

Synthesis of the first metal-free phosphanylphosphonate and its use in the “phospha-Wittig-Horner” Reaction

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Supporting Information

Experimental Details

Synthesis of phosphaalkene 4 via the phospha-Wittig-Horner reaction – pWH reagent (0.25 mmol, 104 mg) was exposed to the reaction conditions as specified in the main manuscript and the reaction mixture purified by column chromatography (silica gel, heptane/ethyl acetate, 4:1) to give a mixture of isomers (yellow solid). Yield: 69 mg, 71%. The chemical shifts of *E*-4 match the NMR data reported in the literature.¹ ¹H NMR and ³¹P NMR data for *Z*-4 were however detectable from the spectrum of the pure isomeric mixture (Appendices, Figure S4): ¹H NMR (CDCl₃, 399.8 MHz): δ 7.79 (d, ²J_{H-P} = 36.9 Hz, 1H, P=CH), 7.63 (s, 2H), 7.19 (d, ³J_{H-H} = 8.4 Hz, 2H, ArH), 6.20 (d, ⁴J_{H-P} = 8.5, 2.2 Hz, 2H, ArH), 1.43 (s, 18H), 1.40 (s, 9H). ³¹P{¹H} NMR (CDCl₃, 161.8 MHz): δ 265.3 (s).

Both column chromatography and oxalyl chloride treatment were tested as the purification processes. The former afforded pure phosphaalkene 4 as a mixture of isomers but the proportion of the *Z* isomer increased after the purification. The latter gave the phosphaalkene products with high *E*-selectivity without changing the *E/Z* ratio through the purification process (Figure S1), although some traces of unknown impurities was detectable. It is noteworthy that 4-cyanobenzaldehyde does not dissolve in pentane and was filtered off before the addition of oxalyl chloride.

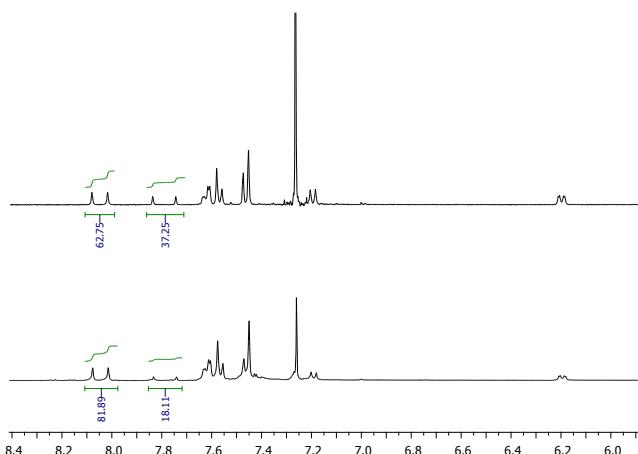


Figure S1 – ¹H NMR spectra of 4 in two different reactions, purified with column chromatography (top) or oxalyl chloride treatment (bottom). The doublets at 8.04 and 7.79 ppm correspond to the P=CH of *E*-4 and *Z*-4, respectively. According to the integration of these protons, it is clearly shown that oxalyl chloride addition keeps the *E*-stereoselectivity high even after the purification.

Synthesis of phosphaalkene **6² via the phospha-Wittig-Horner reaction** – pWH reagent (0.25 mmol, 104 mg) was applied to the reaction conditions to give a crude mixture which was chromatographed on silica gel using pentane/ethyl acetate, 95:5, as the eluent. Final product was isolated as yellow solid with small impurity of Z-**6**. Yield: 53 mg, 65%. ¹H NMR (CDCl₃, 399.8 MHz): δ 8.12 (d, ²J_{H,P} = 24.0 Hz, 1H, P=CH), 7.44 (s, 2H, ArH), 7.17 (d, ³J_{H,H} = 5.4 Hz, 1H), 6.92–6.99 (m, 2H), 1.53 (s, 18H, *ortho*-C(CH₃)), 1.35 (s, 9H, *para*-C(CH₃)). ³¹P{¹H} NMR (CDCl₃, 161.8 MHz): δ 246.8 (s). ¹³C{¹H} NMR (CDCl₃, 100.5 MHz): δ 166.2 (d, ¹J_{C,P} = 30.8 Hz, P=C), 154.3 (d, ³J_{C,P} = 1.2 Hz, *ortho*-ArC), 150.0 (s, *para*-ArC), 145.3 (d, ²J_{C,P} = 17.3 Hz, Thienyl-C), 138.5 (d, ¹J_{C,P} = 53.1 Hz, *ipso*-ArC), 127.8 (d, J = 4.2 Hz), 125.6 (d, J = 22.7 Hz), 125.0 (d, J_{C,P} = 14.2 Hz, Thienyl-C), 121.9 (d, ³J_{C,P} = 1.4 Hz, *meta*-ArC), 38.4 (s, *ortho*-C(CH₃)₃), 35.1 (s, *para*-C(CH₃)₃), 34.0 (d, ⁴J_{C,P} = 6.9 Hz, *ortho*-C(CH₃)₃), 31.5 (s, *para*-C(CH₃)₃).

