

Catalytic Asymmetric Sulfenylation to Structurally Diverse Dithioketals

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Supporting Information

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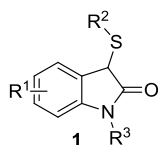
General: Reactions were monitored by thin layer chromatography using UV light to visualize the course of reaction. Purification of reaction products was carried out by flash chromatography on silica gel. Chemical yields refer to pure isolated substances. NMR spectra were obtained using a Bruker DPX-400 spectrometer. Chemical shifts are reported in ppm from tetramethylsilane with the solvent resonance as the internal standard. The following abbreviations were used to designate chemical shift multiplicities: s = singlet, d = doublet, t = triplet, q = quartet, h = heptet, m = multiplet, br = broad.

All reactions were run in air except noted. Anhydrous THF and toluene were prepared by distillation over sodium-benzophenone ketyl prior to use. Anhydrous CH₂Cl₂ and CH₃CN were prepared by first distillation over P₂O₅ and then from CaH₂. Anhydrous EtOAc and acetone was prepared by first distillation over activated CaSO₄ and then stored in 5 Å molecular sieves. The powdered 13X molecular sieves was purchased from Alfa Aesar and activated by heating at 150 °C under vacuum for 8 hours before using. The S-based nucleophiles **1**¹ and **5**² were prepared according to literature reports. Sulfenylation reagent **2**³ and **7**⁴ were synthesized according to literature methods.

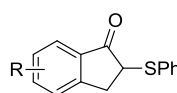
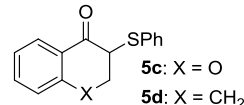
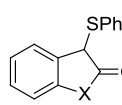
List of abbreviation:

Entry	Chemical name	Abbreviation	Entry	Chemical name	Abbreviation
1	Petroleum ether	PE	3	Dimethylsulfoxide	DMSO
2	Tetrahydrofuran	THF	4	Molecular sieve	MS

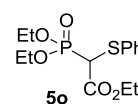
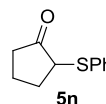
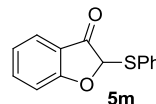
List of starting material:



- 1a:** R¹ = H, R² = *p*-ClC₆H₄, R³ = H
1b: R¹ = 5-OMe, R² = *p*-ClC₆H₄, R³ = H
1c: R¹ = 6-OMe, R² = *p*-ClC₆H₄, R³ = H
1d: R¹ = 7-Cl, R² = *p*-ClC₆H₄, R³ = H
1e: R¹ = 5,7-Me₂, R² = *p*-ClC₆H₄, R³ = H
1f: R¹ = H, R² = *p*-MeC₆H₄, R³ = H
1g: R¹ = H, R² = *o*-FC₆H₄, R³ = H
1h: R¹ = H, R² = 2-Naphthyl, R³ = H
1i: R¹ = H, R² = Bn, R³ = H
1j: R¹ = H, R² = Allyl, R³ = H
1k: R¹ = H, R² = *p*-ClC₆H₄, R³ = Me
1l: R¹ = H, R² = *p*-ClC₆H₄, R³ = Bn
1m: R¹ = 5-F, R² = *p*-ClC₆H₄, R³ = Bn
1n: R¹ = 5-Me, R² = *p*-ClC₆H₄, R³ = Bn



- 5i:** R = 6-Br
5j: R = 6-Me
5k: R = 5-OMe
5l: R = 5,6-(OMe)₂



¹ (a) F. Zhou, X.-P. Zeng, C. Wang, X.-L. Zhao and J. Zhou, *Chem. Commun.*, 2013, **49**, 2022; (b) W.-M. Gao, J.-S. Yu, Y.-L. Zhao, Y.-L. Liu, F. Zhou, H.-H. Wu and J. Zhou, *Chem. Commun.*, 2014, **50**, 15179.

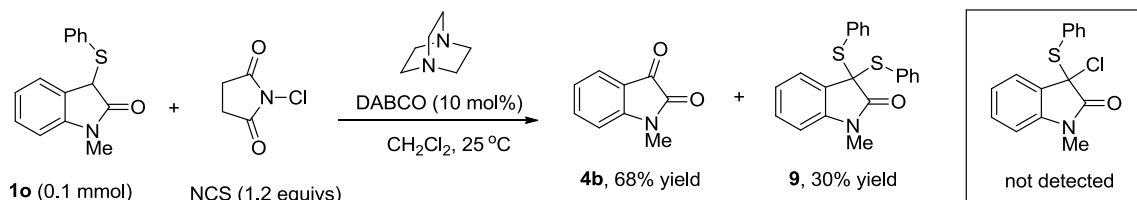
² (a) T. Bootwicha, X. Liu, R. Pluta, I. Atodiresei and M. Rueping, *Angew. Chem., Int. Ed.*, 2013, **52**, 12856; (b) X. Wang, T. Yang, X. Cheng and Q. Shen, *Angew. Chem., Int. Ed.*, 2013, **52**, 12860.

³ P. Saravanan and P. Anbarasan, *Org. Lett.*, 2014, **16**, 848.

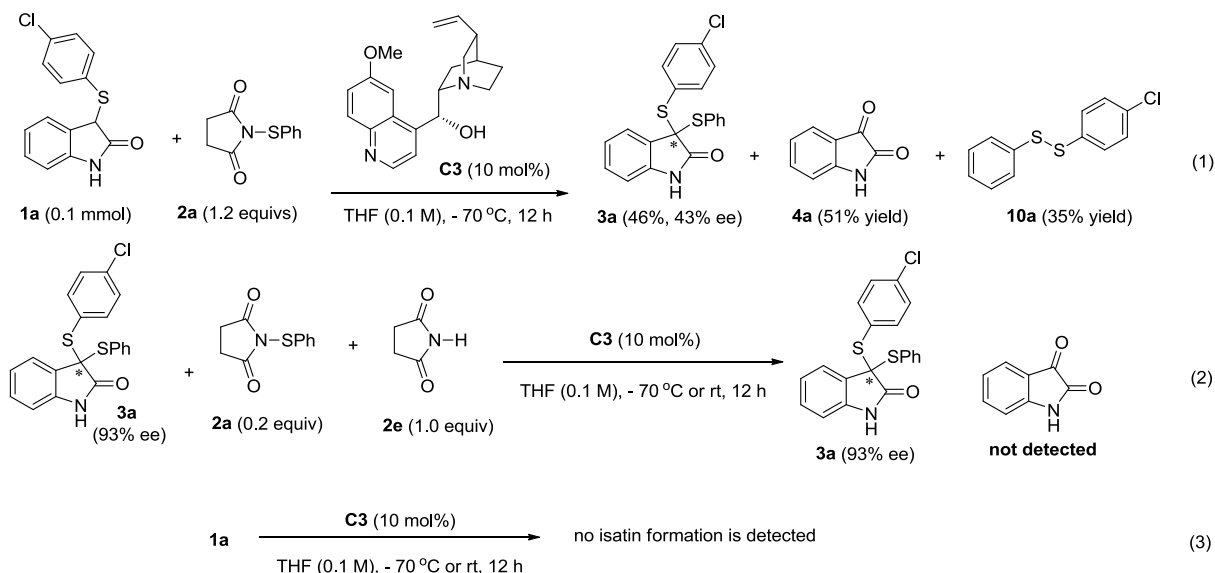
⁴ (a) G. Teverovskiy, D. S. Surry and S. L. Buchwald, *Angew. Chem., Int. Ed.*, 2011, **50**, 7312; (b) C. Xu and Q. Shen, *Org. Lett.*, 2014, **16**, 2046.

1) Mechanistic consideration

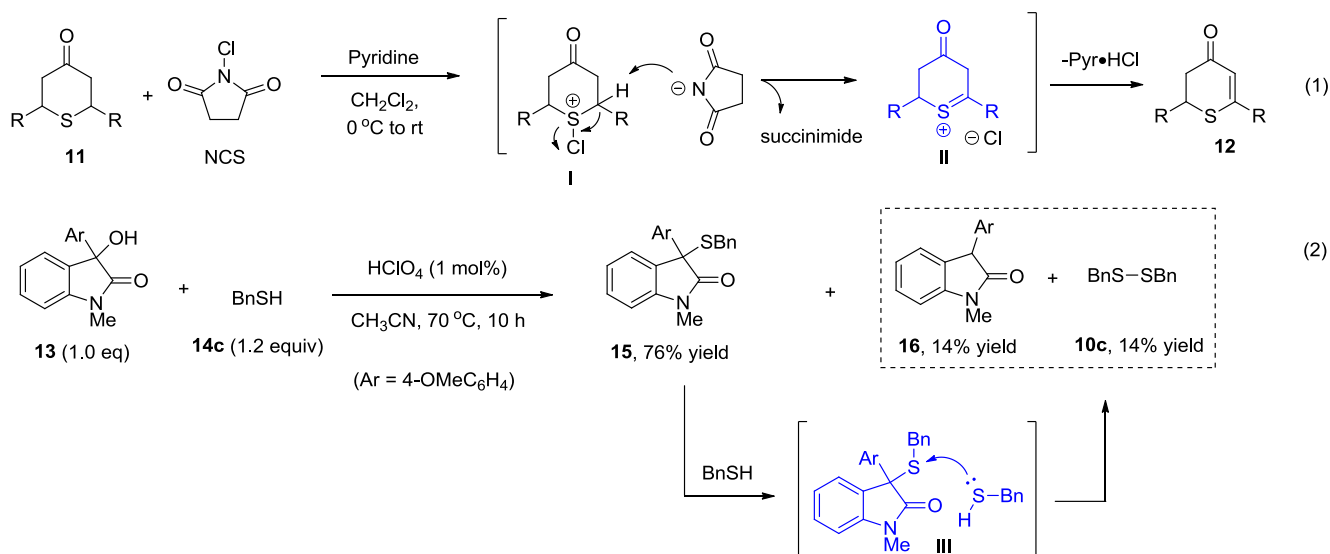
During our studies in base catalyzed reactions of 3-thiooxindoles with some electrophiles, we had noticed the formation of isatin as side products, the amount of which depended on the electrophiles used. For example, the reaction of 3-thiooxindole **1o** with *N*-chlorosuccinimide (NCS) catalyzed by DABCO at 25 °C did not afford the desired 3-chloro-3-thiooxindole, but gave symmetric dithioketal **9** and isatin **4b**. The same phenomena were also observed in the reaction of **1o** with other *N*-halosuccinimides.



In the DABCO catalyzed sulfenylation of 3-thiooxindole **1a** using *N*-SPh succinimide **2a** at room temperature, isatin formation was not observed. However, in the corresponding asymmetric studies, isatin formation proved to be troublesome when we tried lowering temperature to -70 °C to improve enantioselectivity. For example, when using quinine **C3** as the catalyst, dithioketal **3a** was obtained in only 46% yield, but isatin was isolated in 51% yield, along with unsymmetrical disulfane **10a** in 35% yield (eq 1). Dithioketal **3a** proved to be stable under reaction condition or during the purification by column chromatography. Even **3a** was stirred with quinine **C3**, **2a** and **2e** at -70 °C or room temperature for 12 h, no isatin was detected, and it was recovered in unchanged ee value (eq 2). Moreover, without **2a**, no isatin formed if stirring 3-thiooxindole **1a** with catalyst **C3** at the same condition for 12 h (eq 3).



To account for these observations, an investigation into literature report promoted us to consider that 3-thiooxindoles might attack electrophiles in an S-nucleophilic way to form a sulfonium salt that further underwent side reactions, owing to their ambident nucleophilicity. For example, in the synthesis of dihydro-4H-thiopyran-4-one **12** from tetrahydro-4H-thiopyran-4-one **11** via oxidative elimination reaction, in the presence of pyridine, Chen et al proposed that the S-nucleophilic attack of **11** to NCS led to the formation of chloro-sulfonium salt **I**, which was then deprotonated to form a thionium ion **II**, with the dissociation of a chloride (eq 1 of Scheme S1).⁵ It should be noted that thionium ion intermediate was also proposed in the Pummerer reaction.⁶



Scheme S1. Useful literature information

On the other hand, the attack of a sulfur atom on electrophilic sulfur species is possible.⁷ For example, the nucleophilic attack of a mercaptan on sulfenimides had also been reported.⁸ In addition, we had previously observed that in the acid-catalyzed substitution of BnSH with 3-substituted hydroxyoxindoles **13**, 3-monoalkylthiooxindole **16** and a disulfane **10c** were formed as side products, possibly through the attack of a mercaptan to the sulfur of 3-alkylthiooxindole **15** (eq 2 of Scheme S1).⁹ Such a phenomenon was also reported by Magnus.¹⁰

Accordingly, it was possible that in the reaction of 3-thiooxindole and NCS, the S-nucleophilic

⁵ C. H. Chen, G. A. Reynolds and J. A. V. Allan, *J. Org. Chem.*, 1977, **42**, 2777.

⁶ L. H. S. Smith, S. C. Coote, H. F. Sneddon and D. J. Procter, *Angew. Chem., Int. Ed.*, 2010, **49**, 5832 and references therein.

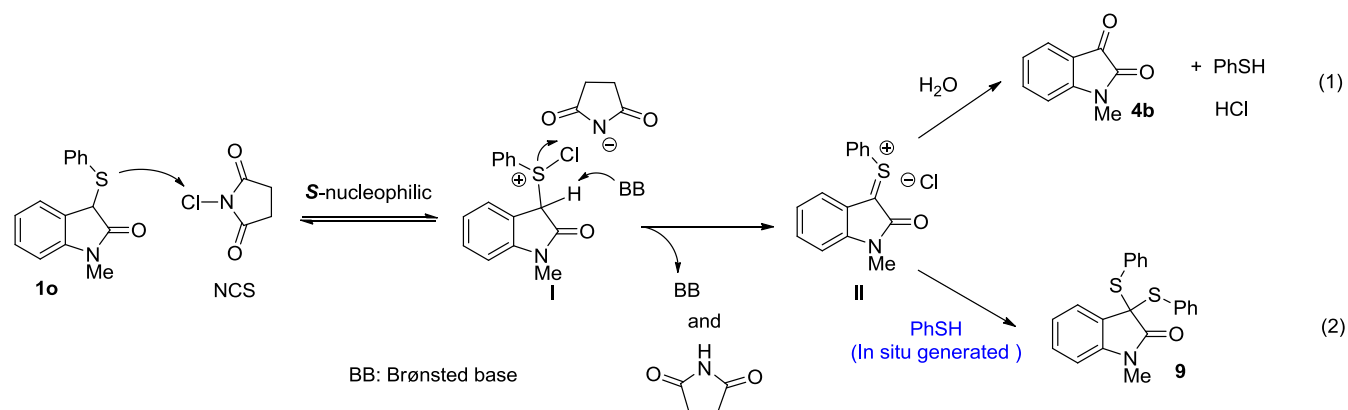
⁷ P. Chauhan, S. Mahajan and D. Enders, *Chem. Rev.*, 2014, **114**, 8807 and reference therein.

⁸ K. S. Boustany and A. B. Sullivan, *Tetrahedron Lett.*, 1970, **11**, 3547.

⁹ F. Zhu, F. Zhou, Z.-Y. Cao, C. Wang, Y.-X. Zhang, C.-H. Wang and J. Zhou, *Synthesis*, 2012, **44**, 3129.

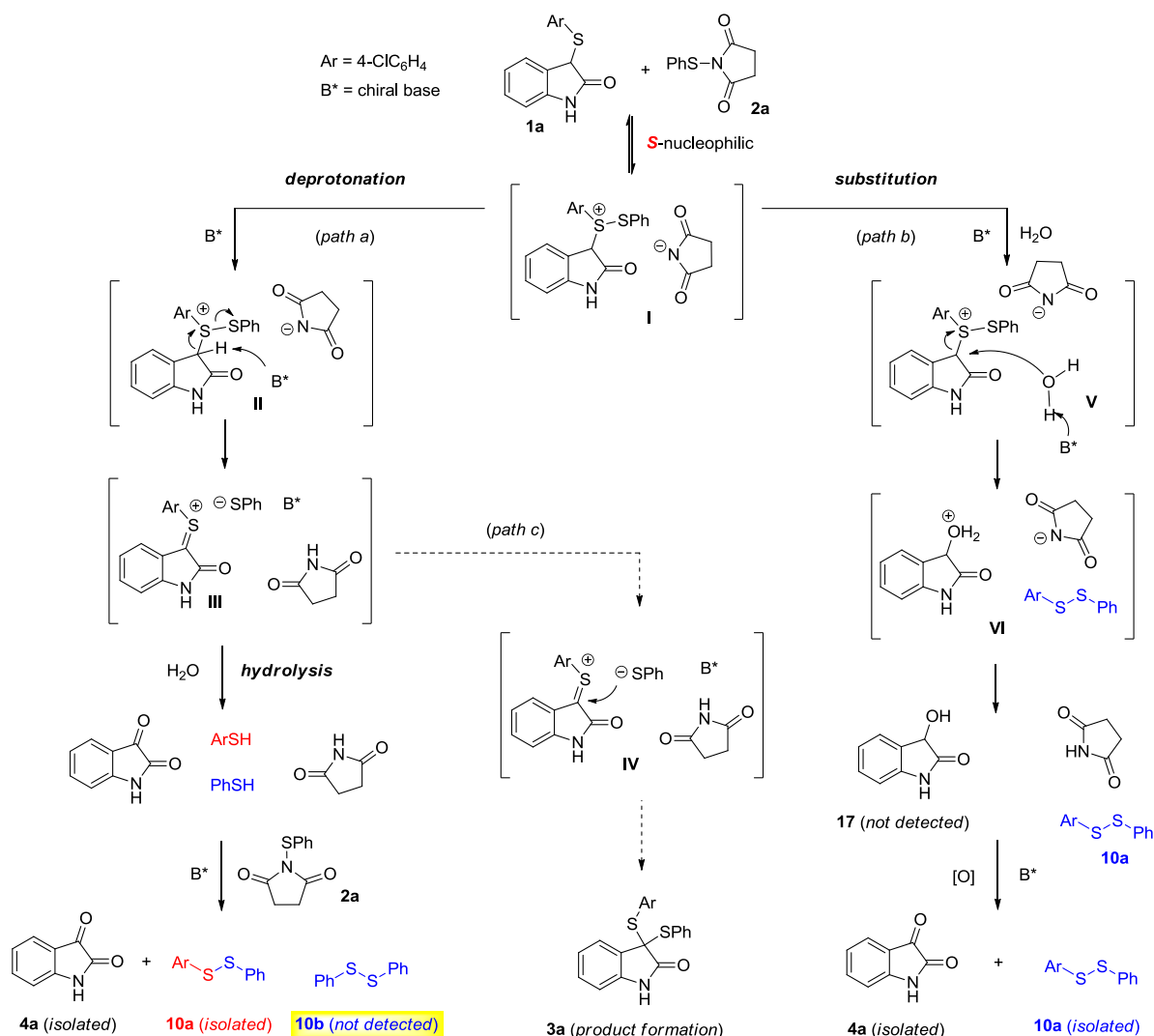
¹⁰ P. Magnus, M. Ladlow and J. Elliott, *J. Am. Chem. Soc.*, 1987, **109**, 7929.

attack of 3-thiooxindole **1o** to NCS would form a sulfonium salt **I**, which then produced a thionium ion intermediate **II** upon deprotonative activation by the base. The following hydrolysis readily afforded isatin **4b** and thiophenol. The in situ generated thiophenol may undergo a nucleophilic addition to sulfur stabilized carbonium ion **II** to give dithioketal **9**, owing to its high nucleophilicity (Scheme S2).



Scheme S2. Possible mechanism for the reaction of 3-thiooxindole **1o** and NCS

While this mechanism reasonably explained the formation of symmetric dithioketal **9** and isatin **4b** in the reaction of 3-thiooxindole **1o** and NCS catalyzed by 10 mol% DABCO, it was insufficient to explain isatin formation in the sulfenylation of 3-thiooxindole. For a detailed discussion, see Scheme S3. If the initial S-nucleophilic attack of **1a** to **2a** afforded sulfonium salt **I** that was deprotonated to give thionium ion **III** (path a). The hydrolysis of thionium ion **III** should produce both *p*-chlorophenyl and phenyl mercaptan. Therefore, both unsymmetrical and symmetric disulfane **10a-b** might be produced via the reaction of the two different thiophenols with *N*-SPh succinimide **2a**, as shown by the control experiments (eq 1 of Scheme S4). Judged by this analysis, both disulfanes **10a-b** should be detected. However, although we isolated isatin in 51% yield and unsymmetrical disulfane **10a** in 35% yield by column chromatography, we failed to isolate symmetric disulfane **10b**. We further tried GC-MS analysis of the reaction mixture to detect whether symmetric disulfane **10b** was formed or not. Unfortunately, since dithioketal **3a** easily decomposed under the EI-analysis condition, and the characteristic peak of unsymmetric disulfane **10a**, symmetric disulfane **10b**, along disulfane **10d** derived from *p*-chlorophenyl mercaptan were all detected, no matter using pure sample of dithioketal **3a** or the reaction mixture for GC-MS analysis (see Figure S2-S3). The LC-MS analysis was also tried, but failed to afford useful information, due to the low molecular weight of disulfane compounds.

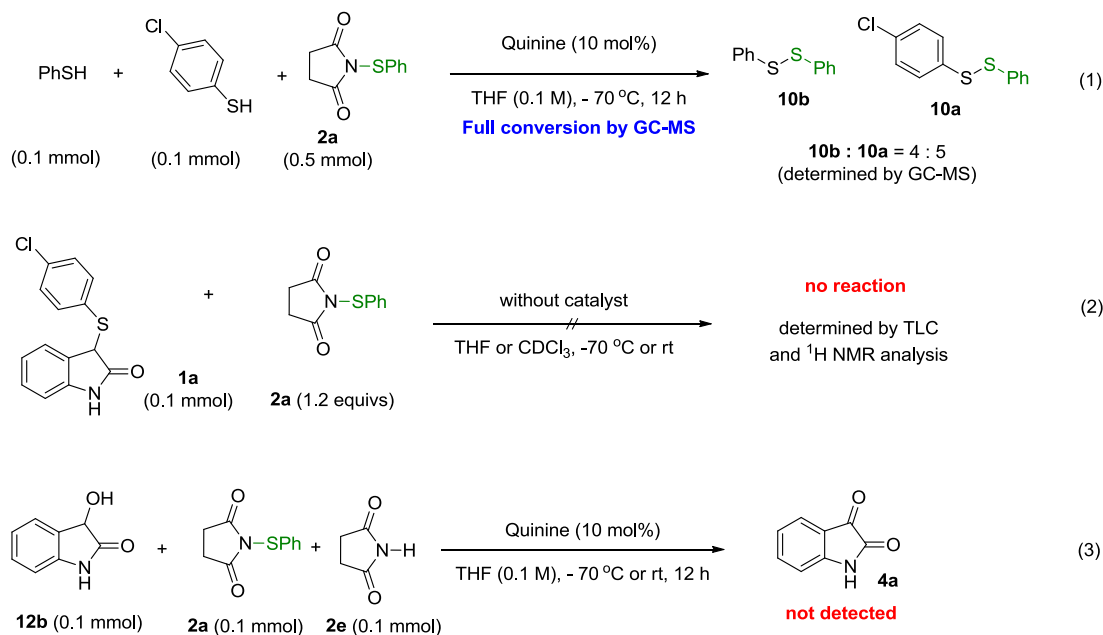


Scheme S3. Possible mechanism for isatin formation in the sulfenylation of 3-thiooxindole **1a**.

To account for why only unsymmetrical disulfane **10a** was produced, along with isatin, in the sulfenylation of 3-thiooxindole **1a**, we considered an alternative substitution reaction pathway (path b, Scheme S3). This possible reaction pathway was based on literature report that dioxindole could be oxidized under basic condition,¹¹ and no reaction occurred if just mixing 3-thiooxindole **1a** and *N*-SPH succinimide **2a** without the addition of base catalyst (eq 2 of Scheme S4). Once the S-nucleophilic attack of **1a** to **2a** produced the sulfonium salt **I**, a further base-assisted nucleophilic attack by water generated **10a** and dioxindole **17**. Then dioxindole was further oxidized to isatin. However, although this route could reasonably explain why only unsymmetrical disulfane **10a** was isolated, it was not in accordance

¹¹ (a) G. Bergonzini and P. Melchiorre, *Angew. Chem., Int. Ed.*, 2012, **51**, 971; (b) Q. Gui, F. Dai, J. Liu, P. Chen, Z. Yang, X. Chen and Z. Tan, *Org. Biomol. Chem.*, 2014, **12**, 3349; (c) G. A. Russell, C. L. Myers, P. Bruni, F. A. Neugebauer and R. Blankespoor, *J. Am. Chem. Soc.*, 1970, **92**, 2762; (d) O. J. Sonderegger, T. Bürgi, L. K. Limbach and A. Baiker, *Journal of Molecular Catalysis A*, 2004, **217**, 93.

with our control experiment that no isatin was detected if stirring a mixture of dioxindole **17**, *N*-SPh succinimide **2a**, succinamide **2e** and 10 mol% quinine in THF at -70 °C or room temperature for 12 hours (eq 3 of Scheme S4). Whether the in situ generated dioxindole might be more reactive to be oxidized under our reaction condition was not clear at current stage.



Scheme S4. Control experiments

Currently, although two possible reaction pathways had been proposed to account for the side isatin formation reaction in the sulfenylation of 3-thiooxindole **1a**, both of them were not in perfect agreement with the experimental data. The detailed mechanism waited for further investigation. Nevertheless, data provided above suggested that side reaction might result from the undesired *S*-nucleophilic attack of 3-thiooxindole **1a** to *N*-SPh succinimide **2a**.

The above analysis also raised an issue whether the thionium ion intermediate **III** might directly convert the desired chiral dithioketal **3a**, in the presence of the chiral catalyst (path c, Scheme S3). To get more information, we monitored the reaction by NMR analysis, as shown in Figure S1.

The NMR analysis of the reaction course provided some useful information. First, without the addition of extra base, no change was monitored even after **1a** and *N*-SPh succinimide **2a** was mixed for 12 h (*a*, *b*, *e* and *f* of Figure S1), which was in accordance with the control experiment that no reaction took place without a base catalyst (eq 2, Scheme S4). Second, the proton at C3 position of 3-thiooxindole **1a** could be easily deprotonated by **C4**, as the characteristic peak of C3 proton of **1a** at δ

4.55 ppm immediately broadened after the addition of 5 mol% catalyst **C4** for 5 min (*a* vs *g*). Third, the reaction proceeded very fast at room temperature, as the characteristic peak of C3 proton of **1a** became very weak after the addition of 5 mol% catalyst **C4** to the mixture of **1a** and *N*-SPh succinimide **2a** for 5 minutes, and then completely disappeared after 20 minutes (*h* and *i* of Figure S1). We also tried lowering the catalyst loading to 1.0 mol% to detect the possible intermediates, but we still failed to detect the characteristic C3 proton of dioxindole (*d* vs *j-l* of Figure S1).

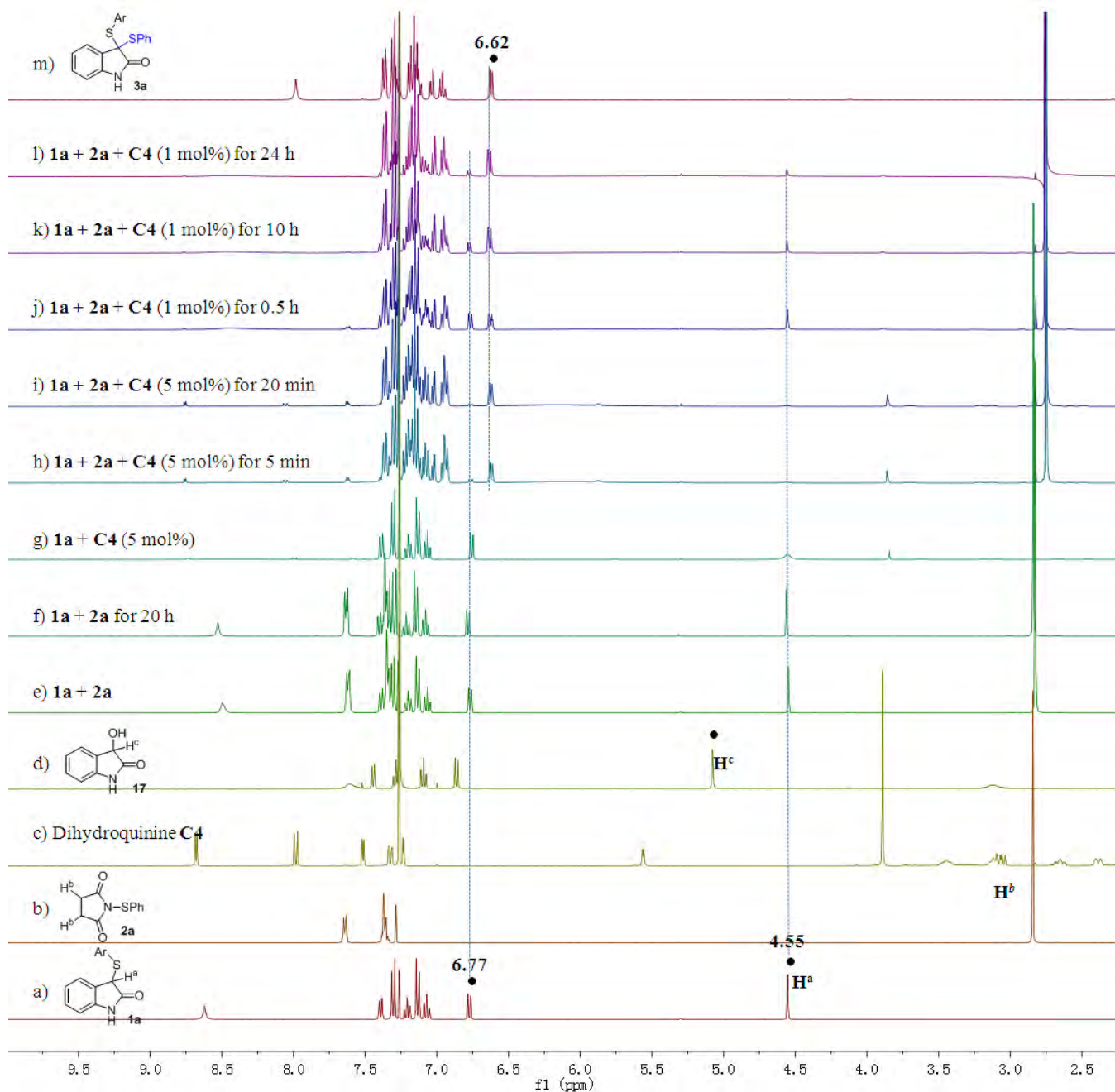
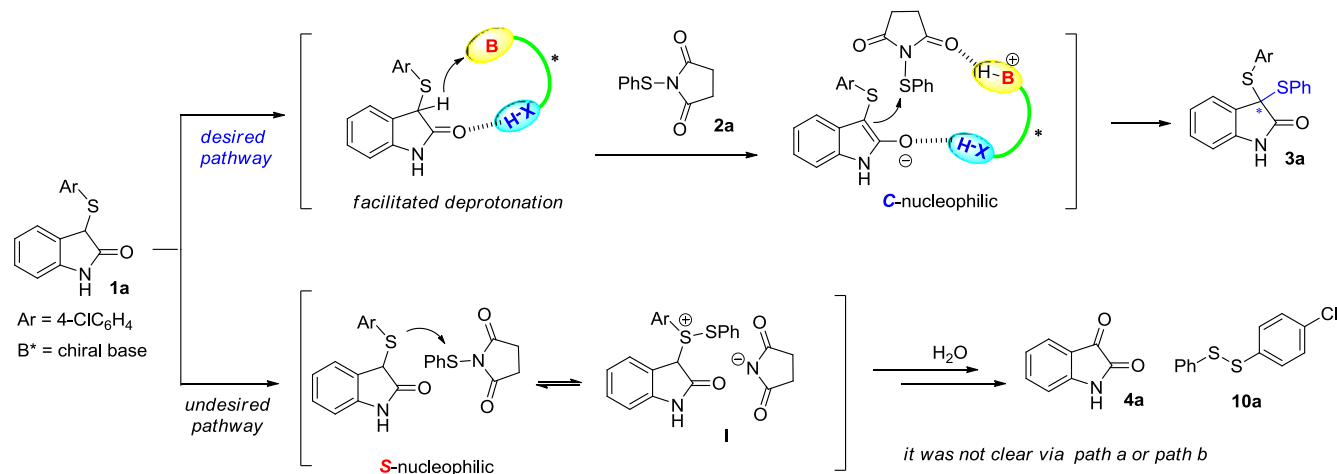


Figure S1. NMR analysis of the reaction course.

Owing to the facile deprotonative activation of C3 proton of 3-thiooxindole **1a**, we tended to believe that the desired chiral dithioketal **3a** was generated via the C-nucleophilic reaction pathway (Scheme S5), rather than via the pathway c shown in Scheme S3, but the detailed mechanism studies waited for further investigation. In addition, the excellent enantioselectivity of dithioketal **3a** implied that chiral catalyst could achieve effective enantiofacial control in the transition state, but it seemed difficult for catalyst **C4** to realize good control over conversion of thionium ion intermediate **III** that included a sulfur stabilized carbonium ion and a benzenethiolate (Scheme S3).

Based on the aforementioned analyses, although the details waited for further studies, we believed that in the base-catalyzed sulfenylation of 3-thiooxindole, the C- and S-nucleophilic pathways were two competitive reaction paths. When reaction was carried out at room temperature, the deprotonative activation of 3-thiooxindole was very easy, so C-nucleophilic pathway dominated to afford the desired dithioketal **3a**, without isatin formation detected. On the other hand, when the catalytic asymmetric reaction was run in low temperature to improve the enantioselectivity, the S-nucleophilic pathway could not be ignored, as the deprotonative activation of 3-thiooxindoles was slowed down greatly. By data accumulated to date, we believed that the formation of a sulfonium salt **I** in the S-nucleophilic pathway was responsible for the formation of isatin side product, but further detailed mechanistic studies were required. Based on this analysis, it was reasonably explained why the use of bifunctional tertiary amine catalysis was effective in suppressing the isatin formation at low reaction temperature, as the H-bonding interactions between 3-thiooxindoles and H-bond donors might facilitate the deprotonative activation, for the desired C-nucleophilic reaction pathway (Scheme S5).



Scheme S5. Bifunctional catalysis in the sulfenylation reaction.

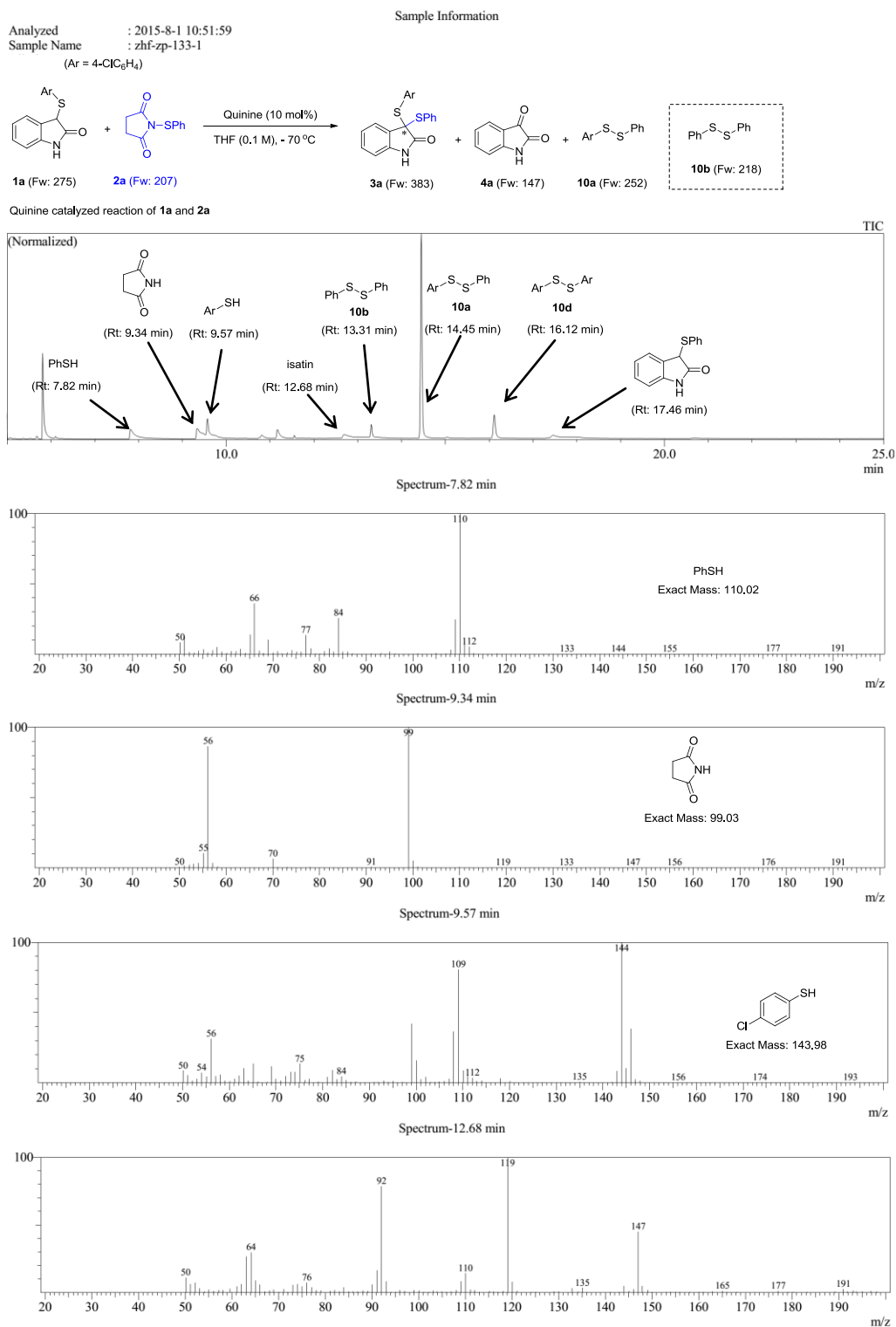


Figure S2-1. GC-MS analysis of the reaction mixture of **1a** and **2a**.

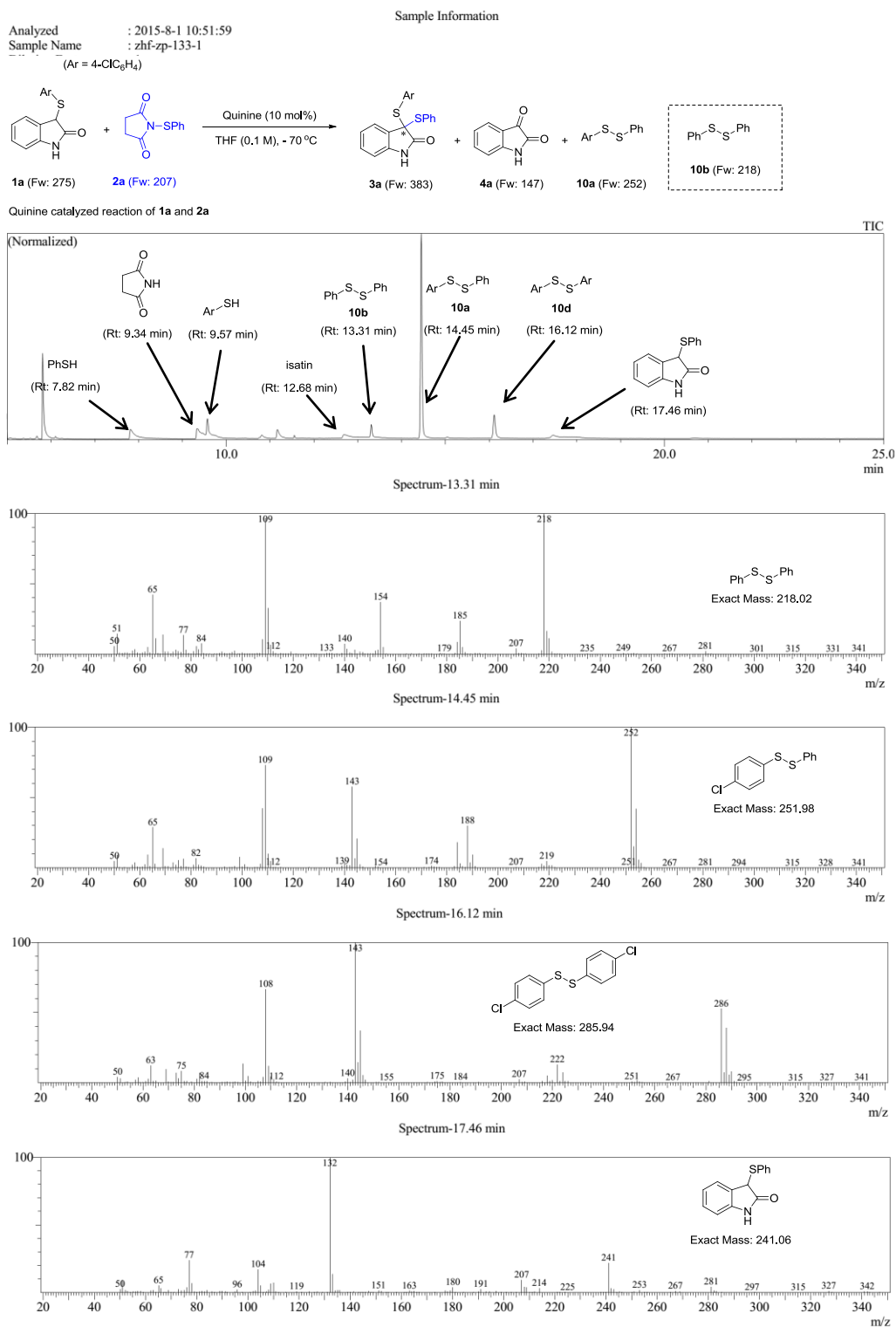


Figure S2-2. GC-MS analysis of the reaction mixture of **1a** and **2a**.

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 Injection Volume : 1.00
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 Org Method File : E:\Data\zhf\Method\0-12程序升温50.5-30-250.20-10-300.10 250 10-1.qgm
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Sample Information

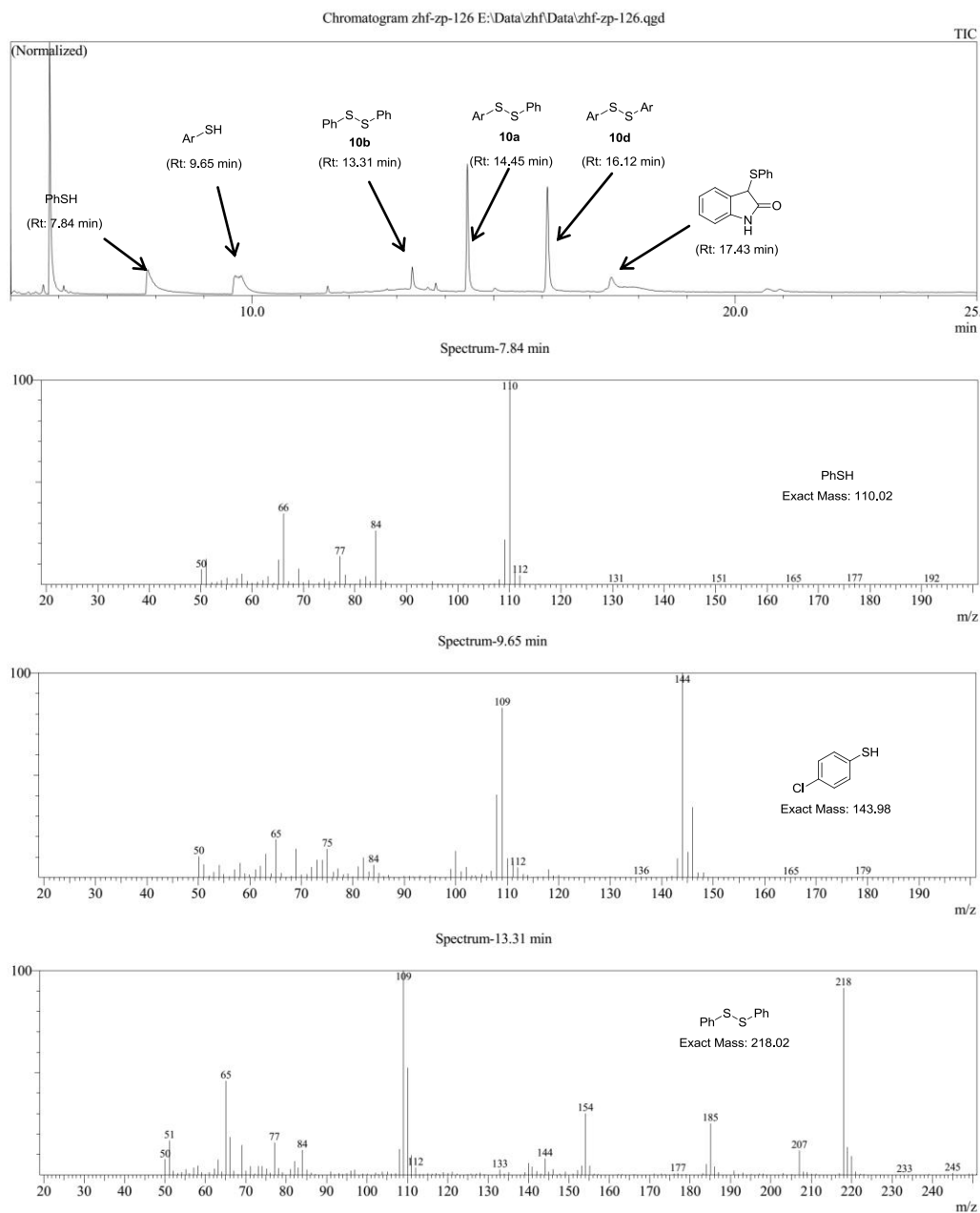
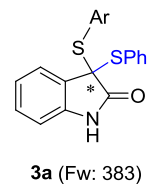


Figure S3-1. GC-MS analysis of the dithioketal **3a**.

Sample Information

Analyzed : 2015-8-1 10:09:37
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 Dilution Factor : 1
 Injection Volume : 1.00
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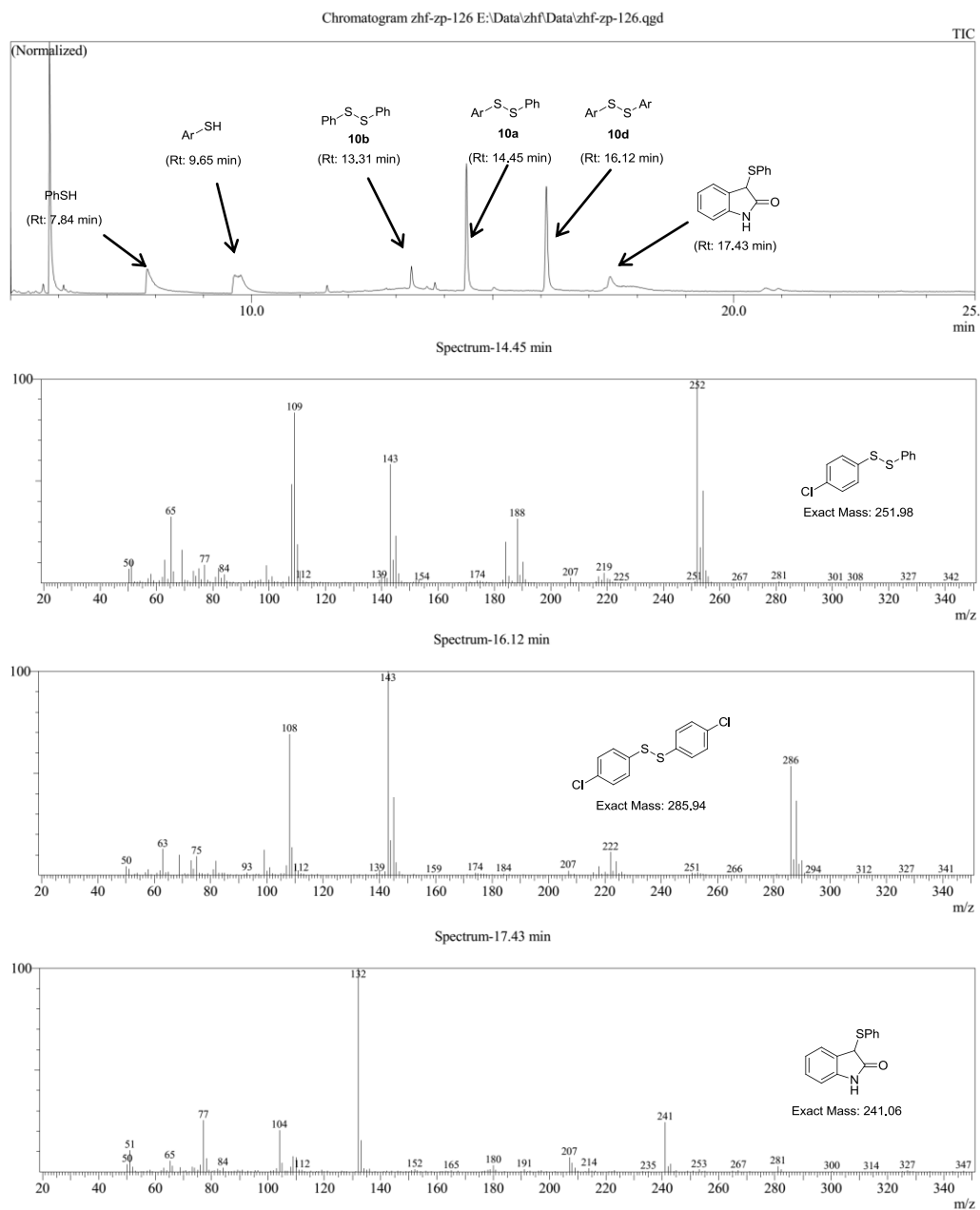
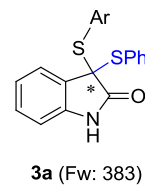


Figure S3-2. GC-MS analysis of the dithioketal **3a**.

2) Condition optimization.

Initially, the reaction of 3-thiooxindole **1a** and *N*-phenylthiosuccinimide **2a** in THF was selected to determine the performance of organocatalysts, typical results were shown in Table S1. With bifunctional catalyst **C1** or **C2**, which we employed for amination^{1a} and Michael addition^{1b} reaction of 3-thiooxindole, respectively, all afforded side product isatin **4a** in significant amount, along with desired product **3a** in low yield and ee value (Table S1, entry 1-2). The isatin formation was also severe when the reaction promoted by secondary amine-phosphoramidate **C2'**, which we developed recently¹² (entry 3). Simple and commercial available quinine **C3** catalyzed the reaction in a much faster rate to give **3a** in 43% ee, together with isatin in 51% yield (entry 4). Then catalysts with a dual H-bond donor were also examined. It was found that isatin formation was almost suppressed when using thiourea **C6** as the catalyst, but **3a** was obtained in only 40% ee (entry 5). The squaramide **C7** as catalyst could afford **3a** in up to 79% ee, albeit with isatin in 43% yield (entry 6). By comparison, **C7** gave a better result in terms of yield and enantioselectivity, but **C3** could promote the reaction much faster than others.

Table S1. Catalyst evaluation.

Clc1ccc(Sc2cc[nH]c2=O)cc1 + O=C1CC(=O)N(Sc2cc[nH]c2=O)C1
 $\xrightarrow[\text{THF (0.1 M), -70 }^{\circ}\text{C}]{\text{Catalyst (10 mol\%)}}$
Clc1ccc(Sc2cc[nH]c2=O)cc1 + O=C1C=CC(=O)Nc2cc[nH]c2=O

1a (0.1 mmol) **2a** (1.2 equivs) **3a** **4a**

C1 **C2** **C2'** **C3** **C6** **C7**

Ar¹ = 3,5-(CF₃)₂C₆H₃

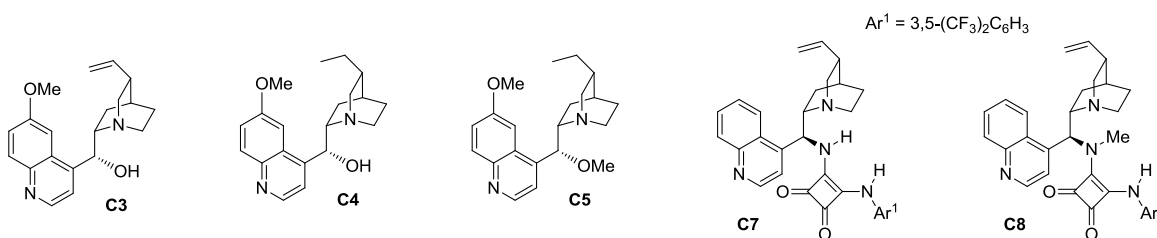
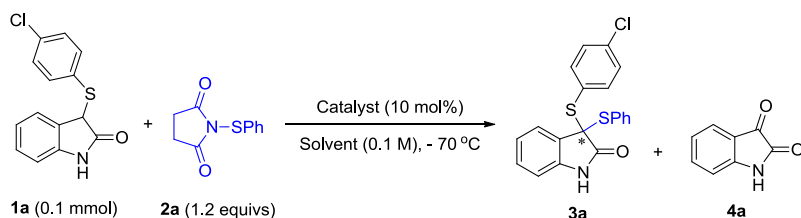
Entry	Cat.	Time(h)	Yield of 3a ^a (%)	Ee of 3a ^b (%)	Yield of 4a ^a (%)
1	C1	84	68	3	23
2	C2	84	20	15	71
3	C2'	84	65	14	30
4	C3	12	46	43	51
5	C6	84	82	40	trace
6	C7	84	54	79	43

^a Isolated yield. ^b Determined by HPLC analysis.

¹² J.-S. Yu, F.-M. Liao, W.-M. Gao, K. Liao, R.-L. Zuo and J. Zhou, *Angew. Chem., Int. Ed.*, 2015, **54**, 7381

Afterward, the effect of solvent for the sulfenylation reaction of 3-thiooxindoles **1a** was examined with squaramide **C7** and quinine **C3** as catalyst respectively, and typical results were shown in Table S2.

Table S2. The solvent effect and control experiments.



Entry	Cat.	Solvent	Time(h)	Yield of 3a ^a (%)	Ee of 3a ^b (%)	Yield of 4a ^a (%)
1	C7	Acetone	84	53	66	22
2	C7	Toluene	84	58	2	trace
3 ^c	C7	CH ₃ CN	84	65	2	trace
4	C7	CH ₂ Cl ₂	84	72	16	21
5 ^d	C7	THF	84	75	79	19
6	C3	Acetone	12	49	68	41
7	C3	Toluene	12	65	67	25
8 ^c	C3	CH ₃ CN	12	81	49	trace
9	C3	CH ₂ Cl ₂	12	82	79	8
10	C4	CH ₂ Cl ₂	12	88	81	7
11 ^e	C4	CH ₂ Cl ₂	24	82	91	7
12 ^f	C4	CH₂Cl₂	36	78	93	trace
13 ^e	C5	CH ₂ Cl ₂	24	70	7	20
14	C8	THF	84	83	29	trace
15	DABCO	THF	24	40	---	56

^a Isolated yield. ^b Determined by HPLC analysis. ^c Run in -40 °C. ^d With 60 mg MS 13X. ^e 0.05 M with 60 mg MS 13X.

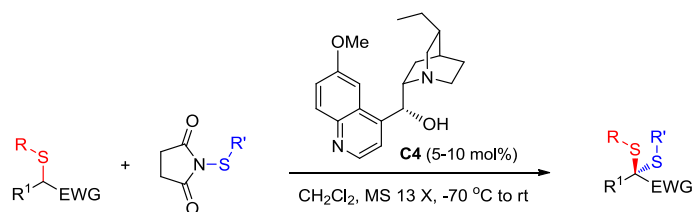
^f 0.05 M with 60 mg MS 13X, 5 mol% of **C4**

With **C7** as catalyst, the reaction using acetone as solvent afforded **3a** in 53% yield with 66% ee (Table S2, entry 1). With toluene and CH₃CN as solvent, the ee decreased dramatically to 2%, despite only trace amount of isatin was detected (entries 2-3). In CH₂Cl₂, the reaction provided **3a** in 16% ee, together with isatin in 21% yield (entry 4). By comparing, squaramide **C8** gave the best result in THF. We further tried using MS 13X as additive, the isatin formation was suppressed in part, but there was no change in the enantioselectivity (entry 5).

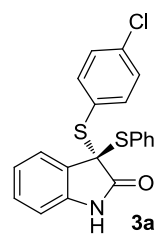
Subsequently, the performance of **C3** in different solvents was examined (Table S2, entries 6-9), and finally CH₂Cl₂ was identified as the best one, which afforded the chiral dithioketal **3a** in 82% yield and 79% ee, with the yield of isatin decreasing to 8% (entries 9). Further exploration revealed that dihydroquinine **C4** could give a better result (entry 10). When the reaction concentration decreased from 0.1 M to 0.05 M, with the addition of MS 13X, the enantioselectivity could be improved to 91% (entry 11). Ultimately, using 5 mol% of dihydroquinine **C4**, along with the addition of MS 13X, allowed **3a** to be obtained in 78% yield and 93% ee, accompanied by trace amount of isatin (entry 12).

In order to declare the importance of bifunctional catalysis in this reaction, control experiments were performed (Table S2, entries 13-15). When the hydroxyl group of dihydroquinine **C4** was changed to methoxyl, the methylated dihydroquinine **C5** not only resulted in poor enantioselectivity, but led to the obvious increase in isatin generation (entry 13 vs 11). With one N-H bond of squamide **C7** was masked by a methyl group, the corresponding catalyst **C8**, could inhibit isatin formation, but the ee value of **3a** dropped from 79% to 29% (Table S2, entry 14 vs Table S1, entry 6). When racemic catalyst DABCO (1,4-diazabicyclo[2.2.2]octane) was used under -70 °C, the achiral dithioketal **3a** could only be obtained in 40% yield, along with large amount of isatin formed (entry 15). The above results revealed that a suitable H-bond donor might be important for the reaction, which could not only restrain the undesired S-nucleophilic pathway, but also contribute to achieving high enantioselectivity.

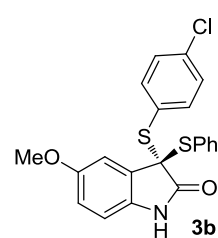
3) General procedure for the catalytic asymmetric sulfenylation reaction.



To a 10.0 mL vial were added dihydroquinine **C4** (0.0125 or 0.025 mmol) and S-based nucleophiles (0.25 mmol), followed by 5.0 mL of CH₂Cl₂ and 150 mg MS 13X. The resulting mixture was stirred at indicated temperature for about 30 min before the corresponding electrophilic sulfenylation reagents (0.30 mmol) was added. After the full conversion of S-based nucleophiles by TLC analysis, the solvent was carefully removed under reduced pressure. The residue was directly subjected to flash column chromatography to afford the desired chiral dithioketals, using the indicated eluent.

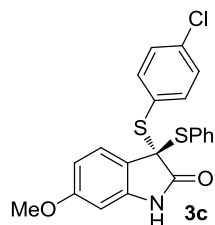


Column chromatography (PE/EtOAc = 6:1 to 3:1) afforded the desired product **3a** in 78% yield as pale pink solid (m. p. 134-136 °C). HPLC analysis (Chiralpak AD-H, *i*PrOH/hexane = 10/90, 1.0 mL/min, 230 nm; *t_r*(minor) = 16.537 min, *t_r*(major) = 15.396 min) gave the isomeric composition of the product: 93% ee, $[\alpha]_D^{28} = -17.0$ (*c* = 0.91, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.98 (s, 1H), 7.38-7.36 (m, 2H), 7.32-7.28 (m, 3H), 7.20-7.11 (m, 5H), 7.04-7.03 (m, 1H), 6.96 (t, *J* = 7.6 Hz, 1H), 6.62 (d, *J* = 7.6 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃): δ 175.72, 139.60, 137.71, 136.50, 136.27, 129.81, 129.57, 129.19, 128.77, 128.58, 127.97, 127.91, 125.51, 122.52, 110.34, 63.06; IR (KBr): 3240, 1735, 1701, 1616, 1470, 1093, 746, 685; HRMS (ESI): Exact mass calcd for C₂₀H₁₄NOS₂³⁵Cl²³Na [M+Na]⁺: 406.0103, Found: 406.0108.

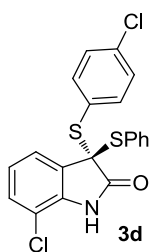


Column chromatography (PE/EtOAc = 6:1 to 3:1) afforded the desired product **3b** in 73% yield as pale pink solid (m. p. 146-148 °C). HPLC analysis (Chiralpak OZ-H, *i*PrOH/hexane = 3/97, 1.0 mL/min, 254 nm; *t_r*(minor) = 30.920 min, *t_r*(major) = 39.240 min) gave the isomeric composition of the product: 96% ee, $[\alpha]_D^{27} = -34.1$ (*c* = 1.00, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.72 (s, 1H), 7.40-7.38 (m, 2H), 7.33-7.31 (m, 3H), 7.23-7.19 (m, 2H), 7.18-7.16 (m, 2H), 6.68 (dd, *J* = 8.4 Hz, *J* = 2.8 Hz, 1H),

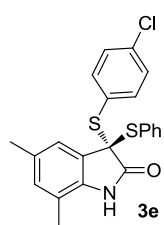
6.57-6.52 (m, 2H), 3.69 (s, 3H); ^{13}C NMR (100 MHz, CDCl_3): δ 175.63, 155.51, 137.62, 136.49, 136.25, 132.99, 129.82, 129.25, 129.02, 128.80, 128.63, 128.02, 115.51, 111.16, 110.99, 63.42, 55.67; IR (KBr): 3448, 3178, 1690, 1484, 1304, 1205, 748, 691; HRMS (EI): Exact mass calcd for $\text{C}_{21}\text{H}_{16}\text{NO}_2\text{S}_2^{35}\text{Cl}$ $[\text{M}]^+$: 413.0311, Found: 413.0307.



Column chromatography (PE/EtOAc = 6:1 to 3:1) afforded the desired product **3c** in 70% yield as brown solid (m. p. 157-159 °C). HPLC analysis (Chiralpak OZ-H, *i*PrOH/hexane = 3/97, 1.0 mL/min, 230 nm; t_r (minor) = 26.940 min, t_r (major) = 29.887 min) gave the isomeric composition of the product: 87% ee, $[\alpha]_D^{29} = -15.7$ ($c = 1.04$, CHCl_3); ^1H NMR (400 MHz, CDCl_3): δ 9.14 (s, 1H), 7.37-7.35 (m, 2H), 7.31-7.26 (m, 3H), 7.19-7.13 (m, 4H), 6.82 (d, $J = 8.4$ Hz, 1H), 6.45 (dd, $J = 8.4$ Hz, $J = 2.4$ Hz, 1H), 6.30 (d, $J = 2.0$ Hz, 1H), 3.74 (s, 3H); ^{13}C NMR (100 MHz, CDCl_3): δ 176.43, 160.98, 140.92, 137.70, 136.50, 136.20, 129.75, 129.46, 128.79, 128.61, 128.20, 126.42, 119.41, 108.09, 96.88, 63.19, 55.47; IR (KBr): 3159, 1734, 1622, 1455, 1311, 1158, 1018, 752; HRMS (EI): Exact mass calcd for $\text{C}_{21}\text{H}_{16}\text{NO}_2\text{S}_2^{35}\text{Cl}$ $[\text{M}]^+$: 413.0311, Found: 413.0312.

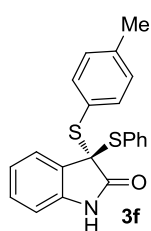


Column chromatography (PE/EtOAc = 6:1 to 3:1) afforded the desired product **3d** in 85% yield as pink solid (m. p. 116-118 °C). HPLC analysis (Chiralpak OZ-H, *i*PrOH/hexane = 3/97, 1.0 mL/min, 230 nm; t_r (minor) = 17.500 min, t_r (major) = 20.433 min) gave the isomeric composition of the product: 91% ee, $[\alpha]_D^{26} = -11.5$ ($c = 1.05$, CHCl_3); ^1H NMR (400 MHz, CDCl_3): δ 7.40 (s, 1H), 7.39-7.38 (m, 1H), 7.37-7.36 (m, 1H), 7.35-7.34 (m, 1H), 7.33-7.31 (m, 2H), 7.24-7.18 (m, 4H), 7.12 (dd, $J = 7.6$ Hz, $J = 2.0$ Hz, 1H), 6.96-6.89 (m, 2 H); ^{13}C NMR (100 MHz, CDCl_3): δ 174.20, 137.97, 137.36, 136.74, 136.67, 130.17, 129.60, 129.46, 129.00, 128.91, 128.79, 127.61, 123.89, 123.35, 115.19, 63.47; IR (KBr): 3447, 1715, 1617, 1473, 1457, 1091, 753, 505; HRMS (EI): Exact mass calcd for $\text{C}_{20}\text{H}_{13}\text{NOS}_2^{35}\text{Cl}_2$ $[\text{M}]^+$: 416.9816, Found: 416.9817.



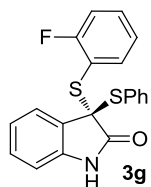
Column chromatography (PE/EtOAc = 6:1 to 3:1) afforded the desired product **3e** in 87% yield as pink solid (m. p. 146-148 °C). HPLC analysis (Chiralpak OZ-H, *i*PrOH/hexane = 3/97, 1.0 mL/min, 230 nm; t_r (minor) = 20.853 min, t_r (major) = 24.233 min) gave the isomeric composition of the product: 93% ee, $[\alpha]_D^{27} = -22.9$ (c = 1.00, CHCl₃);

¹H NMR (400 MHz, CDCl₃): δ 8.50 (s, 1H), 7.39-7.38 (m, 1H), 7.37-7.36 (m, 1H), 7.34-7.32 (m, 2H), 7.31-7.29 (m, 1H), 7.21-7.15 (m, 4H), 6.77 (s, 1H), 6.59 (s, 1 H), 2.21 (s, 3 H), 2.04 (s, 3 H); ¹³C NMR (100 MHz, CDCl₃): δ 176.14, 137.77, 136.63, 136.15, 136.08, 131.98, 131.57, 129.76, 129.69, 128.71, 128.54, 128.48, 127.59, 123.47, 119.52, 63.39, 21.06, 16.22; IR (KBr): 3173, 3076, 2919, 1705, 1626, 1475, 828, 508; HRMS (EI): Exact mass calcd for C₂₂H₁₈NOS₂³⁵Cl [M]⁺: 411.0518, Found: 411.0515.



Column chromatography (PE/EtOAc = 6:1 to 3:1) afforded the desired product **3f** in 70% yield as white solid (m. p. 151-153 °C). HPLC analysis (Chiralpak AD-H, *i*PrOH/hexane = 10/90, 1.0 mL/min, 230 nm; t_r (minor) = 19.452 min, t_r (major) = 15.965 min) gave the isomeric composition of the product: 88% ee, $[\alpha]_D^{28} = 5.4$ (c = 1.00, CHCl₃);

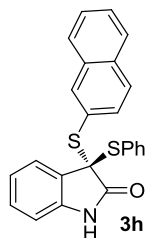
¹H NMR (400 MHz, CDCl₃): δ 8.37 (s, 1H), 7.36-7.34 (m, 2H), 7.28-7.24 (m, 3H), 7.17-7.13 (m, 2H), 7.10 (td, *J* = 7.6 Hz, *J* = 1.6 Hz, 1 H), 7.01-6.97 (m, 3H), 6.93 (td, *J* = 7.6 Hz, *J* = 1.2 Hz, 1 H), 6.62 (d, *J* = 7.6 Hz, 1H), 2.27 (s, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 176.13, 140.14, 139.83, 136.71, 136.43, 129.79, 129.68, 129.47, 129.34, 128.61, 128.53, 125.98, 125.71, 122.39, 110.30, 63.28, 21.34; IR (KBr): 3410, 3188, 1736, 1693, 1469, 1324, 747, 687; HRMS (EI): Exact mass calcd for C₂₁H₁₇NOS₂ [M]⁺: 363.0752, Found: 363.0754.



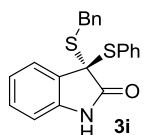
Column chromatography (PE/EtOAc = 6:1 to 3:1) afforded the desired product **3g** in 57% yield as white solid (m. p. 192-194 °C). HPLC analysis (Chiralpak OZ-H, *i*PrOH/hexane = 10/90, 1.0 mL/min, 230 nm; t_r (minor) = 11.751 min, t_r (major) = 10.054 min) gave the isomeric composition of the product: 90% ee, $[\alpha]_D^{28} = -5.9$ (c = 0.60, CHCl₃);

¹H NMR (400 MHz, CDCl₃): δ 8.62 (s, 1H), 7.50 (td, *J* = 7.6 Hz, *J* = 1.6 Hz, 1 H), 7.38-7.35 (m, 2H), 7.29-7.26 (m, 2H), 7.19-7.15 (m, 2H), 7.09 (td, *J* = 7.6 Hz, *J* = 1.6 Hz, 1 H), 7.00 (td, *J* = 7.6 Hz, *J* = 1.2 Hz, 1 H), 6.95-6.86 (m, 3H), 6.65 (d, *J* = 7.6 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃): δ

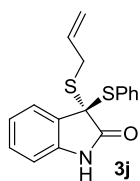
175.32, 163.54 ($J = 248$ Hz), 139.57, 138.39, 136.60, 132.36 ($J = 8$ Hz), 129.83, 129.59, 129.53, 128.63, 127.83, 125.81, 124.19 ($J = 4$ Hz), 122.42, 116.94 ($J = 8$ Hz), 115.78 ($J = 24$ Hz), 110.89, 63.01; IR (KBr): 3415, 3187, 1731, 1728, 1469, 1227, 824, 746; HRMS (EI): Exact mass calcd for $C_{20}H_{14}NOS_2F$ $[M]^+$: 367.0501, Found: 367.0507.



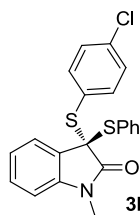
Column chromatography (PE/EtOAc = 6:1 to 3:1) afforded the desired product **3h** in 67% yield as pale pink solid (m. p. 130-132 °C). HPLC analysis (Chiralpak IC, i PrOH/hexane = 10/90, 1.0 mL/min, 230 nm; t_r (minor) = 15.332 min, t_r (major) = 18.502 min) gave the isomeric composition of the product: 87% ee, $[\alpha]_D^{28} = -6.0$ ($c = 1.00$, $CHCl_3$); 1H NMR (400 MHz, $CDCl_3$): δ 7.89 (s, 1H), 7.75-7.72 (m, 2H), 7.67-7.65 (m, 1H), 7.62-7.60 (m, 1H), 7.47-7.42 (m, 2 H), 7.40-7.38 (m, 3H), 7.29-7.27 (m, 1H), 7.20-7.16 (m, 2H), 7.08-7.05 (m, 2H), 6.94-6.91 (m, 1H), 6.52-6.50 (m, 1H); ^{13}C NMR (100 MHz, $CDCl_3$): δ 175.93, 139.76, 136.91, 136.56, 133.45, 133.17, 132.64, 129.80, 129.64, 129.50, 128.67, 128.42, 128.16, 128.07, 127.62, 127.22, 126.90, 126.37, 125.73, 122.49, 110.32, 63.34; IR (KBr): 3448, 3186, 1751, 1701, 1467, 1182, 746, 481; HRMS (EI): Exact mass calcd for $C_{24}H_{17}NOS_2$ $[M]^+$: 399.0752, Found: 399.0755.



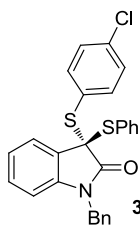
Column chromatography (PE/EtOAc = 10:1 to 5:1) afforded the desired product **3i** in 84% yield as white solid (m. p. 137-139 °C). HPLC analysis (Chiralpak AS-H, i PrOH/hexane = 20/80, 1.0 mL/min, 230 nm; t_r (minor) = 18.567 min, t_r (major) = 16.320 min) gave the isomeric composition of the product: 91% ee, $[\alpha]_D^{26} = -0.6$ ($c = 0.90$, $CHCl_3$); 1H NMR (400 MHz, $CDCl_3$): δ 7.70 (s, 1H), 7.34-7.33 (m, 1H), 7.32-7.31 (m, 1H), 7.30-7.29 (m, 1H), 7.28-7.27 (m, 3H), 7.26-7.25 (m, 1H), 7.24-7.22 (m, 1H), 7.21-7.20 (m, 1H), 7.19-7.16 (m, 2H), 7.15-7.14 (m, 1H), 7.00 (td, $J = 7.6$ Hz, $J = 1.2$ Hz, 1H), 6.71 (d, $J = 7.6$ Hz, 1H), 4.22, 4.19 (AB, $J = 12.0$ Hz, 2H); ^{13}C NMR (100 MHz, $CDCl_3$): δ 175.15, 139.31, 136.53, 136.12, 129.78, 129.60, 129.56, 129.43, 128.62, 128.47, 128.43, 127.31, 125.29, 122.83, 109.95, 60.38, 34.83; IR (KBr): 3449, 3241, 1724, 1494, 1232, 747, 702, 498; HRMS (EI): Exact mass calcd for $C_{21}H_{17}NOS_2$ $[M]^+$: 363.0752, Found: 363.0748.



Column chromatography (PE/EtOAc = 10:1 to 5:1) afforded the desired product **3j** in 56% yield as white solid (m. p. 51-53 °C). HPLC analysis (Chiralpak OZ-H, *i*PrOH/hexane = 3/97, 1.0 mL/min, 230 nm; t_r (minor) = 10.453 min, t_r (major) = 11.473 min) gave the isomeric composition of the product: 83% ee, $[\alpha]_D^{29} = -7.2$ ($c = 1.09$, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 8.91 (s, br, 1H), 7.34-7.28 (m, 3H), 7.20-7.15 (m, 3H), 7.11 (d, $J = 6.8$ Hz, 1H), 7.00 (td, $J = 7.6$ Hz, $J = 0.8$ Hz, 1H), 6.80 (d, $J = 7.6$ Hz, 1H), 5.86-5.76 (m, 1H), 5.08-5.00 (m, 2H), 3.53 (d, $J = 6.8$ Hz, 2H); ¹³C NMR (100 MHz, CDCl₃): δ 176.11, 139.56, 136.66, 132.83, 129.89, 129.64, 129.47, 128.63, 128.50, 125.29, 122.82, 118.44, 110.26, 60.18, 33.79; IR (KBr): 3448, 1733, 1685, 1522, 1472, 1458, 1384, 690; HRMS (EI): Exact mass calcd for C₁₇H₁₅NOS₂ [M]⁺: 313.0595, Found: 313.0597.

