

ESI

Elucidating the mechanism of interaction between peptides and inorganic surfaces

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Peptide sequences

The protected peptide sequences that were used for tip functionalization were

1. Fmoc-Gln(Trt)-Pro-Ala-Ser(tBu)-Ser(tBu)-Arg(pbf)-Tyr(tBu)-COOH
2. Fmoc-Ala-Pro-Ala-Ser(tBu)-Ser(tBu)Arg(pbf)-Tyr(tBu)-COOH
3. Fmoc-Gln(Trt)-Ala-Ala-Ser(tBu)-Ser(tBu)-Arg(pbf)-Tyr(tBu)-COOH
4. Fmoc-Gln(Trt)-Pro-Ala-Ala-Ser(tBu)-Arg(pbf)-Tyr(tBu)-COOH
5. Fmoc-Gln(Trt)-Pro-Ala-Ser(tBu)-Ala-Arg(pbf)-Tyr(tBu)-COOH
6. Fmoc-Gln(Trt)-Pro-Ala-Ser(tBu)-Ser(tBu)-Ala-Tyr(tBu)-COOH
7. Fmoc-Gln(Trt)-Pro-Ala-Ser(tBu)-Ser(tBu)-Arg(pbf)-Ala-COOH

Detailed version of Fig. 2 (this figure contains the number of measurements and MPF for each experiment).

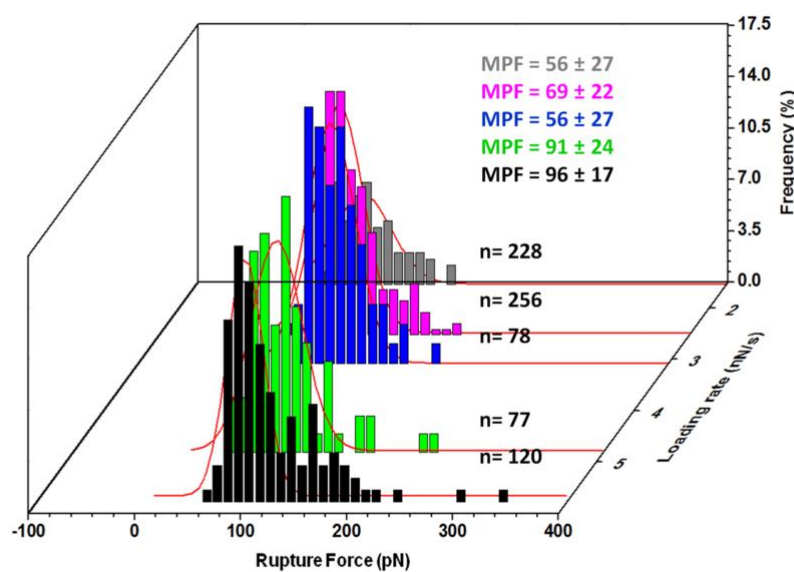


Fig 2. Waterfall representation of a pull-off force histogram for the Q4 interaction between peptide 1 and mica at different apparent loading rates (1.54 nN, 2.52 nN, 3.08 nN, 4.82 nN and 5.80 nN) at 298 K in TDW. The red lines represent the Gaussian fit for each histogram. n is the number of curves used for generating a histogram.

