

Electronic Supplementary Information

Fluorine-Controlled C–H Borylation of Arenes Catalyzed by PSiN-Pincer Platinum Complex

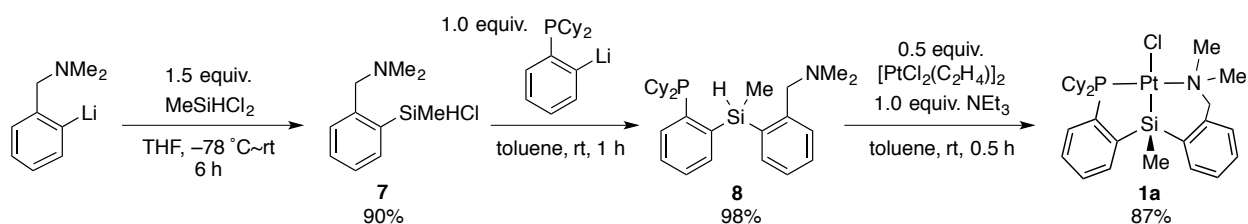
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General: All operations were performed under an argon atmosphere. ^1H , ^{13}C , ^{19}F and ^{31}P -NMR spectra were recorded on a JEOL ECX-400 (400 MHz for ^1H , 100 MHz for ^{13}C , 160 MHz for ^{31}P and 368 MHz for ^{19}F) or a JEOL ECS-400 (400 MHz for ^1H , 100 MHz for ^{13}C , 160 MHz for ^{31}P and 368 MHz for ^{19}F) or a Bruker DRX-500 (500 MHz for ^1H , 125 MHz for ^{13}C) spectrometer. ^1H and $^{13}\text{C}\{^1\text{H}\}$ NMR chemical shifts are expressed in parts per million (ppm) downfield from tetramethylsilane and are referenced to residual solvents (δ_{H} 7.26 and δ_{C} 77.0 for chloroform, δ_{H} 7.15 and δ_{C} 128.6 for benzene). ^{31}P NMR chemical shifts are expressed in ppm downfield from an 85% H_3PO_4 external standard. ^{19}F NMR spectra are referenced relative to hexafluorobenzene (−162.9 ppm). IR spectra were recorded on an FT/IR-VIR-200 (JASCO Co., Ltd.). Silica Gel 60 (Kanto Chemical Co., Inc.) was used for flash column chromatography. Merck Kieselgel 60 F254 (0.25 mm thickness, coated on glass 20×20 cm²) plate was used for analytical thin layer chromatography (TLC), and Wakogel B-5F coated on glass in a thickness of 0.9 mm was used for preparative TLC. High resolution mass analyses (FD^+ and EI^+) were performed on a JEOL JMS-T100 GSV mass spectrometer or a JEOL JMS-700 mass spectrometer. Elemental analyses were performed on an Elementar Vario MICRO. Unless otherwise noted, arenes, diboron, and mesitylene were used after distillation or recrystallization. THF, Et_2O , pentane and toluene were purified by solvent purification system of Glass-Contour. PSiP-platinum complex **1e** were prepared according to the reported procedures.¹

Preparation of PSiN-PtCl complex **1a**



Synthesis of chlorosilane **7**

To a stirred solution of *o*-(*N,N*-dimethylaminomethyl)phenyllithium² (571 g, 4.05 mmol) in THF (20 ml) was added dichloromethylsilane (0.6 mL, 5.76 mmol) at −78 °C. The mixture was allowed to stand at room temperature and was stirred for 6 h. The solvent was removed under reduced pressure, and the mixture was filtered with toluene through a short pad of Celite. Evaporation of the solvent afforded a pale yellow solid, which was purified by recrystallization from Et_2O /pentane to give **7** as a white solid (0.774 g, 3.63 mmol) in 90% yield.³

^1H NMR (CDCl_3 , 400 MHz) δ = 0.59 (s, 3H), 1.49 (s, 6H), 2.77 (d, J = 14.0 Hz, 1H), 2.95 (d, J = 14.0 Hz, 1H), 5.20–5.30 (m, 1H), 6.71 (d, J = 7.6 Hz, 1H), 7.10–7.50 (m, 2H), 8.66 (d, J = 6.8 Hz, 1H).

Synthesis of PN-mixed silane **8**

o-(Dicyclohexylphosphino)phenyllithium-diethylether (1.06 g, 3.0 mmol), which was prepared from (*o*-bromophenyl)dicyclohexylphosphine and *n*-BuLi, was added to a stirred solution of chlorosilane **7** (643 mg, 3.0 mmol) in toluene (5.0 mL) at room temperature. After 1 h, the mixture was filtered through a short pad of Celite to give P,N-mixed silane **8** (1.33 g, 2.94 mmol) in 98% yield, which was used for the next step without further purification.

IR (ATR) 2921, 2844, 2155, 1445 cm^{-1} ; ^1H NMR (CDCl_3 , 400 MHz) δ = 0.91 (s, 3H), 1.05–2.00 (m, 22H), 2.14 (s, 6H), 3.40 (d, J = 12.8 Hz, 1H), 3.90 (d, J = 12.8 Hz, 1H), 5.80–5.90 (m, 1H), 7.01–7.32 (m, 4H), 7.38–7.42 (m, 1H), 7.49–7.56 (m, 1H), 7.66–7.72 (m, 1H), 7.74–7.80 (m, 1H); ^{13}C NMR (CDCl_3 , 125 MHz) δ = 146.5 (d, J = 44.4 Hz), 145.5, 142.9 (d, J = 16.0 Hz), 136.9 (d, J = 3.9 Hz), 136.2 (d, J = 2.4 Hz), 135.3 (d, J = 14.8 Hz), 132.0 (d, J = 2.5 Hz), 128.4, 128.1, 127.9, 127.3, 125.6, 63.9, 44.4, 34.9 (d, J = 7.8 Hz), 34.8 (d, J = 7.6 Hz), 30.3 (d, J = 16.5 Hz),

30.1 (d, $J = 15.6$ Hz), 29.4 (d, $J = 3.8$ Hz), 29.3 (d, $J = 3.8$ Hz), 26.8-27.7 (m), 26.1 (d, $J = 7.9$ Hz), -3.2 (d, $J = 9.6$ Hz); ^{31}P NMR (CDCl_3 , 160 MHz) = -7.48 (s); HRMS (FD^+) Calcd for $\text{C}_{28}\text{H}_{42}\text{PNSi}$, M 451.2824. Found m/z 451.2819.

Synthesis of PSiN-platinum complex **1a**

To a stirred solution of silane **8** (0.678 g, 1.50 mmol) in toluene (9 ml) was added $[\text{PtCl}_2(\text{C}_2\text{H}_4)]_2$ (0.44 g, 0.75 mmol) and NEt_3 (250 μL , 1.79 mmol) at room temperature. After 1 h, the mixture was filtered through a short pad of Celite, and the solvent was removed under reduced pressure. Extraction with Et_2O and reprecipitation from Et_2O /pentane afforded **1a** (0.676 g, 0.99 mmol) in 66% yield.

IR (ATR) 2928, 1466 cm^{-1} ; ^1H NMR (CDCl_3 , 500 MHz) δ = 0.73 (s, 3H), 0.82-1.84 (m, 18H), 2.25-2.40 (m, 4H), 2.56 (d, $J = 1.5$ Hz, 3H), 2.99 (d, $J = 1.5$ Hz, 3H), 3.15-3.25 (m, 1H), 4.29 (d, $J = 12.3$ Hz, 1H), 6.82 (d, $J = 8.0$ Hz, 1H), 6.95-7.20 (m, 3H), 7.25-7.30 (m, 1H), 7.43 (t, $J = 7.1$, 1H), 7.80 (d, $J = 7.0$ Hz, 1H), 7.87 (d, $J = 7.3$ Hz, 1H); ^{13}C NMR (CD_2Cl_2 , 100 MHz) δ = 152.7 (d, $J = 36.4$ Hz), 141.8, 141.6, 141.2, 134.1 (d, $J = 20.3$ Hz), 131.7, 131.6, 131.1 (d, $J = 6.4$ Hz), 130.7, 129.4 (d, $J = 7.5$ Hz), 128.7, 128.4, 71.1, 51.8, 47.7, 36.3 (d, $J = 7.5$), 36.0 (d, $J = 7.5$ Hz), 30.0-26.7 (m), 2.3; ^{31}P NMR (C_6D_6 , 160 MHz) = 52.4 (s, $J_{\text{PPt}} = 4174$ Hz); HRMS (FD^+) Calcd for $\text{C}_{28}\text{H}_{41}\text{PPtSiNCl}$, M 681.2082. Found m/z 680.2115. *Anal.* Calcd for $\text{C}_{28}\text{H}_{41}\text{PPtSiNCl}$: C, 49.37; H, 6.07; N, 2.06. Found: C, 48.70; H, 6.06; N, 2.06.

Formation of PSiN-pincer type structure was clearly supported by observation of the J_{PtP} coupling and separately observed two methyl protons on the nitrogen and benzylic methylene protons. Furthermore, the structure of a PSiN-platinum triflate complex **1a'**, derived from **1a** and AgOTf , was confirmed by X-ray analysis (Figure S1, CCDC 1419522).

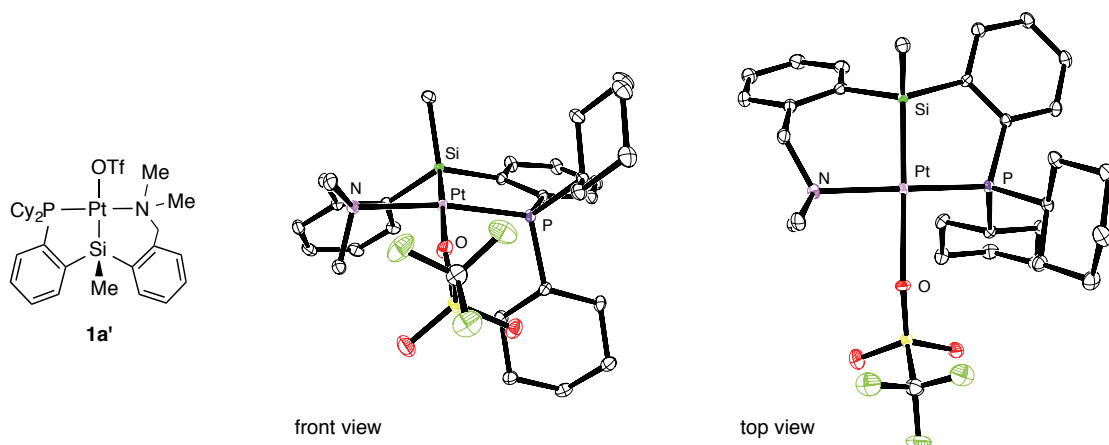
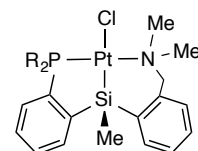


Figure S1. ORTEP drawings of the PSiN-platinum triflate complex **1a'** at 30% probability level. Hydrogen atoms are omitted for clarity.



Other platinum complexes were also synthesized according to the procedure for **1a**.

1b ($\text{R} = i\text{-Pr}$)

IR (ATR) 2961, 1448, 1234 cm^{-1} ; ^1H NMR (CDCl_3 , 400 MHz) δ = 0.79 (t, $J_{\text{PtH}} = 27.2$, 3H), 0.97 (dd, $J = 16.4$, 6.8 Hz, 3H), 1.04 (dd, $J = 13.6$, 6.8 Hz 3H), 1.22 (dd, $J = 14.0$, 6.8 Hz 3H), 1.65 (dd, $J = 16.0$, 6.8 Hz 3H), 2.64 (s, 3H), 3.01 (s, 3H), 3.25-3.40 (m, 1H), 4.23 (d, $J = 12.0$, 3H), 6.92 (d, $J = 7.6$ Hz, 1H), 7.12-7.37 (m, 5H), 7.83 (d, $J = 8.0$

Hz, 1H), 7.92 (d, J = 6.8 Hz, 1H); ^{13}C NMR (CD_2Cl_2 , 125 MHz) δ = 151.6 (d, J = 35.0 Hz), 140.8, 140.6, 140.2, 133.3 (d, J = 19.3 Hz), 130.8, 130.7, 130.0, 129.9 (d, J = 3.6 Hz), 128.6 (d, J = 7.9 Hz), 127.8, 127.6, 70.2, 50.8, 46.7, 25.9 (d, J = 33.5 Hz), 25.5 (d, J = 34.6 Hz), 18.9 (d, J = 1.4 Hz), 18.7 (d, J = 2.3 Hz), 18.2 (d, J = 2.1 Hz), 17.2 (d, J = 3.6 Hz), 1.19; ^{31}P NMR (C_6D_6 , 202 MHz) = 59.5 (s, $J_{\text{P-Pt}}$ = 4177 Hz); HRMS (FD^+) Calcd for $\text{C}_{22}\text{H}_{33}\text{ClNPtSi}$, M 600.1456. Found m/z 600.1466.

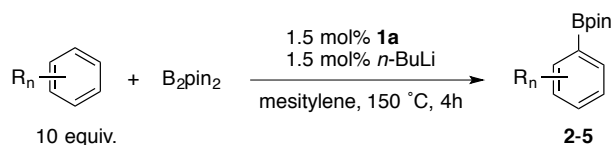
1c (R = *t*-Bu)

IR (ATR) 2890, 1473, 1174, 1107 cm^{-1} ; ^1H NMR (CD_2Cl_2 , 500 MHz) δ = 0.60 (s, 3H), 1.35 (d, J = 14.2 Hz, 9H), 1.43 (d, J = 14.5 Hz, 9H), 2.64 (s, 3H), 3.23 (s, 3H), 3.55 (dd, J = 12.6, 6.2 Hz, 1H), 4.56 (d, J = 12.5 Hz, 1H), 7.20-7.30 (m, 3H), 7.41-7.48 (m, 2H), 7.52-7.57 (m, 1H), 7.90 (d, J = 7.3 Hz, 1H), 7.94 (dd, J = 8.1, 5.1 Hz, 1H); ^{13}C NMR (CD_2Cl_2 , 125 MHz) δ = 151.3 (d, J = 34.8 Hz), 141.1, 140.7 (d, J = 4.4 Hz), 140.6, 133.5 (d, J = 18.9 Hz), 133.1 (d, J = 5.5 Hz), 130.7, 130.5, 129.6 (d, J = 5.9 Hz), 127.7, 127.5, 127.4 (d, J = 6.9 Hz), 69.9, 50.8 (d, J = 2.3 Hz), 47.0 (d, J = 2.0 Hz), 36.9 (d, J = 26.3 Hz), 36.7 (d, J = 24.0 Hz), 30.5 (d, J = 7.9 Hz), 30.4 (d, J = 8.1 Hz), 1.1; ^{31}P NMR (C_6D_6 , 202 MHz) = 76.4; HRMS (FD^+) Calcd for $\text{C}_{24}\text{H}_{37}\text{ClNPtSi}$, M 628.1769. Found m/z 628.1753.

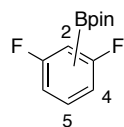
1d (R = Ph)

IR (ATR) 2360, 1431, 1103 cm^{-1} ; ^1H NMR (C_6D_6 , 400 MHz) δ = 0.73 (s, 3H), 2.57 (s, 3H), 2.69 (s, 3H), 2.90-3.10 (m, 1H), 4.20-4.40 (m, 1H), 6.80-7.40 (m, 14H), 7.60-8.20 (m, 6H); ^{31}P NMR (C_6D_6 , 202 MHz) = 36.1 (s, $J_{\text{P-Pt}}$ = 4159 Hz); HRMS (FD^+) Calcd for $\text{C}_{28}\text{H}_{29}\text{ClNPtSi}$, M 668.1143. Found m/z 668.1148.

Procedure for the C–H borylation of arenes



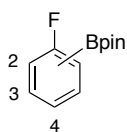
To a solution of **1a** (4.08 mg, 0.0059 mmol) in mesitylene (2 mL) was added a *n*-hexane solution of *n*-BuLi (1.56 M, 3.85 μL , 0.4 mmol) followed by addition of B_2pin_2 (101.3 mg, 0.4 mmol) and arene (4 mmol) at room temperature. The mixture was stirred at 150 $^\circ\text{C}$ for 4 h. After the solvent was removed under reduced pressure, the obtained crude material was purified by silica gel column chromatography to give arylboronic esters as an inseparable mixture of regioisomers. Silica gel column chromatography was carried out by cooling the column at –30 $^\circ\text{C}$ to avoid decomposition of the arylboronic ester on silica gel. The ratios of regioisomers were determined by ^1H NMR or ^{19}F NMR.



2a (82.9 mg, 0.345 mmol, 86%, 2-:4-:5- = 74:22:4 (Entry 1, Table 1)). The ratio of regioisomers was determined by ^{19}F NMR of the isolated product, which was almost identical to that in the crude material. The ratio was slightly improved to 2-:4-:5- = 78:18:4 by carrying out the reaction at 120 $^\circ\text{C}$ (Entry 2, Table 1).

^1H NMR (CDCl_3 , 400 MHz) **2-2a**: δ = 1.38 (s, 12H), 6.81-6.87 (m, 2H), 7.31-7.39 (m, 1H); **4-2a**: δ = 1.35 (s, 12H), 6.73-6.80 (m, 1H), 6.83-6.90 (m, 1H), 7.69-7.76 (m, 1H); ^{19}F NMR (CDCl_3 , 368 MHz) **2-2a**: δ = –101.8; **4-2a**: δ = –99.8, –106.3; **5-2a**: δ = –112.0.

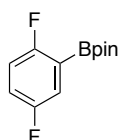
Spectral data were in good agreement with literature values.⁴



2b (45.7 mg, 0.206 mmol, 51%, 2-:3-:4- =80:17:3 (Entry 1, Table 2)). The ratio of regioisomers was determined by ^{19}F NMR of the isolated product, which was almost identical to that in the crude material. The ratio was slightly improved to 2-:3-:4- =84:13:3 by carrying out the reaction at 120 °C (Entry 2, Table 2).

^1H NMR (CDCl_3 , 400 MHz) **2-2b**: δ = 7.71-7.76 (m, 1H), 7.40-7.60 (m, 1H), 7.13 (t, J = 7.6 Hz, 1H), 7.02 (br t, J = 9.4 Hz, 1H), 1.36 (s, 12H); **3-2a**: δ = 7.56 (d, J = 7.2 Hz, 1H), 7.50-7.55 (m, 1H), 7.30-7.37 (m, 1H), 1.35 (s, 12H) (one aromatic proton was overlapped); **4-2a**: δ = 7.77-7.81 (m, 2H), 1.34 (s, 12H) (two aromatic protons were overlapped); ^{19}F NMR (CDCl_3 , 368 MHz) **2-2b**: δ = -103.8; **3-2b**: δ = -115.4; **4-2b**: δ = -109.6.

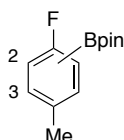
Spectral data were in good agreement with literature values.⁵



2c (66.0 mg, 0.275 mmol, 69%)

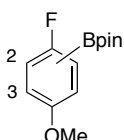
^1H NMR (CDCl_3 , 400 MHz) δ = 7.38 (q, J = 4.0 Hz, 1H), 7.05-7.12 (m, 1H), 6.95-7.00 (m, 1H), 1.36 (s, 12H); ^{19}F NMR (CDCl_3 , 368 MHz) δ = -110.6, -121.8.

Spectral data were in good agreement with literature values.⁶



2d (79.4 mg, 0.33 mmol, 84%, 2-:3- =96:4). The ratio of regioisomers was determined by ^1H NMR of the isolated product, which was almost identical to that in the crude material. The borylated position (2- or 3-) was determined by ^1H NMR, in which the *ortho*-proton to the Bpin appeared to be doublet of doublet with smaller J_{FH} (~5 Hz) in **2-2d** than that in **3-2d** (~10 Hz).

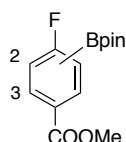
IR (ATR) 2983, 1613, 1492, 1409, 1347, 1216, 1144 cm^{-1} ; ^1H NMR (CDCl_3 , 400 MHz) **2-2d**: δ = 7.52 (dd, J = 5.5, 2.3 Hz, 1H), 7.19-7.24 (m, 1H), 6.91 (t, J = 8.7 Hz, 1H), 1.36 (s, 12H); **3-2d**: δ = 7.42 (dd, J = 9.4, 2.5 Hz, 1H), 7.11 (dd, J = 8.6, 4.9 Hz, 1H), 6.96-7.01 (m, 1H), 1.34 (s, 12H); ^{13}C NMR (CDCl_3 , 100 MHz) **2-2d**: δ = 165.3 (d, J = 284 Hz), 136.9 (dd, J = 60.6, 20.9 Hz), 133.3 (m), 131.1 (d, J = 9.4 Hz), 114.9 (d, J = 24 Hz), 83.6, 24.8, 20.4; ^{19}F NMR (CDCl_3 , 368 MHz) **2-2d**: δ = -108.2; **3-2d**: δ = -119.6; HRMS (FD^+) Calcd for $\text{C}_{13}\text{H}_{18}\text{BFO}_2$, M 236.1384. Found m/z 236.1411.



2e (92.8 mg, 0.39 mmol, 92%, 2-:3- =87:13). The ratio of regioisomers was determined by ^1H NMR of the isolated product, which was almost identical to that in the crude material. The borylated position (2- or 3-) was determined

by analyzing the J_{FH} value as described for **2d**.

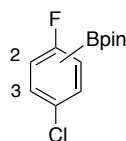
IR (KBr) 2979, 1492, 1418, 1349, 1207, 1145 cm^{-1} ; ^1H NMR (CDCl_3 , 400 MHz) **2-2e**: δ = 7.18-7.21 (m, 1H), 6.92-6.97 (m, 2H), 3.81 (s, 3H), 1.36 (s, 12H); **3-2e**: δ = 7.34 (dd, J = 12.0, 3.36 Hz, 1H), 7.02-7.09 (m, 1H), 6.78 (dd, J = 13.9, 4.03 Hz, 1H), 3.81 (s, 3H), 1.35 (s, 12H); ^{13}C NMR (CDCl_3 , 100 MHz) **2-2e**: δ = 161.6 (d, J = 242 Hz), 155.2, 119.8 (d, J = 8.4 Hz), 119.3 (d, J = 8.4 Hz), 116.0 (d, J = 26 Hz), 83.9, 55.8, 24.8; ^{19}F NMR (CDCl_3 , 368 MHz) **2-2e**: δ = -113.9; **3-2e**: δ = -125.2; HRMS (FD^+) Calcd for $\text{C}_{13}\text{H}_{18}\text{BFO}_3$, M 252.1333. Found m/z 252.1377.



2f (43.9 mg, 0.157 mmol, 78% (from 0.20 mmol B_2pin_2), 2- only). The selective borylation at the 2-position was confirmed by analyzing the J_{FH} value as described for **2d**.

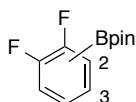
IR (KBr) 2982, 1352 cm^{-1} ; ^1H NMR (CDCl_3 , 400 MHz) δ = 8.44 (dd, J = 3.52, 2.34 Hz, 1H), 8.03-8.08 (m, 1H), 7.07 (t, J = 10.6 Hz, 1H), 3.91 (s, 3H), 1.37 (s, 12H); ^{13}C NMR (CDCl_3 , 100 MHz) δ = 169.9 (d, J = 259 Hz), 166.2, 138.9 (d, J = 9.4 Hz), 135.0 (d, J = 10.5 Hz), 125.9 (d, J = 3.1 Hz), 115.6 (d, J = 25.2 Hz), 84.2, 52.1, 24.8; ^{19}F NMR (CDCl_3 , 368 MHz) δ = -95.2.

HRMS (FD^+) Calcd for $\text{C}_{14}\text{H}_{18}\text{BFO}_4$, M 280.1282. Found m/z 280.1276.



2g (47 mg, 0.18 mmol, 91% (from 0.20 mmol B_2pin_2), 2-:3- =83:17). The ratio of regioisomers was determined by ^1H NMR of the isolated product, which was almost identical to that in the crude material. The borylated position (2- or 3-) was determined by analyzing the J_{FH} value as described for **2d**.

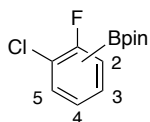
IR (KBr) 2982, 1352, 1299, 1134 cm^{-1} ; ^1H NMR (CDCl_3 , 400 MHz) **2-2g**: δ = 7.68 (dd, J = 5.0, 3.8 Hz, 1H), 7.34-7.40 (m, 1H), 6.97 (t, J = 8.8 Hz, 1H), 1.36 (s, 12H); **3-2g**: δ = 7.34-7.40 (m, 1H), 7.30 (dd, J = 9.2, 4.8 Hz, 1H), 7.00-7.05 (m, 1H), 1.35 (s, 12H); ^{13}C NMR (CDCl_3 , 100 MHz) **2-2g**: δ = 165.5 (d, J = 249 Hz), 136.2 (d, J = 5 Hz), 132.9 (d, J = 8.6 Hz), 116.8 (d, J = 36.3 Hz), 84.3, 24.8; ^{19}F NMR (CDCl_3 , 368 MHz) **2-2g**: δ = -105.8; **3-2g**: δ = -117.1; HRMS (FD^+) Calcd for $\text{C}_{12}\text{H}_{15}\text{BFCIO}_2$, M 256.0838. Found m/z 256.0843.



2h (71.5 mg, 0.29 mmol, 74%, 2-:3- =91:9). The ratio of regioisomers was determined by ^1H NMR of the isolated product, which was almost identical to that in the crude material.

^1H NMR (CDCl_3 , 400 MHz) **2-2h**: δ = 7.43-7.48 (m, 1H), 7.19-7.28 (m, 1H), 7.03-7.10 (m, 1H), 1.36 (s, 12H); **3-2g**: δ = 7.50-7.61 (m, 2H), 7.10-7.20 (m, 1H), 1.36 (s, 12H); ^{19}F NMR (CDCl_3 , 368 MHz) **2-2h**: δ = -130.2, -140.2; **3-2h**: δ = -135.1, -140.8.

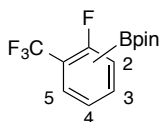
Spectral data were in good agreement with literature values.^{7,8}



2i (35.2 mg, 0.13 mmol, 69% (from 0.2 mmol B₂pin₂), 2-:3-:4-:5-:6-:1=65:6:8:21). The ratio of regioisomers was determined by ¹⁹F NMR of the isolated product, which was almost identical to that in the crude material.

¹H NMR (CDCl₃, 400 MHz) **2-2i**: δ = 7.62 (d, *J* = 1.60, 8.38 Hz, 1H), 7.48 (dt, *J* = 1.20, 7.58 Hz, 1H), 7.08 (t, *J* = 7.98 Hz, 1H), 1.39 (s, 12H); **3-2i**: δ = 7.56 (d, *J* = 9.62 Hz, 1H), 7.5 (dd, *J* = 1.30, 7.80 Hz, 1H), 7.41 (t, *J* = 7.06 Hz, 1H), 1.36 (s, 12H); **4-2i**: δ = 7.88 (d, *J* = 12.6 Hz, 1H), 7.7-7.68 (m, 1H), 7.17-7.13 (m, 1H), 1.36 (s, 12H); **5-2i**: δ = 7.45-7.50 (m, 1H), 7.17-7.22 (m, 2H), 1.37 (s, 12H) (aromatic protons are overlapped); ¹⁹F NMR (CDCl₃, 368 MHz) **2-2i**: δ = -105.8; **3-2i**: δ = -117.1; **4-2i**: δ = -111.1; **5-2i**: δ = -114.4.

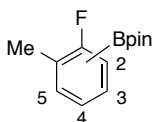
Spectral data were in good agreement with literature values of **2-2i**, **3-2i**, and **4-2i**^{9,10} and those of authentic sample of **5-2i** derived from commercially available corresponding boronic acid.



2j (73%, 2-:3-:4-:5-:6-:1=75:16:9:-). The ratio of regioisomers was determined by ¹⁹F NMR of the crude product due to partial decomposition of the products after column chromatography. The NMR yield of **2-2j** in the crude mixture was shown using 1,1,2,2-tetrachloroethane as an internal standard.

¹H NMR (CDCl₃, 400 MHz) **2-2j**: δ = 7.89 (t, *J* = 8.0 Hz, 1H), 7.68 (t, *J* = 6.0 Hz, 1H), 7.18-7.26 (m, 1H), 1.36 (s, 12H); **3-2j**: δ = 7.55-7.65 (m, 3H), 1.32 (m, 12H); **4-2j**: δ = 8.03 (d, *J* = 7.6 Hz, 1H), 7.88-7.96 (m, 1H), 7.14-7.18 (m, 1H), 1.35 (s, 12H); ¹⁹F NMR (CDCl₃, 368 MHz) **2-2j**: δ = -61.5, -104.0; **3-2j**: δ = -61.5, -116.0; **4-2j**: δ = -61.5, -110.3.

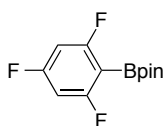
Spectral data were in good agreement with literature values.^{11,12,13}



2k (23.4 mg, 0.99 mmol, 50% (from 0.2 mmol B₂pin₂), 2-:3-:4-:5-:6-:1=71:16:7:6). The ratio of regioisomers was determined by ¹⁹F NMR of the isolated product, which was almost identical to that in the crude material.

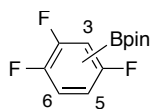
¹H NMR (CDCl₃, 400 MHz) **2-2k**: δ = 7.54 (t, *J* = 5.7 Hz, 1H), 7.27 (t, *J* = 7.8 Hz, 1H), 7.02 (t, *J* = 7.6 Hz, 1H), 2.25 (d, *J* = 2.0 Hz, 1H), 1.36 (s, 12H); **3-2k**: δ = 7.38-7.46 (m, 2H), 7.17 (t, *J* = 7.4 Hz, 1H), 2.28 (d, *J* = 1.8 Hz, 3H), 1.31 (s, 12H); **4-2k**: δ = 7.57-7.65 (m, 2H), 6.95-7.02 (m, 1H), 2.21 (s, 3H), 1.31 (s, 12H); ¹⁹F NMR (CDCl₃, 368 MHz) **2-2k**: δ = -106.8; **3-2k**: δ = -119.0; **4-2k**: δ = -112.8; **5-2k**: δ = -117.3.

Spectral data were in good agreement with literature values of **2-2k**, **3-2k**, and **4-2k**.^{14,15} and those of authentic sample of **5-2k** derived from commercially available corresponding boronic acid.



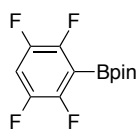
2l (116 mg, 0.449 mmol, 112%)

^1H NMR (CDCl_3 , 400 MHz) δ = 7.55-6.65 (m, 2H), 1.36 (s, 12H); ^{19}F NMR (CDCl_3 , 368 MHz) δ = -98.4, -105.1. Spectral data were in good agreement with literature values.¹⁶



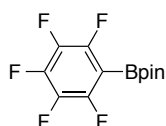
2m (83 mg, 0.32 mmol, 80%, 3-:5-:6- = 73:19:8). The ratio of regioisomers was determined by ^{19}F NMR of the isolated product, which was almost identical to that in the crude material.

IR (KBr) 1634, 1482 cm^{-1} ; ^1H NMR (CDCl_3 , 400 MHz) **3-2m**: δ = 7.10-7.20 (m, 1H), 6.72-6.80 (m, 1H), 1.37 (s, 12H); **5-2m**: δ = 7.10-7.20 (m, 1H), 6.94-7.02 (m, 1H), 1.34 (s, 12H); **6-2m**: δ = 7.49 (ddd, J = 11.2, 9.7, 4.7 Hz, 1H), 6.82-6.90 (m, 1H), 1.33 (s, 12H); ^{19}F NMR (CDCl_3 , 368 MHz) **3-2m**: δ = -107.5 ~ -107.6 (m), -126.1 (dd, J = 21.0, 8.5 Hz), -144.8 ~ -145.0 (m); **5-2m**: δ = -117.4 ~ -117.5 (m), -126.1 (dd, J = 21.3, 8.5 Hz), -135.1 ~ -135.2 (m); **6-2m**: δ = -105.3 ~ -105.4 (m), -129.6 ~ -129.7 (m), -145.2 ~ -145.4 (m); *Anal.* Calcd for $\text{C}_{12}\text{H}_{14}\text{BF}_3\text{O}_2$: C, 55.85; H, 5.47. Found: C, 55.23; H, 5.22. The structure of **3-2m** was confirmed by NOE measurement, which demonstrated the two aromatic protons are next to each other. The identity of **5-2m** and **6-2m** was confirmed by independent synthesis of authentic samples by Pd-catalyzed borylation of 1-bromo-2,4,5-trifluorobenzene or 1-bromo-2,3,5-trifluorobenzene with B_2pin_2 .



2n (74.4 mg, 0.270 mmol, 67%)

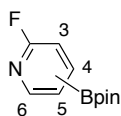
IR (KBr) 3057, 2985, 1479, 1379, 1336 cm^{-1} ; ^1H NMR (CDCl_3 , 400 MHz) δ = 7.05-7.15 (m, 1H), 1.41 (s, 12H); ^{19}F NMR (CDCl_3 , 368 MHz) δ = -110.5, -118.9; ^{13}C NMR (CDCl_3 , 100 MHz) δ = 148.7 (br d, J = 253 Hz), 145.5 (br d, J = 253 Hz), 108.5 (t, J = 22.5 Hz), 84.9, 24.7.



2o (77.9 mg, 0.26 mmol, 66%)

^1H NMR (CDCl_3 , 400 MHz) δ = 1.37 (s, 12H); ^{19}F NMR (CDCl_3 , 368 MHz) δ = -130.6 ~ -130.7 (m, 2F), -150.1 ~ -150.9 (m, 1F), -162.3 ~ -163.0 (m, 2F);

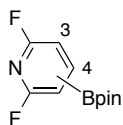
Spectral data were in good agreement with literature values.¹⁷ A small amount of 2,3,5,6- $\text{F}_4\text{C}_6\text{HBpin}$ and 2,3,5,6- F_4 -1,4-(Bpin)₂benzene were observed in ^{19}F NMR (<3%).



3a (77.4 mg, 0.344 mmol, 86%, 3-:4-:5-:6- = 50:38:12:-). The ratio of regioisomers was determined by ^1H NMR of the isolated product. The ^1H NMR of the crude mixture suggested the existence of an unidentified product, which might be **6-3a**, in less than 10% although it disappeared after column chromatography.

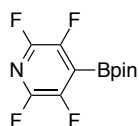
¹H NMR (CDCl₃, 400 MHz) **3-3a**: δ = 8.28-8.31 (m, 1H), 8.14 – 8.20 (m, 1H), 7.15-7.20 (m, 1H), 1.36 (s, 12H); **4-3a**: δ = 8.24 (d, J = 4.7 Hz, 1H), 7.47-7.51 (m, 1H), 7.28 (d, J = 2.0 Hz, 1H), 1.36 (s, 12H); **5-3a**: δ = 8.59 (s, 1H), 8.12-8.20 (m, 1H), 6.91 (dd, J = 4.1, 2.5 Hz, 1H), 1.35 (s, 12H); ¹⁹F NMR (CDCl₃, 368 MHz) **3-3a**: δ = -57.6; **4-3a**: δ = -68.7; **5-3a**: δ = -63.8;

Spectral data were in good agreement with literature values of **3-**, **4-**, and **5-3a**.^{18,19}



3b (69 mg, 0.28 mmol, 72%, 3-:4- = 69:31). The ratio of regioisomers was determined by ¹H NMR of the isolated product, which was almost identical to that in the crude material.

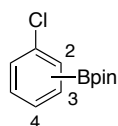
IR (KBr) 1635, 1261 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz) **3-3b**: δ = 8.24 (q, J = 8.2, 1H), 6.79-6.83 (m, 1H), δ = 1.34 (s, 12H); **4-3b**: δ = 7.14 (s, 2H), 1.34 (s, 12H); ¹⁹F NMR (CDCl₃, 368 MHz) **3-3b**: δ = -57.0, -63.6; **4-3b**: δ = -69.2; ¹³C NMR (CDCl₃, 100 MHz) **3-3b**: δ = 165.6 (dd, J = 200, 11.3 Hz), 163.8 (dd, J = 200, 11.3 Hz), 152.4 (t, J = 7.8 Hz), 105.7 (dd, J = 32.8, 5.4 Hz), 84.5, 24.6; **4-3b**: δ = 161.4 (dd, J = 198, 10.8 Hz), 110.8 (dd, J = 21.5, 8.5 Hz), 85.1, 24.6; HRMS (EI⁺) Calcd for C₁₁H₁₄BF₂NO₂, M 241.1086. Found m/z 241.1075.



3c (115mg, 0.41 mmol, 104%)

¹H NMR (C₆D₆, 400 MHz) δ = 1.01 (s, 12H); ¹⁹F NMR (C₆D₆, 368 MHz) δ = -92.5, -132.6.

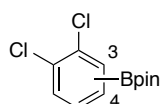
Spectral data were in good agreement with literature values.²⁰



4a (41.2 mg, 0.17 mmol, 43%, 2-:3-:4-:=46:37:17). The ratio of regioisomers was determined by ¹H NMR of the isolated product, which was almost identical to that in the crude material.

