SUPPORTING INFORMATION FOR

Quantum-CORMs: Quantum Dot sensitized CO releasing molecules.

A. Ruggi* and F. Zobi*

Département de Chimie, Université de Fribourg, Chemin du Musée 9, 1700 Fribourg, Switzerland. Email: albert.ruggi@unifr.ch, fabio.zobi@unifr.ch.

INDEX

S1. General experimental section and synthetic procedures (pages 1-14)

S2. Synthesis of QD-Mn conjugates and estimation of [Mn]:[QD] ratio (pages 14-15)

S3. UV-Vis spectra of Mn(I) complexes and integral overlapping with CdSe/ZnS QD emission (pages 15-16)

S4. Calculation of the normalised quenching constants (page 16)

S5. Evidence of CO formation upon irradiation (pages 17-18)

S5. References (page 18)

S1. General experimental section and synthetic procedures

General experimental procedures

Reagents were purchased from commercial sources and used as received. CdSe/ZnS quantum dots, 5-amino-2,2'-bipyridine and 5-bromo-2,2'-bipyridine were synthesized according to literature procedures.^[1] ¹H-NMR, ¹³C-NMR, HMQC and HMBC were performed on a Bruker Avance III spectrometer operating at 500 MHz. High resolution mass spectra were measured with a Bruker FTMS 4.7T BioAPEXII spectrometer. IR spectra were measured with a Bruker Tensor 27. UV-Vis spectroscopy was performed using a Perkin-Elmer Lambda 35. Irradiation at 510 nm was performed using an Edinburgh FS5 fluorimeter (excitation slit: 1 nm) equipped with a 150 W Xe lamp. Time-resolved fluorescence decay were obtained using an Edinburgh LifeSpec II

fluorimeter equipped with a LED excitation source (406 nm, repetition rate 100 ns) and collecting the emission light at 520 nm. An excitation vertical polarization and an emission polarization filter oriented at the magic angle 54.7° was used to remove depolarisation effects. The fitting of the lifetime decay was performed with the software provided by Edinburgh. Photodecomposition kinetics were calculated upon fitting of the monoexponential decay by using the software Origin 7.5.

4-[2,2'-bipyridin]-4-ylethynyl)benzoic acid (Ligand L2). 60 mg (0.26 mmol) of 4-bromo-2,2'-bipyridine are dissolved in 20 ml of dry acetonitrile and 6 mg (0.023 mmol) of Pd(CH₃CN)₂Cl₂, 27 mg (0.057 mmol) of XPhos (XPhos = 2-cyclohexylphosphino-2',4',6'-triisopropylbiphenyl) and 200 mg (0.6 mmol) of Cs₂CO₃ are added. The resulting suspension is degassed with 3 Ar/vacuum cycles and stirred 1 h under Ar atmosphere. 45 mg (0.28 mmol) of methyl 4-ethynylbenzoate are added and the resulting mixture is heated at 80°C for 24 h under Ar atmosphere. The solvent is then evaporated and the crude is dissolved in 50 ml of chloroform and washed 3 times with 30 ml of water, then with 30 ml of brine and the organic phase is eventually dried over Na₂SO₄. The solvent is then removed and the resulting solid is purified by crystallization from boiling methanol. The resulting product is dissolved in 30 ml of THF and treated overnight with 10 ml of an aqueous solution of NaOH 1M at room temperature. After THF evaporation, the free acid is precipitated upon acidification of the aqueous solution with HCl. Obtained 20 mg of pure product (0.07 mmol; 30% overall).

