Supporting information for:

Exploring Environment-dependent effects of Pd nanostructures on reactive oxygen species (ROS) using electron spin resonance (ESR) technique: implications for biomedical applications

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Fig. S1 (A) UV-Vis spectra and typical TEM images for (B) Pd NPs, and (C) Au@Pd NRs.



Fig. S2 (A) STEM-HAADF images and STEM-EDX element maps of (B) Au and (C) Pd for Au@Pd NRs.



Fig. S3 Observation of the dioxygen bubbles generated at 15 min in samples containing 5 mM H_2O_2 and (A) none, (B) 2.5 µg/mL Pd NPs, (C) 2.5 nM Au@Pd NRs. The pH values were (a) 7.4, (b) 9.4, and (c) 10.9.



Fig. S4 ESR spectra of samples in the absence (A-C) and presence (D-F) of 5 mM H_2O_2 . Solutions contained 0.2 mM ¹⁵N-PDT, and 10 mM buffer with different pH values: (A, D) pH 1.09, (B, E) pH 7.4, (C, F) pH 10.9.



Fig. S5 (A) ESR spectra of ¹⁵N-PDT in the presence of Au@Pd NRs, buffer (pH 7.4), and H_2O_2 at different time intervals. (B) Effect of pH on linewidth for NRs after incubation for 1 min. The solutions contain 10 mM buffer, 0.2 mM ¹⁵N-PDT, 5 mM H_2O_2 and 2.5 nM NRs.



Fig. S6 (A) ESR spectra of BMPO/•OH adducts in the absence and presence of 2.5 nM Au@Pd NRs. Samples contain 10 mM buffer (pH 1.09 or pH 3.9), 5 mM H_2O_2 , 25 mM BMPO. (B) Effect of pH on the generation of •OH. The solutions contain 25 mM BMPO, 5 mM H_2O_2 , 10 mM different pH buffer and 2.5 nM NRs. ESR spectra were collected after 1 min of incubation and averaged from 9 scans.



Fig. S7 (A) UV-Vis spectra of (a) 2.5 mM K_2PdCl_4 , (b) 100 µg/mL Pd NPs (1 mM) in water. (B) Difference of UV-Vis spectra in the presence and absence of 20 µg/mL Pd NPs and reaction for (a) 0 min and (b) 60 min. Sample solutions contain 10 mM buffer (pH 1.09) and 1 mM H_2O_2 .

To investigate the possible mechanism for the generation of •OH in the presence of Pd NPs, we measured the UV-vis extinction spectra of Pd NPs before and after reacting with H₂O₂. Owing to poor dispersibility, Pd NPs solutions show strong scattering and give a gradually increased scattering signal from 800 nm to short wavelength regions (Fig. S7 A). To observe the change after reaction, we obtained the difference of UV-Vis spectra in the presence and absence of 20 μ g/mL Pd NPs before and after reaction for 1 h in 1 mM H₂O₂ and 10 mM buffer (pH 1.09). After the decomposition of H₂O₂ in buffer from Pd NPs and H₂O₂ in buffer, we obtained a qualitative view of reaction products from the change in UV-vis spectra. As Fig. S7 B shows, a spectrum similar to that of Pd²⁺ ions was clearly visible after decomposition of H₂O₂ for 1 h in the presence of Pd NPs. Data obtained from ICP-MS analysis of the supernatant shows the existence of Pd²⁺ ions after reaction for 1 h (data not shown). It is well known that both the oxygen and H₂O₂ have high oxidation abilities under acidic conditions, which can be evidenced from their reduction potential:

O₂ + 4 H⁺+ 4 e⁻ 2 H₂O (+1.229 V) H₂O₂ + 2 H⁺+ 2 e⁻ 2 H₂O (+1.78 V)

Thus, we propose that Pd NPs were oxidized to Pd^{2+} ($Pd^{2+}/Pd = + 0.951$ V) in the present of H_2O_2 under acidic condition:

 $Pd + H_2O_2 + 2 H^+ Pd^{2+} + H_2O$



Fig. S8 ESR spectra of BMPO/•OH adducts obtained from samples containing 25 mM BMPO. ESR spectra were collected after 3 min of incubation. $[K_2PdCl_4] = 2.5 \text{ mM}, [H_2O_2] = 5 \text{ mM}, \text{ and}$ [DMSO] = 10%.