X-ray diffraction data

All measurements were performed using graphite-monochromatized Mo K_α radiation at 100K using a Bruker D8 APEX-II equipped with a CCD camera. The structure was solved by direct methods (SHELXS-2014) and refined by full-matrix least-squares techniques against F² (SHELXL-2014/7).^[1] The non-hydrogen atoms were refined with anisotropic displacement parameters. The H atoms of the CH₂ groups were refined with common isotropic displacement parameters for the H atoms of the same group and idealized geometry with approximately tetrahedral angles and C-H distances of 0.99 Å. The H atoms of the methyl groups were refined with common isotropic displacement parameters for the H atoms of the same group and idealized geometry with tetrahedral angles, enabling rotation around the X-C bond, and C-H distances of 0.98 Å.

Table 1. Crystal data and structure refinement for compounds **2-H** and **Z-5**.

Crystal data	Compound 2-H		Compound Z-5	
CCDC-No.	1423585		1423584	
Empirical formula	C ₂₂ H ₄₀ O ₃ P ₂		C ₃₀ H ₃₉ P ₁	
Formula weight	414.48		430.58	
Crystal description	colorless block		colorless needle	
Crystal size	0.3x0.26x0.22		0.30x0.12x0.12	
Crystal system, space group	triclinic, P-1		triclinic, P-1	
Unit cell dimensions:	a b c	9.5128(3) 10.5246(4) 12.6133(5)	10.3513(12) 10.3810(13) 24.524(3)	
α, β, γ	74.848(2), 74.761(2), 88.062(2)		94.404(3), 98.422(4), 94.918(4)	
Volume	1175.24(8)		2586.6(6)	
Z	2		4	
Calculated density	1.171		1.106	
F(000)	452		936	
Linear absorption coefficient μ	0.203		0.121	
Absorption correction	multi-scan, SADABS 2008		multi-scan, SADABS 2008	
Max. and min. transmission	0.5095, 0.7456		0.6037 and 0.7454	

Unit cell determination	$1.7 < \theta < 25.2^\circ$	$2.4 < \theta < 25.2^\circ$
	3964 reflections used at 100K	4389 reflections used at 100K
Data collection		
Temperature	100(2)K	100(2)K
Diffractometer	Bruker APEX-II CCD	Bruker APEX-II CCD
Radiation source	fine-focus sealed tube	fine-focus sealed tube
Radiation and wavelength	MoK _a , 0.71073 Å	MoK _a , 0.71073 Å
Monochromator	Graphite	Graphite
Scan type	ω scans	ω scans
Θ range for data collection	1.73 to 27.96°	2.41 to 26.45°
Index ranges	$-12 \leq h \leq 11, -13 \leq k \leq 13, -16 \leq l \leq 16$	$-12 \leq h \leq 12, -12 \leq k \leq 12, -30 \leq l \leq 30$
Reflections collected / unique	18027 / 5613	21308 / 10361
Significant unique reflections	3964 with $I > 2\sigma(I)$	4389 with $I > 2\sigma(I)$
R(int), R(sigma)	0.0492, 0.0758	0.1187, 0.2347
Completeness to Θ_{\max}	99.2%	97.5%
Refinement		
Refinement method	Full-matrix least-squares on F ²	Full-matrix least-squares on F ²
Data / parameters / restraints	5613/ 259/ 0	10361/ 559/ 0
Goodness-of-fit on F ²	1.067	0.956
Final R indices [$I > 2\sigma(I)$]	R1 = 0.0480, wR2 = 0.1106	R1 = 0.0828, wR2 = 0.1658
R indices (all data)	R1 = 0.0810, wR2 = 0.1237	R1 = 0.2216, wR2 = 0.2228
Weighting scheme	w=1/[$\sigma^2(F_o^2)+(aP)^2+bP$] where P=(F _o ² +2F _c ²)/3	w=1/[$\sigma^2(F_o^2)+(aP)^2$], where P=(F _o ² +2F _c ²)/3
Weighting scheme parameters a, b	0.0564, 0.0511	0.0936
Largest Δ/σ in last cycle	0.000	0.001
Largest difference peak and hole	0.470 and -0.590 e/Å ³	0.383 and -0.322 e/Å ³
Structure Solution Program	SHELXS-2014 (Sheldrick, 2008)	SHELXS-2014 (Sheldrick, 2008)
Structure Refinement Program	SHELXL-2014 (Sheldrick, 2008)	SHELXL-2014 (Sheldrick, 2008)

References

- 1 A. Termaten, M. van der Sluis and F. Bickelhaupt, *Eur. J. Org. Chem.*, 2003, **2003**, 2049–2055.
- 2 M. van der Sluis, A. Klootwijk, J. B. M. Wit, F. Bickelhaupt, N. Veldman, A. L. Spek and P. W. Jolly, *J. Organomet. Chem.*, 1997, **529**, 107–119.