¹H NMR (CDCl₃, 400 MHz) **2-4a**: δ = 7.65 (d, J = 7.6 Hz, 1H), 7.30-7.36 (m, 2H), 7.19-7.24 (m, 1H), 1.36 (s, 12H); **3-4a**: δ = 7.77 (br s, 1H), 7.67 (d, J = 7.6 Hz, 1H), 7.38-7.43 (m, 1H), 7.26-7.32 (m, 1H), 1.33 (s, 12H); **4-4a**: δ = 7.71 (d, J = 8.4 Hz, 2H), 7.30-7.34 (m, 2H), 1.33 (s, 12H);

Spectral data were in good agreement with literature values.²¹

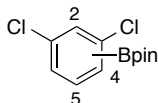


4b (82.8 mg, 0.30 mmol, 76%, 3-:4- = 60:40). The ratio of regioisomers was determined by ¹H NMR of the isolated product, which was almost identical to that in the crude material.

¹H NMR (CDCl₃, 400 MHz) **3-4b**: δ = 7.55 (dd, J = 7.6, 1.6 Hz, 1H), 7.50 (dd, J = 7.6, 1.6 Hz, 1H), 7.18 (t, J = 7.6 Hz, 1H), 1.37 (s, 12H); **4-4b**: δ = 7.86 (d, J = 1.4 Hz, 1H), 7.60 (d, J = 8.0, 1.4, 1H), 7.44 (d, J = 8.0, 1H), 1.34 (s,

12H);

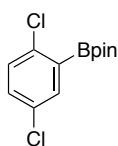
Spectral data were in good agreement with literature values.^{22,23}



4c (99.7 mg, 0.36 mmol, 92%, 2-:4-:5- = 23:40:37). The ratio of regioisomers was determined by ¹H NMR of the isolated product, which was almost identical to that in the crude material.

¹H NMR (CDCl₃, 400 MHz) **2-4c**: δ = 7.20-7.24 (m, 3H), 1.43 (s, 12H); **4-4c**: δ = 7.62 (d, J = 8.2 Hz, 1H), 7.36 (d, J = 2.0 Hz, 1H), 7.20-7.24 (m, 1H), 1.34 (s, 12H); **5-4c**: 7.64 (d, J = 2.0 Hz, 2H), 7.43 (t, J = 2.0 Hz, 1H), 1.36 (s, 12H);

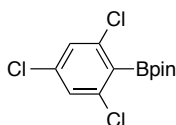
Spectral data were in good agreement with literature values.^{24,25,26}



4d (93.7 mg, 0.34 mmol, 86%)

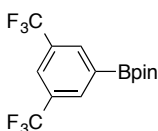
¹H NMR (CDCl₃, 400 MHz) δ = 7.65 (d, J = 2.5 Hz, 1H), 7.34-7.27 (m, 2H), 1.35 (s, 12H);

Spectral data were in good agreement with literature values.²⁷



4e (70.8 mg, 0.23 mmol, 58%)

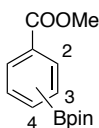
IR(ATR) 1577, 1535, 1323, 1109; ¹H NMR (CDCl₃, 400 MHz) δ = 7.25 (s, 2H), 1.41 (s, 12H); ¹³C NMR (CDCl₃, 100 MHz) δ = 138.6, 136.2, 127.0, 85.2, 24.7; *Anal.* Calcd for C₁₂H₁₄BCl₃O₂: C, 46.89; H, 4.59. Found: C, 47.10; H, 4.77;



4f (83.0 mg, 0.244 mmol, 58%).

¹H NMR (CDCl₃, 400 MHz) δ = 8.23 (s, 1H), 7.93 (d, J = 14.8 Hz, 2H), 1.38 (s, 12H);

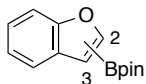
Spectral data were in good agreement with literature values.²⁸



4g (49.6 mg, 0.189 mmol, 47%, 2-:3-:4- = -:58:42). The ratio of regioisomers was determined by ¹H NMR of the isolated product, which was almost identical to that in the crude material.

¹H NMR (CDCl₃, 400 MHz) **3-4g**: δ = 8.47 (s, 1H), 8.13 (dt, *J* = 8.0, 1.6 Hz, 1H), 7.98 (d, *J* = 8.0 Hz, 1H), 7.45 (t, *J* = 8.0 Hz, 1H), 3.92 (s, 3H), 1.36 (s, 3H); **4-4g**: δ = 8.02 (d, *J* = 8.0 Hz, 2H), 7.87 (d, *J* = 8.0 Hz, 2H), 3.92 (s, 3H), 1.36 (s, 3H);

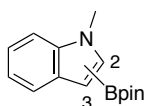
Spectral data were in good agreement with literature values.^{29,30}



5a (91.9 mg, 0.37 mmol, 94%, 2-:3- = 86:14). The ratio of regioisomers was determined by ¹H NMR of the isolated product, which was almost identical to that in the crude material.

¹H NMR (CDCl₃, 400 MHz) **2-5a**: δ = 7.63 (d, *J* = 7.8 Hz, 1H), 7.57 (dd, *J* = 8.3, 0.9 Hz, 1H), 7.40 (d, *J* = 0.9 Hz, 1H), 7.32-7.36 (m, 1H), 7.20-7.26 (m, 1H), 1.39 (s, 12H); **3-5a**: δ = 7.95 (s, 1H), 7.91-7.94 (m, 1H), 7.47-7.52 (m, 1H), 7.23-7.29 (m, 2H), 1.37 (s, 12H);

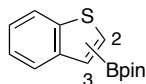
Spectral data were in good agreement with literature values.³¹



5b (69.7 mg, 0.27 mmol, 68%, 2-:3- = 91:9). The ratio of regioisomers was determined by ¹H NMR of the isolated product, which was almost identical to that in the crude material.

¹H NMR (CDCl₃, 400 MHz) **2-5b**: δ = 7.64 (d, *J* = 8.3 Hz, 1H), 7.35 (dd, *J* = 8.3, 0.9 Hz, 1H), 7.26 (td, *J* = 8.3, 0.9 Hz, 1H), 7.13 (s, 1H), 7.08 (td, *J* = 8.3, 0.9 Hz, 1H), 3.97 (s, 3H), 1.37 (s, 12H); **3-5a**: δ = 8.04 (br d, *J* = 7.6 Hz, 1H), 7.52 (s, 1H), 7.25-7.35 (m, 3H), 3.80 (s, 3H), 1.36 (s, 12H);

Spectral data were in good agreement with literature values.^{32,33}

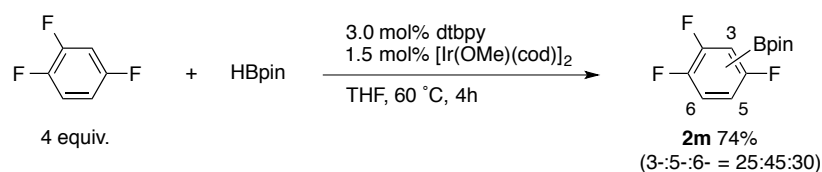


5c (79.2 mg, 0.304 mmol, 76%, 2-:3- = 84:16). The ratio of regioisomers was determined by ¹H NMR of the isolated product, which was almost identical to that in the crude material.

¹H NMR (CDCl₃, 400 MHz) **2-5c**: δ = 7.82-7.92 (m, 3H), 7.32-7.40 (m, 2H), 1.38 (s, 12H); **3-5c**: δ = 8.37 (d, *J* = 7.8 Hz, 1H), 8.07 (s, 1H), 7.88-7.92 (m, 1H), 7.36-7.42 (m, 2H), 1.38 (s, 12H);

Spectral data were in good agreement with literature values.^{34,35}

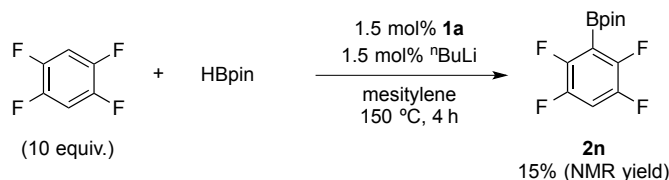
Iridium-catalyzed borylation of 1,2,4-trifluorobenzene (Concerning the footnote d in Figure 1)



To a solution of [Ir(OMe)(cod)]₂ (5.0 mg, 0.0075 mmol) and dtbpy (4.0 mg, 0.015 mmol) in THF (2 mL) was added HBpin (73 μL, 0.5 mmol) and 1,2,4-trifluorobenzene (208 μL, 2.0 mmol) at room temperature. After the

mixture was stirred at 60 °C for 4 h, the solvent was removed under reduced pressure to give a crude product. The yield and ratio of regioisomers were determined by ^{19}F NMR of the crude product using C_6F_6 (10 μL , 0.087 mmol) as an internal standard.

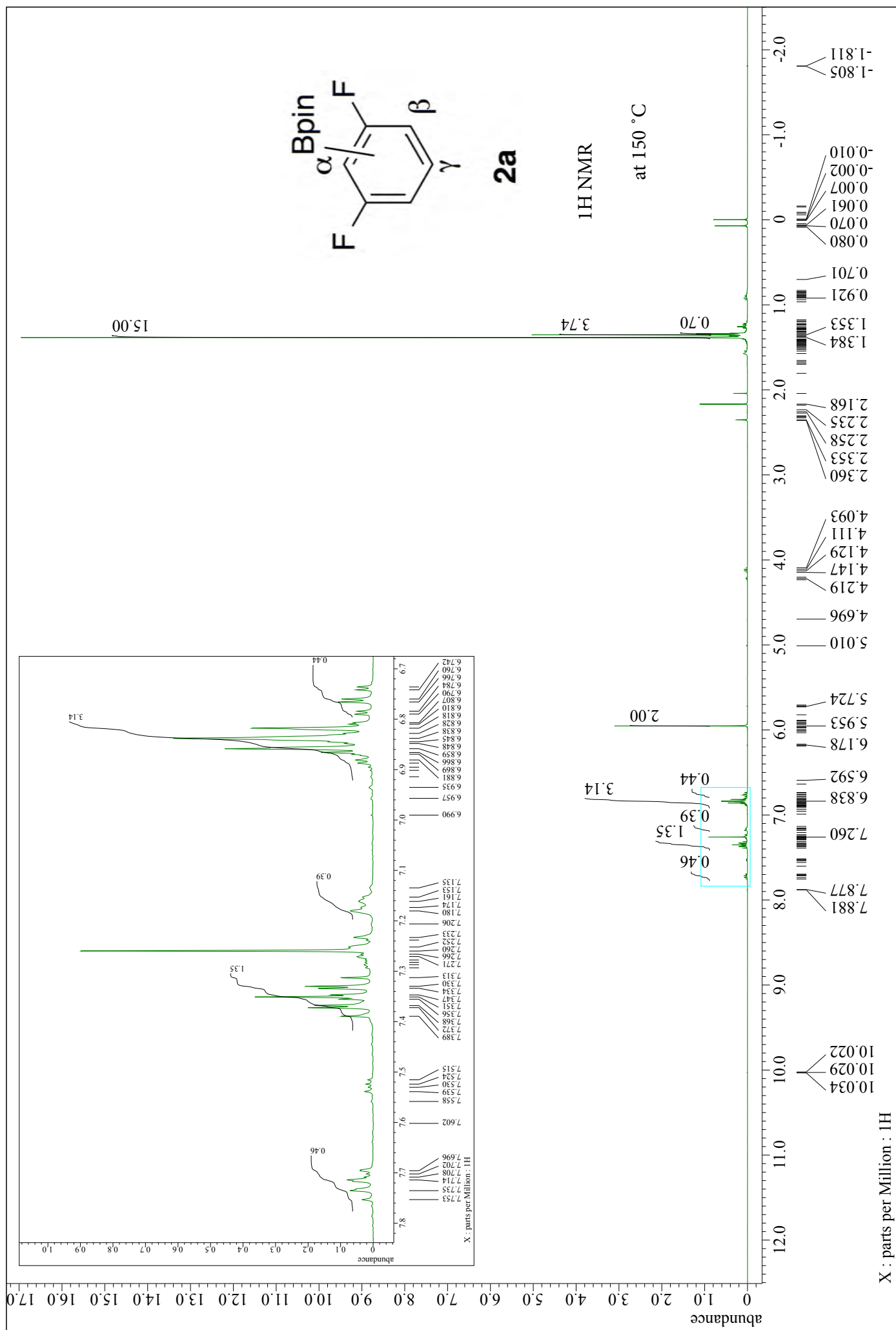
Borylation of 1,2,4,5-tetrafluorobenzene with HBpin instead of B_2pin_2 (Concerning the comment in ref 16.)



To a solution of **1a** (6.81 mg, 0.01 mmol) in mesitylene (2 mL) was added a *n*-hexane solution of *n*-BuLi (1.56 M, 6.4 μL , 0.01 mmol) followed by addition of HBpin (25.9 mg, 0.2 mmol) and 1,2,4,5-tetrafluorobenzene (2 mmol) at room temperature. The mixture was stirred at 150 °C for 4 h. After the solvent was removed under reduced pressure, formation of **2n** in 15% yield was confirmed by ^{19}F NMR. Therefore, it was demonstrated that HBpin instead of B_2pin_2 could be utilized as an atom economical boron source for this reaction, and further investigation on this is in progress.

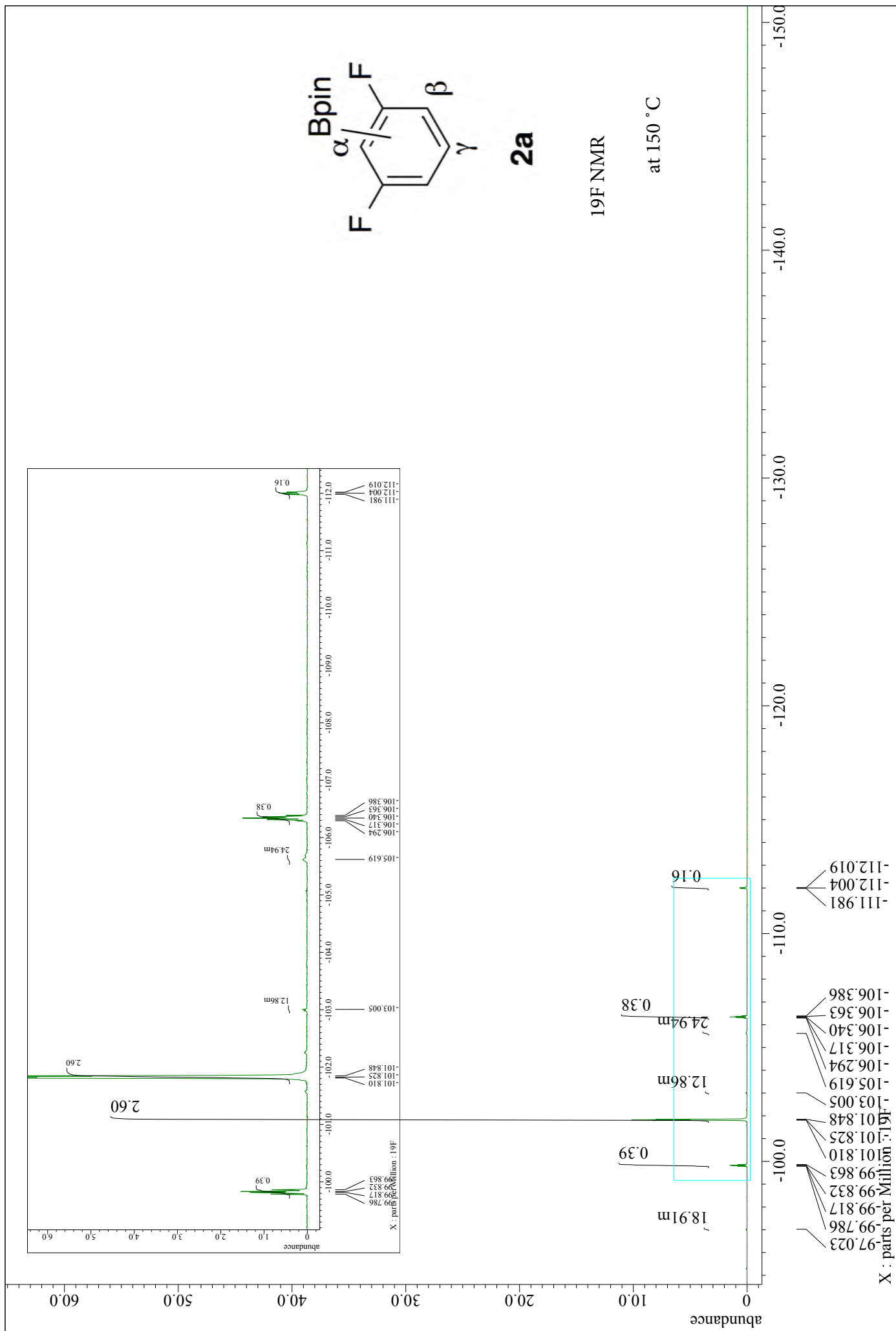
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- ¹⁰ A. Del Grosso, J. Ayuso Carrillo and M. J. Ingleson, *Chem. Commun.*, 2015, **51**, 2878.
- ¹¹ Ref 8.
- ¹² C.-W. Hsu, S.-T. Ho, K.-L. Wu, Y. Chi, S.-H. Liu and P.-T. Chou, *Energy Environ. Sci.*, 2012, **5**, 7549.
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- ¹⁴ Ref 8.
- ¹⁵ Ref 10.
- ¹⁶ J.-Y. Cho, C. N. Iverson and M. R. Smith, *J. Am. Chem. Soc.*, 2000, **122**, 12868.
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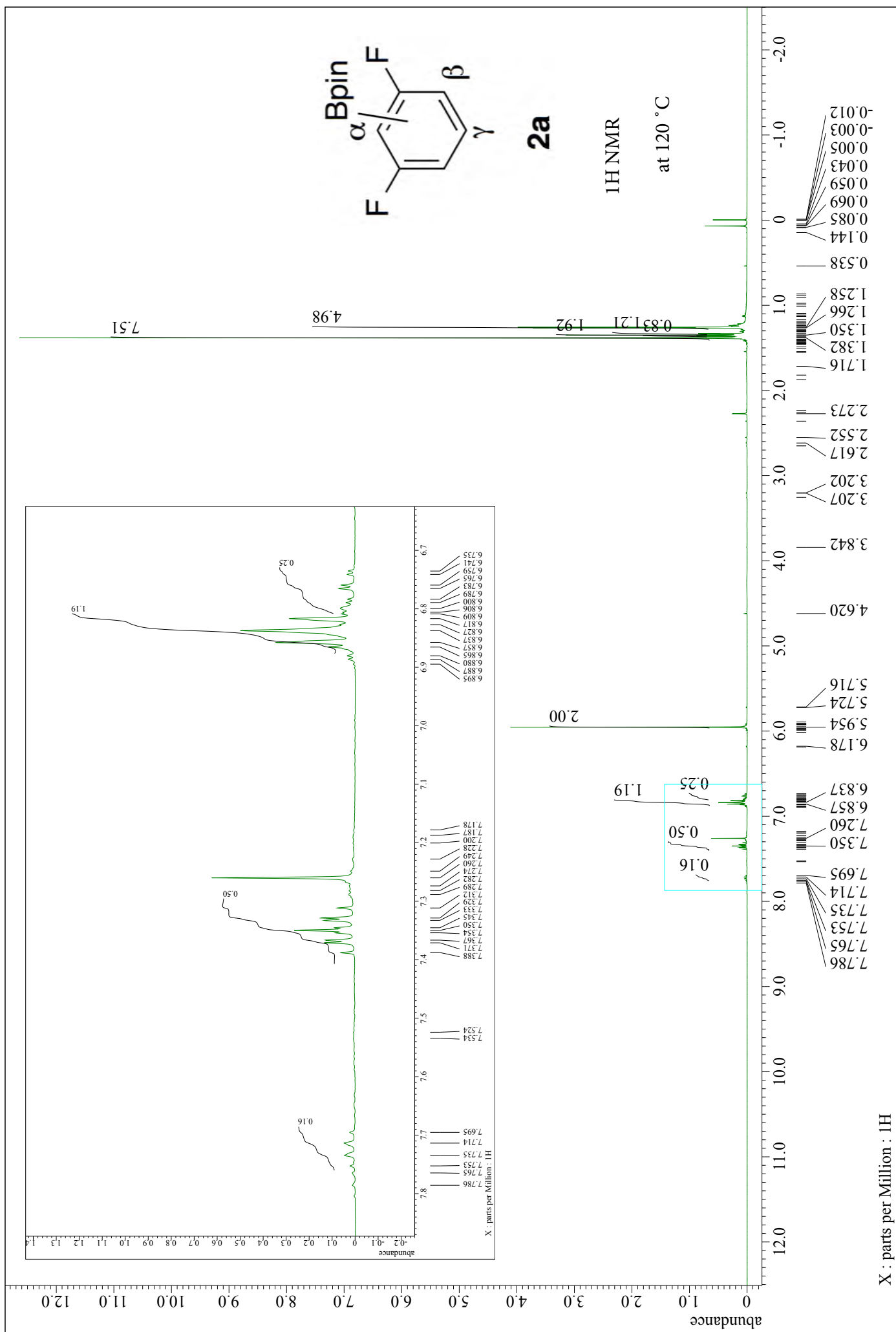
-
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with 1,1,2,2-tetrachloroethane (5.9 ppm)
as an internal standard

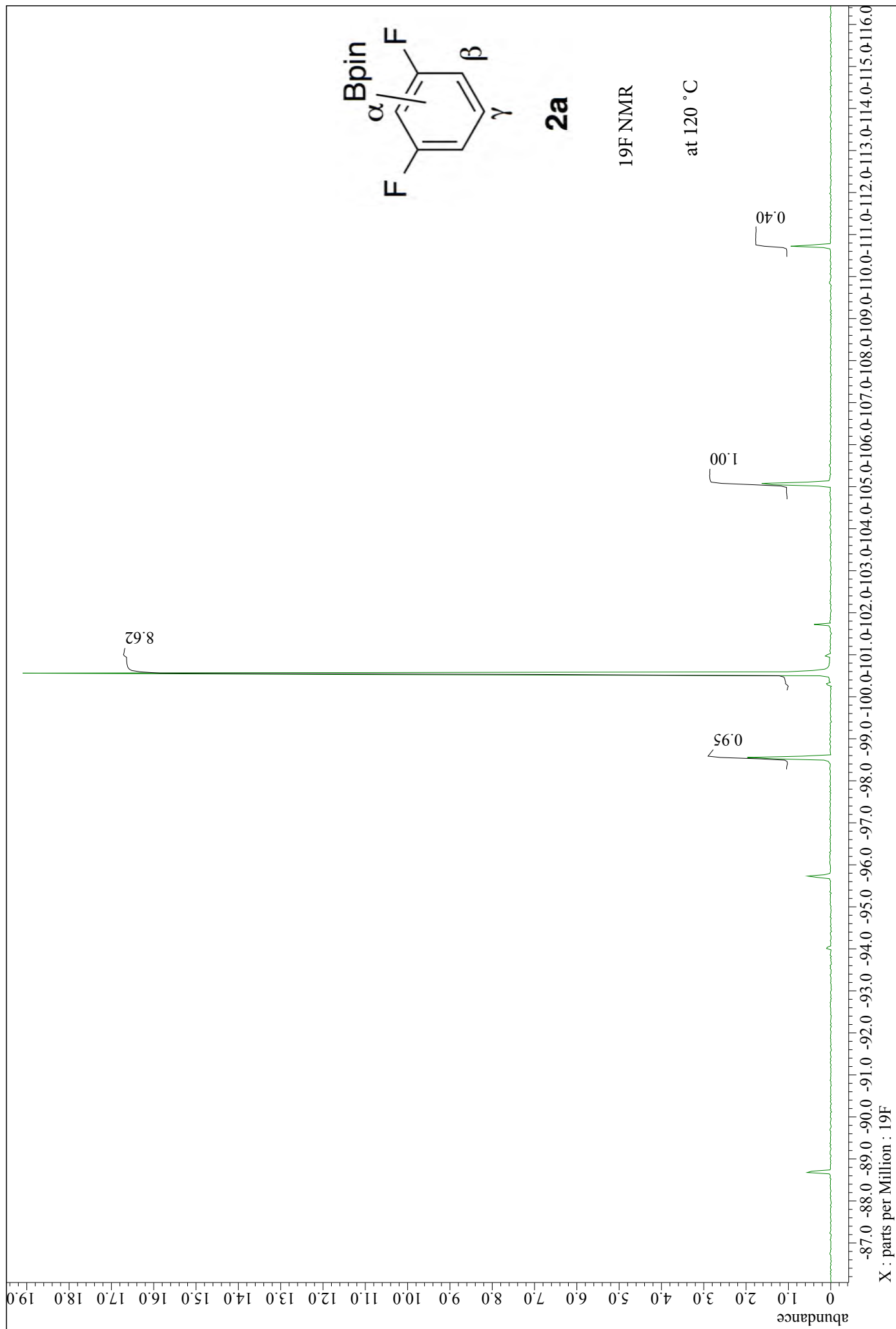
X : parts per Million : 1H

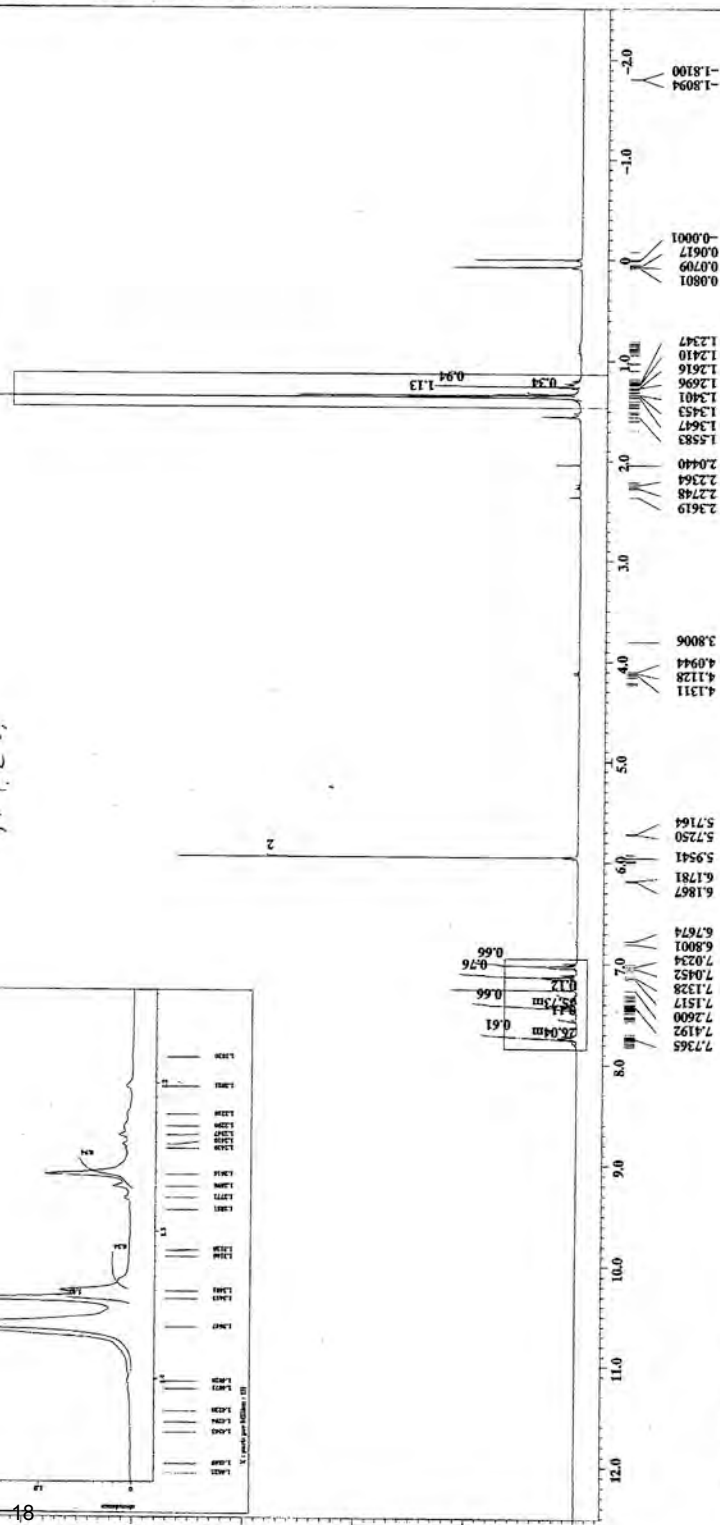
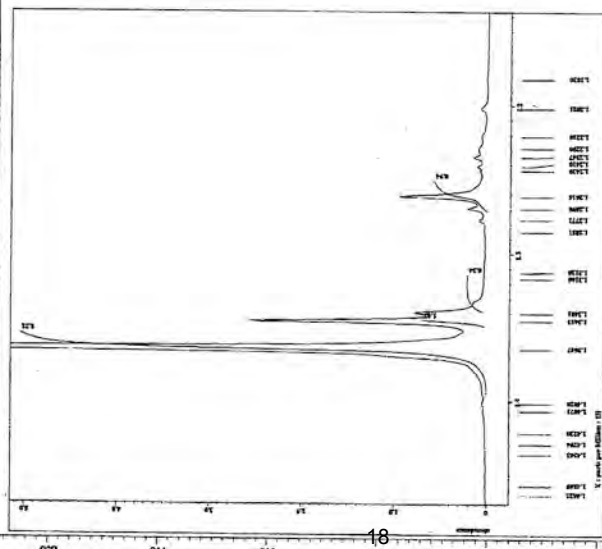
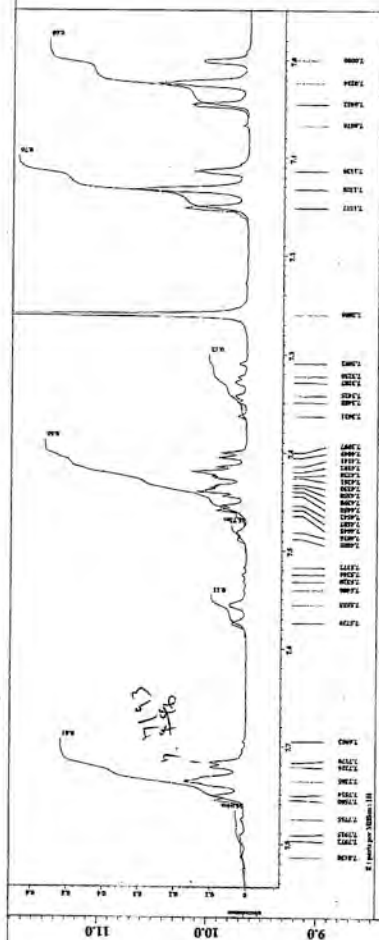




with 1,1,2,2-tetrachloroethane (5.9 ppm)
as an internal standard

X : parts per Million : 1H



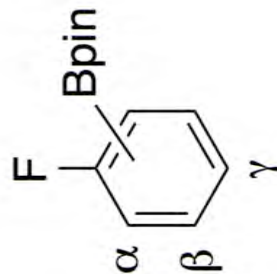


```

PROCESSING PARAMETERS
dc_balance : 0 : FALSE
sawp : 0.2[Hz] : 0.0[s]
trapoids : 0 [%] : 80 [%] : 100 [%]
zerofold : 1
fit : 1 : TRUE
machinephase
ppm
phase : 0 : 0 : 50 [%]
Derived from: ex110 columnmk-1.1.job

```

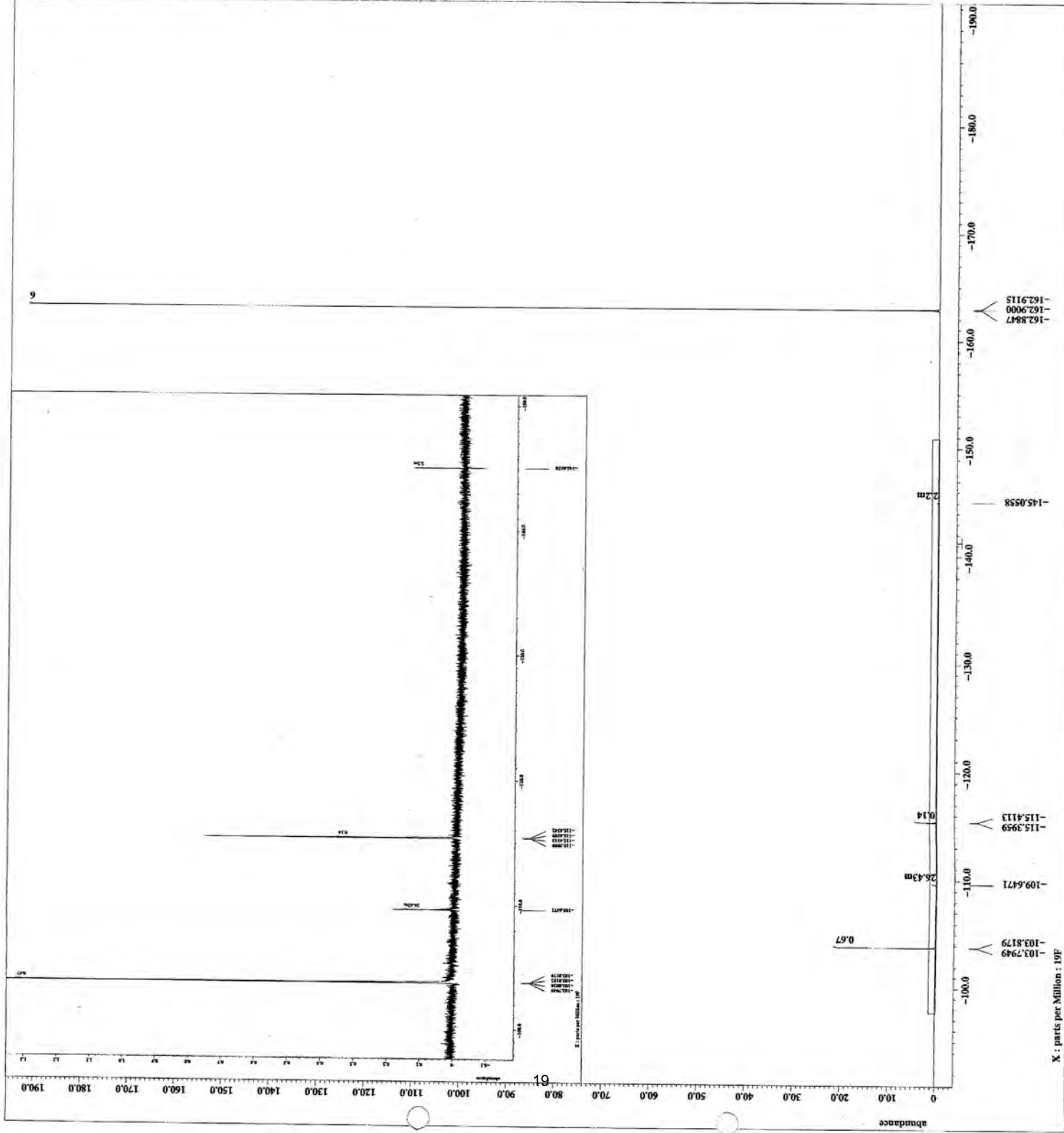
Filename	ex10 column-H-5.fdr
Author	
Experiment	delta_pulse.es2
Sample_id	A
Solvent	CHLOROFORM-D
Time	8-OCT-2014 11:25:17
Revision	8-OCT-2014 11:25:17
Revision_time	8-OCT-2014 11:25:17
Current_time	
Comment	
Data_format	single_pulse
Dia_title	NO COMPLEX
Dia_units	Hz
Dimensions	1D
Site	X
Spectrometer	ECK 400
Field_strength	MAGNA_NMR
X_acq_duration	9.201297668[s] (390 MHz)
X_domain	4.4556448[s]
X_offset	1H
X_pulses	371.78655441 [MHz]
X_resolution	32768
X_swept	1
X_tan	0.2449396 [Hz]
Xir_freq	15.3529411 [kHz]
Xir_offset	15
F1_domain	391.78655441 [MHz]
F1_offset	5 [ppm]
F1_freq	1H
F1_resolution	391.78655441 [MHz]
F1_swept	1
F1_tan	FALSE
Mod_return	1
Scans	8
Total_scans	8
X_acq_time	9.7[us]
X_angle	4.45648[s]
X_atn	45[deg]
X_pulse	3[dB]
X_phase	4.98[us]
Pri_mode	OFF
Delta_presat	FALSE
Initial_wait	1[s]
Acsvr_gain	42
Demodulation_delay	9.456448[s]
Demodulation_time	9.456448[s]
Pump_get	25[GC]

¹H NMR

2b at 150 °C

X : parts per Million : 1H

with 1,1,2,2-tetrachloroethane (5.9 ppm) as an internal standard

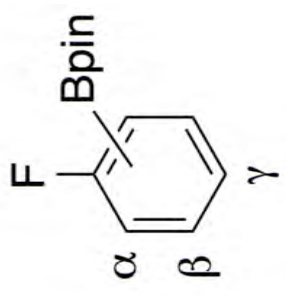


X : parts per Million : 19F
with hexafluorobenzene (-162.9 ppm)
as an internal standard

