Electronic Supplementary Information (ESI)

Higher-Order Human Telomeric G-Quadruplex DNA

Metalloenzyme Catalyzed Diels-Alder Reaction: An Unexpected

Inversion of Enantioselectivity Modulated by $K^{\scriptscriptstyle +}$ and $NH_4^{\scriptscriptstyle +}$ ions†

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Materials

All DNA oligodeoxynucleotides were purchased from Sangon (Shanghai, China). 3-(N-morpholino)propanesulfonic acid (MOPS) was purchased from Sangon (Shanghai, China). Cu(NO₃)₂·3H₂O (>99.5%), KCl (>99.5%) and NH₄Cl (>99.5%) were purchased from the Shanghai Chemical Reagent Company of the Chinese Medicine Group. Water was distilled and deionized (specific resistance of 18.2 M Ω at 25 °C) using a Milli-Q A10 water purification system. All experiments were carried out in 20 mM MOPS buffer (pH 6.5) unless otherwise stated. Other reagents and solvents were obtained from commercial sources and used without further purification. Dienophiles **1a-f** were prepared according to the literature.¹ All of the racemic products were prepared by the Diels-Alder reaction using Cu(NO₃)₂ as catalyst in methanol.



Methods

Circular dichroism (CD) spectra were recorded on a dual beam DSM 1000 CD spectrophotometer (Olis, Bogart, GA) with a 10 mm path-length quartz cell. Each measurement was recorded from 220 to 320 nm at 4 °C under N₂ purge. The scan rate was 0.5 nm per second. The average scan for each sample was subtracted by a background CD spectrum of corresponding buffer solution. The molarities are related to G-quadruplex unit. CD samples of all G-quadruplexes were prepared at a concentration (G4 unit) of 5 μ M by using a MOPS buffer solution (20 mM, pH = 6.5) consisting of different concentration of KCl or NH₄Cl as required.

The precise DNA strand concentrations were determined by measuring the UV absorbance of sample at 260 nm using the molar extinction coefficient values provided by the manufacturer. UV-Vis experiments were carried out on a Shimadzu 2450 spectrophotometer (Shimadzu, Japan) equipped with a Peltier temperature control accessory. All UV-Vis spectra were measured using a sealed quartz cell with a path length of 1.0 cm.

¹H NMR spectra were recorded on 400 MHz in CDCl₃ and ¹³C NMR spectra were recorded on 100 MHz in CDCl₃ using TMS or residual protic solvent signals as internal standard. Data for ¹H NMR are recorded as follows: chemical shift (δ , ppm), multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet or unresolved, coupling constant(s) in Hz, integration).

The enantioselectivity was determined by chiral HPLC analysis using Daicel chiralcel ODH, OJ-H column or chiralpak ADH, AD column with a UV-detector by using isopropanol and n-hexane as eluents at 25 °C.

General procedure



To a MOPS buffer (0.5 mL, 20 mM, pH 6.5) containing KCl (150 mM) or NH₄Cl (25 mM), an aqueous solution of HT45 (final G4 unit conc. of 50 μ M) was added. After stirred for a half hour at 4 °C, a solution of Cu(NO₃)₂ (final conc. of 10 μ M for K⁺ case and 16.7 μ M for NH₄⁺ case) was added. Then, aza-chalcone **1** in CH₃CN (final conc. of 1 mM for K⁺ case and 0.5 mM for NH₄⁺ case, 1 equiv.) was added. The reaction was initiated by the addition of freshly distilled cyclopentadiene **2** (100 equiv.). The mixture was stirred for 12 hours (**1a**), 48 hours (**1b-e**) and 72 hours (**1f**) at 4 °C, respectively. Followed by the extraction with diethyl ether (3 × 5 mL), the solvent was removed under reduced pressure. The residue was directly analyzed by ¹H-NMR and HPLC. The conversions were determined by ¹H-NMR and HPLC (only for **3a**) of the crude product.² The diastereoselectivity (*endo/exo*) and enantiomeric excess (*ee*) were determined by chiral HPLC.