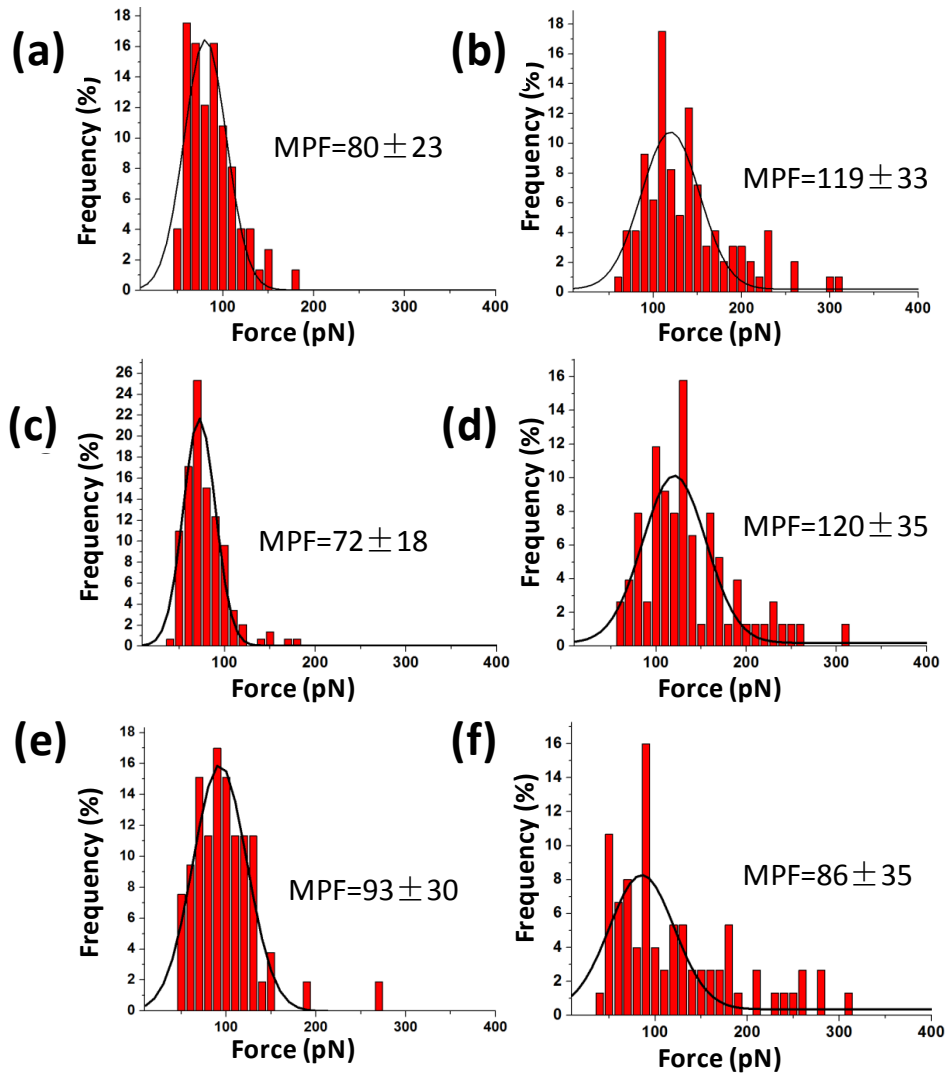


Fig. S1 Force histogram with Gaussian fit (black curve) for the different peptides and mica at different apparent loading rate at 298K. (a) peptide 1 at 3.1 ± 0.6 nN/s ($n=78$) (b) peptide 2 at 3.8 ± 0.9 nN/s ($n=107$) (c) peptide 4 at 3.2 ± 0.7 nN/s ($n=140$) (d) peptide 5 at 3.3 ± 0.8 nN/s ($n=122$) (e) peptide 6 at 2.9 ± 0.7 nN/s ($n=112$) and (f) peptide 7 at 3.5 ± 1.1 nN/s ($n=71$). n is the number of curve used for preparing histogram.

