

**Supporting Information**

**Hydrogen Bonding Play a Significant Role in Binding of Coomassie Brilliant Blue-R to Hemoglobin: FT-IR, Fluorescence and Molecular Dynamics Studies**

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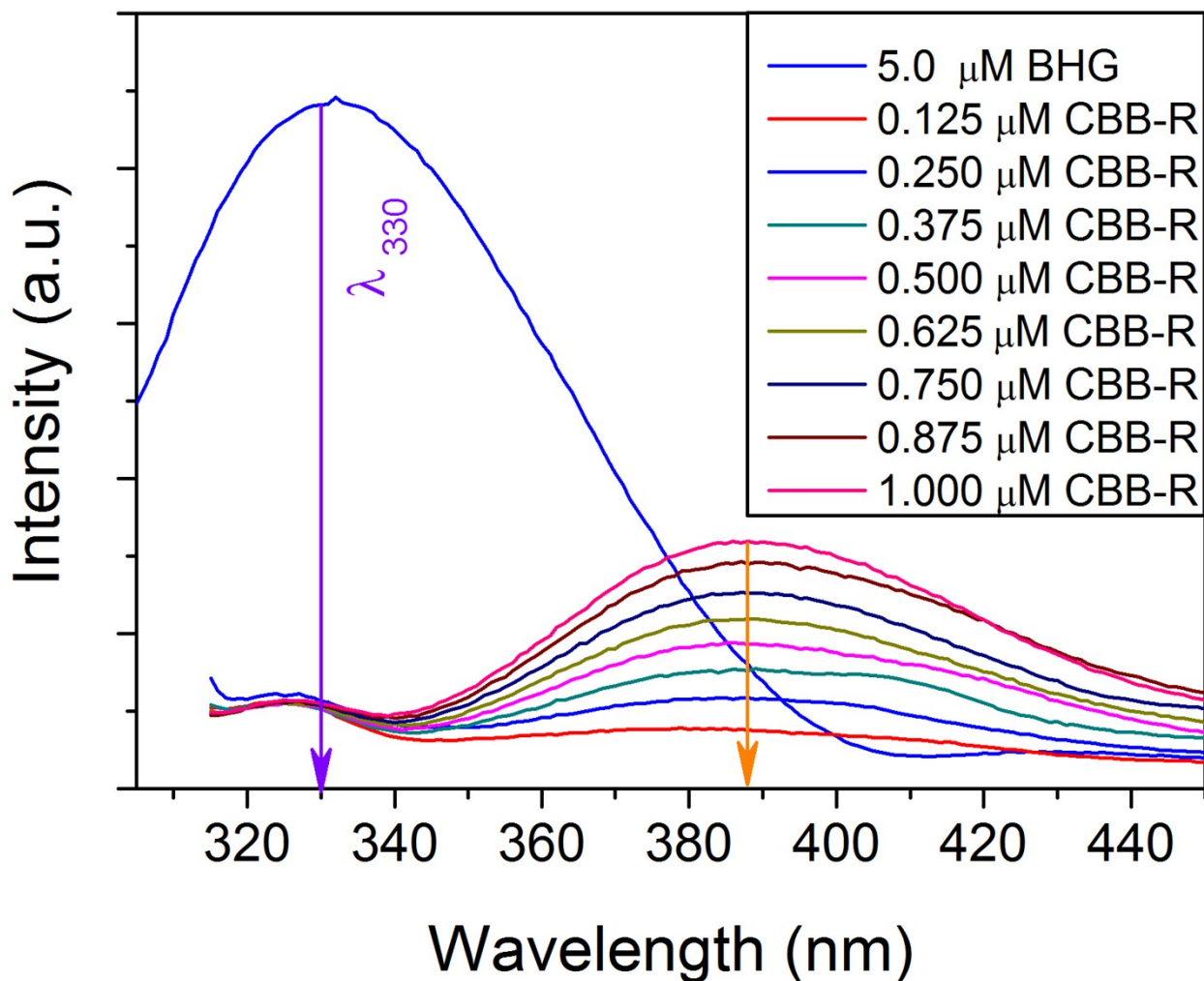
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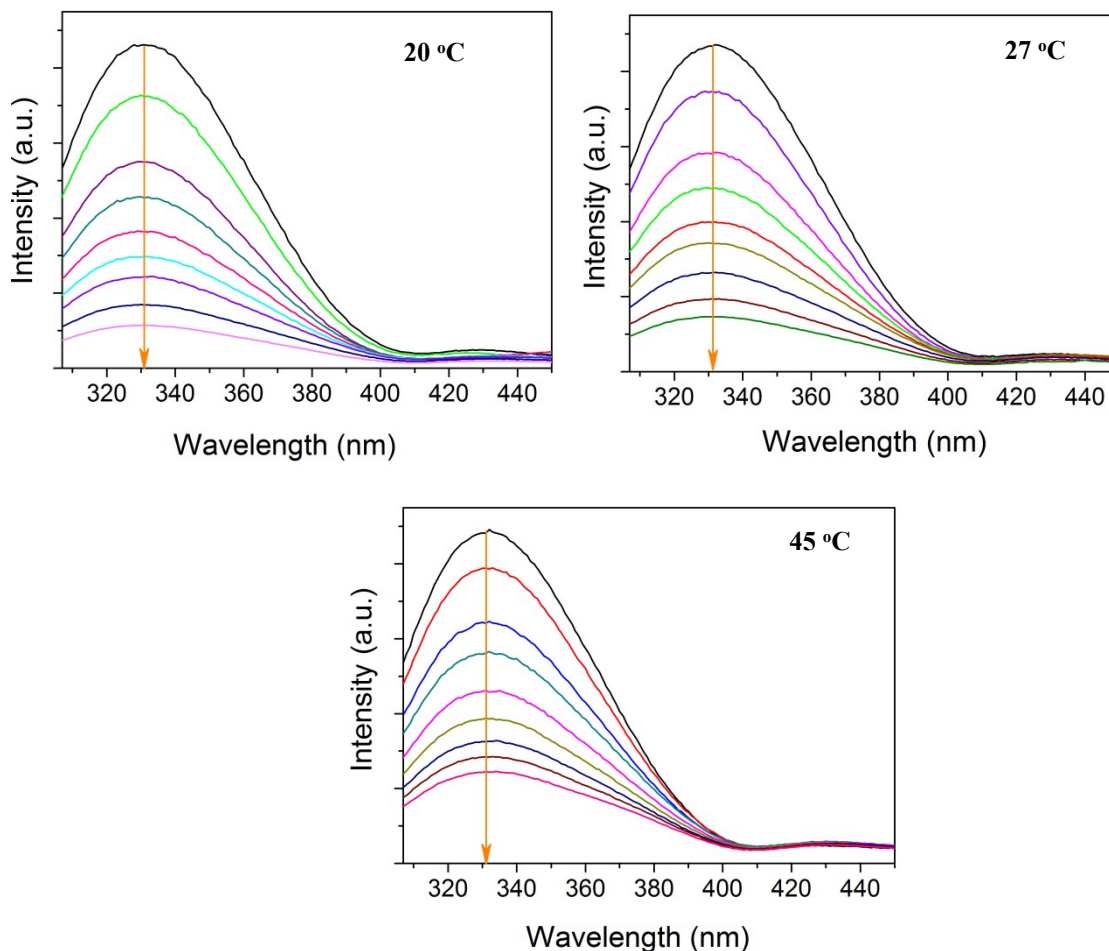
Fluorescence spectra of free BHG (5mM) and free CBB-R



