Total Synthesis and Stereochemical Assignment of Scytonemin A

Junyang Liu,^{a,b} Lei Wang,^a Juefei Zhang,^a Zhengshuang Xu, ^{*a} and Tao Ye^{*a,b}

a. Laboratory of Chemical Genomics, Peking University Shenzhen Graduate School, University Town, Xili, Shenzhen, China, 518055.

b. Department of Applied Biology & Chemical Technology, The Hong Kong Polytechnic University, Hong Kong.

Email: yet@pkusz.edu.cn; xuzs@pkusz.edu.cn

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1. General Experimental

All reactions were conducted in flame-dried or oven-dried glassware under an atmosphere of dry nitrogen or argon. Oxygen and/or moisture sensitive solids and liquids were transferred appropriately. Concentration of solutions in vacuo was accomplished using a rotary evaporator fitted with a water aspirator. Residual solvents were removed under high vacuum (0.1-0.2 mm Hg). All reaction solvents were purified before use: Tetrahydrofuran were distilled from sodium benzophenone ketyl. Toluene was distilled over molten sodium metal. Dichloromethane, dimethylformamide, diethylamine, triethylamine and diisoproylethylamine were distilled from CaH₂. Methanol was distilled from Mg/I₂. Flash column chromatography was performed using the indicated solvents on E. Qingdao silica gel 60 (230 - 400 mesh ASTM). TLC was carried out using pre-coated sheets (Qingdao silica gel 60-F250, 0.2 mm). Compounds were visualized with UV light, iodine, p-anisaldehyde stain, ceric ammonium molybdate stain, or phosphomolybdic acid in EtOH. 1H NMR spectra were recorded on Bruker DPX 300 MHz, AV 500 MHz or AV 600 MHz spectrometers. Chemical shifts were reported in parts per million (ppm), relative to either a tetramethylsilane (TMS) internal standard or the signals due to the solvent. The following abbreviations are used to describe spin multiplicity: s = singlet, d = doublet, t = triplet, q = quartet, qn = quintet, m = multiplet, br = broad, dd = doubletdoublet of doublets, dt = doublet of triplets, dq = doublet of quartets, ddd = doublet of doublets; other combinations are derived from those listed above. Coupling constants (J) are reported in Hertz. 13C NMR spectra were completely heterodecoupled and measured at 75, 125, or 150 MHz. High resolution mass spectra were measured on ABI Q-star Elite. Optical rotations were recorded on a Perkin-Elmer 351 polarimeter at 589 nm, 100 mm cell at 20°C. Data were reported as follow: optical rotation (c (g/100 mL), solvent).

2. Experimental procedures

2.1. Synthesis of pentapeptide 2

2.1.1 Synthesis of Boc-*D*-(2*R*,3*S*)-threo-3-hydroxyleucine (HyLeu) and dipeptide 5



Alcohol $5a^{[1]}$ (1.50 g, 5.49 mmol) was dissolved in AcOH-H₂O (25 mL, 5/1, v/v) and stirred at room temperature for 24 h. The reaction was cooled to 0 °C and quenched by addition of NaHCO₃(s) to adjust pH 10–11. The aqueous residue was extracted with ethyl acetate (2 x 100 mL). The combined organic phases were washed with brine (30 mL), dried over Na₂SO₄ (s) and concentrated under reduced pressure. The residue was purified by column chromatography (silica gel, ethyl acetate / hexane, 1:2 to 1:1) to give 1,3-diol **5b** (1.15 g, 90%) as a colorless oil.

*R*_f 0.5 (ethyl acetate / hexane, 2:1); $[\alpha]_D^{20} = +16.8$ (*c* 1.10, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 5.21 (brs, 1H), 3.89 – 3.69 (m, 3H), 3.49 (d, *J* = 8.1 Hz, 1H), 2.66 (d, *J* = 3.8 Hz, 1H), 2.47 (t, *J* = 5.1 Hz, 1H), 1.81 – 1.69 (m, 1H), 1.45 (s, 9H), 1.00 (d, *J* = 6.7 Hz, 3H), 0.93 (d, *J* = 6.7 Hz, 3H) ; ¹³C NMR (75 MHz, CDCl₃) δ 156.4, 79.6, 77.5, 64.9, 52.0, 30.9, 28.3, 18.9, 18.8; HRMS (ESI): *m*/*z* calcd for C₁₁H₂₃NO₄Na⁺ [M+Na]⁺ 256.1519, found 256.1527.

1,3-Diol **5b** (1.50 g, 6.43 mmol) was dissolved in MeCN (100 mL) and aqueous solution of NaH₂PO₄-Na₂HPO₄ (60 mL, pH 6.7). To this solution, TEMPO (300.0 mg, 1.93 mmol), NaClO₂ (2.36 g, 19.30 mmol) and bleach solution (1.4 mL, 10% in water) were sequentially added. The reaction mixture was gently heated at 35 °C and stirred for 3 h (monitored by TLC). The reaction was then quenched at 0 °C by acidification to pH 1–2 with HCl (1M, aqueous solution). The aqueous solution was extracted with ethyl acetate (2 x 300 mL). The combined organic layers were washed with saturated aqueous solution of Na₂S₂O₃ (20 mL), brine (20 mL), dried over anhydrous Na₂SO₄ (s) and concentrated under reduced pressure to give Boc-*D*-(2*R*,3*S*)-threo-3-hydroxyleucine (HyLeu) **5c** (1.45 g), which was used in the next

^[1]Willams, L.; Zhang, Z.; Shao, F.; Carroll, P. J.; Joullit, M. M. Tetrahedron 1996, 52, 11673–11694.

step of reaction without further purification.



SOCl₂ (1.9 mL, 25.72 mmol) was added to a solution of acid **5c** (1.45 g) in MeOH (100 mL) at 0 °C. The reaction mixture was refluxed for 10 h before it was concentrated under reduced pressure. To a solution of the crude amine hydrochloride salt and Cbz-Gly-OH (1.70 g, 7.71 mmol) in CH₂Cl₂ (30 mL), DIPEA (4.5 mL, 25.72 mmol) and PyAOP (5.03 g, 9.65 mmol) were added sequentially at 0 °C. The reaction mixture was warmed to room temperature and stirred for 24 h. The reaction was quenched by addition of water (1 mL) and diluted with ethyl acetate (200 mL). The organic phase washed with saturated aqueous solution of NH₄Cl (20 mL) and brine (20 mL), dried over anhydrous Na₂SO₄ (s) and concentrated under reduced pressure. The residue was purified by column chromatography (silica gel, ethyl acetate / hexane, 1:1 to 2:1) to afford dipeptide **5d** (1.71 g, 73% 3 steps) as a colorless oil.

*R*_f 0.3 (ethyl acetate / hexane, 2:1); $[\alpha]_D^{20} = -2.0$ (*c* 0.87, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.40 - 7.29 (m, 5H), 7.03 (d, *J* = 9.1 Hz, 1H), 5.76 (brs, 1H), 5.12 (s, 2H), 4.81 (dd, *J* = 9.1, 1.9 Hz, 1H), 3.94 (brs, 2H), 3.73 (s, 4H), 3.08 (brs, 1H), 1.70 - 1.51 (m, 1H), 0.98 (d, *J* = 6.5 Hz, 3H), 0.91 (d, *J* = 6.3 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 172.2, 169.6, 156.6, 136.0, 128.4, 128.1, 128.0, 77.3, 67.1, 54.3, 52.6, 44.2, 30.8, 18.8; HRMS (ESI) *m/z* calcd for C₁₇H₂₅N₂O₆⁺[M+H]⁺ 353.1707, found 353.1704.



To a stirred solution of **5d** (88.0 mg, 0.25 mmol) in THF (5 mL), NaOH (2.5 mL, 1 M in water) was added at room temperature. The reaction mixture was stirred at room temperature for 4 h. Then it was acidified to pH 1–2 with HCl (1.0 M in water) and extracted with ethyl acetate (3 x 30 mL). The combined organic layers were washed with brine (15 mL), dried over anhydrous Na_2SO_4 (s) and concentrated under reduced pressure to afford the desired acid **5** (81.0 mg, 92%), which was used directly in the next step without further purification.

2.1.2 Synthesis of Ahda fragment and pentapeptide 2



DIBAL-H (85.0 mL, 85.00 mmol, 1.0 M in toluene) was added to a solution of ester **8** (20.00 g, 67.92 mmol) in CH₂Cl₂ (120 mL) at -78 °C. The reaction mixture was stirred at -78 °C for 5 h and quenched by addition of MeOH (2 mL) with careful. The reaction solution was then treated by saturated aqueous solution of Rochelle salt (150 mL) with vigorous stirring for 2 h. Layers were separated. The aqueous phase was extracted with CH₂Cl₂ (2 x 300 mL). The combined organic layers were successively washed with water (30 mL), saturated aqueous solution of NaHCO₃ (30 mL) and brine (20 mL), dried over anhydrous Na₂SO₄ (s) and concentrated under reduced pressure to produce the corresponding crude aldehyde **28**, which was subjected to next step of reaction without further purifications.

CBr₄ (39.8 g, 120.00 mmol) was slowly added to a solution of Ph₃P (62.94 g, 240.00 mmol) in CH₂Cl₂ (250 mL) at 0 $\$ The reaction mixture was then stirred at room temperature for 30 min. A solution of aldehyde **28** and DIPEA (15.7 mL, 90.00 mmol) in CH₂Cl₂ (100 mL) was added at 0 $\$ The reaction mixture was allowed to warm to room temperature and stirred for 12 h. The solution was poured into hexane (500 mL), the precipitation formed was removed by filtration. The filtrate was collected and the solid was dissolved in CH₂Cl₂ (250 mL), poured into hexanes (250 mL) and filtered again to remove the precipitation. The above procedures were repeated once with CH₂Cl₂ (125 mL)-hexane (125 mL). The combined filtrate was concentrated under reduced pressure to yield a yellow oil contaminated by solid precipitations. The residue was purified by column chromatography (silica gel, hexane) to give the dibromoalkene **29** (21.5 g, 76% in two steps) as a colorless oil.

 $R_{f} 0.6 \text{ (Hexane); } [\alpha]_{D}^{20} = +2.1 \text{ (} c 2.05, \text{ CHCl}_{3}\text{); }^{1}\text{H NMR} (300 \text{ MHz, CDCl}_{3}) \delta 7.35 - 7.17 \text{ (m, 5H), } 6.46 \text{ (d, } J = 7.9 \text{ Hz, 1H), } 4.46 \text{ (td, } J = 7.9, 4.6 \text{ Hz, 1H), } 2.90 - 2.70 \text{ (m, 2H), } 0.83 \text{ (s, 9H), } -0.08 \text{ (s, 3H), } -0.18 \text{ (s, } 3\text{H); }^{13}\text{C NMR} (75 \text{ MHz, CDCl}_{3}) \delta 141.6, 137.4, 130.0, 128.1, 126.4, 88.4, 75.0, 43.1, 25.7, 18.0, -4.9, -5.4; \text{HRMS (ESI) } m/z \text{ calcd for } C_{16}\text{H}_{24}\text{Br}_2\text{OSiNa}^+ \text{ [M+Na]}^+ 440.9855, \text{ found } 440.9837.$

ⁿBuLi (44.6 mL, 111.50 mmol, 2.5 M in hexane) was added dropwise to a solution of dibromoalkene **29** (21.20 g, 50.71 mmol) in THF (250 mL) at -78 °C. 1 h later, the reaction was quenched by addition of

saturated aqueous solution of NH_4Cl (5 mL). Volatiles were removed under reduced pressure, the residue was dissolved in ethyl acetate (500 mL). The solution was successively washed with H_2O (25 mL) and brine (25 mL), dried over anhydrous Na_2SO_4 (s) and concentrated under reduced pressure. The residue was purified by column chromatography (silica gel, ethyl acetate / hexane, 1:60 to 1:40) afforded **9** (11.87 g, 90%) as a colorless oil.

*R*_f 0.5 (hexane); $[\alpha]_D^{20} = -9.1$ (*c* 1.61, CHCl₃); ¹H NMR (CDCl₃, 300 MHz) δ 7.38-7.19 (m, 5H), 4.49 (dd, *J* = 7.7, 5.9, 2.1 Hz, 1H), 3.07 – 2.91 (m, 2H), 2.44 (d, *J* = 2.1 Hz, 1H), 0.86 (s, 9H), -0.01 (s, 3 H), -0.06 (s, 3 H); ¹³C NMR (CDCl₃, 75 MHz) δ 137.4, 129.9, 128.1, 126.6, 85.1, 72.7, 64.1, 45.1, 25.7, 18.2, -5.1, -5.3; HRMS (ESI) *m/z* calcd for C₁₆H₂₄OSiNa⁺ [M+Na]⁺ 283.1489, found: 283.1480.



To a solution of alkyne **9** (3.12 g, 12.00 mmol) in THF (50 mL) at -78 C, LiHMDS (24.0 mL, 24.00 mmol, 1.0 M in THF) was added dropwise and the reaction mixture was stirred for 1 h. BF₃ Et₂O (2.2 mL, 17.50 mmol) was added and stirred for 30 min at -78 C. A solution of epoxide **10** (2.00 g, 9.60 mmol) in THF (20 mL) was added slowly. The reaction was stirred for further 3 h at -78 C and quenched by addition of saturated aqueous solution of NH₄Cl (1 mL). Volatiles were removed under reduced pressure, the residue was diluted with water (15 mL) and extracted with ethyl acetate (3 x 100 mL). The organic layers were combined and washed with brine (15 mL), dried over anhydrous Na₂SO₄ (s) and concentrated under reduced pressure. The residue was purified by column chromatography (silica gel, ethyl acetate / hexane, 1:10 to 1:4) to afford compound **11** (4.27 g, 95% from **10**) as a colorless oil.

*R*_f 0.4 (ethyl acetate / hexane, 1:4); $[\alpha]_D^{20} = -24.2$ (*c* 1.22, CHCl₃);¹H NMR (CDCl₃, 500 MHz) δ 7.24 – 7.13 (m, 7H), 6.86 – 6.81 (m, 2H), 4.49 – 4.43 (m, 1H), 4.39 (s, 2H), 3.88 – 3.80 (m, 1H), 3.73 (s, 3H), 3.61 (ddd, *J* = 9.3, 6.3, 4.7 Hz, 1H), 3.53 (ddd, *J* = 9.4, 7.6, 4.5 Hz, 1H), 2.95 – 2.86 (m, 3H), 2.40 – 2.27 (m, 2H), 1.79 (dddd, *J* = 14.2, 6.4, 4.5, 3.2 Hz, 1H), 1.70 (dddd, *J* = 14.4, 8.5, 7.6, 4.7 Hz, 1H), 0.81 (s, 9H), -0.07 (s, 3H), -0.09 (s, 3H) ; ¹³C NMR (CDCl₃, 150 MHz) δ 159.2, 137.7, 130.0, 129.7, 129.1, 127.9, 126.3, 113.7, 83.6, 81.4, 72.8, 69.3, 68.0, 64.3, 55.1, 45.3, 35.2, 27.3, 25.6, 18.1, -5.1, -5.3; HRMS (ESI) *m/z* calcd for C₂₈H₄₀O₄SiNa⁺ [M+Na]⁺ 491.2588, found 491.2595.



