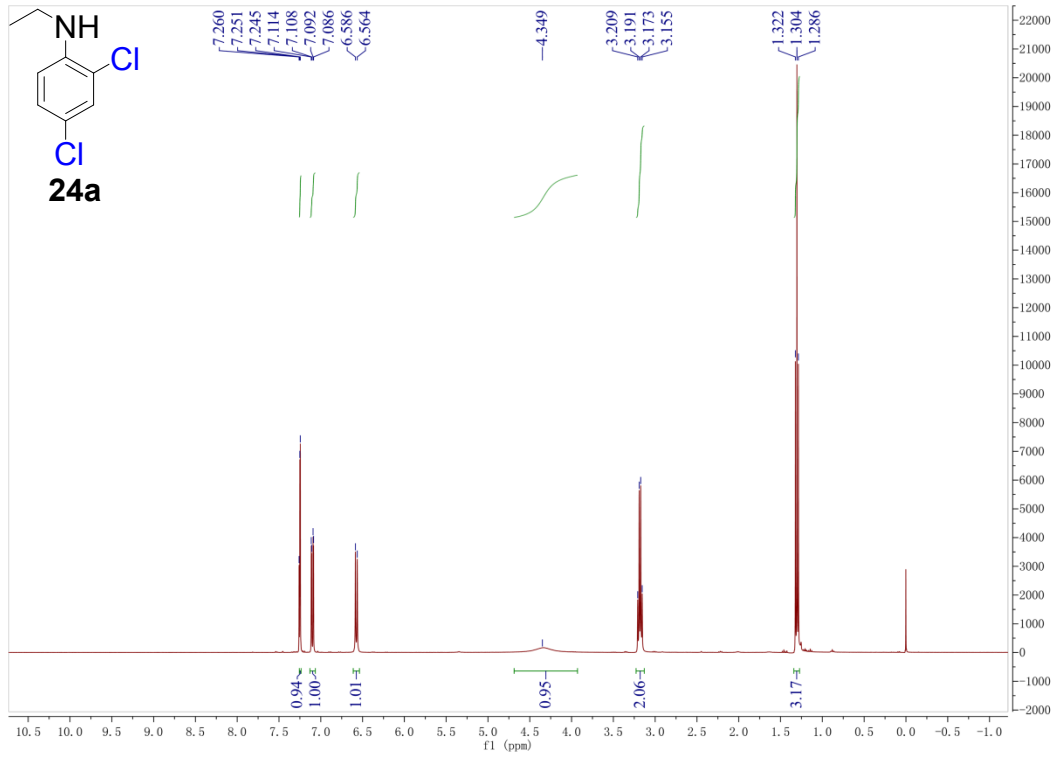


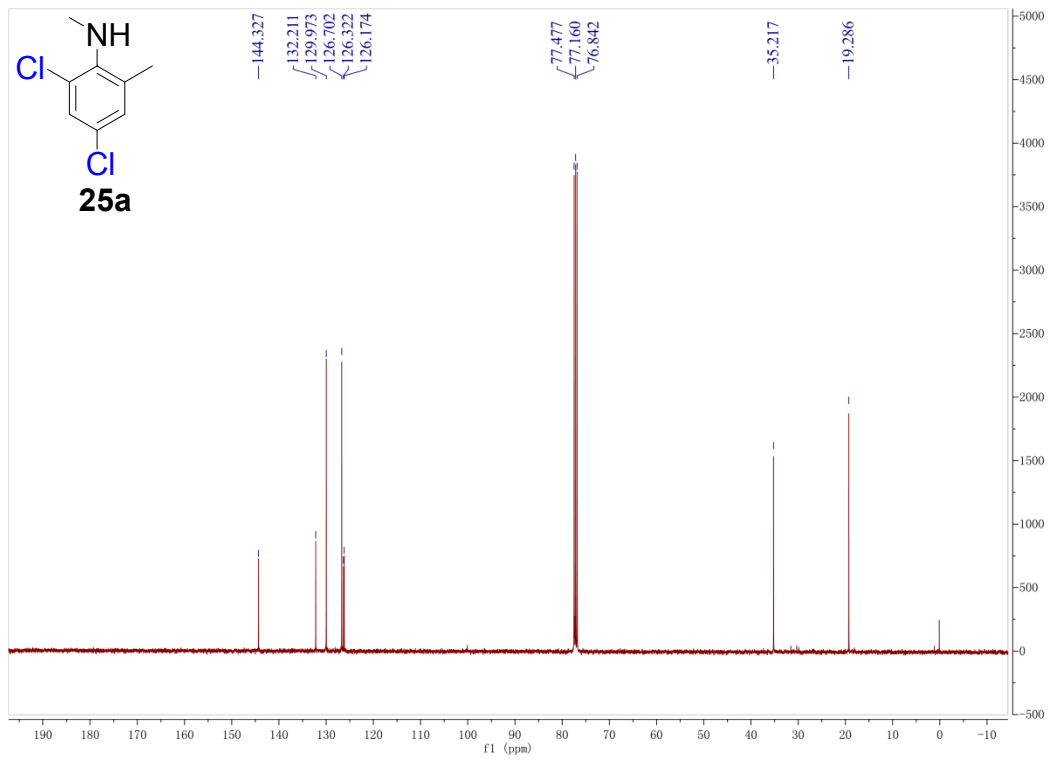
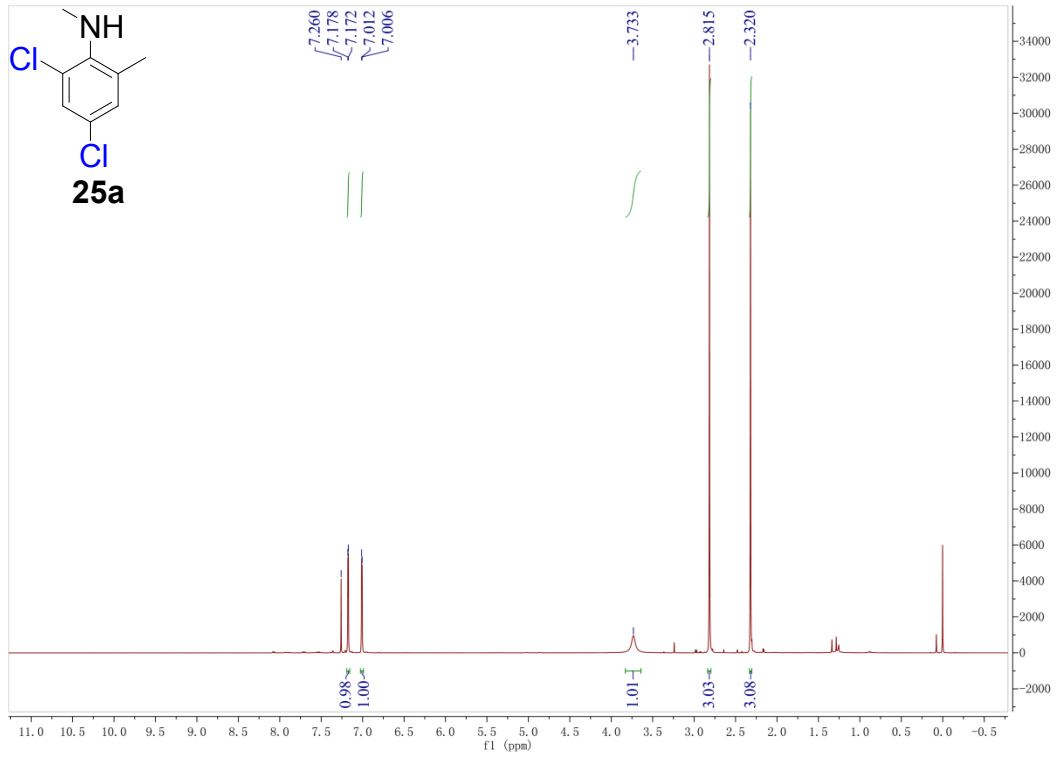


