Electronic supplementary information

Pyrene-biimidazole based Ru(II) and Os(II) complexes as highly efficient probes for visible and near-infrared detection of cyanide in aqueous medium

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Physical measurements

Elemental analyses of the compounds were performed with a Vario-Micro V2.0.11 elemental (CHNSO) analyzer. NMR spectra were collected on either a Bruker 300 or Bruker 500 spectrometer in DMSO- d_6 , and high resolution mass spectroscopy was performed on a Waters Xevo G2 QTOf mass spectrometer. The UV/vis absorption spectra were recorded with a Shimadzu UV 1800 spectrometer. A matched pair of quartz cuvettes (path length 1 cm) was employed. Steady state luminescence spectra were obtained either by a Perkin–Elmer LS55 or Spex fluorolog-2 spectrofluorometer equipped with DM3000F software. Luminescence quantum yields were determined by using literature method taking [Ru(bpy)₃]²⁺ as the standard. The quantum yields were calculated by using eq S1.

$$\Phi_{\rm r} = \Phi_{\rm std} \cdot A_{\rm std} / A_{\rm r} \cdot I_{\rm r} / I_{\rm std} \cdot \eta_{\rm r}^2 / \eta_{\rm std}^2$$
(S1)

where Φ r and Φ std are the quantum yields of unknown and standard samples [Φ std=0.089 (at 298 K) and 0.35 (at 77 K) in ethanol-methanol (4:1) at λ ex=450 nm], Ar and Astd (<0.1) are the solution absorbances at the excitation wavelength (λ ex), Ir and Istd are the integrated emission intensities, and η r and η std are the refractive indices of the solvent.^{S1}

Luminescence lifetime measurements were carried out by using time–correlated single photon counting set up from Horiba Jobin-Yvon. The luminescence decay data were collected on a Hamamatsu MCP photomultiplier (R3809) and were analyzed by using IBH DAS6 software. Cyclic and square-wave voltammetric experiments were performed in deaerated acetonitrile with a BAS epsilon electrochemistry system and a three-electrode set up consisting of a platinum or glassy carbon working electrode, a platinum counter electrode, and Ag/AgCl reference electrode. Tetraethylammonium perchlorate (TEAP) was used as background electrolyte. The potentials reported in this study were referenced against the Ag/AgCl electrode, which under the given experimental conditions gave a value of 0.36 V for the Fc/Fc⁺ couple.

Experimental uncertainties were as follows: absorption maxima ± 2 nm; molar absorption coefficients, 10%; emission maxima, ± 5 nm; excited-state lifetimes, 10%; luminescence quantum yields, 20%; redox potentials ± 10 mV.

Anion sensing experiments

For a typical absorption and emission titration experiment, aliquots of a TBA salts of F⁻, Cl⁻, Br⁻, I⁻ and AcO⁻ and H₂PO₄⁻ (5.0×10^{-3} M) were added incrementally to a 2.5 mL solution of the complexes (2.0×10^{-5} M). In aqueous medium the experiments were carried out in

HEPES buffer medium (pH=7.00) and the concentration of the metal complexes were taken as 1.5×10^{-5} M for study. The equilibrium constants were evaluated from the absorbance data using equation (S2)^{S2}.

(S2)

 $A_{\rm obs} = (A_0 + A_\infty K[G]_{\rm T})/(1 + K[G]_{\rm T})$

where A_{obs} is the observed absorbance, A_0 is the absorbance of the free receptor, A_{∞} is the maximum absorbance induced by the presence of a given anionic guest, $[G]_T$ is the total concentration of the guest, and *K* is the equilibrium constant of the host–guest entity. Binding constants were performed in duplicate, and the average value is reported. The lifetimes of the receptors were recorded as a function of different anions and solvents.