¹H-NMR (500 MHz, d6-DMSO): δ (ppm)= 13.26 (br s, 1H), 8.78 (d, 2H, J = 5 Hz), 8.73 (d, 2H, J = 4 Hz), 8.52 (s, 1H), 8.44 (d, 1H, J = 7 Hz), 8.06-7.99 (m, 3H), 7.77 (d, 2H, J = 6 Hz), 7.66 (d, 1H, J = 6 Hz), 7.54 (t, 1H, J = 6 Hz). ¹³C-NMR (125 Hz, d6-DMSO) δ = 156.5, 155.9, 153.6, 149.6, 148.9, 138.2, 132.0, 131.4, 131.0, 129.6, 125.7, 126.4, 124.9, 92.9, 89.9. HR-ESI (m/z): Calc. for C₁₉H₁₃N₂O₂ [M + H]⁺: 301.0977. Found: 301.0973.





Figure S1. ¹H NMR (top) and ¹³C NMR (bottom) of ligand L2 in d6-DMSO.

4-[2,2'-bipyridin]-4-ylethynyl)aniline (Ligand L4). 540 mg of 4-bromo-2,2'-bipyridine (2.3 mmol), (236 mg, (0.5 mmol) of XPhos, 1.7 mg of Cs₂CO₃ (5 mmol) and 49 mg of Pd(CH₃CN)₂Cl₂ (0.19 mmol) are dissolved in 50 ml of CH₃CN. The mixture is degassed with three Ar/vacuum cycles and stirred at room temperature for 1 hour before adding 292 mg of 4-ethynylaniline (2.5 mmol). The mixture is refluxed for 2 days under N₂. The solvent is removed and the brownish oil is purified by column chromatography on a SiO₂ column using CH₂Cl₂/MeOH, 98:2 as eluting solvent to yield ligand **L4** (370 mg, 1.38 mmol; 60%). 100 mg. ¹H-NMR (500 MHz, CDCl₃): δ (ppm): 8.71 (d, 1H, J = 5 Hz), 8.63 (d, 1H, J = 5 Hz), 8.48 (s, 1H), 8.41 (d, 1H, J = 8 Hz), 7.64 (t, 1H, J = 8 Hz), 7.38-7.33 (m, 4H), 6.66 (d, 2H, J = 8 Hz). ¹³C-NMR (125 Hz, CDCl₃): δ (ppm) = 156.0, 155.7, 149.2, 149.0, 147.5, 137.0, 133.5, 133.2, 125.0, 124.0, 123.0, 121.1, 114.7, 111.4, 95.4, 85.4. HR-ESI (m/z) : Calc. for C₁₈H₁₄N₃ [M + H]⁺: 272.1188. Found: 272.1180.



Figure S2. ¹H NMR (top) and ¹³C NMR (bottom) of ligand L4 in CDCl₃.

Synthesis of *fac*-[Mn(CO)₃LBr] complexes.

Species 1-4 were synthesized according to the procedure described by Sampson et al. with slight modifications.^[2] Briefly. $Mn(CO)_5Br$ (100 mg, 0.37 mmol) was added to 15 mL of diethyl ether (Et₂O) in ambient air. The selected ligand (0.37 mmol) was added to the mixture and heated to reflux. For species 2-4 the solution turned orange within 60 min, and the product precipitated out of solution. After 3 h, the mixture was cooled to room temperature, and the precipitate was filtered off and washed with Et₂O. The orange solid was then dried overnight under vacuum. For species 1 the precipitate was identified as a mixture of unreacted ligand and $Mn(CO)_5Br$. The reaction mixture was filtered off and the solution containing 1 was dried under vacuum. Compounds were then purified via HPLC. All compounds show a dynamic equilibrium

in the presence of coordinating solvents. Such equilibrium can be shifted towards the formation of a single species upon addition of an excess of KCN, as evidenced by the NMR shown in Figure S8. Therefore when needed (complexes 1, 2, and 4) KCN was added to the NMR solutions.

