

Supplementary Information for

J. Malina, P. Scott, V. Brabec: Recognition of DNA/RNA bulges by antimicrobial and antitumor metallohelicenes

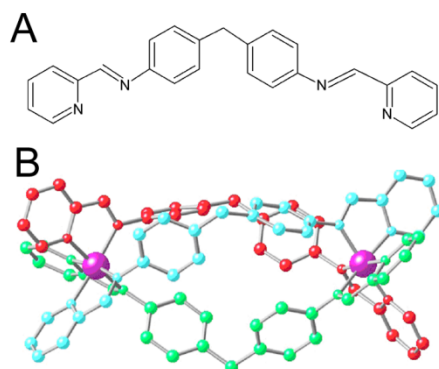


Figure S1. (A) Structure of **I** ($\text{I} = \text{C}_{25}\text{H}_{20}\text{N}_4$) and (B) 3D structure of $M\text{-}[\text{Fe}_2(\text{I})_3]^{4+}$.

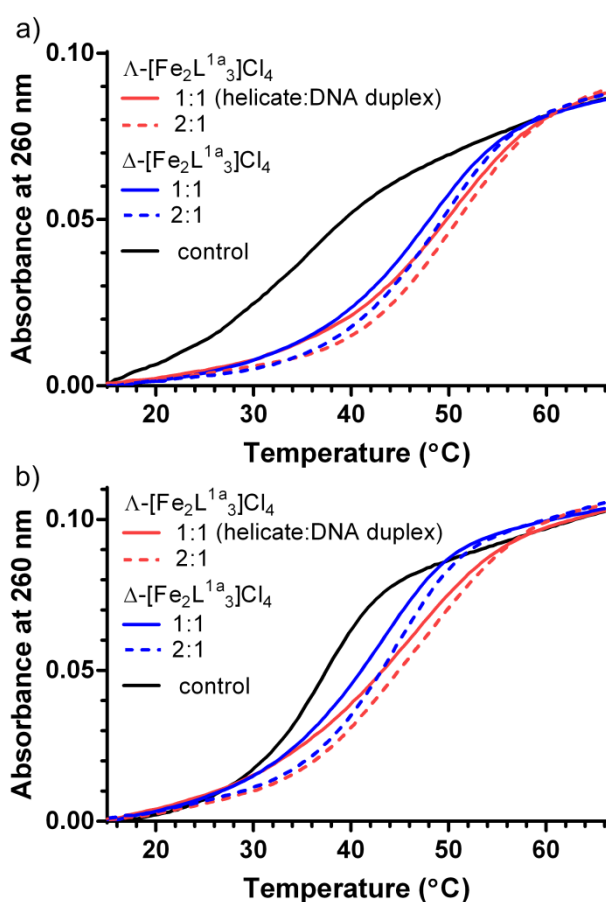


Figure S2. Melting curves for G-C2-G (a) and C-A2-C (b) DNA bulges in the presence of Λ - and Δ - $[\text{Fe}_2\text{L}^{\text{1a}_3}]\text{Cl}_4$.

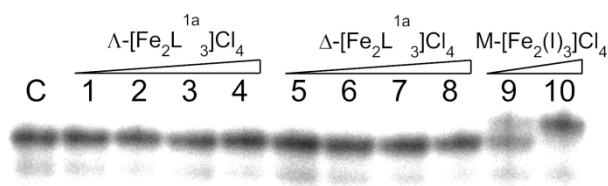


Figure S6. Autoradiogram of the gel run at 5 °C. Lane C: control containing the TAR RNA in the buffer in the absence of the flexicates. Lanes 1-4: TAR RNA mixed with Λ -[Fe₂L^{1a}₃]Cl₄ at 0.5:1, 1:1, 1.5:1 and 2:1 (flexicate:TAR RNA) ratios, respectively. Lanes 5-8: TAR RNA mixed with Δ -[Fe₂L^{1a}₃]Cl₄ at 0.5:1, 1:1, 1.5:1 and 2:1 (flexicate:TAR RNA) ratios, respectively. Lanes 9, 10: TAR RNA mixed with the *M*-[Fe₂(I)₃]Cl₄ at 0.5:1 and 1:1 (flexicate:TAR RNA) ratios, respectively.

The sample of control TAR RNA yielded an extra faint band migrating slightly faster than the major strong band. The presumed cause of this observation is a lower stability of RNA (in comparison with DNA). RNA decomposes relatively rapidly and had to be prepared for each experiment fresh by its isolation from the polyacrylamide gel. It implies that the second band in the Figure S6 very likely corresponds to degraded TAR RNA from which one nucleotide on the 3' side was cut off. Nevertheless, the results indicate that the flexicates do not bind TAR RNA strongly enough to ensure intactness of the resulting complex during its migration in the gel.