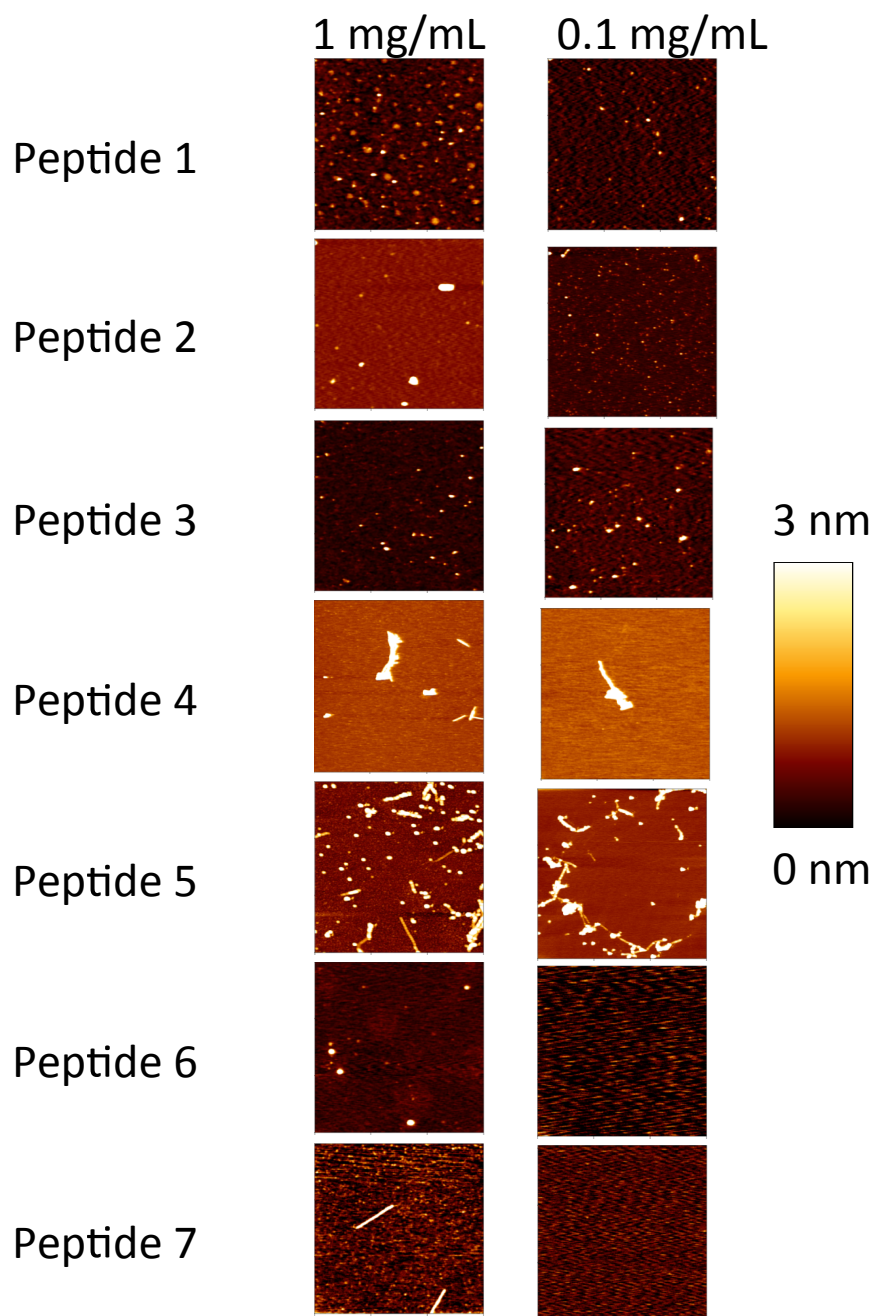


Fig. S2 AFM topography analysis of peptides 1-7 at different concentrations after one hour of incubation. The size scan of all images is 3x3 μm .