<u>Analytical HPLC method.</u> Instrument: MERCK HITACHI LaChrom with a D-7000 interface coupled with a Diode Array detector L-7455 and a pump L-7100 system. Column: Macherey-Nagel, EC250/3 Nucleosil 100-5 C18. Flow rate 0.5 mL/min. Absorbance monitored at 250 nm. Solutions: A: 0.1% trifluoroacetic acid in water; B: methanol. Chromatographic method: 0-5 min: isocratic flow of 75% A - 25% B; 5-35 min: linear gradient to 100% B; 35-45 min: isocratic flow of 100% B.

Analytical data for 1: Yield 65 mg (24%). HPLC retention time (min): 21.17. ¹H-NMR (500 MHz, CD₃OD + KCN): δ (ppm) = 9.11 (d, 2H, J = 6 Hz), 8.88 (s, 2H), 8.02 (d, 2H, , J = 6 Hz). ¹³C-NMR (125 MHz, CD₃OD + KCN): see figure S3. IR (KBr) vCO: 2027, 1943, 1913, 1733 cm⁻¹. HR-ESI (m/z) : Calc. for C₁₅H₇BrMn N₂O₇ [M - H]⁻: 460.8823. Found: 460.8826.



Sample: AR.MnbbyCCNH2 / Solvent: CD3OD / Temp.: 293.0 K



6



Figure S3. (from top to bottom) ¹H-NMR, ¹³C-NMR, HMQC/HMBC-NMR (aromatic region) spectra (CD₃OD + KCN) with ¹H and ¹³C signals assignment of 1 and HPLC chromatogram. * = KCN methanolysis product, ** = KCN.

Analytical data for **2**: Yield 150 mg (78%). HPLC retention time (min): 27.99. IR (KBr) vCO: 2026, 1925, 1736 cm⁻¹. HR-ESI (m/z) : Calc. for $C_{22}H_{11}BrMn N_2O_5$ [M - H]⁻: 518.9217. Found: 518.9218. Although this species was isolated as a single HPLC peak, it undergoes a rapid equilibrium (see Figure S5) giving two HPLC peaks having identical mass (m/z : 439.0 [M-Br]⁺). This equilibrium mixture cannot be completely shifted by addition of KCN, therefore the ¹H-NMR spectrum of the unresolved mixture is given, along with the tentative peak assignment of the major and minor species (relative ratio: 1.4:1).

Major species: ¹H-NMR (500 MHz, CD₃OD + KCN): δ (ppm) = 9.10 (d, 1H, J= 5Hz), 9.06 (d, 1H, J = 6 Hz), 8.67-8.65 (m, 2H), 8.21 (t, 1H, J = 7 Hz), 8.0 (d, 2H, J = 8 Hz), 7.73 (d, 1H, J = 6 Hz), 7.70 (t, 1H, J = 7 Hz), 7.64 (d, 2H, J = 8 Hz).

Minor species: ¹H-NMR (500 MHz, CD₃OD + KCN): δ (ppm) = 8.60 (d, 2H, J = 8 Hz), 8.40 (s, 1H), 8.34 (d, 1H, J = 8Hz), 7.99-7.94 (m, 3H), 7.58 (d, 2H, J = 8Hz), 7.53 (d, 1H, J = 5 Hz), 7.47 (t, 1H, J = 7 Hz).

MnbpyCCCO2H + KCN / Solvent: CD3OD / Temp.: 293.0 K / after cooling to r.t.



Figure S4 ¹H-NMR of 2 (CD₃OD + KCN). * = KCN methanolysis product.



HPLC chromatogram of purified 2.



HPLC chromatogram of same compound above (i.e. purified **2**) left in solution for 10 min and then re-injected.



Analytical data for **3**: Yield 90 mg (62%). HPLC retention time (min): 21.95. ¹H-NMR (500 MHz, d6-DMSO + D₂O): δ (ppm) = 9.02 – 9.00 (m, 2H), 8.83 (d, 1H, J = 6 Hz), 8.77 (d, 1H, J = 8 Hz), 8.29 (t, 1H, J = 8 Hz), 7.97 (dd, 1H, J = 6 Hz), 7.75 (t, 1H, J = 6 Hz.). ¹³C-NMR (125 MHz, d6-DMSO+D₂O): see figure S6. IR (KBr) vCO: 2022, 1940 cm⁻¹. HR-ESI (m/z) : Calc. for C₁₃H₉Mn N₃O₃ [M - Br]⁺: 310.0024. Found: 310.0024.