X-ray crystallographic analyses

The crystallographic data, details of data collection, and refinement parameter for the complexes **1** is summarized in Table S1. Single crystals of suitable size were obtained by diffusing toluene to 1:1 acetonitrile-dichloromethane solution of the complexes. The crystal was immersed in paratone oil and then mounted on the tip of a glass fiber and cemented using epoxy resin. Intensity data for the crystal was collected using MoK α ($\lambda = 0.7107$ Å) radiation on a Bruker SMART APEX II diffractometer equipped with CCD area detector at room temperature. The data integration and reduction were processed with SAINT^{S3} software provided with the software package of SMART APEX II. An empirical absorption correction was applied to the collected reflections with SADABS.^{S3} The structure was solved by direct methods using SHELXTL ^{S4} and was refined on F² by the full-matrix least-squares technique using the SHELXL-97 ^{S5} program package. Graphics were generated using PLATON. ^{S6} Nonhydrogen atoms were refined anisotropically until the convergence. All the hydrogen atoms were geometrically positioned and treated as riding atoms. In the complex **1** the hydrogen atoms of the lattice water molecules could not be located from the difference Fourier map.

CCDC reference numbers is 1409760 for 1

Theoretical computational methods

Quantum chemical calculations were performed with the Gaussian 09 program ^{S7} employing the DFT method with Becke's three-parameter hybrid functional and Lee-Yang-Parr's gradient corrected correlation functional B3LYP level of theory.^{S8-S9} The 6-31G(d) basis set was employed for the C, H and N while SDD basis set was used for Ru and Os atoms. ^{S10} Geometries were fully optimized using the criteria of the respective programs. To compute the UV-vis transition of the compounds, the singlet excited state geometries corresponding to

the vertical excitations were optimized using the time-dependent DFT (TDDFT) scheme starting with the ground state geometries optimized in solution phase.^{S11-S14}The excitation energies, computed within the acetonitrile solvent simulated by the CPCM model,^{S15} has been determined by using the so-called nonequilibrium approach, which has been designed for the study of the absorption process.^{S16-S17} Only singlet-singlet transitions, that is, the spin-allowed transitions, have been taken into account. The geometries of the lowest energy triplet states of the complexes were also optimized in CH₃CN using the CPCM model. Orbital analysis was completed with Gauss View^{S18} and Gauss sum 2.2.^{S19}

Compound	1					
Formula	C43 H35 N9 Cl2 O10 Ru					
fw	1009.77					
T (K)	273(2)					
Cryst. Syst.	Monoclinic					
Space group	P2(1)/c					
$a(\text{\AA})$	11.4961(4)					
b(A)	30.8004(10)					
<i>c</i> (Å)	12.1865(4)					
α (deg)	90.00					
β (deg)	90.00					
γ (deg)	90.00					
$V(Å^3)$	4315.1(2)					
$Dc(g \text{ cm}^{-3})$	1.554					
Ζ	4					
μ (mm ⁻¹)	0.557					
<i>F</i> (000)	2056.0					
θ range (deg)	2.21-27.50					
Data/restraints/params	9382/0/588					
GOF on F ²	0.703					
$R_1 [I > 2\sigma(I)]^a,$	0.0590					
wR_2 (all data) ^b	0.1800					
$\Delta \rho_{\text{max}} / \Delta \rho_{\text{min}} (e \text{ Å})$	0.568/-0.693					
^a R1(F) = [$\sum F_0 - F_C / \sum F_0 $], ^b wR2 (F ²)						
$= \left[\sum w(F_0^2 - F_C^2)^2 / \sum w(F_0^2)^2\right]^{1/2}$						

Table S1 Crystallographic data for 1

		1					
	Exptl.	Soln.(1)	Soln.(1a)	Soln.(1b)	Triplet(1)	Triplet (1a)	Triplet(1b)
Ru1-N1	2.075(4)	2.102	2.094	2.083	2.099	2.012	2.068
Ru1-N3	2.187(4)	2.264	2.259	2.200	2.268	2.251	2.226
Ru1-N5	2.057(5)	2.100	2.106	2.110	2.100	2.144	2.096
Ru1-N6	2.057(4)	2.103	2.100	2.095	2.104	2.142	2.098
Ru1-N7	2.030(4)	2.086	2.078	2.073	2.087	2.034	2.089
Ru1-N8	2.008(4)	2.067	2.064	2.077	2.066	2.072	2.078
	· · · ·				•		•
N1-Ru1-N3	76.85(15)	76.95	77.60	77.54	76.84	78.88	77.65
N1-Ru1-N5	173.91(17)	173.52	172.33	171.27	173.63	170.66	173.00
N1-Ru1-N6	97.68(17)	96.00	94.86	94.02	96.14	95.47	96.65
N1-Ru1-N7	88.31(17)	88.74	89.19	89.92	88.64	93.06	89.13
N1-Ru1-N8	89.54(16)	92.25	91.90	91.98	92.27	92.20	91.08
N3-Ru1-N5	107.39(15)	104.60	103.68	104.10	104.74	104.48	106.25
N3-Ru1-N6	88.12(16)	87.25	87.13	87.53	87.12	80.03	88.54
N3-Ru1-N7	96.80(16)	96.42	96.03	95.36	96.52	96.80	95.14
N3-Ru1-N8	166.14(16)	168.30	168.36	167.88	168.26	170.44	167.57
N5-Ru1-N6	78.31(17)	77.87	77.72	77.56	77.86	76.26	77.82
N5-Ru1-N7	95.43(17)	97.28	98.13	98.39	97.24	95.13	96.21
N5-Ru1-N8	86.38(17)	86.62	87.38	87.17	86.55	84.84	85.49
N6-Ru1-N7	173.01(17)	174.57	175.32	175.53	174.55	171.38	173.70
N6-Ru1-N8	96.39(18)	98.53	98.88	99.04	98.55	98.45	98.08
N7-Ru1-N8	79.97(18)	78.59	78.61	78.35	78.60	80.08	79.25

