

Site-selective multi-porphyrin attachment enables the formation of a next-generation antibody-based photodynamic therapeutic

Antoine Maruani,^a Huguette Savoie,^b Francesca Bryden,^b Stephen Caddick,^a Ross Boyle^{*b} and Vijay Chudasama^{*a}

^a *Department of Chemistry, University College London, 20 Gordon Street, London, WC1H 0AJ, UK*

^b *Department of Chemistry, University of Hull, Cottingham Road, Hull, HU6 7RX, UK.*

General Experimental

All reagents were purchased from Sigma-Aldrich, Promega, AlfaAesar, Invitrogen and were used as received. Where described below petrol refers to petroleum ether (40–60 °C). All reactions were monitored by thin-layer chromatography (TLC) on pre-coated SIL G/UV254 silica gel plates (254 μm) purchased from VWR. Flash column chromatography was carried out with Kiesegel 60M 0.04/0.063mm (200–400 mesh) silica gel. ¹H and ¹³C NMR spectra were recorded at ambient temperature on a Jeol JNMLA400 spectrometer instrument operating at a frequency of 400 MHz for ¹H and 100 MHz for ¹³C or on a Bruker Avance 600 instrument operating at a frequency of 600 MHz for ¹H and 150 MHz for ¹³C in DMSO-d, CDCl₃ or CD₃OD (as indicated below). The chemical shifts (δ) for ¹H and ¹³C are quoted relative to residual signals of the solvent on the ppm scale. ¹H NMR peaks are reported as singlet (s), doublet (d), triplet (t), m (multiplet), br. (broad). Coupling constants (*J* values) are reported in Hertz (Hz) and are H-H coupling constants unless otherwise stated. Signal multiplicities in ¹³C NMR were determined using the distortionless enhancement by phase transfer (DEPT) spectral editing technique. Infrared spectra were obtained on a Perkin Elmer Spectrum 100 FTIR Spectrometer operating in ATR mode with frequencies given in reciprocal centimetres (cm⁻¹). Melting points were measured with a Gallenkamp apparatus and are uncorrected. Mass spectra of organic compounds were obtained on a VG70-SE mass spectrometer.

UV-Vis spectroscopy

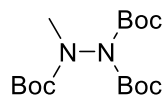
UV-Vis spectra were recorded on a Varian Cary 100 Bio UV/Visible spectrophotometer, operating at room temperature. Sample buffer was used as blank for baseline correction. Calculation of molecule over antibody ratio, *r*, follows the formula below with ε₂₈₀ = 215380 M⁻¹ cm⁻¹ for trastuzumab, ε₂₈₀ = 68590 M⁻¹ cm⁻¹ for trastuzumab Fab, ε₃₄₅ = 9100 M⁻¹ cm⁻¹ for Mestra-PD, ε₄₂₂ = 165175 M⁻¹ cm⁻¹ for the porphyrin, 0.28, as a correction factor (CF) for Mestra-PD for the absorbance at 280 nm.

$$r = \frac{A_{\lambda}/\epsilon_{\lambda}}{(A_{280} - \sum CF_{\lambda} \times A_{\lambda})/\epsilon_{280}}$$

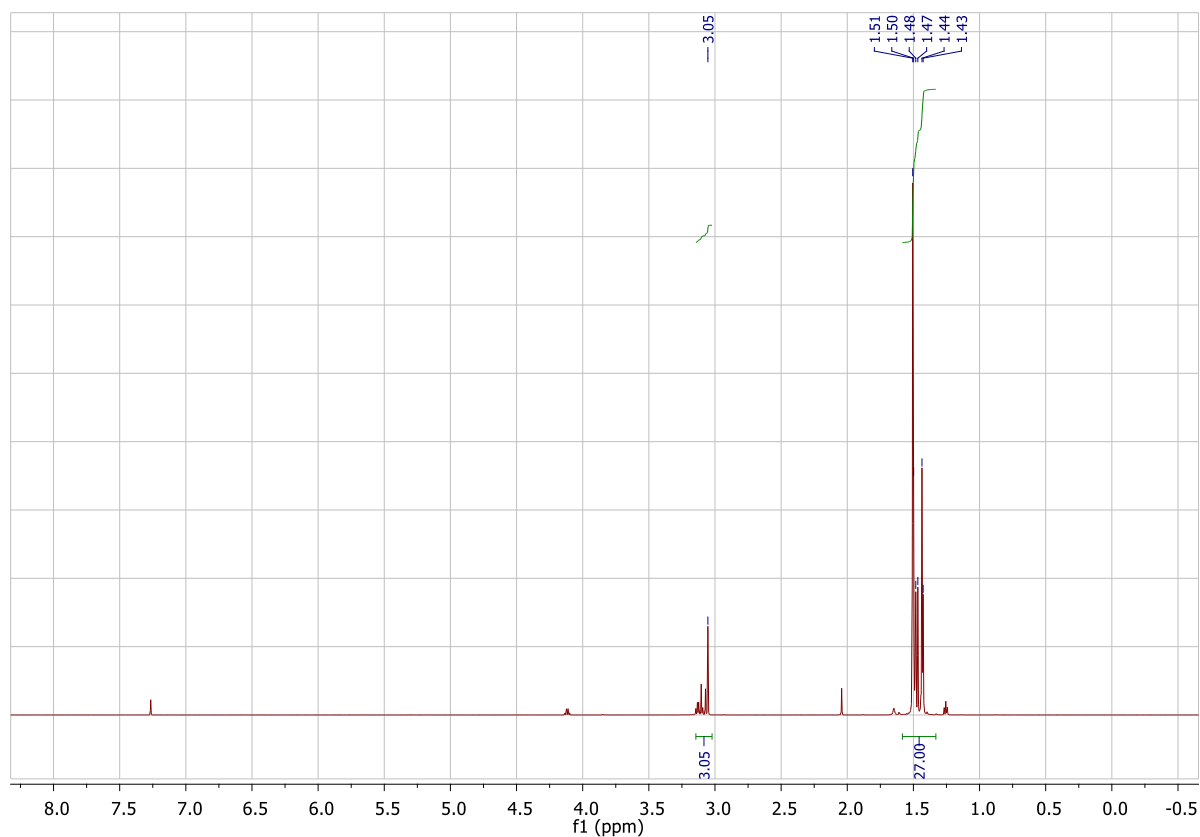
With A_λ the absorbance at the wavelength λ , and ϵ_λ extinction coefficient of the relevant molecule.

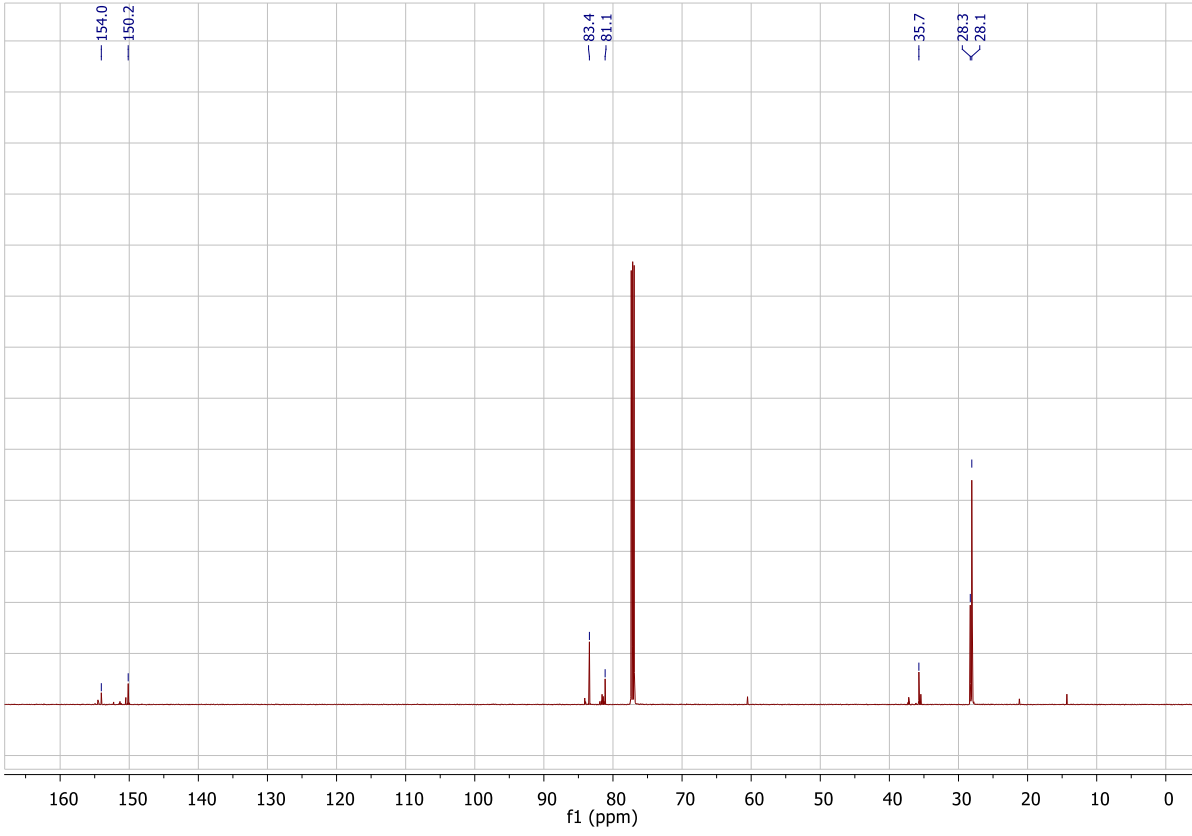
Synthesis of compounds

Tri-*tert*-butyl 2-methylhydrazine-1,1,2-tricarboxylate **2**¹

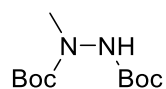


To a solution of methylhydrazine **1** (1.00 g, 1.14 mL 21.7 mmol), NEt_3 (4.34 g, 6.04 mL, 43.4 mmol) and DMAP (260 mg, 2.17 mmol) in CH_2Cl_2 (75 mL) was added Boc_2O (18.9 g, 86.8 mmol) and the reaction mixture stirred at 20 °C for 72 h. Then the reaction mixture was diluted with H_2O (80 mL), extracted with EtOAc (3×60 mL), the combined organic layers were dried (MgSO_4) and concentrated *in vacuo*. The crude residue was purified by flash column chromatography (20% EtOAc/petrol) to afford tri-*tert*-butyl 2-methylhydrazine-1,1,2-tricarboxylate **2** (7.43 g, 21.5 mmol, 99%) as a yellowish oil: ^1H NMR (600 MHz, CDCl_3) (major rotamer) δ 3.05 (s, 3H), 1.51–1.43 (m, 27H); ^{13}C NMR (150 MHz, CDCl_3) (major rotamer) δ 154.0 (C), 150.1 (C) 83.4 (C), 81.4 (C), 35.7 (CH_3), 28.3 (CH_3), 28.1 (CH_3); LRMS (ES^+) 369 (100, $[\text{M}+\text{Na}]^+$).