HPLC analysis condition

Product **3a**: Daicel Chiralcel-ODH, *n*-hexane/*i*-PrOH 98:2, flow rate 0.5 mL/min, $\lambda = 212$ nm). Product **3b**: Daicel Chiralcel-ODH, *n*-hexane/*i*-PrOH 98:2, flow rate 0.5 mL/min, $\lambda = 212$ nm). Product **3c**: Daicel Chiralpak-ADH, *n*-hexane/*i*-PrOH 95:5, flow rate 1.0 mL/min, $\lambda = 212$ nm). Product **3d**: Daicel Chiralcel-OJH, *n*-hexane/*i*-PrOH 90:10, flow rate 0.5 mL/min, $\lambda = 212$ nm). Product **3e**: Daicel Chiralpak-AD, *n*-hexane/*i*-PrOH 90:10, flow rate 1.0 mL/min, $\lambda = 254$ nm). Product **3f**: Daicel Chiralpak-ADH, *n*-hexane/*i*-PrOH 90:10, flow rate 1.0 mL/min, $\lambda = 254$ nm).

Calculation the conversion of 1a

The procedure to determine the conversion of 1a by HPLC was according to the literature.²

Conversions of **1a** were calculated using the formula:

Conversion (%) = $A_{3a} / (A_{3a} + A_{1a} / f)$

Where A_{Ia} and A_{3a} are the HPLC peak areas of **1a** and **3a**, respectively. And *f* is the correction factor determined to be 0.73 from a calibration curve.

Kinetic measurements

All kinetic measurements were performed using UV-Vis spectroscopy (Shimadzu 2450) at 298 K by monitoring the disappearance of the absorption of **1a** at 326 nm.^{2b} Typical procedure is described as follows: HT45 (final G4 unit conc. 50 μ M) was added to MOPS (20 mM, pH 6.5) containing KCl (150 mM) or NH₄Cl (25 mM) in a quartz cuvette. After stirring for 15 min, Cu²⁺ ion (final conc. 10 μ M in K⁺ media or 16.7 μ M in NH₄⁺ media) was added. After stirring for another 15 min, 4 μ L of a fresh solution of **1a** in CH₃CN was added, resulting in a final concentration of 20 μ M. The determination was made after **2** (final conc. 4 mM) was added with the cuvette sealed tightly.

The following equations were used to calculate k_{app} and k_{rel} :

$$k_{app} = d[\mathbf{A}_{1\mathbf{a}}]/d\mathbf{t} \cdot (\mathbf{d} \cdot (\varepsilon_{1\mathbf{a}} - \varepsilon_{3\mathbf{a}}) \cdot [\mathbf{1a}]^0 \cdot [\mathbf{2}]^0)^{-1}$$
$$k_{rel} = k_{app-catalyst} / k_{app-no\ catalyst}$$

Where $d[A_{1a}]/dt$ is the slope of the absorption of 1a *vs*. time during the first 15% of the reaction, and d is the path length of the cuvette. ε_{1a} (22173 M⁻¹cm⁻¹) and ε_{3a} (177 M⁻¹cm⁻¹) are the molar extinction coefficient of 1a and 3a at 326 nm, respectively.

References

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- (a) N. S. Oltra and G. Roelfes, *Chem. Commun.* 2008, 6039; (b) C. H. Wang, G. Q. Jia, J. Zhou, Y. H. Li, Y. Liu, S. M. Lu and C. Li, *Angew. Chem. Int. Ed.* 2012, 51, 9352.