----- PROCESSING PARAMETERS -----
ac_balance : 0 : FALSE
sweep : 0.2 [Hz] : 0.0 [s]
sweep_start : 0 [Hz] : 80 [Hz] : 100 [Hz]
sweep_end : 0 [Hz] : 80 [Hz] : 100 [Hz]
fft : 1 : TRUE : TRUE
machinphase :
ppm
Derived from: ex110 column 19F-1.drf

=====
File Name : ex110 column 19F-5.f
Author :
Experiment :
Sample ID :
Solvent : CHLOROFORM-D
Creation Time : 8-OCT-2014 17:11:00
Revision Time : 8-OCT-2014 18:17:00
Current Time : 8-OCT-2014 18:17:09
Comment :
Data Format : 1D COMPLEX
Dir Size : 26214
Dir Title : 19F
Dir Units : [ppm]
Dir Dimensions :
Dir Size : 26214
Spectrometer : DELTA2_NMR
Field Strength : 9.20157068 [T] (390 [MHz])
X_acq_duration : 12.7077888 [s]
X_acq_start : 368.64763285 [MHz]
X_freq : -140 [ppm]
X_offset : 32768
X_points : 1
X_prescans : 1
X_resolution : 4328584 [Hz]
X_sweep : 46.2952563 [MHz]
X_domain : 19F
Irr_freq : 368.64763285 [MHz]
Irr_offset : 5 [ppm]
Irr_domain : 19F
Tri_domain : 19F 64763285 [MHz]
Tri_offset : 5 [ppm]
Clipped : FALSE
Mod_return : 1
Scans : 16
Total_scans : 16
X_90_width : 11.5 [us]
X_acq_time : 0.7077888 [s]
X_angle : 45 [deg]
X_atn : 3.4 [dB]
X_pulse : 8.75 [us]
X_pulse_prog : OFP
Tri_mode : OFP
Data_presat : FALSE
Initial_wait : 1 [s]
Recvr_gain : 58
Relaxation_delay : 5 [s]
Relaxation_time : 1.077888 [s]
Temp_set : 24.7 [degC]

19F NMR

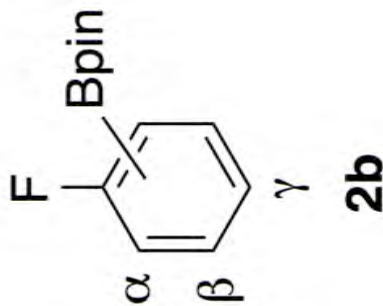


2b

at 150 °C

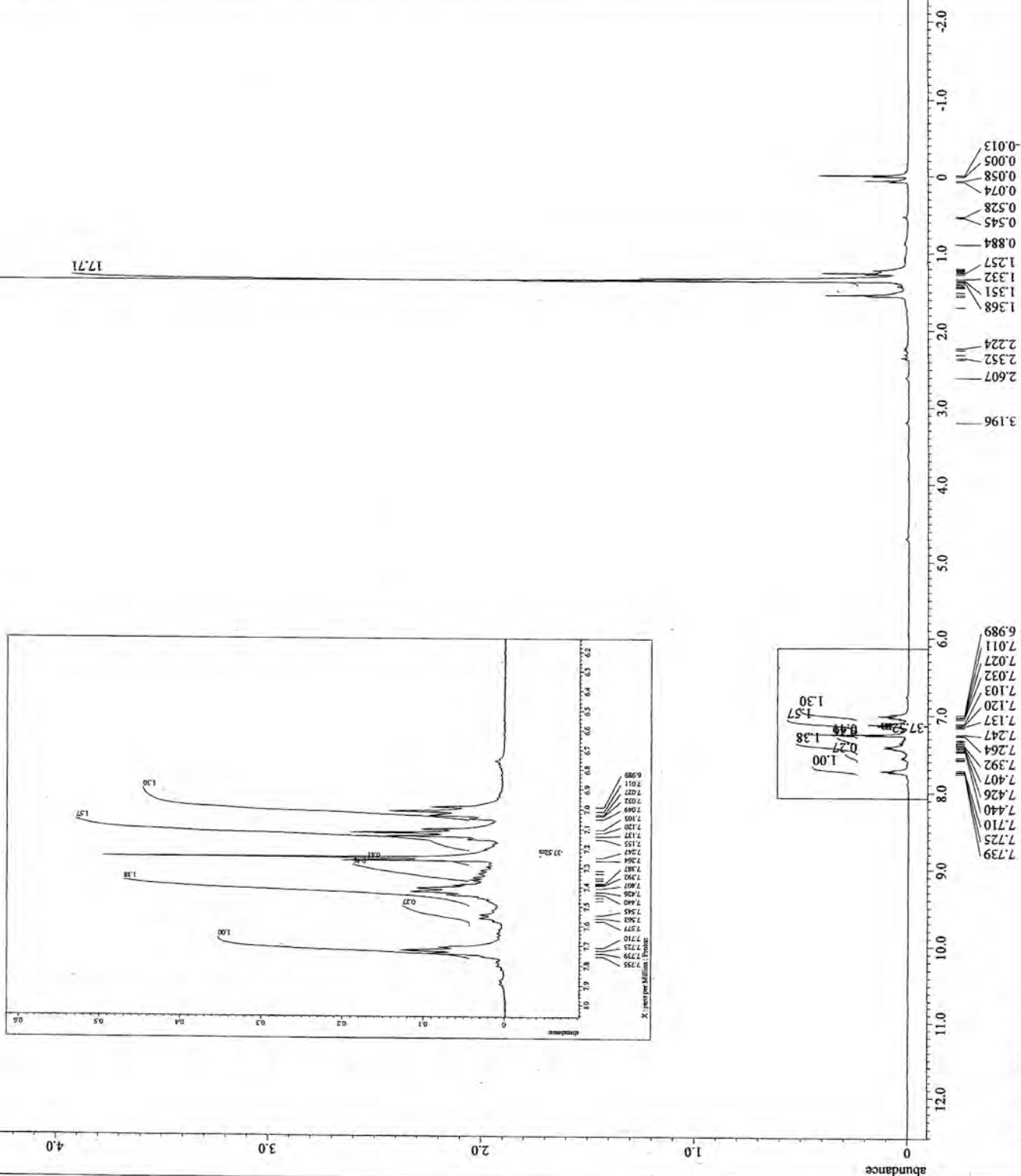
Filename = ex282 column_Proton-1-3.j
 Sample = 1
 Experiment = proton_1xp
 Sample Id = ex282 column
 Solvent = CHLOROFORM-D
 Creation_Time = 21-JUL-2015 14:15:46
 Revision_Time = 21-JUL-2015 14:17:28
 Current_Time = 21-JUL-2015 14:17:34
 Comment = single pulse
 Data Format = 1D COMPLEX
 Dim Size = 26214
 Dim Title = Proton
 Dim Units = (ppm)
 Dimensions = 1
 File Name = 30A-RCS400
 Spectrometer = DELTA2_NMR
 Field Strength = 9.389766[T] (400[MHz])
 X_Acq_Duration = 4.36731904[s]
 X_Domain = 1H
 X_Freq = 399.78219838[MHz]
 X_P1 = 32768
 X_P2 = 32768
 X_P3 = 1
 X_Resolution = 0.22897343[Hz]
 X_Sweep_Clip = 7.5030012[MHz]
 X_Sweep_Domain = 6.00240096[MHz]
 X_Sweep_Time = 5.17978219838[MHz]
 X_T1_Offset = 5.17978219838[MHz]
 X_T1_Domain = Proton
 X_T1_Freq = 399.78219838[MHz]
 X_T1_Offset_Clip = 5.17978219838[MHz]
 X_T1_Offset_Clip = FALSE
 X_T1_Offset_Clip = 8
 Total_Scans = 8
 Relaxation_Delay = 5[s]
 Recvr Gain = 40
 Temp_Get = 26.7[deg]
 X_90_Width = 6.15[us]
 X_Acq_Time = 4.36731904[s]
 X_Angle = 45[deg]
 X_Pulse = 1[us]
 X_P2 = 3.075[us]
 X_P3 = Off
 X_T1_Mode = Off
 X_T1_Offset = FALSE
 X_T1_Offset = 1[s]
 Repetition_Time = 9.36731904[s]

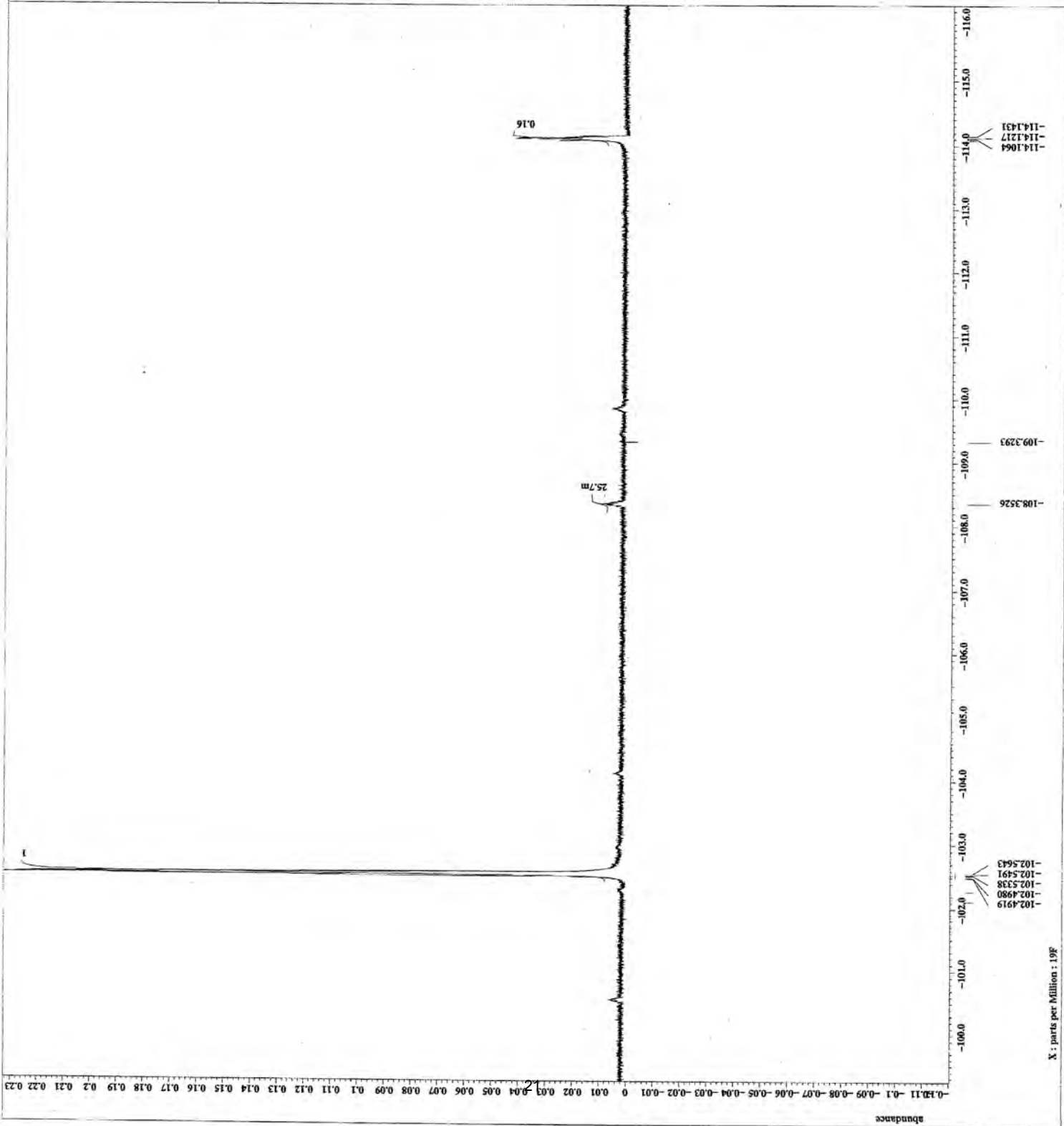
¹H NMR



at 120 °C

X : parts per Million : Proton



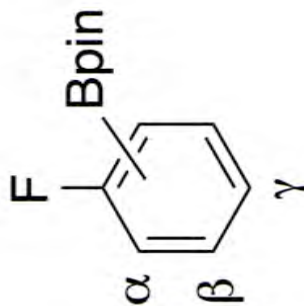


----- PROCESSING PARAMETERS -----
 dc_balance : 0 : FALSE
 smp : 0.2 [Hz] : 0.0 [s]
 trapd3 : 0 [Hz] : 80 [Hz] : 100 [Hz]
 RecvOff : 0.000000
 #f1 : 1 : TRUE : TRUE
 machinephase
 ppm
 Derived from: sm338 date 19F-1.jdc

28

File: sm338 date 19F-1.jdc
 Author: delta
 Experiment: single_pulse.acq
 SampleId: CHLOROFORM-D
 Solvent: CHLOROFORM-D
 Creation_time: 29-JUL-2015 16:23:33
 Revision_time: 29-JUL-2015 18:35:29
 Current_time: 29-JUL-2015 18:35:41
 Comment: single_pulse
 Data_format: 1D COMPLEX
 Dia_size: 52428
 Dia_title: 19F
 Dia_units: [ppm]
 Dimensions: 1
 Size: 824 400
 Spectrometer: DELTA3_NMR
 Field_strength: 9.2017068 [T] (390 [MHz])
 X_acq_duration: 3.55467266 [s]
 X_delay: 1.00 [s]
 X_freq: 368.64763285 [MHz]
 X_offset: -108 [ppm]
 X_points: 65536
 X_prescans: 1
 X_resolution: 0.2811986 [Hz]
 X_sweep_rate: 4.3057817 [Hz]
 Irr_domain: 19F
 Irr_freq: 368.64763285 [MHz]
 Irr_offset: 5 [ppm]
 Tri_domain: 19F
 Tri_freq: 368.64763285 [MHz]
 Tri_offset: 5 [ppm]
 Mod_return: FALSE
 Total_scans: 1
 Scans: 84
 X_90_width: 11.5 [us]
 X_acq_time: 3.55467266 [s]
 X_angle: 45 [deg]
 X_atn: 3.4 [dB]
 X_pulse: 5.75 [us]
 Irr_mode: Off
 Tri_mode: Off
 Densit_presat: FALSE
 Initial_wait: 1 [s]
 Recv_gain: 54
 Relaxation_delay: 5 [s]
 Spectrometer: DELTA3_NMR
 Temperature: 23.6 [°C]
 Temp_set: 23.6 [°C]

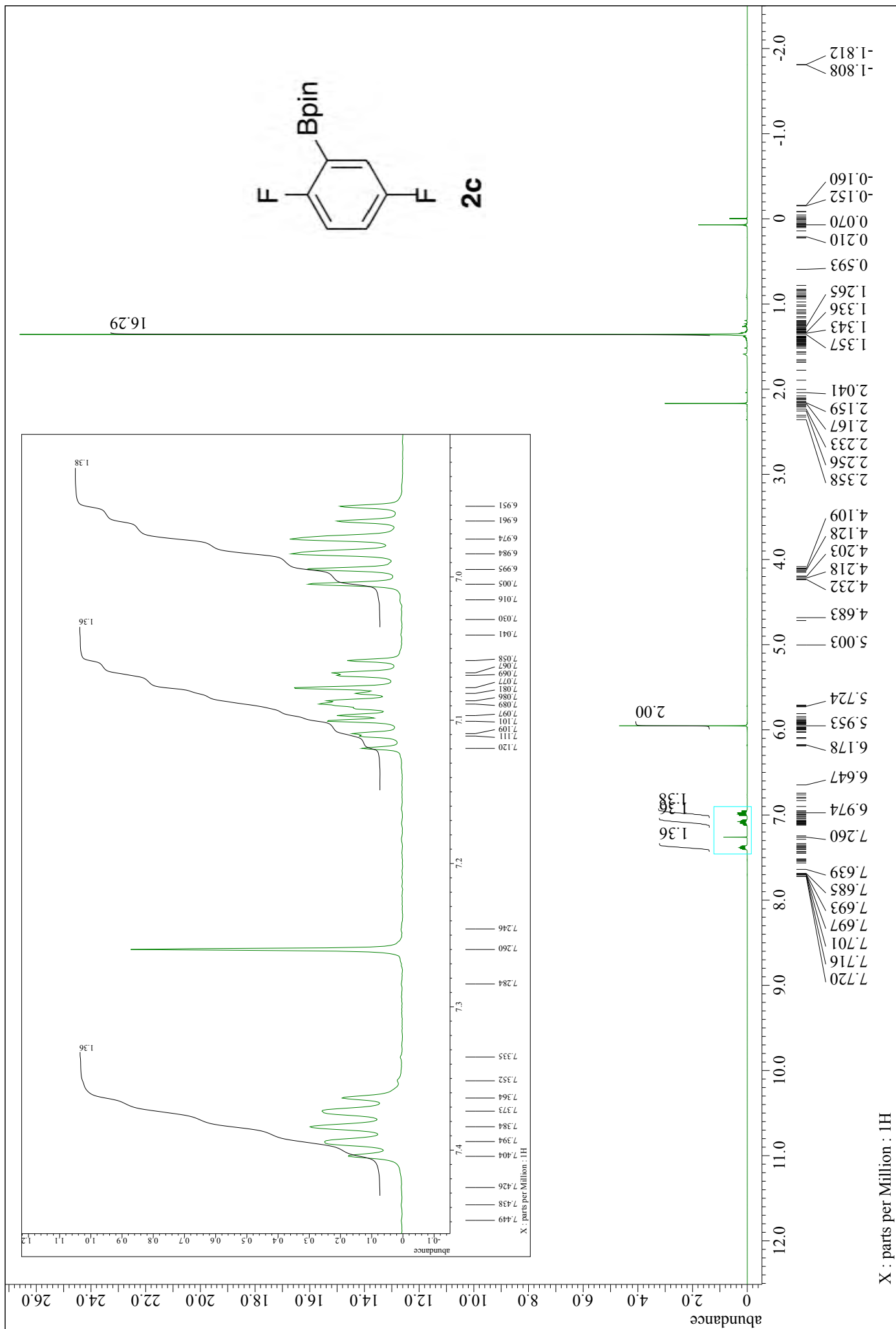
19F NMR

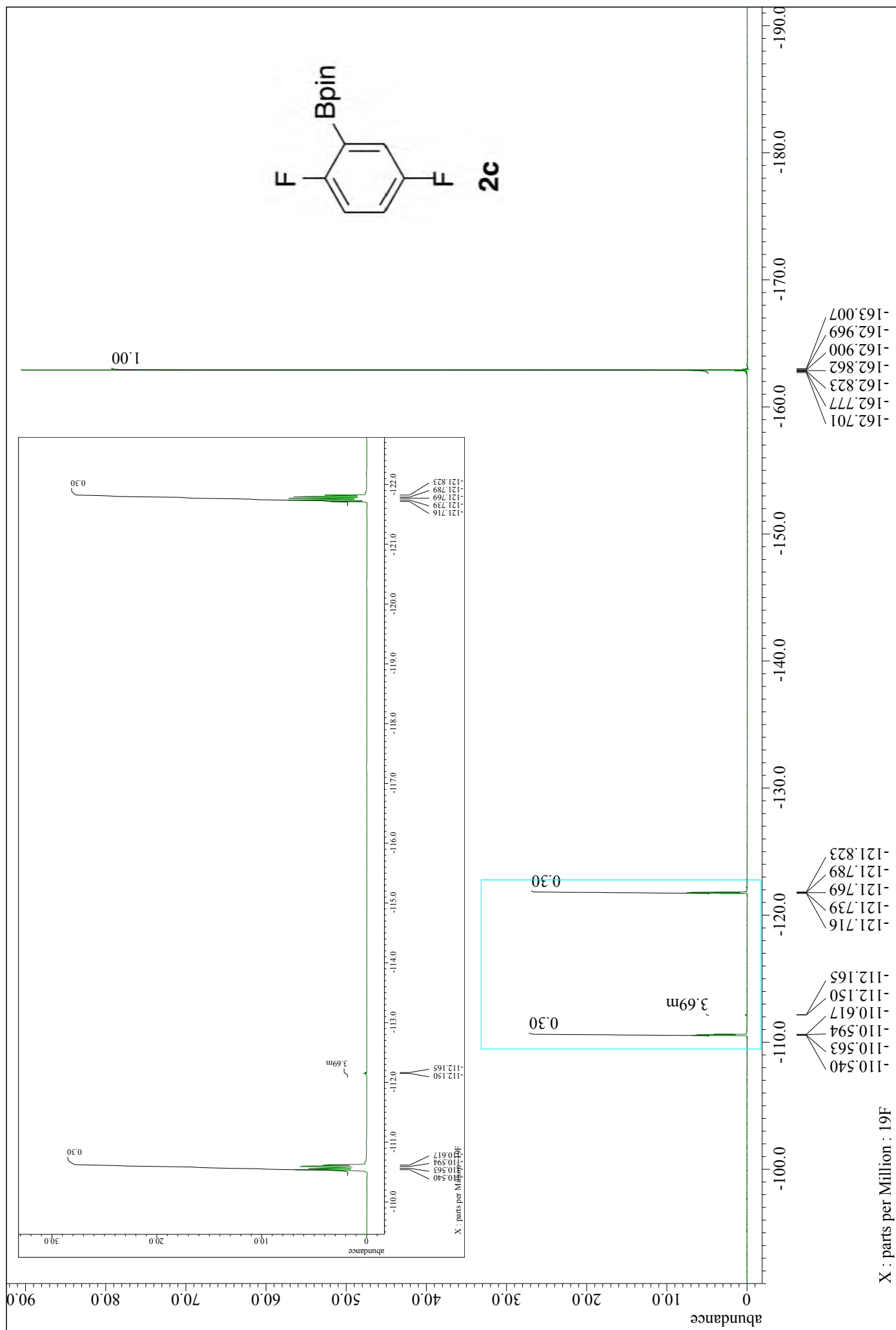


2b

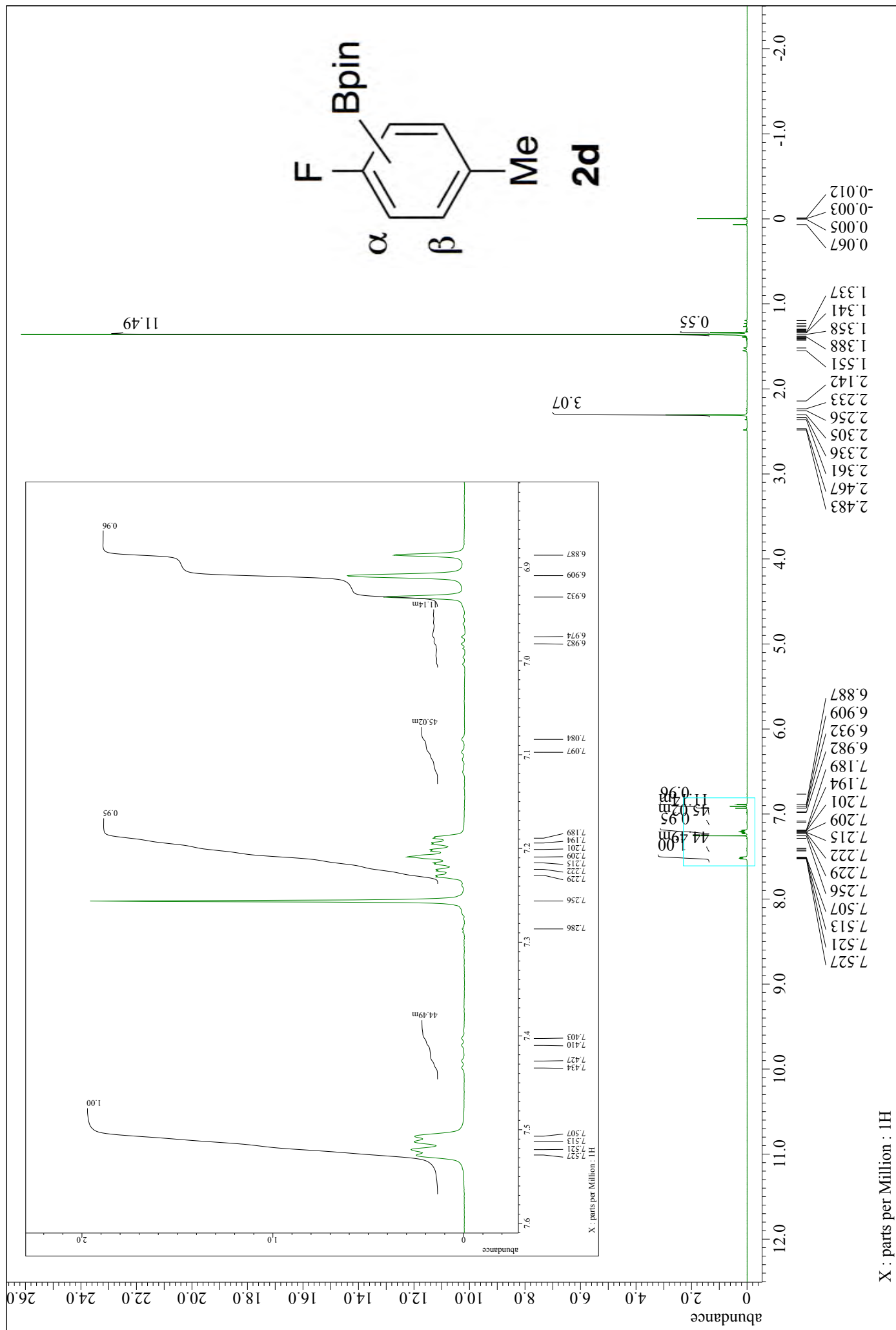
at 120 °C

with 1,1,2,2-tetrachloroethane (5.9 ppm)
as an internal standard





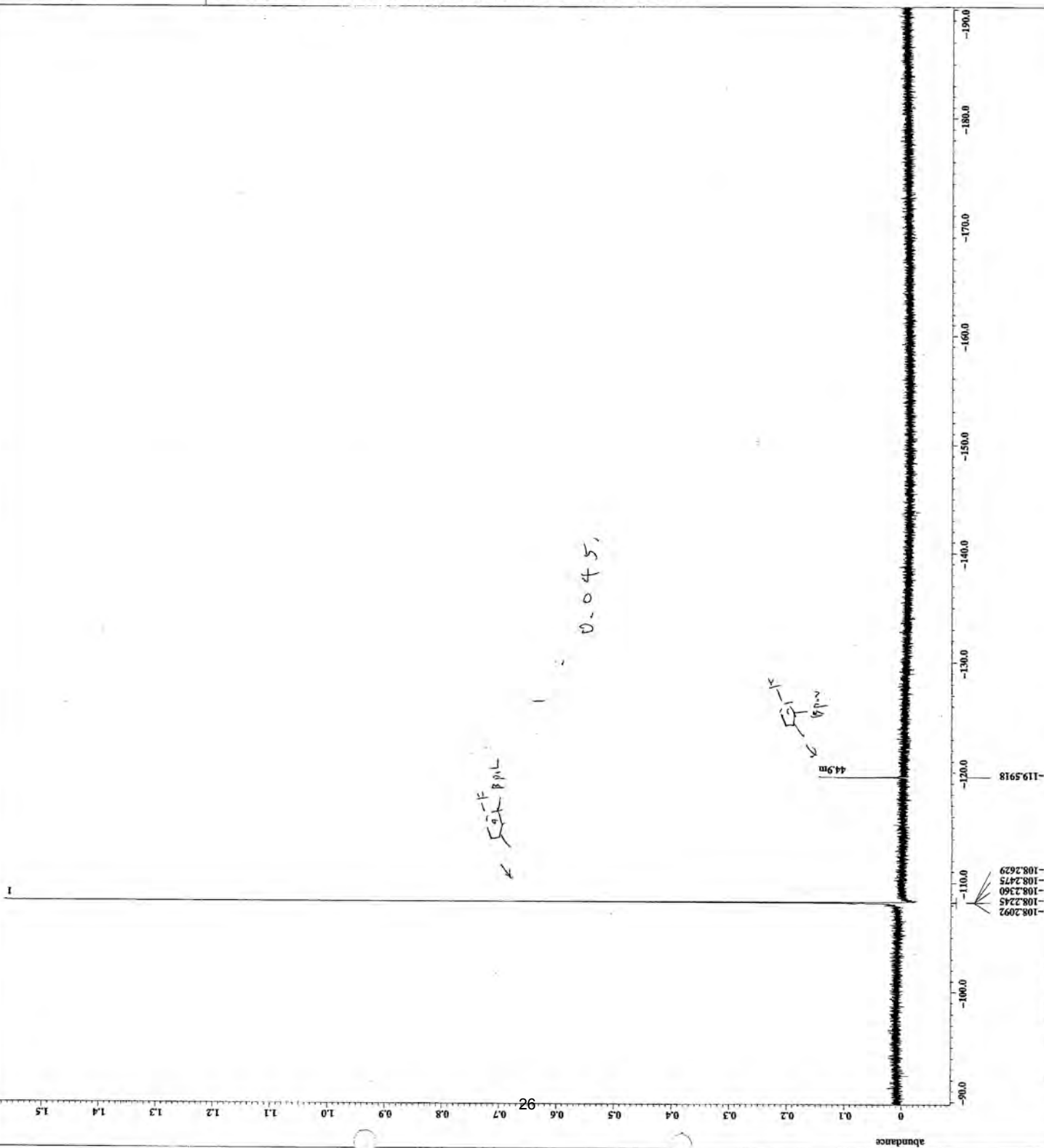
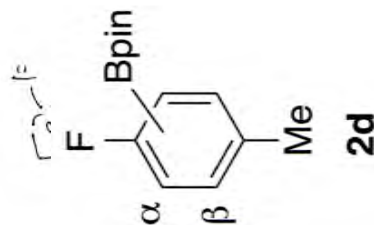
with hexafluorobenzene (-162.9 ppm)
as an internal

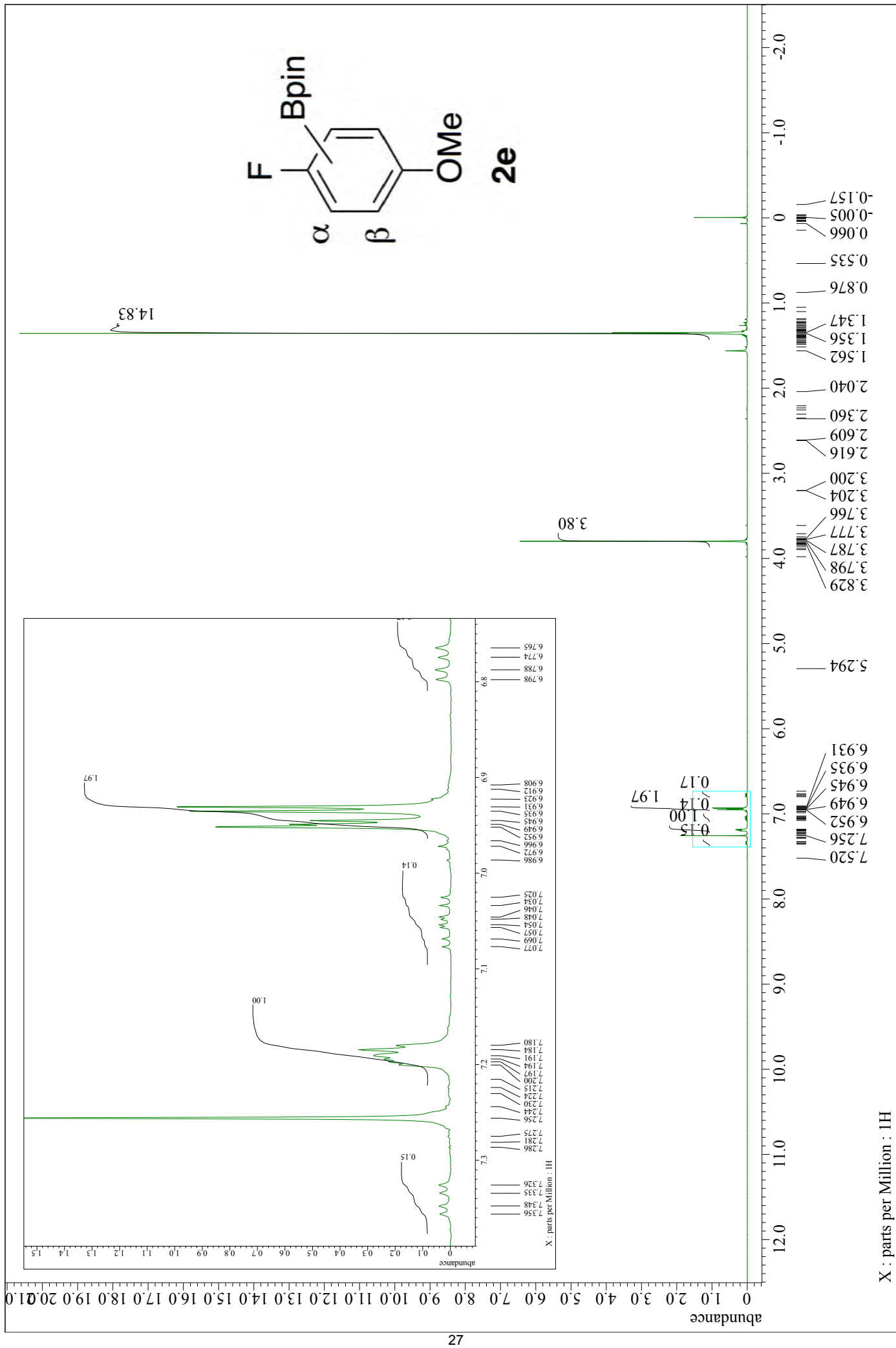


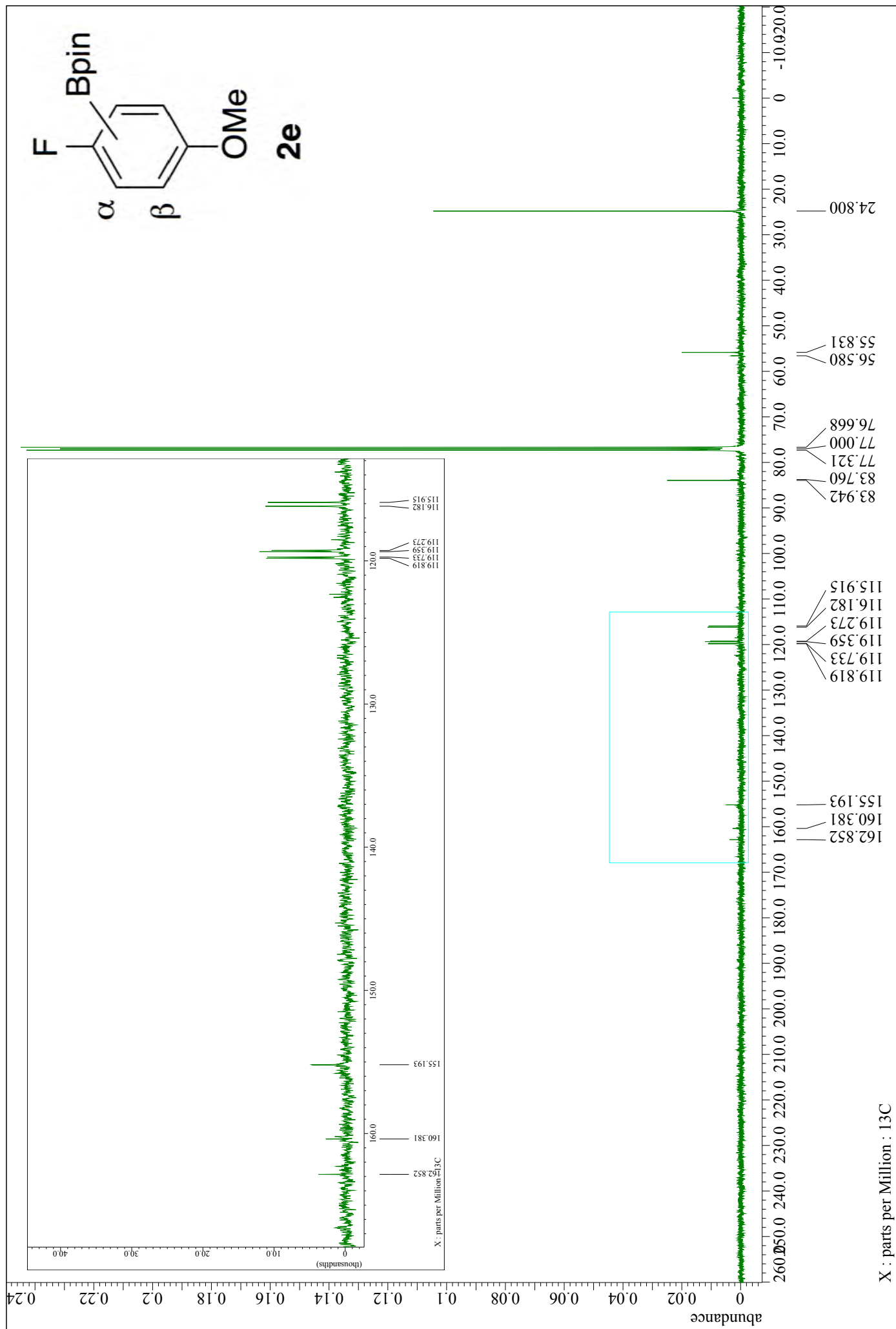
----- PROCESSING PARAMETERS -----
 Ac_balance : 0 : FALSE
 keep : 0.2 [Hz] : 0.0 [s]
 threshold : 0 [Hz] : 80 [Hz] : 100 [Hz]
 zerofill : 1
 zft : 1 : TRUE : TRUE
 machinphase
 ppm
 phase : 3.75 : 0 : 18.38331 [Hz]
 Derived from: ex166 crude 19F-14.jff

ex166 crude 19F-14.jf
 File name
 Author
 Experiment
 Sample_id
 Solvent
 Creation_time
 Revision_time
 Current_time
 Comment
 Data_format
 Dia_size
 Dia_title
 Dia_units
 Dimensions
 Site
 Spectrometer
 Field_strength
 X_acq_time
 X_domain
 X_freq
 X_offset
 X_points
 X_prescans
 X_resolution
 X_sweep
 Irr_domain
 Irr_freq
 Irr_offset
 Trf_domain
 Trf_freq
 Trf_offset
 Clipped
 Mod_return
 Scans
 Total_scans
 X_90_width
 X_acq_time
 X_angle
 X_atn
 X_phase
 X_pulse
 Trf_mode
 Dante_preset
 Initial_wait
 Recvz_gain
 Recvz_delay
 Repetition_time
 Temp_get