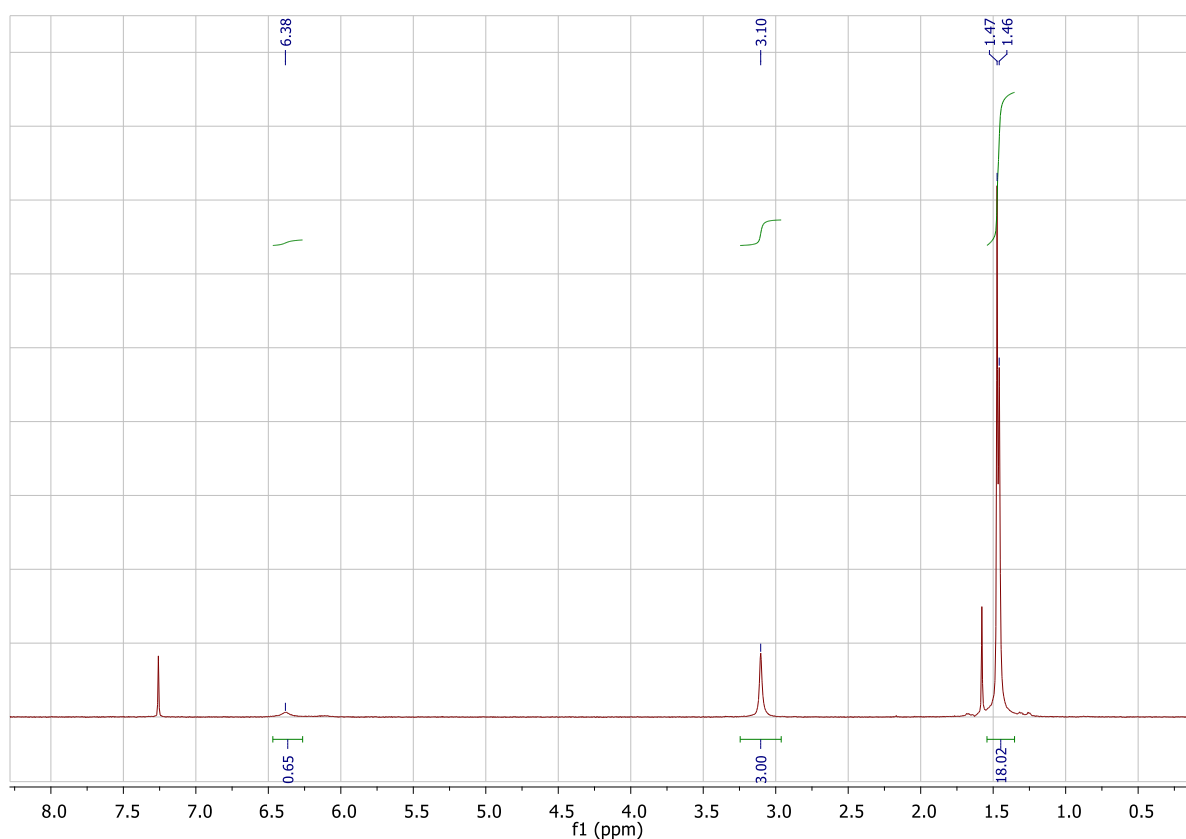


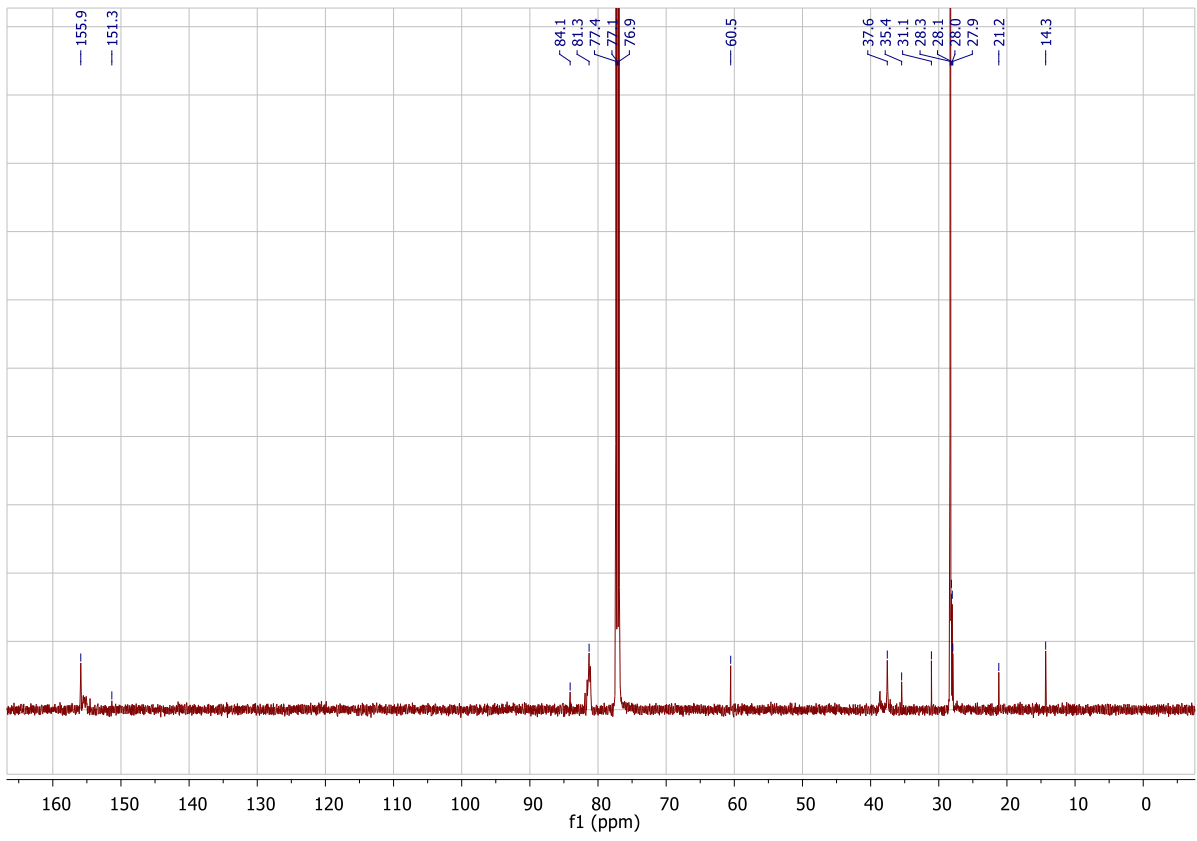


Di-*tert*-butyl 1-methylhydrazine-1,2-dicarboxylate **3**¹

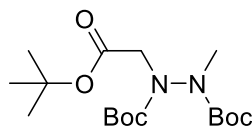


To a solution of tri-*tert*-butyl 2-methylhydrazine-1,1,2-tricarboxylate **2** (2.0 g, 5.8 mmol), in dry MeCN (15 mL) was added Mg(ClO₄)₂ (0.27 g, 1.2 mmol) and the reaction mixture stirred at 20 °C for 1 h. Then the reaction mixture was diluted with 10% aq. citric acid (20 mL) and Et₂O (15 mL), extracted with Et₂O (3 × 20 mL), the combined organic layers were dried (MgSO₄) and concentrated *in vacuo*. The crude residue was purified by flash column chromatography (15% EtOAc/petrol) to afford di-*tert*-butyl 1-methylhydrazine-1,2-dicarboxylate **3** (1.3 g, 5.2 mmol, 89%) as a white solid: m.p. 54–55 °C (*lit. m.p.* 55–56 °C); ¹H NMR (600 MHz, CDCl₃) (major rotamer) δ 6.40 (br. s, 1H), 3.11 (s, 3H), 1.48–1.45 (m, 18H); ¹³C NMR (150 MHz, CDCl₃) (major rotamer) δ 155.9 (C), 155.3 (C) 81.3 (C), 81.1 (C), 37.6 (CH₃), 28.3 (CH₃), 28.1 (CH₃); IR (solid) 3316, 2978, 2932, 1701 cm⁻¹; LRMS (ES⁺) 269 (100, [M+Na]⁺); HRMS (ES⁺) calcd. for C₁₁H₂₂N₂O₄Na [M+Na]⁺ 269.1477, observed 269.1476.

