

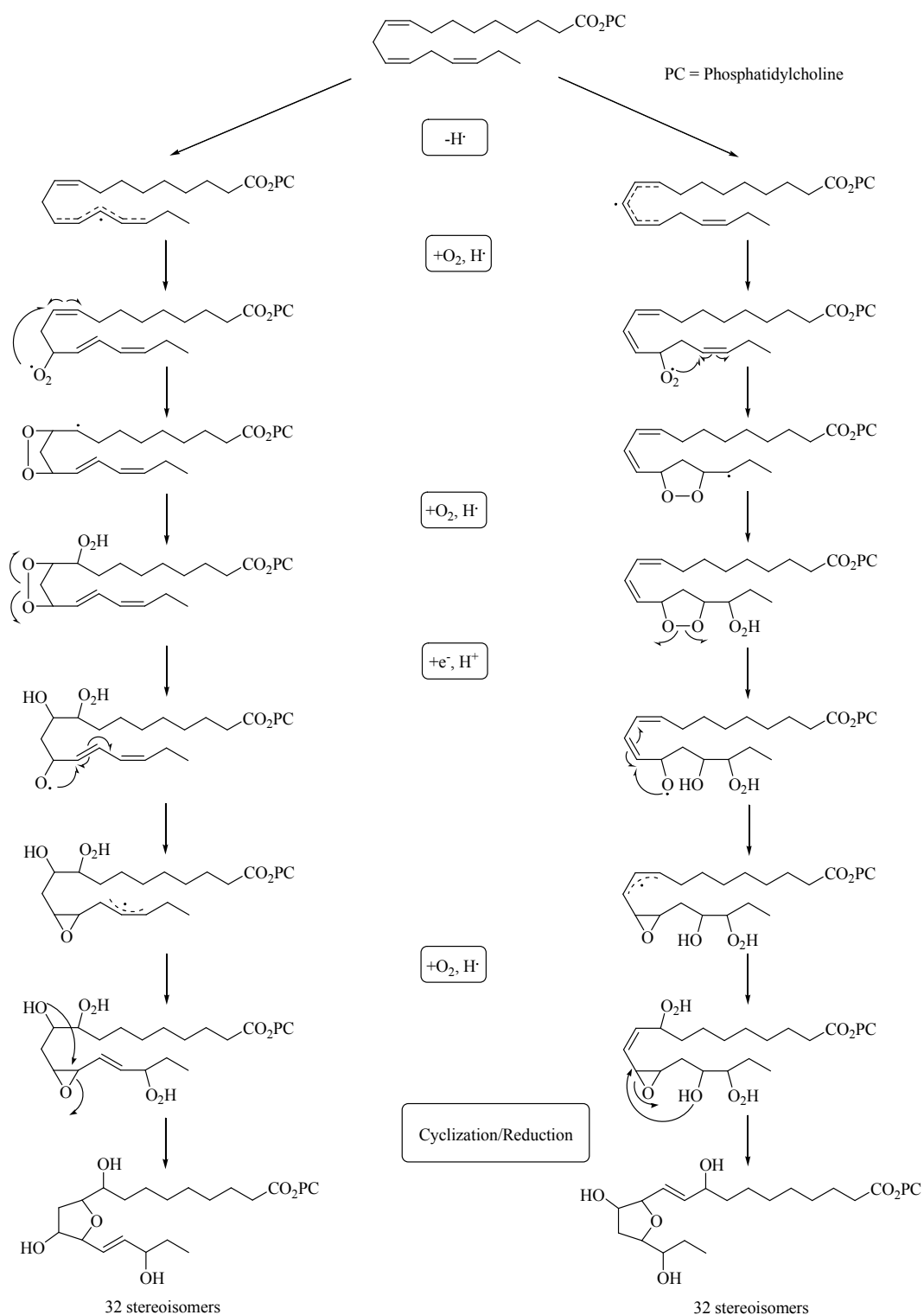
Supporting Information
**Synthesis and Discovery of Phytofurans: Metabolites of α -
Linolenic Acid Peroxidation**

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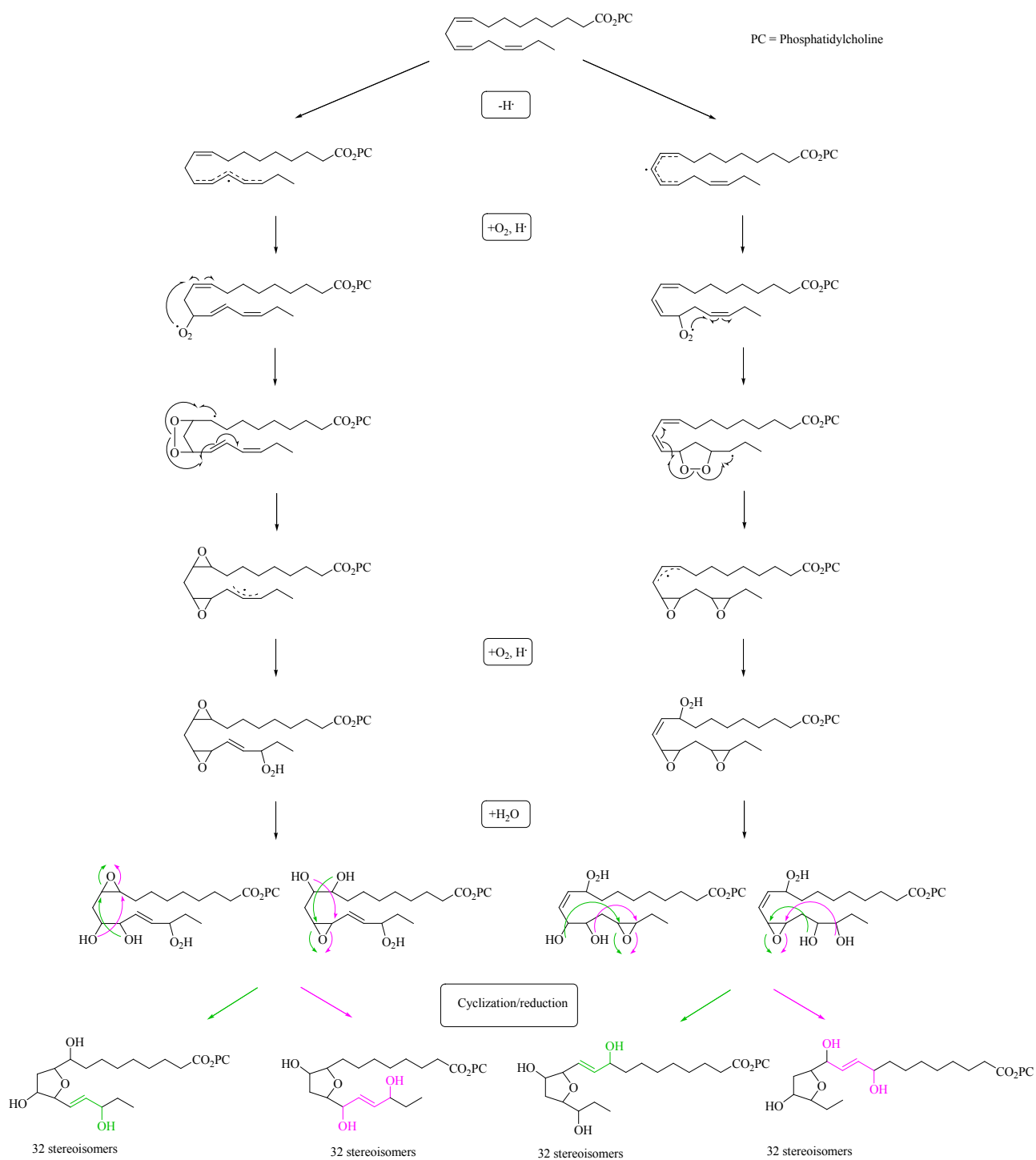
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1. The biosynthesis of Phytofurans (PhytoFs) based on the Isofuran pathways by Fessel and coworkers,¹ and Jahn and co-workers².



Scheme 1: Biosynthesis mechanism of alkenyl PhytoFs by reductive endoperoxide cleavage; 3-exo cyclization, and epoxide ring opening



Scheme 2: Alternative mechanism based on 1,3-S_Hi reaction, 3-exo cyclization, ring epoxide opening

2. General Information

All reactions requiring anhydrous conditions were conducted in oven-dried (120 °C) glassware with magnetic stirring under an atmosphere of nitrogen, unless mentioned otherwise. Syringes and needles for the transfer of reagents were dried at 120 °C and allowed to cool in a desiccators with CaCl₂ before use. Anhydrous THF and DCM were obtained from the Innovative Technology PS-Micro solvent purification system. Other solvents and reagents were used as obtained from the supplier unless otherwise noted. The reactions were monitored by TLC, using plates pre-coated Silica Gel 60 (Merck). Visualization of reaction components was achieved with 254 nm, and treatment with acidic *p*-anisaldehyde stain, followed by gentle heating. Organic layers were dried using MgSO₄ unless otherwise stated. Column chromatography was carried out on silica gel Kieselgel 60 (40-63 μm). Optical rotations ($[\alpha]_D^{20}$) were recorded on Perkin Elmer Polarimeter 341 ($\lambda = 589$ nm, 20 °C, concentration *c* in mg/mL, CHCl₃ or MeOH). Infrared spectra were obtained using a Perkin-Elmer Spectrum One spectrophotometer. They were reported as wavenumber (cm⁻¹) of significant peaks. Mass spectra were obtained by positive or negative electrospray ionization and electronic impact methods. Unless otherwise stated, ¹H NMR and ¹³C NMR spectra were recorded at 303 K with 300 or 500 MHz on Bruker spectrometers. For the ¹H NMR, the peak due to residual CHCl₃ was used as the internal reference (fixed at 7.26 ppm) or for MeOD: the peak due to residual MeOH was used as the internal reference (fixed at 3.31 ppm) or for D₂O: the peak due to residual H₂O was used as the internal reference (fixed at 4.79 ppm). The ¹H NMR spectra are reported as follows: chemical shift in parts per million (multiplicity, coupling constant(s) *J* (Hz), relative integral, assignment) where multiplicity is defined as: br = broad, m = multiplet, s = singlet, d = doublet, t = triplet, or combination thereof). Selected ¹³C NMR spectra were conducted using a *J* modulated sequence and in CDCl₃ the central peak of the CDCl₃ triplet was used as an internal reference (77.16 ppm), MeOD the central peak of the MeOD multiplet was used as an internal reference (49.00 ppm). The assignments of NMR spectra were assisted by homonuclear (¹H-¹H) and heteronuclear (¹H-¹³C) correlation spectroscopy (COSY45, HMQC, HMBC) and are reported as follows: CH₃, CH₂, CH, and Cq (for quaternary carbon atoms).

3. Experimental procedures

3.1 Chemical synthesis

Preparation of hepta-2,5-diyne-1,7-diol **4**:

To a solution of but-2-yne-1,4-diol (86 g, 1 mol, 1 eq) in 100 mL of dry benzene was added dry pyridine (87 mL, 1 mol, 1 eq). Dry thionyl chloride (80 mL, 1 mol, 1 eq) was added dropwise using syringe pump (over 3h). The temperature was maintained between 10-20 °C with cold bath. The mixture was stirred overnight at rt. The reaction was quenched with dropwise addition by syringe pump of 135 mL of water. After separation, the aqueous layer was extracted with ether (3 x 100 mL). The ether extracts were combined with the original benzene layer. The combined organic layers were stirred with saturated NaHCO₃ (150 mL) and solid NaHCO₃ (pH=8) for 20 min. After separation, the organic layers were dried over MgSO₄, filtered and the solvent was removed under vacuum. The product was purified by distillation under vacuum to give 4-chlorobut-2-yne-1-ol (48.50 g, 46%).

R_f = 0.69 (Cyclohexane/Et₂O: 3/7)

¹H NMR (300 MHz, CDCl₃) δ 4.31 (s, 2H, CH₂OH), 4.17 (s, 2H, CH₂Cl), 2.06 (br, 1H, OH)

¹³C NMR (75 MHz, CDCl₃) δ 84.8 (1C, CH), 80.6 (1C, CH), 51.0 (1C, CH₂OH), 30.4 (1C, CH₂Cl)

EI: 69.2 [M-Cl], 87.2 [M-OH]

IR (ν_{max} cm⁻¹): 3324, 1430, 1263, 1142, 1009, 691

To a suspension of dry K₂CO₃ (17.82 g, 0.13 mol, 1.5 eq) in 60 mL of dry DMF, were successively added, dry NaI (25.8 g, 0.17 mol, 2 eq), dry CuI (32.7 g, 0.17 mol, 2 eq) and prop-2-yne-1-ol (7.02 mL, 0.12 mol, 1.4 eq). The mixture was stirred at rt for 1h. A solution of 4-chlorobut-2-yne-1-ol (9 g, 0.09 mol, 1 eq) in 40 mL of dry DMF was added by cannulation. The mixture was stirred overnight at rt. The reaction was quenched with saturated NH₄Cl (180 mL) and the pH was adjusted to 10 with NH_{3(aq)}. The product was extracted with EtOAc (9 x 150 mL). The combined organic layers were dried over MgSO₄, filtered and the solvent was evaporated under vacuum. The crude product was purified on silica gel (pentane/EtOAc : 50/50 to 0/100) to give hepta-2,5-diyne-1,7-diol **4** as a yellow solid (8.6 g, 81%).

R_f = 0.46 (Cyclohexane/EtOAc: 1/7)

¹H NMR (300 MHz, MeOD) δ 4.20-4.10 (m, 4H, CH₂OH), 3.30-3.20 (m, 2H, CH₂)

¹³C NMR (75 MHz, MeOD) δ 79.9 (2C, Cq), 79.8 (2C, Cq), 50.8 (2C, CH₂OH), 9.9 (1C, CH₂)

EI: 106.2 [M-H₂O]

IR (ν_{max} cm⁻¹): 3245, 2918, 1362, 1312, 1145, 1011

Mp: 84.6-85.7 °C

Preparation of (2S,3S,5S,6S)-2,3:5,6-diepoxyheptane-1,7-diol **3**:

To a solution of LiAlH_4 (100 mL, 2.4 M/THF, 240 mmol, 4.9 eq) in 100 mL of THF at $-5\text{ }^\circ\text{C}$, 2-methoxyethanol (32 mL, 406 mmol, 8.4 eq) was added drop wise, using syringe pump over 1 h. After cooling to $-40\text{ }^\circ\text{C}$ a solution of the diynediol **4** (6 g, 48.4 mmol, 1 eq) in 45 mL of THF was added dropwise, using syringe pump, over 1h. The mixture was stirred at $0\text{ }^\circ\text{C}$ for 1h and at rt for 15 h. After the installation of mechanic stirrer, the mixture was cooled at $0\text{ }^\circ\text{C}$ and hydrolyzed by a slow addition (1.5h) of 30 mL of water. The pasty mixture was filtered on fritted glass and the residue was washed with ethanol (3 x 500 mL). The filtrates were concentrated under vacuum. A flash column chromatography of the residue, eluting with pentane/ Et_2O (50/50 to 0/100), gave the (*E,E*)-hepta-2,5-diene-1,7-diol (3.44 g, 55%).

$R_f = 0.19$ (Et_2O :100%)

$^1\text{H NMR}$ (300 MHz, MeOD) δ 5.80-5.50 (m, 4H, CH), 4.01 (d, $J = 4.6$, 4H, CH_2OH), 2.79 (t, $J = 5.1$, 2H, CH_2)

$^{13}\text{C NMR}$ (75 MHz, MeOD) δ 131.3 (2C, CH), 130.7 (2C, CH), 63.4 (2C, CH_2OH), 35.7 (1C, CH_2)

EI: 81.2 [$\text{M}-2\text{H}_2\text{O}$]

IR ($\nu_{\text{max}}\text{ cm}^{-1}$): 3293, 2864, 1667, 1425, 1089, 967

To a suspension of powdered molecular sieves (4Å, 1.5g) in 30 mL of dry DCM and 10 mL of dry CHCl_3 at $-10\text{ }^\circ\text{C}$, distilled (2*R*,3*R*)-(+)-diethyl tartrate (221 μL , 1.29 mmol, 0.15 eq), distilled titanium tetraisopropoxide (253 μL , 0.86 mmol, 0.1 eq) and a 5 M solution of *tert*-butyl hydroperoxide in decane (5.14 mL, 25.7 mmol, 3 eq) were successively added. After stirring for 30 min at $-10\text{ }^\circ\text{C}$, the mixture was cooled to $-30\text{ }^\circ\text{C}$. A solution of the (*E,E*)-heptadienediol (1.10 g, 8.59 mmol, 1 eq) in 10 mL of CHCl_3 was added. The mixture was stirred for 1 h at $-30\text{ }^\circ\text{C}$ and stored in a deep freeze for 48 h at $-20\text{ }^\circ\text{C}$. A solution of citric acid monohydrate (192 mg, 0.91 mmol) in 3 mL of acetone and 20 mL of diethyl ether was added at this temperature. After reaching rt, the mixture was filtered and the residue was washed with 30 mL of THF and 80 mL of CH_3CN . The filtrate was concentrated in vacuo. The residue was washed with 80 mL of water to extract the product. The solution was concentrated by freeze drying to give (2*S*,3*S*,5*S*,6*S*)-2,3:5,6-diepoxyheptane-1,7-diol **3** (1.05 g, 76%, ee 90% determined below) as a white powder.

$R_f = 0.23$ (DCM/MeOH: 9/1)

$^1\text{H NMR}$ (500 MHz, MeOD) δ 3.76 (dd, $J = 12.5, 3.1$, 2H, CH_2OH), 3.53 (dd, $J = 12.5, 5.1$, 2H, CH_2OH), 3.03(td, $J = 5.7, 2.2$, 2H, CHO), 2.95-2.90 (m, 2H, CHO), 1.80(t, $J = 5.7$, 2H, CH_2)

$^{13}\text{C NMR}$ (155 MHz, MeOD) δ 62.8 (2C, CH_2OH), 59.6 (2C, CH), 54.0 (2C, CH), 35.4 (1C, CH_2)

ES+ : 183.1 [$\text{M}+\text{Na}$] $^+$, 343.1 [$\text{M}+\text{M}+\text{Na}$] $^+$

HRMS (ESI+): calculated for $\text{C}_7\text{H}_{12}\text{O}_4\text{Na}$ [$\text{M}+\text{Na}$] $^+$ 183.0633, found 183.0633

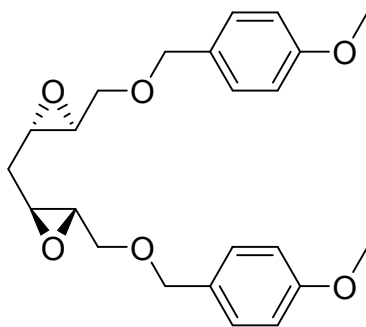
IR ($\nu_{\text{max}}\text{ cm}^{-1}$): 3120, 2873, 2338, 2169, 2028, 1479, 1334, 1254, 1018, 983, 956, 901, 875, 854, 715

$M_p = 153.1\text{-}154.6\text{ }^\circ\text{C}$

$[\alpha]_D^{20}(\text{H}_2\text{O}) = -48.1$ (c 4.33); lit: 3 $[\alpha]_D^{20}(\text{H}_2\text{O}) = -49.8$ (c 0.64).

Determination of the enantiomeric excess

a) Preparation of bis(4-methoxybenzyl) derivative



To a solution bisepoxy diol **3** (50 mg, 0.31 mmol, 1 eq) in 3 mL of DMF, 4-methoxybenzyl chloride (93 μL , 0.69 mmol, 2.2 eq) and NaH (60% in grease, 30 mg, 0.75 mmol, 2.4 eq) were added. After 20 min at 100 $^{\circ}\text{C}$, the reaction quenched with water (10 mL) and extracted with Et_2O (3 x 6 mL). The organics layers were washed with brine (10 mL), dried over MgSO_4 , filtered and concentrated under vacuum. Fast column chromatography (pentane/ EtOAc : 60/40) afforded bis(4-methoxybenzyl) derivative (34 mg, 30%).

$R_f = 0.41$ (Cyclohexane/ EtOAc : 9/1)

$^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.28-7.23 (m, 4H, CH_{Ph}), 6.87 (d, $J = 7.5$, 4H, CH_{Ph}), 4.49 (q_{app}, $J = 11.6$, 4H, CH_2O), 3.8 (s, 6H, CH_3O), 3.70 (d, $J = 11.3$, 2H, CH_2O), 3.47 (dd, $J = 5.2, 11.5$, 2H, CH_2O), 3.05-2.90 (m, 4H, CHO), 1.80 (t, $J = 5.5$, 2H, CH_2)

$^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 159.4 (2C, Cq), 130.0 (2C, Cq), 129.6 (4C, CH), 113.9 (4C, CH), 73.1 (2C, CH_2O), 69.8 (2C, CH_2O), 56.9 (2C, CHO), 55.4 (2C, CH_3O), 53.0 (2C, CHO), 34.6 (1C, CH_2)

ES+ : 401.20 $[\text{M}+\text{H}]^+$, 423.18 $[\text{M}+\text{Na}]^+$

HRMS (ESI+): calculated for $\text{C}_{23}\text{H}_{29}\text{O}_6$ $[\text{M}+\text{H}]^+$ 401.1964, found 401.1967

b) Preparation of the racemic reference

Racemic bis(4-methoxybenzyl) derivative was prepared following the above procedure from a 1:1 mixture of (d,l)-racemic and meso 2,3:5,6-diepoxyheptane-1,7-diol prepared from corresponding (*E,E*)-hepta-2,5-diene-1,7-diol and mCPBA.

c) HPLC analysis of bis(4-methoxybenzyl) derivative for ee determination

HPLC Perkin Helmer series 200

Chiral column Chiralcel OD 0.46 cm x 25 cm

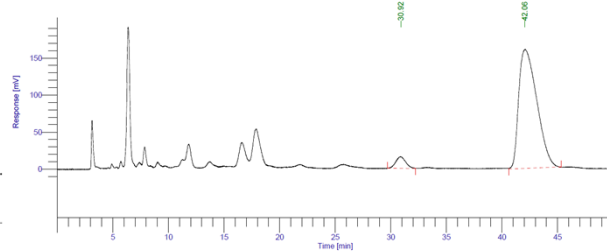
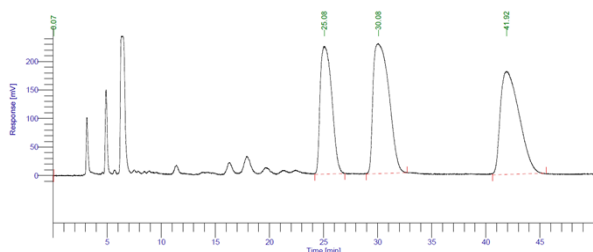
Diode Array Detector ($\lambda=210$ nm)

Flow: 1.0 mL/min;

Eluent: Hexane/*i*-PrOH: 85/15

Bis(4-methoxybenzyl) derivative from (2R,3R,5R,6R)-2,3:5,6-diepoxyheptane-1,7-diol $t_r = 30.92$ min.

Bis(4-methoxybenzyl) derivative from (2S,3S,5S,6S)-2,3:5,6-diepoxyheptane-1,7-diol $t_r = 41.92$ min.



Racemic/Meso-bis(4-methoxybenzyl) derivative bisepoxy diol

Peak #	Component Name	Time [min]	Height [uV]	Area [%]	Norm. Area [%]	BL	RT relatif	Rel. RT
1		0.068	586.39	0.00	0.00	*BB	1.00	1.00 0.07
2		25.080	224191.96	27.04	27.04	*MM	367.93	367.93 25.08
3		30.081	227950.22	37.58	37.58	*MM	441.30	441.30 30.08
4		41.917	180595.13	35.38	35.38	*MM	614.93	614.93 41.92
		633323.69	100.00	100.00				97.15

(2S,3S,5S,6S)- bis(4-methoxybenzyl) derivative bisepoxy diol

Peak #	Component Name	Time [min]	Height [uV]	Area [%]	Norm. Area [%]	BL	RT relatif	Rel. RT
1		30.917	15673.57	5.08	5.08	*MM	1.00	1.00 30.92
2		42.064	160608.67	94.92	94.92	*MM	1.36	1.36 42.06
			176282.24	100.00	100.00			72.98

Preparation of tetraol **2**:

To a solution of bisepoxy diol **3** (100 mg, 0.63 mmol, 1 eq) in 5 mL of water, a solution of KOH (175 mg, 3.12 mmol, 5 eq) in 5 mL of distilled water was added. The mixture was heated at 80 °C for 2 h. The mixture was neutralized by a solution of 1 M HCl. The solution was concentrated by freeze-drying. Column chromatography (DCM/MeOH: 95/5 to 80/20) afforded furanic tetraol **2** in mixture of two diastereoisomers (dr = 4/1; 72 mg, 65%) as a white solid.