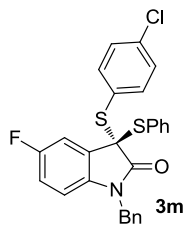


Column chromatography (PE/EtOAc = 20:1 to 15:1) afforded the desired product **3k** in 97% yield as pale pink solid (m. p. 102-104 °C). HPLC analysis (Chiralpak AD-H, *i*PrOH/hexane = 3/97, 1.0 mL/min, 230 nm; t_r (minor) = 19.953 min, t_r (major) = 18.033 min) gave the isomeric composition of the product: 90% ee, $[\alpha]_D^{29} = -26.5$ ($c = 1.00$, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.35-7.32 (m, 2H), 7.30-7.27 (m, 3H), 7.20-7.14 (m, 5H), 7.03-7.01 (m, 1H), 6.96 (td, $J = 7.2$ Hz, $J = 0.8$ Hz, 1H), 6.49 (d, $J = 7.6$ Hz, 1H), 2.90 (s, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 173.05, 142.26, 137.89, 136.75, 136.26, 129.85, 129.60, 129.27, 128.70, 128.51, 128.06, 127.73, 125.41, 122.61, 108.05, 62.63, 26.37; IR (KBr): 3421, 3082, 1705, 1607, 1508, 1093, 759, 540; HRMS (EI): Exact mass calcd for C₂₁H₁₆NOS₂³⁵Cl [M]⁺: 397.0362, Found: 397.0360.

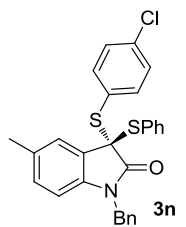


Column chromatography (PE/EtOAc = 20:1 to 15:1) afforded the desired product **3l** in 98% yield as yellow oil. HPLC analysis (Chiralpak AD-H, *i*PrOH/hexane = 3/97, 1.0 mL/min, 230 nm; t_r (minor) = 67.035 min, t_r (major) = 57.884 min) gave the isomeric composition of the product: 94% ee, $[\alpha]_D^{29} = -36.5$ ($c = 0.97$, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.38-7.36 (m, 2H), 7.33-7.31 (m, 1H), 7.28-7.27 (m, 1H), 7.27-7.26 (m, 2H), 7.20 (t, $J = 1.6$ Hz, 1H), 7.19-7.18 (m, 3H), 7.17-7.16 (m, 1H), 7.13-7.11 (m, 2H), 7.05 (td, $J = 7.6$ Hz, $J = 1.2$ Hz, 1H), 6.97 (td, $J = 7.6$ Hz, $J = 1.2$ Hz, 1H), 6.72-6.70 (m, 2H), 6.35 (d, $J = 7.6$ Hz, 1H), 4.69, 4.55 (AB, $J = 16.0$ Hz, 2H); ¹³C NMR (100 MHz, CDCl₃): δ 172.62, 141.40, 137.88, 136.81, 136.21, 134.71, 129.80, 129.44, 129.03, 128.73, 128.52, 128.46, 127.87, 127.66, 127.30, 126.62, 125.34, 122.52, 109.11, 62.68,

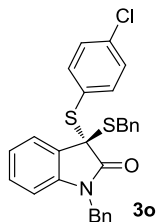
43.85; IR (KBr): 3503, 3038, 1710, 1608, 1467, 1092, 749, 692; HRMS (ESI): Exact mass calcd for $C_{27}H_{21}^{35}ClNOS_2$ $[M+H]^+$: 474.0748, Found: 474.0749.



Column chromatography (PE/EtOAc = 20:1 to 15:1) afforded the desired product **3m** in 97% yield as pink oil. HPLC analysis (Chiralpak AD-H, *i*PrOH/hexane = 10/90, 1.0 mL/min, 230 nm; t_r (minor) = 20.867 min, t_r (major) = 18.487 min) gave the isomeric composition of the product: 91% ee, $[\alpha]_D^{28} = -46.2$ ($c = 1.04$, $CHCl_3$); 1H NMR (400 MHz, $CDCl_3$): δ 7.39-7.36 (m, 3H), 7.30-7.28 (m, 2H), 7.23-7.14 (m, 7H), 6.92 (dd, $J = 8.0$ Hz, $J = 2.8$ Hz, 1H), 6.75 (td, $J = 8.8$ Hz, $J = 2.8$ Hz, 1H), 6.69-6.67 (m, 2H), 6.24 (dd, $J = 8.8$ Hz, $J = 4.4$ Hz, 1H), 4.68, 4.53 (AB, $J = 16.0$ Hz, 2H); ^{13}C NMR (100 MHz, $CDCl_3$): δ 172.65, 158.92 ($J = 240$ Hz, 1C), 138.08, 137.48, 137.05, 136.66, 134.55, 130.24, 129.65 ($J = 2.0$ Hz, 1C), 129.07, 128.86, 128.72, 127.63, 116.06 ($J = 23$ Hz, 1C), 113.18 ($J = 25$ Hz, 1C), 110.04, 109.96, 62.91, 44.18; IR (KBr): 3064, 1720, 1610, 1167, 747, 693, 597, 505; HRMS (EI): Exact mass calcd for $C_{27}H_{19}NOS_2^{35}ClF$ $[M]^+$: 491.0581, Found: 491.0583.

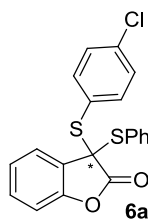


Column chromatography (PE/EtOAc = 20:1 to 15:1) afforded the desired product **3n** in 99% yield as yellow oil. HPLC analysis (Chiralpak AD-H, *i*PrOH/hexane = 10/90, 1.0 mL/min, 230 nm; t_r (minor) = 19.753 min, t_r (major) = 18.200 min) gave the isomeric composition of the product: 94% ee, $[\alpha]_D^{27} = -48.1$ ($c = 1.06$, $CHCl_3$); 1H NMR (400 MHz, $CDCl_3$): δ 7.38-7.32 (m, 3H), 7.28-7.27 (m, 1H), 7.26-7.25 (m, **1H**), 7.21-7.11 (m, 7H), 6.95 (s, 1H), 6.85 (dd, $J = 8.0$ Hz, $J = 0.8$ Hz, 1H), 6.72-6.69 (m, 2H), 6.25 (d, $J = 8.0$ Hz, 1H), 4.68, 4.54 (AB, $J = 16.0$ Hz, 2H), 2.25 (s, 3H); ^{13}C NMR (100 MHz, $CDCl_3$): δ 172.69, 139.18, 138.01, 137.02, 136.31, 135.03, 132.23, 129.94, 129.92, 129.38, 128.86, 128.65, 128.60, 128.25, 127.72, 127.42, 126.81, 126.12, 109.04, 62.89, 44.03, 21.10; IR (KBr): 3427, 3061, 2919, 1723, 1494, 1092, 747, 504; HRMS (EI): Exact mass calcd for $C_{28}H_{22}NOS_2^{35}Cl$ $[M]^+$: 487.0831, Found: 487.0828.

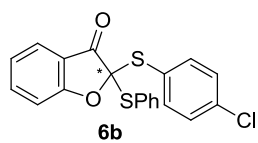


Column chromatography (PE/EtOAc = 20:1 to 15:1) afforded the desired product **3o** in 70% yield as orange solid (m. p. 45-47 °C). HPLC analysis (Chiralpak AD-H, *i*PrOH/hexane = 10/90, 1.0 mL/min, 230 nm; t_r (minor) = 24.613 min, t_r (major) = 13.920

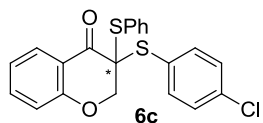
min) gave the isomeric composition of the product: 81% ee, $[\alpha]_D^{29} = -16.6$ ($c = 1.05$, CHCl_3); ^1H NMR (400 MHz, CDCl_3): δ 7.37 (dd, $J = 7.6$ Hz, $J = 1.2$ Hz, 1H), 7.32-7.28 (m, 4H), 7.24-7.19 (m, 6H), 7.15-7.08 (m, 3H), 7.04 (td, $J = 7.6$ Hz, $J = 1.2$ Hz, 1H) 6.83-6.80 (m, 2H), 6.50 (d, $J = 7.6$ Hz, 1H), 4.95, 4.45 (AB, $J = 15.6$ Hz, 2H), 4.34, 4.29 (AB, $J = 12.0$ Hz, 2H); ^{13}C NMR (100 MHz, CDCl_3): δ 172.85, 141.41, 137.85, 136.36, 136.20, 135.03, 129.79, 129.51, 129.04, 128.74, 128.60, 128.42, 127.93, 127.62, 127.39, 126.87, 124.95, 123.09, 109.52, 58.89, 44.10, 34.61; IR (KBr): 3448, 2922, 1716, 1606, 1485, 1355, 1092, 696; HRMS (EI): Exact mass calcd for $\text{C}_{28}\text{H}_{22}\text{NOS}_2^{35}\text{Cl}$ $[\text{M}]^+$: 487.0831, Found: 487.0828.



Column chromatography (PE/EtOAc = 20:1 to 15:1) afforded the desired product **6a** in 95% yield as orange solid (m. p. 60-62 °C). HPLC analysis (Chiralpak AD-H, i PrOH/hexane = 3/97, 1.0 mL/min, 230 nm; t_r (minor) = 11.040 min, t_r (major) = 13.853 min) gave the isomeric composition of the product: 87% ee, $[\alpha]_D^{29} = +13.6$ ($c = 0.98$, CHCl_3); ^1H NMR (400 MHz, CDCl_3): δ 7.38-7.30 (m, 5H), 7.24-7.23 (m, 1H), 7.22-7.19 (m, 5H), 7.13-7.09 (m, 1H), 6.82-6.80 (m, 1H); ^{13}C NMR (100 MHz, CDCl_3): δ 172.37, 151.94, 138.03, 137.04, 136.83, 130.60, 130.45, 129.20, 128.98, 128.51, 127.20, 126.45, 125.48, 124.49, 110.76, 60.95; IR (KBr): 3448, 1797, 1637, 1463, 1230, 1092, 748, 685; HRMS (EI): Exact mass calcd for $\text{C}_{20}\text{H}_{13}\text{O}_2\text{S}_2^{35}\text{Cl}$ $[\text{M}]^+$: 384.0046, Found: 384.0048.

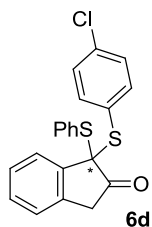


Column chromatography (PE/EtOAc = 20:1 to 15:1) afforded the desired product **6b** in 85% yield as yellow solid (m. p. 69-71 °C). HPLC analysis (Chiralpak OD-H, i PrOH/hexane = 1/99, 1.0 mL/min, 230 nm; t_r (minor) = 16.600 min, t_r (major) = 18.247 min) gave the isomeric composition of the product: 95% ee, $[\alpha]_D^{29} = -1.3$ ($c = 1.04$, CHCl_3); ^1H NMR (400 MHz, CDCl_3): δ 7.55-7.53 (m, 2H), 7.51-7.46 (m, 3H), 7.41 (d, $J = 7.6$ Hz, 1H), 7.29-7.26 (m, 1H), 7.22-7.18 (m, 4H), 6.95-6.90 (m, 2H); ^{13}C NMR (100 MHz, CDCl_3): δ 194.05, 169.39, 138.61, 137.79, 136.62, 136.55, 130.04, 129.01, 128.80, 127.80, 126.57, 124.78, 122.72, 119.66, 112.59, 99.24; IR (KBr): 3421, 3080, 1750, 1647, 1474, 1092, 758, 518; HRMS (EI): Exact mass calcd for $\text{C}_{20}\text{H}_{13}\text{O}_2\text{S}_2^{35}\text{Cl}$ $[\text{M}]^+$: 384.0046, Found: 384.0049.



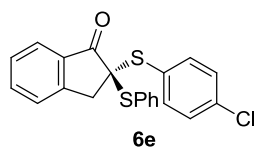
Column chromatography (PE/EtOAc = 20:1 to 15:1) afforded the desired product **6c** in 97% yield as pale yellow solid (m. p. 52-54 °C). HPLC analysis (Chiralpak IC, *i*PrOH/hexane = 1/99, 1.0 mL/min, 230 nm; t_r (minor) = 17.033

min, t_r (major) = 19.213 min) gave the isomeric composition of the product: 94% ee, $[\alpha]_D^{29} = +22.6$ ($c = 0.96$, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.89 (dd, $J = 8.0$ Hz, $J = 1.6$ Hz, 1H), 7.59-7.57 (m, 2H), 7.51-7.48 (m, 3H), 7.42-7.39 (m, 1H), 7.35-7.28 (m, 4H), 7.06 (td, $J = 7.6$ Hz, $J = 1.2$ Hz, 1H), 6.94 (dd, $J = 8.0$ Hz, $J = 0.8$ Hz, 1H), 4.42, 4.36 (AB, $J = 12.4$ Hz, 2H); ¹³C NMR (100 MHz, CDCl₃): δ 184.13, 160.08, 138.16, 136.92, 136.57, 136.14, 130.05, 129.18, 129.02, 128.56, 128.46, 127.33, 122.21, 119.50, 117.67, 73.28, 66.48; IR (KBr): 3447, 3074, 2865, 1681, 1601, 1062, 842, 540; HRMS (EI): Exact mass calcd for C₂₁H₁₅O₂S₂³⁵Cl [M]⁺: 398.0202, Found: 398.0203



Column chromatography (PE/EtOAc = 20:1 to 15:1) afforded the desired product **6d** in 85% yield as yellow oil. HPLC analysis (Chiralpak OD-H, *i*PrOH/hexane = 1/99, 1.0 mL/min, 230 nm; t_r (minor) = 11.677 min, t_r (major) = 12.746 min) gave the isomeric composition of the product: 91% ee, $[\alpha]_D^{27} = +24.5$ ($c = 1.06$, CHCl₃); ¹H NMR (400

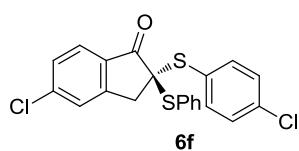
MHz, CDCl₃): δ 7.45 (d, $J = 7.2$ Hz, 1H), 7.30-7.29 (m, 1H), 7.29-7.28 (m, 2H), 7.27-7.26 (m, 1H), 7.26-7.25 (m, 1H), 7.23-7.22 (m, 2H), 7.20-7.13 (m, 4H), 7.08 (d, $J = 7.6$ Hz, 1H), 3.24 (s, 2H); ¹³C NMR (100 MHz, CDCl₃): δ 206.83, 139.85, 137.66, 136.50, 136.12, 135.71, 129.99, 129.69, 129.35, 128.89, 128.81, 128.71, 127.85, 126.52, 124.65, 68.58, 40.80; IR (KBr): 3481, 3058, 1752, 1474, 1092, 1014, 745, 550; HRMS (EI): Exact mass calcd for C₂₁H₁₅OS₂³⁵Cl [M]⁺: 382.0253, Found: 382.0249.



Column chromatography (PE/EtOAc = 20:1 to 15:1) afforded the desired product **6e** in 95% yield as orange solid (m. p. 90-92 °C). HPLC analysis (Chiralpak OD-H, *i*PrOH/hexane = 1/99, 1.0 mL/min, 230 nm; t_r (minor) =

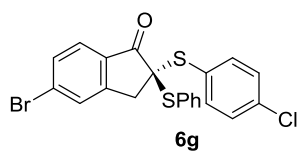
16.760 min, t_r (major) = 15.387 min) gave the isomeric composition of the product: 95% ee, $[\alpha]_D^{29} = +26.4$ ($c = 0.95$, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.78 (d, $J = 7.6$ Hz, 1H), 7.58-7.50 (m, 5H), 7.40-7.23 (m, 7H), 3.44, 3.35 (AB, $J = 18.0$ Hz, 2H); ¹³C NMR (100 MHz, CDCl₃): δ 196.66, 148.89, 137.21, 136.01, 135.71, 135.35, 133.72, 130.94, 129.48, 129.45, 129.02, 128.87, 128.19, 125.86, 125.35, 67.00, 41.87; IR (KBr): 3448, 1708, 1647, 1465, 1092, 1025, 746, 688;

HRMS (EI): Exact mass calcd for $C_{21}H_{15}OS_2^{35}Cl$ $[M]^+$: 382.0253, Found: 382.0250.



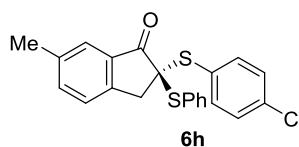
Column chromatography (PE/EtOAc = 20:1 to 15:1) afforded the desired product **6f** in 96% yield as yellow solid (m. p. 121-123 °C). HPLC analysis (Chiralpak AD-H, *i*PrOH/hexane = 3/97, 1.0 mL/min, 230 nm; t_r (minor) =

23.873 min, t_r (major) = 22.227 min) gave the isomeric composition of the product: 90% ee, $[\alpha]_D^{29} = +42.9$ ($c = 0.96$, $CHCl_3$); 1H NMR (400 MHz, $CDCl_3$): δ 7.70 (d, $J = 8.4$ Hz, 1H), 7.56-7.54 (m, 2H), 7.51-7.49 (m, 2H), 7.37-7.35 (m, 2H), 7.33-7.28 (m, 4H), 7.24 (s, 1H), 3.39, 3.30 (AB, $J = 18.0$ Hz, 2H); ^{13}C NMR (100 MHz, $CDCl_3$): δ 195.33, 150.39, 141.93, 137.35, 136.27, 135.86, 132.23, 130.69, 129.72, 129.23, 129.18, 129.07, 129.04, 126.49, 126.22, 67.02, 41.49; IR (KBr): 3449, 1703, 1600, 1477, 1261, 1069, 744, 486; HRMS (EI): Exact mass calcd for $C_{21}H_{14}OS_2^{35}Cl_2$ $[M]^+$: 415.9863, Found: 415.9860.



Column chromatography (PE/EtOAc = 20:1 to 15:1) afforded the desired product **6g** in 93% yield as orange solid (m. p. 155-157 °C). HPLC analysis (Chiralpak AD-H, *i*PrOH/hexane = 3/97, 1.0 mL/min, 230 nm; t_r (minor) =

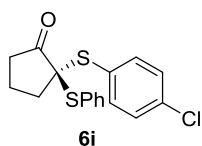
28.926 min, t_r (major) = 26.484 min) gave the isomeric composition of the product: 92% ee, $[\alpha]_D^{29} = +41.1$ ($c = 1.0$, $CHCl_3$); 1H NMR (400 MHz, $CDCl_3$): δ 7.63 (d, $J = 8.4$ Hz, 1H), 7.56-7.49 (m, 5H), 7.42 (s, 1H), 7.38-7.36 (m, 1H), 7.33-7.28 (m, 4H), 3.39, 3.31 (AB, $J = 18.0$ Hz, 2H); ^{13}C NMR (100 MHz, $CDCl_3$): δ 195.49, 150.45, 137.34, 136.29, 135.86, 132.63, 131.92, 130.76, 130.66, 129.72, 129.26, 129.18, 129.03, 126.55, 66.91, 41.39; IR (KBr): 3448, 1704, 1592, 1474, 1024, 968, 825, 746; HRMS (EI): Exact mass calcd for $C_{21}H_{14}OS_2^{35}Cl^{79}Br$ $[M]^+$: 459.9358, Found: 459.9354.



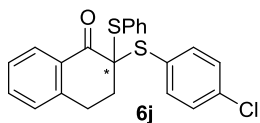
Column chromatography (PE/EtOAc = 20:1 to 15:1) afforded the desired product **6h** in 95% yield as pink solid (m. p. 71-73 °C). HPLC analysis (Chiralpak AD-H, *i*PrOH/hexane = 3/97, 1.0 mL/min, 230 nm; t_r (minor) =

22.556 min, t_r (major) = 24.207 min) gave the isomeric composition of the product: 94% ee, $[\alpha]_D^{28} = +27.5$ ($c = 1.02$, $CHCl_3$); 1H NMR (400 MHz, $CDCl_3$): δ 7.58-7.55 (m, 3H), 7.52-7.50 (m, 2H), 7.39-7.26 (m, 6H), 7.13 (d, $J = 7.6$ Hz, 1H), 3.39, 3.30 (AB, $J = 17.6$ Hz, 2H), 2.39 (s, 3H); ^{13}C NMR

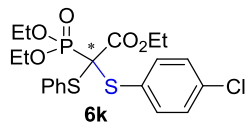
(100 MHz, CDCl₃): δ 196.73, 146.26, 138.33, 137.23, 136.70, 135.98, 135.71, 133.83, 131.13, 129.68, 129.45, 129.06, 128.93, 125.67, 125.32, 67.41, 41.60, 21.20; IR (KBr): 3449, 1705, 1495, 1281, 1091, 815, 743, 688; HRMS (EI): Exact mass calcd for C₂₂H₁₇OS₂³⁵Cl [M]⁺: 396.0409, Found: 396.0407.



Column chromatography (PE/EtOAc = 20:1 to 15:1) afforded the desired product **6i** in 80% yield as yellow oil. HPLC analysis (Chiralpak OD-H, *i*PrOH/hexane = 5/95, 1.0 mL/min, 230 nm; t_r (minor) = 8.477 min, t_r (major) = 7.632 min) gave the isomeric composition of the product: 86% ee, $[\alpha]_D^{29} = -8.7$ ($c = 0.92$, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.59-7.57 (m, 2H), 7.53-7.51 (m, 2H), 7.40-7.31 (m, 5H), 2.36 (t, $J = 7.6$ Hz, 2H), 2.16-2.05 (m, 2H), 2.02-1.95 (m, 2H); ¹³C NMR (100 MHz, CDCl₃): δ 206.71, 137.57, 136.20, 136.08, 130.46, 129.60, 129.16, 129.11, 128.95, 69.27, 36.92, 35.63, 18.32; IR (KBr): 3448, 2968, 1733, 1474, 1024, 823, 745, 492; HRMS (EI): Exact mass calcd for C₁₇H₁₅OS₂³⁵Cl [M]⁺: 334.0253, Found: 334.0256.

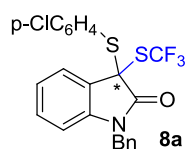


Column chromatography (PE/EtOAc = 20:1 to 15:1) afforded product **6j** in 80% yield as light green oil. HPLC analysis (Chiralpak AD-H, *i*PrOH/hexane = 3/97, 1.0 mL/min, 230 nm; t_r (minor) = 15.927 min, t_r (major) = 16.753 min) gave the isomeric composition of the product: 97% ee, $[\alpha]_D^{28} = +8.5$ ($c = 0.95$, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 8.02 (dd, $J = 8.0$ Hz, $J = 1.2$ Hz, 1H), 7.60-7.57 (m, 2H), 7.53-7.48 (m, 3H), 7.40-7.38 (m, 1H), 7.36-7.29 (m, 5H), 7.21 (d, $J = 7.6$ Hz, 1H), 3.11 (t, $J = 6.0$ Hz, 2H), 2.40-2.36 (m, 2H); ¹³C NMR (100 MHz, CDCl₃): δ 188.45, 142.18, 138.06, 136.80, 136.11, 133.62, 130.92, 130.09, 129.64, 129.02, 128.88, 128.83, 128.52, 127.11, 70.57, 35.62, 26.71; IR (KBr): 3446, 3057, 2926, 1684, 1474, 1234, 744, 511; HRMS (EI): Exact mass calcd for C₂₂H₁₇OS₂³⁵Cl [M]⁺: 396.0409, Found: 396.0410.

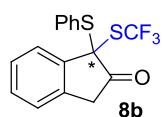


Column chromatography (PE/EtOAc = 4:1 to 2:1) afforded the desired product **6k** in 94% yield as colorless oil. HPLC analysis (Chiralpak AD-H, *i*PrOH/hexane = 10/90, 1.0 mL/min, 205 nm; t_r (minor) = 8.751 min, t_r (major) = 9.755 min) gave the isomeric composition of the product: 70% ee, $[\alpha]_D^{28} = -18.0$ ($c = 1.02$, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.68-7.64 (m, 4H), 7.36-7.25 (m, 5H), 4.31-4.20 (m, 4H), 3.88 (q, $J = 7.2$ Hz, 2H), 1.33-1.28

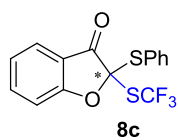
(m, 6H), 1.00 (t, $J = 7.2$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3): δ 165.58 (d, $J = 3$ Hz, 1C), 137.70, 135.98, 130.71, 130.66, 129.60, 129.57, 129.54, 128.54, 65.16 (d, $J = 149$ Hz, 1C), 64.74 (d, $J = 2$ Hz, 1C), 64.67 (d, $J = 2$ Hz, 1C), 62.81, 16.38, 16.33, 13.56; ^{31}P NMR (161.7 MHz, CDCl_3): 13.91 (s, 1P); IR (KBr): 3448, 2982, 2929, 1723, 1475, 1258, 1026, 748; HRMS (EI): Exact mass calcd for $\text{C}_{20}\text{H}_{24}\text{O}_5\text{S}_2\text{P}^{35}\text{Cl} [\text{M}]^+$: 474.0491, Found: 474.0488.



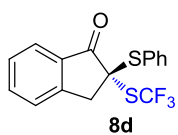
Column chromatography (PE/EtOAc = 20:1 to 15:1) afforded the desired product **8a** in 65% yield as pale brown solid (m. p. 92-94 °C). HPLC analysis (Chiralpak AD-H, i PrOH/hexane = 5/95, 1.0 mL/min, 230 nm; t_r (minor) = 13.313 min, t_r (major) = 14.893 min) gave the isomeric composition of the product: 85% ee, $[\alpha]_D^{29} = +0.4$ ($c = 0.94$, CHCl_3); ^1H NMR (400 MHz, CDCl_3): δ 7.35-7.32 (m, 3H), 7.31-7.25 (m, 4H), 7.24-7.20 (m, 3H), 7.06-7.04 (m, 1H), 6.99 (td, $J = 7.6$ Hz, $J = 0.8$ Hz, 1H), 6.71 (d, $J = 8.0$ Hz, 1H), 4.94, 4.86 (AB, $J = 15.6$ Hz, 2H); ^{13}C NMR (100 MHz, CDCl_3): δ 171.78, 141.32, 139.13, 137.83, 135.01, 130.65, 129.23, 128.87, 128.13 (q, $J = 311$ Hz, 1C), 127.89, 127.22, 125.82, 125.75, 125.36, 122.94, 109.90, 59.27, 44.57; ^{19}F NMR (376 MHz, CDCl_3): -39.11 (s, 3F). IR (KBr): 3447, 3064, 2922, 1728, 1522, 1097, 748, 500; HRMS (ESI): Exact mass calcd for $\text{C}_{22}\text{H}_{19}^{35}\text{ClF}_3\text{N}_2\text{OS}_2 [\text{M}+\text{NH}_4]^+$: 483.0574, Found: 483.0577.



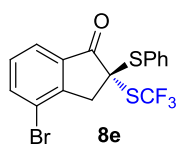
Column chromatography (PE/ CH_2Cl_2 = 10:1 to 5:1) afforded the desired product **8b** in 65% yield as yellow oil. HPLC analysis (Chiralpak OD-H, i PrOH/hexane = 1/99, 1.0 mL/min, 230 nm; t_r (minor) = 8.276 min, t_r (major) = 6.826 min) gave the isomeric composition of the product: 83% ee, $[\alpha]_D^{29} = +15.4$ ($c = 1.10$, CHCl_3); ^1H NMR (400 MHz, CDCl_3): δ 7.48-7.44 (m, 1H), 7.39-7.31 (m, 6H), 7.28-7.22 (m, 2H), 3.87, 3.61 (AB, $J = 22.4$ Hz, 2H); ^{13}C NMR (100 MHz, CDCl_3): δ 203.72, 137.89, 137.68, 135.15, 131.02, 130.06, 129.06, 128.30 (q, $J = 311$ Hz, 1C), 127.96, 127.81, 126.40, 125.00, 65.61, 39.55; ^{19}F NMR (376 MHz, CDCl_3): -38.23 (s, 3F). IR (KBr): 3447, 3064, 2919, 1762, 1473, 1109, 749, 493; HRMS (EI): Exact mass calcd for $\text{C}_{16}\text{H}_{11}\text{OF}_3\text{S}_2 [\text{M}]^+$: 340.0203, Found: 340.0201.



Column chromatography (PE/CH₂Cl₂ = 10:1 to 5:1) afforded the desired product **8c** in 48% yield as colorless oil. HPLC analysis (Chiralpak OJ-H, *i*PrOH/hexane = 20/80, 1.0 mL/min, 230 nm; *t_r* (minor) = 10.706 min, *t_r* (major) = 12.356 min) gave the isomeric composition of the product: 95% ee, $[\alpha]_D^{27} = +95.8$ (*c* = 0.26, CHCl₃); ¹H NMR (400 MHz, acetone-*d*₆): δ 7.90-7.86 (m, 1H), 7.77-7.75 (m, 1H), 7.66-7.63 (m, 2H), 7.55-7.51 (m, 1H), 7.46-7.43 (m, 2H), 7.35-7.30 (m, 2H); ¹³C NMR (100 MHz, acetone-*d*₆): δ 192.11, 169.38, 140.72, 138.08, 132.00, 130.25, 129.12 (q, *J* = 310 Hz, 1C), 127.18, 126.17, 125.22, 118.93, 114.46, 96.22; ¹⁹F NMR (376 MHz, acetone-*d*₆): 138.26. IR (KBr): 3447, 1750, 1647, 1522, 1322, 1112, 750, 490; HRMS (EI): Exact mass calcd for C₁₅H₉O₂F₃S₂ [M]⁺: 341.9996, Found: 341.9995.

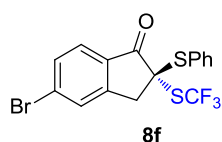


Column chromatography (PE/CH₂Cl₂ = 10:1 to 5:1) afforded the desired product **8d** in 64% yield as yellow oil. HPLC analysis (Chiralpak OD-H, *i*PrOH/hexane = 1/99, 1.0 mL/min, 230 nm; *t_r* (minor) = 7.707 min, *t_r* (major) = 6.733 min) gave the isomeric composition of the product: 92% ee, $[\alpha]_D^{27} = -117.9$ (*c* = 1.03, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.85 (d, *J* = 7.6 Hz, 1H), 7.66 (td, *J* = 7.6 Hz, *J* = 1.2 Hz, 1H), 7.51-7.44 (m, 4H), 7.40-7.34 (m, 3H), 3.87, 3.58 (AB, *J* = 18.0 Hz, 2H); ¹³C NMR (100 MHz, CDCl₃): δ 194.33, 148.37, 137.25, 135.88, 132.53, 130.73, 129.44 (q, *J* = 310 Hz, 1C), 129.00, 128.89, 128.62, 126.09, 125.77, 64.96, 42.95; ¹⁹F NMR (376 MHz, CDCl₃): -37.14 (s, 3F). IR (KBr): 3448, 3059, 2927, 1729, 1606, 1109, 747, 471; HRMS (EI): Exact mass calcd for C₁₆H₁₁OF₃S₂ [M]⁺: 340.0203, Found: 340.0206.

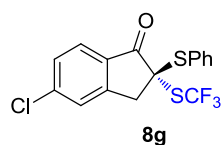


Column chromatography (PE/CH₂Cl₂ = 10:1 to 5:1) afforded the desired product **8e** in 64% yield as yellow oil. HPLC analysis (Chiralpak OD-H, *i*PrOH/hexane = 1/99, 1.0 mL/min, 230 nm; *t_r* (minor) = 7.873 min, *t_r* (major) = 6.633 min) gave the isomeric composition of the product: 93% ee, $[\alpha]_D^{28} = -96.7$ (*c* = 0.96, CHCl₃); ¹H NMR (400 MHz, acetone-*d*₆): δ 7.99 (dd, *J* = 7.6 Hz, *J* = 0.8 Hz, 1H), 7.84 (dd, *J* = 7.2 Hz, *J* = 0.4 Hz, 1H), 7.57-7.50 (m, 4H), 7.46-7.42 (m, 2H), 3.78, 3.60 (AB, *J* = 18.4 Hz, 2H); ¹³C NMR (100 MHz, acetone-*d*₆): δ 193.96, 148.74, 139.75, 138.10, 135.47, 131.94, 131.89, 130.42 (q, *J* = 309 Hz, 1C) 130.15, 129.26, 125.30, 121.86, 65.68, 44.31; ¹⁹F NMR (376 MHz, acetone-*d*₆): -139.46 (s, 3F). IR (KBr): 3447, 1716, 1653, 1541, 1522, 1489, 1386, 1108; HRMS (EI): Exact mass calcd for C₁₆H₁₀OF₃S₂⁷⁹Br[M]⁺: 417.9309,

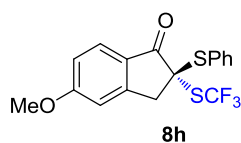
Found: 417.9307.



Column chromatography (PE/CH₂Cl₂ = 10:1 to 5:1) afforded the desired product **8f** in 71% yield as white solid (m. p. 45-47 °C). HPLC analysis (Chiralpak AD-H, *i*PrOH/hexane = 1/99, 1.0 mL/min, 230 nm; *t_r* (minor) = 8.193 min, *t_r* (major) = 6.847 min) gave the isomeric composition of the product: 93% ee, $[\alpha]_D^{27} = -117.8$ (*c* = 0.25, CHCl₃); ¹H NMR (400 MHz, acetone-*d*₆): δ 7.84-7.83 (m, 1H), 7.77-7.73 (m, 2H), 7.56-7.52 (m, 3H), 7.46-7.42 (m, 2H), 3.92, 3.74 (AB, *J* = 18.8 Hz, 2H); ¹³C NMR (100 MHz, acetone-*d*₆): δ 193.77, 151.35, 138.06, 133.19, 132.25, 131.80, 131.59, 130.78, 130.45 (q, *J* = 309 Hz, 1C), 130.09, 129.47, 127.57, 65.90, 43.10; ¹⁹F NMR (376 MHz, acetone-*d*₆): -139.34 (s, 3F). IR (KBr): 3421, 2924, 1717, 1596, 1318, 1105, 753, 484; HRMS (ESI): Exact mass calcd for C₁₆H₁₄⁷⁹BrF₃NOS₂ [M+NH₄]⁺: 435.9647, Found: 435.9649.

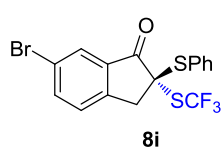


Column chromatography (PE/CH₂Cl₂ = 10:1 to 5:1) afforded the desired product **8g** in 70% yield as pale yellow solid (m. p. 68-70 °C). HPLC analysis (Chiralpak AD-H, *i*PrOH/hexane = 1/99, 1.0 mL/min, 230 nm; *t_r* (minor) = 9.935 min, *t_r* (major) = 8.172 min) gave the isomeric composition of the product: 93% ee, $[\alpha]_D^{27} = -67.6$ (*c* = 0.24, CHCl₃); ¹H NMR (400 MHz, acetone-*d*₆): δ 7.82 (d, *J* = 8.0 Hz, 1H), 7.66-7.65 (m, 1H), 7.60-7.58 (m, 1H), 7.55-7.52 (m, 3H), 7.46-7.42 (m, 2H), 3.91, 3.73 (AB, *J* = 18.4 Hz, 2H); ¹³C NMR (100 MHz, acetone-*d*₆): δ 193.56, 151.29, 142.75, 138.05, 131.89, 131.79, 130.45 (q, *J* = 308 Hz, 1C) 130.30, 130.08, 129.48, 127.70, 127.54, 65.98, 43.18; ¹⁹F NMR (376 MHz, acetone-*d*₆): -139.36 (s, 3F). IR (KBr): 3421, 2963, 1710, 1601, 1109, 887, 752, 492; HRMS (ESI): Exact mass calcd for C₁₆H₁₄³⁵ClF₃NOS₂ [M+NH₄]⁺: 392.0152, Found: 392.0154.



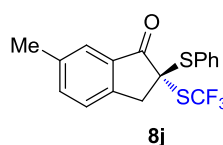
Column chromatography (PE/EtOAc = 15:1 to 10:1) afforded the desired product **8h** in 60% yield as pale yellow solid (m. p. 74-76 °C). HPLC analysis (Chiralpak OD-H, *i*PrOH/hexane = 0.5/99.5, 1.0 mL/min, 230 nm; *t_r* (minor) = 24.200 min, *t_r* (major) = 19.933 min) gave the isomeric composition of the product: 93% ee, $[\alpha]_D^{29} = -198.7$ (*c* = 1.03, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.78 (d, *J* = 8.4 Hz, 1H), 7.52-7.50 (m, 2H), 7.47-7.44 (m, 1H), 7.39-7.35 (m, 2H), 6.98 (dd, *J* = 8.4 Hz, *J* = 2.0 Hz, 1H), 6.82 (s, 1H), 3.90 (s, 3H), 3.81, 3.54 (AB, *J* =

18.0 Hz, 2H); ^{13}C NMR (100 MHz, CDCl_3): δ 193.04, 166.32, 151.50, 137.13, 130.57, 129.44 (q, J = 310 Hz, 1C), 129.15, 128.96, 127.53, 125.37, 116.47, 109.46, 65.16, 55.83, 42.89; ^{19}F NMR (376 MHz, CDCl_3): -37.35 (s, 3F). IR (KBr): 3421, 2944, 1716, 1599, 1262, 1107, 752, 436; HRMS (EI): Exact mass calcd for $\text{C}_{17}\text{H}_{13}\text{O}_2\text{F}_3\text{S}_2$ $[\text{M}]^+$: 370.0309, Found: 370.0305.



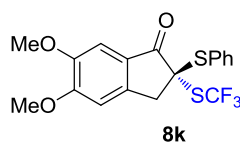
Column chromatography (PE/ CH_2Cl_2 = 10:1 to 5:1) afforded the desired product **8i** in 60% yield as pale yellow solid (m. p. 82-84 °C). HPLC analysis (Chiralpak AD-H+AD-H, i PrOH/hexane = 0.5/99.5, 1.0 mL/min, 230 nm; t_r (minor) = 25.140

min, t_r (major) = 24.202 min) gave the isomeric composition of the product: 92% ee, $[\alpha]_D^{29}$ = -115.9 (c = 0.93, CHCl_3); ^1H NMR (400 MHz, acetone- d_6): δ 7.93-7.91 (m, 2H), 7.58-7.52 (m, 4H), 7.46-7.42 (m, 2H), 3.87, 3.69 (AB, J = 18.8 Hz, 2H); ^{13}C NMR (100 MHz, acetone- d_6): δ 193.36, 148.40, 139.64, 138.12, 135.19, 131.84, 130.44 (q, J = 309 Hz, 1C), 130.09, 129.59, 129.38, 128.60, 123.00, 66.11, 43.23; ^{19}F NMR (376 MHz, acetone- d_6): -139.42 (s, 3F). IR (KBr): 3451, 2921, 1726, 1467, 1104, 895, 753, 489; HRMS (EI): Exact mass calcd for $\text{C}_{16}\text{H}_{10}\text{OF}_3\text{S}_2^{79}\text{Br}$ $[\text{M}]^+$: 417.9309, Found: 417.9312.



Column chromatography (PE/ CH_2Cl_2 = 10:1 to 5:1) afforded the desired product **8j** in 68% yield as pale yellow solid (m. p. 47-49 °C). HPLC analysis (Chiralpak OD-H+OD-H, i PrOH/hexane = 1/99, 1.0 mL/min, 230 nm; t_r (minor) = 13.367 min,

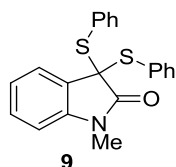
t_r (major) = 11.600 min) gave the isomeric composition of the product: 94% ee, $[\alpha]_D^{27}$ = -156.7 (c = 0.95, CHCl_3); ^1H NMR (400 MHz, CDCl_3): δ 7.65 (s, 1H), 7.50-7.45 (m, 4H), 7.39-7.36 (m, 2H), 7.29 (d, J = 7.6 Hz, 1H), 3.82, 3.53 (AB, J = 18.0 Hz, 2H), 2.44 (s, 3H); ^{13}C NMR (100 MHz, CDCl_3): δ 194.41, 145.68, 138.77, 137.25, 137.15, 132.59, 130.68, 129.44 (q, J = 309 Hz, 1C), 128.98, 125.80, 125.68, 65.26, 42.63, 21.18; ^{19}F NMR (376 MHz, CDCl_3): δ -37.24 (s, 3F). IR (KBr): 3422, 2918, 1727, 1278, 1102, 896, 796, 491; HRMS (EI): Exact mass calcd for $\text{C}_{17}\text{H}_{13}\text{OF}_3\text{S}_2$ $[\text{M}]^+$: 354.0360, Found: 354.0359.



Column chromatography (PE/EtOAc = 10 to 5:1) afforded the desired product **8k** in 68% yield as orange solid (m. p. 99-101 °C). HPLC analysis (Chiralpak AD-H, i PrOH/hexane = 10/90, 1.0 mL/min, 230 nm; t_r (minor) = 10.253 min, t_r (major) =

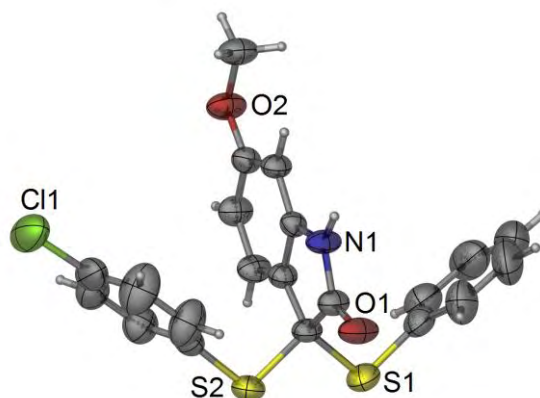
8.840 min) gave the isomeric composition of the product: 94% ee, $[\alpha]_D^{27}$ = -162.6 (c = 0.96, CHCl_3); ^1H

NMR (400 MHz, CDCl₃): δ 7.54-7.52 (m, 2H), 7.45-7.44 (m, 1H), 7.39-7.36 (m, 2H), 7.25 (s, 1H), 6.80 (s, 1H), 3.98 (s, 3H), 3.94 (s, 3H), 3.78, 3.52 (AB, J = 18.0 Hz, 2H); ¹³C NMR (100 MHz, CDCl₃): δ 193.63, 156.60, 150.27, 143.96, 137.12, 130.53, 129.38 (q, J = 310 Hz, 1C), 129.15, 128.93, 124.99, 107.01, 105.79, 64.94, 56.39, 56.22, 42.65; ¹⁹F NMR (376 MHz, CDCl₃): δ -37.50 (s, 3F). IR (KBr): 3449, 2940, 1711, 1590, 1311, 1268, 756, 464; HRMS (EI): Exact mass calcd for C₁₈H₁₅O₃F₃S₂ [M]⁺: 400.0415, Found: 400.0414.



The reaction of 3-thiooxindole **1k** with N-chlorosuccinimide (NCS) catalyzed by DABCO at 25 °C afforded **9** in 30% yield as orange solid. ¹H NMR (400 MHz, CDCl₃): δ 7.36-7.33 (m, 4H), 7.29-7.25 (m, 2H), 7.18-7.09 (m, 5H), 6.99-6.90 (m, 2H), 6.45 (d, J = 7.6 Hz, 1H), 2.87 (s, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 173.28, 142.30, 136.71, 129.69, 129.57, 129.30, 128.43, 128.08, 125.52, 122.43, 107.81, 62.81, 26.30; HRMS (EI): Exact mass calcd for C₂₁H₁₇NOS₂ [M]⁺: 363.0752, Found: 363.0749.

4) Single-Crystal X-ray Crystallography of **3c**.¹³



Data intensity of **3c** was collected using a Bruker SMART APEX II (Mo radiation) at 296 K in a nitrogen stream. Data collection and reduction were done by using the Bruker ApexII software package. The structures were solved by direct methods and refined by full-matrix least-squares on F^2 with anisotropic displacement parameters for non-H atoms using SHELX-97. Hydrogen atoms were added at their geometrically ideal positions and refined isotropically. Crystal data for **3c**: $C_{21}H_{16}ClNO_2S_2$, $T = 296(2)$ K, orthorhombic, space group $P2(1)2(1)2(1)$, $a = 8.7074(4)$ Å, $b = 12.6241(6)$ Å, $c = 18.0261(8)$ Å, $\alpha = 90$ deg, $\beta = 90$ deg, $\gamma = 90$ deg. $V = 1981.48(16)$ Å³. $Z = 4$, $d_{\text{calc}} = 1.387$ mg/m³. Total number of reflections 23460 [$R(\text{int}) = 0.0900$], $R_1 = 0.0423$ ($I > 2\sigma(I)$), $wR_2 = 0.0939$ (all data), GOF = 0.943, and 244 parameters.

Table 1. Crystal data and structure refinement for **3c**.

Identification code	3c
Empirical formula	$C_{21}H_{16}ClNO_2S_2$
Formula weight	413.92
Temperature	296(2) K
Wavelength	0.71073 Å
Crystal system, space group	Orthorhombic, $P2(1)2(1)2(1)$
Unit cell dimensions	$a = 8.7074(4)$ Å $\alpha = 90$ deg. $b = 12.6241(6)$ Å $\beta = 90$ deg. $c = 18.0261(8)$ Å $\gamma = 90$ deg.
Volume	$1981.48(16)$ Å ³
Z , Calculated density	4, 1.387 Mg/m ³
Absorption coefficient	0.420 mm ⁻¹
$F(000)$	856
Crystal size	$0.27 \times 0.18 \times 0.17$ mm
Theta range for data collection	1.97 to 25.01 deg.

¹³ Supplementary crystallographic data have been deposited at the Cambridge Crystallographic Data Center. (CCDC 1413662).

Limiting indices	-10<=h<=10, -15<=k<=14, -21<=l<=21
Reflections collected / unique	23460 / 3490 [R(int) = 0.0900]
Completeness to theta = 25.01	100.0 %
Absorption correction	Semi-empirical from equivalents
Max. and min. transmission	0.9321 and 0.8952
Refinement method	Full-matrix least-squares on F ²
Data / restraints / parameters	3490 / 0 / 244
Goodness-of-fit on F ²	0.943
Final R indices [I>2sigma(I)]	R1 = 0.0423, wR2 = 0.0848
R indices (all data)	R1 = 0.0631, wR2 = 0.0939
Absolute structure parameter	-0.03(7)
Largest diff. peak and hole	0.216 and -0.256 e.A ⁻³

Table 2. Atomic coordinates (x 10⁴) and equivalent isotropic displacement parameters (A² x 10³) for z.

U(eq) is defined as one third of the trace of the orthogonalized Uij tensor.

	x	y	z	U(eq)
S(1)	2349(1)	630(1)	-615(1)	57(1)
S(2)	2504(1)	1099(1)	929(1)	55(1)
Cl(1)	-1441(1)	1178(1)	3828(1)	100(1)
O(1)	525(2)	2728(2)	-61(1)	60(1)
O(2)	-3812(2)	-1864(2)	806(1)	60(1)
N(1)	-1382(3)	1531(2)	195(1)	46(1)
C(1)	240(5)	1208(4)	-1670(2)	81(1)
C(2)	-752(6)	1005(6)	-2256(2)	104(2)
C(3)	-971(6)	-16(6)	-2513(3)	109(2)
C(4)	-230(6)	-814(5)	-2172(3)	101(2)
C(5)	764(4)	-637(3)	-1579(2)	73(1)
C(6)	989(4)	387(3)	-1329(2)	56(1)
C(7)	1092(3)	847(2)	182(2)	41(1)
C(8)	82(3)	1839(2)	85(2)	44(1)
C(9)	-1495(3)	442(2)	356(2)	36(1)
C(10)	-2811(3)	-117(2)	501(2)	43(1)
C(11)	-2636(3)	-1193(2)	655(2)	42(1)
C(12)	-1193(3)	-1661(2)	645(2)	51(1)
C(13)	101(3)	-1073(2)	488(2)	48(1)
C(14)	-47(3)	-7(2)	350(2)	40(1)
C(15)	-5334(3)	-1455(3)	792(2)	64(1)
C(16)	1306(4)	1109(3)	1719(2)	50(1)
C(17)	574(5)	2006(3)	1948(2)	83(1)

C(18)	-295(6)	2024(4)	2580(3)	95(2)
C(19)	-401(4)	1147(4)	3009(2)	68(1)
C(20)	307(5)	245(3)	2792(2)	72(1)
C(21)	1167(5)	226(3)	2157(2)	68(1)

Table 3. Bond lengths [Å] and angles [deg] for z.

S(1)-C(6)	1.777(4)
S(1)-C(7)	1.827(3)
S(2)-C(16)	1.765(3)
S(2)-C(7)	1.852(3)
Cl(1)-C(19)	1.731(4)
O(1)-C(8)	1.215(3)
O(2)-C(11)	1.357(3)
O(2)-C(15)	1.423(4)
N(1)-C(8)	1.348(4)
N(1)-C(9)	1.408(3)
N(1)-H(1A)	0.8600
C(1)-C(6)	1.370(5)
C(1)-C(2)	1.389(6)
C(1)-H(1B)	0.9300
C(2)-C(3)	1.383(7)
C(2)-H(2A)	0.9300
C(3)-C(4)	1.344(7)
C(3)-H(3A)	0.9300
C(4)-C(5)	1.394(6)
C(4)-H(4A)	0.9300
C(5)-C(6)	1.383(5)
C(5)-H(5A)	0.9300
C(7)-C(14)	1.495(4)
C(7)-C(8)	1.541(4)
C(9)-C(10)	1.371(4)
C(9)-C(14)	1.382(4)
C(10)-C(11)	1.395(4)
C(10)-H(10A)	0.9300
C(11)-C(12)	1.390(4)
C(12)-C(13)	1.379(4)
C(12)-H(12A)	0.9300
C(13)-C(14)	1.375(4)
C(13)-H(13A)	0.9300
C(15)-H(15A)	0.9600
C(15)-H(15B)	0.9600

C(15)-H(15C)	0.9600
C(16)-C(17)	1.362(5)
C(16)-C(21)	1.372(5)
C(17)-C(18)	1.368(5)
C(17)-H(17A)	0.9300
C(18)-C(19)	1.354(6)
C(18)-H(18A)	0.9300
C(19)-C(20)	1.352(5)
C(20)-C(21)	1.367(5)
C(20)-H(20A)	0.9300
C(21)-H(21A)	0.9300
C(6)-S(1)-C(7)	101.31(14)
C(16)-S(2)-C(7)	101.33(13)
C(11)-O(2)-C(15)	118.2(2)
C(8)-N(1)-C(9)	112.2(2)
C(8)-N(1)-H(1A)	123.9
C(9)-N(1)-H(1A)	123.9
C(6)-C(1)-C(2)	119.8(5)
C(6)-C(1)-H(1B)	120.1
C(2)-C(1)-H(1B)	120.1
C(3)-C(2)-C(1)	120.8(5)
C(3)-C(2)-H(2A)	119.6
C(1)-C(2)-H(2A)	119.6
C(4)-C(3)-C(2)	118.7(5)
C(4)-C(3)-H(3A)	120.7
C(2)-C(3)-H(3A)	120.7
C(3)-C(4)-C(5)	121.9(5)
C(3)-C(4)-H(4A)	119.0
C(5)-C(4)-H(4A)	119.0
C(6)-C(5)-C(4)	119.2(4)
C(6)-C(5)-H(5A)	120.4
C(4)-C(5)-H(5A)	120.4
C(1)-C(6)-C(5)	119.6(4)
C(1)-C(6)-S(1)	120.7(3)
C(5)-C(6)-S(1)	119.5(3)
C(14)-C(7)-C(8)	103.3(2)
C(14)-C(7)-S(1)	116.7(2)
C(8)-C(7)-S(1)	112.0(2)
C(14)-C(7)-S(2)	114.6(2)
C(8)-C(7)-S(2)	108.8(2)
S(1)-C(7)-S(2)	101.53(13)
O(1)-C(8)-N(1)	126.8(3)

O(1)-C(8)-C(7)	126.5(3)
N(1)-C(8)-C(7)	106.7(2)
C(10)-C(9)-C(14)	123.6(2)
C(10)-C(9)-N(1)	126.8(3)
C(14)-C(9)-N(1)	109.6(2)
C(9)-C(10)-C(11)	116.6(3)
C(9)-C(10)-H(10A)	121.7
C(11)-C(10)-H(10A)	121.7
O(2)-C(11)-C(12)	114.8(2)
O(2)-C(11)-C(10)	124.5(3)
C(12)-C(11)-C(10)	120.7(3)
C(13)-C(12)-C(11)	120.8(3)
C(13)-C(12)-H(12A)	119.6
C(11)-C(12)-H(12A)	119.6
C(14)-C(13)-C(12)	119.2(3)
C(14)-C(13)-H(13A)	120.4
C(12)-C(13)-H(13A)	120.4
C(13)-C(14)-C(9)	119.0(3)
C(13)-C(14)-C(7)	132.8(3)
C(9)-C(14)-C(7)	108.1(2)
O(2)-C(15)-H(15A)	109.5
O(2)-C(15)-H(15B)	109.5
H(15A)-C(15)-H(15B)	109.5
O(2)-C(15)-H(15C)	109.5
H(15A)-C(15)-H(15C)	109.5
H(15B)-C(15)-H(15C)	109.5
C(17)-C(16)-C(21)	117.4(3)
C(17)-C(16)-S(2)	121.8(3)
C(21)-C(16)-S(2)	120.7(3)
C(16)-C(17)-C(18)	121.6(4)
C(16)-C(17)-H(17A)	119.2
C(18)-C(17)-H(17A)	119.2
C(19)-C(18)-C(17)	120.0(4)
C(19)-C(18)-H(18A)	120.0
C(17)-C(18)-H(18A)	120.0
C(20)-C(19)-C(18)	119.4(4)
C(20)-C(19)-Cl(1)	120.3(4)
C(18)-C(19)-Cl(1)	120.3(4)
C(19)-C(20)-C(21)	120.5(4)
C(19)-C(20)-H(20A)	119.8
C(21)-C(20)-H(20A)	119.8
C(20)-C(21)-C(16)	121.0(4)

C(20)-C(21)-H(21A)	119.5
C(16)-C(21)-H(21A)	119.5

Symmetry transformations used to generate equivalent atoms:

Table 4. Anisotropic displacement parameters ($\text{\AA}^2 \times 10^3$) for z.

The anisotropic displacement factor exponent takes the form:

$$-2\pi^2 [h^2 a^{*2} U_{11} + \dots + 2 h k a^* b^* U_{12}]$$

	U11	U22	U33	U23	U13	U12
S(1)	40(1)	73(1)	59(1)	4(1)	12(1)	3(1)
S(2)	35(1)	68(1)	61(1)	3(1)	-3(1)	-3(1)
Cl(1)	88(1)	152(1)	59(1)	-20(1)	14(1)	-37(1)
O(1)	36(1)	44(1)	100(2)	18(1)	0(1)	0(1)
O(2)	42(1)	45(1)	92(2)	8(1)	1(1)	-7(1)
N(1)	28(1)	37(2)	74(2)	8(1)	2(1)	4(1)
C(1)	99(3)	81(3)	62(3)	13(2)	-5(2)	7(3)
C(2)	108(4)	142(5)	61(3)	27(3)	-11(3)	15(4)
C(3)	95(4)	166(6)	65(3)	18(4)	-1(3)	-27(4)
C(4)	102(4)	129(5)	71(3)	-16(3)	8(3)	-45(4)
C(5)	68(2)	76(3)	74(3)	3(2)	13(2)	-7(2)
C(6)	49(2)	65(2)	54(2)	6(2)	14(2)	0(2)
C(7)	28(1)	44(2)	52(2)	4(1)	3(1)	3(1)
C(8)	35(2)	40(2)	57(2)	5(2)	1(2)	0(1)
C(9)	34(2)	34(2)	40(2)	2(1)	-3(1)	1(1)
C(10)	30(2)	46(2)	52(2)	4(1)	1(1)	2(1)
C(11)	38(2)	42(2)	48(2)	1(1)	-3(2)	-2(2)
C(12)	48(2)	34(2)	69(2)	6(2)	0(2)	2(2)
C(13)	38(2)	41(2)	66(2)	5(2)	2(2)	9(2)
C(14)	33(2)	42(2)	46(2)	3(2)	1(1)	3(1)
C(15)	40(2)	64(2)	87(3)	2(2)	3(2)	-8(2)
C(16)	48(2)	50(2)	51(2)	-1(2)	-10(2)	-5(2)
C(17)	119(3)	57(3)	73(3)	5(2)	26(3)	6(3)
C(18)	136(4)	71(3)	80(3)	-10(3)	30(3)	14(3)
C(19)	63(2)	95(3)	47(2)	-10(2)	-2(2)	-19(2)
C(20)	76(3)	75(3)	66(3)	16(2)	-1(2)	-14(2)
C(21)	73(3)	62(2)	70(2)	7(2)	1(2)	7(2)

Table 5. Hydrogen coordinates ($\times 10^4$) and isotropic displacement parameters ($\text{\AA}^2 \times 10^3$) for z.

	x	y	z	U(eq)
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H(1A)	-2159	1950	170	55
H(1B)	396	1900	-1509	97
H(2A)	-1276	1562	-2479	125
H(3A)	-1617	-148	-2913	130
H(4A)	-386	-1504	-2337	121
H(5A)	1270	-1201	-1355	87
H(10A)	-3772	205	497	51
H(12A)	-1098	-2381	744	61
H(13A)	1063	-1393	477	58
H(15A)	-6048	-2011	907	95
H(15B)	-5554	-1179	308	95
H(15C)	-5429	-899	1153	95
H(17A)	668	2621	1667	100
H(18A)	-814	2638	2714	114
H(20A)	208	-366	3076	87
H(21A)	1666	-396	2021	82

Table 6. Torsion angles [deg] for z.

C(6)-C(1)-C(2)-C(3)	-1.5(7)
C(1)-C(2)-C(3)-C(4)	1.6(8)
C(2)-C(3)-C(4)-C(5)	-1.1(7)
C(3)-C(4)-C(5)-C(6)	0.5(6)
C(2)-C(1)-C(6)-C(5)	0.9(6)
C(2)-C(1)-C(6)-S(1)	176.3(3)
C(4)-C(5)-C(6)-C(1)	-0.4(5)
C(4)-C(5)-C(6)-S(1)	-175.9(3)
C(7)-S(1)-C(6)-C(1)	80.0(3)
C(7)-S(1)-C(6)-C(5)	-104.6(3)
C(6)-S(1)-C(7)-C(14)	54.5(2)
C(6)-S(1)-C(7)-C(8)	-64.2(2)
C(6)-S(1)-C(7)-S(2)	179.91(16)
C(16)-S(2)-C(7)-C(14)	-44.9(2)
C(16)-S(2)-C(7)-C(8)	70.2(2)
C(16)-S(2)-C(7)-S(1)	-171.60(15)
C(9)-N(1)-C(8)-O(1)	-179.9(3)
C(9)-N(1)-C(8)-C(7)	0.0(4)
C(14)-C(7)-C(8)-O(1)	-179.8(3)
S(1)-C(7)-C(8)-O(1)	-53.4(4)
S(2)-C(7)-C(8)-O(1)	58.0(4)
C(14)-C(7)-C(8)-N(1)	0.4(3)

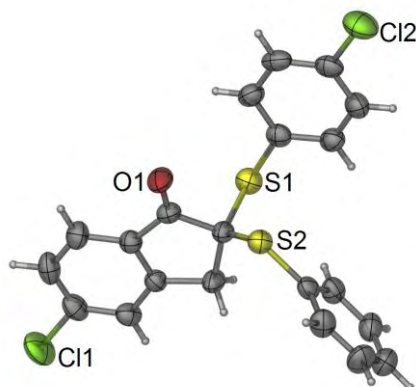
S(1)-C(7)-C(8)-N(1)	126.8(2)
S(2)-C(7)-C(8)-N(1)	-121.8(2)
C(8)-N(1)-C(9)-C(10)	179.4(3)
C(8)-N(1)-C(9)-C(14)	-0.4(4)
C(14)-C(9)-C(10)-C(11)	0.5(4)
N(1)-C(9)-C(10)-C(11)	-179.2(3)
C(15)-O(2)-C(11)-C(12)	-177.1(3)
C(15)-O(2)-C(11)-C(10)	1.4(4)
C(9)-C(10)-C(11)-O(2)	-179.5(3)
C(9)-C(10)-C(11)-C(12)	-1.2(4)
O(2)-C(11)-C(12)-C(13)	179.1(3)
C(10)-C(11)-C(12)-C(13)	0.6(5)
C(11)-C(12)-C(13)-C(14)	0.7(5)
C(12)-C(13)-C(14)-C(9)	-1.3(5)
C(12)-C(13)-C(14)-C(7)	178.5(3)
C(10)-C(9)-C(14)-C(13)	0.7(5)
N(1)-C(9)-C(14)-C(13)	-179.5(3)
C(10)-C(9)-C(14)-C(7)	-179.2(3)
N(1)-C(9)-C(14)-C(7)	0.6(3)
C(8)-C(7)-C(14)-C(13)	179.5(3)
S(1)-C(7)-C(14)-C(13)	56.2(4)
S(2)-C(7)-C(14)-C(13)	-62.3(4)
C(8)-C(7)-C(14)-C(9)	-0.6(3)
S(1)-C(7)-C(14)-C(9)	-124.0(2)
S(2)-C(7)-C(14)-C(9)	117.6(2)
C(7)-S(2)-C(16)-C(17)	-86.5(3)
C(7)-S(2)-C(16)-C(21)	98.2(3)
C(21)-C(16)-C(17)-C(18)	-1.4(6)
S(2)-C(16)-C(17)-C(18)	-176.9(4)
C(16)-C(17)-C(18)-C(19)	2.3(7)
C(17)-C(18)-C(19)-C(20)	-2.7(7)
C(17)-C(18)-C(19)-Cl(1)	177.6(4)
C(18)-C(19)-C(20)-C(21)	2.3(6)
Cl(1)-C(19)-C(20)-C(21)	-178.1(3)
C(19)-C(20)-C(21)-C(16)	-1.4(6)
C(17)-C(16)-C(21)-C(20)	1.0(5)
S(2)-C(16)-C(21)-C(20)	176.5(3)

Symmetry transformations used to generate equivalent atoms:

Table 7. Hydrogen bonds for z [Å and deg.].

D-H...A	d(D-H)	d(H...A)	d(D...A)	<(DHA)
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5) Single-Crystal X-ray Crystallography of **6f**.¹⁴



Data intensity of **6f** was collected using a Bruker SMART APEX II (Mo radiation) at 296 K in a nitrogen stream. Data collection and reduction were done by using the Bruker ApexII software package. The structures were solved by direct methods and refined by full-matrix least-squares on F^2 with anisotropic displacement parameters for non-H atoms using SHELX-97. Hydrogen atoms were added at their geometrically ideal positions and refined isotropically. Crystal data for **6f**: $C_{21}H_{14}Cl_2OS_2$, $T = 296(2)$ K, Triclinic, space group P1, $a = 6.8482(4)$ Å, $b = 7.9264(4)$ Å, $c = 8.7986(5)$ Å, $\alpha = 92.069(2)$ deg, $\beta = 99.774(2)$ deg, $\gamma = 96.704(2)$ deg. $V = 466.69(4)$ Å³. $Z = 1$, $d_{\text{calc}} = 1.485$ mg/m³. Total number of reflections 5443 [$R(\text{int}) = 0.0167$], $R_1 = 0.0306$ ($I > 2\sigma(I)$), $wR_2 = 0.0769$ (all data), GOF = 1.033, and 235 parameters.

Table 1. Crystal data and structure refinement for **z**.

Identification code	z	
Empirical formula	C₂₁ H₁₄ Cl₂ O S₂	
Formula weight	417.34	
Temperature	296(2) K	
Wavelength	0.71073 Å	
Crystal system, space group	Triclinic, P1	
Unit cell dimensions	$a = 6.8482(4)$ Å	$\alpha = 92.069(2)$ deg.
	$b = 7.9264(4)$ Å	$\beta = 99.774(2)$ deg.
	$c = 8.7986(5)$ Å	$\gamma = 96.704(2)$ deg.
Volume	$466.69(4)$ Å ³	
Z, Calculated density	1, 1.485 Mg/m ³	

¹⁴ Supplementary crystallographic data have been deposited at the Cambridge Crystallographic Data Center. (CCDC 1413670).

Absorption coefficient	0.579 mm ⁻¹
F(000)	214
Crystal size	0.32 x 0.23 x 0.15 mm
Theta range for data collection	2.35 to 25.00 deg.
Limiting indices	-8<=h<=7, -9<=k<=9, -10<=l<=10
Reflections collected / unique	5443 / 2607 [R(int) = 0.0167]
Completeness to theta = 25.00	99.1 %
Absorption correction	Semi-empirical from equivalents
Max. and min. transmission	0.9182 and 0.8364
Refinement method	Full-matrix least-squares on F ²
Data / restraints / parameters	2607 / 3 / 235
Goodness-of-fit on F ²	1.033
Final R indices [I>2sigma(I)]	R1 = 0.0306, wR2 = 0.0758
R indices (all data)	R1 = 0.0319, wR2 = 0.0769
Absolute structure parameter	0.03(6)
Largest diff. peak and hole	0.417 and -0.190 e.A ⁻³

Table 2. Atomic coordinates (x 10⁴) and equivalent isotropic displacement parameters (A² x 10³) for z.

U(eq) is defined as one third of the trace of the orthogonalized U_{ij} tensor.

	x	y	z	U(eq)
S(1)	10503(1)	4483(1)	4796(1)	47(1)
S(2)	9767(1)	2596(1)	7738(1)	42(1)
O(1)	6610(3)	1598(3)	4900(2)	52(1)
Cl(1)	12122(2)	-3358(1)	1263(1)	79(1)
Cl(2)	5459(2)	9077(1)	8110(1)	75(1)
C(1)	8992(4)	164(4)	3788(3)	39(1)
C(2)	7890(5)	-934(4)	2619(3)	48(1)
C(3)	8857(6)	-2021(4)	1836(4)	56(1)
C(4)	10911(5)	-1983(4)	2259(3)	49(1)
C(5)	12042(5)	-893(4)	3428(3)	43(1)
C(6)	11041(4)	185(4)	4180(3)	38(1)
C(7)	11931(4)	1463(4)	5498(3)	46(1)
C(8)	10185(4)	2430(4)	5759(3)	38(1)
C(9)	8306(4)	1399(4)	4812(3)	38(1)

C(10)	12192(5)	2919(4)	8868(3)	40(1)
C(11)	13834(5)	3870(4)	8444(3)	46(1)
C(12)	15645(5)	4074(4)	9400(4)	53(1)
C(13)	15874(6)	3360(5)	10814(4)	62(1)
C(14)	14236(7)	2448(5)	11259(4)	66(1)
C(15)	12433(6)	2203(4)	10288(3)	53(1)
C(16)	9021(5)	5784(4)	5671(3)	41(1)
C(17)	9952(5)	6882(4)	6932(3)	46(1)
C(18)	8860(5)	7903(4)	7646(4)	51(1)
C(19)	6842(5)	7844(4)	7127(4)	48(1)
C(20)	5909(5)	6807(4)	5868(4)	51(1)
C(21)	6993(5)	5770(4)	5132(4)	48(1)

Table 3. Bond lengths [Å] and angles [deg] for z.

S(1)-C(16)	1.771(3)
S(1)-C(8)	1.869(3)
S(2)-C(10)	1.770(3)
S(2)-C(8)	1.814(3)
O(1)-C(9)	1.205(4)
Cl(1)-C(4)	1.741(3)
Cl(2)-C(19)	1.742(4)
C(1)-C(2)	1.380(4)
C(1)-C(6)	1.385(4)
C(1)-C(9)	1.476(4)
C(2)-C(3)	1.379(5)
C(2)-H(2A)	0.9300
C(3)-C(4)	1.389(5)
C(3)-H(3A)	0.9300
C(4)-C(5)	1.383(4)
C(5)-C(6)	1.374(4)
C(5)-H(5A)	0.9300
C(6)-C(7)	1.508(4)
C(7)-C(8)	1.538(4)
C(7)-H(7A)	0.9700
C(7)-H(7B)	0.9700
C(8)-C(9)	1.536(4)
C(10)-C(15)	1.383(4)
C(10)-C(11)	1.392(4)
C(11)-C(12)	1.364(5)
C(11)-H(11A)	0.9300
C(12)-C(13)	1.378(5)

C(12)-H(12A)	0.9300
C(13)-C(14)	1.384(5)
C(13)-H(13A)	0.9300
C(14)-C(15)	1.366(5)
C(14)-H(14A)	0.9300
C(15)-H(15A)	0.9300
C(16)-C(21)	1.388(5)
C(16)-C(17)	1.399(4)
C(17)-C(18)	1.369(5)
C(17)-H(17A)	0.9300
C(18)-C(19)	1.375(5)
C(18)-H(18A)	0.9300
C(19)-C(20)	1.372(5)
C(20)-C(21)	1.383(5)
C(20)-H(20A)	0.9300
C(21)-H(21A)	0.9300
C(16)-S(1)-C(8)	103.69(13)
C(10)-S(2)-C(8)	104.59(13)
C(2)-C(1)-C(6)	121.0(3)
C(2)-C(1)-C(9)	129.2(3)
C(6)-C(1)-C(9)	109.8(2)
C(3)-C(2)-C(1)	119.0(3)
C(3)-C(2)-H(2A)	120.5
C(1)-C(2)-H(2A)	120.5
C(2)-C(3)-C(4)	118.8(3)
C(2)-C(3)-H(3A)	120.6
C(4)-C(3)-H(3A)	120.6
C(5)-C(4)-C(3)	123.0(3)
C(5)-C(4)-Cl(1)	118.5(3)
C(3)-C(4)-Cl(1)	118.5(2)
C(6)-C(5)-C(4)	116.9(3)
C(6)-C(5)-H(5A)	121.5
C(4)-C(5)-H(5A)	121.5
C(5)-C(6)-C(1)	121.2(3)
C(5)-C(6)-C(7)	126.8(3)
C(1)-C(6)-C(7)	111.9(2)
C(6)-C(7)-C(8)	104.3(2)
C(6)-C(7)-H(7A)	110.9
C(8)-C(7)-H(7A)	110.9
C(6)-C(7)-H(7B)	110.9
C(8)-C(7)-H(7B)	110.9
H(7A)-C(7)-H(7B)	108.9

C(9)-C(8)-C(7)	105.6(2)
C(9)-C(8)-S(2)	106.72(19)
C(7)-C(8)-S(2)	115.5(2)
C(9)-C(8)-S(1)	105.54(19)
C(7)-C(8)-S(1)	106.8(2)
S(2)-C(8)-S(1)	115.80(15)
O(1)-C(9)-C(1)	127.7(3)
O(1)-C(9)-C(8)	125.5(3)
C(1)-C(9)-C(8)	106.8(2)
C(15)-C(10)-C(11)	118.5(3)
C(15)-C(10)-S(2)	116.8(2)
C(11)-C(10)-S(2)	124.7(2)
C(12)-C(11)-C(10)	120.7(3)
C(12)-C(11)-H(11A)	119.6
C(10)-C(11)-H(11A)	119.6
C(11)-C(12)-C(13)	120.5(3)
C(11)-C(12)-H(12A)	119.7
C(13)-C(12)-H(12A)	119.7
C(12)-C(13)-C(14)	119.1(3)
C(12)-C(13)-H(13A)	120.5
C(14)-C(13)-H(13A)	120.5
C(15)-C(14)-C(13)	120.5(3)
C(15)-C(14)-H(14A)	119.7
C(13)-C(14)-H(14A)	119.7
C(14)-C(15)-C(10)	120.6(3)
C(14)-C(15)-H(15A)	119.7
C(10)-C(15)-H(15A)	119.7
C(21)-C(16)-C(17)	119.3(3)
C(21)-C(16)-S(1)	122.4(2)
C(17)-C(16)-S(1)	118.3(2)
C(18)-C(17)-C(16)	120.2(3)
C(18)-C(17)-H(17A)	119.9
C(16)-C(17)-H(17A)	119.9
C(17)-C(18)-C(19)	119.9(3)
C(17)-C(18)-H(18A)	120.0
C(19)-C(18)-H(18A)	120.0
C(20)-C(19)-C(18)	120.8(3)
C(20)-C(19)-Cl(2)	120.0(3)
C(18)-C(19)-Cl(2)	119.2(3)
C(19)-C(20)-C(21)	119.9(3)
C(19)-C(20)-H(20A)	120.0
C(21)-C(20)-H(20A)	120.0

C(20)-C(21)-C(16)	119.8(3)
C(20)-C(21)-H(21A)	120.1
C(16)-C(21)-H(21A)	120.1

Symmetry transformations used to generate equivalent atoms:

Table 4. Anisotropic displacement parameters ($\text{\AA}^2 \times 10^3$) for z.

The anisotropic displacement factor exponent takes the form:

$$-2 \pi^2 [h^2 a^{*2} U_{11} + \dots + 2 h k a^* b^* U_{12}]$$

	U11	U22	U33	U23	U13	U12
S(1)	49(1)	50(1)	45(1)	8(1)	16(1)	5(1)
S(2)	42(1)	44(1)	41(1)	4(1)	11(1)	7(1)
O(1)	34(1)	60(1)	59(1)	-6(1)	3(1)	10(1)
Cl(1)	105(1)	70(1)	70(1)	-19(1)	28(1)	30(1)
Cl(2)	67(1)	68(1)	98(1)	7(1)	27(1)	26(1)
C(1)	39(2)	39(2)	40(1)	3(1)	7(1)	4(1)
C(2)	42(2)	50(2)	48(2)	-3(1)	6(1)	-5(1)
C(3)	69(3)	48(2)	46(2)	-10(1)	11(2)	-7(2)
C(4)	65(2)	42(2)	45(2)	0(1)	20(2)	11(2)
C(5)	47(2)	46(2)	39(1)	5(1)	12(1)	13(1)
C(6)	42(2)	37(2)	35(1)	3(1)	6(1)	7(1)
C(7)	37(2)	53(2)	45(2)	-10(1)	1(1)	13(1)
C(8)	36(2)	41(2)	37(1)	-1(1)	4(1)	9(1)
C(9)	34(2)	42(2)	40(1)	7(1)	6(1)	7(1)
C(10)	49(2)	35(2)	37(1)	-2(1)	8(1)	8(1)
C(11)	55(2)	44(2)	37(1)	2(1)	10(1)	3(2)
C(12)	47(2)	54(2)	53(2)	-9(2)	6(2)	0(2)
C(13)	58(2)	62(2)	60(2)	-4(2)	-8(2)	6(2)
C(14)	81(3)	65(2)	45(2)	12(2)	-8(2)	7(2)
C(15)	63(2)	51(2)	44(2)	8(1)	9(2)	0(2)
C(16)	44(2)	36(2)	46(2)	12(1)	12(1)	3(1)
C(17)	39(2)	43(2)	54(2)	7(1)	4(1)	3(1)
C(18)	51(2)	40(2)	60(2)	-1(1)	6(2)	2(2)
C(19)	48(2)	39(2)	62(2)	16(1)	12(2)	9(1)
C(20)	36(2)	48(2)	68(2)	23(2)	5(2)	8(1)
C(21)	43(2)	49(2)	49(2)	12(1)	-1(1)	6(2)

Table 5. Hydrogen coordinates ($\times 10^4$) and isotropic displacement parameters ($\text{\AA}^2 \times 10^3$) for z.

	x	y	z	U(eq)
H(2A)	6514	-941	2362	58
H(3A)	8148	-2767	1038	67
H(5A)	13416	-891	3693	51
H(7A)	13013	2230	5227	55
H(7B)	12434	896	6417	55
H(11A)	13695	4373	7499	55
H(12A)	16733	4700	9095	63
H(13A)	17113	3489	11460	75
H(14A)	14363	1997	12227	79
H(15A)	11358	1550	10585	63
H(17A)	11316	6919	7286	55
H(18A)	9483	8636	8482	61
H(20A)	4550	6801	5510	61
H(21A)	6365	5066	4278	58

Table 6. Torsion angles [deg] for z.