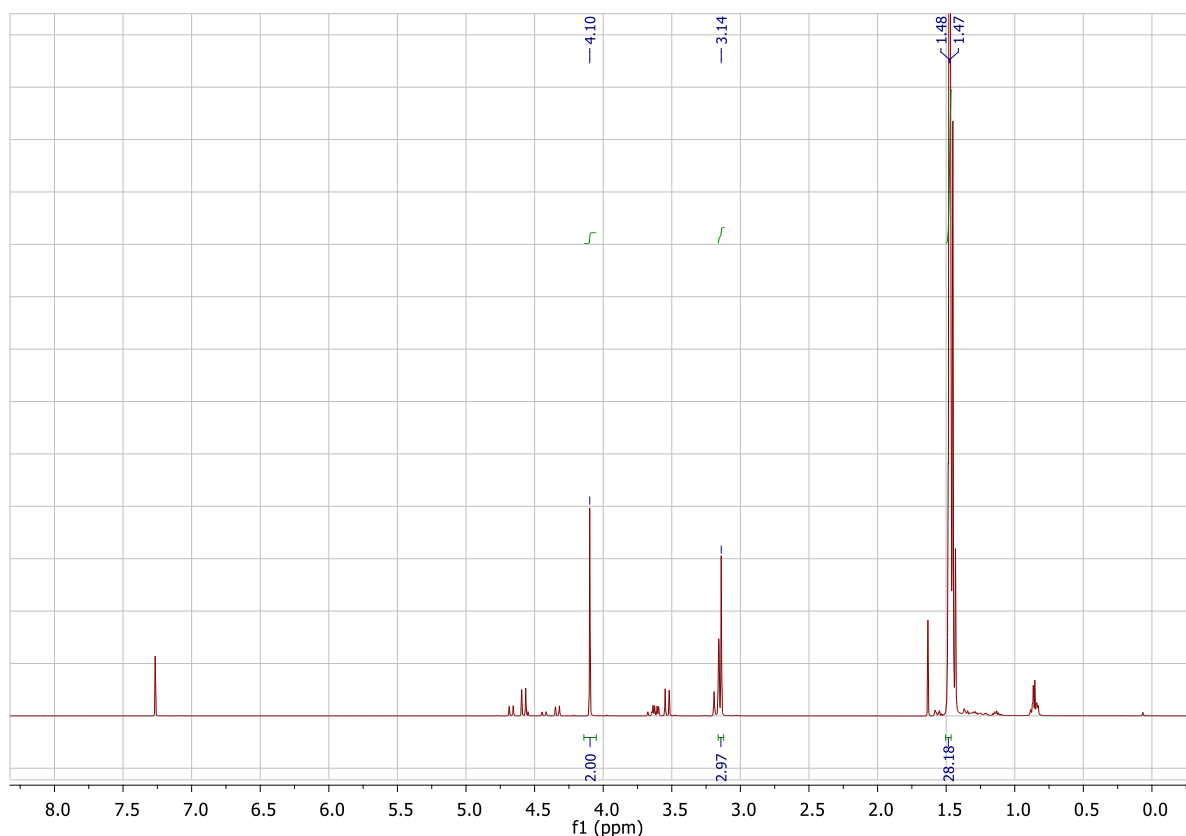


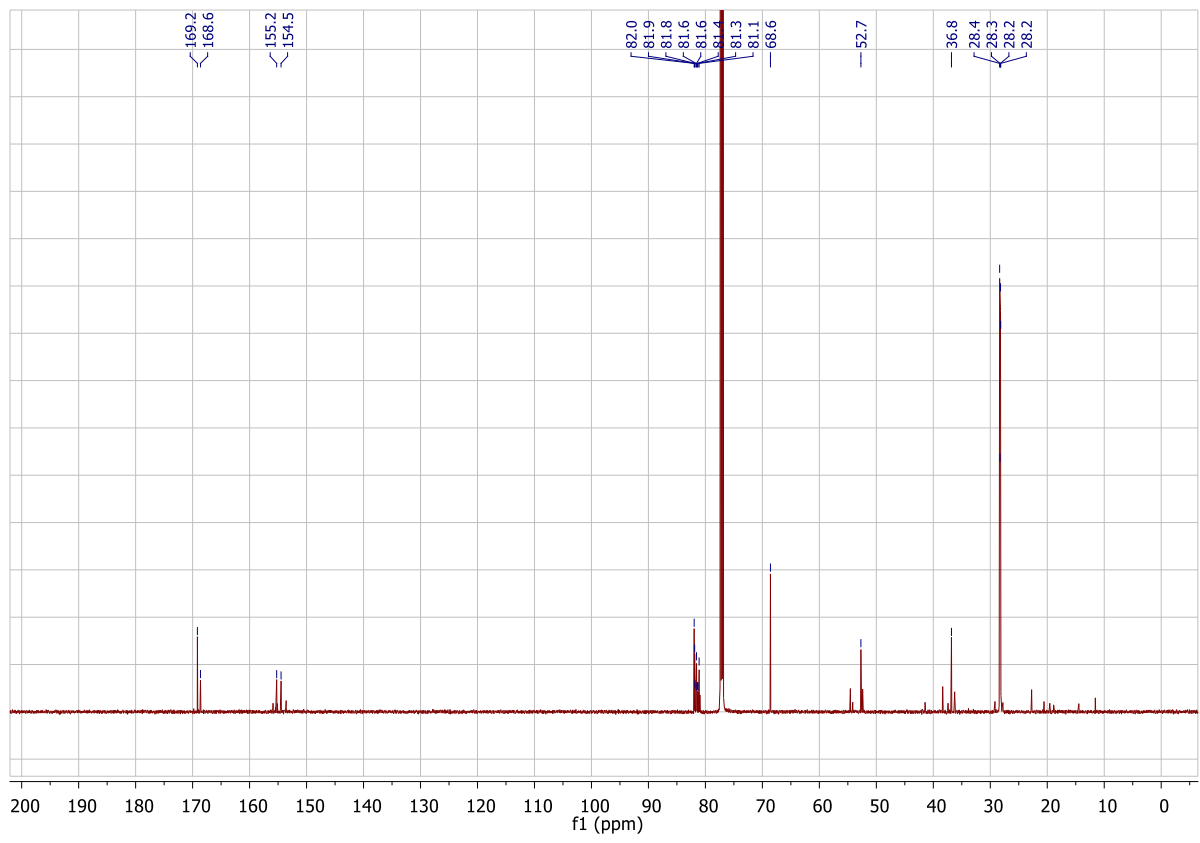


Di-*tert*-butyl 1-(2-(*tert*-butoxy)-2-oxoethyl)-2-methylhydrazine-1,2-dicarboxylate **4**

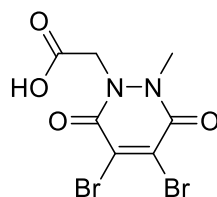


To a solution of di-*tert*-butyl 1-methylhydrazine-1,2-dicarboxylate **3** (0.94 g, 3.8 mmol) in DMF (20 mL) was added caesium carbonate (1.86 g, 5.7 mmol) and *tert*-butyl bromoacetate (1.1 g, 0.84 mL, 5.7 mmol) and the reaction mixture stirred at 20 °C for 16 h. After this time, the reaction mixture was diluted with H₂O (50 mL), extracted with Et₂O (4 × 50 mL), the combined organic layers washed with sat. aq. LiCl (2 × 30 mL), dried (MgSO₄), and concentrated *in vacuo*. Purification by flash column chromatography (10% Et₂O/petrol) yielded di-*tert*-butyl 1-(2-(*tert*-butoxy)-2-oxoethyl)-2-methylhydrazine-1,2-dicarboxylate **4** (1.3 g, 3.7 mmol, 98%) as a colourless oil: ¹H NMR (600 MHz, CDCl₃) δ 4.73–4.04 (m, 2H), 3.68–3.10 (m, 3H), 1.54–1.39 (m, 27H); ¹³C NMR (150 MHz, CDCl₃) (major rotamer) δ 169.2 (C), 155.2 (C), 81.9 (C), 81.6 (C), 81.1 (C), 52.7 (CH₂), 36.8 (CH₃), 28.4 (CH₃), 28.3 (CH₃), 28.2 (CH₃); IR (thin film) 2978, 1748 cm⁻¹; LRMS (ES⁺) 361 (100, [M+H]⁺); HRMS (ES⁺) calcd for C₁₇H₃₃O₆N₂ [M+H]⁺ 361.2339, observed 361.2333.