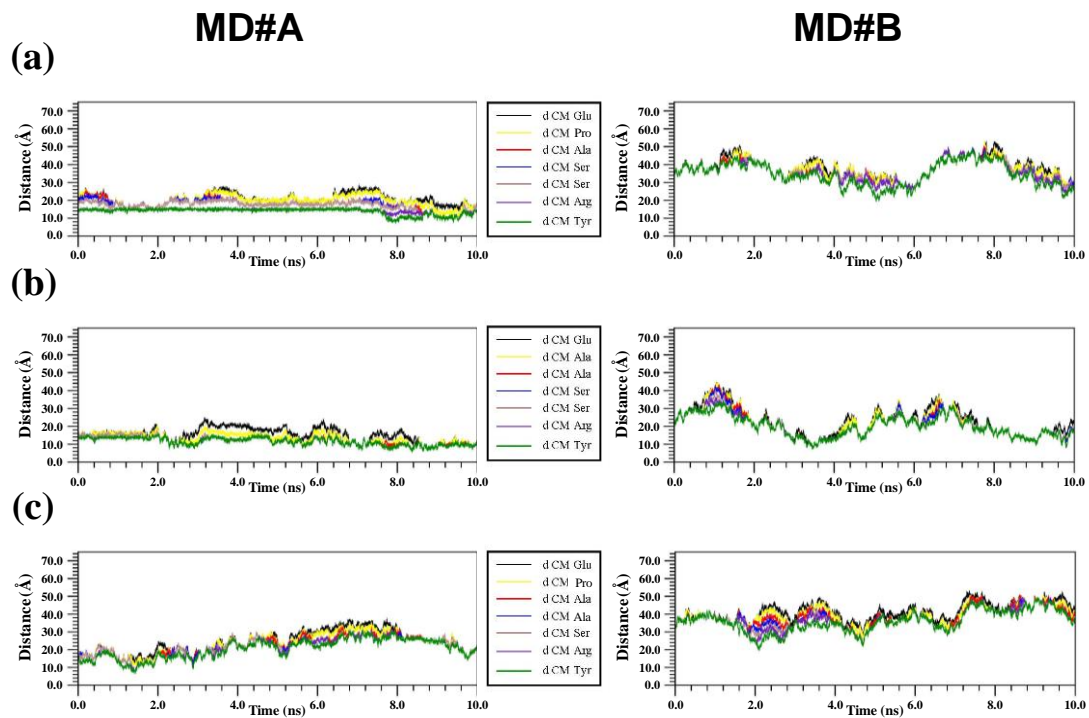


Fig. S3 Temporal evolution of the distance between the center of masses of each residue and the mica surface, calculated using the normal vector, for MD#A and MD#B simulations with #= (a) 1; (b) 2 and (c) 3.