C(6)-C(1)-C(2)-C(3)	0.0(5)
C(9)-C(1)-C(2)-C(3)	-178.1(3)
C(1)-C(2)-C(3)-C(4)	0.5(5)
C(2)-C(3)-C(4)-C(5)	-0.4(5)
C(2)-C(3)-C(4)-Cl(1)	-179.9(2)
C(3)-C(4)-C(5)-C(6)	-0.1(4)
Cl(1)-C(4)-C(5)-C(6)	179.4(2)
C(4)-C(5)-C(6)-C(1)	0.6(4)
C(4)-C(5)-C(6)-C(7)	179.0(3)
C(2)-C(1)-C(6)-C(5)	-0.6(4)
C(9)-C(1)-C(6)-C(5)	177.9(3)
C(2)-C(1)-C(6)-C(7)	-179.2(3)
C(9)-C(1)-C(6)-C(7)	-0.8(3)
C(5)-C(6)-C(7)-C(8)	174.2(3)
C(1)-C(6)-C(7)-C(8)	-7.2(3)
C(6)-C(7)-C(8)-C(9)	11.8(3)
C(6)-C(7)-C(8)-S(2)	129.4(2)
C(6)-C(7)-C(8)-S(1)	-100.2(2)
C(10)-S(2)-C(8)-C(9)	155.4(2)
C(10)-S(2)-C(8)-C(7)	38.4(2)
C(10)-S(2)-C(8)-S(1)	-87.44(18)

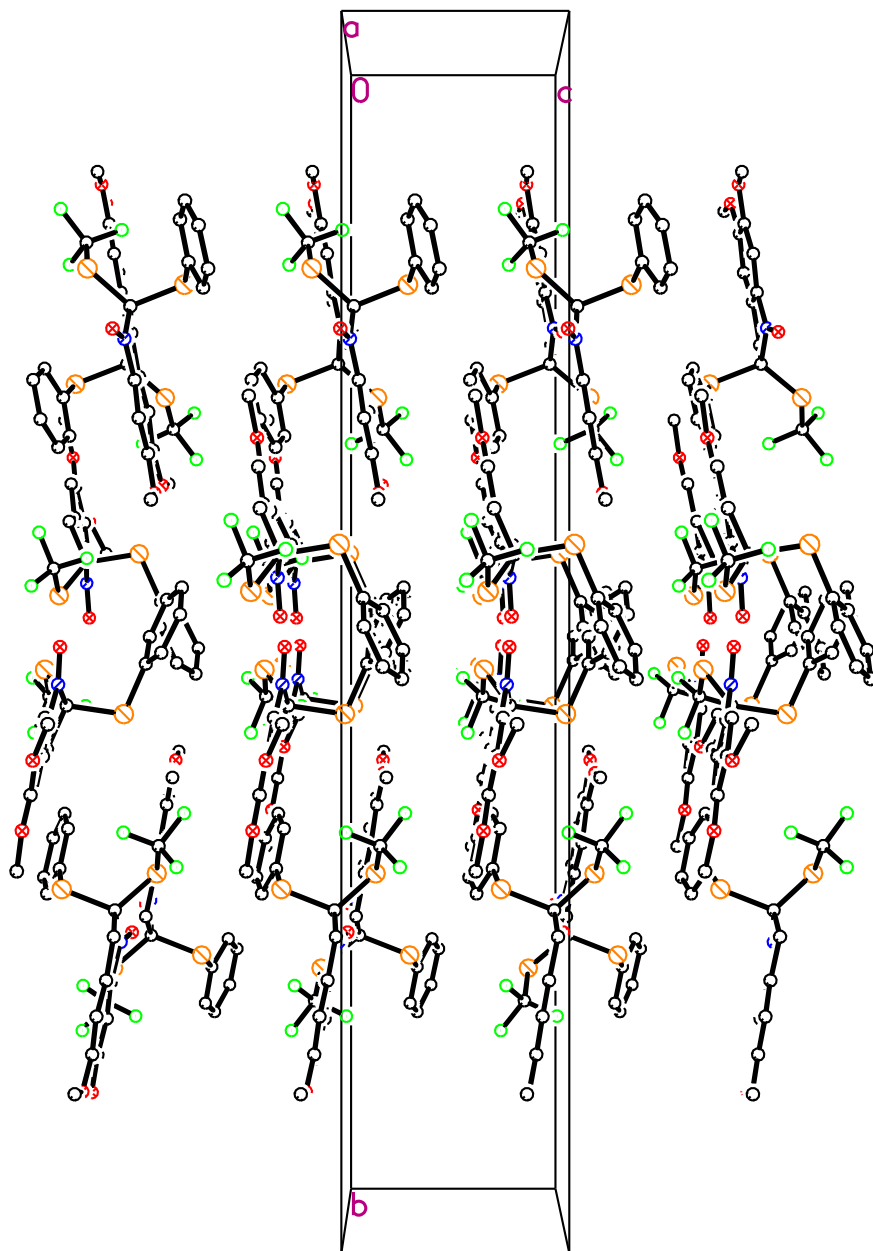
C(16)-S(1)-C(8)-C(9)	87.0(2)
C(16)-S(1)-C(8)-C(7)	-160.96(19)
C(16)-S(1)-C(8)-S(2)	-30.8(2)
C(2)-C(1)-C(9)-O(1)	6.1(5)
C(6)-C(1)-C(9)-O(1)	-172.2(3)
C(2)-C(1)-C(9)-C(8)	-173.2(3)
C(6)-C(1)-C(9)-C(8)	8.5(3)
C(7)-C(8)-C(9)-O(1)	168.1(3)
S(2)-C(8)-C(9)-O(1)	44.7(4)
S(1)-C(8)-C(9)-O(1)	-79.1(3)
C(7)-C(8)-C(9)-C(1)	-12.6(3)
S(2)-C(8)-C(9)-C(1)	-136.0(2)
S(1)-C(8)-C(9)-C(1)	100.3(2)
C(8)-S(2)-C(10)-C(15)	-145.6(2)
C(8)-S(2)-C(10)-C(11)	36.2(3)
C(15)-C(10)-C(11)-C(12)	0.8(5)
S(2)-C(10)-C(11)-C(12)	179.0(2)
C(10)-C(11)-C(12)-C(13)	-0.9(5)
C(11)-C(12)-C(13)-C(14)	-0.7(6)
C(12)-C(13)-C(14)-C(15)	2.4(6)
C(13)-C(14)-C(15)-C(10)	-2.6(6)
C(11)-C(10)-C(15)-C(14)	0.9(5)
S(2)-C(10)-C(15)-C(14)	-177.4(3)
C(8)-S(1)-C(16)-C(21)	-86.3(3)
C(8)-S(1)-C(16)-C(17)	94.9(2)
C(21)-C(16)-C(17)-C(18)	1.5(4)
S(1)-C(16)-C(17)-C(18)	-179.7(2)
C(16)-C(17)-C(18)-C(19)	0.3(5)
C(17)-C(18)-C(19)-C(20)	-1.9(5)
C(17)-C(18)-C(19)-Cl(2)	176.9(2)
C(18)-C(19)-C(20)-C(21)	1.8(5)
Cl(2)-C(19)-C(20)-C(21)	-177.1(2)
C(19)-C(20)-C(21)-C(16)	0.0(5)
C(17)-C(16)-C(21)-C(20)	-1.6(4)
S(1)-C(16)-C(21)-C(20)	179.6(2)

Symmetry transformations used to generate equivalent atoms:

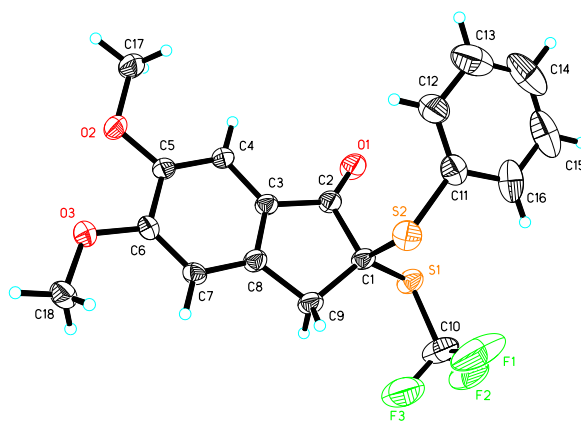
Table 7. Hydrogen bonds for z [A and deg.].

D-H...A	d(D-H)	d(H...A)	d(D...A)	<(DHA)
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6) Single-Crystal X-ray Crystallography of 8k.¹⁵



¹⁵ Supplementary crystallographic data have been deposited at the Cambridge Crystallographic Data Center. (CCDC 1413672).



Data intensity of **8k** was collected using a 'Bruker APEX-II CCD' diffractometer. Single crystals of $C_{18}H_{15}F_3O_3S_2$ [dm15546] is shown above. A suitable crystal was selected and analyzed on a 'Bruker APEX-II CCD' diffractometer. The crystal was kept at 296.15 K during data collection. Using Olex2, the structure was solved with the ShelXS structure solution program using Direct Methods and refined with the XL refinement package using Least Squares minimization. Crystal Data for $C_{18}H_{15}F_3O_3S_2$ ($M = 400.42$ g/mol): Orthorhombic, space group P 21 21 2, $a = 13.1691(14)$ Å, $b = 38.663(4)$ Å, $c = 7.1029(8)$ Å, $V = 3616.5(7)$ Å³, $Z = 8$, $T = 296.15$ K, $\mu(\text{MoK}\alpha) = 0.339$ mm⁻¹, $D_{\text{calc}} = 1.471$ g/cm³, 35999 reflections measured ($1.634 \leq 2\theta \leq 30.474$), 10915 unique ($R_{\text{int}} = 0.0451$) which were used in all calculations. The final R_1 was 0.0672 ($I > 2\sigma(I)$) and wR_2 was 0.2020 (all data).

Table 1. Crystal data and structure refinement for DM15546.

Identification code dm15546

Empirical formula $C_{18}H_{15}F_3O_3S_2$

Formula weight 400.42

Temperature 296.15 K

Wavelength 0.71073 Å

Crystal system Orthorhombic

Space group P 21 21 2

Unit cell dimensions $a = 13.1691(14)$ Å $= 90^\circ$.

$b = 38.663(4)$ Å $= 90^\circ$.

$c = 7.1029(8)$ Å $= 90^\circ$.

Volume 3616.5(7) Å³
 Z 8
 Density (calculated) 1.471 Mg/m³
 Absorption coefficient 0.339 mm⁻¹
 F(000) 1648
 Crystal size ? x ? x ? mm³
 Theta range for data collection 1.634 to 30.474°.
 Index ranges -18 ≤ h ≤ 18, -54 ≤ k ≤ 49, -9 ≤ l ≤ 10
 Reflections collected 35999
 Independent reflections 10915 [R(int) = 0.0451]
 Completeness to theta = 25.242° 99.5 %
 Absorption correction Semi-empirical from equivalents
 Max. and min. transmission 0.7461 and 0.6495
 Refinement method Full-matrix least-squares on F²
 Data / restraints / parameters 10915 / 90 / 474
 Goodness-of-fit on F² 1.143
 Final R indices [I > 2σ(I)] R1 = 0.0672, wR2 = 0.1789
 R indices (all data) R1 = 0.1130, wR2 = 0.2020
 Absolute structure parameter 0.02(12)
 Extinction coefficient n/a
 Largest diff. peak and hole 0.734 and -0.536 e.Å⁻³

Table 2. Atomic coordinates (x 104) and equivalent isotropic displacement parameters (Å² x 10³) for DM15546. U(eq) is defined as one third of the trace of the orthogonalized U_{ij} tensor.

x	y	z	U(eq)
S(1)	8319(1)	316(1)	3661(2) 56(1)
S(2)	7689(1)	682(1)	-29(2) 59(1)
F(1)	9843(5)	620(2)	2075(12) 177(3)
F(2)	10154(3)	392(1)	4596(10) 118(2)
F(3)	9431(4)	852(1)	4382(13) 144(2)
O(1)	6270(3)	124(1)	2591(7) 62(1)
O(2)	3045(2)	1011(1)	3145(6) 54(1)

O(3) 3966(3) 1583(1) 3799(6) 54(1)
 C(1) 7452(3) 608(1) 2518(8) 45(1)
 C(2) 6399(4) 431(1) 2693(8) 44(1)
 C(3) 5670(3) 715(1) 3005(7) 41(1)
 C(4) 4614(3) 689(1) 2927(7) 42(1)
 C(5) 4070(3) 987(1) 3211(7) 40(1)
 C(6) 4586(3) 1308(1) 3556(7) 41(1)
 C(7) 5621(3) 1320(1) 3598(7) 40(1)
 C(8) 6171(3) 1016(1) 3336(7) 43(1)
 C(9) 7296(3) 969(1) 3377(8) 49(1)
 C(10) 9455(6) 544(2) 3691(15) 87(2)
 C(11) 7549(5) 264(1) -1038(8)58(1)
 C(12) 6663(6) 175(2) -1865(10) 81(2)
 C(13) 6587(9) -147(3)-2720(13) 110(3)
 C(14) 7391(11) -369(2)-2703(12) 113(4)
 C(15) 8272(10) -271(2)-1900(13) 106(3)
 C(16) 8359(6) 44(2)-1103(10) 83(2)
 C(17) 2516(4) 721(1) 2403(10) 61(1)
 C(18) 4421(5) 1912(1) 4109(9) 59(1)
 S(3)3589(1) 2976(1) 11393(3) 68(1)
 S(4)3287(1) 2850(1) 7109(2) 62(1)
 F(4)1715(4) 3070(1) 12530(10) 118(2)
 F(5)2703(5) 3494(1) 12757(10) 132(2)
 F(6)2071(6) 3346(2) 10116(10) 148(2)
 O(4) 4949(3) 2444(1) 10055(8) 71(1)
 O(5) 3674(4) 1133(1) 8411(7) 76(1)
 O(6) 1757(4) 1223(1) 8322(7) 74(1)
 C(19) 3258(4) 2674(1) 9526(8) 51(1)
 C(20) 4063(4) 2387(1) 9674(8) 52(1)
 C(21) 3561(3) 2063(1) 9294(7) 48(1)
 C(22) 3990(4) 1743(2) 9043(7) 54(1)
 C(23) 3370(4) 1463(2) 8717(8) 59(1)
 C(24) 2292(4) 1509(2) 8664(8) 55(1)
 C(25) 1886(4) 1836(1) 8922(8) 51(1)
 C(26) 2518(3) 2113(1) 9256(7) 48(1)
 C(27) 2240(4) 2483(1) 9652(9) 55(1)
 C(28) 2485(7) 3226(2) 11641(13) 93(2)
 C(29) 4553(4) 2994(1) 6772(8) 48(1)
 C(30) 4820(4) 3327(2) 7105(10) 62(1)
 C(31) 5788(5) 3440(2) 6763(10) 70(2)
 C(32) 6499(5) 3217(2) 6087(10) 72(2)
 C(33) 6247(5) 2879(2) 5710(10) 76(2)

C(34) 5283(5) 2762(2) 6039(9) 64(2)
 C(35) 4729(5) 1067(2) 8402(10) 76(2)
 C(36) 686(6) 1248(2) 8184(13) 87(2)

Table 3. Bond lengths [Å] and angles [°] for DM15546.

S(1)-C(1) 1.801(5)
 S(1)-C(10) 1.738(7)
 S(2)-C(1) 1.858(6)
 S(2)-C(11) 1.776(6)
 F(1)-C(10) 1.290(11)
 F(2)-C(10) 1.268(9)
 F(3)-C(10) 1.286(10)
 O(1)-C(2) 1.201(5)
 O(2)-C(5) 1.353(5)
 O(2)-C(17) 1.423(6)
 O(3)-C(6) 1.351(5)
 O(3)-C(18) 1.426(6)
 C(1)-C(2) 1.552(6)
 C(1)-C(9) 1.536(6)
 C(2)-C(3) 1.474(6)
 C(3)-C(4) 1.396(6)
 C(3)-C(8) 1.359(6)
 C(4)-H(4) 0.9300
 C(4)-C(5) 1.372(6)
 C(5)-C(6) 1.435(6)
 C(6)-C(7) 1.364(6)
 C(7)-H(7) 0.9300
 C(7)-C(8) 1.392(6)
 C(8)-C(9) 1.494(6)
 C(9)-H(9A) 0.9700
 C(9)-H(9B) 0.9700
 C(11)-C(12) 1.351(10)
 C(11)-C(16) 1.366(9)
 C(12)-H(12) 0.9300
 C(12)-C(13) 1.388(11)
 C(13)-H(13) 0.9300
 C(13)-C(14) 1.364(15)
 C(14)-H(14) 0.9300
 C(14)-C(15) 1.346(15)
 C(15)-H(15) 0.9300
 C(15)-C(16) 1.347(12)

C(16)-H(16) 0.9300
 C(17)-H(17A) 0.9600
 C(17)-H(17B) 0.9600
 C(17)-H(17C) 0.9600
 C(18)-H(18A) 0.9600
 C(18)-H(18B) 0.9600
 C(18)-H(18C) 0.9600
 S(3)-C(19) 1.819(6)
 S(3)-C(28) 1.756(9)
 S(4)-C(19) 1.847(6)
 S(4)-C(29) 1.773(5)
 F(4)-C(28) 1.339(10)
 F(5)-C(28) 1.336(10)
 F(6)-C(28) 1.297(10)
 O(4)-C(20) 1.218(6)
 O(5)-C(23) 1.352(7)
 O(5)-C(35) 1.413(8)
 O(6)-C(24) 1.335(7)
 O(6)-C(36) 1.417(9)
 C(19)-C(20) 1.538(7)
 C(19)-C(27) 1.534(7)
 C(20)-C(21) 1.443(7)
 C(21)-C(22) 1.372(8)
 C(21)-C(26) 1.387(6)
 C(22)-H(22) 0.9300
 C(22)-C(23) 1.375(8)
 C(23)-C(24) 1.431(8)
 C(24)-C(25) 1.384(8)
 C(25)-H(25) 0.9300
 C(25)-C(26) 1.379(7)
 C(26)-C(27) 1.500(7)
 C(27)-H(27A) 0.9700
 C(27)-H(27B) 0.9700
 C(29)-C(30) 1.356(7)
 C(29)-C(34) 1.415(8)
 C(30)-H(30) 0.9300
 C(30)-C(31) 1.370(8)
 C(31)-H(31) 0.9300
 C(31)-C(32) 1.360(10)
 C(32)-H(32) 0.9300
 C(32)-C(33) 1.374(10)
 C(33)-H(33) 0.9300

C(33)-C(34) 1.369(9)
 C(34)-H(34) 0.9300
 C(35)-H(35A) 0.9600
 C(35)-H(35B) 0.9600
 C(35)-H(35C) 0.9600
 C(36)-H(36A) 0.9600
 C(36)-H(36B) 0.9600
 C(36)-H(36C) 0.9600
 C(10)-S(1)-C(1)103.4(3)
 C(11)-S(2)-C(1)103.7(2)
 C(5)-O(2)-C(17) 116.5(4)
 C(6)-O(3)-C(18) 118.0(4)
 S(1)-C(1)-S(2) 115.4(3)
 C(2)-C(1)-S(1)104.7(3)
 C(2)-C(1)-S(2)107.2(3)
 C(9)-C(1)-S(1)118.4(4)
 C(9)-C(1)-S(2)105.7(3)
 C(9)-C(1)-C(2) 104.5(4)
 O(1)-C(2)-C(1) 123.9(4)
 O(1)-C(2)-C(3) 130.8(4)
 C(3)-C(2)-C(1) 105.4(3)
 C(4)-C(3)-C(2) 126.1(4)
 C(8)-C(3)-C(2) 110.4(4)
 C(8)-C(3)-C(4) 123.5(4)
 C(3)-C(4)-H(4) 121.5
 C(5)-C(4)-C(3) 117.0(4)
 C(5)-C(4)-H(4) 121.5
 O(2)-C(5)-C(4) 125.0(4)
 O(2)-C(5)-C(6) 114.7(4)
 C(4)-C(5)-C(6) 120.2(4)
 O(3)-C(6)-C(5) 114.6(4)
 O(3)-C(6)-C(7) 125.0(4)
 C(7)-C(6)-C(5) 120.5(4)
 C(6)-C(7)-H(7) 120.4
 C(6)-C(7)-C(8) 119.2(4)
 C(8)-C(7)-H(7) 120.4
 C(3)-C(8)-C(7) 119.6(4)
 C(3)-C(8)-C(9) 112.3(4)
 C(7)-C(8)-C(9) 128.1(4)
 C(1)-C(9)-H(9A)111.0
 C(1)-C(9)-H(9B) 111.0
 C(8)-C(9)-C(1) 103.6(4)

C(8)-C(9)-H(9A) 111.0
 C(8)-C(9)-H(9B) 111.0
 H(9A)-C(9)-H(9B) 109.0
 F(1)-C(10)-S(1) 116.4(6)
 F(2)-C(10)-S(1) 113.3(6)
 F(2)-C(10)-F(1) 105.6(8)
 F(2)-C(10)-F(3) 104.7(7)
 F(3)-C(10)-S(1) 117.0(7)
 F(3)-C(10)-F(1) 98.0(8)
 C(12)-C(11)-S(2) 119.8(5)
 C(12)-C(11)-C(16) 120.0(7)
 C(16)-C(11)-S(2) 120.0(6)
 C(11)-C(12)-H(12) 120.6
 C(11)-C(12)-C(13) 118.8(9)
 C(13)-C(12)-H(12) 120.6
 C(12)-C(13)-H(13) 119.9
 C(14)-C(13)-C(12) 120.3(10)
 C(14)-C(13)-H(13) 119.9
 C(13)-C(14)-H(14) 120.1
 C(15)-C(14)-C(13) 119.8(8)
 C(15)-C(14)-H(14) 120.1
 C(14)-C(15)-H(15) 119.9
 C(14)-C(15)-C(16) 120.3(10)
 C(16)-C(15)-H(15) 119.9
 C(11)-C(16)-H(16) 119.6
 C(15)-C(16)-C(11) 120.8(10)
 C(15)-C(16)-H(16) 119.6
 O(2)-C(17)-H(17A) 109.5
 O(2)-C(17)-H(17B) 109.5
 O(2)-C(17)-H(17C) 109.5
 H(17A)-C(17)-H(17B) 109.5
 H(17A)-C(17)-H(17C) 109.5
 H(17B)-C(17)-H(17C) 109.5
 O(3)-C(18)-H(18A) 109.5
 O(3)-C(18)-H(18B) 109.5
 O(3)-C(18)-H(18C) 109.5
 H(18A)-C(18)-H(18B) 109.5
 H(18A)-C(18)-H(18C) 109.5
 H(18B)-C(18)-H(18C) 109.5
 C(28)-S(3)-C(19) 103.2(3)
 C(29)-S(4)-C(19) 105.1(2)
 C(23)-O(5)-C(35) 117.5(5)

C(24)-O(6)-C(36) 118.7(5)
 S(3)-C(19)-S(4) 115.9(3)
 C(20)-C(19)-S(3) 104.3(4)
 C(20)-C(19)-S(4) 108.4(4)
 C(27)-C(19)-S(3) 118.4(4)
 C(27)-C(19)-S(4) 104.5(4)
 C(27)-C(19)-C(20) 104.5(4)
 O(4)-C(20)-C(19) 123.0(5)
 O(4)-C(20)-C(21) 129.7(5)
 C(21)-C(20)-C(19) 107.3(4)
 C(22)-C(21)-C(20) 128.2(4)
 C(22)-C(21)-C(26) 122.2(5)
 C(26)-C(21)-C(20) 109.5(5)
 C(21)-C(22)-H(22) 120.4
 C(21)-C(22)-C(23) 119.2(5)
 C(23)-C(22)-H(22) 120.4
 O(5)-C(23)-C(22) 126.4(5)
 O(5)-C(23)-C(24) 114.0(5)
 C(22)-C(23)-C(24) 119.6(5)
 O(6)-C(24)-C(23) 115.1(5)
 O(6)-C(24)-C(25) 125.3(5)
 C(25)-C(24)-C(23) 119.6(5)
 C(24)-C(25)-H(25) 120.0
 C(26)-C(25)-C(24) 120.0(5)
 C(26)-C(25)-H(25) 120.0
 C(21)-C(26)-C(27) 111.9(4)
 C(25)-C(26)-C(21) 119.4(5)
 C(25)-C(26)-C(27) 128.7(4)
 C(19)-C(27)-H(27A) 111.1
 C(19)-C(27)-H(27B) 111.1
 C(26)-C(27)-C(19) 103.5(4)
 C(26)-C(27)-H(27A) 111.1
 C(26)-C(27)-H(27B) 111.1
 H(27A)-C(27)-H(27B) 109.0
 F(4)-C(28)-S(3) 115.1(6)
 F(5)-C(28)-S(3) 108.0(7)
 F(5)-C(28)-F(4) 103.5(7)
 F(6)-C(28)-S(3) 117.4(6)
 F(6)-C(28)-F(4) 103.7(8)
 F(6)-C(28)-F(5) 108.1(7)
 C(30)-C(29)-S(4) 121.2(4)
 C(30)-C(29)-C(34) 119.4(5)

C(34)-C(29)-S(4) 119.3(4)
 C(29)-C(30)-H(30) 119.6
 C(29)-C(30)-C(31) 120.9(6)
 C(31)-C(30)-H(30) 119.6
 C(30)-C(31)-H(31) 120.0
 C(32)-C(31)-C(30) 120.1(6)
 C(32)-C(31)-H(31) 120.0
 C(31)-C(32)-H(32) 119.8
 C(31)-C(32)-C(33) 120.4(6)
 C(33)-C(32)-H(32) 119.8
 C(32)-C(33)-H(33) 119.8
 C(34)-C(33)-C(32) 120.4(6)
 C(34)-C(33)-H(33) 119.8
 C(29)-C(34)-H(34) 120.5
 C(33)-C(34)-C(29) 118.9(6)
 C(33)-C(34)-H(34) 120.5
 O(5)-C(35)-H(35A) 109.5
 O(5)-C(35)-H(35B) 109.5
 O(5)-C(35)-H(35C) 109.5
 H(35A)-C(35)-H(35B) 109.5
 H(35A)-C(35)-H(35C) 109.5
 H(35B)-C(35)-H(35C) 109.5
 O(6)-C(36)-H(36A) 109.5
 O(6)-C(36)-H(36B) 109.5
 O(6)-C(36)-H(36C) 109.5
 H(36A)-C(36)-H(36B) 109.5
 H(36A)-C(36)-H(36C) 109.5
 H(36B)-C(36)-H(36C) 109.5

Symmetry transformations used to generate equivalent atoms:

Table 4. Anisotropic displacement parameters ($\text{\AA}^2 \times 10^3$) for DM15546. The anisotropic displacement factor exponent takes the form: $-2 \left[h^2 a^{*2} U_{11} + \dots + 2 h k a^* b^* U_{12} \right]$

U11U22 U33U23U13U12

S(1)44(1) 48(1)76(1) 9(1)-8(1) 2(1)
 S(2)65(1) 47(1)65(1) 10(1)5(1) -2(1)
 F(1)97(4) 263(6) 172(5) 42(5)-21(4) -86(4)
 F(2)64(2) 109(3) 182(5) 20(3)-51(3) 8(2)
 F(3)83(3) 95(3)254(6) -15(4)-37(4) -15(3)
 O(1) 49(2) 35(2)101(3) 4(2)3(2) -6(1)
 O(2) 34(2) 48(2)81(3) -7(2)-2(2) 0(1)

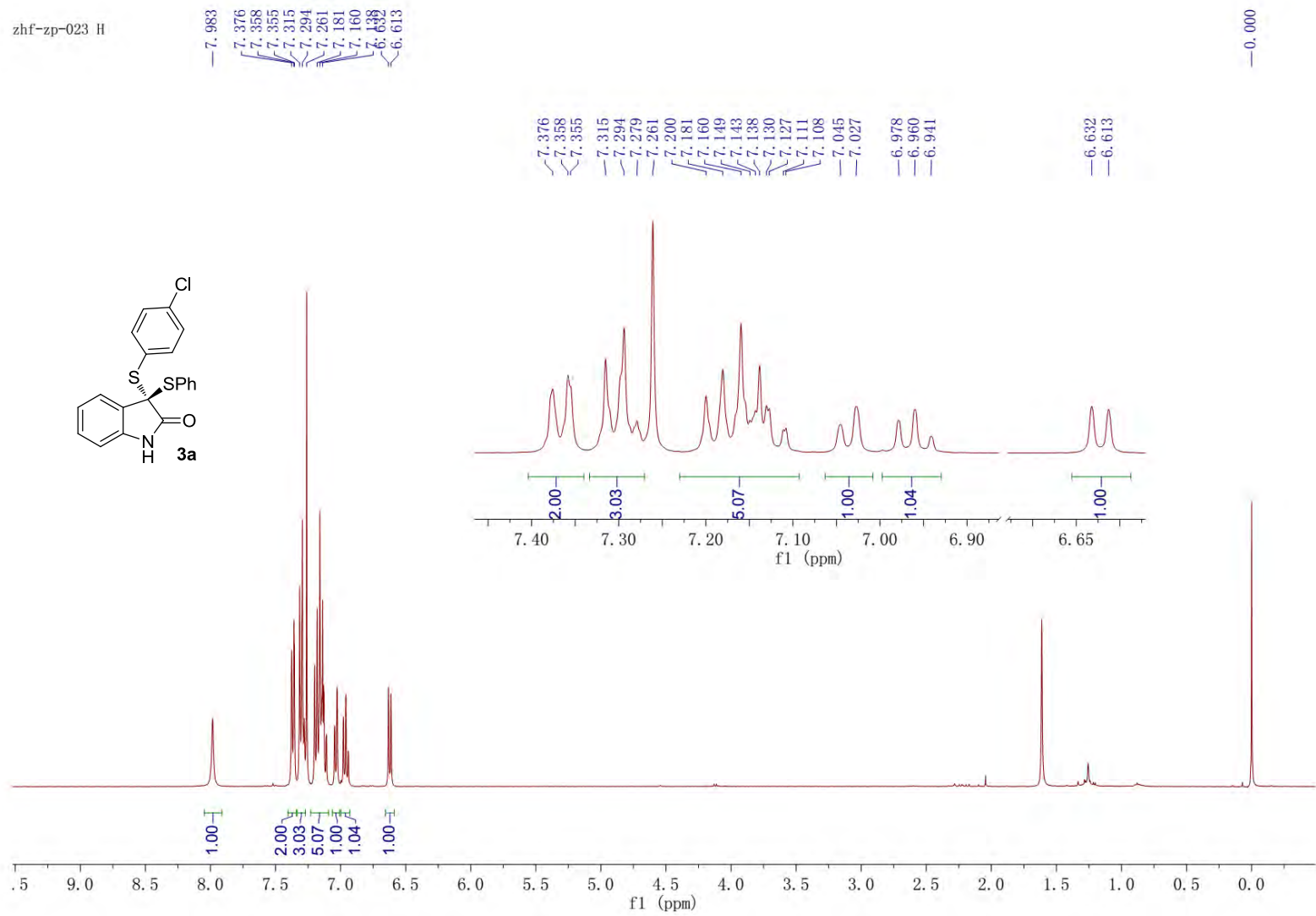
O(3) 48(2) 41(2)74(2) -10(2)-2(2) 5(1)
 C(1) 36(2) 33(2)66(3) 0(2)-6(2) -3(2)
 C(2) 40(2) 33(2)60(3) 1(2)0(2) -4(2)
 C(3) 37(2) 35(2)50(2) 1(2)-2(2) -5(2)
 C(4) 35(2) 41(2)50(2) -4(2) 1(2) -5(2)
 C(5) 36(2) 40(2)44(3) -1(2) 1(2) -5(2)
 C(6) 45(2) 34(2)44(2) -4(2) 2(2) 4(2)
 C(7) 45(2) 28(2)48(2) 0(2)3(2) -6(2)
 C(8) 37(2) 38(2)55(3) 3(2)-1(2) -2(2)
 C(9) 39(2) 38(2)71(3) 0(2)-5(2) -5(2)
 C(10) 60(3) 79(3)122(4) 14(3)-24(3) -17(3)
 C(11) 70(3) 50(3)56(3) 4(2)15(3) -2(3)
 C(12) 80(4) 91(5)71(4) -23(4)6(4) -13(4)
 C(13) 140(9) 106(7) 86(6) -18(5)-1(6) -44(7)
 C(14) 216(13) 62(5)61(5) -5(4) 33(7) -18(7)
 C(15) 175(10) 68(5)75(5) 7(4)42(6) 37(6)
 C(16) 100(5) 82(4)69(4) 18(3)30(4) 36(4)
 C(17) 42(3) 56(3)84(4) -8(3) -3(3) -6(2)
 C(18) 65(3) 41(3)70(4) -8(2) -6(3) 4(2)
 S(3)57(1) 76(1)70(1) -3(1)-4(1) -11(1)
 S(4)42(1) 73(1)71(1) 19(1)-10(1) -14(1)
 F(4)87(3) 107(3) 159(5) -24(3)32(3) -1(3)
 F(5)144(4) 94(3)160(5) -58(3)8(4) -3(3)
 F(6)171(5) 146(4) 128(4) 14(4)-6(4) 86(4)
 O(4) 31(2) 84(3)99(3) 16(3)-6(2) -9(2)
 O(5) 75(3) 64(2)90(3) -6(2) -3(3) 14(2)
 O(6) 70(3) 61(2)90(3) 5(2)-10(2) -12(2)
 C(19) 36(2) 50(3)68(3) 6(2)1(2) -6(2)
 C(20) 36(2) 60(3)60(3) 7(3)2(2) -3(2)
 C(21) 35(2) 60(3)50(3) 13(2)0(2) -2(2)
 C(22) 45(2) 67(3)49(3) 12(2)0(2) 0(2)
 C(23) 61(3) 63(3)53(3) 9(3)2(3) 7(3)
 C(24) 56(3) 58(3)50(3) 5(2)-3(2) -11(2)
 C(25) 39(2) 59(3)55(3) 5(2)2(2) -8(2)
 C(26) 36(2) 60(3)47(2) 8(2)3(2) -3(2)
 C(27) 34(2) 57(3)75(4) 5(3)4(2) -7(2)
 C(28) 102(4) 82(4)96(4) -13(3)5(4) 13(3)
 C(29) 40(2) 50(3)56(3) 6(2)-3(2) -7(2)
 C(30) 52(3) 52(3)82(4) -3(3) 2(3) -6(2)
 C(31) 70(4) 60(3)81(4) 3(3)-2(3) -26(3)
 C(32) 49(3) 92(5)75(4) 24(4)6(3) -12(3)
 C(33) 62(4) 98(5)69(4) 12(4)16(3) 13(4)

C(34) 76(4) 46(3)69(4) -4(3) 10(3) -5(3)
 C(35) 76(4) 84(4)69(4) 5(4)2(3) 18(4)
 C(36) 74(4) 78(4)109(6) -8(4) -10(4) -32(4)

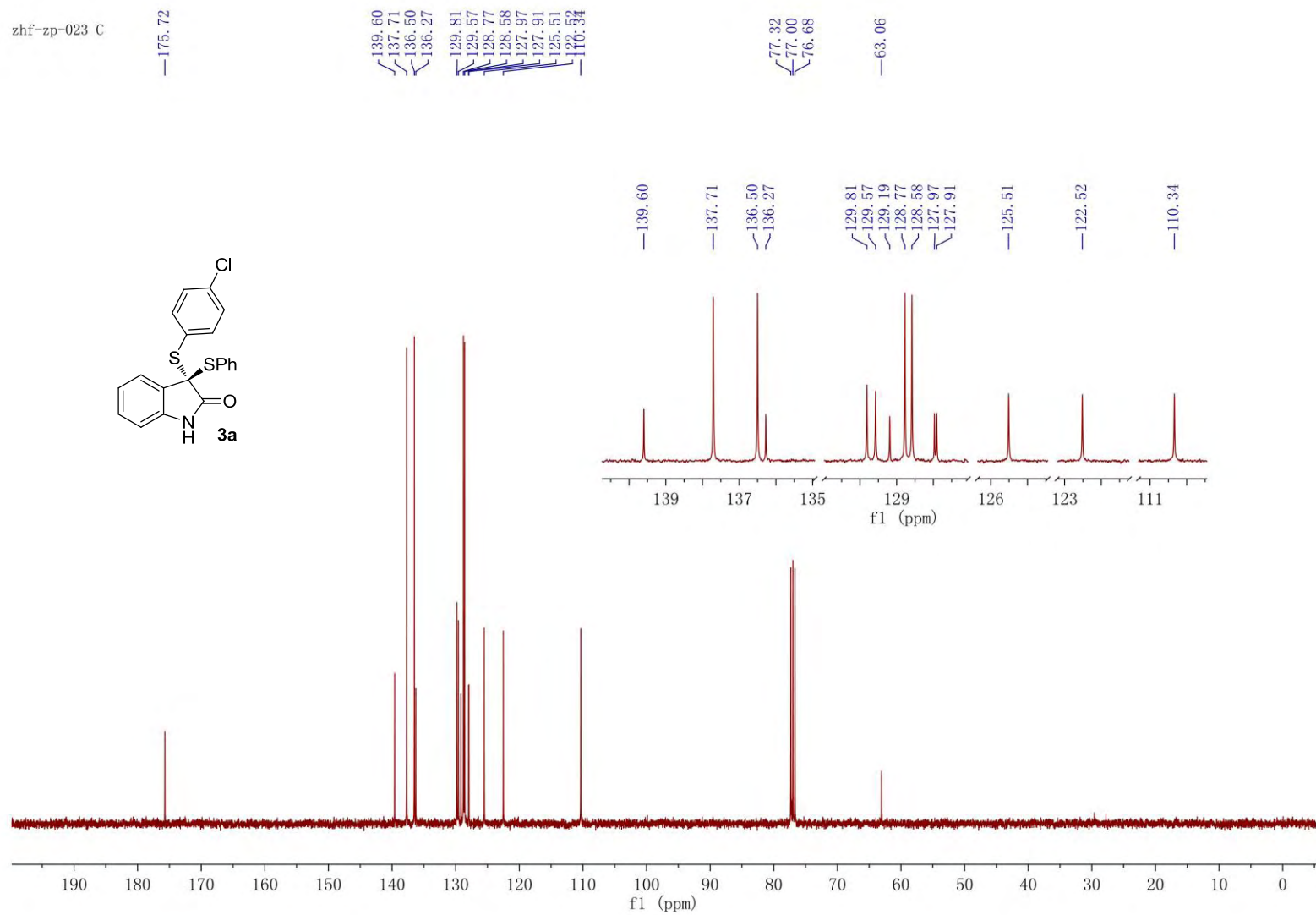
Table 5. Hydrogen coordinates (x 104) and isotropic displacement parameters ($\text{\AA}^2 \times 10^3$) for DM15546.

x	y	z	U(eq)
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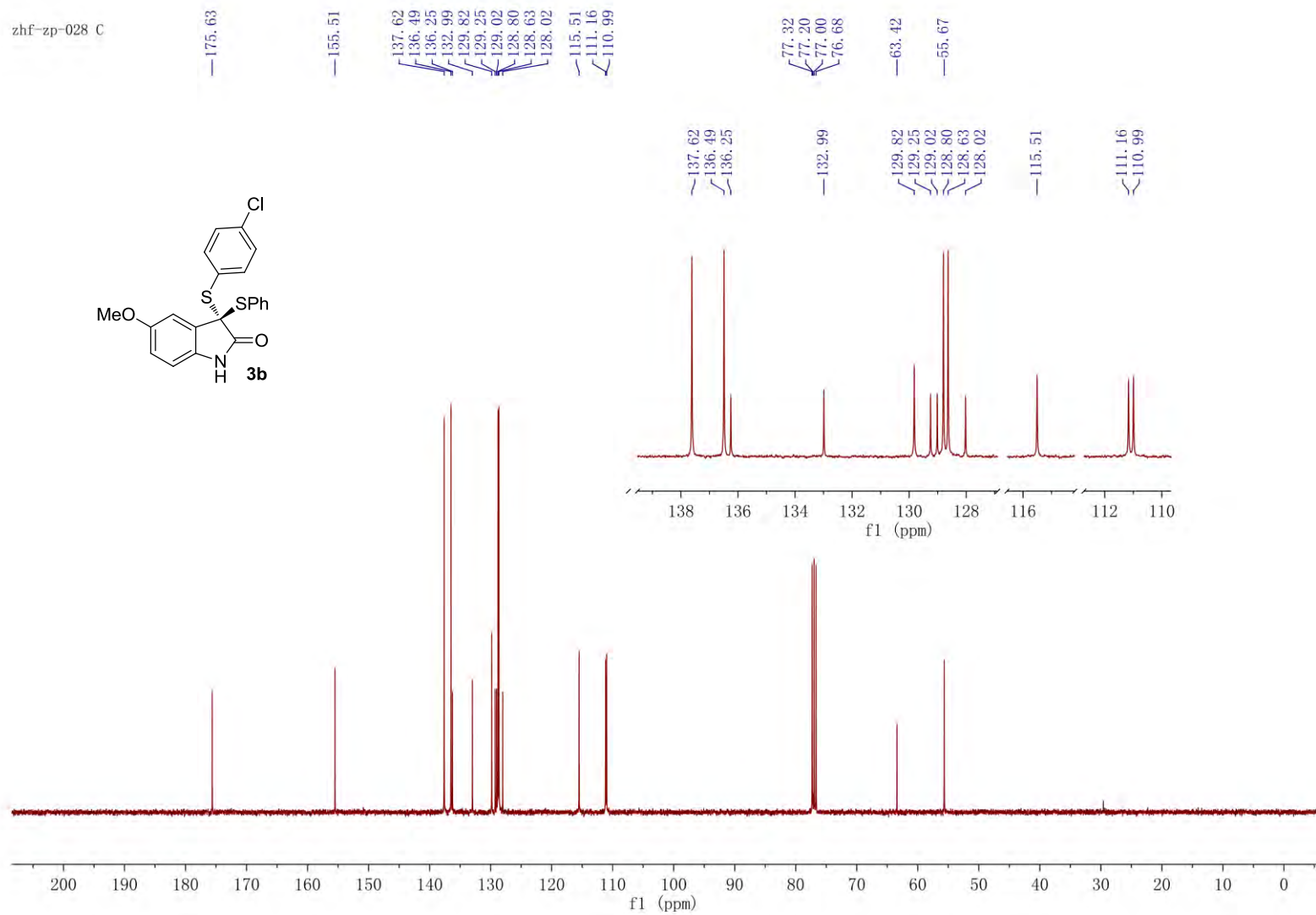
H(4)	4293	479	2693	51
H(7)	5955	1529	3799	48
H(9A)	7553	978	4656	59
H(9B)	7634	1145	2631	59
H(12)	6115	327	-1862	97
H(13)	5984	-211	-3307	132
H(14)	7332	-587	-3244	136
H(15)	8823	-422	-1895	127
H(16)	8977	111	-590	100
H(17A)	2829	649	1247	91
H(17B)	1821	783	2168	91
H(17C)	2539	534	3294	91
H(18A)	4869	1966	3084	88
H(18B)	4800	1908	5264	88
H(18C)	3901	2086	4189	88
H(22)	4691	1715	9093	64
H(25)	1187	1868	8871	61
H(27A)	1944	2506	10896	66
H(27B)	1763	2569	8724	66
H(30)	4339	3481	7573	75
H(31)	5960	3670	6993	84
H(32)	7159	3294	5879	87
H(33)	6735	2729	5229	92
H(34)	5110	2533	5784	76
H(35A)	5037	1185	7356	115
H(35B)	4844	823	8288	115
H(35C)	5024	1149	9554	115
H(36A)	434	1065	7397	130
H(36B)	506	1467	7646	130
H(36C)	391	1228	9416	130

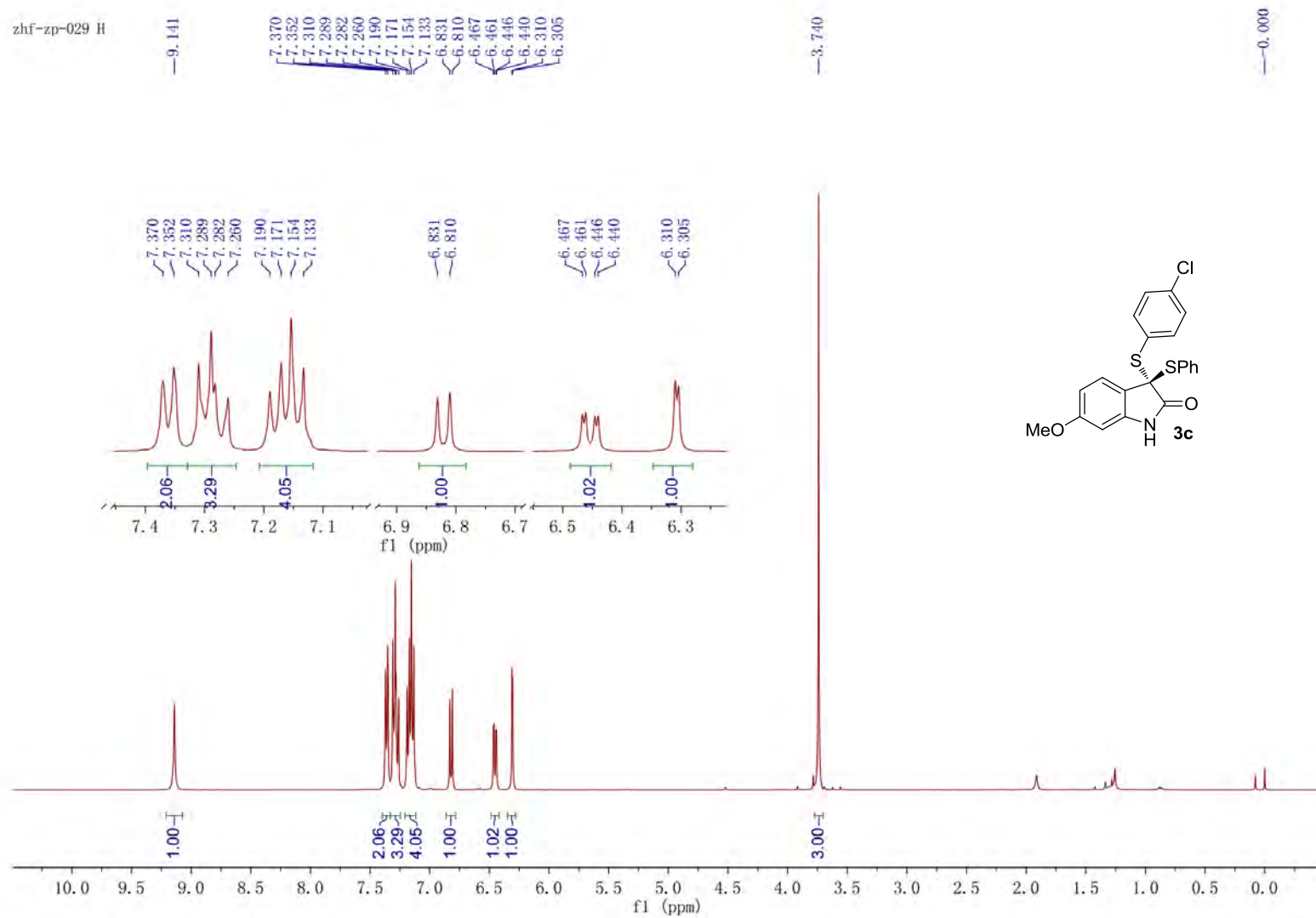


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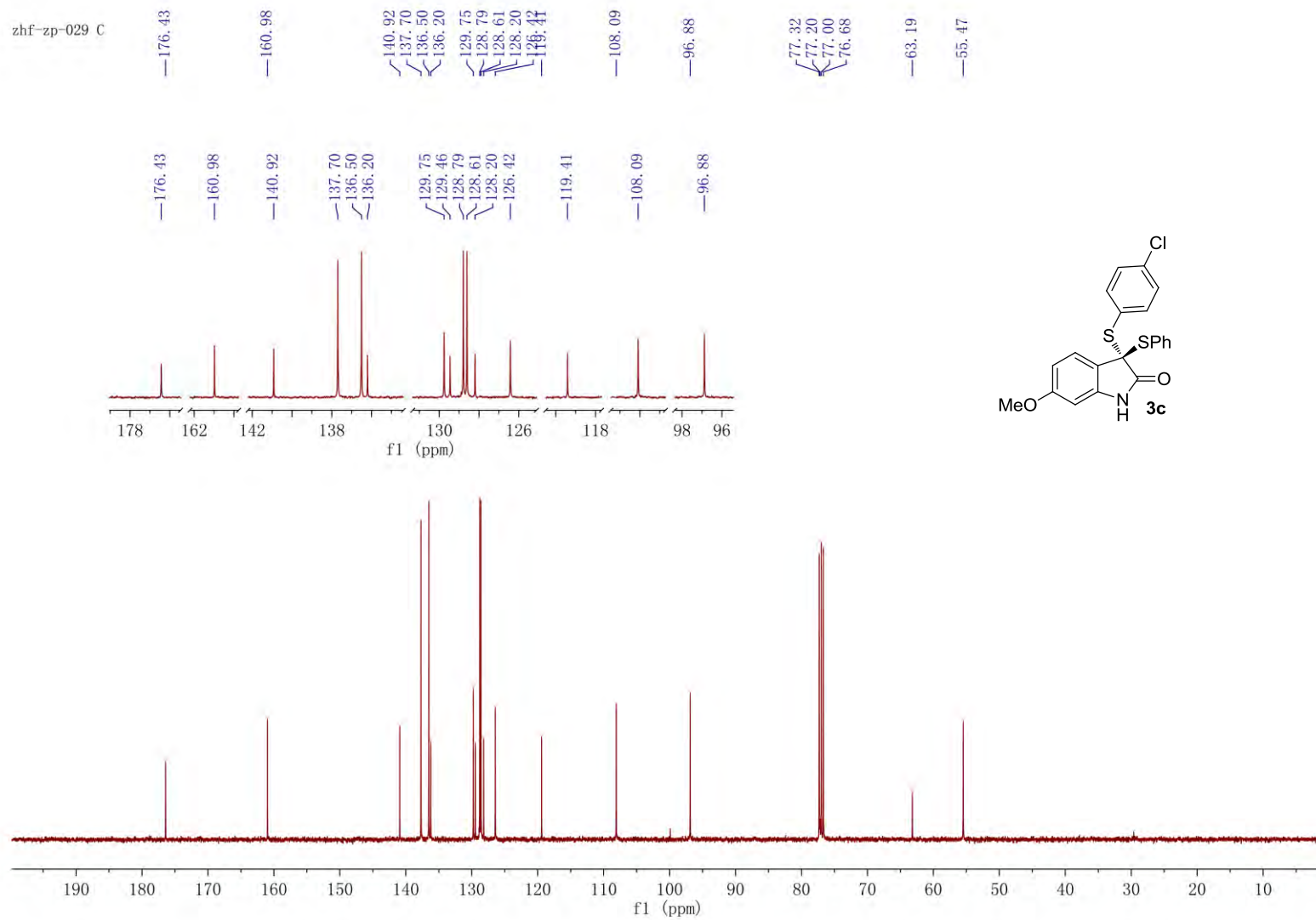


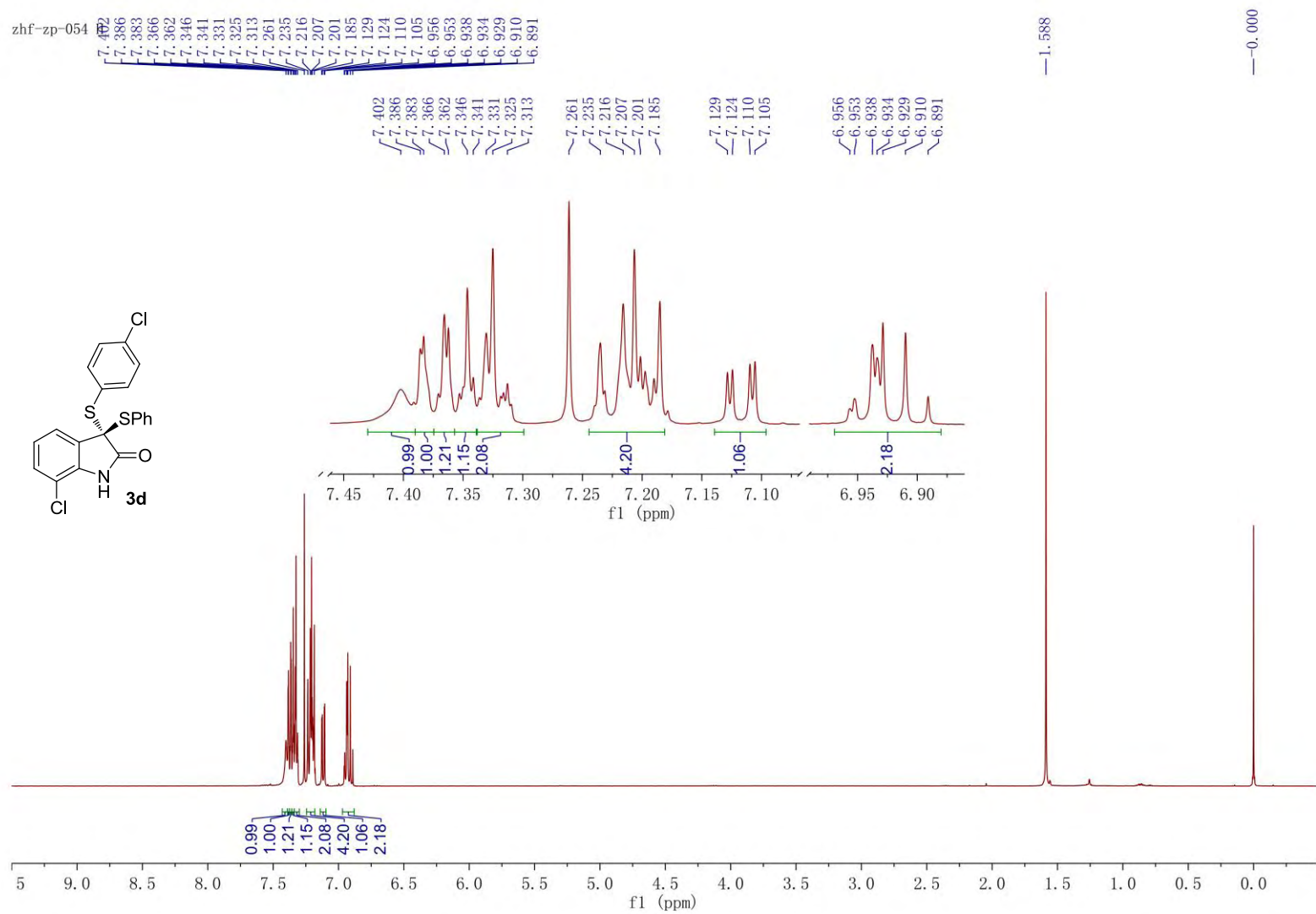
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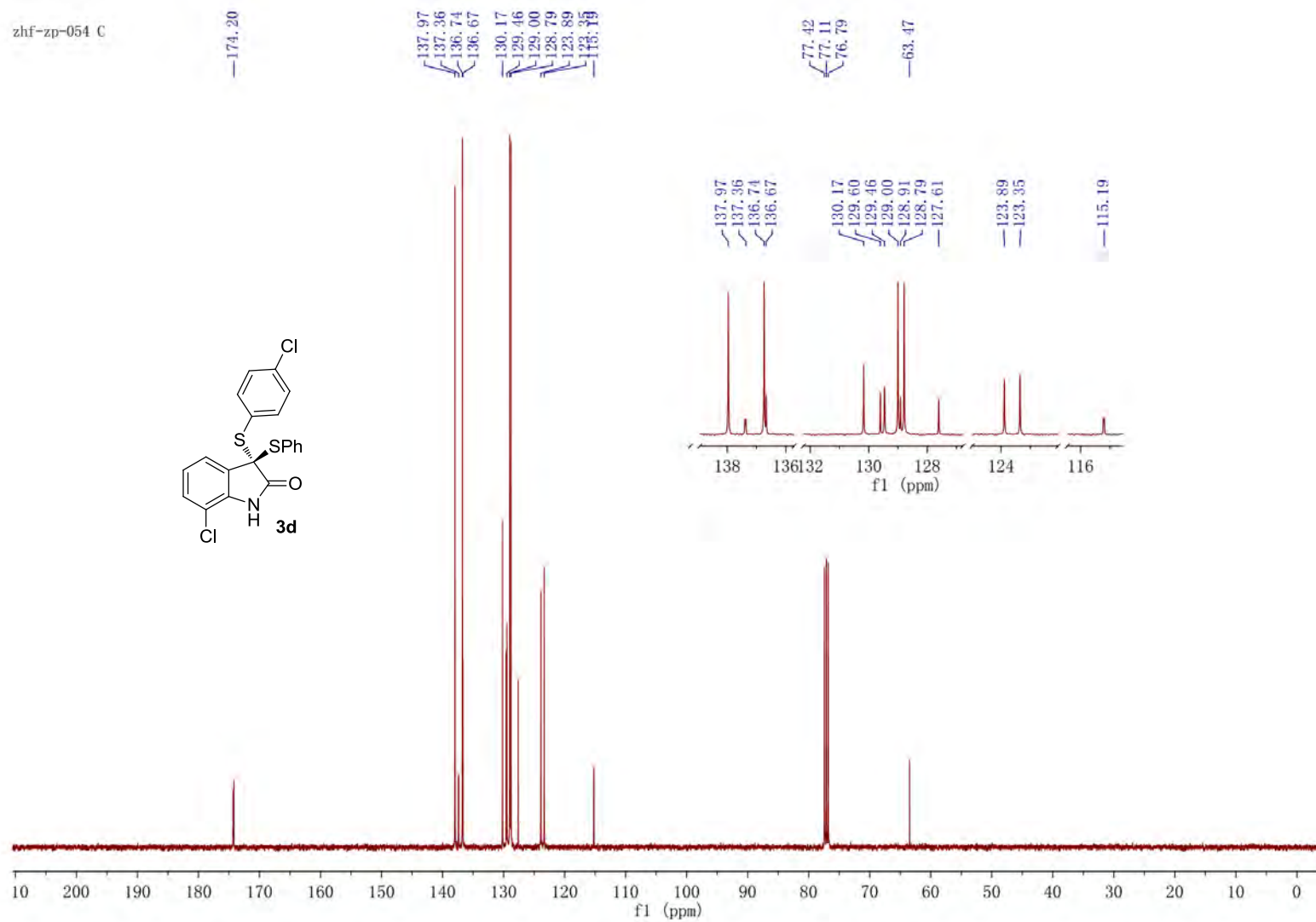


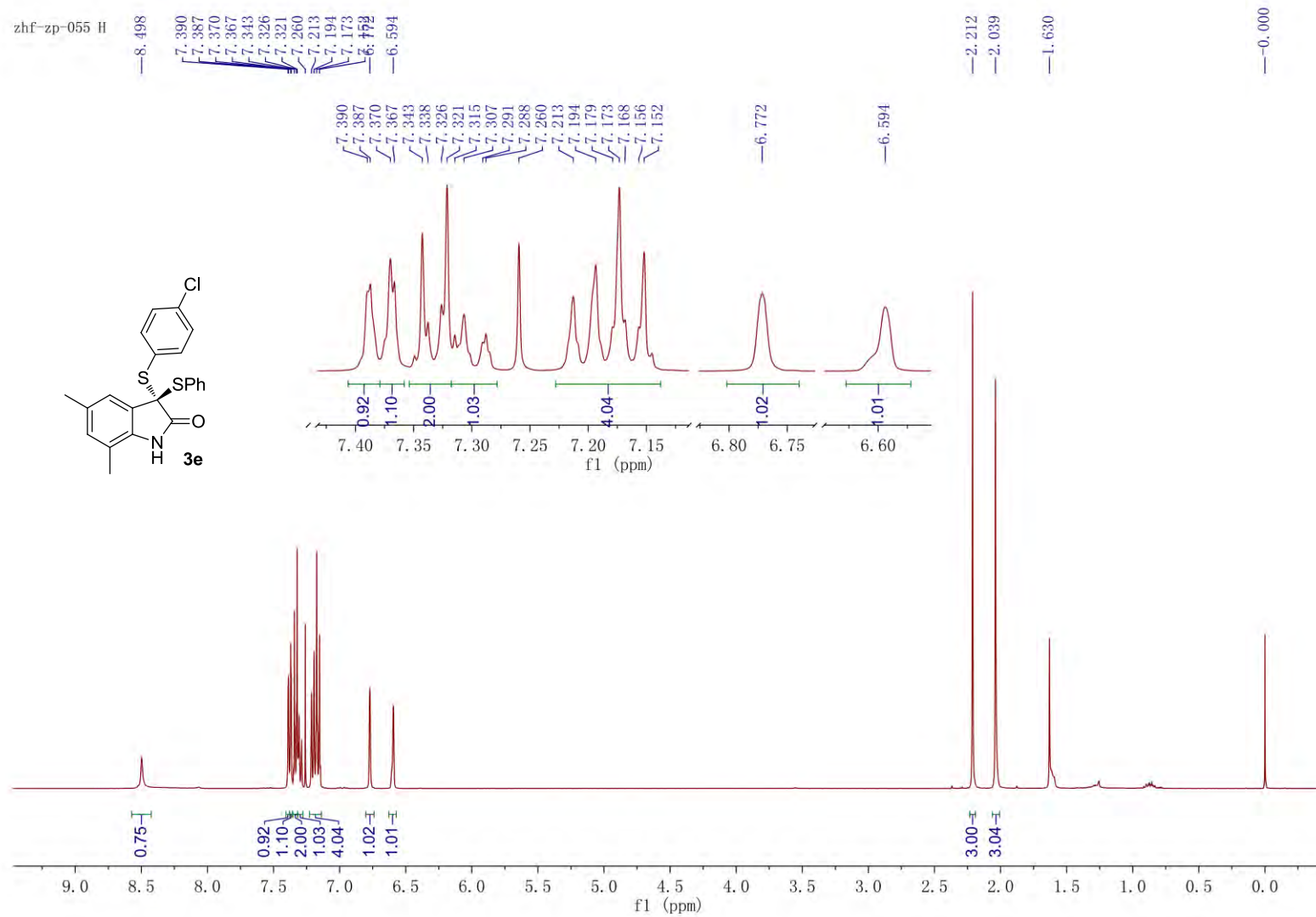
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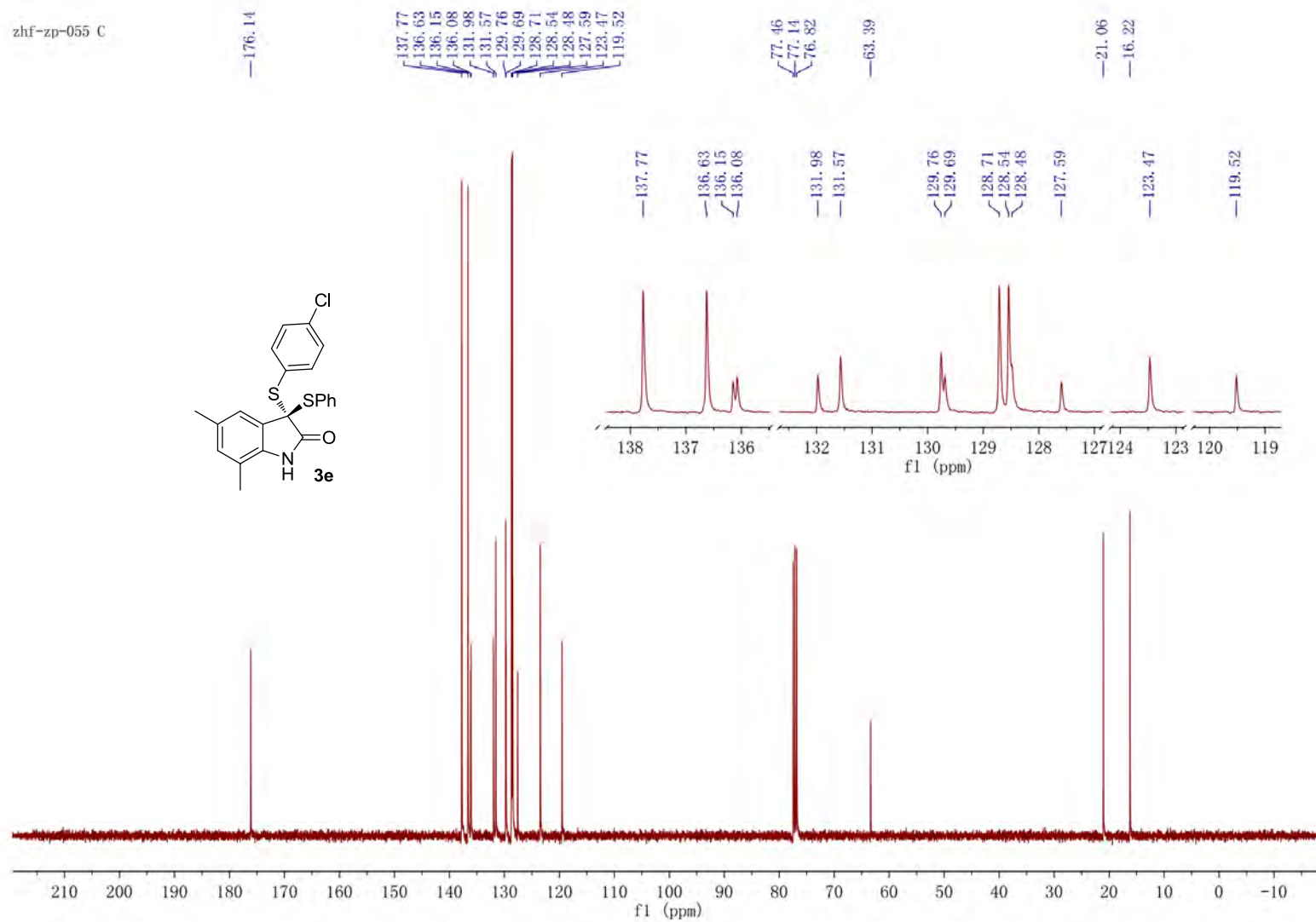
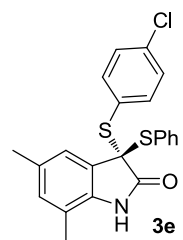


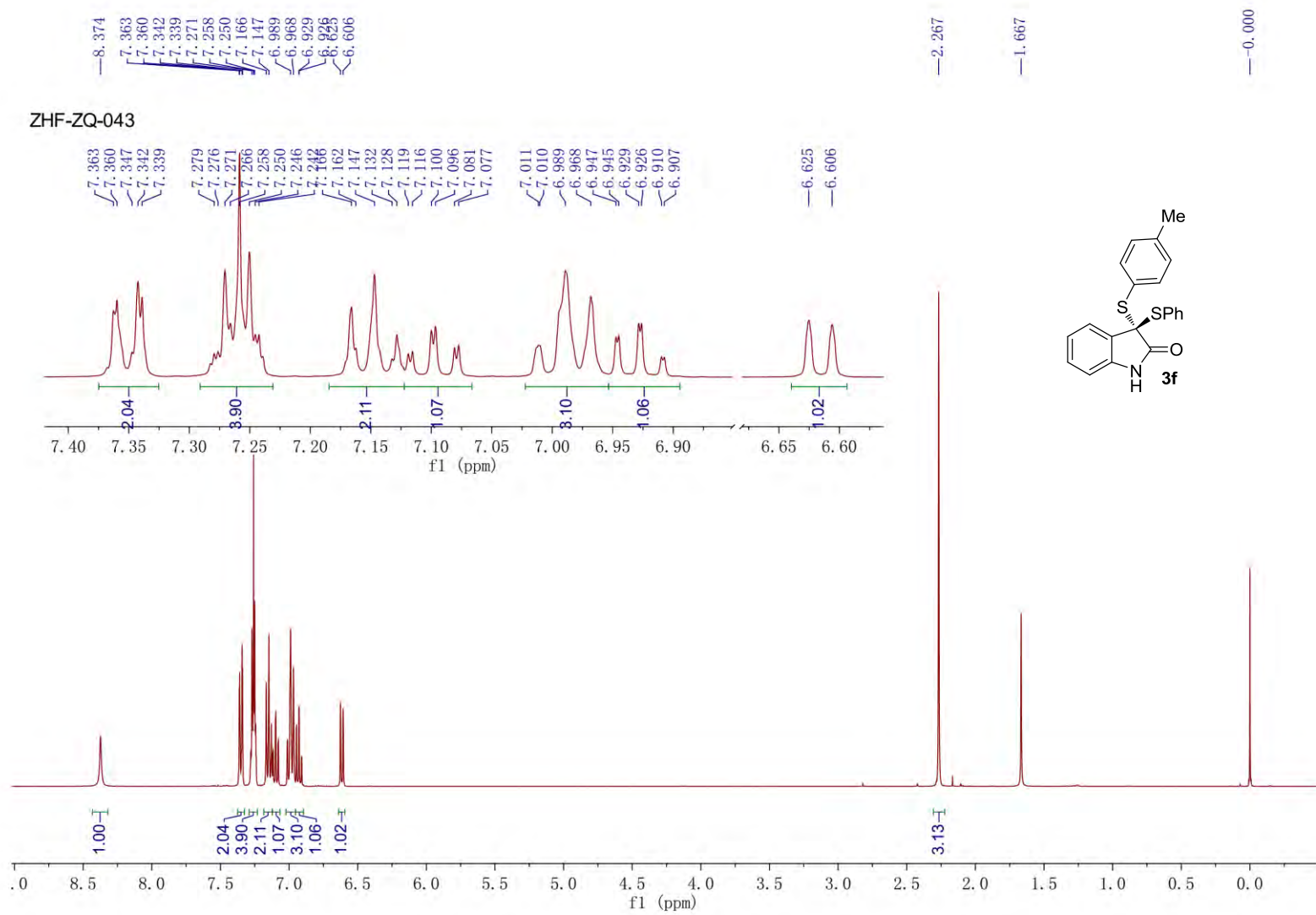
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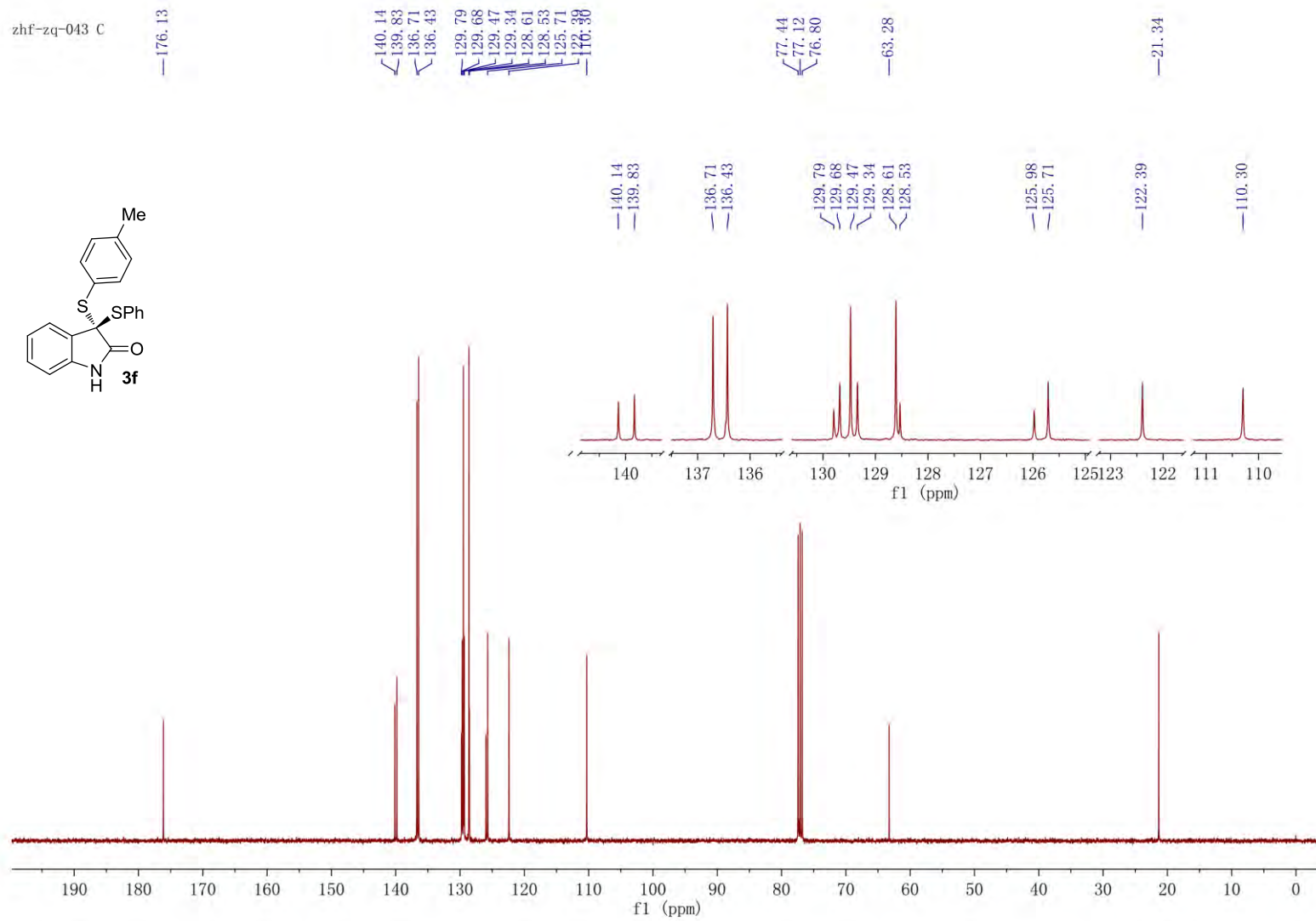