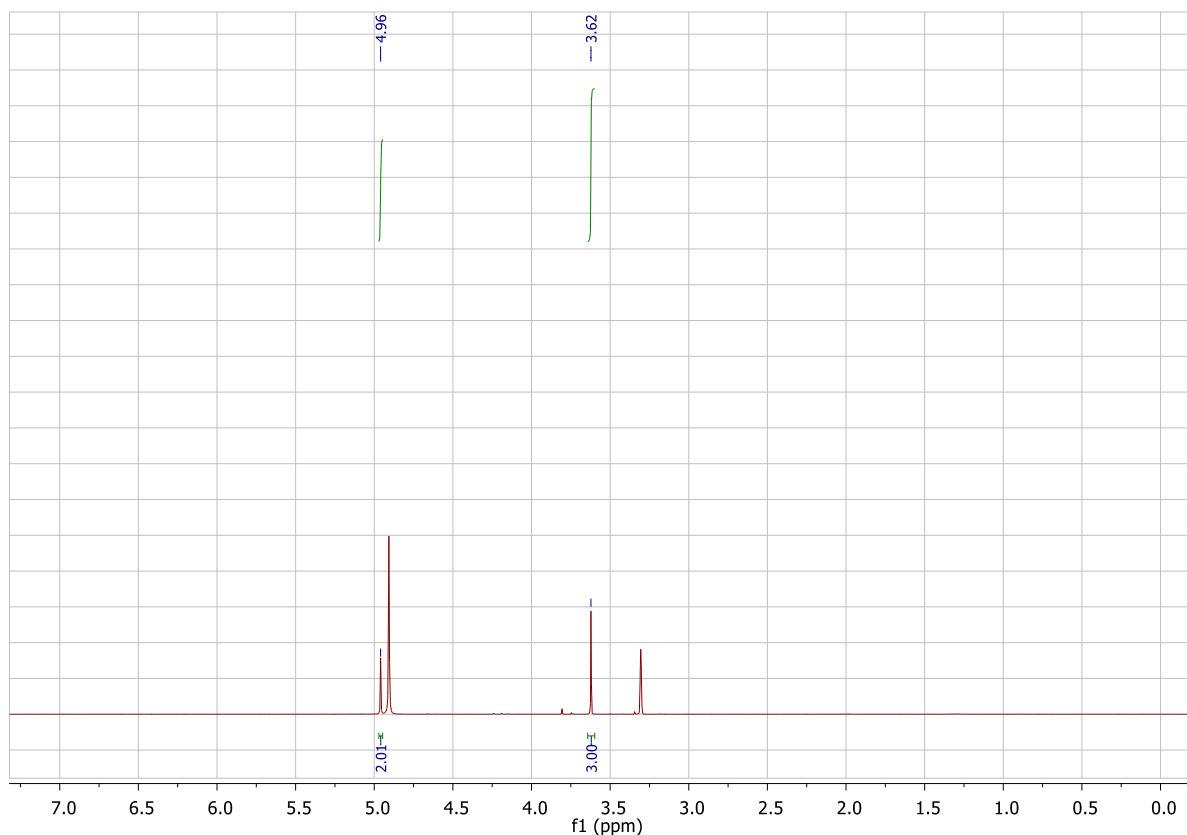


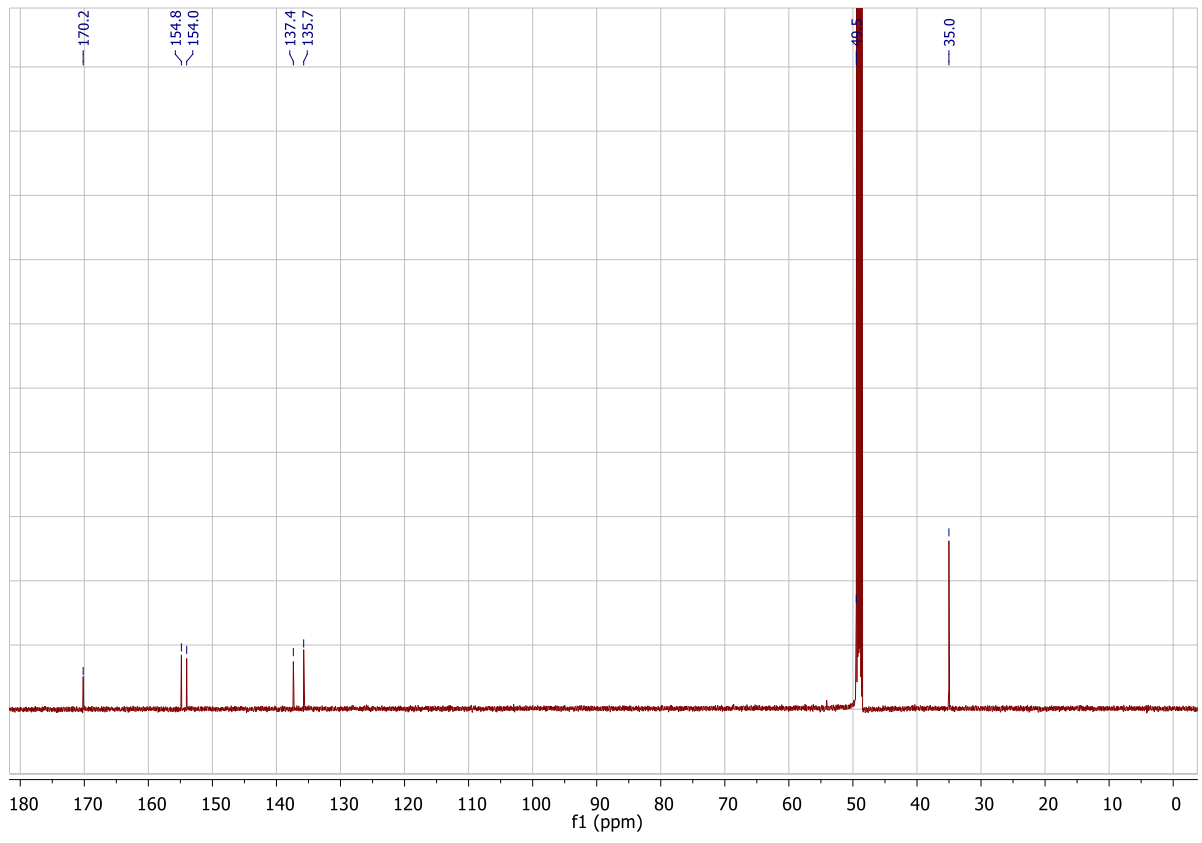


2-(4,5-Dibromo-2-methyl-3,6-dioxo-3,6-dihydropyridazin-1(2H)-yl)acetic acid **5**

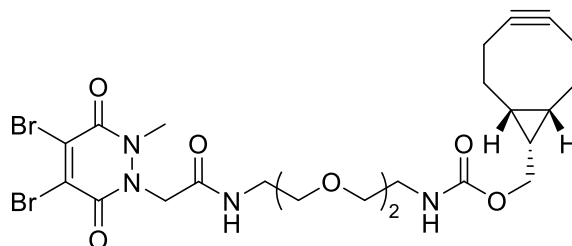


To a solution of di-*tert*-butyl 1-(2-(*tert*-butoxy)-2-oxoethyl)-2-methylhydrazine-1,2-dicarboxylate **4** (1.0 g, 2.8 mmol) in CH₂Cl₂ (10 mL) was added TFA (10 mL) and the reaction mixture stirred at 20 °C for 2 h. After this time, all volatile materials were removed *in vacuo*. The crude residue was added to a solution of 2,3-dibromomaleic anhydride (0.75 g, 2.8 mmol) in glacial AcOH (40 mL), and the reaction mixture stirred at 20 °C for 16 h heated at 130 °C for 16 h. Then the reaction mixture was concentrated *in vacuo*, and purification by flash column chromatography (3% MeOH/CH₂Cl₂ with 1% AcOH) yielded 2-(4,5-dibromo-2-methyl-3,6-dioxo-3,6-dihydropyridazin-1(2H)-yl)acetic acid **5** (0.65 g, 1.9 mmol, 73%) as a white solid: m.p. 210–214 °C; ¹H NMR (600 MHz, MeOD) δ 4.96 (s, 2H), 3.62 (s, 3H); ¹³C NMR (150 MHz, MeOD) δ 170.2 (C), 154.8 (C), 154.0 (C), 137.4 (C), 135.7 (C), 49.5 (CH₂), 35.0 (CH₃); IR (solid) 3023, 2969, 1731, 1662 cm⁻¹; LRMS (ES⁻) 338 (50, [M⁸¹Br⁸¹Br-H]⁻), 340 (100, [M⁸¹Br⁷⁹Br-H]⁻), 342 (50, [M⁷⁹Br⁷⁹Br-H]⁻); HRMS (ES⁻) calcd for C₇H₅N₂O₄⁷⁹Br₂ [M⁷⁹Br⁷⁹Br-H]⁻ 337.8538, observed 337.8540.

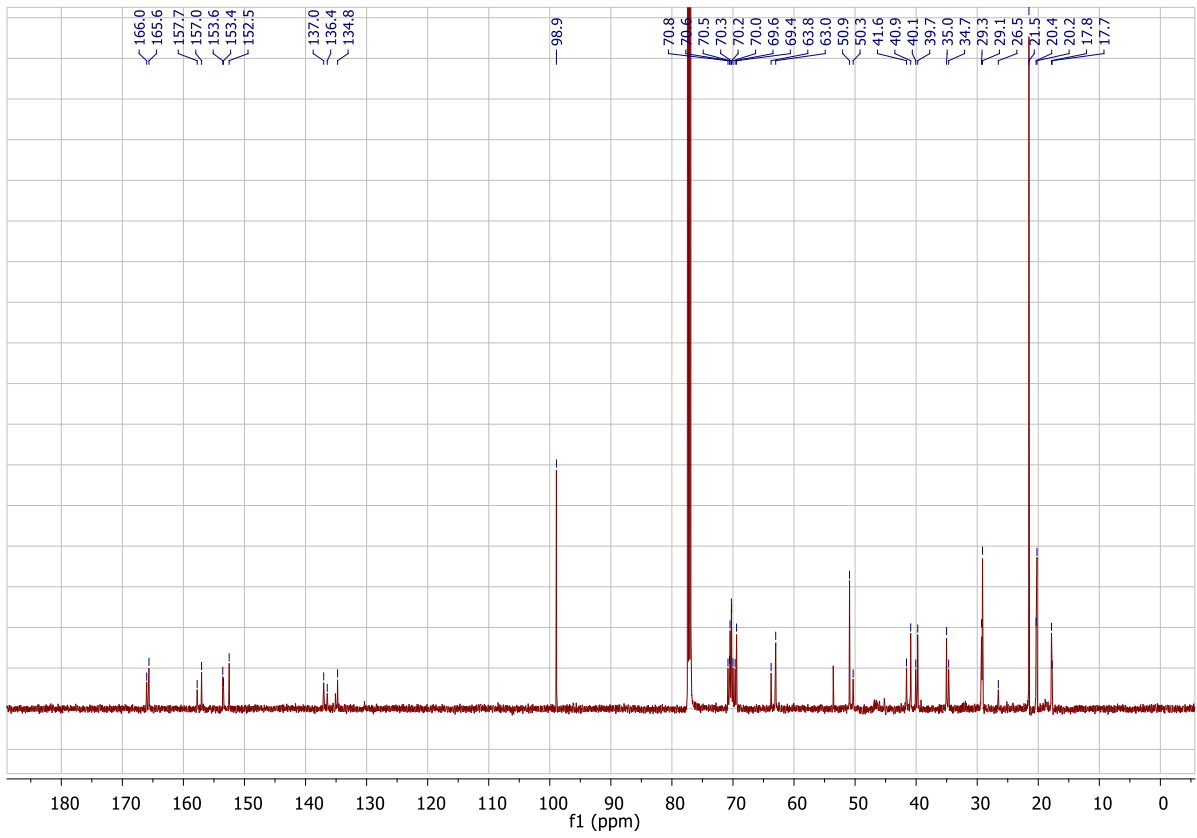
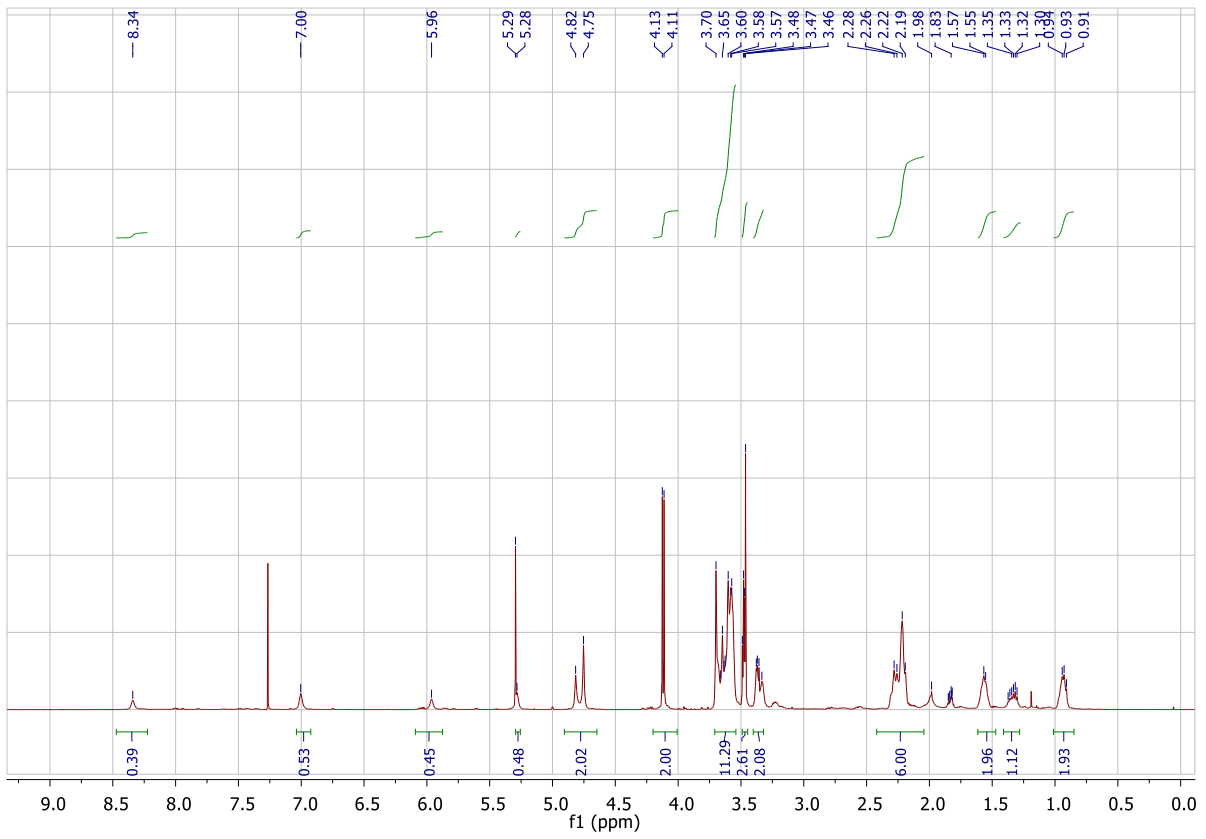