$R_f = 0.2$ (DCM/MeOH: 8/2)

¹H NMR (500 MHz, D₂O) δ 4.20-4.12 (m, 1H, CHOH), 4.12-4.00 (m, 1H, CHO + *diast*, 0.4H, CHO), 3.76-3.80 (m, 1H, CHO + *diast*, 0.2H, CHO), 3.71-3.64 (m, 1H, CHOH + *diast*, 0.3H, CHOH), 3.60-3.50 (m, 2H, CH₂OH + *diast*, 0.2H, CHOH), 3.50-3.48 (*diast*, m, 0.4H, CH₂OH), 3.45-3.35 (m, 2H, CH₂OH + *diast*, 0.5H, CHOH), 2.25-2.10 (*diast*, m, 0.2H, CH₂), 2.00-1.90 (m, 1H, CH₂), 1.85-1.70 (m, 1H, CH₂ + *diast*, m, 0.3H, CH₂)

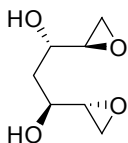
¹³C NMR (125 MHz, D₂O) δ 88.1 (1C, CH), 87.3 (*diast*, 1C, CH), 80.4 (1C, CH), 80.0 (*diast*, 1C, CH), 74.6 (*diast*, 1C, CH), 74.4 (1C, CH), 73.8 (1C, CH), 72.8 (*diast*, 1C, CH), 64.6 (*diast*, 1C, CH₂OH), 64.4 (1C, CH₂OH), 63.7 (1C, CH₂OH), 63.2 (*diast*, 1C, CH₂OH), 36.1 (1C, CH₂ + *diast*)

ES⁺ : 201.1 [M+Na]⁺

HRMS (ESI⁺): calculated for C₇H₁₄O₅Na [M+Na]⁺ 201.0739, found 201.0742

IR (ν_{\max} cm⁻¹): 3347, 2947, 2885, 1453, 1075, 1026, 925

Column chromatography of the above reaction also afforded bisepoxy diol **B** resulting of double-Payne rearrangement of **3**:



Bisepoxy diol **B** isolated (7 mg, 7%)

$R_f = 0.46$ (DCM/MeOH: 9/1)

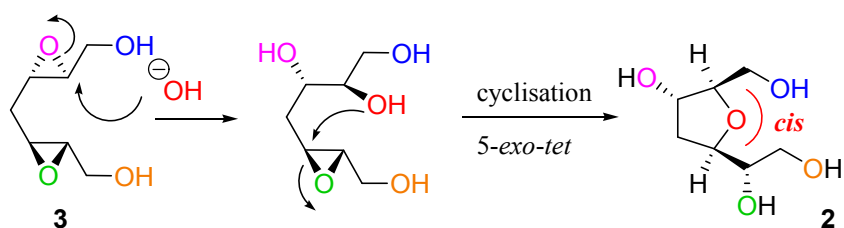
$^1\text{H NMR}$ (500 MHz, MeOD) δ 4.48 (s, 2H, CHO), 3.89 (t, $J = 6.8$, 2H, CHO), 3.78 (dd, $J = 10.9, 7.2$, 2H, CH_2O), 3.71 (dd, $J = 10.9, 6.2$, 2H, CH_2O), 2.12 (s, 2H, CH_2)

$^{13}\text{C NMR}$ (150 MHz, MeOD) δ 86.2 (2C, CHO), 78.0 (2C, CHO), 60.6 (2C, CH_2O), 39.0 (1C, CH_2)

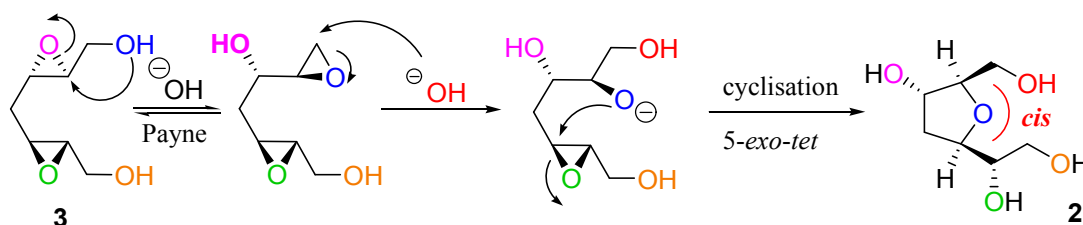
IR (ν_{max} cm^{-1}): 3347, 2947, 2885, 1649, 1453, 1210, 1075, 1026, 925

It is important to note that bisepoxy diol **B** in the same condition as above do not furnish any THF ring but only recovered SM and very polar compounds (probably by opening of the epoxides) already observed for the reaction of **3** to give **2**.

Justification of the Payne rearrangement reaction over a direct opening of the epoxide.



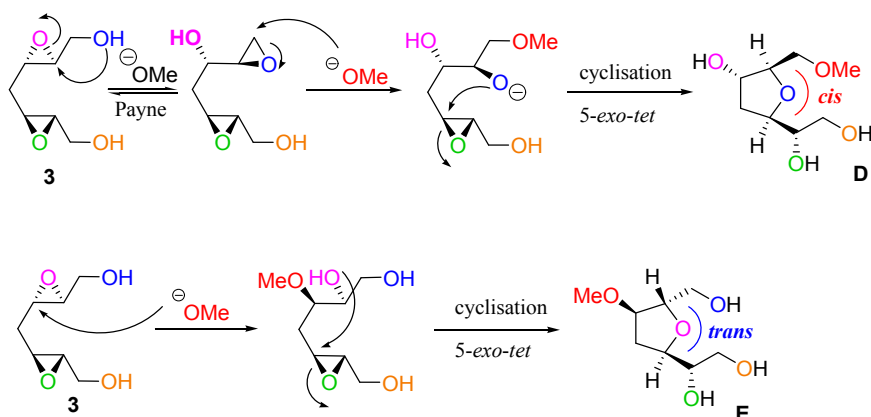
Direct opening of **3**



Payne reaction of **3** to give **2**

A direct opening of **3** on the C2 atom follow by cyclization will give the same stereoisomer **2** which can also be obtained via a mono Payne rearrangement/opening of epoxide and cyclization. The following experiment in MeONa/MeOH condition reported almost the same ratio of THF ring products which permitted to assign the position of the reactive MeO group similarly as the OH group should have reacted.

Preparation of methoxy-tetraol **D** and **E**.



The bisepoxy diol **3** (100 mg, 0.63 mmol, 1 eq) was dissolved in 4 mL of NaOMe freshly prepared (0.2M in MeOH). The mixture was heated at 80°C for 16 h. The solvent was evaporated and the crude product was purified by column chromatography (DCM/MeOH: 95/5 to 80/20) to afford a mixture of furanic methoxy cycle **D** and **E** in a 7/3 ratio (69 mg, 58%). Relative configurations of **D** and **E** were assigned based on the position of the MeO group following a Payne rearrangement for **D** and a direct opening of **3** for **E**.

$R_f = 0.4$ (DCM/MeOH: 8/2)

$^1\text{H NMR}$ (500 MHz, CDCl_3) δ 4.69 (br, 1H, OH),

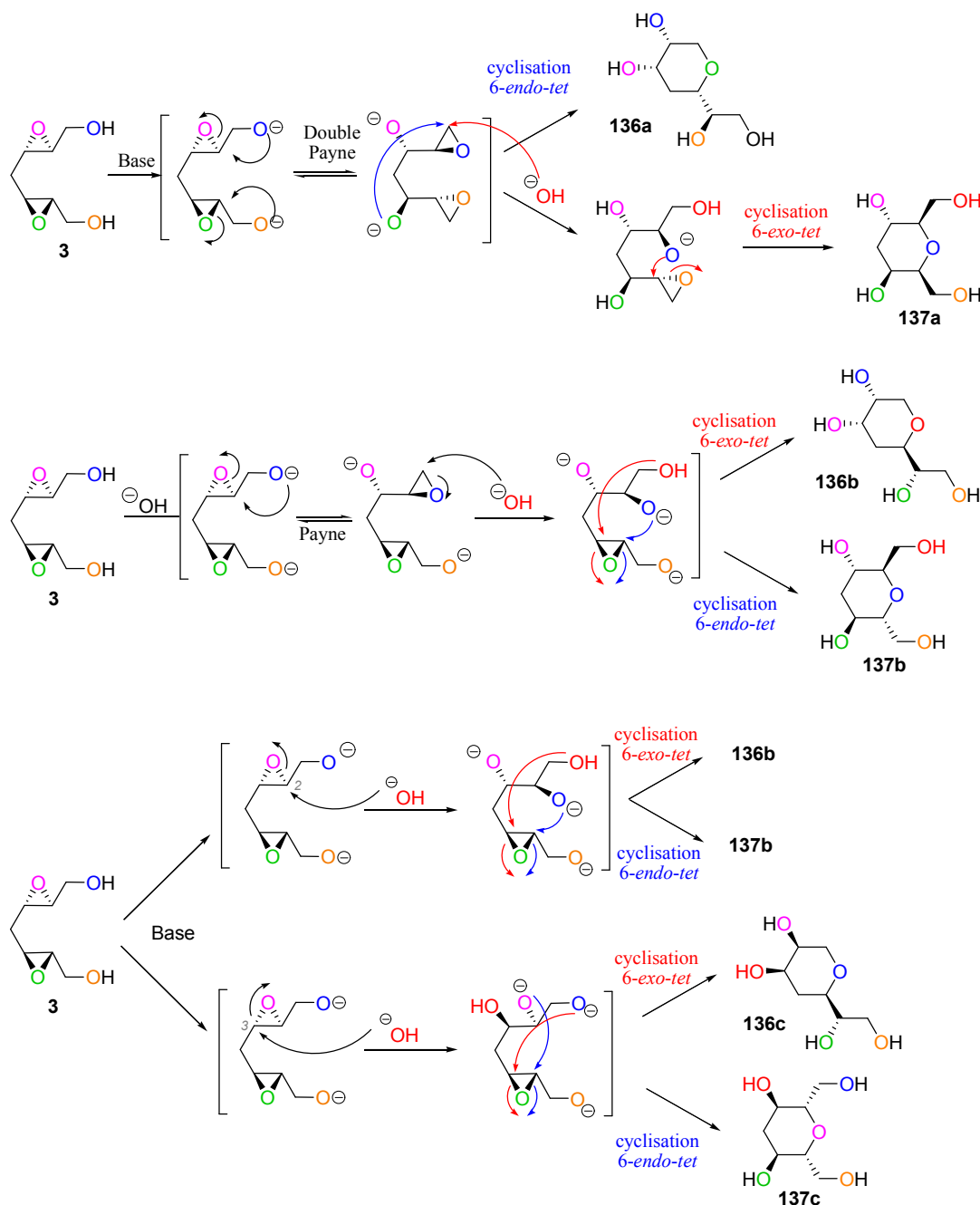
4.33-4.30 (m, 1H, $\text{CHOH}_{(\text{cycle})}$), 4.27-4.23 (m, 1H, CH), 4.14-4.10 (*minor*, m, 1H, CH), 4.04-4.01 (*minor*, m, 1H, CH), 3.94-3.92 (m, 1H, CH), 3.86-3.83 (m, 1H, $\text{CHOH} + \text{minor}$, 1H, CHOMe), 3.79-3.76 (*minor*, m, 1H, $\text{CHOH} + \text{minor}$, 1H, CH_2OH), 3.71-3.61 (m, 1H, $\text{CH}_2\text{OH} + \text{minor}$, 1H, CH_2OH), 3.60-3.52 (m, 1H, $\text{CH}_2\text{OH} + \text{minor}$, 1H, CH_2OH), 3.51-3.44 (m, 2H, $\text{CH}_2\text{OMe} + \text{minor}$, 1H, CH_2OH), 3.38 (s, 3H, CH_3), 3.34 (*minor*, s, 3H, CH_3), 1.13 (br, 2H, OH), 2.27-2.23 (*minor*, m, 1H, CH_2), 2.17-2.10 (m, 1H, CH_2), 2.07-2.02 (*minor*, m, 1H, CH_2), 1.90-1.85 (m, 1H, CH_2)

$^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 84.8 (1C, CH), 84.0 (*minor*, 1C, CH), 82.1 (*minor*, 1C, CHOMe), 79.9 (1C, CH), 79.2 (*minor*, 1C, CH), 74.0 (1C, $\text{CHOH}_{(\text{cycle})}$), 73.4 (1C, CH_2OMe), 73.3 (*minor*, 1C, CHOH), 73.1 (1C, CHOH), 63.7 (*minor*, 1C, CH_2OH), 63.6 (1C, CH_2OH), 63.1 (*minor*, 1C, CH_2OH), 59.4 (1C, OCH_3), 57.3 (*minor*, 1C, OCH_3), 35.7 (1C, CH_2), 33.1 (*minor*, 1C, CH_2)

ES+ : 2015.09 $[\text{M}+\text{Na}]^+$

HRMS (ESI+): calculated for $\text{C}_8\text{H}_{16}\text{O}_5\text{Na}$ $[\text{M}+\text{Na}]^+$ 215.0895, found 2015.0897

Possible formation of pyran derivatives



Formation of pyran derivatives such as above was a possibility but clearly refuted by the non-observation after acetonide formation of a bisprotected diol derivatives (for **136 a-c** potential compounds). For compounds **137a-c** the symmetrical nature of such compounds would have stood up by NMR.

Preparation of acetonide **5**:

To a solution of the furanic tetraol **2** (639 mg, 5.59 mmol, 1 eq) in 30 mL of acetone, 2,2-dimethoxypropane (490 μ L, 3.95 mmol, 1,1 eq) and para-toluenesulfonic acid (68 mg, 0.36 mmol, 0.1 eq) were added. The mixture was stirred at reflux for 3 h. Solid NaHCO_3 was added and the solvent was concentrated under reduced pressure prior to purification by column chromatography (DCM/MeOH: 98/2 to 94/6), to yield the acetonide **5** with its diastereoisomer (614 mg, 79%).

$R_f = 0.12$ (DCM/MeOH: 9.5/0.5)

Mixture of diastereoisomers :

$^1\text{H NMR}$ (500 MHz, $\text{CDCl}_3 + \text{D}_2\text{O}$) δ 4.45-4.40 (m, 1H, CHOH), 4.38-4.32 (*diast*, m, 0.2H, CHOH), 4.32-4.28 (m, 1H, CH), 4.27-4.16 (m, 1H, CH + *diast*, 0.4H, CH), 4.12-4.06 (m, 1H, CH_2O + *diast*, 0.2H, CH_2O), 4.05-4.00 (*diast*, m, 0.3H, CH), 3.93-3.91 (m, 1H, CH), 3.75 (dd, $J = 11.9, 3.2$, 1H, CH_2OH), 3.69 (dd, $J = 8.5, 6.0$, 1H, CH_2O), 3.65-3.62 (*diast*, m, 0.3H, CH_2O), 3.63-3.59 (m, 1.2H uncluded (dd, $J = 11.9, 3.5$, 1H, CH_2OH + *diast*, m, 0.2H, CH_2OH)), 3.55-3.50 (*diast*, m, 0.2H, CH_2OH), 2.50-2.25 (br, 2H, OH + *diast*, m, 1H, CH_2), 2.20-2.10 (m, 1H, CH_2), 1.99-1.90 (m, 1H, CH_2 + *diast*, 1H, CH_2), 1.47 (*diast*, s, 0.7H, CH_3), 1.43 (s, 3H, CH_3), 1.37 (*diast*, s, 0.7H, CH_3), 1.35 (s, 3H, CH_3)

$^{13}\text{C NMR}$ (125 MHz, $\text{CDCl}_3 + \text{D}_2\text{O}$) δ 109.8 (1C, Cq + *diast*), 87.6 (*diast*, 1C, CH), 87.3 (1C, CH), 79.3 (1C, CH), 79.3 (*diast*, 1C, CH), 77.6 (*diast*, 1C, CH), 77.1 (1C, CH), 74.1 (1C, CHOH), 72.9 (*diast*, 1C, CHOH), 66.7 (*diast*, 1C, CH_2), 66.6 (1C, CH_2), 63.3 (1C, CH_2), 63.1 (*diast*, 1C, CH_2), 36.3 (1C, CH_2), 35.3 (*diast*, 1C, CH_2), 26.3 (*diast*, 1C, CH_3), 26.2 (1C, CH_3), 25.1 (1C, CH_3), 25.0 (*diast*, 1C, CH_3)

To 30 mg of the mixture of diastereoisomeric acetonides, 3 mL of pentane was added. The solution was warmed at 40 °C and DCM was added until complete dissolution of the solid. Pentane was added until the solution become turbid (2 drops). The solution was allowed to reach rt and the solvent was slowly evaporated for 2 days. The crystals obtained were washed with cold cyclohexane then cold Et_2O to remove the non crystallized diastereoisomer. The washed crystals were dissolved in Et_2O (1 mL) and the solvent was slowly evaporated in 2 days to give pure crystals of the major acetonide **5** (10 mg).

Single cristal :

$^1\text{H NMR}$ (500 MHz, $\text{CDCl}_3 + \text{D}_2\text{O}$) δ Cq is missing, 4.45-4.40 (m, 1H, CHOH), 4.35-4.30 (m, 1H, CH), 4.30-4.20 (m, 1H, CH), 4.10 (dd, $J = 8.5, 6.9$, 1H, CH_2O), 3.98-3.90 (m, 1H, CH), 3.76 (dd, $J = 12, 3.3$, 1H, CH_2OH), 3.69 (dd, $J = 8.5, 6.1$, 1H, CH_2O), 3.59 (dd, $J = 11.9, 3.4$, 1H, CH_2OH), 2.22-2.10 (m, 1H, CH_2), 1.99-1.90 (m, 1H, CH_2), 1.44 (s, 3H, CH_3), 1.36 (s, 3H, CH_3)

$^{13}\text{C NMR}$ (125 MHz, $\text{CDCl}_3 + \text{D}_2\text{O}$) δ 88.6 (1C, CH), 80.6 (1C, CH), 78.3 (1C, CH), 75.3 (1C, CHOH), 67.9 (1C, CH_2), 64.9 (1C, CH_2), 37.5 (1C, CH_2), 27.4 (1C, CH_3), 26.3 (1C, CH_3)

ES+: 241.11 $[\text{M}+\text{Na}]^+$

HRMS (ESI+): calculated for $\text{C}_{10}\text{H}_{18}\text{O}_5\text{Na}$ $[\text{M}+\text{Na}]^+$ 241.1052, found 241.1055

IR (ν_{max} cm^{-1}): 3297, 2987, 2904, 2879, 1373, 1207, 1052, 995, 850

$[\alpha]_{\text{D}}^{20}(\text{CHCl}_3) = +33.1$ ($c = 5.23$)

X-ray diffraction:

Crystal evaluation and data collection were done at the ID29 beamline of the European Synchrotron Radiation Facility in Grenoble, France, with monochromatic X-rays ($\lambda = 0.72932 \text{ \AA}$) and a Pilatus-6M detector. Data reduction was performed using the XDS package.⁴ The structure was solved using the *ab initio* iterative charge flipping method with parameters described elsewhere,⁵ with use of the SUPERFLIP program.⁶ The structure was then refined using full-matrix least-squares procedures as implemented in CRYSTALS⁷ on all independent reflections with $I > 2\sigma(I)$. The hydrogen atoms were refined with riding constraints, except the two hydrogens involved in hydrogen bonding, which were refined with geometric restraints. The absolute configuration of the crystal structure was arbitrarily assigned, since the used wavelength does not permit to refine the Flack parameter for a structure that contains only carbon, hydrogen and oxygen atoms.

Crystal data for acetonide **5**: Formula= C₁₀H₁₈O₅, Moiety Formula= C₁₀H₁₈O₅, $T = 100$ K, $M_r = 218.24$ g.mol⁻¹, crystal size = 0.030x0.050x0.110 mm³, monoclinic, space group C2, $a = 8.8970(13)$, $b = 6.7680(15)$, $c = 18.304(2)$ Å, $\alpha = 90^\circ$, $\beta = 90.365(6)^\circ$, $\gamma = 90^\circ$, $V = 1102.2(3)$ Å³, $Z = 4$, $\rho_{\text{calcd}} = 1.315$ gcm⁻³, $\mu = 0.105$ mm⁻¹, $\theta_{\text{max}} = 31.361^\circ$, 10485 reflections measured, 1593 unique, 1589 with $I > 2\sigma(I)$, $R_{\text{int}} = 0.059$, 143 refined parameters, $R_1(I > 2\sigma(I)) = 0.05210$, $wR_2(I > 2\sigma(I)) = 0.0491$, $R_1(\text{all data}) = 0.0521$, $wR_2(\text{all data}) = 0.0491$, GOF = 1.0434, $\Delta\rho(\text{min/max}) = -0.55/0.42$ eÅ⁻³.

CCDC-1403506 contains the supplementary crystallographic data for this paper. These data can be obtained free of charge via www.ccdc.cam.ac.uk/conts/retrieving.html (or from the Cambridge Crystallographic Data Centre, 12, Union Road, Cambridge CB21EZ, UK; fax: (+44)1223-336-033; or deposit@ccdc.cam.ac.uk).

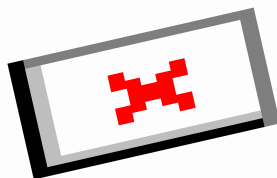


Figure 1 : Ortep representation of the structure of molecule acetonide **5 with ellipsoids at the 50% probability level**

Preparation of mono protected TBS-ether **6**:

To a solution of the previously prepared acetonide **5** and its diastereoisomer (600 mg, 2.77 mmol, 1 eq) in DCM (30 mL) at 0 °C were successively added TBSCl (1.67 g, 11.09 mmol, 4 eq), imidazole (1.51 g, 22.19 mmol, 8 eq) and DMAP (33 mg, 0.27 mmol, 0.1 eq), and the reaction was allowed to stir at rt overnight. A saturated NH₄Cl aqueous solution (ca. 20 mL) was added, the organic phase was separated and the aqueous phase extracted with DCM (3 x 15 mL). The combined organic phases were washed with NaCl_(Sat) (20 mL), dried over MgSO₄ and concentrated under vacuum. The crude product was purified by column chromatography (pentane/EtOAc: 95:5) to afford the intermediate diTBS-acetonide (1.09 g, 88%) as a colourless oil.

$R_f = 0.45$ (Cyclohexane/ EtOAc: 90/10)

¹H NMR (500 MHz, CDCl₃) δ 4.35-4.34 (*diast*, m, 0.2H, CH), 4.32-4.30 (m, 1H, CH), 4.17-4.12 (*diast*, m, 0.2H, CH), 4.10-4.04 (m, 2H, CH₂O, CH + *diast*, 0.3H, CH₂O), 3.97-3.93 (m, 1H, CH + *diast*, 0.2H, CH), 3.87-3.83 (m, 1.2H, included (dd, $J = 7.9, 5.9$, 1H, CH₂O + *diast*, m, 0.2H, CH₂O)), 3.83-3.81 (*diast*, m, 0.2H, CH), 3.80-3.77 (m, 1H, CH), 3.62-3.55 (m, 1.2H, included (dd, $J = 10.8, 4.1$, 1H, CH₂O + *diast*, m, 0.2H, CH₂O), 3.52-3.48 (*diast*, m, 0.2H, CH₂O), 3.46 (dd, $J = 10.8, 5.8$, 1H, CH₂O), 2.30-2.20 (*diast*, m, 0.2H, CH₂), 1.96-1.89 (m, 1H, CH₂ + *diast*, 0.2H, CH₂), 1.88-1.83 (m, 1H, CH₂), 1.41 (s, 3H, CH₃), 1.39 (*diast*, s, 0.8H, CH₃), 1.34 (s, 3H, CH₃), 1.33 (*diast*, s, 0.8H, CH₃), 0.90-1.10 (m, 18H, CH₃(TBS)⁺ *diast*, 5H, CH₃(TBS)), 0.10-0.00 (m, 12H, CH₃Si + *diast*, 3H, CH₃Si)

¹³C NMR (125 MHz, CDCl₃) δ 109.4 (1C, Cq), 109.2 (*diast*, 1C, Cq), 87.6 (1C, CH), 87.2 (*diast*, 1C, CH), 80.1 (*diast*, 1C, CH), 79.2 (1C, CH), 78.4 (1C, CH), 78.3 (*diast*, 1C, CH), 73.5 (1C, CH), 73.4

(*diast*, 1C, CH), 67.9 (*diast*, 1C, CH₂O), 67.5 (1C, CH₂O), 63.6 (1C, CH₂O), 63.5 (*diast*, 1C, CH₂O), 38.2 (1C, CH₂), 38.0 (*diast*, 1C, CH₂), 26.9 (*diast*, 1C, CH₃), 26.7 (1C, CH₃), 26.1 (3C, CH₃(TBS)), 26.0 (*diast*, 3C, CH₃(TBS)), 25.9 (3C, CH₃(TBS)), 25.8 (*diast*, 3C, CH₃(TBS)), 25.5 (1C, CH₃), 25.4 (*diast*, 1C, CH₃), 18.5 (1C, Cq + *diast*), 18.1 (1C, Cq + *diast*), -4.5 (1C, CH₃Si + *diast*), -4.6 (1C, CH₃Si + *diast*), -5.2 (1C, CH₃Si + *diast*), -5.3 (1C, CH₃Si + *diast*)

ES+: 447.30 [M+H]⁺, 389.25 [M+H-(CH₃)₂CO]⁺

HRMS (ESI⁺): calculated for C₂₂H₄₇O₅Si₂ [M+H]⁺ 447.2962, found 447.2966

IR (ν_{max} cm⁻¹): 2954, 2930, 2858, 1473, 1463, 1380, 1370, 1252, 1214, 1064, 831, 774

To a solution of diTBS-acetonide (67 mg, 0.15 mmol, 1 eq) in EtOH 96% (3 mL), at 0 °C, PPTS (38 mg, 0.15 mmol, 1 eq) was added and the reaction was stirred at the same temperature for 4 days. At this temperature a saturated NaHCO₃ aqueous solution (ca. 2 mL) was added, the organic phase was separated and the aqueous phase extracted with EtOAc (3 x 2 mL). The combined organic phases were washed with water and NaCl_(Sat), dried over MgSO₄ and concentrated under vacuum. The crude product was purified by column chromatography (pentane/EtOAc: 90:10 to 85:15) to afford the intermediate alcohol **6** (36 mg, 73%, 85 brsm) as a colourless oil, as well as starting material (10 mg).