Appendices

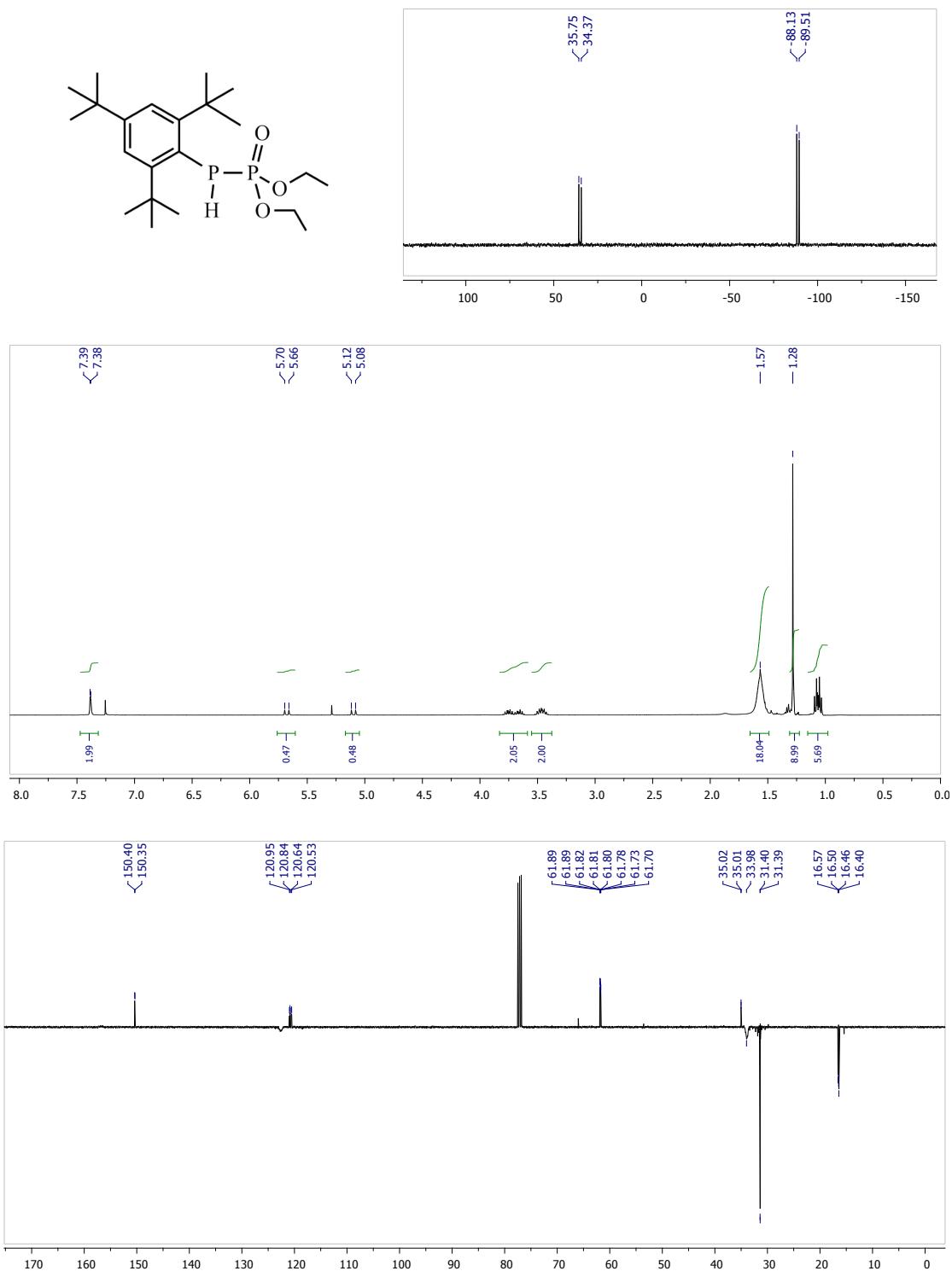
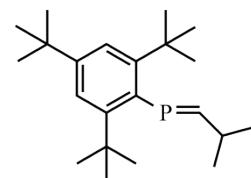


Figure S2 – ^{31}P NMR (top), ^1H NMR (middle) and APT (bottom) spectra of phosphanylphosphonate **2-H.**