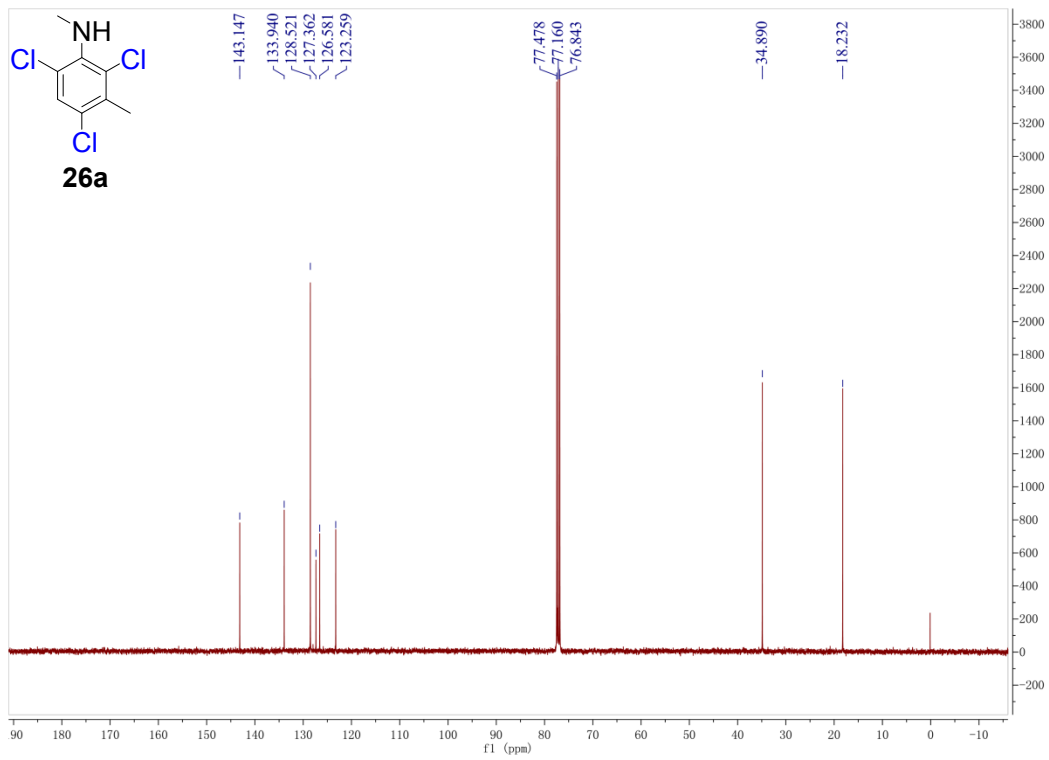
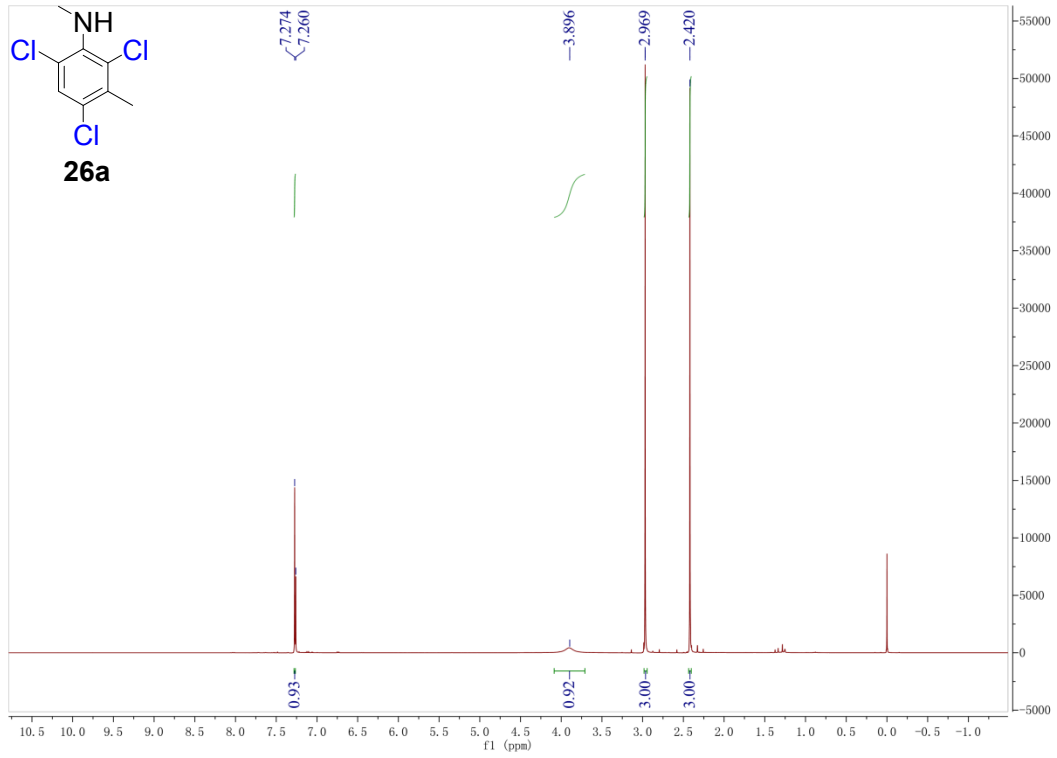


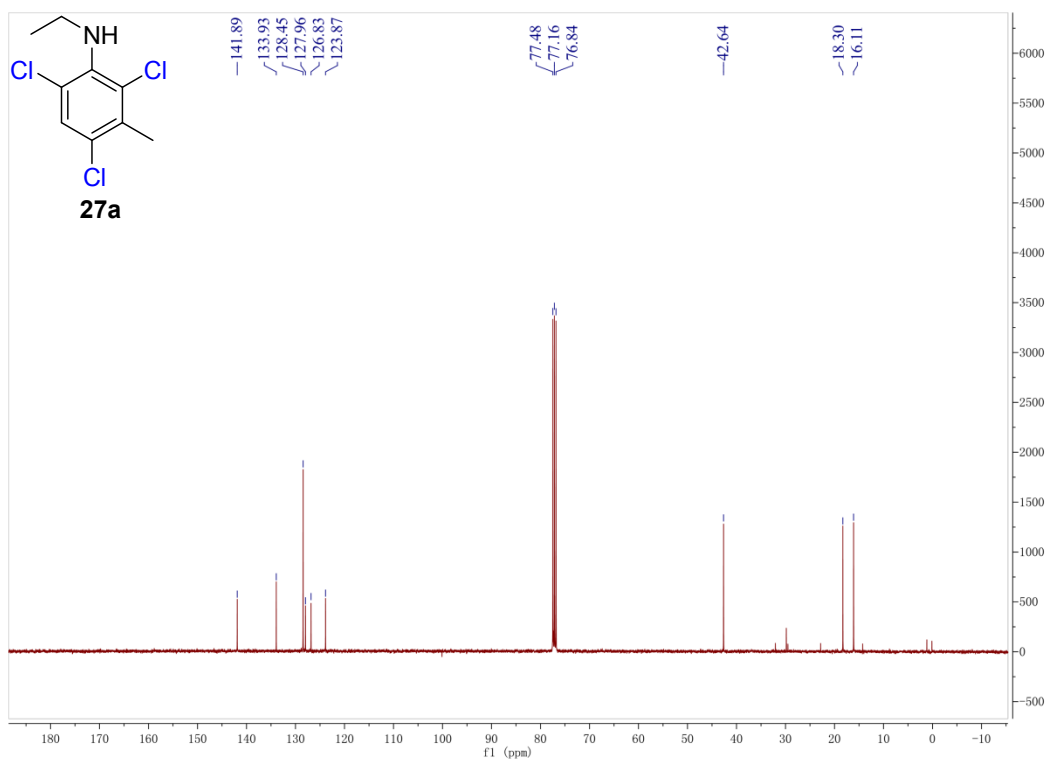
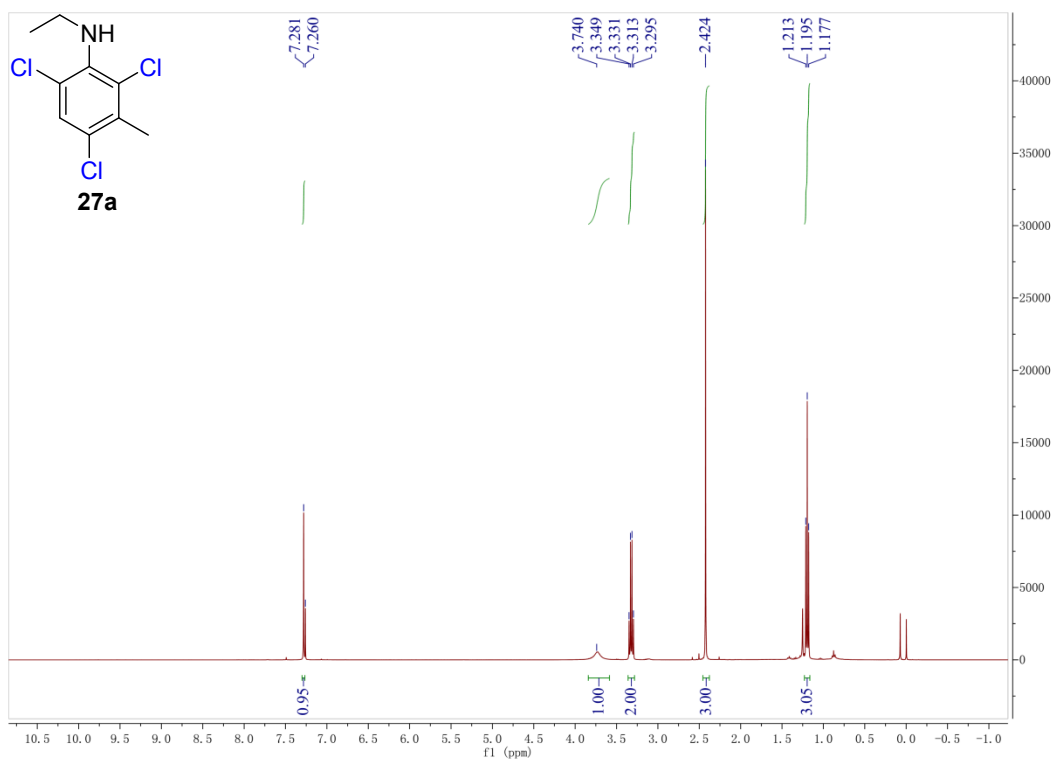