R_f = 0.12 (Cyclohexane/EtOAc: 80:20)

¹H NMR (300 MHz, CDCl₃) δ 4.35-4.30 (m, 1H, CHO), 4.30-4.10 (m, 2H, CHO + *diast*, 0.5H, CH), 4.10-4.05 (m, 1H, CH₂O + *diast*, 0.4H, CH), 4.05-3.90 (*diast*, 0.2H, CH), 3.90-3.80 (m, 1H, CH + *diast*, 0.3H, CH₂O), 3.82-3.70 (*diast*, m, 0.2H, CH), 3.73-3.72 (m, 1H, CH₂O + *diast*, 0.4H, CH₂O), 3.70-3.68 (m, 1H, CH₂O), 3.56-3.50 (m, 1H, CH₂O + *diast*, 0.2H, CH₂O), 2.33-2.24 (*diast*, m, 0.2H, CH₂), 2.11-2.02 (m, 2H, CH₂ OH + *diast*, 0.2H, CH₂), 1.91-1.84 (m, 1H, CH₂), 1.43 (s, 3H, CH₃), 1.41 (*diast*, s, 0.9H, CH₃), 1.35 (s, 3H, CH₃), 1.34 (*diast*, s, 0.9H, CH₃), 0.95-0.80 (m, 9H, CH₃(TBS) + *diast*, 3H, CH₃(TBS)), 0.10- -0.10 (m, 6H, CH₃Si + *diast*, 1.6H, CH₃Si)

¹³C NMR (125 MHz, CDCl₃) δ 109.7 (1C, Cq), 87.7 (1C, CH), 85.9 (*diast*, 1C, CH), 79.4 (1C, CH), 79.3 (*diast*, 1C, CH), 78.2 (*diast*, 1C, CH), 77.4 (1C, CH), 74.0 (1C, CHOH), 72.6 (*diast*, 1C, CHOH), 67.5 (*diast*, 1C, CH₂), 66.7 (1C, CH₂), 63.4 (1C, CH₂), 62.4 (*diast*, 1C, CH₂), 38.3 (*diast*, 1C, CH₂), 36.8 (1C, CH₂), 26.9 (*diast*, 1C, CH₃), 26.2 (1C, CH₃), 25.9 (3C, CH₃(TBS)), 25.8 (*diast*, 3C, CH₃(TBS)), 25.4 (*diast*, 1C, CH₃), 25.1 (1C, CH₃), 18.1 (1C, Cq + *diast*), -4.5 (1C, CH₃Si + *diast*), -4.7 (1C, CH₃Si + *diast*)

ES+: 355.19 [M+Na]⁺, 275.17 [M+H-(CH₃)₂CO]⁺

HRMS (ESI⁺): calculated for C₁₆H₃₂O₅NaSi [M+Na]⁺ 355.1917, found 355.1916

IR (ν_{max} cm⁻¹): 3456, 2930, 2859, 1473, 1463, 1371, 1252, 1213, 1104, 1058, 832, 775

Preparation of Ester **7**:

To a solution of **6** (200 mg, 0.60 mmol, 1 eq) in DCM (12 mL) and H₂O (24 μL) were added NaHCO₃ solid (152mg, 1.80 mmol, 3 eq) and DMP (5.12 mL, 2.4 mmol, 0.47 M in DCM, 4 eq) and the reaction was stirred at rt overnight. The reaction was quenched by adding a 10% NaHCO₃/Na₂S₂O₃ (10 mL, 1/1, v/v) aqueous solution, and stirred for 2 h. The aqueous phase was extracted with DCM (3 x 10 mL), washed with water, dried over MgSO₄ and concentrated under vacuum to afford the corresponding aldehyde, which is used for the next step without purification.

To a -78 °C cold suspension of the phosphonium salt (1.2 g, 2.4 mmol, 4 eq) in THF (30 mL) was slowly added NaHMDS (1.11 mL, 2.2 mmol, 2 M in THF, 3.7 eq). The solution coloured in orange. After 1 h stirring, a solution of the previously prepared aldehyde in THF (20 mL) was slowly added at -

78 °C by cannulation, and the reaction was allowed to reach rt, then left to stir for 24 h. The reaction was quenched by addition of water (20 mL) and Et₂O (20 mL), extracted with Et₂O (3 x 10 mL), washed with brine, dried over MgSO₄ and concentrated under vacuum. The crude product was purified by chromatography column (pentane/EtOAc: 90/10) to afford the corresponding alkene THF derivative (211 mg, 75%) as colourless oil.

R_f = 0.45 (Cyclohexane/EtOAc: 5/5)

¹H NMR (500 MHz, CDCl₃) δ 5.60-5.51 (m, 1H, CH=), 5.29-5.22 (m, 1H, CH=), 4.49-4.39 (*diast*, m, 0.03H, CH + dd, *J* = 9.6, 4.3, 1H, CH), 4.15-4.02 (m, 4H, CH₂O, CH + *diast*, 0.6H, CH₂O, CH), 4.01-3.97 (m, 2H, CH + *diast*, 0.3H, CH), 3.89-3.79 (m, 1.15H, included (*diast*, dd, *J* = 8.1, 5.3, 0.15H, CH₂O + dd, *J* = 8.3, 5.6, 1H, CH₂O)), 2.31-2.24 (m, 2.5H, included (t, *J* = 7.5, 2H, CH₂CO + *diast*, m, 0.5H, CH₂)), 2.18-2.11 (m, 1H, CH₂ + *diast*, 0.3H, CH₂), 2.11-1.92 (m, 3H, CH₂ + *diast*, 0.3H, CH₂), 1.68-1.57 (m, 2H, CH₂ + *diast*, 0.3H, CH₂), 1.43-1.27 (m, 10H, CH₃, CH₂ + *diast*, 1.5H, CH₃, CH₂), 1.25 (t, *J* = 7.1, 3H, CH₃ + *diast*, 0.6H, CH₃), 0.90-0.84 (m, 9H, CH₃(TBS) + *diast*, 1.7H, CH₃(TBS)), 0.10- -0.05 (m, 6H, CH₃Si + *diast*, 1H, CH₃Si)

¹³C NMR (75 MHz, CDCl₃) δ 173.9 (1C, Cq + *diast*), 134.3 (*diast*, 1C, CH=), 134.2 (1C, CH=), 128.5 (1C, CH=), 129.3 (*diast*, 1C, CH=), 109.5 (1C, Cq + *diast*), 82.2 (1C, CH), 81.2 (*diast*, 1C, CH), 78.9 (1C, CH), 78.6 (*diast*, 1C, CH), 78.5 (*diast*, 1C, CH), 78.4 (1C, CH), 77.7 (*diast*, 1C, CH), 77.5 (1C, CH), 67.9 (*diast*, 1C, CH₂O), 67.6 (1C, CH₂O), 60.5 (*diast*, 1C, CH₂O), 60.3 (1C, CH₂O), 38.3 (*diast*, 1C, CH₂), 38.2 (1C, CH₂), 34.4 (1C, CH₂COO), 29.5 (1C, CH₂), 29.5 (*diast*, 1C, CH₂), 29.0 (1C, CH₂), 28.9 (*diast*, 1C, CH₂), 27.9 (1C, CH₂), 27.8 (*diast*, 1C, CH₂), 26.9 (*diast*, 1C, CH₃), 26.7 (1C, CH₃), 25.9 (3C, CH₃(TBS)), 25.8 (*diast*, 3C, CH₃(TBS)), 25.4 (1C, CH₃), 25.4 (*diast*, 1C, CH₃), 25.0 (*diast*, 1C, CH₂), 25.0 (1C, CH₂), 18.2 (1C, Cq (TBS) + *diast*), 14.4 (1C, CH₃), 14.3 (*diast*, 1C, CH₃), -4.6 (1C, CH₃Si + *diast*), -4.7 (1C, CH₃Si + *diast*)

ES⁺: 471.31 [M+H]⁺, 488.34 [M+NH₄]⁺

HRMS (ESI⁺): calculated for C₂₅H₄₇O₆Si [M+H]⁺ 471.3142, found 471.3141

IR (ν_{max} cm⁻¹): 2931, 2858, 1736, 1463, 1370, 1252, 1211, 1150, 1110, 1061, 834, 776

To a solution of the previously synthesized alkene THF derivative (200 mg, 0.3 mmol, 1 eq) in 20 mL of EtOH, Pd/C 5% (50 mg, 120 mg/mmol) was added. Under H₂ atmosphere, the suspension was stirred at rt overnight. The mixture was filtered over a Celite[®] pad and rinsed with EtOH. The solvents were removed under reduced pressure and the crude reaction mixture was purified by flash chromatography (pentane/EtOAc: 90/10) to obtain ester 7 as colourless oil (175 mg, 87%).

R_f = 0.21 (Cyclohexane/EtOAc: 9/1)

¹H NMR (500 MHz, CDCl₃) δ 4.15-4.09 (m, 2.4H, included (q, *J* = 7.2, 2H, CH₂O + *diast*, 0.4H, CH₂O), 4.09-4.04 (m, 1.1H, included (dd, *J* = 8.3, 6.4, 1H, CH₂O + *diast*, m, 0.1H, CH₂O)), 4.04-3.98 (m, 1H, CH + *diast*, 0.3H, CH), 3.98-3.90 (m, 2H, CH + *diast*, 1H, CH), 3.90-3.87 (*diast*, m, 0.17H, CH), 3.86-3.79 (m, 1.17H, included (dd, *J* = 8.3, 5.9, 1H, CH₂O + *diast*, m, 0.17H, CH₂O)), 3.77-3.71 (*diast*, m, 0.3H, CH), 3.70-3.63 (*diast*, m, 0.16H, CH), 3.63-3.58 (m, 1H, CH), 2.29-2.23 (m, 2.6H, included (t, *J* = 7.5, 2H, CH₂CO + *diast*, m, 0.6H, CH₂CO)), 1.89 (dd, *J* = 7.2, 5.0, 2H, CH₂) 1.87-1.81 (*diast*, m, 0.4H, CH₂), 1.65-1.55 (m, 2H, CH₂ + *diast*, 0.6H, CH₂), 1.45-1.18 (m, 22.9H included (s, 3H, CH₃ + s, 3H, CH₃ + m, 10H, CH₂ + *diast*, 3.9H, CH₃, CH₂ + t, *J* = 7.2, 3H, CH₃)), 1.00-0.80 (m, 9H, CH₃(TBS) + *diast*, 1.8H, CH₃(TBS)), 0.10- -0.10 (m, 6H, CH₃Si + *diast*, 1.1H, CH₃Si)

¹³C NMR (125 MHz, CDCl₃) δ 174.0 (1C, Cq + *diast*), 109.4 (1C, Cq + *diast*), 86.6 (1C, CH), 85.7 (*diast*, 1C, CH), 78.6 (1C, CH), 78.5 (1C, CH), 78.4 (*diast*, 1C, CH), 78.4 (*diast*, 1C, CH), 76.6 (*diast*,

1C, CH), 76.4 (1C, CH), 67.8 (*diast*, 1C, CH₂O), 67.5 (1C, CH₂O), 65.9 (*diast*, 1C, CH₂O), 60.3 (1C, CH₂O), 38.1 (1C, CH₂), 38.1 (*diast*, 1C, CH₂), 34.6 (1C, CH₂COO), 34.0 (1C, CH₂), 33.6 (*diast*, 1C, CH₂), 29.6 (*diast*, 1C, CH₂), 29.5 (1C, CH₂), 29.3 (1C, CH₂), 29.2 (*diast*, 1C, CH₂), 29.2 (1C, CH₂), 26.9 (*diast*, 1C, CH₃), 26.7 (1C, CH₃), 26.1 (1C, CH₂), 26.0 (*diast*, 1C, CH₂), 25.9 (1C, CH₃), 25.8 (*diast*, 1C, CH₃), 25.5 (3C, CH₃(TBS)), 25.1 (*diast*, 3C, CH₃(TBS)), 25.0 (1C, CH₂ + *diast*), 18.1 (1C, C_q(TBS) + *diast*), 14.4 (1C, CH₃ + *diast*), -4.4 (1C, CH₃Si + *diast*), -4.6 (1C, CH₃Si + *diast*)

ES+: 473.33 [M+H]⁺, 490.36 [M+NH₄]⁺

HRMS (ESI+): calculated for C₂₅H₄₉O₆Si [M+H]⁺ 473.3298, found 473.3299

IR (ν_{max} cm⁻¹): 2930, 2858, 1737, 1464, 1370, 1251, 10212, 1179, 1156, 1108, 1062, 834, 775

Preparation of 1-methyl-1-cyclopropyl hydroxyl derivative **8**:

To a solution of ester **7** (85 mg, 0.18 mmol, 1 eq) in 3 mL of DCM was added freshly distilled *i*-Pr₂NEt (61 μL, 0.36 mmol, 1 eq) and was cooled to 0 °C. Then, freshly distilled TMSOTf (55 μL, 0.31 mmol, 1.7 eq) was added dropwise and the mixture was warmed to rt and heated over reflux overnight. Then, the mixture was diluted with hexane, filtered through a plug of neutral alumina (activity III; 6% of water), rinsed with hexane/EtOAc: 90/10, and concentrated under reduced pressure.

To the resulting enol ether, in 2 mL of Et₂O, was added Et₂Zn (539 μL, 0.54 mmol, 1 M in decane, 3 eq) and distilled CH₂I₂ (73 μL, 0.90 mmol, 5 eq) over 10 min. The mixture was stirred overnight at rt. The reaction mixture was diluted with 10 mL of 1 N NaOH, extracted with Et₂O (3 x 7 mL), dried, and concentrated under reduced pressure.

The crude oil was treated with 2 mL of MeOH and 7 mg of K₂CO₃ for 15 min at rt to remove the remaining TMS group. The mixture was quenched with saturated NH₄Cl, extracted with Et₂O (3 x 2 mL), dried over MgSO₄ and concentrated under reduced pressure. Purification by flash chromatography (pentane/EtOAc: 90/10) gave 1-methyl-1-cyclopropyl (MCP) hydroxyl derivative **8** free from its diastereoisomer (54 mg, 62%) as a colorless oil.

R_f = 0.22 (Cyclohexane/EtOAc: 80/20)

¹H NMR (300 MHz, CDCl₃) δ 4.17-4.00 (m, 3H, included (1H, CH + q, *J* = 7.1, 2H, CH₂OCO)), 3.94-3.86 (m, 1H, CH), 3.71-3.66 (m, 2H, CH₂OH), 3.66-3.55 (m, 2H, CH), 2.40 (br, 1H, OH), 2.27 (t, *J* = 7.1, 2H, CH₂CO₂), 1.87-1.79 (m, 2H, CH₂), 1.64-1.54 (m, 3H, CH₂), 1.45-1.20 (m, 15H, included (s, 3H, CH₃ + m, 9H, CH₂ + t, *J* = 7.1, 3H, CH₃)), 0.90-0.83 (m, 11H, included (s, 9H, CH₃(TBS) + m, 2H, CH₂(MCP))), 0.47-0.36 (m, 2H, CH₂(MCP)), 0.10- -0.03 (m, 6H, CH₃Si)

¹³C NMR (75 MHz, CDCl₃) δ 174.0 (1C, C_q), 86.8 (1C, CH), 79.5 (1C, CH), 78.6 (1C, CH), 76.4 (1C, CH), 63.8 (1C, CH₂OH), 60.3 (1C, CH₂OCO), 58.4 (1C, C_q(MCP)), 38.2 (1C, CH₂), 34.5 (1C, CH₂), 34.2 (1C, CH₂), 29.6 (1C, CH₂), 29.3 (1C, CH₂), 29.2 (1C, CH₂), 26.2 (1C, CH₂), 25.0 (3C, CH₃(TBS)), 25.1 (1C, CH₂), 22.7 (1C, CH₃(MCP)), 18.1 (1C, C_q(TBS)), 14.5 (1C, CH₂(MCP)), 14.4 (1C, CH₃), 13.5 (1C, CH₂(MCP)), -4.4 (1C, CH₃Si), -4.6 (1C, CH₃Si)

ES+ : 509.33 [M+Na]⁺

HRMS (ESI+): calculated for C₂₆H₅₀O₆SiNa [M+Na]⁺ 509.3274, found 509.3275

IR (ν_{max} cm⁻¹): 3446, 2929, 2857, 1737, 1464, 1384, 1255, 1179, 1109, 1047, 834, 775

[α]_D²⁰(CHCl₃) = +26.4 (c=10)

Preparation of enone **9**:

To a solution of alcohol **8** (54 mg, 0.11 mmol, 1 eq) in DCM (2 mL) and 2 drops of water was added, at 0 °C, the DMP (353 μ L, 0.17 mmol, 0.47 M in DCM, 1.5 eq). After stirring for 1.5 h, the reaction was quenched by adding a 10% NaHCO₃/Na₂S₂O₃ (2 mL, 1/1, v/v) aqueous solution, and stirred for 30 min. The aqueous phase was extracted with DCM (3 x 1 mL), washed with brine, dried over MgSO₄ and concentrated under vacuum to afford the crude aldehyde.

To a solution of the crude aldehyde in 1 mL of DCM was added the 1-(triphenylphosphoranylidene)-butan-2-one (96 mg, 0.28 mmol, 2.6 eq). After stirring at rt for 64 h, the reaction was quenched by addition of water (1 mL), extracted with DCM (3 x 1 mL), dried over MgSO₄ and concentrated under vacuum. The crude product was purified by chromatography column (pentane/EtOAc: 90/10) to afford enone **9** (30 mg, 51%) as a colourless oil.

R_f = 0.44 (Cyclohexane/EtOAc: 80/20)

¹H NMR (500 MHz, CDCl₃) δ 6.76 (dd, J = 16.1, 5.6, 1H, CH=), 6.30 (dd, J = 16.1, 1.5, 1H, CH=), 4.23-4.19 (m, 1H, CH), 4.11 (q, J = 7.2, 2H, CH₂OCO), 4.09-4.02 (m, 1H, CH), 3.87-3.82 (m, 1H, CH), 3.63-3.53 (m, 1H, CH), 2.58 (q, J = 7.3, 2H, CH₂CO), 2.27 (t, J = 7.5, 2H, CH₂CO₂), 1.86-1.78 (m, 1H, CH₂), 1.64-1.54 (m, 3H, CH₂ + 1H, CH₂), 1.40-1.28 (m, 9H, CH₂ + s, 3H, CH₃), 1.24 (t, J = 7.2, 3H, CH₃), 1.09 (t, J = 7.3, 3H, CH₃), 0.95-0.90 (m, 1H, CH₂(MCP)), 0.85 (s, 9H, CH₃(TBS)), 0.83-0.76 (m, 1H, CH₂(MCP)), 0.42-0.31 (m, 2H, CH₂(MCP)), 0.02 (m, 6H, CH₃Si)

¹³C NMR (125 MHz, CDCl₃) δ 200.9 (1C, Cq), 174.0 (1C, Cq), 145.3 (1C, CH=), 130.0 (1C, CH=), 86.6 (1C, CH), 79.4 (1C, CH), 78.8 (1C, CH), 76.2 (1C, CH), 60.3 (1C, CH₂OCO), 59.7 (1C, Cq(MCP)), 35.5 (1C, CH₂), 34.5 (1C, CH₂CO), 34.0 (1C, CH₂), 33.9 (1C, CH₂CO), 29.6 (1C, CH₂), 29.3 (1C, CH₂), 29.2 (1C, CH₂), 26.2 (1C, CH₂), 25.9 (3C, CH₃(TBS)), 25.1 (1C, CH₂), 22.6 (1C, CH₃(MCP)), 18.1 (1C, Cq(TBS)), 14.4 (1C, CH₂(MCP)), 14.4 (1C, CH₃), 13.4 (1C, CH₂(MCP)), 8.1 (1C, CH₃), -4.4 (1C, CH₃Si), -4.6 (1C, CH₃Si)

ES+ : 539.38 [M+H]⁺, 556.40 [M+NH₄]⁺, 467.32 [(M+H)- C₄H₇O]⁺

HRMS (ESI⁺): calculated for C₃₀H₅₅O₆Si [M+H]⁺ 539.3768, found 539.3769

IR (ν_{\max} cm⁻¹): 2931, 2857, 1735, 1702, 1679, 1463, 1374, 1254, 1190, 1108, 1041, 834, 775

[α]_D²⁰(CHCl₃) = +26.9 (c = 10)

Preparation of enone **10**:

To a solution of the methylcyclopropyl protected alcohol **9** (30 mg, 0.06 mmol, 1 eq) in 0.5 mL of THF and 50 μ L of water was added NBS (12 mg, 0.07 mmol, 1.2 eq). After stirring 1 h at rt, the mixture was quenched with saturated Na₂S₂O₃, extracted with Et₂O (3 x 0.5 mL) washed with brine, dried over MgSO₄, and concentrated under vacuum. The crude product was purified by chromatography column (pentane/EtOAc: 80/20) to afford the allylic alcohol **10** (23 mg, 85%) as colourless oil.

R_f = 0.26 (Cyclohexane/EtOAc: 80/20)

¹H NMR (300 MHz, CDCl₃) δ 6.70 (dd, J = 15.9, 4.1, 1H, CH=), 6.42 (dd, J = 15.9, 1.9, 1H, CH=), 4.60-4.50 (m, 1H, CH), 4.27-4.17 (m, 1H, CH), 4.11 (q, J = 7.1, 2H, CH₂OCO), 3.95-3.87 (m, 1H, CH), 3.73-3.67 (m, 1H, CH), 2.58 (q, J = 7.3, 2H, CH₂CO), 2.35 (d, J = 2.2, 1H, OH), 2.28 (t, J = 7.4, 2H, CH₂CO₂), 1.97-1.86 (m, 1H, CH₂), 1.65-1.49 (m, 3H, CH₂), 1.49-1.20 (m, 13H, included (10H, CH₂ + t, J = 7.1, 3H, CH₃)), 1.10 (t, J = 7.3, 3H, CH₃), 0.90-0.80 (m, 9H, CH₃(TBS)), 0.10- -0.10 (m, 6H, CH₃Si)

¹³C NMR (75 MHz, CDCl₃) δ 200.7 (1C, Cq), 174.0 (1C, Cq), 145.3 (1C, CH=), 129.1 (1C, CH=), 87.0 (1C, CH), 79.9 (1C, CH), 76.6 (1C, CH), 71.1 (1C, CH), 60.3 (1C, CH₂OCO), 34.5 (1C, CH₂), 34.4 (1C, CH₂CO), 34.2 (1C, CH₂), 34.0 (1C, CH₂CO), 29.5 (1C, CH₂), 29.3 (1C, CH₂), 29.2 (1C, CH₂), 26.1 (1C, CH₂), 25.9 (3C, CH₃(TBS)), 25.1 (1C, CH₂), 18.1 (1C, Cq(TBS)), 14.4 (1C, CH₃), 8.1 (1C, CH₃), -4.4 (1C, CH₃Si), -4.6 (1C, CH₃Si)

ES+ : 485.33 [M+H]⁺, 507.31 [M+Na]⁺

HRMS (ESI+): calculated for C₂₆H₄₉O₆Si [M+H]⁺ 485.3298, found 485.3304

IR (ν_{max} cm⁻¹): 3443, 2932, 2855, 2361, 1737, 1676, 1460, 1371, 1256, 1194, 1113, 1059, 836, 777

[α]_D²⁰(CHCl₃) = +30.1 (c = 9)

Preparation of *ent*-16-(*RS*)-13-*epi*-ST-Δ¹⁴-9-PhytoF **1**:

To a solution of enone **10** (18 mg, 0.04 mmol, 1 eq) in MeOH (1.8 mL) was added CeCl₃·7H₂O (14 mg, 0.04 mmol, 1.01 eq), the reaction was cooled to 0 °C and NaBH₄ (1 mg, 0.03 mmol, 0.7 eq) was added. After 1 h stirring at rt., water (ca. 1 mL) was added and the aqueous phase was extracted with EtOAc (3 x 1 mL). The combined organic phases were washed with brine, dried over MgSO₄ and concentrated under vacuum. The crude product was purified by flash chromatography (pentane/ EtOAc: 80/20) to give the enediol compound (16 mg, 89%) as colourless oil.

R_f = 0.35 (Cyclohexane/ EtOAc: 50/50)

¹H NMR (500 MHz, CDCl₃) δ 5.80 (ddd, *J* = 17.1, 11.3, 6.3, 1H, CH= in addition with a shift of 1.4Hz linked to the *epimer*), 5.60 (ddd, *J* = 15.6, 5.7, 3.6, 1H, CH= in addition with a shift of 1.2Hz linked to the *epimer*), 4.37-4.30 (m, 1H, CH), 4.16-4.09 (m, 1H, CH + q, *J* = 7.5, 2H, CH₂OCO), 4.09-4.02 (m, 1H, CH), 3.93-3.87 (m, 1H, CH), 3.71-3.64 (m, 1H, CH), 2.27 (t, *J* = 7.50, 2H, CH₂CO₂), 2.21 (br, 1H, OH), 2.00-1.92 (m, 1H, CH₂), 1.64-1.50 (m, 5H, CH₂), 1.50-1.30 (m, 10H, CH₂ + t, *J* = 7.15, 3H, CH₃), 0.93-0.90 (td, *J* = 7.45, 3.15, 3H, CH₃), 0.90-0.80 (m, 9H, CH₃), 0.10- -0.10 (m, 6H, CH₃Si)

¹³C NMR (125 MHz, CDCl₃) δ 174.0 (1C, Cq), 135.2 (1C, CH=), 135.1 (*epi*, 1C, CH=), 128.6 (1C, CH=), 128.5 (*epi*, 1C, CH=), 86.8 (1C, CH), 80.7 (1C, CH), 80.6 (*epi*, 1C, CH), 76.6 (1C, CH), 76.6 (*epi*, 1C, CH), 73.8 (1C, CH), 73.8 (*epi*, 1C, CH), 71.8 (1C, CH), 71.7 (*epi*, 1C, CH), 60.3 (1C, CH₂OCO), 34.5 (1C, CH₂CO), 34.2 (1C, CH₂), 34.1 (*epi*, 1C, CH₂), 34.0 (1C, CH₂), 34.0 (*epi*, 1C, CH₂), 30.2 (1C, CH₂), 30.1 (*epi*, 1C, CH₂), 29.5 (1C, CH₂), 29.3 (1C, CH₂), 29.2 (1C, CH₂), 26.1 (1C, CH₂), 25.9 (3C, CH₃(TBS)), 25.1 (1C, CH₂), 18.1 (1C, Cq(TBS)), 14.4 (1C, CH₃), 9.8 (1C, CH₃), -4.4 (1C, CH₃Si), -4.6 (1C, CH₃Si)

ES+: 509.33 [M+Na]⁺, 504.34 [M+NH₄]⁺

HRMS (ESI+): calculated for C₂₆H₅₀O₆SiNa [M+Na]⁺ 509.3274, found 509.3275

IR (ν_{max} cm⁻¹): 3416, 2931, 1737, 1463, 1250, 1112, 1056, 969, 837, 776

[α]_D²⁰(CHCl₃) = +30.6 (c = 7.85)

A solution of TBAF (1 M/THF, 241 μL, 0.24 mmol, 7.6 eq) was added to the solution of enediol (15.5 mg, 0.03 mmol) in 0.5 mL of THF. The reaction was allowed to stir at rt for 3 h. CaCO₃ (46 mg), DOWEX-50W resin (16 mg) and MeOH (1 mL) were added, the mixture was allowed to stir at rt for 1 h and was filtrated through a pad of Celite® and concentrated under vacuum. The crude product was obtained and the triol, in mixture with transesterified methylated product (7/3 Ethyl/Methyl ratio) (11.3 mg) was directly put in the next step.

