

Palladium(II) complexes with electron-poor, 4,5-disubstituted diimidazol-2-ylidene ligands: synthesis, characterization and catalytic activity.

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Supporting Information

Additional X-ray crystallographic details pp. S2-S4

NMR spectra of the compounds pp. S5-S20

Structure Determination

Preliminary examination and data collection were carried out on a NONIUS κ -CCD diffraction system equipped with an Oxford Cryosystems cooler at the window of a fine-focus sealed tube using graphite-monochromated Mo K α radiation ($\lambda = 0.71073 \text{ \AA}$).

The reflections were merged and corrected from Lorentz, polarization and decay effects. An absorption correction was applied using SADABS.¹ The structures were solved by a combination of direct methods^{2,3} with the aid of difference Fourier synthesis and were refined against all data using SHELXL-97.⁴ Hydrogen atoms were assigned to calculated positions and then refined using the SHELXL-97 riding model. All non-hydrogen atoms were refined with anisotropic displacement parameters. Full-matrix least-squares refinements were carried out by minimizing $\Sigma w(F_o^2 - F_c^2)^2$ with the SHELXL-97 weighting scheme. Details of the structure determinations are given in the Supporting Information. Neutral-atom scattering factors for all atoms and anomalous dispersion corrections for the non-hydrogen atoms were taken from the International Tables for Crystallography. All calculations were performed with the programs COLLECT,⁵ DIRAX,⁶ EVALCCD,⁷ SIR92,² SIR97,³ SHELXLE,⁸ SADABS,¹ the SHELXL-97 package,⁴ and ENCIFER.⁹ Images of the solid state structures were generated with ORTEP-3.¹⁰

References

- (1) Sheldrick, G. M. *SADABS*, Version 2.10; University of Goettingen, Goettingen, Germany, 2002.
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- (3) Altomare, A.; Burla, M. C.; Camalli, M.; Cascarano, G. L.; Giacovazzo, C.; Guagliardi, A.; Moliterni, A. G. G.; Polidori, G.; Spagna, R. J. *Appl. Crystallogr.* **1999**, 32, 115-119.
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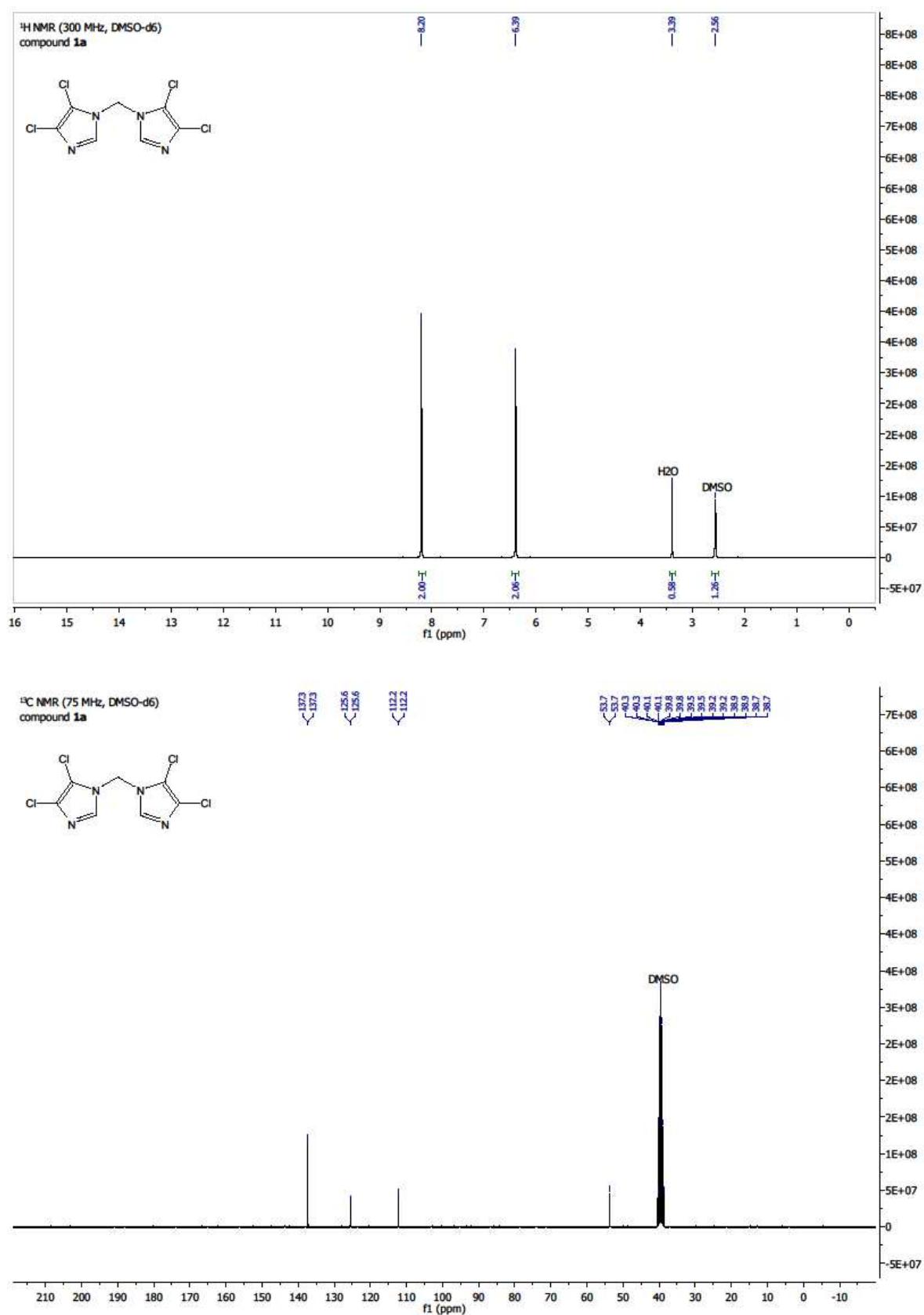
Table S1. Crystallographic Data for Compounds **3a** and **3b**.

	3a	3b		
empirical formula	C ₂₉ H ₂₇ Br ₆ Cl ₁₂ N ₁₃ Pd ₃	C ₁₇ H ₂₆ Br ₂ Cl ₄ N ₆ O ₂ Pd		
formula weight	1781.70	754.46		
temperature (K)	198(2)	198(2)		
wavelength (Å)	0.71073	0.71073		
crystal system	orthorhombic	triclinic		
space group	Pbca	P-1		
unit cell dimensions (in Å and °)	a = 19.767(5) b = 15.478(10) c = 34.001(9)	α = 90 β = 90 γ = 90	a = 8.0460(14) b = 11.572(2) c = 15.7320(13)	α = 72.144(9) β = 89.760(14) γ = 74.103(15)
volume (in Å ³)	10403(8)	1336.0(4)		
Z	8	2		
density (g/cm ³ , calculated)	2.275	1.875		
absorptioncoeff. (mm ⁻¹)	6.292	4.112		
F(000)	6752	740		
crystal size (mm)	0.48 x 0.18 x 0.17	0.73 x 0.53 x 0.38		
θ range for data collection (°)	2.06 to 23.25	2.64 to 26.40		
index ranges	-21 ≤ h ≤ 21 -17 ≤ k ≤ 17 -37 ≤ l ≤ 37	-9 ≤ h ≤ 10 -14 ≤ k ≤ 13 -19 ≤ l ≤ 18		
reflections collected	177002	13850		
independent reflections	7462 [R(int) = 0.1558]	4963 [R(int) = 0.0242]		
absorption correction	semi-empirical from equivalents	semi-empirical from equivalents		
refinement method	full-matrix least-squares on <i>F</i> ²	full-matrix least-squares on <i>F</i> ²		
data/restraints/parameters	7462/0/575	4963/0/295		
goodness of fit on <i>F</i> ²	1.077	1.159		
final R indices [<i>I</i> >2σ(<i>I</i>)]	R1 = 0.0423, wR2 = 0.1044	R1 = 0.0341, wR2 = 0.0746		
R indices (all data)	R1 = 0.0849, wR2 = 0.1297	R1 = 0.0585, wR2 = 0.0809		
largest diff. peak and hole (e·Å ⁻³)	0.1134 and -1.119	0.590 and -0.521		

Table S2. Crystallographic Data for Compounds **3c** and **3e**.

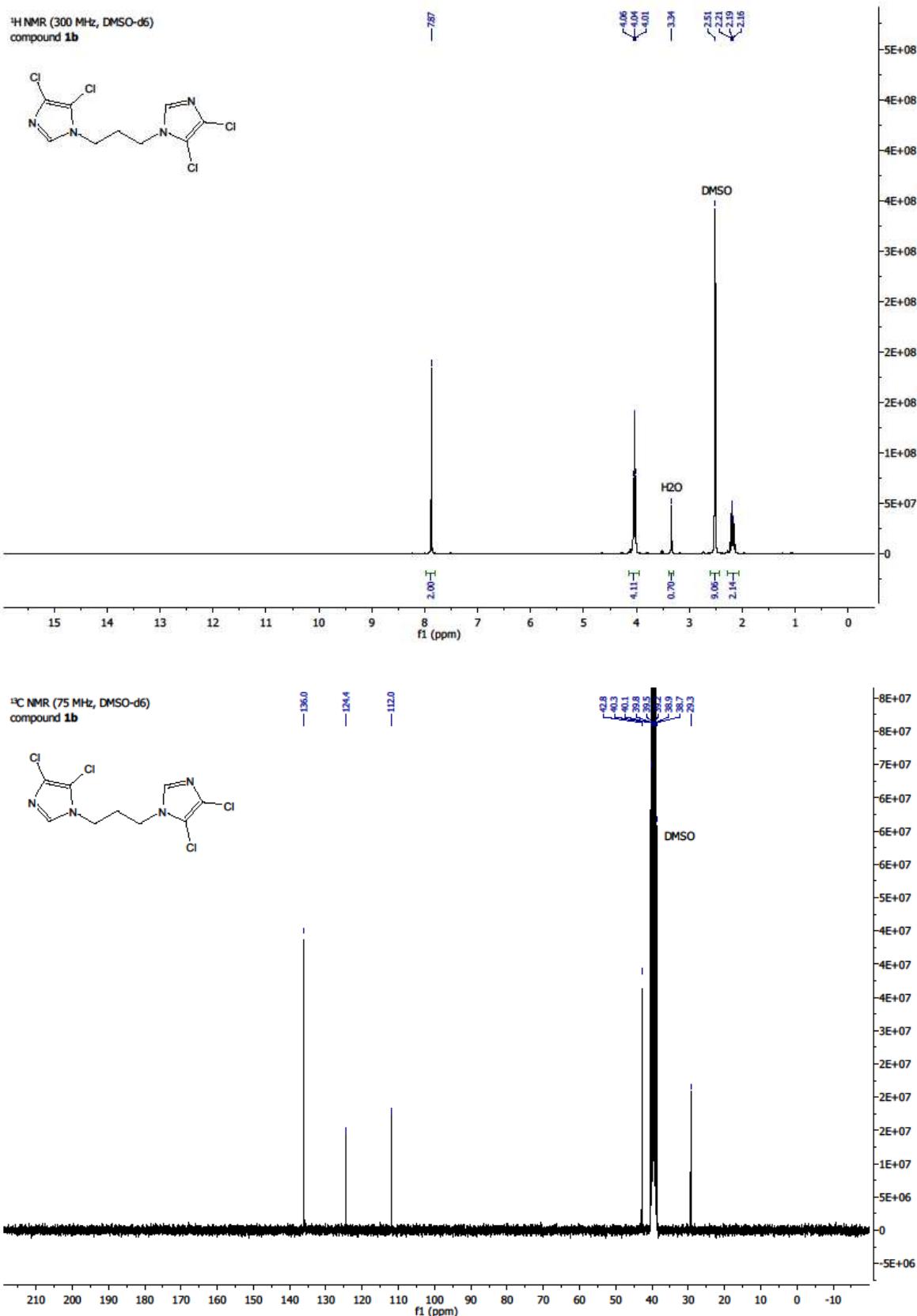
	3c	3e
empirical formula	C ₁₆ H ₁₄ Br ₂ Cl ₄ N ₄ Pd	C ₁₅ H ₁₂ Br ₂ N ₈ Pd
formula weight	670.33	570.55
temperature (K)	198(2)	198(2)
wavelength (Å)	0.71073	0.71073
crystal system	orthorhombic	tetragonal
space group	<i>Pnma</i>	<i>I41cd</i>
unit cell dimensions (in Å and °)	a = 13.430(3) b = 18.758(2) c = 8.310(6)	α = 90 β = 90 γ = 90
volume (in Å ³)	2093.5(16)	7833.5(16)
Z	4	16
density (g/cm ³ , calculated)	2.127	1.935
absorptioncoeff. (mm ⁻¹)	5.225	5.044
F(000)	1288	4384
crystal size (mm)	0.53 x 0.36 x 0.24	0.25x 0.22 x 0.22
θ range for data collection (°)	2.17 to 23.25	3.21 to 25.20
index ranges	-16≤ <i>h</i> ≤ 16 -23≤ <i>k</i> ≤ 23 -10≤ <i>l</i> ≤ 10	-28≤ <i>h</i> ≤ 28 -28≤ <i>k</i> ≤ 28 -16≤ <i>l</i> ≤ 16
reflections collected	43521	72663
independent reflections	2215 [<i>R</i> (int) = 0.0669]	3514 [<i>R</i> (int) = 0.0793]
absorption correction	semi-empirical from equivalents	semi-empirical from equivalents
refinement method	full-matrix least-squares on <i>F</i> ²	full-matrix least-squares on <i>F</i> ²
data/restraints/parameters	2215/0/125	3514/1/237
goodness of fit on <i>F</i> ²	1.084	1.056
final R indices [<i>I</i> >2σ(<i>I</i>)]	<i>R</i> 1 = 0.0299, w <i>R</i> 2 = 0.0608	<i>R</i> 1 = 0.0331, w <i>R</i> 2 = 0.0736
R indices (all data)	<i>R</i> 1 = 0.0472, w <i>R</i> 2 = 0.0659	<i>R</i> 1 = 0.0394, w <i>R</i> 2 = 0.0764
largest diff. peak and hole (e·Å ⁻³)	0.621 and -0.643	0.625 and -0.355