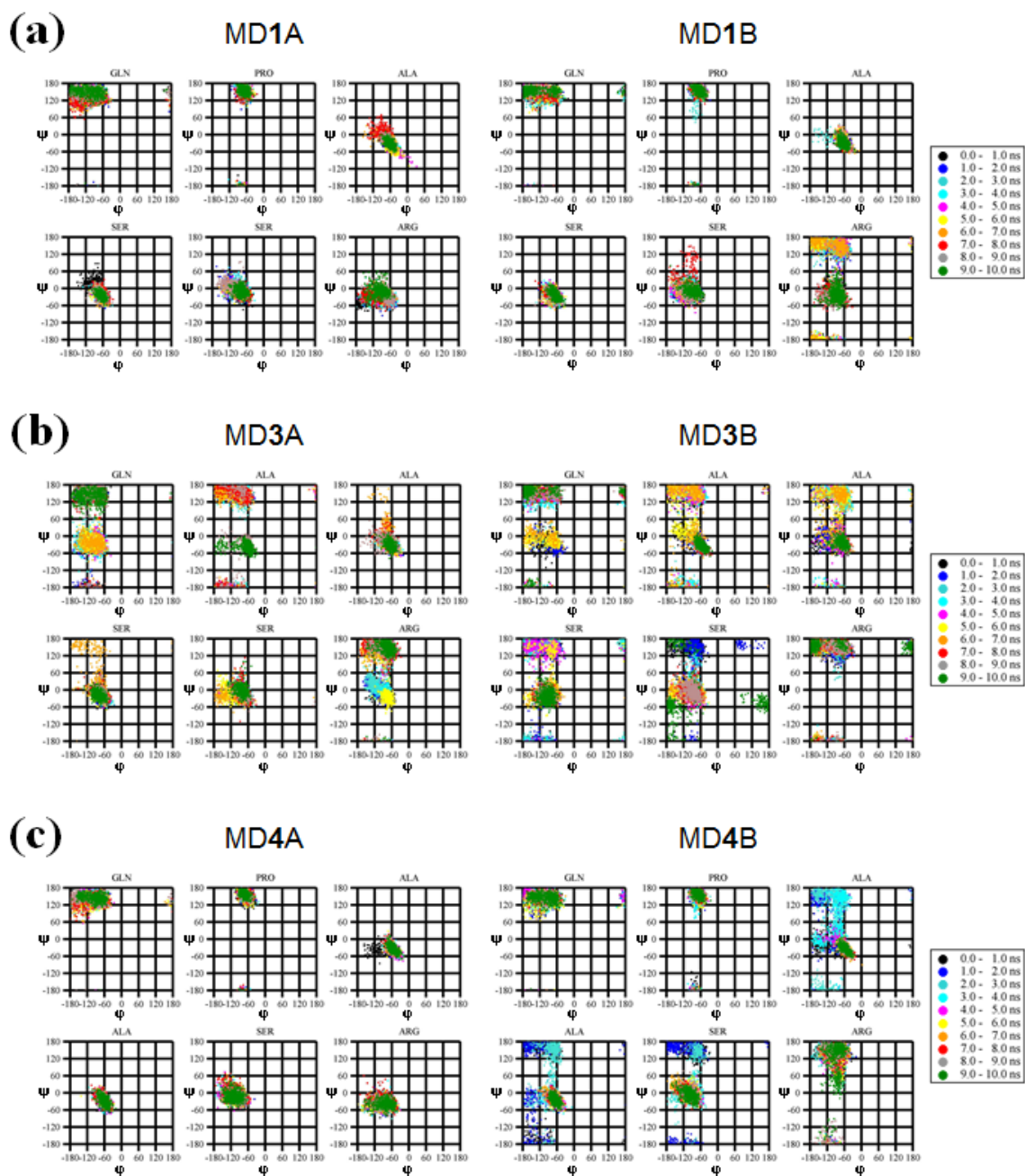


Fig. S4 Time dependent accumulated Ramachandran plots for (a) 1, (b) 3 and (c) 4 depicted using snapshots recorded from MD#A and MD#B simulations. Each different color represents a time period of 1 ns.