19F NMR

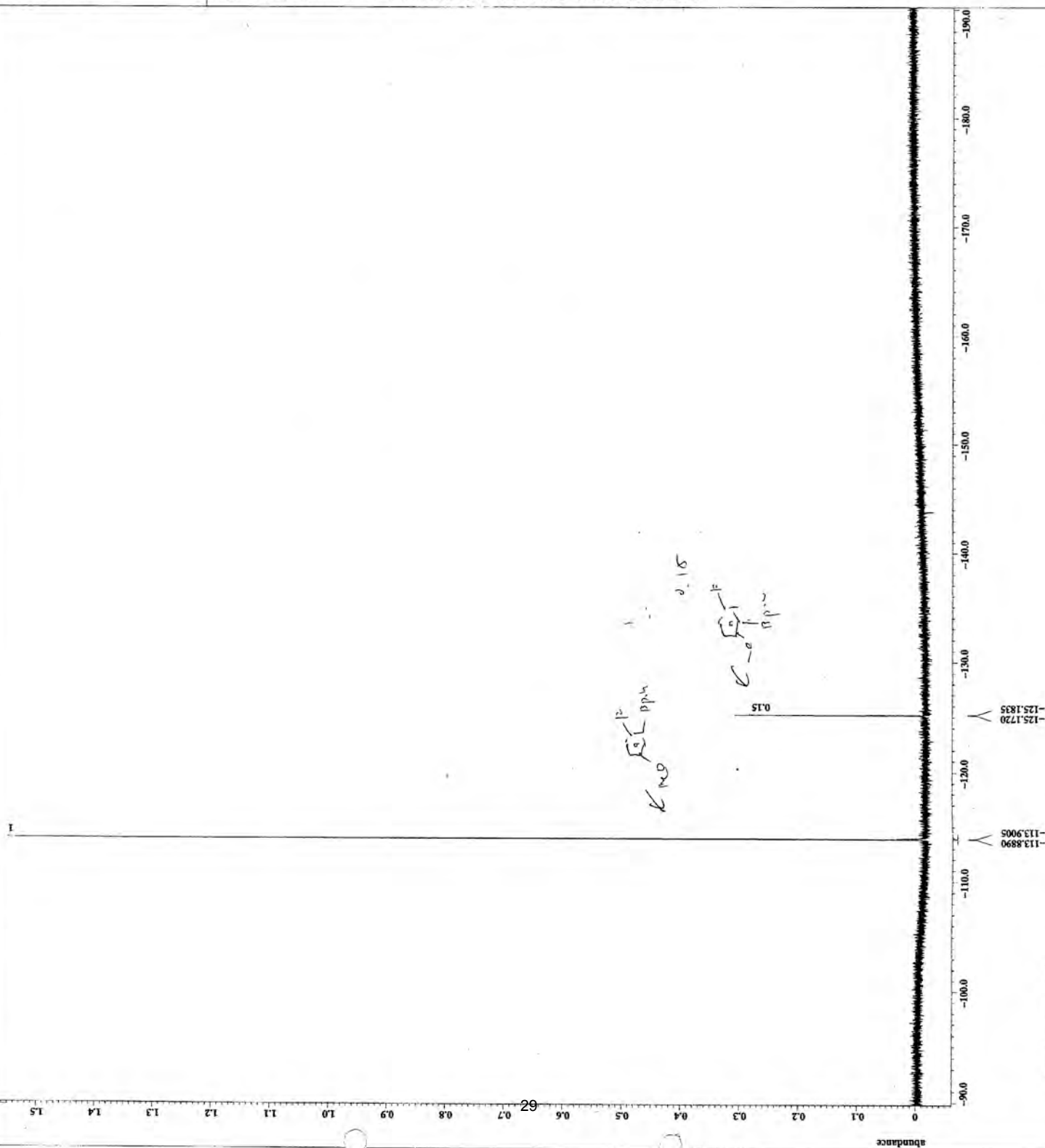
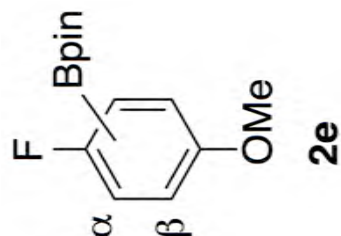






colated

¹⁹F NMR



```

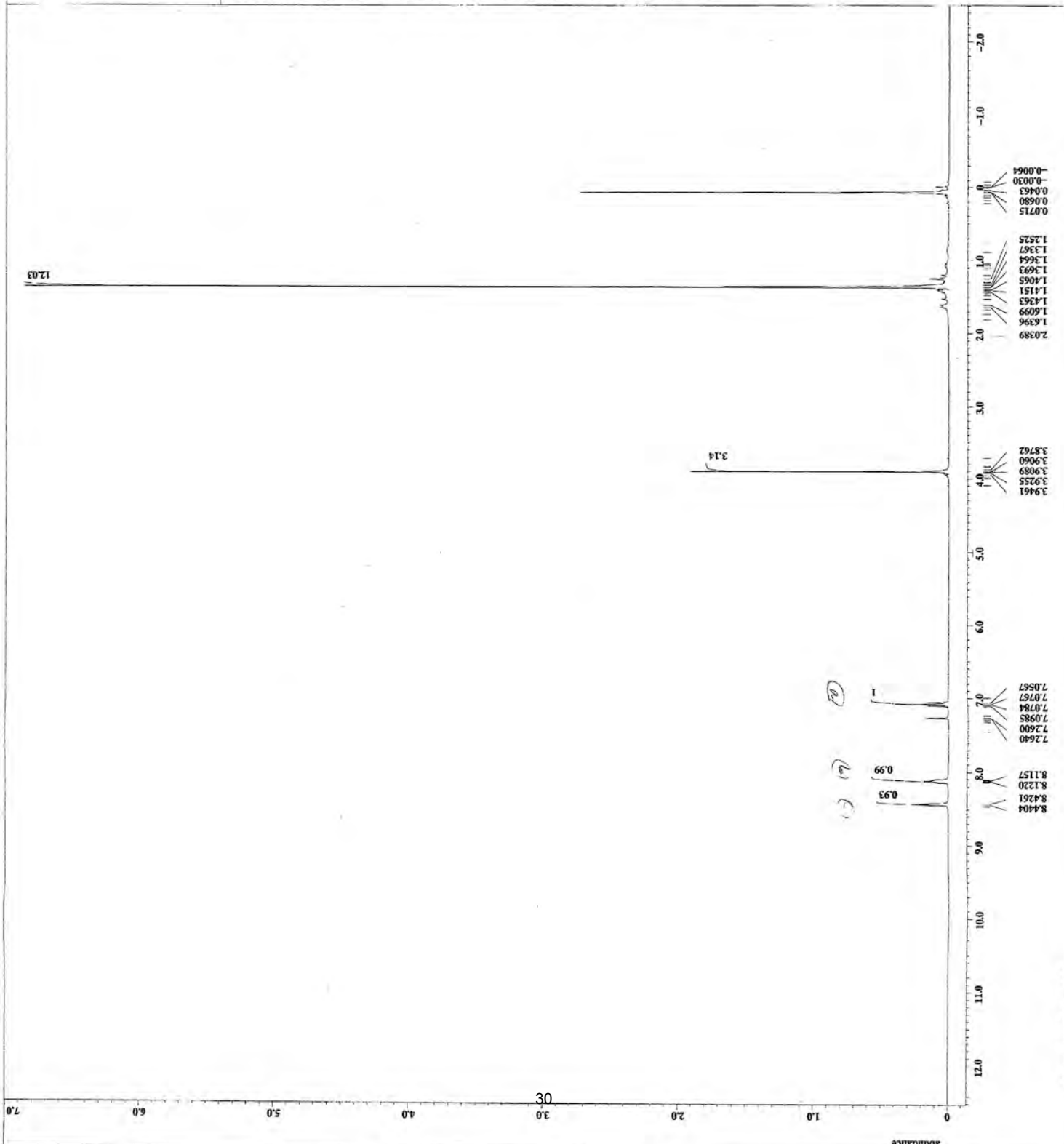
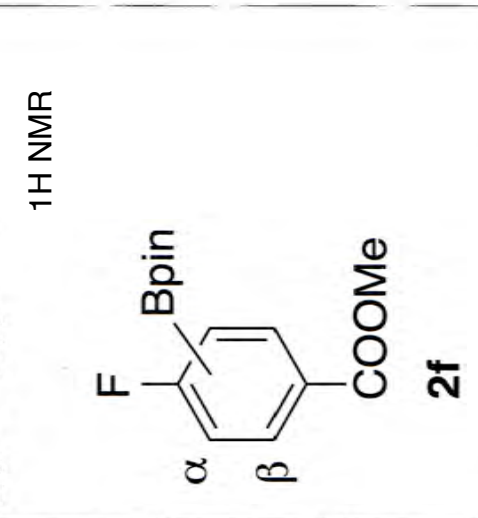
--- PROCESSING PARAMETERS ---
dc_balance : 0 : FALSE
temp : 300.2 [K]
tempco : 0.01 [K]
sfs : 100 [Hz]
sfsco : 0.01 [Hz]
sfsf11 : 1
fft : 1 : TRUE
machinephase :
Phase : 0 : 0 : 50 [Hz]
Derived from: ex247 date 1H-1.jdf

```

```

File: 1H-1.jdf
Acq: 1H-1.jdf
Experiment:
Sample: A
Solvent: CHLOROFORM-D
Creation_time: 1-MAY-2015 16:48:23
Current_time: 1-MAY-2015 18:24:28
Comment:
Data_format: single_pulse
Data_size: 16384
Data_units: [ppm]
Dimensions: X
Site: ECK 400
Spectrometer: DELTA2_NMR
Field_strength: 9.20197069 [T] (390.136 MHz)
X_acq_duration: 4.456448 [s]
X_domain: 1H
X_freq: 391.78655441 [MHz]
X_offset: 5 [ppm]
X_points: 32768
X_resolution: 0.22439396 [Hz]
X_sweep: 7.35294118 [kHz]
X1_domain: 1H
X1_freq: 391.78655441 [MHz]
X1_offset: 5 [ppm]
X1_resolution: 0.22439396 [Hz]
X1_sweep: 7.35294118 [kHz]
X2_domain: 1H
X2_freq: 391.78655441 [MHz]
X2_offset: 5 [ppm]
X2_resolution: 0.22439396 [Hz]
X2_sweep: 7.35294118 [kHz]
Mod_return: FALSE
Total_scans: 1
X90_width: 9.33 [us]
X90_time: 4.456448 [s]
X_angle: 45 [deg]
X_coupling: 1.965 [us]
X_pulse: 1.965 [us]
X1_mode: Off
X2_mode: Off
Data_preset: F2
Initial_wait: 1 [s]
Relaxation_delay: 5 [s]
Repetition_time: 9.456448 [s]
Temp_get: 25 [deg]

```



```

----- PROCESSING PARAMETERS -----
dc_balance : 0 : FALSE
sweep : 0.2 [Hz] : 0.0 [s]
tempset1 : 1 : 0 [°C] : 80 [°C] : 100 [°C]
sacch1 : 1 : 0 [°C] : 80 [°C] : 100 [°C]
fft : 1 : TRUE : TRUE
machinename :
ppm
Derived from: ex247 date 19f-1.jdf

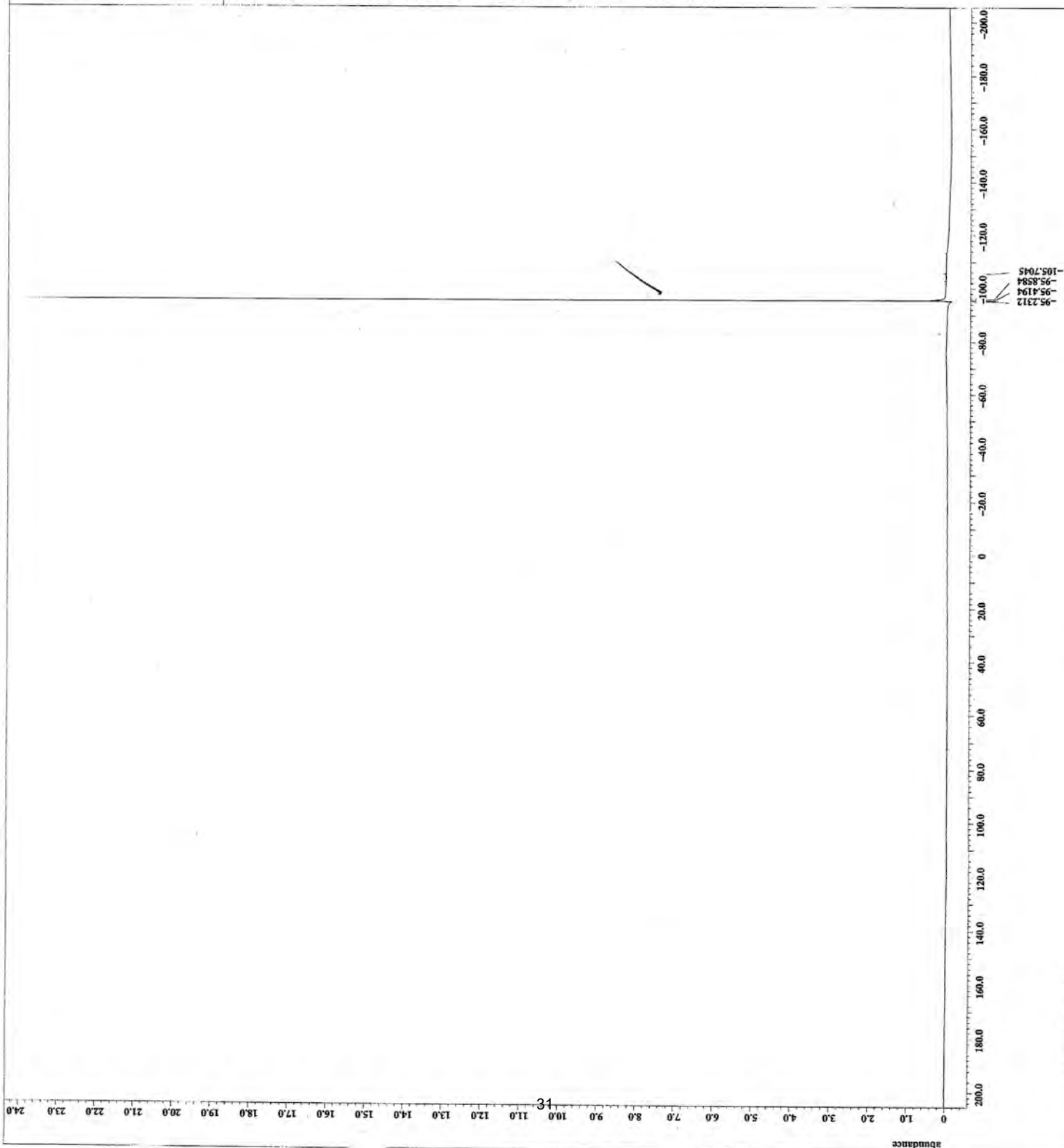
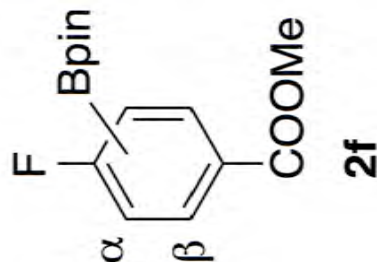
```

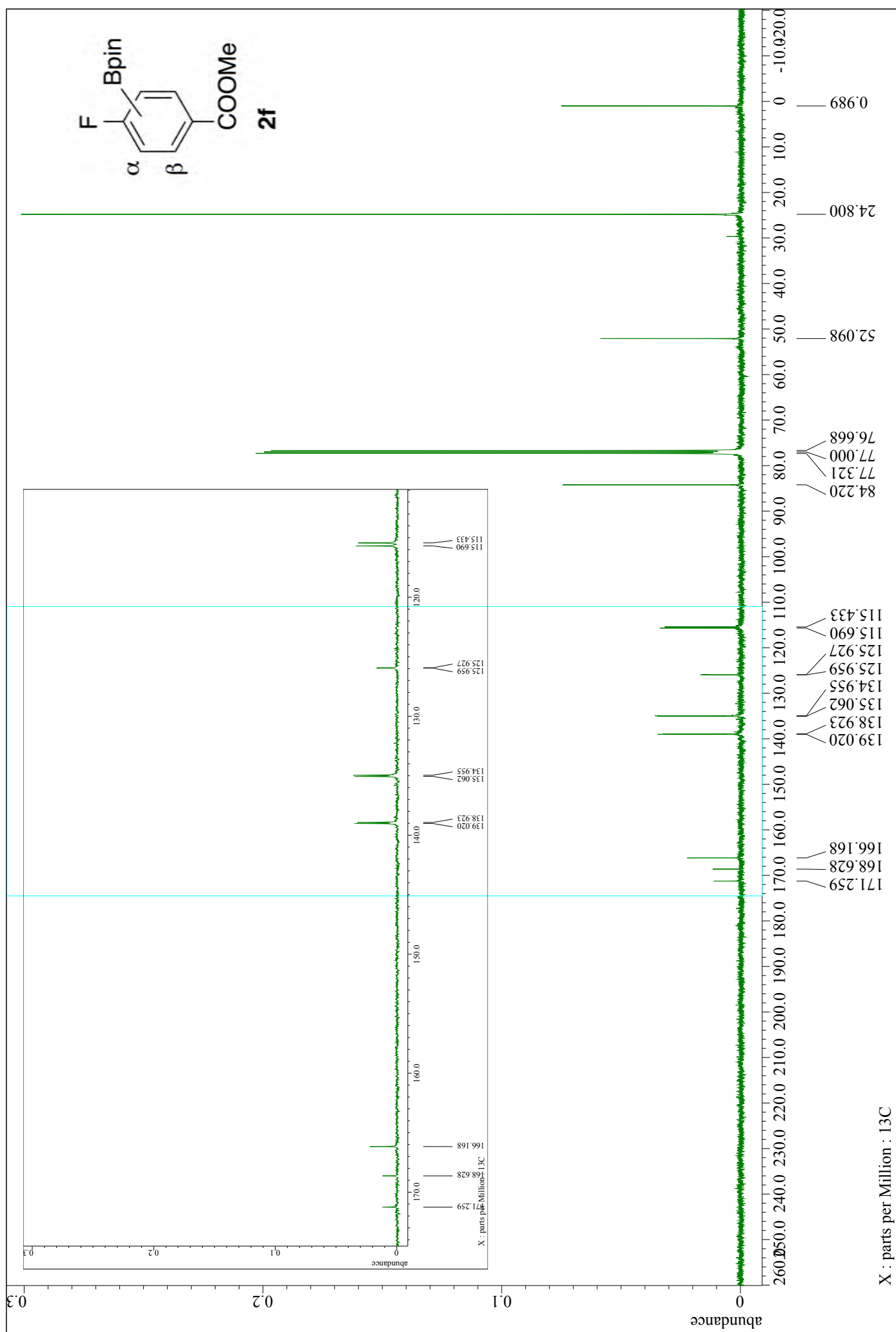
```

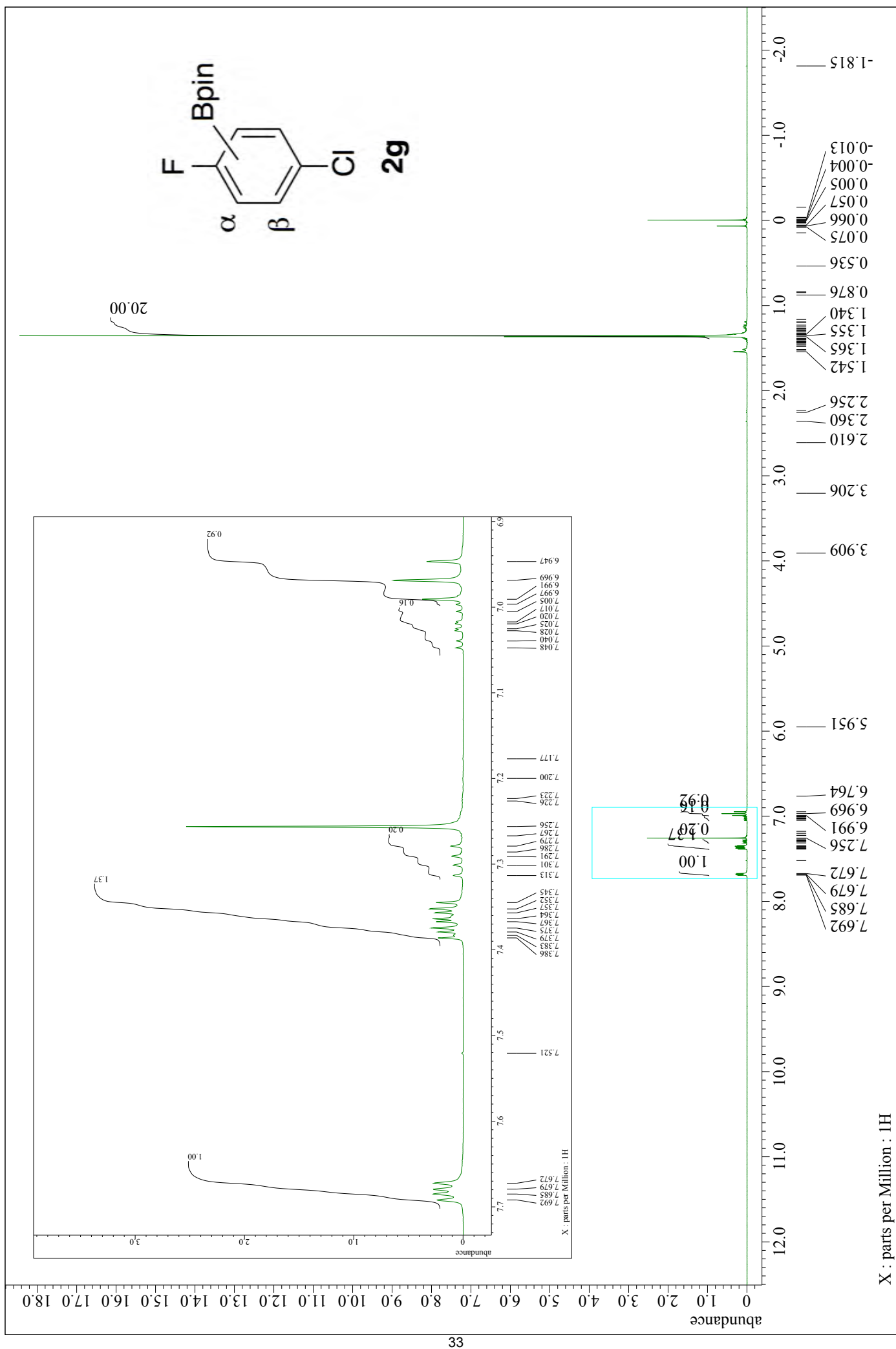
Filename = ex247 date 19f-1.jdf
Author =
Experiment =
Sample_id = A
Solvent = CHLOROFORM-D
Creation_time = 1-MAY-2015 17:15:44
Revision_time = 1-MAY-2015 18:28:02
Current_time = 1-MAY-2015 18:28:16
Comment = single pulse
Data_format = 1D COMPLEX
Dia_size = 13107
Dia_title = 19F
Dimensions = X
Site = EXC 400
Spectrometer = DELTA3_MMR
Field_strength = 9.20397068 [T] (390 [MH]
X_acq_time = 86.50752 [ms]
X_domain = 19F
X_freq = 368.64763285 [MHz]
X_offset = 0 [ppm]
X_points = 16384
X_prescan = 1
X_resolution = 11.5586868 [Hz]
X_sweep = 189.38393939 [kHz]
Irr_domain = 19F
Irr_freq = 368.64763285 [MHz]
Irr_offset = 5 [ppm]
Tri_domain = 19F
Tri_freq = 368.64763285 [MHz]
Tri_offset = 5 [ppm]
Clipped = FALSE
Mod_return = 1
Scans = 305
Total_scans = 305
X_90_width = 11.5 [us]
X_acq_time = 86.50752 [ms]
X_angle = 45 [deg]
X_atn = 3.4 [dB]
X_phase = 0.5 [us]
Irr_phase = 0.5 [us]
Tri_phase = 0.5 [us]
Dante_preset = OFF
Dante_preset = FALSE
Initial_wait = 1 [s]
Recvr_gain = 22
Reference_delay = 5 [s]
Repetition_time = 5.08650752 [s]
Temp_get = 24.7 [°C]

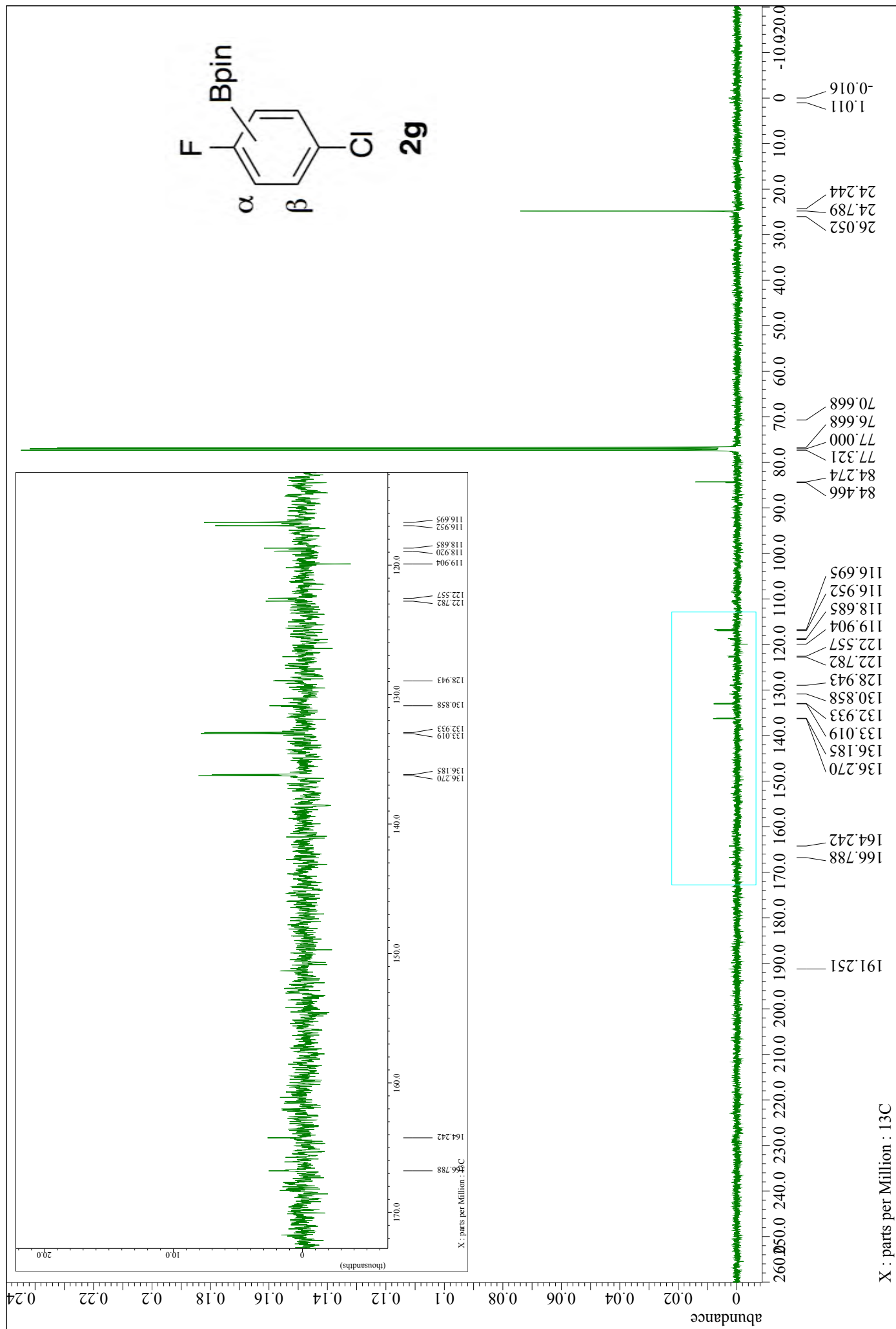
```

19F NMR





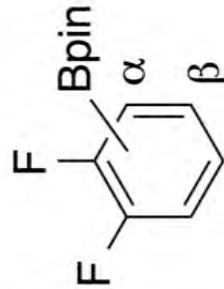




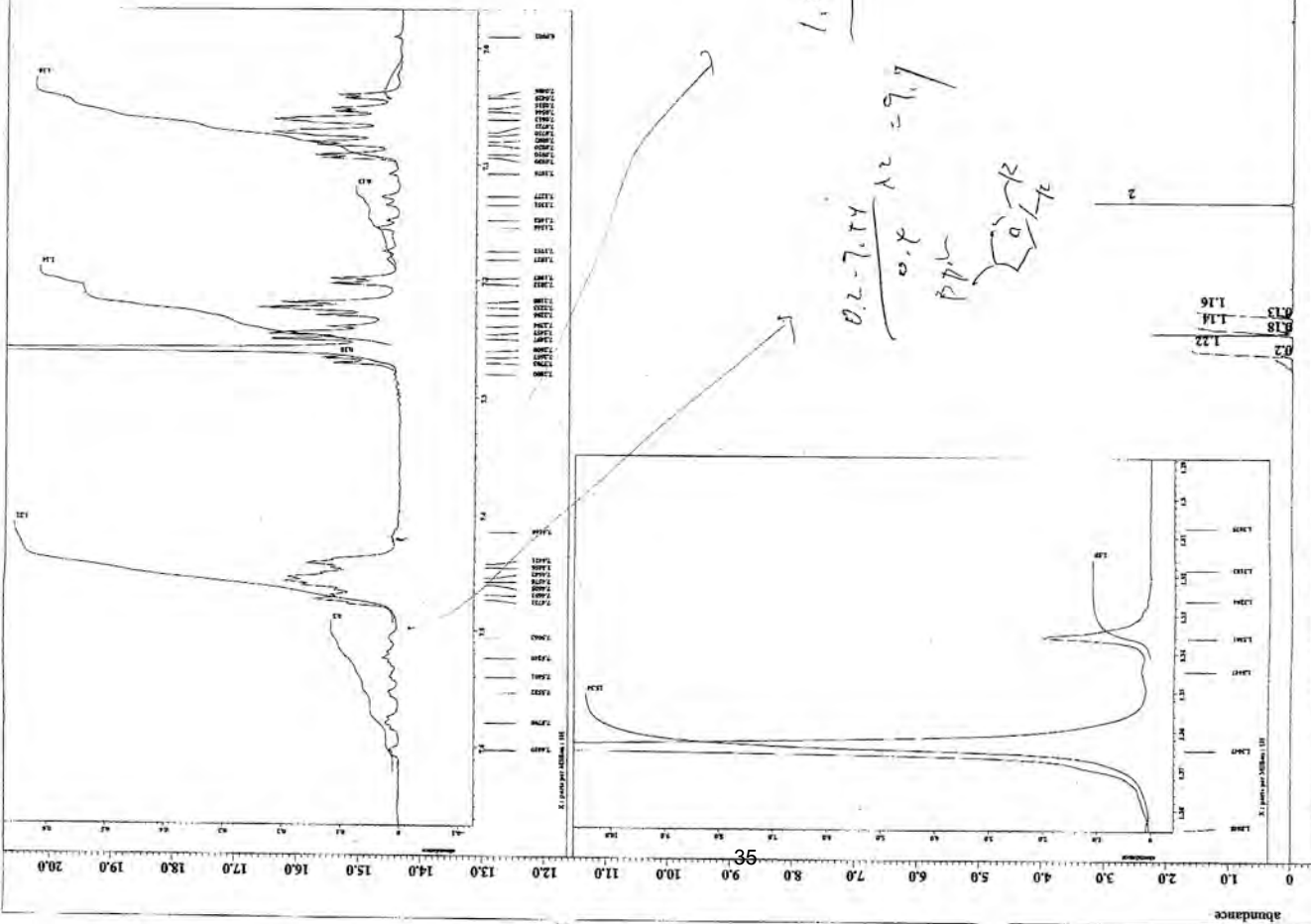
----- PROCESSING PARAMETERS -----
 dc_balance : 0 : FALSE
 freq : 300.136 [MHz]
 gamma : 1.000000000
 hetero : 1 : 0 [N] : 80 [N] : 100 [N]
 resolution : 1 : 0 [N] : 80 [N] : 100 [N]
 zft : 1 : TRUE : TRUE
 machinephase :
 phase : 0 : 0 : 50 [N]
 Derived from: ex111 crude 1H-4.jde

 File Name : ex111 crude 1H-4.jde
 Author :
 Experiment :
 Sample ID :
 Solvent :
 Creation Time : 10-OCT-2014 11:39:54
 Acquisition Time : 10-OCT-2014 11:39:54
 Current Time : 10-OCT-2014 11:39:54
 Comment :
 Data Format : 1D_COMPLEX
 Dir Size : 25214
 Dir Units :
 Dir Name :
 Dimensions :
 Site :
 Spectrometer :
 P1 (s) : 9.2017056 [s] (350 [MHz])
 X_acq duration : 4.456448 [s]
 X_domain : 1H
 X_freq : 391.78655441 [MHz]
 X_offset : 5 [ppm]
 X_pulses : 1
 X_resolution : 0.22439396 [Hz]
 X_sweep : 7.35284118 [MHz]
 Irr_domain : 1H
 Irr_freq : 391.78655441 [MHz]
 Irr_offset : 5 [ppm]
 Tri_domain : 1H
 Tri_freq : 391.78655441 [MHz]
 Tri_offset : 5 [ppm]
 Clipped : FALSE
 Mod_return : 1
 Scan : 8
 Total_scans : 8
 X_90_width : 9.7 [us]
 X_acq_time : 4.456448 [s]
 X_angle : 45 [deg]
 X_pulse : 4.85 [us]
 Irr_mode : OFF
 Tri_mode : OFF
 Dante_present : FALSE
 Initial_wait : 1 [s]
 Relaxation_delay : 5 [s]
 Repetition_time : 9.456448 [s]
 Temp_get : 24.8 [degC]