**Figure S1:** The fluorescence emission spectra of free BHG (5mM) and free CBB-R at different concentration. These concentrations of CBB-R were used for quenching study of BHG. From the above spectra it was clear that the contribution of CBB-R fluorescence in BHG fluorescence at 330 nm was minimum even at maximum concentration of CBB-R.

### Normalization and inner-filter correction:

Each and individual fluorescence spectrum of the protein in the presence of different concentration of CBB-R was multiplied with a normalized factor (fluorescence intensity of the protein alone solution at 440 nm divided by fluorescence intensity of the protein/CBB-R complex.) The resultant spectrum was the contribution of CBB-R to the protein/CBB-R complex and it was subtracted from the original spectrum. Then the inner-filter correction was done by using equation 1 (in main manuscript).



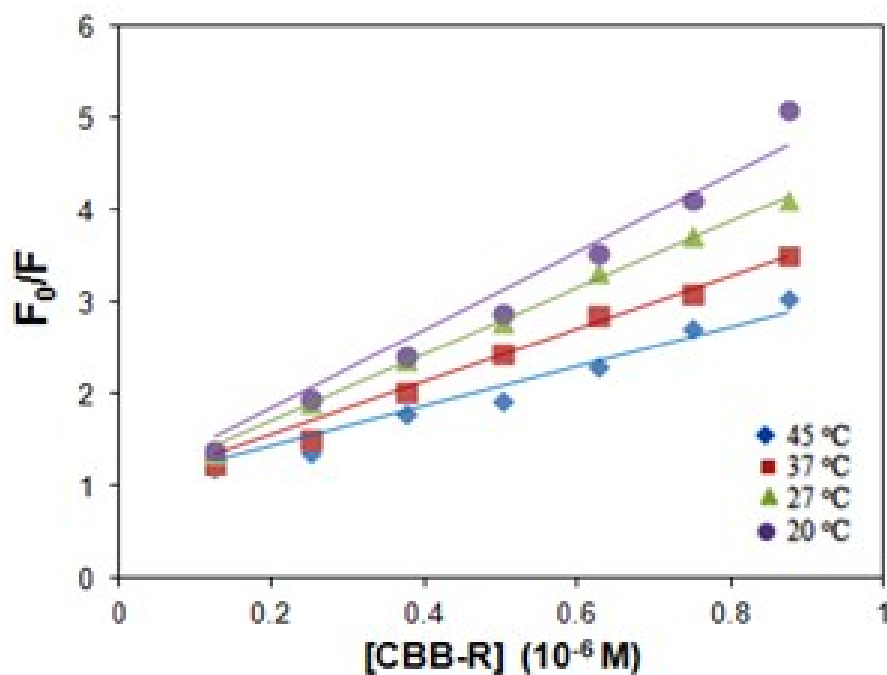
**Figure S2:** Change of fluorescence spectrum of 5.0  $\mu\text{M}$  BHG solutions upon the addition of different concentration of CBB-R in 10.0 mM phosphate buffer at pH 7.4 at other three different temperatures (20 °C, 27 °C and 45 °C), with the increase of CBB-R concentration the intrinsic fluorescence intensity of BHG decreases. The concentration of CBB-R was varied from 0.125  $\mu\text{M}$  to 2.0  $\mu\text{M}$ . The excited wavelength was ( $\lambda_{\text{ex}}$ ) 290 nm and the path length of the cell was 10 mm. All spectrums are normalized by using the emission values at 440 nm and inner-filter corrected.

The plot of  $F_0/F_{\text{corr}}$  against different concentration of CBB-R

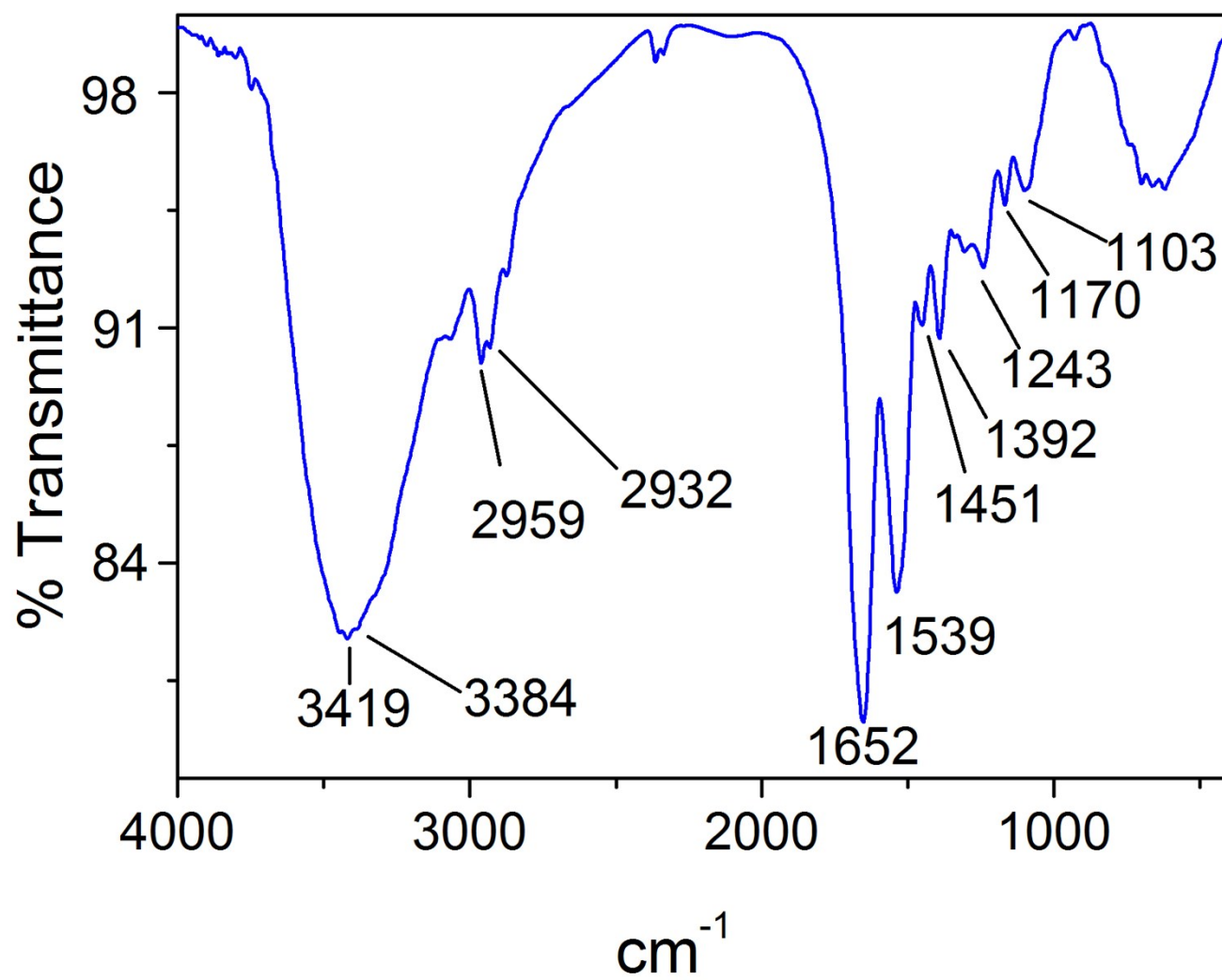
The corrected fluorescence data were analyzed by using Stern-volmer equation.