(*E*)-3

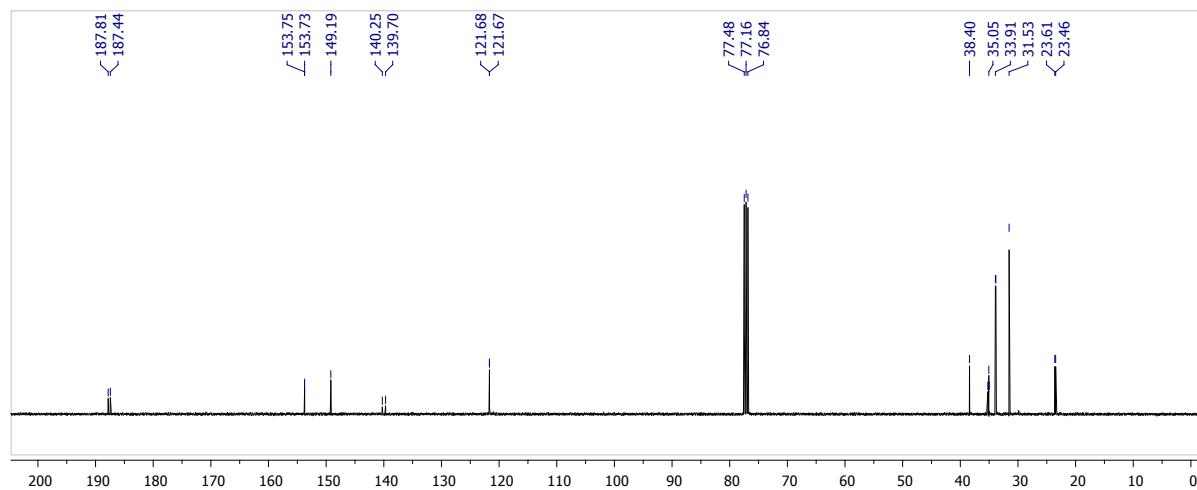
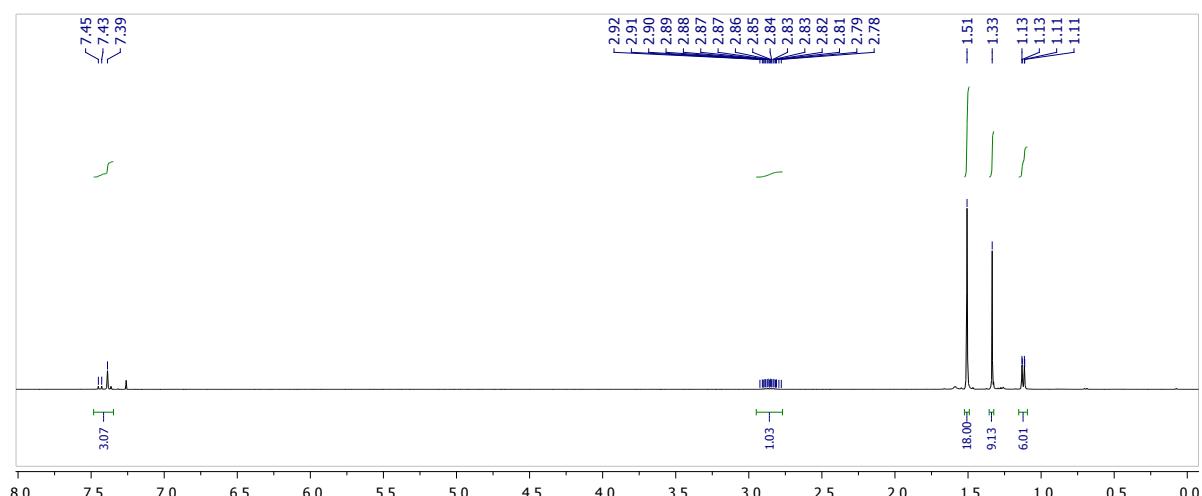
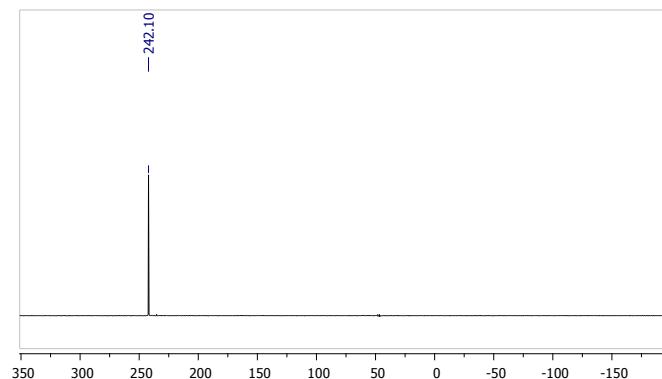
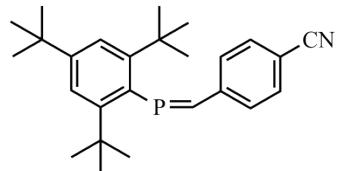


Figure S3 – ^{31}P NMR (top), ^1H NMR (middle) and $^{13}\text{C}\{^1\text{H}\}$ NMR (bottom) spectra of phosphaalkene *E*-3.