To a solution of **11** (8.99 g, 19.18 mmol) in ethyl acetate (200 mL), palladium on charcoal (1 g, 10% Pd) and *N*,*N*-diisopropylethylamine (5.0 mL, 28.70 mmol) were added. The reaction vessel was sealed and the inner atmosphere was switched to H₂ and stirred for 48 h at room temperature. The catalyst was removed by filtration through a pad of Celite and washed with ethyl acetate (3 x 50 mL). The filtrate was concentrated under reduced pressure. The residue was purified by column chromatography (silica gel, ethyl acetate / hexane, 1:4) to afford product **30** (8.88 g, 98%) as a colorless oil.

*R*_f 0.4 (ethyl acetate / hexane, 1:4); $[\alpha]_D^{20} = -10.3$ (*c* 1.54, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 7.22 – 7.15 (m, 4H), 7.13 – 7.07 (m, 3H), 6.83 – 6.78 (m, 2H), 4.38 (s, 2H), 3.77 (dq, *J* = 7.2, 5.6 Hz, 1H), 3.73 (s, 3H), 3.71 (dd, *J* = 4.5, 2.7 Hz, 1H), 3.62 (dt, *J* = 9.3, 5.2 Hz, 1H), 2.80 (d, *J* = 3.2, 1H), 2.71 – 2.59 (m, 2H), 1.69 – 1.62 (m, 2H), 1.52 – 1.26 (m, 6H), 0.78 (s, 9H), -0.12 (s, 3H), -0.27 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 159.3, 139.5, 130.1, 129.8, 129.3, 128.1, 126.0, 113.9, 73.7, 73.0, 71.4, 69.0, 55.3, 43.8, 37.6, 37.0, 36.4, 25.9, 21.2, 18.1, -4.7, -4.9; HRMS (ESI) *m*/*z* calcd for C₂₈H₄₄NaO₄Si⁺ [M+Na]⁺ 495.2901, found 495.2887.

Alcohol **30** (10.01 g, 21.15 mmol) in THF (50 mL) was added to NaH (2.50 g, 62.50 mmol, 60 wt% in mineral oil) suspended in THF (200 mL) at 0 $^{\circ}$ C. 20 min later, BnBr (3.8 mL, 31.99 mmol) was added slowly. The reaction mixture was allowed to warm to 45 $^{\circ}$ C and stirred for 12 h. The reaction was quenched with saturated aqueous solution of NH₄Cl (1 mL) at 0 $^{\circ}$ C. Volatiles were removed under reduced pressure, the residue was diluted with water (20 mL) and extracted with ethyl acetate (2 x 200 mL). The combined organic phases were washed successively with water (15 mL), saturated aqueous solution of NH₄Cl (15 mL) and brine (15 mL), dried over anhydrous NaSO₄ (s) and concentrated reduced pressure. The residue was purified by column chromatography (silica gel, ethyl acetate / hexane, 1:50 to 1:10) to give product **12** (10.71 g, 90%) as a colorless oil.

 $R_{\rm f}$ 0.6 (ethyl acetate / hexane, 1:10); $[\alpha]_{\rm D}^{20} = +7.4$ (c 1.62, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 7.34 – 7.17 (m, 9H), 7.17 – 7.08 (m, 3H), 6.91 – 6.79 (m, 2H), 4.45 (d, J = 11.5 Hz, 1H), 4.39 (d, J = 11.5 Hz, 1H), 4.38 (d, J = 11.5 Hz, 1H), 4.35 (d, J = 11.5 Hz, 1H), 3.82 – 3.76 (m, 1H), 3.74 (s, 3H), 3.56 – 3.45 (m, 3H), 2.67 (d, J = 6.3 Hz, 2H), 1.84 – 1.71 (m, 2H), 1.54 – 1.28 (m, 6H), 0.81 (s, 9H), -0.09 (s, 3H), -0.23 (s, 3H);

¹³C NMR (125 MHz, CDCl₃) δ 159.2, 139.4, 139.0, 130.7, 129.8, 129.3, 128.3, 128.1, 127.8, 127.4, 126.0, 113.8, 76.4, 73.8, 72.7, 71.3, 66.8, 55.3, 44.0, 37.2, 34.5, 34.5, 25.9, 21.1, 18.1, -4.6, -4.8; HRMS (ESI) *m/z* calcd for C₃₅H₅₀O₄SiNa⁺ [M+Na]⁺ 585.3371, found 585.3354.



DDQ (3.16 g, 13.92 mmol) was added to a solution of compound **12** (6.02 g, 10.70 mmol) in CH₂Cl₂-H₂O (180 mL, 8/1, v/v) at 0 °C. 10 min later, the reaction mixture was brought to ambient temperature and stirred for 3 h. The reaction mixture was diluted with CH₂Cl₂ (500 mL) and washed with a 1:1 mixture of saturated aqueous solution of NaHCO₃ and saturated aqueous solution of Na₂S₂O₃ (3 x 100 mL) and brine (20 mL), dried over anhydrous Na₂SO₄. The organic layer was concentrated under reduced pressure. The residue was purified by column chromatography (silica gel, ethyl acetate / hexane, 1:8 to 1:4) to yield alcohol **31** (4.07 g, 86%) as a colorless oil.

*R*_f 0.4 (ethyl acetate / hexane, 1:4); $[\alpha]_D^{20} = +19.6$ (*c* 1.51, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 7.37 – 7.30 (m, 4H), 7.30 – 7.23 (m, 3H), 7.20 – 7.13 (m, 3H), 4.56 (d, *J* = 11.4 Hz, 1H), 4.46 (d, *J* = 11.5 Hz, 1H), 3.88 – 3.74 (m, 2H), 3.75 – 3.67 (m, 1H), 3.66 – 3.57 (m, 1H), 2.78 – 2.65 (m, 2H), 2.38 (brs, 1H), 1.86 – 1.66 (m, 2H), 1.66 – 1.55 (m, 1H), 1.55 – 1.30 (m, 5H), 0.85 (s, 9H), -0.04 (s, 3H), -0.17 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 139.3, 138.4, 129.7, 128.5, 128.1, 127.9, 127.7, 126.0, 78.5, 73.7, 71.0, 60.7, 44.0, 37.0, 36.0, 33.7, 25.9, 20.9, 18.1, -4.6, -4.8; HRMS (ESI) *m/z* calcd for C₂₇H₄₂O₃SiNa⁺ [M+Na]⁺ 465.2795, found 465.2786.

Trichloroisocyanuric acid (2.02 g, 8.70 mmol) and TEMPO (31.0 mg, 0.20 mmol) were subsequently added to a solution of alcohol **31** (3.19 g, 7.20 mmol) in CH_2Cl_2 (70 mL) at 0°C. The reaction mixture was stirred at 0 °C for 30 min before it was filtered through a pad of Celite and washed with CH_2Cl_2 (2 x 100 mL). The combined filtrate was washed successively with saturated aqueous solution of NaHCO₃ (30 mL) and brine (30 mL), dried over anhydrous Na₂SO₄(s) and concentrated under reduced pressure to give the

crude aldehyde 32 (ca. 3.06 g), which was used directly in the next step without further purification.

To a solution of phosphate **13** (3.62 g, 10.90 mmol) and 18-crown-6 (3.82 g, 14.50 mmol) in THF (100 mL) at -78 °C, KHMDS (21.8 mL, 10.90 mmol, 0.5 M) was added. 30 min later, a solution of the above aldehyde **32** in THF (50 mL) was slowly added via a syringe. The reaction mixture was stirred for 2 h at -78 °C, then it was carefully quenched by addition of saturated aqueous solution of NH₄Cl (2 mL) and allowed to warm to room temperature. Volatiles were removed under reduced pressure. The residue was dissolved in ethyl acetate (500 mL). The organic solution was washed successively with water (15 mL), saturated aqueous solution of NH₄Cl (15 mL) and brine (15 mL), dried over anhydrous NaSO₄ (s) and concentrated under reduced pressure. The residue was purified by column chromatography (silica gel, ethyl acetate / hexane, 1:10) to give the desired product **14** (2.68 g, 73% 2 steps) as a colorless oil and the undesired *E* configuration byproduct (ca. 0.2g).

*R*_f 0.7 (ethyl acetate / hexane, 1:10); $[\alpha]_D^{20} = -10.7$ (*c* 0.57, CHCl₃); ¹H NMR (CDCl₃, 300 MHz) δ7.43 – 7.26 (m, 7H), 7.26 – 7.15 (m, 3H), 6.39 (dt, *J* = 11.6, 7.3 Hz, 1H), 5.89 (dt, *J* = 11.6, 1.7 Hz, 1H), 4.59 (d, *J* = 11.6 Hz, 1H), 4.52 (d, *J* = 11.6 Hz, 1H), 4.20 (q, *J* = 7.1 Hz, 2H), 3.92 – 3.78 (m, 1H), 3.64 – 3.49 (m, 1H), 3.11 – 2.92 (m, 2H), 2.74 (d, *J* = 6.3 Hz, 2H), 1.70 – 1.36 (m, 6H), 1.32 (t, *J* = 7.1 Hz, 3H), 0.89 (s, 9H), -0.01 (s, 3H), -0.16 (s, 3H); ¹³C NMR (CDCl₃, 75 MHz) δ 166.3, 146.2, 139.3, 138.6, 129.7, 128.3, 128.0, 127.7, 127.5, 125.9, 121.1, 78.1, 73.7, 70.8, 59.8, 43.9, 37.0, 34.3, 32.9, 25.9, 21.3, 18.0, 14.2, -4.8, -4.9; HRMS (ESI) *m/z* calcd for C₃₁H₄₆O₄SiNa⁺ [M+Na]⁺ 533.3058, found 533.3042.



DIBAL-H (27.5 mL, 27.50 mmol, 1.0 M in toluene) was added slowly to a solution of ester **14** (4.01 g, 7.85 mmol) in THF (50 mL) at -78 °C. The reaction mixture was then warmed to -36 °C and stirred for 4 h, it was quenched by careful addition of methanol (1 mL). The reaction mixture was stirred for 30 min, then it was poured into saturated aqueous solution of Rochelle salt (50 mL). The mixture was virgously stirred at room temperature for 2 h. Volatiles were removed under reduced pressure. The aqueous residue was extracted with ethyl acetate (2 x 200 mL). The combined organic extracts were washed with saturated aqueous solution of NH₄Cl (30 mL) and brine (30 mL), dried over anhydrous Na₂SO₄ (s) and concentrated

under reduced pressure. The residue was purified by column chromatography (silica gel, ethyl acetate / hexane, 1:10 to 1:5) gave alcohol **33** (3.38 g, 92%) as a colorless oil.

*R*_f 0.3 (ethyl acetate / hexanes, 1:5); $[\alpha]_D^{20} = -9.3 (c \ 1.00, CHCl_3)$; ¹H NMR (CDCl₃, 500 MHz) δ 7.31 – 7.25 (m, 4H), 7.25 – 7.17 (m, 3H), 7.16 – 7.09 (m, 3H), 5.74 – 5.70 (m, 1H), 5.58 – 5.52 (m, 1H), 4.45 (s, 2H), 4.11 – 4.01 (m, 2H), 3.78 (dddd, *J* = 5.8, 5.8, 5.8, 5.8 Hz, 1H), 3.36 (dddd, *J* = 5.4, 5.4, 5.4, 5.4 Hz, 1H), 2.70 – 2.63 (m, 2H), 2.35 – 2.30 (m, 1H), 2.26 – 2.20 (m, 1H), 1.97 (brs, 1H), 1.20 – 1.53 (m, 6H), 0.82 (s, 9H), -0.08 (s, 3H), -0.21 (s, 3H); ¹³C NMR (CDCl₃, 125 MHz) δ 139.3, 138.4, 130.9, 129.8, 128.9, 128.4, 128.2, 127.9, 127.7, 126.1, 78.4, 73.7, 71.4, 58.2, 44.0, 37.0, 34.1, 32.0, 25.9, 21.2, 18.1, -4.6, -4.8; HRMS (ESI) *m*/*z* calcd for C₂₉H₄₄O₃SiNa⁺ [M+Na]⁺ 491.2952, found: 491.2941.

D-(-)-diisopropyl tartrate (1.8 mL, 8.65 mmol) was added to a solution of Ti(O'Pr)₄ (2.1 mL, 7.73 mmol) and 4Å molecular sieves (3.00 g) in CH₂Cl₂ (20 mL) at -20 °C. 30 min later, *tert*-butyl hydroperoxide (3.7 mL, ~5.5 M in decane) was dropwise added at -20 °C. After being stirred for another 20 min, a solution of allylic alcohol **33** (3.02 g, 6.44 mmol) in CH₂Cl₂ (80 mL) was dropwise added to the solution via a syringe. The reaction mixture was then stirred slowly for 24 h at -20 °C and maintained in a refrigerator at -20 °C for 6 days. The reaction was quenched by addition of an aqueous solution of 3 M NaOH (*aq.*) -brine (100 mL, 3/7, v/v). The reaction mixture was allowed to warm to room temperature, diluted with CH₂Cl₂ (500 mL) and stirred for another 1.5 h. The reaction mixture was then filtered through a pad of Celite, the filtration residue was eluted with CH₂Cl₂ (100 mL). The filtrate was partitioned and layers were seperated. The organic phase was washed with brine (30 mL), dried over anhydrous Na₂SO₄ (s) and concentrated under reduced pressure. The residue was purified by column chromatography (silica gel, ethyl acetate / CH₂Cl₂, 1:10) to afford the desired epoxide **15** (2.68 g, 86%, *dr* = 12:1) as a colorless oil.

*R*_f 0.6 (ethyl acetate / CH₂Cl₂, 1:10) ; $[\alpha]_D^{20} = +0.3$ (*c* 0.63, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 7.35 – 7.24 (m, 7H), 7.20 – 7.14 (m, 3H), 4.53 (d, *J* = 11.6 Hz, 1H), 4.49 (d, *J* = 11.6 Hz, 1H), 3.82 (dddd, *J* = 6.0, 6.0, 6.0, 11.6 Hz, 1H), 3.74 – 3.69 (m, 1H), 3.64 – 3.59 (m, 1H), 3.56 (dddd, *J* = 5.5, 5.5, 5.5, 11.0 Hz, 1H), 3.25-3.09 (m, 2H), 2.76 – 2.65 (m, 2H), 2.36 (dt, *J* = 8.1, 4.4 Hz, 1H), 1.93 (ddd, *J* = 5.5, 5.5, 16.6 Hz, 1 H), 1.73 (ddd, *J* = 5.5, 7.6, 16.6 Hz, 1H), 1.69 – 1.28 (m, 6H), 0.85 (s, 9H),-0.04 (s, 3H), -0.16 (s, 3H); ¹³C NMR (CDCl₃, 125 MHz) δ 139.2, 138.0, 129.7, 128.5, 128.2, 128.0, 127.9, 126.1, 76.8, 73.6, 71.3, 60.5, 55.9, 53.6, 44.1, 36.9, 33.5, 31.6, 25.9, 21.5, 18.1, -4.6, -4.8; HRMS (ESI) *m*/*z* calcd forC₂₉H₄₄O₄SiNa⁺ [M+Na]⁺ 507.2901, found 507.2899.