1a



¹H-NMR (300 MHz, DMSO-d₆) δ 8.20 (s, 2H), 6.39 (s, 2H).

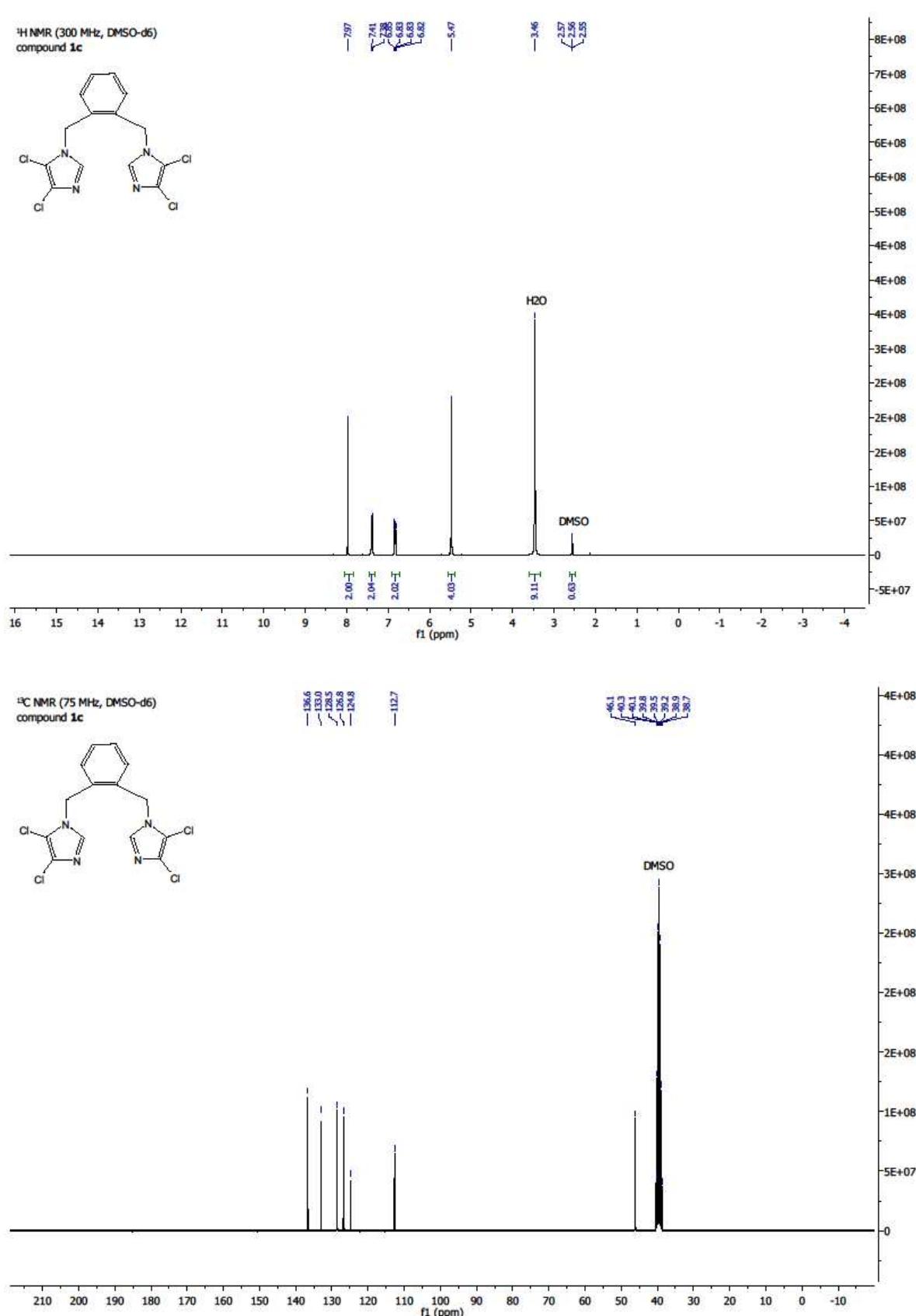
¹³C NMR (75 MHz, DMSO) δ 137.3 (s), 125.57 (s), 112.2 (s), 53.8 (s).

1b

¹H-NMR (300 MHz, DMSO) δ 7.87 (s, 2H), 4.04 (t, *J* = 7.2 Hz, 4H), 2.29 (t, *J* = 7.2 Hz, 2H).

¹³C-NMR (75 MHz, DMSO) δ 136.0 (s), 124.4 (s), 112.0 (s), 42.8 (s), 29.3 (s).

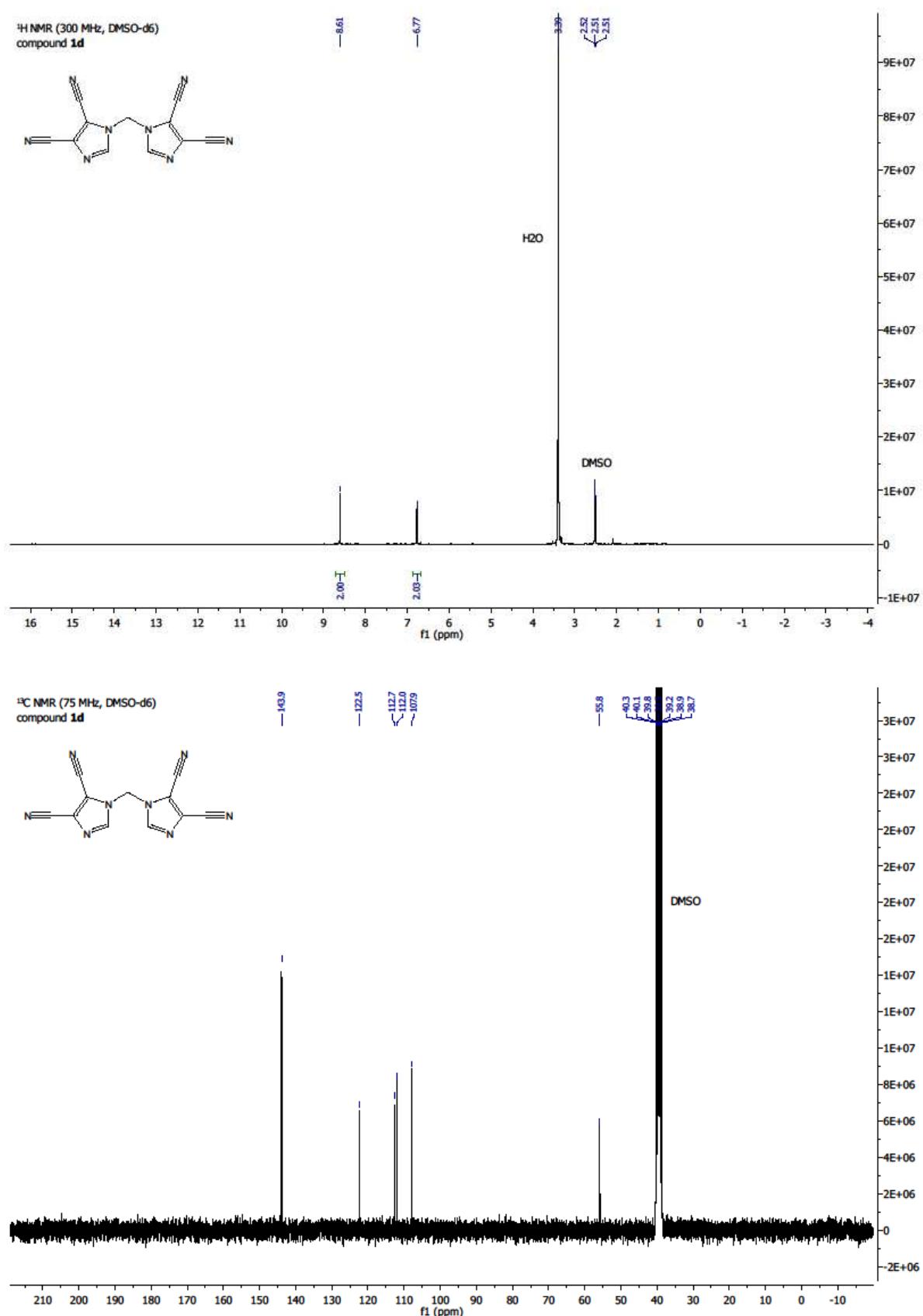
1c



¹H-NMR (300 MHz, DMF) δ 7.97 (s, 2H), 7.39 (dd, *J* = 5.7, 3.4 Hz, 2H), 6.83 (dd, *J* = 5.6, 3.4 Hz, 2H), 5.47 (s, 4H).

¹³C-NMR (75 MHz, DMSO) δ 136.6 (s), 133.0(s), 128.52 (s), 126.8 (s), 124.8 (s), 112.7 (s), 46.1 (s).