1H NMR spectra

(3-phenylbicyclo[2.2.1]hept-5-en-2-yl)(pyridin-2-yl)methanone (3a)



¹H NMR (400 MHz, CDCl₃): δ 8.67 (m, 1H), 8.00 (m, 1H), 7.81 (m, 1H), 7.49 – 7.39 (m, 1H), 7.36 – 7.23 (m, 4H), 7.17 (m, 1H), 6.49 (m, 1H), 5.82 (m, 1H), 4.53 (m, 1H), 3.54 (s, 1H), 3.45 (d, J = 3.9 Hz, 1H), 3.09 (s, 1H), 2.07 (d, J = 8.4 Hz, 1H), 1.62 (m, 1H).



pyridin-2-yl(3-p-tolylbicyclo[2.2.1]hept-5-en-2-yl)methanone (3b)



¹H NMR (400 MHz, CDCl₃, *endo* isomer): δ 8.67 (d, J = 4.6 Hz, 1H), 8.00 (d, J = 7.8 Hz, 1H), 7.81 (td, J = 7.7, 1.3 Hz, 1H), 7.44 (dd, J = 7.0, 5.3 Hz, 1H), 7.21 (d, J = 8.0 Hz, 2H), 7.08 (d, J = 7.9 Hz, 2H), 6.49 (dd, J = 5.4, 3.3 Hz, 1H), 5.81 (dd, J = 5.6, 2.7 Hz, 1H), 4.52 (dd, J = 4.9, 3.7 Hz, 1H), 3.53 (s, 1H), 3.41 (d, J = 5.0 Hz, 1H), 3.05 (s, 1H), 2.30 (s, 3H), 2.06 (d, J = 8.4 Hz, 1H), 1.59 (dd, J = 8.5, 1.1 Hz, 1H).



(3-(4-methoxyphenyl)bicyclo[2.2.1]hept-5-en-2-yl)(pyridin-2-yl)methanone (3c)



¹H NMR (400 MHz, CDCl₃, *endo* isomer): δ 8.74 – 8.60 (m, 1H), 8.00 (d, *J* = 7.9 Hz, 1H), 7.81 (td, *J* = 7.7, 1.7 Hz, 1H), 7.44 (ddd, *J* = 7.5, 4.8, 1.1 Hz, 1H), 7.28 – 7.18 (m, 2H), 6.86 – 6.78 (m, 2H), 6.48 (dd, *J* = 5.5, 3.2 Hz, 1H), 5.81 (dd, *J* = 5.6, 2.7 Hz, 1H), 4.49 (dd, *J* = 5.1, 3.5 Hz, 1H), 3.77 (s, 3H), 3.53 (s, 1H), 3.39 (d, *J* = 4.5 Hz, 1H), 3.02 (d, *J* = 1.2 Hz, 1H), 2.06 (d, *J* = 8.4 Hz, 1H), 1.60 (dd, *J* = 8.5, 1.6 Hz, 1H).



(3-(4-chlorophenyl)bicyclo[2.2.1]hept-5-en-2-yl)(pyridin-2-yl)methanone (3d)



¹H NMR (400 MHz, CDCl₃, *endo* isomer): δ 8.67 (d, J = 4.6 Hz, 1H), 8.00 (d, J = 7.8 Hz, 1H), 7.83 (dd, J = 8.5, 6.9 Hz, 1H), 7.46 (dd, J = 7.4, 4.9 Hz, 1H), 7.25 – 7.10 (m, 4H), 6.48 (dd, J = 5.4, 3.2 Hz, 1H), 5.83 (dd, J = 5.5, 2.7 Hz, 1H), 4.46 (dd, J = 5.1, 3.5 Hz, 1H), 3.54 (s, 1H), 3.40 (d, J = 5.1 Hz, 1H), 3.05 (s, 1H), 2.01 (d, J = 8.5 Hz, 1H), 1.62 (d, J = 8.6 Hz, 1H).



(3-(4-nitrophenyl)bicyclo[2.2.1]hept-5-en-2-yl)(pyridin-2-yl)methanone (3e)



¹H NMR (400 MHz, *endo* isomer): δ 8.66 (d, *J* = 4.5 Hz, 1H), 8.07 (m, 3H), 7.84 (t, *J* = 7.6 Hz, 1H), 7.47 (m, 3H), 6.52 – 6.44 (m, 1H), 5.88 (dd, *J* = 5.3, 2.5 Hz, 1H), 4.51 – 4.44 (m, 1H), 3.66 – 3.50 (m, 2H), 3.13 (s, 1H), 2.00 (d, *J* = 8.6 Hz, 1H), 1.67 (d, *J* = 8.4 Hz, 1H).