1H NMR

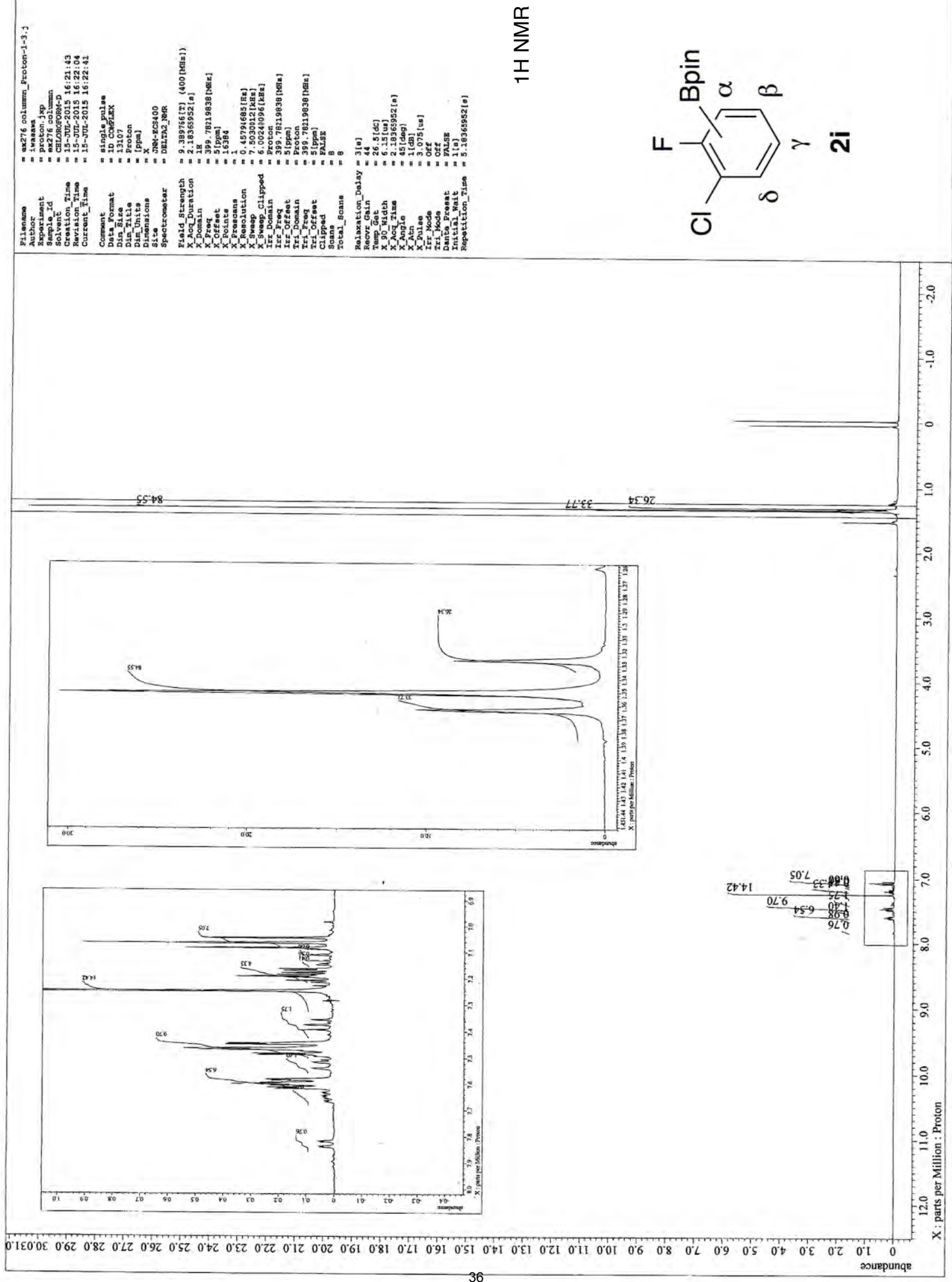


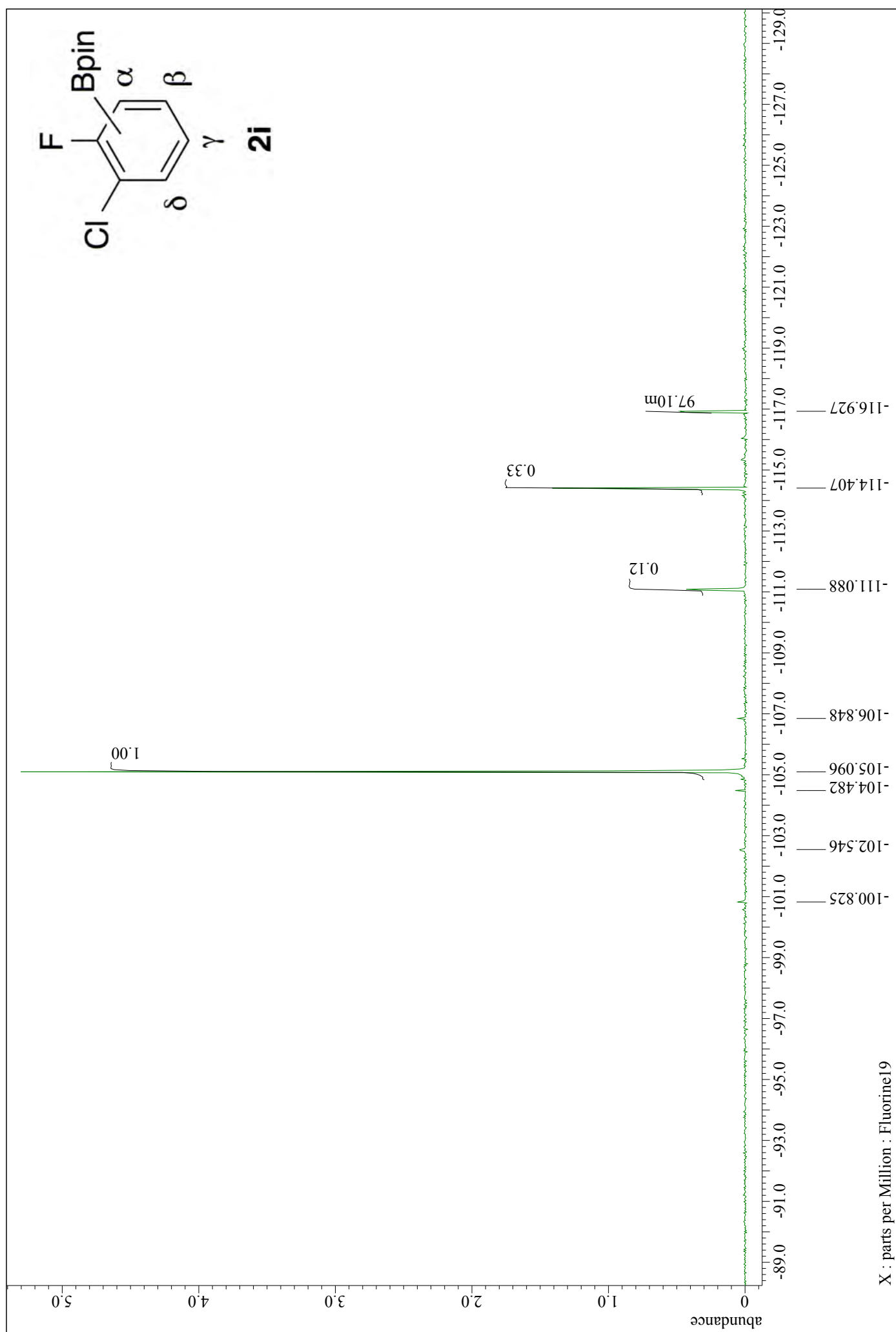
2h



X: parts per Million : 1H

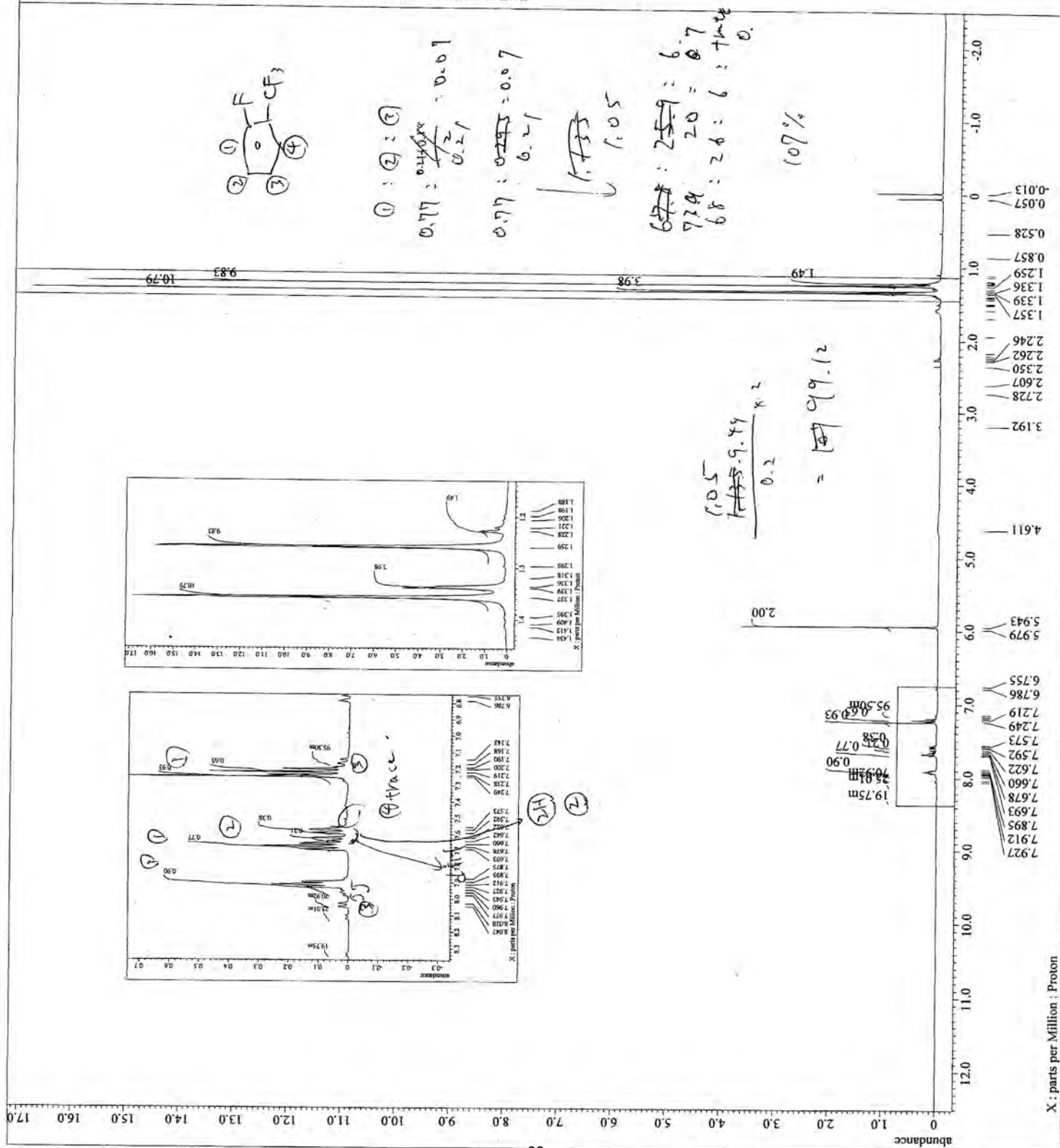
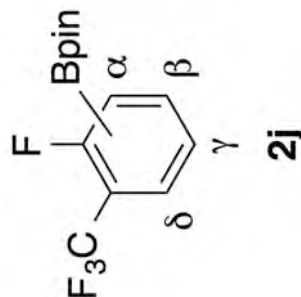
with 1,1,2,2-tetrachloroethane (5.9 ppm)
 as an internal standard





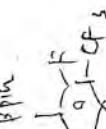
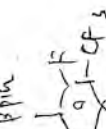
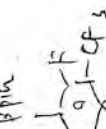
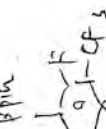
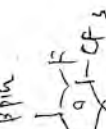
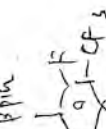
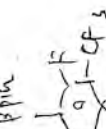
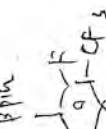
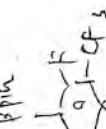
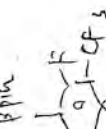
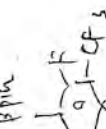
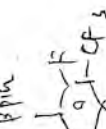
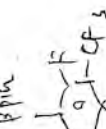
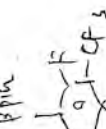
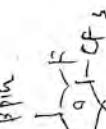
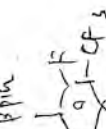
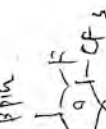
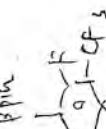
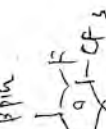
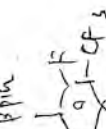
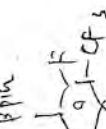
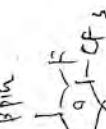
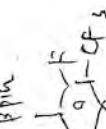
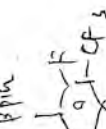
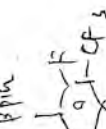
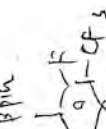
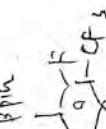
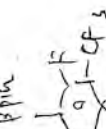
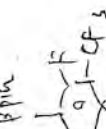
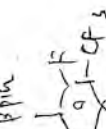
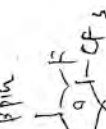
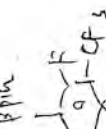
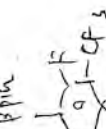
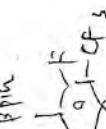
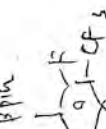
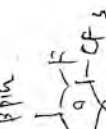
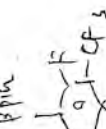
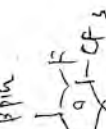
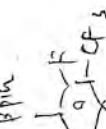
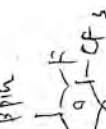
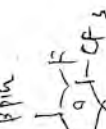
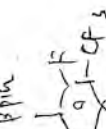
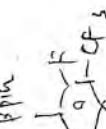
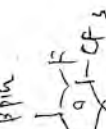
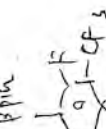
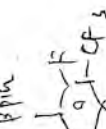
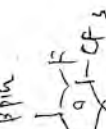
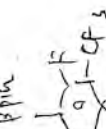
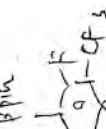
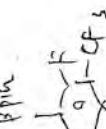
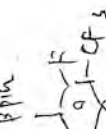
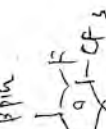
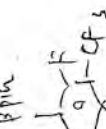
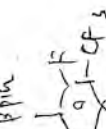
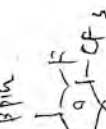
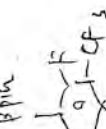
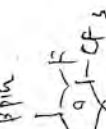
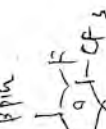
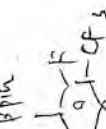
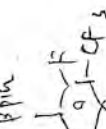
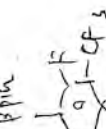
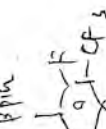
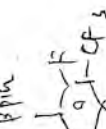
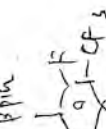
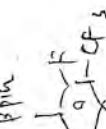
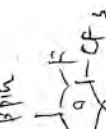
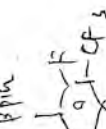
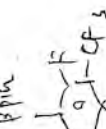
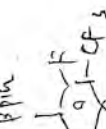
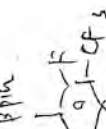
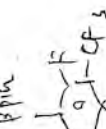
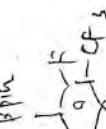
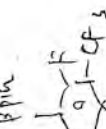
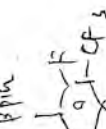
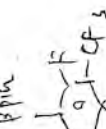
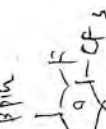
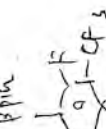
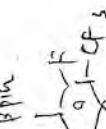
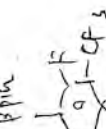
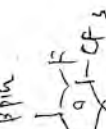
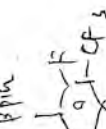
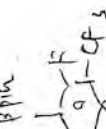
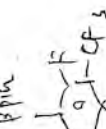
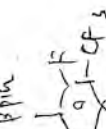
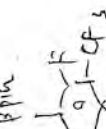
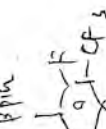
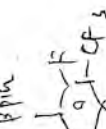
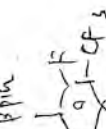
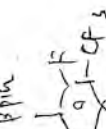
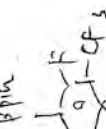
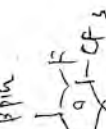
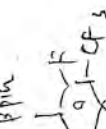
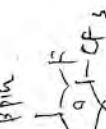
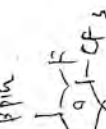
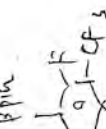
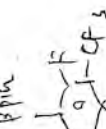
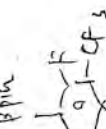
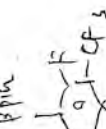
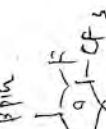
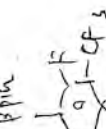
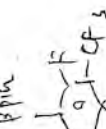
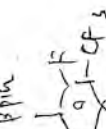
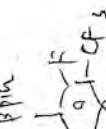
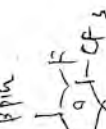
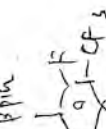
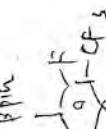
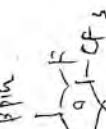
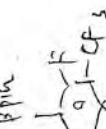
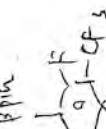
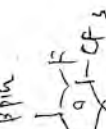
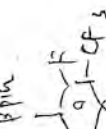
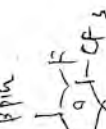
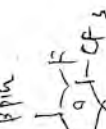
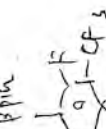
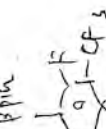
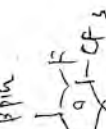
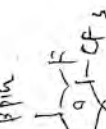
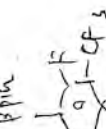
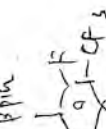
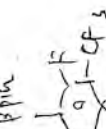
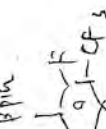
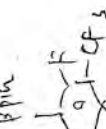
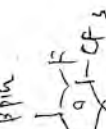
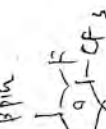
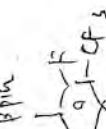
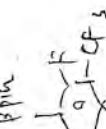
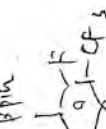
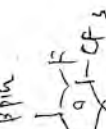
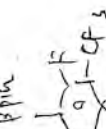
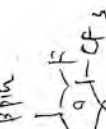
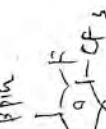
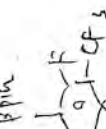
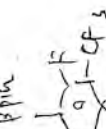
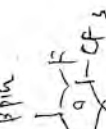
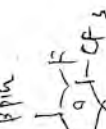
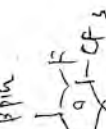
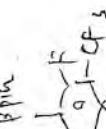
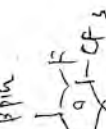
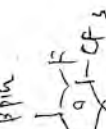
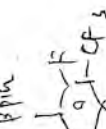
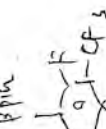
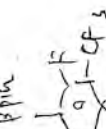
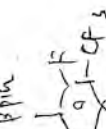
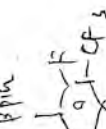
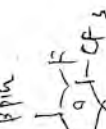
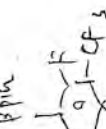
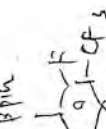
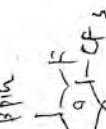
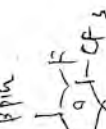
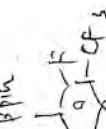
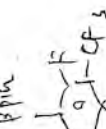
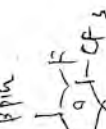
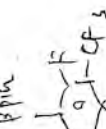
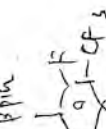
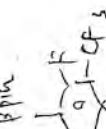
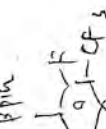
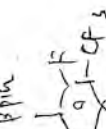
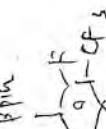
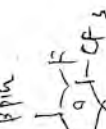
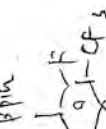
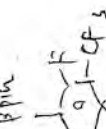
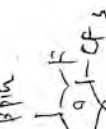
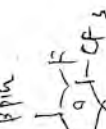
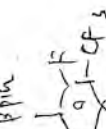
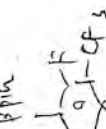
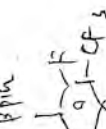
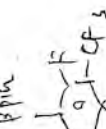
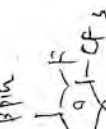
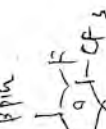
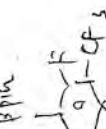
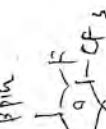
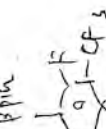
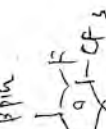
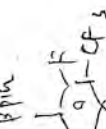
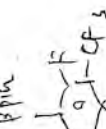
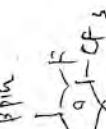
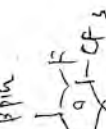
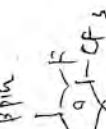
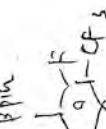
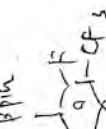
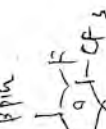
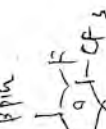
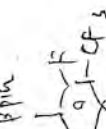
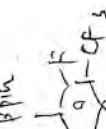
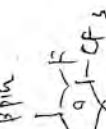
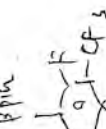
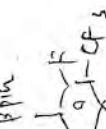
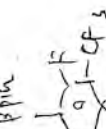
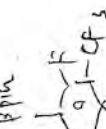
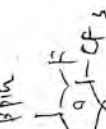
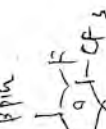
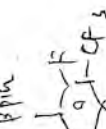
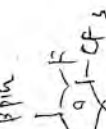
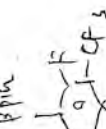
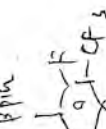
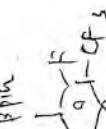
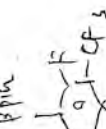
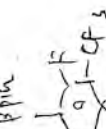
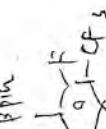
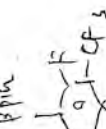
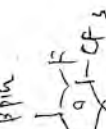
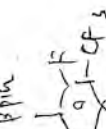
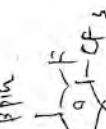
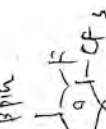
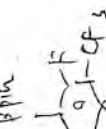
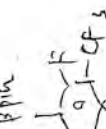
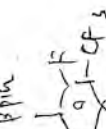
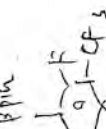
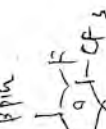
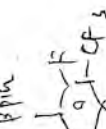
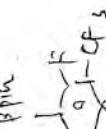
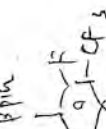
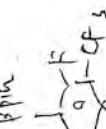
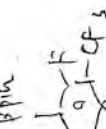
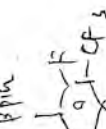
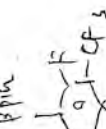
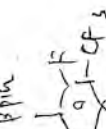
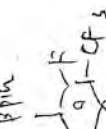
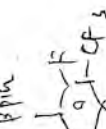
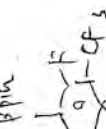
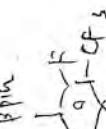
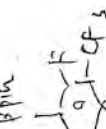
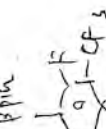
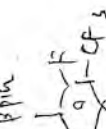
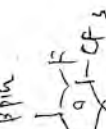
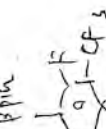
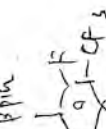
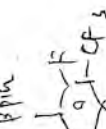
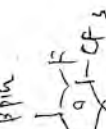
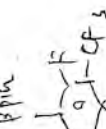
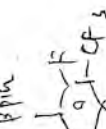
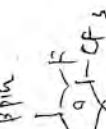
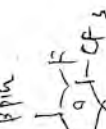
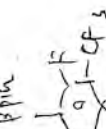
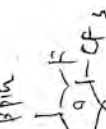
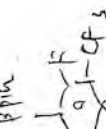
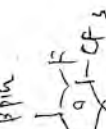
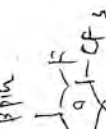
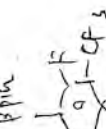
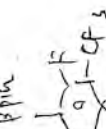
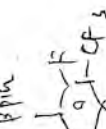
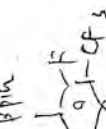
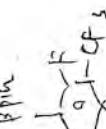
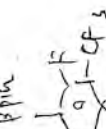
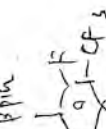
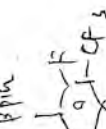
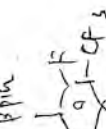
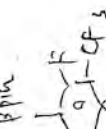
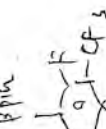
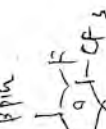
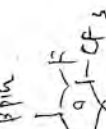
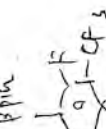
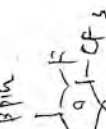
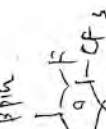
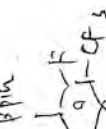
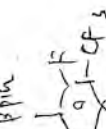
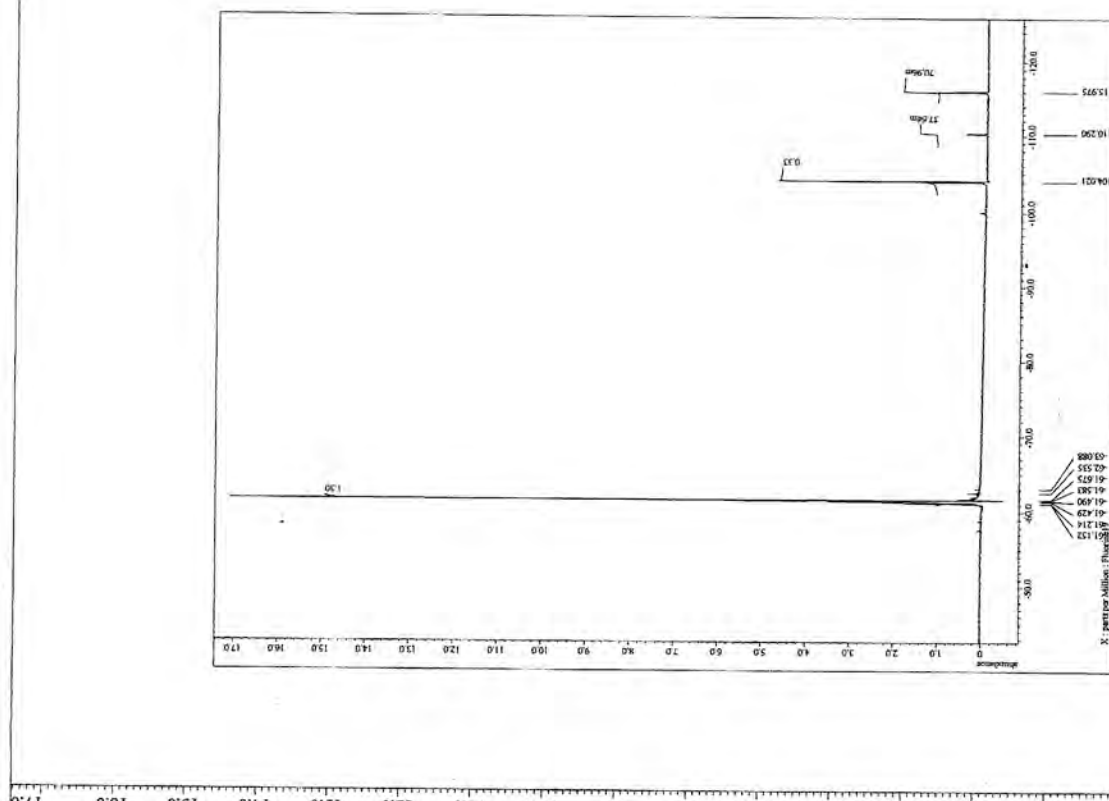
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Author	
Experiment	
Sample_ID	
Solvent	
Revision	
Current_F ₁	
Comment	
Data Format	
Din Size	
Dim Units	
Dimensions	
Size	
Spectrometers	
Field Strength	
X Domain	
Freq	
X Offset	
X Points	
X PrescansName	
X Resolution	
X Sweep	
Y Domain	
Irr Freq	
Irr OffsetsName	
Y Domain	
Tot Freq	
Cycled	
Scans	
Total_Scans	
Relaxation	
Recvr Gain	
X 90 Angle	
AQc Time	
XAngle	
XAtn	
Pulse	
Irr Mode	
Date Processed	
Initial Wdr	
Repetition	

¹H NMR



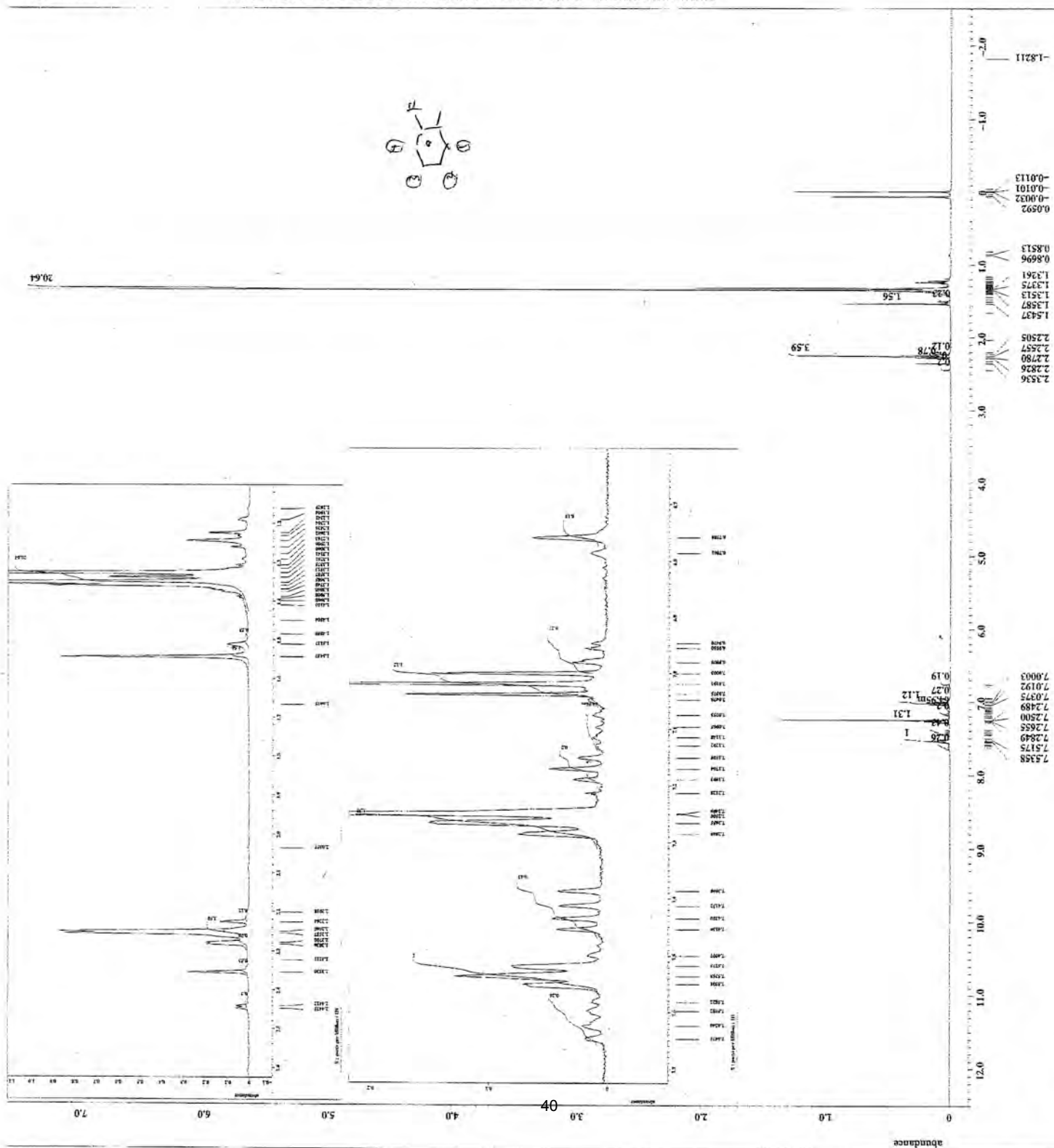
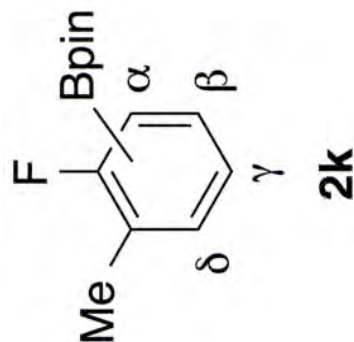
with 1,1,2,2-tetrachloroethane (5.9 ppm) as an internal standard

X: parts per Million : Proton



19.64

Filename	ex2770rev04 IH-5_50f	
Author	data	
Experiment	single pulse.ex2	
Sample_id	A	
Solvent	CHLOROFORM-D	
Acq_time	14-JUN-2015 13:04:39	
Revision_time	14-JUN-2015 14:19:53	
Current_time	14-JUN-2015 14:30:37	
Comment	single pulse	
dataformat	COMPLEX	
bits	262144	
dia_title	1H	
dia_units	[ppm]	
dimensions	XZ 400	
site	DMF44_gbr	
spectrometer		
Field_strength	9.201379668 [T] 390 [MHz]	
X_acq_duration	4.456448 [s]	
X_domain	1H	
X_offset	7.8655441 [MHz]	
X_points	51000	
X_pos	32768	
X_prescans	1	
X_resolution	0.22439396 [Hz]	
Y_acq_duration	1.35294118 [Hz]	
Y_domain	391.78655441 [MHz]	
Y_offset	5 [ppm]	
Yi_domain	1H	
Yi_freq	391.78655441 [MHz]	
Yi_offset	FALSE	
Yi_clip	FALSE	
Mod_return	1	
Scans	8	
Total_scans	8	
X90_width	9.93 [us]	
Xacq_time	4.456448 [s]	
X_sample	45 [deg]	
X_atn	3 [dB]	
X_pulse	0.985 [us]	
X_delay	46	
Pri_mode	Off	
Pulse_presat	FALSE	
Initial_wait	1 [s]	
Recvr_gain	46	
Delay	46	
Relaxation_time	9.456448 [s]	
Temp_set	23 [C]	

¹H NMR

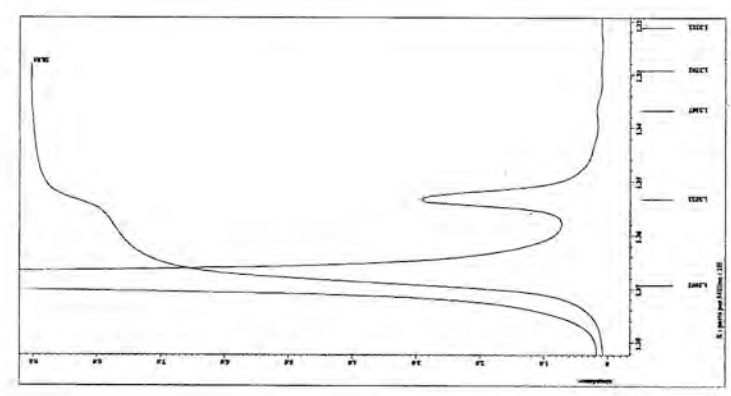
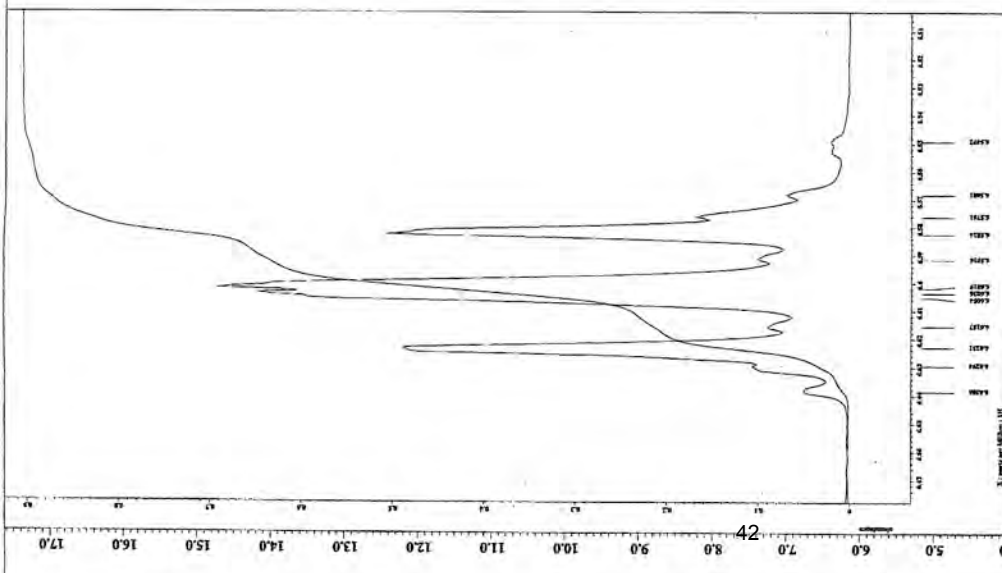
X : parts per Million : 1H

valer



A chemical structure of a benzene ring with four substituents: a Bpin group at the top position, a fluorine atom (F) at the top-left position, a methyl group (Me) at the bottom-left position, and a hydrogen atom at the bottom-right position. The positions are labeled with Greek letters: α for the Bpin group, β for the fluorine atom, γ for the methyl group, and δ for the hydrogen atom.