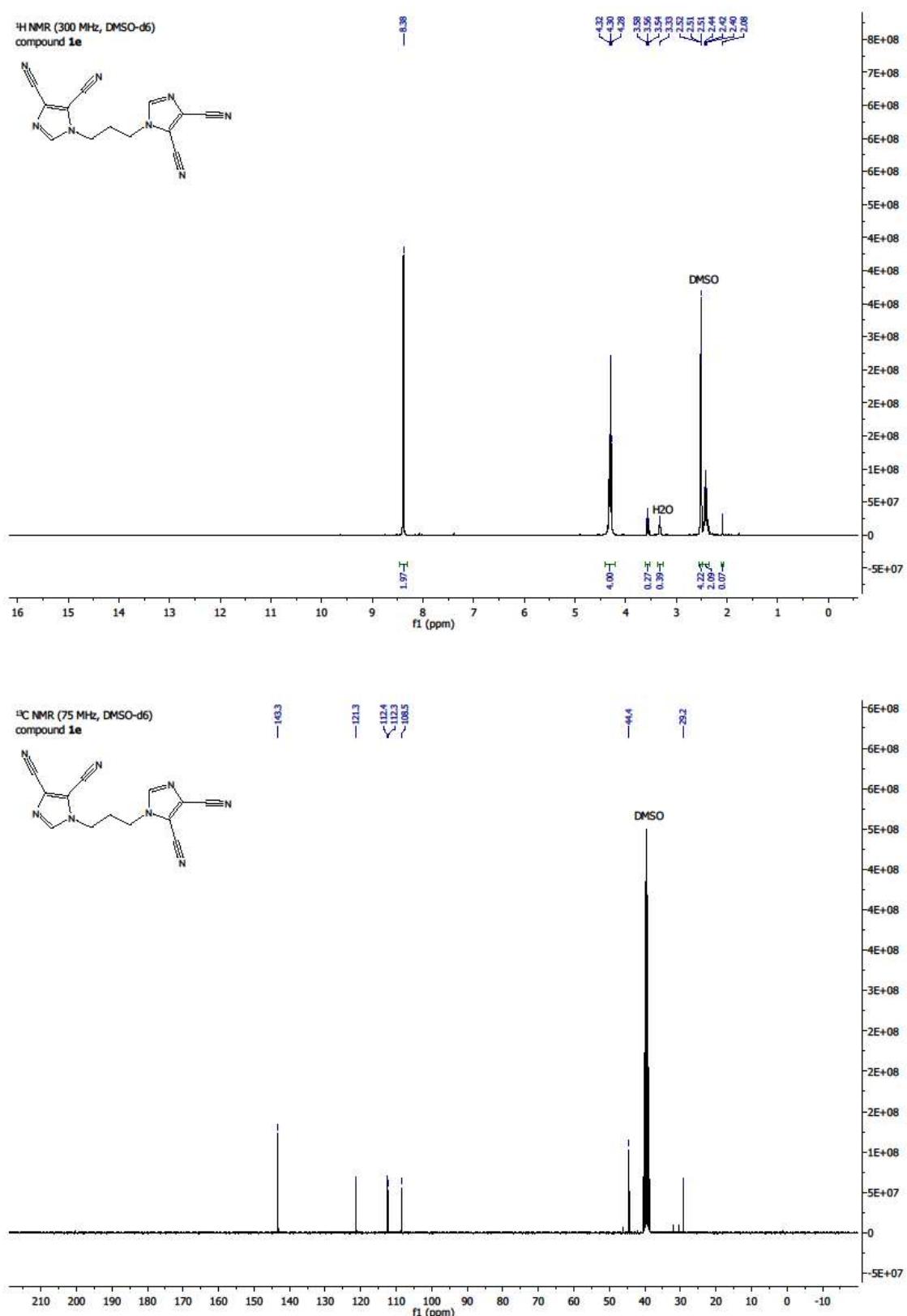
1d



¹H-NMR (300 MHz, DMSO) δ 8.61 (s, 2H), 6.77 (s, 2H).

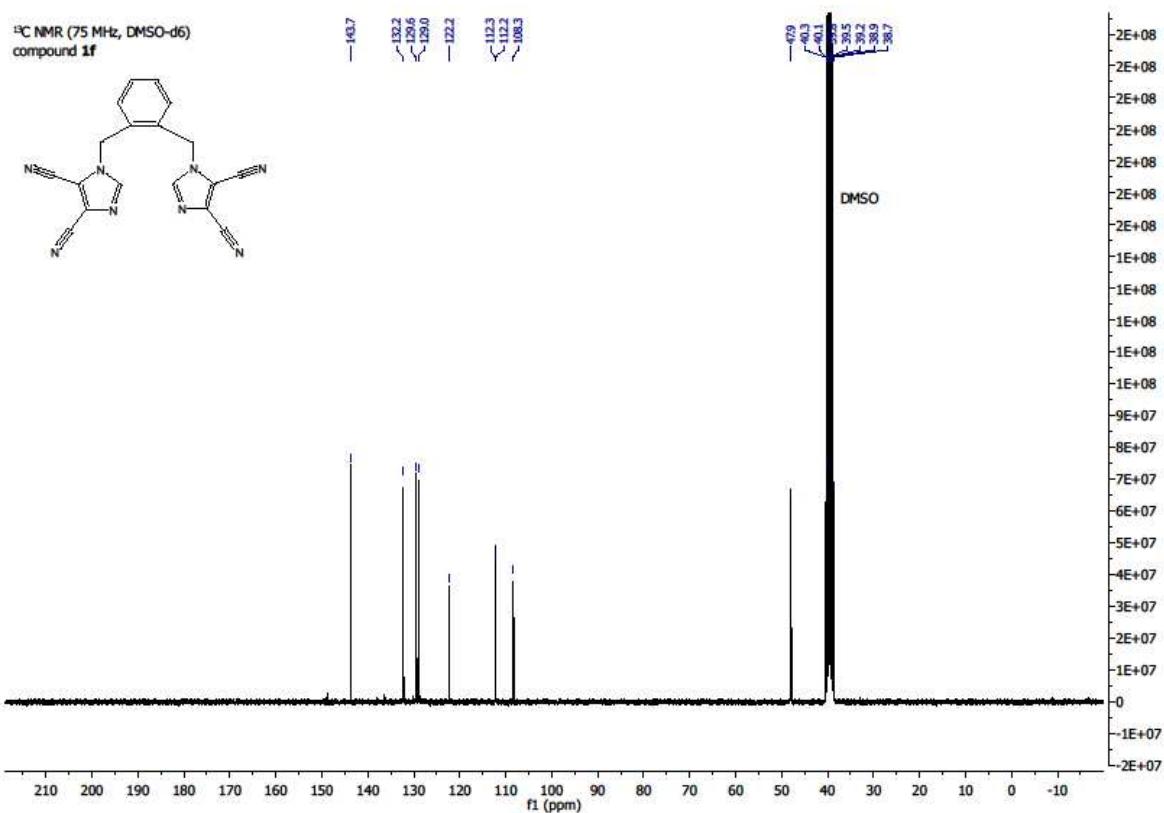
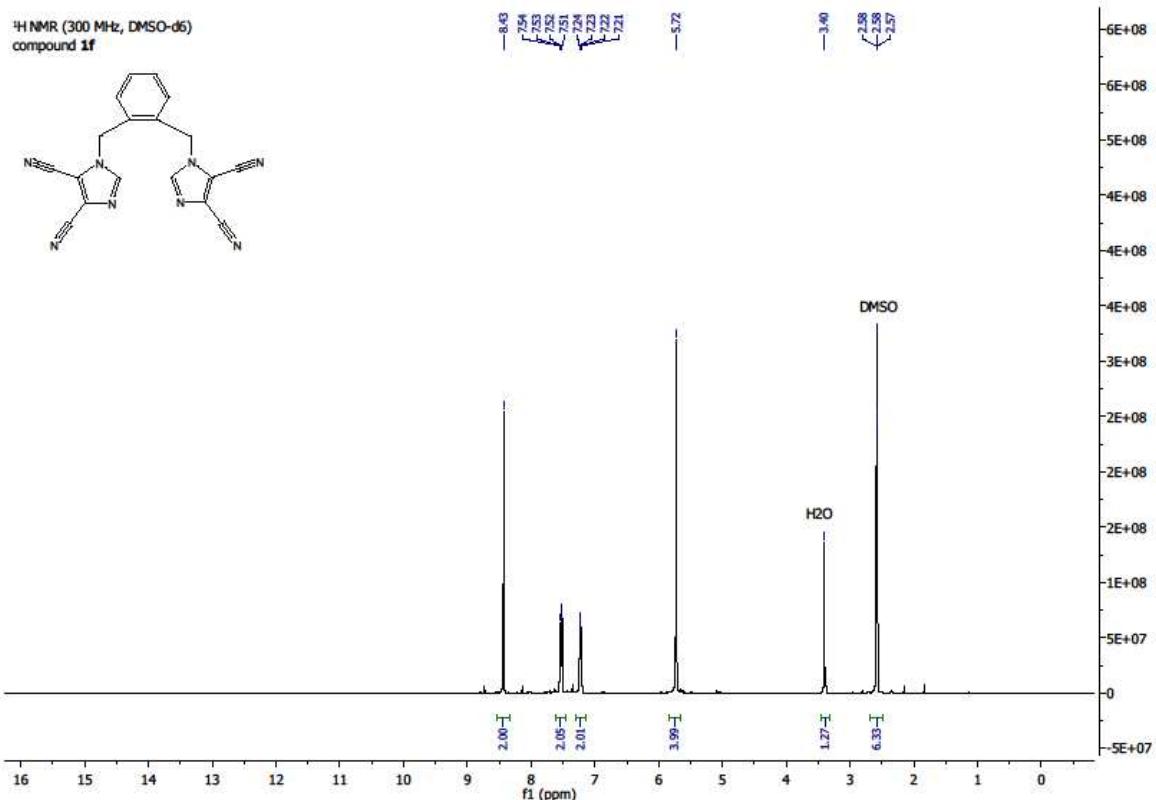
¹³C-NMR (75 MHz, DMSO) δ 143.9 (s), 122.5 (s), 112.7 (s), 112.0 (s), 107.9 (s), 55.8 (s).

1e



¹H-NMR (300 MHz, DMSO) δ 8.38 (s, 2H), 4.30 (t, J = 7.1 Hz, 4H), 2.48 – 2.33 (m, 2H).