(3-(naphthalen-1-yl)bicyclo[2.2.1]hept-5-en-2-yl)(pyridin-2-yl)methanone (3f)



¹H NMR (400 MHz, CDCl₃): δ 8.70 – 8.57 (m, 1H), 8.18 – 7.98 (m, 2H), 7.87 – 7.76 (m, 2H), 7.70 (d, *J* = 8.2 Hz, 1H), 7.62 (d, *J* = 7.2 Hz, 1H), 7.49 – 7.37 (m, 4H), 6.68 – 6.57 (m, 1H), 5.97 – 5.89 (m, 1H), 4.74 – 4.65 (m, 1H), 4.07 (d, *J* = 4.8 Hz, 1H), 3.53 (s, 1H), 3.21 (s, 1H), 2.19 (dd, *J* = 8.4 Hz, 1H), 1.69 (dd, *J* = 8.3 Hz, 1H).



HPLC traces

1. Product 3a

Ee's were determined by HPLC analysis (Daicel Chiralcel-ODH, n-hexane/i-PrOH 98:2, flow rate

- 0.5 mL/min, $\lambda = 212$ nm).
- (1) Racemic 3a

Retention times: 13.0, 14.7 (exo isomer) and 17.0, 22.0 (endo isomer) mins



(2) Product 3a from the Diels-Alder reaction catalyzed by HT45-K-Cu

Retention times: 12.9, 14.4 (exo isomer) and 16.6, 21.3 (endo isomer) mins (92% ee)



(3) Product **3a** from the Diels-Alder reaction catalyzed by HT45-NH₄-Cu

Retention times: 12.8, 14.3 (exo isomer) and 16.0, 20.8 (endo isomer) mins (-73% ee)



2. Product 3b

Ee's were determined by HPLC analysis (Daicel Chiralcel-ODH, n-hexane/i-PrOH 98:2, flow rate

 $0.5 \text{ mL/min}, \lambda = 212 \text{ nm}).$

(1) Racemic 3b

Retention times: 11.0, 12.6 (exo isomer) and 13.4, 18.1 (endo isomer) mins



(2) Product 3b from the Diels-Alder reaction catalyzed by HT45-K-Cu

VWD1 A, Wavelength=254 nm (LYH131030\DEF_LC 2013-10-30 09-52-22\001-0101.D) mAU – 18.757 140 -120 -100 -14.045 80 -60 -12.878 40 -11.204 20 -0 -10 20 15 mir _____ Area Percent Report _____ _____ Sorted By : Signal 1.0000 Multiplier: : 1.0000 Dilution: • Use Multiplier & Dilution Factor with ISTDs Signal 1: VWD1 A, Wavelength=254 nm Area Peak RetTime Type Width Height Area [mAU*s] [mAU] # [min] [min] 8 0.2469 120.00939 7.14752 0.2698 217.48340 12.08158 0.2117 1488.25195 71.81748 7.181748 ---- | ----- | ----- | -----1 11.204 VV 1.9482 2 12.878 VV 3.5305 3 14.045 VB 24.1596 4 18.757 BB 0.4035 4334.34521 163.78825 70.3617 6160.08996 254.83484 Totals : _____ *** End of Report ***

Retention times: 11.2, 12.9 (exo isomer) and 14.0, 18.8 (endo isomer) mins (50% ee)

