Supporting Information

Efficient one-pot synthesis of *trans*-Pt(II)(salicylaldimine)(4picoline)Cl complexes: Effective agents for enhanced expression of p53 tumor suppressor genes

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Synthesis and characterization of ligands L1-8



General procedure for the synthesis of ligands L1-8

1 mmol of amine/aniline, particular salicylaldehyde and anhyd. Na_2SO_4 (200 mg) were taken in 20 mL of CH_2Cl_2 . The mixture was stirred under reflux for 4h (checked by TLC). On completion it is cooled to room temperature and filtered to remove undissolved solid which is further washed with CH_2Cl_2 . The filtrate was vacuum evaporated to get L1 to L8 which was dried and used in the next step without further purification.

Characterization of ligand L1.



Yellow liquid, ¹H NMR (400 MHz, CDCl₃) δ 13.45 (s, 1H), 8.26 (s, 1H), 7.11 (d, J = 8.3 Hz, 1H), 7.05 (s, 1H), 6.87 (d, J = 8.3 Hz, 1H), 3.41 (d, J = 7.5 Hz, 2H), 2.29 (s, 3H), 2.02–1.92 (m, 1H), 0.98 (d, J = 6.7 Hz, 6H). ¹³C NMR (100 MHz, CDCl₃) δ 164.7, 159.1, 132.9, 131.2, 127.4, 118.5, 116.8, 67.6, 29.6, 20.5, 20.4.

Characterization of ligand L2.



Orange solid, ¹H NMR (400 MHz, CDCl₃) δ 13.01 (s, 1H), 8.56 (s, 1H), 7.41 (t, *J* = 7.7 Hz, 2H), 7.27 (t, *J* = 7.6 Hz, 3H), 7.18 (d, *J* = 7.0 Hz, 2H), 6.93 (d, *J* = 8.6 Hz, 1H), 2.31 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 162.7, 159.0, 148.7, 134.1, 132.3, 129.4, 128.2, 126.8, 121.2, 118.9, 117.0, 20.4.

Characterization of ligand L3.



Light yellow solid, ¹H NMR (400 MHz, CDCl₃) δ 12.85 (s, 1H), 8.54 (s, 1H), 7.28–7.22 (m, 2H), 7.22–7.16 (m, 2H), 7.15–7.07 (m, 2H), 6.94 (d, *J* = 8.1 Hz, 1H), 2.32 (s, 3H). ¹⁹F NMR (376 MHz, CDCl₃) δ –116.17. ¹³C NMR (100 MHz, CDCl₃) δ 162.5, 161.6 (d, *J*_{C-F} = 247.45 Hz), 158.9, 144.8, 134.2, 132.3, 128.2, 122.58 (d, *J*_{C-F} = 8.3 Hz), 118.7, 117.0, 116.2 (d, *J*_{C-F} = 22.7 Hz), 20.4.

Characterization of ligand L4.¹



Light yellow solid, ¹H NMR (400 MHz, CDCl₃) δ 13.27 (s, 1H), 8.64 (s, 1H), 7.48–7.36 (m, 4H), 7.34–7.27 (m, 3H), 7.04 (d, *J* = 8.1 Hz, 1H), 6.96 (t, *J* = 7.5 Hz, 1H). ¹³C NMR (100 MHz, CDCl₃) δ 162.7, 161.2, 148.5, 133.2, 132.3, 129.4, 126.9, 121.2, 119.2, 119.1, 117.3.

Characterization of ligand L5.²



Light yellow solid, ¹H NMR (400 MHz, CDCl₃) δ 13.10 (s, 1H), 8.57 (s, 1H), 7.42–7.33 (m, 2H), 7.26 (dd, J = 5.1, 3.0 Hz, 1H), 7.23 (dd, J = 5.0, 3.0 Hz, 1H), 7.15–7.06 (m, 2H), 7.02 (d, J = 8.8 Hz, 1H), 6.94 (t, J = 7.5 Hz, 1H). ¹⁹F NMR (376 MHz, CDCl₃) δ –115.93. ¹³C NMR (100 MHz, CDCl₃) δ 162.5, 161.7 (d, $J_{C-F} = 247.45$ Hz), 161.1, 160.4, 144.7, 133.2, 132.3, 122.6 (d, $J_{C-F} = 8.3$ Hz), 119.2, 117.3, 116.2 (d, $J_{C-F} = 22.7$ Hz).

Characterization of ligand L6.



Yellow liquid, ¹H NMR (400 MHz, CDCl₃) δ 13.41 (s, 1H), 8.25 (s, 1H), 7.06–6.98 (m, 1H), 6.95 (dd, J = 8.4, 3.1 Hz, 1H), 6.90 (dd, J = 9.0, 4.5 Hz, 1H), 3.43 (d, J = 6.5 Hz, 2H), 2.03–1.93 (m, 1H), 0.98 (d, J = 6.7 Hz, 6H). ¹⁹F NMR (376 MHz, CDCl₃) δ –

126.64. ¹³C NMR (100 MHz, CDCl₃) δ 163.7, 157.4, 155.3 (d, $J_{C-F} = 236.2$ Hz), 119.0 (d, $J_{C-F} = 23.2$ Hz), 118.5 (d, $J_{C-F} = 7.2$ Hz), 118.0 (d, $J_{C-F} = 7.4$ Hz), 116.2 (d, $J_{C-F} = 23.1$ Hz), 67.6, 29.6, 20.4.