 Table S2 Selected calculated bond distances (Å) and angles (deg) for 1, 1a, 1b in ground

 state and triplet excited state along with available X-ray crystal data

In Acetonitrile Medium										
	From Absorption spectra									
	-	1			2					
anion	$K_1(10^6 \text{ M}^{-1})$	$K_2(10^6 \text{ M}^{-1})$		$K_1(10^6 \text{ M}^{-1})$	$K_2(10^6 \text{ M}^{-1})$					
F-	4.36	2.63		3.52	2.98					
CN-	3.60	2.33		3.07	2.79					
AcO-	3.01	2.31		2.82	2.30					
H ₂ PO ₄ -	2.33			1.89						
		From Emiss	sio	n spectra						
	-	1			2					
anion	$K_1(10^6 \text{ M}^{-1})$	$K_2(10^6 \text{ M}^{-1})$		$K_1(10^6 \text{ M}^{-1})$	$K_2(10^6 \text{ M}^{-1})$					
F-	3.76	3.33		3.39	2.14					
CN-	3.62	2.80		3.06	1.83					
AcO-	3.16	2.68		2.59	1.68					
H ₂ PO ₄ -	3.02			2.89						
		In Aqueou	s I	Medium						
		From	Absorption spectra							
	-	1		2						
anion	$K_1(10^5 \text{ M}^{-1})$	$K_2(10^4 \text{ M}^{-1})$		$K_1(10^5 \text{ M}^{-1})$	$K_2(10^4 \text{ M}^{-1})$					
CN-	3.45 1.34			6.77	1.07					
From Emission Spectra										
		1			2					
anion	$K_1(10^5 \text{ M}^{-1})$ $K_2(10^4 \text{ M}^{-1})$			$K_1(10^5 \text{ M}^{-1})$	$K_2(10^4 \text{ M}^{-1})$					
CN-	2.03 1.03			3.20	1.37					

Table S3 Equilibrium constants^{a,b} for 1 and 2 towards various anions in acetonitrile and pure aqueous medium at 298 K

Table S4 Selected calculated bond distances (Å) and angles (deg) for 2, 2a, 2b in ground state and triplet excited state