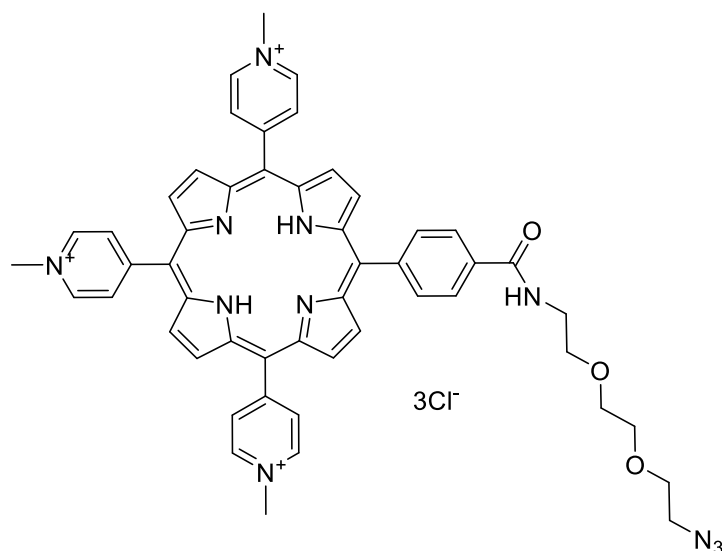
((1*R*,8*S*,9*s*)-Bicyclo[6.1.0]non-4-yn-9-yl)methyl (2-(2-(2-(2-(4,5-dibromo-2-methyl-3,6-dioxo-3,6-dihydropyridazin-1(2*H*)-yl)acetamido)ethoxy)ethoxy)ethyl)carbamate **6**



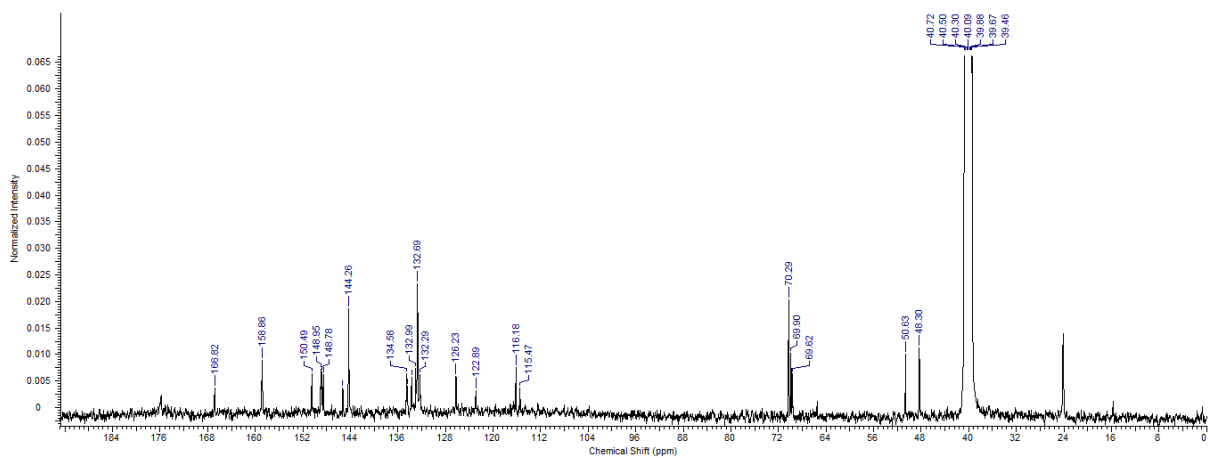
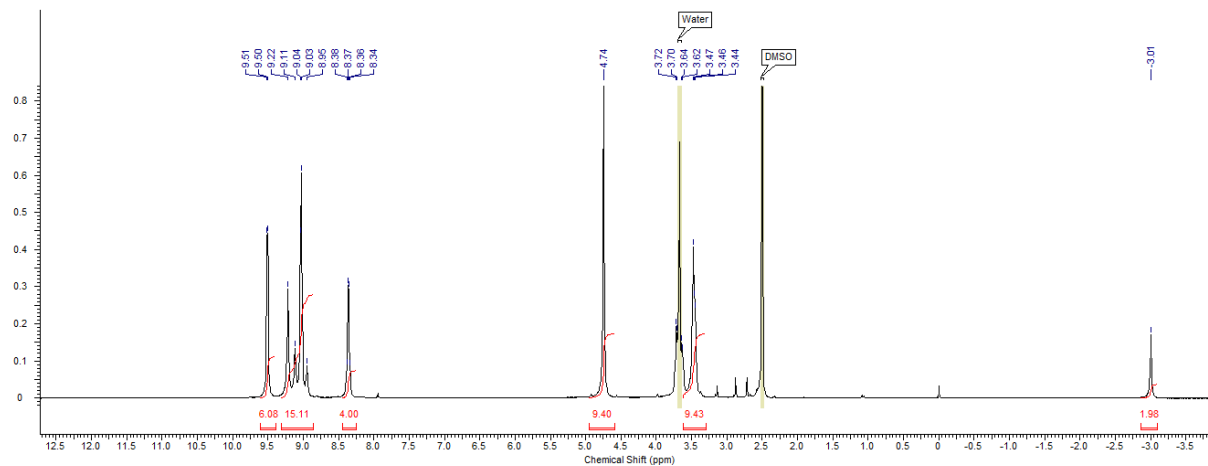
To a solution of 2-(4,5-dibromo-2-methyl-3,6-dioxo-3,6-dihydropyridazin-1(2*H*)-yl)acetic acid **5** (86 mg, 0.25 mmol), PyBOP (0.14 g, 0.28 mmol), and DIPEA (36 mg, 0.28 mmol) in CH₂Cl₂ (5 mL). The resulting solution was stirred at 21 °C for 16 h. Then the reaction mixture was diluted with H₂O (15 mL), extracted with EtOAc (3 × 15 mL), the combined organic layers were dried (MgSO₄) and concentrated *in vacuo*. The crude residue was purified by flash column chromatography (neat EtOAc) to afford ((1*R*,8*S*,9*s*)-bicyclo[6.1.0]non-4-yn-9-yl)methyl (2-(2-(2-(2-(4,5-dibromo-2-methyl-3,6-dioxo-3,6-dihydropyridazin-1(2*H*)-yl)acetamido)ethoxy)ethoxy)ethyl)carbamate **6** (87 mg, 0.14 mmol, 54%) as a yellowish oil: ¹H NMR (600 MHz, CDCl₃) δ 8.34 (br. s, 0.5H), 7.00 (br. s, 0.5H), 5.96 (br. s, 0.5H), 5.29 (br. s, 0.5H), 4.85–4.73 (m, 2H), 4.12 (d, *J* = 8.2 Hz, 2H), 3.76–3.50 (m, 11H), 3.50–3.43 (m, 2H), 3.40–3.30 (m, 2H), 2.31–2.17 (m, 6H), 1.61–1.51 (m, 2H), 1.41–1.24 (m, 1H), 1.01–0.85 (m, 2H); ¹³C NMR (150 MHz, CDCl₃) (major rotamer) δ 165.6 (C), 157.8 (C), 153.4 (C), 152.5 (C), 137.0 (C), 134.8 (C), 98.9 (C), 77.4 (CH₂), 77.2 (CH₂), 77.0 (CH₂), 70.8 (CH₂), 70.6 (CH₂), 70.5 (CH₂), 70.3 (CH₂), 70.2 (CH₂), 70.0 (CH₂), 69.6 (CH₂), 69.4 (CH₂), 63.0 (CH₂), 50.9 (CH₂), 50.3 (CH₂), 40.9 (CH₂), 39.7 (CH₂), 35.0 (CH₃), 29.1 (CH), 21.5 (CH₂), 20.2 (CH), 17.8 (CH₂); IR (thin film) 3338, 2925, 1685, 1633 cm⁻¹; LRMS (ES⁺) 651 (50, [M⁸¹Br⁸¹Br+H]⁺), 649 (100, [M⁸¹Br⁷⁹Br+H]⁺), 647 (50, [M⁷⁹Br⁷⁹Br+H]⁺); HRMS (ES⁺) calcd for C₂₄H₃₃N₄O₇⁷⁹Br₂ [M⁷⁹Br⁷⁹Br+H]⁺ 647.0716, observed 647.0713.