Sample: AR.MnbpyNH2 / Solvent: DMSO-d6 + trace of D2O / Temp.: 298.0 K







Figure S6. (from top to bottom) ¹H-NMR, ¹³C-NMR, HMQC/HMBC-NMR (aromatic region) spectra (d6-DMSO + trace of D_2O) with ¹H and ¹³C signals assignment of **3** and HPLC chromatogram.

Analytical data for 4: Yield 108 mg (60%). HPLC retention time (min): 26.45. ¹H-NMR (500 MHz, CD₃OD+KCN): δ (ppm) = 9.09 (d, 1H, J = 5 Hz), 8.97 (d, 1H, J = 6 Hz), 8.57 (d, 1H, J = 8 Hz), 8.53 (s, 1H), 8.20 (t, 1H, J = 8 Hz), 7.69 (t, 1H, J = 7 Hz), 7.61 (d, 1H, J = 6 Hz), 7.35 (d, 2H, J = 7 Hz), ¹³C-NMR (125 MHz, CD3OD + KCN): see figure S7. IR (KBr) vCO: 2023, 1928, 1910 cm⁻¹. HR-ESI (m/z) : Calc. for C₂₁H₁₃Mn N₃O₃ [M - Br]⁺: 410.0337. Found: 410.0331.





Figure S7. (from top to bottom) ¹H-NMR, ¹³C-NMR, HMQC/HMBC-NMR (aromatic region) spectra $(CD_3OD + KCN)$ with ¹H and ¹³C signals assignment of **4** and HPLC chromatogram.. ** = KCN.



Figure S8. Evidence of ligand exchange equilibrium upon addition of KCN to a CD₃OD solution of 4.

S2. Synthesis of QD-Mn conjugates and estimation of [Mn]:[QD] ratio.

0.5 ml of the stock solution0.038 mM of QD in THF were added to 5 equivalents of Mn complex suspended in 1.5 ml of THF (in the case of complexes **3** and **4**, the compounds were previously treated with 0.1 ml of CS_2 for 1h and then dried under vacuum). The resulting mixture was stirred overnight in the dark and then centrifuged to remove unreacted complex. The conjugated were then characterised by UV-Vis and used without further purification. For the irradiation test, 0.1 ml of this solution were dissolved in 1.8 ml of THF and 0.1 ml of MeOH.

To evaluate the [Mn]:[QD] ratio, the UV-Vis of the conjugated was compared with an equimolar solution of QD prepared by following the same procedure but without addition of Mn complexes (Figure S9). The estimation of the [Mn]:[QD] ratio in conjugate **QD-1** is here given as example.

By assuming the additivity of absorption spectra, the Mn contribution to the absorption at 313 nm 313 Abs(Mn)_{QD-1} was evaluated using the equation

 $^{313}Abs(Mn)_{QD-1} = ^{313}Abs_{QD-1} - ^{313}Abs_{QD}$

where ${}^{313}Abs_{QD-1}$ and ${}^{313}Abs_{QD}$ are the absorption of the QD-1 conjugate solution and of the QD solution at 313 nm, respectively.

[Mn] concentration is then evaluated using Lambert-Beer equation and the previously determined ε of **1**. [QD] is evaluated using the literature procedure.^[3]

In the case of **QD-1**, the obtained values are ${}^{313}Abs(Mn)_{QD-1} = 0.13$ $[Mn]_{QD-1} = 0.13 / 15063 = 8.6 \cdot 10^{-5} M.$ $[QD] = 5 \cdot 10^{-6} M$



Figure S9. UV-Vis absorption spectrum of equimolar solutions of QD and of QD-Mn used for the estimation of [Mn]:[QD] ratio.