		2 (sol)		2 (triplet)					
	Soln.(2)	Soln.(2a)	Soln.(2b)	Triplet(2)	Triplet (2a)	Triplet(2b)			
Os1-N1	2.107	2.105	2.097	2.078	2.035	2.019			
Os1-N3	2.249	2.248	2.198	2.243	2.244	2.197			
Os1-N5	2.102	2.104	2.103	2.115	2.137	2.134			
Os1-N6	2.106	2.101	2.095	2.137	2.138	2.128			
Os1-N7	2.093	2.082	2.076	2.051	2.051	2.062			
Os1-N8	2.074	2.069	2.079	2.076	2.088	2.104			
		•	•						
N1-Os1-N3	76.18	76.71	76.66	76.87	77.86	77.98			
N1-Os1-N5	173.07	171.96	170.89	172.14	170.42	170.45			
N1-Os1-N6	96.15	94.99	94.07	97.03	95.59	95.50			
N1-Os1-N7	88.54	88.86	89.56	89.39	92.62	92.36			
N1-Os1-N8	93.12	92.77	92.70	93.09	92.76	92.84			
N3-Os1-N5	105.24	104.26	104.53	106.74	105.74	105.45			
N3-Os1-N6	86.70	86.54	86.99	85.05	85.45	85.59			
N3-Os1-N7	96.90	96.41	95.45	97.29	97.13	96.83			
N3-Os1-N8	168.31	168.26	167.52	169.54	169.87	169.77			
N5-Os1-N6	77.25	77.16	77.05	76.56	76.06	76.07			
N5-Os1-N7	97.95	98.91	99.24	96.95	95.67	96.00			
N5-Os1-N8	85.93	86.88	87.00	83.57	84.11	84.33			
N6-Os1-N7	174.66	175.60	176.00	173.51	171.73	172.08			
N6-Os1-N8	99.28	99.72	100.43	99.28	99.44	99.93			
N7-Os1-N8	77.87	77.93	77.72	79.42	79.27	78.85			

Fable S5 Selected molecular orbital along with their energies and compositions for 1, 1a, 1b and 2, 2a and 2b in triplet excited state (using 6-
B1G(d) and SDD basis sets)

	Energy/	eV		(%) Composition														
				1 (%) Composition					1a (%) Composition					1b (%) Composition				
	1	1a	1b	Ru ^Ⅱ	pyrene	imida	bpy		Ru ^{II}	pyrene	imida	bpy		Ru ^{II}	pyrene	imida	bpy	
MO					imida					imida					imida			
LUMO+3	-1.95	-1.47	-1.16	3.87	62.23	31.63	2.25		2.64	1.07	0.27	96.00		1.70	79.97	0.05	18.26	
LUMO+2	-2.18	-1.64	-1.34	0.48	91.23	7.72	0.55		0.61	97.83	0.46	1.07		2.90	1.48	0.16	95.45	
LUMO+1	-2.46	-2.27	-2.15	6.04	0.38	0.05	93.50	[4.83	0.44	0.09	94.63		6.75	0.54	0.08	92.61	
LUMO	-2.52	-2.62	-2.5	3.10	0.44	0.53	95.91	[7.15	0.10	0.45	92.29		5.70	0.07	0.30	93.91	
HOMO	-5.43	-5.05	-4.12	4.38	88.79	6.22	0.58	[26.62	38.23	29.95	5.17		3.44	59.85	36.14	0.55	
HOMO-1	-5.92	-5.42	-5.17	75.38	9.67	4.40	10.53		43.27	32.04	15.96	8.72		62.08	13.58	14.47	9.85	
HOMO-2	-6.11	-5.76	-5.3	75.22	9.09	1.65	14.02	[74.56	2.73	5.12	17.57		59.06	30.72	2.81	7.39	
HOMO-3	-6.17	-5.78	-5.43	76.77	2.70	3.55	16.96		71.04	19.08	0.23	9.63		48.82	26.26	10.49	14.41	
				2 (%) Composition				2a (%) Composition					2b (%) Composition					
	2	2a	2b	Os ^{II}	pyrene	imida	bpy		OsII	pyrene	imida	bpy		Os ^{II}	pyrene	imida	bpy	
MO					imida					imida					imida			
LUMO+3	-1.79	-1.47	-1.2	2.00	92.85	4.38	0.75	[2.66	0.76	0.16	96.40		0.99	95.81	0.05	3.14	
LUMO+2	-2.05	-1.66	-1.31	3.53	59.59	34.09	2.77	[0.61	97.80	0.52	1.05		2.88	1.03	0.17	95.90	
LUMO+1	-2.46	-2.27	-2.1	6.73	0.77	0.29	92.19	[7.69	0.58	0.14	91.57		8.63	0.56	0.16	90.63	
LUMO	-2.82	-2.6	-2.42	7.04	0.68	1.03	91.23	[8.66	0.29	0.88	90.15		9.56	0.25	1.01	89.16	
НОМО	-5.47	-4.95	-4.35	55.50	25.47	7.84	11.16		48.94	17.17	21.71	12.17		17.17	49.04	28.89	4.88	
HOMO-1	-5.78	-5.33	-4.82	24.44	64.56	5.46	5.52		28.35	45.43	19.24	6.96		49.21	22.30	15.43	13.03	
HOMO-2	-5.9	-5.56	-5.09	73.07	10.83	1.23	14.85	ľ	74.80	5.31	4.39	15.49	Ī	62.22	25.95	2.50	9.31	
HOMO-3	-5.98	-5.6	-5.25	68.05	5.81	3.66	22.46		59.51	20.17	1.94	18.36		64.26	6.03	5.81	23.88	