$$\frac{F_0}{F_{\text{corr}}} = 1 + K_{\text{SV}} [Q]$$

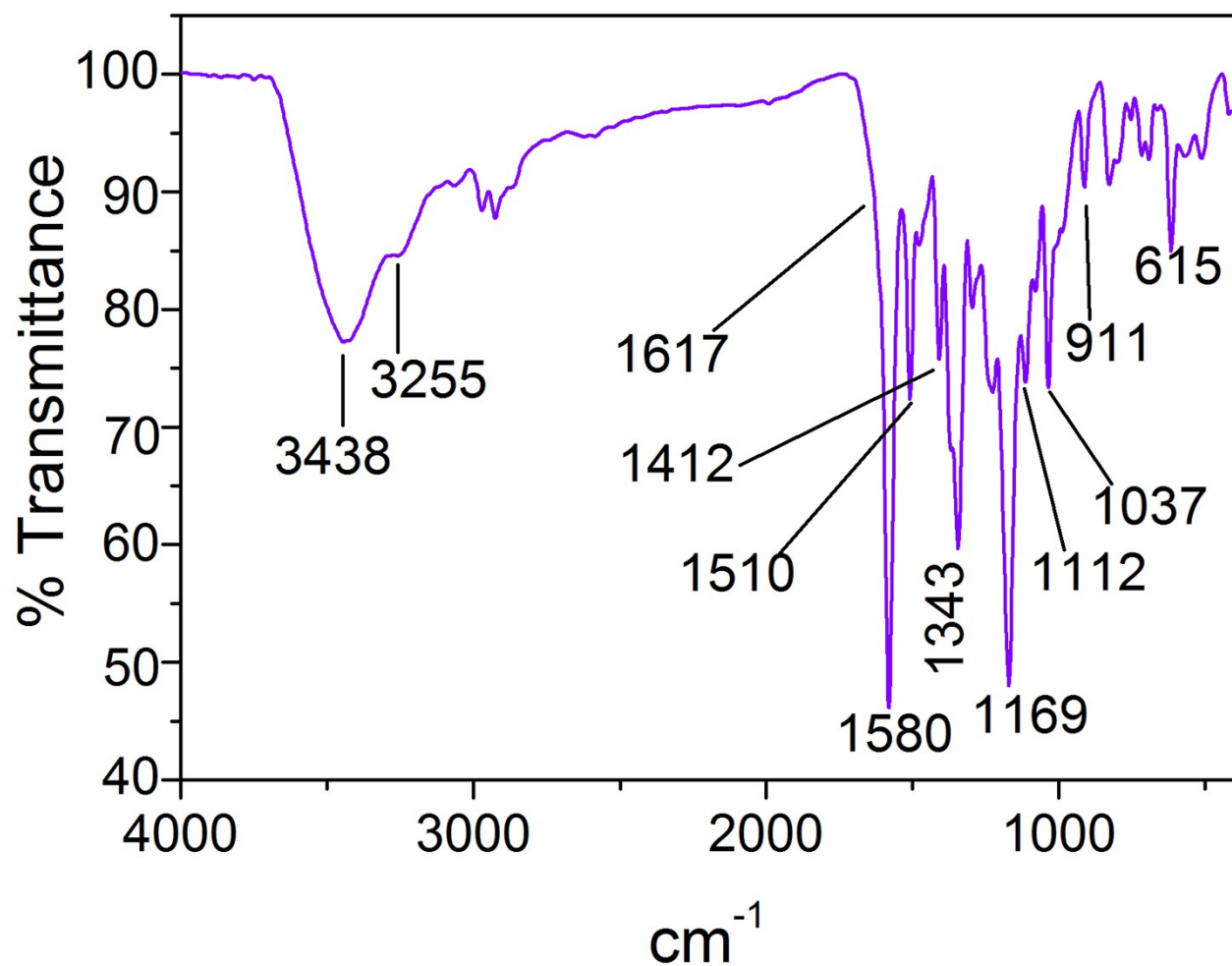
Where  $F_0$  and  $F_{\text{corr}}$  are the fluorescence intensity of BHG in the absence and presence of CBB-R, respectively.  $K_{\text{SV}}$  is the stern-volmer constant and  $[Q]$  is the concentration of CBB-R in the protein solution. Figure S3 shows the fitted Stern–Volmer plots.



**Figure S3:** The panel shows the S–V plot obtained using the fluorescence intensity at 330 nm of BHG and in the presence of the different concentrations of CBB-R at four different temperatures.