5-[4-[2-(2-(2-Azidoethoxy)ethoxy)ethanaminocarbonyl]phenyl]-10,15,20-tris(*N*-methyl-4-pyridinium)porphyrin trichloride 9



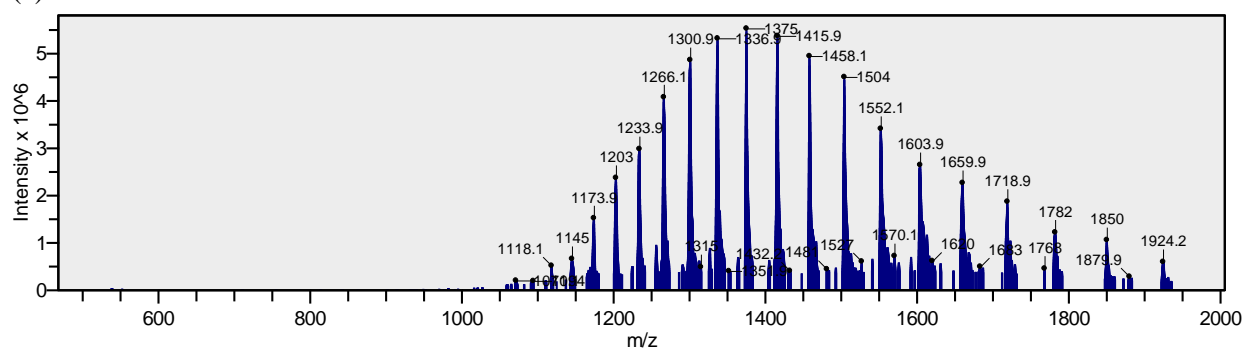
To a stirred solution of 5-[4-[2-(2-(2-azidoethoxy)ethoxy)ethanaminocarbonyl]phenyl]-10,15,20-tris(4-pyridyl)porphyrin² (87 mg, 0.11 mmol) in DMF (10ml) was added methyl iodide (2.0 mL, 32 mmol) *via* syringe. The reaction mixture was stirred at 40 °C overnight. The mixture was cooled to room temperature and cold diethyl ether (100 mL) was added. The mixture was filtered through cotton wool, and the residue redissolved in methanol. The mixture was stirred at room temperature for 3 h, and ammonium hexafluorophosphate added. The resulting solution was filtered and the precipitate redissolved in acetone. Tetrabutylammonium chloride was added, and the resulting solution filtered. The product was precipitated from diethyl ether over MeOH to yield the product as a purple solid (107 mg, 0.37 mmol, 82 %): Rf: 0.49 (silica, 1:1:8 sat. KNO₃ solution:water:MeCN); UV-vis (H₂O) λ_{max} (nm) 422, 519, 558, 584, 643; $\epsilon_{422} = 165,175 \text{ M}^{-1} \text{ cm}^{-1}$; ¹H NMR (400 MHz, DMSO-d₆) δ -2.98 (s, 2H, -NH), 3.45 (m, 2H, CH₂-N₃), 3.56–3.80 (m, 10H), 4.76 (s, 9H, N-CH₃), 8.30–8.43 (m, 4H, 5-*m*-Ph, 5-*o*-Ph), 8.96–9.25 (m, 14H, β H and 10,15,20-*o*-Py), 9.44–9.61 (m, 6H, 10,15,20-*m*-Py); ¹³C NMR (100 MHz, DMSO-d₆) δ 48.3 (CH₂-NH), 50.6 (N-CH₃), 69.6 (O-CH₂), 69.9 (O-CH₂), 70.3 (O-CH₂), 115.5, 116.2, 122.9, 126.2, 132.3, 132.7 (β -C), 133.0, 133.7, 134.6, 144.3 (β -C), 145.3, 148.5, 148.8, 149.0, 150.5, 158.9, 168.8 (C=O); LRMS (MALDI-TOF) *m/z* 287 (100[M-3Cl]³⁺); HRMS(MALDI-TOF) calcd. for C₅₁H₄₆N₁₁O₃: 287.4642, observed 287.4643.



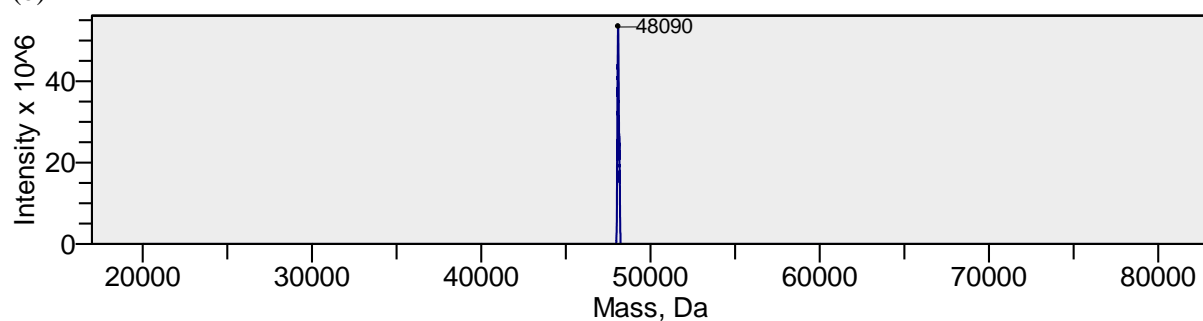
Control conjugation reactions involving trastuzumab Fab

To a solution of trastuzumab Fab (100 μ L, 23 μ M, 1.0 mg/mL, 1 eq) in borate buffer (25 mM sodium borate, 25 mM NaCl, 0.5 mM EDTA, pH 8.0) was added TCEP (final concentration 69 μ M, 3 eq) and Mestra-PD **6** in DMSO (final concentration 115 μ M, 5 eq) and the reaction mixture incubated at 4 $^{\circ}$ C for 16 h. The excess reagents were then removed by repeated diafiltration into fresh buffer using VivaSpin sample concentrators (GE Healthcare, 10000 MWCO). Following this, analysis by LCMS revealed conversion to the desired Fab-Mestra conjugate as the sole product (expected mass: 48089 Da, observed mass: 48090 Da). Then, porphyrin-N₃ **9** (5 eq from a 20 mM solution in DMF) was added and the reaction mixture incubated at 37 $^{\circ}$ C for 4 h. The excess reagents were then removed by repeated diafiltration into fresh PBS using VivaSpin sample concentrators (GE Healthcare, 10000 MWCO). Following this, analysis by LCMS and UV-Vis revealed conversion to the desired Fab-Mestra-Porphyrin conjugate with a porphyrin-to-antibody ratio of 1 (expected mass: 49022 Da, observed mass: 49021 Da).

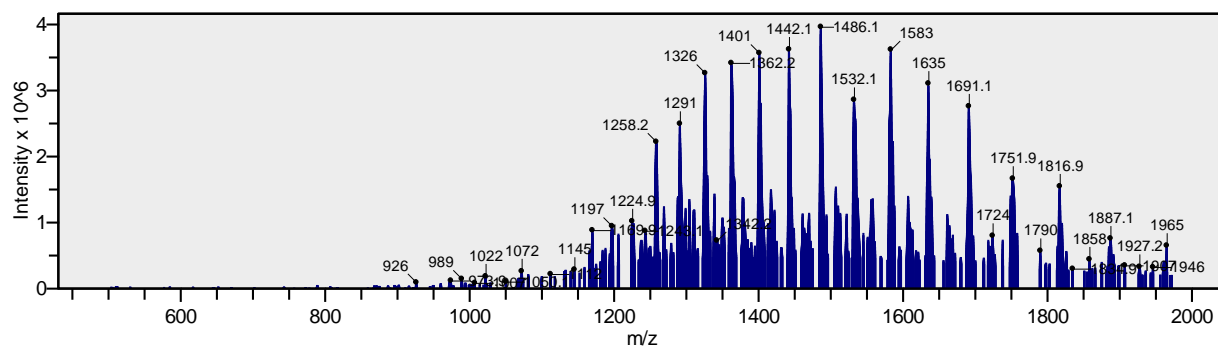
(a)



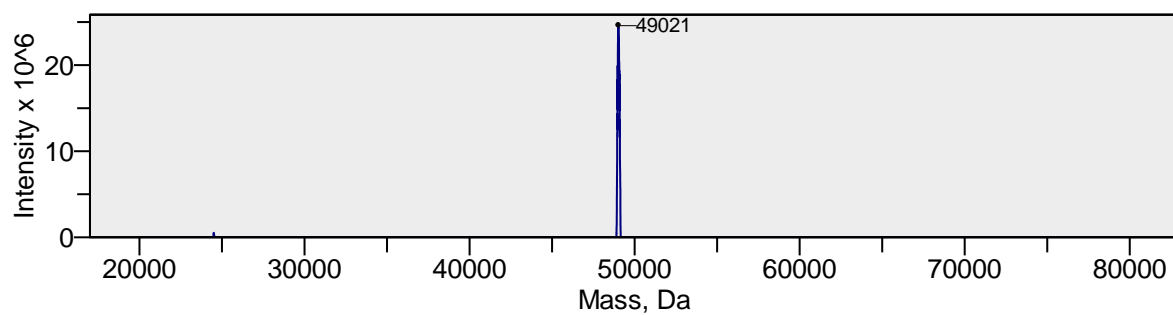
(b)



(c)



(d)



(e)