Table S6 Phosphorescence emissions of **1**, **1a**, **1b** and **2**, **2a** and **2b** in acetonitrile solution according to TD-DFT calculation and associated experimental values (using 6-31G(d) and SDD basis sets)

Compounds	Acetonilrile					
	Expt.	Theo. (Transition)				
1	712 nm	743 nm				
		$L+2 \to H (84\%),$				
		$L+2 \rightarrow H-1 (3\%),$				
		$L+3 \rightarrow H(8\%)$				
1a	712 nm	849 nm				
		$L \rightarrow H-1 (30\%),$				
		$L \rightarrow H (65\%)$				
1b	793 nm	1161 nm				
		$L \to H (96\%), L$				
		\rightarrow H-1(3%)				
2	786 nm	790 nm				
		$L \rightarrow H-1 (12\%),$				
		$L \rightarrow H (80\%)$				
2a	806 nm	988 nm				
		$L \rightarrow H (83\%)$				
		$L \to H-1 (9\%),$				
2b	834 nm	1200 nm				
		$L \rightarrow H-1$ (29%).				
		$L \rightarrow H(63\%)$				



Fig. S1 ¹H NMR spectra of 1 (a) and 2 (b) (300 MHz) in DMSO- d_6 .



Fig. S2 ¹H-¹H COSY NMR spectrum of 1 in DMSO- d_6 .



Fig. S3 ¹H-¹H COSY NMR spectrum of 2 in DMSO- d_6 .



Fig. S4 ESI-MS (positive) for the complex cation $[(bpy)_2Ru(Py-BiimzH_2)]^{2+}$ (*m*/*z* = 361.07) in acetonitrile showing the observed and isotopic distribution patterns.



Fig. S5 ESI-MS (positive) for the complex cation $[(bpy)_2Os(Py-BiimzH_2)]^{2+}$ (*m*/*z* = 405.58) in acetonitrile showing the observed and isotopic distribution patterns.



Fig. S6 Strong hydrogen bonding interactions between the N-H imidazole protons of receptor **1** with water and dimethylformamide solvents.



Fig. S7 Changes in UV-vis absorption (a) and emission spectral profiles (b) (normalised) of **2** in few selective solvents.



Fig. S8 Luminescence spectra of **1** (a) and **2** (b) in MeCN at room temperature and at 77K in MeOH: EtOH (1:4) glass.



Fig. S9 Decay profiles of 1 (a) and 2 (b) in few selected solvents. Insets show the corresponding lifetimes.



Fig. S10 Cyclic voltammograms and square wave voltammograms of 1 (a) and 2 (b) in acetonitrile at a scan rate of 100 mV/s showing both oxidation and reduction of the complexes.



Fig. S11 Changes in absorption, emission and excited state lifetimes of **2** within the pH range 2.5-7.6 (a, b and e) and 7.6-12 (c, d and f) respectively in MeCN/ H_2O (1/9 v/v) mixture. Insets show the variation of absorption, emission and decay profiles with pH.



Fig. S12 Changes in UV-vis absorption (a and b) and luminescence (c and d) spectral profiles of **2** in acetonitrile solution upon the addition of CN⁻ ion. The inset shows the fit of the experimental absorbance and luminescence data to a 1:1 binding profile.



Fig. S13 Changes in UV-vis absorption (a and c) and luminescence (b and d) spectra of complex **1** in acetonitrile solution upon the addition of F⁻ ion. The inset shows the fit of the experimental absorbance and luminescence data to a 1:1 binding profile.



Fig. S14 Changes in UV-vis absorption (a and b) and luminescence (c and d) spectra of complex **1** in acetonitrile solution upon the addition of AcO⁻ ion. The inset shows the fit of the experimental absorbance and luminescence data to a 1:1 binding profile.



Fig. S15 Changes in UV-vis absorption (a and b) and luminescence (c and d) spectra of complex **2** in acetonitrile solution upon the addition of F⁻ ion. The inset shows the fit of the experimental absorbance and luminescence data to a 1:1 binding profile.