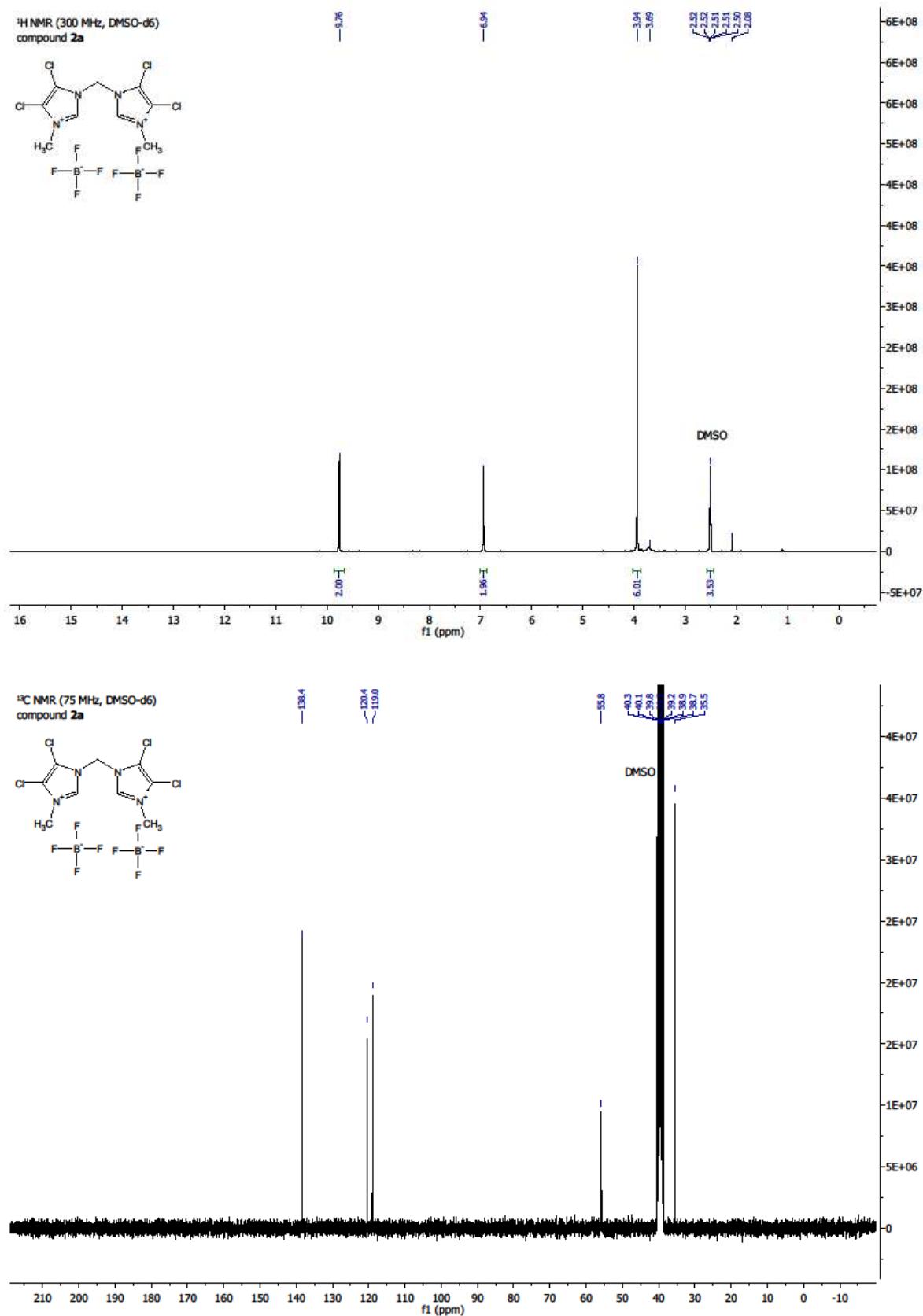
¹³C-NMR (75 MHz, DMSO) δ 143.3 (s), 121.3 (s), 112.4(s), 112.3(s), 108.5 (s), 44.4 (s), 29.2 (s).

1f

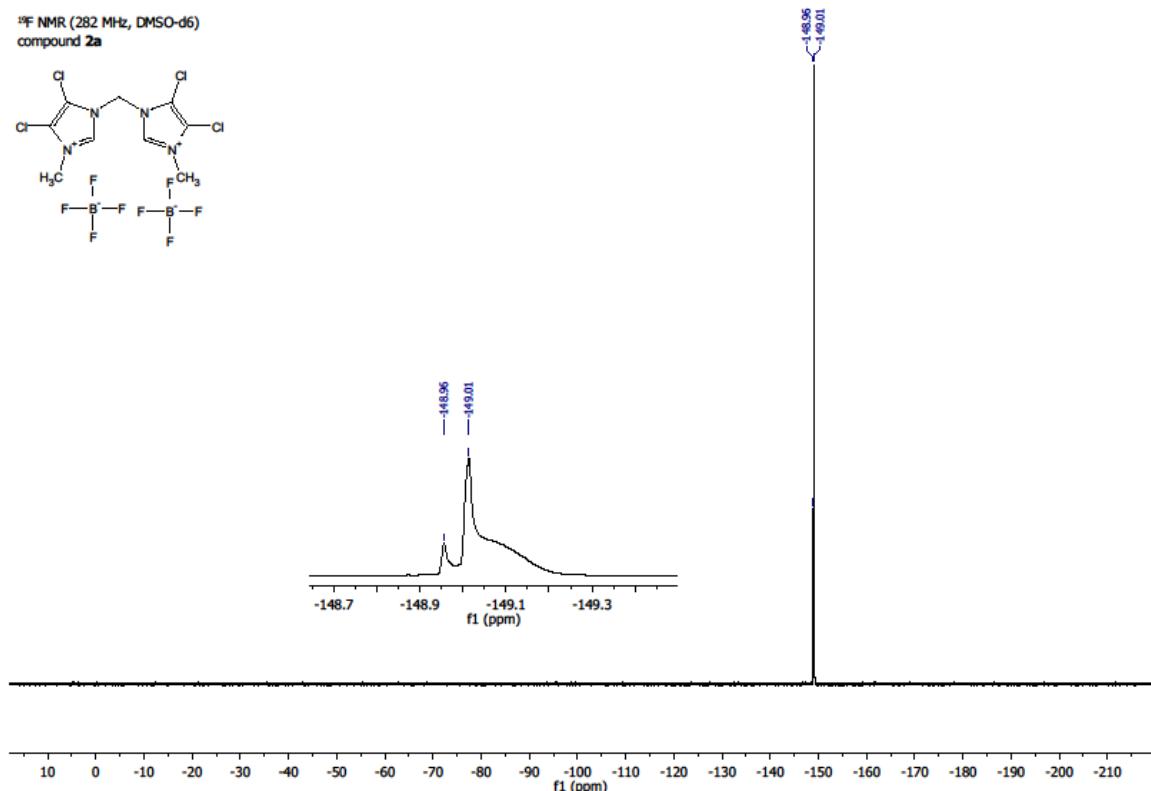
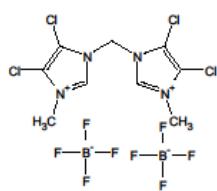
¹H-NMR (300 MHz, DMSO) δ 8.43 (s, 2H), 7.53 (dd, *J* = 5.7, 3.3 Hz, 2H), 7.22 (dd, *J* = 5.6, 3.4 Hz, 2H), 5.72 (s, 4H).

¹³C-NMR (75 MHz, DMSO) δ 143.7 (s), 132.23 (s), 129.6 (s), 129.0 (s), 122.20 (s), 112.3 (s), 112.2 (s), 108.3 (s), 47.9 (s).

2a



¹⁹F-NMR (282 MHz, DMSO-d₆)
compound 2a

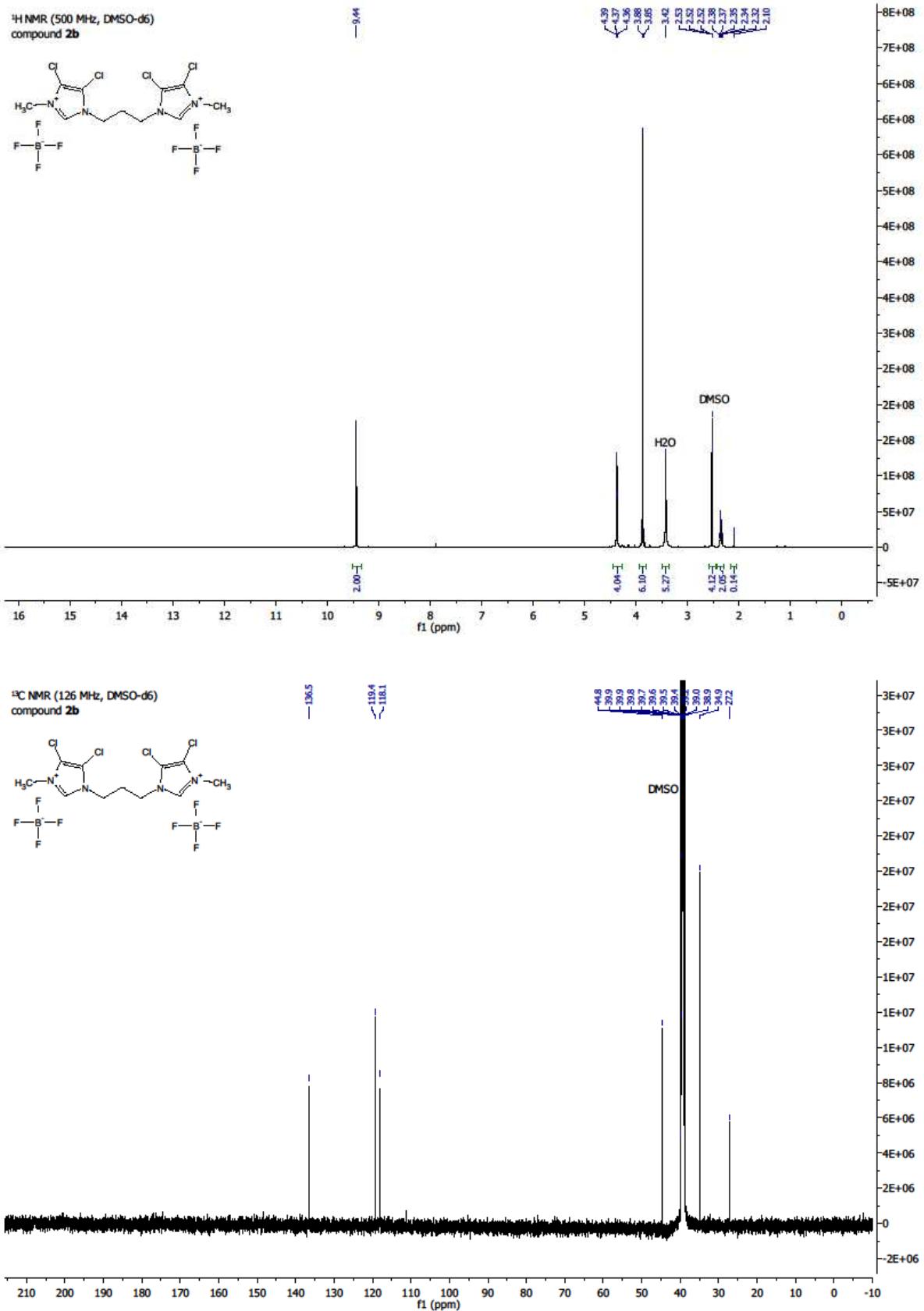


¹H-NMR (300 MHz, DMSO) δ 9.76 (s, 2H), 6.94 (s, 2H), 3.94 (s, 6H).