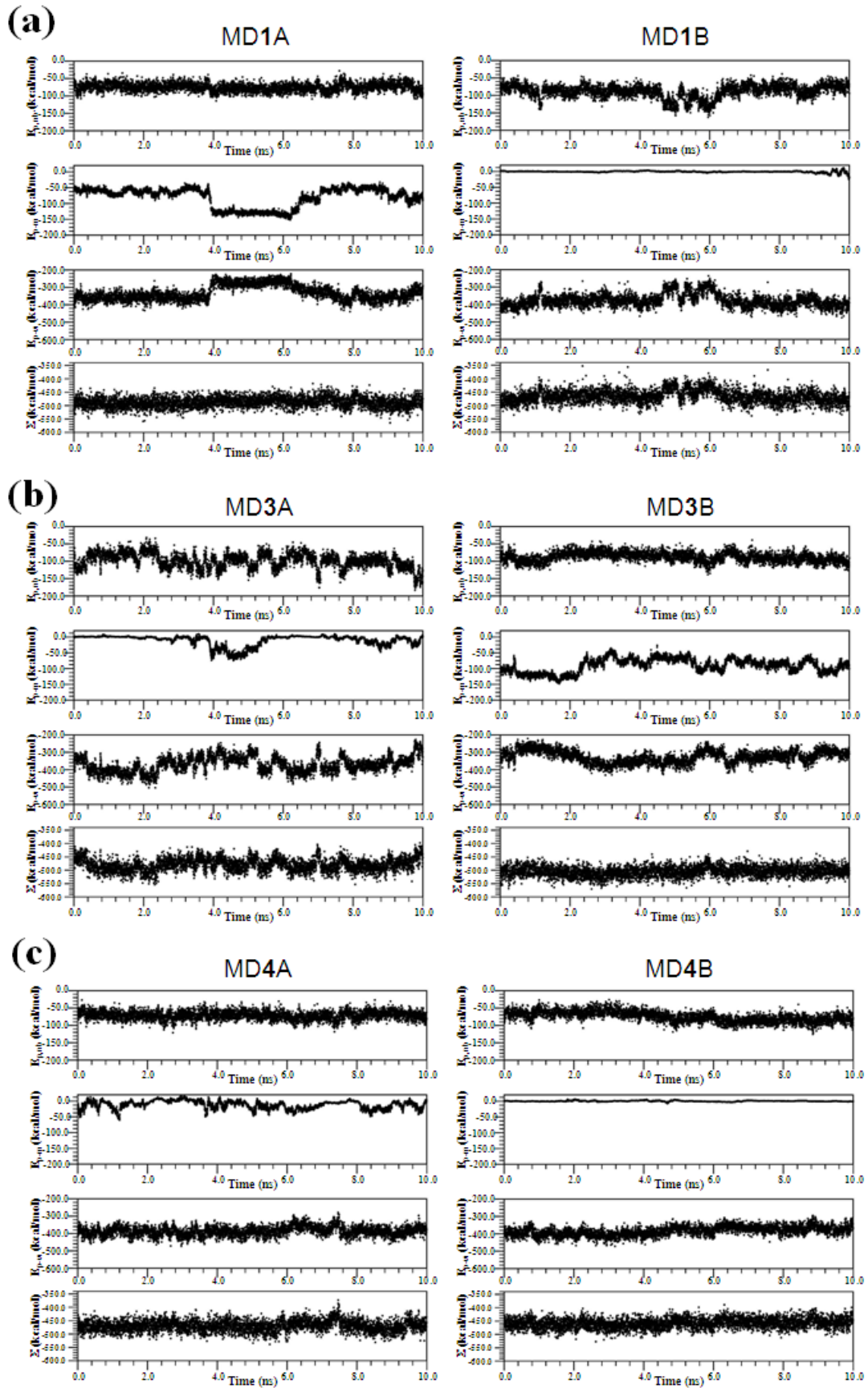


Fig. S5 Temporal evolution of $E_{p,nb}$, $E_{p,m}$, $E_{p,w}$ and Σ (which is the sum of the three previous contributions) for MD#A and MD#B: (a) 1; (b) 2 and (c) 3.