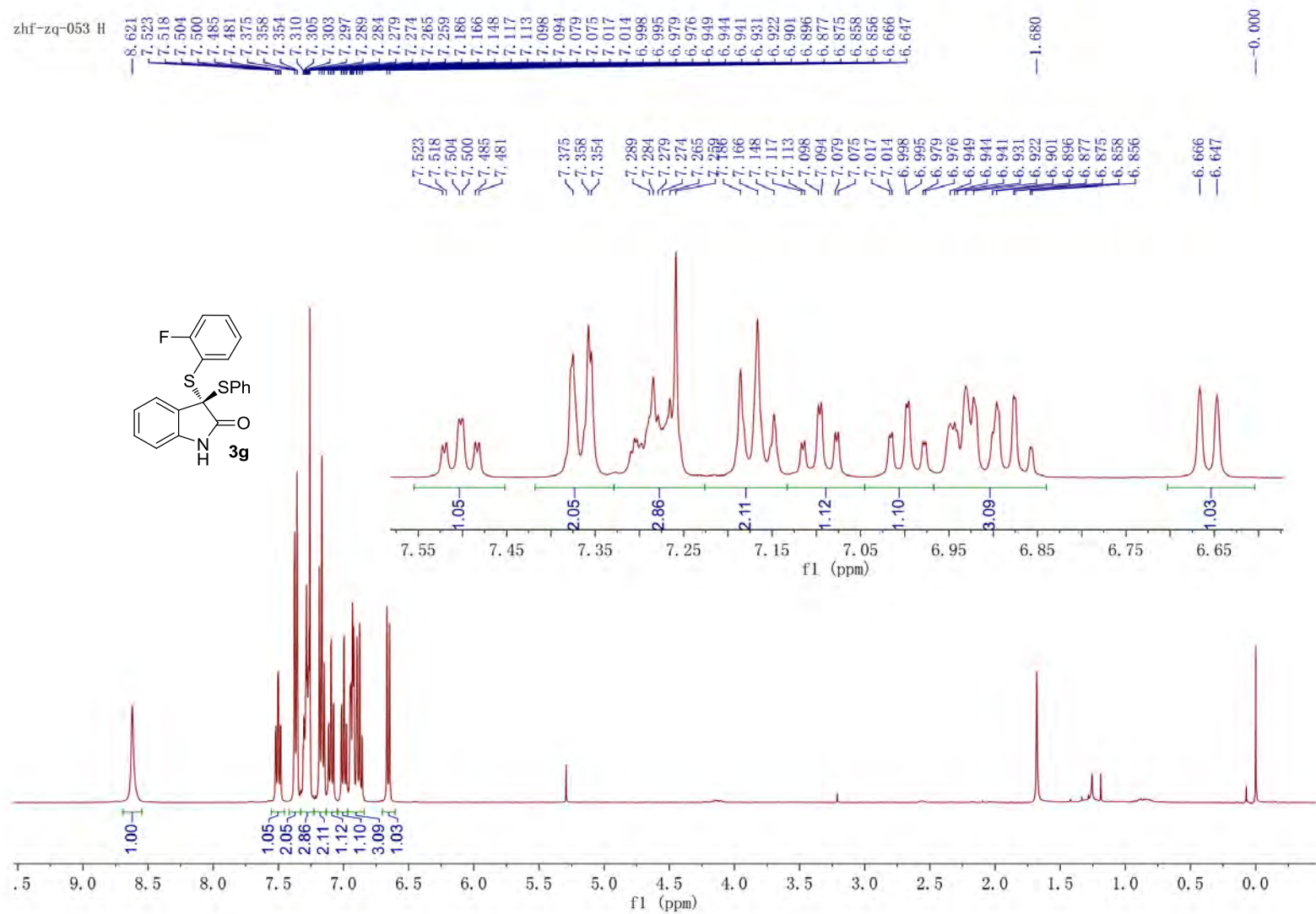
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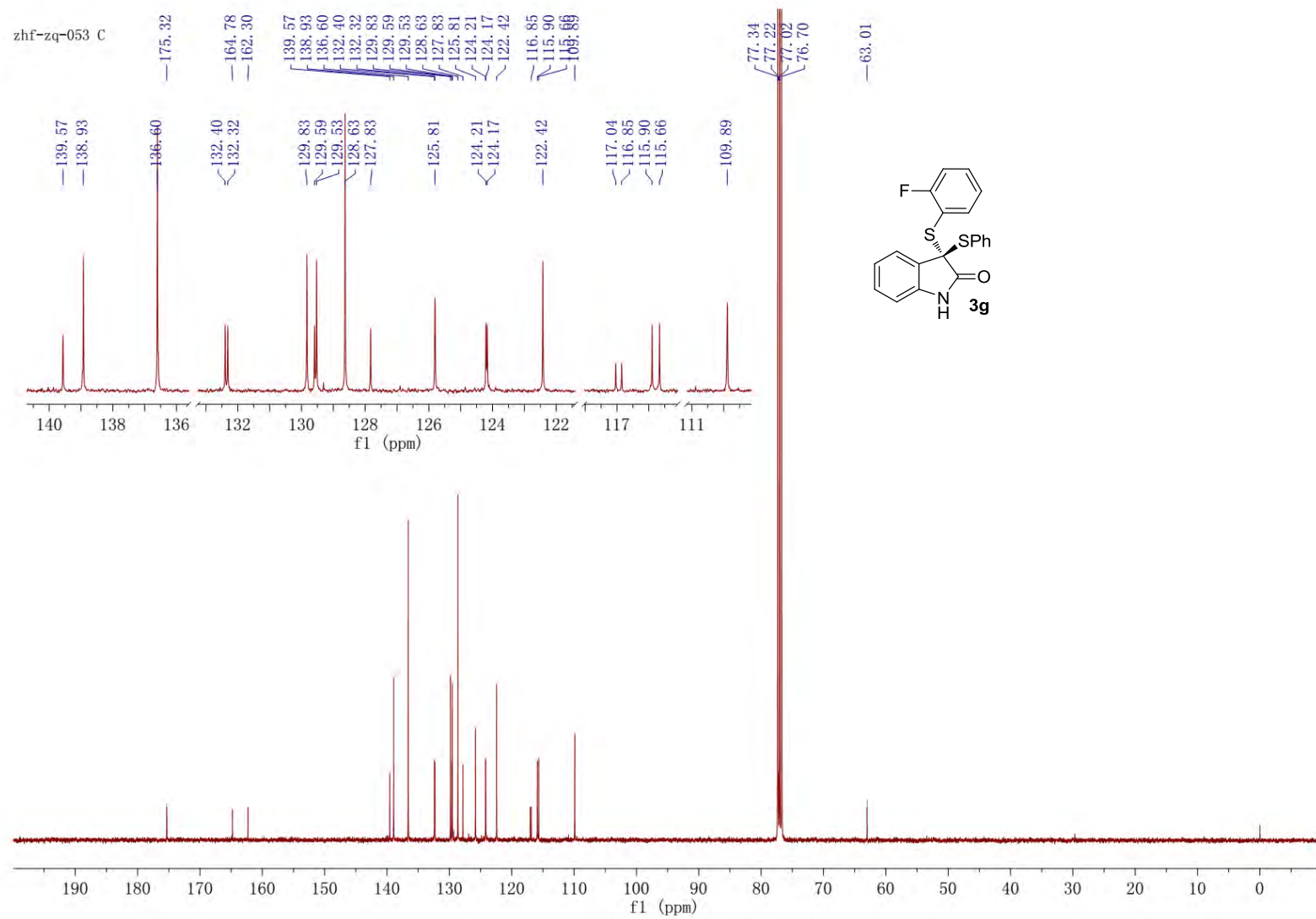


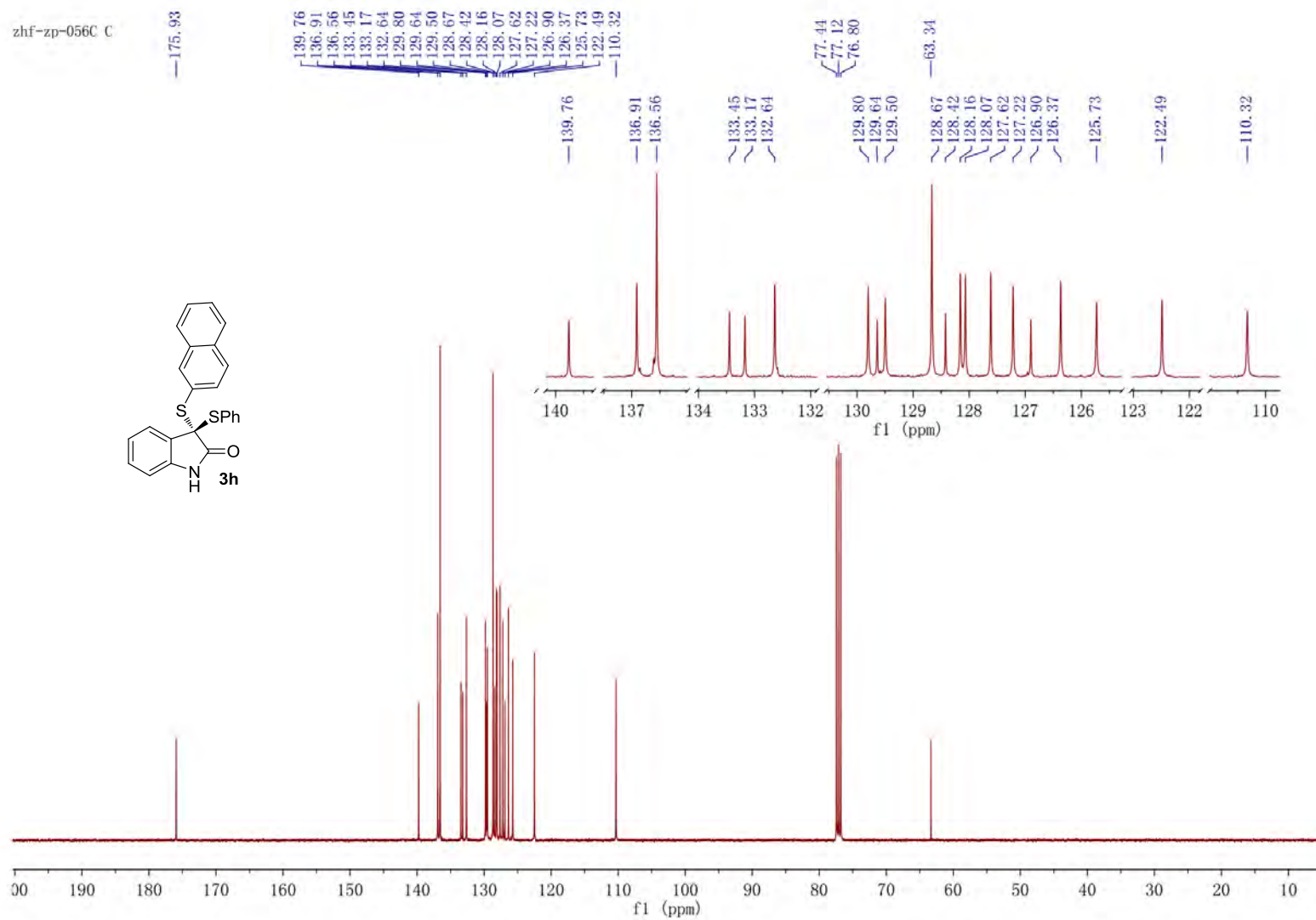


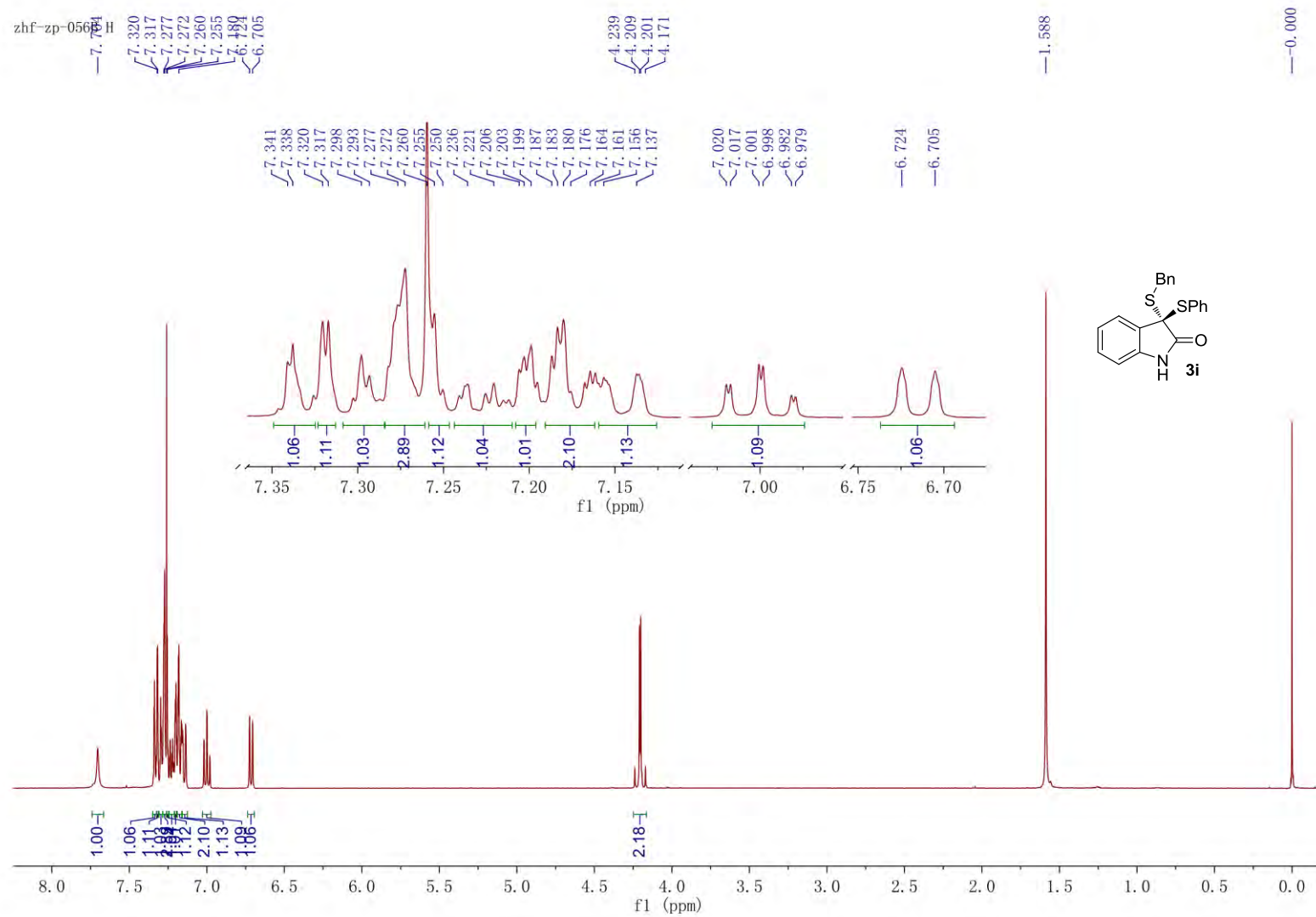
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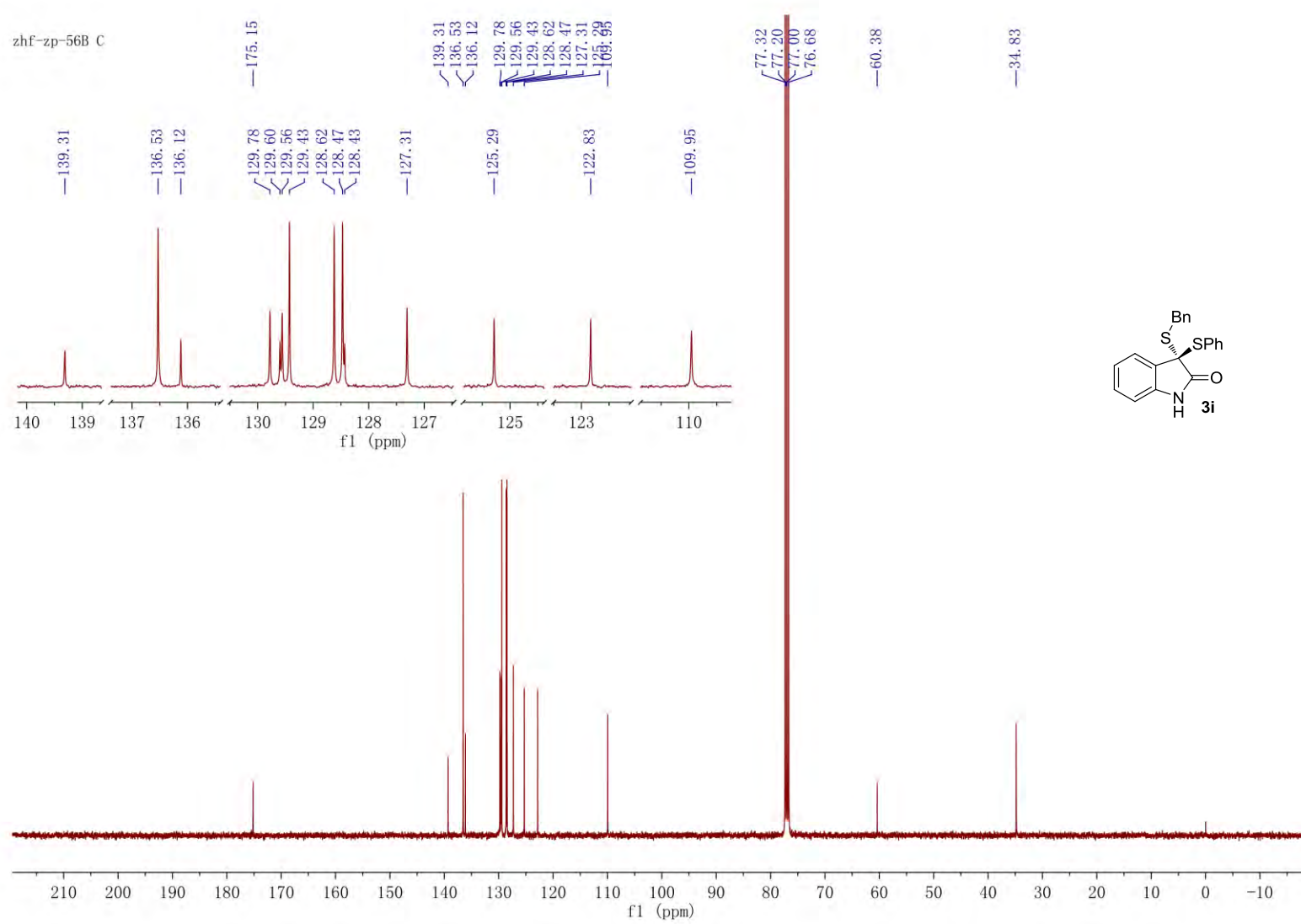


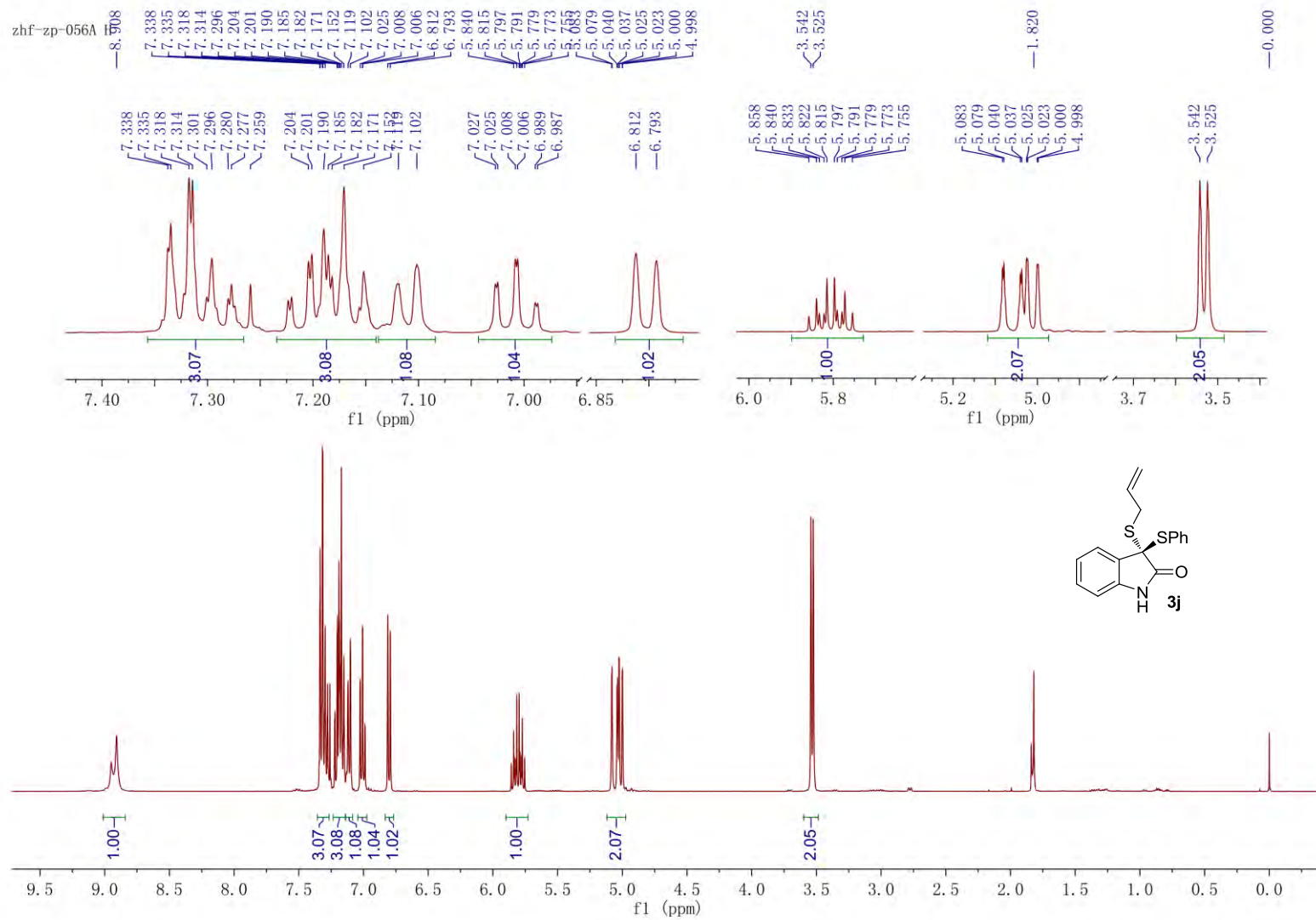




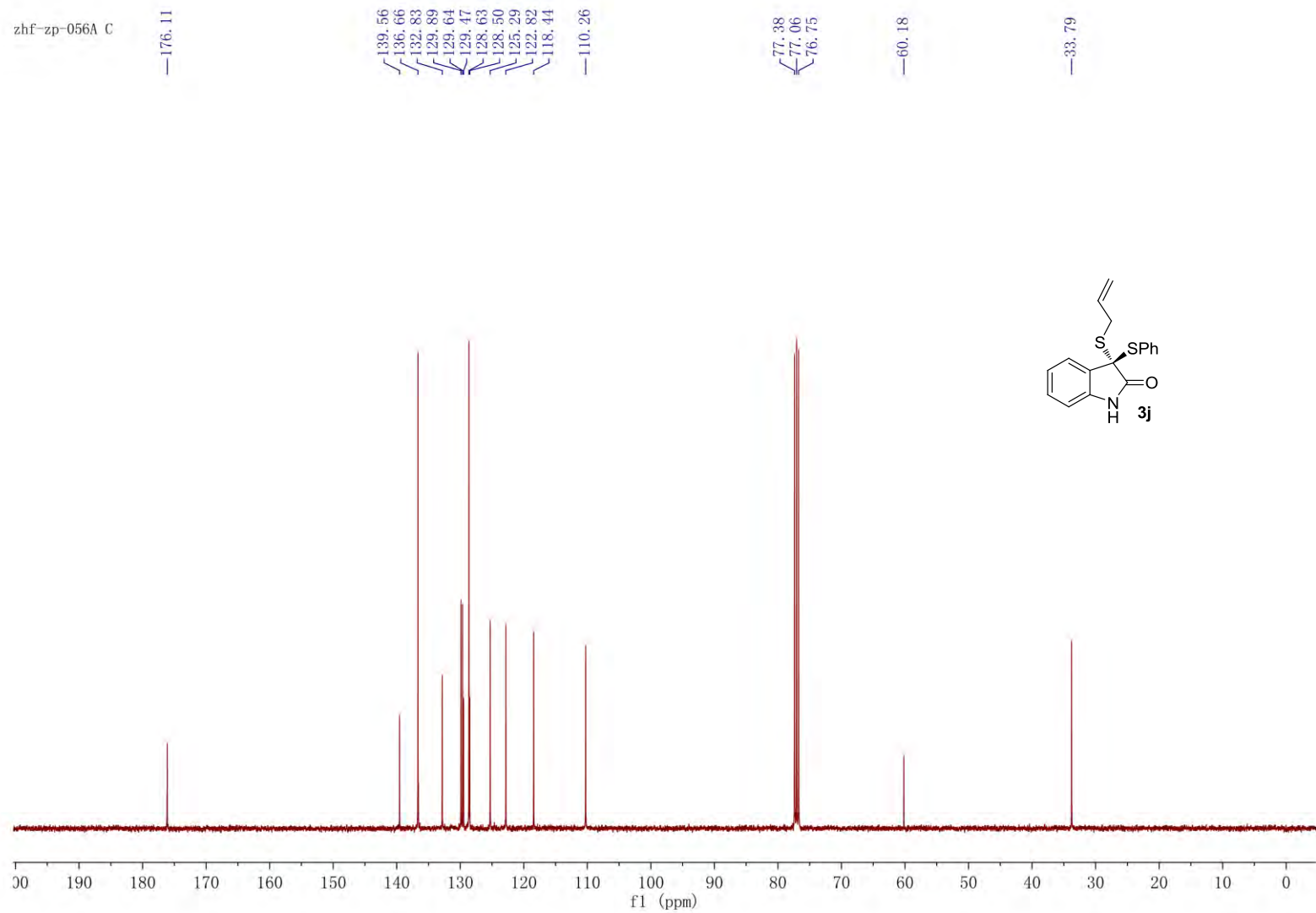


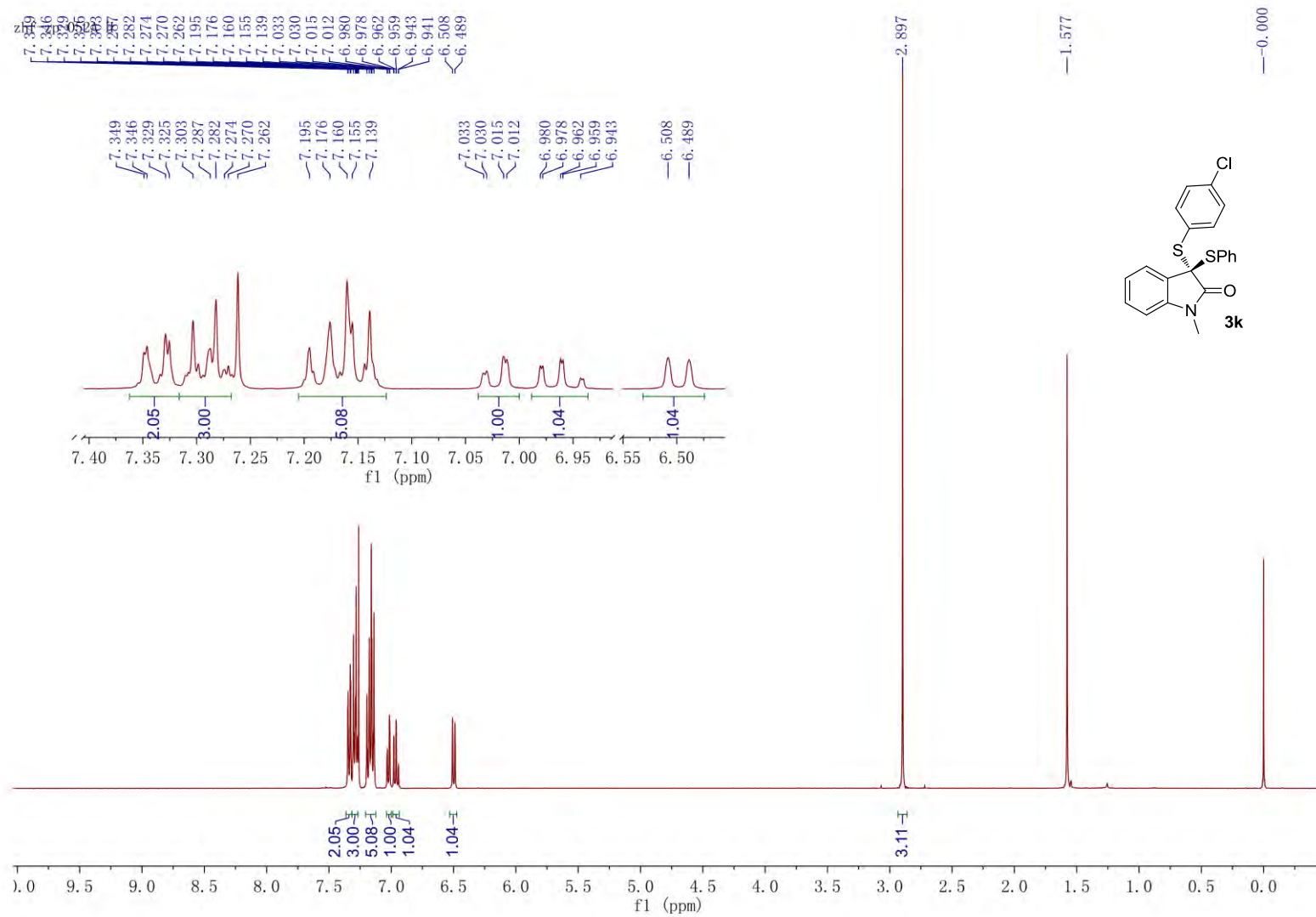




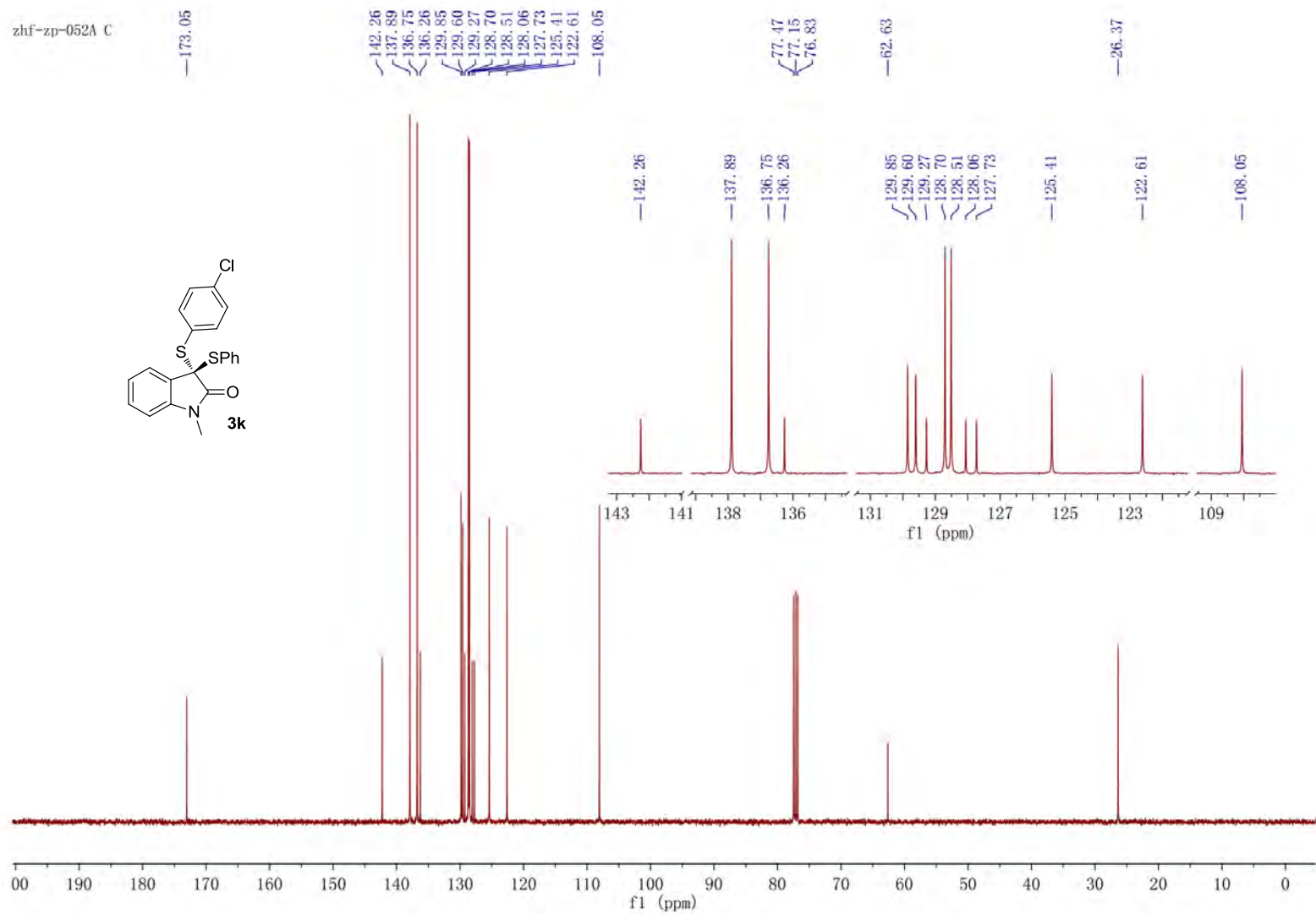


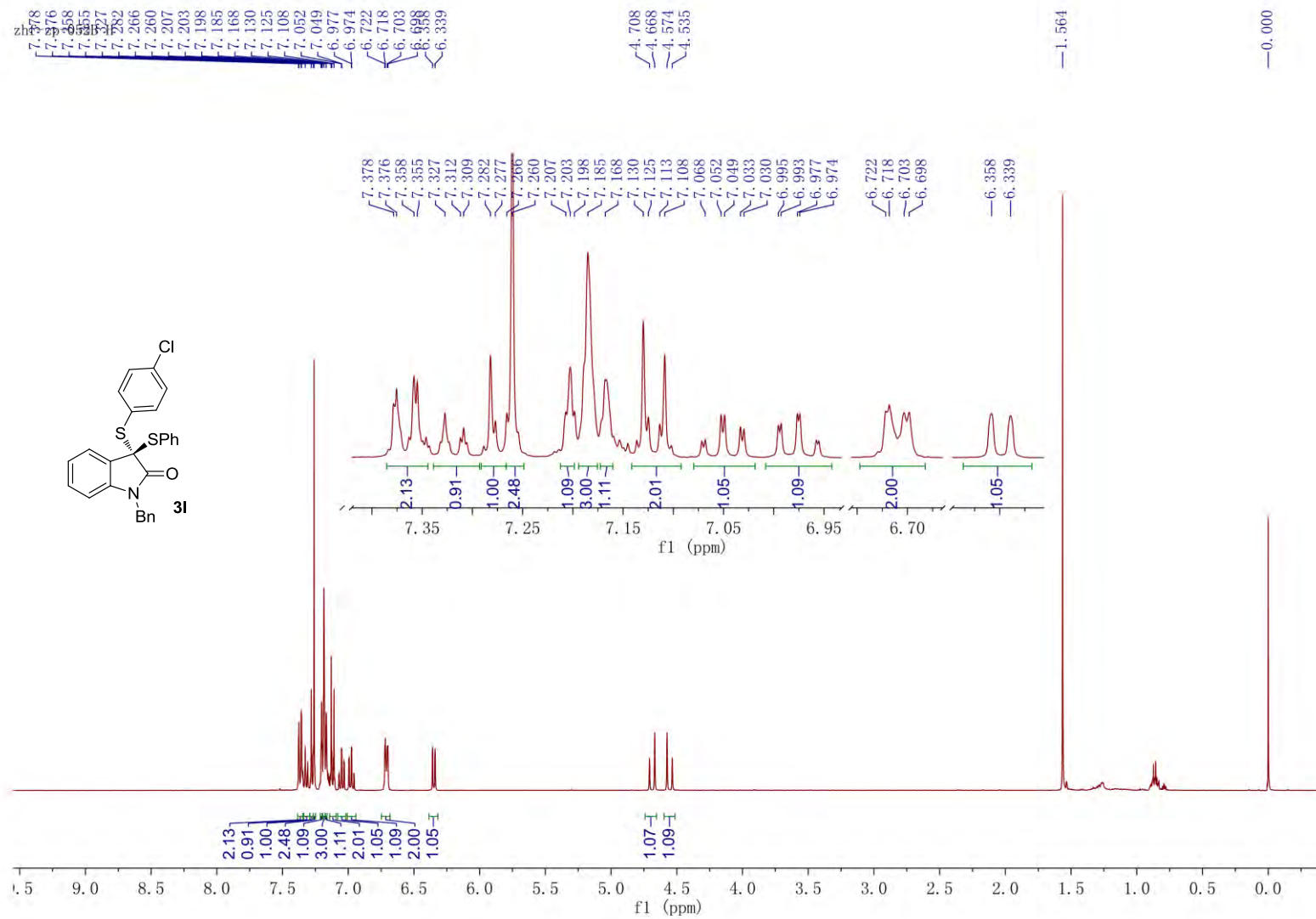
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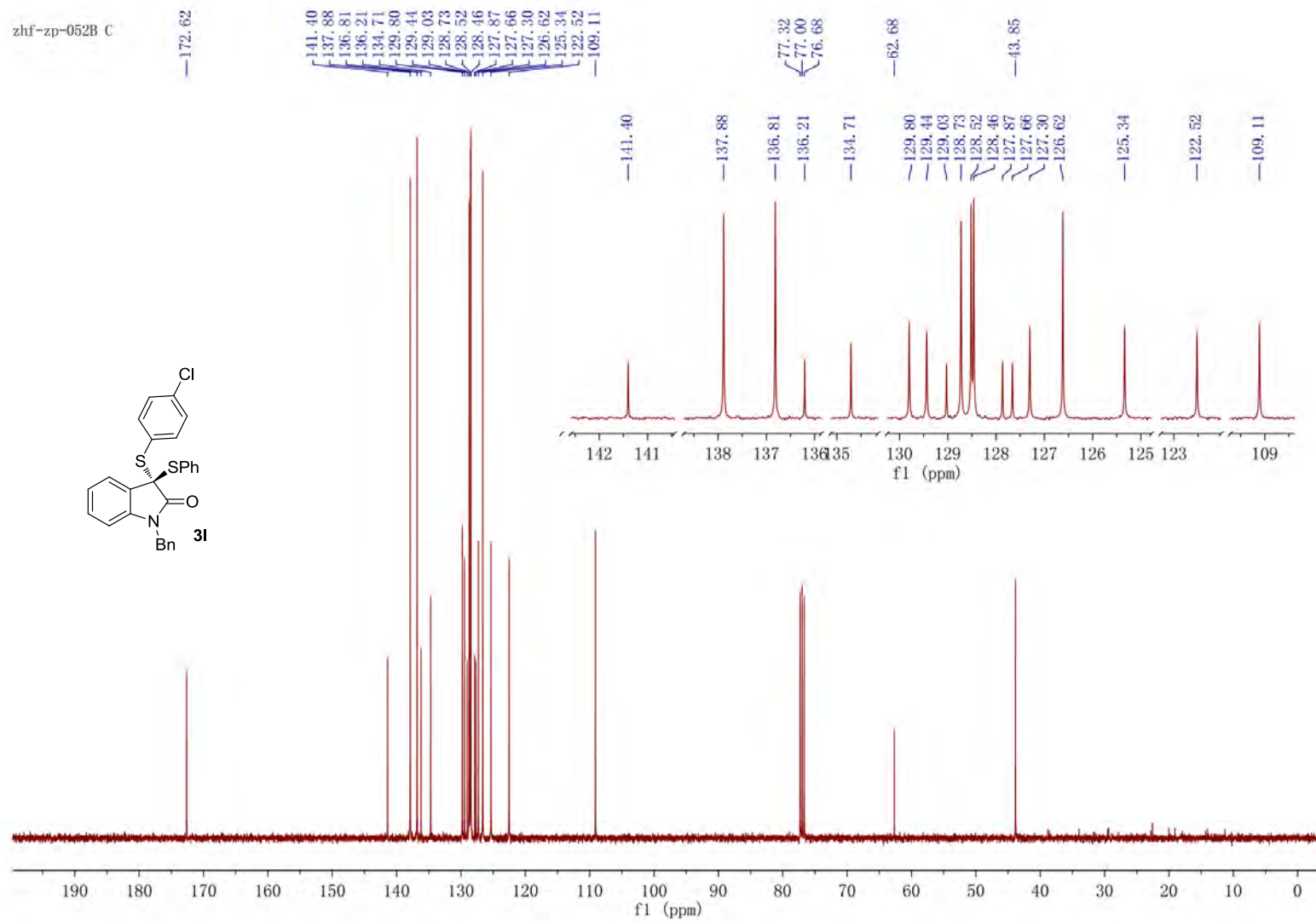


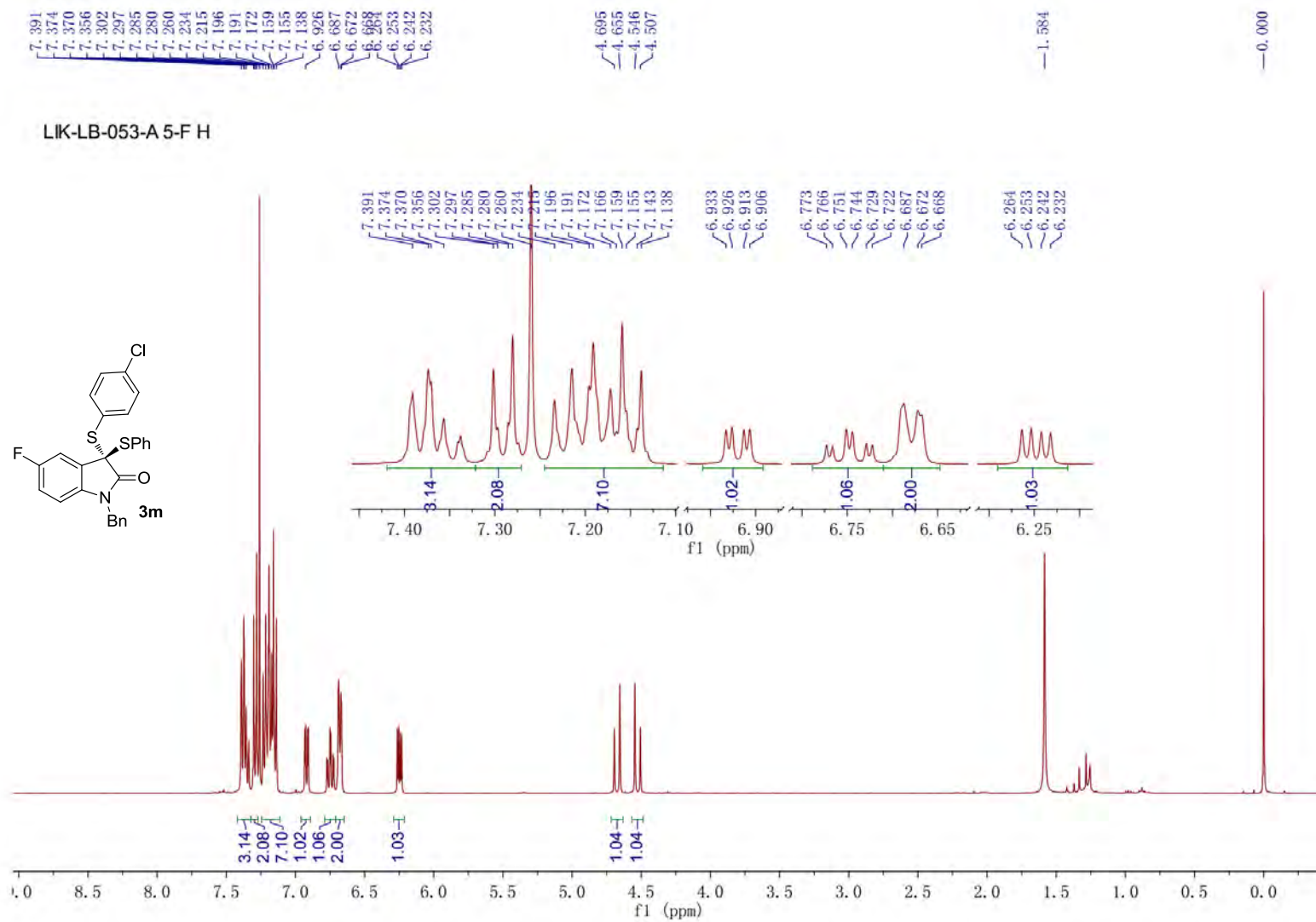
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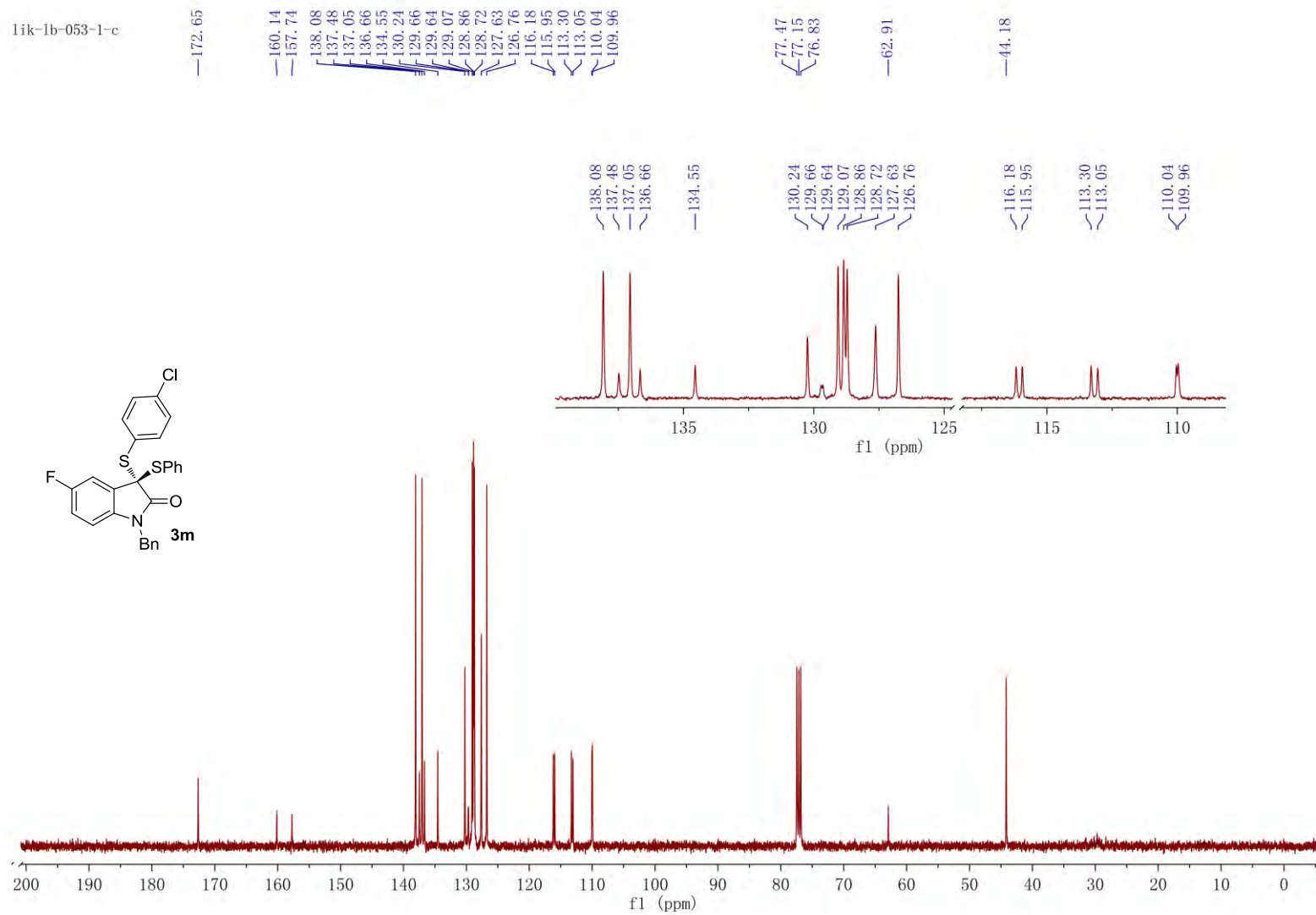


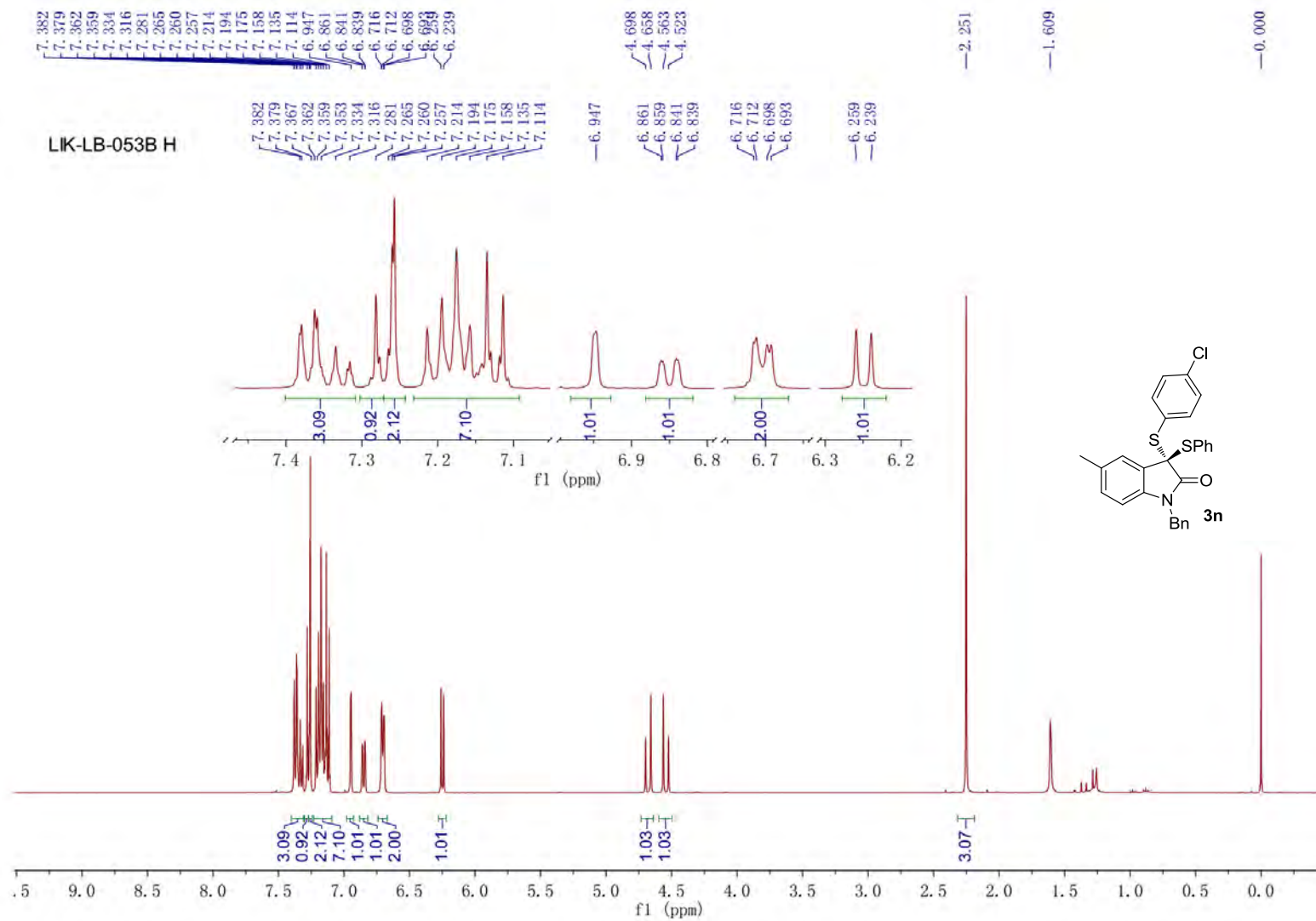
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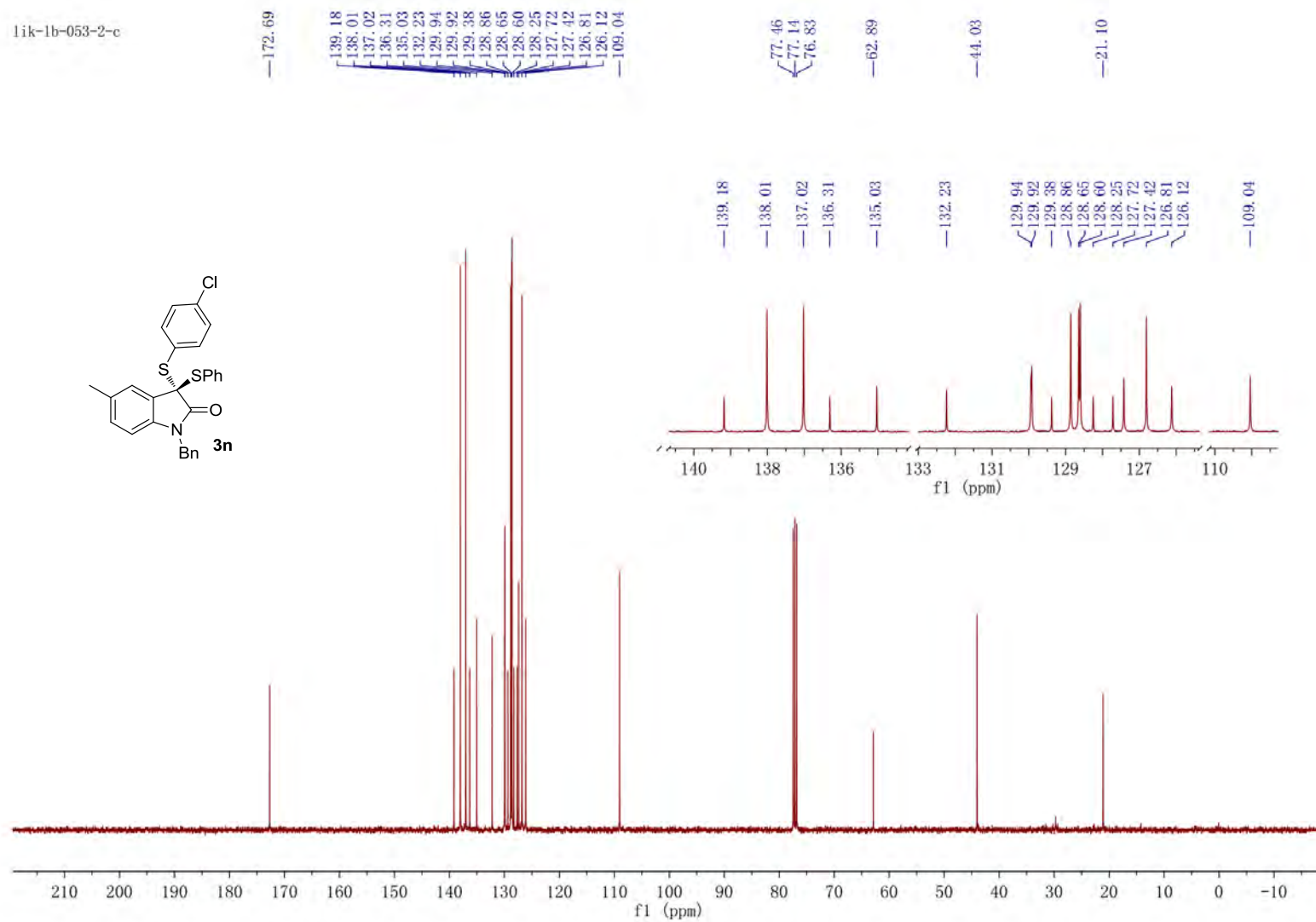


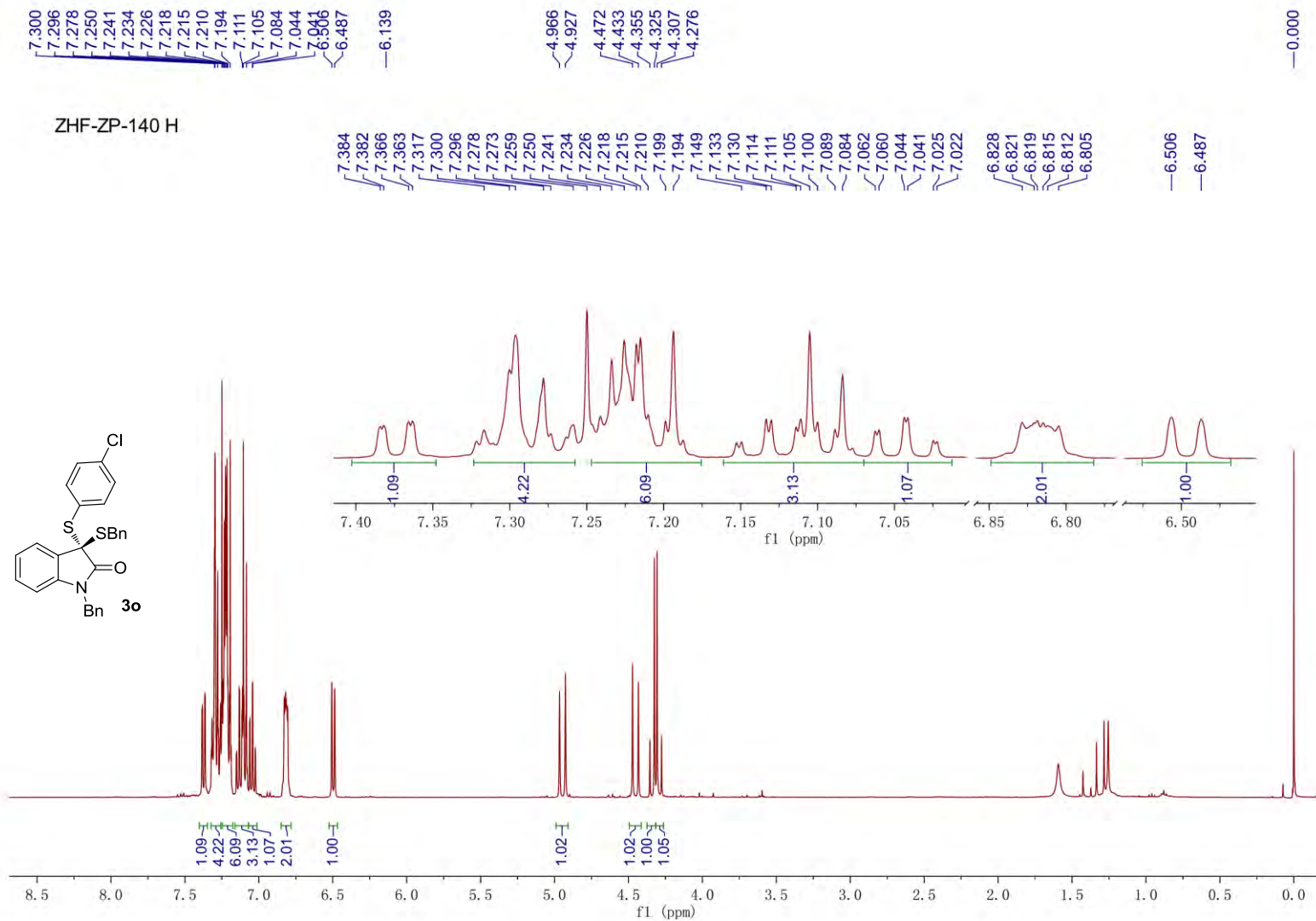
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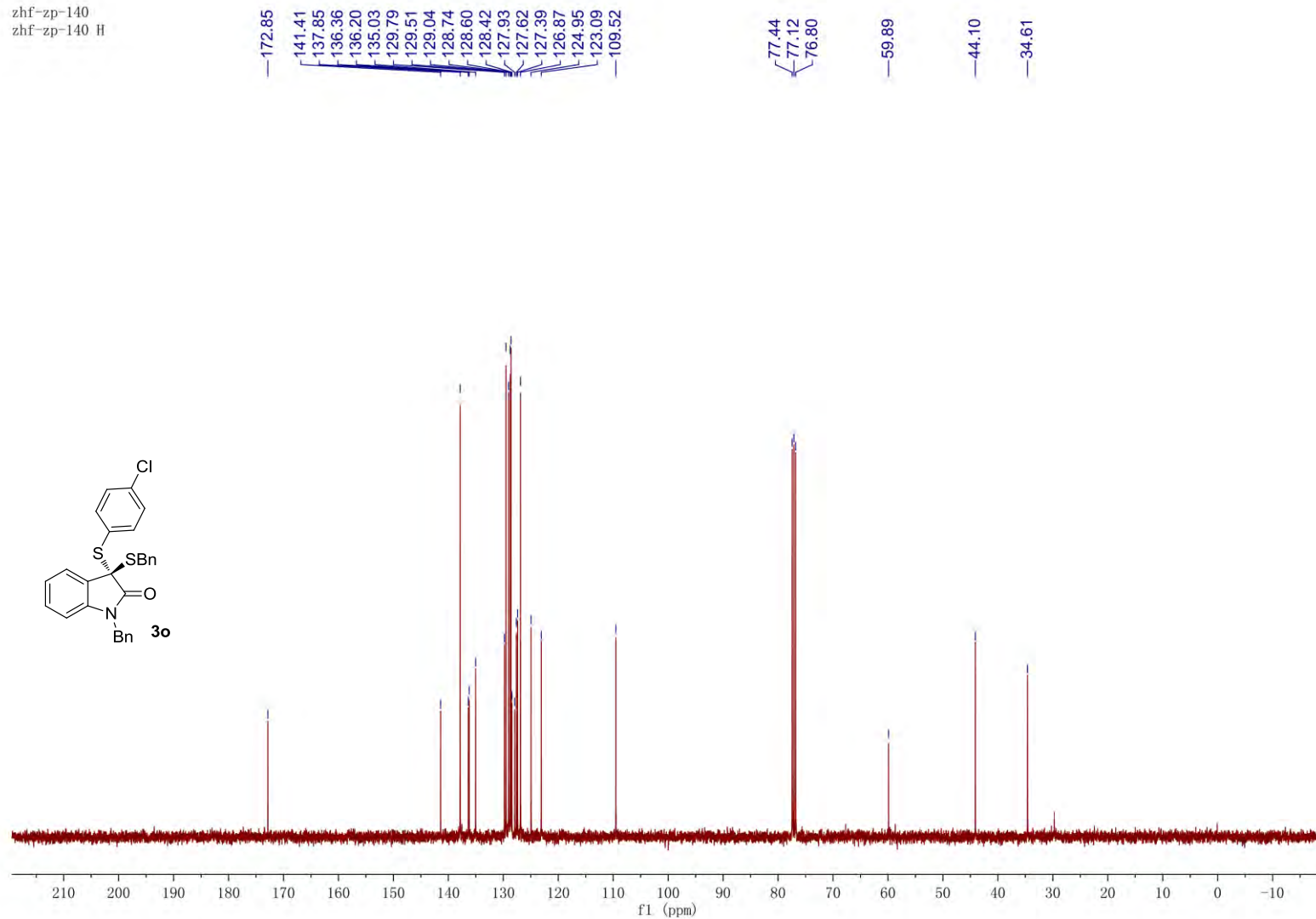


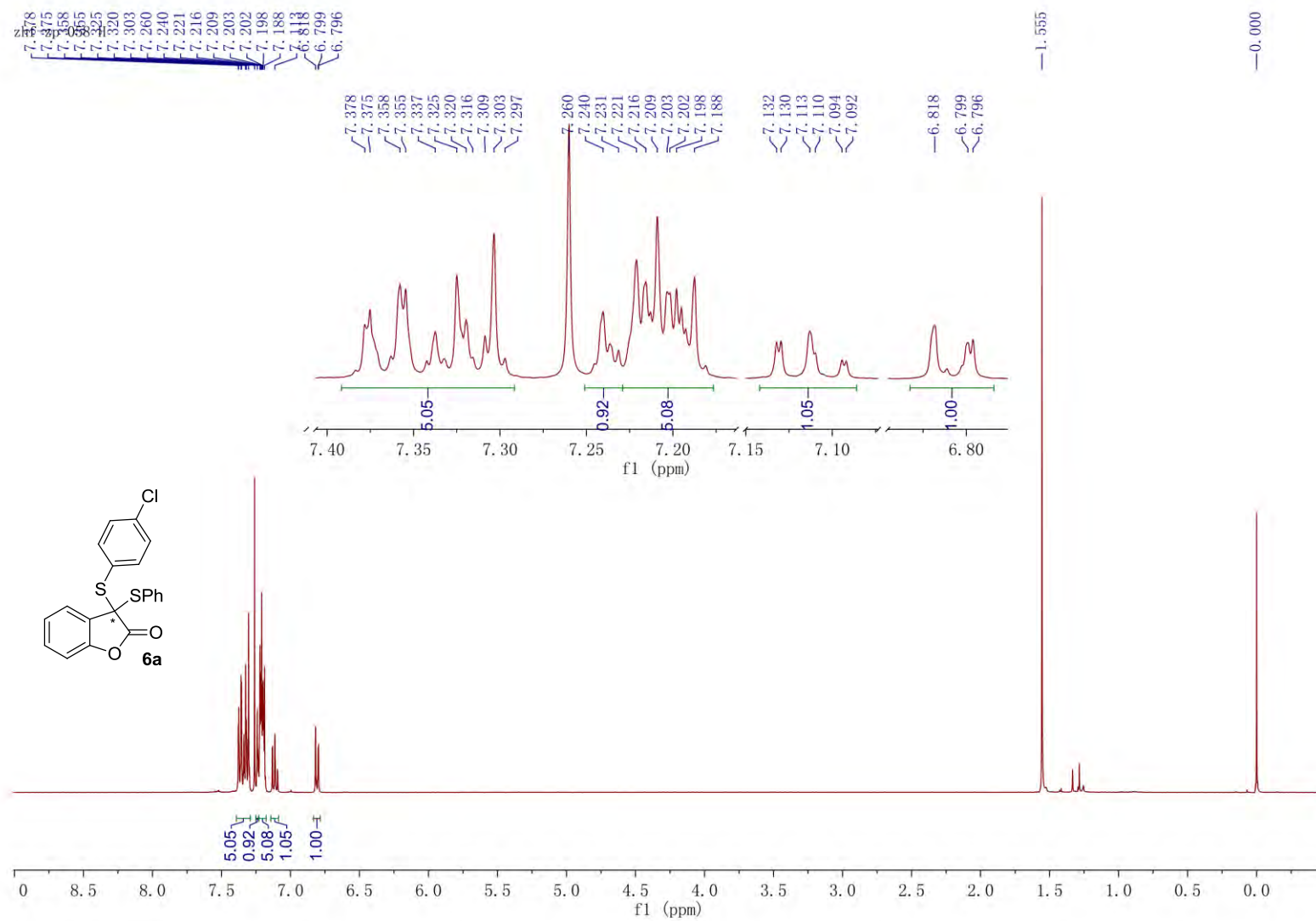
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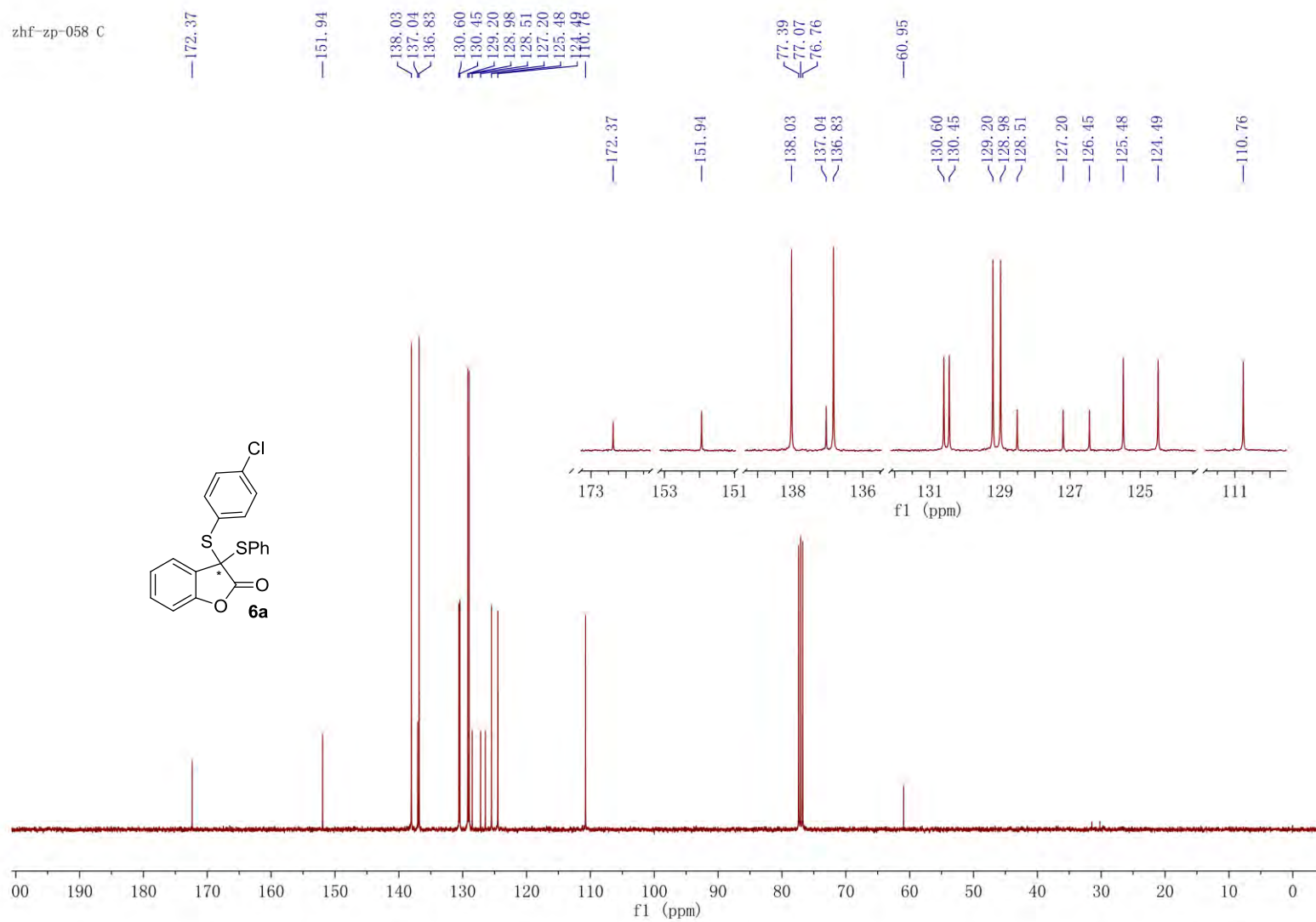


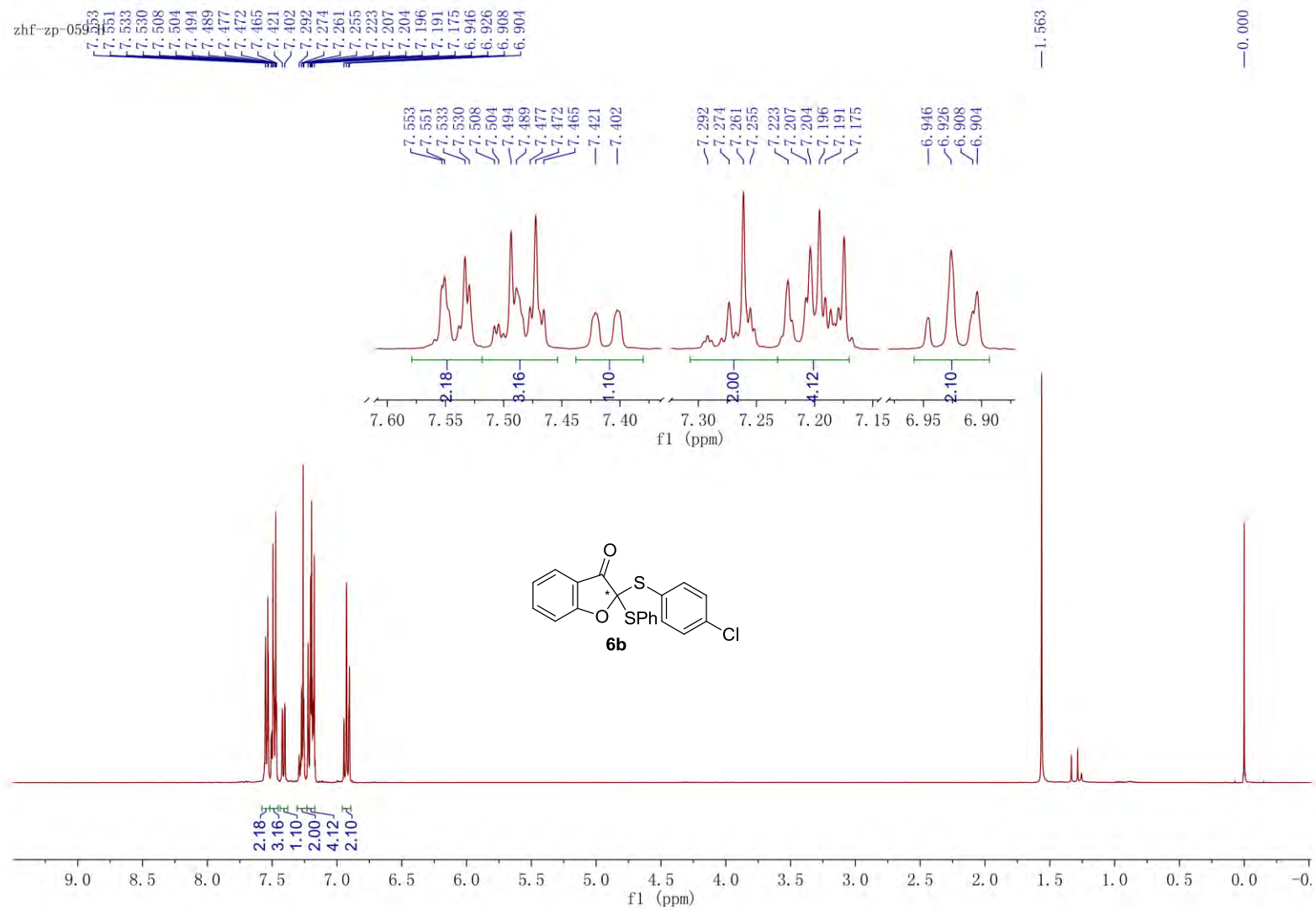
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zhf-zp-140 H