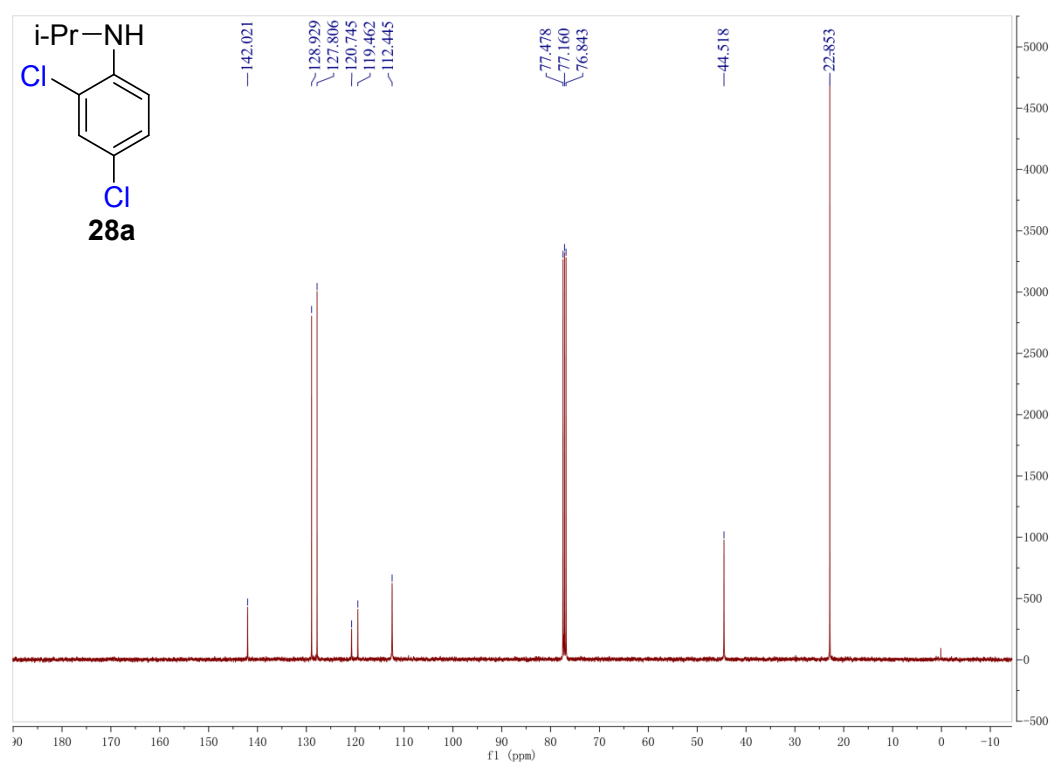
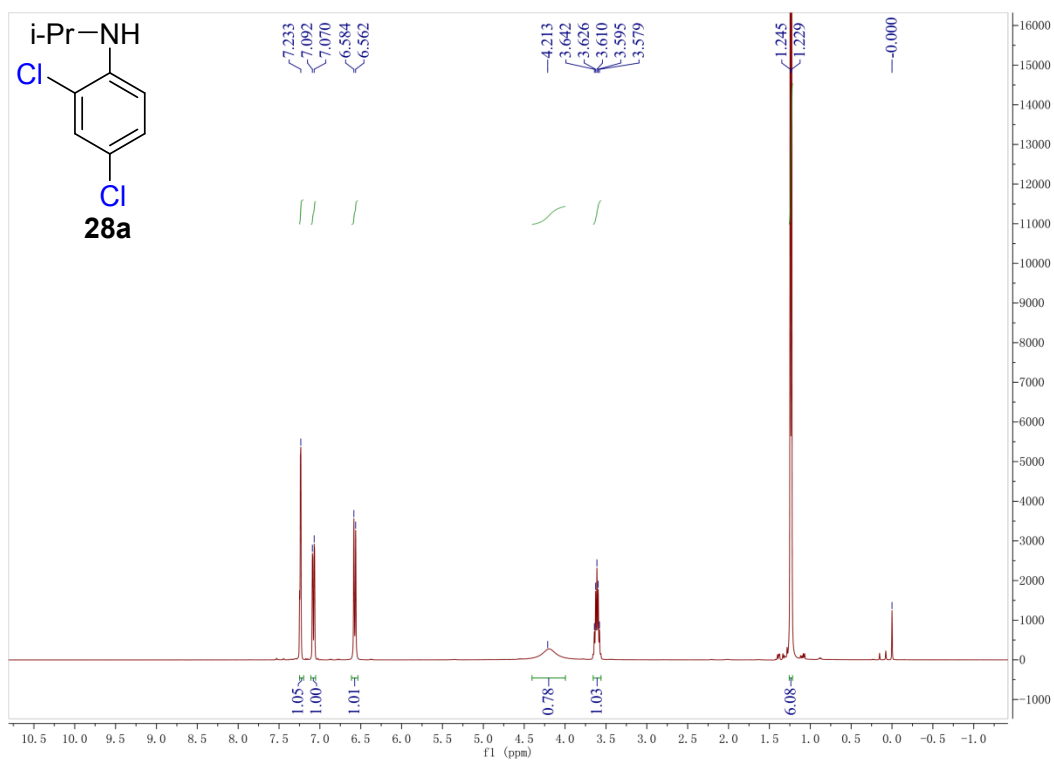