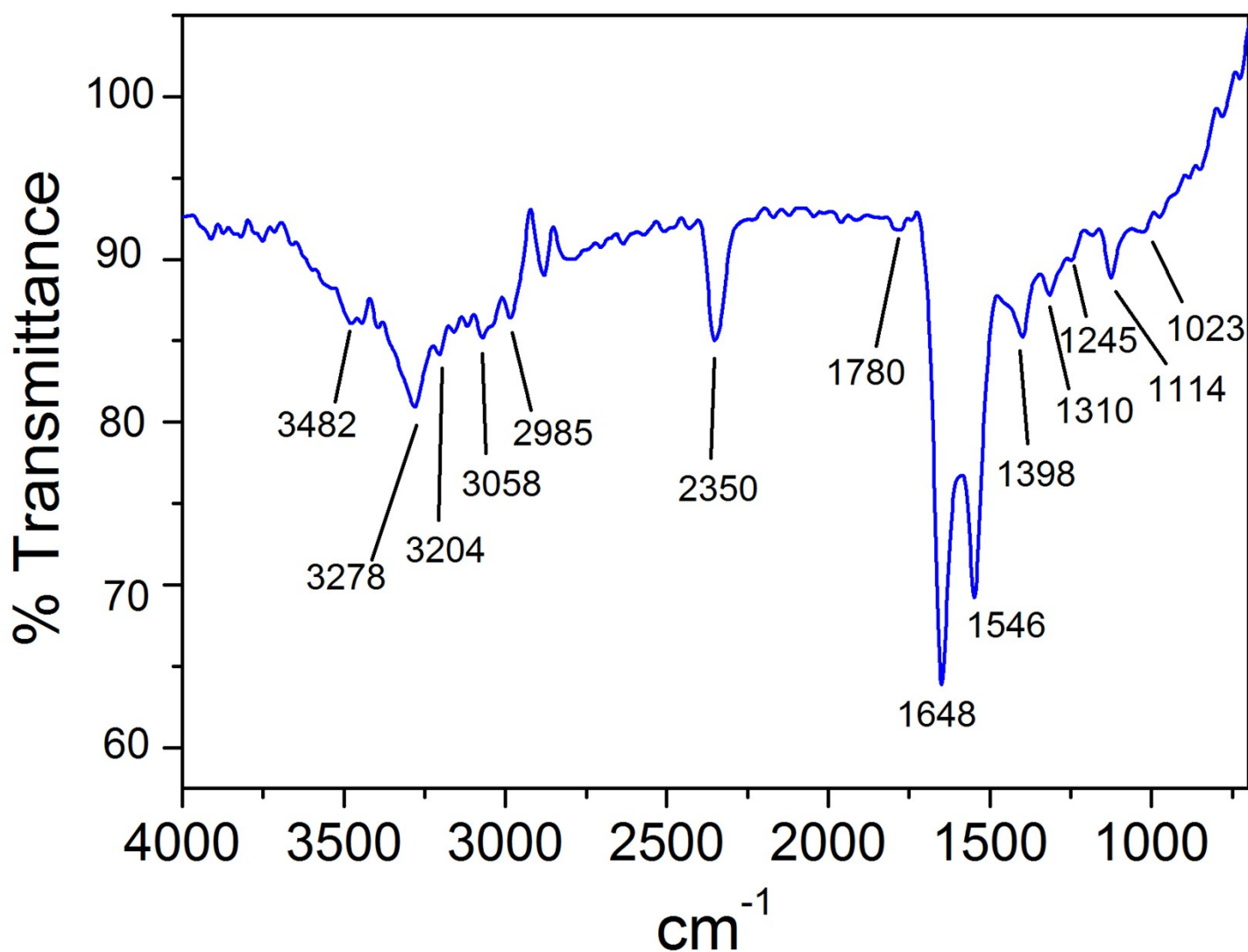


**Figure S4:** FT-IR spectra of bovine hemoglobin collected from KBr pellet techniques

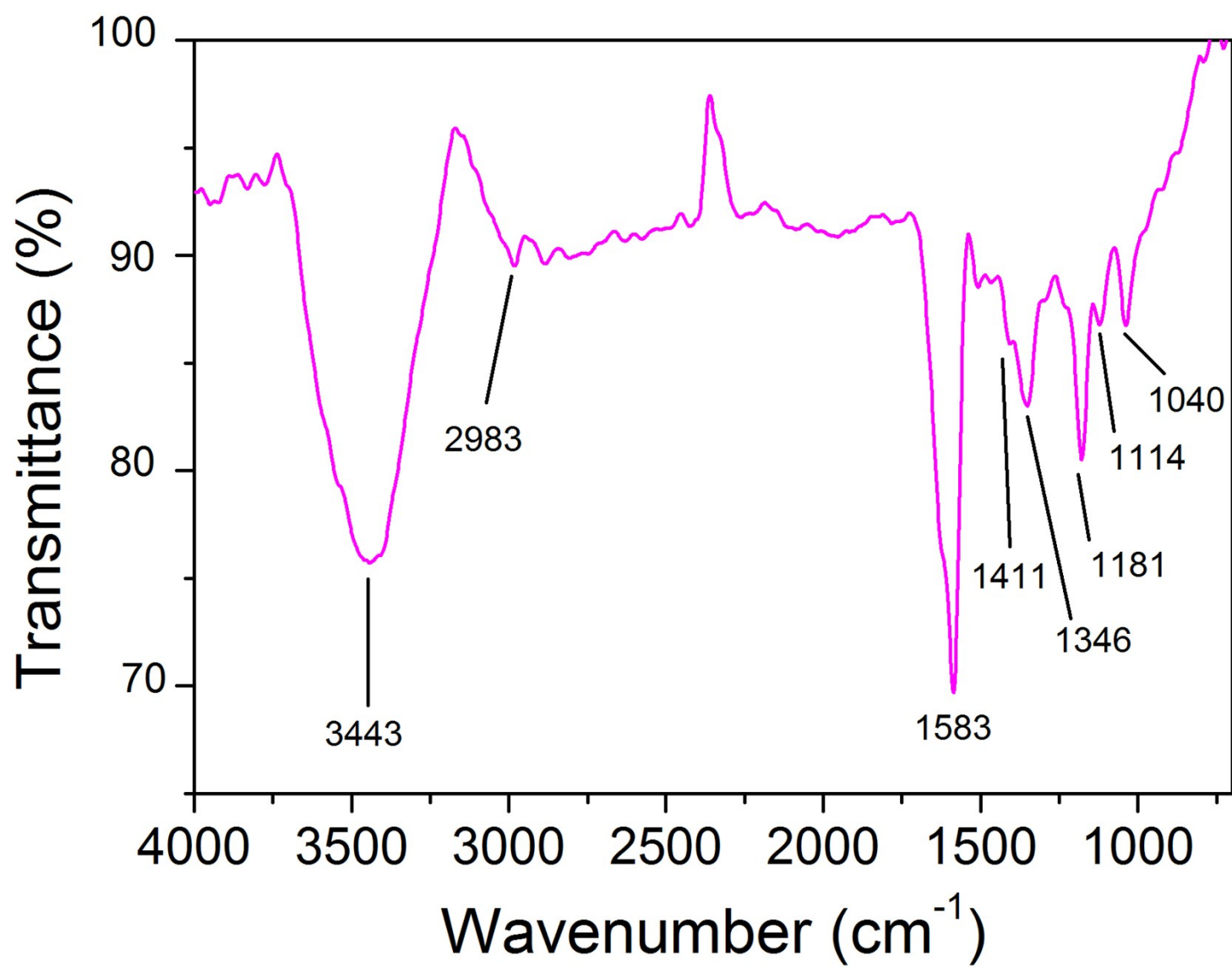


**Figure S5:** FT-IR spectra of coomassie brilliant blue R (CBB-R). The spectrum was recorded using KBr pellet technique.

Full range FT-IR spectra of BHG and CBB-R in aqueous buffer solutions. Different panels represent different solution conditions. All the samples were prepared in 5mM phosphate buffer at pH 7.4. The instrumental base was corrected with same buffer solution.