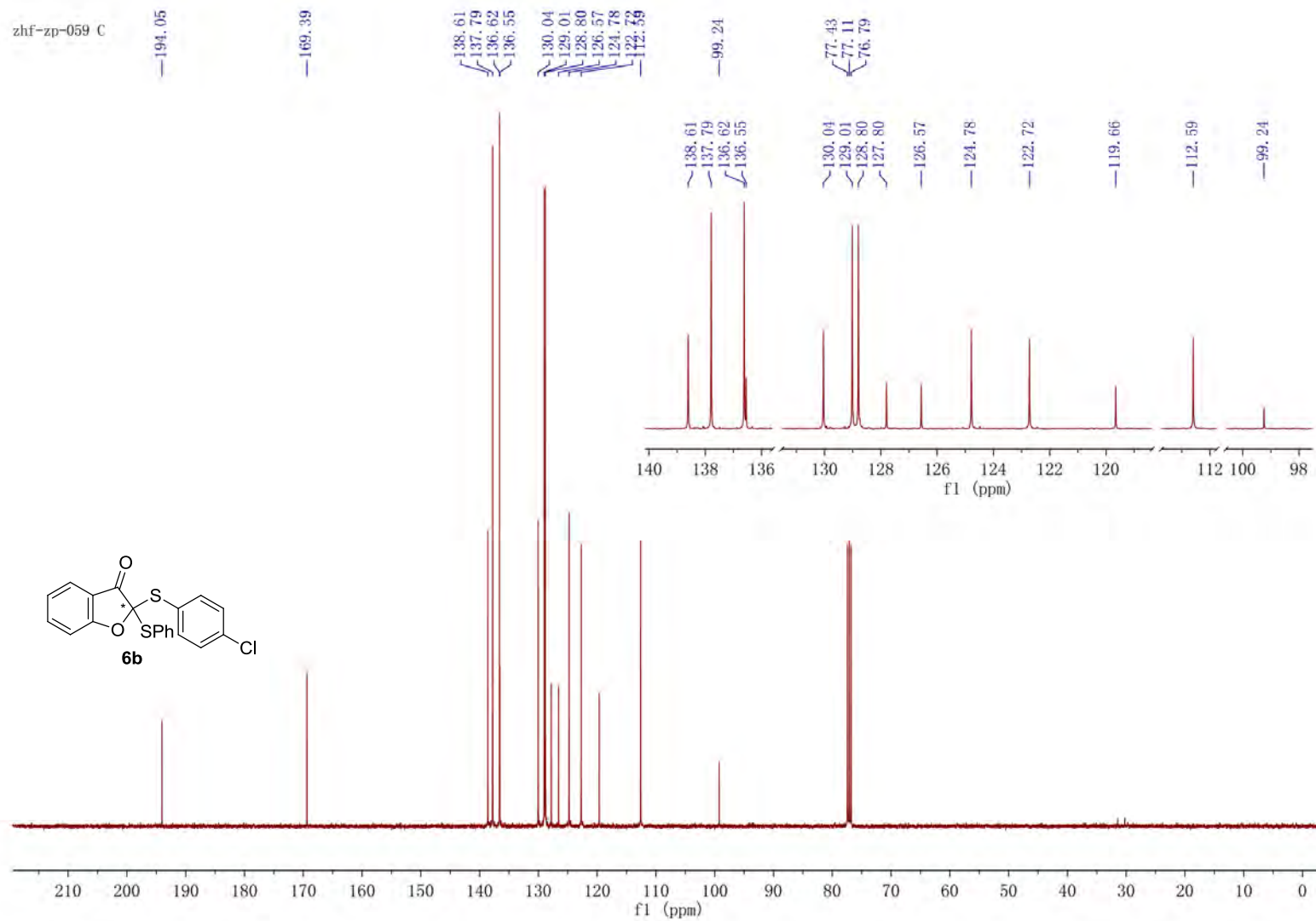


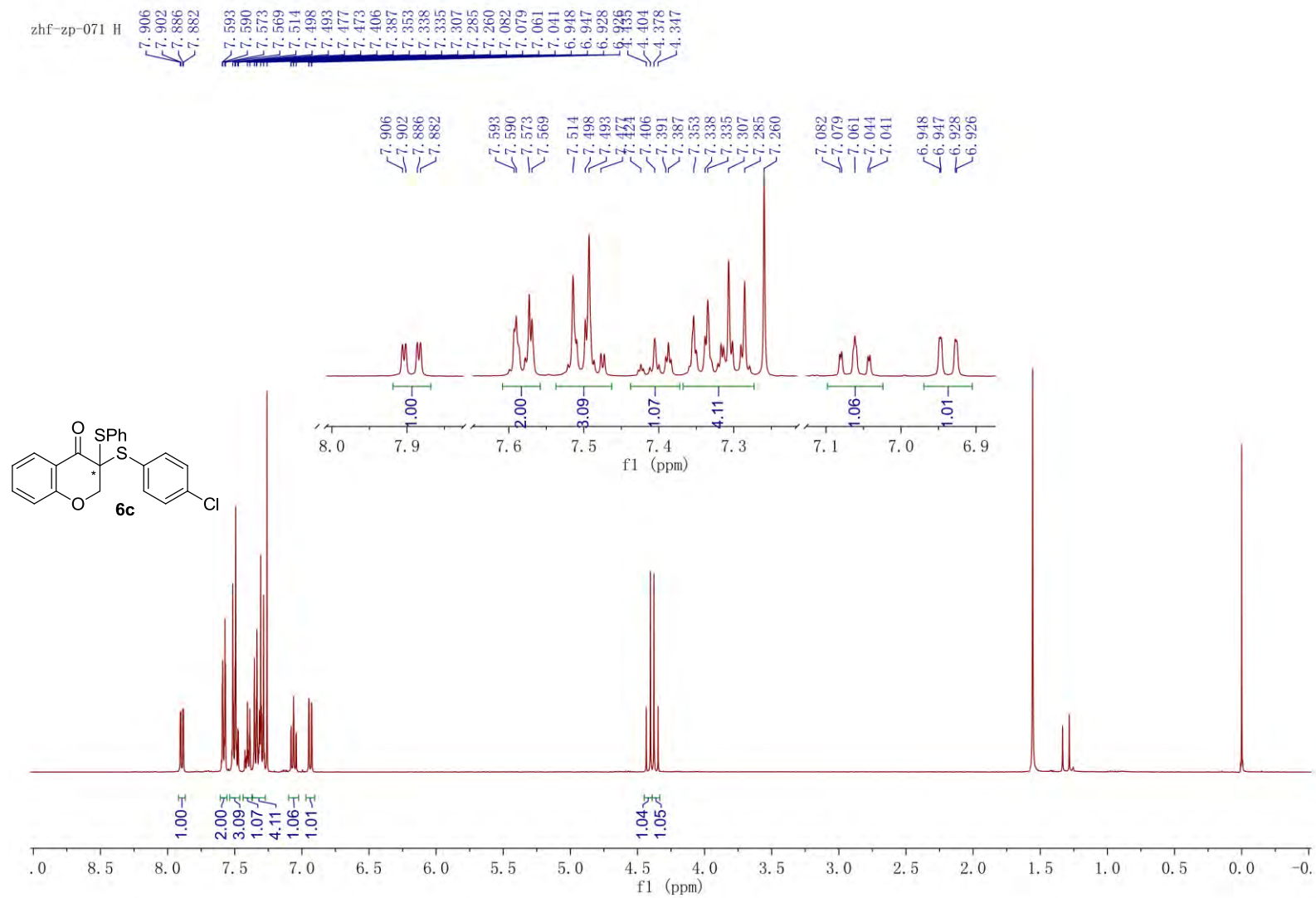
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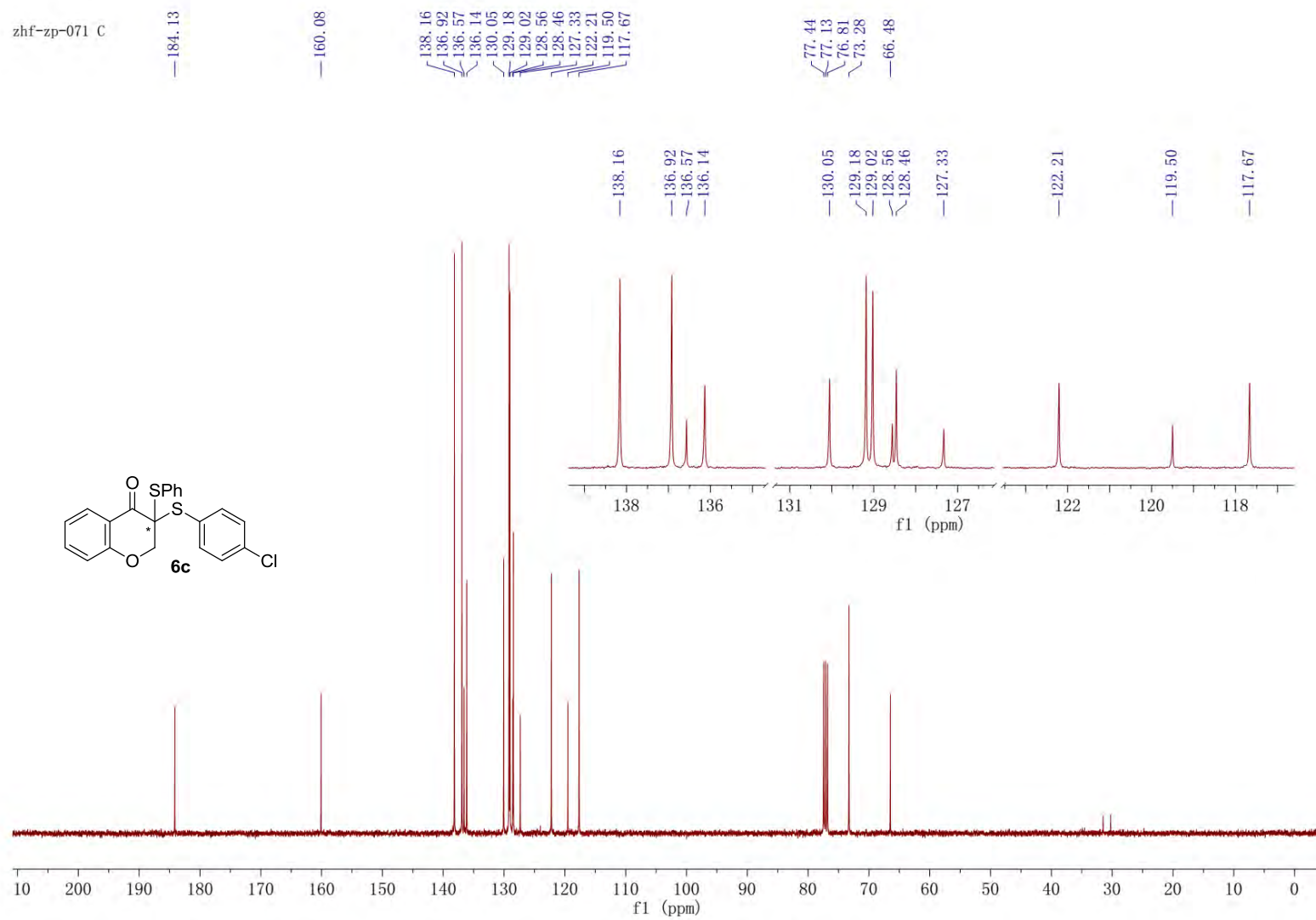


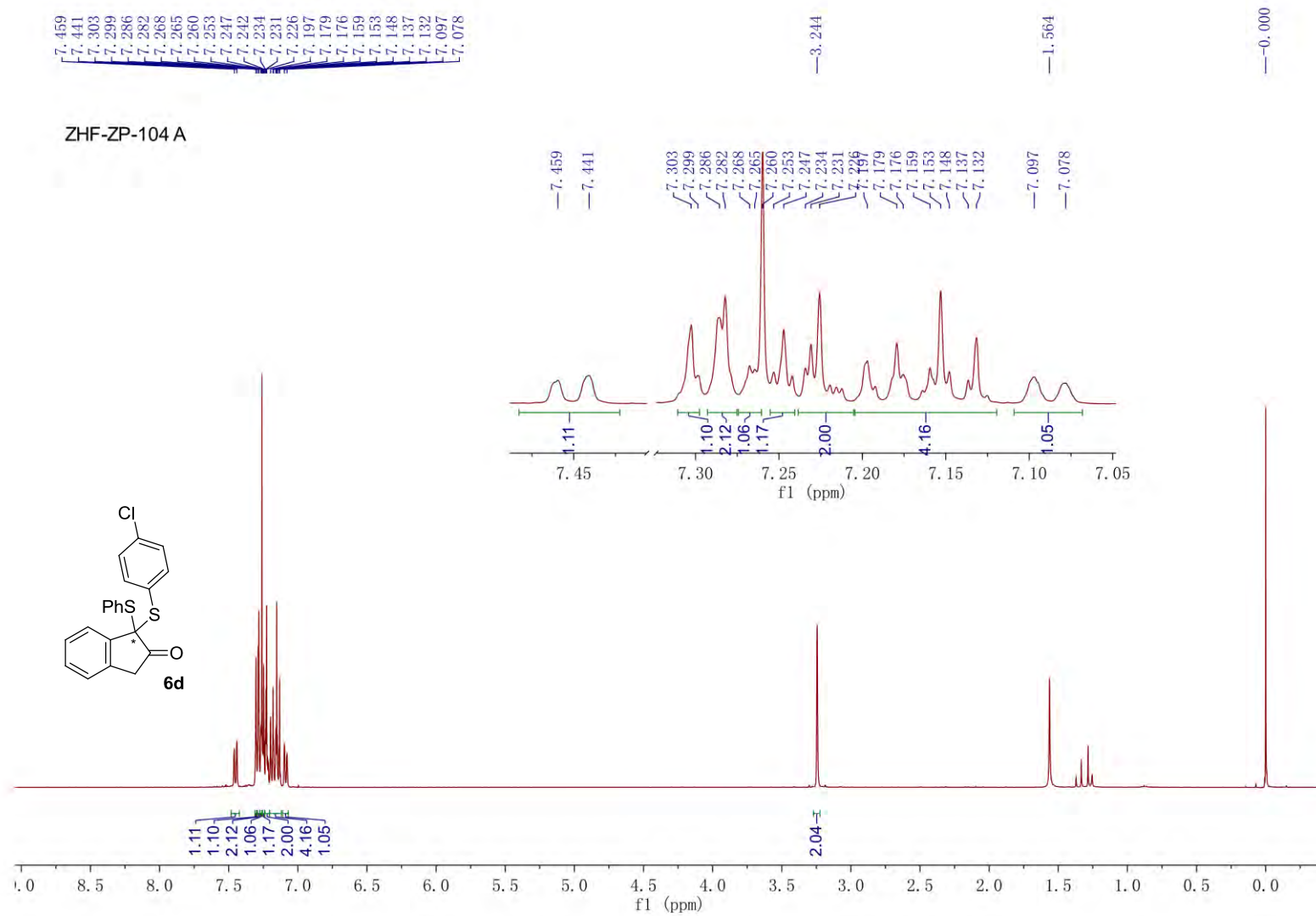
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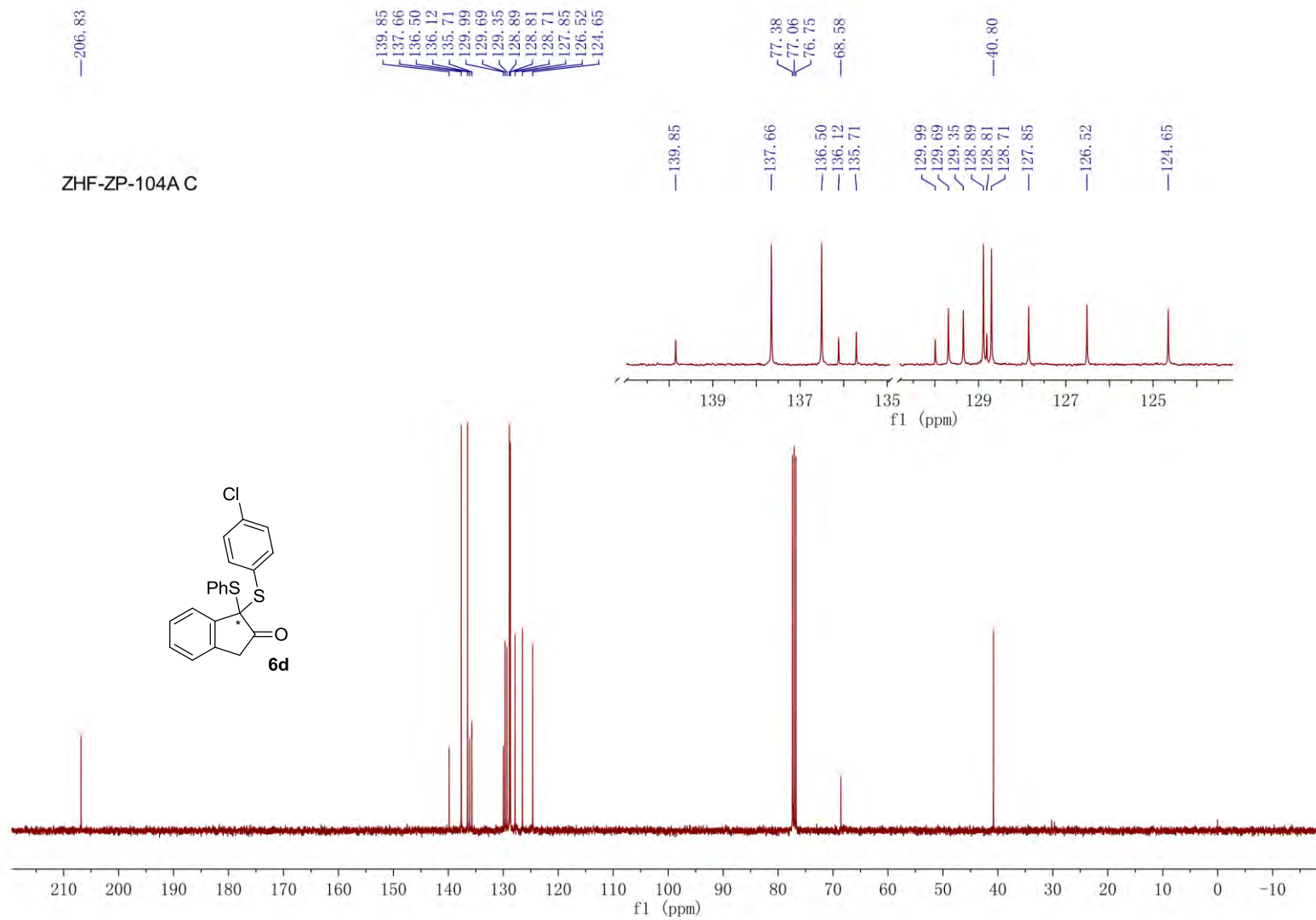


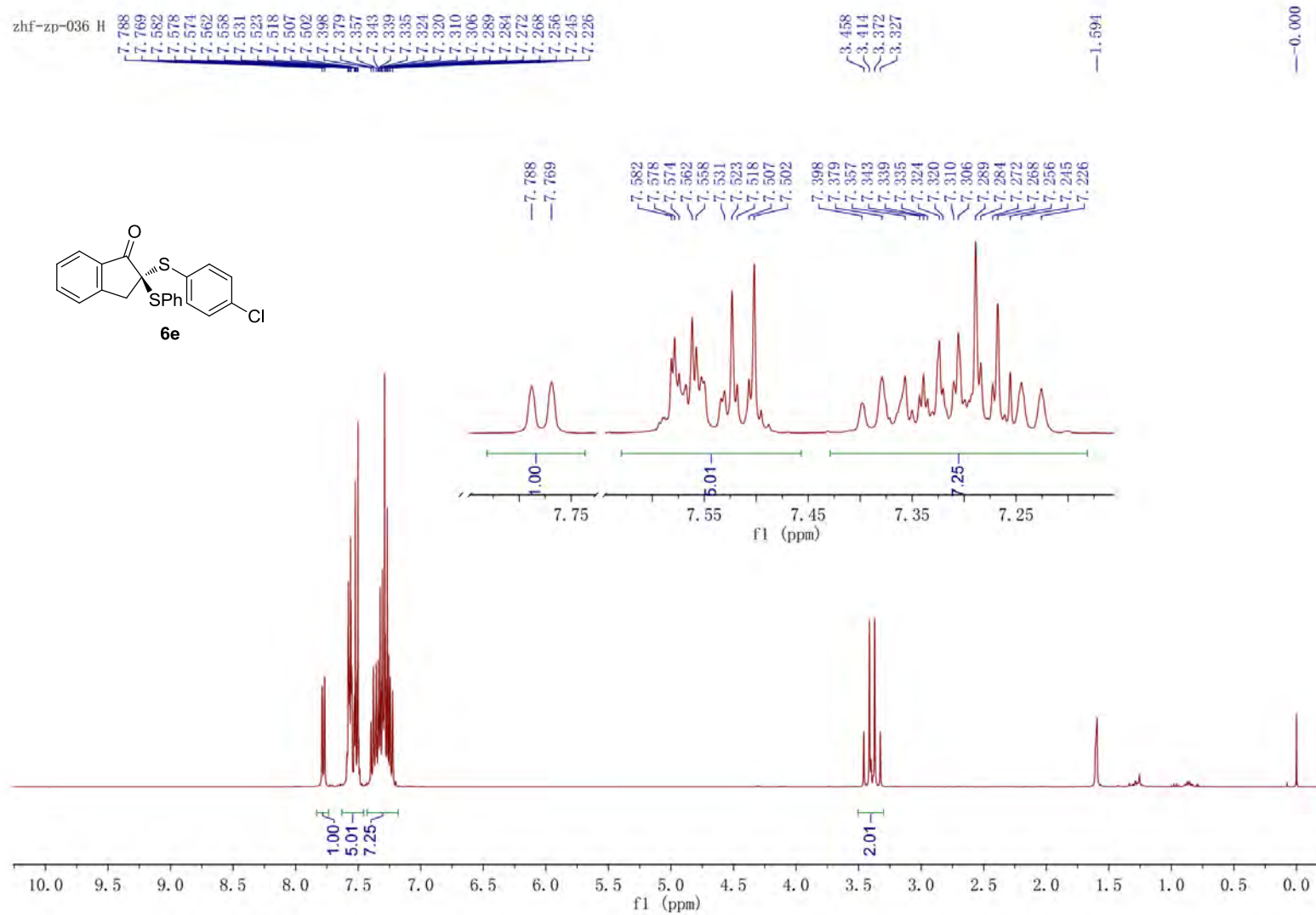


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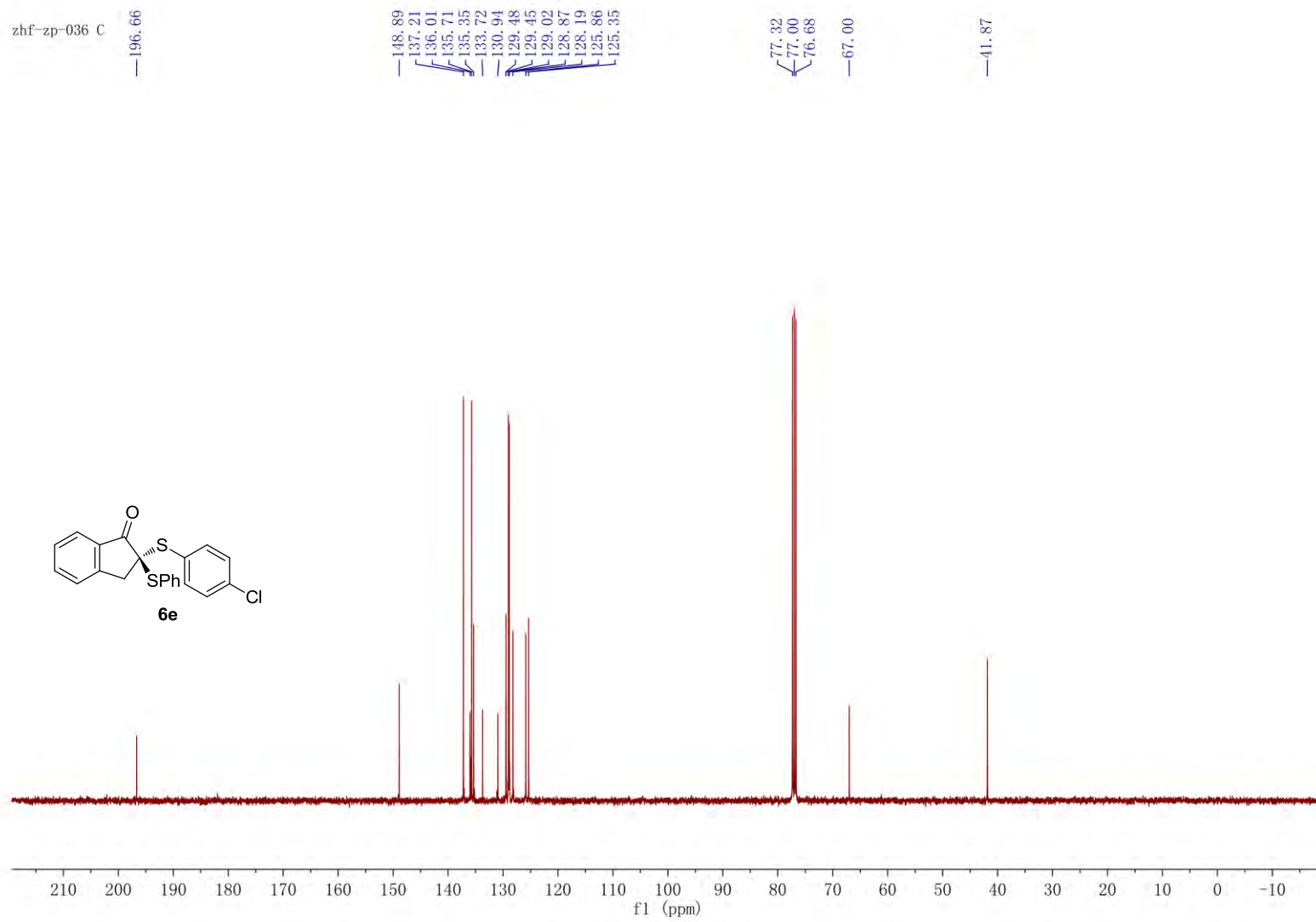


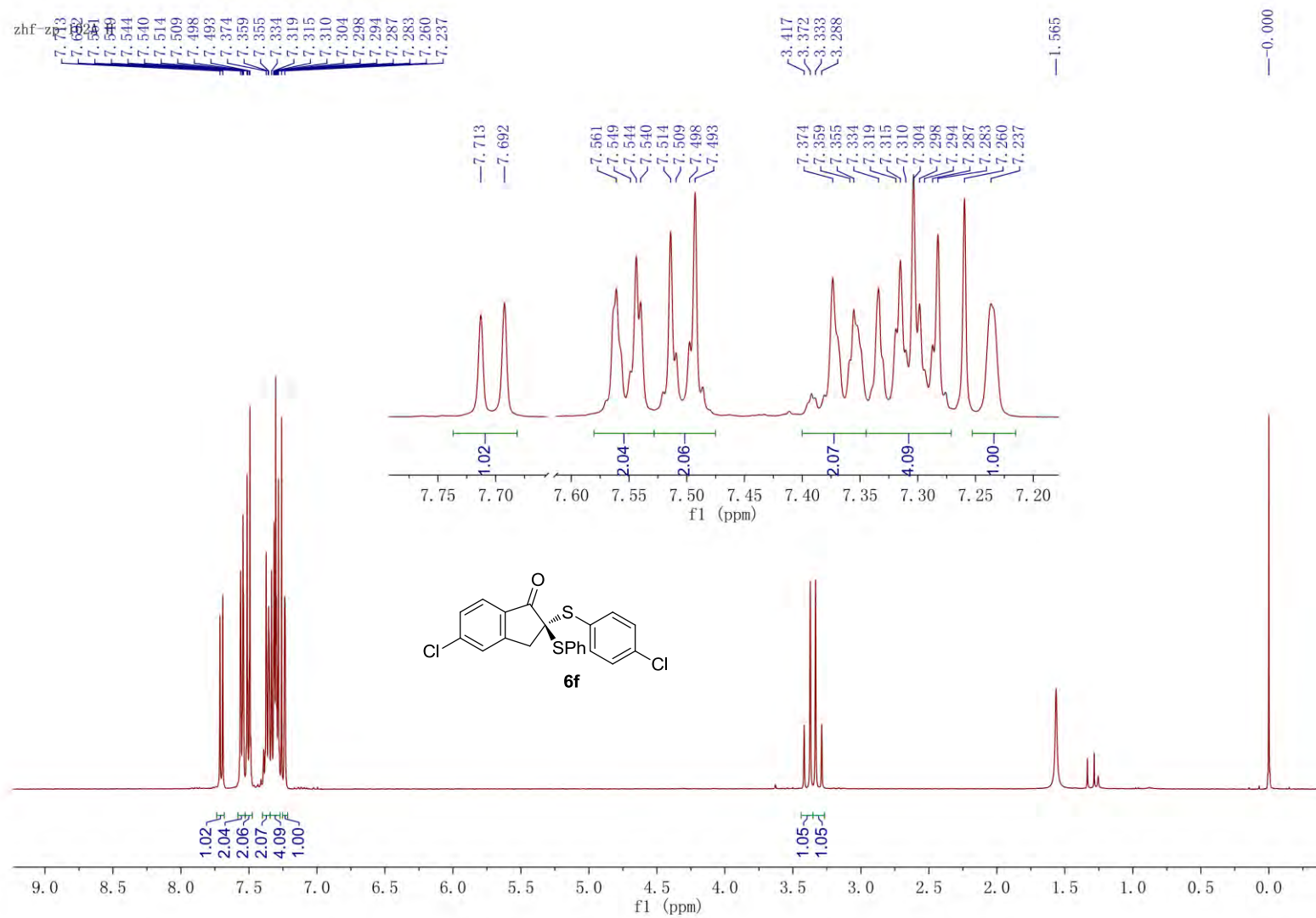


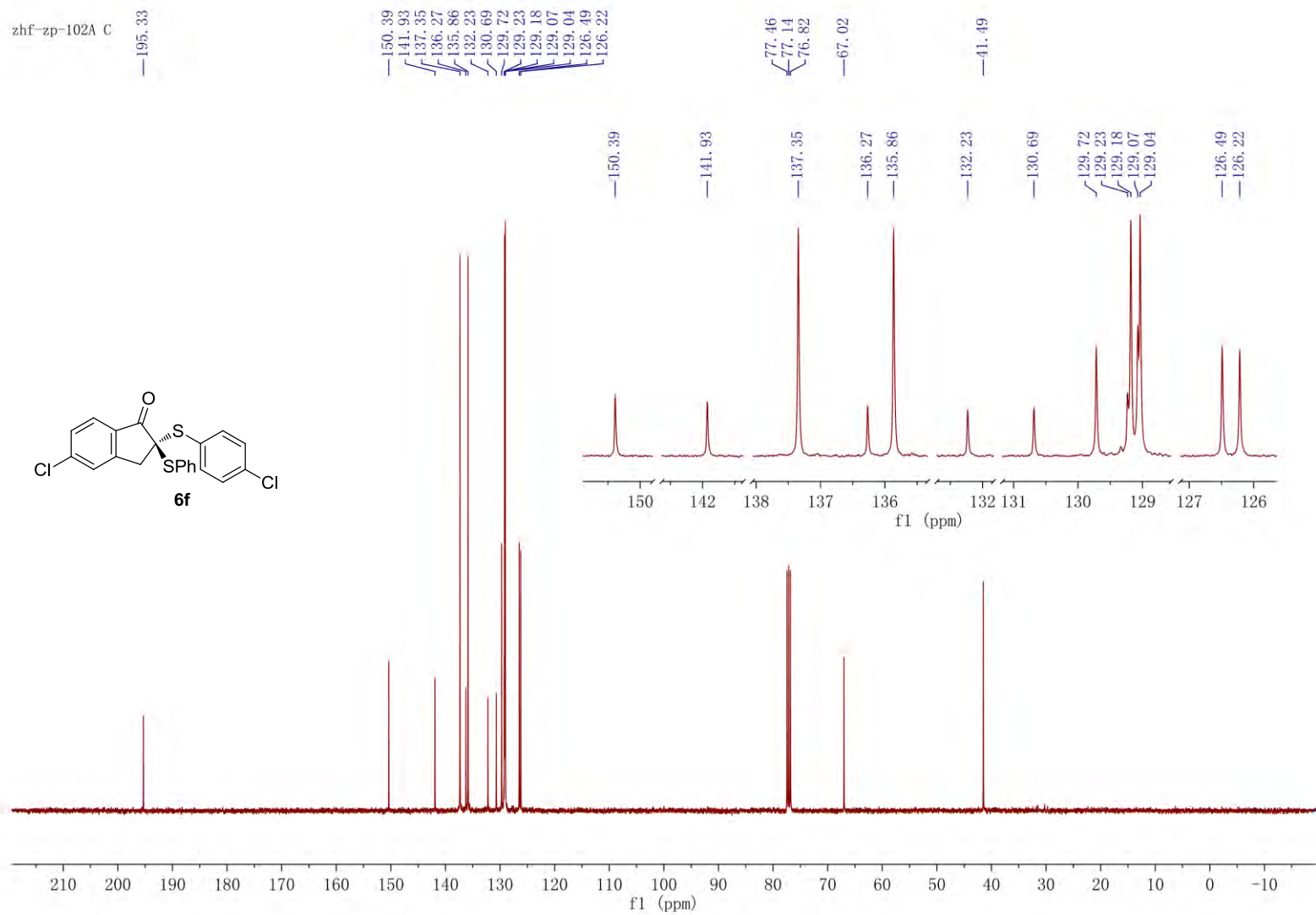


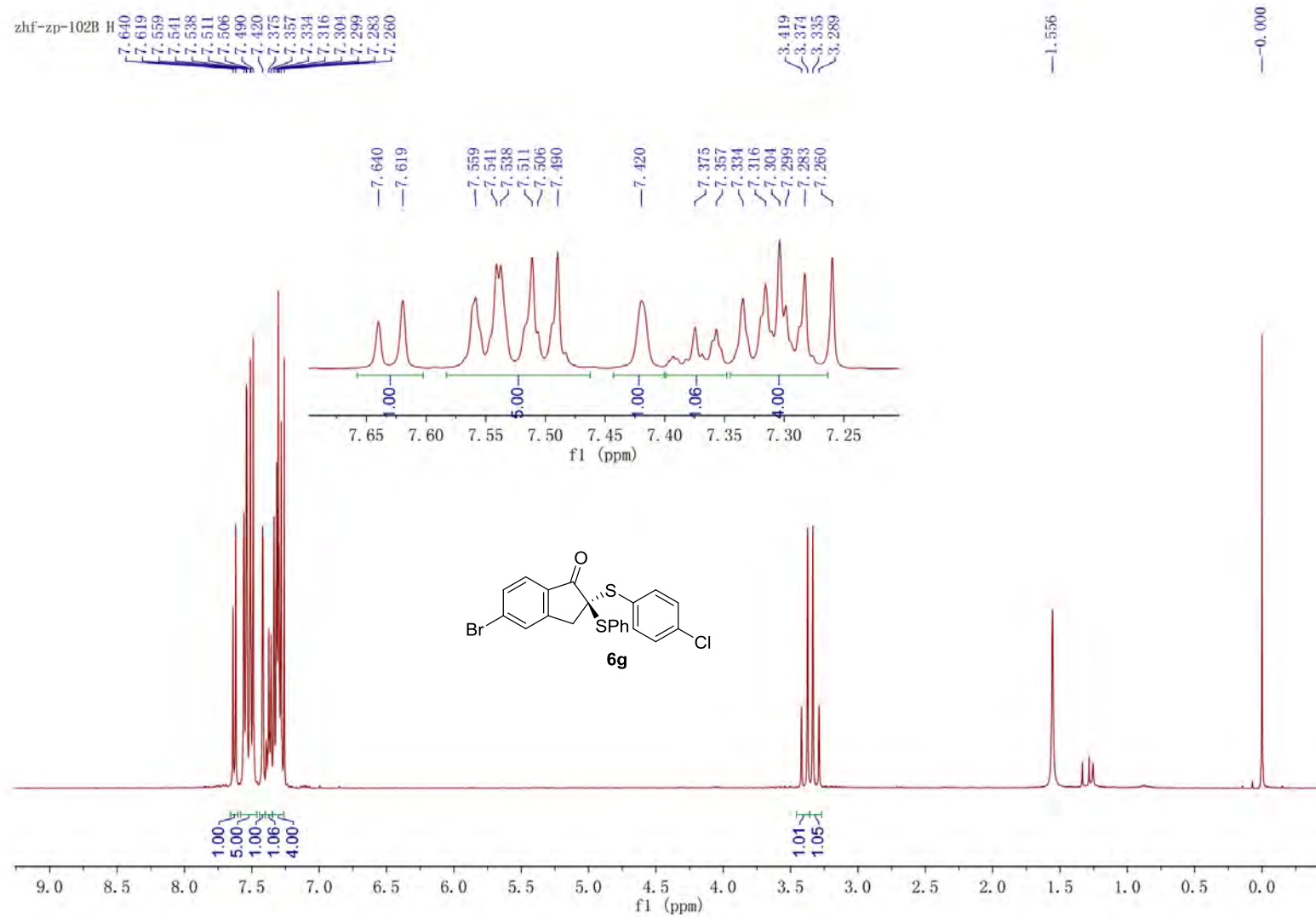


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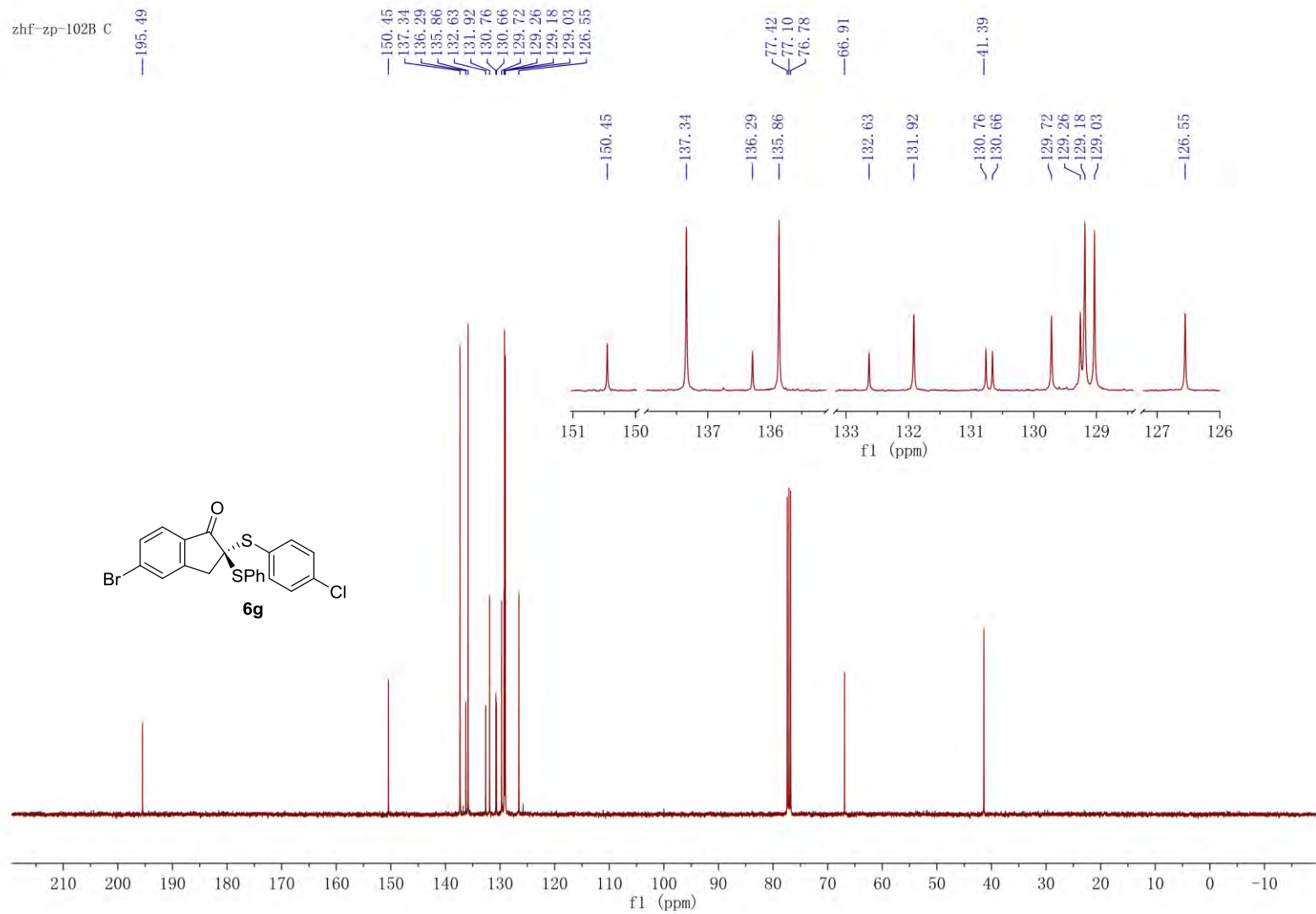


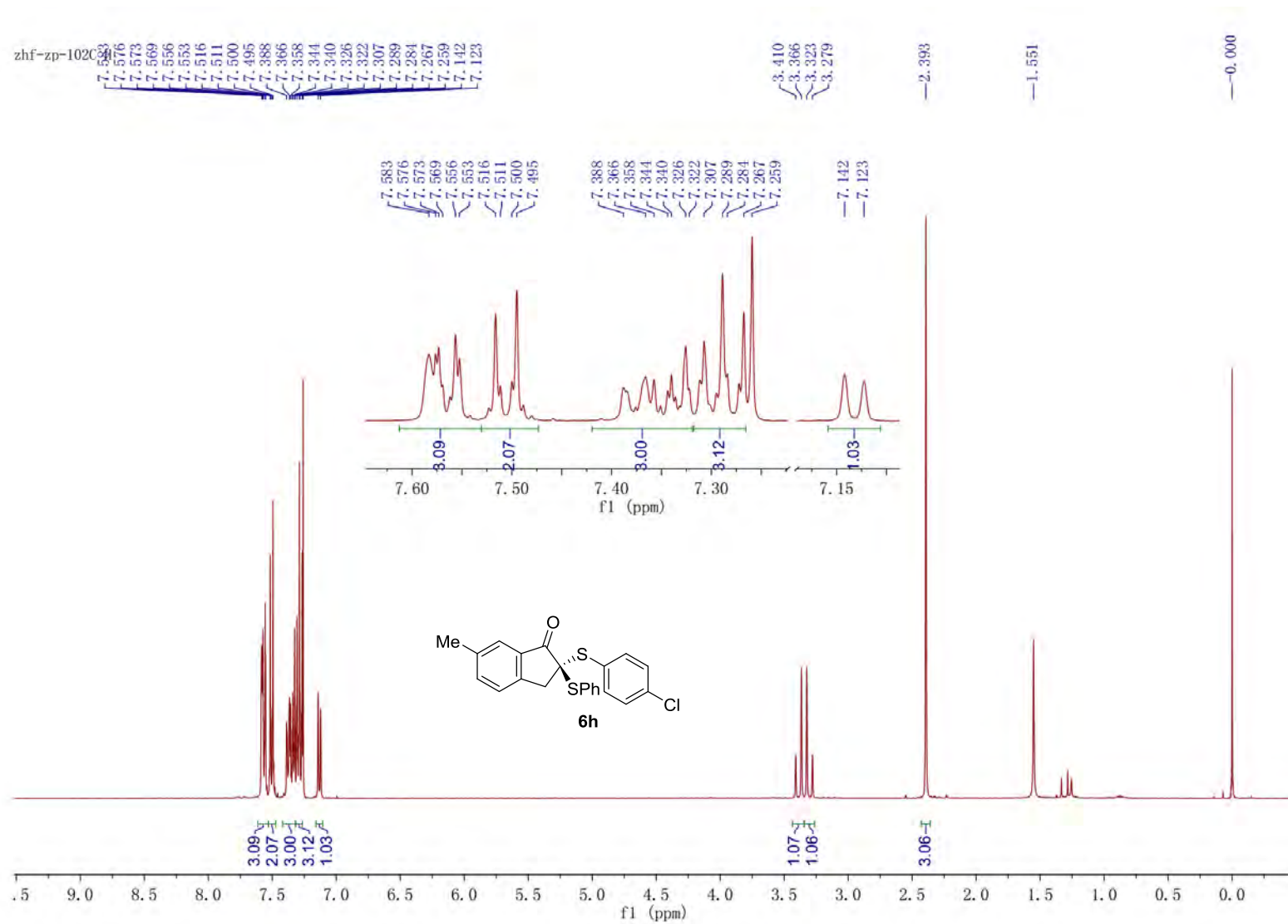




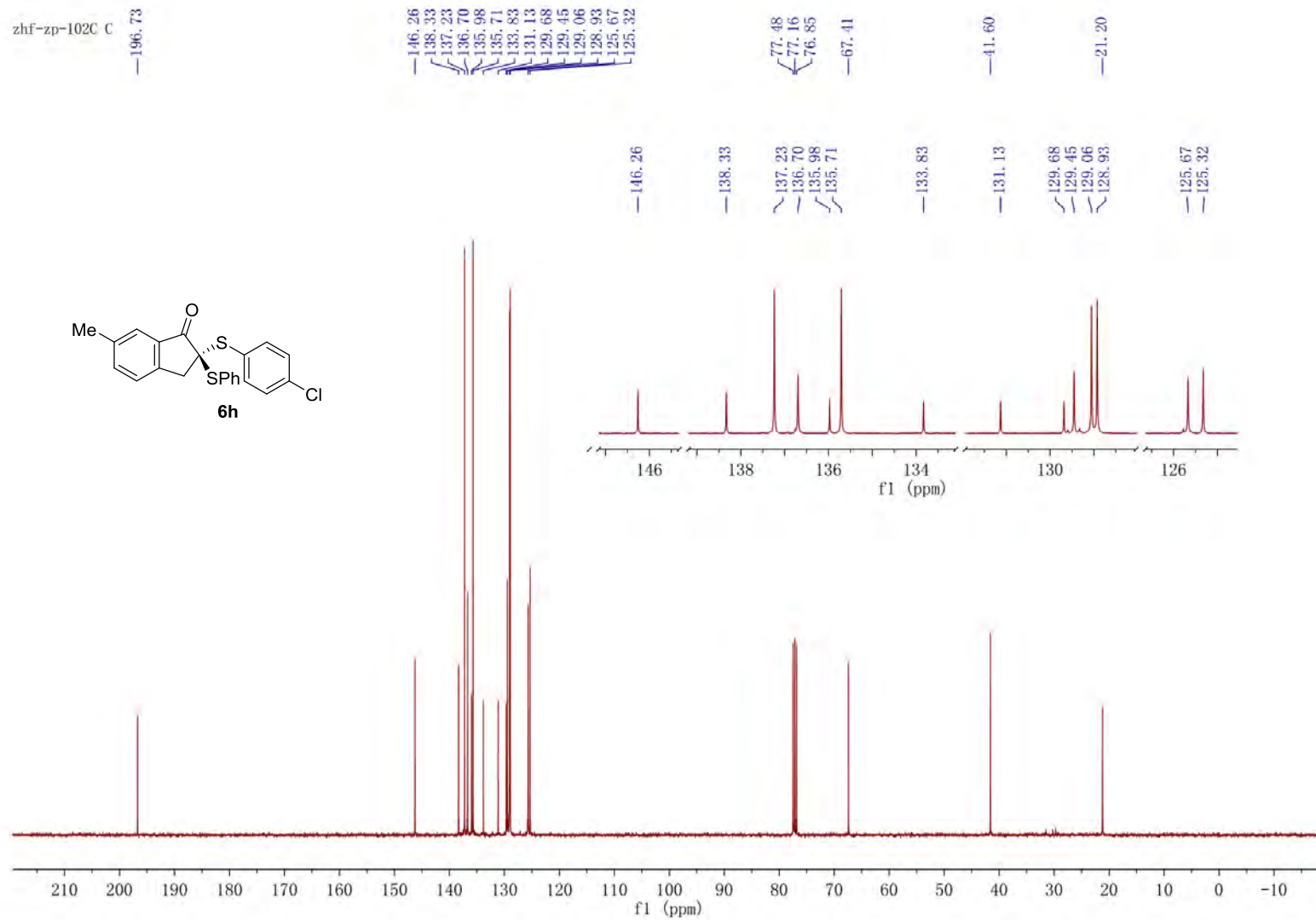
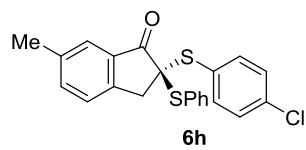


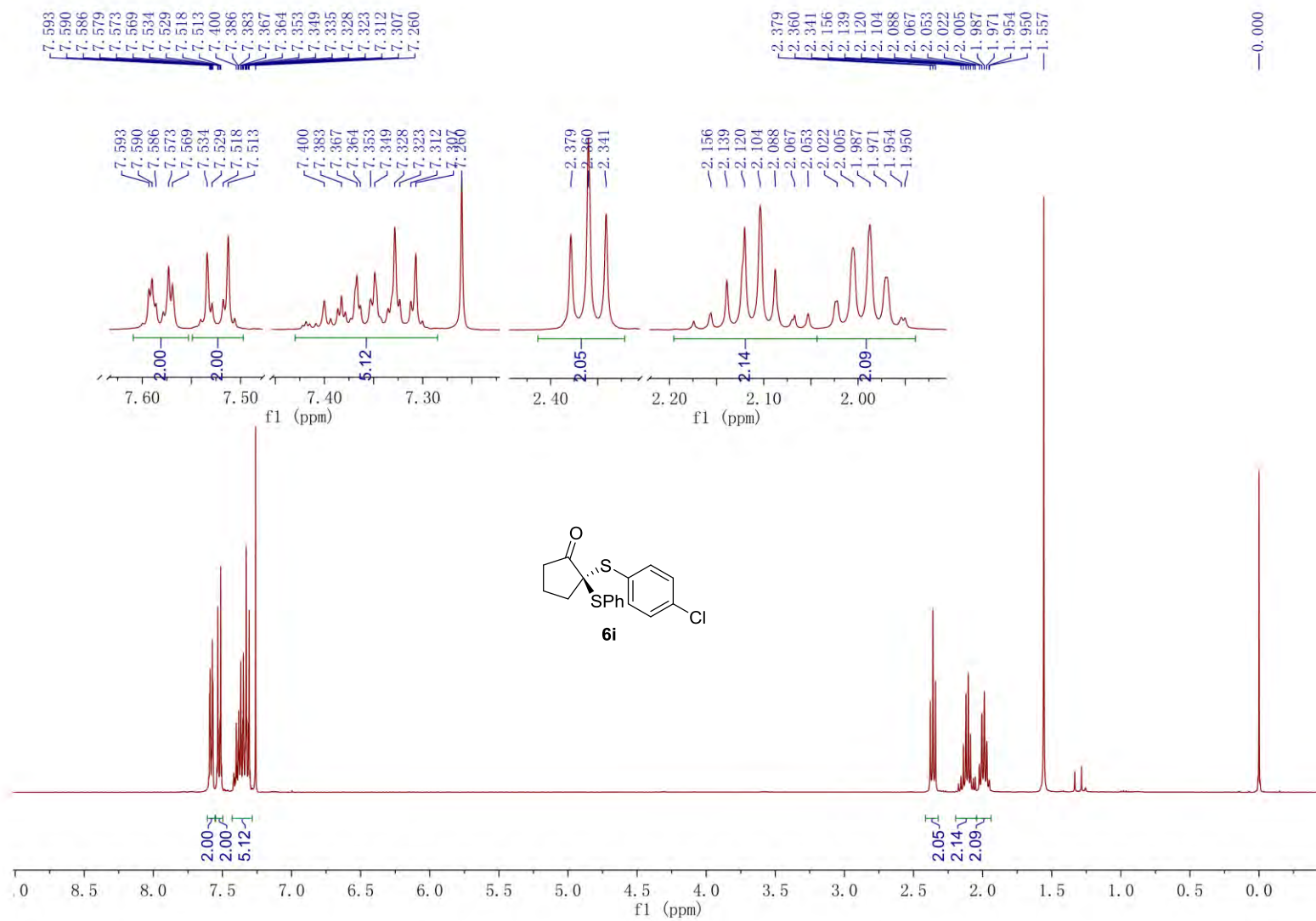
zhf-zp-102B C

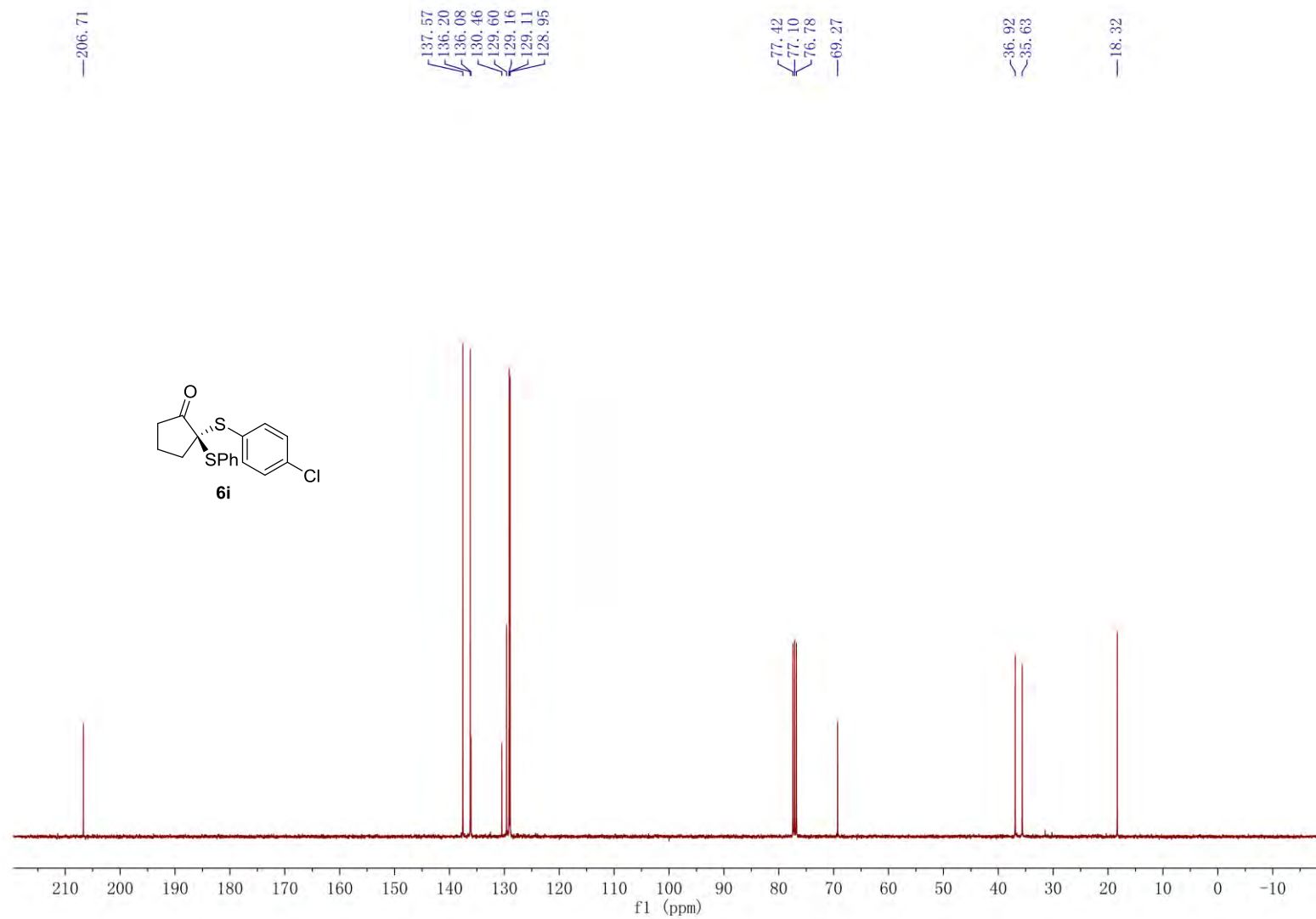


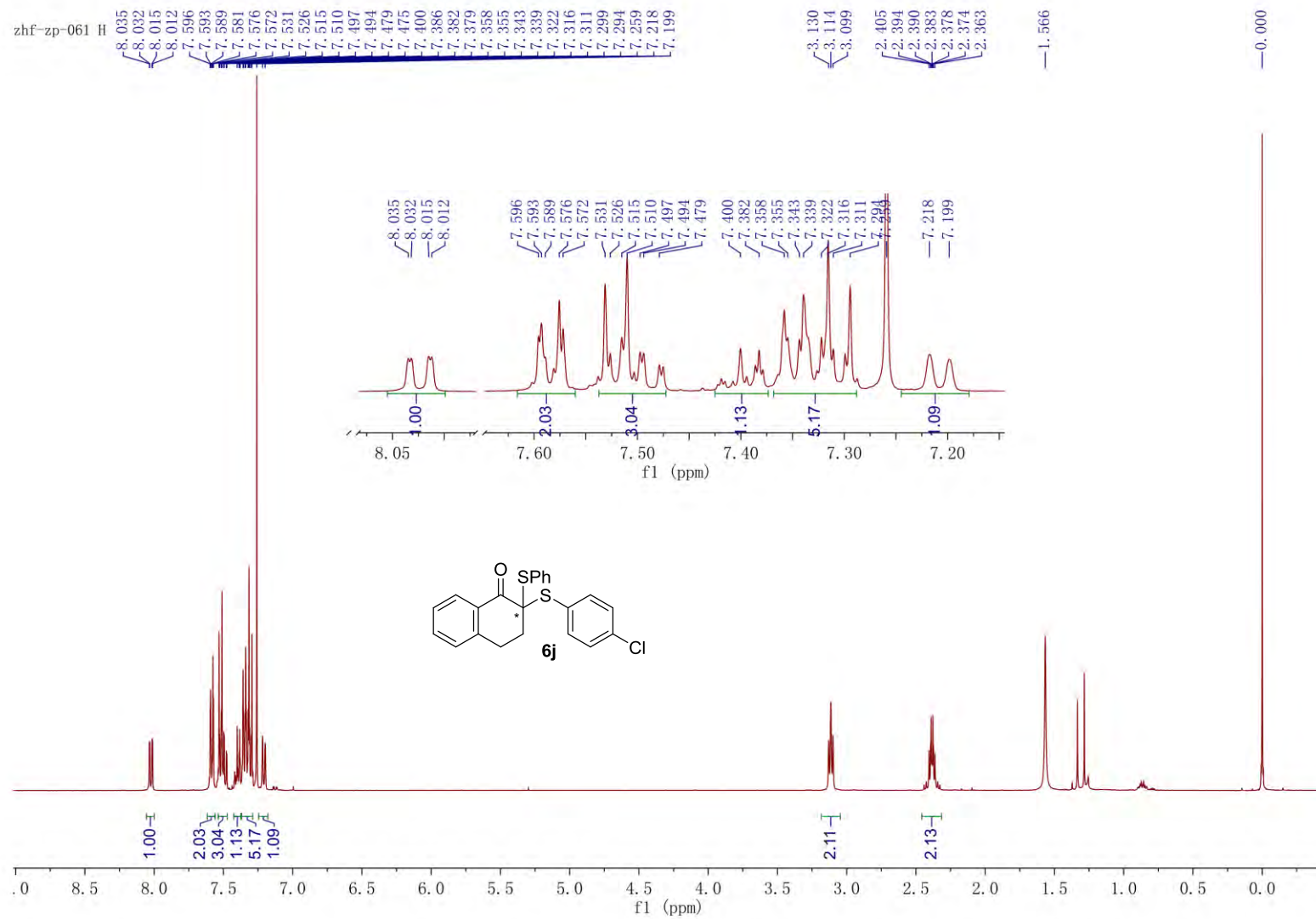


zhf-zp-102C C

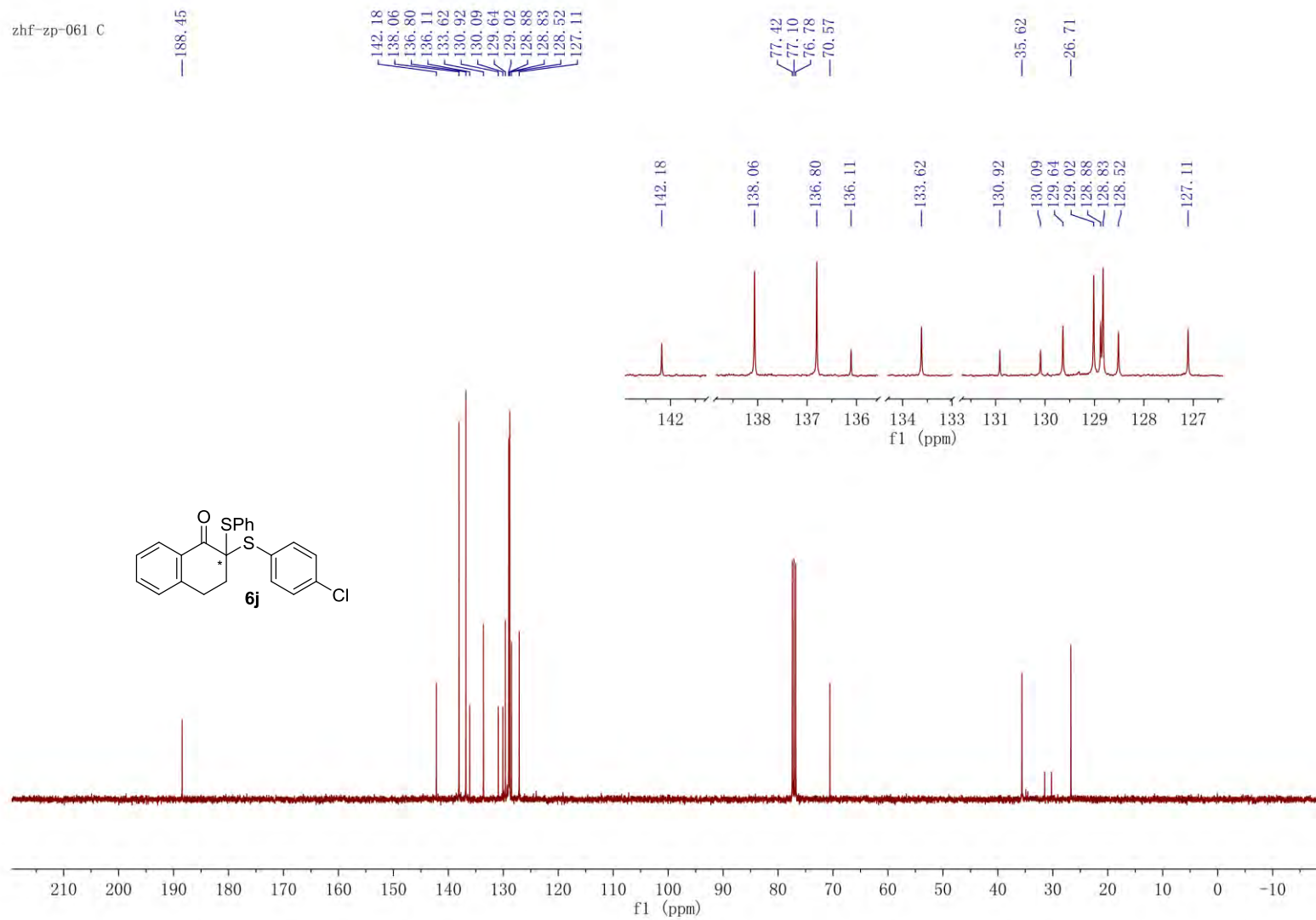


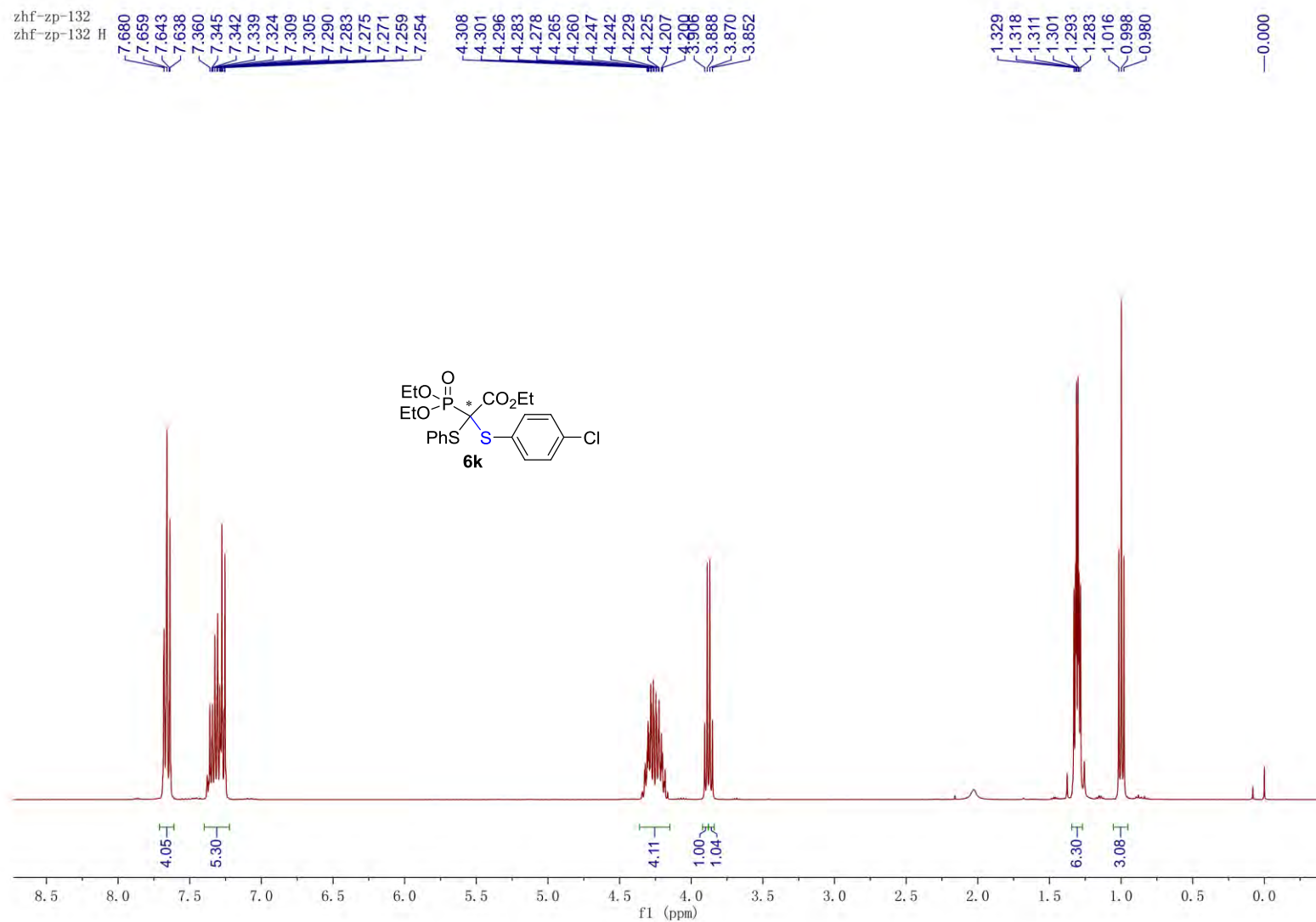




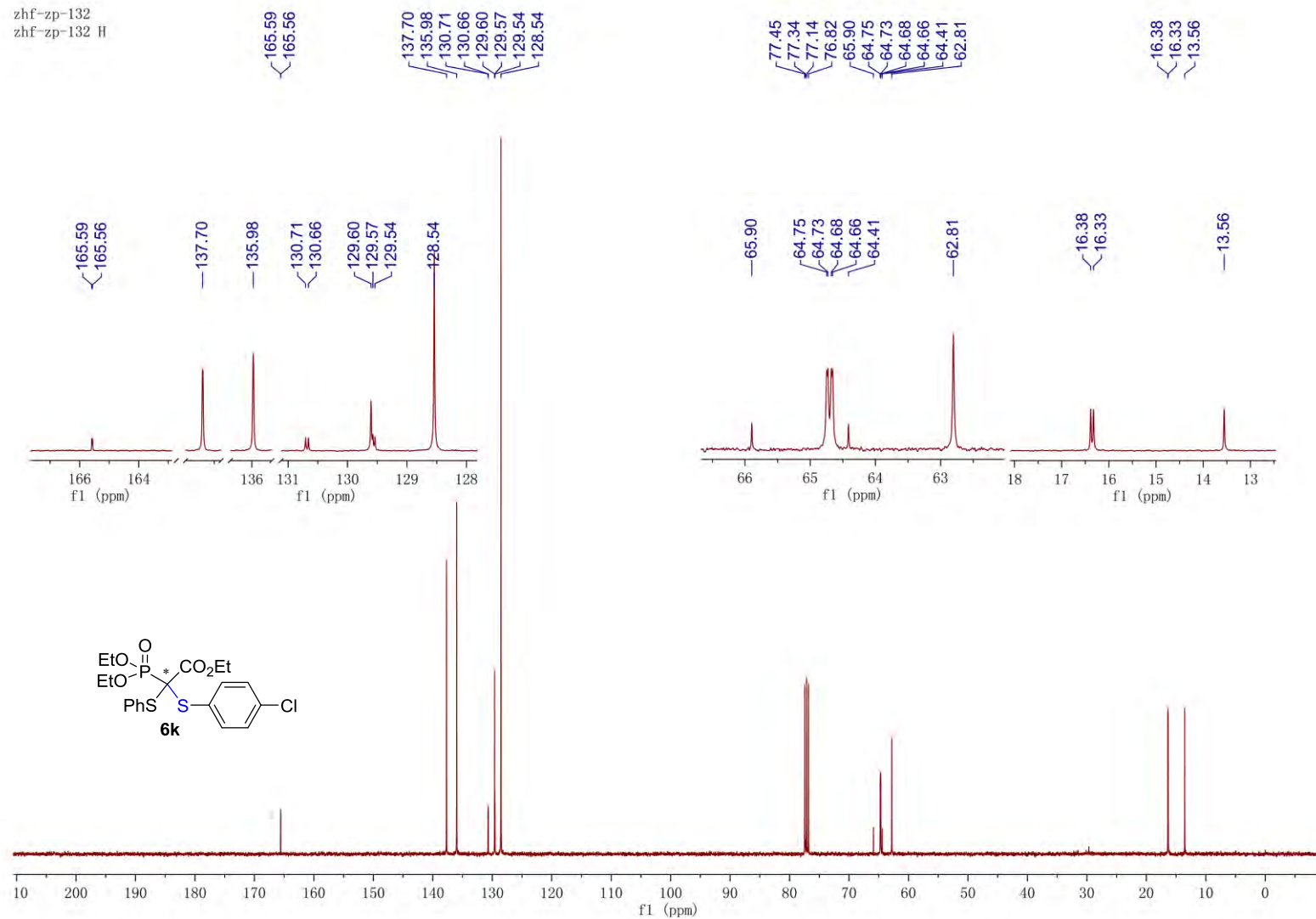


zhf-zp-061 C



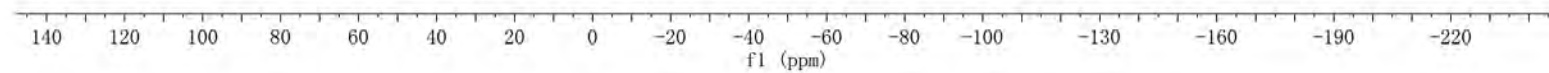
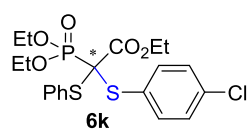