(Z)-4

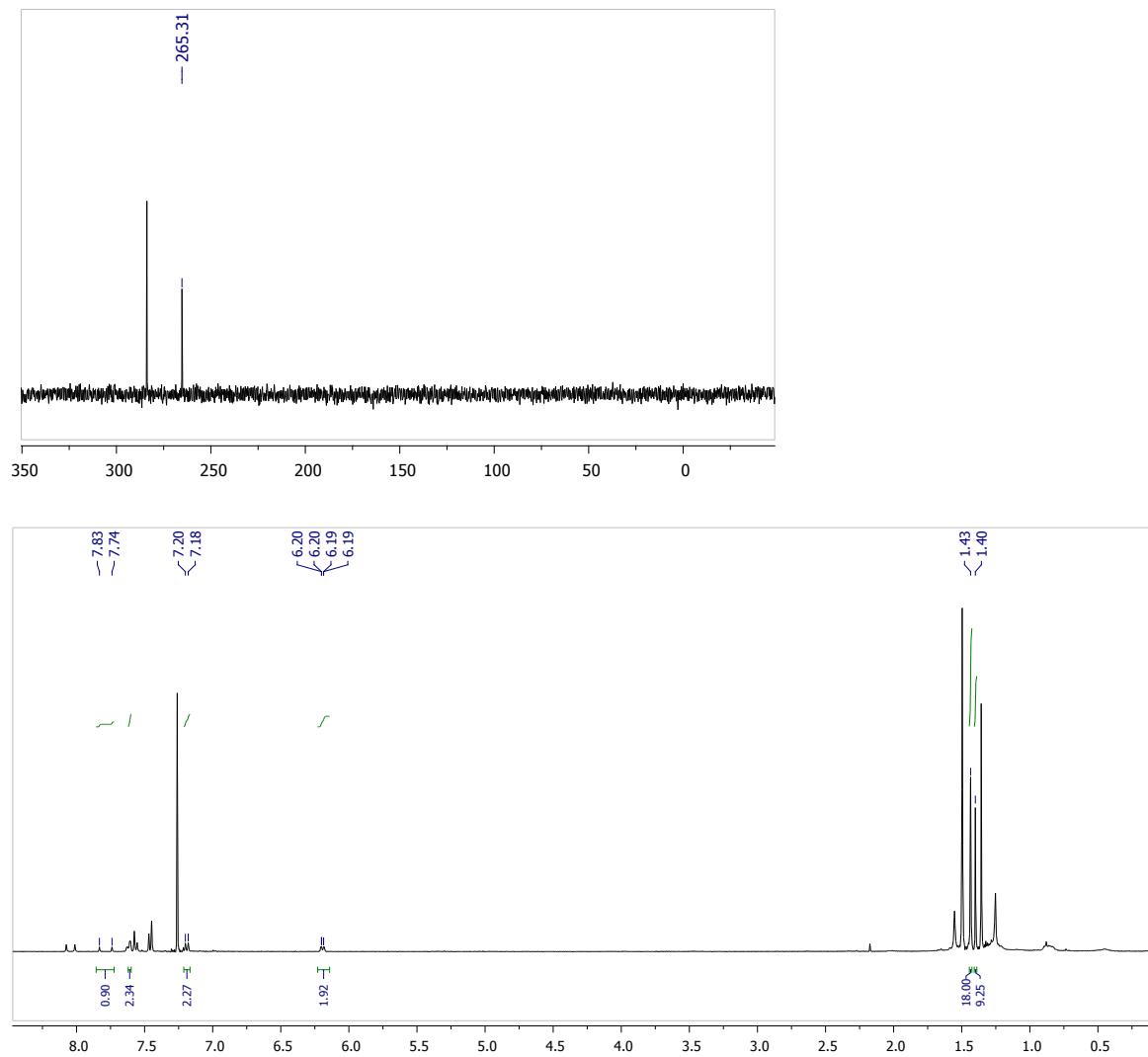
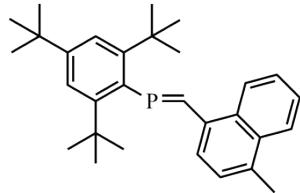


Figure S4 – ^{31}P NMR (top), ^1H NMR (bottom) spectra of phosphaalkene **4**, as mixture of the two isomers. The