S3. UV-Vis spectra of Mn(I) complexes and integral overlapping with CdSe/ZnS QD emission.





The overlapping integral (J) was calculated using the software a/e UV-Vis-IR Spectral Software (v. 2.0) available on www.fluorotools.com.

S4. Calculation of the normalised quenching constants

Quenching constants normalised according to the number of Mn(I) complexes attached to the CdSe/ZnS quantum dots were obtained by using the equation^[4]

$$nk_q = \frac{1}{\tau} - \frac{1}{\tau_0}$$

where τ and τ_0 are the average emission lifetimes in the presence and in the absence of quencher, respectively and n is the number of Mn complexes.

S5. Evidence of CO evolution upon irradiation.

Qualitative evidence of CO evolution was obtained via myoglobin (Mb) assay.^[5] Complexes **1-4** were dissolved in DMSO and added as a 10-fold molar excess to an aqueous solution of reduced horse heart Mb having a 26 μ M concentration in the case of complexes **1** and **4** and a 31 μ M concentration in the case of complexes **2** and **3**. In all cases denaturation of the protein (and subsequent scattering) occurs within the first 15 minutes of irradiation. Prolonged irradiation leads to a complete denaturation and precipitation of the protein with consequent loss of signal. The lower Mb concentration in the case of complexes **1** and **4** is due to our efforts to minimize the effect outlined above.



Figure S10. Changes occurring in the UV-Vis spectra of a reduced myoglobin solution (black) upon addition of Mn(I) complexes 1-4 and subsequent irradiation at 510 nm (red).

In the case of QD-Mn systems a THF solution was used due to the insolubility of such systems in DMSO. A pronounced scattering was observed upon addition the QD-Mn system to the 26 μ M aqueous Mb solution (as evidenced in Figure S11, see black and blue lines) as a consequence of QD-Mn precipitation in water. However, qualitative evidence of CO release upon irradiation (red line) could still be obtained within the timeframe of the experiment (15 min.). Analogously to what observed for complexes 1-4, prolonged irradiation leads to a complete denaturation and precipitation of the protein with consequent loss of signal. The typical spectral changes are shown in Figure S11 for QD-3. All other species show similar behaviours.



Figure S11. Changes occurring in the UV-Vis spectra of a reduced myoglobin solution (black) upon addition of **QD-3** system (blue) and subsequent irradiation at 510 nm (red).

S6. References

- a) A. Baron, C. Herrero, A. Quaranta, M.-F. Charlot, W. Leibl, B. Vauzeilles, A. Aukauloo, *Inorg. Chem.* 2012, *51*, 5985-5987; b) M. Zalas, B. Gierczyk, M. Klein, K. Siuzdak, T. Pedzinski, T. Luczak, *Polyhedron* 2014, *67*, 381-387; c) S. Impellizzeri, S. Monaco, I. Yildiz, M. Amelia, A. Credi, F. M. Raymo, *J. Phys. Chem. C* 2010, *114*, 7007-7013.
- [2] M. D. Sampson, A. D. Nguyen, K. A. Grice, C. E. Moore, A. L. Rheingold, C. P. Kubiak, J. Am. Chem. Soc. 2014, 136, 5460-5471.
- [3] W. W. Yu, L. H. Qu, W. Z. Guo, X. G. Peng, Chem. Mater. 2003, 15, 2854-2860.
- [4] P. T. Burks, A. D. Ostrowski, A. A. Mikhailovsky, E. M. Chan, P. S. Wagenknecht, P. C. Ford, *J. Am. Chem. Soc.* **2012**, *134*, 13266-13275.
- [5] R. Motterlini, B. E. Mann, T. R. Johnson, J. E. Clark, R. Foresti and C. J. Green, *Curr. Pharm. Des.*, **2003**, *9*, 2525-2539.