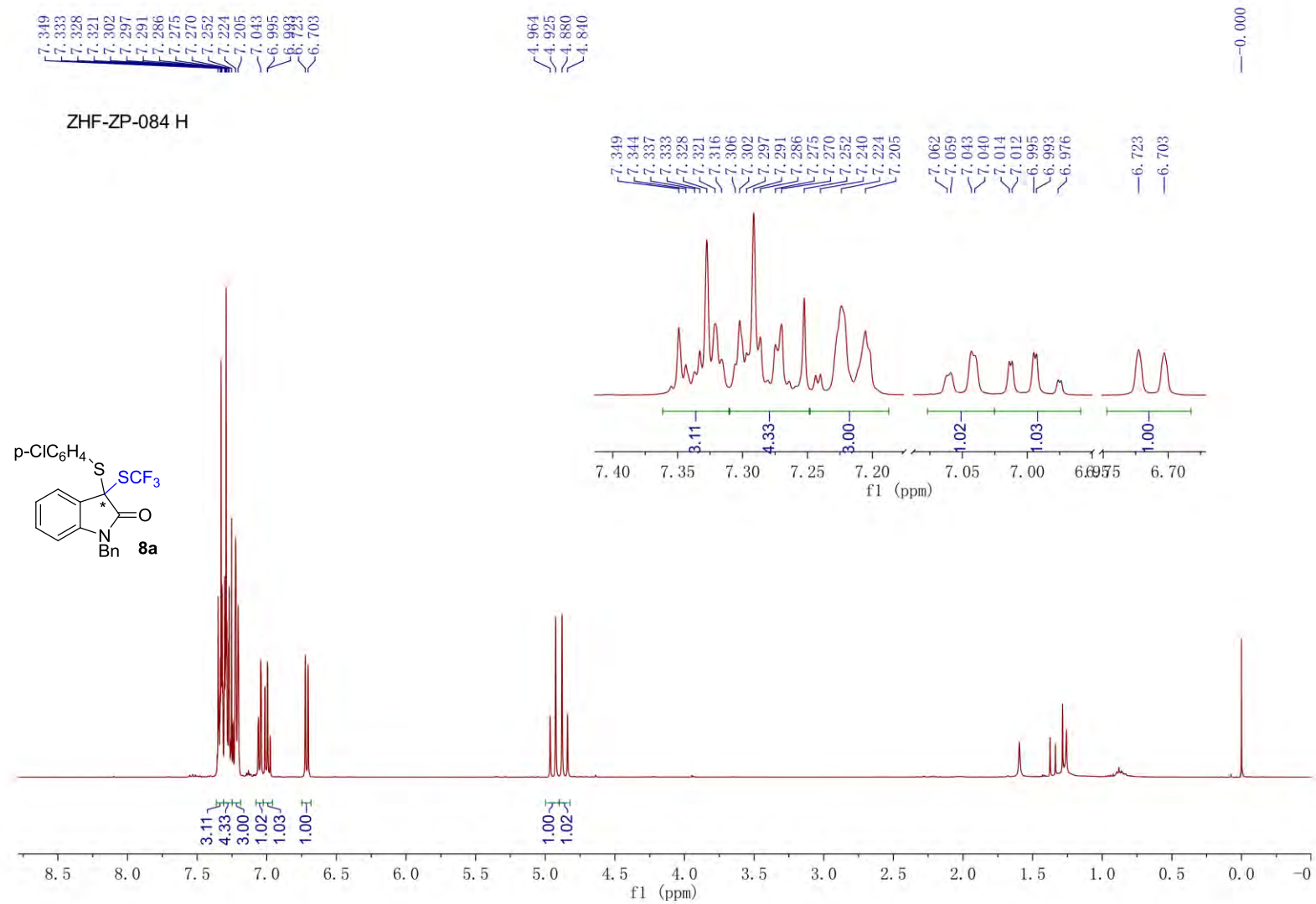
zhf-zp-132
zhf-zp-132 H



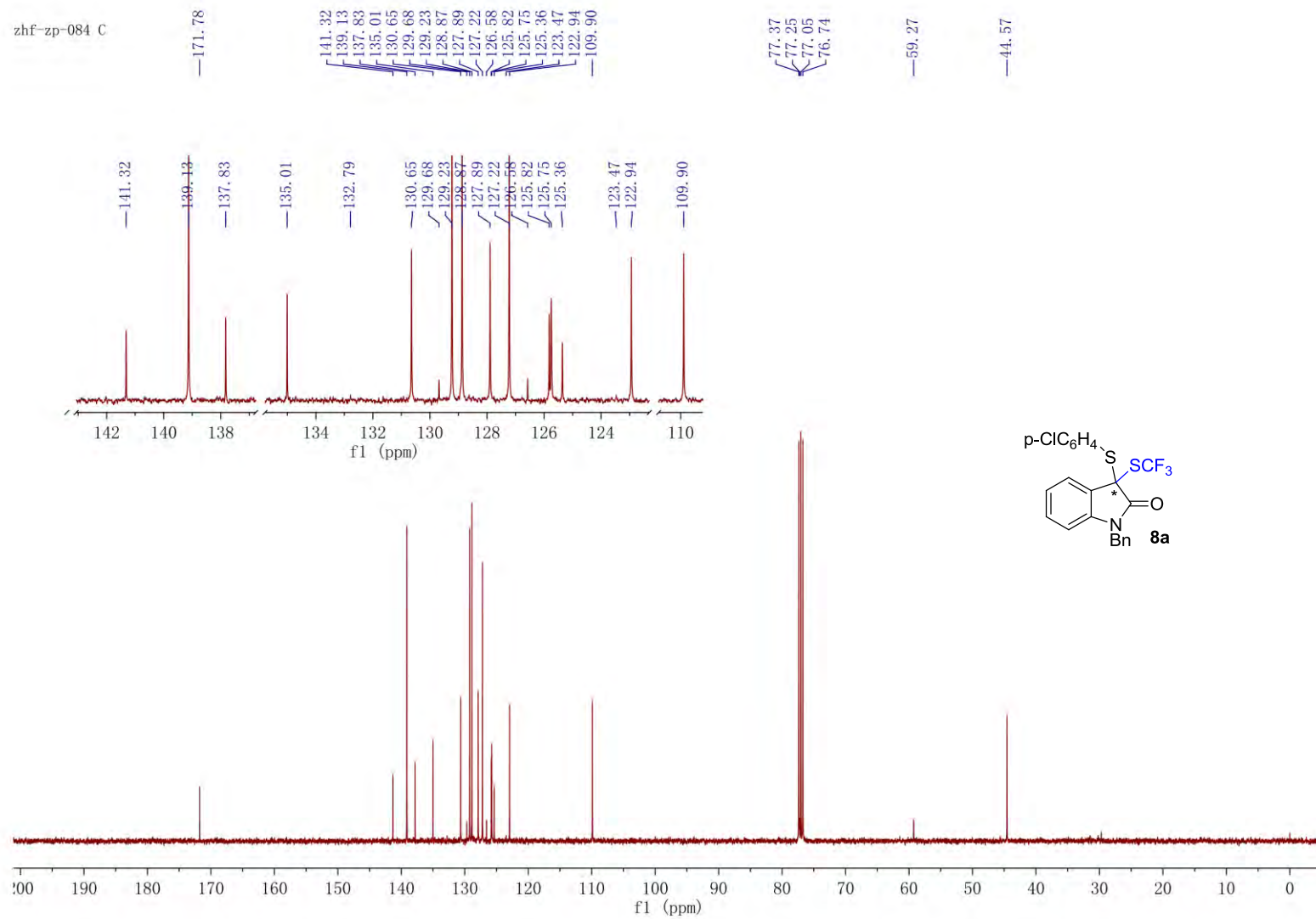
zhf-zp-132-P

—13.913



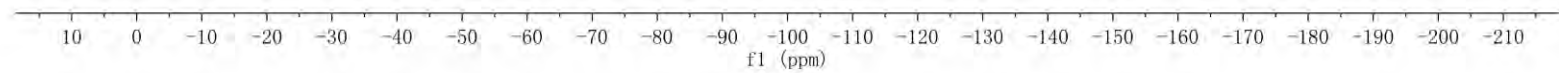
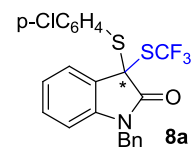


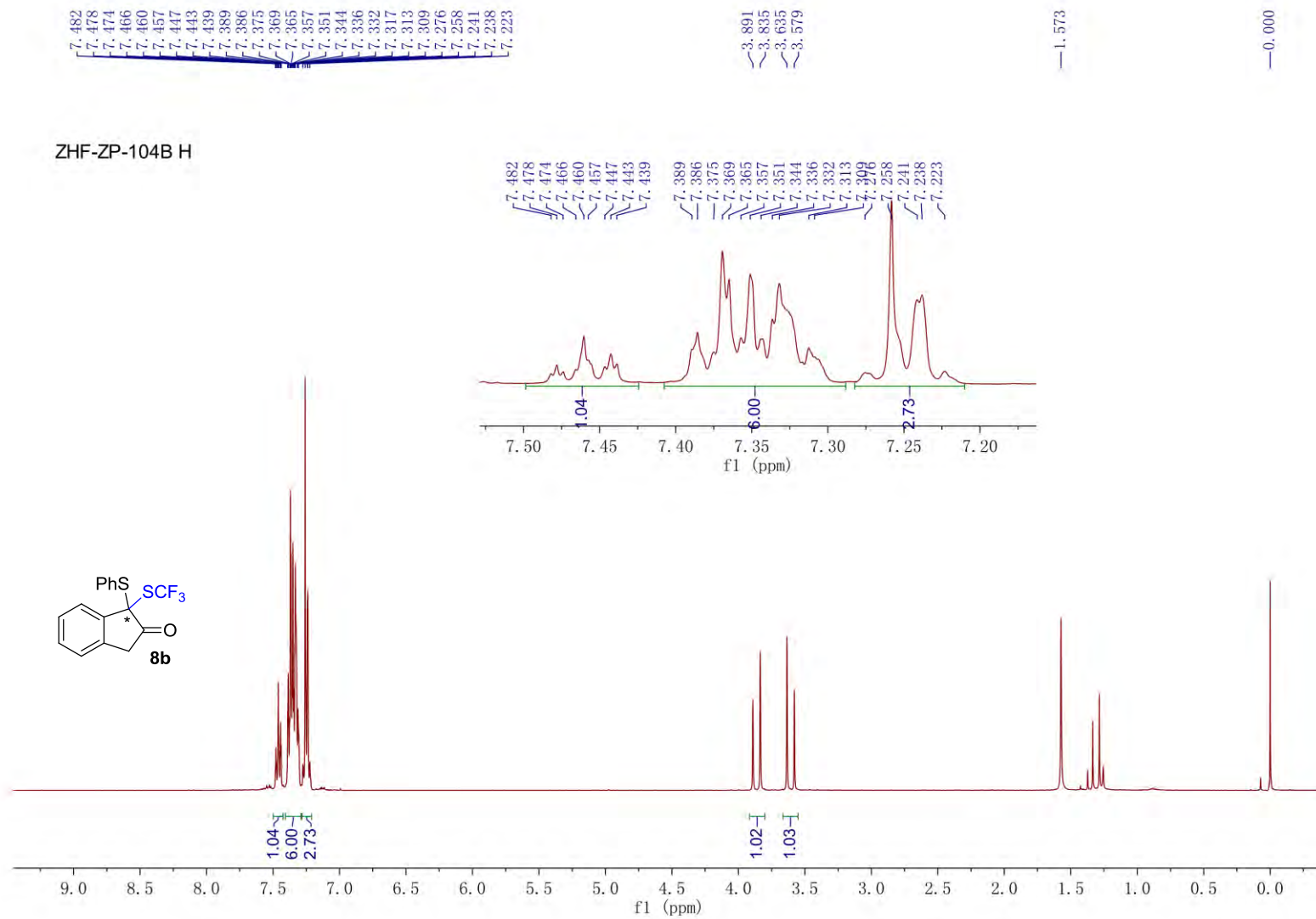
zhf-zp-084 C

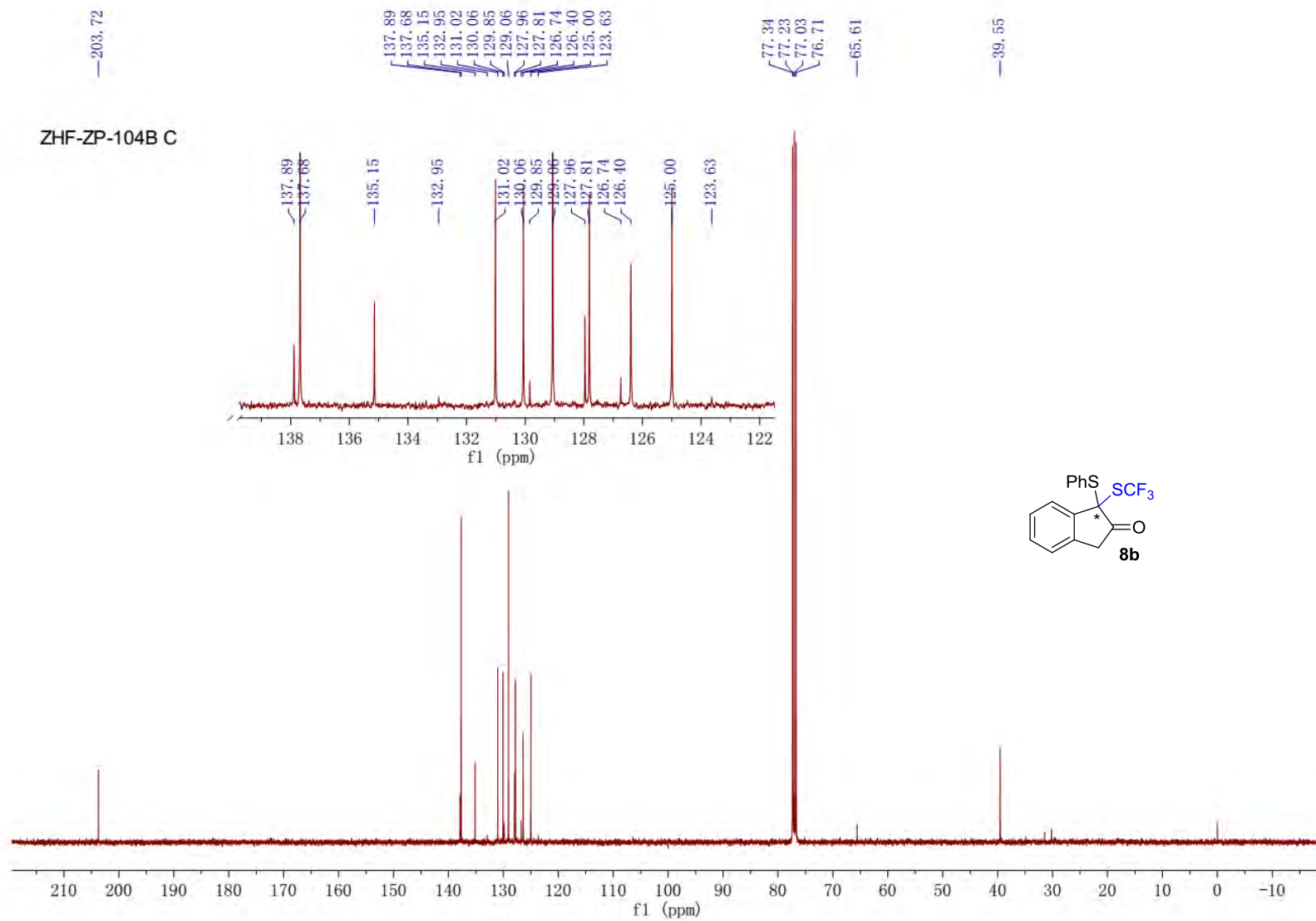


zhf-zp-084 F

-39.110

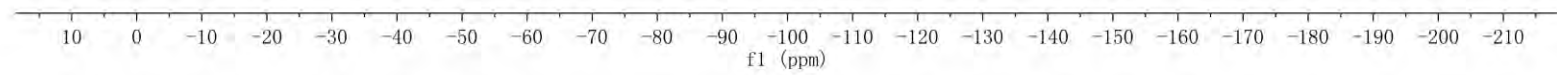
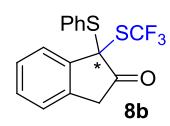


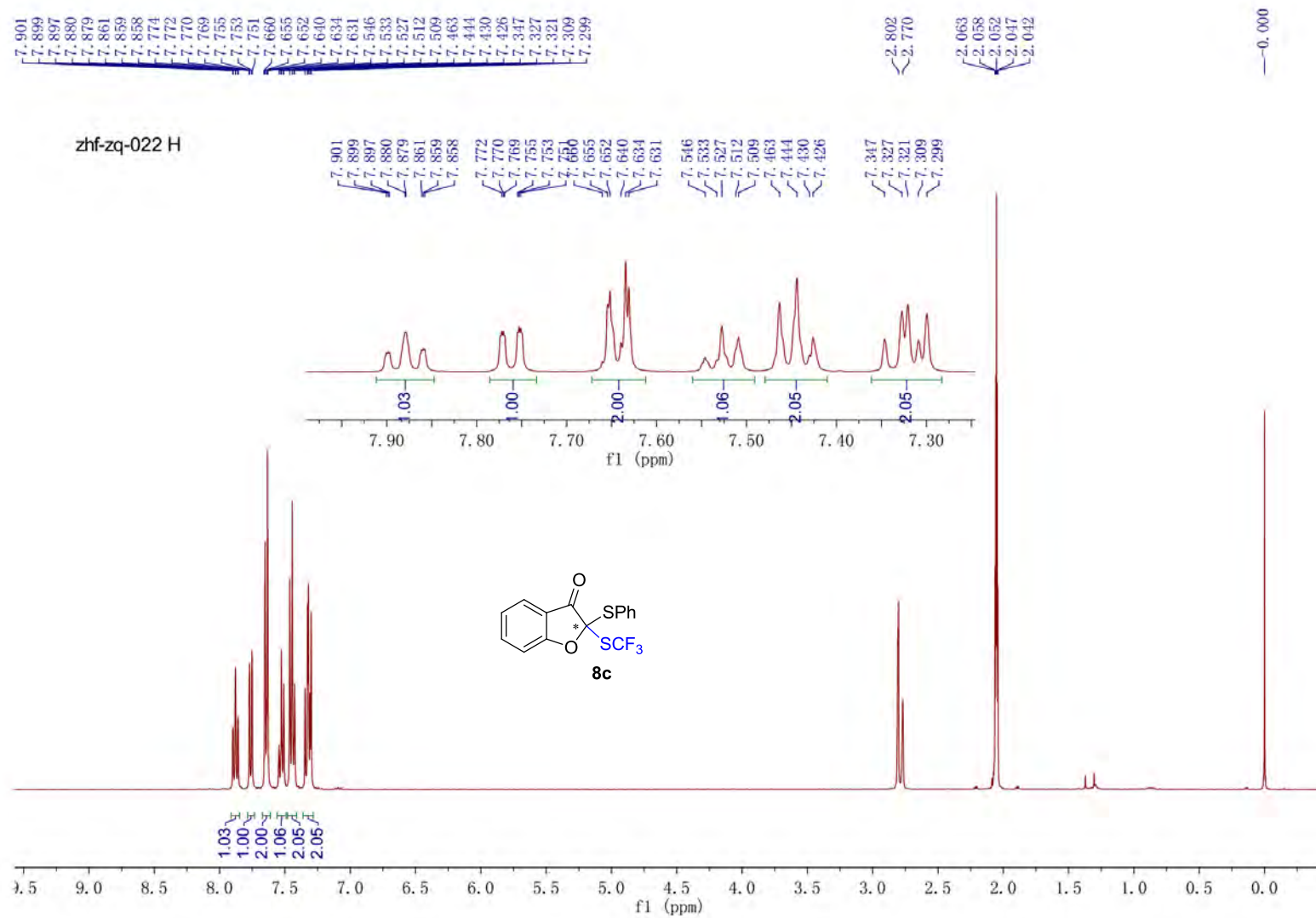


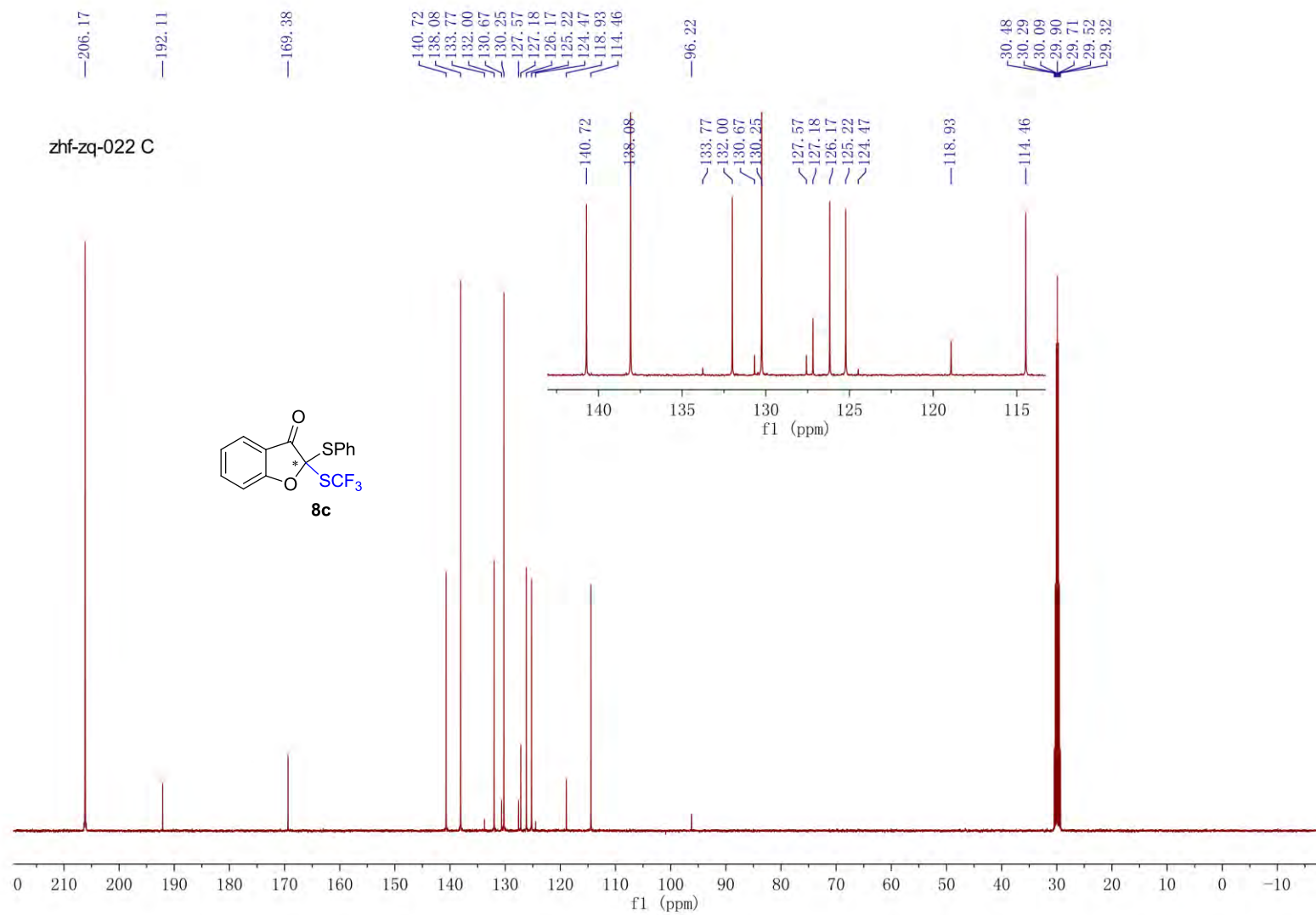


zhf-zp-104B F

— -38.227

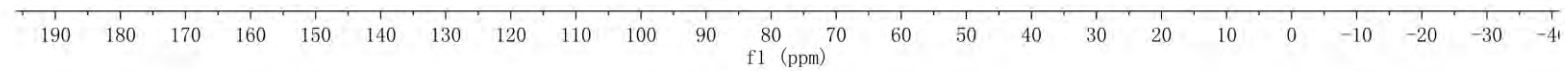
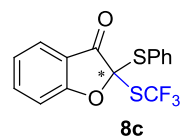


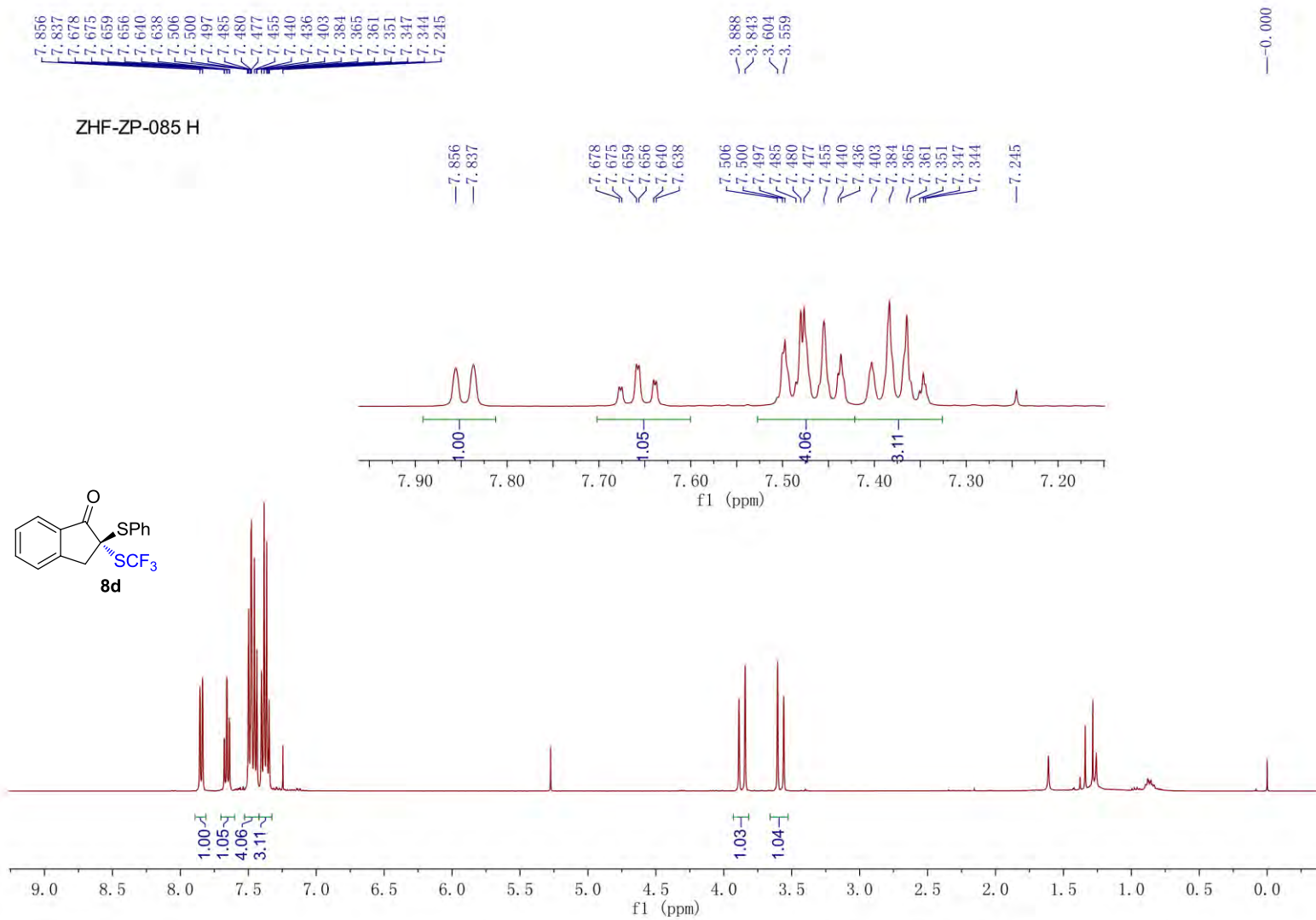


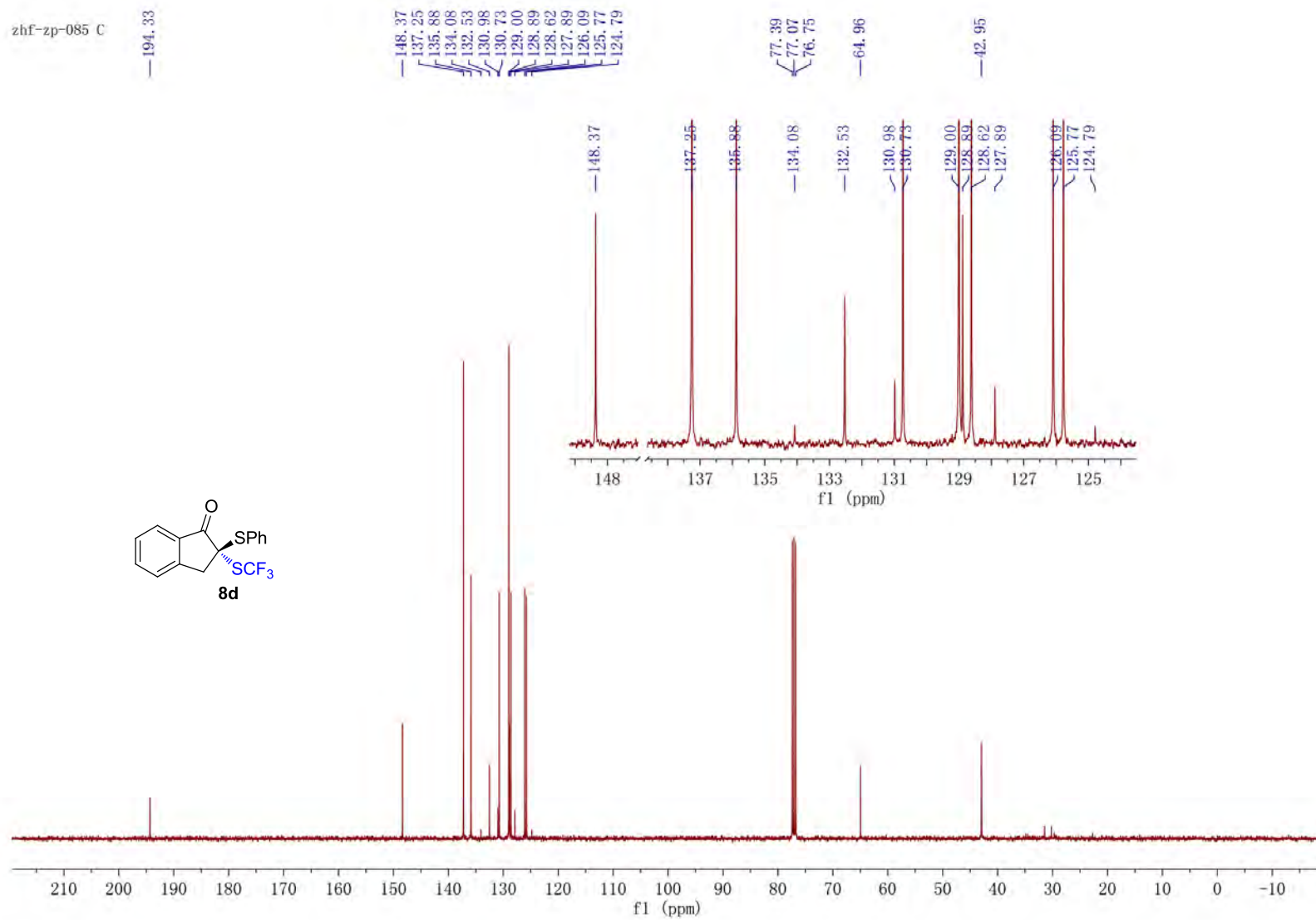


zhf-zq-022 F

—138.263

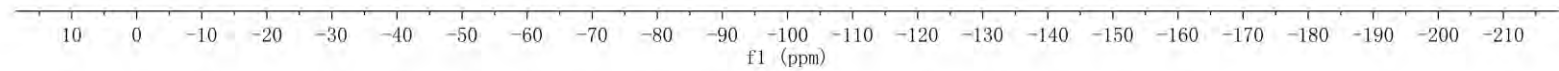
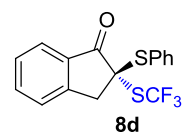


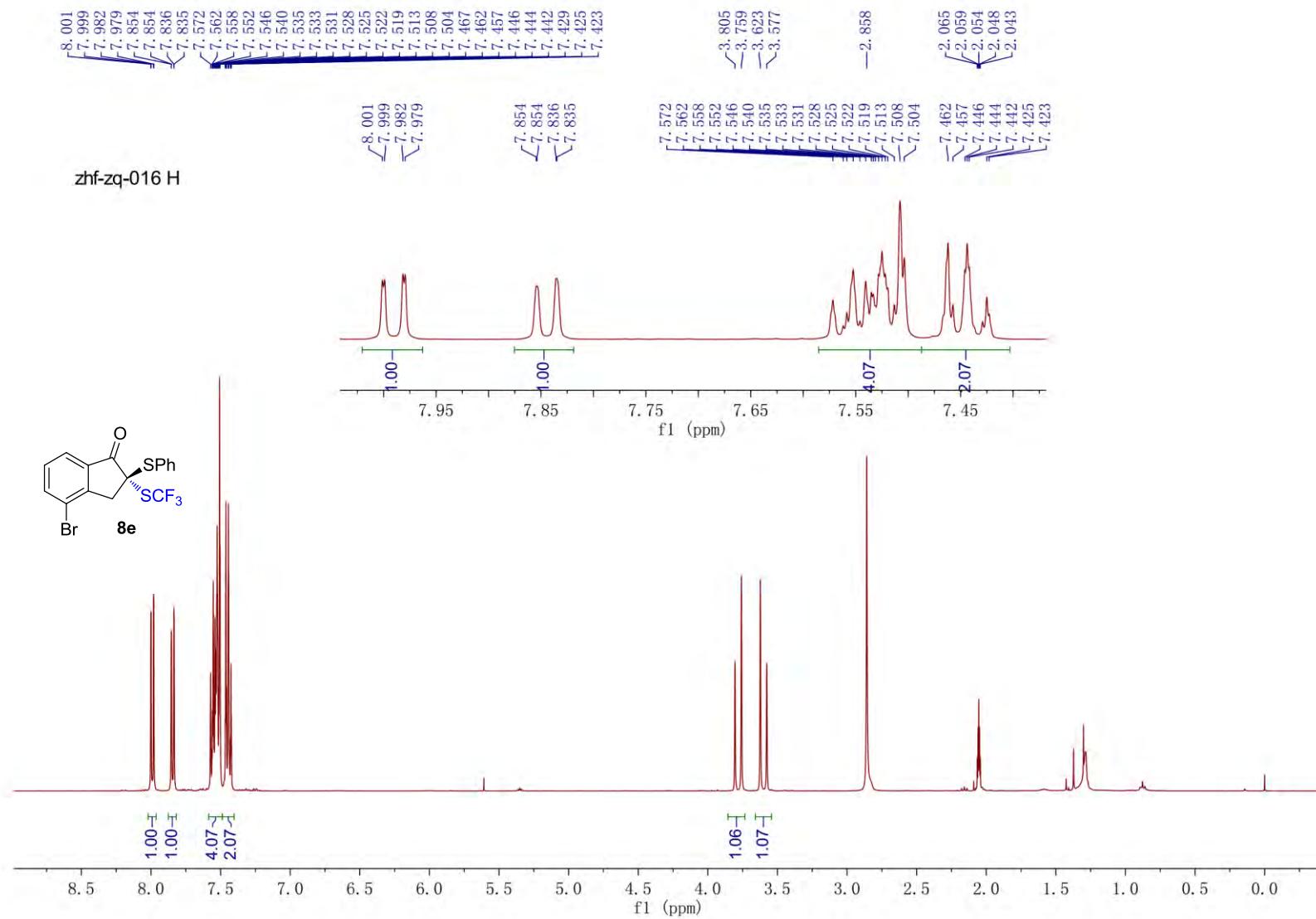


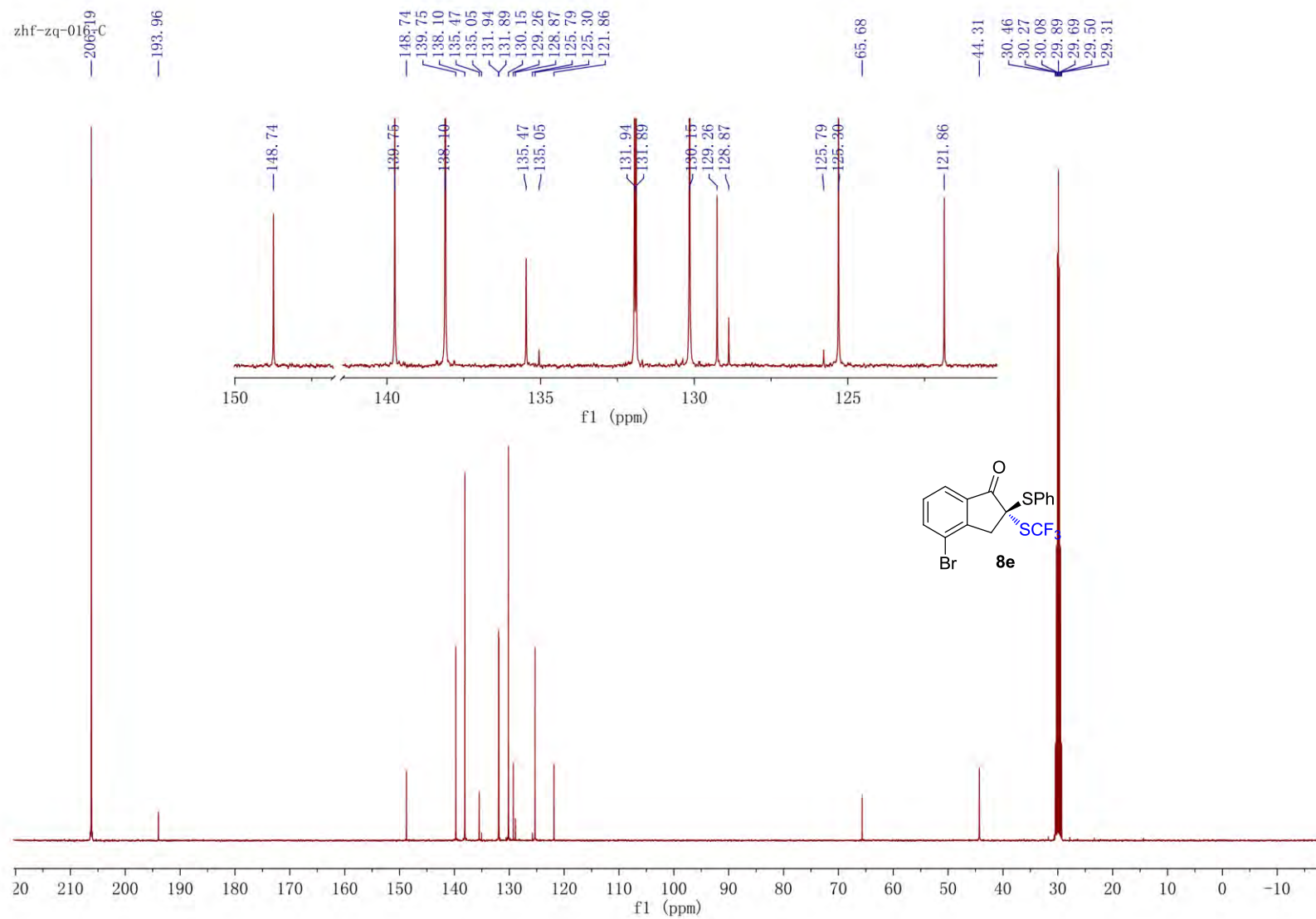


zhf-zp-085 F

— -37.145

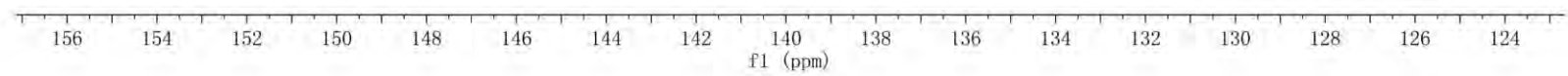
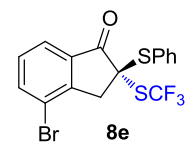


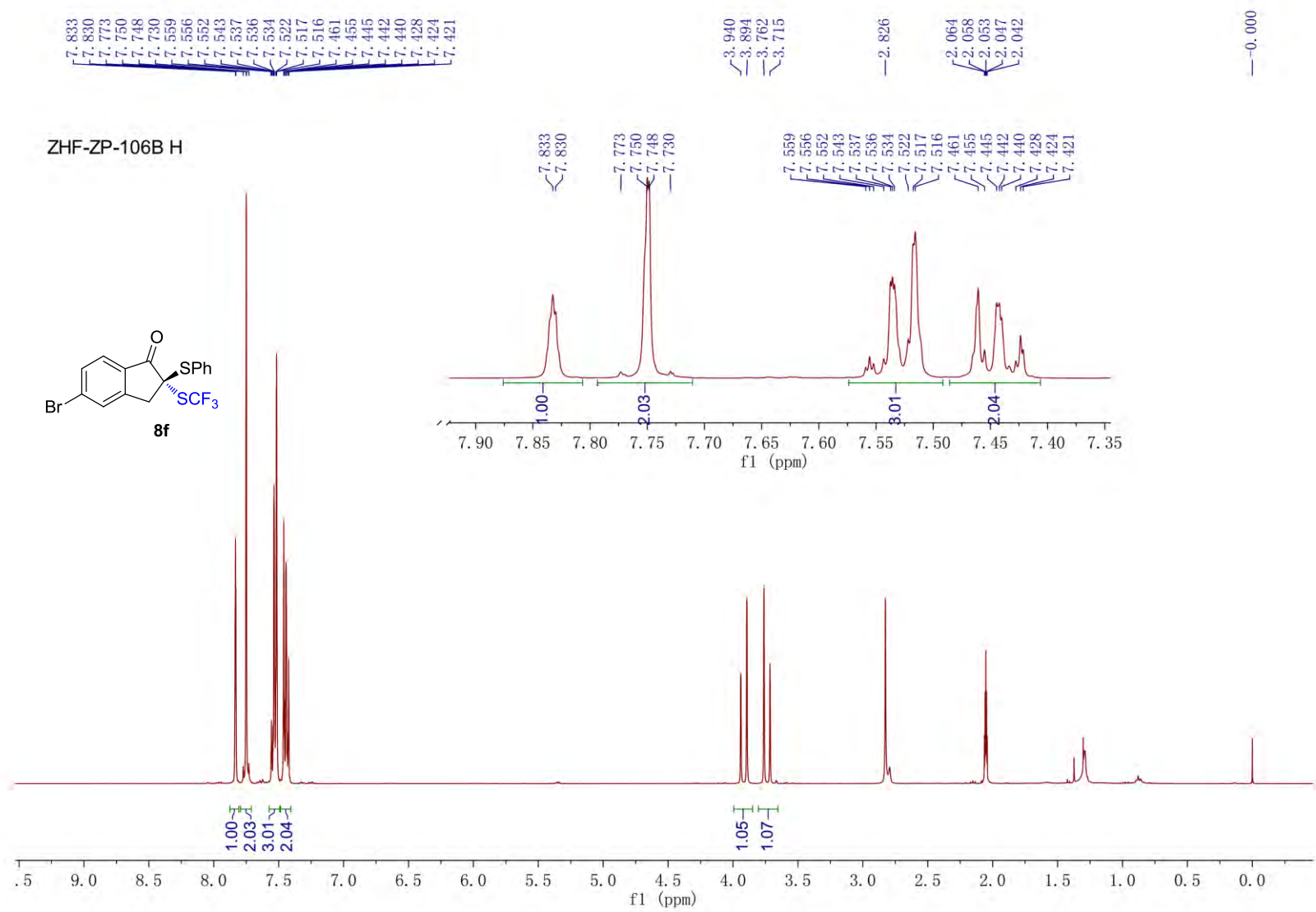


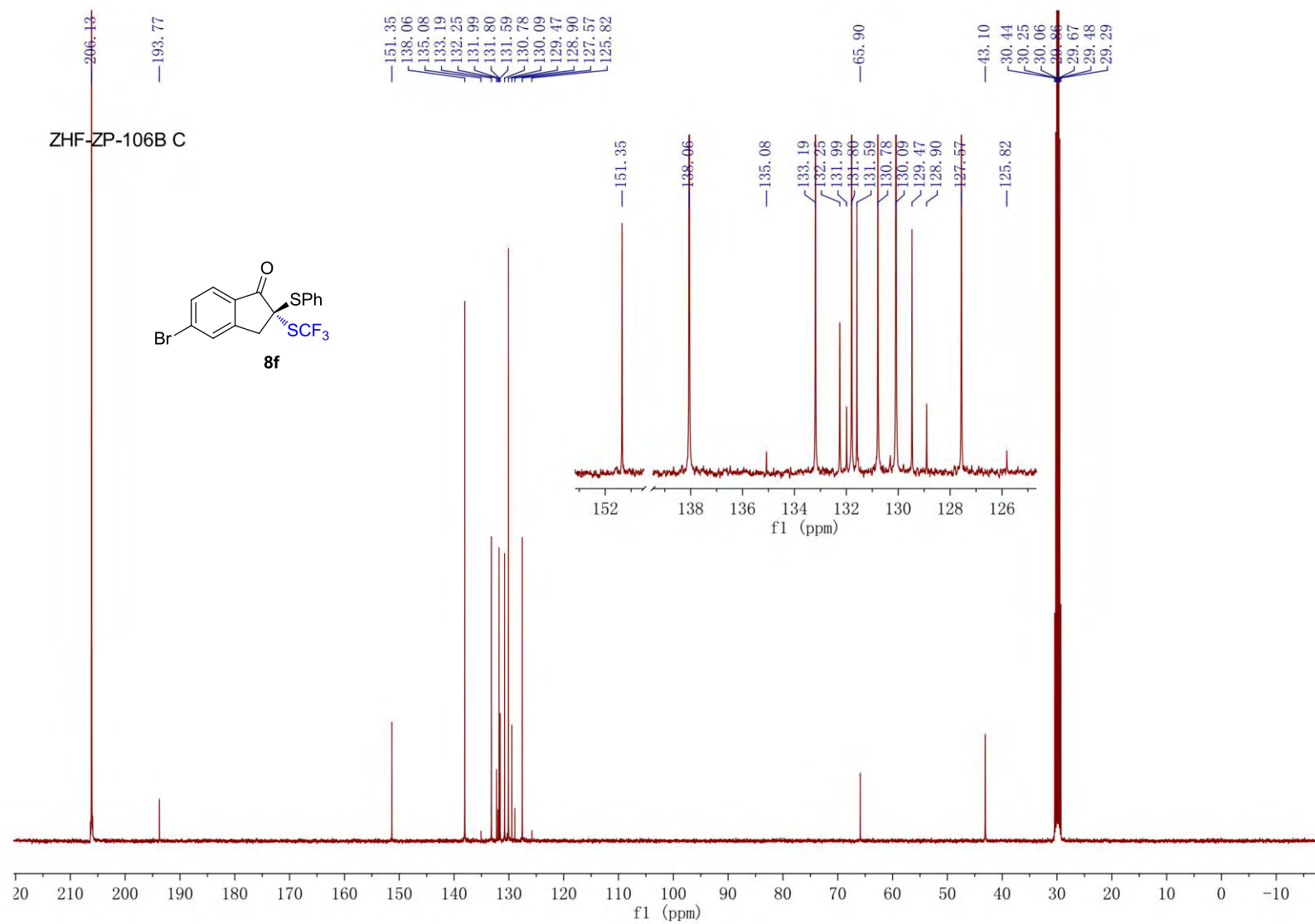


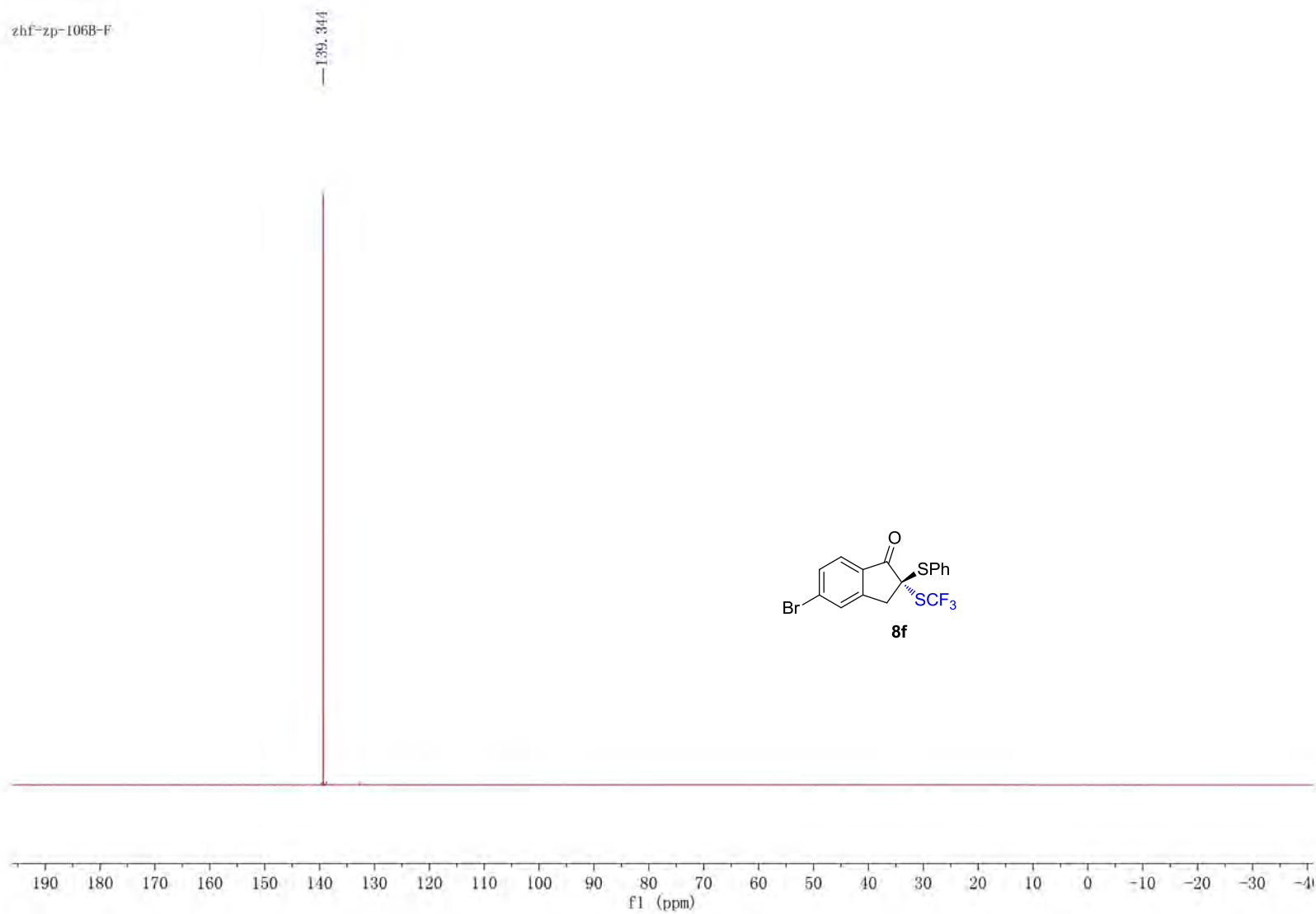
zhf-zq-016-F

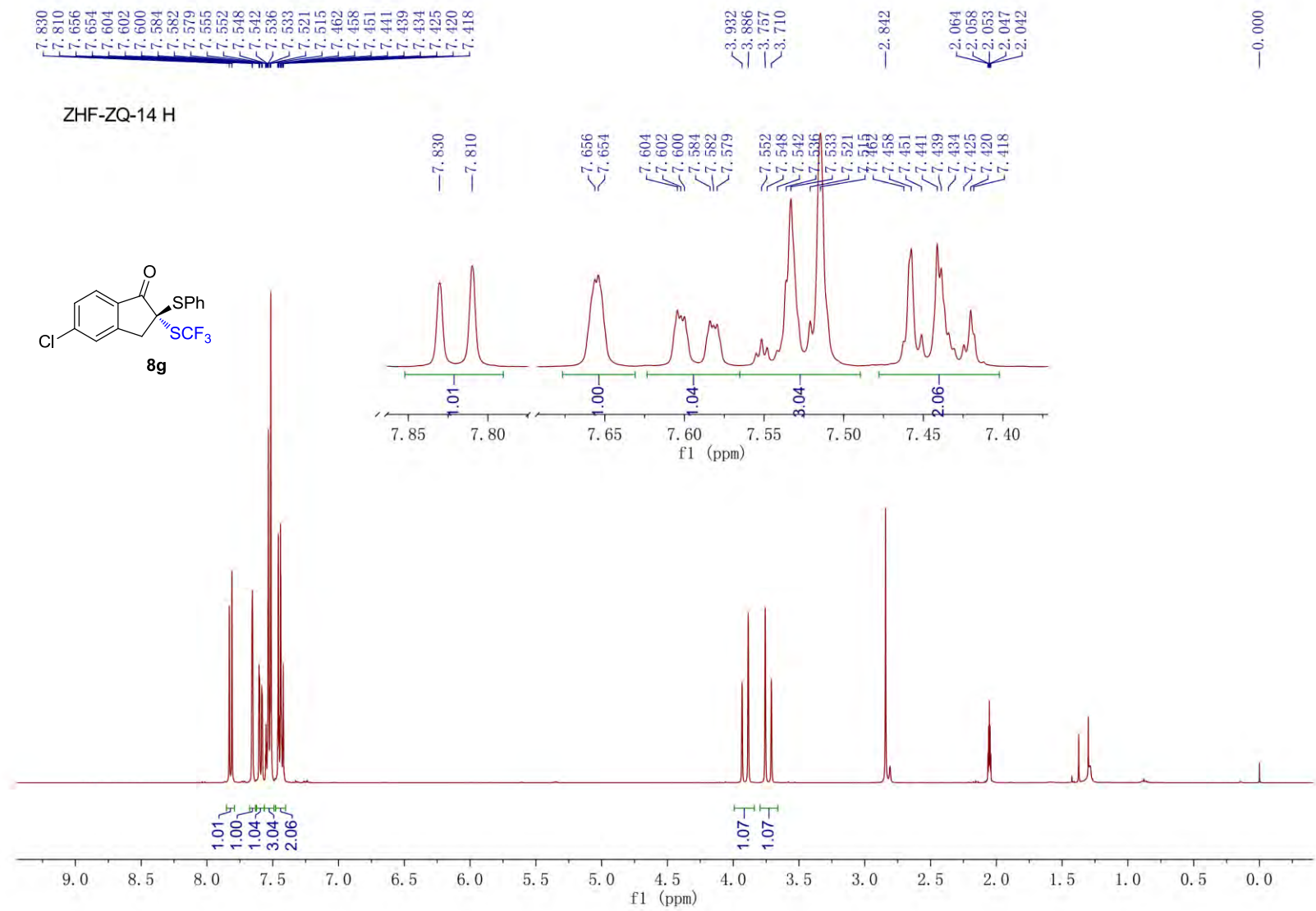
139.460

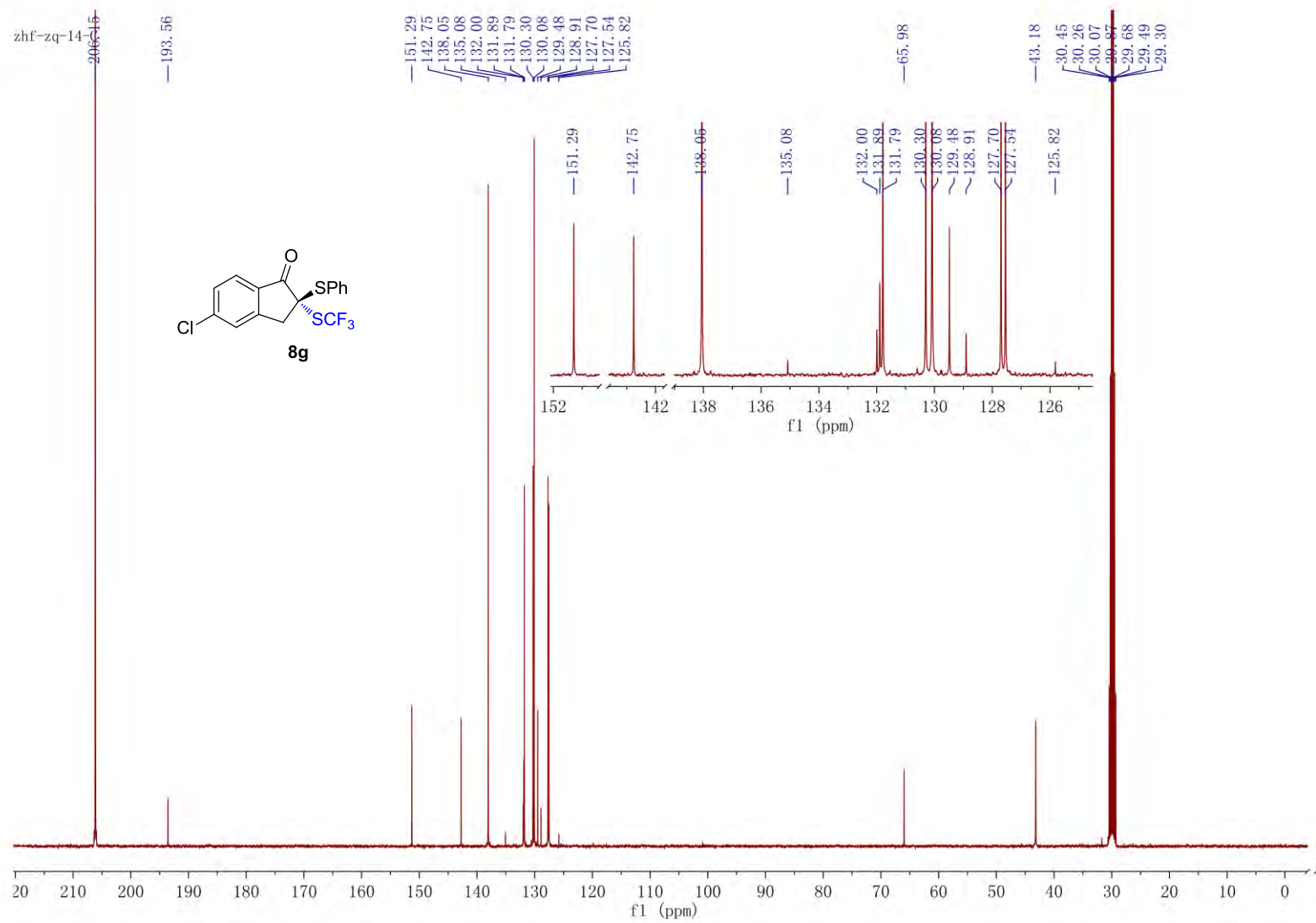






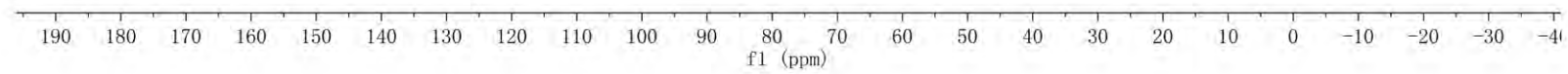
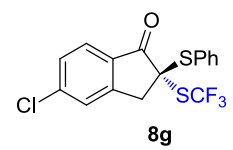


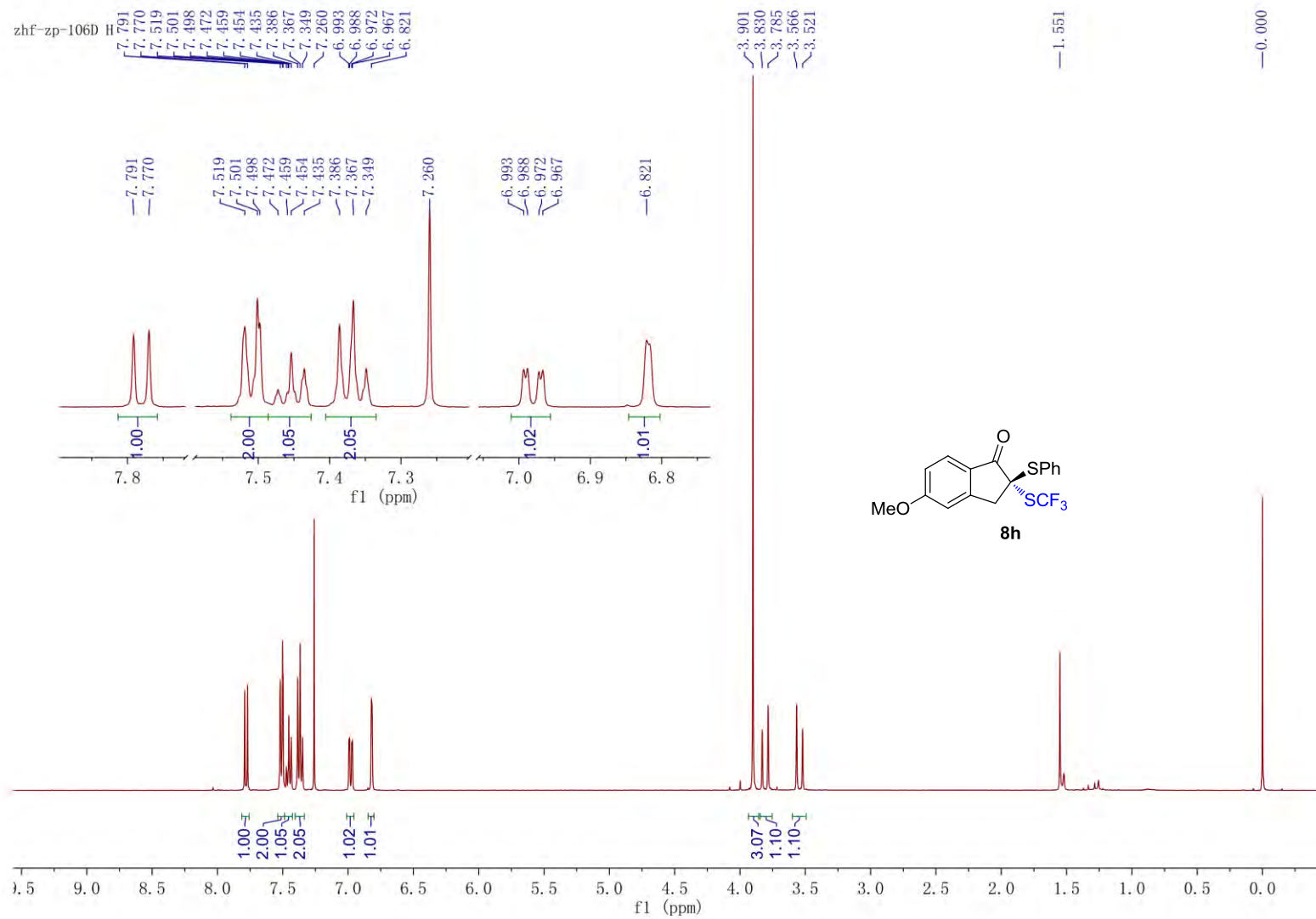


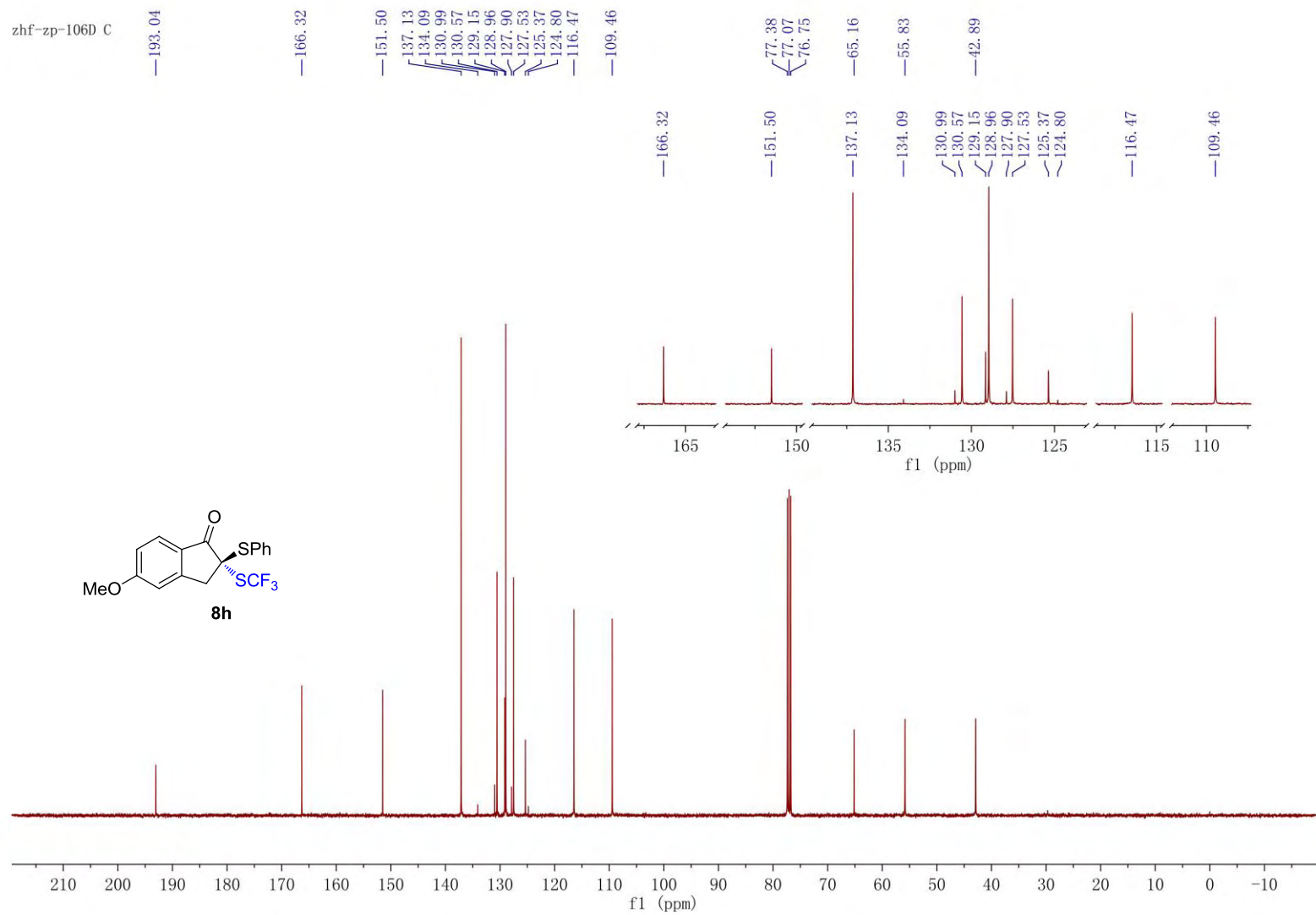


zhf-zq-14-F

—139.356

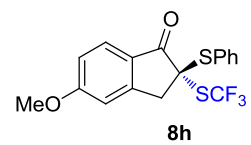




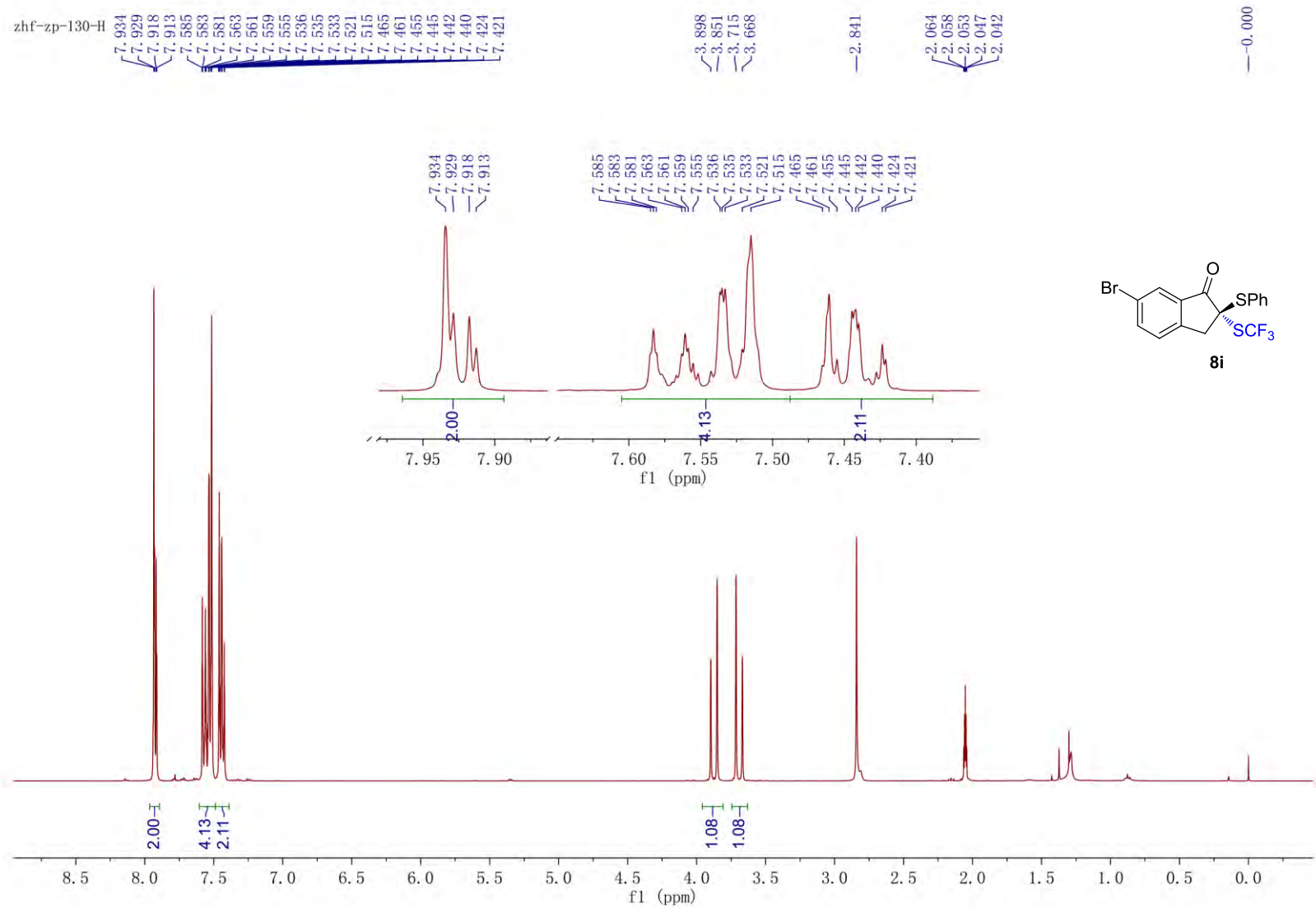


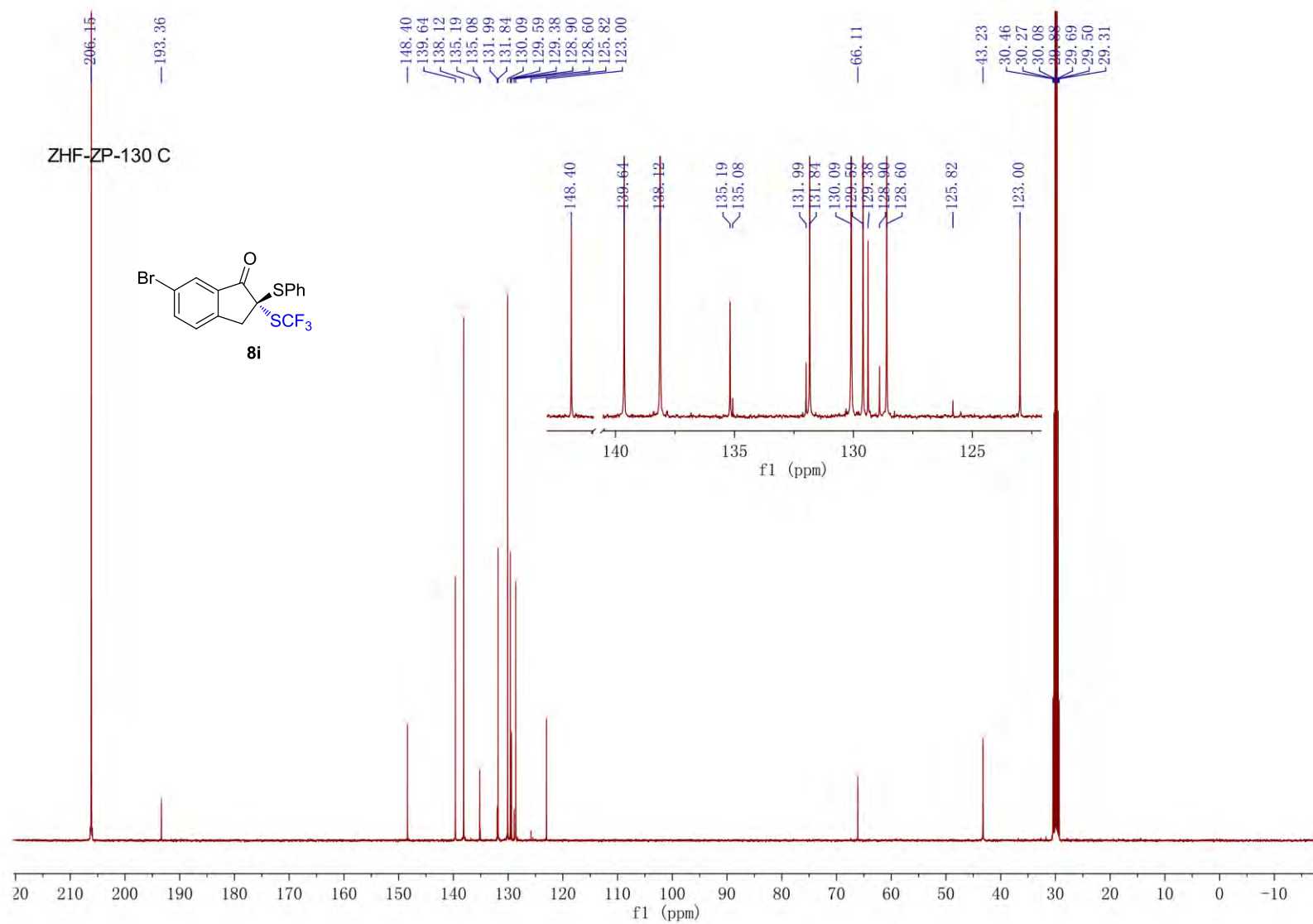
zhf-zp-106D F

-37.352



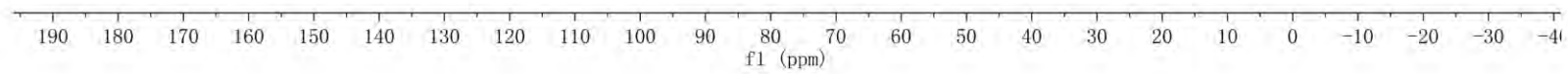
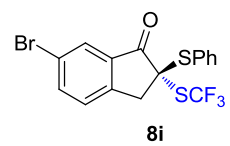
f1 (ppm)