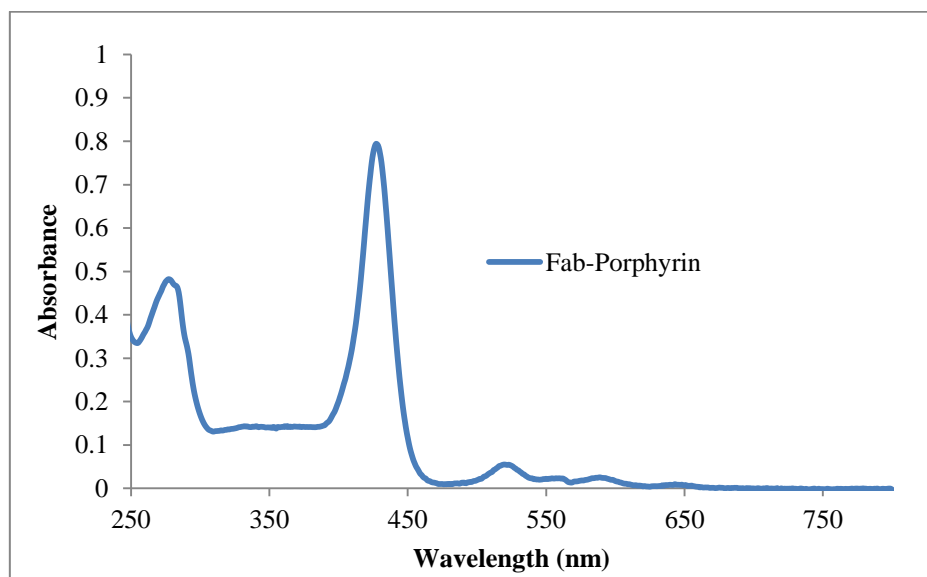
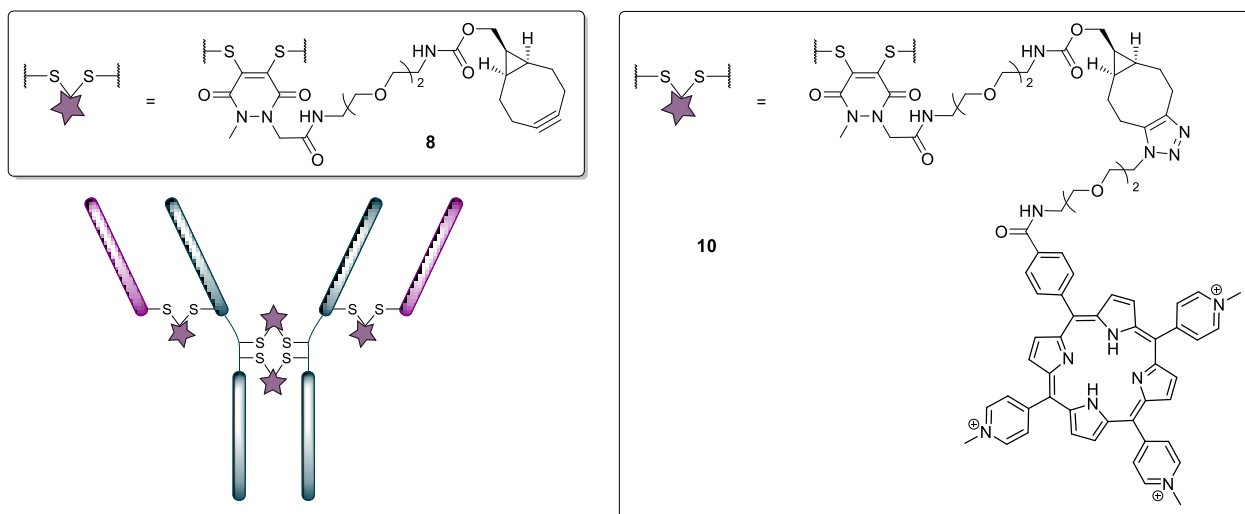


Figure S1. (a) non-deconvoluted and (b) deconvoluted MS data for Fab fragment of trastuzumab reacted with Mestra-PD **6**; (c) non-deconvoluted and (d) deconvoluted MS data for Fab-Mestra reacted with porphyrin **9**; (e) UV-Vis trace for Fab-Mestra reacted with porphyrin **9**.

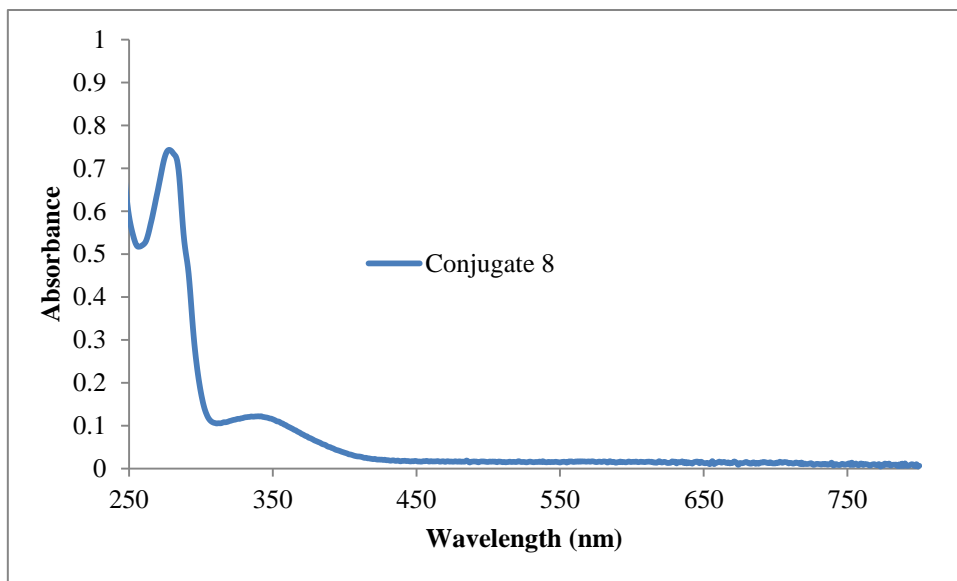
Conjugation reactions involving trastuzumab mAb

Formation of conjugates **8** and **10**



To a solution of trastuzumab (100 μ L, 50 μ M, 7.3 mg/mL, 1 eq) in borate buffer (25 mM sodium borate, 25 mM NaCl, 0.5 mM EDTA, pH 8.0) was added TCEP (final concentration 500 μ M, 10 eq) and Mestra-PD **6** in DMSO (final concentration 1.0 mM, 25 eq) and the reaction mixture incubated at 4 $^{\circ}$ C for 16 h. The excess reagents were then removed by repeated diafiltration into fresh buffer using VivaSpin sample concentrators (GE Healthcare, 10000 MWCO). Following this, analysis by SDS-PAGE gel and UV-Vis revealed conversion to the desired Her-Mestra conjugate **8** with a PD-to-antibody ratio of *ca.* 4. Then, porphyrin-N₃ **9** (20 eq from a 20 mM solution in DMF) was added and the reaction mixture incubated at 37 $^{\circ}$ C for 4 h. The excess reagents were then removed by repeated diafiltration into fresh PBS using VivaSpin sample concentrators (GE Healthcare, 10000 MWCO). Following this, analysis by SDS-PAGE gel and UV-Vis revealed conversion to the desired Her-Mestra-Porphyrin conjugate **10** with a porphyrin-to-antibody ratio of *ca.* 4.1.

(a)



(b)

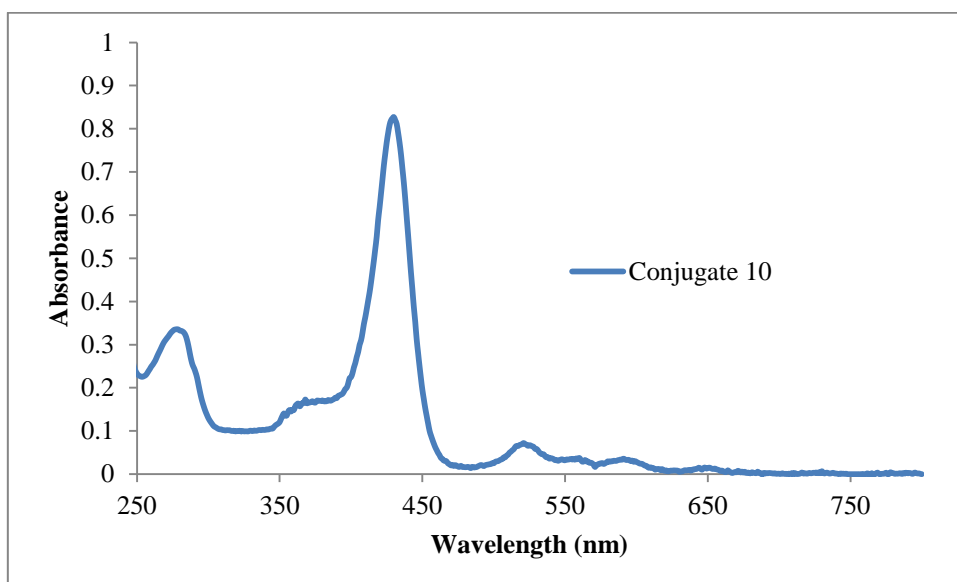


Figure S2. UV-Vis trace for (a) Conjugate **8**, (b) Conjugate **10**.

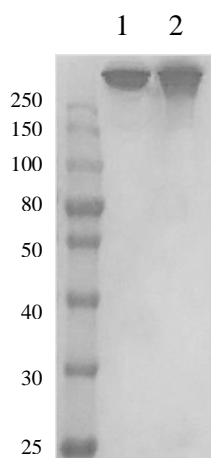


Figure S3. SDS-PAGE of conjugate **8** (lane 1) and conjugate **10** (lane 2).

Cytotoxicity Methodology

Each conjugate was diluted in the appropriate medium, but without FCS, to give a range of five concentrations varying between 2.5×10^{-7} M and 3.0×10^{-6} M. 400 μ L of each cell line adjusted to a concentration of 10^6 cells/mL was added to the dilutions and incubated for 1 h at 37 °C and 5% CO₂. After incubation, the cells were washed with a 4 \times excess of serum-free medium to eliminate any unbound conjugate. The pellets were re-suspended in 0.5 mL of the appropriate serum free medium and 100 μ L aliquoted into 5 wells of a 96-well plate. Each plate was irradiated (IRR) with 20 J/cm² of light (400–700 nm) in two equal doses separated by 10 min delivered using an Oriel, 1000 W quartz tungsten halogen lamp fitted with water filter and diffuser. After irradiation, 5 μ L of fetal bovine serum was added to each well and the plates were returned to the incubator overnight. After 24 h, an MTT (Thiazolyl blue; Sigma M5655) cell viability assay was performed and the results expressed as % of cell viability versus porphyrin–conjugate concentration.