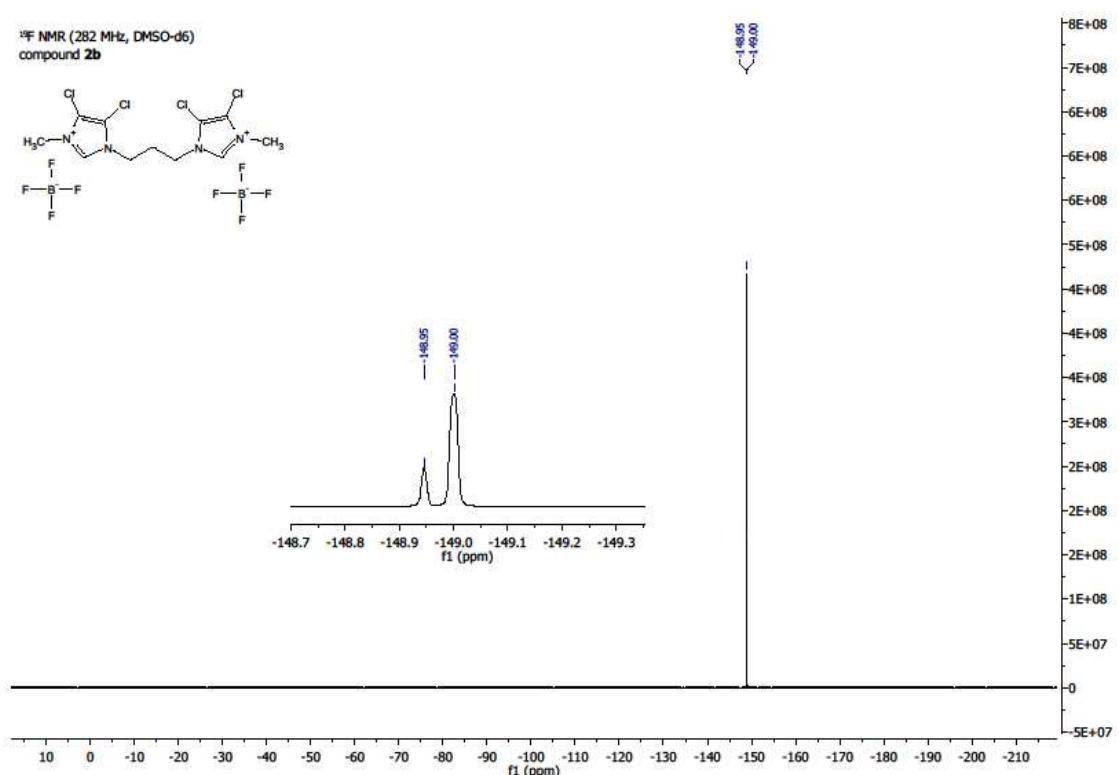
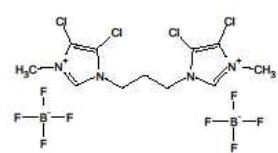
¹³C-NMR (75 MHz, DMSO) δ 138.5 (s), 120.4 (s), 119.0 (s), 55.8 (s), 35.5 (s).

¹⁹F-NMR (282 MHz, DMSO) δ -148.96(s), -149.01(s).

2b



¹⁹F NMR (282 MHz, DMSO-d6)
compound 2b

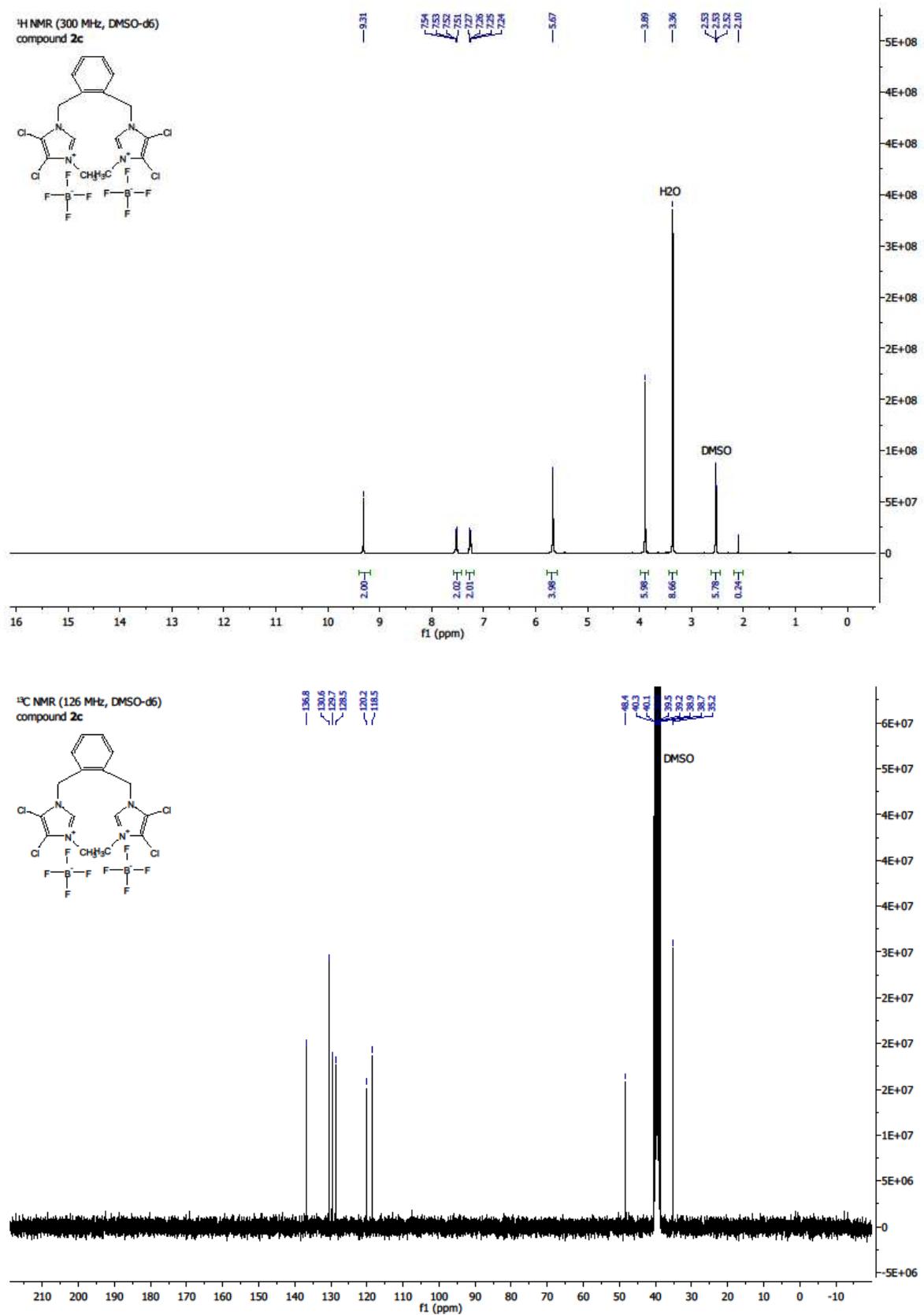


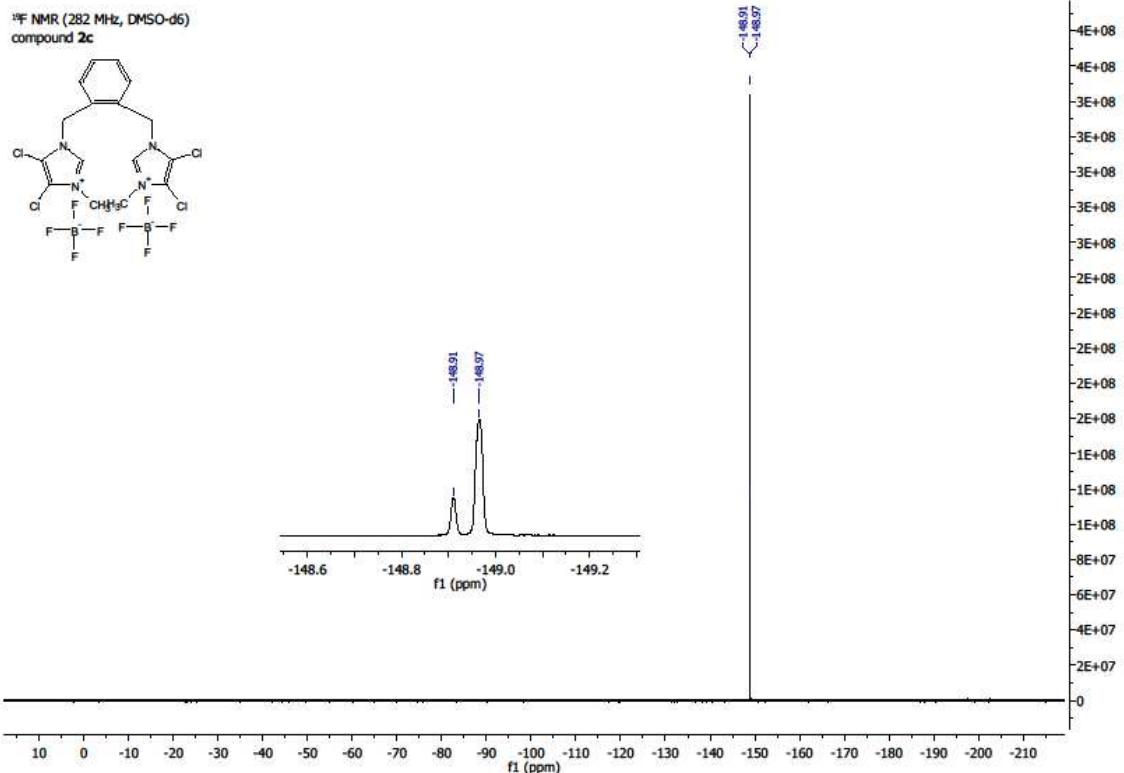
¹H-NMR (500 MHz, DMSO) δ 9.44 (s, 2H), 4.37 (t, *J* = 7.2 Hz, 4H), 3.86 (s, 6H), 2.42 – 2.29 (m, 2H).

¹³C-NMR (126 MHz, DMSO) δ 136.5 (s), 119.4 (s), 118.1 (s), 44.8 (s), 34.9 (s), 27.2 (s).

¹⁹F-NMR (282 MHz, DMSO) δ -148.95(s), -149.00(s).