2k

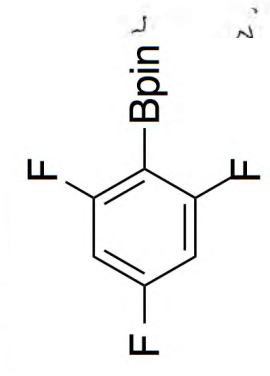


28.84

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 acq_freq : 400.146 [MHz]
 zerooffset : 0.000 [Hz]
 zerooffset : 0.000 [Hz]
 zft : 1 : TRUE : TRUE
 machinename :
 phase : 0 : 0 : 50 [Hz]
 Derived from: ex34 column1 1H-1.jcf

 File : ex34 column1 1H-1.jcf
 Acq :
 Experiment :
 Sample :
 Solvent : CHLOROFORM-D
 Creation_time : 20-SEP-2014 14:37:51
 Revision : 1
 Current_time : 20-SEP-2014 11:58:11
 Comment :
 Data :
 Data_format : 1D COMPLEX
 Dia_size : 26214
 Dia_unit : mm
 Dia_unit : mm
 Dimensions :
 Site :
 Spectrometer : ECK 400
 DELTA :
 Field_strength : 9.2017068 [T] (399.999 MHz)
 X_acq_duration : 4.456448 [s]
 X_domain : 1H
 X_freq : 391.78655441 [MHz]
 X_offset : 51 [ppm]
 X_points : 32788
 X_resolution : 0.22439396 [Hz]
 X_sweep : 7.35294128 [MHz]
 Irr_domain : 1H
 Irr_freq : 391.78655441 [MHz]
 Irr_offset : 51 [ppm]
 Irr_sweep : 7.35294128 [MHz]
 Trf_freq : 391.78655441 [MHz]
 Trf_offset : 51 [ppm]
 Mod_return : FALSE
 Clipped :
 Total_scans : 1
 X_90_width : 9.7 [us]
 X_acq_time : 4.456448 [s]
 X_angle : 45 [deg]
 X_pulse : 4.98 [us]
 Trf_pulse : 4.98 [us]
 Trf_mode : Off
 Dntc_preset : FALSE
 Initial_wait : 1 [s]
 Relaxation_delay : 5 [s]
 Repetition_time : 9.456448 [s]
 Temp_get : 25.1 [deg]

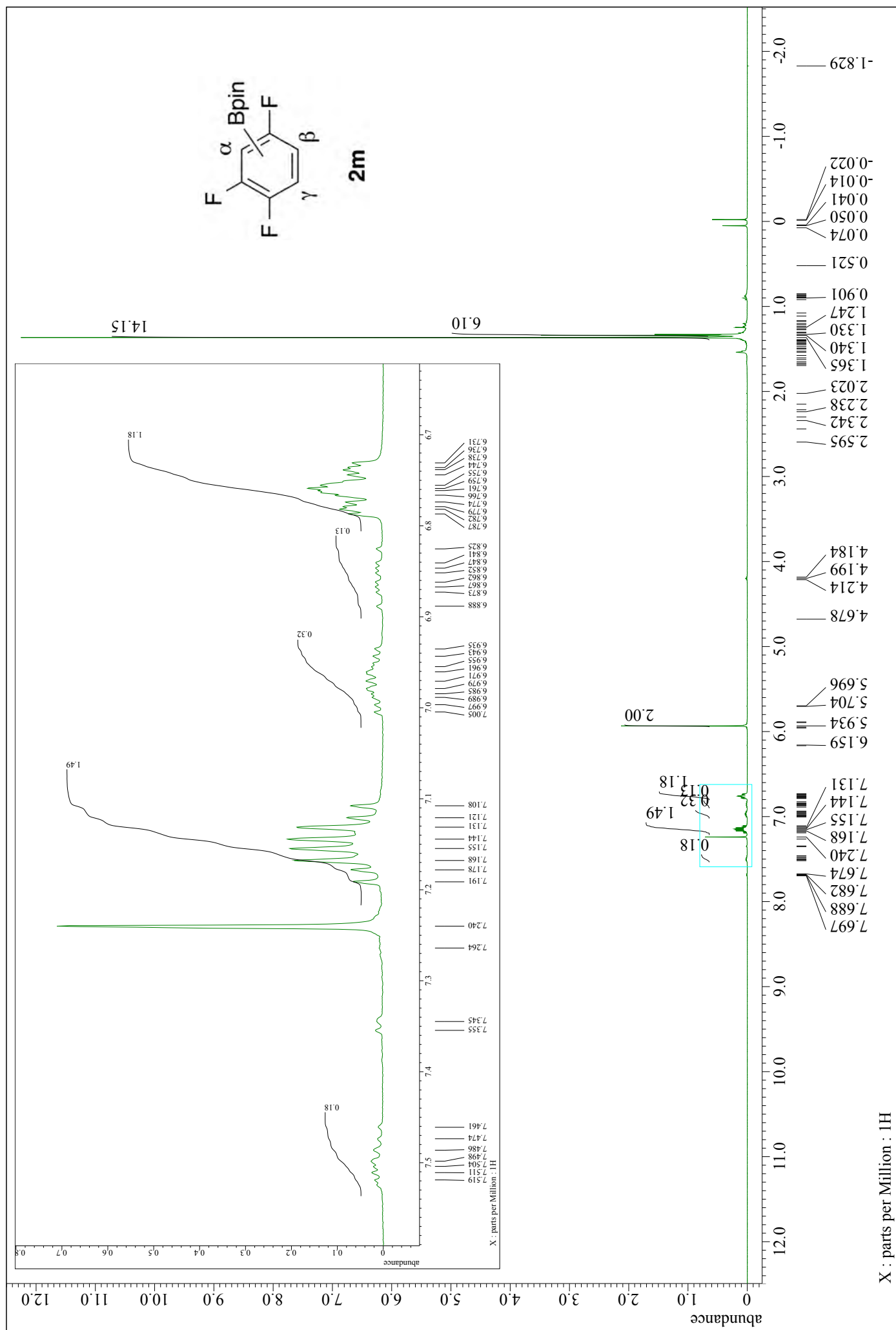
1H NMR



21

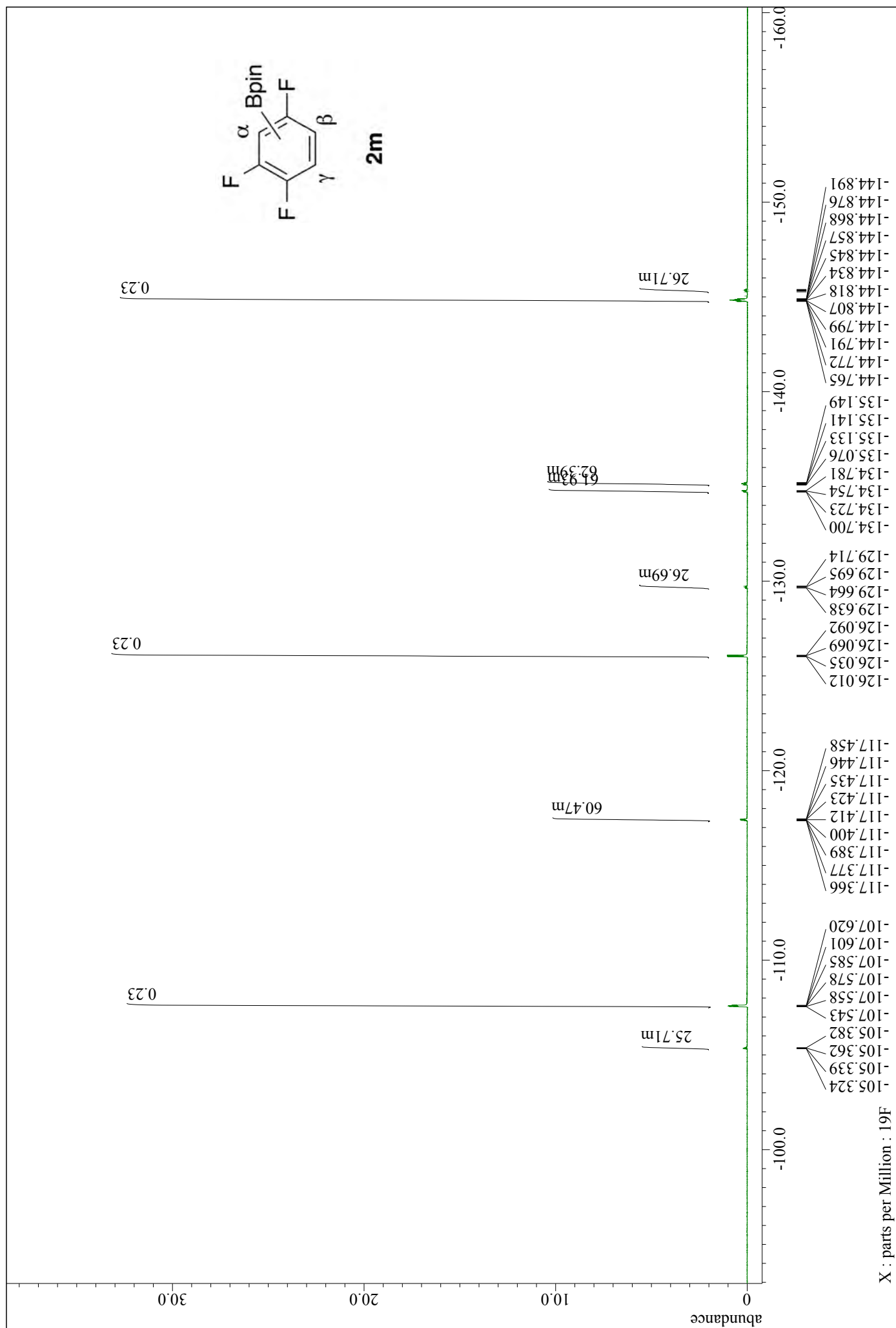
X : parts per Million : 1H

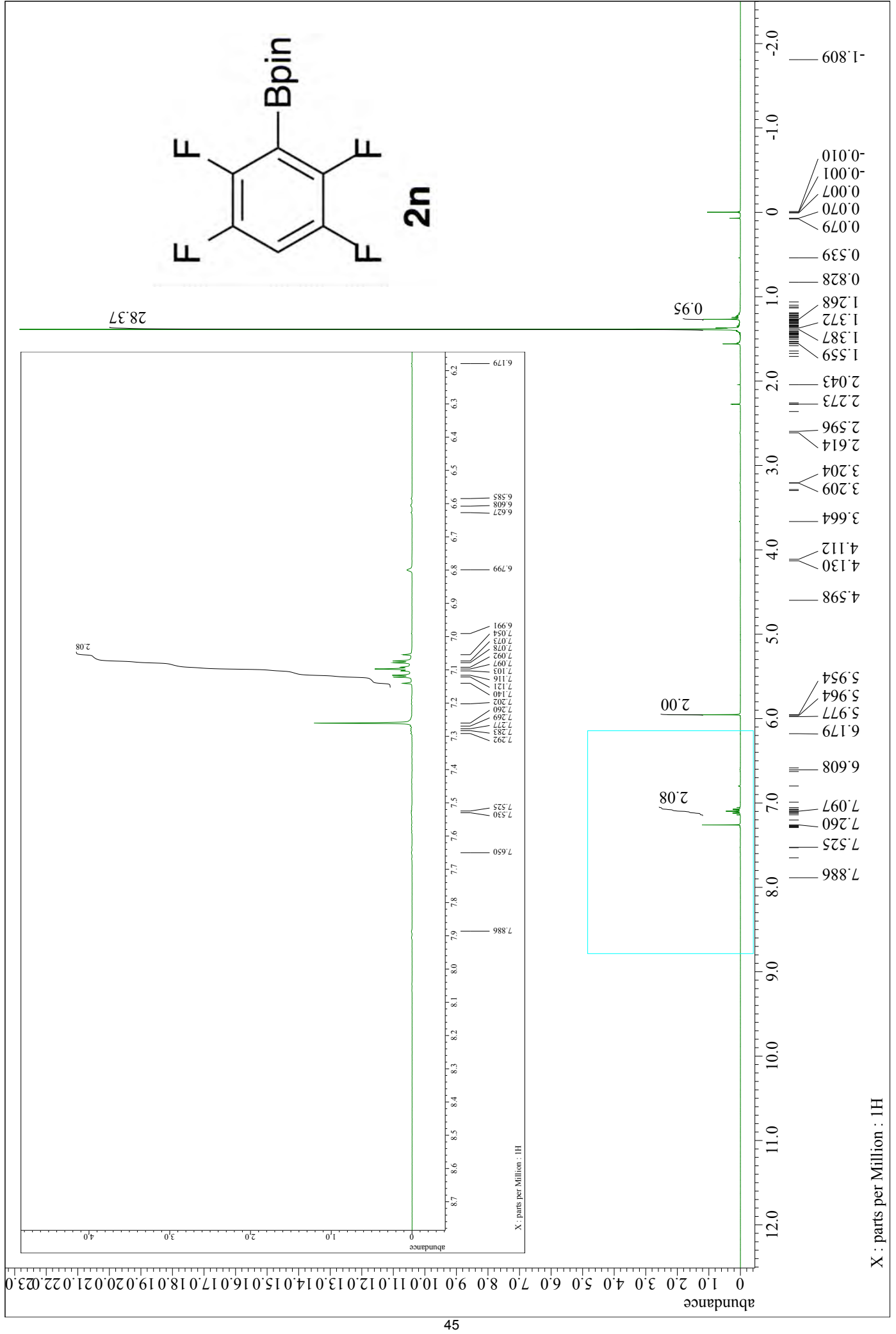
with 1,1,2,2-tetrachloroethane (5.9 ppm)
 as an internal standard



with 1,1,2,2-tetrachloroethane (5.9 ppm)
as an internal standard

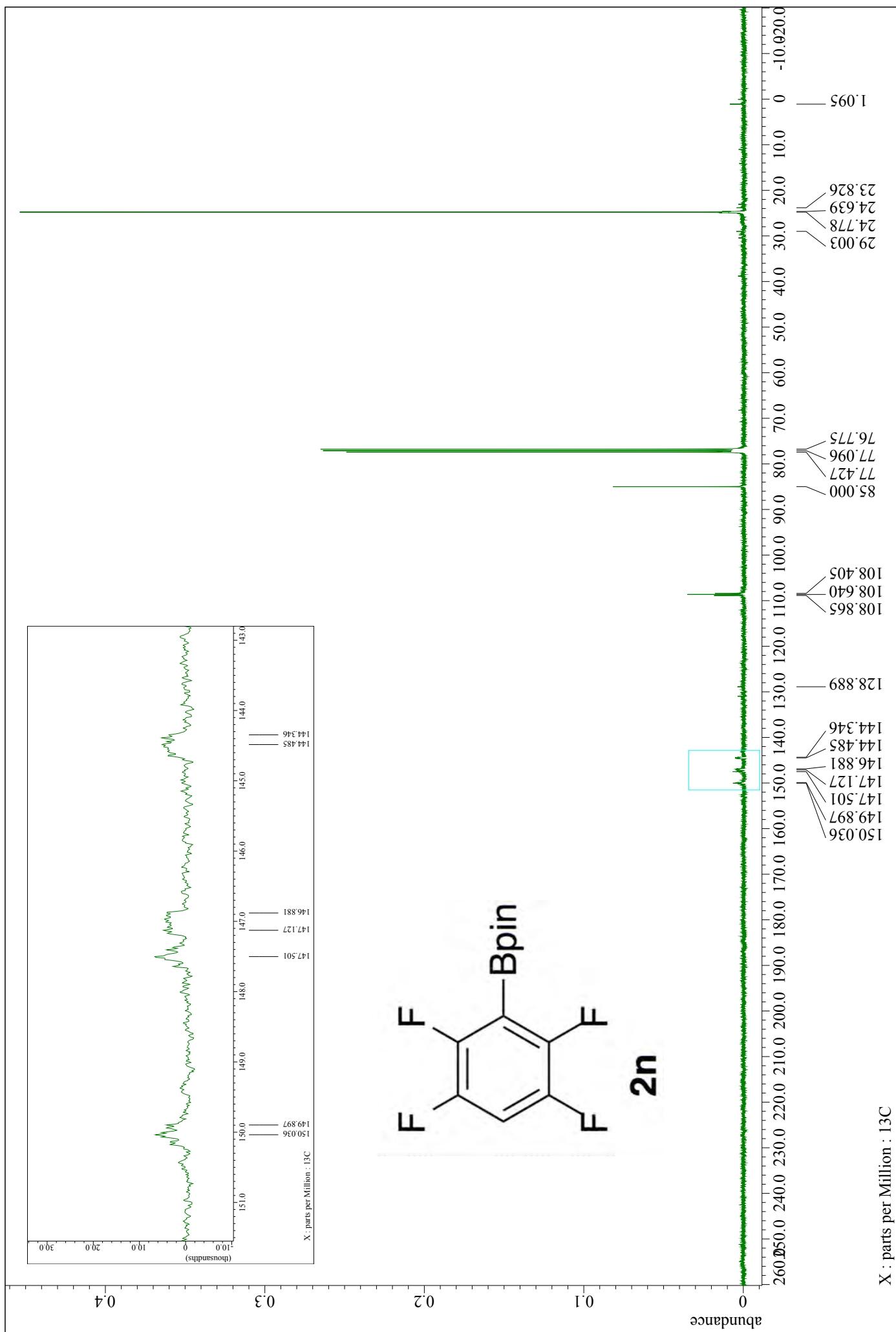
X : parts per Million : 1H





with 1,1,2,2-tetrachloroethane (5.9 ppm)
as an internal standard

X : parts per Million : 1H

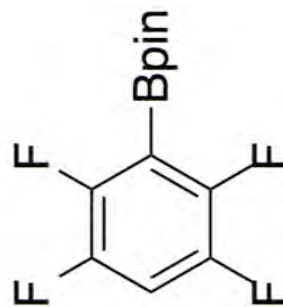


9

47



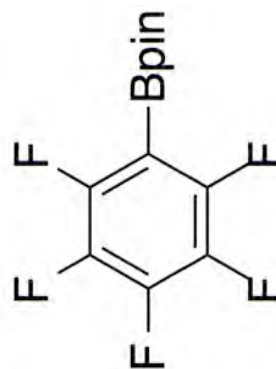
2n



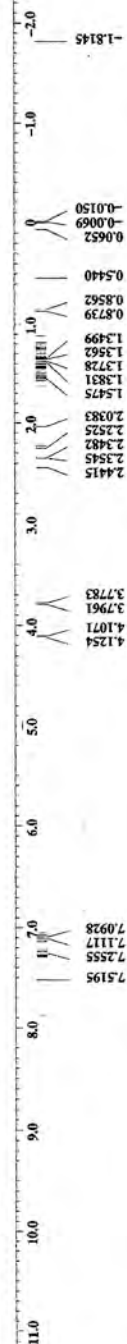
^{19}F NMR

48

¹H NMR

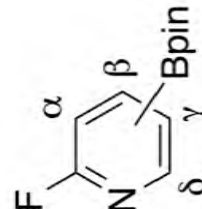
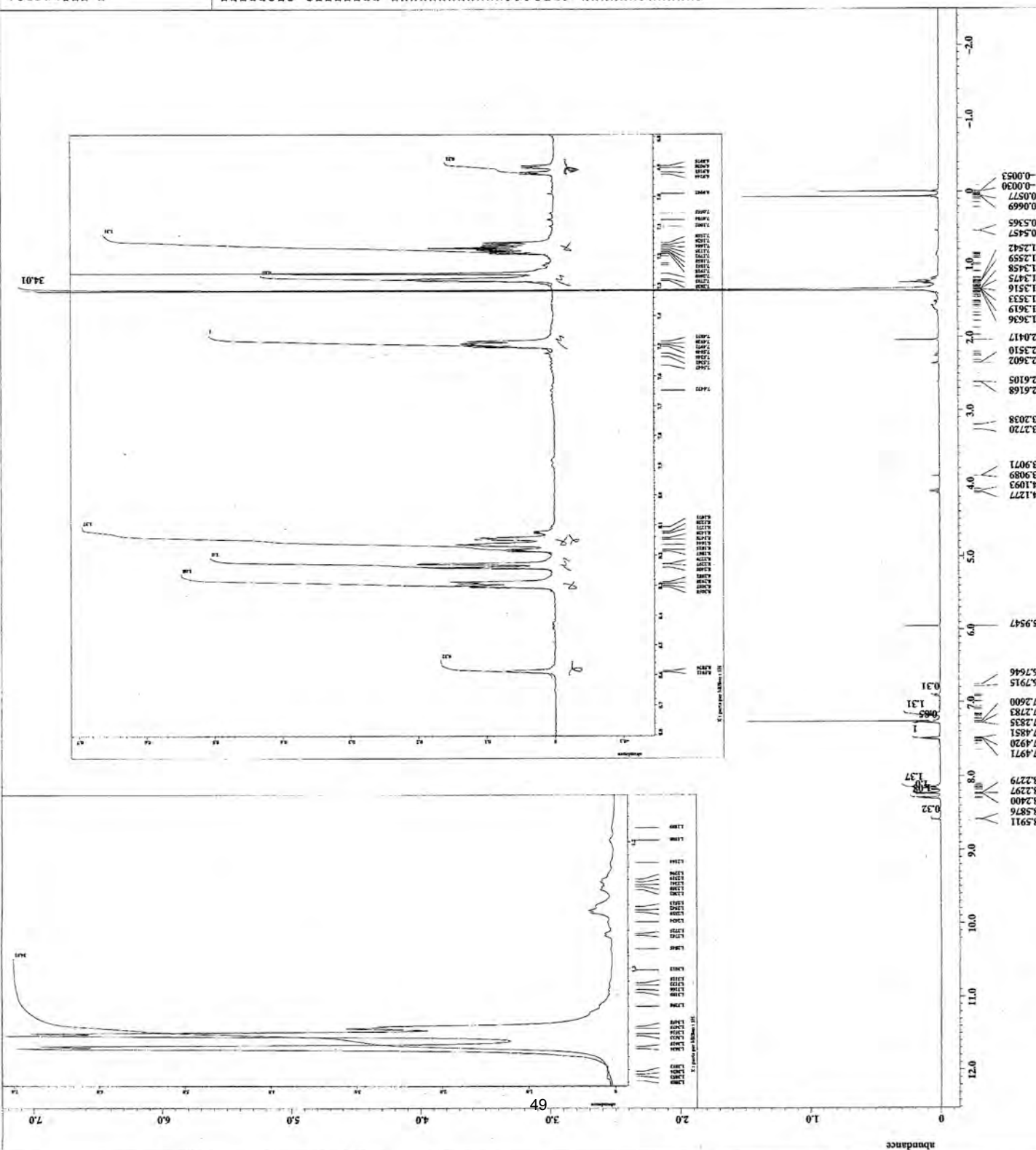


20



X : parts per Million : 1H

----- PROTECTING PARAMETERS

[illegible]

3a

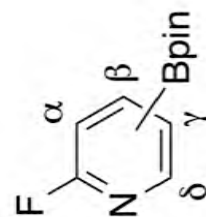
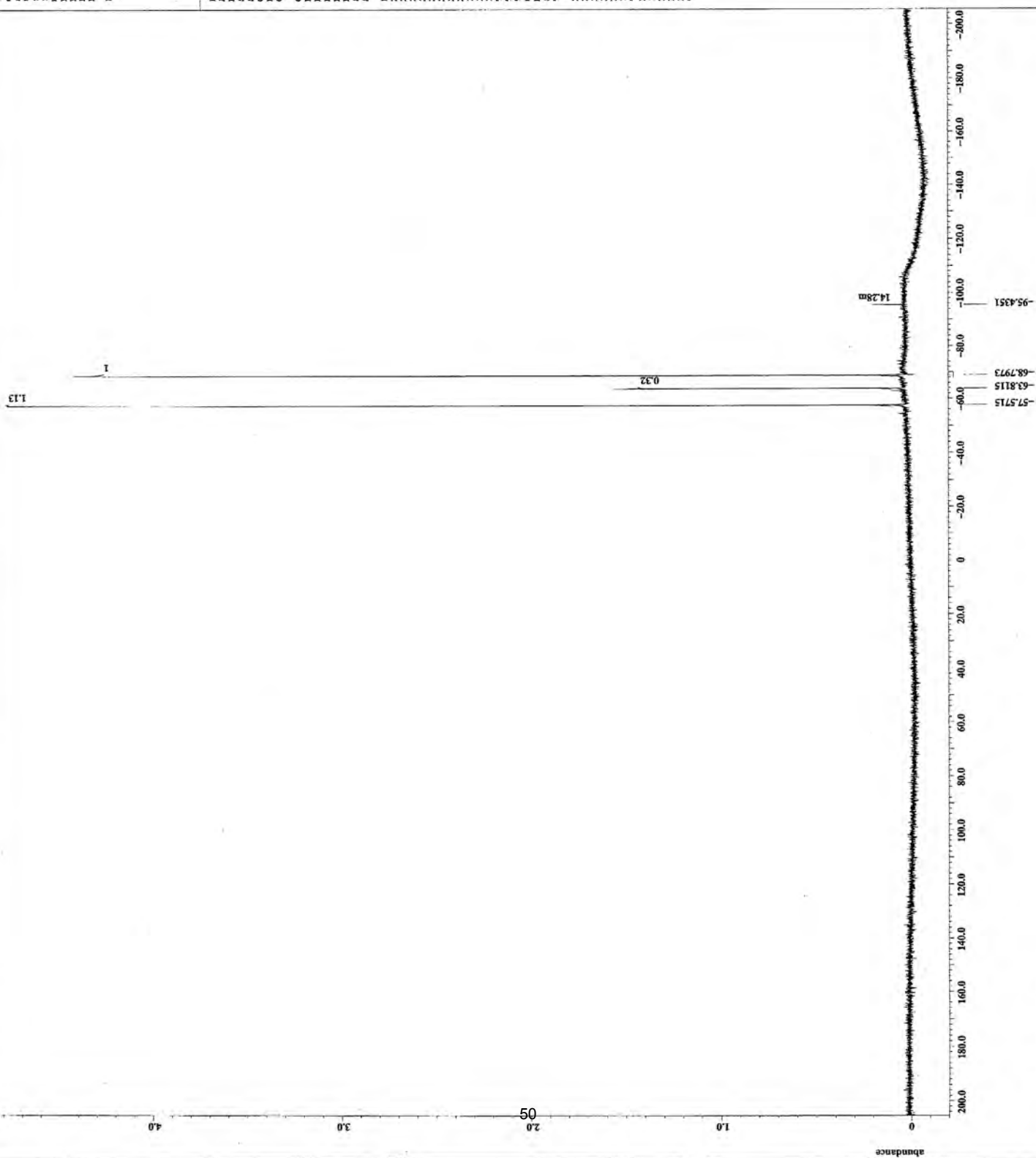
X : parts per Million : 1H

with 1,1,2,2-tetrachloroethane (5.9 ppm) as an internal standard

```

PROCESSING PARAMETERS -----
Balance: 0 FALSE
Comp: 0.2 [Hz] 0.0 [s]
Resperiods: 0 [Hz] 80 [s] 100 [Hz]
zerofill: 1
fft: 1 TRUE
machinephase
pygm
phase: 16.5 0 66.75059 [Hz]
phase: 35 220 66.75059 [Hz]
phase: -20 -100 66.75059 [Hz]
Derived from: ex149 column 129

```

[illegible]

3a

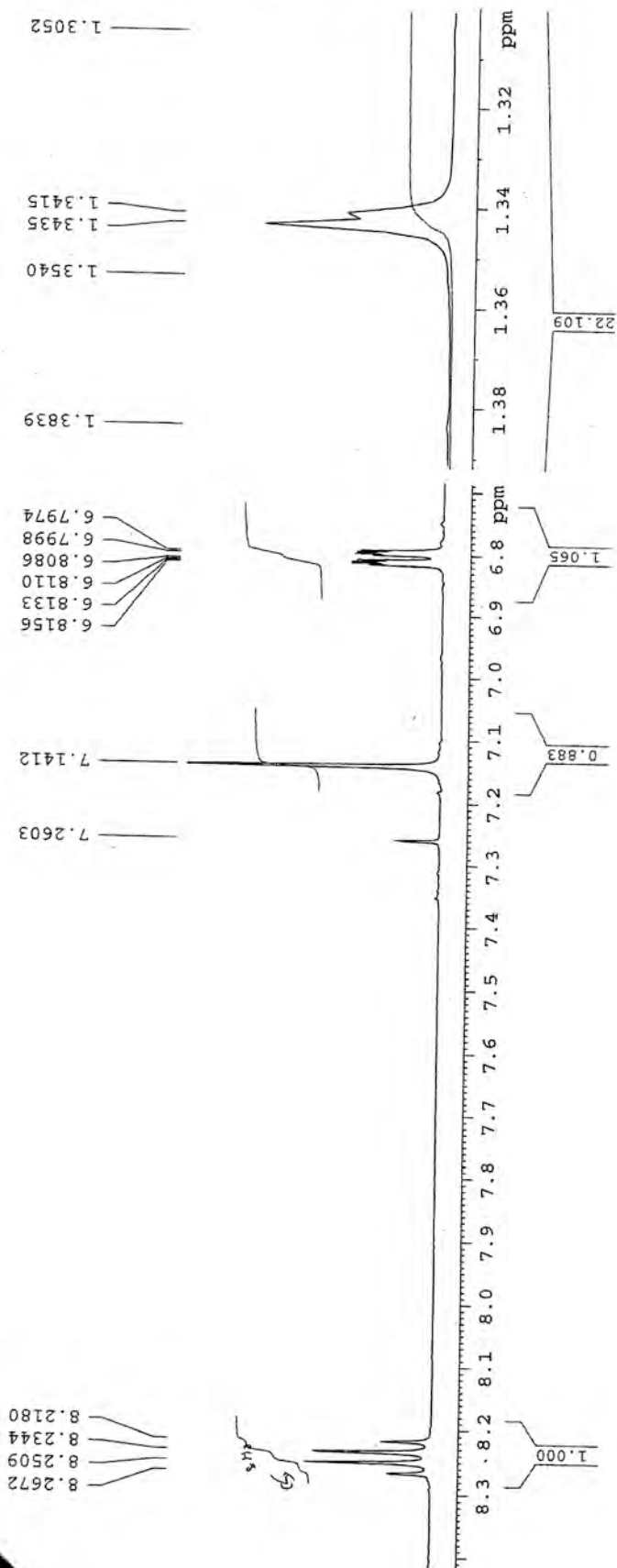
X : parts per Million : 19F

Current Data Parameters
NAME Shisei
EXPNO 1
PROCNO 1

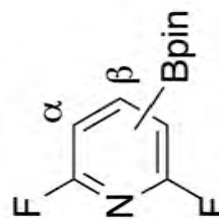
F2 - Acquisition Parameters
Date_ 20150129
Time 22.31
INSTRUM spect
PROBHD 5 mm Multinucl
PULPROG zg
TD 159996
SOLVENT CDCl3
NS 10
DS 2
SWH 20000.000 Hz
FIDRES 0.125003 Hz
AQ 3.9999750 sec
RG 28.5
DE 25.000 usec
TE 300.0 K
D1 3.00000000 sec
MCREST 0.00000000 sec
MCWRK 0.01500000 sec

==== CHANNEL f1 =====
NUC1 1H
P1 10.00 usec
PL1 -3.50 dB
SFO1 500.1274993 MHz

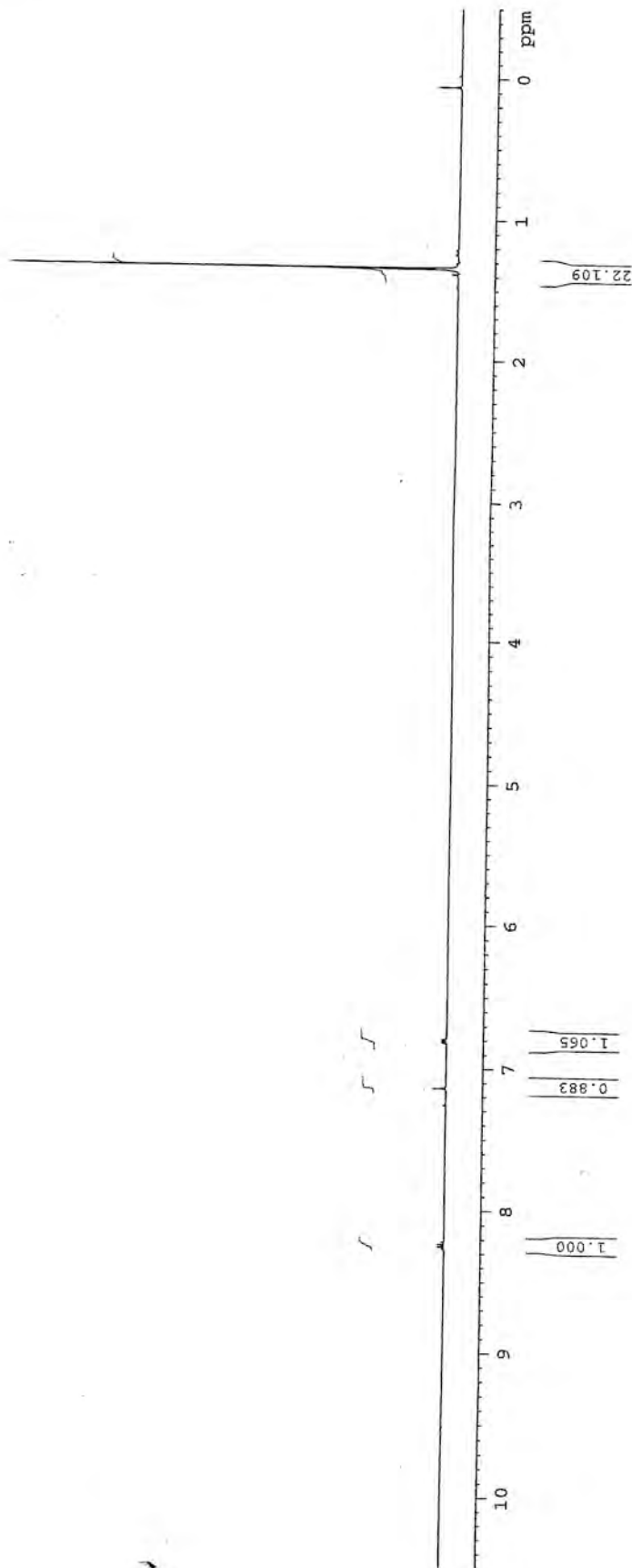
F2 - Processing parameters
SI 32768
SF 500.1300128 MHz
WDW EM
SSB 0
LB 0.20 Hz
GB 0
PC 1.00

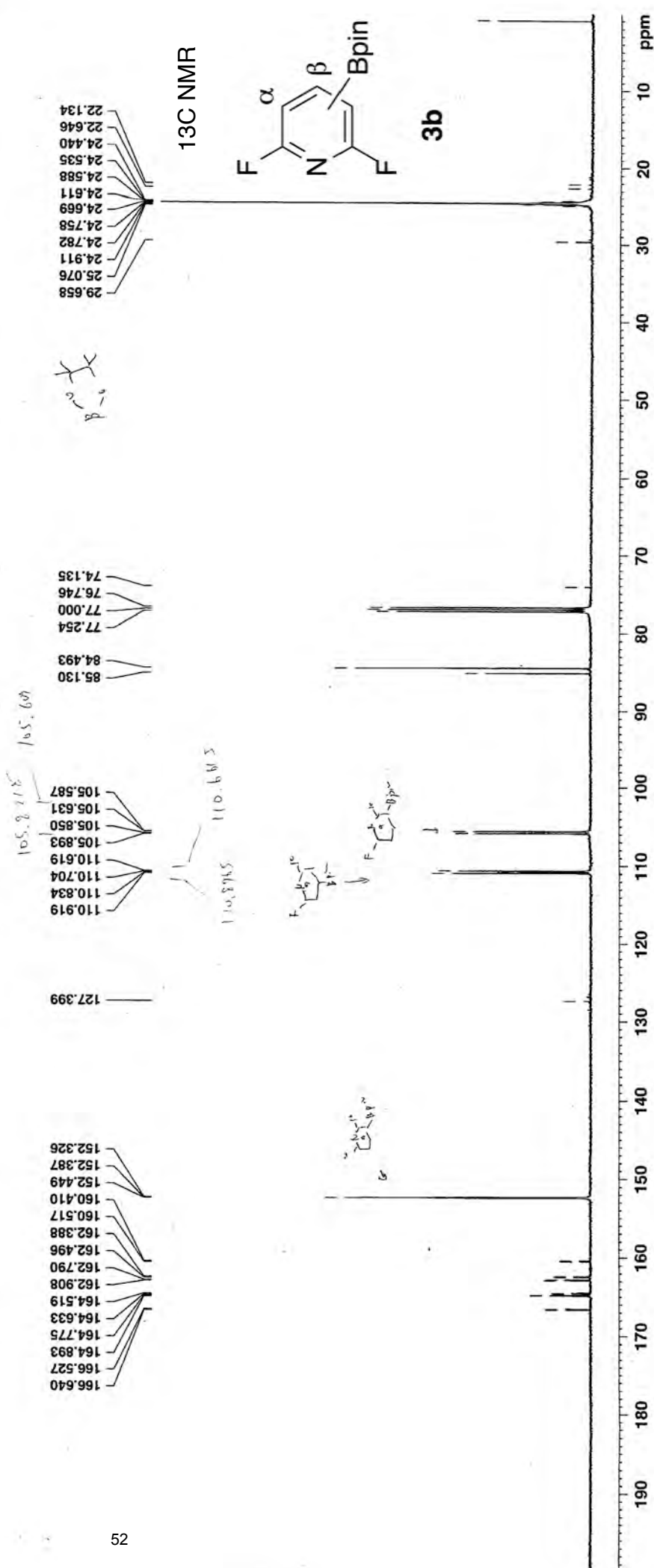
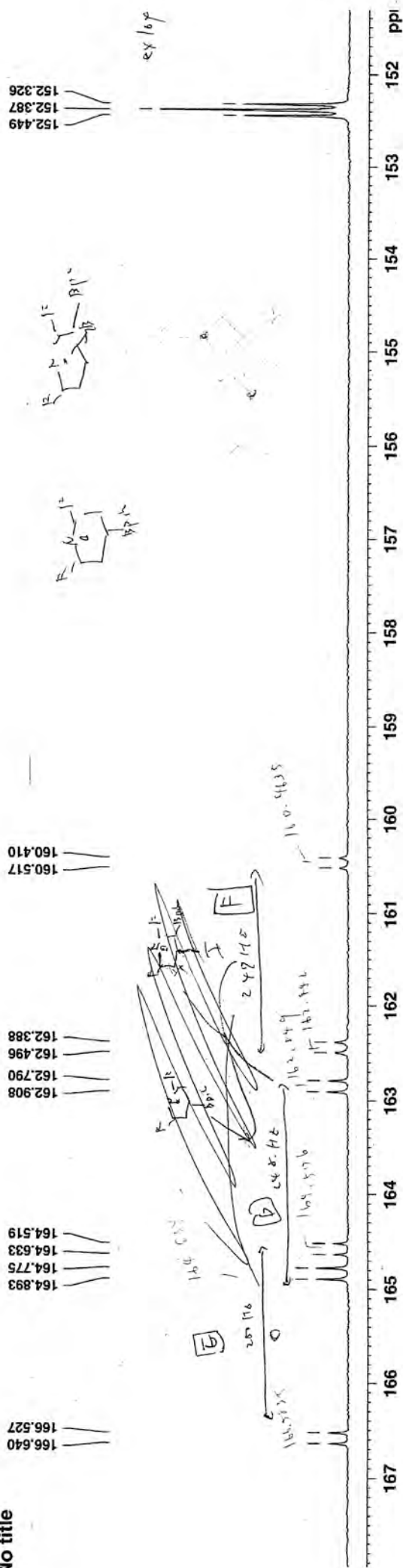