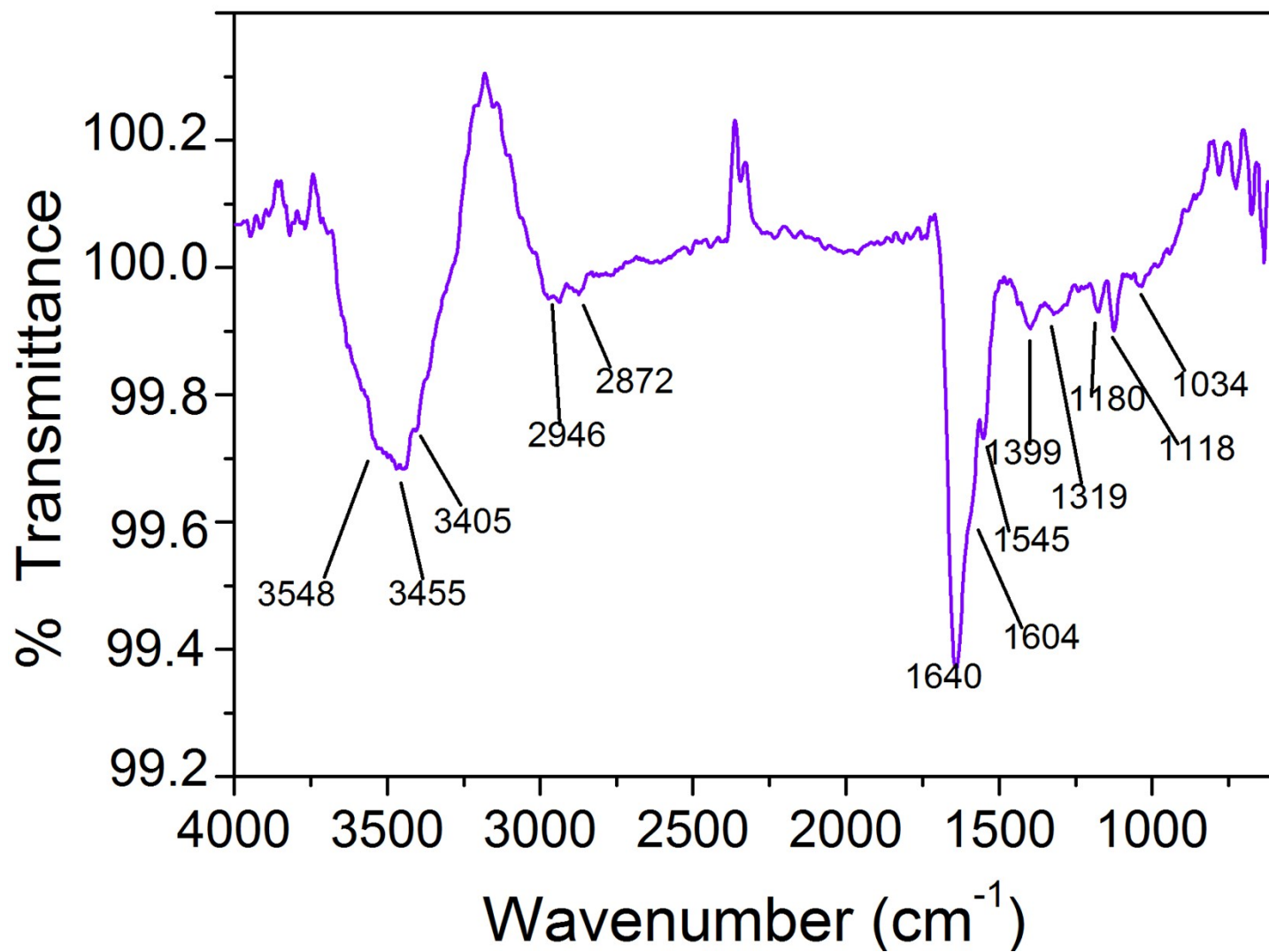


**Figure S6:** panel A. FT-IR spectra of BHG (10 μM) in aqueous buffer solution.

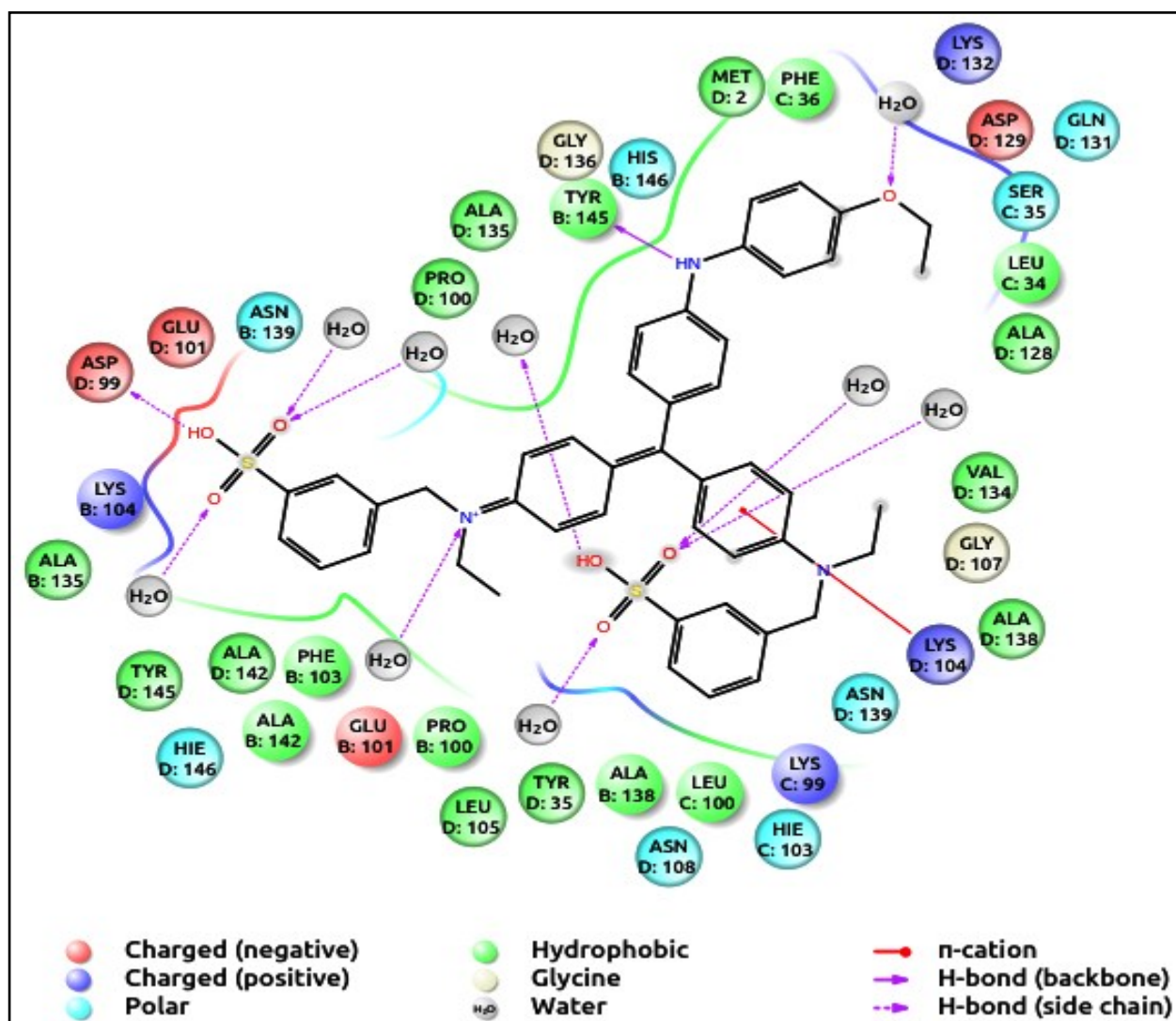


**Figure S6:** Panel B. FT-IR spectra of CBB-R (20.0  $\mu\text{M}$ ) in aqueous buffer solution.

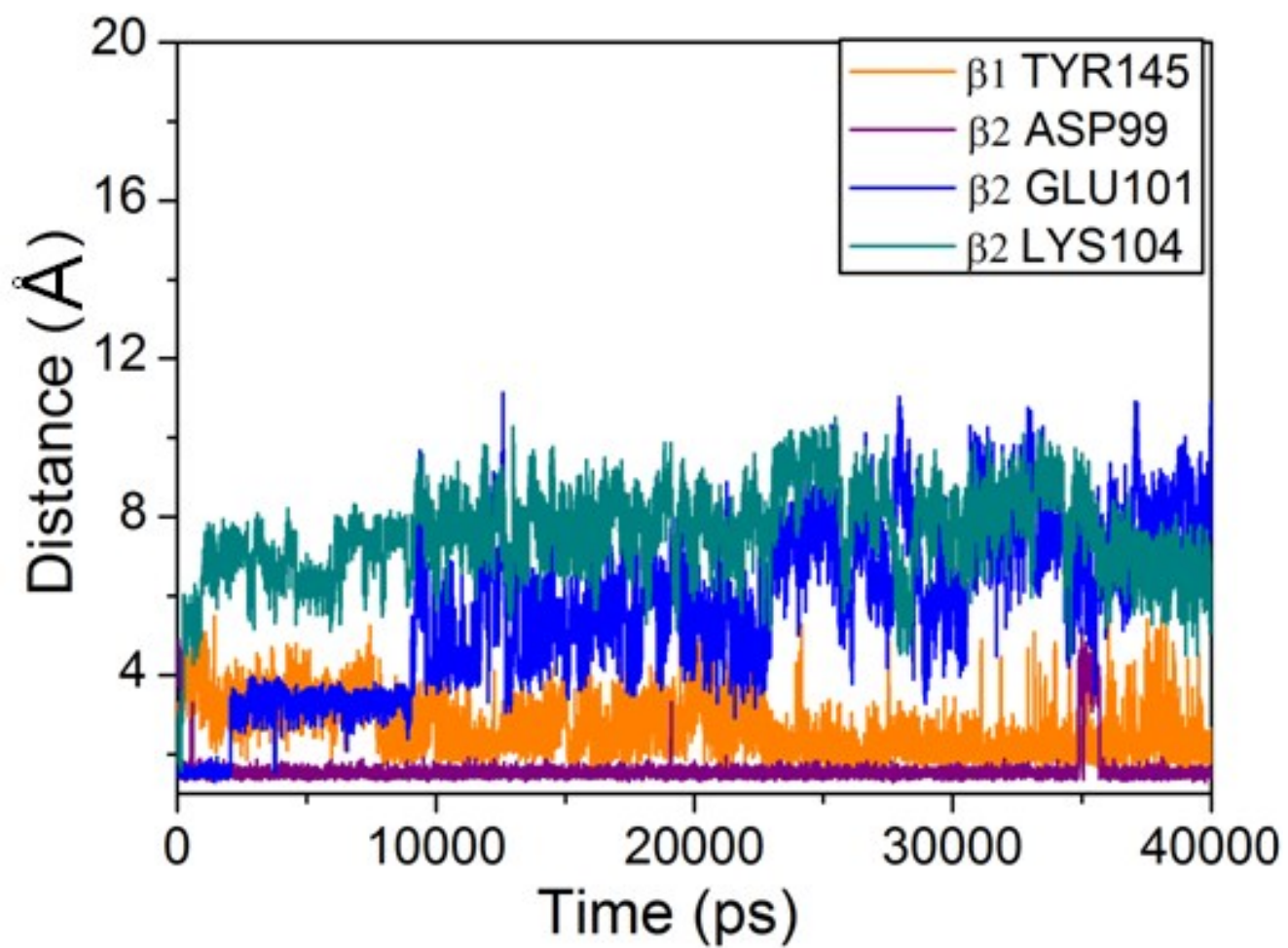




**Figure S6:** Panel C. FT-IR spectra of BHG (10 $\mu$ M) in the presence of CBB-R (7.5  $\mu$ M) in aqueous buffer solution.