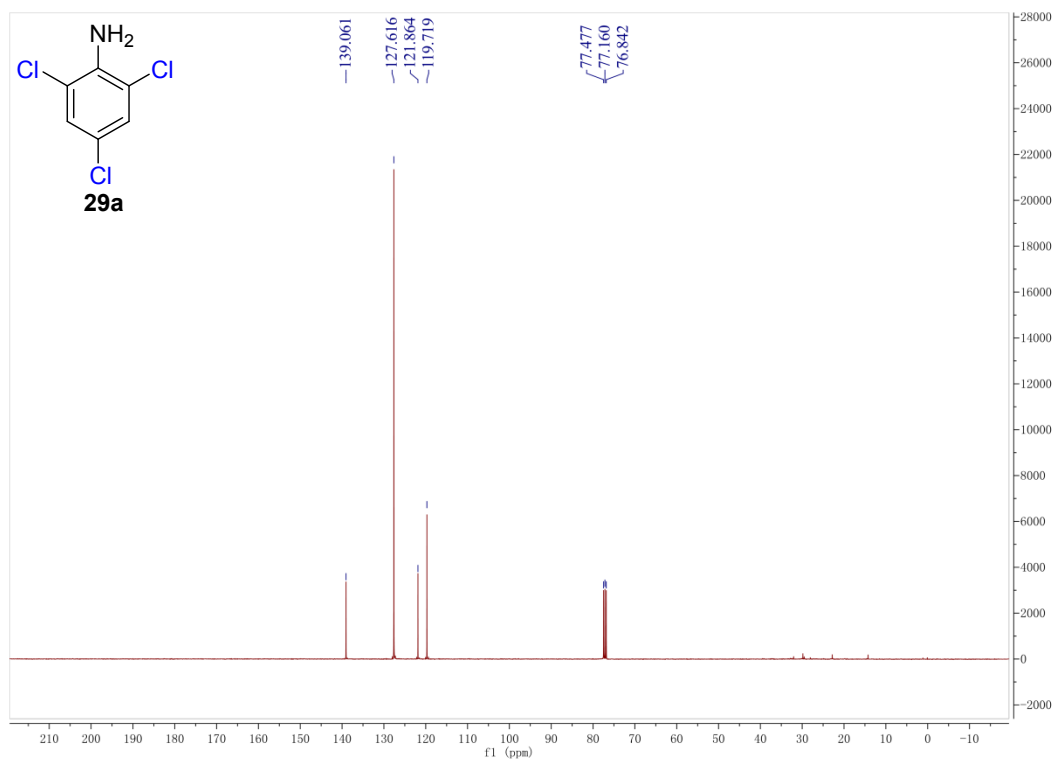
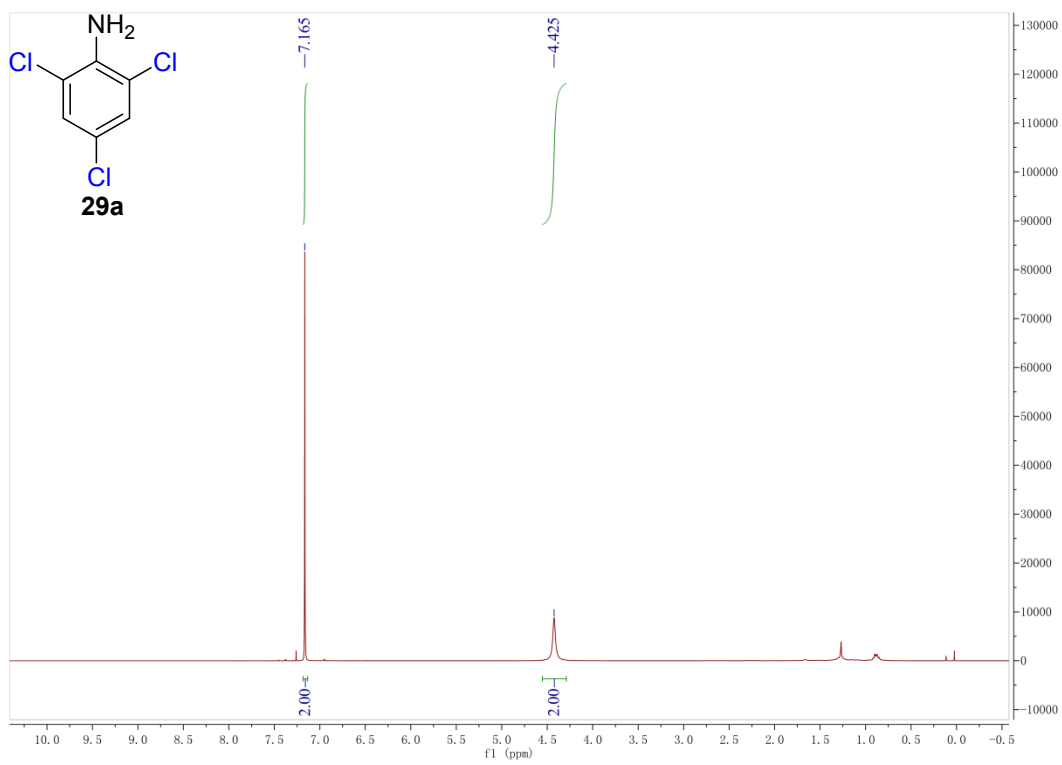


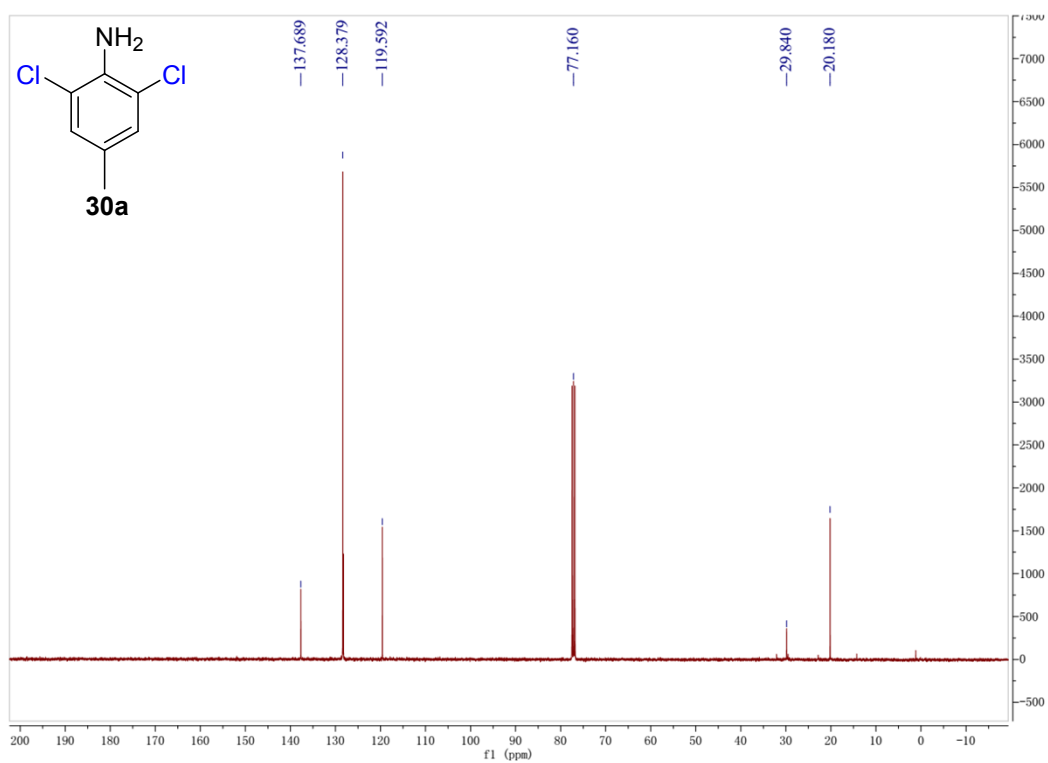
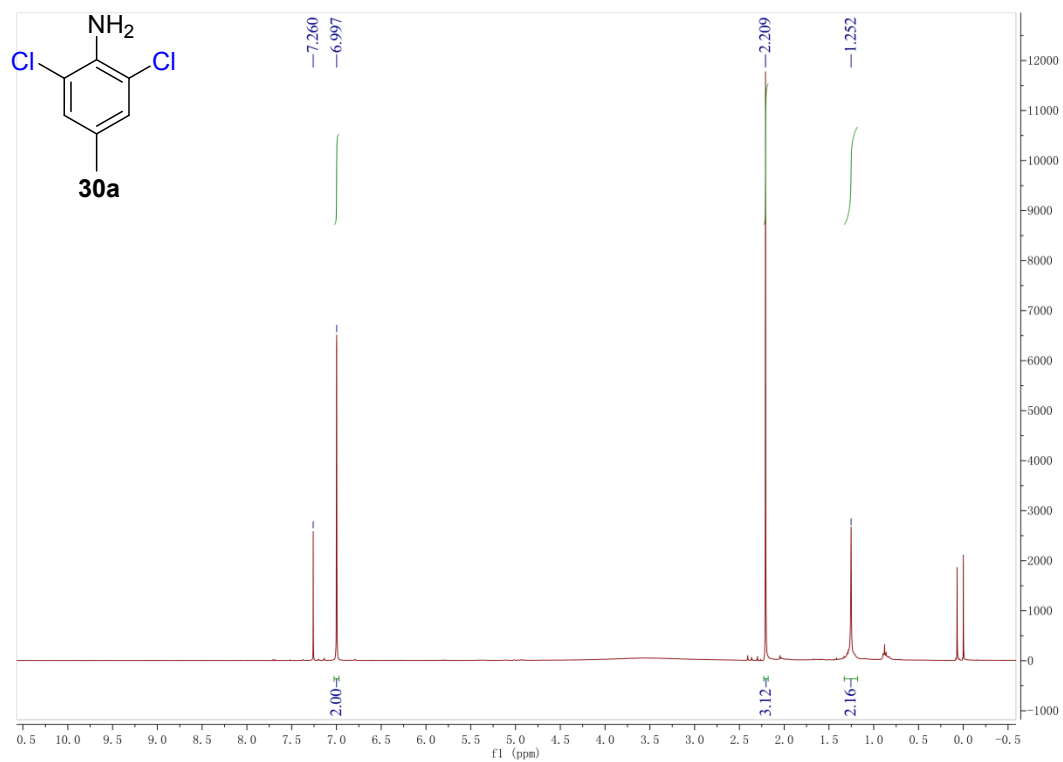