$R_f = 0.47$ (DCM/MeOH: 9/1)

To a solution the triol in THF (1 mL) and water (1 mL), LiOH.H₂O (7 mg, 0.17 mmol, 6 eq) was added and the reaction was allowed to stir at rt overnight. At 0 °C, the reaction was quenched by slow addition of an aqueous solution of HCl (1 M), until acidic pH and the aqueous phase was extracted with EtOAc (4 x 2 mL), washed with brine, dried and concentrated under vacuum. The crude product was purified by column chromatography (EtOAc/MeOH: 97/3) to afford the *ent*-16-(*RS*)-13-*epi*-ST- Δ^{14} -9-PhytoF **1** (8 mg, 76% in 2 steps) as colourless oil.

$R_f = 0.12$ (DCM/MeOH: 9/1)

¹H NMR (500 MHz, MeOD) δ 5.79-5.60 (m, 2H, CH=), 4.18-4.00 (m, 2H, CHO), 4.00-3.91 (m, 2H, CHO), 3.69-3.62 (m, 1H, CHO), 2.28 (t, $J = 7.4$, 2H, CH₂CO), 2.03-1.95 (m, 1H, CH₂), 1.83-1.76 (m, 1H, CH₂), 1.65-1.41 (m, 7H, CH₂), 1.35 (br, 7H, CH₂), 0.92 (td, $J = 7.4, 2.1$, 3H, CH₃)

¹³C NMR (125 MHz, MeOD) δ 177.8 (1C, Cq), 136.0 (1C, CH=), 135.8 (*epi*, CH=), 131.1 (1C, CH=), 131.1 (*epi*, CH=), 87.8 (*epi*, CH), 87.7 (1C, CH), 82.3 (*epi*, CH), 82.3 (1C, CH), 76.6 (1C, CH), 74.7 (*epi*, CHOH), 74.6 (1C, CHOH), 74.5 (*epi*, CHOH), 74.4 (1C, CHOH), 36.6 (1C, CH₂), 36.4 (*epi*, CH₂), 35.1 (1C, CH₂), 35.0 (1C, CH₂), 31.1 (1C, CH₂), 31.1 (*epi*, CH₂), 30.6 (1C, CH₂), 30.4 (1C, CH₂), 30.4 (*epi*, CH₂), 30.2 (1C, CH₂), 27.1 (1C, CH₂), 26.1 (1C, CH₂), 10.2 (1C, CH₃), 10.2 (*epi*, CH₃)

ES- : 343.21 [M-H]⁻

HRMS (ESI-): calculated for C₁₈H₃₁O₆ [M-H]⁻ 343.2121, found 343.2117

$[\alpha]_D^{20}$ (MeOH) = + 16.5 (c = 4)

3.2 Quantitation

Extraction of lipids from nut or seed samples

Raw, unprocessed, organic nut (Noberasco, Savona, Italy) and seed (Organic Gardens Co., Melbourne, Australia) samples (4 g), imported from local food market were obtained and finely grounded using electronic grinder (Kenwood AT320A). The ground sample was then extracted in a Soxhlet extractor with 100 mL n-hexane/ diethyl ether (80:20, v/v) for 6 hours⁸. The extracted oil was dried completely and purged using nitrogen gas and stored at -80°C until analysis.

Sample preparation for LC-MS/MS analysis

The prepared samples (n=7) were thawed at room temperature. The lipid component of the samples was extracted by Folch method with slight modifications^{9,10}. Briefly, 20 mL of ice-cold chloroform/ methanol (2:1, v/v) containing 0.01% butylated hydroxytoluene was added into each samples and agitated at room temperature for 15 min by an orbital shaker. Phase separation was introduced into the samples by adding 4 mL of 0.9% NaCl solution. Samples were mixed for 10-30 seconds and then centrifuged at 3,000 x g for 10 min at 4°C. The lower organic phase was collected into a glass vial and dried under a stream of nitrogen gas. The dried Folch extract was hydrolyzed with potassium hydroxide

in methanol (1:1) with heavy labeled isotopic isoprostane (derived from arachidonic acid) internal standard, 15-F_{2t}-IsoP-d₄ and α -linolenic acid-d₁₄ (Cayman Chemicals, MI, USA). After overnight hydrolysis in room temperature and neutralized by the addition of hydrochloric acid, equal volume of formic acid (pH 4.5) was further added and the samples were purified by solid phase extraction (SPE) using 60 mg mixed anionic exchange cartridges (MAX Oasis, Waters, USA) ¹¹. In brief, the cartridges were washed and preconditioned with 2 mL methanol and formic acid (pH 4.5) respectively. After loading the sample, it was further cleaned with 2 mL 2% ammonium hydroxide and then 2 mL hexane. The analytes were eluted with 2 mL hexane/ethanol/acetic acid mix (70/29.95/0.05), immediately dried under nitrogen and reconstituted in methanol for liquid chromatography tandem mass spectrometry (LC-MS/MS) analysis. To a part of the samples (n=3) prepared, 5 ng of *ent*-16-(*RS*)-13-*epi*-ST- Δ ¹⁴-9-PhytoF was added to evaluate the relative retention time of the chromatogram with the pure standard. All solvents used in the analysis were high performance liquid chromatography (HPLC) grade.

Liquid chromatography tandem mass spectrometry analysis

The α -linolenic acid and *ent*-16-(*RS*)-13-*epi*-ST- Δ ¹⁴-9-PhytoF were analyzed using LC-MS/MS system, 1290 Infinity LC (Agilent, USA) with Hillic C₁₈ column (2.6 mm particle size, 150 x 3.0 mm, Phenomenex, CA USA) maintained at 30°C. The mobile phase consisted of 5 mM ammonium formate in 90 v/v % acetonitrile (A) and 5 mM ammonium formate in 50 v/v % acetonitrile (B) and set at a gradient elution from 100 % A for 2.5 min, followed by an increase to 100 % B from 2.5 to 10 min and then kept constant for 3 min. The flow rate was constant at 100 μ l/min throughout the analysis. The triple quad mass spectrometer (Sciex Applied Biosystems, MA USA) was operated in a negative atmospheric pressure chemical ionization (APCI) mode. The spray voltage was set to -4200 V and nitrogen gas was used as the curtain gas. The analytes were detected by MS/MS using multiple reaction monitoring (MRM) and the transitions determined from synthesized *ent*-16-(*RS*)-13-*epi*-ST- Δ ¹⁴-9-PhytoF, and commercially available ALA, ALA and 15-F_{2t}-IsoP-d₄ (Cayman Chemicals, USA) were used (as shown in Table 1). A dwell time of 800 ms was used for each ion transition for a total scan time of 3.0 s. Quantitation of the compounds was achieved by relating the peak area with its corresponding deuterated internal standard peak ALA-d₁₄, and 15-F_{2t}-IsoP-d₄ was used to quantitate *ent*-16-(*RS*)-13-*epi*-ST- Δ ¹⁴-9-PhytoF. The response ratio of 15-F_{2t}-IsoP-d₄ to *ent*-16-(*RS*)-13-*epi*-ST- Δ ¹⁴-9-PhytoF (1:0.07) was adjusted accordingly in the calculation. The chromatogram and fragmentations of ALA and *ent*-16-(*RS*)-13-*epi*-ST- Δ ¹⁴-9-PhytoF are depicted in Figure 1. The relative retention time for the chromatogram of spiked samples and pure standard of *ent*-16-(*RS*)-13-*epi*-ST- Δ ¹⁴-9-PhytoF was 0.98 to 1.05. Concentrations of the *ent*-16-(*RS*)-13-*epi*-ST- Δ ¹⁴-9-PhytoF are expressed per gram of powdered nuts or seeds.

Statistical analysis

Statistical analysis was performed by GraphPad Prism version 6.0 for Macintosh (GraphPad Software, CA USA). All values are annotated as mean \pm SD. One-way analysis of variance (ANOVA) and Tukey's multiple comparison tests were performed between the compounds measured in the nuts and seeds. Only significant level of $p < 0.05$ was noted in the results.

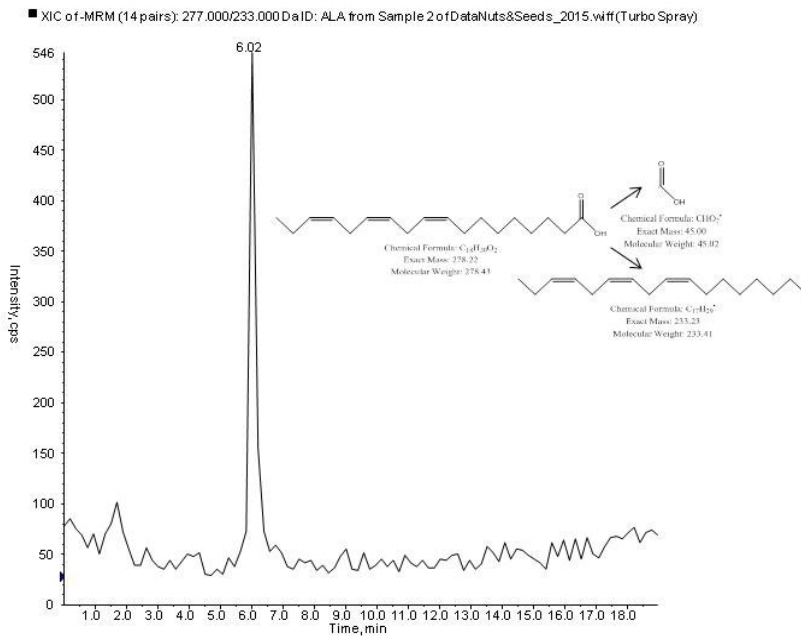
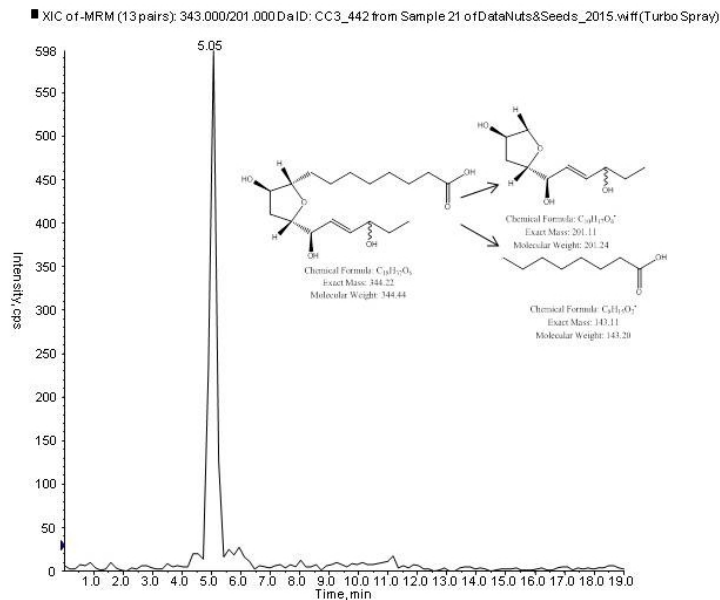
(A) α -Linolenic acid**(B) *ent*-16-(*RS*)-13-*epi*-ST- Δ^{14} -9-PhytoF**

Figure 1. Mass ion transition of pure (A) α -Linolenic acid (m/z 277 \rightarrow 233) and synthesized (B) *ent*-16-(*RS*)-13-*epi*-ST- Δ^{14} -9-PhytoF (m/z 343 \rightarrow 201) standard. The precursor and fragmentation ions were used to detect and quantify α -Linolenic acid (ALA) and *ent*-16-(*RS*)-13-*epi*-ST- Δ^{14} -9-PhytoF by the LC-MS/MS in lipid extract of nuts and seeds. Typical chromatograms of ALA and *ent*-16-(*RS*)-13-*epi*-ST- Δ^{14} -9-PhytoF elucidated in lipid extract of flaxseed are depicted.

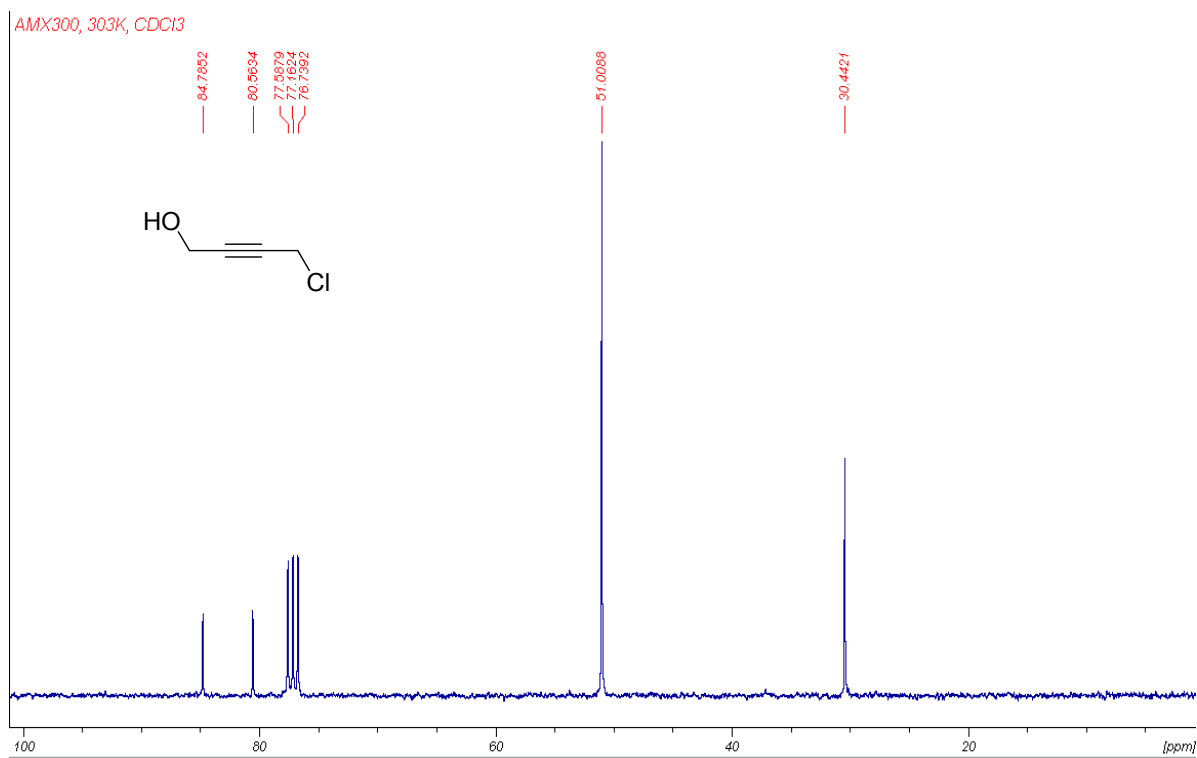
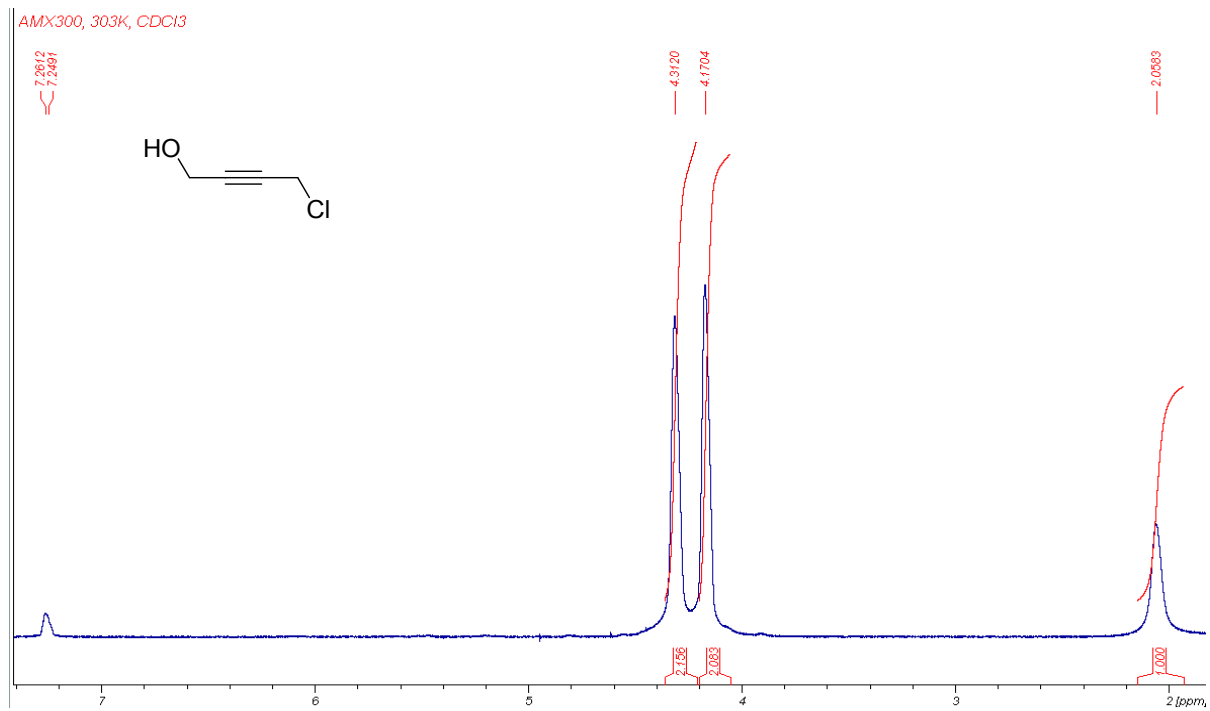
Table 1. Multiple reaction monitoring transition measured by the LC-MS/MS of the internal standards commercially available (α -Linolenic acid, α -Linolenic acid-d₁₄, 15-F_{2t}-IsoP-d₄) and 16(*R,S*)-13-*epi*-ST- Δ^{14} -9-PhytoF synthesized in the study.

<i>Analyte</i>	<i>Q1 m/z</i>	<i>Q3 m/z</i>	<i>CE</i>	<i>DP</i>
α -Linolenic acid	277	233	-50	-16
α -Linolenic acid-d ₁₄	291	247	-50	-26
15-F _{2t} -IsoP-d ₄	357	197	-50	-25
16(<i>R,S</i>)-13- <i>epi</i> -ST- Δ^{14} -9-PhytoF	343	201	-55	-28

Q1 *m/z*: precursor ion; Q3 *m/z*: fragmented ion; DP: declustering potential; CE: collision energy

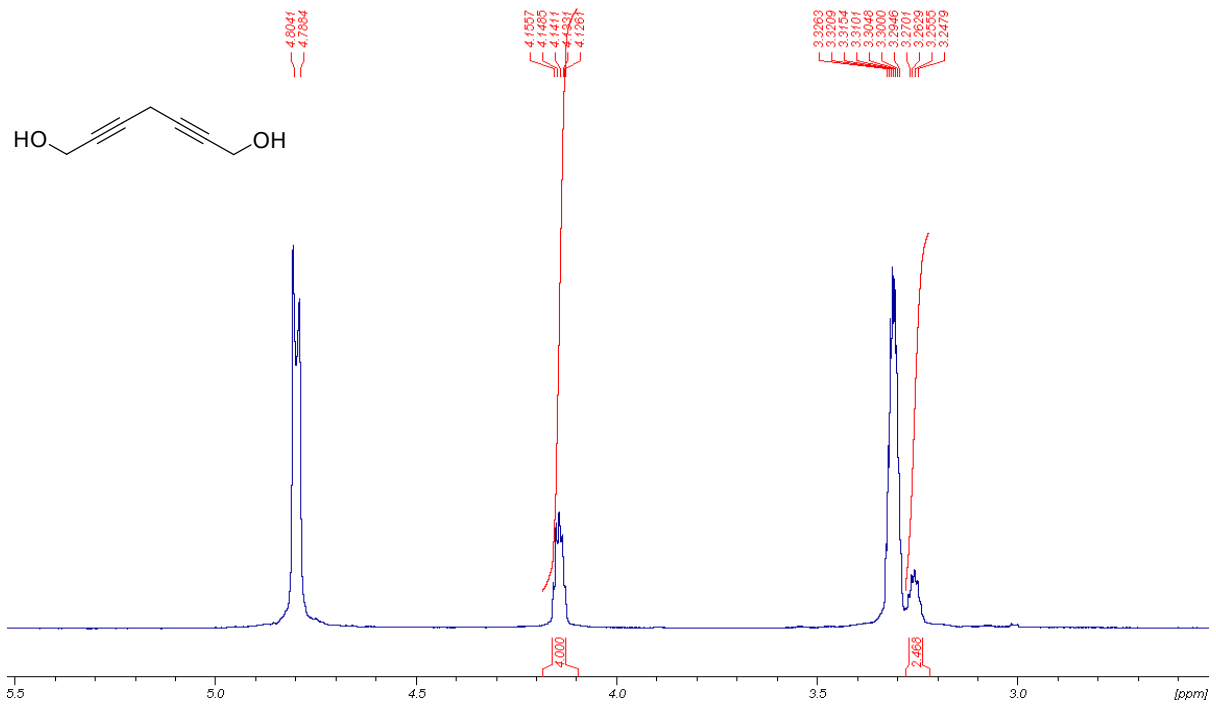
4. ^1H , ^{13}C and selected 2D NMR spectra

4-chlorobut-2-yn-1-ol:

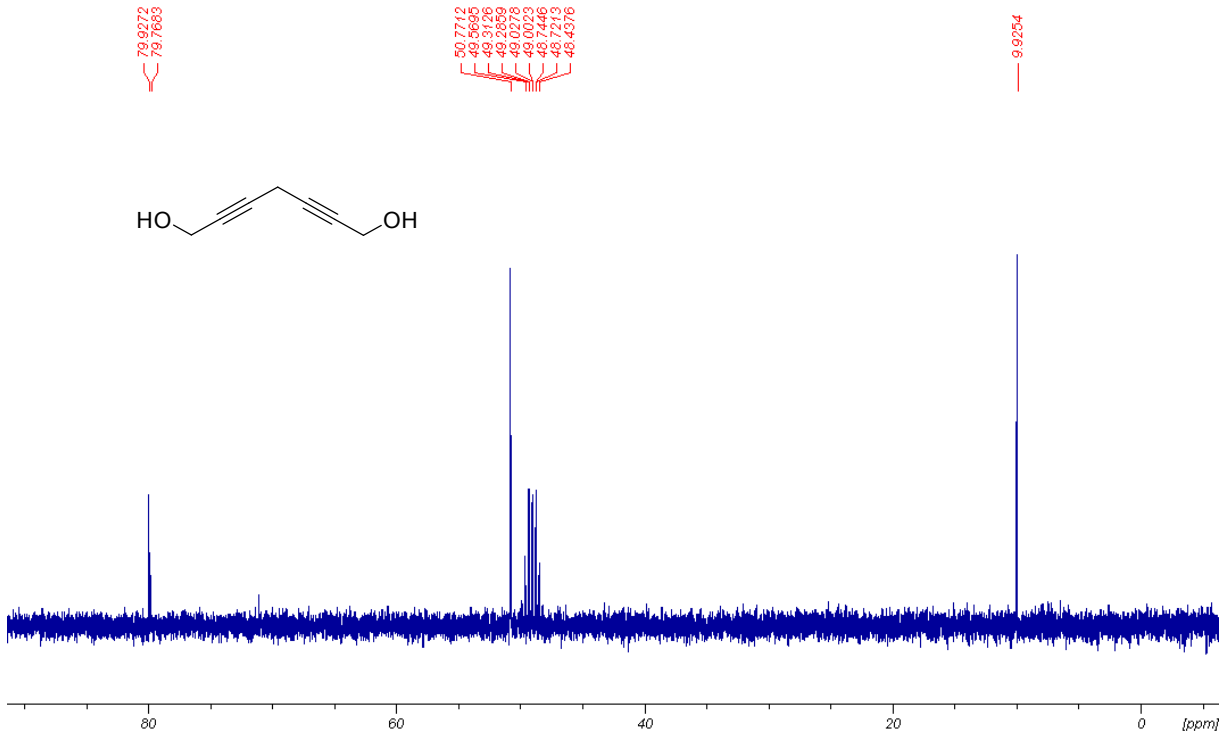


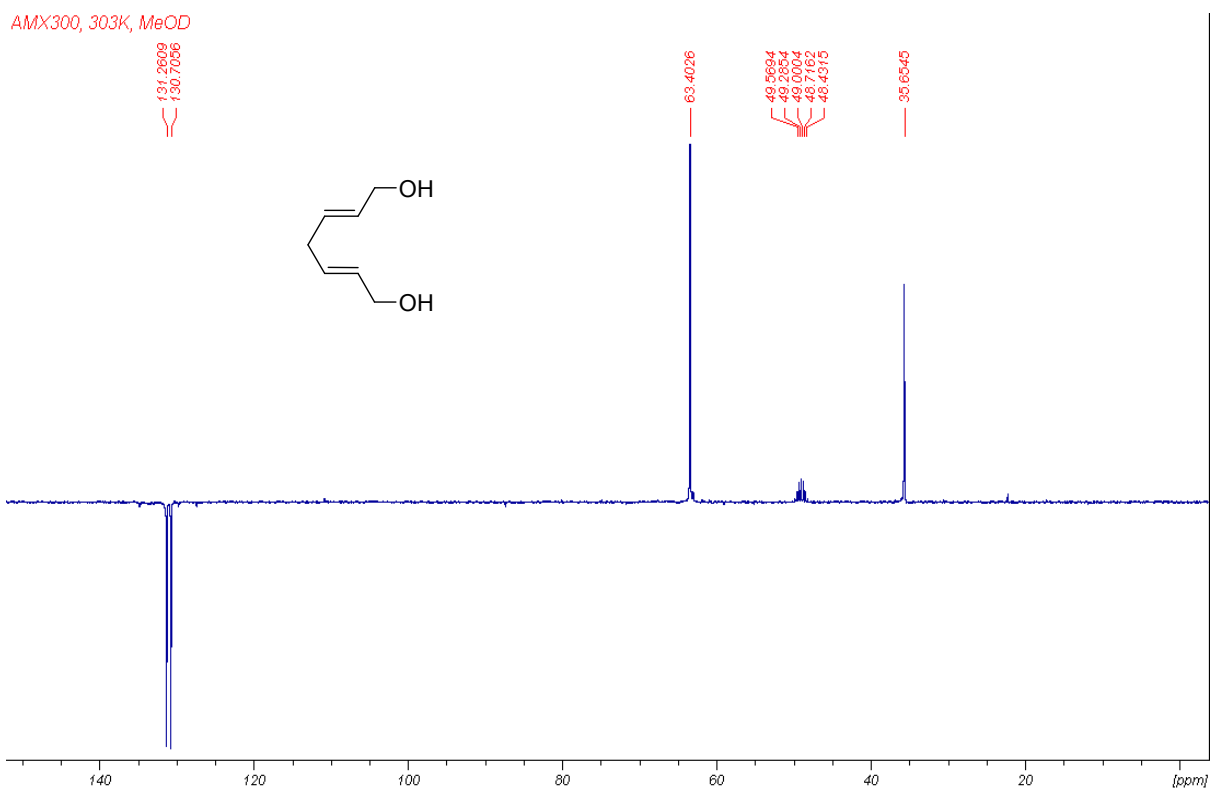
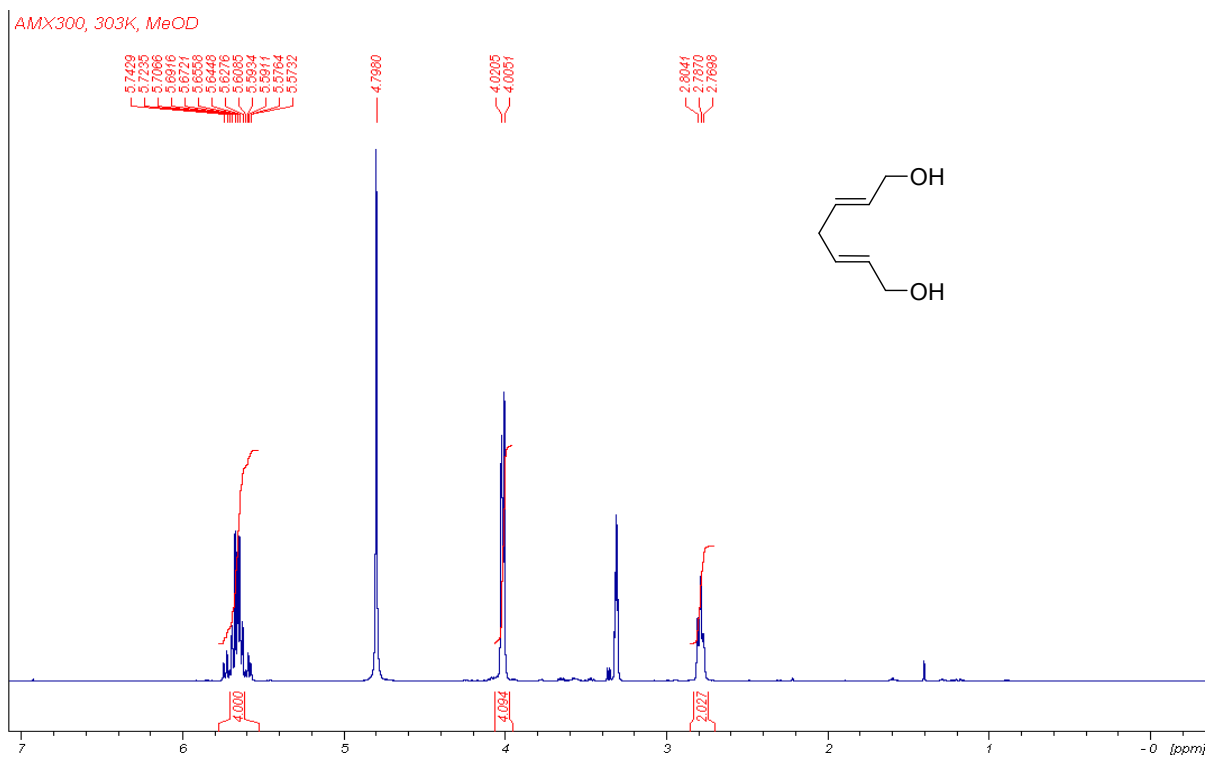
hepta-2,5-diyne-1,7-diol **4**:

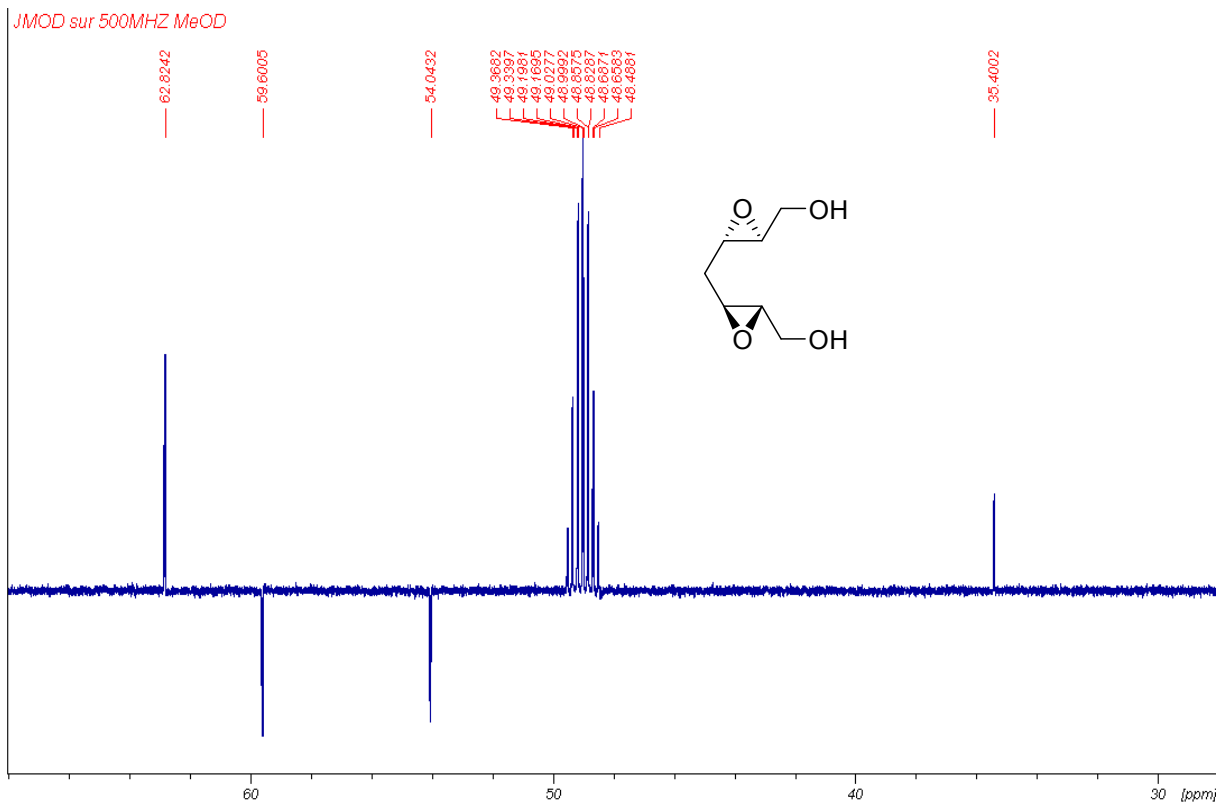
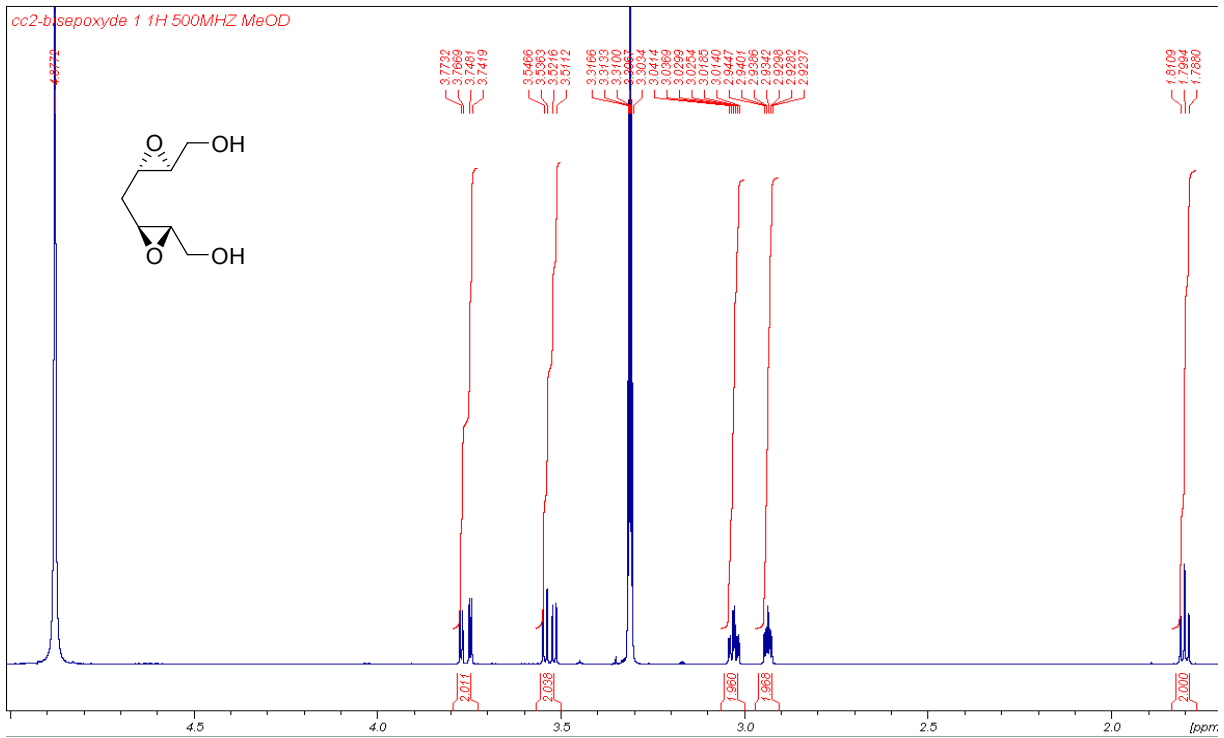
AMX300, 303K, MeOD



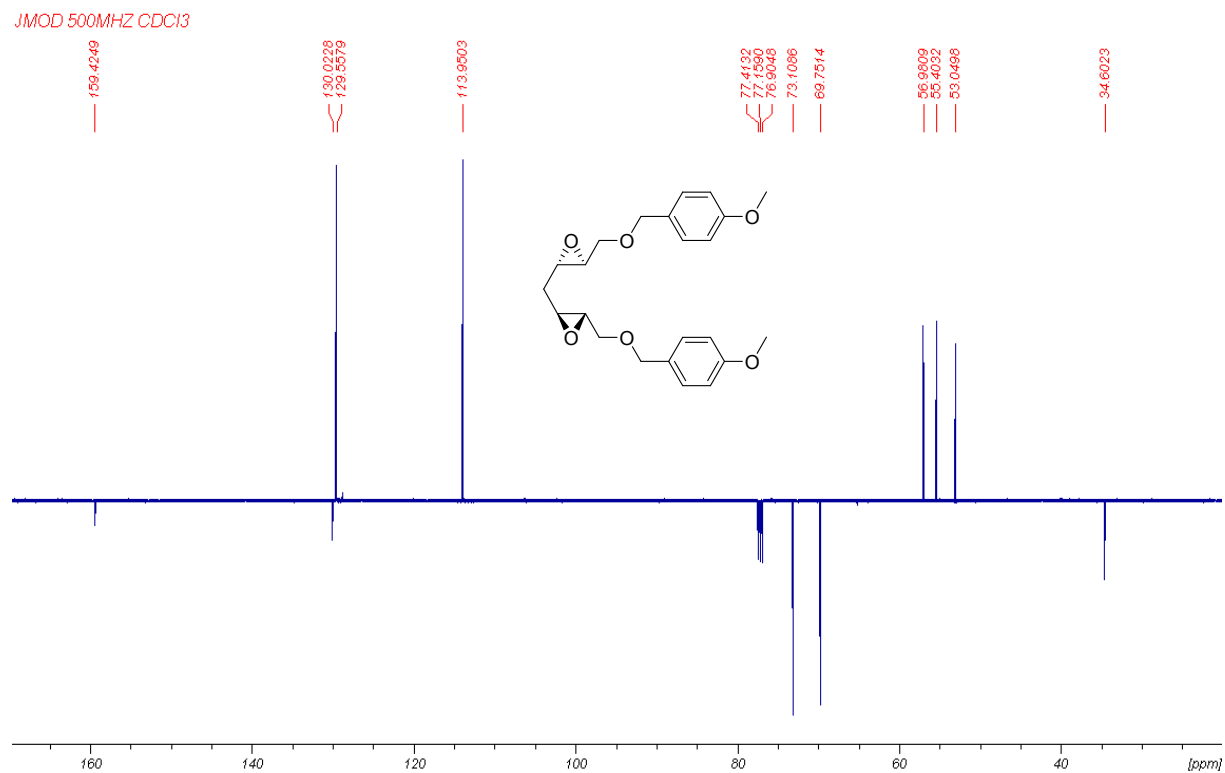
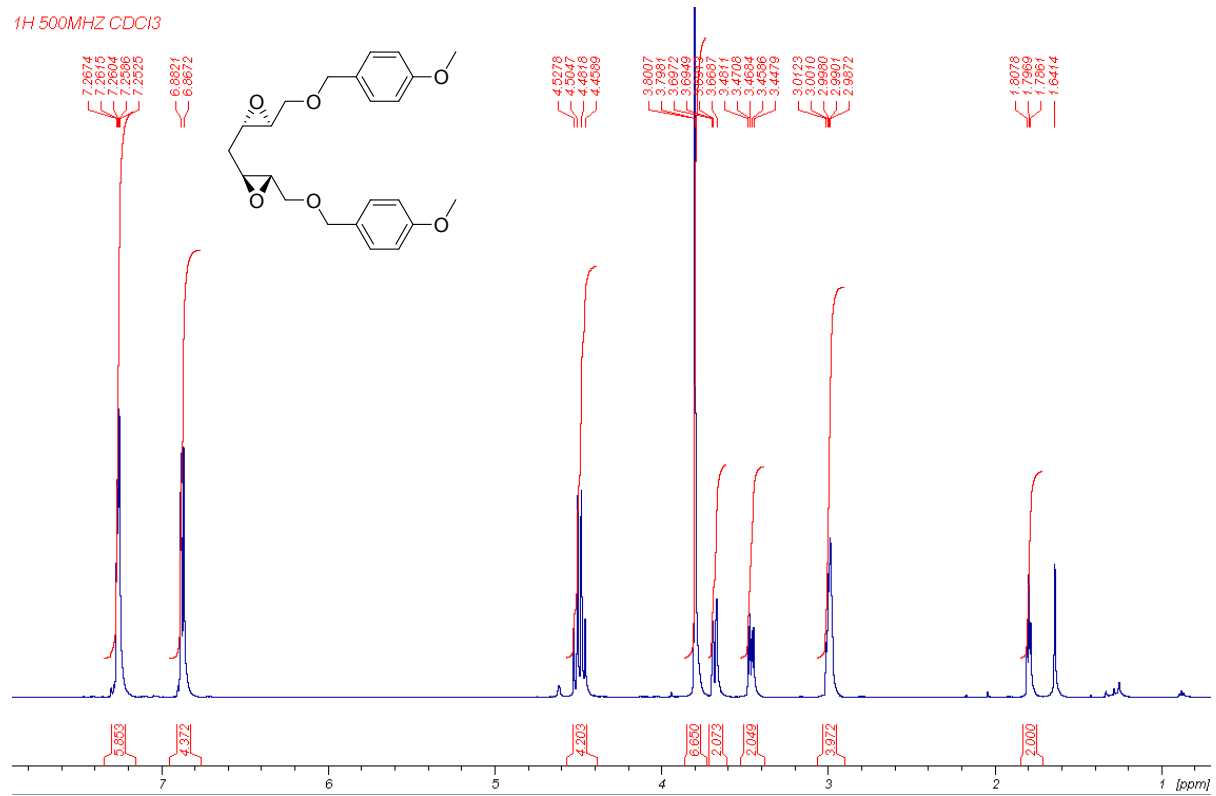
AMX300, 303K, MeOD

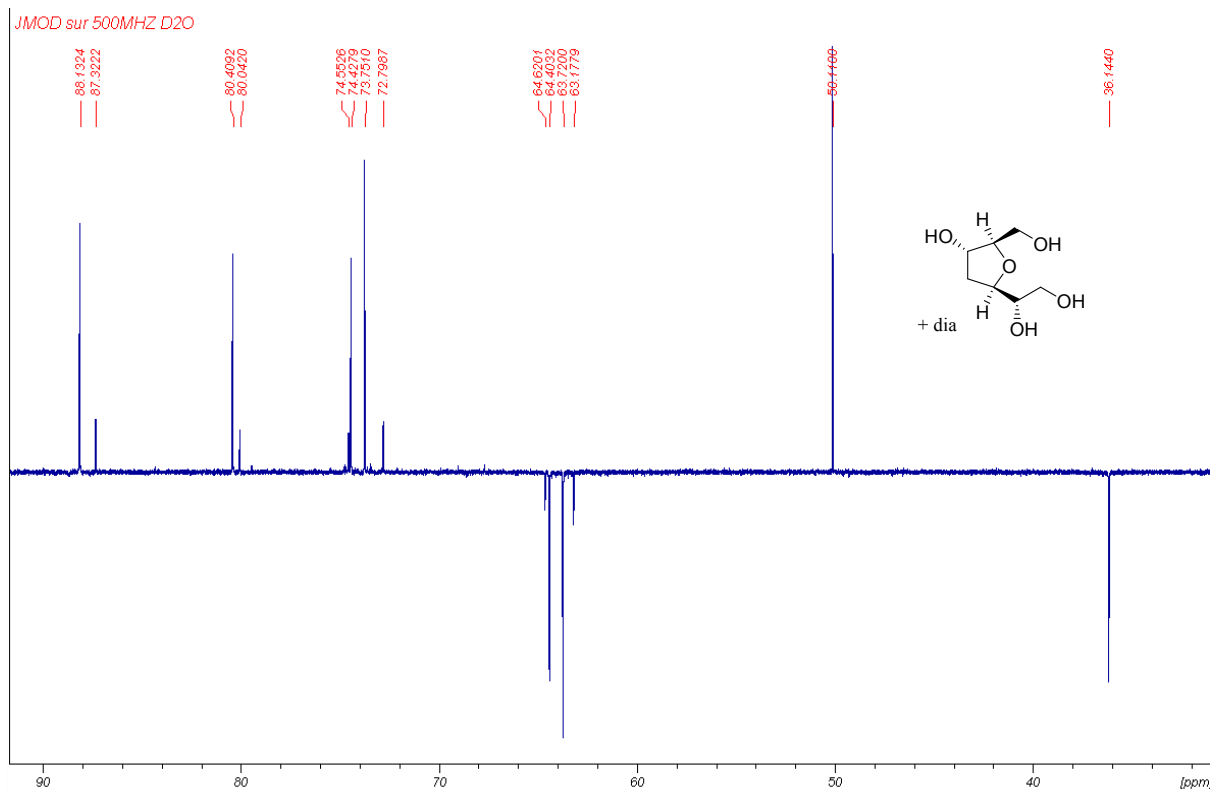
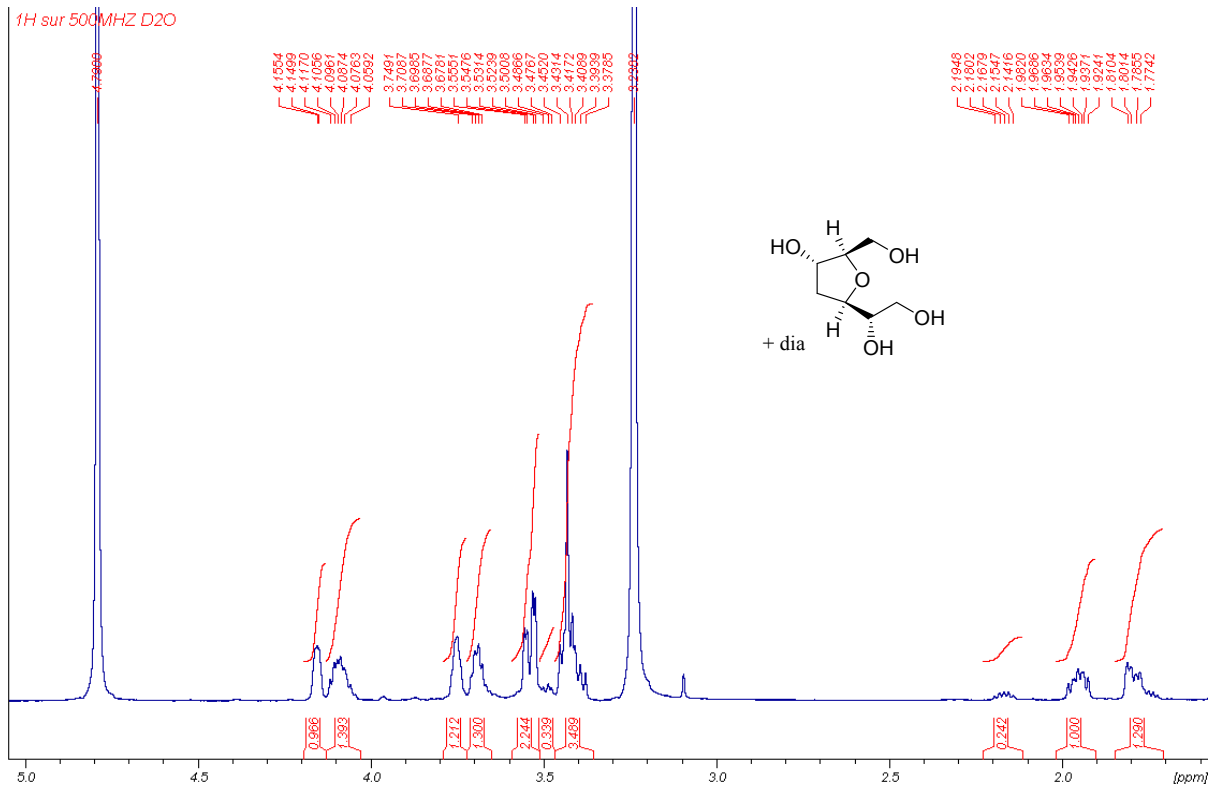


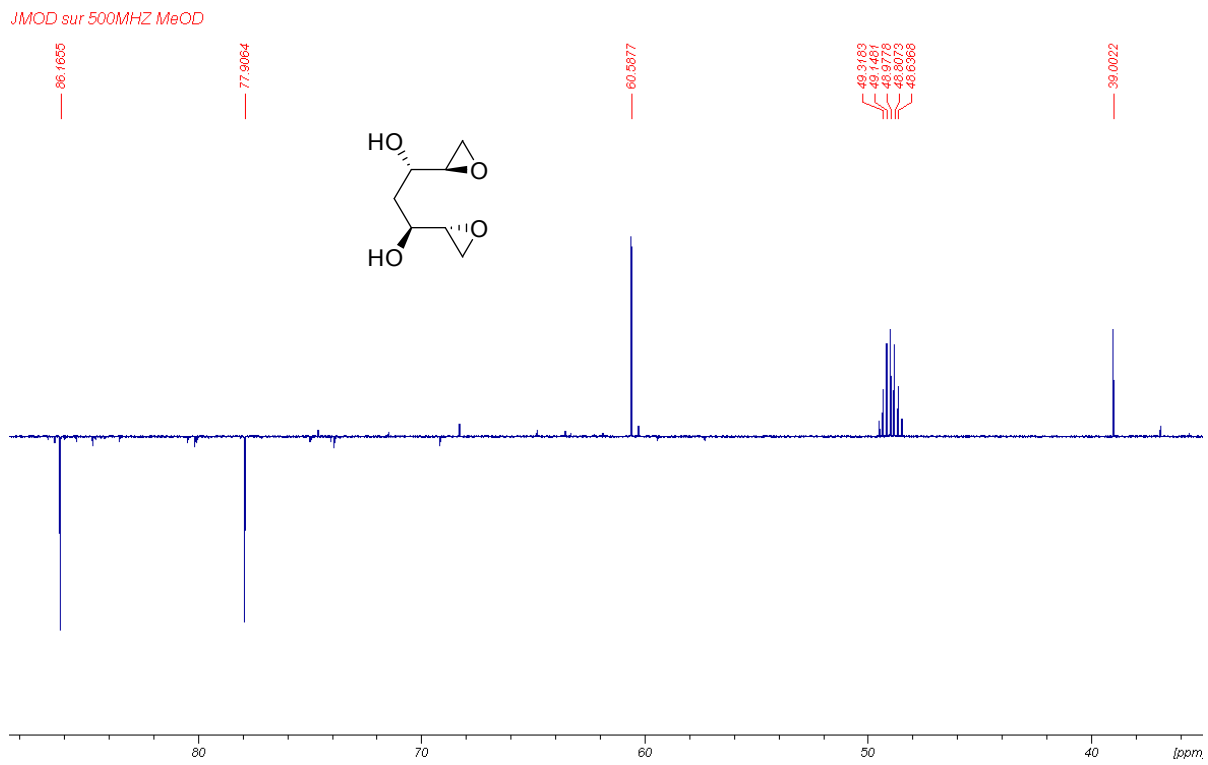
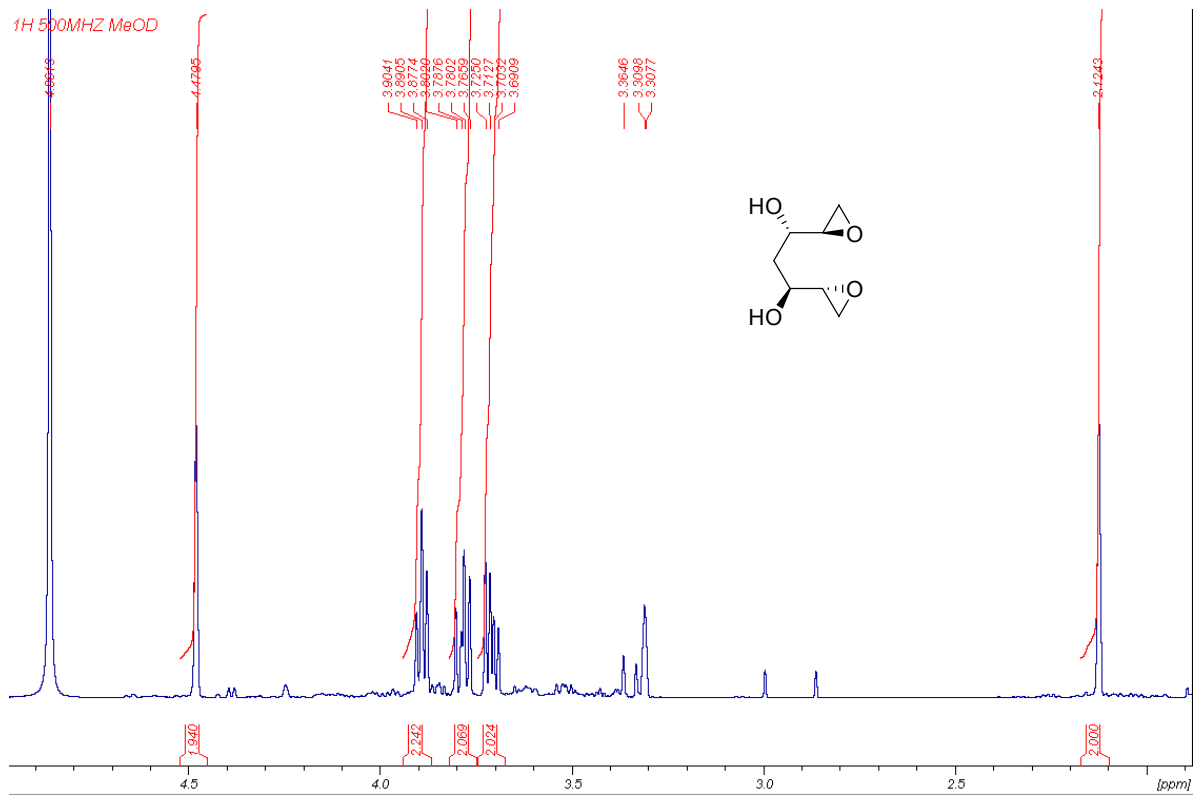
(E,E)-hepta-2,5-diene-1,7-diol:

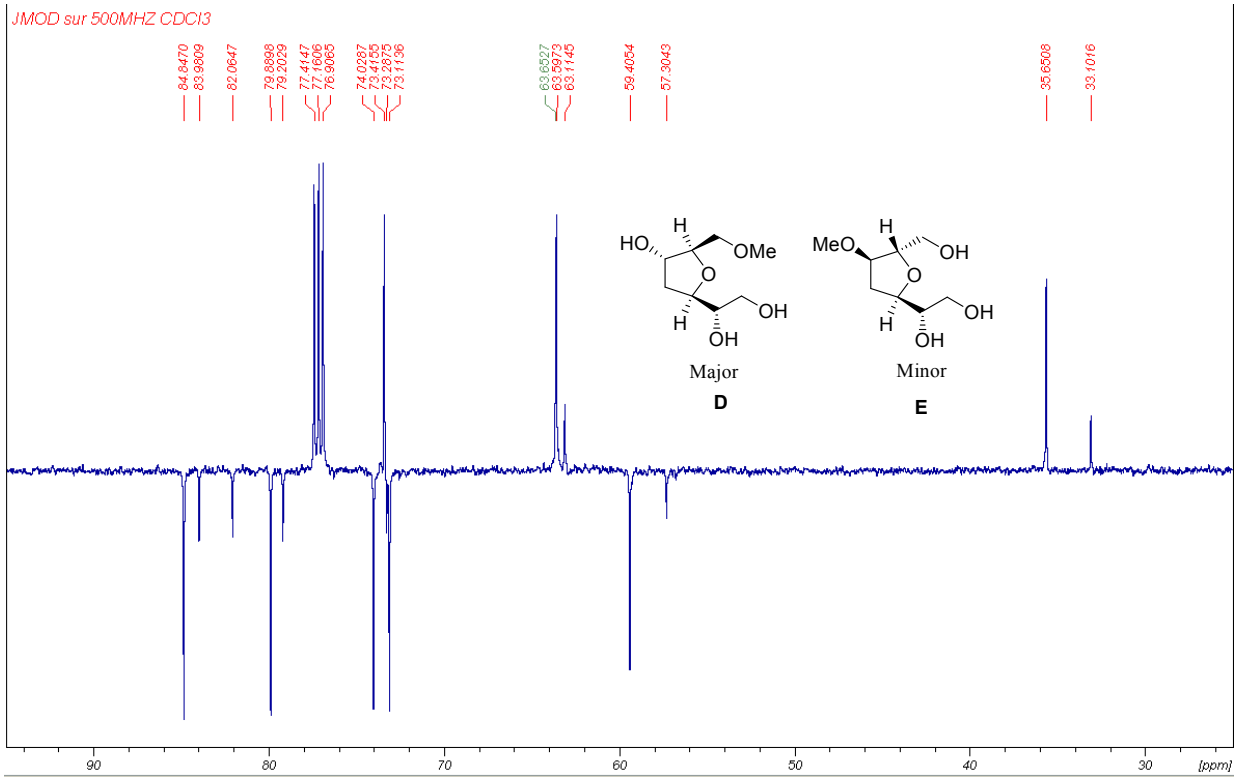
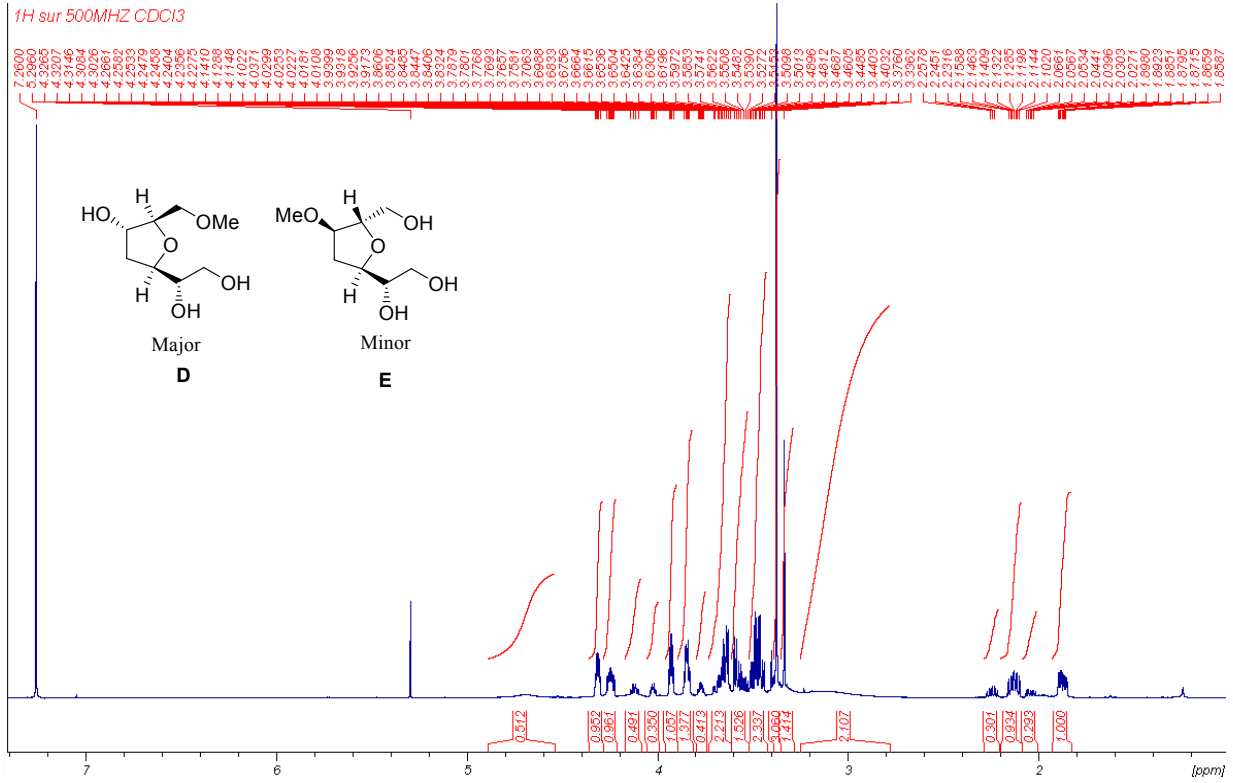
(2S,3S,5S,6S)-2,3:5,6-diepoxyheptane-1,7-diol **3**:

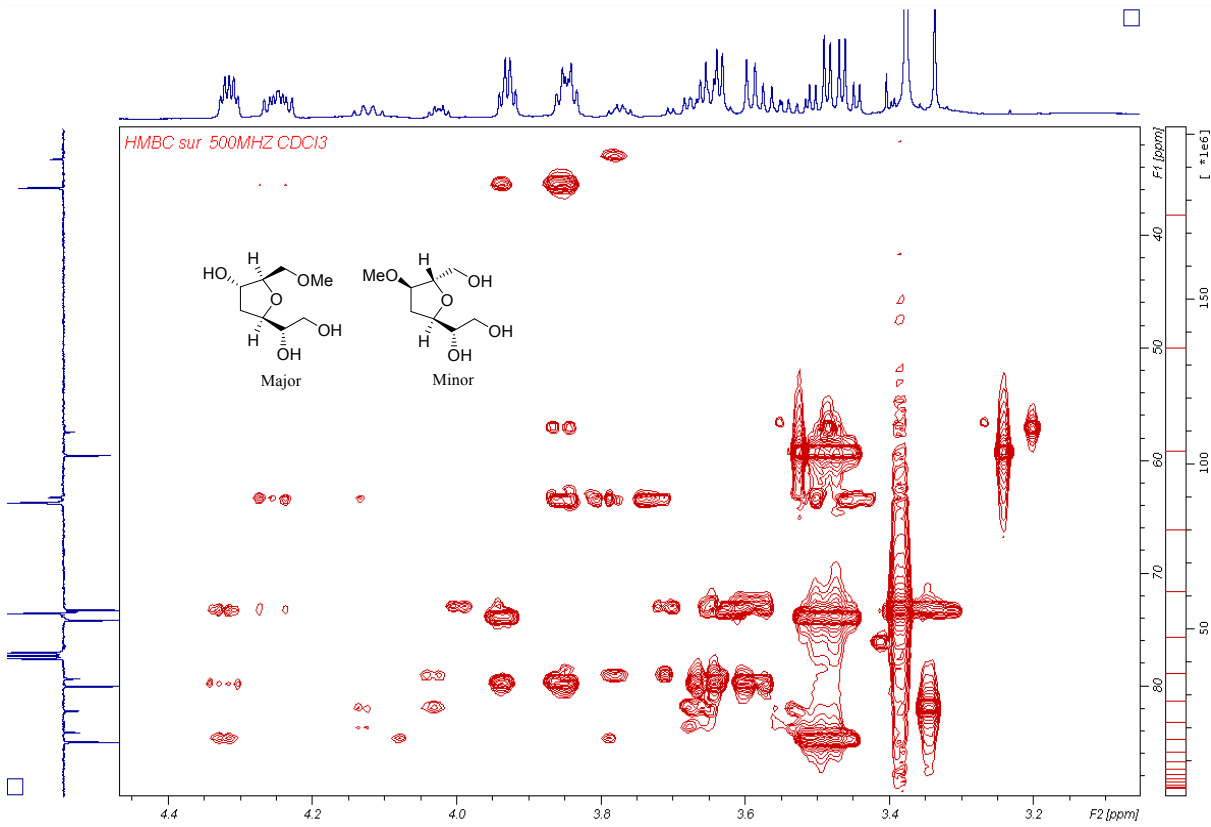
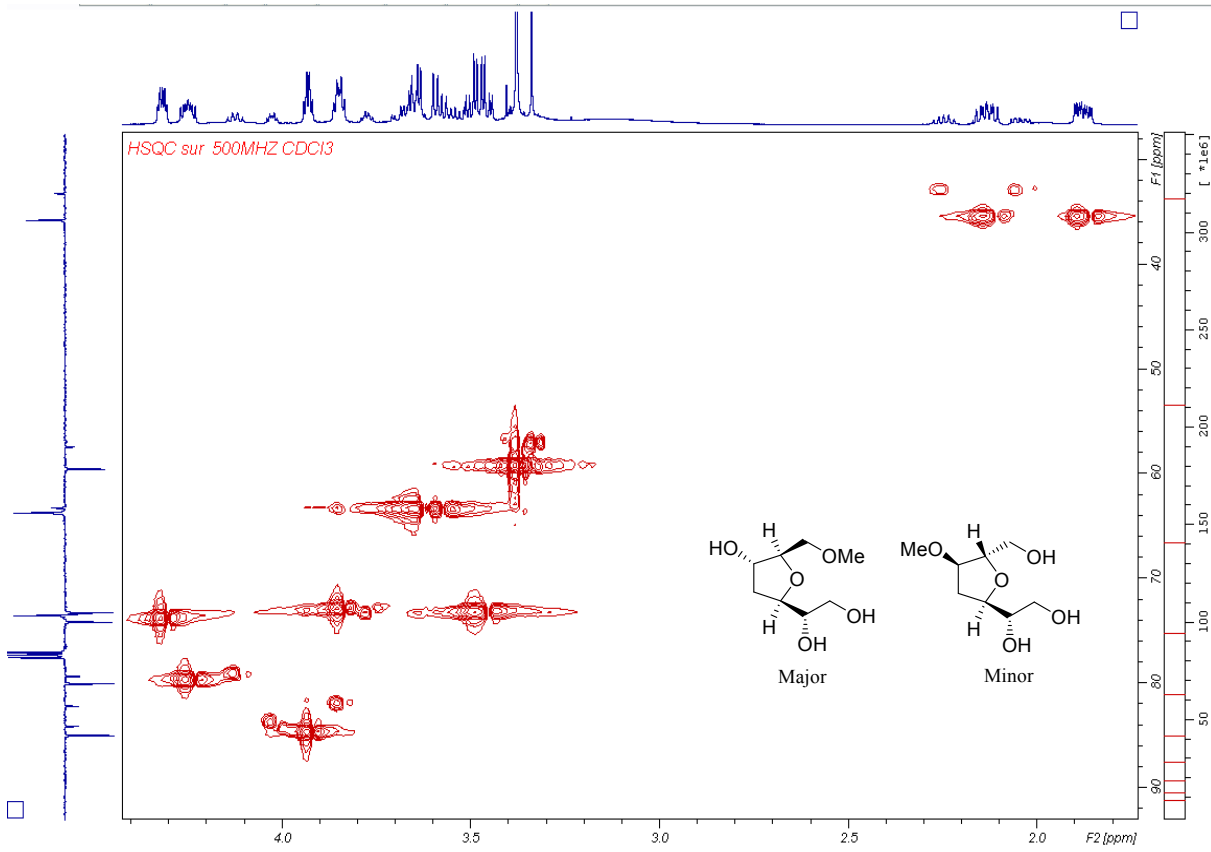
bis((2S,3S)-3-((4-methoxybenzyloxy)methyl)oxiran-2-yl)methane:

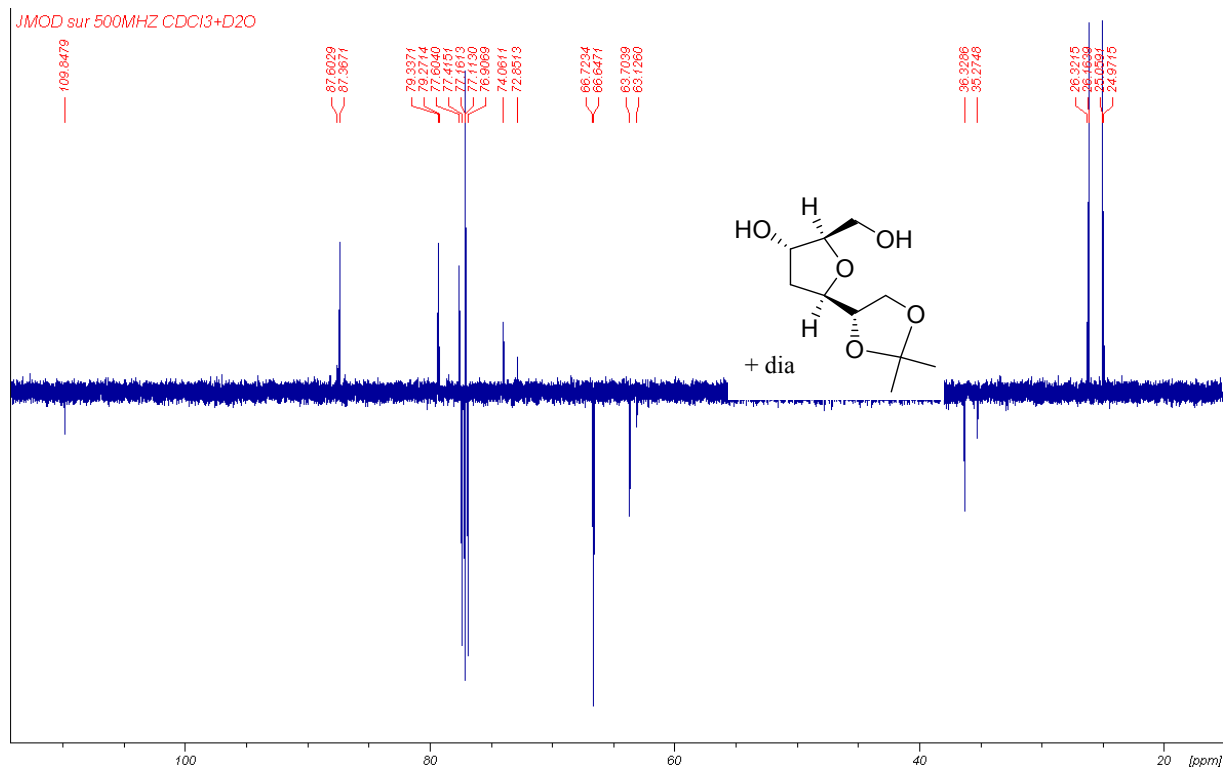
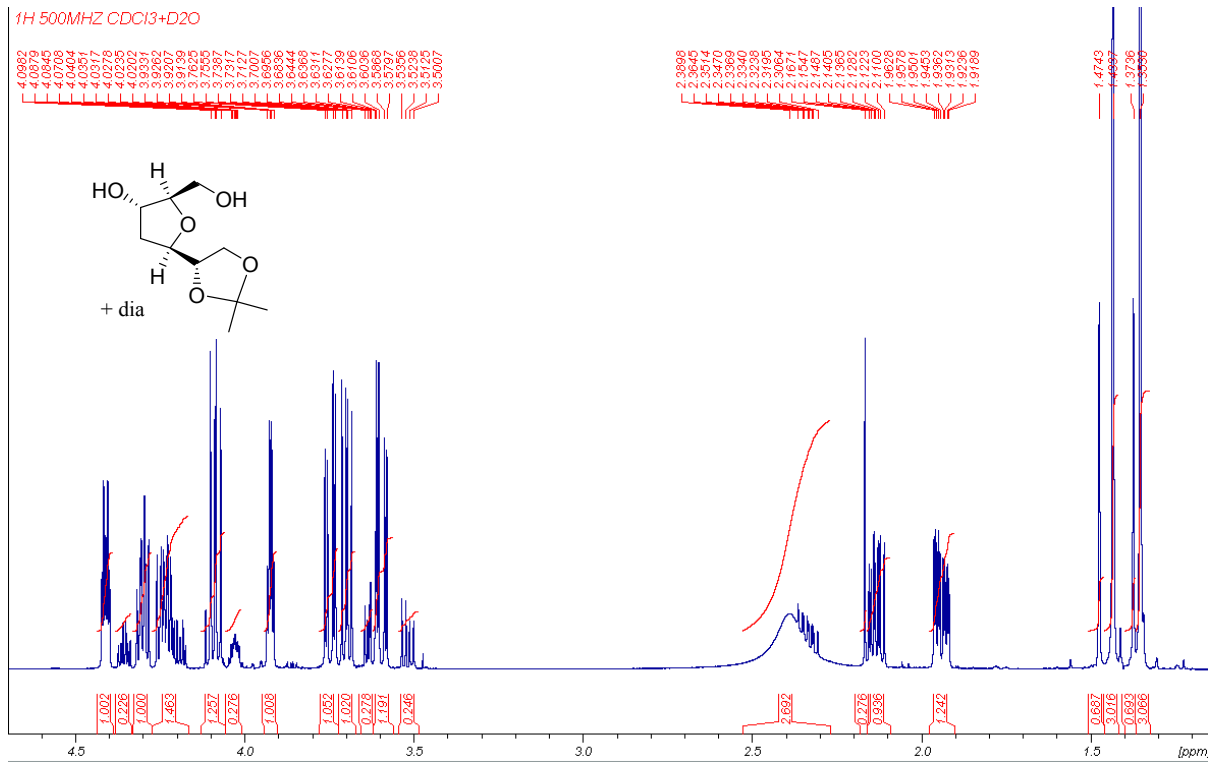


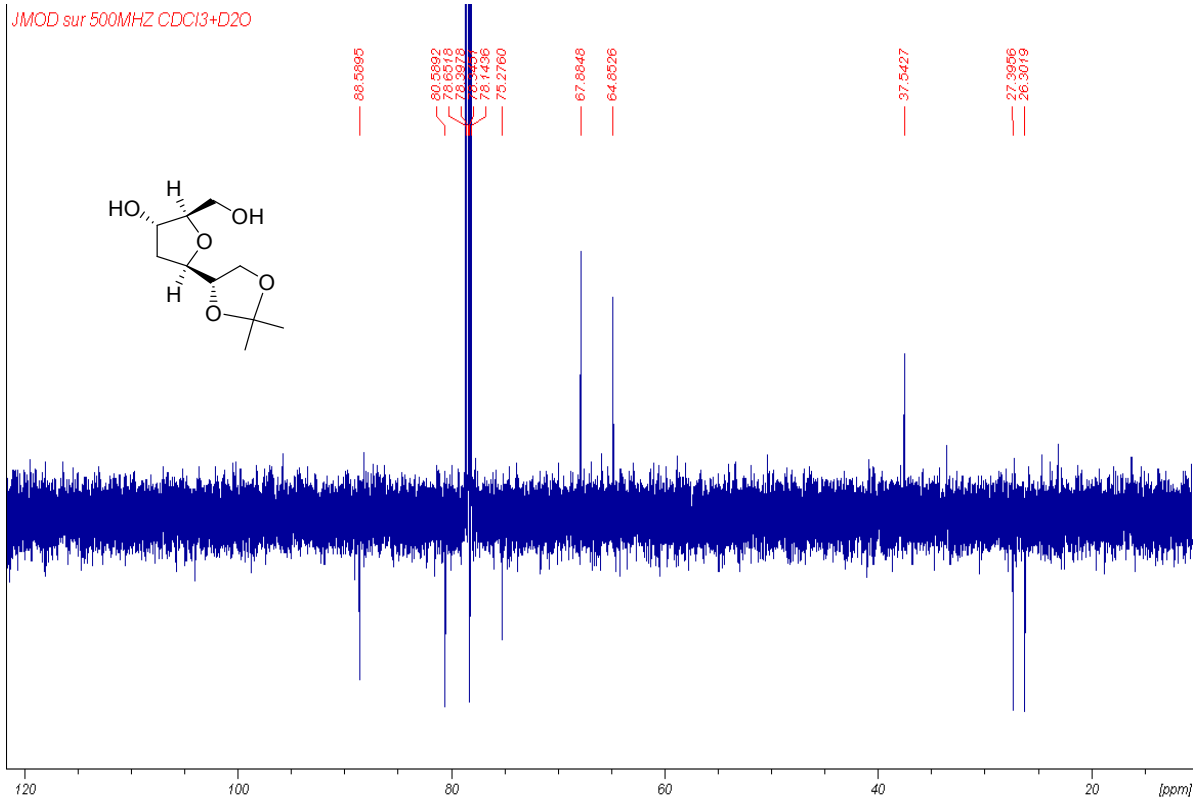
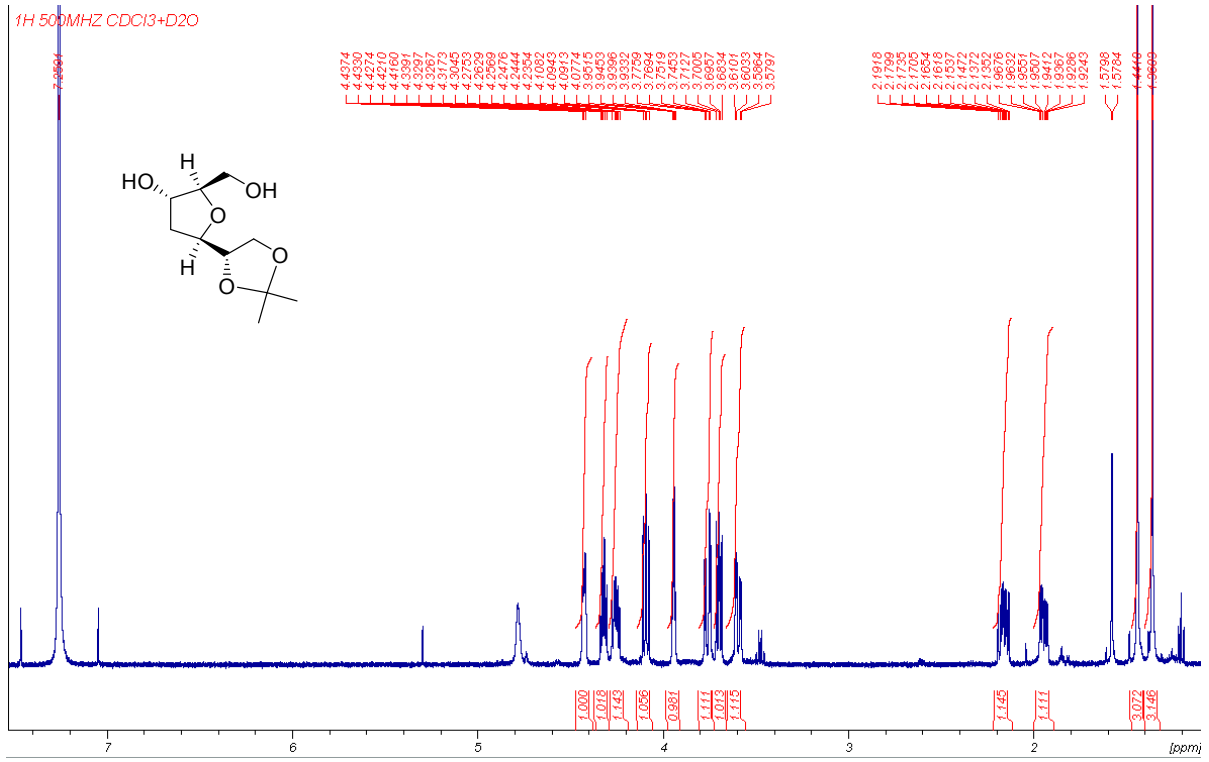
tetraol **2**:

Bisepoxy diol resulting of double-Payne rearrangement **B**:

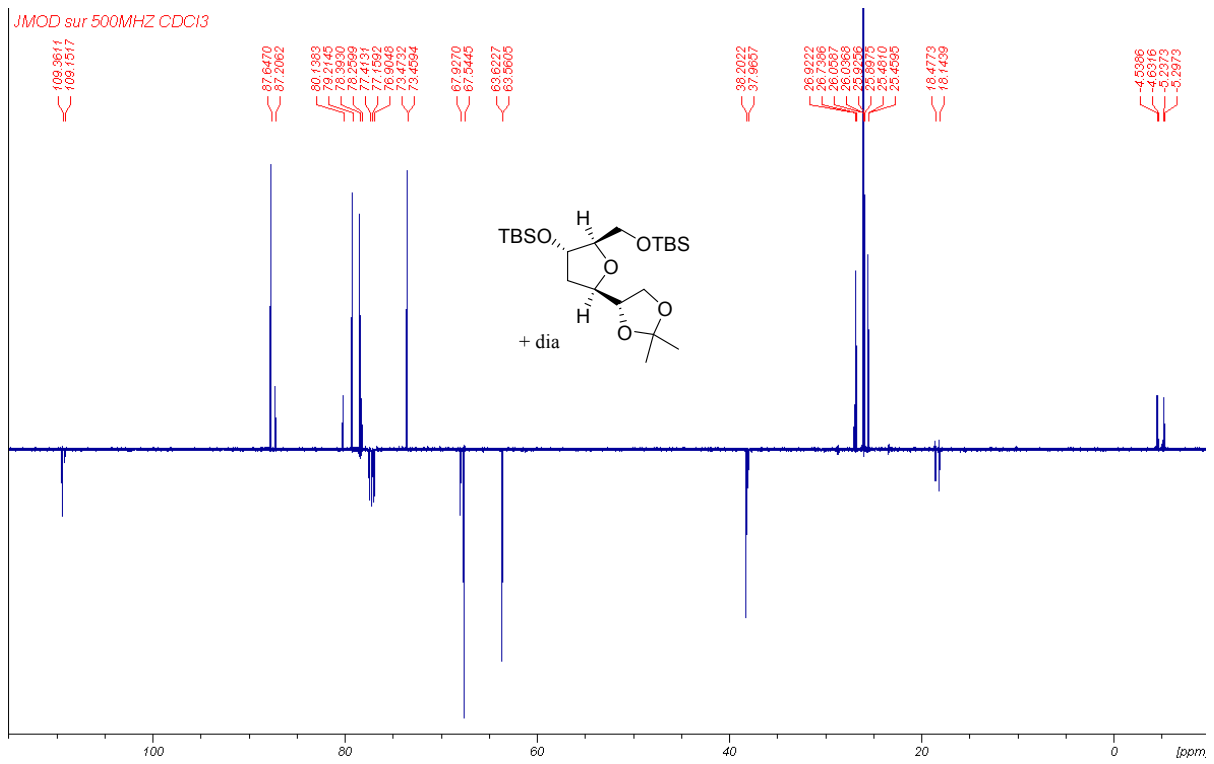
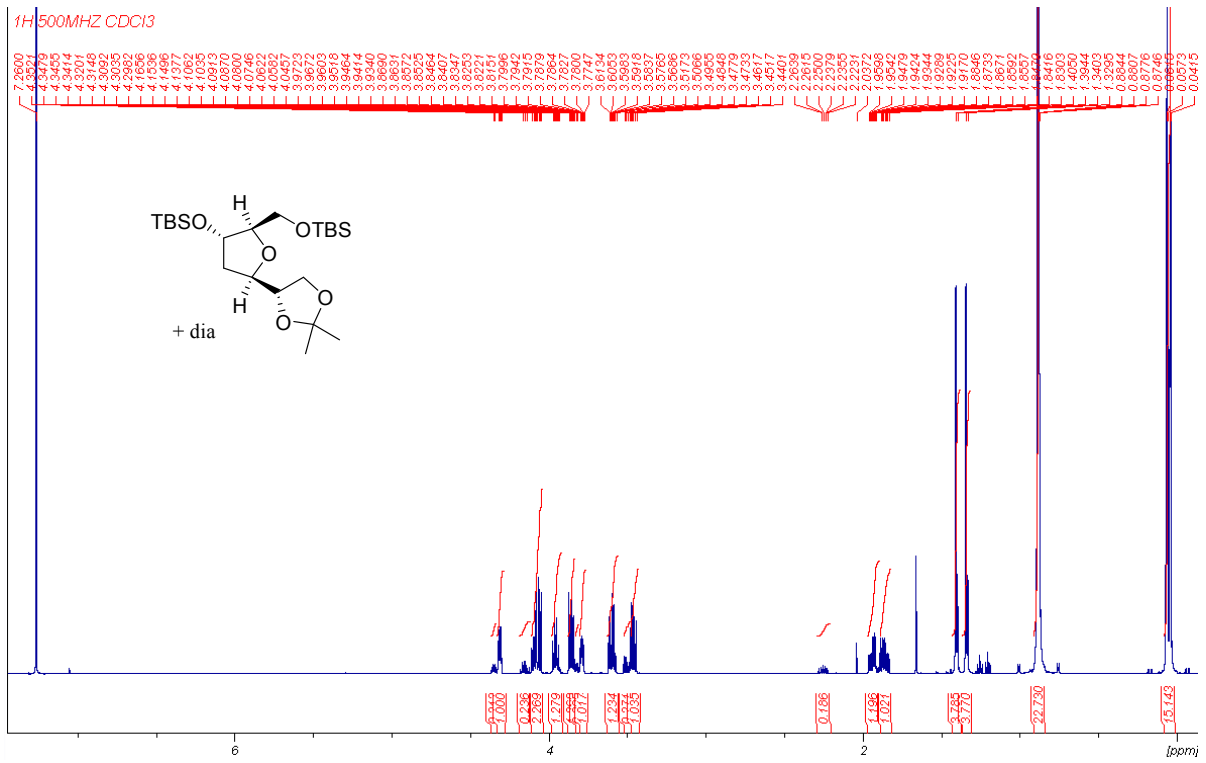
Methoxy-tetraol D and E:

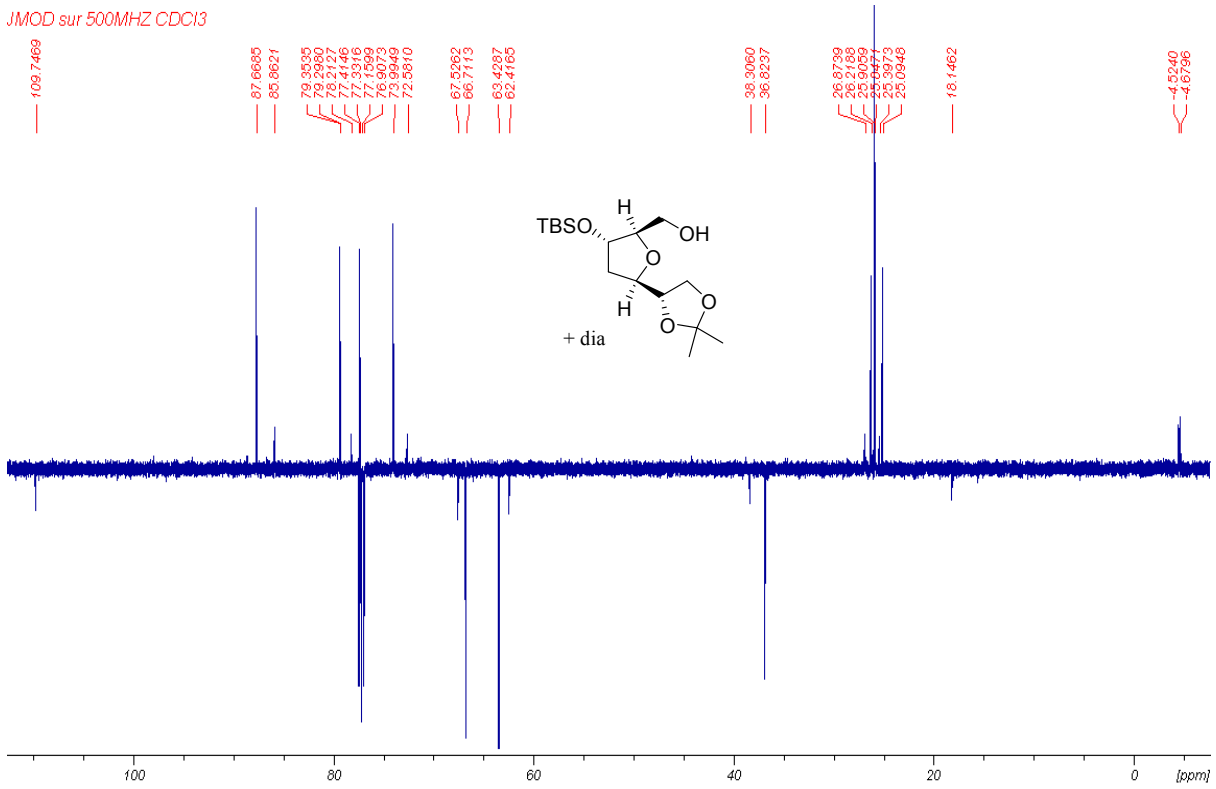
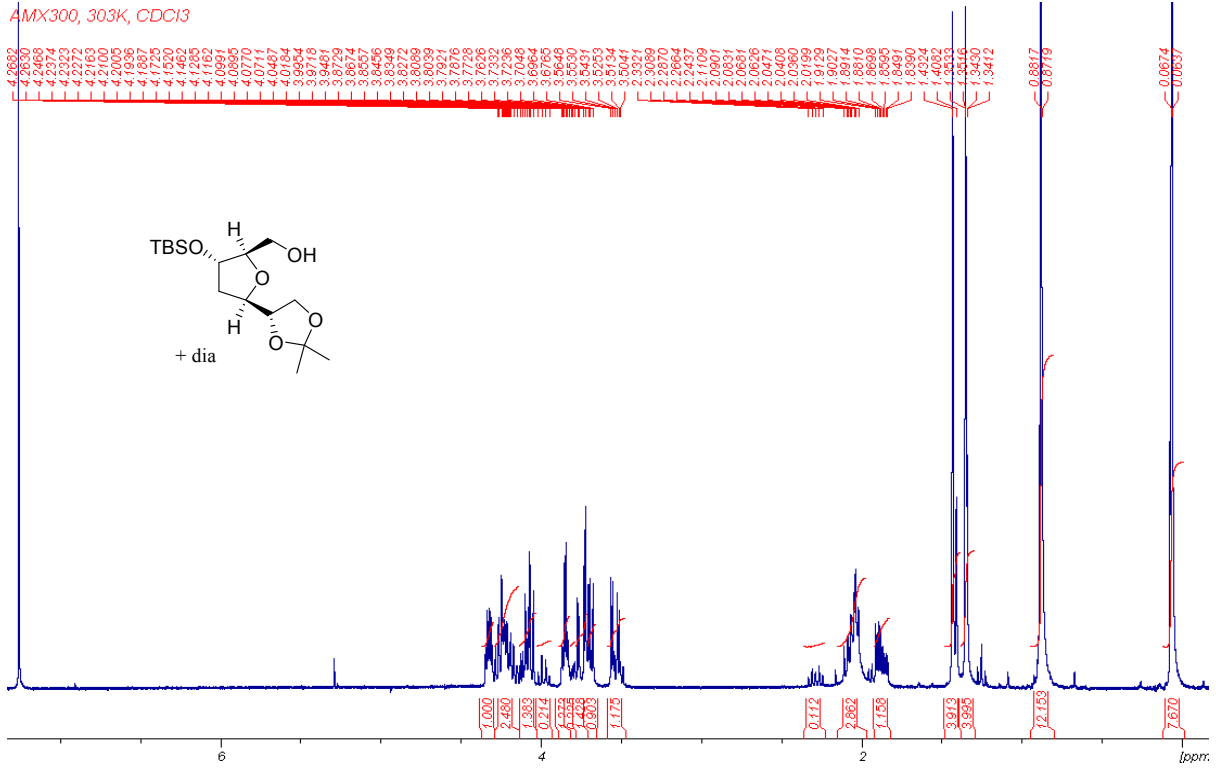


Acetonide **5**:

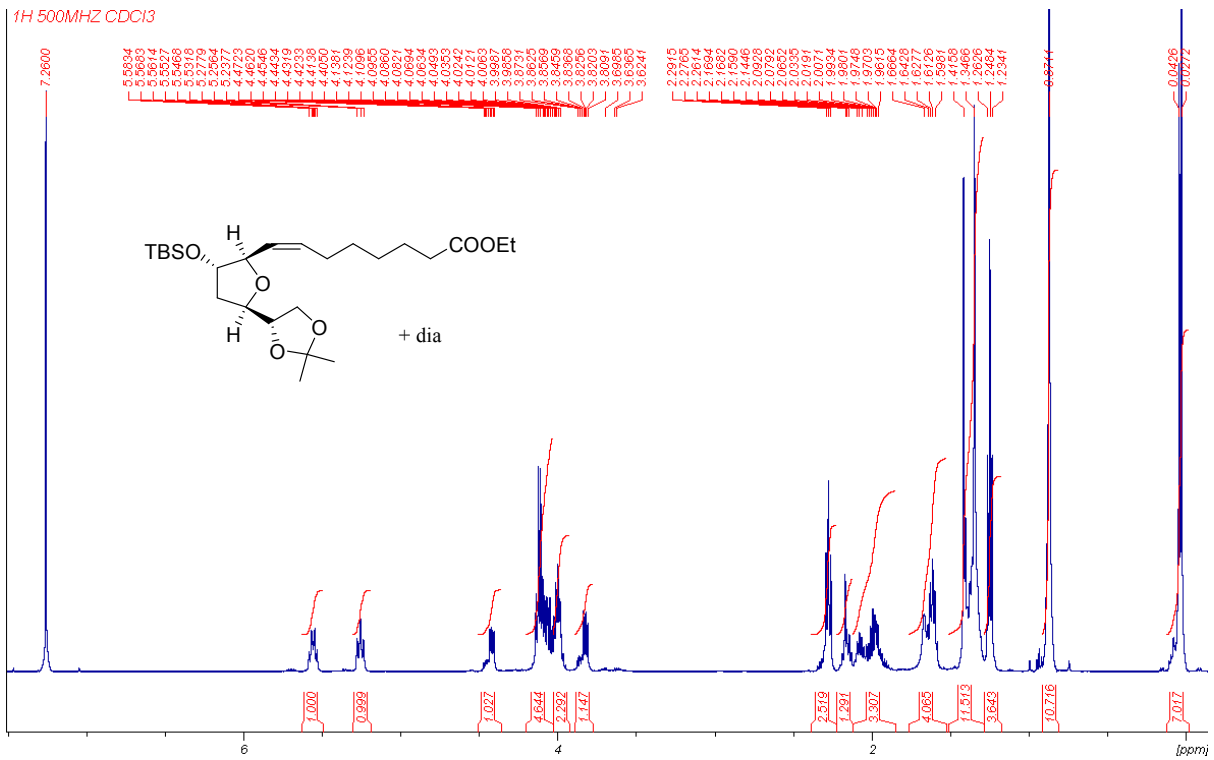
Crystals of acetonide **5**:

diTBS-acetonide:

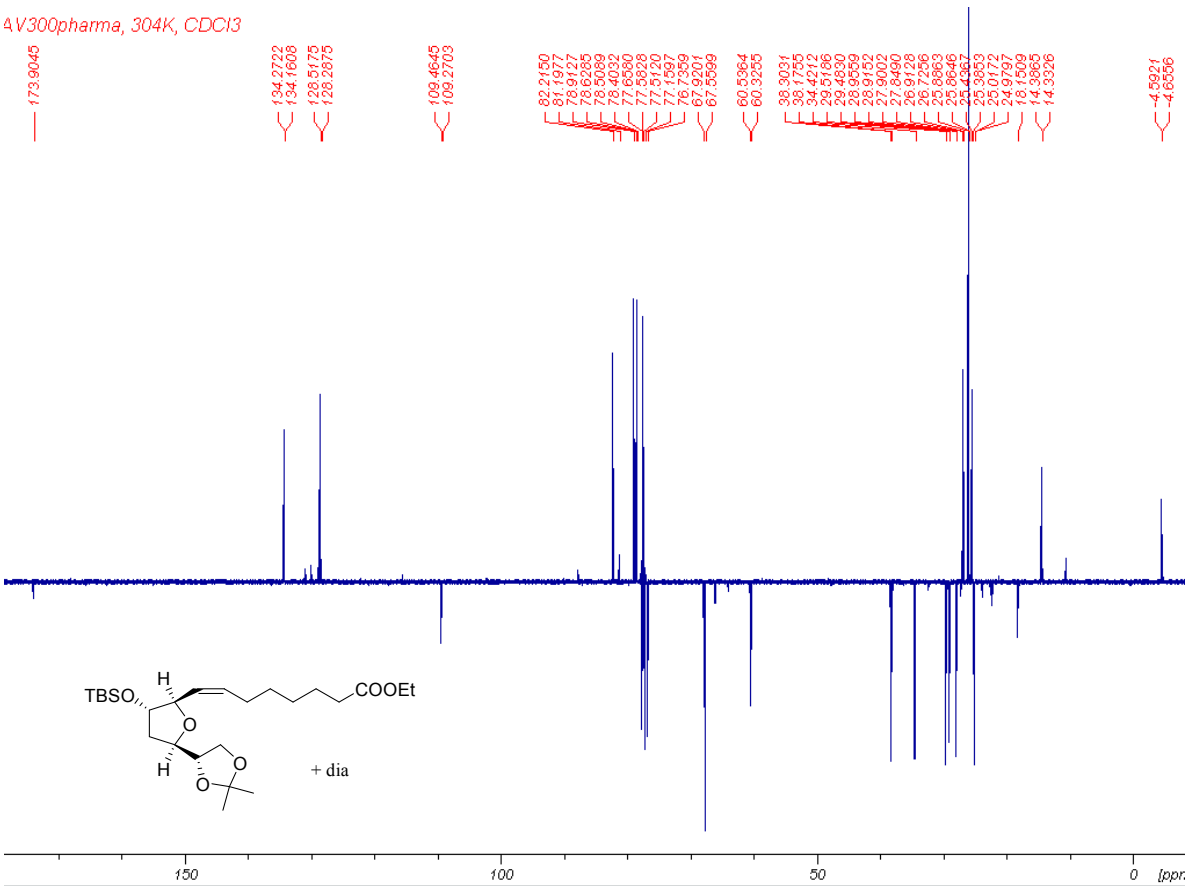


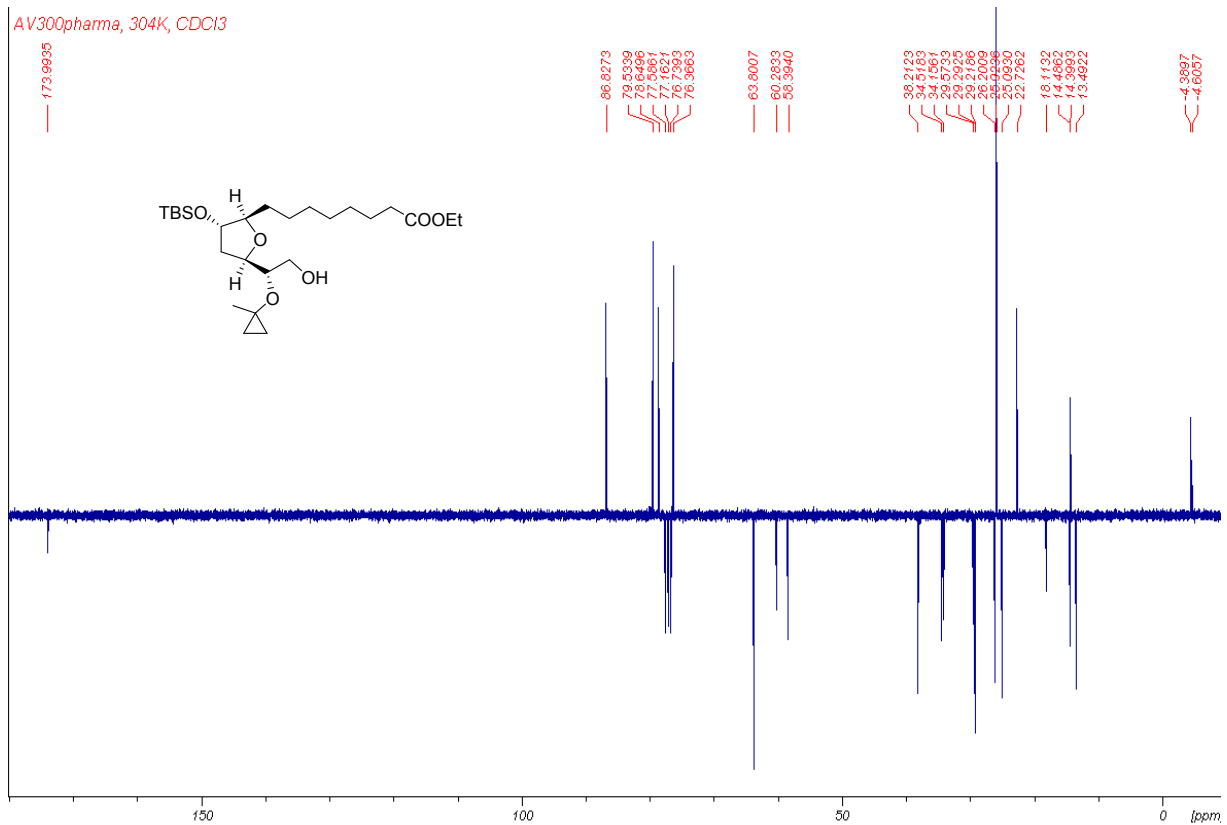
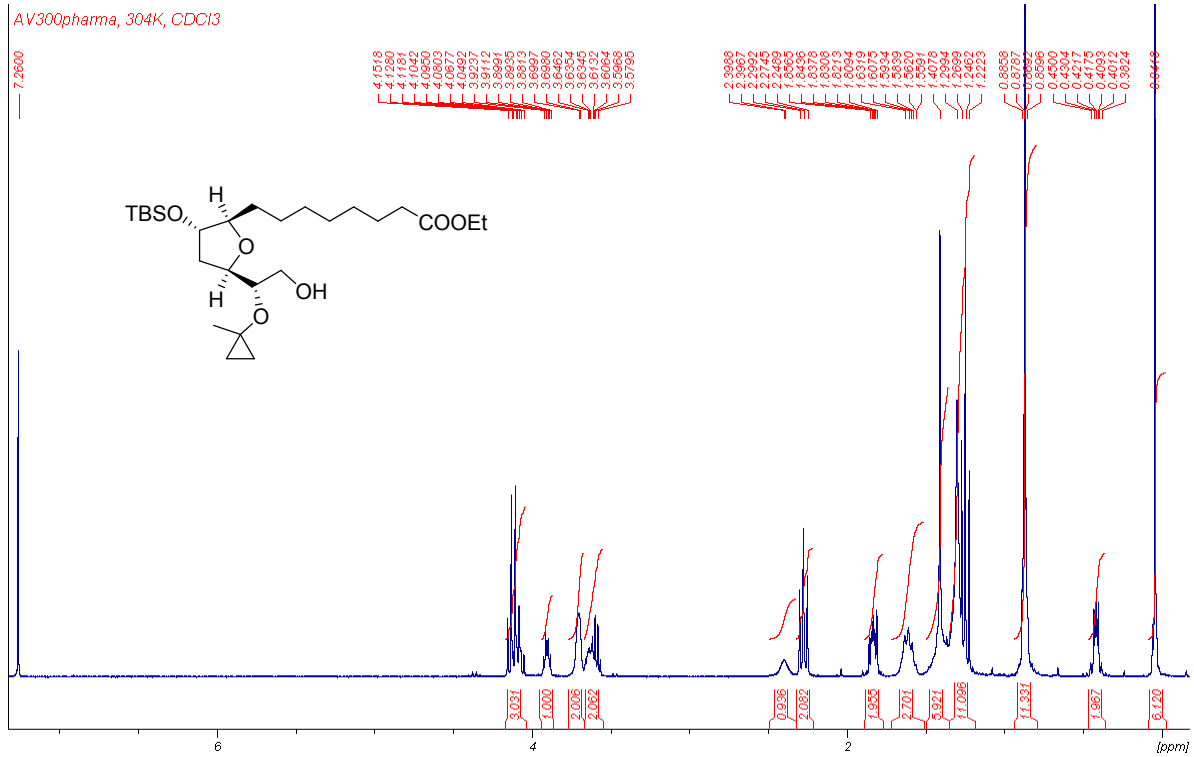
mono protected TBS-ether **6**:

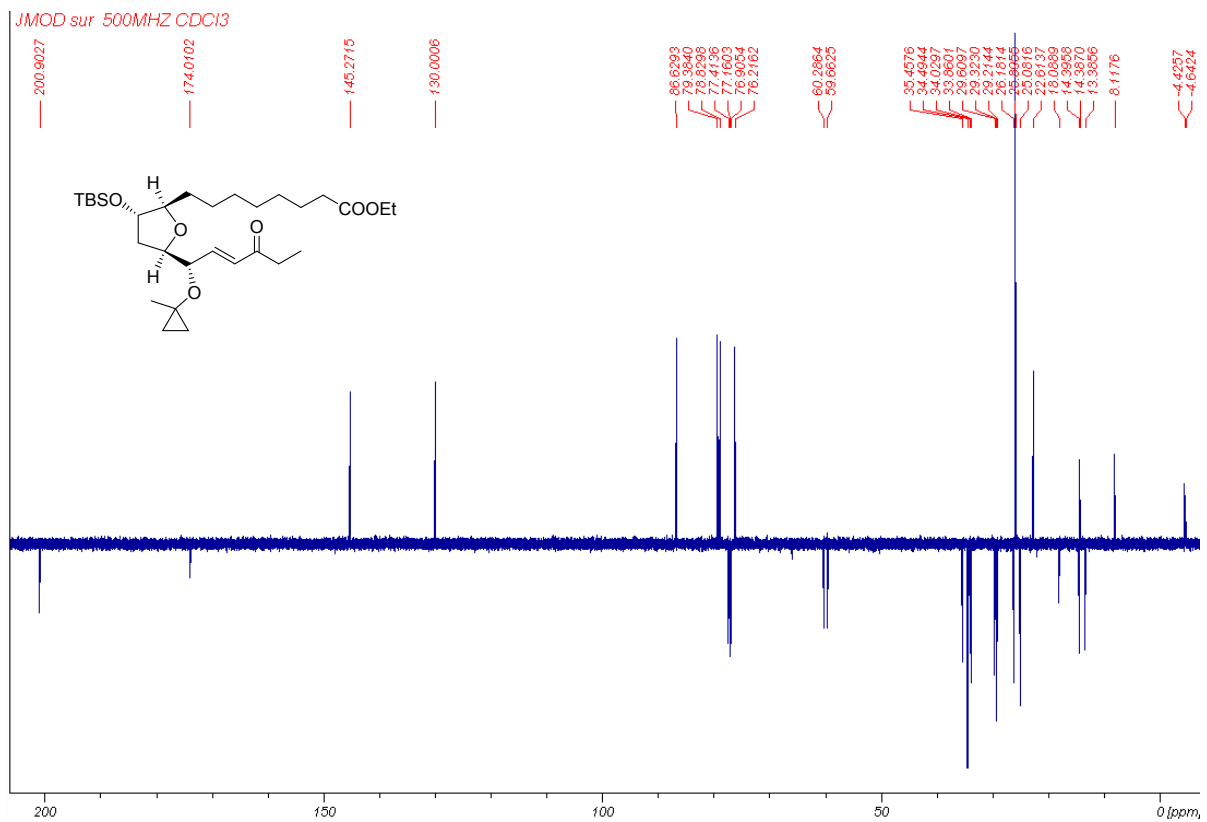
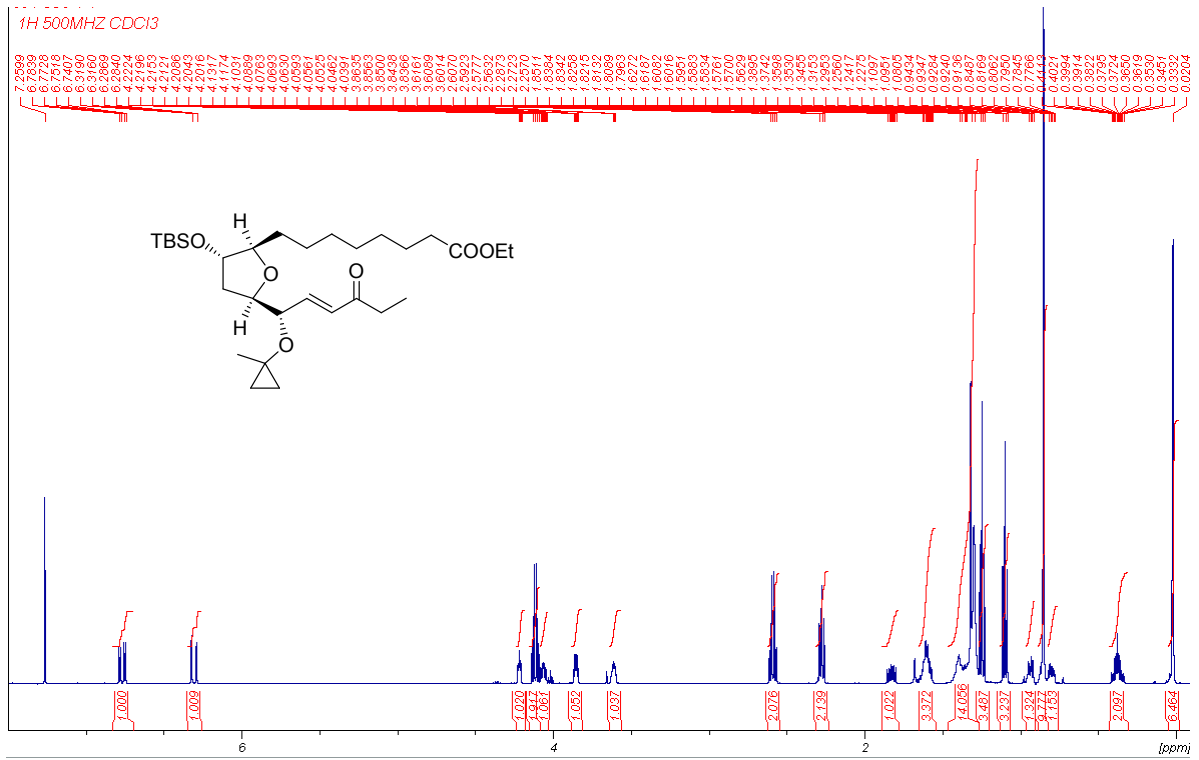
Alkene:

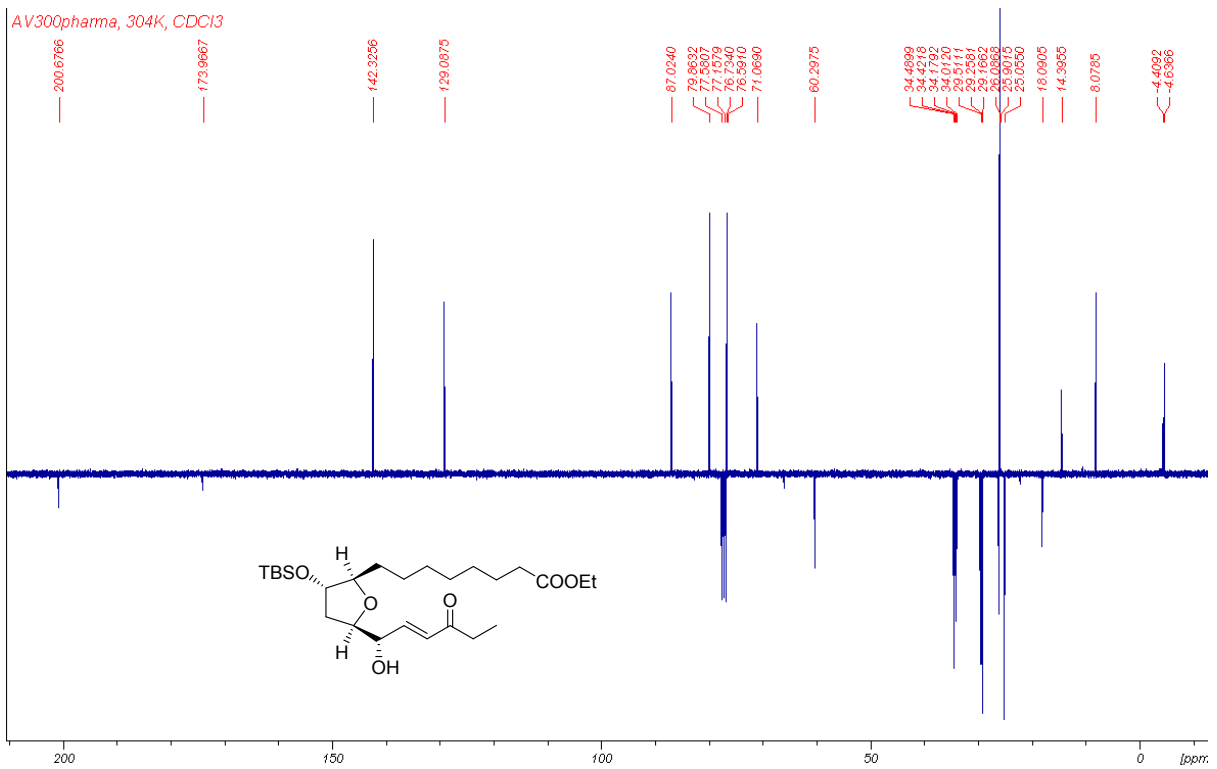
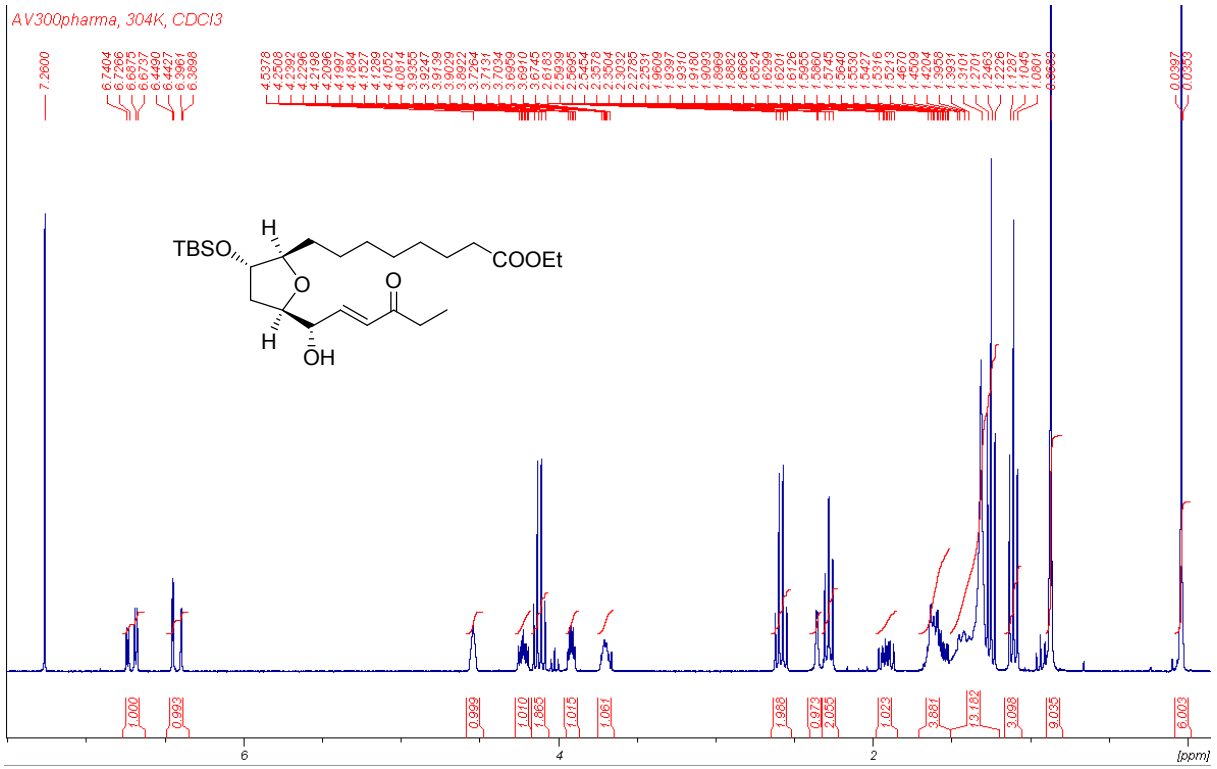


4V300pharma, 304K, CDC13

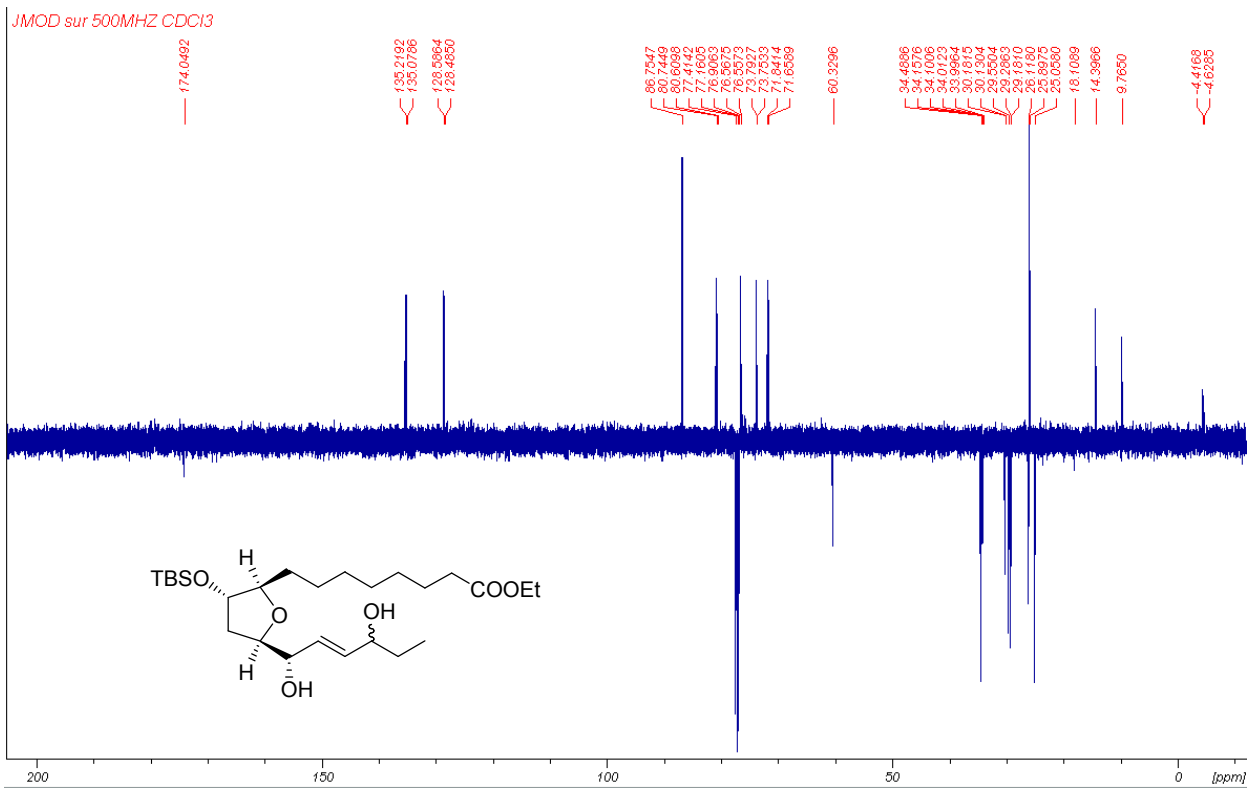
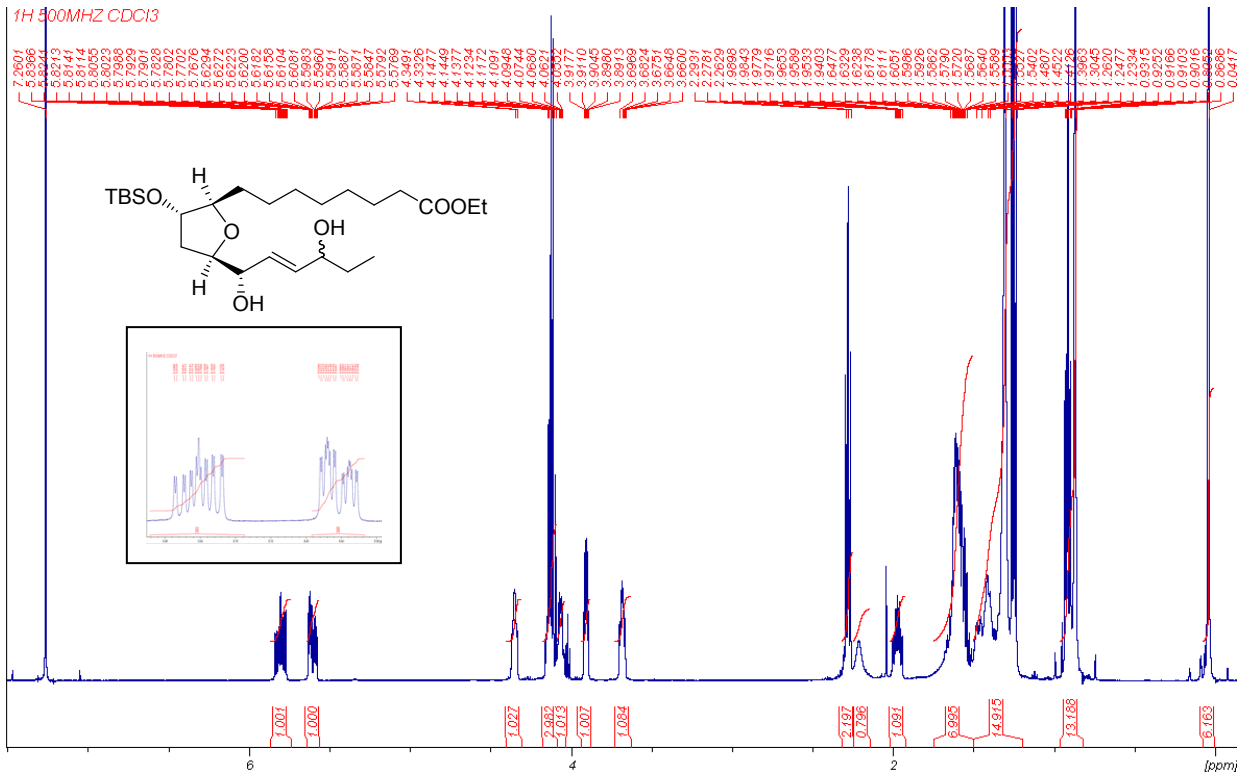


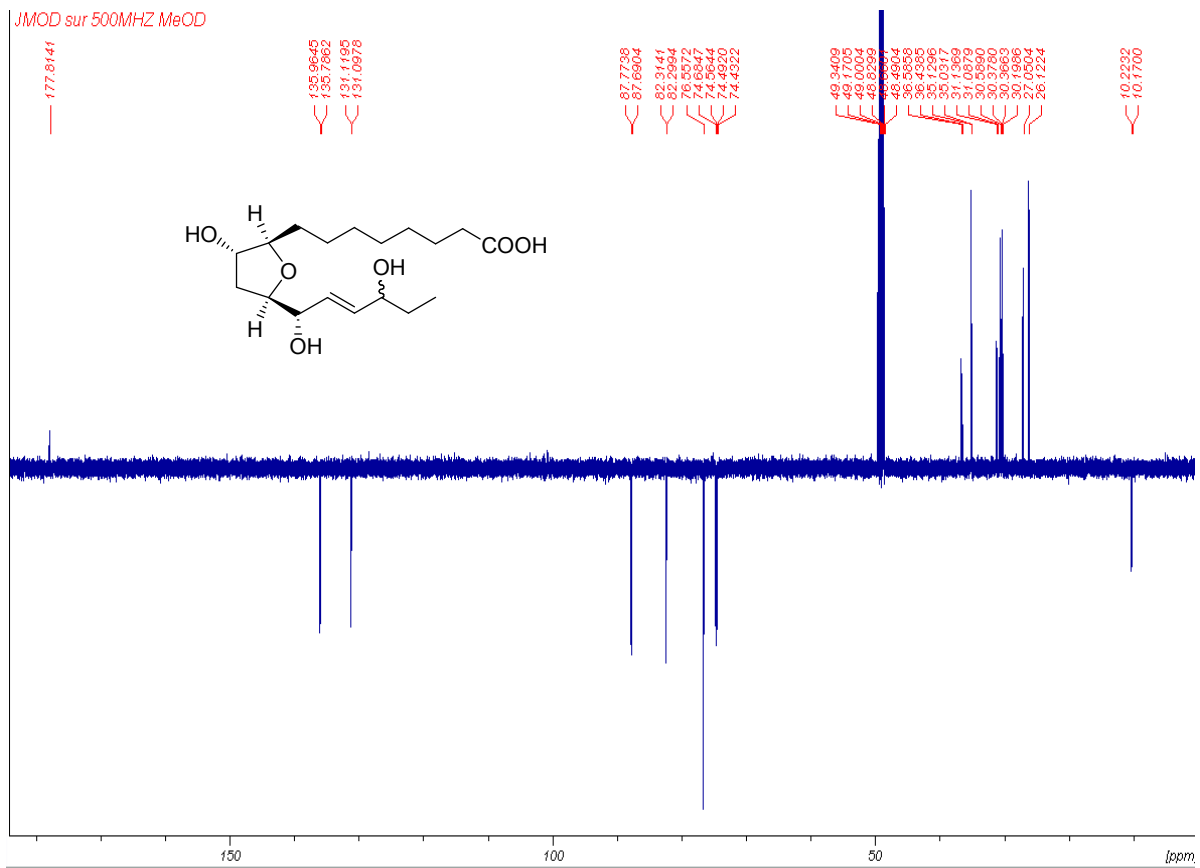
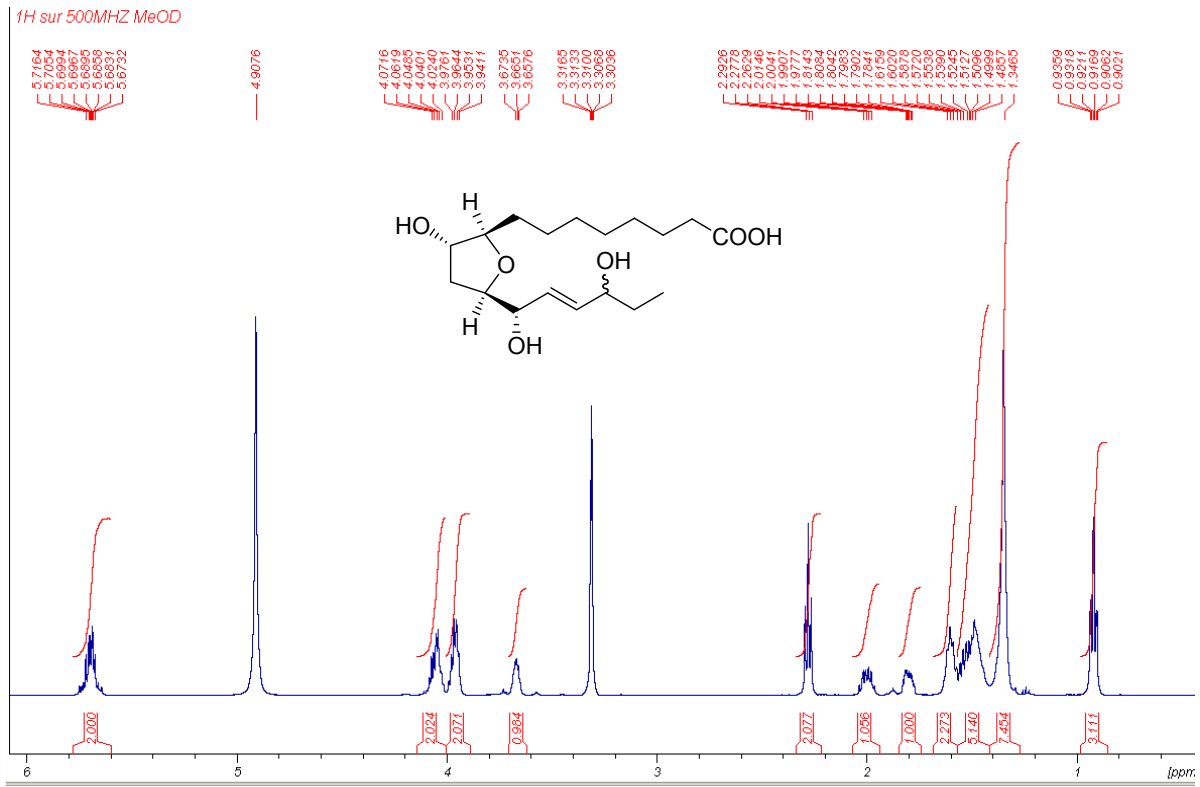
1-methyl-1-cyclopropyl hydroxyl derivative **8**:

enone **9**:

Enone **10**:

enediol:



ent-16-(*RS*)-13-*epi*-ST- Δ^14 -9-PhytoF **1**:

References

- 1 J. P. Fessel, N. A. Porter, K. P. Moore, J. R. Sheller and L. J. Roberts, *Proc. Natl. Acad. Sci.*, 2002, **99**, 16713–16718.
- 2 U. Jahn, J.-M. Galano and T. Durand, *Angew Chem Int Ed.*, 2008, **47**, 5894–5955.
- 3 R. W. Hoffmann, B. C. Kahrs, J. Schiffer and J. Fleischhauer, *J. Chem. Soc. Perkin Trans. 2*, 1996, 2407–2414.
- 4 W. Kabsch, *Acta Crystallogr. D Biol. Crystallogr.*, 2010, **66**, 125–132.
- 5 A. van der Lee, *J. Appl. Crystallogr.*, 2013, **46**, 1306–1315.
- 6 L. Palatinus and G. Chapuis, *J. Appl. Crystallogr.*, 2007, **40**, 786–790.
- 7 P. W. Betteridge, J. R. Carruthers, R. I. Cooper, K. Prout and D. J. Watkin, *J. Appl. Crystallogr.*, 2003, **36**, 1487–1487.
- 8 H. K. Mangold, *Fresenius Z. Für Anal. Chem.*, 1988, **332**, 679–684.
- 9 S. T. Girisha, K. Ravikumar, B. R. Mrunalini and V. Girish, *Asian J. Plant Sci. Res.*, 2014, **4**, 28–35.
- 10 K. S. Leung, X. Chen, W. Zhong, A. C. H. Yu and C.-Y. J. Lee, *Chem. Phys. Lipids*, 2014, **180**, 53–60.
- 11 C.-Y. J. Lee, R. C. S. Seet, S. H. Huang, L. H. Long and B. Halliwell, *Antioxid. Redox Signal.*, 2009, **11**, 407–420.