2c



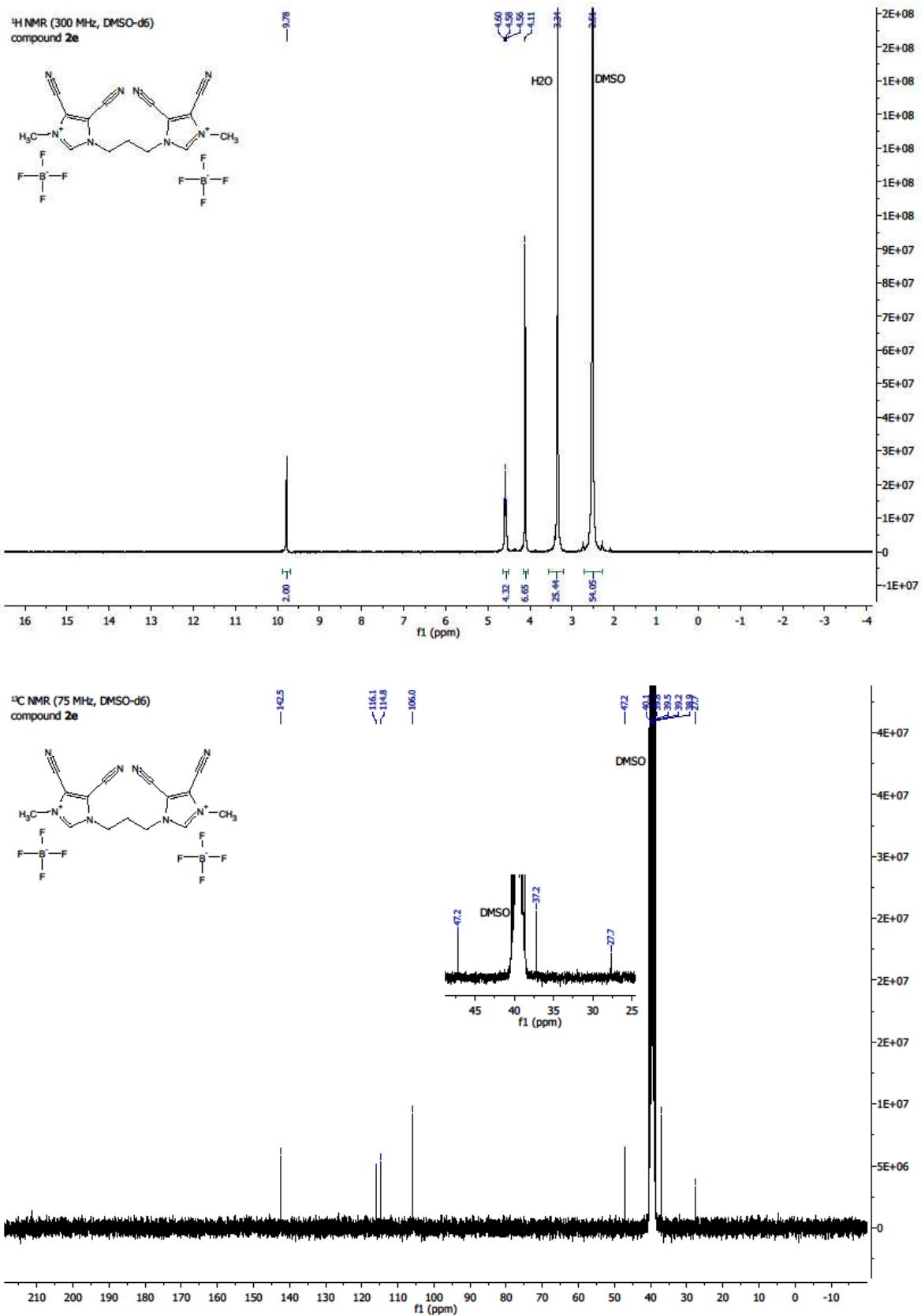


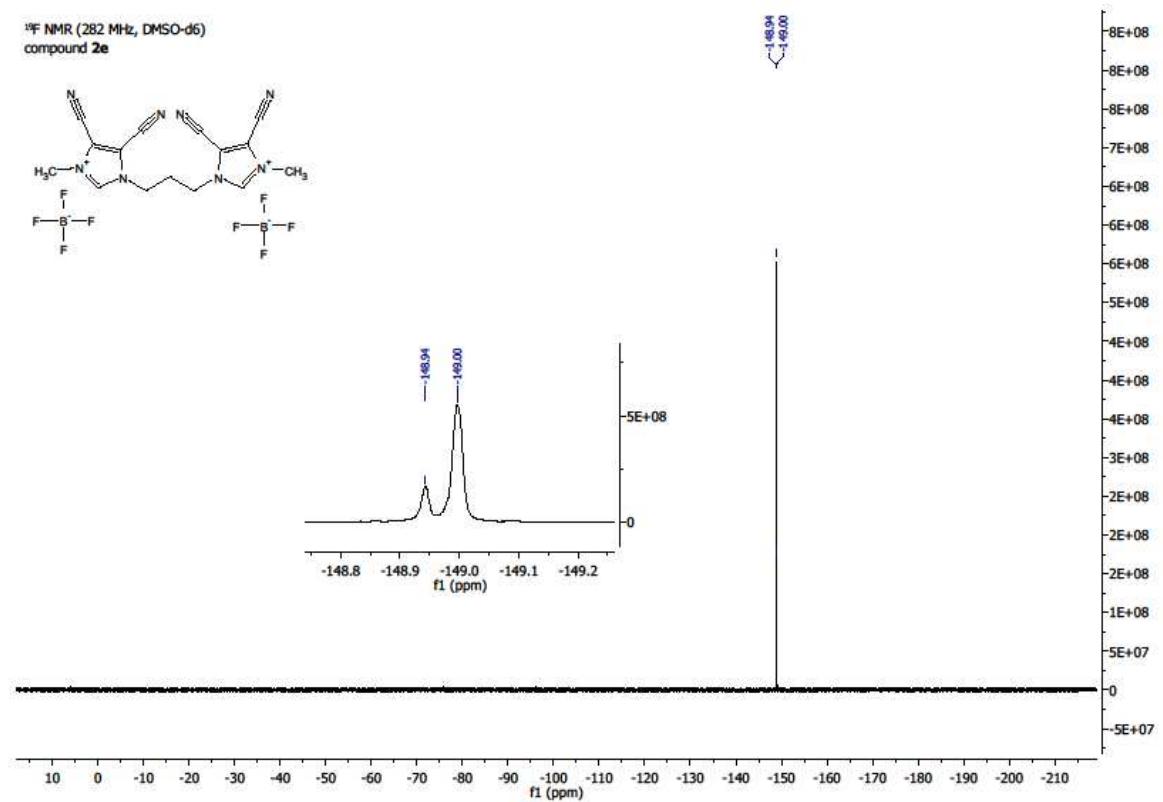
¹H-NMR (300 MHz, DMSO) δ 9.31 (s, 2H), 7.53 (dd, J = 5.7, 3.3 Hz, 2H), 7.26 (dd, J = 5.6, 3.4 Hz, 2H), 5.67 (s, 4H), 3.89 (s, 6H).

¹³C-NMR (75 MHz, DMSO) δ 136.8 (s), 130.62 (s), 129.7 (s), 128.5 (s), 120.2 (s), 118.5 (s), 48.4 (s), 35.2 (s).

¹⁹F-NMR (282 MHz, DMSO) δ -148.91(s), -148.97(s).

2e



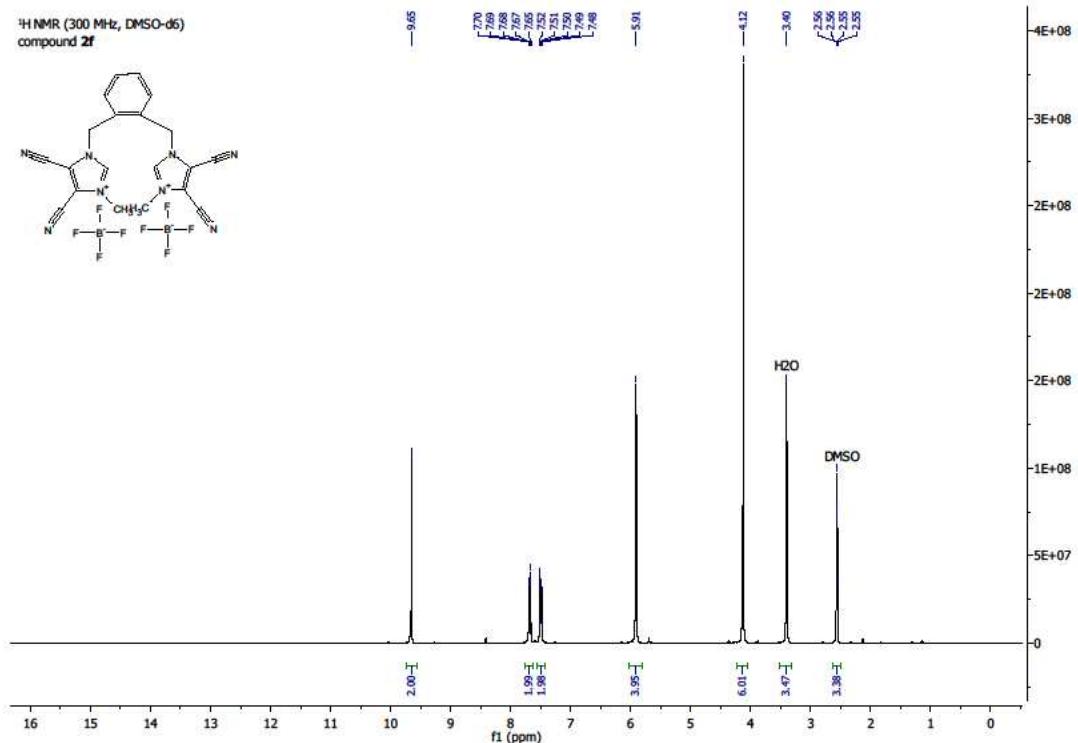


¹H-NMR (300 MHz, DMSO) δ 9.78 (s, 2H), 4.58 (t, *J* = 7.1 Hz, 4H), 4.11 (s, 6H).

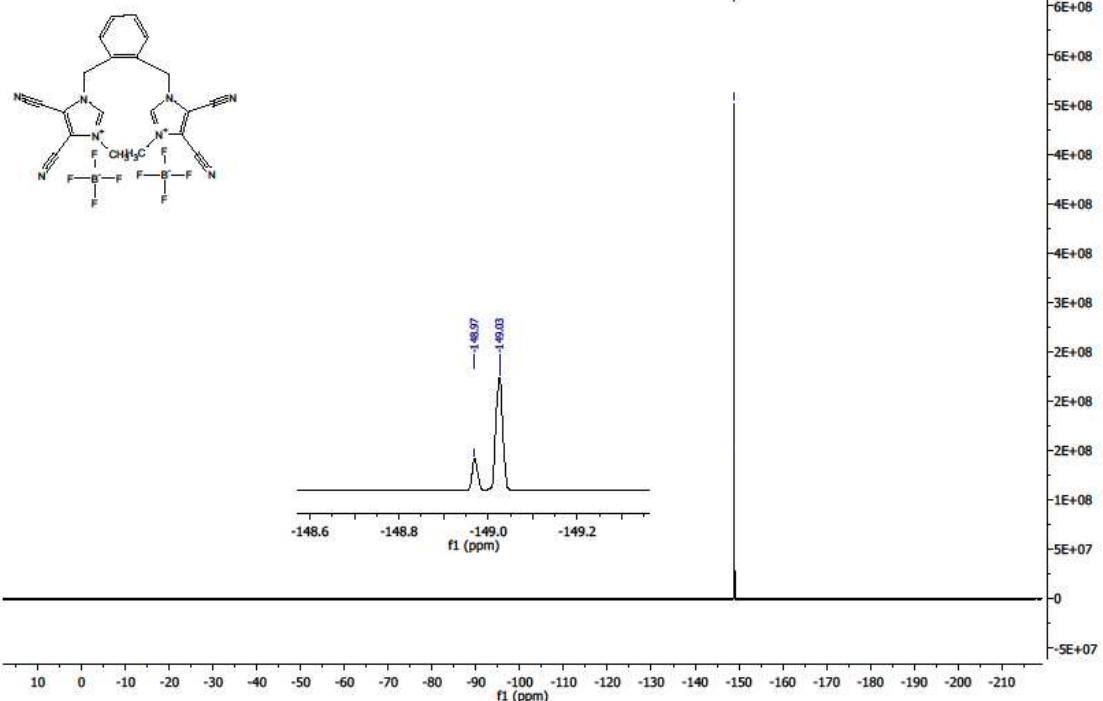
¹³C-NMR (75 MHz, DMSO) δ 142.5 (s), 116.1 (s), 114.8 (s), 106.0 (s), 47.2 (s), 37.2 (s), 27.7 (s).

¹⁹F NMR (282 MHz, DMSO) δ -148.94(s), -149.00(s).