assigned signals belong to Z-4.



(E)-5

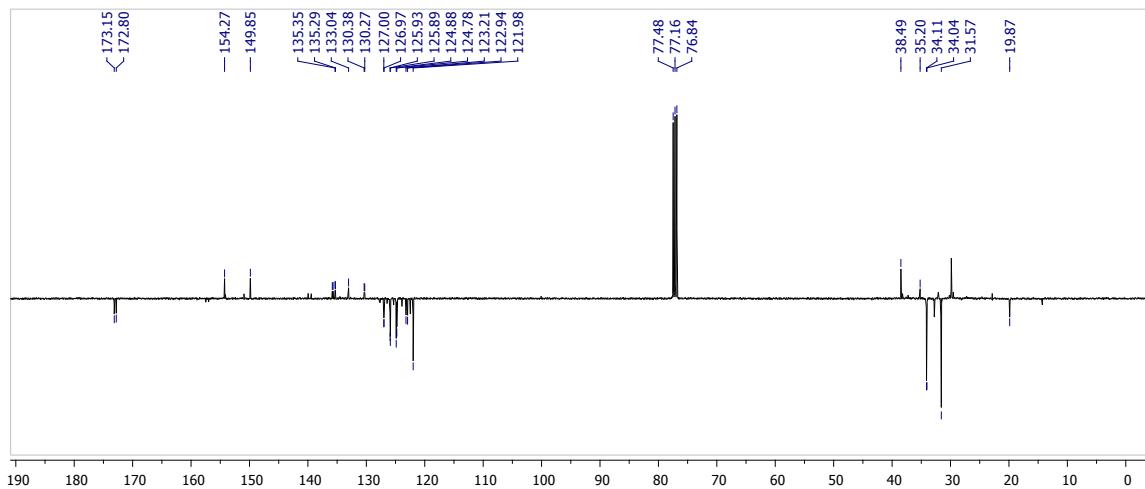
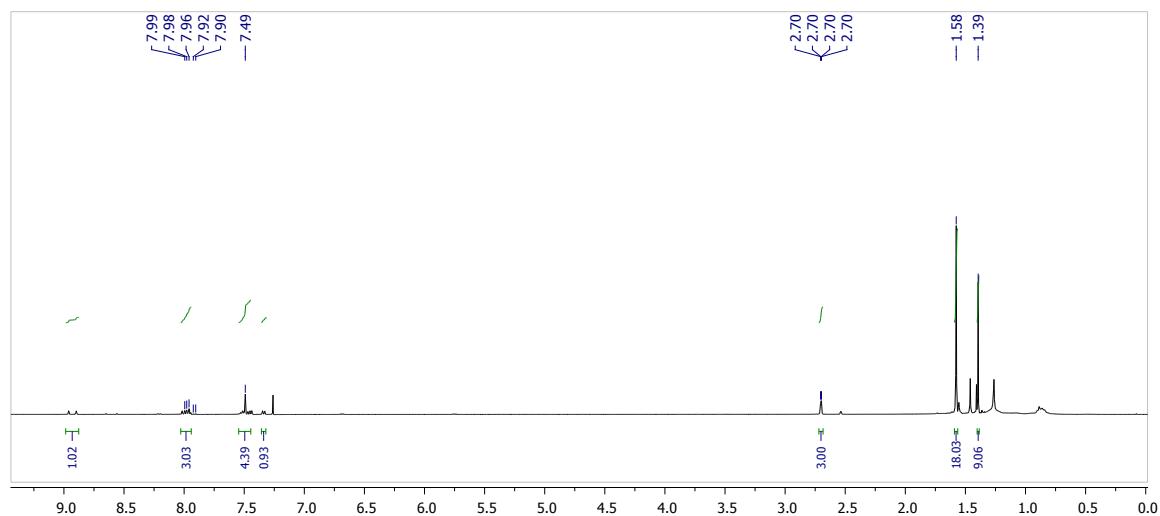
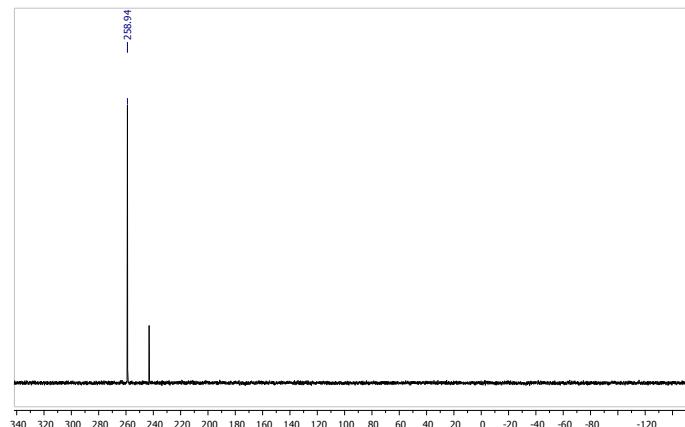
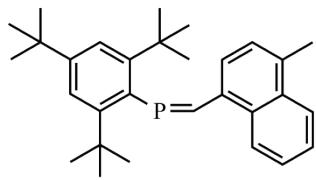