1H NMR

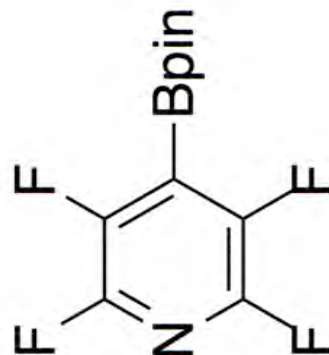


3b

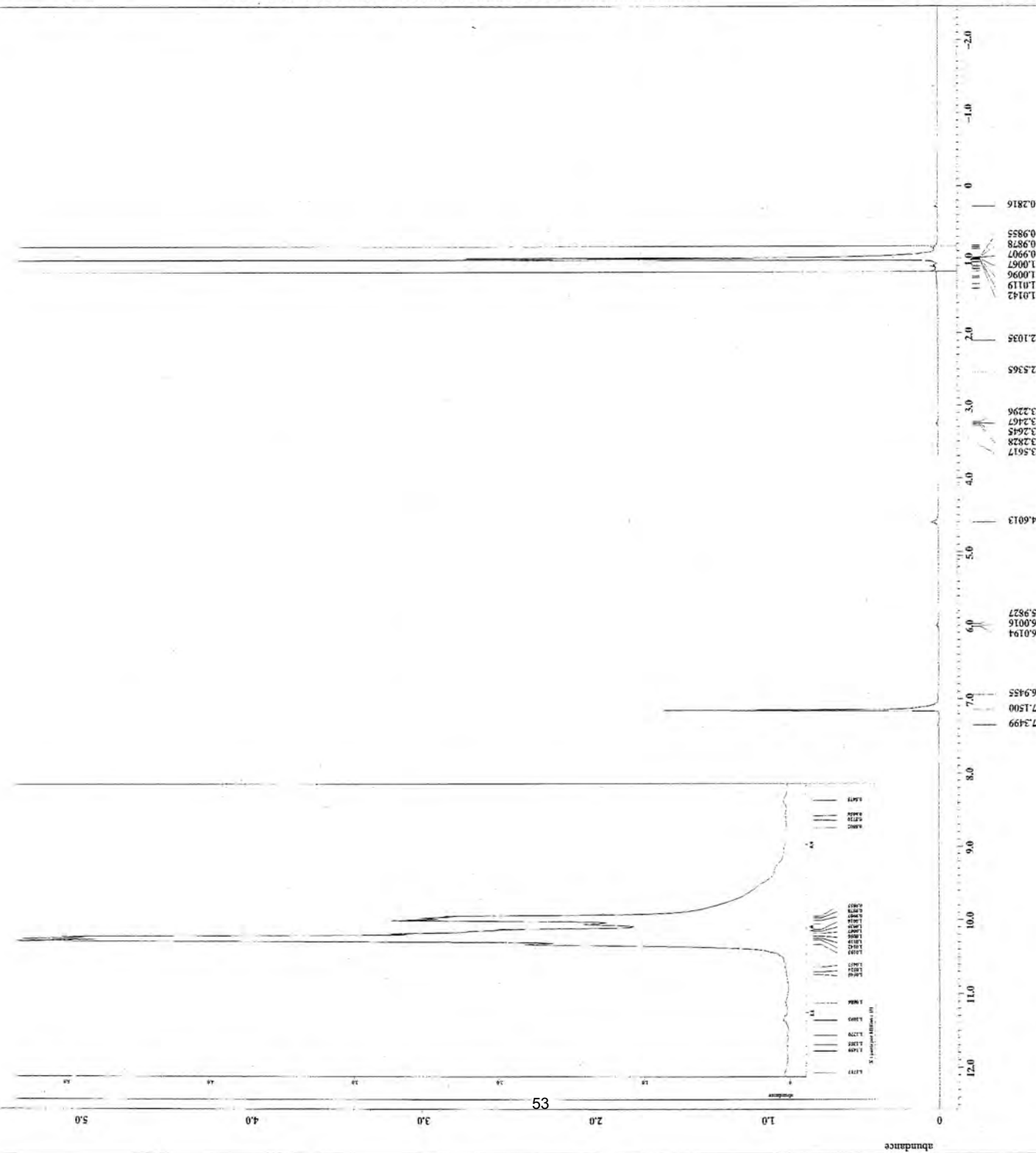




File name	n fda spEn -4.3df
Author	m dolia
Experiment	"single_pulse_ex2"
Sample_id	"A"
Solvent	BENZENE-D6
Time	3-AUG-2015 18:39:11
Acquisition time	3-AUG-2015 20:49:17
Revision time	3-AUG-2015 20:49:18
Durant time	
Comment	"single_pulse"
Data format	2021A
Bin title	1H
bin_units	[ppm]
dimensions	X CK 400
Site	DMZFA_NMR
Operator	
Field strength	"9.201797068 [T] (390 NMR)"
K _{acq} _duration	4.4564448 [s]
K _{decoupl}	1H
K _{domain}	5 [ppm]
K _{offset}	7.76655441 [MHz]
K _{points}	32768
K _{prescan}	1
K _{resolution}	0.22439396 [Hz]
K _{sweep}	1
K _{gain}	35294118 [Hz]
K _{f1_freq}	391.78655441 [MHz]
K _{txr_offset}	5 [ppm]
K _{r1_domain}	1H
K _{r1_freq}	391.78655441 [MHz]
K _{r1_offset}	0
K _{tilt_poset}	FALSE
K _{mod_return}	1
K _{scans}	8
Total_acquis	9.93 [us]
K _{90_width}	4.4564448 [s]
K _{acq_time}	45 [deg]
K _{angle}	3 [dB]
K _{atn}	4.965 [us]
K _{pulse}	OFF
K _{mode}	OFF
K _{presente}	PALZE
K _{initial_wait}	1[s]
K _{relaxation_delay}	5 [s]
K _{recov_gain}	40
K _{relaxation_time}	23.5 [sec]
K _{temp_set}	

¹H NMR

36



N : parts per Million : 1H

```

----- PROCESSING PARAMETERS -----
dc_balance : 0 : FALSE
temp : 0.2[Hz] : 0.0[s]
processed : 1 : 0[%] : 00[%] : 100[%]
xrefid1 : 1
fft : 1 : TRUE : TRUE
machinename
ppm
Derived from: N F4 Spin-1.jdf

```

```

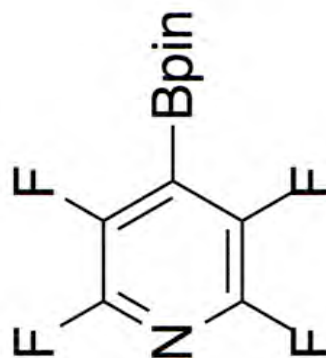
Filename = N F4 Spin-1.jdf
Author = deita
Experiment = single_pulse.ex2
Sample_id = A
Solvent = Benzene-D6
Acquisition_time = 18.45:46
Revision_time = 3-AUG-2015 20:56:19
Current_time = 3-AUG-2015 20:56:45

Comment = single_pulse
Data_format = 13007
Data_title = 13007
Dia_units = [Dpm]
Dimensions = X
Site = ECR 400
Spectrometer = spectra_200

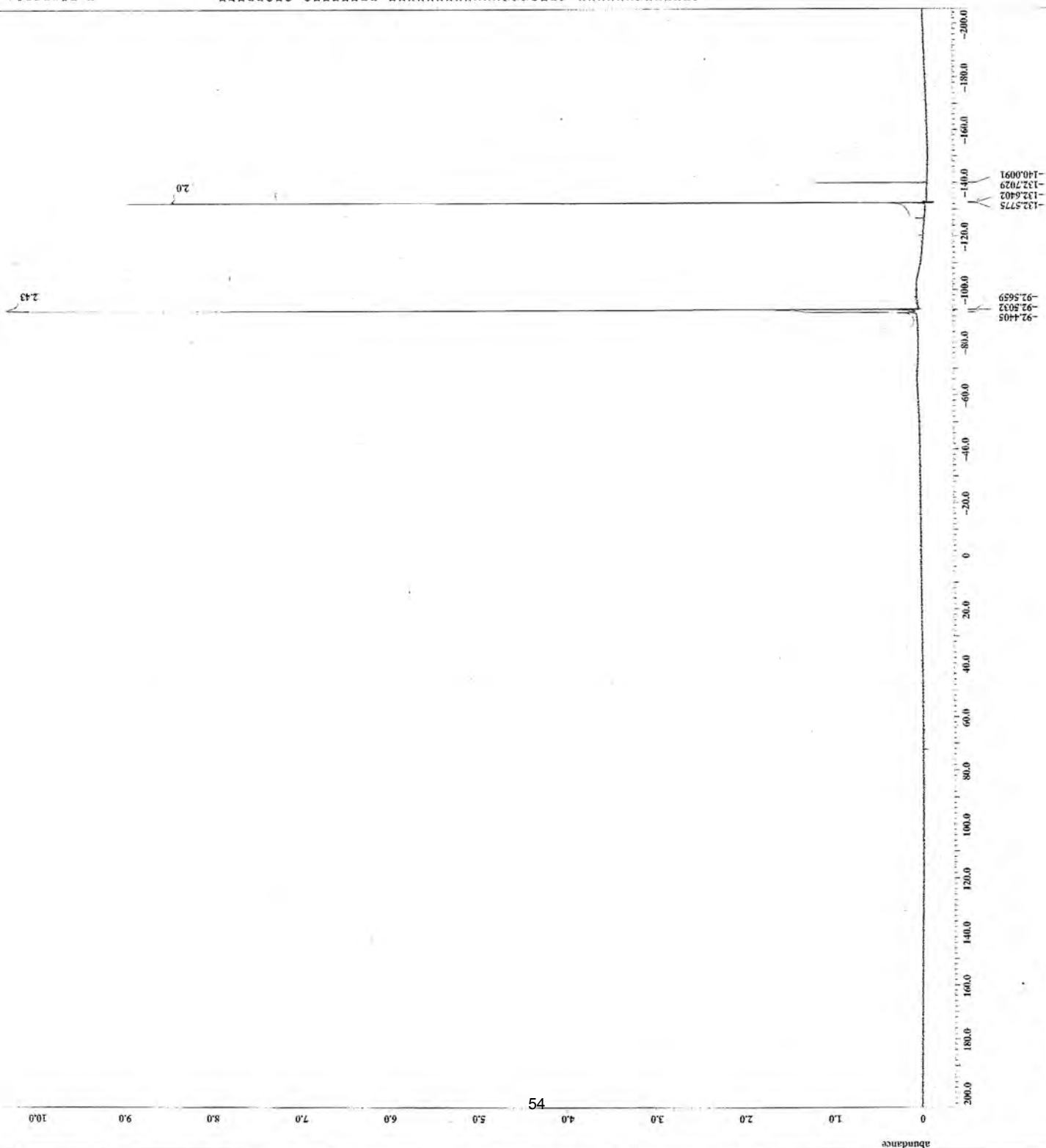
Field_strength = 9.20197068[T] (390[MHz])
X_acq_duration = 86.50752[ms]
X_domain = 19F
X_freq = 368.64763285[MHz]
X_offset = 0[Dpm]
X_points = 16384
X_resolution = 11.5596868[Hz]
X_sweep = 189.3593939[MHz]
X_gain = 19F
Irr_freq = 368.64763285[MHz]
Irr_offset = 5[Dpm]
Tri_domain = 19F
Tri_freq = 368.64763285[MHz]
Tri_offset = 5[Dpm]
Clipped = FALSE
Mod_return = 1
Scans = 51
Total_scans = 51
X_90_width = 11.5[us]
X_acq_time = 86.50752[ms]
X_angle = 45[deg]
X_atn = 3.4[dB]
X_pulse = 5.75[us]
X_offset = 0[Dpm]
Tri_mode = OFF
Data_preset = FALSE
Initial_wait = 1[s]
Recvr_gain = 50
Mixdown_delay = 5[us]
Preamplifier_time = 5.08650752[s]
Temp_get = 23.5[CC]

```

19F NMR



3C

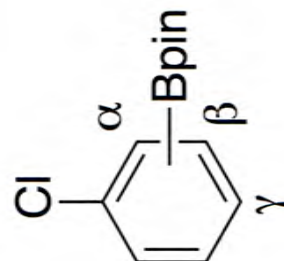


X: parts per Million : 19F

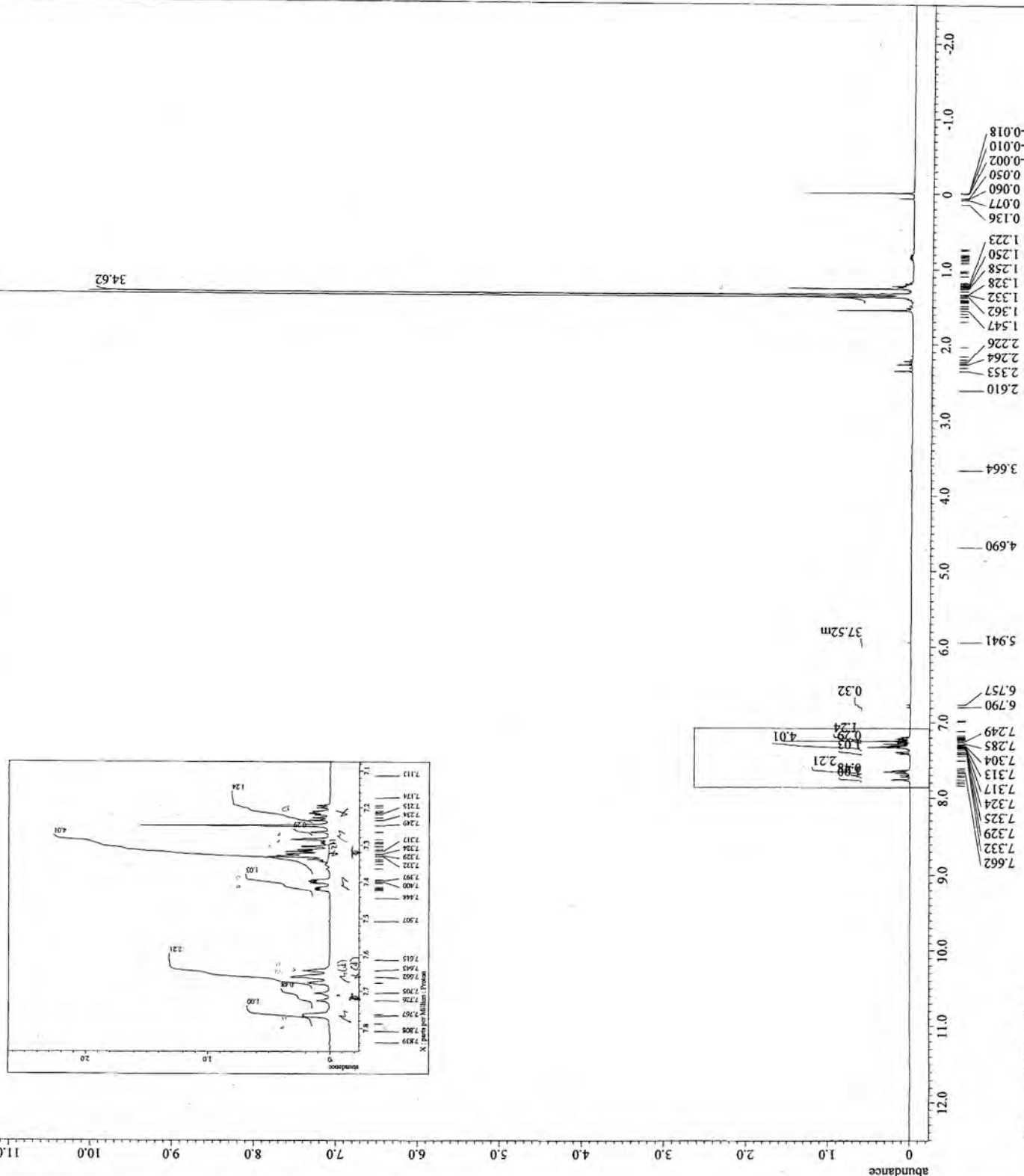
¹H NMR



$$t = 13 = \gamma = 46 = 37217$$



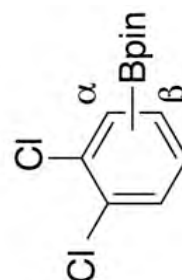
4a



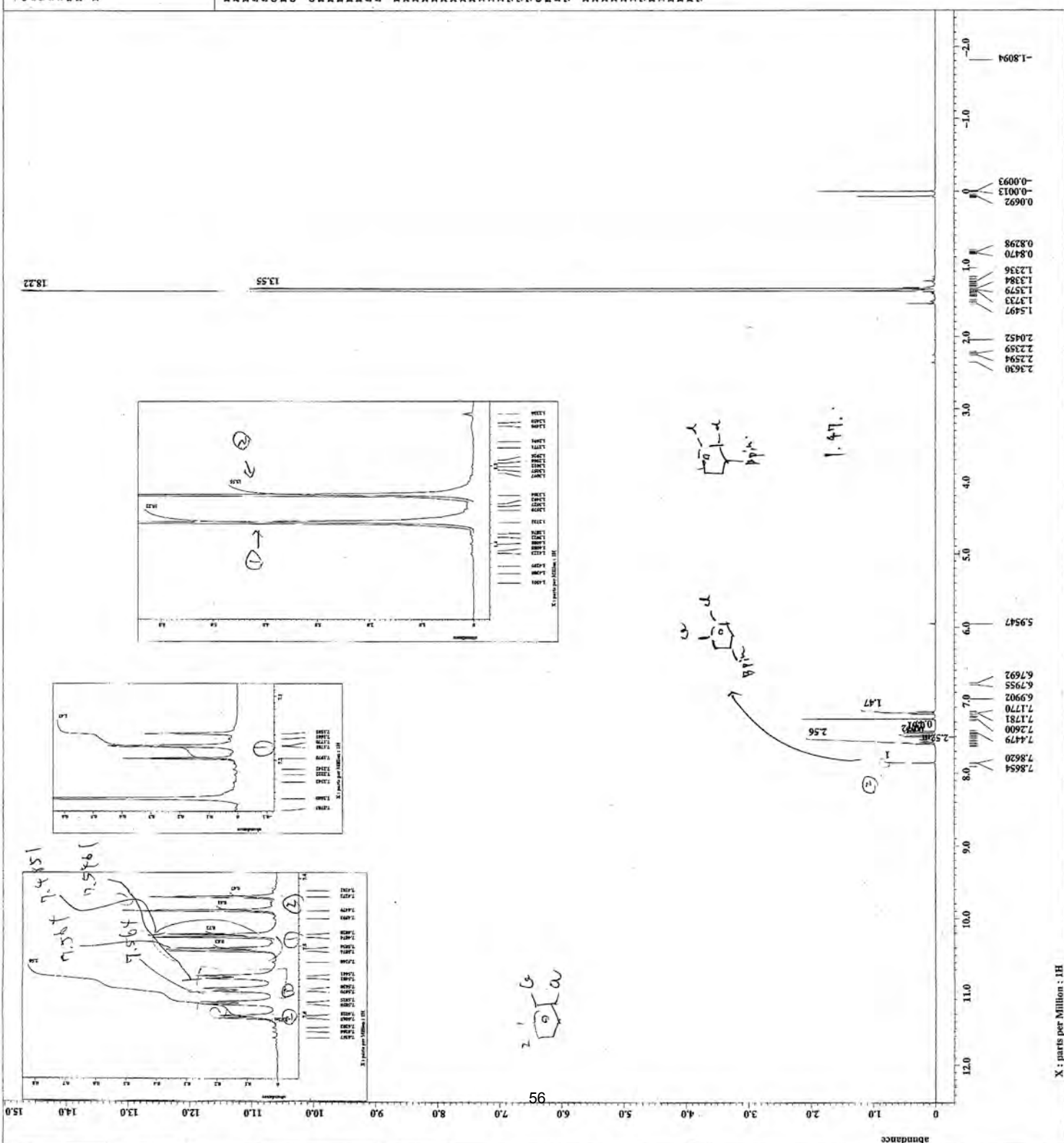
X : parts per Million : Proton

18.22

Derived from: ex148 column 1H-1.jdr

[illegible]¹H NMR

4b



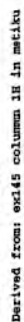
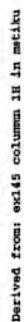
X : parts per Million : 1H



Derived from: ex145 column 1H in metaku

Derived from: ex145 column 1H in metaku

Derived from: ex145 column 1H in metaku



Derived from: ex145 column 1H in metaku

Derived from: EX157 DATE 13C-1. jdc

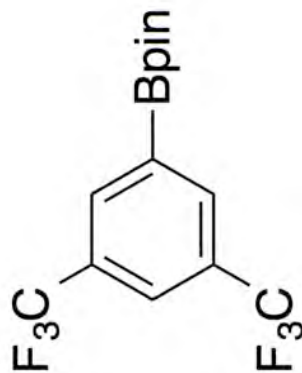
Derived from: EX157 DATE 13C-1.1.jdc



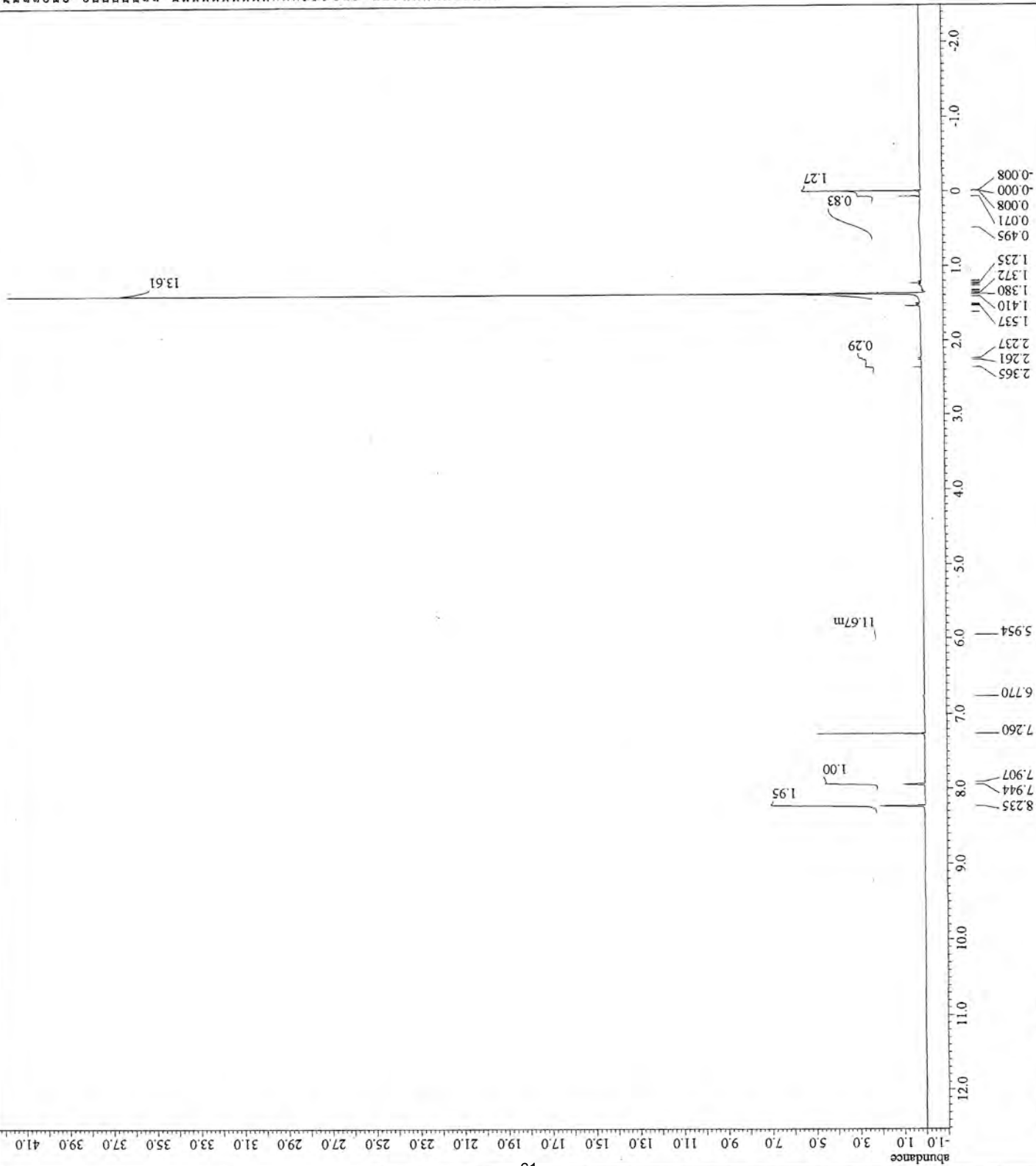
Derived from: EX157 DATE 13C-1.1.jdc

Derived from: EX157 DATE 13C-1.1.jdc

13.61

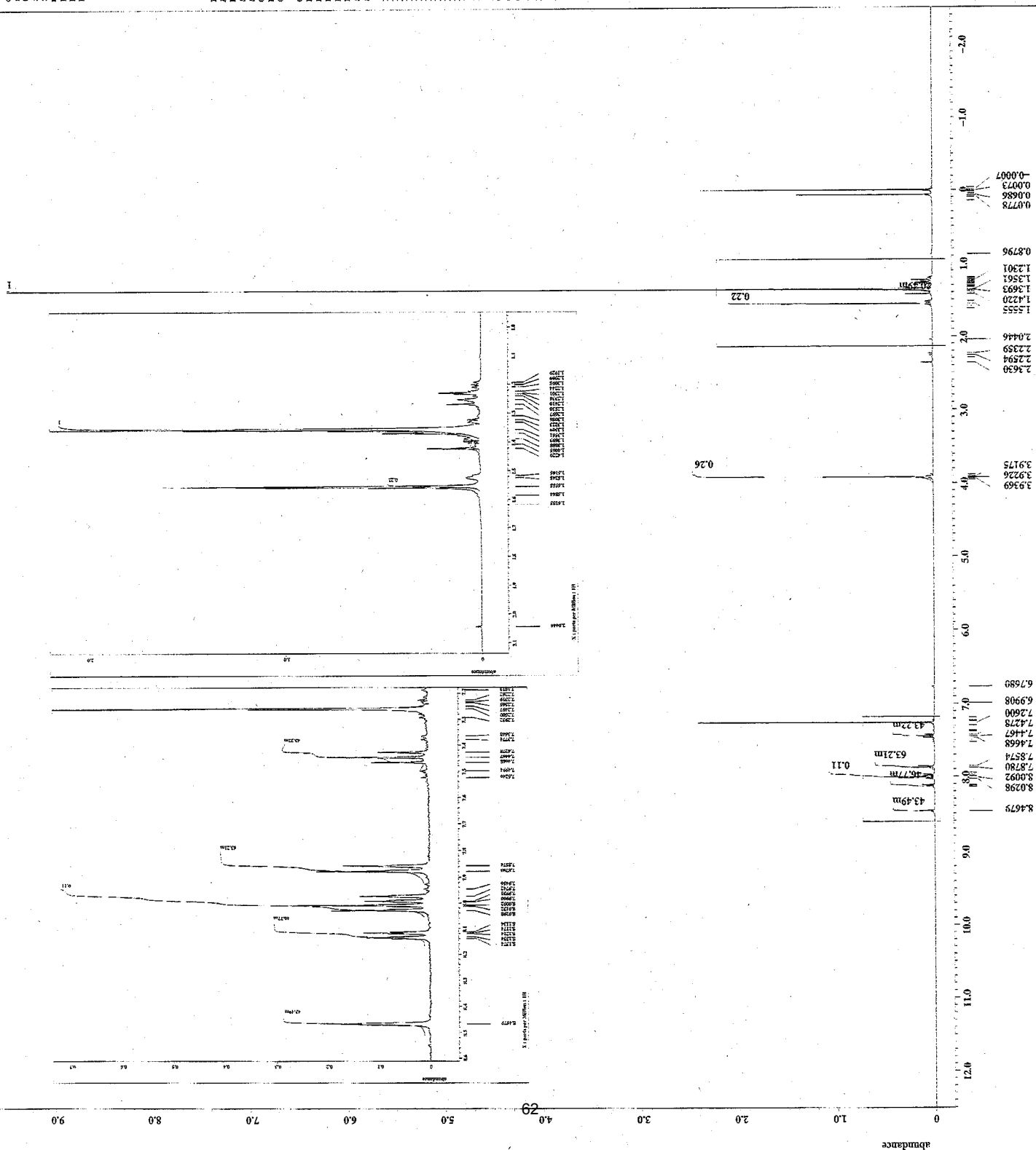
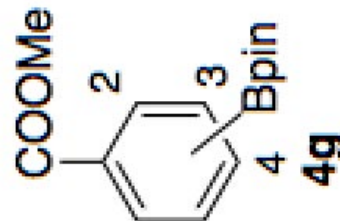
C1=CC(=C(C=C1)C(F)(F)F)C(F)(F)F

4f



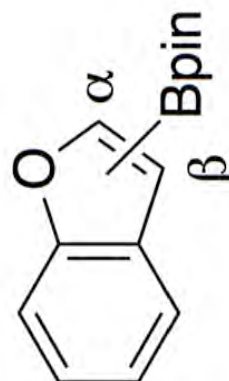
X : parts per Million : Proton

Filename	= ex275_crude_1h-18_161
Author	= GoLisa
Experiment	= single_pulse_cw2
Sample_id	= A
Acq_date	= 2015-09-08
Acq_time	= 20:17:59
Acq_start_time	= 17-SEP-2015 22:51:59
Acq_end_time	= 17-SEP-2015 22:52:50
Current_time	= 17-SEP-2015 22:53:28
Comment	= single_pulse
Experiment	= CW2
Bin_size	= 26344
Bin_start	= 1H
Bin_end	= 1H
Bin_units	= [ppm]
Dimensions	= X CX 400
Site	=
Spectrometer	= DEVA2_NMR
Field_strength	= 9.201370668 [T] [390] [MH]
X_acq_duration	= 4.4564448 [s]
X_domain	= 1H
X_offset	= 391.78655441 [MHz]
X_points	= 510 [ppm]
X_prescans	= 32766
X_resolution	= 1
Y_acq_duration	= 0.2249399 [Hz]
Y_domain	= 1H
Y_offset	= 391.78655441 [MHz]
Y_points	= 510 [ppm]
Y_prescans	= 1
Y_resolution	= 1H
Z_acq_duration	= 0.2249399 [Hz]
Z_domain	= 1H
Z_offset	= 391.78655441 [MHz]
Z_points	= 510 [ppm]
Z_prescans	= 1
Z_resolution	= 1H
Ir_freq	= 500.136051 [MHz]
Ir_domain	= 1H
Ir_offset	= 391.78655441 [MHz]
Ir_points	= 510 [ppm]
Ir_prescans	= 1
Ir_resolution	= 1H
Mod_return	= 1
Scans	= 8
Total_scans	= 8
X_90_width	= 9.93 [us]
X_acq_time	= 4.4564448 [s]
X_angle	= 45 [deg]
X_tau	= 3 [ns]
X_pulse	= 100 [us]
X_mode	= Off
T1_mode	= Off
Dance_preset	= FALSE
Initial_val	= 1 [s]
Recur_gain	= 5 [dB]
Recur_delay	= 5 [us]
Repetition_time	= 9.4564448 [s]
Temp_get	= 23.3 [C]