2f



¹⁹F NMR (282 MHz, DMSO-d6)
compound 2f

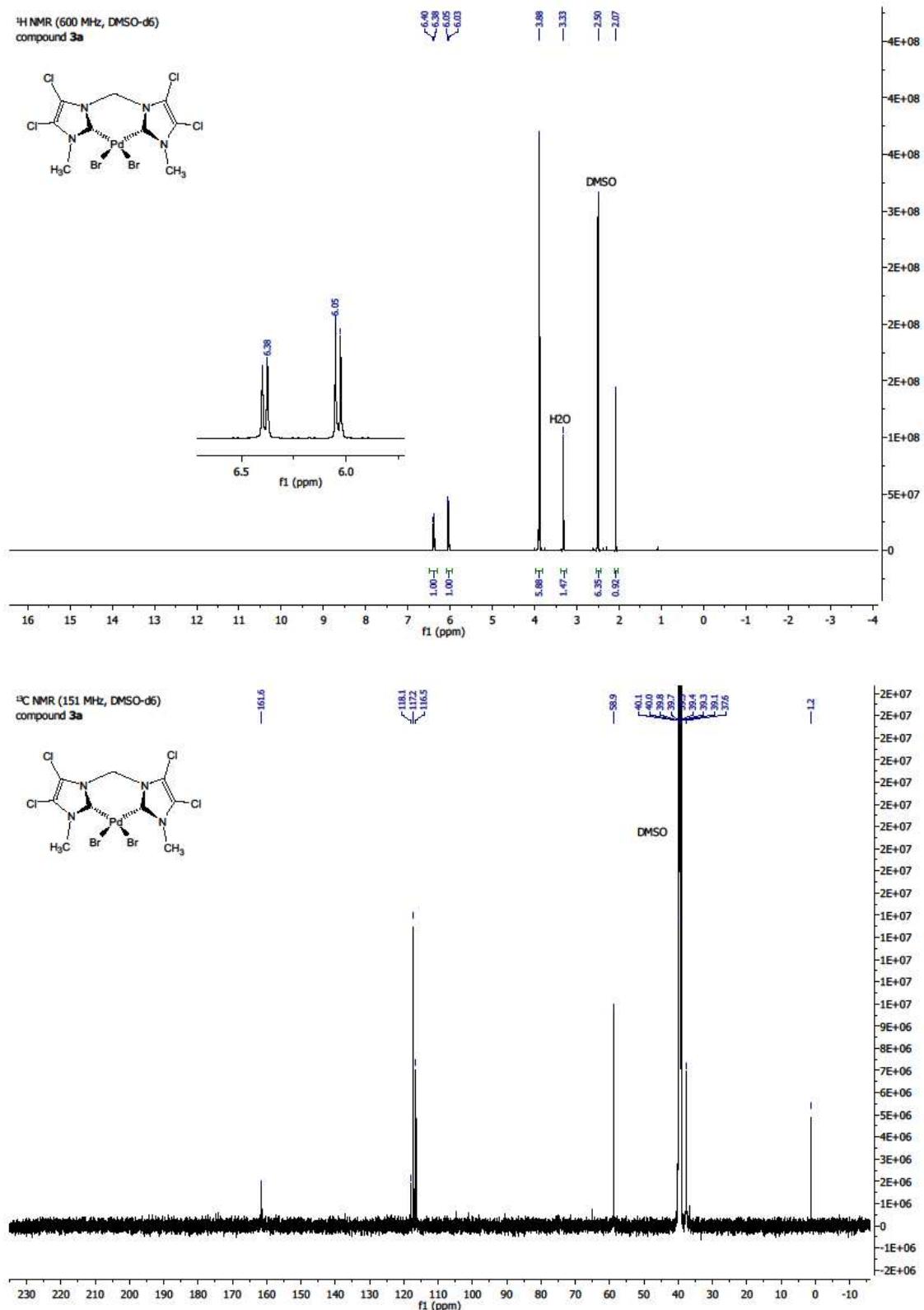


¹H-NMR (300 MHz, DMSO) δ 9.65 (s, 2H), 7.68 (dt, *J* = 7.4, 3.7 Hz, 2H), 7.57 – 7.42 (m, 2H), 5.91 (s, 4H), 4.12 (s, 6H).

¹³C-NMR (75 MHz, DMSO) δ 142.6 (s), 130.7 (s), 130.3 (s), 130.2 (s), 117.1 (s), 114.3 (s), 106.0 (s), 105.9 (s), 50.5 (s), 37.2 (s).

¹⁹F-NMR (282 MHz, DMSO) δ -148.97(s), -149.03(s).

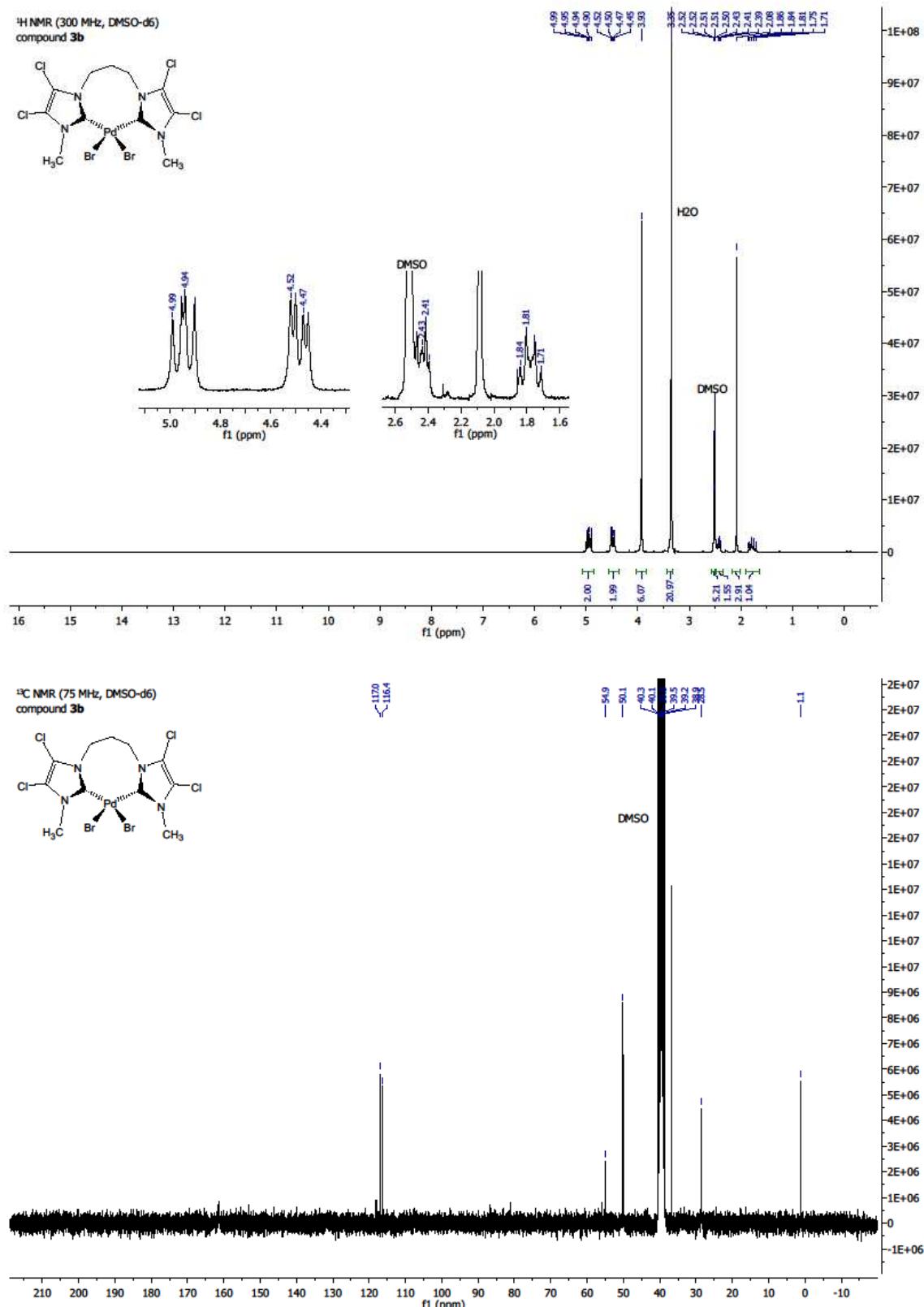
3a



¹H-NMR (600 MHz, DMSO) δ 6.39 (d, *J* = 14.4 Hz, 1H), 6.04 (d, *J* = 14.4 Hz, 1H), 3.88 (s, 6H).

¹³C-NMR (151 MHz, DMSO) δ 161.6 (s), 117.2 (s), 116.5 (s), 58.9 (s), 37.6 (s).

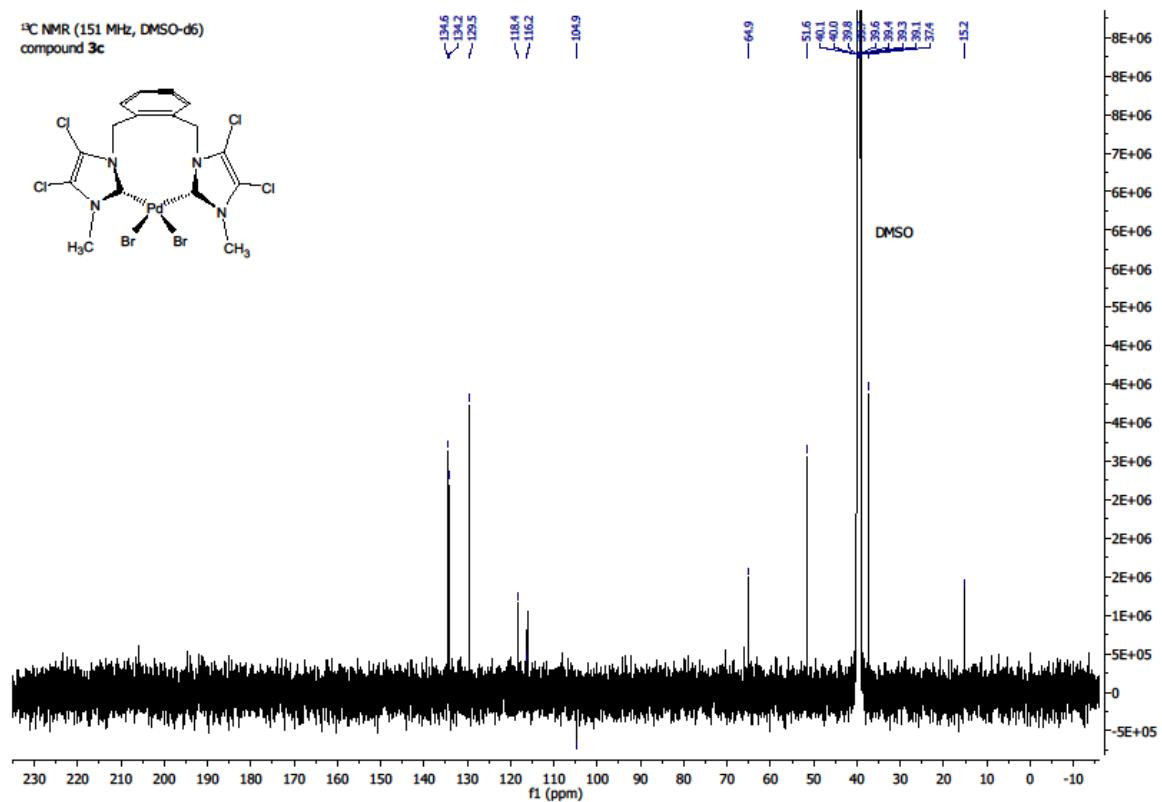
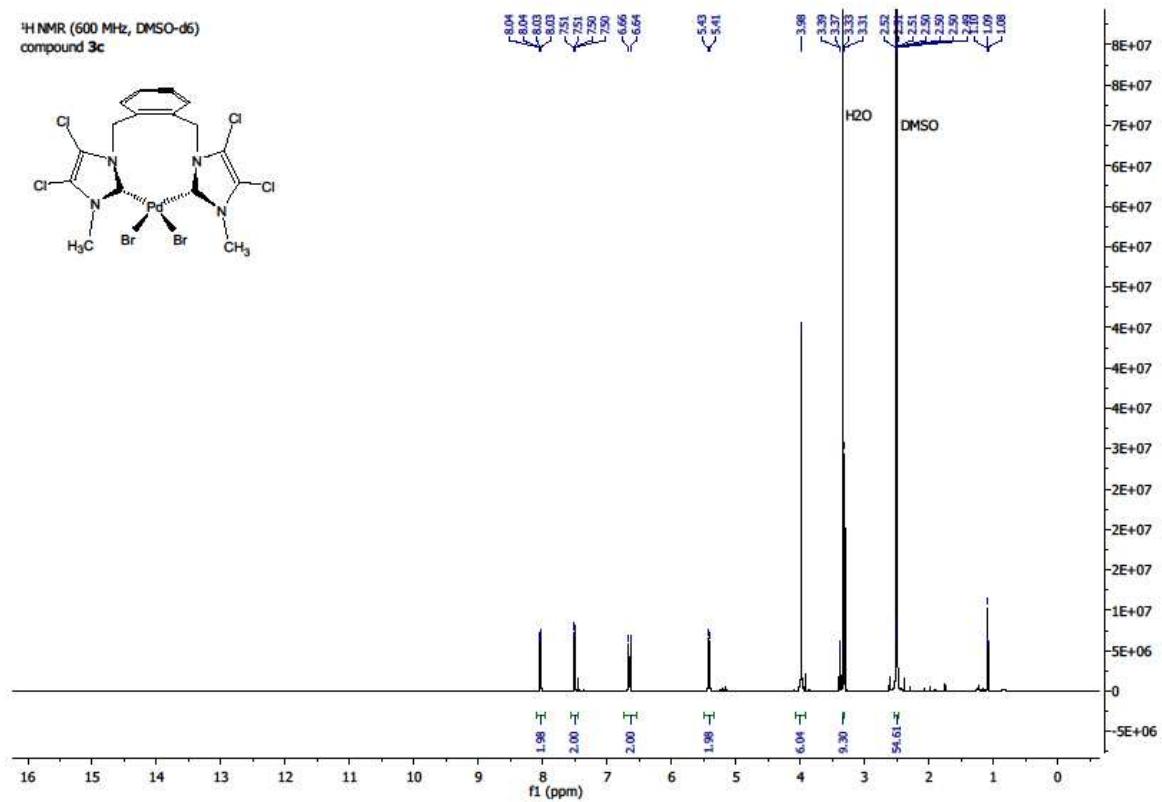
3b