Figure S5 – 31P NMR (top), 1H NMR (middle) and APT NMR (bottom) spectra of phosphaalkene *E*-5. The small peaks observable in these spectra belong to the other isomer, *Z*-5.



(*Z*)-5

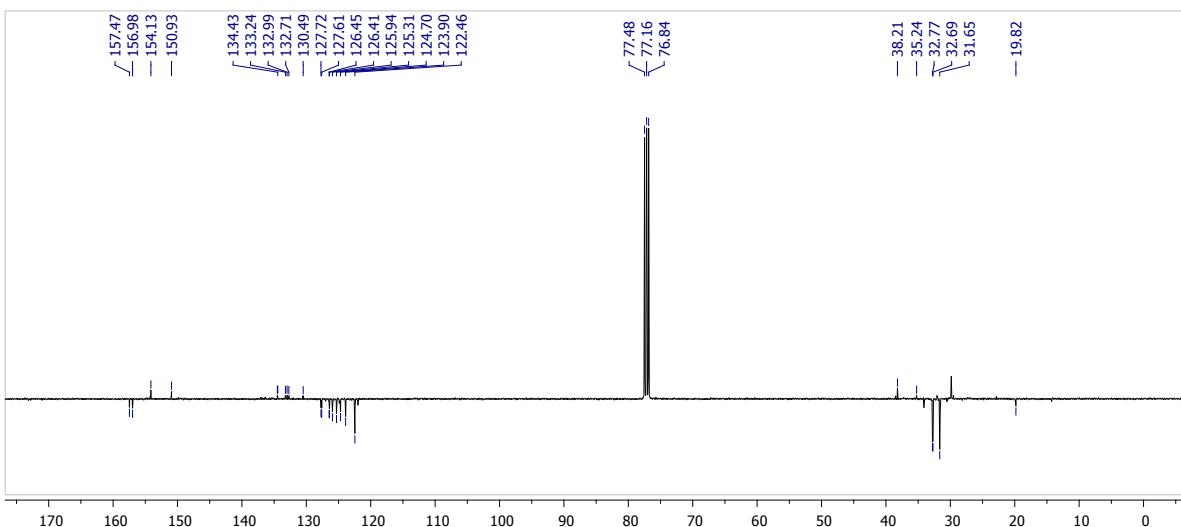
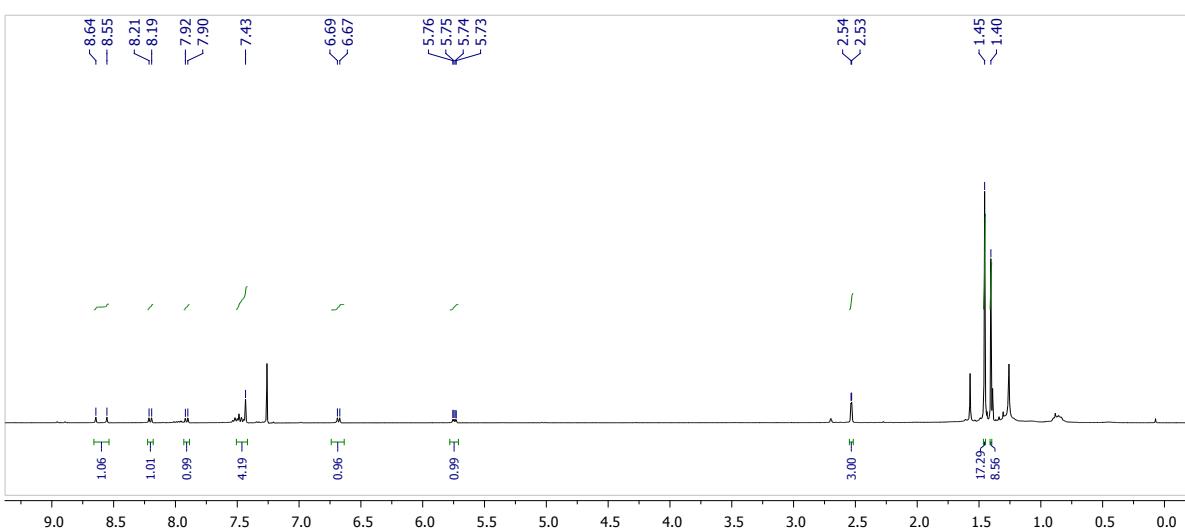
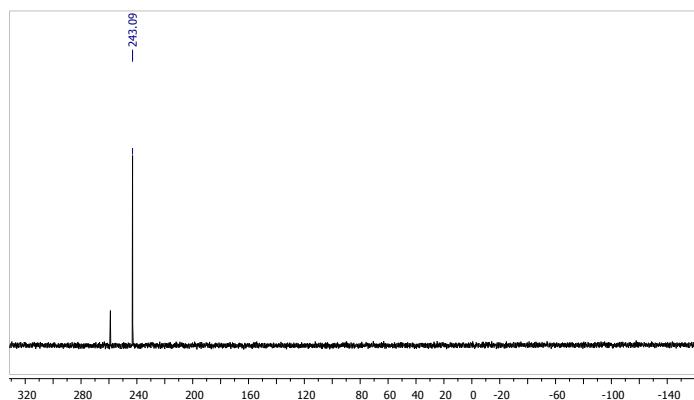