In the presence of Pd^{2+} , •OH can be generated during decomposition of H_2O_2 . A BMPO/•OH signal is not observed from controls (Fig. S8). However, the solution has a strong BMPO/•OH signal in the presence of Pd^{2+} (Fig. S8). Again, adding 10% DMSO reduces signal significantly. As reported in previous research, hydrogen peroxide can produce •OH mainly in two ways: 1) using UV irradiation, and 2) Fenton or Fenton-like reactions involving transition metal ions such as Fe²⁺ and Cu²⁺.^{S1} Herein, we suppose the generation of •OH by Pd NPs under acidic condition is mainly attributed to Pd^{2+} , formed in a reaction with H_2O_2 , and a subsequent Fenton-like reaction ($Pd^{2+} + 2 H_2O_2 Pd^{4+} + 2 \cdot OH + 2 OH^-$).



Fig. S9 SOD-like activity of Au NRs and Au@Pd NRs in scavenging superoxide under neutral conditions in Xan/XOD system. (A) ESR spectra of BMPO/•OOH adducts obtained from samples containing different concentration of Au@Pd NRs or SOD in Xan/XOD system. (B) Concentration effect on O₂⁻⁻ scavenging activity of Au NRs and Au@Pd NRs. Xan/XOD system contains 1 mM Xan, 0.05 U/mL XOD, 0.05 mM DTPA, and 10 mM PBS buffer (pH 7.4). ESR spectra were collected after incubation with 25 mM BMPO for 5 min.



Fig. S10 Concentration effect on singlet oxygen scavenging activity of Au NRs and Au@Pd NRs in ZnO nanoparticles (0.02 mg/mL) system. ESR spectra were collected after irradiation for 1 min containing 10 mM TEMP.



Fig. S11 (A) ESR spectrum of ascorbyl radical. (B) Evolution of UV-Vis spectra for a sample containing 2 U/mL AAO and 5 mM AA in 10 mM PBS (pH 7.4).



Fig. S12 Catalytic activity of NRs during oxidation of AA. (A) AA absorbance at 265 nm as a function of time with different concentrations of Au NRs and Au@Pd NRs. UV-Vis spectra for solutions of NRs with 0.25 mM AA in 10 mM PBS (pH 7.4). (B) ESR spectra for samples containing CTPO and NRs after mixing 5 min. The concentrations are: [CTPO] = 100 μ M, [NR] = 2.5 nM, [AA] = 5 mM, and [PBS] = 10 mM.

Effect on CPH



Fig. S13 Catalytic role of Pd NSs on CPH oxidation. (A) ESR spectra of CP• obtained from samples containing 100 μ M CPH, and different concentration of Pd NPs. Effects of (B) Pd NPs and (C) NRs concentration on ESR signal intensity. ESR spectra were collected after incubation for 3 min. [CPH] = 100 μ M.

We use 1-Hydroxy-3-carboxy-2,2,5,5-tetramethylpyrrolidine- hydrochloride (CPH) as an ESR probe to demonstrate the catalytic role of NSs acting as an oxidase mimic. CPH, itself, is ESR silent, but can be oxidized to form CP-nitroxide radicals (CP•) which show a characteristic ESR signal having three lines with intensity ratios of 1:1:1 (Fig. S13A).^{S2} We observe nearly linear concentration-dependence in CP• intensity from solutions containing CPH and Pd NSs (Fig. S13B and S13C). Both Pd NPs and Au@Pd NRs show catalytic activity on CPH oxidation. Compared to Au NRs, Au@Pd NRs show an enhanced activity for catalyzing the oxidization of CPH at same particle concentration, indicating that Pd is better than Au in catalyzing CPH oxidation by oxygen molecules.

References

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