Characterization of ligand L7.



Orange solid, ¹H NMR (400 MHz, CDCl₃) δ 13.02 (s, 1H), 8.56 (s, 1H), 7.49–7.38 (m, 2H), 7.36–7.26 (m, 3H), 7.16–7.05 (m, 2H), 7.02–6.94 (m, 1H). ¹⁹F NMR (376 MHz, CDCl₃) δ –125.82. ¹³C NMR (100 MHz, CDCl₃) δ 161.5, 157.3, 155.6 (d, $J_{C-F} = 237.5$ Hz), 148.1, 129.5, 127.3, 121.2, 120.3 (d, $J_{C-F} = 23.3$ Hz), 118.9 (d, $J_{C-F} = 6.8$ Hz), 118.4 (d, $J_{C-F} = 7.3$ Hz), 117.1 (d, $J_{C-F} = 23.2$ Hz).

Characterization of ligand L8.³



Orange solid, ¹H NMR (400 MHz, CDCl₃) δ 12.87 (s, 1H), 8.53 (s, 1H), 7.33–7.22 (m, 2H), 7.18–7.05 (m, 4H), 6.98 (dd, J = 8.7, 4.5 Hz, 1H). ¹⁹F NMR (376 MHz, CDCl₃) δ –115.27, – 125.68. ¹³C NMR (100 MHz, CDCl₃) δ 161.9 (d, $J_{C-F} = 248.5$ Hz), 161.2, 157.2, 155.6 (d, $J_{C-F} = 237.4$ Hz), 144.3, 122.7 (d, $J_{C-F} = 8.4$ Hz), 120.35 (d, $J_{C-F} = 23.3$ Hz), 118.8 (d, $J_{C-F} = 5.7$ Hz), 118.4 (d, $J_{C-F} = 7.4$ Hz), 117.1 (d, $J_{C-F} = 23.1$ Hz), 116.4 (d, $J_{C-F} = 22.8$ Hz).

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¹H, ¹³C and ¹⁹F NMR Spectra of Ligands



Fig. S1 ¹H NMR of L1 in CDCl₃ at 25°C



Fig. S2 ¹³C NMR of L1 in CDCl₃ at 25°C







Fig. S4 ^{13}C NMR of L2 in CDCl3 at 25°C



Fig. S6 ^{13}C NMR of L3 in CDCl3 at 25°C



Fig. S8 ¹H NMR of L4 in CDCl₃ at 25° C







Fig. S12 $^{19}\mathrm{F}$ NMR of L5 in CDCl3 at 25°C







Fig. S14 $^{\rm 13}\rm C$ NMR of L6 in CDCl3 at 25°C







Fig. S18 $^{19}\mathrm{F}$ NMR of L7 in CDCl3 at 25°C



Fig. S20 13 C NMR of L8 in CDCl₃ at 25°C



Fig. S21 $^{19}\mathrm{F}$ NMR of L8 in CDCl3 at 25°C

¹H, ¹³C and ¹⁹F NMR Spectra of platinum complexes







Fig. S23 ^{13}C NMR of C1 in CDCl3 at 25°C







Fig. S25 $^{\rm 13}\rm C$ NMR of C2 in CDCl3 at 25°C



Fig. S26 ¹H NMR of C3 in CDCl₃ at 25°C



Fig. S27 $^{13}\mathrm{C}$ NMR of C3 in CDCl3 at 25°C







Fig. S31 ¹H NMR of C5 in CDCl₃ at 25°C



Fig. S33 $^{19}\mathrm{F}$ NMR of C5 in CDCl3 at 25°C







Fig. S35 ^{13}C NMR of C6 in CDCl3 at 25°C





Fig. S37 $^1\mathrm{H}$ NMR of C7 in CDCl3 at 25°C



Fig. S39 $^{19}\mathrm{F}$ NMR of C7 in CDCl3 at 25°C



Fig. S40 ¹H NMR of C8 in CDCl₃ at 25°C



Fig. S41 ^{13}C NMR of C8 in CDCl₃ at 25°C



Fig. S42 $^{19}\mathrm{F}$ NMR of C8 in CDCl3 at 25°C

Mass Spectra of all platinum complexes



Fig. S43 Mass Spectrum of C1



Fig. S44 Mass Spectrum of C2



Fig. S45 Mass Spectrum of C3



Fig. S46 Mass Spectrum of C4



Fig. S47 Mass Spectrum of C5



Fig. S48 Mass Spectrum of C6



Fig. S49 Mass Spectrum of C7



Fig. S50 Mass Spectrum of C8

Stabliltiy study

Stability of metal complex studied for cytotoxic analysis is always important. Therefore we perfomed stability analysis experiments for C2, C3 and C5 as model complexes using ¹H NMR spectroscopy in 10–15% D₂O–DMSO- d_6 mixture under room temperature at 0, 24, 48, and 72 h. No changes were observed in ¹H chemical shifts and also in peak number (Fig. S51 for C2, Fig. S52 for C3 and Fig. S53 for C5 below). It is concluded that C2, C3 and C5 are highly stable under these conditions.