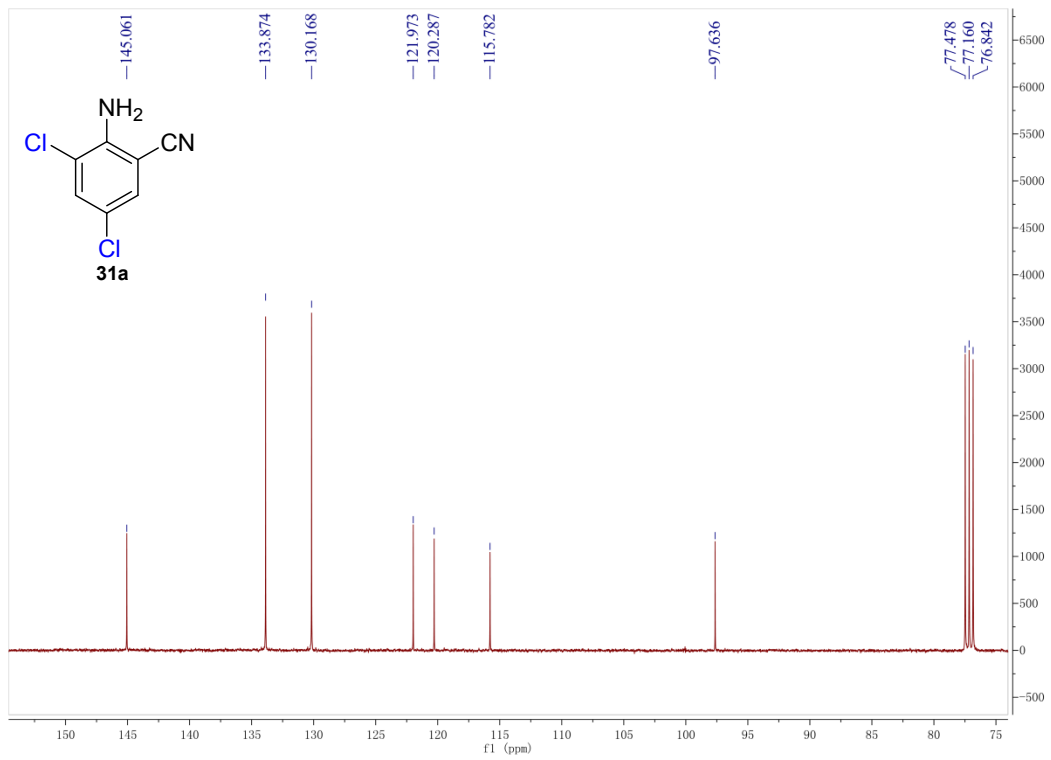
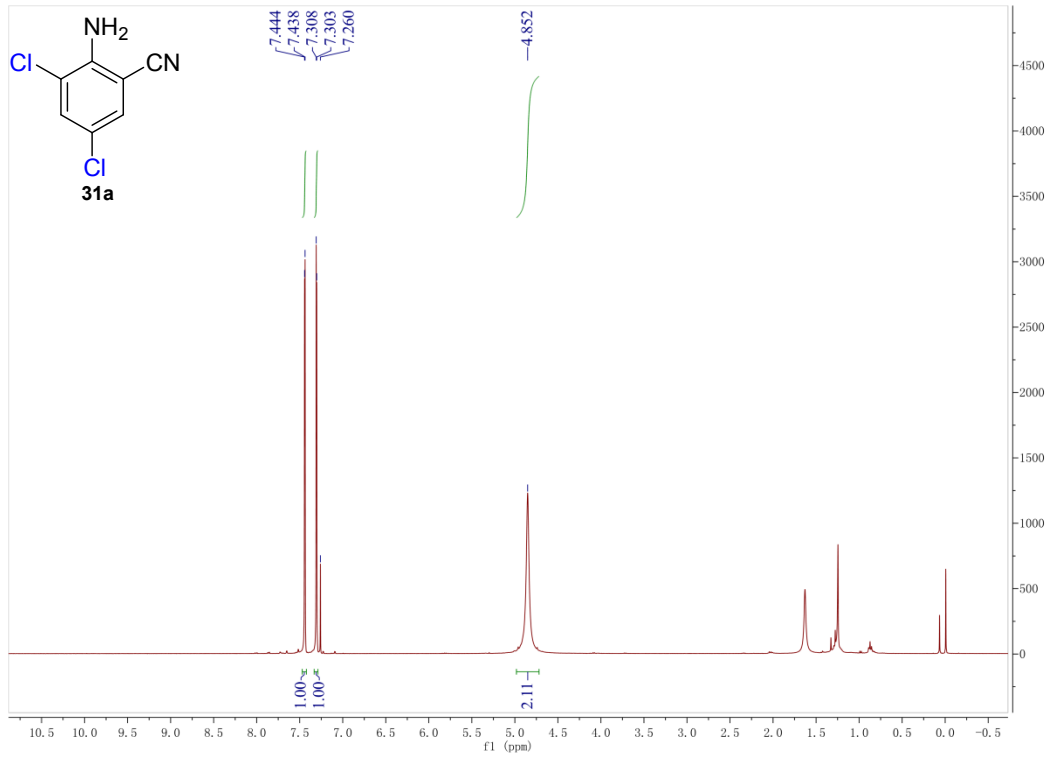