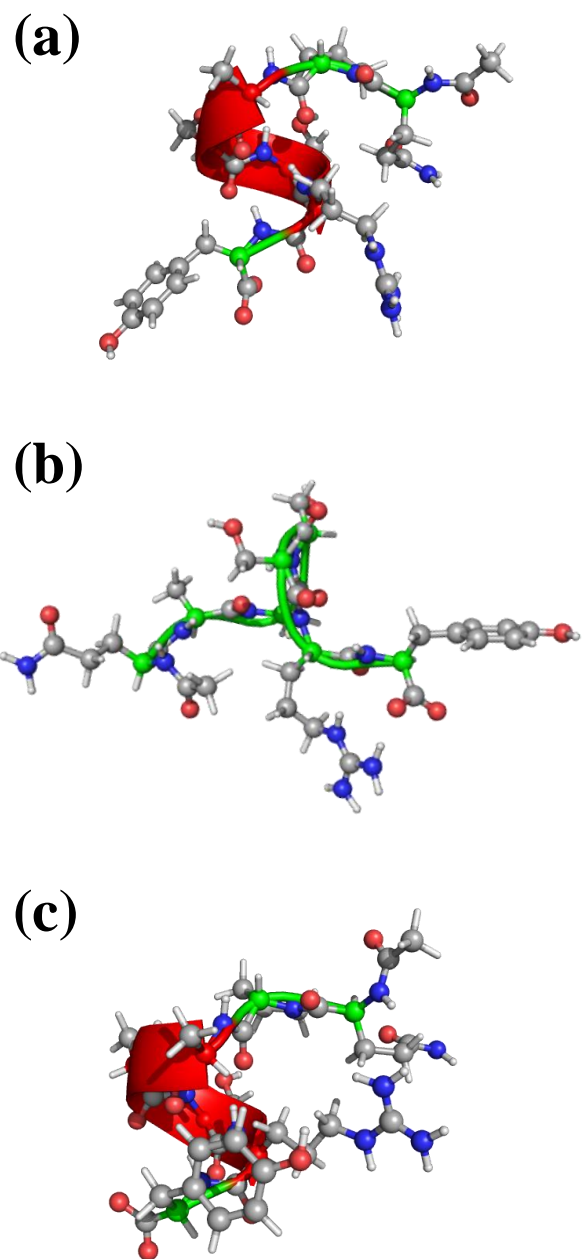


Fig. S6 Representative snapshots of peptides (a) **1**, (b) **3** and (c) **4** extracted from MD#A simulations.

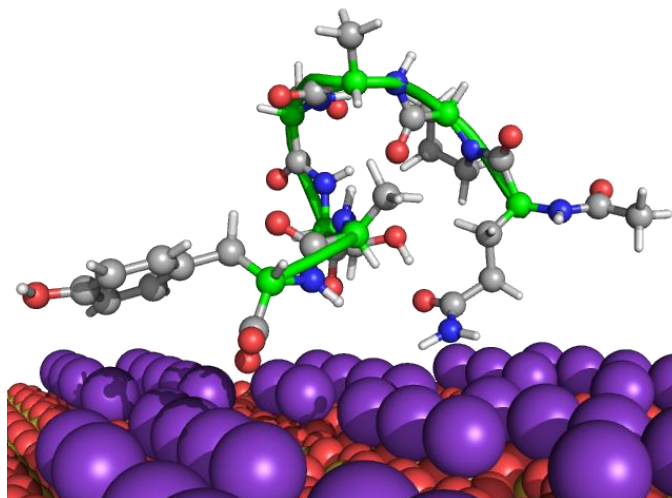


Fig. S7 Representative snapshot of peptides **6** adsorbed onto mica extracted from preliminary MD simulations performed according to the explanation given in the text.

Table S1. Results of the clustering analysis for MD#A and MD#B simulations, where # corresponds to peptides **1, 3** and **4**.

	MD1A	MD1B	MD3A	MD3B	MD4A	MD4B
Total number of unique minima	361	695	758	1554	315	565
Number of minima with hydrogen bonds or salt bridges ($N_{\text{hb/sb}}$)^a	353	672	706	1267	311	416
Number of interactions per minimum included in $N_{\text{hb/sb}}$	3.04	2.40	2.29	2.46	2.89	2.50
Number of clusters found in the $N_{\text{hb/sb}}$ minima	455	209	342	536	225	253
Number of clusters with more than 10 unique minima	11	13	17	11	9	13
Percentage of main chain – main chain hydrogen bonds (%)	53.92	87.63	79.88	53.46	80.69	60.41
Percentage of main chain – side chain hydrogen bonds (%)	42.30	8.59	14.06	43.41	14.18	36.99
Percentage of side chain – side chain hydrogen bonds (%)	3.80	3.81	6.07	3.14	2.16	2.62

^a Salt bridges between the side chain of Arg and the COO⁻ groups of the C-terminal have not been found in any studied system.