 NH_4Cl (s) (1.00 g, 18.58 mmol) and NaN_3 (s) (2.01 g, 31.0 mmol) were sequentially added to a solution of **15** (1.50 g, 3.10 mmol) in MeOH-H₂O (27 mL, 8/1, v/v) at room temperature. The reaction mixture was refluxed for 24 h. After it was cooled to room temperature, the reaction mixture was concentrated under reduced pressure. The residue was dissolved in ethyl acetate (200 mL) and then washed with brine (20 mL), dried over anhydrous Na_2SO_4 (s) and concentrated under reduced pressure. The residue was purified by column chromatography (silica gel, ethyl acetate / CH_2Cl_2 : 1:4) to give the required product **17** (1.10 g, 67%) along with its regio-isomer **16** (0.26 g, 16%) as colorless oils.

Analytical data for 1,3-diol 16:

*R*_f 0.7 (ethyl acetate / CH₂Cl₂, 1:4); $[\alpha]_D^{20} = +16.6$ (*c* 0.50, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 7.41 – 7.24 (m, 7H), 7.24 – 7.12 (m, 3H), 4.64 (d, J = 11.2 Hz, 1H), 4.40 (d, J = 11.2 Hz, 1H), 4.05 – 3.97 (m, 1H), 3.94 – 3.79 (m, 4H), 3.76 – 3.65 (m, 1H), 3.32 – 3.24 (m, 1H), 2.80 – 2.69 (m, 2H), 2.37 (t, J = 5.4 Hz, 1H), 1.87 (dt, J = 14.6, 9.8 Hz, 1H), 1.72 – 1.53 (m, 3H), 1.53 – 1.35 (m, 4H), 0.88 (s, 9H), -0.01 (s, 3H), -0.13 (s, 3H). ¹³C NMR (CDCl₃, 125 MHz) δ 139.2, 137.7, 129.7, 128.6, 128.2, 128.0, 127.9, 126.1, 79.4, 73.6, 72.8, 70.7, 66.1, 63.2, 44.1, 37.4, 37.0, 33.6, 25.9, 20.3, 18.1, -4.6, -4.7; HRMS (ESI): *m/z* calcd for C₂₉H₄₆N₃O₄Si⁺ [M+H]⁺ 528.3252, found 528.3259.

Analytical data for 1,2-diol 17:

*R*_f 0.5 (ethyl acetate / CH₂Cl₂, 1:4); $[\alpha]_D^{20}$ = +36.8 (*c* 0.67, CHCl₃); ¹H NMR (CDCl₃, 500 MHz) δ 7.25-7.36 (m, 7 H), 7.15-7.25 (m, 3 H), 4.60 (d, *J* = 11.1 Hz, 1 H), 4.40 (d, *J* = 11.1 Hz, 1 H), 3.83-3.85 (m, 1 H), 3.60-3.66 (m, 4 H), 3.53-3.58 (m, 1 H), 2.68-2.76 (m, 2 H), 2.21 (brs, 2 H), 1.69-1.79 (m, 2 H), 1.66-1.68 (m, 1 H), 1.64-1.65 (m, 1 H), 1.36-1.55 (m, 4 H), 0.86 (s, 9H), -0.03 (s, 3 H), -0.16 (s, 3 H); ¹³C NMR (CDCl₃, 125 MHz) δ 139.2, 138.3, 129.7, 128.5, 128.1, 127.9, 127.8, 126.0, 75.9, 74.5, 73.6, 71.2, 64.0, 61.1, 44.0, 37.0, 36.0, 33.9, 25.9, 20.5, 18.1, -4.7, -4.8; HRMS (ESI) *m*/*z* calcd for C₂₉H₄₅N₃O₄SiNa⁺ [M+Na]⁺ 550.3072, found 550.3061.



1,2-diol **17** (200.0 mg, 0.38 mmol) was dissolved in saturated aqueous solution of Na₂HPO₄-MeCN (7 mL, 1/1, v/v), then the solution was adjusted to pH 8–9 with NaOH (1 M aqueous solution). TEMPO (3.0 mg, 0.019 mmol), NaClO₂ (91.0 mg, 1.10 mmol) and NaBr (103.0 mg, 1.00 mmol) were sequentially added to the solution at room temperature. A solution of bleach (40 µL, 10% in water) was added to the reaction mixture every 3 h until all the starting material was consumed (monitored by TLC). After being acidified to pH 1–2 with HCl (1.0 M aqueous solution) at 0 °C, the reaction mixture was extracted with ethyl acetate (2 x 50 mL). The combined organic layers were washed with brine (10 mL), dried over anhydrous Na₂SO₄ (s) and concentrated under reduced pressure. The residue contained an inseparable mixture of carboxylic acids **18** and **34** (*ca.* 185.0 mg in total), which was directly subjected to next step of reaction.

To a solution of the carboxylic acids obtained above and amine **19** (135.0 mg, 0.49 mmol) in CH₂Cl₂ (5 mL), DIPEA (260 μ L, 1.52 mmol) and PyAOP (297.2 mg, 0.57 mmol) were added sequentially at 0 °C. The reaction mixture was warmed to room temperature and stirred for 12 h. The reaction was then quenched with H₂O (1 mL) and diluted with ethyl acetate (100 mL). The organic solution was washed sequentially with saturated aqueous solution of NH₄Cl (10 mL) and brine (10 mL), dried over anhydrous Na₂SO₄ and concentrated under reduced pressure. The residue was purified by column chromatography (silica gel, ethyl acetate / hexane, 1:8 to 1:4) to afford the desired dipeptide **20** (227.6 mg, 75%) and **35** (35.1 mg, 12%) as colorless oils.

Analytical data for compound **20**:

*R*_f 0.4 (ethyl acetate / hexane, 1:4); $[α]_D^{20} = -2.3$ (*c* 1.03, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 7.39 - 7.26 (m, 7H), 7.22 - 7.17 (m, 3H), 4.65 - 4.54 (m, 2H), 4.40 (d, *J* = 11.1 Hz, 1H), 4.14 (s, 1H), 4.12 - 4.04 (m, 2H), 3.93 - 3.79 (m, 2H), 3.60 (d, *J* = 4.3 Hz, 1H), 3.33 (brs, 1H), 2.83 - 2.68 (m, 2H), 1.95 - 1.79 (m, 2H), 1.62 - 1.63 (m, 2H), 1.49 (s, 14H), 0.88 (s, 9H), 0.88 (s, 9H), 0.05 (s, 3H), 0.04 (s, 3H), -0.01 (s, 3H), -0.15 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 170.5, 168.9, 139.2, 138.0, 129.7, 128.5, 128.1, 127.9, 127.8,

126.1, 82.2, 76.0, 73.6(2), 71.0, 63.6, 61.1, 54.7, 44.0, 37.1, 35.4, 33.7, 28.0, 25.9, 25.7, 20.4, 18.1, 18.1, -4.7, -4.8, -5.6; **HRMS (ESI)** *m*/*z* calcd for C₄₂H₇₀N₄O₇Si₂Na⁺ [M+Na]⁺ 821.4675, found 821.4709. Analytical data for compound **35**:

*R*_f 0.6 (ethyl acetate / hexane, 1:4); ¹H NMR (300MHz, CDCl₃) δ 7.41 – 7.15 (m, 10H), 7.11 (d, *J* = 10.3 Hz, 1H), 4.64 (d, *J* = 11.3 Hz, 1H), 4.51 (dt, *J* = 8.1, 2.7 Hz, 1H), 4.44 (d, *J* = 11.3 Hz, 1H), 4.21 (dd, *J* = 10.8, 3.0 Hz, 1H), 4.06 (dd, *J* = 10.0, 2.5 Hz, 1H), 3.91 – 3.84 (m, 1H), 3.81 (dd, *J* = 10.0, 3.0 Hz, 1H), 3.73 – 3.60 (m, 1H), 2.74 (d, *J* = 6.3 Hz, 2H), 2.27 – 2.17 (m, 1H), 1.86 – 1.55 (m, 3H), 1.50 (s, 13H), 0.91 (s, 9H), 0.88 (s, 9H), 0.065 (s, 3H), 0.061 (s, 3H), -0.02 (s, 3H), -0.16 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 169.5, 168.8, 139.2, 138.3, 129.7, 128.3, 128.1, 127.7, 127.6, 126.0, 82.2, 75.0, 73.5, 70.7, 63.4, 61.3, 54.6, 43.8, 37.4, 37.0, 33.6, 27.9, 25.8, 25.6, 20.4, 18.1, 18.0, -4.8, -4.9, -5.6; HRMS (ESI) *m*/*z* calcd for $C_{41}H_{68}N_4O_6Si_2Na^+$ [M+Na]⁺ 791.4570, found 791.4583.



To a solution of azide **20** (150.0 mg, 0.19 mmol) in ethyl acetate (5 mL), DIPEA (40 μ L, 0.23 mmol), palladium on charcoal (20.0 mg, 10% Pd) was added at room temperature. The reaction vessel was sealed and the inner atmosphere was switched to H₂ and stirred for 24 h at room temperature. The catalyst was removed by filtration through a pad of Celite and washed with ethyl acetate (3 x 30 mL). The filtrate was concentrated under reduced pressure to produce the corresponding amine (*ca.* 137.0 mg), which was used in next step of reaction without further purifications.

To a solution of amine (derived from **20** as indicated above) and carboxylic acid **36** (89.0 mg, derived from sponification of 100 mg correspoding methyl ester) in 5 mL CH₂Cl₂, DIPEA (130 μ L, 0.76 mmol) and PyAOP (188.0 mg, 0.36 mmol) were sequentially added at 0 °C. The reaction mixture was warmed to room

temperature and stirred for 12 h, then it was quenched addition of water (1 mL) and diluted with ethyl acetate (100 mL). The organic solution was washed with saturated aqueous solution of NH_4Cl (10 mL) and brine (10 mL), dried over anhydrous Na_2SO_4 (s) and concentrated under reduced pressure. The residue was purified by column chromatography (silica gel, ethyl acetate / hexane, 1:4 to 1:2) to give dipeptide **37** (145.1 mg, 75% from **20**) as a colorless oil.

*R*_f 0.5 (ethyl acetate / hexane, 1:2); $[\alpha]_D^{20} = -14.5$ (*c* 1.00, CHCl₃); ¹H NMR (300 MHz, CDCl₃) exists as rotational conformers: δ 7.66 – 7.52 (m, 1H), 7.40 – 7.12 (m, 15H), 6.88 – 6.62 (m, 1H), 5.54 – 5.41 (m, 0.6H), 5.21 – 5.01 (m, 2H), 4.97 – 4.85 (m, 0.4H), 4.59 – 4.39 (m, 2H), 4.30 – 4.11 (m, 2H), 4.09 – 3.97 (m, 1H), 3.87 – 3.72 (m, 2H), 3.70 – 3.41 (m, 4H), 2.77 – 2.62 (m, 2H), 2.47 – 2.26 (m, 1H), 2.13 – 1.92 (m, 2H), 1.84 – 1.72 (m, 1H), 1.61 – 1.31 (m, 17H), 1.08 – 0.99 (m, 3H), 0.87 (s, 9H), 0.84 (s, 9H), 0.03 (s, 6H), -0.06 (s, 3H), -0.22 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) exists as rotational conformers: δ 173.2, 172.9, 172.0, 171.8, 168.6, 155.1, 154.8, 139.3, 138.4, 137.9, 136.4, 136.4, 129.7, 128.4, 128.4, 128.3, 128.0, 127.9, 127.6, 125.9, 81.8, 81.6, 76.1, 74.5, 73.6, 71.2, 67.6, 67.0, 63.5, 54.3, 54.1, 51.4, 50.9, 46.3, 45.9, 43.7, 39.6, 37.5, 37.3, 35.3, 34.6, 33.9, 33.5, 32.4, 31.6, 28.0, 25.9, 25.7, 20.3, 19.9, 18.6, 18.3, 18.1, 18.0, -4.8, -5.0, -5.7; HRMS (ESI) *m/z* calcd for C₅₆H₈₈N₃O₁₀Si₂⁺ [M+H]⁺ 1018.6003, found 1018.5999.

2,6-Lutidine (210.0 μ L, 1.80 mmol) was added to a stirred solution of **37** (190.0 mg, 0.18 mmol) in CH₂Cl₂ (5 mL) at -78 °C, followed by dropwise addition of TESOTF (210.0 μ L, 0.94 mmol) at -78 °C. The reaction mixture was stirred at -78 °C for 4 h before it was quenched with water (0.5 mL) and diluted with ethyl acetate (150 mL). The organic solution was then washed successively with citric acid (3 x 10 mL, 10wt% in water), saturated aqueous solution of NaHCO₃ (2 x 10 mL) and brine (10 mL), dried over anhydrous Na₂SO₄ (s) and concentrated under reduced pressure.The residue was purified by column chromatography (silica gel, ethyl acetate / hexane, 1:8 to 1:2) to obtain **21** (192.6 mg, 94%) as a colorless oil.

*R*_f 0.7 (ethyl acetate / hexane, 1:2); $[α]_D^{20} = -22.5$ (*c* 1.05, CHCl₃); ¹H NMR (300 MHz, CDCl₃) exists as rotational conformers: δ 7.62 – 7.51 (m, 1H), 7.44 – 7.13 (m, 15H), 7.03 – 6.96 (m, 0.5H), 6.92 – 6.85 (m, 0.5H), 5.26 (d, *J* = 12.9 Hz, 0.5H), 5.14 (d, *J* = 5.9 Hz, 1H), 4.98 (d, *J* = 13.0 Hz, 0.5H), 4.59 – 4.39 (m, 3H), 4.37 – 4.21 (m, 1.5H), 4.15 – 4.04 (m, 1.5H), 3.85 (d, *J* = 5.0 Hz, 1H), 3.82 – 3.55 (m, 4H), 3.46 – 3.25 (m, 1H), 2.77 – 2.59 (m, 2H), 2.52 – 2.42 (m, 1H), 2.16 – 2.05 (m, 1H), 1.96 – 1.83 (m, 1H), 1.59 – 1.24 (m, 17H), 1.19 – 1.09 (m, 3H), 0.98 – 0.80 (m, 27H), 0.73 – 0.55 (m, 6H), 0.04 (s, 6H), -0.09 (s, 3H),

-0.24 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) exists as rotational conformers: δ 172.2, 172.0, 171.5, 171.2, 168.5, 168.4, 155.0, 139.3, 139.0, 138.8, 136.7, 136.3, 129.7, 128.3, 128.2, 128.0, 127.7, 127.5, 127.1, 125.8, 81.8, 76.1, 76.0, 73.7, 73.2, 73.0, 72.2, 72.0, 68.3, 66.9, 63.6, 54.0, 49.1, 46.4, 45.9, 43.7, 39.6, 38.1, 35.1, 34.8, 34.0, 33.6, 32.1, 31.4, 28.0, 25.8, 25.7, 21.1, 20.9, 18.8, 18.5, 18.1, 18.0, 6.6, -4.8, -5.1, -5.7; HRMS (ESI) *m*/*z* calcd for C₆₂H₁₀₂N₃O₁₀Si₃⁺ [M+H]⁺ 1132.6868, found 1132.6877.