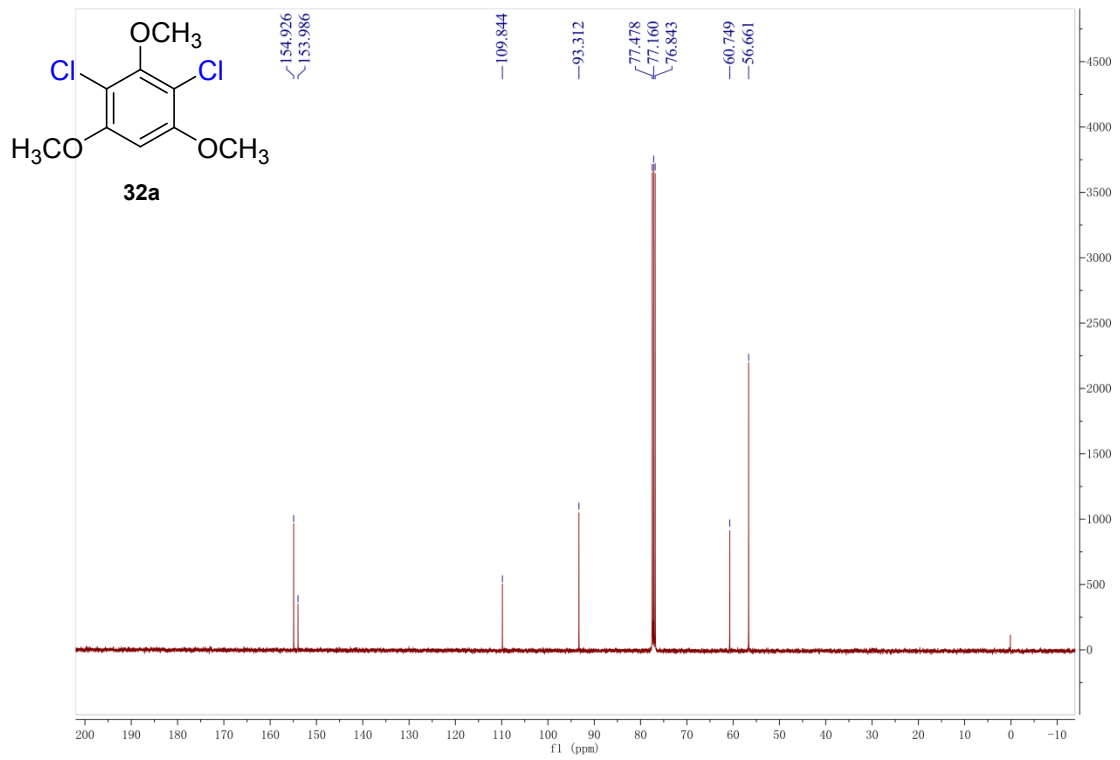
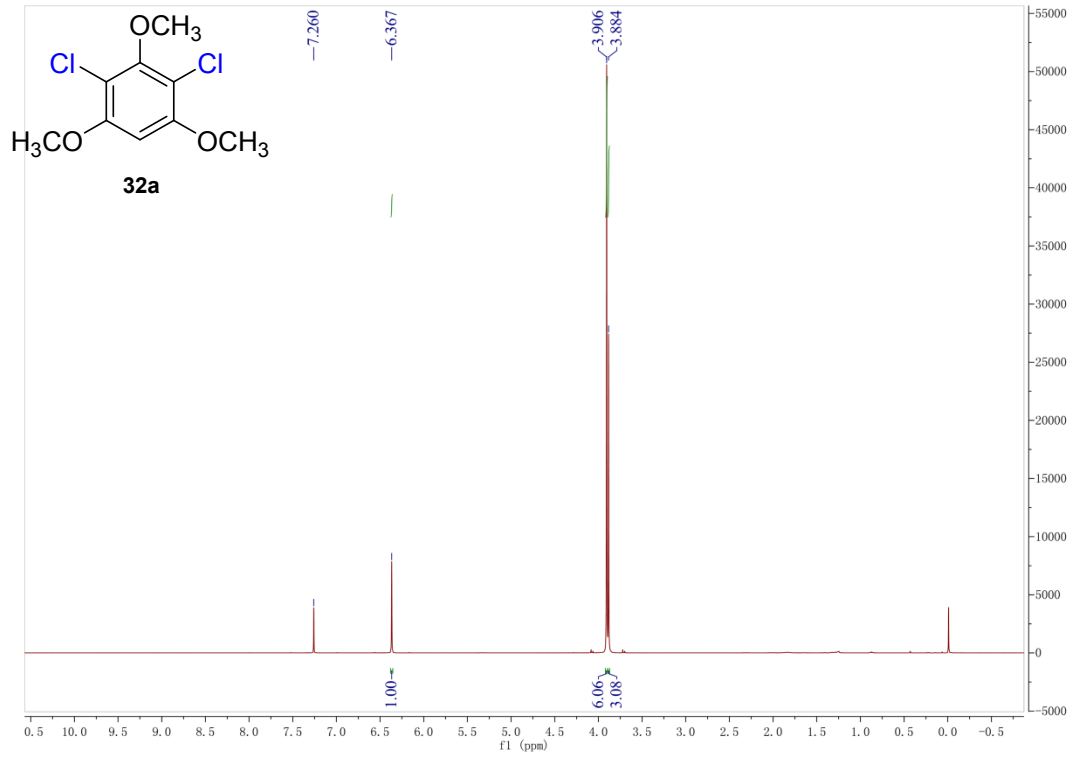


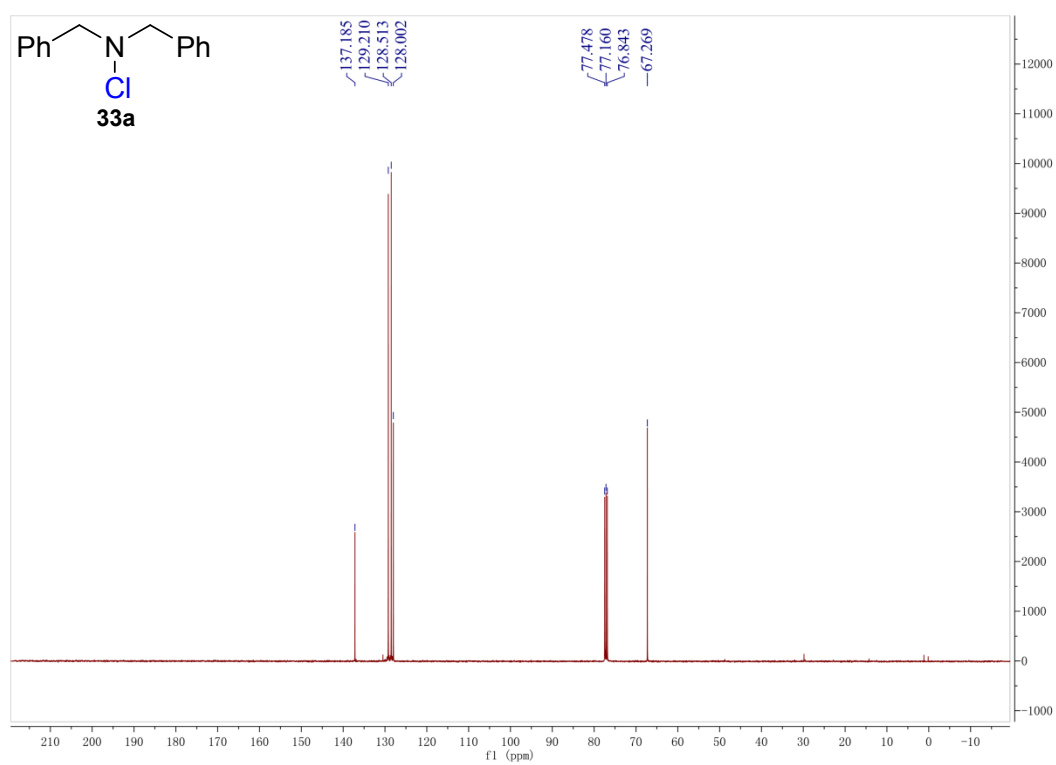
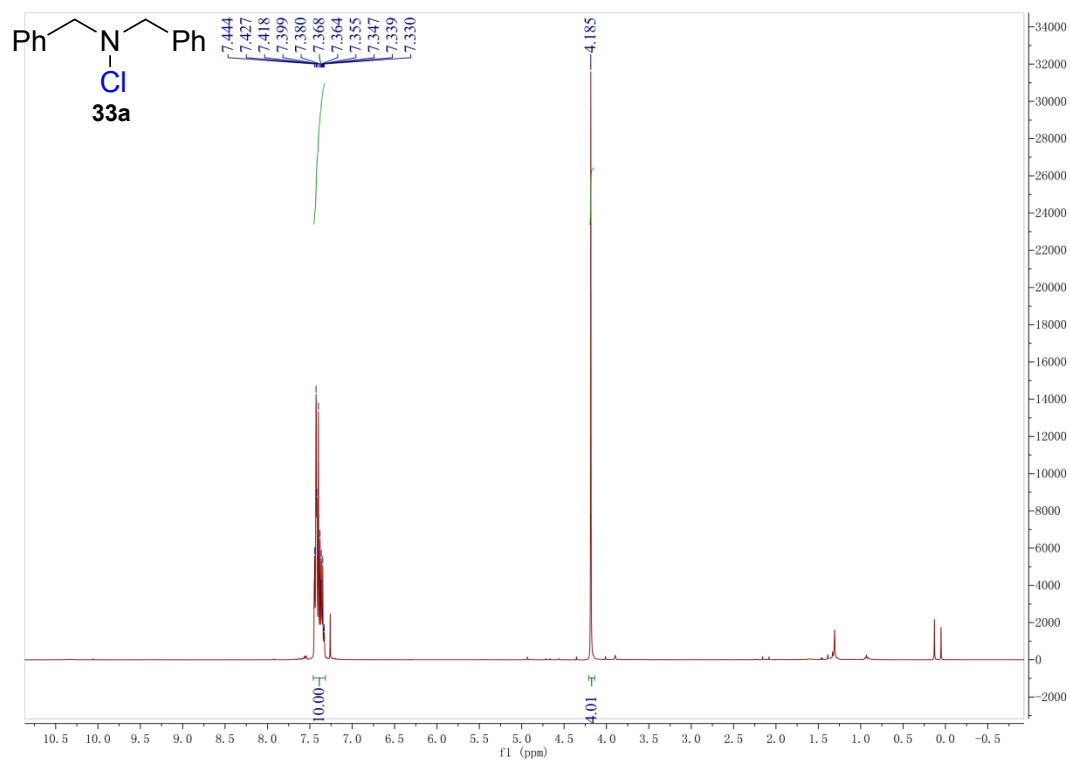