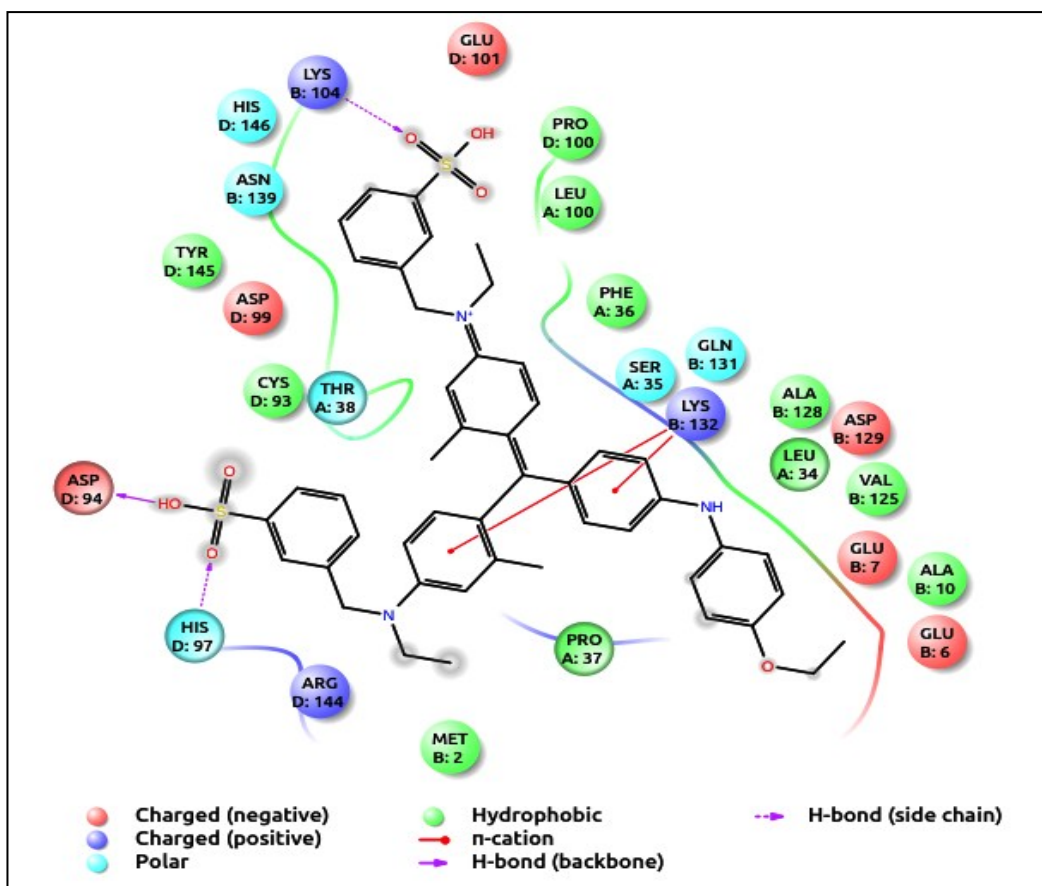
**Figure S7:** 2D plot of CBB-R with BHG complex observed after 40 ns of MD simulation. The H-bonds between Asp99 of  $\beta 2$  subunit, Tyr145 of  $\beta 1$  subunits, and  $\pi$ -cation interactions between Lys104 of  $\beta 1$  subunits (the notation as A, B, C and D below the three letter code of amino acids indicates  $\alpha 1$ ,  $\beta 1$ ,  $\alpha 2$  and  $\beta 2$  subunit, respectively).



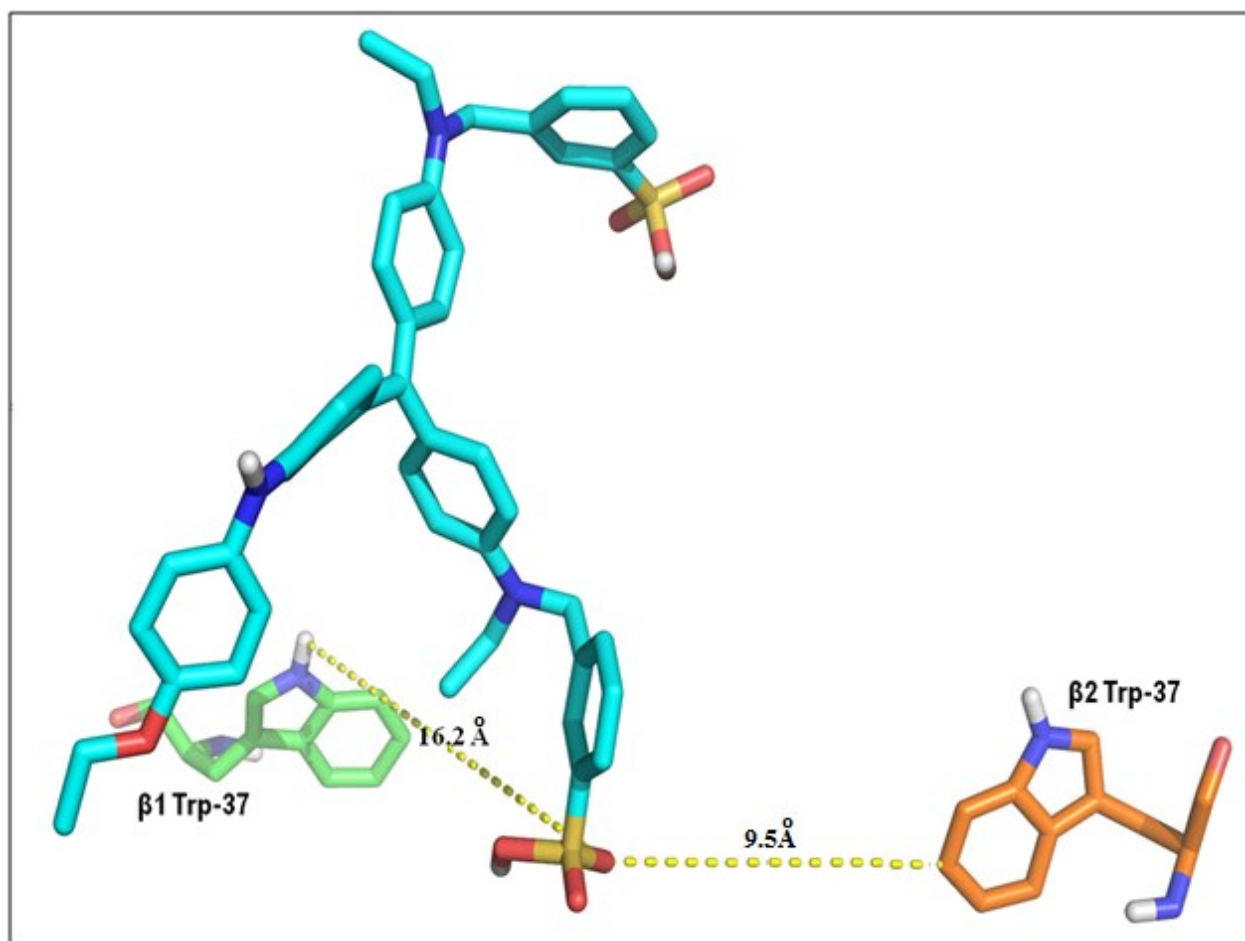
**Figure S8:** Hydrogen bonding donar-acceptor distances during 40000 ps MD simulation.

