

## Supporting Information for:

### 5 Substrate channel evolution of an esterase for the synthesis of Cilastatin

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### 15 Contents

**Table S1.** Primers used for site-directed mutagenesis.

**Table S2.** Data collection and refinement statistics of *RhEst1*.

**Fig. S1** <sup>1</sup>H NMR spectrum of product (S)-(+)-DmCpCa.

**Table S1.** Primers used for site-directed mutagenesis.

Library	Primers	Oligonucleotide sequences (5'-3') <sup>a</sup>
Library A		
	I128-F	TGGTCGGGGCG <u>NDT</u> ACCAGC <u>NDT</u> GGGCGCG
	I128-R	CGCGCCCA <u>HNG</u> CTGGTA <u>HNC</u> GCCCCGACCA
	I131-F	TGGTCGGGGCG <u>NDT</u> ACCAGC <u>NDT</u> GGGCGCG
	I131-R	CGCGCCCA <u>HNG</u> CTGGTA <u>HNC</u> GCCCCGACCA
	G138-F	GCGAAAAAGGT <u>NNK</u> AAGGTGGCTGGCTGGCGA
	G138-R	TCGCCGAGCCGACCTT <u>MNN</u> ACCTTTTCGC
	V140-F	GCGAAAAAGGTGGCAAG <u>NNK</u> GGCTGGCGA
	V140-R	GCGCTACGCATGCCGAGCC <u>MNN</u> CTGCCACCTTTT
	M144-F	TGGCAAGGTGGCTCGGC <u>NNK</u> CGTAGCGCAGTCCC
	M144-R	GCCGGGAACTGCGTACGMNNCGCCGAGCCGACCTTGCCA
	A147/V148-F	TGCGTAGC <u>NDT</u> CCGGCGCGATGTCCG
	A147/V148-R	GCCGGGA <u>HNA</u> HNGCTACGCATGCCGAGCC
	F200-F	GGGTGCGCGCAGCACTG <u>NNK</u> AACCGTGCTGTCGG
	F200-R	TCGTGACCGACAGCACGGT <u>MNN</u> CAGTGCTGCGCG
	V227/V228-F	CACGGAACCGACGACTC <u>NDT</u> NTGACGTG
	V227/V228-R	TACCTGCGTCACGT <u>CAHNA</u> HNGGAGTCGT
Library B		
	W33/A34-F	GGTGTGCTGCACGGCTGG <u>NDT</u> NTTCTCGCAG
	W33/A34-R	CCAGCACTGCGAGGA <u>AHN</u> CCAGCCGTG
	W100-F	ACGCGATCCTGCTCGC <u>NNK</u> CGTACGGCG
	W100-R	ATCACCA <u>GGC</u> GTACGAMNNNGCCGAGC
	F166-F	CGTGC <u>ACTCG</u> CGCT <u>NNK</u> GGCAATGCTCTCACC
	F166-R	GGTGAGAGCATTGCC <u>MNN</u> AGCGCCGAGTCACG
	L170-F	GCTTCGGCAATGCT <u>NNK</u> ACCGGCCACCGA
	L170-R	TCGGGTGGGCCGGT <u>MNN</u> AGCATTGCCAAAGC
	L184-F	GGGTGCGTC <u>GGC</u> AGGCC <u>NNK</u> TTGGTA
	L184-R	CGAGAGGCTGTACCCGAAMNNNGCCTGCGA

<sup>a</sup> The nucleotide sequences corresponding to the mutated amino acids are underline.