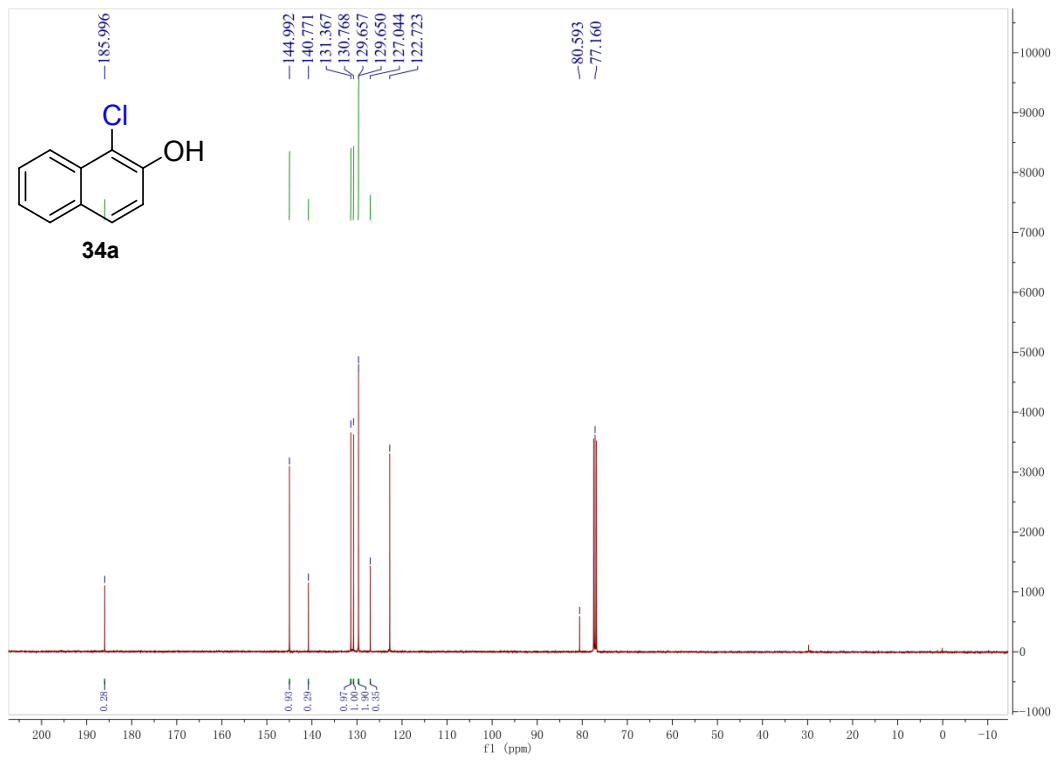
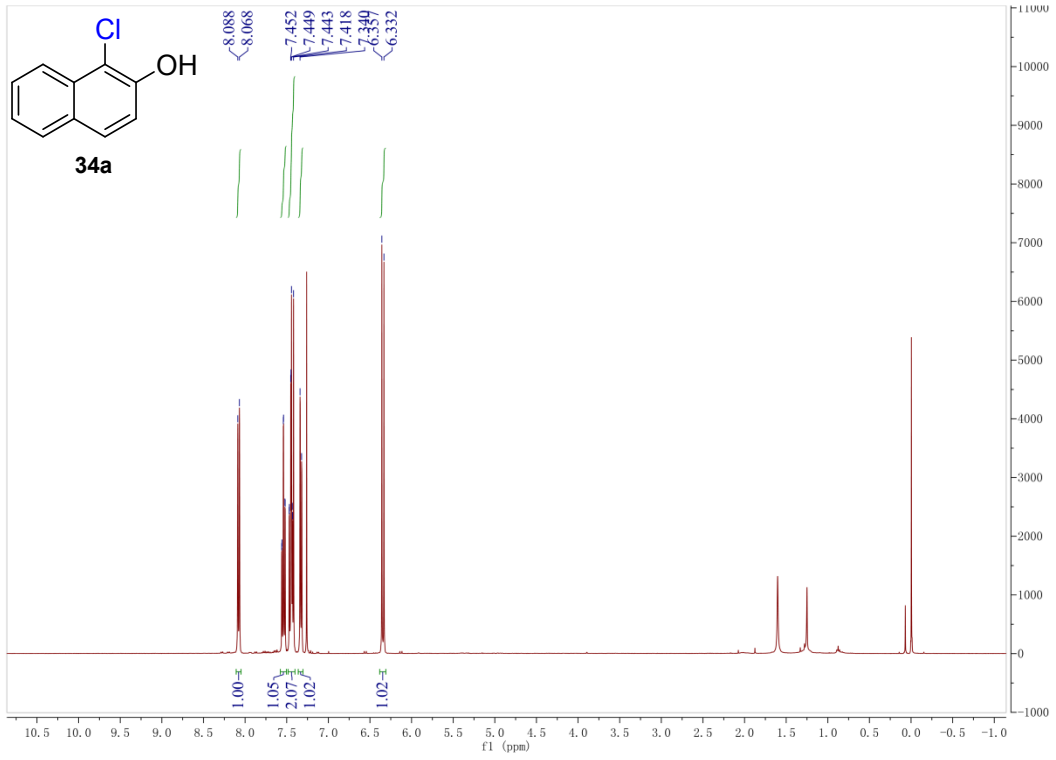