## Molecular docking of CBB-G with BHG:

The  $\Delta G^0$  value of best energy conformer as docked was  $-26.33 \text{ kJ mol}^{-1}$ . The best energy ranked structure revealed that CBB-G located near the cavity formed among the  $\alpha_1$ ,  $\beta_1$  and  $\beta_2$  subunits. It was observed that CBB-G was stabilized by hydrogen bond interaction with Lys104 of  $\beta_1$  subunit, Asp94 and His97 of  $\beta_2$  subunits of the protein. The two  $\pi$ -cation interactions were also observed with Lys104 of  $\beta_1$  subunit.



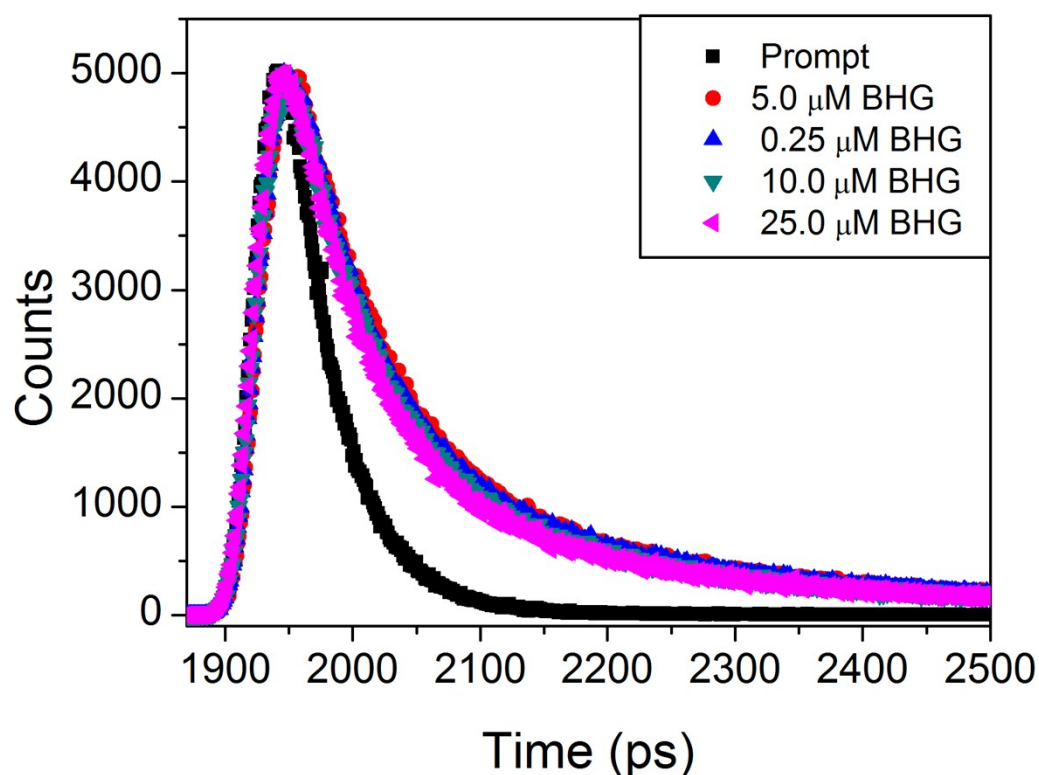
**Figure S9:** 2D plot of docking conformation observed during docking simulation. The H-bonds between Lys104 of  $\beta_1$  subunit, His97 and Asp94 of  $\beta_2$  subunit, and two  $\pi$ -cation interactions between Lys132 of  $\beta_1$  subunit (the notation as A, B, C and D below the three letter code of amino acids indicates  $\alpha_1$ ,  $\beta_1$ ,  $\alpha_2$  and  $\beta_2$  subunit, respectively).



**Figure S10:** The distance between CBB-R and subunit of  $\beta 1$  Trp37 and  $\beta 2$  Trp37 after 40 ns MD simulation, presented by stick model using pymol.

### Time-resolved fluorescence measurements:

Fluorescence lifetime measurement was performed by time correlated single-photon counting (TCSPC) method on FluoroCube-01-NL spectrometer (Horiba Jobin Yvon) using a Laser-Diode has an output of 375 nm, The signals were collected at the magic angle at  $54.7^\circ$  and the details of the method are given elsewhere.<sup>1 2</sup> The Excitation wavelength was 290 nm.



**Figure S11:** Time resolved fluorescence decay of BHG in solution and in the presence of CBB-R. ‘Prompt’ is the laser pulse profile.

**Table TS1:** Fluorescence lifetime of BHG as a function of concentrations of CBB-R

samples	$\tau_1(\text{ns})$	$\tau_2(\text{ns})$	$\tau_3(\text{ns})$	$A_1$	$A_2$	$A_3$	$\tau$ (ns)	$\chi^2$
BHG	1.2	5.4	0.08	0.05	0.01	0.94	~0.2	1.07
BHG:CBB-R (1:2)	1.3	5.4	0.08	0.04	0.01	0.95	~0.2	1.01

### References:

1. B. K. Paul and N. Guchhait, *J. Phys. Chem. B*, 2011, 115, 10322-10334.
2. W. Peng, F. Ding, Y.-K. Peng and Y. Sun, *Mol. BioSyst.*, 2014, 10, 138-148.