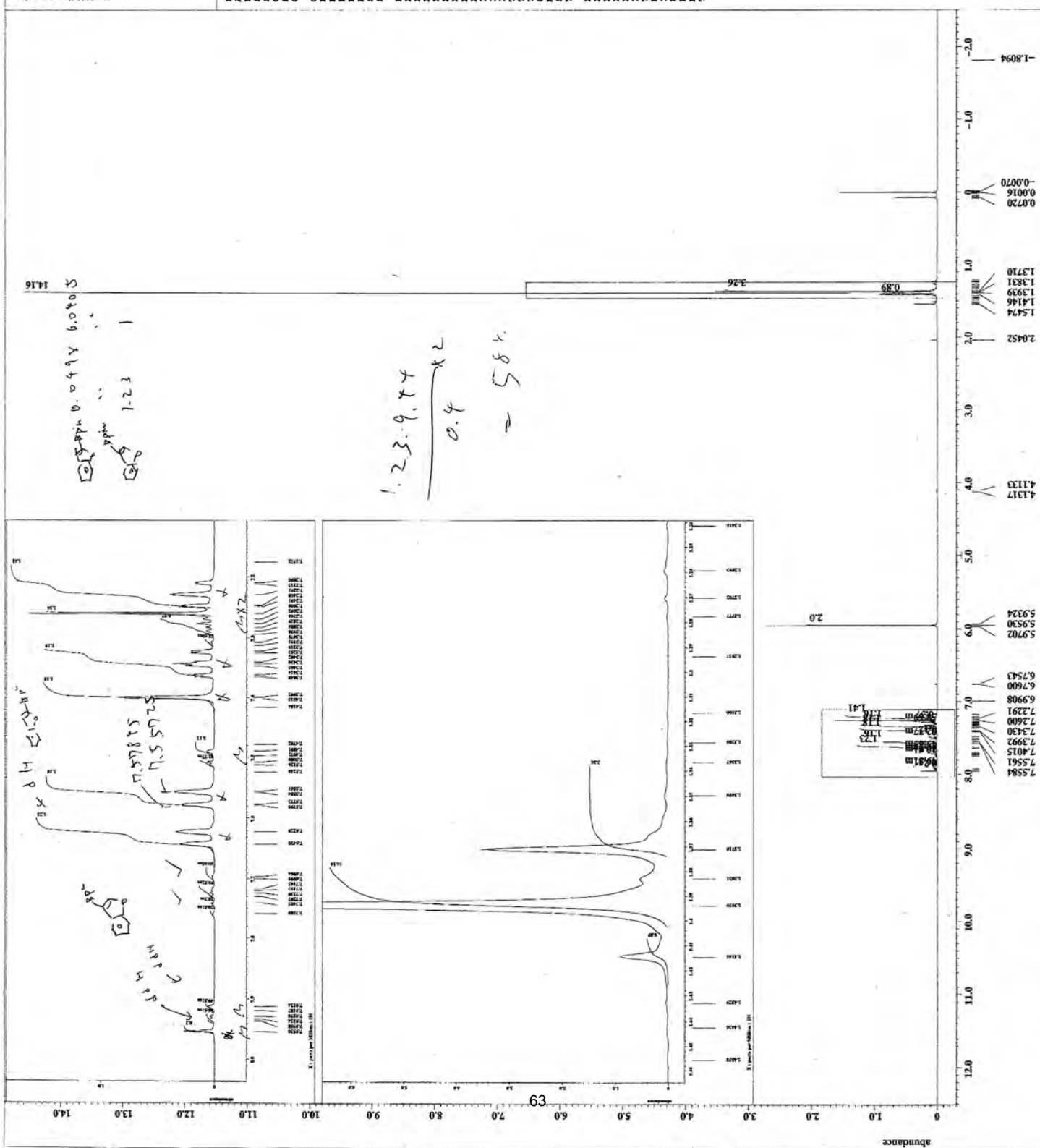


62

Filename	= ex_109 column=5.fdr
Author	= delta
Experiment	= single_pulse
Sample_id	= A
Solvent	= DMSO-D6
Acq_time	= 13:08:14
Rev_time	= 14-OCT-2014 14:09:13
Revision_time	= 14-OCT-2014 14:09:13
Current_time	=
Comment	= single_pulse
Format	= DELTA2
bin_size	= 26214
bin_title	= 1H
bin_units	= [ppm]
dimensions	= X 400
Site	= DELTA2_00R
Spectrometer	=
Field_strength	= 9.201701068 [T] 390 [MH]
Acq_duration	= 4.456448 [s]
Domain	= 1
X_offset	= 391.78655441 [MHz]
X_points	= 32768
X_resolution	= 1
Y_resolution	= 0.22439396 [Hz]
F1_domain	= 1
F1_freq	= 391.7865441 [MHz]
F1_offset	= 5 [ppm]
F1_domain	= 1H
F1_freq	= 391.78655441 [MHz]
F1_offset	= 5 [ppm]
Clipped	= FALSE
Modo_return	= 1
Scans	= 8
Total_scans	= 8
Acq_width	= 9.7 [us]
Acq_time	= 4.456448 [s]
X_angle	= 45 [deg]
X_atn	= 3 [dB]
X_pulse	= 0 [us]
Pri_mode	= Off
Element_present	= FALSE
Initial_wait	= 1 [s]
Acq_gain	= 44
Delay	= 24.5 [us]
Resolving_time	= 9.456448 [s]
Temp_get	= 24.5 [C]

$$X = \sqrt{118} = 10.86$$


5a



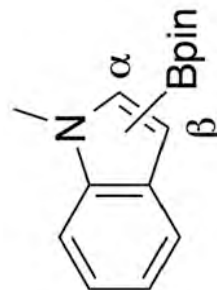
X : parts per Million : 1H

with 1,1,2,2-tetrachloroethane (5.9 ppm) as an internal standard

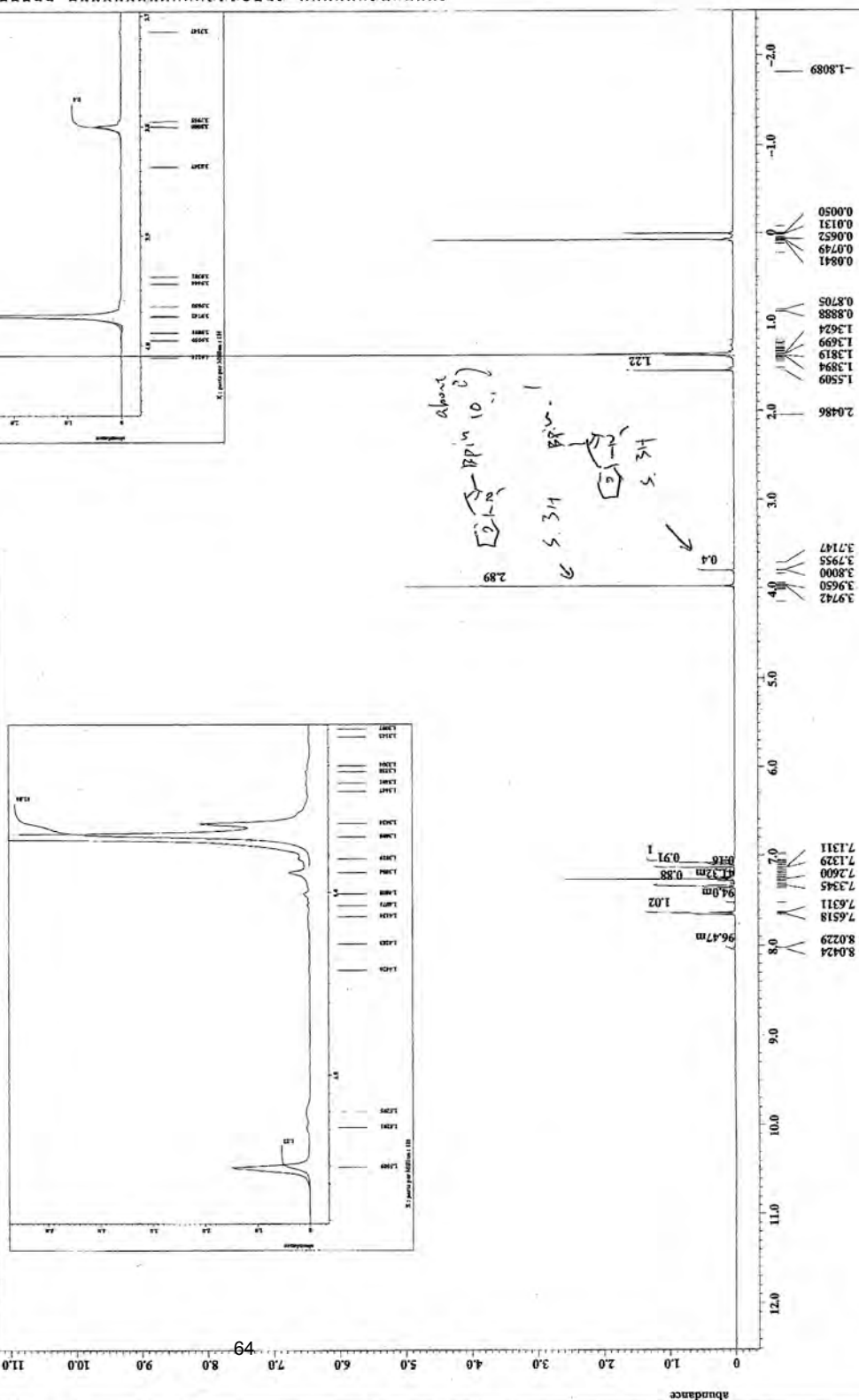
derived from: ex135 column IH-6.jdd

Filespec	=	ex135 column 1h-10.1
Author	=	edra3 pulse.exe2
Experiment	=	CHLOROPHY-a
Sample_id	=	15-51-15
Solvent	=	28-NOV-2014 15:52:31
Acq_time	=	28-NOV-2014 15:52:31
Revision_time	=	28-NOV-2014 15:53:13
Current_time	=	single_pulse
Comment	=	COMPLEX
Data format	=	2024.4
File title	=	1h
File units	=	[ppm]
Dimensions	=	X
Site	=	NOX 400
Apparatus	=	DMXAI_NMR
Acq_strength	=	9.201570668 [W] 390 [MHz]
X_acq_duration	=	4.456448 [s]
X_domain	=	33.78655441 [MHz]
X_offset	=	33766
X_points	=	33766
X_prescan	=	1
X_resolution	=	0.22439396 [Hz]
Acq_freq	=	33.78655441 [MHz]
Trf_domain	=	33.78655441 [MHz]
Trf_offset	=	5 [ppm]
Trf_domain	=	1h
Trf_freq	=	33.78655441 [MHz]
Trf_offset	=	5 [ppm]
Clipped	=	FALSE
Mod_return	=	1
Mod_return	=	8
Scans	=	8
Total_scans	=	8
X0_width	=	9.7 [us]
X_acq_time	=	4.456448 [s]
X_angle	=	45 [deg]
X_atn	=	3 [dB]
X_pulse	=	0.8 [us]
Trf_mode	=	Off
Trf_prescan	=	Off
Trf_offset	=	FALSE
Initial_wait	=	1 [s]
Recvr_gain	=	5 [s]
Repetition_time	=	9.456448 [s]
Relaxation_time	=	24.4 [s]
Temp_get	=	24.4 [C]

¹H NMR



5b



X : parts per Million : 1H

The image displays two NMR spectra of a polymer sample. The top spectrum is a ^1H NMR spectrum, and the bottom spectrum is a ^{13}C NMR spectrum. Both spectra include a list of chemical shifts on the right side.

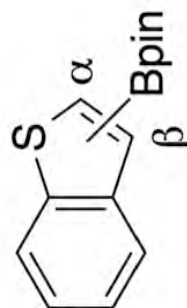
^1H NMR Spectrum (Top):

- Chemical shifts (ppm): 11.9, 6.9, 5.7.
- Integration values: 1.00, 1.00, 1.00.
- Chemical shift range: 0 to 12 ppm.

^{13}C NMR Spectrum (Bottom):

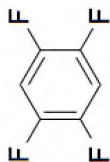
- Chemical shifts (ppm): 161.2, 151.2, 141.2, 131.2, 121.2, 111.2, 101.2, 91.2, 81.2, 71.2, 61.2, 51.2, 41.2, 31.2, 21.2, 11.2, 0.2.
- Integration values: 1.00, 1.00, 1.00, 1.00, 1.00, 1.00, 1.00, 1.00, 1.00, 1.00, 1.00, 1.00, 1.00, 1.00, 1.00, 1.00, 1.00.
- Chemical shift range: 0 to 170 ppm.

OK



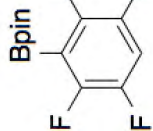
55

X : parts per Million : 1H



HBpin

1.5 mol% **1a**
1.5 mol% ⁿBuLi
mesitylene
150 °C, 4 h

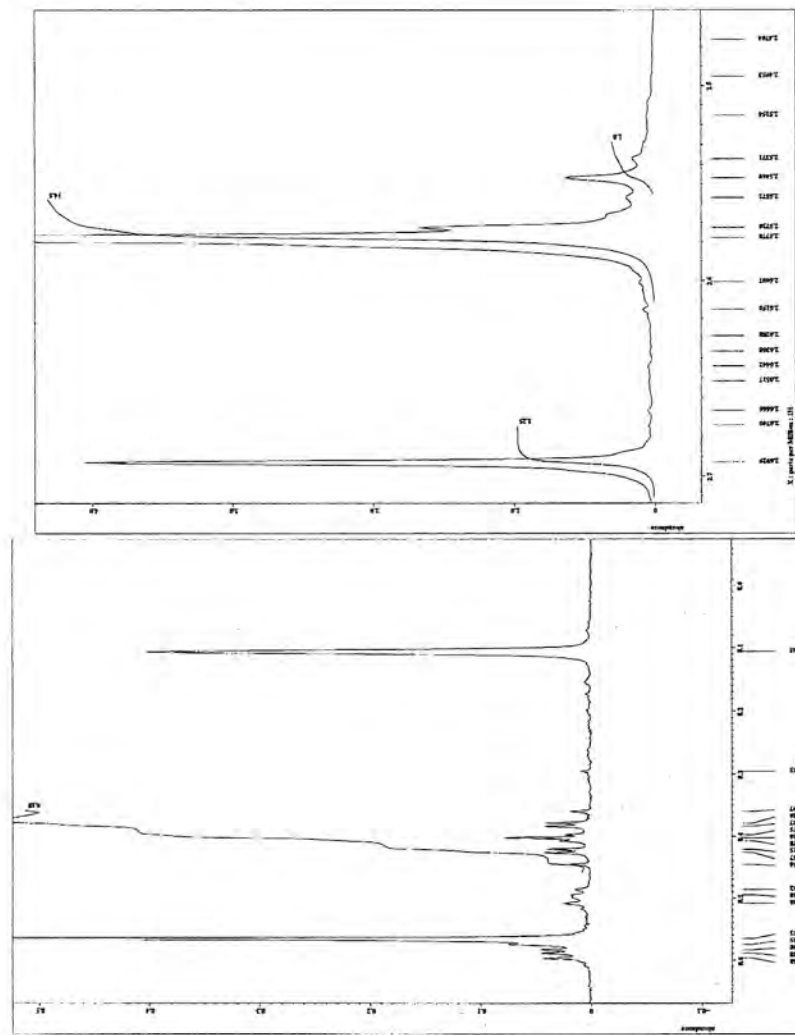


(10 equiv.)

2n

15% (NMR yield)

14.5



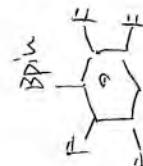
66

```

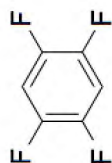
=====
File      : 2021 crude 1H-4.jdx
Date      : 2021-01-14
Sample    : 2021-01-14
Solvent   : CHLOROFORM-D
Creation  : 2021-01-14 19:27:39
AcqTime   : 2021-01-14 20:21:22
Current   : 2021-01-14 20:26:02
=====
Data      : single_pulse
Format    : 1D
File      : 2021-01-14
Dir       : 2021-01-14
Dim       : 1
Unit      : [ppm]
Dim       : 1
Unit      : [ppm]
Dimensions : 1
Site      : ECH 400
Spectrum  : 2021-01-14_NMR
=====
Field      : 9.20197058 [T] (390 MHz)
AcqDur     : 4.456448 [s]
X1         : 1H
X2         : 1H
X3         : 391.78655441 [MHz]
X4         : 327.66
X5         : 1
X6         : 0.22439396 [Hz]
X7         : 7.35294118 [kHz]
X8         : 1H
X9         : 391.78655441 [MHz]
X10        : 5 [ppm]
X11        : 1H
X12        : 391.78655441 [MHz]
X13        : 5 [ppm]
X14        : 20.00
X15        : 8
TotalScans : 8
=====
X10Width   : 9.7 [um]
X10Time    : 4.456448 [s]
X10Date    : 2021-01-14
X10Attn    : 3 [dB]
X10Pulse   : 4.85 [us]
X10Mode    : OFF
X10Phase    : 0.00
X10Offset  : 0.00
X10Gain    : 11.0
X10RecvrGain : 46
X10RelaxationDelay : 5 [s]
X10RepetitionTime : 9.456448 [s]
X10TempGet : 25.1 [dC]
=====

```

1H NMR

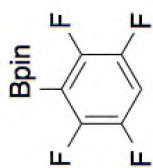


X : parts per Million : 1H



HBpin

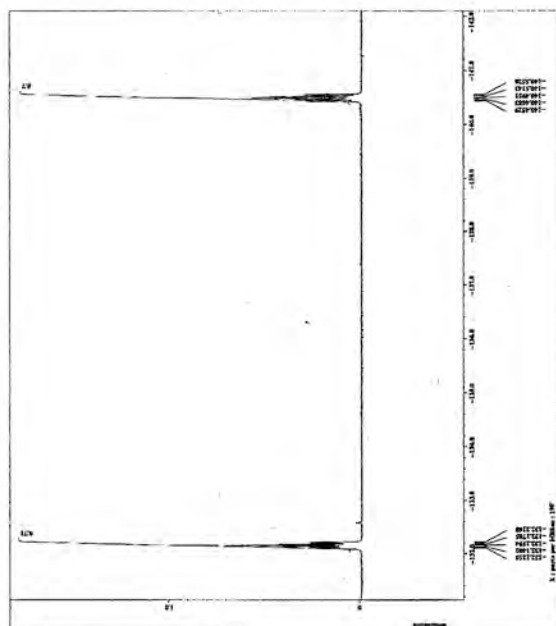
(10 equiv.)



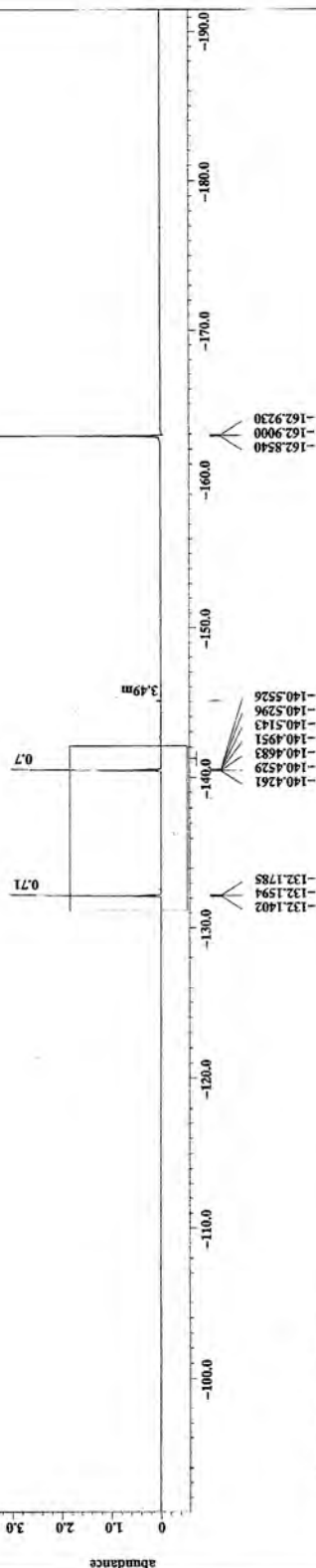
1.5 mol% 1a 1.5 mol% ⁿ BuLi	mesitylene 150 °C, 4 h
--	---------------------------

2n

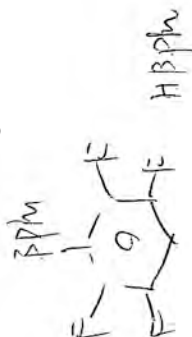
15% (NMR yield)



$$\frac{0.7 \cdot 8.622 \cdot 10^{-4}}{2} = 15\%$$



19F NMR



```

=sw72 crudo 19F-5-JAR
= dola
= single_pulse.exe2
= A
= CHLOROFORM-D
= 1-SEP-2014 19:21:37
= 2-SEP-2014 20:20:21
= 3-SEP-2014 20:20:21
= 4-SEP-2014 20:20:25
= single_pulse
= COMPLEX
= 202.44
= 19F
= [ppm]
= X
= KX 400
= MZRA12345678
= 9.201379668[F] (390[MHz]
= 0.7077888[s]
= 19F
= 64763285 [MHz]
= -94.0 [ppm]
= 337.69
= 1
= 1.41285084 [Hz]
= 16.2926265 [Hz]
= 368.64763285 [MHz]
= 5 [ppm]
= 19F
= 368.64763285 [MHz]
= FALSE
= FALSE
= 1
= 28
= 28
= 11.5[us]
= 0.7077888[s]
= 45[deg]
= 3
= 3.4[dd]
= 5.7[us]
= 0
= Off
= 0
= 0
= Date present
= Initial wait
= 1[s]
= 54
= Relaxation delay
= 5.7077888[s]
= 25.1[sec]
= Temp set

```

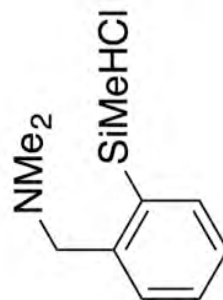
X : parts per Million : 19F

Ito ex 3

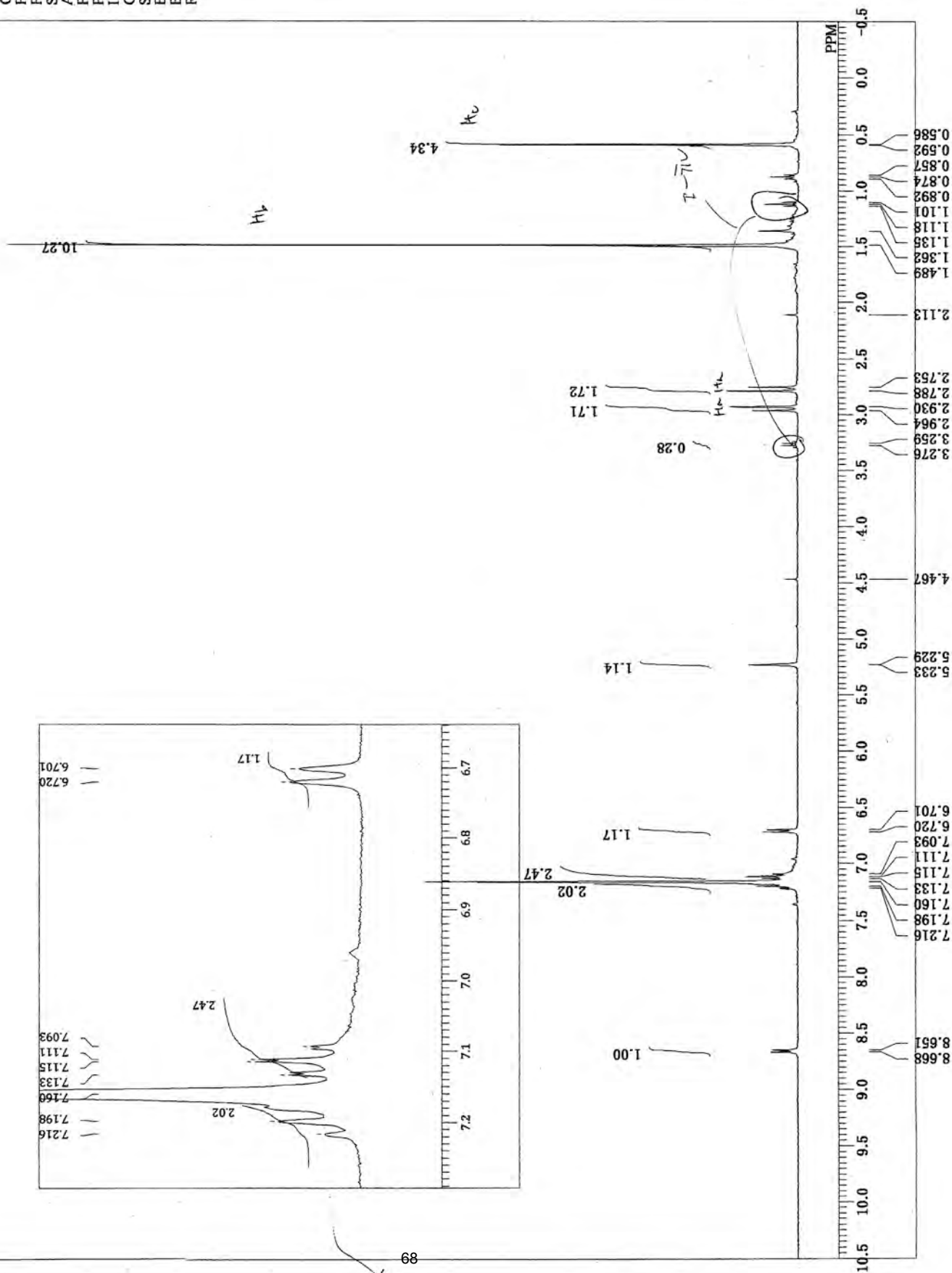
_DEFAULT.ALS
 auto
 Mon Apr 21 19:18:29 :
 1H
 NON
 399.65 MHz
 124.00 KHz
 10500.00 Hz
 16384
 7992.01 Hz
 16
 2.0500 sec
 1.5000 sec
 6.80 usec
 1H
 26.9 c
 C6D6
 7.16 ppm
 0.12 Hz
 19
 RGAIN

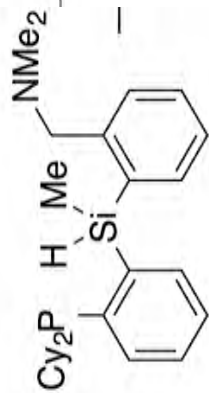
exp 3
9 1/2 分钟

1H NMR

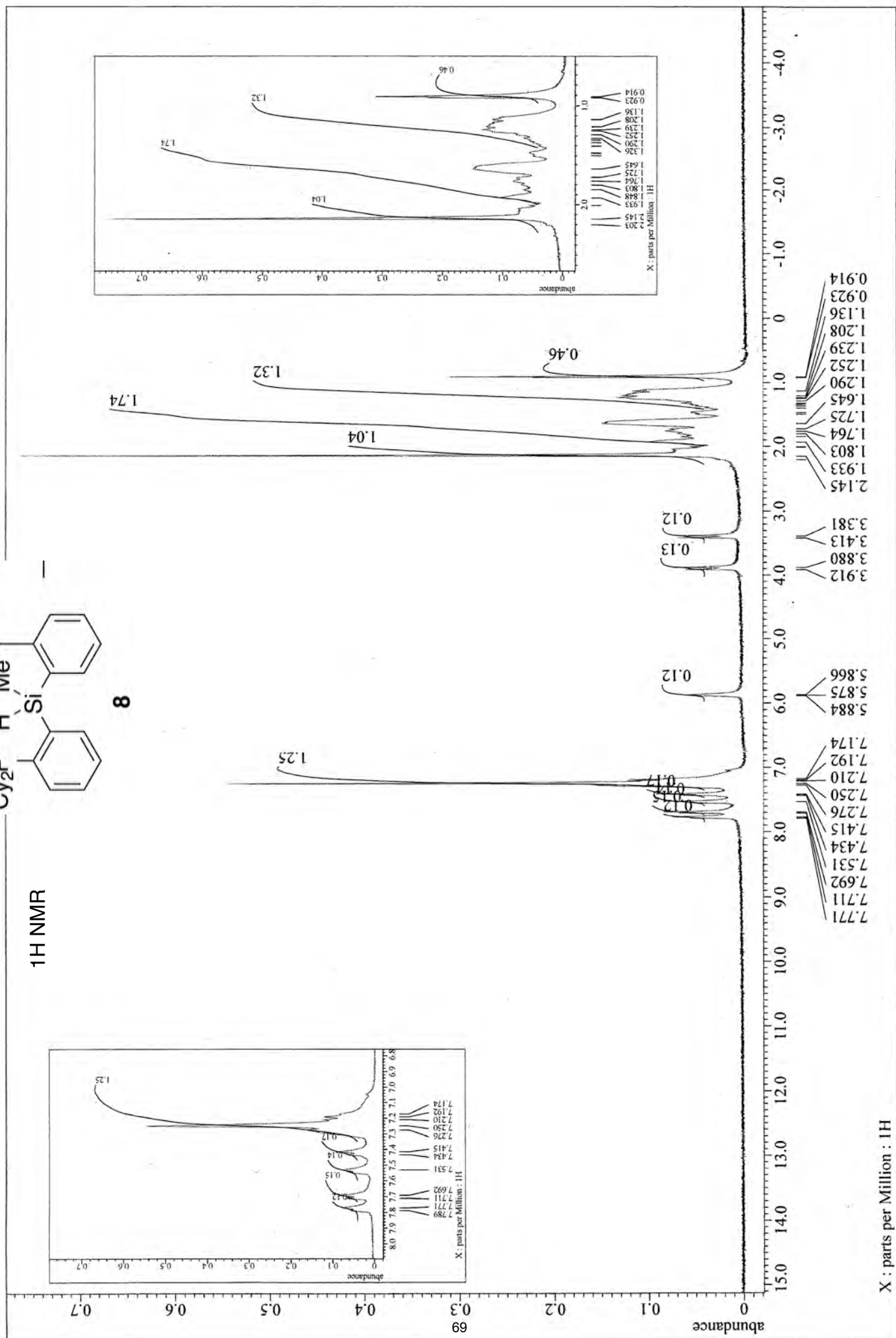


7



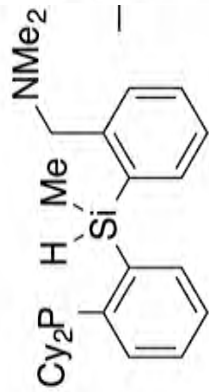


¹H NMR



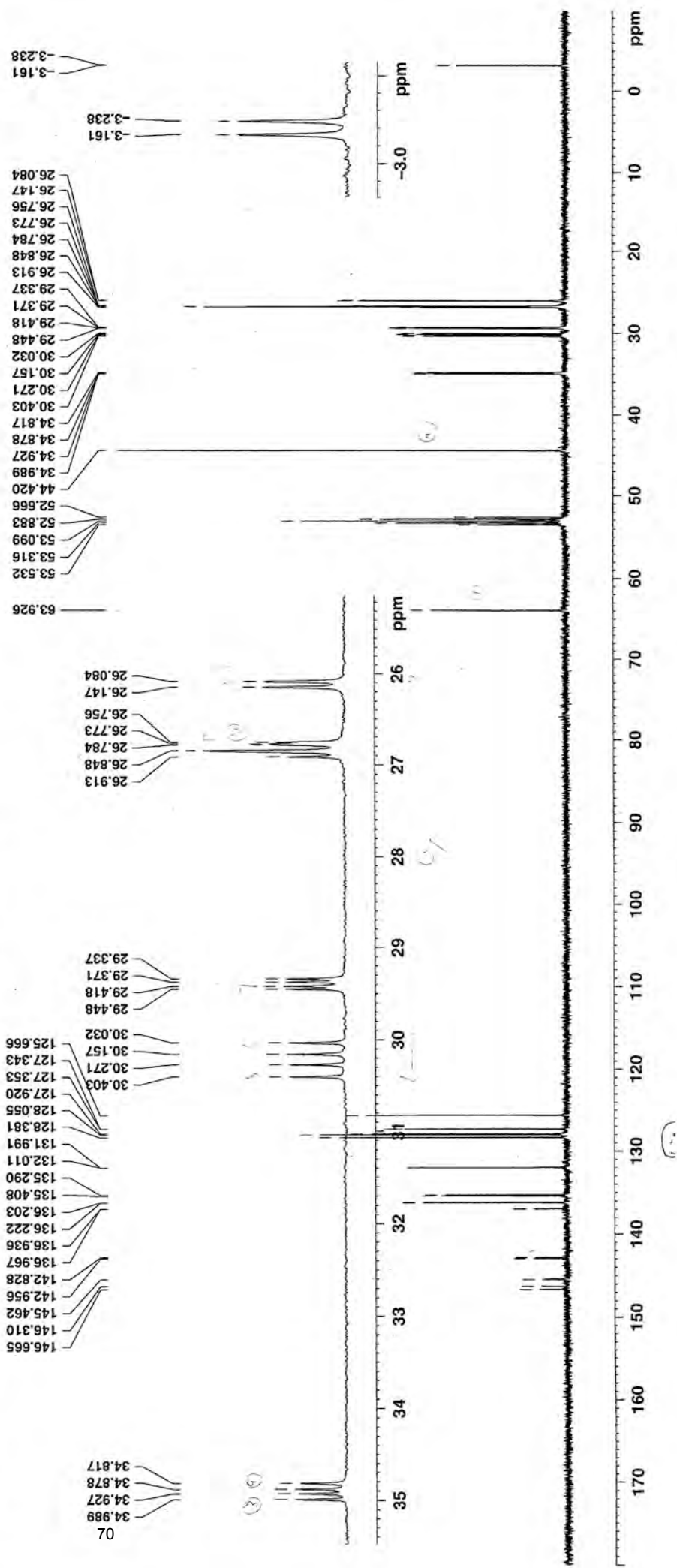
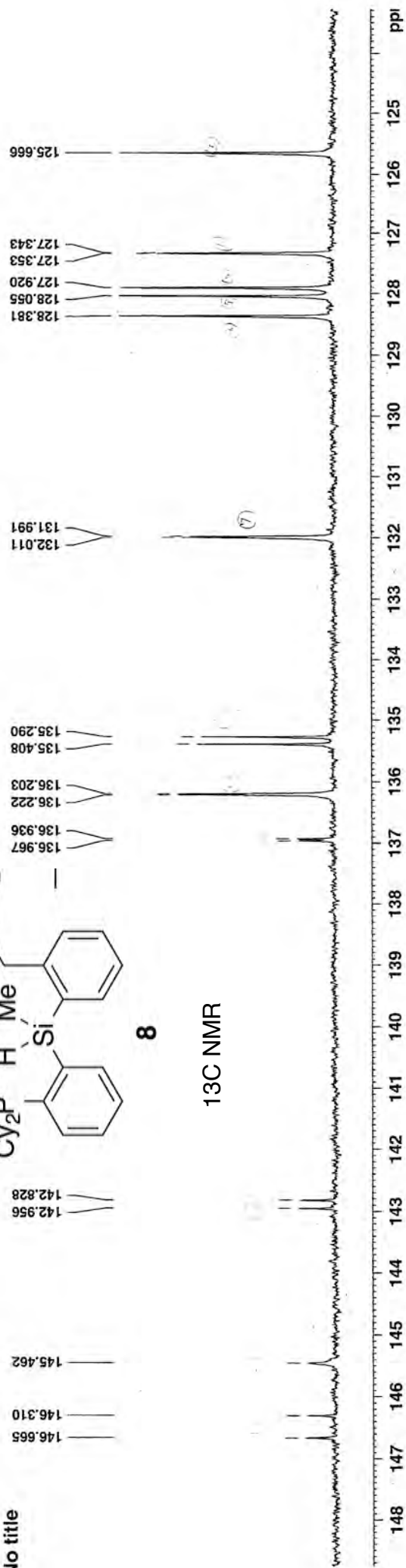
X : parts per Million : 1H

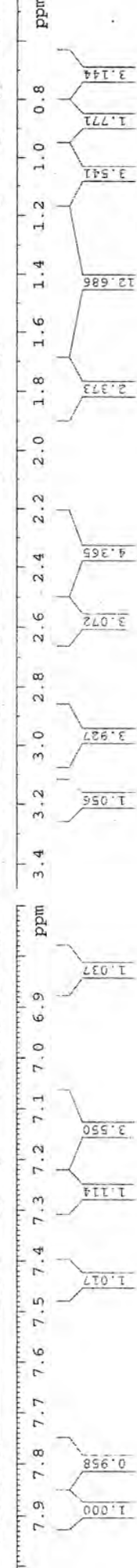
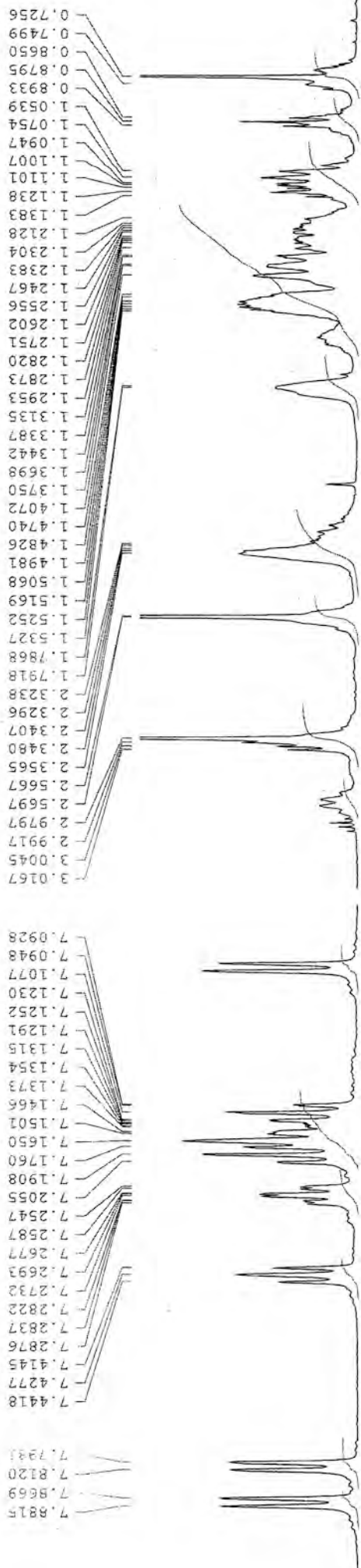
No title



8

¹³C NMR





Current Data Parameters
 NAME Shisei
 EXPNO 6
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20150630
 Time 23.00
 INSTRUM spect
 PROBHD 5 mm Multinucl
 PULPROG zg
 TD 159996
 SOLVENT CDCl3
 NS 16
 DS 2

SWH 20000.000 Hz
 FIDRES 0.125003 Hz
 AQ 3.9999750 sec
 RG 143.7
 DW 25.000 usec
 DE 6.00 usec
 TE 300.0 K

D1 3.00000000 sec
 MCREST 0.00000000 sec
 MCWREK 0.01500000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 10.00 usec
 PL1 -3.50 dB
 SFO1 500.1274993 MHz

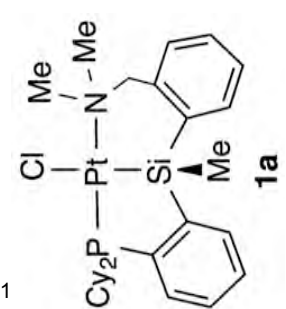
F2 - Processing parameters
 SI 32768
 SF 500.1300538 MHz
 WDW EM
 SSB 0
 LB 0.20 Hz
 GB 0
 PC 1.00

===== CHANNEL f1 =====
 NUC1 1H
 P1 10.00 usec
 PL1 -3.50 dB
 SFO1 500.1274993 MHz

F2 - Processing parameters
 SI 32768
 SF 500.1300538 MHz
 WDW EM
 SSB 0
 LB 0.20 Hz
 GB 0
 PC 1.00

===== CHANNEL f1 =====
 NUC1 1H
 P1 10.00 usec
 PL1 -3.50 dB
 SFO1 500.1274993 MHz

F2 - Processing parameters
 SI 32768
 SF 500.1300538 MHz
 WDW EM
 SSB 0
 LB 0.20 Hz
 GB 0
 PC 1.00



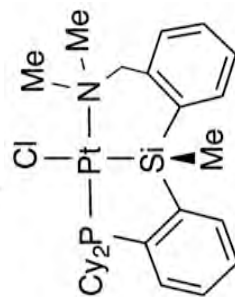
¹H NMR

10 9 8 7 6 5 4 3 2 1 0

7.91
7.89
7.87
7.85
7.83
7.81
7.79
7.77
7.75
7.73
7.71
7.69
7.67
7.65
7.63
7.61
7.59
7.57
7.55
7.53
7.51
7.49

1.99
1.97
1.95
1.93
1.91
1.89
1.87
1.85
1.83
1.81
1.79
1.77
1.75
1.73
1.71
1.69
1.67
1.65
1.63
1.61
1.59
1.57
1.55
1.53
1.51
1.49

1H NMR



1a

¹³C NMR