Fig. S51 Stability analysis of C2 using ¹H NMR, taken in 15% D_2O –DMSO- d_6 at room temperature during (bottom to top) 0, 24, 48 and 72 h.



Fig. S52 Stability analysis of C3 using ¹H NMR, taken in 10% D₂O–DMSO- d_6 at room temperature during (bottom to top) 0, 24, 48 and 72 h.



Fig. S53 Stability analysis of C5 using ¹H NMR, taken in 10% D_2O –DMSO- d_6 at room temperature during (bottom to top) 0, 24, 48 and 72 h.

Table S1 data and structure refinement of the complex C1, C2 and C3				
	C1	C2	C3	
Empirical formula	C ₁₈ H ₂₃ ClN ₂ OPt	$C_{20}H_{19}CIN_2OPt$	$C_{20}H_{18}ClN_2OPt$	
Temperature (K)	293(2)	233(2)	293(2)	
Crystal system	Monoclinic	Monoclinic	Triclinic	
Space group	P 21/c	P 21/c	P -1	
Unit cell dimensions				
a (Å)	14.219(7)	13.766(2)	7.961(3)	
b (Å)	11.945(6)	12.267(2)	10.420(4)	
c (Å)	10.835(6)	11.094(2)	12.674(5)	
α (°)	90	90	84.941(5)	
β (°)	93.962(6)	93.872	74.687(5)	
γ(°)	90	90	/2.304(5)	
Volume (Å ³)	1836.0(16)	1869.1(6)	966.0(7)	
	4	4	2	
Density (calculated) (mg/m^3)	1.859	1.897	1.897	
Absorption coefficient (mm ⁻)	7.793	7.660	7.421	
F(000)	992	1040	528	
Crystal size (mm ³)	$0.600 \times 0.500 \times 0.200$	$0.700 \times 0.200 \times 0.140$	$0.200\times0.100\times0.050$	
Theta range for data collection (°)	2.229 to 26.000	1.483 to 27.397	1.666 to 25.006	
Index ranges	$-17 \le h \le 17,$ $-12 \le k \le 14,$ $-12 \le l \le 13$ 8007	$-17 \le h \le 17,$ $-15 \le k \le 12,$ $-14 \le 1 \le 14$ 12631	$-4 \le h \le 9,$ $-12 \le k \le 12,$ $-14 \le 1 \le 14$ 4009	
Independent reflections	3582 [R(int) = 0.1028]	12031 12031	3327 [R(int) = 0.0484]	
Completeness to theta =	00.4	4228 [R(int) = 0.0493]	05.2	
25.242° (%)	99.4	99.5	95.5	
Absorption correction	Semi-empirical from equivalents	Semi-empirical from equivalents	Semi-empirical from equivalents	
Max. and min. transmission	1.000 and 0.314	0.746 and 0.252	1.000 and -0.099	
Refinement method	Full-matrix least-squares on F ²	Full-matrix least-squares on F ²	Full-matrix least-squares on F ²	
Data / restraints / parameters	3582 / 0 / 212	4228 / 0 / 228	3327 / 12 / 237	
Goodness-of-fit on F ² Final R indices [I>2sigma(I)]	1.041 R1 ^[a] = 0.0592, wR2 ^[b] = 0.1491	1.054 R1 ^[a] = 0.0444, wR2 ^[b] = 0.1320	1.094 R1 ^[a] = 0.0580, wR2 ^[b] = 0.1699	
R indices (all data)	$R1^{[a]} = 0.0674, wR2^{[b]} = 0.1533$	$R1^{[a]} = 0.0523, wR2^{[b]} = 0.1385$	$R1^{[a]} = 0.0655, wR2^{[b]} = 0.1736$	
Extinction coefficient	n/a	n/a	n/a	
Largest diff. peak and hole	4.420 and -2.775 e.Å ⁻³	1.763 and -2.859 e.Å ⁻³	2.342 and -3.095 e.Å ⁻³	

Single crystal structure study and refinement data of C1, C2 and C3

^[a] R1 = $\Sigma_{all \ reflections} |F_0 - F_c| / \Sigma_{all \ reflections} |F_0|$, ^[b] wR2 = $[\Sigma w (F_0^2 - F_c^2)^2 / \Sigma (w (F_0^2)^2)]^{1/2}$.

Packing plot of C1



Fig. S54 1D array made by intermolecular bond between chloride attached to Pt center and H of the next molecule in C1

Packing plot of C2



Fig. S55 3D arrangement of molecules in crystal packing; made by intermolecular bonds between chloride attached to Pt center and H of the next molecule (blue bonds) in C2

Packing plot of C3



Fig. S56 3D arrangement of molecules in crystal packing; made by intermolecular bonds between chloride attached to Pt center and H of the neighbor molecule in C3