Figure S6 – ^{31}P NMR (top), ^1H NMR (middle) and APT NMR (bottom) spectra of phosphaalkene Z-5. The small peaks observable in these spectra belong to the other isomer, E-5.

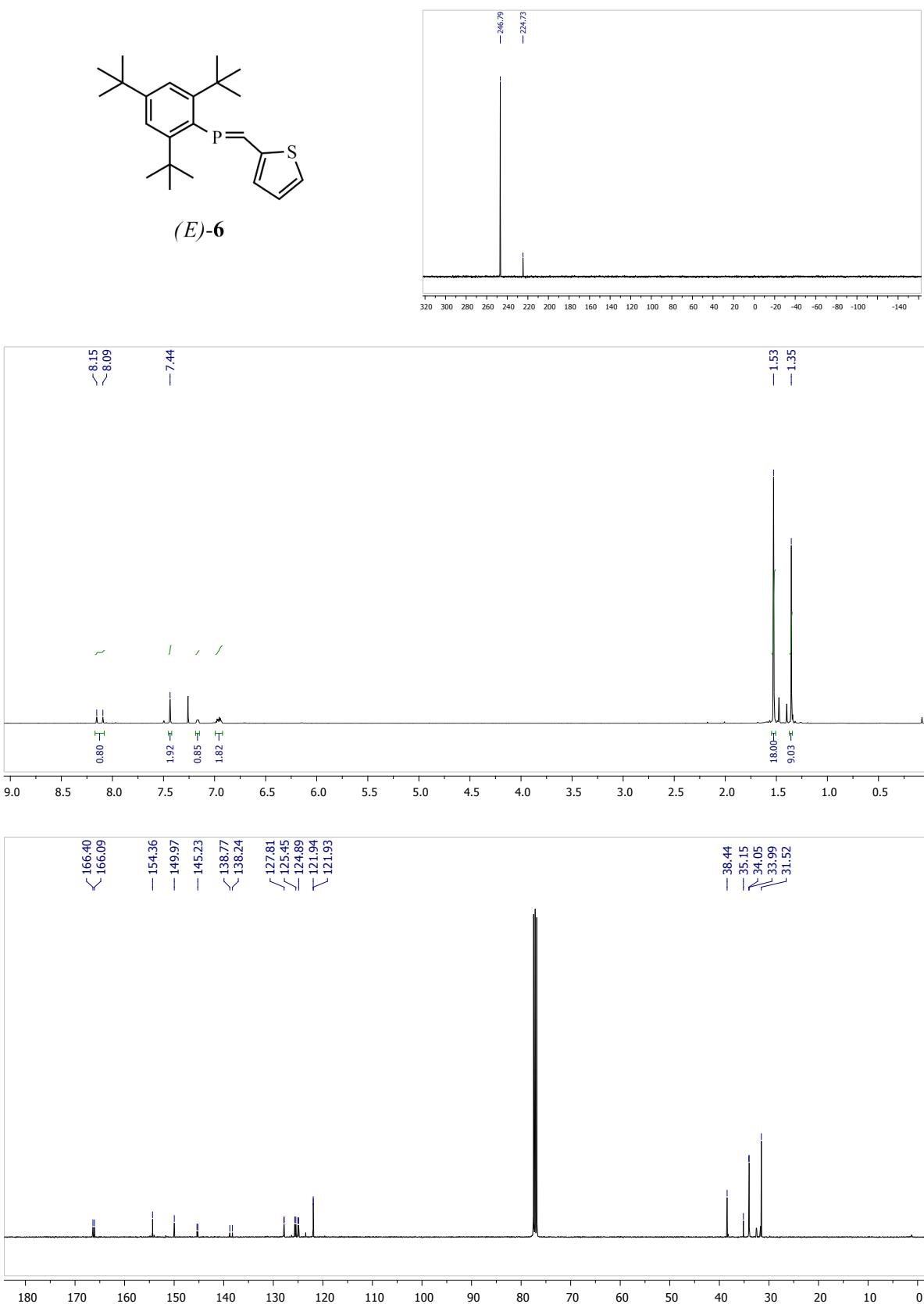


Figure S7 – ^{31}P NMR (top), ^1H NMR (middle) and $^{13}\text{C}\{^1\text{H}\}$ NMR (bottom) spectra of phosphaalkene E -6. The small peaks observable in these spectra belong to the other isomer, E -6.

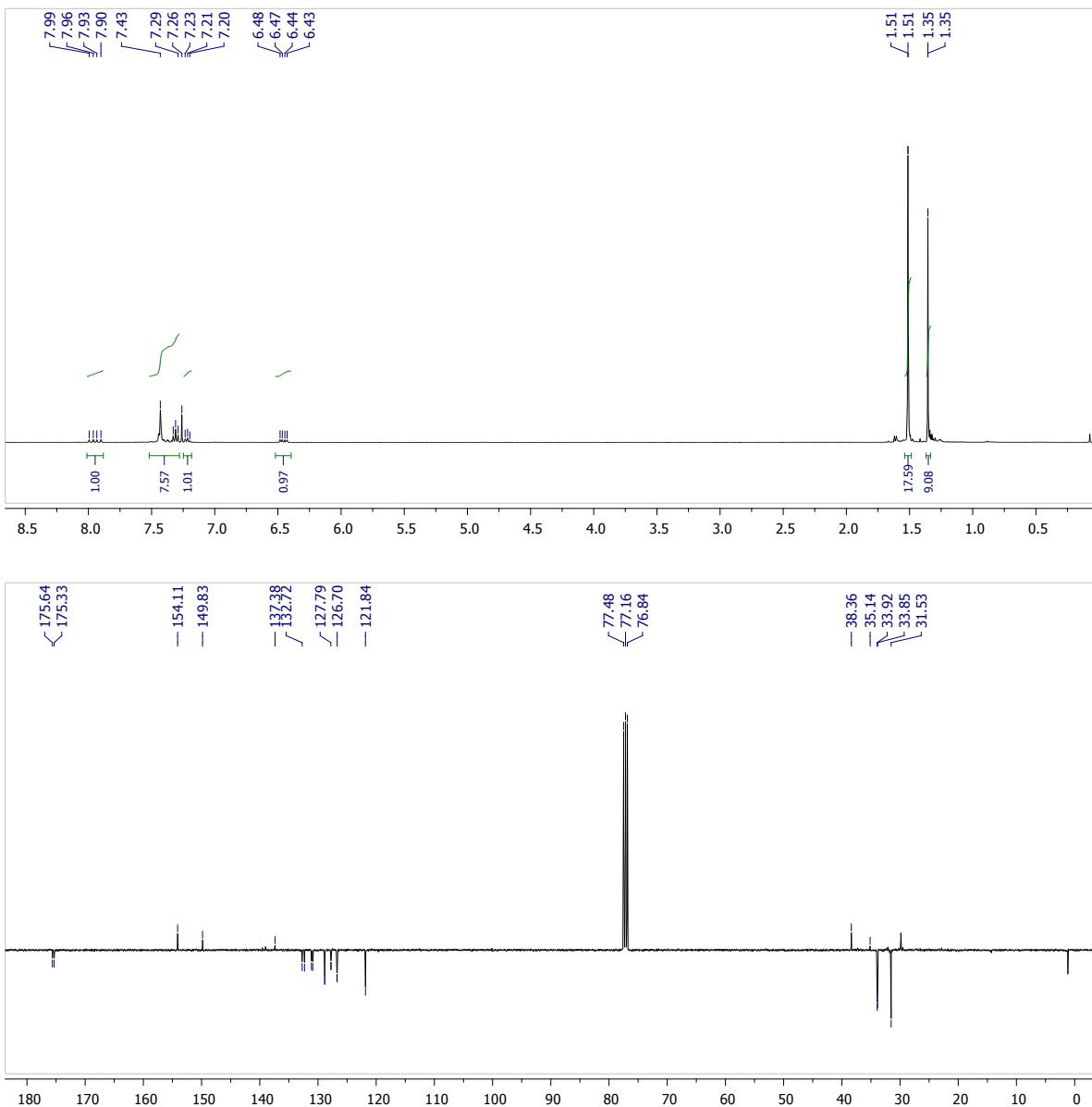
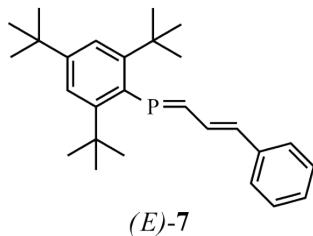


Figure S8 – ^{31}P NMR (top), ^1H NMR (middle) and APT NMR (bottom) spectra of phosphaalkene *E-7*.