To a solution of tripeptide **21** (192.6 mg, 0.17 mmol) in CH_2Cl_2 (7 mL, A.R. grade, $H_2O < 0.5$ wt%), DDQ (115.8 mg, 0.51 mmol) was added at room temperature. The reaction mixture was stirred for 10 h. Volatiles were removed under reduced pressure, the residue was purified by column chromatography (silica gel, ethyl acetate / hexane, 1:4 to 1:2) to give **38** (134.7 mg, 76%) as a colorless oil.

*R*_f 0.6 (EA / Hex, 1:2); $[\alpha]_D^{20} = -6.9$ (*c* 2.50, CHCl₃); ¹H NMR (300 MHz, CDCl₃) exists as rotational conformers: δ 7.71 – 7.53 (m, 2H), 7.40 – 7.08 (m, 10H), 5.21 – 5.05 (m, 2H), 4.51 (d, *J* = 8.3 Hz, 1H), 4.40 – 4.02 (m, 4H), 3.87 (d, *J* = 3.1 Hz, 1H), 3.78 (dd, *J* = 10.0, 2.7 Hz, 2H), 3.68 – 3.54 (m, 2H), 3.52 – 3.22 (m, 1H), 2.80 – 2.54 (m, 2H), 2.51 – 2.39 (m, 1H), 2.12 – 1.98 (m, 1H), 1.73 – 1.31 (m, 16H), 1.21 – 1.09 (m, 4H), 1.01 – 0.94 (m, 9H), 0.89 (s, 9H), 0.81 (s, 9H), 0.67 (dq, *J* = 15.6, 7.7 Hz, 6H), 0.05 (s, 6H), -0.12 (s, 3H), -0.28 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) exists as rotational conformers: δ 173.13, 172.86, 172.15, 172.08, 168.34, 168.28, 155.03, 154.83, 139.54, 136.60, 136.34, 129.75, 128.33, 127.88, 127.77, 127.67, 127.15, 125.75, 82.08, 73.78, 72.80, 72.72, 67.89, 67.81, 66.95, 66.89, 66.66, 63.54, 54.13, 49.46, 49.34, 46.32, 45.89, 43.51, 39.76, 38.36, 37.35, 37.06, 32.26, 31.49, 27.97, 25.80, 25.71, 21.93, 18.87, 18.56, 18.13, 17.95, 6.64, 4.46, -4.84, -5.18, -5.72; HRMS (ESI) *m*/*z* calcd for C₅₅H₉₆N₃O₁₀Si₃⁺ [M+H]⁺ 1042.6398, found 1042.6396.



To a solution of alcohol **38** (134.7 mg, 0.13 mmol) in dry toluene (10 mL) *N*-methylmorpholine (57 μ L, 5.20 mmol) and DMAP (79.4 mg, 0.65 mmol) were added at 0 °C, followed by dropwise addition of **22** (428.7 mg, 1.3 mmol) in dry toluene (10 mL). After being stirred for 0.5 h at 0 °C, the reaction mixture was allowed to warm to room temperature and stirred for another 4 h. The reaction was quenched by addition of

 H_2O (1 mL) and diluted with ethyl acetate (150 mL). The organic solution was washed successively with saturated aqueous solution of NaHCO₃ (10 mL) and brine (10 mL), dried over anhydrous Na₂SO₄(s) and concentrated under reduced pressure. The residue was purified by column chromatography (silica gel, ethyl acetate / hexane, 1:4 to 1:2) to provide **23** (149.4 mg, 86%) as a colorless oil.

*R*_f 0.6 (ethyl acetate / hexane, 1:2); $[\alpha]_D^{20} = -2.2$ (*c* 1.00, CHCl₃); ¹H NMR (300 MHz, CDCl₃) exists as rotational conformers: δ 7.77 (d, *J* = 7.4 Hz, 2H), 7.66 – 7.52 (m, 3H), 7.44 – 7.10 (m, 15H), 7.07 – 6.86 (m, 1H), 5.78 (d, *J* = 6.7 Hz, 0.5H), 5.60 (d, *J* = 7.3 Hz, 0.5H), 5.36 – 5.21 (m, 1H), 5.20 – 4.81 (m, 2H), 4.56 – 3.97 (m, 8H), 3.85 – 3.52 (m, 5H), 2.77 – 2.60 (m, 2H), 2.53 – 2.40 (m, 1H), 2.17 – 2.00 (m, 2H), 1.63 – 1.33 (m, 19H), 1.14 (d, *J* = 6.8 Hz, 3H), 1.05 – 0.93 (m, 9H), 0.90 (s, 9H), 0.83 (s, 9H), 0.77 – 0.60 (m, 6H), 0.06 (s, 6H), -0.08 (s, 3H), -0.23 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) exists as rotational conformers: δ 172.1, 171.6, 168.3, 155.7, 154.9, 144.0, 143.8, 141.2, 139.2, 136.8, 129.7, 128.3, 128.0, 127.5, 127.3, 127.0, 125.9, 125.1, 119.8, 82.0, 73.5, 73.0, 71.5, 71.3, 68.3, 68.1, 66.8, 66.7, 63.4, 54.0, 49.9, 48.5, 48.3, 47.1, 46.4, 46.1, 43.8, 39.4, 37.8, 37.0, 35.0, 32.4, 31.5, 28.0, 25.8, 25.7, 20.9, 18.8, 18.1, 17.9, 6.7, 4.4, -4.8, -5.1, -5.7, -5.7; HRMS (ESI) *m*/*z* calcd for C₇₃H₁₁₁N₄O₁₃Si₃⁺ [M+H]⁺ 1335.7450, found 1335.7440.



DEA (0.4 mL) was added to a solution of compound **23** (149.3 mg, 0.11 mmol) in MeCN (5 mL) at room temperature. The reaction mixture was stirred at room temperature for 8 h and then concentrated under reduced pressure. The residue was re-dissolved in CH_2Cl_2 (5 mL) and concentrated under reduced pressure. The crude amine was used in next step of reaction without further purification.

To a solution of above amine in CH_2Cl_2 (5 mL), pyridine (88 µL, 1.10 mmol), acetic anhydride (47 µL, 0.50 mmol) and DMAP (72.0 mg, 0.59 mmol) were sequentially added at room temperature. After being stirred for 1 h, the reaction was quenched by dilution with ethyl acetate (100 mL). The organic solution was washed with saturated aqueous solution of NaHCO₃ (20 mL) and brine (10 mL), dried over anhydrous

Na₂SO₄ (s) and concentrated under reduced pressure. The residue was purified by column chromatography (silica gel, ethyl acetate / hexane, 1:2 to 1:1) to produce compound **4** (117.0 mg, 92%) as a colorless oil. *R*_f 0.6 (ethyl acetate / hexane, 1:1); $[\alpha]_D^{20} = -4.9$ (*c* 0.4, CHCl₃); ¹**H** NMR (300 MHz, CDCl₃) exists as rotational conformers: δ 7.64 – 7.48 (m, 1H), 7.37 – 7.09 (m, 10H), 7.05 (d, *J* = 8.9 Hz, 0.5H), 6.94 (d, *J* = 9.5 Hz, 0.5H), 6.64 (d, *J* = 6.5 Hz, 0.5H), 6.50 (d, *J* = 6.5 Hz, 0.5H), 5.27 (d, *J* = 13.0 Hz, 0.5H), 5.12 (s, 1H), 5.05 (d, *J* = 13.1 Hz, 0.5H), 4.86 (brs, 1H), 4.65 – 4.54 (m, 1H), 4.49 (d, *J* = 7.0 Hz, 1H), 4.38 – 4.20 (m, 1.5H), 4.16 – 4.05 (m, 1H), 3.96 (s, 0.5H), 3.82 – 3.64 (m, 5H), 2.66 (d, *J* = 6.3 Hz, 2H), 2.49 – 2.36 (m, 1H), 2.13 – 2.01 (m, 2H), 1.98 (s, 3H), 1.58 – 1.31 (m, 20H), 1.13 (d, *J* = 6.8 Hz, 3H), 0.98-0.93 (m, 9H), 0.88 (s, 9H), 0.82 (s, 9H), 0.77 – 0.56 (m, 6H), 0.05 (s, 6H), -0.08 (s, 3H), -0.23 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) exists as rotational conformers: δ 172.0, 171.6, 169.4, 168.4, 168.2, 154.9, 139.2, 136.8, 129.7, 128.3, 128.0, 127.5, 127.2, 125.9, 82.0, 73.4, 73.0, 71.2, 68.2, 68.0, 66.7, 63.4, 54.0, 48.3, 46.4, 46.1, 43.9, 39.4, 37.9, 36.9, 35.0, 32.3, 31.5, 28.0, 25.8, 25.7, 23.1, 20.9, 18.7, 18.4, 18.2, 18.1, 17.9, 6.6, 4.4, -4.8, -5.1, -5.7; HRMS (ESI) *m*/*z* calcd for C₆₀H₁₀₃N₄O₁₂Si₃⁺ [M+H]⁺ 1155.6875, found 1155.6881.



To a solution of **4** (117.0 mg, 0.10 mmol) in ethyl acetate (5 mL), palladium on charcoal (ca. 20 mg, 10% Pd) was added. The reaction vessel was sealed and the inner atmosphere was switched to H_2 and stirred for 24 h at room temperature. The catalyst was removed by filtration through a pad of Celite and washed with ethyl acetate (3 x 30 mL). The filtrate was concentrated under reduced pressure to produce the corresponding amine **4a** (98.0 mg), which was used in next step of reaction without further purifications.

To a solution of amine **4a** and carboxylic acid **5** in CH_2Cl_2 (5 mL), DIPEA (130 μ L, 0.76 mmol) and PyAOP (188.0 mg, 0.36 mmol) were added at 0 °C. The reaction mixture was warmed to room temperature and stirred for 12 h. The reaction was quenched by addition of water (1 mL) and diluted with ethyl acetate

(100 mL). The organic solution was washed with saturated aqueous solution of NH_4Cl (10 mL) and brine (10 mL), dried over anhydrous Na_2SO_4 (s) and concentrated under reduced pressure. The residue was purified by column chromatography (silica gel, ethyl acetate / hexane / MeOH, 3:1:0.05) to produce the pentapeptide **2** (91.3 mg, 68% from compound **4**) as a colorless oil.

*R*_f 0.4 (ethyl acetate / hexane / MeOH, 3:1:0.05); $[\alpha]_D^{20} = -1.4$ (*c* 0.53, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.86 (brs, 1H), 7.56 (brs, 1H), 7.38 – 7.23 (m, 7H), 7.22 – 7.11 (m, 3H), 6.84 (s, 1H), 6.53 (brs, 1H), 6.20 (s, 1H), 5.13 (s, 2H), 4.75 (s, 1H), 4.50 (s, 2H), 4.28 (brs, 2H), 4.16 (brs, 1H), 4.09 – 3.99 (m, 1H), 3.97 – 3.73 (m, 5H), 3.69 – 3.48 (m, 3H), 2.69 (d, *J* = 6.1 Hz, 2H), 2.30 (s, 1H), 2.12 (brs, 4H), 1.83 – 1.64 (m, 3H), 1.65 – 1.51 (m, 3H), 1.50 – 1.28 (m, 16H), 1.13 (d, *J* = 5.8 Hz, 3H), 1.06 – 0.95 (m, 12H), 0.94 – 0.82 (m, 21H), 0.82 – 0.71 (m, 6H), 0.02 (s, 6H), -0.06 (s, 3H), -0.19 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 172.3, 172.1, 171.2, 170.3, 169.5, 168.2, 168.2, 156.4, 139.2, 136.2, 129.7, 128.4, 128.0, 127.9, 127.8, 125.8, 82.1, 82.0, 77.4, 77.0, 76.6, 76.2, 73.4, 71.8, 67.5, 66.9, 63.4, 54.0, 51.1, 48.7, 48.3, 46.8, 44.1, 43.8, 37.0, 36.3, 35.1, 34.7, 32.5, 29.7, 27.9, 25.8, 25.7, 23.0, 20.8, 19.0, 18.9, 18.4, 18.1, 17.9, 6.7, 4.4, -4.8, -5.1, -5.7; HRMS (ESI) *m/z* calcd for C₆₈H₁₁₇N₆O₁₅Si₃⁺ [M+H]⁺ 1341.7879, found 1341.7872.

2.2. Synthesis of hexapeptide 24



To a solution of acid **26** (1.10 g, 3.00 mmol) and H-*D*-Phe-OMe (663.0 mg, 3.70 mmol) in CH₂Cl₂ (15 mL), DIPEA (2.1 mL, 12.00 mmol), HOBt (730.0 mg, 5.4 mmol) and DIC (0.84 mL, 5.4 mmol) were sequentially added at 0 °C. The reaction mixture was stirred at room temperature for 16 h. and then quenched with saturated aqueous solution of NH₄Cl (1 mL) and diluted with ethyl acetate (150 mL). The organic solution was successively washed with saturated aqueous solution of NH₄Cl (20 mL) and brine (10 mL), dried over anhydrous Na₂SO₄ (s) and evaporated under reduced pressure. The residue was purified by column chromatography (silica gel, ethyl acetate/hexane = 1:2 to 1:1) to give the desired dipeptide **27** (1.38 g, 87%) as a white foam.

 $R_{\rm f}$ 0.6 (ethyl acetate / hexane, 1:1); $[\alpha]_{\rm D}^{20} = -35.0$ (c 1.55, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 7.41 –

7.31 (m, 5H), 7.27 – 7.20 (m, 3H), 7.11 (d, J = 6.0 Hz, 2H), 6.84 (d, J = 5.5 Hz, 1H), 6.31 (s, 1H), 5.14 (s, 2H), 4.88 (dd, J = 12.3, 5.5 Hz, 1H), 4.37 (d, J = 4.4 Hz, 1H), 3.83 – 3.75 (m, 1H), 3.72 (s, 3H), 3.71 – 3.66 (m, 1H), 3.12 (d, J = 3.9 Hz, 2H), 2.05 – 1.89 (m, 2H), 0.89 (s, 9H), 0.05 (s, 3H), 0.04 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 171.5, 171.3, 156.2, 136.3, 135.7, 129.2, 128.6, 128.5, 128.1, 127.9, 127.1, 66.9, 60.7, 54.5, 53.0, 52.1, 37.9, 33.8, 25.8, 18.0, -5.7; **HRMS (ESI)** m/z calcd for C₂₈H₄₁N₂O₆Si⁺ [M+H⁺] 529.2728, found 529.2735.