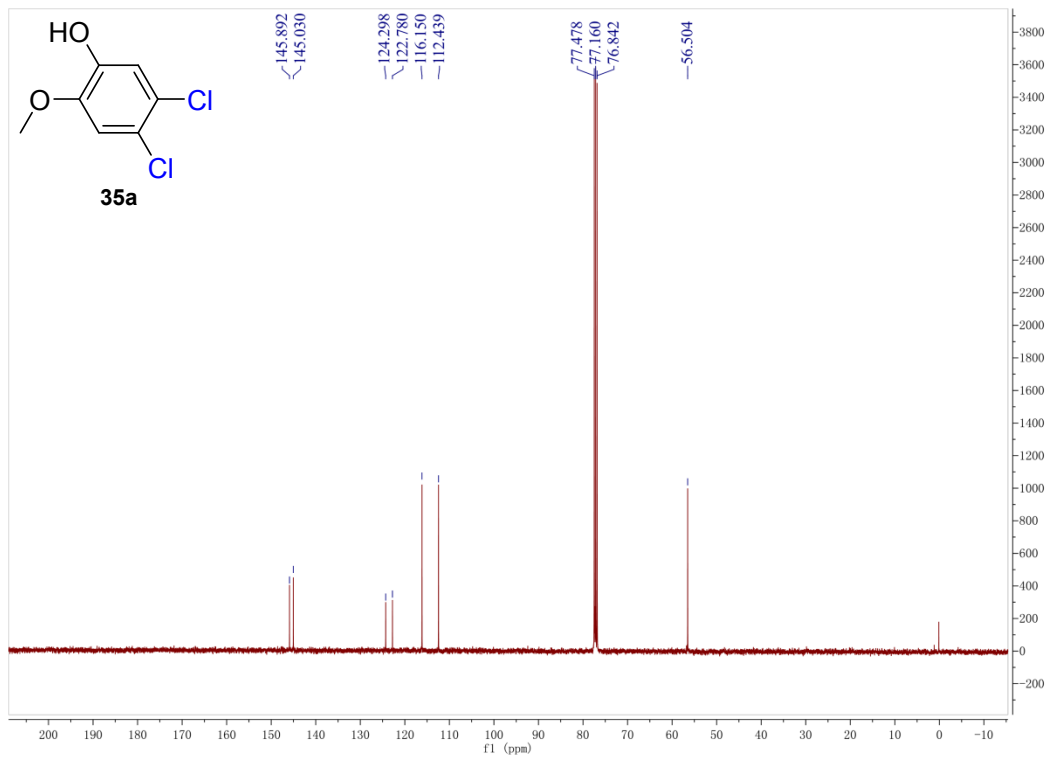
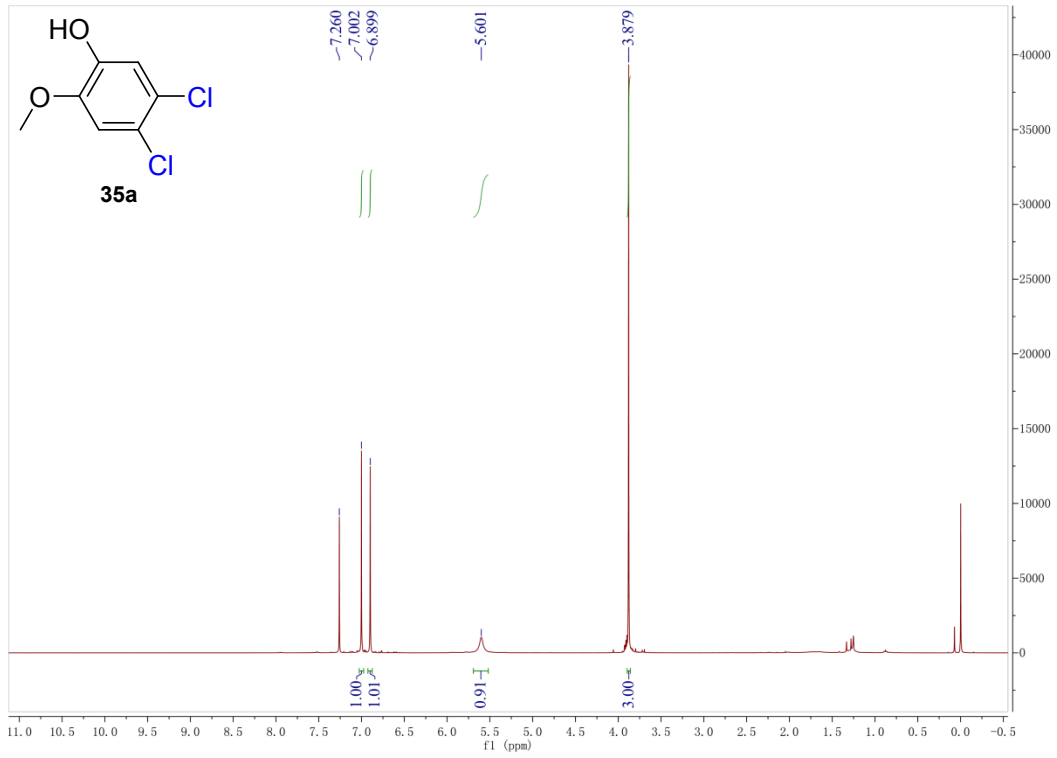






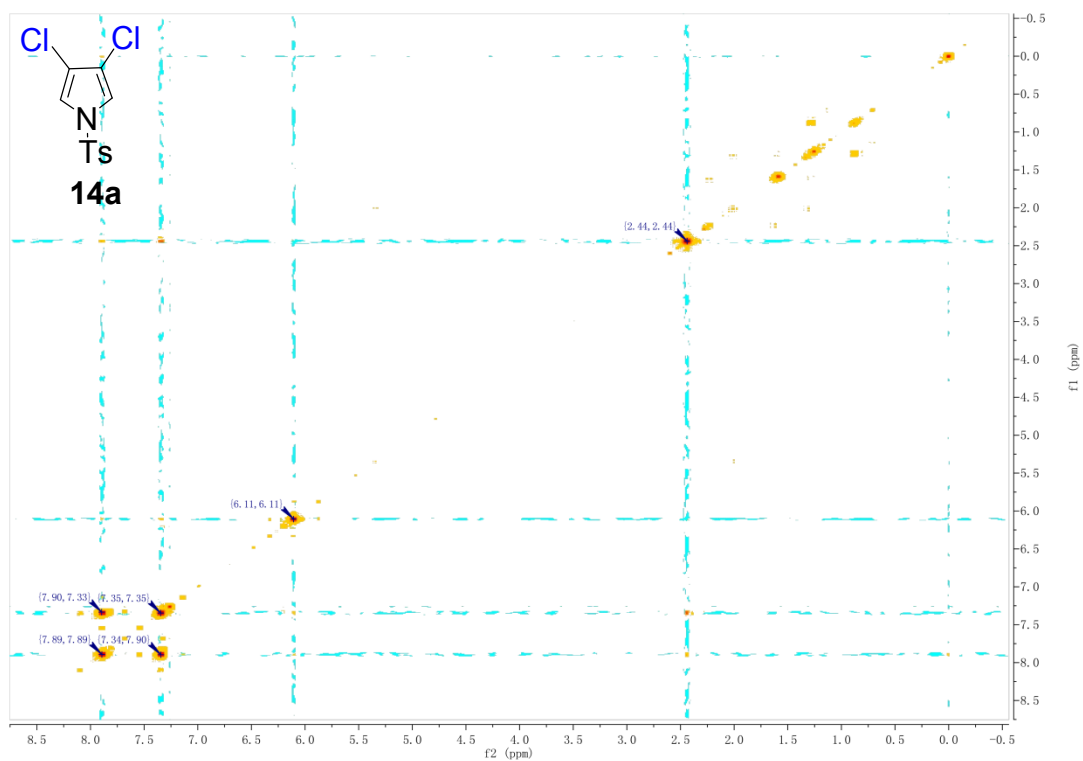






(Homonuclear chemical shift) Correlation Spectroscopy

^1H - ^1H COSY of 14a



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