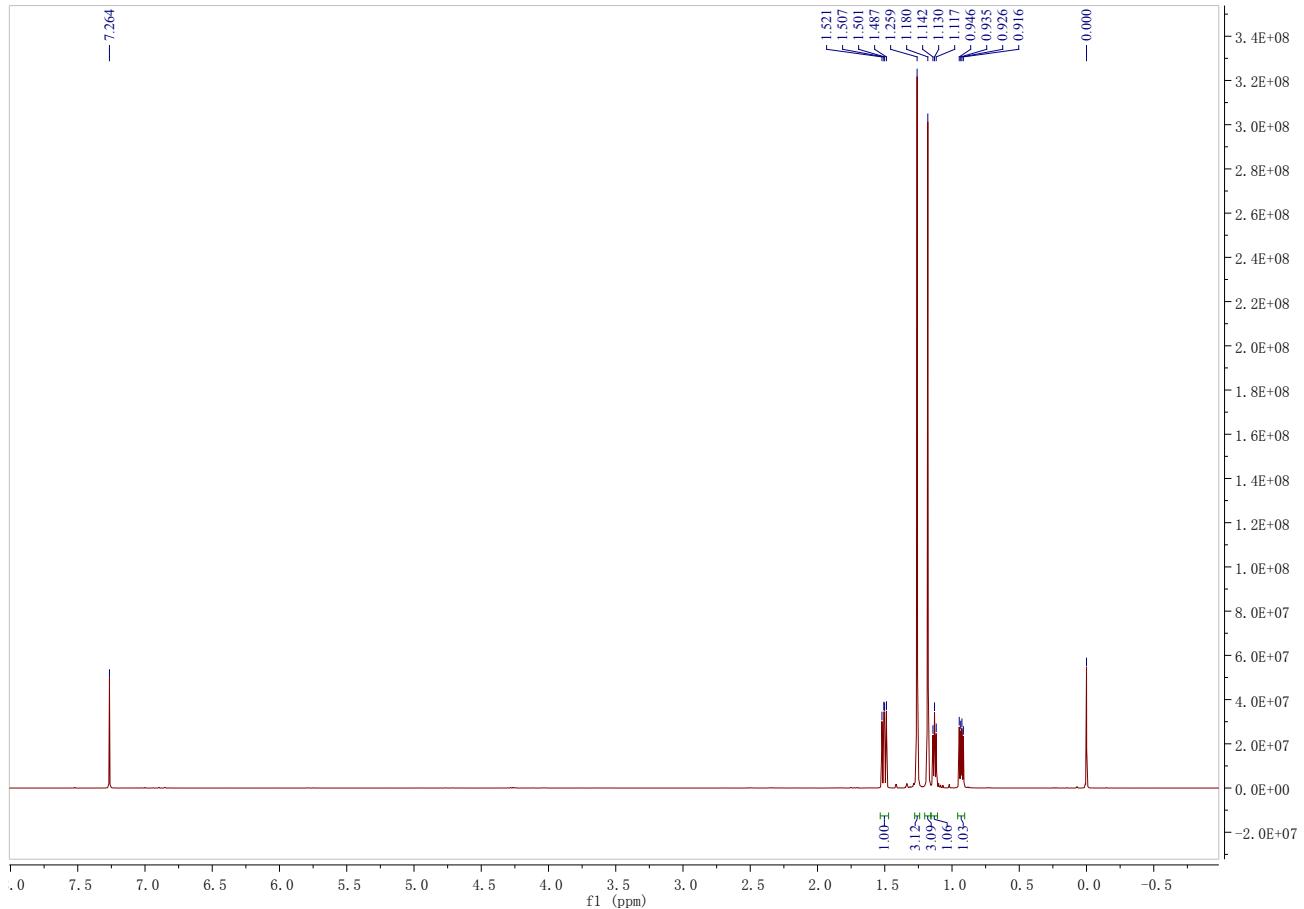
**Table S2.** Data collection and refinement statistics of *RhEst1*.

<b>Data collection</b>	<b><i>RhEst1-native</i></b>	<b><i>Se-RhEst1</i></b>
Wavelength (Å)	1.5418	0.98
Space group	<i>C</i> <sub>2</sub>	<i>P</i> <sub>3121</sub>
a (Å)	213	96.0
b (Å)	45.4	96.0
c (Å)	77.4	206
α, β, γ (°)	90, 106, 90	90, 90, 120
Resolution range (Å) <sup>a</sup>	44.3-1.95 (2.02-1.95)	35.1-2.40 (2.49-2.40)
Total/Unique reflections	52476 / 13119	43537 / 3721
Redundancy <sup>a</sup>	4 (3.8)	11.7 (11.9)
Average (I/δ) <sup>a</sup>	15.33 (2.3)	46.27 (13.4)
Completeness (%) <sup>a</sup>	100 (100)	99.9 (100)
Rmerge (%) <sup>b</sup>	0.043 (0.594)	0.082 (0.514)
<b>Refinement</b>	<b><i>RhEst1-native</i></b>	<b><i>Se-RhEst1</i></b>
Resolution range (Å)	44.35-1.95	35.09-2.40
No. of reflections	49674	42836
R-factor <sup>c</sup> , R <sub>free</sub> (%) <sup>c</sup>	0.1650, 0.2155	0.2504, 0.2861
No. of amino acid residues	830	539
No. of water	592	291
Rmsd bond lengths (Å)	0.007	0.010
Rmsd bond angles (°)	1.059	1.303
Favored (%)	95.9	96.61
Allowed (%)	3.5	2.64
Disallowed (%)	0.6	0.75

<sup>a</sup> Numbers in parentheses are values for the highest-resolution shell.

<sup>b</sup> R<sub>merge</sub> =  $\sum_{hk\ell} |I_i - I_m| / \sum_{hk\ell} I_m$ , where  $I_i$  and  $I_m$  are the observed intensity and the mean intensity of related reflections, respectively. Values in parentheses indicate high-resolution shell.

<sup>c</sup> R-factor =  $\sum ||F_o - F_c|| / \sum ||F_o||$ , R<sub>free</sub> =  $\sum_T ||F_o - F_c|| / \sum_T ||F_o||$ , where T is a test data set of 5% of the total reflections randomly chosen and set aside prior to refinement.



$^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ),  $\delta/\text{ppm}$ : 0.92-0.95 (m, 1H, CH), 1.11-1.14 (t, 1H, CH,  $J = 5.1$  Hz), 1.18 (s, 3H,  $\text{CH}_3$ ), 1.26 (s, 3H,  $\text{CH}_3$ ), 1.48-1.52 (m, 1H, CH).

**Fig. S1**  $^1\text{H}$  NMR spectrum of product (*S*)-(+)-DmCpCa.