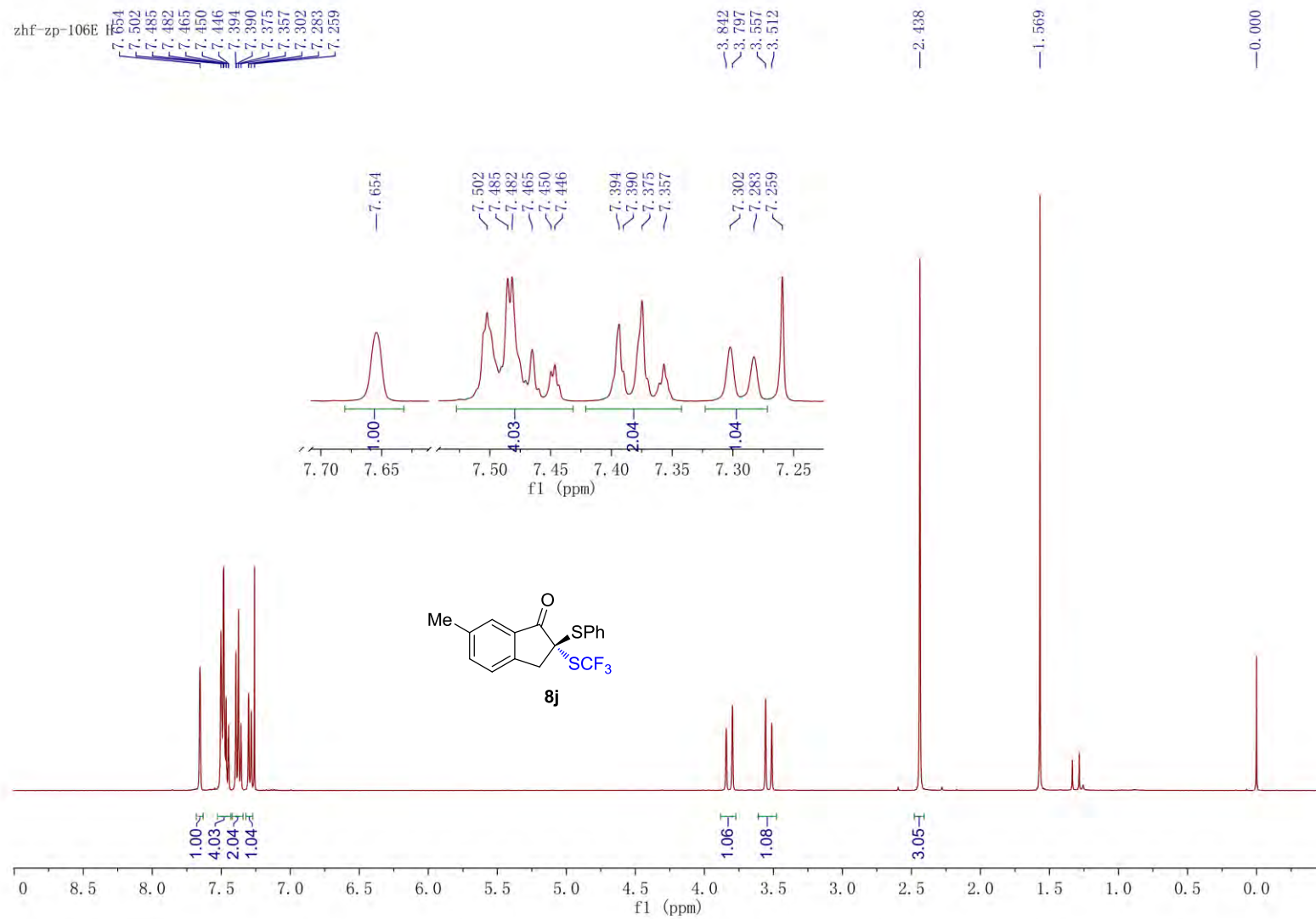




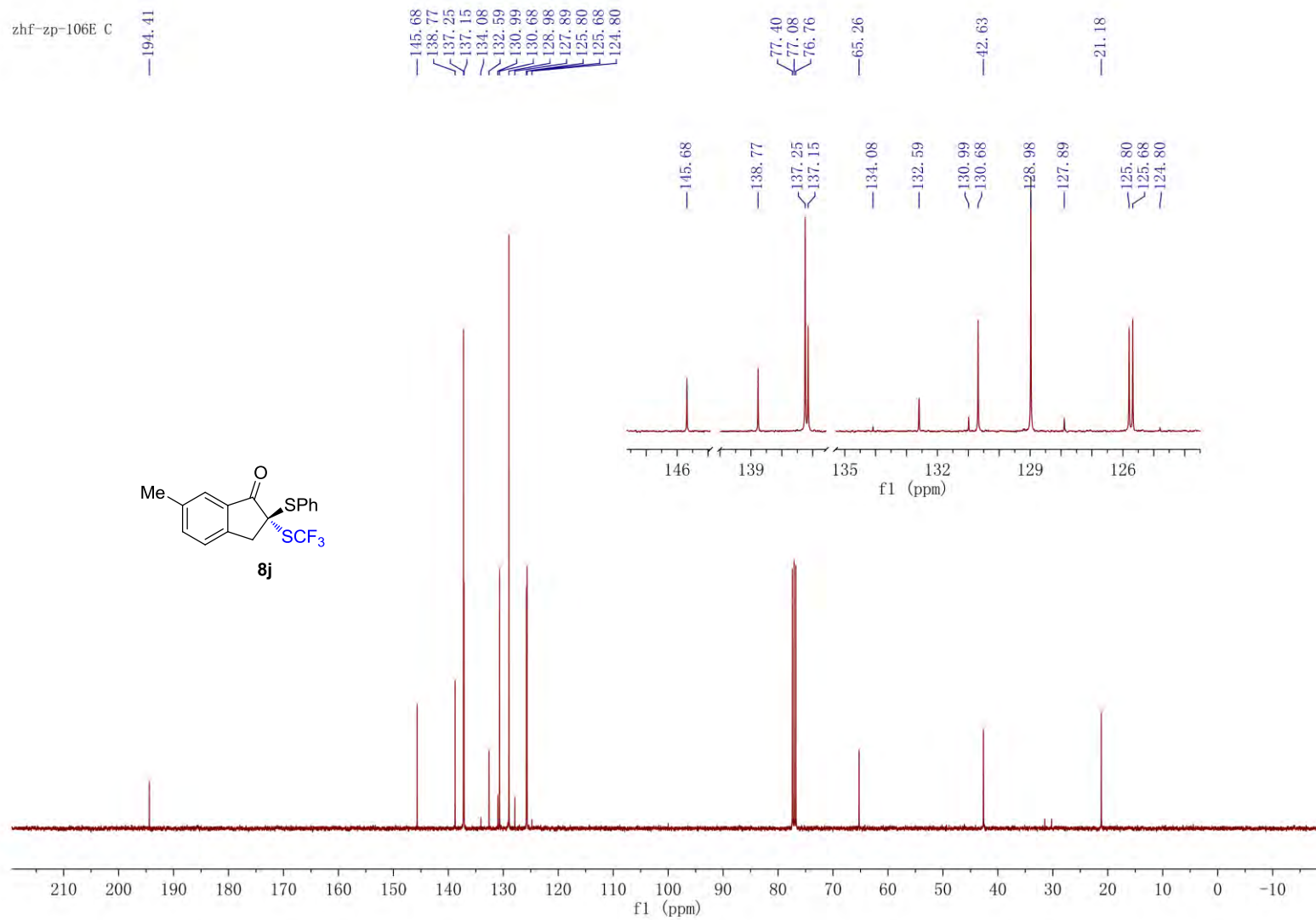
zhf-zp-130-F

—139.424



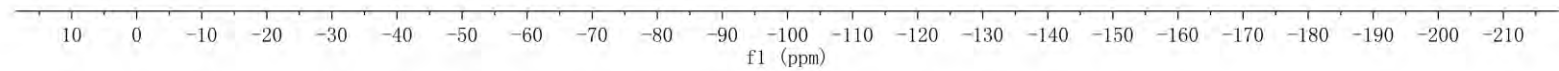
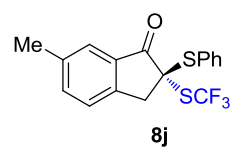


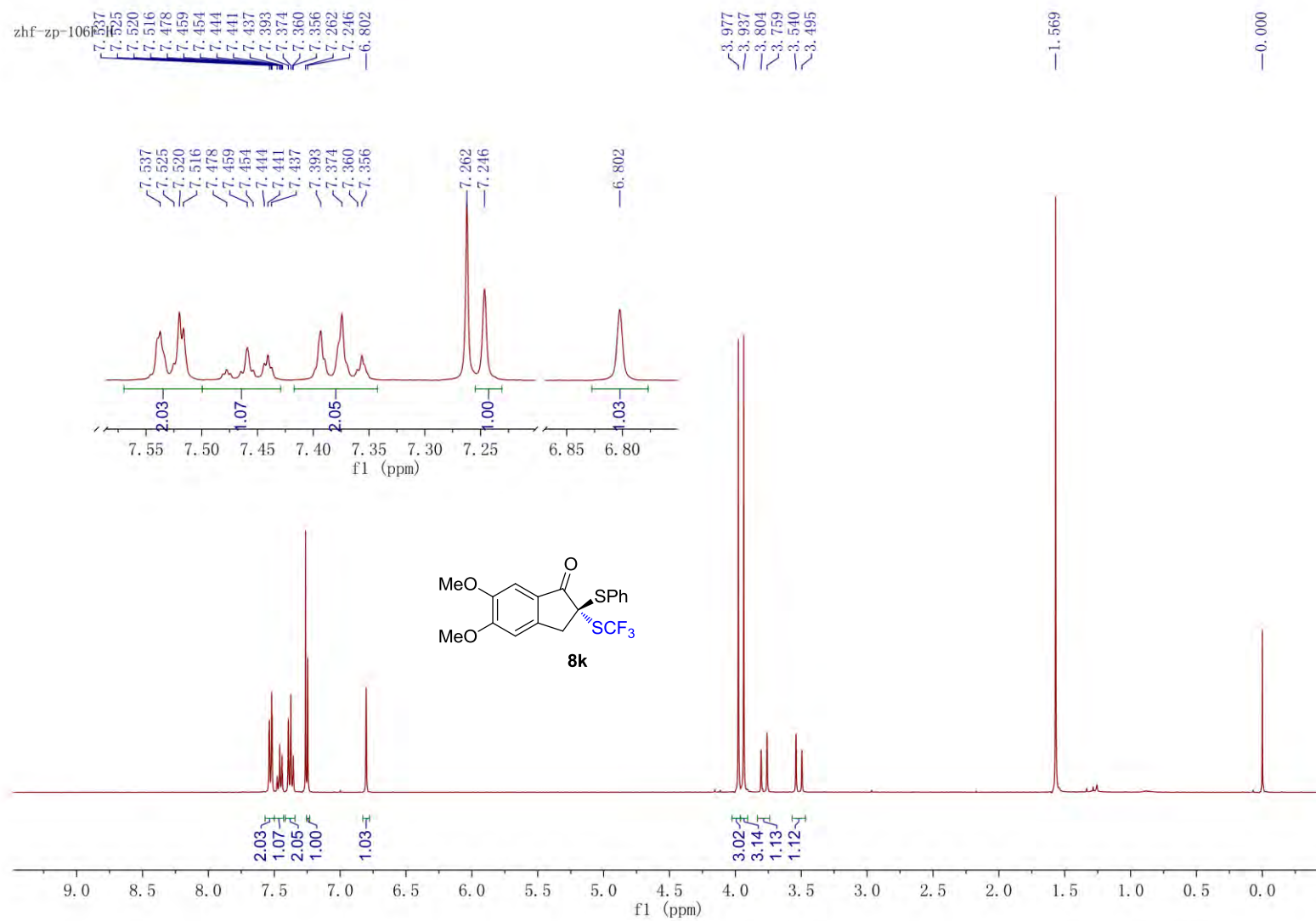
zhf-zp-106E C

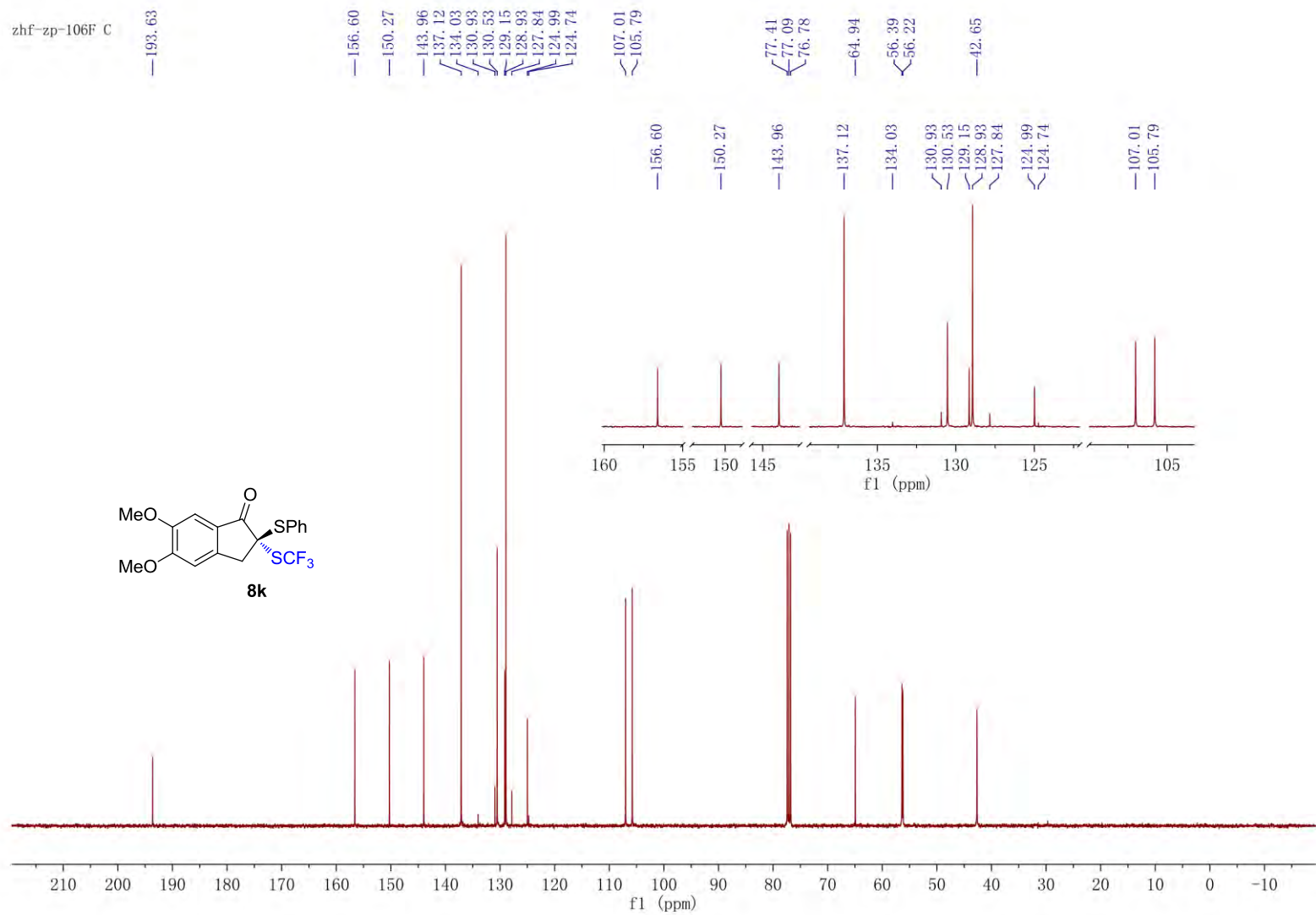


zhf-zp-106E F

-37.243

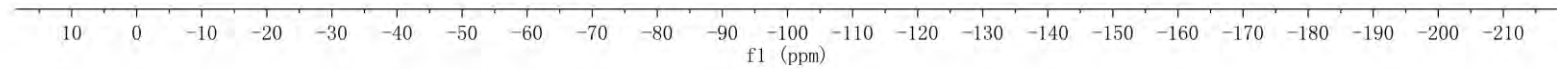
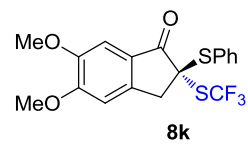


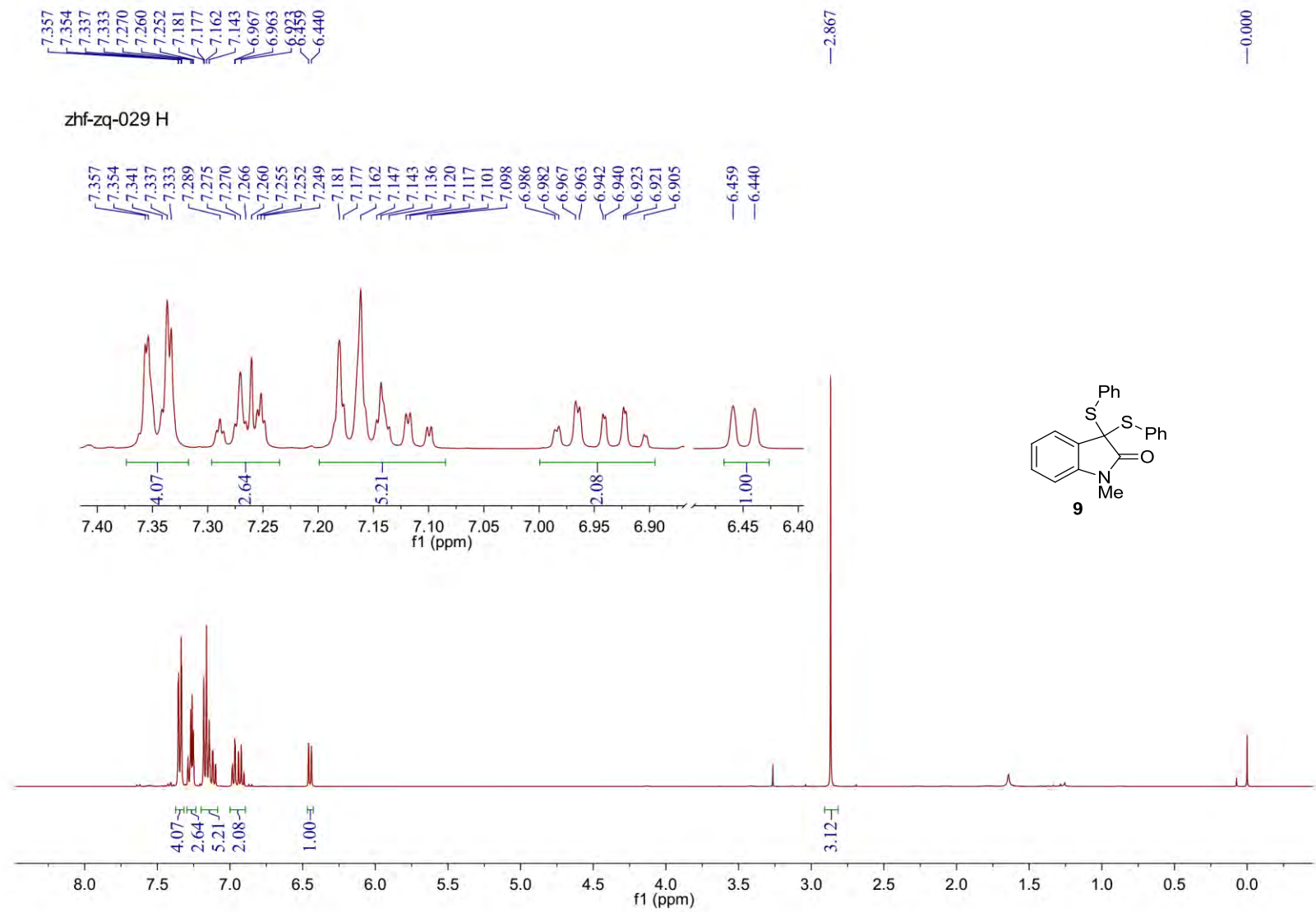




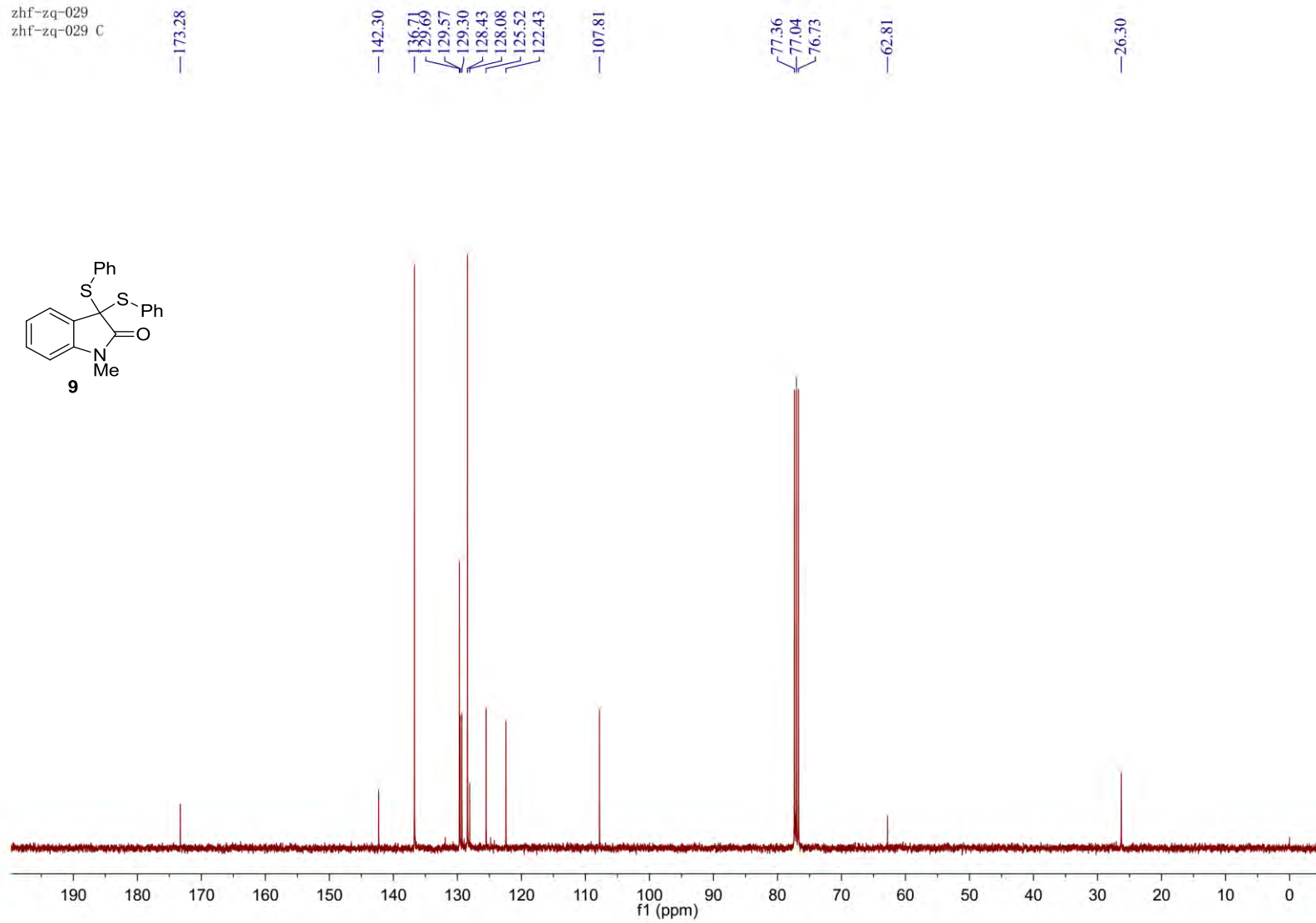
zhf-zp-106F F

— -37.504





zhf-zq-029
zhf-zq-029 C

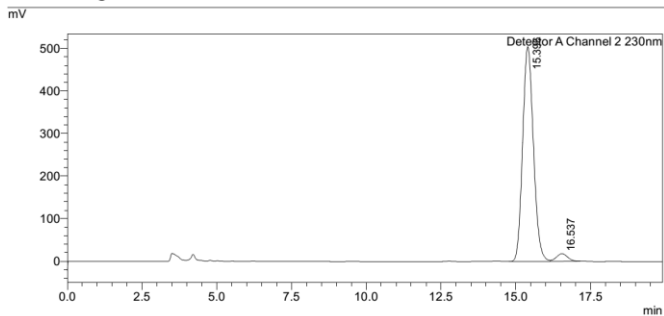


Analysis Report

<Sample Information>

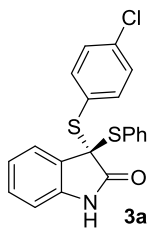
Sample Name : zhf-zp-011A
 Sample ID :
 Data Filename : zhf-zp-011A-asy-ADH-90-10-230-1.0.lcd
 Method Filename : RUN-1.0.lcm
 Batch Filename :
 Vial # : 1-1
 Injection Volume : 5 uL
 Date Acquired : 2015/4/27 15:03:59
 Date Processed : 2015/5/1 19:41:18
 Sample Type : Unknown
 Acquired by : System Administrator
 Processed by : System Administrator

<Chromatogram>



<Peak Table>

Peak#	Ret. Time	Area	Height	Conc.
1	15.396	12648667	504109	96.512
2	16.537	457088	17735	3.488
Total		13105756	521844	



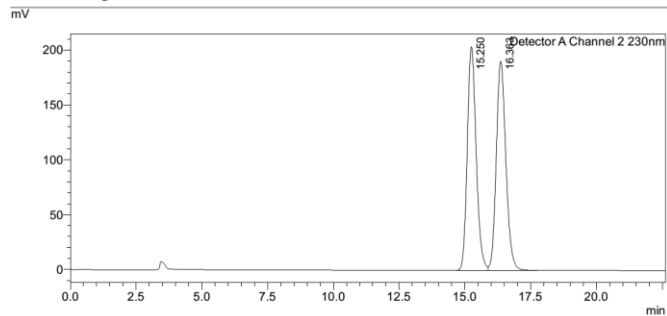
D:\Data\zhofeng\zhf-zp-011A-asy-ADH-90-10-230-1.0.lcd

Analysis Report

<Sample Information>

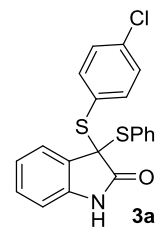
Sample Name : LIK-LA-021
 Sample ID :
 Data Filename : LIK-LA-021-rac-ADH-90-10-230-1.0.lcd
 Method Filename : RUN-1.0.lcm
 Batch Filename :
 Vial # : 1-1
 Injection Volume : 5 uL
 Date Acquired : 2015/4/27 15:48:55
 Date Processed : 2015/4/27 16:11:34
 Sample Type : Unknown
 Acquired by : System Administrator
 Processed by : System Administrator

<Chromatogram>



<Peak Table>

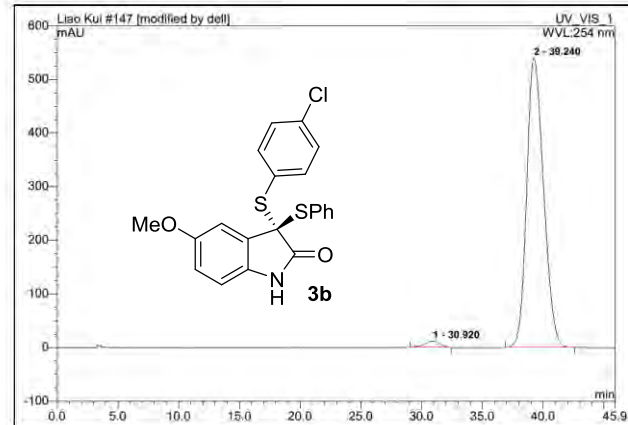
Peak#	Ret. Time	Area	Height	Conc.
1	15.250	4800165	203441	49.872
2	16.363	4824850	190159	50.128
Total		9625015	393601	



D:\Data\zhofeng\LIK-LA-021-rac-ADH-90-10-230-1.0.lcd

147 ZHF-ZP-28-ASY-OZH-97-3-254-1.0

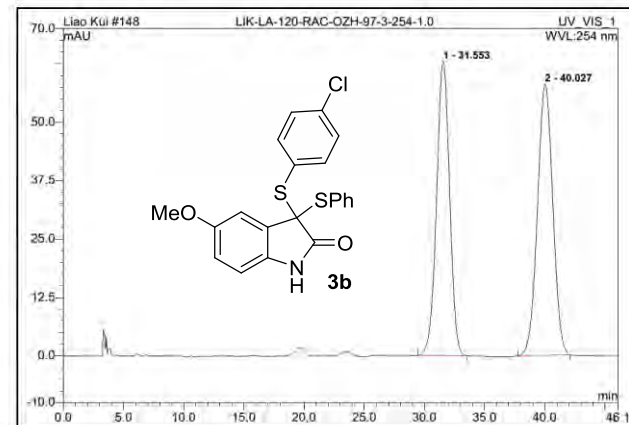
Sample Name:	ZHF-ZP-28-ASY-OZH-97-3-254-1.0	Injection Volume:	20.0
Vial Number:	245	Channel:	UV_VIS_1
Sample Type:	standard	Wavelength:	254
Control Program:	Liao Kui	Bandwidth:	n.a.
Quantif. Method:	Liao Kui	Dilution Factor:	1.0000
Recording Time:	2015-5-7 16:07	Sample Weight:	1.0000
Run Time (min):	45.92	Sample Amount:	1.0000



No.	Ret. Time min	Peak Name	Height mAU	Area mAU*min	Rel. Area %	Amount	Type
1	30.92	n.a.	11.518	16.741	1.89	n.a.	BMB*
2	39.24	n.a.	539.793	869.473	98.11	n.a.	BMB*
Total:			551.311	886.214	100.00	0.000	

148 LIK-LA-120-RAC-OZH-97-3-254-1.0

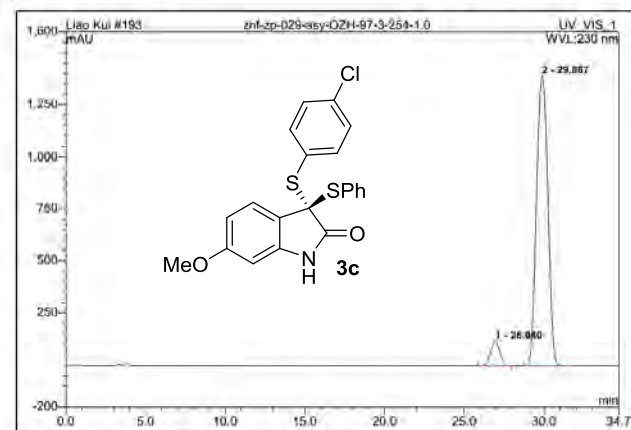
Sample Name:	LIK-LA-120-RAC-OZH-97-3-254-1.0	Injection Volume:	20.0
Vial Number:	246	Channel:	UV_VIS_1
Sample Type:	standard	Wavelength:	254
Control Program:	Liao Kui	Bandwidth:	n.a.
Quantif. Method:	Liao Kui	Dilution Factor:	1.0000
Recording Time:	2015-5-7 16:55	Sample Weight:	1.0000
Run Time (min):	46.09	Sample Amount:	1.0000



No.	Ret. Time min	Peak Name	Height mAU	Area mAU*min	Rel. Area %	Amount	Type
1	31.55	n.a.	62.897	79.213	48.01	n.a.	BMB
2	40.03	n.a.	57.942	85.785	51.99	n.a.	BMB
Total:			120.839	164.998	100.00	0.000	

193 zhf-zp-029-asy-OZH-97-3-254-1.0

Sample Name:	zhf-zp-029-asy-OZH-97-3-254-1.0	Injection Volume:	20.0
Vial Number:	291	Channel:	UV_VIS_1
Sample Type:	standard	Wavelength:	230
Control Program:	Liao Kui	Bandwidth:	n.a.
Quantif. Method:	Liao Kui	Dilution Factor:	1.0000
Recording Time:	2015-5-29 19:53	Sample Weight:	1.0000
Run Time (min):	34.69	Sample Amount:	1.0000



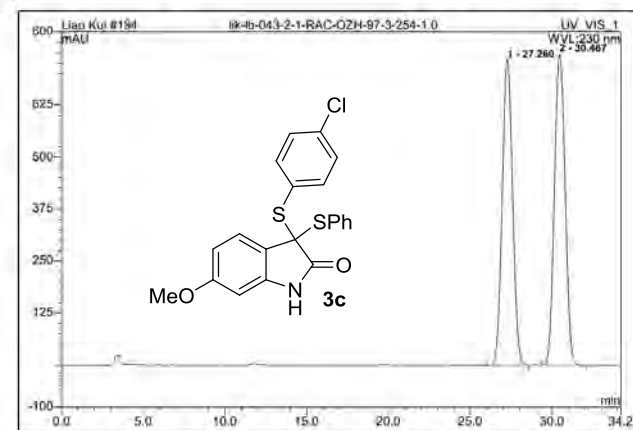
No.	Ret. Time min	Peak Name	Height mAU	Area mAU*min	Rel. Area %	Amount	Type
1	26.94	n.a.	115.776	77.925	6.35	n.a.	BMB
2	29.89	n.a.	1389.806	1148.396	93.65	n.a.	BMB
Total:			1505.583	1226.321	100.00	0.000	

default/Integration

Chromeleon (c) Dionex 1996-2006
Version 6.80 SRBa Build 2643 (158226)

194 lik-lb-043-2-1-RAC-OZH-97-3-254-1.0

Sample Name:	lik-lb-043-2-1-RAC-OZH-97-3-254-1.0	Injection Volume:	20.0
Vial Number:	292	Channel:	UV_VIS_1
Sample Type:	standard	Wavelength:	230
Control Program:	Liao Kui	Bandwidth:	n.a.
Quantif. Method:	Liao Kui	Dilution Factor:	1.0000
Recording Time:	2015-5-29 20:31	Sample Weight:	1.0000
Run Time (min):	34.21	Sample Amount:	1.0000



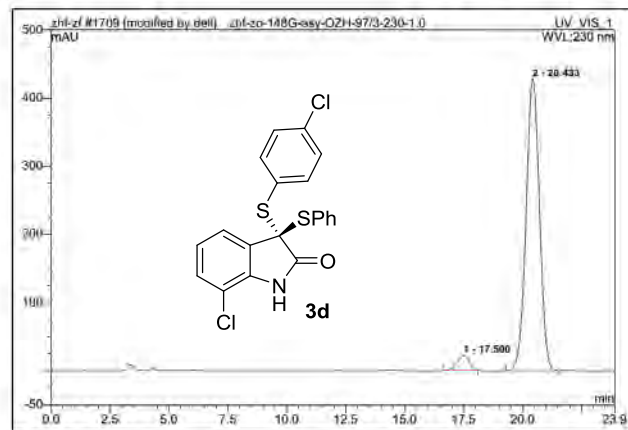
No.	Ret. Time min	Peak Name	Height mAU	Area mAU*min	Rel. Area %	Amount	Type
1	27.26	n.a.	732.877	539.876	48.76	n.a.	BMB
2	30.47	n.a.	745.651	567.273	51.24	n.a.	BMB
Total:			1478.528	1107.148	100.00	0.000	

default/Integration

Chromeleon (c) Dionex 1996-2006
Version 6.80 SRBa Build 2643 (158226)

1709 zhf-zo-148G-asy-OZH-97/3-230-1.0

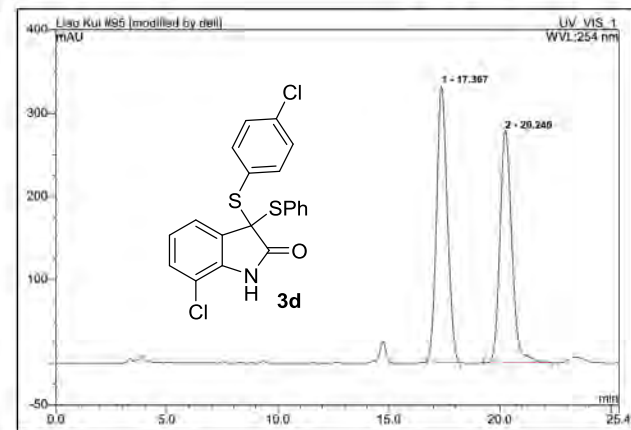
Sample Name:	zhf-zo-148G-asy-OZH-97/3-230-1.0	Injection Volume:	20.0
Vial Number:	816	Channel:	UV_VIS_1
Sample Type:	unknown	Wavelength:	230
Control Program:	zhf	Bandwidth:	n.a.
Quantif. Method:	zhf	Dilution Factor:	1.0000
Recording Time:	2015-4-27 14:15	Sample Weight:	1.0000
Run Time (min):	23.91	Sample Amount:	1.0000



No.	Ret. Time min	Peak Name	Height mAU	Area mAU*min	Rel. Area %	Amount	Type
1	17.50	n.a.	22.016	13.625	4.62	n.a.	BMB*
2	20.43	n.a.	427.902	281.258	95.38	n.a.	BMB*
Total:			449.918	294.883	100.00	0.000	

95 lik-la-122-rac-OZH-97-03-254-1.0

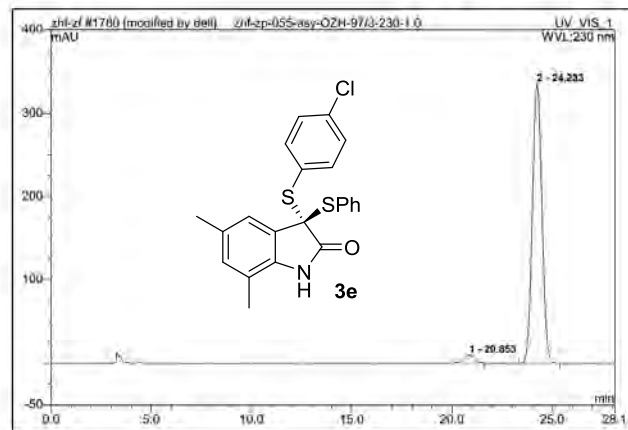
Sample Name:	lik-la-122-rac-OZH-97-03-254-1.0	Injection Volume:	20.0
Vial Number:	193	Channel:	UV_VIS_1
Sample Type:	standard	Wavelength:	254
Control Program:	Liao Kui	Bandwidth:	n.a.
Quantif. Method:	Liao Kui	Dilution Factor:	1.0000
Recording Time:	2015-4-8 14:40	Sample Weight:	1.0000
Run Time (min):	25.36	Sample Amount:	1.0000



No.	Ret. Time min	Peak Name	Height mAU	Area mAU*min	Rel. Area %	Amount	Type
1	17.37	n.a.	331.434	186.233	51.74	n.a.	BMB*
2	20.24	n.a.	278.096	173.719	48.26	n.a.	BMB*
Total:			609.530	359.952	100.00	0.000	

1780 zhf-zp-055-asy-OZH-97/3-230-1.0

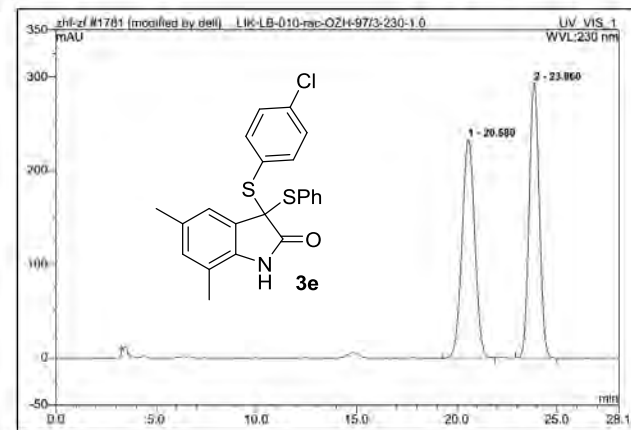
Sample Name:	zhf-zp-055-asy-OZH-97/3-230-1.0	Injection Volume:	20.0
Vial Number:	888	Channel:	UV_VIS_1
Sample Type:	unknown	Wavelength:	230
Control Program:	zhf	Bandwidth:	n.a.
Quantif. Method:	zhf	Dilution Factor:	1.0000
Recording Time:	2015-5-28 18:58	Sample Weight:	1.0000
Run Time (min):	28.11	Sample Amount:	1.0000



No.	Ret. Time min	Peak Name	Height mAU	Area mAU*min	Rel. Area %	Amount	Type
1	20.85	n.a.	9.541	6.548	3.44	n.a.	BMB ⁺
2	24.23	n.a.	334.457	184.060	96.56	n.a.	BMB ⁺
Total:			343.998	190.608	100.00	0.000	

1781 LIK-LB-010-rac-OZH-97/3-230-1.0

Sample Name:	LIK-LB-010-rac-OZH-97/3-230-1.0	Injection Volume:	20.0
Vial Number:	889	Channel:	UV_VIS_1
Sample Type:	unknown	Wavelength:	230
Control Program:	zhf	Bandwidth:	n.a.
Quantif. Method:	zhf	Dilution Factor:	1.0000
Recording Time:	2015-5-28 19:27	Sample Weight:	1.0000
Run Time (min):	28.09	Sample Amount:	1.0000



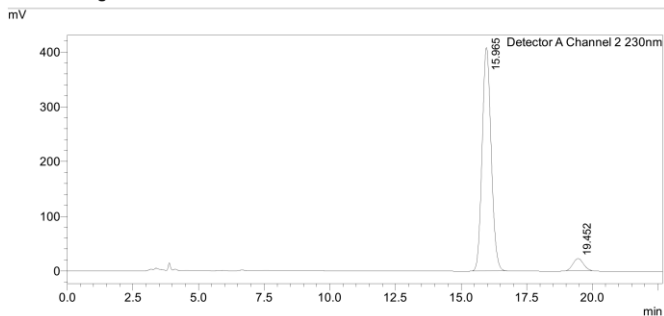
No.	Ret. Time min	Peak Name	Height mAU	Area mAU*min	Rel. Area %	Amount	Type
1	20.58	n.a.	233.228	171.544	49.94	n.a.	BMB
2	23.86	n.a.	293.772	171.984	50.06	n.a.	BMB
Total:			527.001	343.528	100.00	0.000	

Analysis Report

<Sample Information>

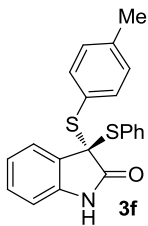
Sample Name : zh-f-zq-043
 Sample ID :
 Data Filename : zh-f-zq-043-asy-2-ADH-90-10-230-1.8.lcd
 Method Filename : zry.lcm
 Batch Filename :
 Vial # : 1-1
 Injection Volume : 10 µL
 Date Acquired : 2015/9/3 19:45:34
 Date Processed : 2015/9/4 11:13:50
 Sample Type : Unknown
 Acquired by : System Administrator
 Processed by : System Administrator

<Chromatogram>



<Peak Table>

Peak#	Ret. Time	Area	Height	Conc.
1	15.965	9726487	407794	94.144
2	19.452	604958	22043	5.856
Total		10331445	429836	



D:\Data\zhoufeng\zhf-zq-043-asy-2-ADH-90-10-230-1.8.lcd

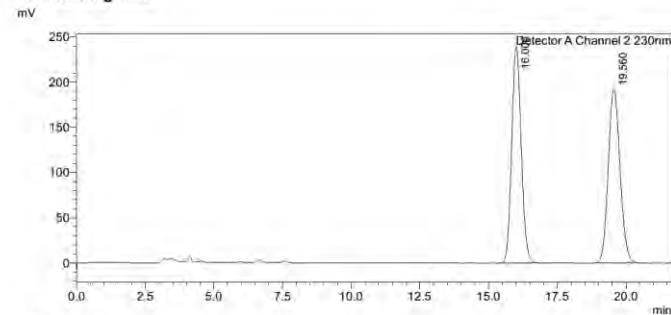


Analysis Report

<Sample Information>

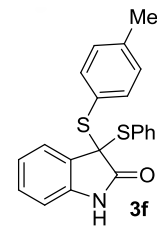
Sample Name : zh-f-zq-043
 Sample ID :
 Data Filename : zh-f-zq-043-rac-ADH-90-10-230-1.1.lcd
 Method Filename : zry.lcm
 Batch Filename :
 Vial # : 1-1
 Injection Volume : 10 µL
 Date Acquired : 2015/9/3 16:07:26
 Date Processed : 2015/9/4 11:14:26
 Sample Type : Unknown
 Acquired by : System Administrator
 Processed by : System Administrator

<Chromatogram>



<Peak Table>

Peak#	Ret. Time	Area	Height	Conc.	Unit	Mark	Name
1	16.006	5762079	239758	50.140		M	
2	19.560	5729797	192790	49.860		M	
Total		11491877	432548				



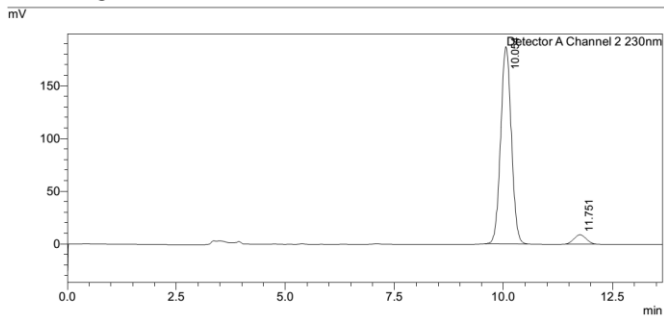
D:\Data\zhoufeng\zhf-zq-043-rac-ADH-90-10-230-1.1.lcd

Analysis Report

<Sample Information>

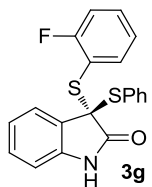
Sample Name : ZHF-ZQ-053
Sample ID :
Data Filename : zhf-zq-053-asy-OZH-90-10-230-1.0.lcd
Method Filename : chl-0128.lcm
Batch Filename :
Vial # : 1-1
Injection Volume : 10 uL
Date Acquired : 2015/9/6 13:51:09
Date Processed : 2015/9/7 9:26:48
Sample Type : Unknown
Acquired by : System Administrator
Processed by : System Administrator

<Chromatogram>



<Peak Table>

Peak#	Ret. Time	Area	Height	Conc.
1	10.054	3175326	186759	94.923
2	11.751	169841	8750	5.077
Total		3345167	195509	



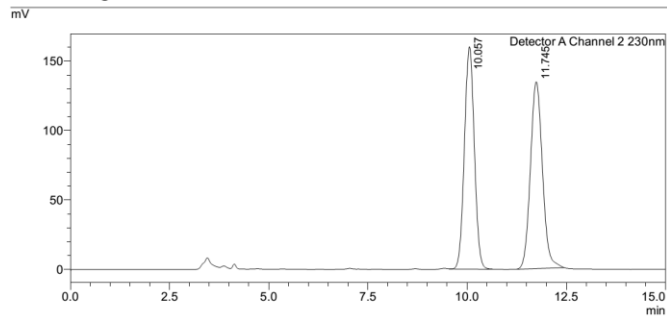
D:\Data\zhoufeng\zhf-zq-053-asy-OZH-90-10-230-1.0.lcd

Analysis Report

<Sample Information>

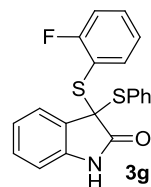
Sample Name : ZHF-ZQ-056A
Sample ID :
Data Filename : zhf-zq-056A-rac-OZH-90-10-230-1.0.lcd
Method Filename : chl-0128.lcm
Batch Filename :
Vial # : 1-1
Injection Volume : 10 uL
Date Acquired : 2015/9/6 14:05:36
Date Processed : 2015/9/7 9:29:00
Sample Type : Unknown
Acquired by : System Administrator
Processed by : System Administrator

<Chromatogram>



<Peak Table>

Peak#	Ret. Time	Area	Height	Conc.
1	10.057	2722740	160035	48.908
2	11.745	2844300	134440	51.092
Total		5567040	294475	



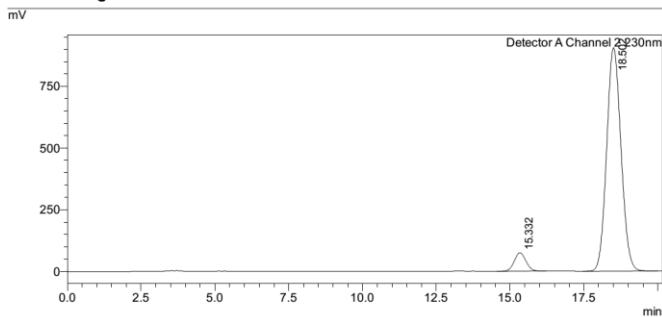
D:\Data\zhoufeng\zhf-zq-056A-rac-OZH-90-10-230-1.0.lcd

Analysis Report

<Sample Information>

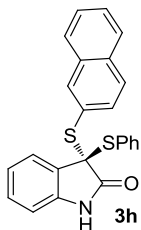
Sample Name : zhf-zp-71C-ASY-IC-90-10-230-1.0
 Sample ID :
 Data Filename : zhf-zp-71C-ASY-IC-90-10-230-1.0.lcd
 Method Filename : RUN-1.0.lcm
 Batch Filename :
 Vial # : 1-1
 Injection Volume : 10 uL
 Date Acquired : 2015/7/18 16:18:57
 Date Processed : 2015/7/18 17:15:24
 Sample Type : Unknown
 Acquired by : System Administrator
 Processed by : System Administrator

<Chromatogram>



<Peak Table>

Peak#	Ret. Time	Area	Height	Conc.
1	15.332	2054306	74595	6.289
2	18.502	30608564	904672	93.711
Total		32662870	979267	



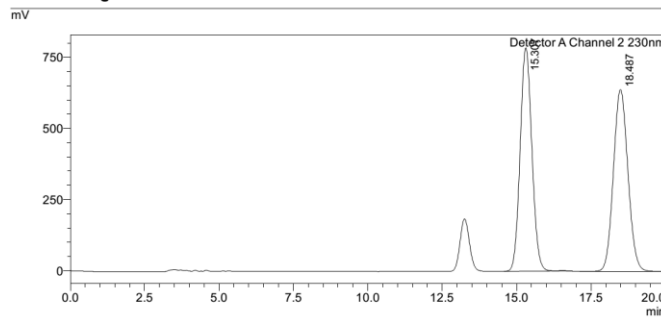
D:\Data\LiaoKui\zhf-zp-71C-ASY-IC-90-10-230-1.0.lcd

Analysis Report

<Sample Information>

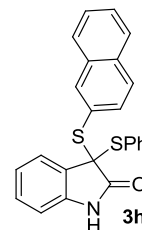
Sample Name : LIK-LB-091-7-RAC-IC-90-10-230-1.0
 Sample ID :
 Data Filename : LIK-LB-091-7-RAC-IC-90-10-230-1.0.lcd
 Method Filename : RUN-1.0.lcm
 Batch Filename :
 Vial # : 1-1
 Injection Volume : 10 uL
 Date Acquired : 2015/7/18 16:40:06
 Date Processed : 2015/7/18 22:29:27
 Sample Type : Unknown
 Acquired by : System Administrator
 Processed by : System Administrator

<Chromatogram>



<Peak Table>

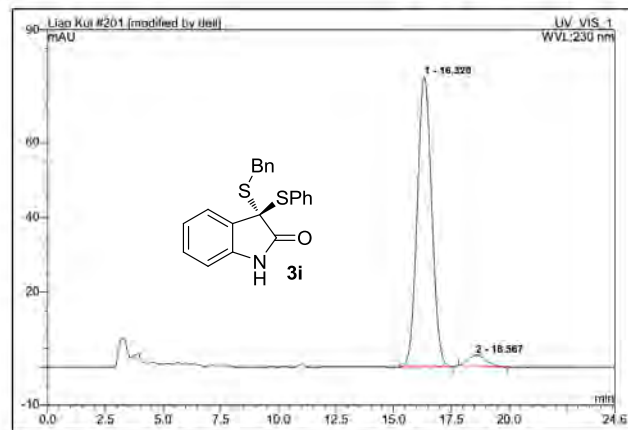
Peak#	Ret. Time	Area	Height	Conc.
1	15.307	21631578	784472	50.057
2	18.487	21582148	638126	49.943
Total		43213726	1422599	



D:\Data\LiaoKui\LIK-LB-091-7-RAC-IC-90-10-230-1.0.lcd

201 zhf-zp-056B-asy-ASH-80-20-254-1.0

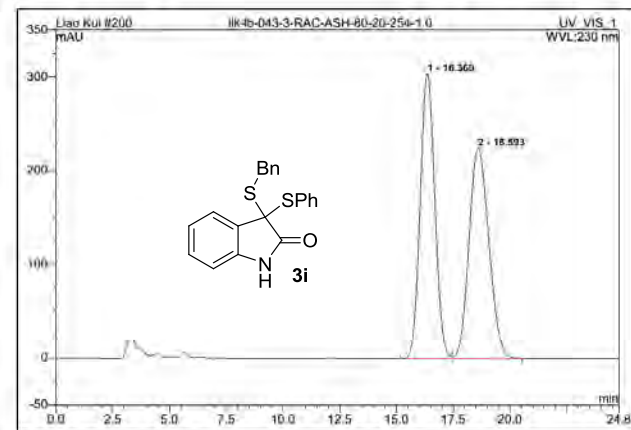
Sample Name:	zhf-zp-056B-asy-ASH-80-20-254-1.0	Injection Volume:	20.0
Vial Number:	299	Channel:	UV_VIS_1
Sample Type:	standard	Wavelength:	230
Control Program:	Liao Kui	Bandwidth:	n.a.
Quantif. Method:	Liao Kui	Dilution Factor:	1.0000
Recording Time:	2015-5-30 10:38	Sample Weight:	1.0000
Run Time (min):	24.58	Sample Amount:	1.0000



No.	Ret. Time min	Peak Name	Height mAU	Area mAU*min	Rel. Area %	Amount	Type
1	16.32	n.a.	77.309	56.202	95.58	n.a.	BMB
2	16.57	n.a.	2.947	2.598	4.42	n.a.	BMB*
Total:			80.256	58.800	100.00	0.000	

200 lik-lb-043-3-RAC-ASH-80-20-254-1.0

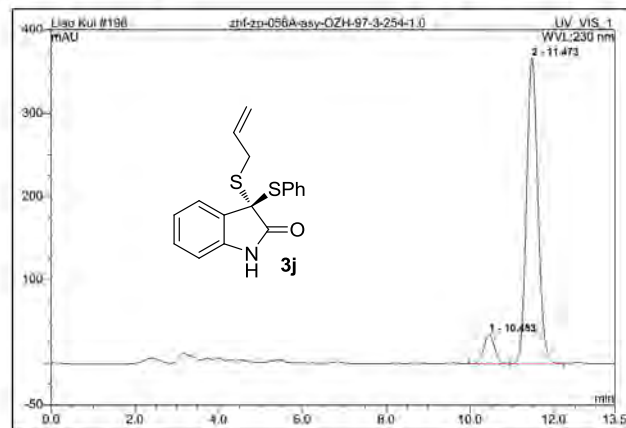
Sample Name:	lik-lb-043-3-RAC-ASH-80-20-254-1.0	Injection Volume:	20.0
Vial Number:	298	Channel:	UV_VIS_1
Sample Type:	standard	Wavelength:	230
Control Program:	Liao Kui	Bandwidth:	n.a.
Quantif. Method:	Liao Kui	Dilution Factor:	1.0000
Recording Time:	2015-5-30 10:12	Sample Weight:	1.0000
Run Time (min):	24.80	Sample Amount:	1.0000



No.	Ret. Time min	Peak Name	Height mAU	Area mAU*min	Rel. Area %	Amount	Type
1	16.36	n.a.	304.018	226.959	49.93	n.a.	BM
2	16.59	n.a.	224.443	227.629	50.07	n.a.	MB
Total:			528.462	454.588	100.00	0.000	

196 zhf-zp-056A-asy-OZH-97-3-254-1.0

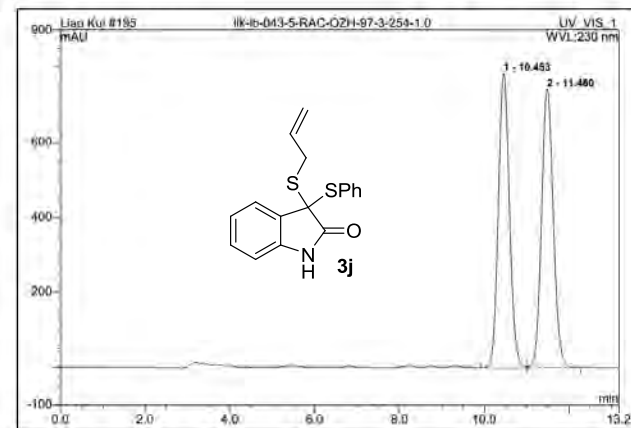
Sample Name:	zhf-zp-056A-asy-OZH-97-3-254-1.0	Injection Volume:	20.0
Vial Number:	294	Channel:	UV_VIS_1
Sample Type:	standard	Wavelength:	230
Control Program:	Liao Kui	Bandwidth:	n.a.
Quantif. Method:	Liao Kui	Dilution Factor:	1.0000
Recording Time:	2015-5-29 21:42	Sample Weight:	1.0000
Run Time (min):	13.45	Sample Amount:	1.0000



No.	Ret. Time min	Peak Name	Height mAU	Area mAU*min	Rel. Area %	Amount	Type
1	10.45	n.a.	35.852	10.501	8.34	n.a.	BMB
2	11.47	n.a.	366.514	115.408	91.66	n.a.	BMB
Total:			402.165	125.909	100.00	0.000	

195 lik-lb-043-5-RAC-OZH-97-3-254-1.0

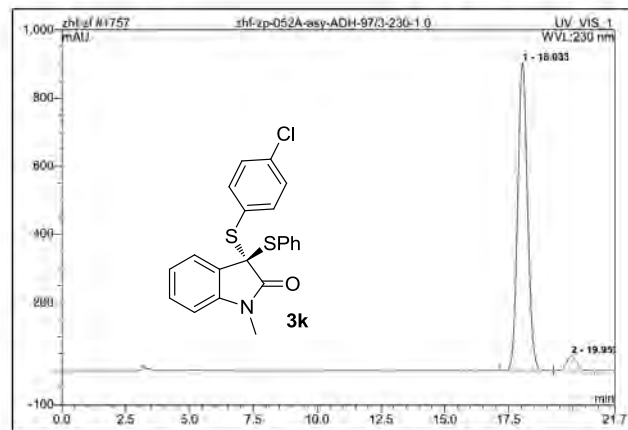
Sample Name:	lik-lb-043-5-RAC-OZH-97-3-254-1.0	Injection Volume:	20.0
Vial Number:	293	Channel:	UV_VIS_1
Sample Type:	standard	Wavelength:	230
Control Program:	Liao Kui	Bandwidth:	n.a.
Quantif. Method:	Liao Kui	Dilution Factor:	1.0000
Recording Time:	2015-5-29 21:26	Sample Weight:	1.0000
Run Time (min):	13.17	Sample Amount:	1.0000



No.	Ret. Time min	Peak Name	Height mAU	Area mAU*min	Rel. Area %	Amount	Type
1	10.45	n.a.	784.714	238.873	50.01	n.a.	BM
2	11.48	n.a.	743.434	238.819	49.99	n.a.	MB
Total:			1528.148	477.692	100.00	0.000	

1757 zhf-zp-052A-asy-ADH-97/3-230-1.0

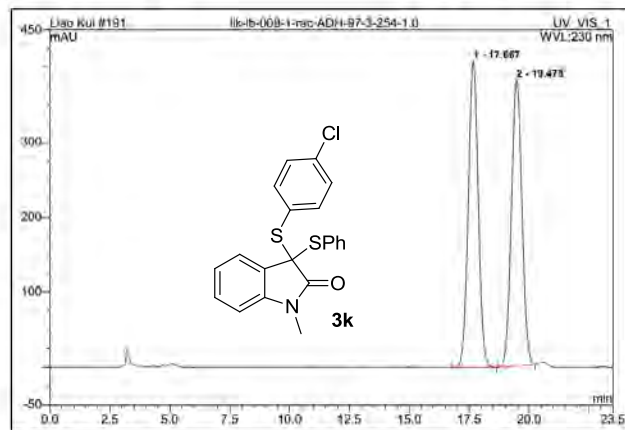
Sample Name:	zhf-zp-052A-asy-ADH-97/3-230-1.0	Injection Volume:	20.0
Vial Number:	865	Channel:	UV_VIS_1
Sample Type:	unknown	Wavelength:	230
Control Program:	zhf	Bandwidth:	n.a.
Quantif. Method:	zhf	Dilution Factor:	1.0000
Recording Time:	2015-5-15 14:03	Sample Weight:	1.0000
Run Time (min):	21.85	Sample Amount:	1.0000



No.	Ret. Time min	Peak Name	Height mAU	Area mAU*min	Rel. Area %	Amount	Type
1	18.03	n.a.	902.885	405.599	95.06	n.a.	BMB
2	19.95	n.a.	43.903	21.056	4.94	n.a.	BMB
Total:			946.588	426.654	100.00	0.000	

191 lik-lb-009-1-rac-ADH-97-3-254-1.0

Sample Name:	lik-lb-009-1-rac-ADH-97-3-254-1.0	Injection Volume:	20.0
Vial Number:	289	Channel:	UV_VIS_1
Sample Type:	standard	Wavelength:	230
Control Program:	Liao Kui	Bandwidth:	n.a.
Quantif. Method:	Liao Kui	Dilution Factor:	1.0000
Recording Time:	2015-5-29 14:24	Sample Weight:	1.0000
Run Time (min):	23.49	Sample Amount:	1.0000



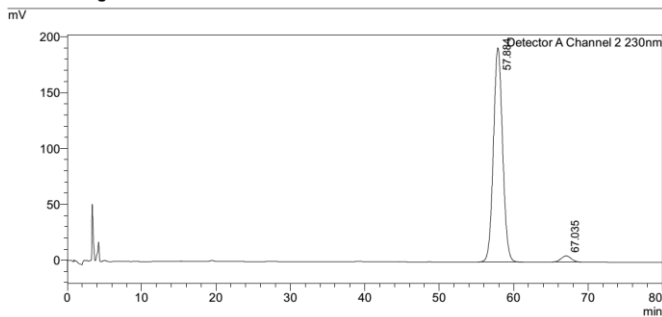
No.	Ret. Time min	Peak Name	Height mAU	Area mAU*min	Rel. Area %	Amount	Type
1	17.67	n.a.	408.591	197.741	50.40	n.a.	BMB
2	19.47	n.a.	381.953	194.606	49.60	n.a.	BMB
Total:			790.544	392.346	100.00	0.000	

Analysis Report

<Sample Information>

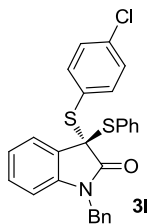
Sample Name : zhf-zp-052B-asy-ADH-97-3-254-1.0
 Sample ID :
 Data Filename : zhf-zp-052B-asy-ADH-97-3-254-1.0.lcd
 Method Filename : method 1.lcm
 Batch Filename :
 Vial # : 1-1
 Injection Volume : 10 uL
 Date Acquired : 2015/5/9 18:05:55
 Date Processed : 2015/5/9 21:57:27
 Sample Type : Unknown
 Acquired by : System Administrator
 Processed by : System Administrator

<Chromatogram>



<Peak Table>

Peak#	Ret. Time	Area	Height	Conc.
1	57.884	16512335	191854	96.940
2	67.035	521141	5525	3.060
Total		17033476	197379	



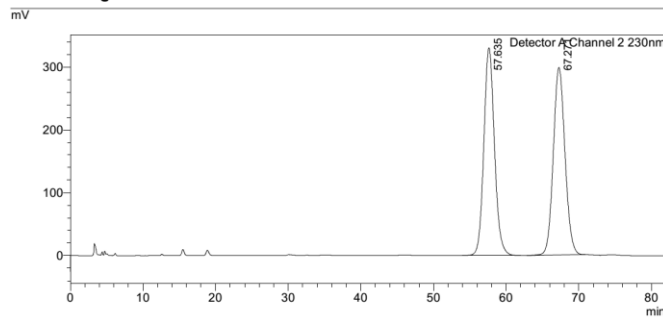
D:\Data\zhoufeng\zhf-zp-052B-asy-ADH-97-3-254-1.0.lcd

Analysis Report

<Sample Information>

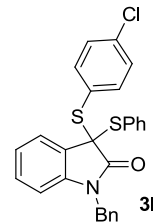
Sample Name : zhf-zp-032B-rac-ADH-97-3-254-1.0
 Sample ID :
 Data Filename : zhf-zp-032B-rac-ADH-97-3-254-1.0.lcd
 Method Filename : RUN-1.0.lcm
 Batch Filename :
 Vial # : 1-1
 Injection Volume : 10 uL
 Date Acquired : 2015/5/27 15:07:56
 Date Processed : 2015/6/9 10:57:01
 Sample Type : Unknown
 Acquired by : System Administrator
 Processed by : System Administrator

<Chromatogram>



<Peak Table>

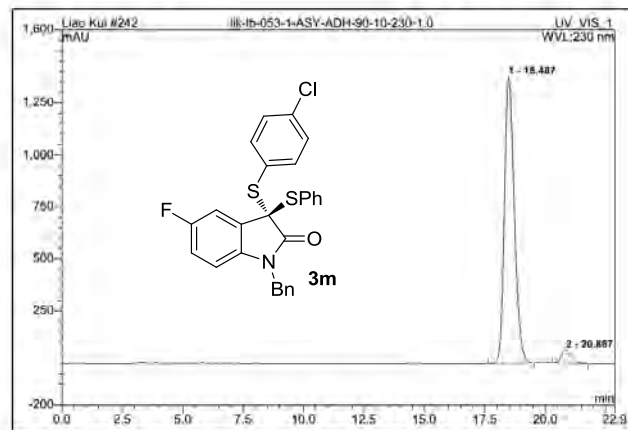
Peak#	Ret. Time	Area	Height	Conc.
1	57.635	33672023	330926	50.150
2	67.271	33470809	298530	49.850
Total		67142832	629456	



D:\Data\zhoufeng\zhf-zp-032B-rac-ADH-97-3-254-1.0.lcd

242 lik-lb-053-1-ASY-ADH-90-10-230-1.0

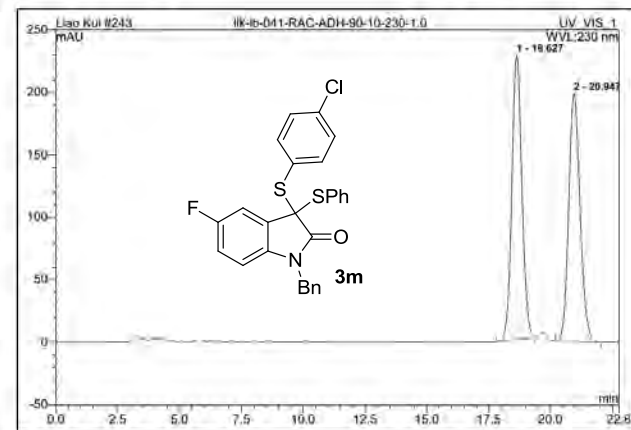
Sample Name:	lik-lb-053-1-ASY-ADH-90-10-230-1.0	Injection Volume:	20.0
Vial Number:	340	Channel:	UV_VIS_1
Sample Type:	standard	Wavelength:	230
Control Program:	Liao Kui	Bandwidth:	n.a.
Quantif. Method:	Liao Kui	Dilution Factor:	1.0000
Recording Time:	2015-7-14 16:34	Sample Weight:	1.0000
Run Time (min):	22.88	Sample Amount:	1.0000



No.	Ret. Time min	Peak Name	Height mAU	Area mAU*min	Rel. Area %	Amount	Type
1	18.49	n.a.	1373.894	852.059	95.47	n.a.	BMB
2	20.87	n.a.	60.703	30.923	4.53	n.a.	BMB
Total:			1434.397	882.983	100.00	0.000	

243 lik-lb-041-RAC-ADH-90-10-230-1.0

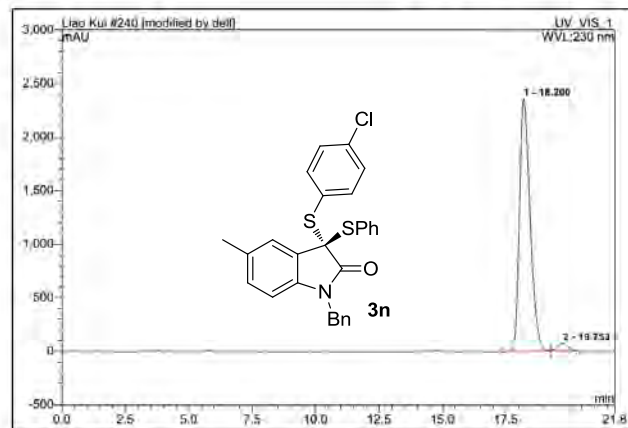
Sample Name:	lik-lb-041-RAC-ADH-90-10-230-1.0	Injection Volume:	20.0
Vial Number:	341	Channel:	UV_VIS_1
Sample Type:	standard	Wavelength:	230
Control Program:	Liao Kui	Bandwidth:	n.a.
Quantif. Method:	Liao Kui	Dilution Factor:	1.0000
Recording Time:	2015-7-14 16:58	Sample Weight:	1.0000
Run Time (min):	22.77	Sample Amount:	1.0000



No.	Ret. Time min	Peak Name	Height mAU	Area mAU*min	Rel. Area %	Amount	Type
1	18.63	n.a.	227.829	106.990	50.36	n.a.	BMB
2	20.95	n.a.	198.638	105.442	49.64	n.a.	BMB
Total:			426.468	212.432	100.00	0.000	

240 lik-lb-053--2--ASY-ADH-90-10-230-1.0

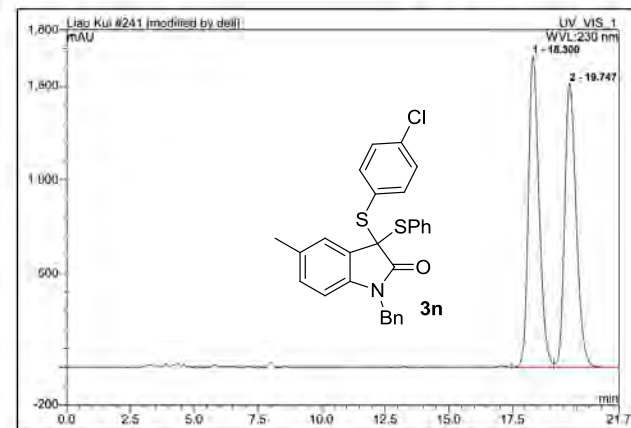
Sample Name:	lik-lb-053--2--ASY-ADH-90-10-230-1.0	Injection Volume:	20.0
Vial Number:	338	Channel:	UV_VIS_1
Sample Type:	standard	Wavelength:	230
Control Program:	Liao Kui	Bandwidth:	n.a.
Quantif. Method:	Liao Kui	Dilution Factor:	1.0000
Recording Time:	2015-7-14 15:47	Sample Weight:	1.0000
Run Time (min):	21.79	Sample Amount:	1.0000



No.	Ret. Time min	Peak Name	Height mAU	Area mAU*min	Rel. Area %	Amount	Type
1	18.20	n.a.	2358.988	1149.672	97.13	n.a.	BMB ⁺
2	19.75	n.a.	71.506	33.951	2.87	n.a.	BMB ⁺
Total:			2430.494	1183.624	100.00	0.000	

241 lik-lb-036-RAC-ADH-90-10-230-1.0

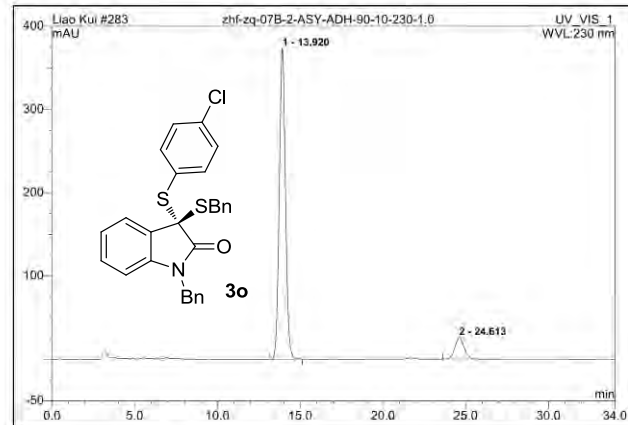
Sample Name:	lik-lb-036-RAC-ADH-90-10-230-1.0	Injection Volume:	20.0
Vial Number:	339	Channel:	UV_VIS_1
Sample Type:	standard	Wavelength:	230
Control Program:	Liao Kui	Bandwidth:	n.a.
Quantif. Method:	Liao Kui	Dilution Factor:	1.0000
Recording Time:	2015-7-14 16:11	Sample Weight:	1.0000
Run Time (min):	21.68	Sample Amount:	1.0000



No.	Ret. Time min	Peak Name	Height mAU	Area mAU*min	Rel. Area %	Amount	Type
1	18.30	n.a.	1660.024	812.019	50.52	n.a.	BM
2	19.75	n.a.	1514.865	795.303	49.48	n.a.	MB
Total:			3174.889	1607.322	100.00	0.000	

283 zh-f-zq-07B-2-ASY-ADH-90-10-230-1.0

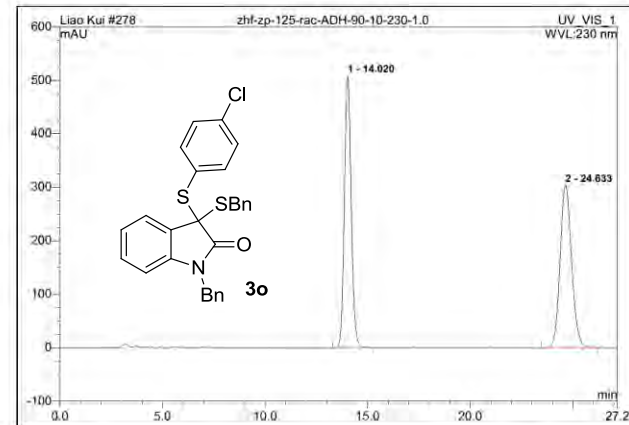
Sample Name:	zhf-zq-07B-2-ASY-ADH-90-10-230-1.0	Injection Volume:	20.0
Vial Number:	381	Channel:	UV_VIS_1
Sample Type:	standard	Wavelength:	230
Control Program:	Liao Kui	Bandwidth:	n.a.
Quantif. Method:	Liao Kui	Dilution Factor:	1.0000
Recording Time:	2015-7-22 17:33	Sample Weight:	1.0000
Run Time (min):	34.02	Sample Amount:	1.0000



No.	Ret. Time min	Peak Name	Height mAU	Area mAU*min	Rel. Area %	Amount	Type
1	13.92	n.a.	373.513	162.485	90.57	n.a.	BMB
2	24.61	n.a.	25.809	16.909	9.43	n.a.	BMB
Total:			399.322	179.394	100.00	0.000	

278 zhf-zp-125-rac-ADH-90-10-230-1.0

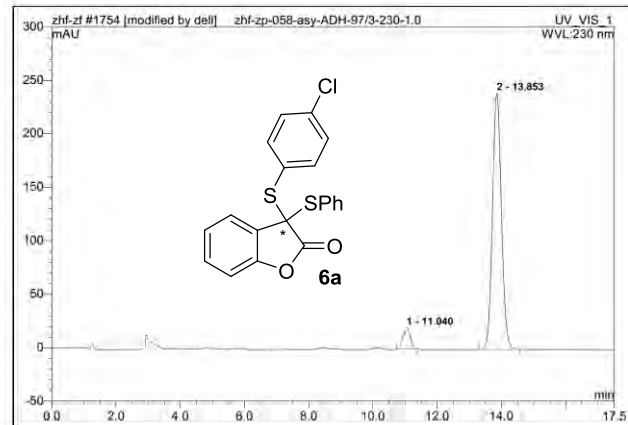
Sample Name:	zhf-zp-125-rac-ADH-90-10-230-1.0	Injection Volume:	20.0
Vial Number:	376	Channel:	UV_VIS_1
Sample Type:	standard	Wavelength:	230
Control Program:	Liao Kui	Bandwidth:	n.a.
Quantif. Method:	Liao Kui	Dilution Factor:	1.0000
Recording Time:	2015-7-22 15:42	Sample Weight:	1.0000
Run Time (min):	27.18	Sample Amount:	1.0000