¹H-NMR (300 MHz, DMSO) δ 4.95 (dd, *J* = 15.1, 11.1 Hz, 2H), 4.49 (dd, *J* = 15.0, 5.8 Hz, 2H), 3.93 (s, 6H), 2.41 (t, *J* = 6.1 Hz, 1H), 1.88 – 1.65 (m, 1H).

¹³C-NMR (75 MHz, DMSO) δ 117.0 (s), 116.4 (s), 54.9 (s), 50.1 (s), 36.8 (s), 28.5 (s).

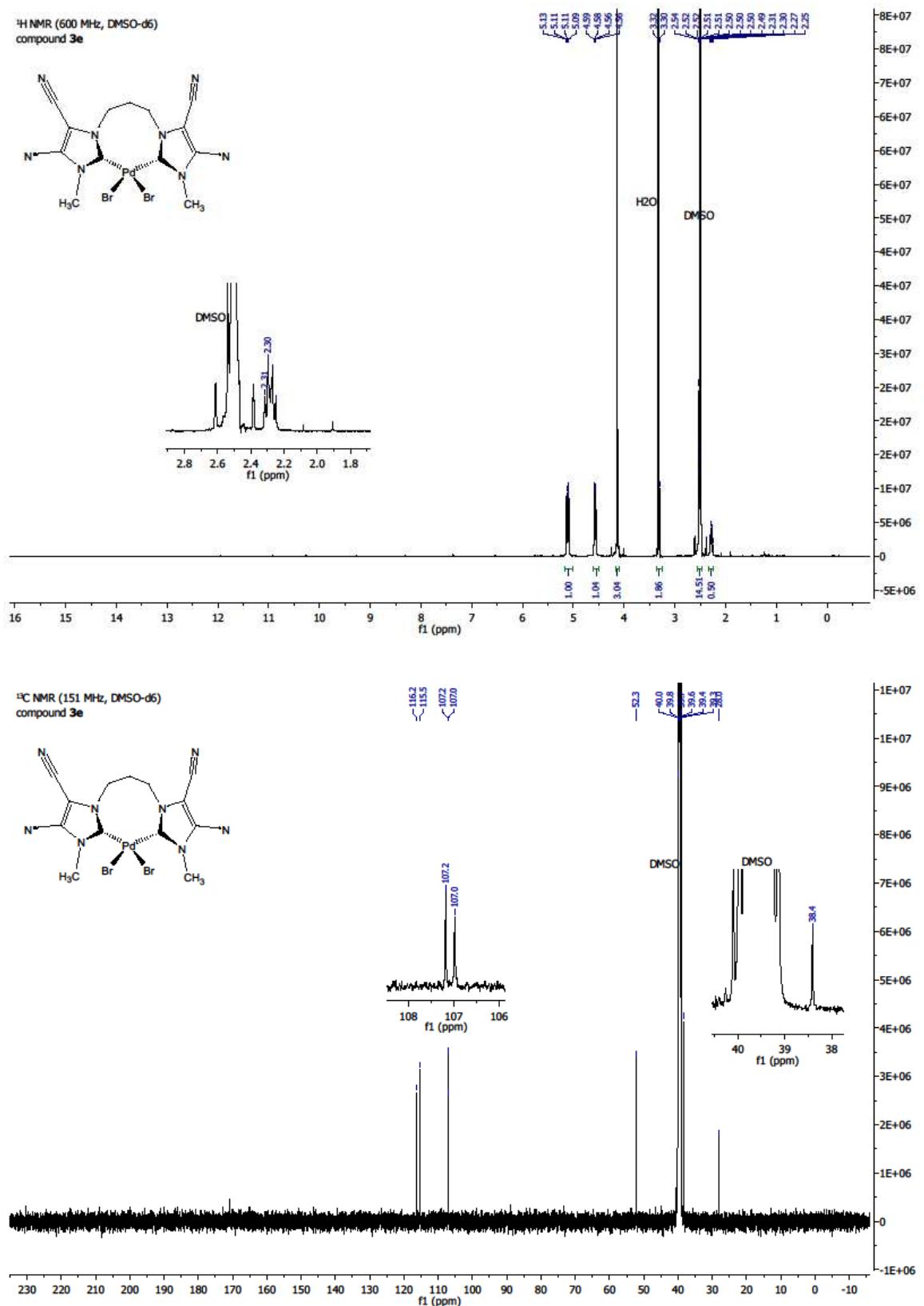
3c



¹H-NMR (600 MHz, DMSO) δ 8.04 (dd, *J* = 5.7, 3.5 Hz, 2H), 7.50 (dd, *J* = 5.7, 3.4 Hz, 2H), 6.65 (d, *J* = 15.2 Hz, 2H), 5.42 (d, *J* = 15.3 Hz, 2H), 3.98 (s, 6H).

¹³C-NMR (151 MHz, DMSO) δ 134.6 (s), 134.2 (s), 118.4 (s), 116.2 (s), 65.0(s), 51.6 (s), 37.4 (s).

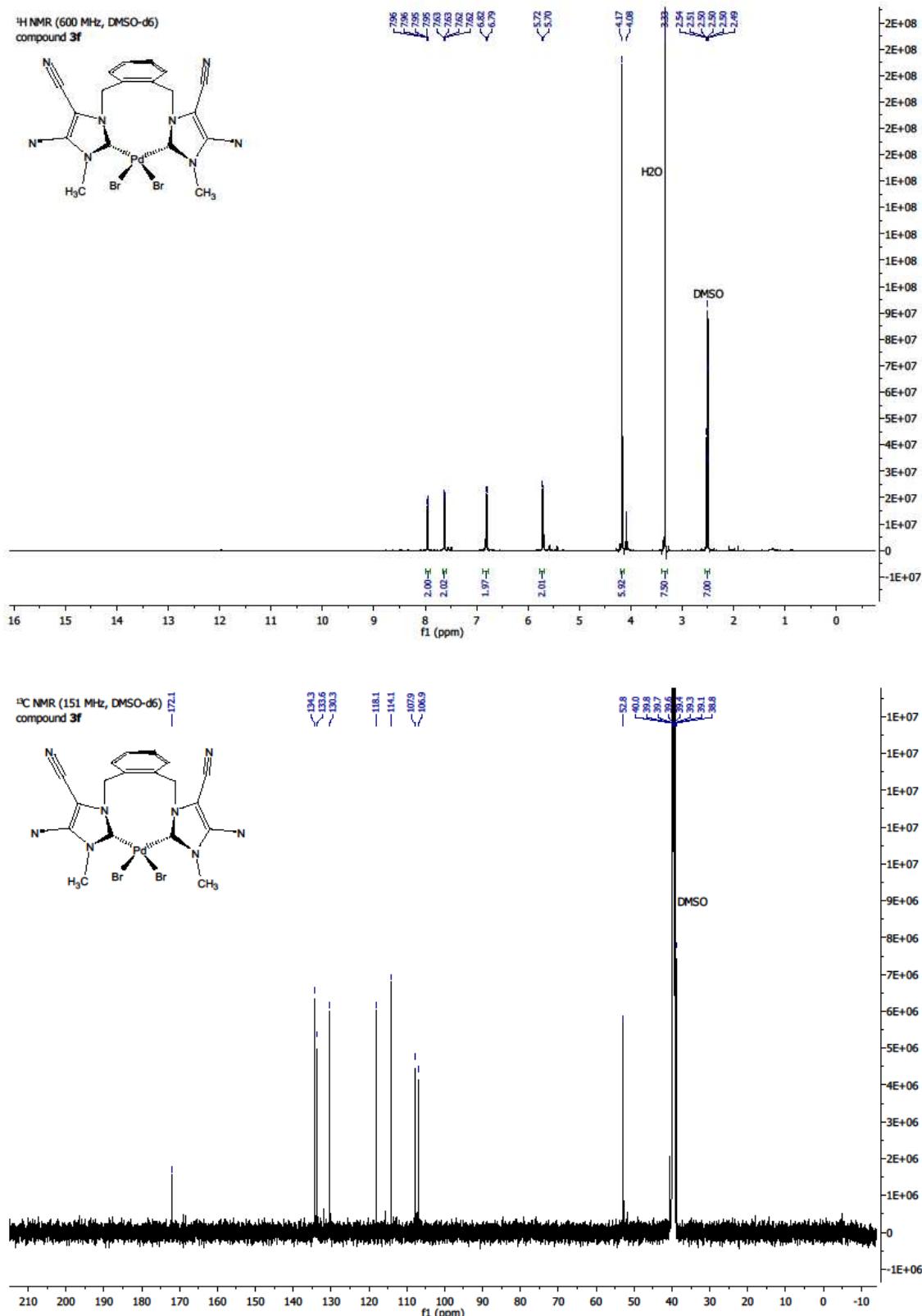
3e



¹H-NMR (600 MHz, DMSO) δ 5.11 (dd, *J* = 15.2, 11.5 Hz, 2H), 4.57 (dd, *J* = 15.0, 5.8 Hz, 2H), 4.13 (s, 6H), 2.28 (dd, *J* = 27.2, 11.4 Hz, 1H).

¹³C-NMR (151 MHz, DMSO) δ 116.2 (s), 115.5 (s), 107.2(s), 107.0 (s), 52.3 (s), 38.4 (s), 28.0 (s).

3f



¹H-NMR (600 MHz, DMSO) δ 7.95 (dd, J = 5.7, 3.5 Hz, 2H), 7.63 (dd, J = 5.7, 3.4 Hz, 2H), 6.81 (d, J = 15.2 Hz, 2H), 5.71 (d, J = 15.3 Hz, 2H), 4.17 (s, 6H).

¹³C-NMR (151 MHz, DMSO) δ 172.1 (s), 134.3 (s), 133.6 (s), 130.3 (s), 118.1 (s), 114.1 (s), 107.9 (s), 106.9 (s), 52.8 (s).