To a solution of dipeptide **27** (500.0 mg, 0.95 mmol) in ethyl acetate (10 mL) at room temperature, palladium on charcoal (ca. 50 mg, 10% Pd) was added. The reaction vessel was sealed and the inner atmosphere was switched to H_2 and stirred for 10 h at room temperature. The catalyst was removed by filtration through a pad of Celite and washed with ethyl acetate (3 x 30 mL). The filtrate was concentrated under reduced pressure to produce the corresponding amine(*ca.* 369.0 mg), which was used in next step of reaction without further purifications.

To a solution of above amine and *D*-Cbz-Leu-OH (604.9 mg, 2.28 mmol) in CH_2Cl_2 (10 mL), TEA (0.53 mL, 3.80 mmol), HOBt (205.4 mg, 1.52 mmol), EDCI (291.4 mg, 1.52 mmol) were sequentially added sequentially at 0 °C. The reaction mixture was stirred at room temperature for 16 h before it was quenched by addition of saturated aqueous solution of NH₄Cl (1 mL) and diluted with ethyl acetate (150 mL). The organic solution was successively washed with saturated aqueous solution of NH₄Cl (20 mL) and brine (10 mL), dried over anhydrous Na₂SO₄ (s) and evaporated under reduced pressure. The residue was purified by column chromatography to give the corresponding product **7** (560.4 mg, 92%) as a white foam.

*R*_f 0.5 (ethyl acetate / hexane, 1:2); $[\alpha]_D^{20} = -17.6$ (c 1.68, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.42 – 7.17 (m, 8H), 7.13 (d, *J* = 7.5 Hz, 2H), 5.21 (d, *J* = 6.8 Hz, 1H), 5.11 (d, *J* = 12.0 Hz, 1H), 4.99 (d, *J* = 12.2 Hz, 1H), 4.78 (dd, *J* = 14.0, 7.0 Hz, 1H), 4.59 (dd, *J* = 11.3, 5.2 Hz, 1H), 4.06 (dd, *J* = 13.6, 7.1 Hz, 1H), 3.84 – 3.73 (m, 1H), 3.69 (s, 1H), 3.65 (s, 3H), 3.09 (qd, *J* = 13.8, 6.7 Hz, 2H), 2.07 – 1.84 (m, 3H), 1.74 – 1.45 (m, 3H), 0.92 (s, 16H), 0.09 (s, 3H), 0.08 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 172.2, 171.7, 170.8, 156.3, 136.2, 136.1, 129.2, 128.5, 128.3, 128.2, 128.0, 127.0, 67.1, 60.8, 54.1, 53.7, 52.1, 52.0, 41.1, 37.8, 33.6, 26.0, 24.7, 23.0, 21.9, 18.3, -5.4, -5.5; HRMS (ESI) m/z calcd for C₃₄H₅₂N₃O₇Si⁺ [M+H]⁺ 642.3569, found 642.3576.



To a solution of tripeptide 7 (352.7 mg, 0.55 mmol) in ethyl acetate (10 mL), palladium on charcoal (ca. 40 mg, 10% Pd) was added. The reaction vessel was sealed and the inner atmosphere was switched to H_2 and stirred for 24 h at room temperature. The catalyst was removed by filtration through a pad of Celite and washed with ethyl acetate (3 x 30 mL). The filtrate was concentrated under reduced pressure to produce the corresponding amine 7a (*ca.* 253.0 mg), which was used in next step of reaction without further purifications.

To a stirred solution of carboxylic acid $6^{[2]}$ (296.1 mg, 0.45 mmol) and the amine **7a** obtained above in MeCN (5 mL), DIPEA (0.32 mL, 1.83 mmol), HOAT (54.4 mg, 0.40 mmol) and HATU (258.6 mg, 0.68 mmol) were sequentially added at 0 °C. The reaction mixture was warmed to room temperature and stirred for 12 h and then quenched by addition of water (1 mL) and diluted with ethyl acetate (100 mL). The organic solution was washed with saturated aqueous solution of NH₄Cl (10 mL) and brine (10 mL), dried over anhydrous Na₂SO₄(s) and concentrated under reduced pressure. The residue was purified by column chromatography (silica gel, ethyl acetate / hexane, 1:4 to 1:2) to afford hexapeptide **24** (397.7 mg, 63%, from **7**) as a colorless oil.

 $R_{\rm f}$ 0.6 (ethyl acetate / hexane, 1:2); $[\alpha]_{\rm D}^{20} = +61.1$ (*c* 0.75, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.44 (d, J = 7.6 Hz, 1H), 7.34 – 7.13 (m, 6H), 5.41 (brs, 1H), 4.95 (d, J = 9.4 Hz, 1H), 4.77 (ddd, J = 11.6, 7.5, 4.3 Hz, 1H), 4.42 – 4.34 (m, 2H), 4.29 (t, J = 3.1 Hz, 1H), 4.21 (brs, 1H), 4.09 – 4.06 (m, 1H), 3.99 – 3.96 (m, 1H), 3.82 – 3.81 (m, 2H), 3.73 (s, 3H), 3.69 – 3.65 (m, 2H), 3.59 – 3.46 (m, 2H), 3.33 (d, J = 10.9 Hz, 1H), 3.13 (dd, J = 15.2, 4.2 Hz, 1H), 2.65 (dd, J = 15.2, 11.5 Hz, 1H), 2.36 – 2.27 (m, 2H), 2.00 – 1.91 (m, 1H), 1.81 – 1.70 (m, 2H), 1.41 (s, 9H), 1.31 – 1.18 (m, 3H), 1.08 – 1.02 (m, 6H), 1.00 (s, 9H), 0.93 (d, J = 6.6

^[2] Tripeptide **6** was prepared following our previous reported procedures: Wang L.; Liu J. Y.; Zhang, Hui,; Xu, Z. X.; Ye, T. *Synlett*, **2010**, 563–566.

Hz, 3H), 0.88 (s, 9H), 0.85 (s, 9H), 0.80 (d, J = 6.5 Hz, 3H), 0.19 (s, 3H), 0.15 (s, 3H), 0.06 (s, 6H), -0.00 (s, 6H); ¹³C NMR (75 MHz, CDCl₃) δ 173.0, 172.5, 171.1, 171.1, 170.8, 166.8, 155.5, 137.2, 129.0, 128.4, 126.1, 79.3, 73.8, 73.5, 72.6, 66.1, 62.5, 62.3, 60.3, 60.0, 56.8, 55.2, 52.3, 51.6, 51.1, 50.3, 43.9, 42.5, 42.4, 38.3, 35.7, 34.8, 28.2, 25.8, 25.7, 24.6, 23.3, 20.5, 18.1, 18.1, 18.0, 11.1, 10.7, -4.71, -4.79, -4.94, -5.03, -5.47, -5.55; **HRMS (ESI)** *m*/*z* calcd for C₅₇H₁₀₃N₆O₁₂Si₃⁺ [M+H]⁺: 1147.6936; found: 1147.6939.

2.3. Total synthesis of 9-(R)-scytonemin A (1a)



To a solution of compound 2 (50.0 mg, 0.037 mmol) in ethyl acetate (5 mL), palladium on charcoal (ca. 10 mg, 10% Pd) was added. The reaction vessel was sealed and the inner atmosphere was switched to H_2 and stirred for 24 h at room temperature. The catalyst was removed by filtration through a pad of Celite and washed with ethyl acetate (3 x 30 mL). The filtrate was concentrated under reduced pressure to produce the corresponding amine **2a** (*ca.* 33.0 mg), which was used in next step of reaction without further purifications.

A solution of NaOH (0.8 mL, 0.5 M) was added to a stirred solution of **24** (50.0 mg, 0.044 mmol) in THF (3 mL) at room temperature. The reaction mixture was stirred at room temperature for 1 h before it was acidified to pH 1–2 with HCl (1.0 M in water) and extracted with ethyl acetate (100 mL). The combined organic layers were washed with brine (15 mL), dried over anhydrous Na₂SO₄ (s) and concentrated under reduced pressure to afford the desired acid **3** (*ca.* 45.0 mg), which was used directly in the next step without further purification.

To a solution of the above amine **2a** and carboxylic acid **3** in dry MeCN (5 mL), *N*-Methylmorpholine (15.0 μ L, 0.14 mmol), HOAT (5.4 mg, 0.04 mmol) and HATU (34.0 mg, 0.09 mmol) were sequentially added at 0 °C. The reaction mixture was warmed to room temperature and stirred for 12 h and then quenched by addition of water (1 mL) and diluted with ethyl acetate (100 mL). The organic solution was washed with saturated aqueous solution of NH₄Cl (10 mL) and brine (10 mL), dried over anhydrous Na₂SO₄ (s) and concentrated under reduced pressure. The residue was purified by column chromatography (silica gel, ethyl acetate / hexane, 3:1) to give the linear undecapeptide **25** (40.4 mg, 47% from **2**) as a colorless oil.

*R*_f 0.3 (ethyl acetate / hexane / MeOH, 2:1:0.05); $[α]_D^{20} = +10.1$ (*c* 0.44, CHCl₃); ¹H NMR (500 MHz, CDCl₃) exists as rotational conformers: δ 7.67 − 7.53 (m, 2H), 7.49 − 7.41 (m, 2H), 7.35 − 7.28 (m, 2H), 7.25 − 7.20 (m, 5H), 7.15 (q, *J* = 5.6 Hz, 3H), 6.85 (d, *J* = 8.6 Hz, 1H), 6.36 (s, 1H), 5.44 (s, 1H), 4.77 (s, 1H), 4.70 (dd, *J* = 15.1, 7.4 Hz, 1H), 4.60 − 4.41 (m, 3H), 4.32 − 4.04 (m, 8H), 4.04 − 3.94 (m, 2H), 3.90 − 3.61 (m, 9H), 3.59 − 3.51 (m, 1H), 3.50 − 3.44 (m, 1H), 3.39 − 3.30 (m, 1H), 3.25 (dd, *J* = 15.0, 7.3 Hz, 1H), 3.17 − 3.08 (m, 1H), 2.90 − 2.80 (m, 1H), 2.73 (dd, *J* = 13.3, 5.1 Hz, 1H), 2.65 (dd, *J* = 13.4, 7.1 Hz, 1H), 2.54 − 2.45 (m, 1H), 2.37 − 2.24 (m, 3H), 2.12 − 2.08 (m, 1H), 2.05 (s, 1H), 2.03 (s, 2H), 2.01 (s, 1H), 1.98 − 1.91 (m, 1H), 1.81 − 1.69 (m, 4H), 1.68 − 1.50 (m, 7H), 1.49 − 1.36 (m, 23H), 1.23 − 1.19 (m, 2H), 1.18 − 1.13 (m, 2H), 1.12 − 1.04 (m, 8H), 1.02 − 0.93 (m, 24H), 0.92 − 0.80 (m, 40H), 0.75 − 0.67 (m, 6H), 0.18 − 0.01 (m, 24H), -0.06 − -0.09 (m, 3H), -0.22 (s, 0.8H), -0.24 (s, 2.2H); ¹³C NMR (125 MHz, CDCl₃) exists as rotational conformers: δ 174.38, 172.63, 172.52, 172.22, 171.74, 171.66, 171.63, 171.15, 170.62, 170.18, 169.99, 168.67, 167.10, 155.72, 139.35, 137.00, 129.80, 129.74, 129.20, 128.74, 128.61, 128.04, 128.00, 126.57, 125.94, 125.89, 81.92, 79.33, 75.58, 73.93, 73.63, 72.93, 68.23, 66.40, 63.68, 62.71, 59.54, 56.61, 55.23, 54.82, 54.24, 51.55, 50.54, 48.75, 48.10, 46.19, 43.91, 43.78, 43.10, 42.60, 38.43, 37.28, 36.86, 35.97, 34.63, 33.85, 31.95, 31.45, 29.66, 28.36, 28.10, 25.93, 25.86, 25.82, 25.79, 24.73, 23.37, 35.

S23

23.18, 23.13, 22.64, 20.99, 20.87, 20.16, 18.87, 18.21, 18.14, 18.01, 17.87, 14.02, 11.27, 11.00, 6.74, 4.60, -4.65, -4.67, -4.86, -4.94, -4.97, -5.43, -5.62, -5.65. **HRMS (ESI)**: *m*/*z* calcd for C₁₁₆H₂₀₉N₁₂O₂₄Si₆⁺ [M+H]⁺ 2322.4113, found 2322.3150.



To a stirred solution of undecapeptide **25** (32.5 mg, 0.014 mmol) in CH_2Cl_2 (5.0 mL), TiCl₄ (70.0 µL, 1.0 M in DCM) was added at 0 °C. The reaction mixture was then allowed to warm to room temperature and stirred for 4 h. The reaction was quenched by addition of aqueous solution of KHSO₄ (5 mL, 0.1 M) and stirred for another 2h. The reaction mixture was diluted with CH_2Cl_2 (100 mL) and washed with NaH₂PO₄-Na₂HPO₄ buffer (2 x 5 mL, 0.1 M, pH 7.1) and brine (5 mL). The organic layer was dried over anhydrous Na₂SO₄ (s) and concentrated under reduced pressure.

The residue was dried under high vacuum for 2h and then dissolved in MeCN-CH₂Cl₂ (40 mL, 9/1, v/v) at 0 $^{\circ}$ C. To this solution, *N*-Methylmorpholine (40.0 µL, 0.36 mmol) and HATU (45.6 mg, 0.12 mmol) were added at 0 $^{\circ}$ C. The reaction mixture was stirred at room temperature for additional 12 h. The reaction was quenched by addition of water (1 mL) and diluted with ethyl acetate (100 mL). The organic solution was washed with saturated aqueous solution of NH₄Cl (5 mL) and brine (5 mL), dried over anhydrous Na₂SO₄ (s) and concentrated under reduced pressure.

The above crude residue was dissolved in THF-HCl (0.1 M, aqueous solution) (9 mL, prepared by addition of 4.3 mL *con*. HCl to 500 mL THF) and stirred for 36 h. The reaction was monitored by HRMS until all silyl protective groups were removed. Volatiles were removed by evaporation under reduced pressure. The

residue was directly subjected to purification by HPLC (Agilent 1200 system, Agilent SB-C18 column (reverse-phase, 21.2×250 mm, 7 µm), linear elution gradient consisting of H₂O-MeCN (from 10% to 98% MeCN over 25 min) at a flow rate of 10 mL/min, retention time: 15.9–16.2 min) to produce 9-(*R*)-scytonemin A **1a** (5.5 mg, 27% 3 steps) as a white powder.