(3) Product **3b** from the Diels-Alder reaction catalyzed by $HT45-NH_4$ -Cu

VWD1 A, Wavelength=254 nm (LYH131030\DEF_LC 2013-10-30 09-52-22\002-0201.D) mAU 긬 786 175 -150 125 100 18.610 75 -50 -11.123 12.761 25 0 12.5 20 2.5 7.5 10 15 17.5 22.5 min _____ _____ Area Percent Report Sorted By Signal : Multiplier: 1.0000 : Dilution: 1.0000 : Use Multiplier & Dilution Factor with ISTDs Signal 1: VWD1 A, Wavelength=254 nm Peak RetTime Type Width Area Height Area # [min] [min] [mAU*s] [mAU] 양 0.2276 129.98175 1 11.123 VV 8.51987 2.4360 2 12.761 VV 1.7545 77.0059 0.2549 93.61814 5.59692 0.3082 4108.95508 199.96890 3 13.786 VB 4 18.610 BB 0.3968 1003.33954 38.57032 18.8036 Totals : 5335.89451 252.65601 _____ _____ *** End of Report ***

Retention times: 11.1, 12.8 (exo isomer) and 13.8, 18.6 (endo isomer) mins (-61% ee)

3. Product 3c

Ee's were determined by HPLC analysis (Daicel Chiralpak-ADH, *n*-hexane/*i*-PrOH 95:5, flow rate 1.0 mL/min, $\lambda = 212$ nm).

(1) Racemic 3c

Retention times: 13.8, 14.2 (exo isomer) and 16.6, 22.8 (endo isomer) mins



(2) Product 3c from the Diels-Alder reaction catalyzed by HT45-K-Cu

Retention times: 14.0, 14.4 (exo isomer) and 16.8, 23.4 (endo isomer) mins (52% ee)



(3) Product **3c** from the Diels-Alder reaction catalyzed by HT45-NH₄-Cu

Retention times: 13.9, 14.2 (exo isomer) and 16.6, 22.9 (endo isomer) mins (-90% ee)



4. Product **3d**

Ee's were determined by HPLC analysis (Daicel Chiralcel-OJH, *n*-hexane/*i*-PrOH 90:10, flow rate 0.5 mL/min, λ = 212 nm).

(1) Racemic 3d

Retention times: 20.8, 26.9 (exo isomer) and 25.4, 44.4 (endo isomer) mins



(2) Product 3d from the Diels-Alder reaction catalyzed by HT45-K-Cu

Retention times: 20.4, 26.4 (exo isomer) and 24.5, 43.4 (endo isomer) mins (60% ee)



(3) Product **3d** from the Diels-Alder reaction catalyzed by HT45-NH₄-Cu

Retention times: 20.3, 26.3 (exo isomer) and 24.6, 43.5 (endo isomer) mins (-90% ee)



5. Product **3e**

Ee's were determined by HPLC analysis (Daicel Chiralpak-AD, *n*-hexane/*i*-PrOH 90:10, flow rate 1.0 mL/min, λ = 254 nm).

(1) Racemic 3e

Retention times: 12.5, 16.0 (exo isomer) and 14.6, 18.1 (endo isomer) mins



(2) Product 3e from the Diels-Alder reaction catalyzed by HT45-K-Cu

Retention times: 12.5, 16.0 (exo isomer) and 14.6, 18.2 (endo isomer) mins (26% ee)



(3) Product **3e** from the Diels-Alder reaction catalyzed by HT45-NH₄-Cu

Retention times: 12.5, 15.9 (exo isomer) and 14.5, 18.1 (endo isomer) mins (-90% ee)



6. Product **3f**

Ee's were determined by HPLC analysis (Daicel Chiralpak-ADH, n-hexane/i-PrOH 90:10, flow rate

1.0 mL/min, λ = 254 nm).

(1) Racemic 3f

Retention times: 7.4 and 10.8 mins



(2) Product 3f from the Diels-Alder reaction catalyzed by HT45-K-Cu

Retention times: 7.4 (*major*) and 10.7 (*minor*) mins (91% *ee*)



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(3) Product **3f** from the Diels-Alder reaction catalyzed by HT45-NH₄-Cu

Retention times: 7.4 (minor) and 10.6 (major) mins (-82% ee)