Fig. S16 Changes in UV-vis absorption (a and b) and luminescence (c and d) spectra of complex **2** in acetonitrile solution upon the addition of AcO⁻ ion. The inset shows the fit of the experimental absorbance and luminescence data to a 1:1 binding profile.



Fig. S17 Changes in UV–vis absorption (a and b) and luminescence (c and d) spectral profiles for 1 and 2, respectively in acetonitrile solution upon the addition of H_2PO_4 - ion. The inset shows the fit of the experimental absorbance and luminescence data to a 1:1 binding profile.



Fig. S18 (a) Absorption spectral change during the titration of 1 with CN⁻ in acetonitrile, inset shows the normalized absorbance between the minimum absorbance (free complex 1) and the maximum absorbance. (b) A plot of $(A-A_{min})/(A_{max}-A_{min})$ vs Log([CN⁻]), the calculated detection limit of receptor is 5.24×10^{-9} M.



Fig. S19 (a) Fluorescence spectral change during the titration of 1 with CN⁻ in acetonitrile, inset shows the normalized intensity between the minimum intensity (free complex 1) and the maximum intensity. (b) A plot of $(I-I_{min})/(I_{max}-I_{min})$ vs Log([CN⁻]), the calculated detection limit of receptor is 4.67×10^{-9} M.



Fig. S20 (a) Absorption spectral change during the titration of 2 with CN^{-} in acetonitrile, inset shows the normalized absorbance between the minimum absorbance (free complex 2) and the maximum absorbance. (b) A plot of $(A-A_{min})/(A_{max}-A_{min})$ vs Log([CN⁻]), the calculated detection limit of receptor is 4.46×10^{-9} M.



Fig. S21 (a) Fluorescence spectral change during the titration of 2 with CN⁻ in acetonitrile, inset shows the normalized intensity between the minimum intensity (free complex 2) and the maximum intensity. (b) A plot of $(I-I_{min})/(I_{max}-I_{min})$ vs Log([CN⁻]), the calculated detection limit of receptor is 4.57×10^{-9} M.



Fig. S22 Changes in the time-resolved luminescence decays for **1** (a and b) with incremental addition of F^- (0-4 equiv) and for **2** (c and d) with F^- (0-3 equiv) in acetonitrile solution. Insets show the lifetime values.



Fig. S23 Changes in the time-resolved luminescence decays for 1 (a) and 2 (b) with incremental addition of $H_2PO_4^-$ in acetonitrile solution. Insets show the lifetime values.



Fig. S24 Changes in the oxidation process of complex 2 in terms of CV (a and b) with incremental addition of CN^{-} ion in its acetonitrile solution.



Fig. S25 Changes in the oxidation process of complex **1** in terms of CV (a and b) with incremental addition of CN⁻ ion in its acetonitrile solution.



Fig. S26 Changes in UV-vis absorption (a and c) and luminescence (b and d) spectra of complex 1 in aqueous buffer solution (pH=7) upon the addition of CO_3^{2-} and S^{2-} ion respectively.



Fig. S27 Changes in UV-vis absorption (a and c) and luminescence (b and d) spectra of complex 2 in aqueous buffer solution (pH=7) upon the addition of CO_3^{2-} and S^{2-} ion respectively.



Fig. S28 Changes in the time-resolved luminescence decays for 1 (a and b) with incremental addition of CN^{-} (0-50 equiv) and for 2 (c and d) with CN^{-} (0-45 equiv) in pure aqueous medium. Insets show the lifetime values.



Fig. S29 (a) Absorption spectral change during the titration of 1 with CN⁻ in water medium, inset shows the normalized absorbance between the minimum absorbance (free complex 1) and the maximum absorbance. (b) A plot of $(A-A_{min})/(A_{max}-A_{min})$ vs Log([CN⁻]), the calculated detection limit of receptor is 1.12×10^{-8} M.



Fig. S30 (a) Fluorescence spectral change during the titration of 1 with CN⁻ in water medium, inset shows the normalized intensity between the minimum intensity (free complex 1) and the maximum intensity. (b) A plot of $(I-I_{min})/(I_{max}-I_{min})$ vs Log([CN⁻]), the calculated detection limit of receptor is 1.03×10^{-8} M.