No.	Ret. Time min	Peak Name	Height mAU	Area mAU*min	Rel. Area %	Amount	Type
1	14.02	n.a.	509.553	196.716	49.95	n.a.	BMB
2	24.63	n.a.	304.527	197.082	50.05	n.a.	BMB
Total:			814.080	393.798	100.00	0.000	

1754 zhf-zp-058-asy-ADH-97/3-230-1.0

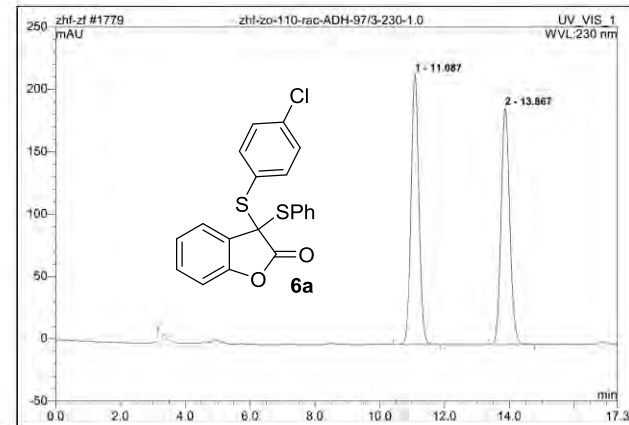
Sample Name:	zhf-zp-058-asy-ADH-97/3-230-1.0	Injection Volume:	20.0
Vial Number:	862	Channel:	UV_VIS_1
Sample Type:	unknown	Wavelength:	230
Control Program:	zhf	Bandwidth:	n.a.
Quantif. Method:	zhf	Dilution Factor:	1.0000
Recording Time:	2015-5-15 10:40	Sample Weight:	1.0000
Run Time (min):	17.52	Sample Amount:	1.0000



No.	Ret. Time min	Peak Name	Height mAU	Area mAU*min	Rel. Area %	Amount	Type
1	11.04	n.a.	19.938	5.360	6.52	n.a.	BMB*
2	13.85	n.a.	239.892	76.808	93.48	n.a.	BMB*
Total:			259.830	82.168	100.00	0.000	

1779 zhf-zo-110-rac-ADH-97/3-230-1.0

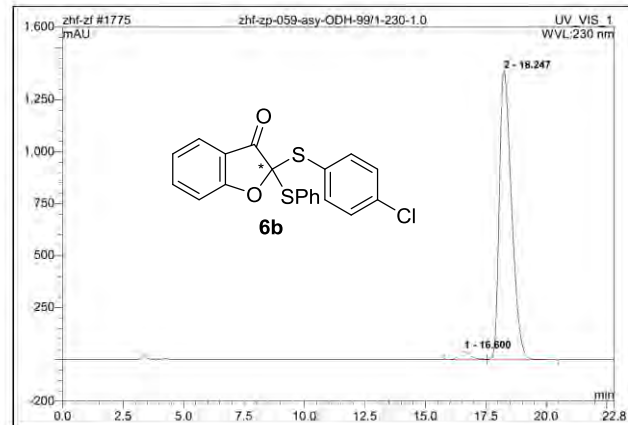
Sample Name:	zhf-zo-110-rac-ADH-97/3-230-1.0	Injection Volume:	20.0
Vial Number:	887	Channel:	UV_VIS_1
Sample Type:	unknown	Wavelength:	230
Control Program:	zhf	Bandwidth:	n.a.
Quantif. Method:	zhf	Dilution Factor:	1.0000
Recording Time:	2015-5-28 9:43	Sample Weight:	1.0000
Run Time (min):	17.34	Sample Amount:	1.0000



No.	Ret. Time min	Peak Name	Height mAU	Area mAU*min	Rel. Area %	Amount	Type
1	11.09	n.a.	216.655	60.716	50.78	n.a.	BMB
2	13.87	n.a.	189.643	58.847	49.22	n.a.	BMB
Total:			406.298	119.563	100.00	0.000	

1775 zhf-zp-059-asy-ODH-99/1-230-1.0

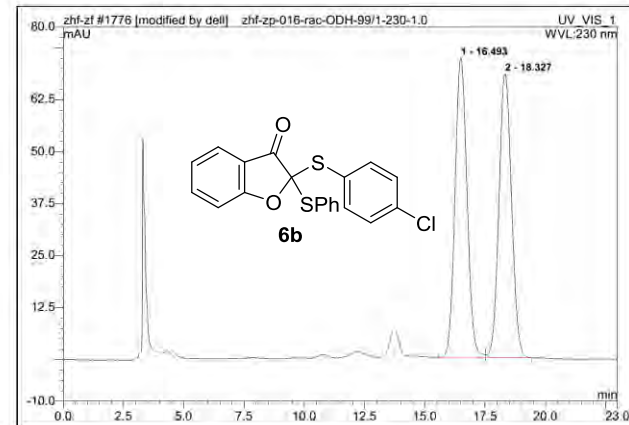
Sample Name:	zhf-zp-059-asy-ODH-99/1-230-1.0	Injection Volume:	20.0
Vial Number:	883	Channel:	UV_VIS_1
Sample Type:	unknown	Wavelength:	230
Control Program:	zhf	Bandwidth:	n.a.
Quantif. Method:	zhf	Dilution Factor:	1.0000
Recording Time:	2015-5-27 17:22	Sample Weight:	1.0000
Run Time (min):	22.82	Sample Amount:	1.0000



No.	Ret. Time min	Peak Name	Height mAU	Area mAU*min	Rel. Area %	Amount	Type
1	16.60	n.a.	40.155	22.489	2.69	n.a.	BM
2	18.25	n.a.	1388.672	813.396	97.31	n.a.	MB
Total:			1428.827	835.885	100.00	0.000	

1776 zhf-zp-016-rac-ODH-99/1-230-1.0

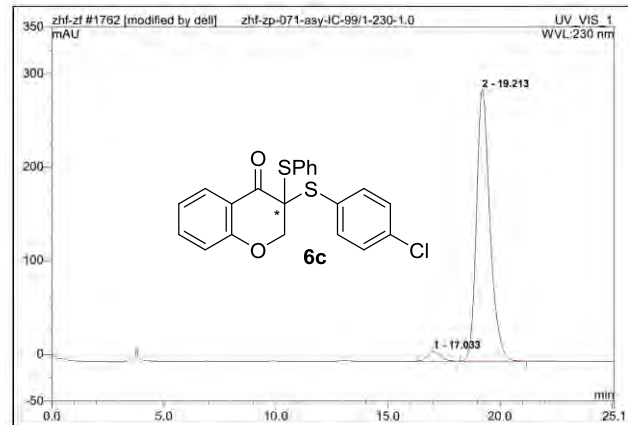
Sample Name:	zhf-zp-016-rac-ODH-99/1-230-1.0	Injection Volume:	20.0
Vial Number:	884	Channel:	UV_VIS_1
Sample Type:	unknown	Wavelength:	230
Control Program:	zhf	Bandwidth:	n.a.
Quantif. Method:	zhf	Dilution Factor:	1.0000
Recording Time:	2015-5-27 17:45	Sample Weight:	1.0000
Run Time (min):	23.00	Sample Amount:	1.0000



No.	Ret. Time min	Peak Name	Height mAU	Area mAU*min	Rel. Area %	Amount	Type
1	16.49	n.a.	72.057	41.507	50.09	n.a.	BM
2	18.33	n.a.	68.315	41.361	49.91	n.a.	MB
Total:			140.372	82.868	100.00	0.000	

1762 zhf-zp-071-asy-IC-99/1-230-1.0

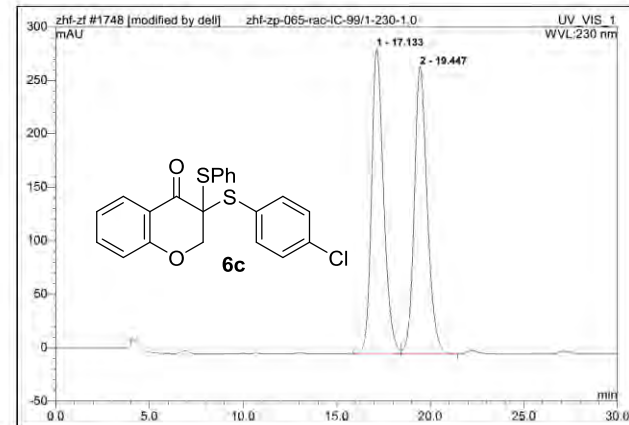
Sample Name:	zhf-zp-071-asy-IC-99/1-230-1.0	Injection Volume:	20.0
Vial Number:	870	Channel:	UV_VIS_1
Sample Type:	unknown	Wavelength:	230
Control Program:	zhf	Bandwidth:	n.a.
Quantif. Method:	zhf	Dilution Factor:	1.0000
Recording Time:	2015-5-15 17:15	Sample Weight:	1.0000
Run Time (min):	25.11	Sample Amount:	1.0000



No.	Ret. Time min	Peak Name	Height mAU	Area mAU*min	Rel. Area %	Amount	Type
1	17.03	n.a.	10.315	6.649	3.19	n.a.	BMB*
2	19.21	n.a.	289.840	201.927	96.81	n.a.	BMB*
Total:			300.155	208.577	100.00	0.000	

1748 zhf-zp-065-rac-IC-99/1-230-1.0

Sample Name:	zhf-zp-065-rac-IC-99/1-230-1.0	Injection Volume:	20.0
Vial Number:	856	Channel:	UV_VIS_1
Sample Type:	unknown	Wavelength:	230
Control Program:	zhf	Bandwidth:	n.a.
Quantif. Method:	zhf	Dilution Factor:	1.0000
Recording Time:	2015-5-12 18:18	Sample Weight:	1.0000
Run Time (min):	30.00	Sample Amount:	1.0000



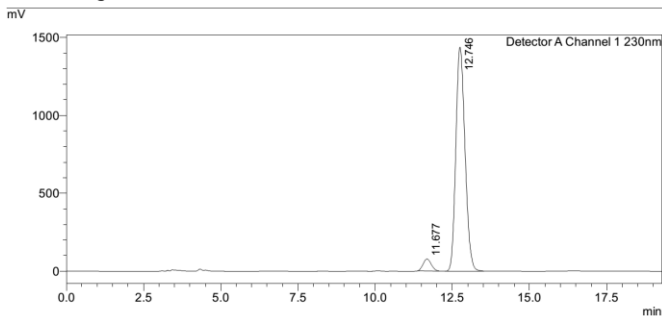
No.	Ret. Time min	Peak Name	Height mAU	Area mAU*min	Rel. Area %	Amount	Type
1	17.13	n.a.	285.620	213.798	49.89	n.a.	BM
2	19.45	n.a.	268.459	214.769	50.11	n.a.	MB
Total:			554.080	428.567	100.00	0.000	

Analysis Report

<Sample Information>

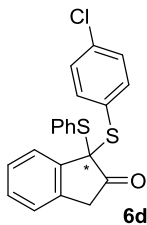
Sample Name : zhf-zq-021
 Sample ID :
 Data Filename : zhf-zq-021-1-asy-ODH-99-1-230-1.0.lcd
 Method Filename : 2.lcm
 Batch Filename :
 Vial # : 1-1
 Injection Volume : 10 uL
 Date Acquired : 2015/7/24 17:37:24
 Date Processed : 2015/7/25 10:16:50
 Sample Type : Unknown
 Acquired by : System Administrator
 Processed by : System Administrator

<Chromatogram>



<Peak Table>

Peak#	Ret. Time	Area	Height	Conc.
1	11.677	1402630	76587	4.503
2	12.746	29747260	1437389	95.497
Total		31149890	1513976	



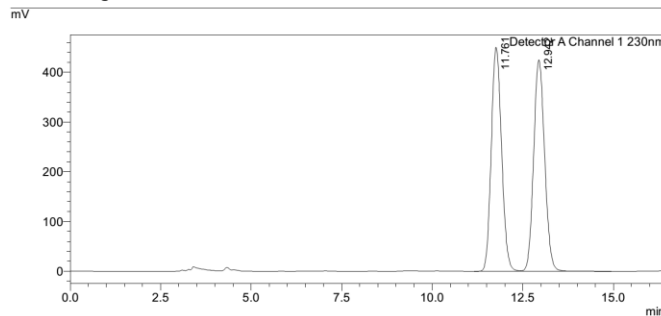
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Analysis Report

<Sample Information>

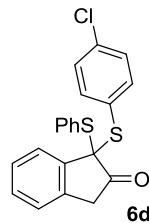
Sample Name : zhf-zq-021
 Sample ID :
 Data Filename : zhf-zq-021-rac-ODH-99-1-230-1.0.lcd
 Method Filename : 2.lcm
 Batch Filename :
 Vial # : 1-1
 Injection Volume : 10 uL
 Date Acquired : 2015/7/24 17:20:26
 Date Processed : 2015/7/24 17:38:11
 Sample Type : Unknown
 Acquired by : System Administrator
 Processed by : System Administrator

<Chromatogram>



<Peak Table>

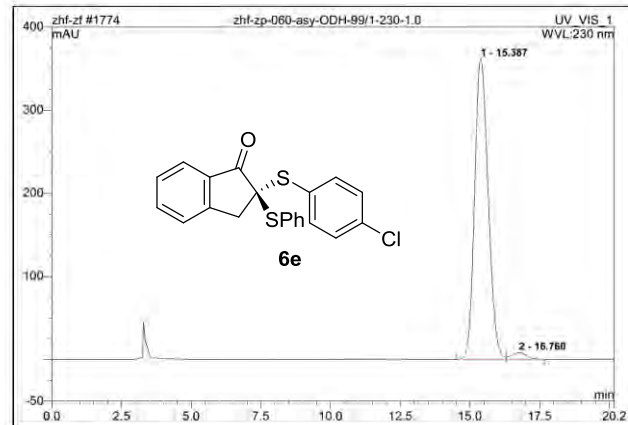
Peak#	Ret. Time	Area	Height	Conc.
1	11.761	8952886	450609	49.822
2	12.942	9016704	425387	50.178
Total		17969590	875996	



D:\Data\zhufeng\zhf-zq-021-rac-ODH-99-1-230-1.0.lcd

1774 zhf-zp-060-asy-ODH-99/1-230-1.0

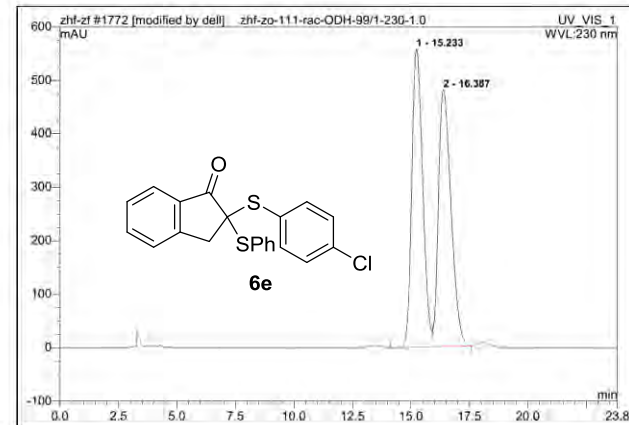
Sample Name:	zhf-zp-060-asy-ODH-99/1-230-1.0	Injection Volume:	20.0
Vial Number:	882	Channel:	UV_VIS_1
Sample Type:	unknown	Wavelength:	230
Control Program:	zhf	Bandwidth:	n.a.
Quantif. Method:	zhf	Dilution Factor:	1.0000
Recording Time:	2015-5-27 17:00	Sample Weight:	1.0000
Run Time (min):	20.19	Sample Amount:	1.0000



No.	Ret. Time min	Peak Name	Height mAU	Area mAU*min	Rel. Area %	Amount	Type
1	15.39	n.a.	361.704	203.477	97.56	n.a.	BM
2	16.76	n.a.	8.047	5.091	2.44	n.a.	MB
Total:			369.750	208.568	100.00	0.000	

1772 zhf-zo-111-rac-ODH-99/1-230-1.0

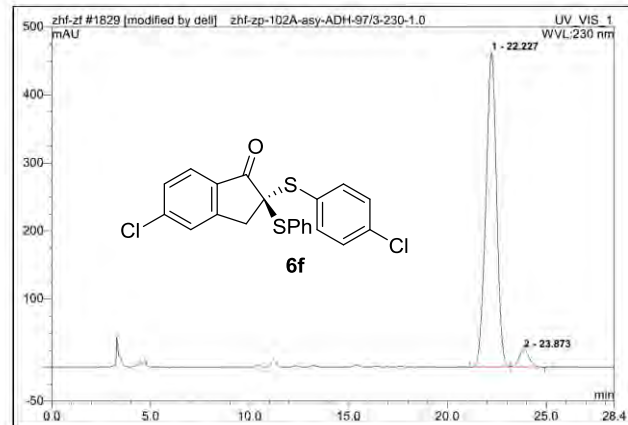
Sample Name:	zhf-zo-111-rac-ODH-99/1-230-1.0	Injection Volume:	20.0
Vial Number:	880	Channel:	UV_VIS_1
Sample Type:	unknown	Wavelength:	230
Control Program:	zhf	Bandwidth:	n.a.
Quantif. Method:	zhf	Dilution Factor:	1.0000
Recording Time:	2015-5-27 16:01	Sample Weight:	1.0000
Run Time (min):	23.83	Sample Amount:	1.0000



No.	Ret. Time min	Peak Name	Height mAU	Area mAU*min	Rel. Area %	Amount	Type
1	15.23	n.a.	557.506	299.879	50.05	n.a.	BM *
2	16.39	n.a.	480.002	299.241	49.95	n.a.	MB *
Total:			1037.508	599.120	100.00	0.000	

1829 zhf-zp-102A-asy-ADH-97/3-230-1.0

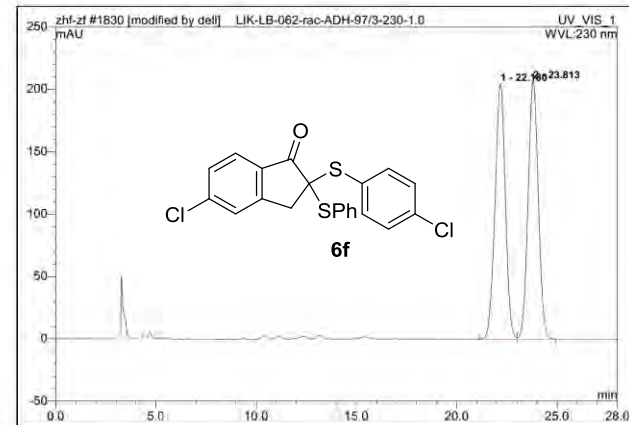
Sample Name:	zhf-zp-102A-asy-ADH-97/3-230-1.0	Injection Volume:	20.0
Vial Number:	938	Channel:	UV_VIS_1
Sample Type:	unknown	Wavelength:	230
Control Program:	zhf	Bandwidth:	n.a.
Quantif. Method:	zhf	Dilution Factor:	1.0000
Recording Time:	2015-6-23 19:01	Sample Weight:	1.0000
Run Time (min):	28.45	Sample Amount:	1.0000



No.	Ret. Time min	Peak Name	Height mAU	Area mAU*min	Rel. Area %	Amount	Type
1	22.23	n.a.	462.502	290.101	94.94	n.a.	BM
2	23.87	n.a.	25.105	15.458	5.06	n.a.	MB
Total:			487.607	305.559	100.00	0.000	

1830 LIK-LB-062-rac-ADH-97/3-230-1.0

Sample Name:	LIK-LB-062-rac-ADH-97/3-230-1.0	Injection Volume:	20.0
Vial Number:	939	Channel:	UV_VIS_1
Sample Type:	unknown	Wavelength:	230
Control Program:	zhf	Bandwidth:	n.a.
Quantif. Method:	zhf	Dilution Factor:	1.0000
Recording Time:	2015-6-23 19:32	Sample Weight:	1.0000
Run Time (min):	28.03	Sample Amount:	1.0000



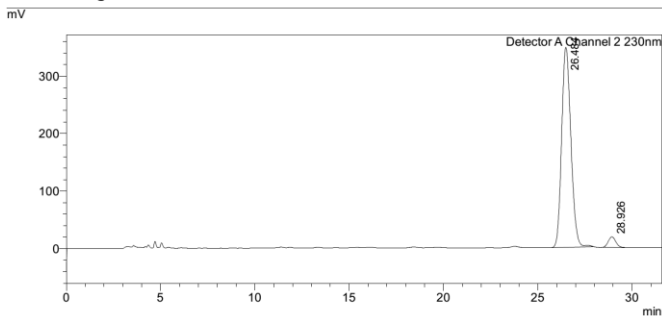
No.	Ret. Time min	Peak Name	Height mAU	Area mAU*min	Rel. Area %	Amount	Type
1	22.18	n.a.	204.803	122.356	50.02	n.a.	BM
2	23.81	n.a.	206.512	122.279	49.98	n.a.	MB
Total:			411.316	244.635	100.00	0.000	

Analysis Report

<Sample Information>

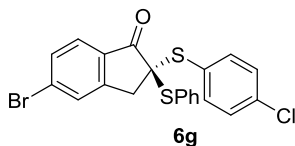
Sample Name : zhf-zp-102B-ASY-ADH-97-3-230-1.0
 Sample ID :
 Data Filename : zhf-zp-102B-ASY-ADH-97-3-230-1.0.lcd
 Method Filename : chl-0128.lcm
 Batch Filename :
 Vial # : 1-1
 Injection Volume : 10 uL
 Date Acquired : 2015/7/27 12:51:30
 Date Processed : 2015/7/27 16:26:49
 Sample Type : Unknown
 Acquired by : System Administrator
 Processed by : System Administrator

<Chromatogram>



<Peak Table>

Peak#	Ret. Time	Area	Height	Conc.
1	26.484	11846569	347614	95.938
2	28.926	501642	18234	4.062
Total		12348211	365848	



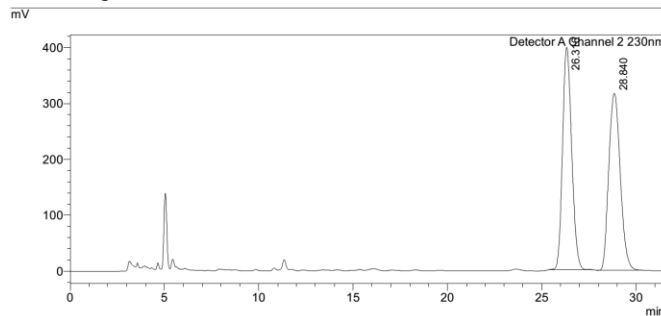
D:\Data\LiaoKui\zhf-zp-102B-ASY-ADH-97-3-230-1.0.lcd

Analysis Report

<Sample Information>

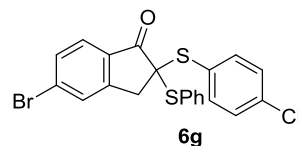
Sample Name : LIK-LB-063-RAC-ADH-97-3-230-1.0
 Sample ID :
 Data Filename : LIK-LB-063-RAC-ADH-97-3-230-1.0.lcd
 Method Filename : chl-0128.lcm
 Batch Filename :
 Vial # : 1-1
 Injection Volume : 10 uL
 Date Acquired : 2015/7/27 13:28:16
 Date Processed : 2015/7/27 16:27:54
 Sample Type : Unknown
 Acquired by : System Administrator
 Processed by : System Administrator

<Chromatogram>



<Peak Table>

Peak#	Ret. Time	Area	Height	Conc.
1	26.316	13481467	398299	50.124
2	28.840	13415018	316822	49.876
Total		26896485	715121	



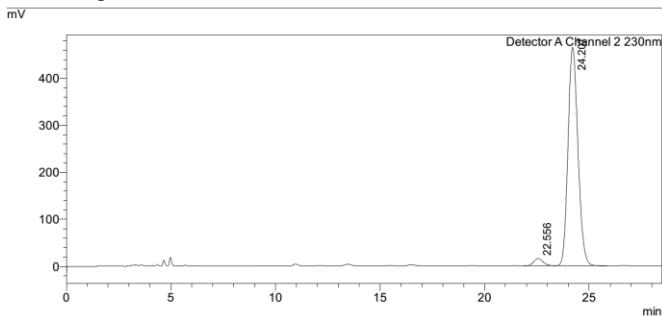
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Analysis Report

<Sample Information>

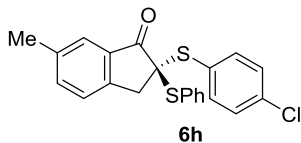
Sample Name : zhf-zp-102C-ASY-ADH-97-3-230-1.0
 Sample ID :
 Data Filename : zhf-zp-102C-ASY-ADH-97-3-230-1.0.lcd
 Method Filename : RUN-1.0.lcm
 Batch Filename :
 Vial # : 1-1
 Injection Volume : 10 uL
 Date Acquired : 2015/7/18 9:32:13
 Date Processed : 2015/7/18 22:35:09
 Sample Type : Unknown
 Acquired by : System Administrator
 Processed by : System Administrator

<Chromatogram>



<Peak Table>

Peak#	Ret. Time	Area	Height	Conc.
1	22.556	487785	15348	3.069
2	24.207	15407457	464695	96.931
Total		15895242	480043	



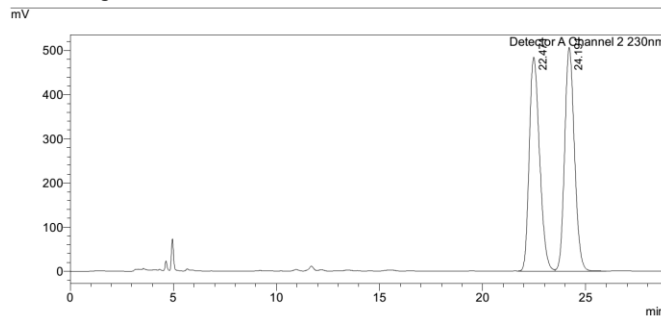
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Analysis Report

<Sample Information>

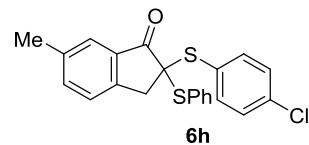
Sample Name : LIK-LB-091-1-RACADH-97-3-230-1.0
 Sample ID :
 Data Filename : LIK-LB-091-1-RACADH-97-3-230-1.0.lcd
 Method Filename : RUN-1.0.lcm
 Batch Filename :
 Vial # : 1-1
 Injection Volume : 10 uL
 Date Acquired : 2015/7/18 10:02:03
 Date Processed : 2015/7/18 22:36:02
 Sample Type : Unknown
 Acquired by : System Administrator
 Processed by : System Administrator

<Chromatogram>



<Peak Table>

Peak#	Ret. Time	Area	Height	Conc.
1	22.471	16852937	484134	50.178
2	24.191	16733333	506547	49.822
Total		33586269	990681	



D:\Data\LiaoKui\LIK-LB-091-1-RACADH-97-3-230-1.0.lcd

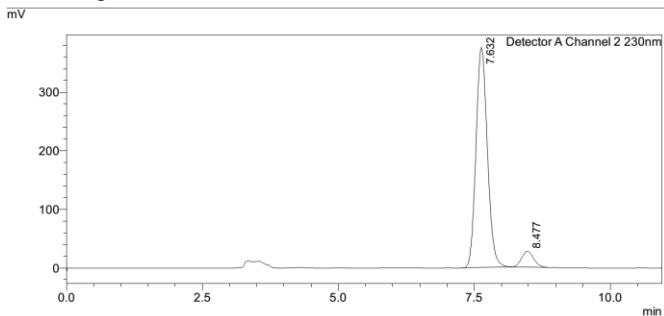
Analysis Report

<Sample Information>

Sample Name : ZHF-ZP-083
 Sample ID :
 Data Filename : zh-f-zp-083-asy-ODH-95-5-230-1.0.lcd
 Method Filename : method 1.lcm
 Batch Filename :
 Vial # : 1-1
 Injection Volume : 10 uL
 Date Acquired : 2015/5/29 11:36:35
 Date Processed : 2015/7/22 16:15:23

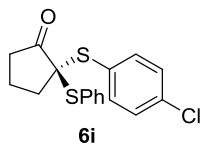
Sample Type : Unknown
 Acquired by : System Administrator
 Processed by : System Administrator

<Chromatogram>



<Peak Table>

Peak#	Ret. Time	Area	Height	Conc.
1	7.632	5478511	375916	92.941
2	8.477	416072	27078	7.059
Total		5894583	402993	



D:\Data\zhofeng\zhf-zp-083-asy-ODH-95-5-230-1.0.lcd

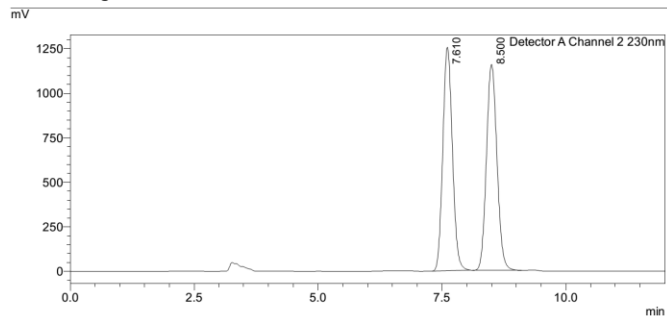
Analysis Report

<Sample Information>

Sample Name : zh-f-zp-076A
 Sample ID :
 Data Filename : zh-f-zp-076A-rac-ODH-95-5-230-1.0.lcd
 Method Filename : chl-0128.lcm
 Batch Filename :
 Vial # : 1-1
 Injection Volume : 10 uL
 Date Acquired : 2015/5/22 10:52:41
 Date Processed : 2015/7/14 9:43:38

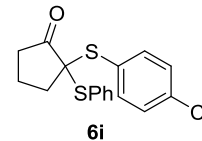
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 Acquired by : System Administrator
 Processed by : System Administrator

<Chromatogram>



<Peak Table>

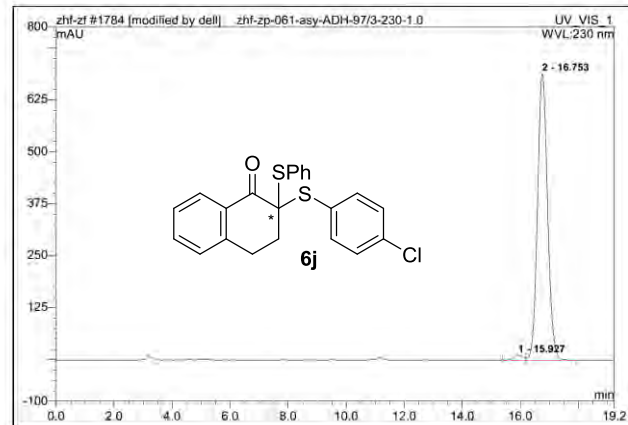
Peak#	Ret. Time	Area	Height	Conc.
1	7.610	16925253	1256144	49.726
2	8.500	17111773	1158380	50.274
Total		34037027	2414523	



D:\Data\zhofeng\zhf-zp-076A-rac-ODH-95-5-230-1.0.lcd

1784 zh-f-zp-061-asy-ADH-97/3-230-1.0

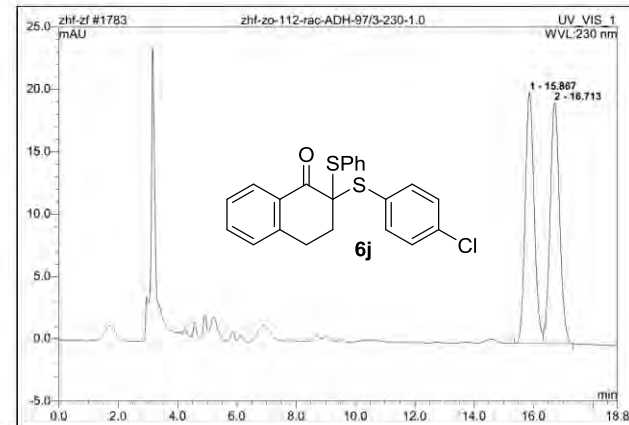
Sample Name:	zhf-zp-061-asy-ADH-97/3-230-1.0	Injection Volume:	20.0
Vial Number:	892	Channel:	UV_VIS_1
Sample Type:	unknown	Wavelength:	230
Control Program:	zhf	Bandwidth:	n.a.
Quantif. Method:	zhf	Dilution Factor:	1.0000
Recording Time:	2015-5-29 9:20	Sample Weight:	1.0000
Run Time (min):	19.25	Sample Amount:	1.0000



No.	Ret. Time min	Peak Name	Height mAU	Area mAU*min	Rel. Area %	Amount	Type
1	15.93	n.a.	11.813	4.299	1.53	n.a.	BM
2	16.75	n.a.	688.598	276.210	98.47	n.a.	MB
Total:			700.411	280.509	100.00	0.000	

1783 zh-f-zo-112-rac-ADH-97/3-230-1.0

Sample Name:	zhf-zo-112-rac-ADH-97/3-230-1.0	Injection Volume:	20.0
Vial Number:	891	Channel:	UV_VIS_1
Sample Type:	unknown	Wavelength:	230
Control Program:	zhf	Bandwidth:	n.a.
Quantif. Method:	zhf	Dilution Factor:	1.0000
Recording Time:	2015-5-29 8:55	Sample Weight:	1.0000
Run Time (min):	18.84	Sample Amount:	1.0000



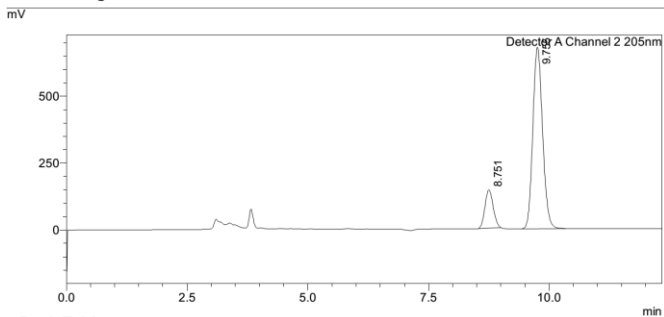
No.	Ret. Time min	Peak Name	Height mAU	Area mAU*min	Rel. Area %	Amount	Type
1	15.87	n.a.	20.134	7.088	49.77	n.a.	BM
2	16.71	n.a.	19.300	7.153	50.23	n.a.	MB
Total:			39.434	14.241	100.00	0.000	

Analysis Report

<Sample Information>

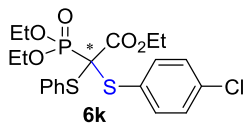
Sample Name : zhf-zq-020
 Sample ID :
 Data Filename : zhf-zq-020-asy-ADH-90-10-230-1.0.lcd
 Method Filename : 2.lcm
 Batch Filename :
 Vial # : 1-1
 Injection Volume : 10 uL
 Date Acquired : 2015/7/24 12:43:52
 Date Processed : 2015/7/24 13:13:11
 Sample Type : Unknown
 Acquired by : System Administrator
 Processed by : System Administrator

<Chromatogram>



<Peak Table>

Peak#	Ret. Time	Area	Height	Conc.
1	8.751	1685426	143049	15.045
2	9.755	9517482	678677	84.955
Total		11202907	821726	



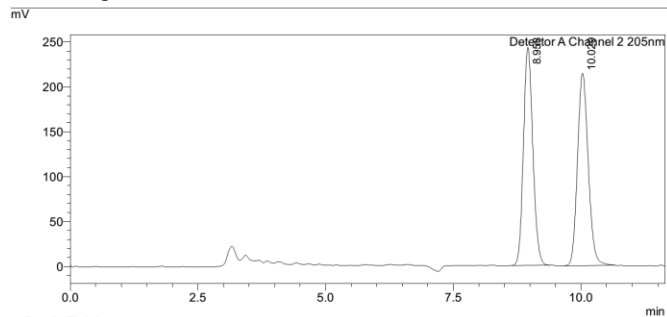
D:\Data\zhoufeng\zhf-zq-020-asy-ADH-90-10-230-1.0.lcd

Analysis Report

<Sample Information>

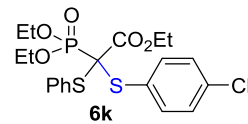
Sample Name : LIK-LB-091B
 Sample ID :
 Data Filename : LIK-LB-091B-1-rac-ADH-90-10-230-1.0.lcd
 Method Filename : 2.lcm
 Batch Filename :
 Vial # : 1-1
 Injection Volume : 10 uL
 Date Acquired : 2015/7/24 15:01:08
 Date Processed : 2015/7/24 15:15:04
 Sample Type : Unknown
 Acquired by : System Administrator
 Processed by : System Administrator

<Chromatogram>



<Peak Table>

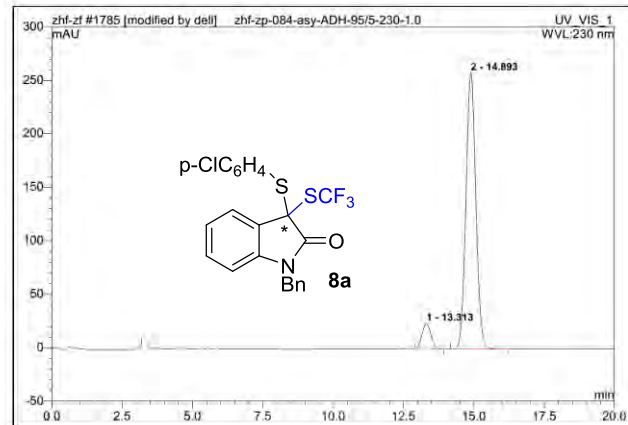
Peak#	Ret. Time	Area	Height	Conc.
1	8.958	3113564	243000	49.831
2	10.029	3134709	214064	50.169
Total		6248273	457064	



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1785 zh1-zp-084-asy-ADH-95/5-230-1.0

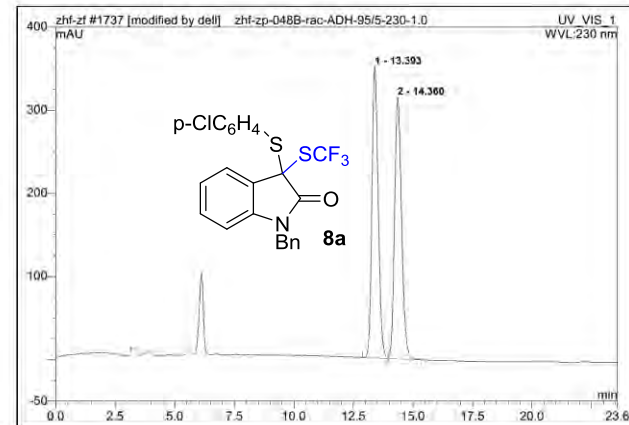
Sample Name:	zh1-zp-084-asy-ADH-95/5-230-1.0	Injection Volume:	20.0
Vial Number:	893	Channel:	UV_VIS_1
Sample Type:	unknown	Wavelength:	230
Control Program:	zh1	Bandwidth:	n.a.
Quantif. Method:	zh1	Dilution Factor:	1.0000
Recording Time:	2015-5-29 12:21	Sample Weight:	1.0000
Run Time (min):	20.00	Sample Amount:	1.0000



No.	Ret. Time min	Peak Name	Height mAU	Area mAU*min	Rel. Area %	Amount	Type
1	13.31	n.a.	23.520	8.697	7.64	n.a.	BMB*
2	14.89	n.a.	257.926	105.102	92.36	n.a.	BMB*
Total:			281.446	113.799	100.00	0.000	

1737 zh1-zp-048B-rac-ADH-95/5-230-1.0

Sample Name:	zh1-zp-048B-rac-ADH-95/5-230-1.0	Injection Volume:	20.0
Vial Number:	845	Channel:	UV_VIS_1
Sample Type:	unknown	Wavelength:	230
Control Program:	zh1	Bandwidth:	n.a.
Quantif. Method:	zh1	Dilution Factor:	1.0000
Recording Time:	2015-5-11 8:33	Sample Weight:	1.0000
Run Time (min):	23.59	Sample Amount:	1.0000



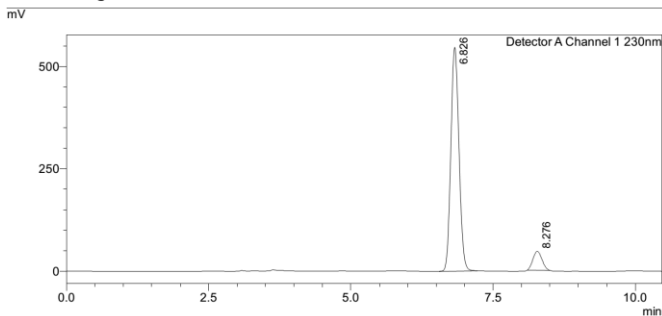
No.	Ret. Time min	Peak Name	Height mAU	Area mAU*min	Rel. Area %	Amount	Type
1	13.39	n.a.	349.898	107.124	49.88	n.a.	BM
2	14.36	n.a.	314.163	107.618	50.12	n.a.	MB
Total:			664.061	214.742	100.00	0.000	

Analysis Report

<Sample Information>

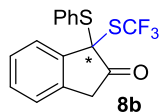
Sample Name : ZHF-ZQ-019
 Sample ID :
 Data Filename : zh-f-zq-019-1-asy-ODH-99-1-230-1.0.lcd
 Method Filename : 2.lcm
 Batch Filename :
 Vial # : 1-1
 Injection Volume : 10 uL
 Date Acquired : 2015/7/24 16:27:35
 Date Processed : 2015/7/25 10:25:50
 Sample Type : Unknown
 Acquired by : System Administrator
 Processed by : System Administrator

<Chromatogram>



<Peak Table>

Peak#	Ret. Time	Area	Height	Conc.
1	6.826	5431898	546877	91.405
2	8.276	510778	46218	8.595
Total		5942676	593095	



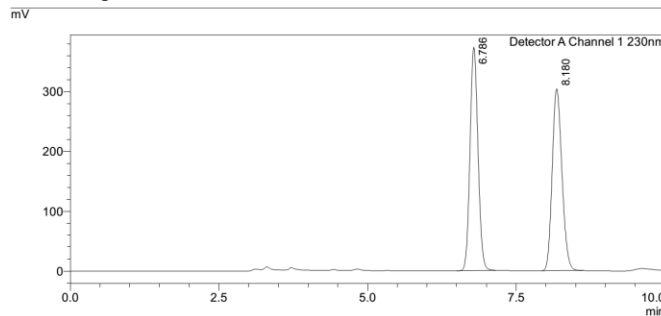
D:\Data\zhufeng\zhf-zq-019-1-asy-ODH-99-1-230-1.0.lcd

Analysis Report

<Sample Information>

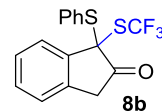
Sample Name : LIK-LB-063
 Sample ID :
 Data Filename : LIK-LB-063-RAC-ODH-99-1-230-1.0.lcd
 Method Filename : 2.lcm
 Batch Filename :
 Vial # : 1-1
 Injection Volume : 10 uL
 Date Acquired : 2015/7/24 16:40:44
 Date Processed : 2015/7/25 10:22:02
 Sample Type : Unknown
 Acquired by : System Administrator
 Processed by : System Administrator

<Chromatogram>



<Peak Table>

Peak#	Ret. Time	Area	Height	Conc.
1	6.786	3501971	373485	50.274
2	8.180	3463745	304108	49.726
Total		6965716	677593	



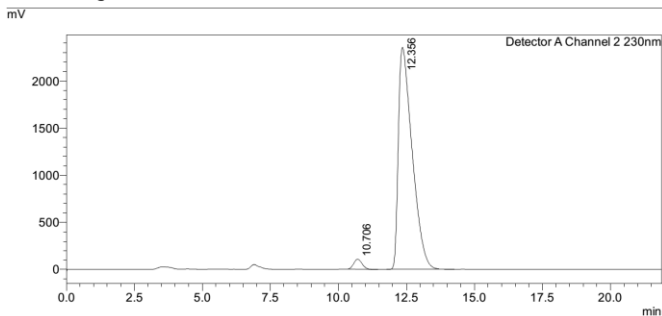
D:\Data\zhufeng\LIK-LB-063-RAC-ODH-99-1-230-1.0.lcd

Analysis Report

<Sample Information>

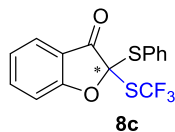
Sample Name : ZHF-ZP-110-ASY-OJH-80-20-230-1.0
 Sample ID :
 Data Filename : zhf-zp-110-asy-OJH-80-20-230-1.1.lcd
 Method Filename : chl-0128.lcm
 Batch Filename :
 Vial # : 1-1
 Injection Volume : 10 uL
 Date Acquired : 2015/7/11 15:41:36
 Date Processed : 2015/7/11 16:04:22
 Sample Type : Unknown
 Acquired by : System Administrator
 Processed by : System Administrator

<Chromatogram>



<Peak Table>

Peak#	Ret. Time	Area	Height	Conc.
1	10.706	2349638	107463	2.741
2	12.356	83358980	2357424	97.259
Total		85708618	2464887	

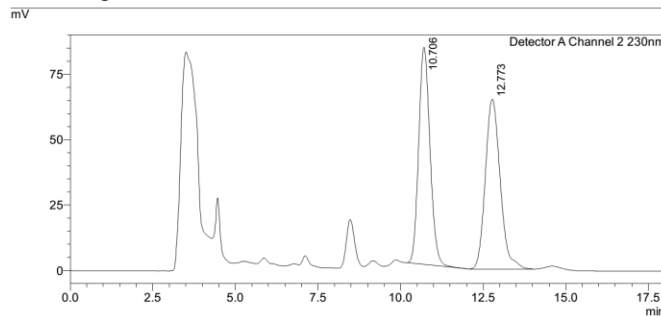


Analysis Report

<Sample Information>

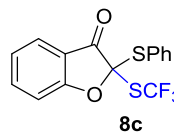
Sample Name : LIK-LB-088-RAC-OJH-80-20-230-1.0
 Sample ID :
 Data Filename : LIK-LB-088-RAC-OJH-80-20-230-1.1.lcd
 Method Filename : chl-0128.lcm
 Batch Filename :
 Vial # : 1-1
 Injection Volume : 10 uL
 Date Acquired : 2015/7/11 16:06:09
 Date Processed : 2015/7/13 21:55:55
 Sample Type : Unknown
 Acquired by : System Administrator
 Processed by : System Administrator

<Chromatogram>



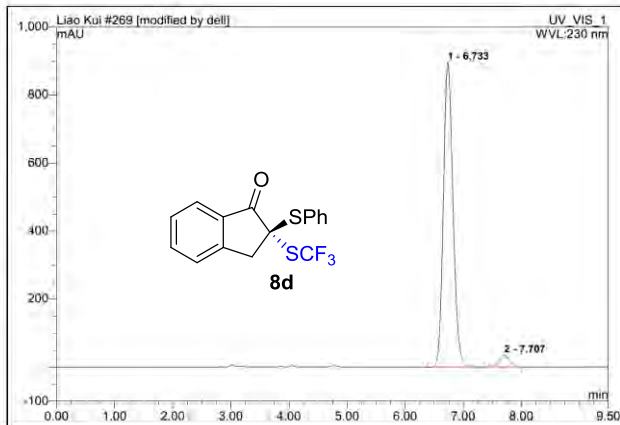
<Peak Table>

Peak#	Ret. Time	Area	Height	Conc.
1	10.706	1985675	83014	49.427
2	12.773	2031748	64888	50.573
Total		4017423	147902	



269 ZHF-ZP-85-ASY-ODH-99-1-230-1.0

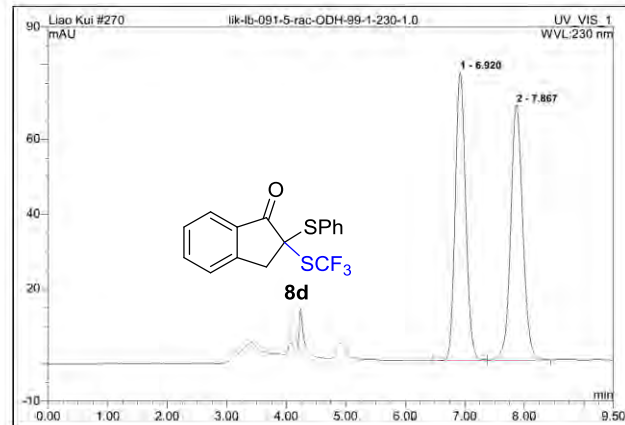
Sample Name:	ZHF-ZP-85-ASY-ODH-99-1-230-1.0	Injection Volume:	20.0
Vial Number:	367	Channel:	UV_VIS_1
Sample Type:	standard	Wavelength:	230
Control Program:	Liao Kui	Bandwidth:	n.a.
Quantif. Method:	Liao Kui	Dilution Factor:	1.0000
Recording Time:	2015-7-17 17:19	Sample Weight:	1.0000
Run Time (min):	13.83	Sample Amount:	1.0000



No.	Ret. Time min	Peak Name	Height mAU	Area mAU*min	Rel. Area %	Amount	Type
1	6.73	n.a.	895.086	172.086	96.00	n.a.	BMB*
2	7.71	n.a.	34.097	7.164	4.00	n.a.	BMB*
Total:			929.183	179.250	100.00	0.000	

270 lik-lb-091-5-rac-ODH-99-1-230-1.0

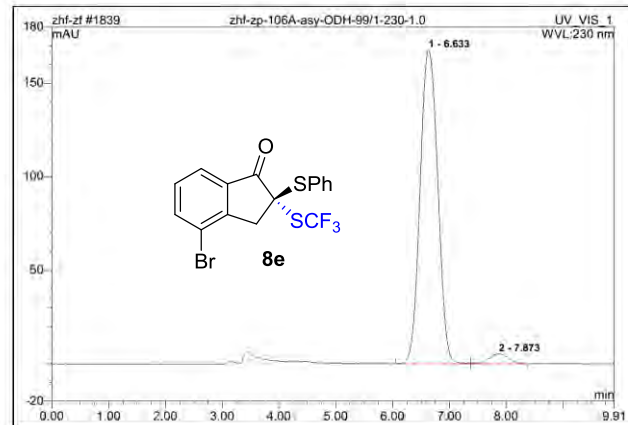
Sample Name:	lik-lb-091-5-rac-ODH-99-1-230-1.0	Injection Volume:	20.0
Vial Number:	368	Channel:	UV_VIS_1
Sample Type:	standard	Wavelength:	230
Control Program:	Liao Kui	Bandwidth:	n.a.
Quantif. Method:	Liao Kui	Dilution Factor:	1.0000
Recording Time:	2015-7-17 17:35	Sample Weight:	1.0000
Run Time (min):	13.49	Sample Amount:	1.0000



No.	Ret. Time min	Peak Name	Height mAU	Area mAU*min	Rel. Area %	Amount	Type
1	6.92	n.a.	77.188	15.705	49.31	n.a.	BM
2	7.87	n.a.	68.251	16.142	50.69	n.a.	MB
Total:			145.439	31.847	100.00	0.000	

1839 zhf-zp-106A-asy-ODH-99/1-230-1.0

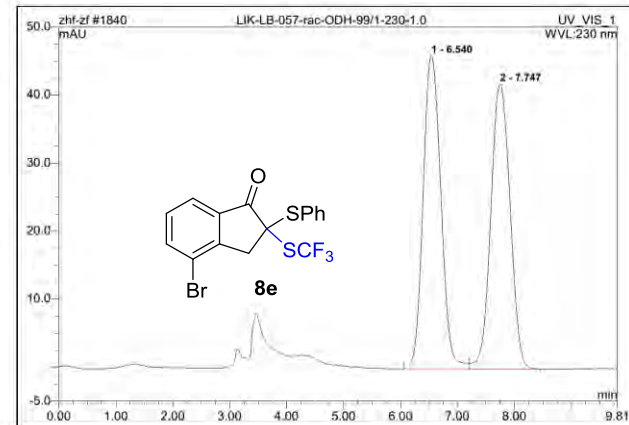
Sample Name:	zhf-zp-106A-asy-ODH-99/1-230-1.0	Injection Volume:	20.0
Vial Number:	948	Channel:	UV_VIS_1
Sample Type:	unknown	Wavelength:	230
Control Program:	zhf	Bandwidth:	n.a.
Quantif. Method:	zhf	Dilution Factor:	1.0000
Recording Time:	2015-6-24 10:37	Sample Weight:	1.0000
Run Time (min):	9.91	Sample Amount:	1.0000



No.	Ret. Time min	Peak Name	Height mAU	Area mAU*min	Rel. Area %	Amount	Type
1	6.63	n.a.	167.556	60.368	96.67	n.a.	BM
2	7.87	n.a.	5.244	2.076	3.33	n.a.	MB
Total:			172.800	62.444	100.00	0.000	

1840 LIK-LB-057-rac-ODH-99/1-230-1.0

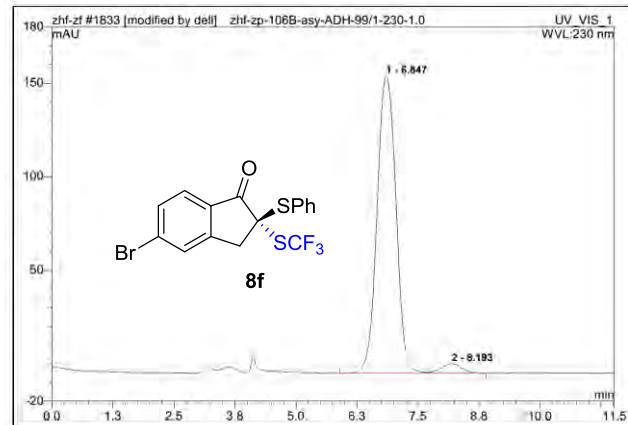
Sample Name:	LIK-LB-057-rac-ODH-99/1-230-1.0	Injection Volume:	20.0
Vial Number:	949	Channel:	UV_VIS_1
Sample Type:	unknown	Wavelength:	230
Control Program:	zhf	Bandwidth:	n.a.
Quantif. Method:	zhf	Dilution Factor:	1.0000
Recording Time:	2015-6-24 10:48	Sample Weight:	1.0000
Run Time (min):	9.81	Sample Amount:	1.0000



No.	Ret. Time min	Peak Name	Height mAU	Area mAU*min	Rel. Area %	Amount	Type
1	6.54	n.a.	46.032	16.613	50.22	n.a.	BM
2	7.75	n.a.	41.888	16.468	49.78	n.a.	MB
Total:			87.920	33.081	100.00	0.000	

1833 zhf-zp-106B-asy-ADH-99/1-230-1.0

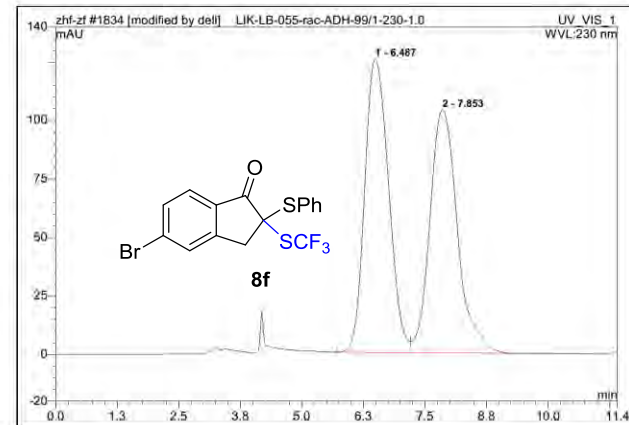
Sample Name:	zhf-zp-106B-asy-ADH-99/1-230-1.0	Injection Volume:	20.0
Vial Number:	942	Channel:	UV_VIS_1
Sample Type:	unknown	Wavelength:	230
Control Program:	zhf	Bandwidth:	n.a.
Quantif. Method:	zhf	Dilution Factor:	1.0000
Recording Time:	2015-6-23 21:29	Sample Weight:	1.0000
Run Time (min):	11.53	Sample Amount:	1.0000



No.	Ret. Time min	Peak Name	Height mAU	Area mAU*min	Rel. Area %	Amount	Type
1	6.85	n.a.	158.828	71.233	96.56	n.a.	BM
2	8.19	n.a.	5.076	2.534	3.44	n.a.	MB
Total:			163.904	73.768	100.00	0.000	

1834 LIK-LB-055-rac-ADH-99/1-230-1.0

Sample Name:	LIK-LB-055-rac-ADH-99/1-230-1.0	Injection Volume:	20.0
Vial Number:	943	Channel:	UV_VIS_1
Sample Type:	unknown	Wavelength:	230
Control Program:	zhf	Bandwidth:	n.a.
Quantif. Method:	zhf	Dilution Factor:	1.0000
Recording Time:	2015-6-23 21:44	Sample Weight:	1.0000
Run Time (min):	11.41	Sample Amount:	1.0000



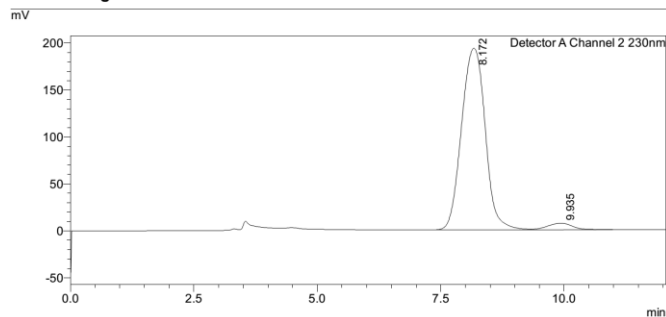
No.	Ret. Time min	Peak Name	Height mAU	Area mAU*min	Rel. Area %	Amount	Type
1	6.49	n.a.	125.457	70.400	50.96	n.a.	BM
2	7.85	n.a.	103.966	67.754	49.04	n.a.	MB
Total:			229.424	138.154	100.00	0.000	

Analysis Report

<Sample Information>

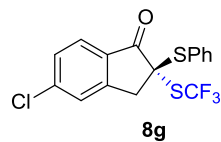
Sample Name : zhf-zp-106-C-ASY-ADH-99-1-230-1.0
 Sample ID :
 Data Filename : zhf-zp-106-C-ASY-ADH-99-1-230-1.0.lcd
 Method Filename : RUN-1.0.lcm
 Batch Filename :
 Vial # : 1-1
 Injection Volume : 10 uL
 Date Acquired : 2015/6/24 9:44:47
 Date Processed : 2015/6/24 10:12:33
 Sample Type : Unknown
 Acquired by : System Administrator
 Processed by : System Administrator

<Chromatogram>



<Peak Table>

Peak#	Ret. Time	Area	Height	Conc.
1	8.172	6456606	193635	96.377
2	9.935	242694	6810	3.623
Total		6699300	200445	



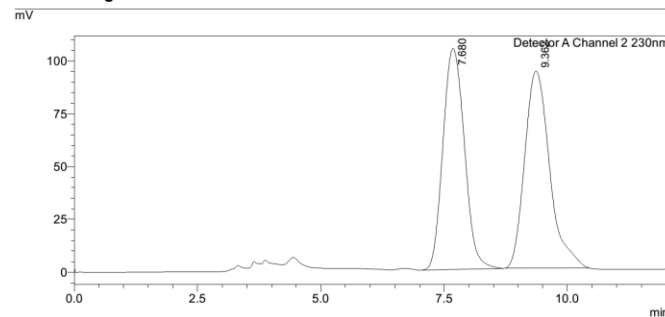
D:\Data\LiaoKu\新建文件夹\zhf-zp-106-C-ASY-ADH-99-1-230-1.0.lcd

Analysis Report

<Sample Information>

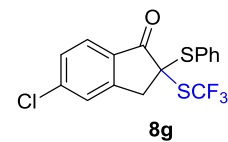
Sample Name : LIK-LB-054-RAC-ADH-99-1-230-1.0
 Sample ID :
 Data Filename : LIK-LB-054-RAC-ADH-99-1-230-1.0.lcd
 Method Filename : RUN-1.0.lcm
 Batch Filename :
 Vial # : 1-1
 Injection Volume : 10 uL
 Date Acquired : 2015/6/24 9:57:52
 Date Processed : 2015/7/13 21:52:11
 Sample Type : Unknown
 Acquired by : System Administrator
 Processed by : System Administrator

<Chromatogram>



<Peak Table>

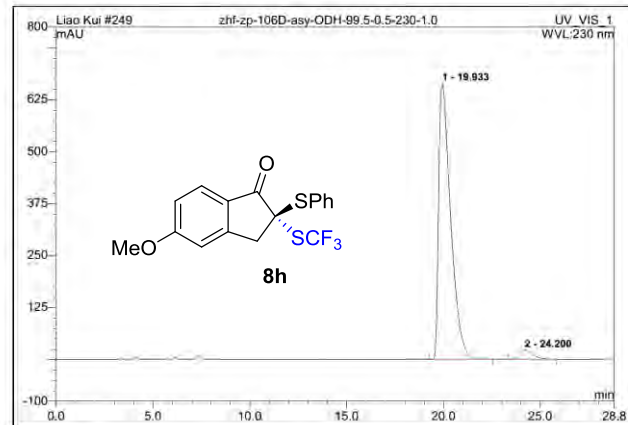
Peak#	Ret. Time	Area	Height	Conc.
1	7.680	3175856	104669	49.167
2	9.362	3283404	93267	50.833
Total		6459260	197936	



D:\Data\LiaoKu\新建文件夹\LIK-LB-054-RAC-ADH-99-1-230-1.0.lcd

249 zhf-zp-106D-asy-ODH-99.5-0.5-230-1.0

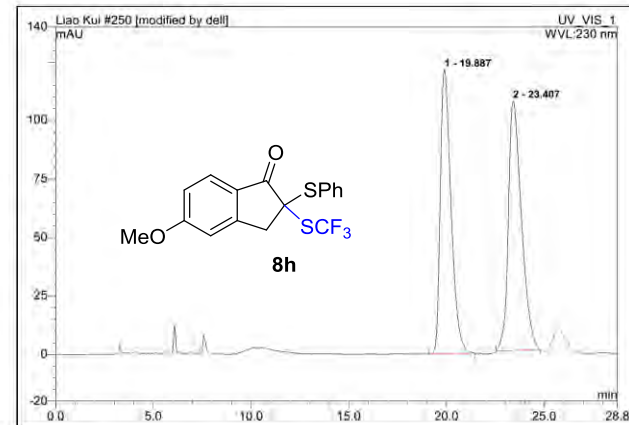
Sample Name:	zhf-zp-106D-asy-ODH-99.5-0.5-230-1.0	Injection Volume:	20.0
Vial Number:	347	Channel:	UV_VIS_1
Sample Type:	standard	Wavelength:	230
Control Program:	Liao Kui	Bandwidth:	n.a.
Quantif. Method:	Liao Kui	Dilution Factor:	1.0000
Recording Time:	2015-7-15 9:07	Sample Weight:	1.0000
Run Time (min):	28.83	Sample Amount:	1.0000



No.	Ret. Time min	Peak Name	Height mAU	Area mAU*min	Rel. Area %	Amount	Type
1	19.93	n.a.	664.302	475.939	96.48	n.a.	BMB
2	24.20	n.a.	22.210	17.350	3.52	n.a.	BMB
Total:			686.512	493.289	100.00	0.000	

250 lik-lb-084-2-rac-ODH-99.5-0.5-230-1.0

Sample Name:	lik-lb-084-2-rac-ODH-99.5-0.5-230-1.0	Injection Volume:	20.0
Vial Number:	348	Channel:	UV_VIS_1
Sample Type:	standard	Wavelength:	230
Control Program:	Liao Kui	Bandwidth:	n.a.
Quantif. Method:	Liao Kui	Dilution Factor:	1.0000
Recording Time:	2015-7-15 9:38	Sample Weight:	1.0000
Run Time (min):	28.75	Sample Amount:	1.0000



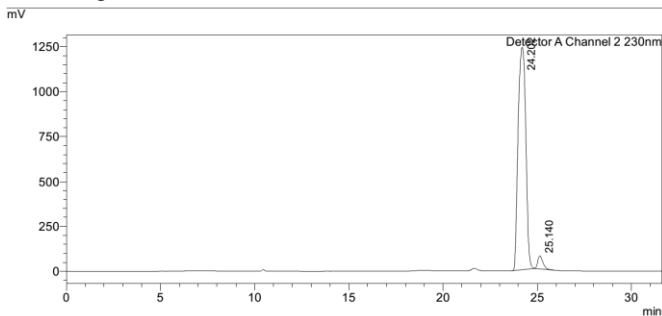
No.	Ret. Time min	Peak Name	Height mAU	Area mAU*min	Rel. Area %	Amount	Type
1	19.89	n.a.	121.386	77.338	48.29	n.a.	BMB*
2	23.41	n.a.	106.758	82.809	51.71	n.a.	BMB*
Total:			228.144	160.148	100.00	0.000	

Analysis Report

<Sample Information>

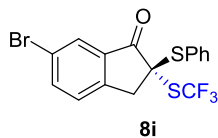
Sample Name : ZHF-ZP-129A--ADH-ADH-99.5-0.5-230-0.5
 Sample ID :
 Data Filename : ZHF-ZP-129A-ADH-ADH-99.5-0.5-230-1.0.lcd
 Method Filename : RUN-1.0.lcm
 Batch Filename :
 Vial # : 1-1
 Injection Volume : 10 uL
 Date Acquired : 2015/7/21 23:05:22
 Date Processed : 2015/7/22 15:28:06
 Sample Type : Unknown
 Acquired by : System Administrator
 Processed by : System Administrator

<Chromatogram>



<Peak Table>

Peak#	Ret. Time	Area	Height	Conc.
1	24.202	35156642	1238540	96.205
2	25.140	1386886	72244	3.795
Total		36543528	1310784	



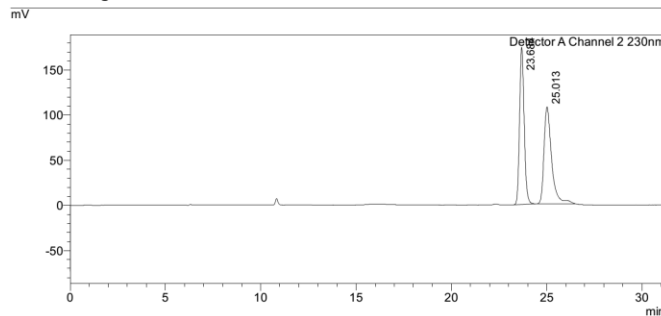
D:\Data\LiaoKui\ZHF-ZP-129A-ADH-ADH-99.5-0.5-230-1.0.lcd

Analysis Report

<Sample Information>

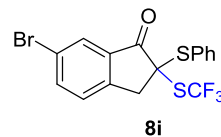
Sample Name : LIK-LB-091-4-RAC-ADH-ADH-99.5-0.5-230-0.5
 Sample ID :
 Data Filename : LIK-LB-091-RAC-ADH-ADH-99.5-0.5-230-1.0.lcd
 Method Filename : RUN-1.0.lcm
 Batch Filename :
 Vial # : 1-1
 Injection Volume : 10 uL
 Date Acquired : 2015/7/21 22:06:01
 Date Processed : 2015/7/22 15:31:07
 Sample Type : Unknown
 Acquired by : System Administrator
 Processed by : System Administrator

<Chromatogram>



<Peak Table>

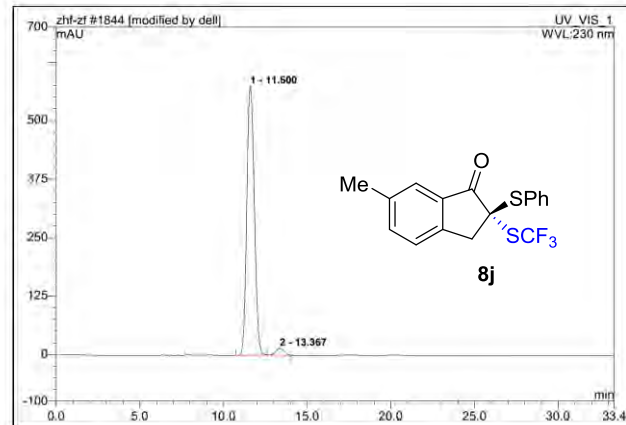
Peak#	Ret. Time	Area	Height	Conc.
1	23.684	2950081	174206	49.596
2	25.013	2998149	107449	50.404
Total		5948230	281656	



D:\Data\LiaoKui\LIK-LB-091-RAC-ADH-ADH-99.5-0.5-230-1.0.lcd

1844 zhf-zp-106E-asy-ODH-ODH-99/1-230-1.0

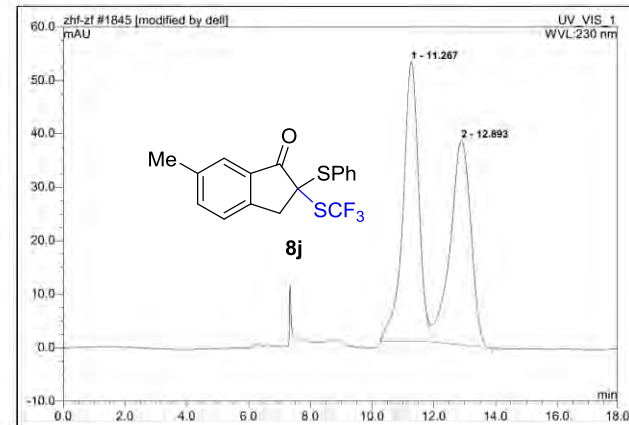
Sample Name:	zhf-zp-106E-asy-ODH-ODH-99/1-230-1.0	Injection Volume:	20.0
Vial Number:	953	Channel:	UV_VIS_1
Sample Type:	unknown	Wavelength:	230
Control Program:	zhf	Bandwidth:	n.a.
Quantif. Method:	zhf	Dilution Factor:	1.0000
Recording Time:	2015-6-24 14:19	Sample Weight:	1.0000
Run Time (min):	33.37	Sample Amount:	1.0000



No.	Ret. Time min	Peak Name	Height mAU	Area mAU*min	Rel. Area %	Amount	Type
1	11.60	n.a.	575.010	299.423	97.16	n.a.	BMb*
2	13.37	n.a.	14.256	8.760	2.84	n.a.	bMB*
Total:			589.266	308.184	100.00	0.000	

1845 LIK-LB-059-rac-ODH-ODH-99/1-230-1.0

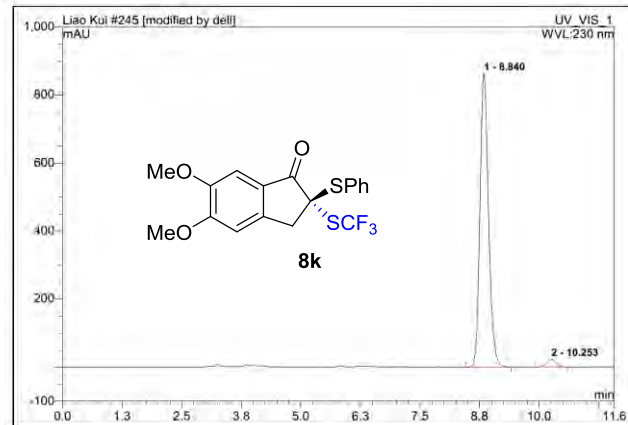
Sample Name:	LIK-LB-059-rac-ODH-ODH-99/1-230-1.0	Injection Volume:	20.0
Vial Number:	954	Channel:	UV_VIS_1
Sample Type:	unknown	Wavelength:	230
Control Program:	zhf	Bandwidth:	n.a.
Quantif. Method:	zhf	Dilution Factor:	1.0000
Recording Time:	2015-6-24 14:56	Sample Weight:	1.0000
Run Time (min):	17.95	Sample Amount:	1.0000



No.	Ret. Time min	Peak Name	Height mAU	Area mAU*min	Rel. Area %	Amount	Type
1	11.27	n.a.	52.302	30.625	50.59	n.a.	BM*
2	12.89	n.a.	38.208	29.916	49.41	n.a.	MB*
Total:			90.511	60.541	100.00	0.000	

245 zhf-zp-106F-asy-ADH-90-10-230-1.0

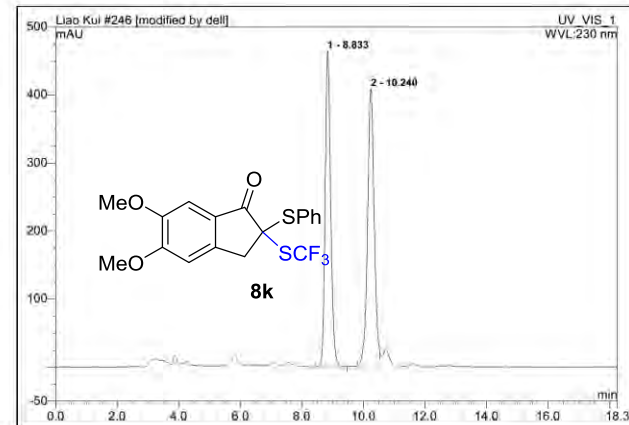
Sample Name:	zhf-zp-106F-asy-ADH-90-10-230-1.0	Injection Volume:	20.0
Vial Number:	343	Channel:	UV_VIS_1
Sample Type:	standard	Wavelength:	230
Control Program:	Liao Kui	Bandwidth:	n.a.
Quantif. Method:	Liao Kui	Dilution Factor:	1.0000
Recording Time:	2015-7-14 19:59	Sample Weight:	1.0000
Run Time (min):	11.58	Sample Amount:	1.0000



No.	Ret. Time min	Peak Name	Height mAU	Area mAU*min	Rel. Area %	Amount	Type
1	8.84	n.a.	864.170	180.929	96.94	n.a.	BMB*
2	10.25	n.a.	24.142	5.706	3.06	n.a.	BMB*
Total:			888.313	186.635	100.00	0.000	

246 lik-lb-084-3-rac-ADH-90-10-230-1.0

Sample Name:	lik-lb-084-3-rac-ADH-90-10-230-1.0	Injection Volume:	20.0
Vial Number:	344	Channel:	UV_VIS_1
Sample Type:	standard	Wavelength:	230
Control Program:	Liao Kui	Bandwidth:	n.a.
Quantif. Method:	Liao Kui	Dilution Factor:	1.0000
Recording Time:	2015-7-14 20:13	Sample Weight:	1.0000
Run Time (min):	18.26	Sample Amount:	1.0000



No.	Ret. Time min	Peak Name	Height mAU	Area mAU*min	Rel. Area %	Amount	Type
1	8.83	n.a.	463.666	96.328	48.57	n.a.	BMB
2	10.24	n.a.	407.595	101.988	51.43	n.a.	BMB*
Total:			871.261	198.316	100.00	0.000	