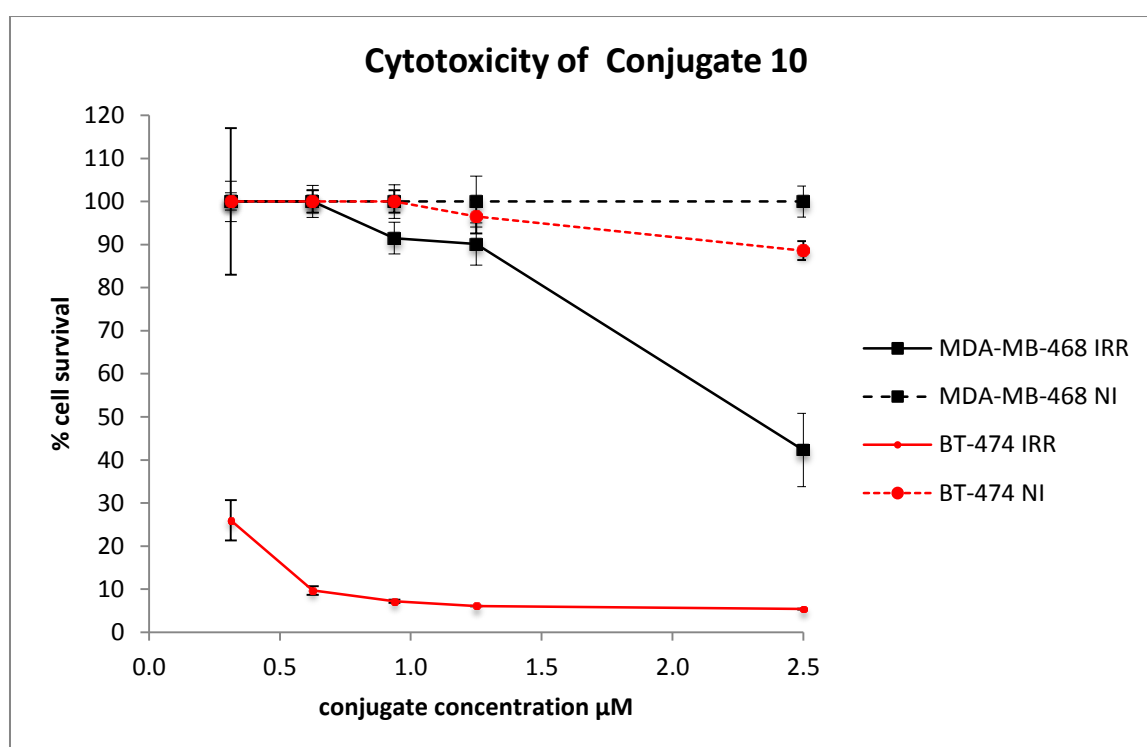


Figure S4. Percentage cell viability for HER2 positive cells (BT-474) and HER2 negative cells (MDA-MB-468), in suspension, determined by MTT assay 24 h after incubation with conjugate **10** and with irradiation (IRR)(20 J/cm² light (400–700 nm) or without irradiation (NI).

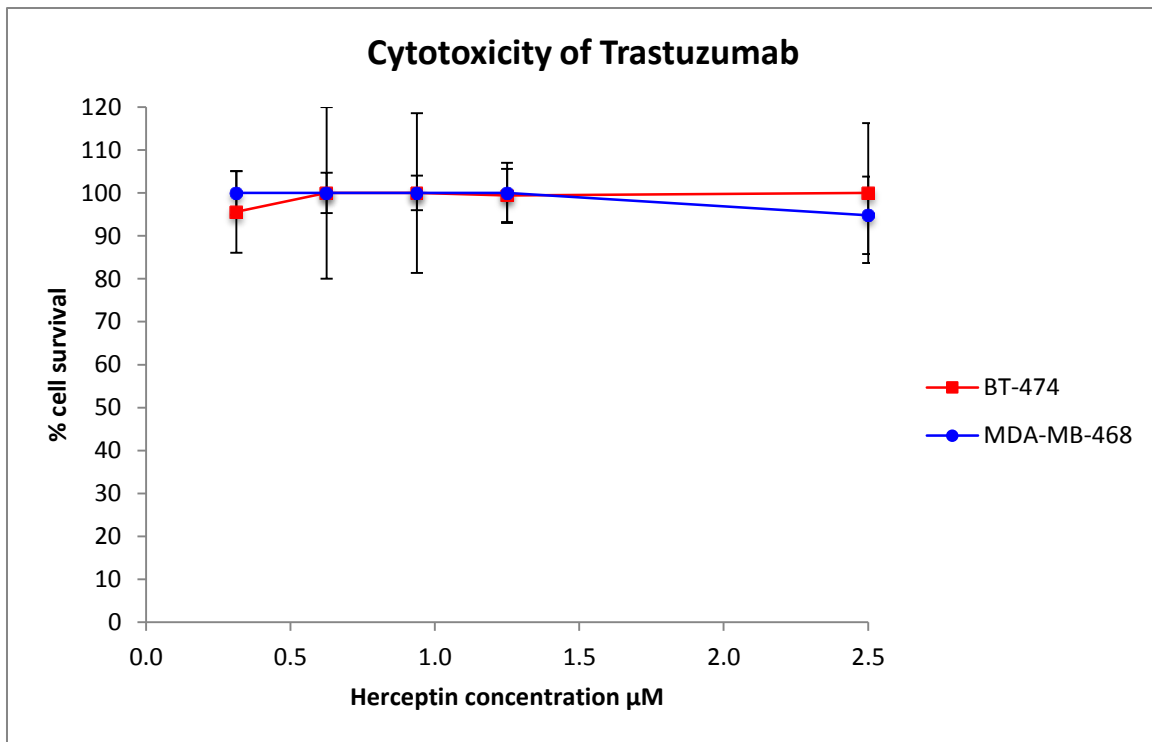


Figure S5. Percentage cell viability for HER2 positive cells (BT-474) and HER2 negative cells (MDA-MB-468), in suspension, determined by MTT assay 24 h after incubation with trastuzumab and irradiation with 20 J/cm^2 light (400–700 nm).

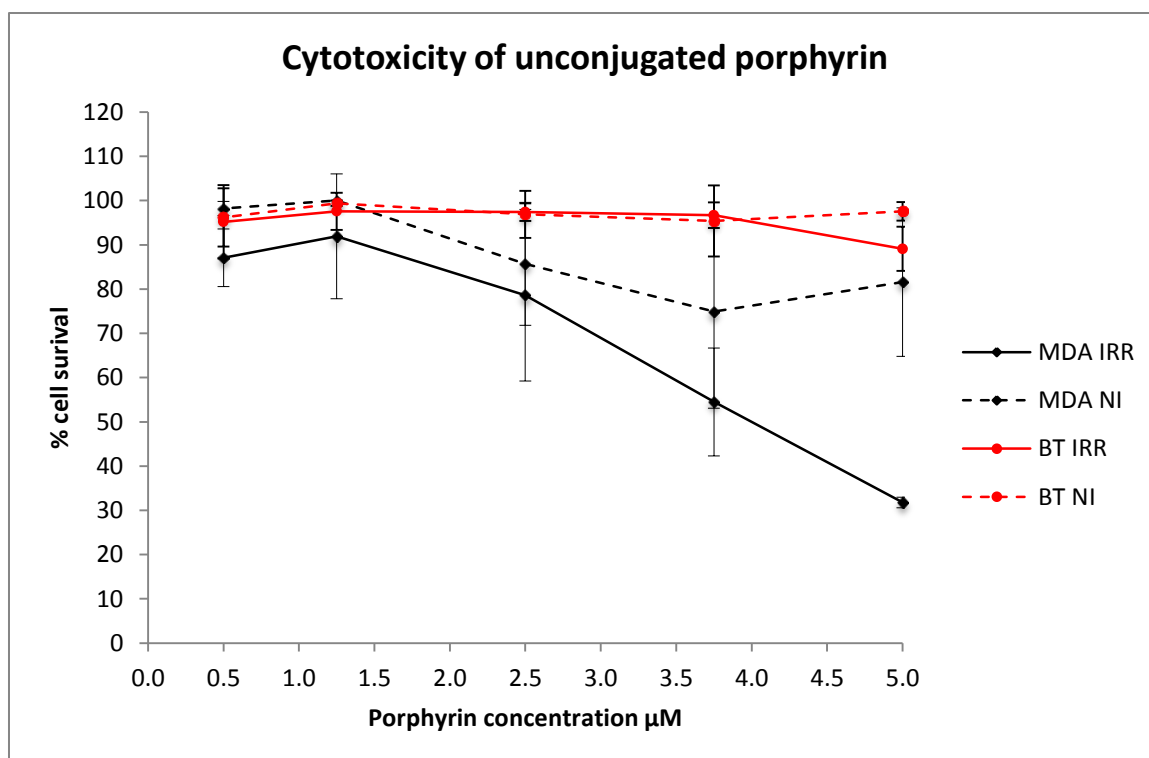


Figure S6. Percentage cell viability for HER2 positive cells (BT-474) and HER2 negative cells (MDA-MB-468), in suspension, determined by MTT assay 24 h after incubation with unconjugated porphyrin and irradiation with 20 J/cm² light (400–700 nm).

References

1. J. A. Stafford, M. F. Brackeen, D. S. Karanewsky and N. L. Valvano, *Tetrahedron Lett.*, 1993, **34**, 7873–7876.
2. F. Bryden, A. Maruani, H. Savoie, V. Chudasama, M. E. B. Smith, S. Caddick and R. W. Boyle, *Bioconjugate Chem.*, **2014**, *25*, 611–617.