 $[\mathbf{\alpha}]_{D}^{20} = +37 (c \ 0.06, \text{ MeOH}); \text{Ref}^{[3]}: [\mathbf{\alpha}]_{D}^{20} = +38.8 (c \ 0.04, \text{MeOH}); {}^{1}\mathbf{H} \mathbf{NMR} (500 \text{ MHz, DMSO-d6}) \delta 8.41 (s, 1H), 8.19 (s, 1H), 8.17 (d, <math>J = 6.6 \text{ Hz}$, 1H), 8.00 (s, 1H), 7.92 (s, 1H), 7.82 (s, 1H), 7.72 (s, 2H), 7.65 (s, 2H), 7.28 – 7.14 (m, 10H), 4.79 – 4.70 (m, 2H), 4.47 (s, 1H), 4.32 – 4.19 (m, 3H), 4.19 – 4.11 (m, 2H), 4.11 – 4.05 (m, 2H), 4.06 – 4.00 (m, 1H), 4.00 – 3.91 (m, 3H), 3.87 – 3.78 (m, 4H), 3.76 – 3.68 (m, 3H), 3.66 – 3.56 (m, 5H), 3.53 – 3.52 (m, 1H), 3.43 – 3.41 (m, 1H), 3.08 (d, J = 13.2 Hz, 1H), 2.89 – 2.82 (m, 1H), 2.63 (brs, 5H), 2.24 – 2.12 (m, 2H), 2.12 – 2.01 (m, 2H), 1.87 – 1.63 (m, 9H), 1.59 – 1.51 (m, 3H), 1.50 – 1.39 (m, 3H), 1.33 – 1.19 (m, 5H), 1.08 – 1.01 (m, 6H), 0.99 (d, J = 6.3 Hz, 3H), 0.91 (s, 3H), 0.90 (s, 3H), 0.89 – 0.88 (m, 3H), 0.81 (d, J = 5.5 Hz, 3H); ¹³C **NMR** (150 MHz, DMSO-d6) δ 172.42,172.17, 172.07, 172.01, 171.43, 171.28, 171.01, 170.17, 169.95, 169.27, 169.10, 168.56, 166.68, 139.81, 138.01, 129.41, 129.05, 128.17, 128.02, 126.26, 125.72, 75.62, 72.94, 71.37, 71.19, 71.04, 70.90, 67.06, 65.58, 62.21, 61.46, 57.62, 56.36, 55.08, 54.25, 54.01, 52.38, 50.74, 50.50, 48.24, 47.88, 46.29, 43.68, 43.68, 42.33, 42.33, 41.77, 41.10, 37.09, 36.64, 36.47, 34.97, 34.29, 32.06, 30.16, 24.41, 23.47, 22.26, 20.72, 20.64, 19.58, 18.50, 18.30, 17.05, 11.38, 10.56; **HRMS (ESI)** *m*/*z* calcd for C₇₁H₁₀₇N₁₂O₂₁⁺ [M+H]⁺ 1463.7668, found 1463.7686.

^[3]Helms, G. L.; Moore, R. E.; Niemczura, W. P.; Patterson, G. M. L.; Tomer, K. B.; Gross, M. L. J. Org. Chem. **1988**, 53, 1298–1307.



2.4. Analytical data for intermediates of 9-(S)-scytonemin A (1b)

Scheme 1 Synthesis of 9-(*S*)-scytonemin A (1b).



epi-11was prepared from epoxide 10 in 95% yield. Analytical data for epi-11:

[α]_D²⁰ = +19.9 (*c* 1.19, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 7.27 – 7.15 (m, 7H), 6.87 (d, *J* = 8.6 Hz, 2H), 4.51 – 4.45 (m, 1H), 4.43 (s, 2H), 3.90 – 3.83 (m, 1H), 3.79 (s, 3H), 3.65 (ddd, *J* = 9.5, 6.2, 4.7 Hz, 1H), 3.57 (ddd, *J* = 9.4, 7.9, 4.5 Hz, 1H), 2.93 – 2.90 (m, 2H), 2.84 (d, *J* = 3.8 Hz, 1H), 2.44 – 2.30 (m, 2H), 1.85 – 1.77 (m, 1H), 1.76 – 1.69 (m, 1H), 0.83 (s, 9H), -0.05 (s, 3H), -0.07 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 159.4, 137.9, 130.1, 129.9, 129.3, 128.0, 126.4, 113.9, 83.8, 81.5, 73.0, 69.6, 68.2, 64.4, 55.3, 45.5, 35.4, 27.4, 25.7, 18.2, -4.9, -5.2; HRMS (ESI): *m*/*z* calcd for C₂₈H₄₀O₄SiNa⁺ [M+Na]⁺ 491.2588, found 491.2573.



epi-30 was prepared from epi-11 in 97% yield. Analytical data for epi-30:

[α]_D²⁰ = +1.20 (*c* 1.09, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 7.28 – 7.21 (m, 4H), 7.20 – 7.13 (m, 3H), 6.90 – 6.84 (m, 2H), 4.44 (s, 2H), 3.84 (t, J = 5.8 Hz, 1H), 3.79 (s, 3H), 3.77 (brs, 1H), 3.72 – 3.64 (m, 1H), 3.63 – 3.57 (m, 1H), 2.84 (d, J = 2.2 Hz, 1H), 2.75 – 2.69 (m, 2H), 1.76 – 1.68 (m, 2H), 1.50 – 1.35 (m, 6H), 0.85 (s, 9H), -0.05 (s, 3H), -0.19 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 159.3, 139.4, 130.1, 129.7, 129.2, 128.0, 125.9, 113.9, 73.7, 72.9, 71.2, 68.8, 55.2, 43.9, 37.6, 37.0, 36.4, 25.9, 21.2, 18.0, -4.7, -4.9; HRMS (ESI) m/z calcd for C₂₈H₄₅O₄Si⁺ [M+H]⁺ 473.3082, found 473.3084.



epi-12 was prepared from epi-30 in 86% yield. Analytical data for epi-12:

 $[α]_D^{20} = +3.30$ (*c* 0.90, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 7.32 – 7.20 (m, 9H), 7.19 – 7.12 (m, 3H), 6.88 – 6.82 (m, 2H), 4.48 (d, *J* = 11.5 Hz, 1H), 4.43 (d, *J* = 11.5 Hz, 1H), 4.38 (d, *J* = 11.5 Hz, 1H), 4.35 (d, *J* = 11.5 Hz, 1H), 3.84 – 3.79 (m, 1H), 3.78 (s, 3H), 3.59 – 3.47 (m, 3H), 2.70 (d, *J* = 6.3 Hz, 2H), 1.86 – 1.74 (m, 2H), 1.53 – 1.38 (m, 6H), 0.85 (s, 9H), -0.04 (s, 3H), -0.19 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 159.2, 139.4, 139.1, 130.8, 129.7, 129.3, 128.3, 128.1, 127.7, 127.4, 126.0, 113.8, 76.3, 73.8, 72.6, 71.2, 66.8, 55.3, 44.0, 37.2, 34.6, 34.5, 25.9, 21.0, 18.1, -4.6, -4.8;

HRMS (ESI) m/z calcd for C₃₅H₅₁O₄Si⁺ [M+H]⁺ 563.3551, found 563.3541.



epi-31 was prepared from epi-12 in 86% yield. Analytical data for epi-31:

 $[α]_D^{20} = +21.4$ (*c* 1.04, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 7.36 – 7.29 (m, 4H), 7.28 – 7.21 (m, 3H), 7.19 – 7.12 (m, 3H), 4.55 (d, *J* = 11.5 Hz, 1H), 4.45 (d, *J* = 11.5 Hz, 1H), 3.86 – 3.79 (m, 1H), 3.78 – 3.74 (m, 1H), 3.73 – 3.67 (m, 1H), 3.64 – 3.57 (m, 1H), 2.76 – 2.64 (m, 2H), 2.31 (s, 1H), 1.83 – 1.68 (m, 2H), 1.68 – 1.56 (m, 1H), 1.53 – 1.30 (m, 5H), 0.84 (s, 9H), -0.05 (s, 3H), -0.18 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 139.3, 138.5, 129.7, 128.4, 128.1, 127.8, 127.7, 126.0, 78.4, 73.6, 71.0, 60.7, 44.0, 37.1, 36.0, 33.8, 25.9, 20.9, 18.1, -4.7, -4.8;

HRMS (ESI) m/z calcd for C₂₇H₄₃O₃Si⁺ [M+H]⁺, 443.2976, found 443.2983.



epi-14 was prepared from epi-31 in 81% yield (Z:E > 10:1). Analytical data for epi-14:

[α]_D²⁰ = -6.5 (*c* 0.58, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 7.34 – 7.27 (m, 4H), 7.27 – 7.21 (m, 3H), 7.18 – 7.11 (m, 3H), 6.32 (dt, *J* = 11.6, 7.3 Hz, 1H), 5.83 (dt, *J* = 11.6, 1.8 Hz, 1H), 4.53 (d, *J* = 11.6 Hz, 1H), 4.47 (d, *J* = 11.6 Hz, 1H), 4.14 (q, *J* = 7.1 Hz, 2H), 3.84 – 3.74 (m, 1H), 3.54 – 3.48 (m, 1H), 2.95 – 2.90 (m, 2H), 2.68 (d, *J* = 6.3 Hz, 2H), 1.61 – 1.55 (m, 1H), 1.49 – 1.37 (m, 5H), 1.26 (t, *J* = 7.1 Hz, 3H), 0.83 (s, 9H), -0.07 (s, 3H), -0.20 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 166.1, 145.9, 139.2, 138.6, 129.5, 128.0, 127.8, 127.5, 127.2, 125.7, 120.9, 78.0, 73.5, 70.6, 59.5, 43.7, 36.8, 34.2, 32.9, 25.7, 21.0, 17.8, 14.0, -4.9, -5.1; HRMS (ESI) m/z calcd for C₃₁H₄₇O₄Si⁺ [M+H]⁺ 511.3238, found 511.3239.



epi-**33** was prepared from *epi*-**14** in 93% yield. Analytical data for *epi*-**33**: [**α**]_D²⁰ = -1.0 (*c* 1.53, CHCl₃); ¹**H** NMR (500 MHz, CDCl₃) δ 7.35 - 7.29 (m, 4H), 7.29 - 7.21 (m, 3H), 7.21 - 7.10 (m, 3H), 5.81 - 5.72 (m, 1H), 5.64 - 5.54 (m, 1H), 4.49 (s, 2H), 4.18 - 4.03 (m, 2H), 3.82 (t, *J* = 5.3 Hz, 1H), 3.43 - 3.35 (m, 1H), 2.74 - 2.66 (m, 2H), 2.36 (dt, *J* = 14.6, 7.0 Hz, 1H), 2.30 - 2.22 (m, 1H), 1.83 (s, 1H), 1.58 - 1.52 (m, 1H), 1.49 - 1.38 (m, 5H), 0.85 (s, 9H), -0.05 (s, 3H), -0.18 (s, 3H); ¹³**C** NMR (125 MHz, CDCl₃) δ 139.3, 138.5, 130.8, 129.7, 128.9, 128.3, 128.1, 127.8, 127.6, 126.0, 78.3, 73.7, 71.3, 58.2, 44.0, 37.0, 34.1, 32.0, 25.9, 21.2, 18.1, -4.7, -4.8; HRMS (ESI) *m/z* calcd for C₂₉H₄₅O₃Si⁺ [M+H]⁺ 469.3132, found 469.3129.



epi-15 was prepared from *epi*-33 in 86% yield. Analytical data for *epi*-15: [α]_D²⁰ = +5.6 (*c* 0.95, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 7.34 - 7.21 (m, 7H), 7.19 - 7.10 (m, 3H), 4.51 (d, *J* = 11.7 Hz, 1H), 4.47 (d, *J* = 11.7 Hz, 1H), 3.86 - 3.75 (m, 1H), 3.77 - 3.65 (m, 1H), 3.64 - 3.57 (m, 1H), 3.53 (dd, J = 10.8, 5.5 Hz, 1H), 3.14 – 3.07 (m, 2H), 2.76 – 2.62 (m, 2H), 2.21 (s, 1H), 1.91 (dt, J = 14.7, 5.5 Hz, 1H), 1.75 – 1.62 (m, 2H), 1.48 – 1.35 (m, 5H), 0.84 (s, 9H), -0.05 (s, 3H), -0.18 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 139.2, 138.2, 129.7, 128.5, 128.2, 128.0, 127.8, 126.1, 73.6, 71.3, 60.5, 55.9, 53.6, 44.1, 36.9, 33.7, 31.8, 25.9, 21.5, 18.1, -4.6, -4.8; HRMS (ESI) m/z calcd for C₂₉H₄₅O₄Si⁺ [M+H]⁺ 485.3082, found 485.3077.



epi-17 was prepared from *epi*-15 in 86% yield. Analytical data for *epi*-17: $[\alpha]_{D}^{20} = +38.6 (c 1.12, CHCl_3); {}^{1}H NMR (500 MHz, CDCl_3) \delta 7.36 - 7.22 (m, 7H), 7.20 - 7.12 (m, 3H), 4.59 (d,$ *J*= 11.1 Hz, 1H), 4.39 (d,*J* $= 11.1 Hz, 1H), 3.87 - 3.80 (m, 1H), 3.67 - 3.52 (m, 5H), 2.76 - 2.64 (m, 2H), 2.54 (s, 1H), 2.06 (s, 1H), 1.75 - 1.68 (m, 2H), 1.54 - 1.30 (m, 6H), 0.84 (s, 9H), -0.05 (s, 3H), -0.18 (s, 3H); {}^{13}C NMR (125 MHz, CDCl_3) \delta 139.2, 138.2, 129.7, 128.5, 128.2, 128.0, 127.8, 126.1, 76.0, 74.5, 73.6, 71.2, 64.1, 61.2, 44.0, 37.1, 36.0, 34.0, 25.9, 20.5, 18.1, -4.7, -4.8; HRMS (ESI):$ *m*/*z* $calcd for <math>C_{29}H_{46}N_3O_4Si^+$ [M+H]⁺ 528.3252, found 528.3248.



epi-20 was prepared from epi-17 in 76% yield. Analytical data for epi-20:

[α]_D²⁰ = +1.7 (*c* 1.20, CHCl₃); ¹**H** NMR (300 MHz, CDCl₃) δ 7.40 – 7.24 (m, 7H), 7.24 – 7.12 (m, 3H), 4.64 – 4.53 (m, 2H), 4.40 (d, *J* = 11.1 Hz, 1H), 4.15 (dd, *J* = 6.4, 2.5 Hz, 1H), 4.07 (dd, *J* = 10.0, 2.5 Hz, 2H), 3.88 – 3.80 (m, 2H), 3.65 – 3.55 (m, 1H), 3.30 (d, *J* = 6.4 Hz, 1H), 2.74 (dd, *J* = 6.3, 2.0 Hz, 2H), 1.93 – 1.80 (m, 2H), 1.70 – 1.31 (m, 16H), 0.88 (s, 9H), 0.88 (s, 9H), 0.04 (s, 3H), 0.03 (s, 3H), -0.02 (s, 3H), -0.16 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 170.3, 168.8, 139.1, 137.8, 129.6, 128.4, 128.1, 127.8, 127.8, 126.0, 82.1, 75.8, 73.5, 73.5, 70.9, 63.4, 60.9, 54.6, 43.9, 37.0, 35.2, 33.6, 27.9, 25.8, 25.6, 20.3, 18.0, 18.0, -4.8, -4.9, -5.7, -5.7; **HRMS (ESI)** *m*/*z* calcd for C₄₂H₇₁N₄O₇Si₂⁺ [M+H]⁺ 799.4856, found 799.4871.



epi-37 was prepared from epi-20 in 73% yield. Analytical data for epi-37:

 $[\alpha]_D^{20} = -9.6 (c \ 0.59, CHCl_3); {}^{1}H \ NMR (300 \ MHz, CDCl_3) exists as rotational conformers: <math>\delta \ 7.59 (dd, J = 22.1, 7.4 \ Hz, 1H), 7.42 - 7.11 (m, 15H), 6.87 (d, J = 6.8 \ Hz, 0.4H), 6.67 (d, J = 7.5 \ Hz, 0.6H), 5.52 (d, J = 6.5 \ Hz, 0.6H), 5.22 - 5.00 (m, 2H), 4.92 (s, 0.4H), 4.62 - 4.39 (m, 2H), 4.33 - 4.10 (m, 2H), 4.10 - 3.99 (m, 1H), 3.81 (s, 2H), 3.73 - 3.40 (m, 4H), 2.78 - 2.63 (m, 2H), 2.50 - 2.27 (m, 1H), 2.15 - 1.96 (m, 1H), 1.84 - 1.66 (m, 1H), 1.60 - 1.30 (m, 17H), 1.12 - 0.96 (m, 3H), 0.87 (s, 9H), 0.85 (s, 9H), 0.03 (s, 6H), -0.05 (s, 3H), -0.20 (brs, 3H); {}^{13}C \ NMR (75 \ MHz, CDCl_3) exists as rotational conformers: <math>\delta \ 173.2, \ 172.9, \ 172.0, \ 171.7, 168.6, 167.0, 155.0, 139.3, 138.5, 136.5, 129.7, 128.4, 128.4, 128.3, 128.0, 127.9, 127.6, 125.9, 81.6, 76.1, 74.5, 73.6, 71.1, 67.6, 67.0, 63.5, 54.3, 51.3, 46.3, 45.9, 43.8, 39.6, 37.5, 37.3, 35.1, 34.5, 33.9, 33.5, 32.2, 31.5, 27.9, 25.8, 25.6, 20.5, 20.1, 18.7, 18.2, 18.1, 18.0, 17.9, -4.8, -5.0, -5.7; \ HRMS (ESI) \ m/z \ calcd for C_{56}H_{88}N_3O_{10}Si_2^+ \ [M+H]^+ \ 1018.6003, \ found \ 1018.6018.$



*epi-***21** was prepared from *epi-***37** in 92% yield. Analytical data for *epi-***21**: $[\alpha]_D^{20} = -17.4 \ (c \ 0.81, \text{CHCl}_3); {}^1\mathbf{H} \mathbf{NMR} \ (300 \text{ MHz, CDCl}_3) \text{ exists as rotational conformers: } \delta 7.65 - 7.49$ (m, 1H), 7.45 - 7.12 (m, 15H), 7.00 (d, <math>J = 9.0 Hz, 0.5H), 6.90 (d, J = 9.6 Hz, 0.5H), 5.26 (d, J = 12.6 Hz, 0.5H), 5.14 (s, 1H), 4.99 (d, J = 13.0 Hz, 0.5H), 4.59 - 4.40 (m, 3H), 4.35 - 4.22 (m, 1.5H), 4.14 - 4.04 (m, 1.5H), 3.85 (d, J = 5.0 Hz, 1H), 3.82 - 3.54 (m, 4H), 3.51 - 3.25 (m, 1H), 2.77 - 2.58 (m, 2H), 2.54 - 2.40 (m, 1H), 2.18 - 2.03 (m, 1H), 1.98 - 1.86 (m, 1H), 1.60 - 1.23 (m, 17H), 1.21 - 1.09 (m, 3H), 0.98 - 0.79 (m, 27H), 0.72 - 0.56 (m, 6H), 0.04 (s, 6H), -0.07 (s, 3H), -0.23 (s, 3H); {}^{13}C \mathbf{NMR} \ (75 \text{ MHz, CDCl}_3) \text{ exists} as rotational conformers: } \delta 172.3, 172.1, 171.5, 171.2, 168.5, 168.4, 155.0, 150.0, 139.3, 138.9, 136.3, 129.7, 128.3, 128.2, 127.9, 127.7, 127.5, 127.2, 127.1, 125.8, 81.8, 75.9, 73.7, 73.0, 72.2, 71.9, 68.3, 66.9, \\ \end{array} 63.6, 54.0, 49.1, 46.4, 45.9, 43.8, 39.7, 38.1, 37.5, 34.9, 34.6, 33.9, 33.5, 32.1, 31.4, 28.0, 25.8, 25.7, 21.1, 20.9, 18.8, 18.5, 18.1, 18.0, 6.6, 4.4, -4.8, -5.1, -5.8; **HRMS (ESI)** *m*/*z* calcd for C₆₂H₁₀₂N₃O₁₀Si₃⁺ [M+H]⁺ 1132.6868, found 1132.6852.



epi-38 was prepared from epi-21 in 76% yield. Analytical data for epi-38:

 $[\alpha]_{D}^{20} = -3.5 (c \ 0.68, CHCl_3); {}^{1}$ H NMR (300 MHz, CDCl₃) exists as rotational conformers: $\delta \ 7.72 - 7.50$ (m, 2H), 7.41 – 7.09 (m, 10H), 5.22 – 5.04 (m, 2H), 4.52 (d, J = 8.3 Hz, 1H), 4.41 – 3.98 (m, 4H), 3.87 (d, J = 5.0 Hz, 1H), 3.84 – 3.26 (m, 4H), 2.82 – 2.55 (m, 2H), 2.50 – 2.39 (m, 1H), 2.11 – 1.99 (m, 1H), 1.73 – 1.32 (m, 17H), 1.22 – 1.10 (m, 4H), 1.05 – 0.92 (m, 9H), 0.88 (s, 9H), 0.82 (s, 9H), 0.76 – 0.60 (m, 6H), 0.05 (s, 6H), -0.10 (s, 3H), -0.26 (s, 3H); {}^{13}C NMR (75 MHz, CDCl₃) exists as rotational conformers: $\delta \ 173.1$, 172.8, 172.1, 168.3, 155.0, 154.8, 139.5, 136.6, 136.4, 129.7, 128.3, 127.9, 127.7, 127.1, 125.7, 82.1, 73.9, 72.8, 67.8, 66.9, 66.6, 63.6, 54.1, 49.5, 49.3, 46.3, 45.9, 43.6, 39.8, 38.3, 37.3, 37.0, 32.2, 31.5, 28.0, 25.8, 25.7, 22.0, 18.9, 18.6, 18.1, 18.0, 6.6, 4.5, -4.8, -5.1, -5.7; HRMS (ESI) m/z calcd for $C_{55}H_{96}N_3O_{10}Si_3^+$ [M+H]⁺ 1042.6398, found 1042.6380.





MHz, CDCl₃) exists as rotational conformers: δ 172.2, 172.1, 172.1, 171.6, 168.4, 168.3, 155.7, 154.9, 144.0, 143.8, 141.2, 139.2, 136.8, 129.7, 128.3, 127.9, 127.5, 127.3, 127.0, 125.8, 125.1, 119.8, 82.0, 73.4, 73.0, 71.5, 71.3, 68.3, 68.1, 66.8, 66.7, 63.5, 54.0, 49.8, 48.5, 48.3, 47.1, 46.4, 46.1, 43.8, 39.4, 37.8, 37.0, 35.0, 32.3, 31.5, 29.6, 28.0, 25.8, 25.7, 20.8, 18.8, 18.5, 18.1, 17.9, 6.7, 4.4, -4.8, -5.1, -5.7. **HRMS (ESI)** *m*/*z* calcd for C₇₃H₁₁₁N₄O₁₃Si₃⁺ [M+H]⁺ 1335.7450, found 1335.7458.



epi-4 was prepared from epi-23 in 91% yield. Analytical data for epi-4:

 $[\alpha]_D^{20} = +0.3 (c \ 0.77, CHCl_3);$ ¹H NMR (300 MHz, CDCl₃) exists as rotational conformers: δ 7.54 (d, J = 8.5 Hz, 1H), 7.40 – 7.10 (m, 10H), 7.05 (d, J = 9.1 Hz, 0.5H), 6.93 (d, J = 9.4 Hz, 0.5H), 6.61 (d, J = 7.3 Hz, 0.5H), 6.47 (d, J = 6.8 Hz, 0.5H), 5.27 (d, J = 12.8 Hz, 0.5H), 5.12 (s, 1H), 5.05 (d, J = 13.1 Hz, 0.5H), 4.86 (brs, 1H), 4.64 – 4.45 (m, 2H), 4.37 – 4.18 (m, 1.5H), 4.16 – 4.04 (m, 1H), 3.96 (s, 0.5H), 3.81 – 3.49 (m, 5H), 2.67 (d, J = 6.3 Hz, 2H), 2.43 (dt, J = 12.7, 6.5 Hz, 1H), 2.14 – 2.01 (m, 2H), 1.98 (s, 3H), 1.57 – 1.29 (m, 20H), 1.13 (d, J = 6.8 Hz, 3H), 1.06 – 0.91 (m, 9H), 0.88 (s, 9H), 0.82 (s, 9H), 0.75 – 0.55 (m, 6H), 0.05 (s, 6H), -0.08 (s, 3H), -0.24 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) exists as rotational conformers: δ 172.1, 172.0, 171.6, 169.3, 168.2, 154.9, 139.2, 136.8, 129.7, 128.3, 128.0, 127.5, 127.2, 125.8, 82.0, 73.4, 73.0, 71.2, 71.2, 68.2, 68.0, 66.7, 63.4, 54.0, 48.4, 46.4, 46.0, 43.9, 39.5, 37.9, 36.9, 34.9, 32.3, 31.5, 28.0, 25.8, 25.7, 23.1, 20.7, 18.7, 18.4, 18.2, 18.1, 17.9, 6.6, 4.4, -4.8, -5.1, -5.73, -5.74; HRMS (ESI) *m/z* calcd for $C_{60}H_{103}N_4O_{12}Si_3^+$ [M+H]⁺ 1155.6875, found 1155.6878.



epi-2 was prepared from *epi-4* in 70% yield. Analytical data for *epi-2*: $[\alpha]_D^{20} = -5.8 (c \ 0.90, \text{ MeOH}); {}^1\text{H} \text{NMR} (500 \text{ MHz}, \text{ MeOD}) \delta 7.38 - 7.27 (m, 5H), 7.24 - 7.21 (m, 2H), 7.18 - 5.12 (m, 3H), 5.13 (d,$ *J*= 5.4 Hz, 2H), 4.87 (s, 1H), 4.52 - 4.38 (m, 3H), 4.34 (dd,*J*= 14.9, 7.4 Hz, 1H), 4.23 (s, 1H), 4.18 (d,*J*= 8.9 Hz, 1H), 4.00 - 3.91 (m, 3H), 3.84 (d,*J*= 7.5 Hz, 2H), 3.65 (d,*J*= 7.8 Hz, 3H), 3.29 (s, 1H), 2.71 (dd,*J*= 13.3, 5.1 Hz, 1H), 2.64 (dd,*J*= 13.2, 7.1 Hz, 1H), 2.38 (brs, 1H), 2.25 (brs, 1H), 2.10 (s, 3H), 1.89 - 1.58 (m, 5H), 1.49 - 1.40 (m, 18H), 1.14 (d,*J*= 6.9 Hz, 1H), 1.04 (t,*J*= 8.0 Hz, 9H), 1.00 (d,*J*= 6.6 Hz, 3H), 0.94 (d,*J* $= 6.5 Hz, 3H), 0.90 - 0.85 (m, 12H), 0.85 - 0.76 (m, 15H), 0.06 (s, 6H), -0.06 (s, 3H), -0.25 (s, 3H); {}^{13}C \text{ NMR} (125 \text{ MHz}, \text{MeOD}) \delta 174.8, 173.3, 172.7, 172.6, 172.3, 171.6, 168.5, 168.2, 157.6, 139.1, 136.3, 129.4, 128.2, 127.9, 127.8, 127.6, 125.7, 82.9, 75.9, 73.9, 73.6, 70.9, 67.3, 67.0, 63.0, 55.8, 55.0, 49.5, 46.1, 44.3, 43.4, 37.1, 36.6, 35.6, 35.0, 31.7, 31.4, 27.2, 25.1, 25.1, 21.1, 20.8, 18.2, 18.1, 17.9, 17.5, 17.4, 15.8, 5.9, 4.3, -5.7, -6.0, -6.6, -6.7;$ **HRMS (ESI)***m*/*z* $calcd for <math>C_{68}H_{112}N_6 O_{15}Si_3^+ [\text{M+H}]^+$ 1341.7879, found 1341.7872.



epi-25 was prepared from epi-2 in 51% yield. Analytical data for epi-25:

 $[\alpha]_{D}^{20}$ = +21.2 (c 0.70, MeOH); ¹H NMR (500 MHz, MeOD) exists as rotational conformers: δ 7.31 – 7.13 (m, 10H), 4.56 – 4.25 (m, 9H), 4.19 (d, J = 9.1 Hz, 1H), 4.17 – 4.12 (m, 1H), 4.10 – 4.05 (m, 1H), 4.00 (d, J = 10.2 Hz, 1H), 3.93 - 3.82 (m, 5H), 3.80 - 3.67 (m, 5H), 3.62 (dd, J = 7.0, 3.3 Hz, 1H), 3.56 - 3.45 (m, 5H), 3.62 (dd, J = 7.0, 3.3 Hz, 1H), 3.56 - 3.45 (m, 5H), 3.62 (dd, J = 7.0, 3.3 Hz, 1H), 3.56 - 3.45 (m, 5H), 3.80 - 3.67 (m, 5H), 3.62 (dd, J = 7.0, 3.3 Hz, 1H), 3.56 - 3.45 (m, 5H), 3.80 - 3.67 (m, 5H), 3.62 (dd, J = 7.0, 3.3 Hz, 1H), 3.56 - 3.45 (m, 5H), 3.80 - 3.67 (m, 5H), 3.80 - 3.67 (m, 5H), 3.62 (m, 5H), 3.80 - 3.67 (m, 5H), 3.62 (dd, J = 7.0, 3.3 Hz, 1H), 3.56 - 3.45 (m, 5H), 3.80 - 3.67 (m, 5H), 3.80 - 3.85 (m, 7H), 3H), 3.27 – 3.20 (m, 1H), 2.76 (dd, J = 13.2, 4.8 Hz, 1H), 2.65 (dd, J = 13.3, 7.2 Hz, 1H), 2.40 – 2.23 (m, 4H), 2.00 (s, 3H), 1.78 – 1.55 (m, 9H), 1.47 (s, 11H), 11.45 – 1.40 (m, 16H), 1.14 (t, J = 7.1 Hz, 6H), 1.10 (d, J = 6.6 Hz, 3H), 1.06 - 1.01 (m, 12H), 1.00 - 0.95 (m, 9H), 0.93 (s, 15H), 0.91 (s, 15H), 0.89 (s, 12H), 0.91 (s, 15H), 0.91 (s, 15H), 0.89 (s, 12H), 0.91 (s, 15H), 0.89 (s, 12H), 0.91 (s, 15H), 0.91 (s, 15H), 0.89 (s, 12H), 0.91 (s, 15H), 0.91 (s, 15H), 0.89 (s, 12H), 0.91 (s, 15H), 00.85 (s, 9H), 0.80 - 0.74 (m, 6H), 0.17 - 0.11 (m, 12H), 0.09 (s, 6H), 0.05 - 0.02 (m, 6H), -0.03 (s, 6H), -0.24 (s, 2.3H), -0.26 (s, 0.7H); ¹³C NMR (125 MHz, MeOD) exists as rotational conformers: δ 173.50, 172.52, 172.41, 172.28, 172.25, 172.21, 172.11, 171.71, 171.69, 171.05, 169.71, 169.68, 168.80, 156.59, 139.27, 137.17, 129.50, 128.87, 128.25, 127.75, 126.48, 125.69, 82.10, 79.18, 75.57, 73.72, 73.64, 73.58, 71.33, 67.94, 66.24, 63.33, 63.22, 63.14, 59.36, 56.37, 55.55, 54.79, 51.51, 51.14, 48.89, 48.53, 46.20, 43.51, 43.13, 42.64, 42.32, 41.84, 39.31, 39.04, 37.50, 37.20, 37.00, 36.55, 34.68, 33.78, 31.83, 31.62, 30.26, 29.29, 29.00, 27.44, 27.23, 25.23, 25.14, 25.06, 25.03, 24.94, 24.66, 22.44, 21.23, 20.20, 19.95, 18.73, 17.86, 17.80, 17.76, 17.63, 17.60, 17.53, 16.47, 12.98, 10.71, 10.34, 5.93, 4.25, -5.55, -5.84, -5.86, -5.98, -6.05, -6.22, -6.39, -6.62, -6.64; **HRMS (ESI)** m/z calcd for $C_{116}H_{209}N_{12}O_{24}Si_6^+[M+H]^+ 2322.4113$, found 2322.4152.



9-(S)-scytonemin A (**1b**) was prepared from *epi*-**25** in 30% yield (11.7 mg). Analytical data for 9-(S)-scytonemin A (**1b**):

[α]²⁰_D = +26 (*c* 0.06, MeOH); ¹H NMR (500 MHz, DMSO-d6) δ 8.38 (s, 1H), 8.16 (d, J = 6.6 Hz, 1H), 7.96 (s, 1H), 7.92 (d, J = 6.7 Hz, 1H), 7.84 (d, J = 8.6 Hz, 1H), 7.74 – 7.57 (m, 3H), 7.26 – 7.13 (m, 10H), 5.29 (s, 1H), 5.19 (s, 1H), 5.12 (s, 1H), 4.81 (s, 1H), 4.74 (s, 1H), 4.66 (s, 1H), 4.43 (d, J = 5.3 Hz, 2H), 4.36 (s, 1H), 4.32 – 4.19 (m, 3H), 4.19 – 4.11 (m, 2H), 4.11 – 4.05 (m, 2H), 4.06 – 4.00 (m, 1H), 4.00 – 3.91 (m, 3H), 3.87 – 3.78 (m, 4H), 3.76 – 3.68 (m, 3H), 3.66 – 3.56 (m, 5H), 3.53 – 3.52 (m, 1H), 3.43 – 3.41 (m, 1H), 3.08 (d, J = 13.2 Hz, 1H), 2.89 – 2.82 (m, 1H), 2.63 (brs, 5H), 2.24 – 2.12 (m, 2H), 2.12 – 2.01 (m, 2H), 1.87 – 1.63 (m, 9H), 1.59 – 1.51 (m, 3H), 1.50 – 1.39 (m, 3H), 1.33 – 1.19 (m, 5H), 1.08 – 1.01 (m, 6H), 0.99 (d, J = 6.3 Hz, 3H), 0.91 (s, 3H), 0.90 (s, 3H), 0.89 – 0.88 (m, 3H), 0.81 (d, J = 5.5 Hz, 3H); ¹³C NMR (150 MHz, DMSO-d6) δ 172.39, 172.17, 172.05, 172.03, 171.44, 171.24, 171.00, 170.18, 169.92, 169.25, 169.07, 168.51, 166.66, 139.78, 137.98, 129.42, 129.03, 128.16, 128.01, 126.26, 125.72, 75.65, 72.97, 71.37, 71.19, 71.05, 71.05, 67.06, 65.58, 62.22, 61.44, 57.61, 56.36, 55.05, 54.22, 54.01, 52.30, 50.74, 50.49, 48.25, 47.85, 46.30, 43.69, 43.69, 42.31, 42.31, 41.75, 41.10, 37.07, 36.63, 36.63, 35.10, 34.30, 32.09, 30.16, 24.40, 23.46, 22.25, 20.86, 20.62, 19.56, 18.50, 18.28, 17.06, 11.37, 10.54; HRMS (ESI) *m/z* calcd for C₇₁H₁₀₇N₁₂O₂₁⁺ [M+H]⁺ 1463.7668, found 1463.7686.

C							
HO,, 9 Ahda MePro HyLeu Gly I HN D-Phe 9-(R	NH O	D-Ser OH OH OH OH OH OH D-I OH L-Hser NH D-I	но _Н HyMePro II		h h h h h h h h h h) ^{∞,}) ^{∞,}) ^{∞,})	
Amino unit	Assignment	δ, ppm Natural Scytonemin A (75 M)	δ_{la} , ppm 9-(<i>R</i>)-Scytonemin A (1a) (150M)	$\Delta \delta_a = \delta_{Ia} \cdot \delta$ (ppm)	δ _{lb} , ppm 9-(S)-Scytonemin A (1b) (150M)	$\varDelta \delta_b = \delta_{lb} - \delta$ (ppm)	
	C=O	172.46	172.42	-0.04	172.39	-0.07	
	C=O	172.27	172.17	-0.10	172.17	-0.10	
	C=O	172.18	172.07	-0.11	172.05	-0.13	
	C=O	172.18	172.01	-0.17	172.03	-0.15	
	C=O	171.55	171.43	-0.12	171.44	-0.11	
	C=O	171.39	171.28	-0.11	171.24	-0.15	
Carbonyl	C=O	171.10	171.01	-0.09	171.00	-0.10	
	C=O	170.33	170.17	-0.16	170.18	-0.15	
	C=O	170.09	169.95	-0.14	169.92	-0.17	
	C=O	169.39	169.27	-0.12	169.25	-0.14	
	C=O	169.18	169.10	-0.08	169.07	-0.11	
	C=O	168.65	168.56	-0.09	168.51	-0.14	
	C=O	166.78	166.68	-0.10	166.66	-0.12	
	C1'	139.74	139.81	+0.07	139.78	+0.04	
	C3',5'	128.98	129.05	+0.07	129.03	+0.05	
	C2',6'	128.09	128.17	+0.08	128.16	+0.07	
Ahda	C4'	125.65	125.72	+0.07	125.72	+0.07	
	C2	72.87	72.94	+0.07	72.97	+0.10	
	C3	50.74	50.74	0	50.74	0	

2.5. ¹³C NMR comparison of synthetic scytonemin A (1a, 1b) & natural scytonemin A (in DMSO-d6)
Amino unit	Assignment	δ, ppm Natural Scytonemin A (75 M)	δ _{la} , ppm 9-(R)-Scytonemin A (1a) (150M)	$arDelta\delta_a=\delta_{la}$ - δ (ppm)	δ _{Ib} , ppm 9-(S)-Scytonemin A (1b) (150M)	$arDelta\delta_b=\delta_{lb}$ - δ (ppm)
Ahda	C4	36.45	36.47	+0.02	36.63	+0.18
	C5	71.03	71.04	+0.01	71.05	+0.02
	C6	34.24	34.29	+0.05	34.30	+0.06
	C7	20.62	20.72	+0.10	20.86	+0.24
	C8	39.79	*	-	*	-
	C9	70.86	70.90	+0.04	71.05	+0.19
	C10	43.64	43.68	+0.04	43.69	+0.05
	C2	47.82	47.88	+0.06	47.85	+0.03
Ala	C3	17.02	17.05	+0.03	17.06	+0.04
	CH3	22.21	22.26	+0.05	22.25	+0.04
	C2	62.24	62.21	-0.03	62.22	-0.02
	C3	42.28	42.33	+0.05	42.31	+0.03
MePro	C4	32.05	32.06	+0.01	32.09	+0.04
	C5	43.64	43.68	+0.04	43.69	+0.05
	CH3	18.28	18.30	+0.02	18.28	0
	C2	52.32	52.38	+0.06	52.30	-0.02
	C3	75.58	75.62	+0.04	75.65	+0.07
HyLeu	C4	30.13	30.16	+0.03	30.16	+0.03
	CH3	19.55	19.58	+0.03	19.56	+0.01
	CH3	18.44	18.50	+0.06	18.50	+0.06
Gly I	C2	46.24	46.29	+0.05	46.30	+0.06
	C1'	137.90	138.01	+0.11	137.98	+0.08
D-Phe	C3',C5'	129.33	129.41	+0.08	129.42	+0.09
	C2',6'	127.94	128.02	+0.08	128.01	+0.07
	C4'	126.20	126.26	+0.06	126.26	+0.06
	C2	54.14	54.25	+0.11	54.22	+0.08
	C3	37.06	37.09	+0.03	37.07	+0.01
L-Hser	C2	50.49	50.50	+0.01	50.49	0
	C3	34.93	34.97	+0.04	35.10	+0.17
	C4	57.61	57.62	+0.01	57.61	0
D-Leu	C2	48.18	48.24	+0.06	48.25	+0.07
	C3	41.08	41.10	+0.02	41.10	+0.02
	C4	24.40	24.41	+0.01	24.40	0
	CH3	23.42	23.47	+0.05	23.46	+0.04
	CH3	20.62	20.64	+0.02	20.62	0

Amino unit	Assignment	<i>δ</i> , ppm Natural Scytonemin A (75 M)	δ _{la} , ppm 9-(R)-Scytonemin A (1a) (150M)	$\Delta \delta_a = \delta_{la} - \delta$ (ppm)	δ _{lb} , ppm 9-(S)-Scytonemin A (1b) (150M)	$\varDelta \delta_b = \delta_{lb} - \delta$ (ppm)
HyMePro I	C2	67.02	67.06	+0.04	67.06	+0.04
	C3	36.56	36.64	+0.08	36.63	+0.07
	C4	71.33	71.37	+0.04	71.37	+0.04
	C5	56.31	56.36	+0.05	56.36	+0.05
	CH3	11.36	11.38	+0.02	11.37	+0.01
HyMePro II	C2	65.57	65.58	+0.01	65.58	+0.01
	C3	41.71	41.77	+0.06	41.75	+0.04
	C4	71.17	71.19	+0.02	71.19	+0.02
	C5	53.95	54.01	+0.06	54.01	+0.06
	CH3	10.52	10.56	+0.04	10.54	+0.02
D-Ser	C2	54.91	55.08	+0.17	55.05	+0.14
	C3	61.41	61.46	+0.05	61.44	+0.03
Gly II	C2	42.28	42.33	+0.05	42.31	+0.03

Note: * stands for the peak was hidden by peaks of deuterated solvent.

3. HPLC comparison of synthetic samples and natural scytonemin A

HPLC Analysis Conditions:

Agilent 1200 system, CHIRALPAK AS-RH column (reverse-phase, 4.6 \times 150 mm, 5 μ m), a constant elution consisting of H₂O-MeCN (74:26) at a flow rate of 0.1 mL/min.

HPLC Chromatogram Comparison

Synthetic scytonemin A (1a and 1b) and natural scytonemin A (injected separately)



HPLC Chromatogram Comparison

Co-injection of synthetic and natural scytonemin A

(1 for natural scytonemin A; 1a for 9-(R)-scytonemin A; 1b for 9-(S)-scytonemin A)



4. ¹H & ¹³C NMR Spectra







samole name:2-202
solvent:CDCl3



Avance 500, Bruker, SZPKU sample:3-059, solvent:CDCl3 spectrum:jyliu



Avance 500, Bruker, SZPKU sample:3-067-2, solvent:CDCl3 spectrum:jyliu





Avance 500, Bruker, SZPKU sample:3-148, solvent:CDCl3 spectrum:jyliu



samole name:3-002
solvent:CDCl3





Avance 500, Bruker, SZPKU sample:3-012-p, solvent:CDCl3 spectrum:jyliu









sample name:4-376-stm solvent:CDCl3 Bruker 300M



sample name:4-350 solvent:CDCl3 Bruker 300M



sample name:4-309-2 solvent:CDCl3 Bruker 300M



sample name:4-312 solvent:CDCl3 Bruker 300M













1-55-p CDC13



sample name:4-325 solvent:CDCl3 Bruker 300M



Avance 500, Bruker, SZPKU sample:4-381-stm, solvent:CDCl3 spectrum:jyliu



Avance 500, Bruker, SZPKU sample:4-TM-L-5-DMSO, solvent:DMSO spectrum:jyliu



TM-L-1



Avance 500, Bruker, SZPKU sample:1-143-zjf user:jyliu solvent: CDC13







Avance 500, Bruker, SZPKU sample:1-156-zjf user:jyliu solvent: CDC13






Avance 500, Bruker, SZPKU sample:1-170-zjf user:jyliu solvent: CDCl3





sample name:4-322 solvent:CDCl3 Bruker 300M

1





sample name:4-328 solvent:CDC13 Bruker 300M





sample name:4-329 solvent:CDCl3 Bruker 300M



sample name:4-331-2 solvent:CDCl3 Bruker 300M







Avance 500, Bruker, SZPKU sample:4-11ptd-D-3-MeOD, solvent:MeOD spectrum:jyliu



Avance 500, Bruker, SZPKU sample:4-11ptd-D-MeOD, solvent:MeOD spectrum:jyliu





TM-D-1