Fig. S31 (a) Absorption spectral change during the titration of 2 with CN⁻ in water medium, inset shows the normalized absorbance between the minimum absorbance (free complex 2) and the maximum absorbance. (b) A plot of $(A-A_{min})/(A_{max}-A_{min})$ vs Log([CN⁻]), the calculated detection limit of receptor is 1.10×10^{-8} M.



Fig. S32 (a) Fluorescence spectral change during the titration of 2 with CN⁻ in water medium, inset shows the normalized intensity between the minimum intensity (free complex 2) and the maximum intensity. (b) A plot of $(I-I_{min})/(I_{max}-I_{min})$ vs Log([CN⁻]), the calculated detection limit of receptor is 1.24×10^{-8} M.



Fig. S33 Visible color changes that occur when receptor 1 and 2 (a and b respectively) are grinded mechanochemically with NaCN in solid state.



Fig. S34 Changes in UV-vis absorption (a and b) and luminescence (c and d) spectra of 1 and 2, respectively in solid state upon grinding with CN⁻ ion.



Fig. S35 Changes in the time-resolved luminescence decays for **1** (a) and **2** (b) in solid state when grinded with CN⁻. Insets show the lifetime values.



Fig. S36 Optimised geometries and labelling scheme for $[(bpy)_2Ru(Py-BiimzH_2)]^{2+}$ (1), $[(bpy)_2Ru(Py-BiimzH)]^+$ (1a) and $[(bpy)_2Ru(Py-Biimz)]^0$ (1b) in solution phase (using 6-31G(d) and SDD basis sets).



Fig. S37 Optimised geometries and labelling scheme for $[(bpy)_2Os(Py-BiimzH_2)]^{2+}$ (2), $[(bpy)_2Os(Py-BiimzH)]^+$ (2a) and $[(bpy)_2Os(Py-Biimz)]^0$ (2b) in solution phase (using 6-31G(d) and SDD basis sets).



Fig. S38 Molecular electrostatic potential mapped on the isodensity surface in the ground state for $[(bpy)_2Ru(Py-BiimzH_2)]^{2+}$ (1), $[(bpy)_2Ru(Py-BiimzH)]^+$ (1a) and $[(bpy)_2Ru(Py-Biimz)]^0$ (1b) in the range from -4.000e-2 (red) to +4.000e-2 (blue).



Fig. S39 Molecular electrostatic potential mapped on the isodensity surface in the ground state for $[(bpy)_2Os(Py-BiimzH_2)]^{2+}$ (2), $[(bpy)_2Os(Py-BiimzH)]^+$ (2a) and $[(bpy)_2Os(Py-Biimz)]^0$ (2b) in the range from -4.000e-2 (red) to +4.000e-2 (blue).



Fig. S40 Schematic drawings of the selective frontier molecular orbitals for $[(bpy)_2Ru(Py-BiimzH_2)]^{2+}$ (1), $[(bpy)_2Ru(Py-BiimzH)]^+$ (1a) and $[(bpy)_2Ru(Py-Biimz)]^0$ (1b) in solution phase (using 6-31G(d) and SDD basis sets).



Fig. S41 Schematic drawings of the selective frontier molecular orbitals for $[(bpy)_2Os(Py-BiimzH_2)]^{2+}$ (1), $[(bpy)_2Os(Py-BiimzH)]^+$ (1a) and $[(bpy)_2Os(Py-Biimz)]^0$ (1b) in solution phase(using -31G(d) and SDD basis sets).



Fig. S42 Energy level diagrams depicting the dominant transitions that comprise the lowest-energy absorption band for $[(bpy)_2Os(Py-BiimzH_2)]^{2+}$ (2), $[(bpy)_2Os(Py-BiimzH)]^+$ (2a) and $[(bpy)_2Os(Py-Biimz)]$ (2b) in acetonitrile (using 6-31G(d) and SDD basis sets).



Fig. S43 Optimised geometries and labelling scheme for $[(bpy)_2Ru(Py-BiimzH_2)]^{2+}$ (1), $[(bpy)_2Ru(Py-BiimzH)]^+$ (1a) and $[(bpy)_2Ru(Py-Biimz)]^0$ (1b) in triplet excited state (using 6-31G(d) and SDD basis sets).



Fig. S44 Optimised geometries and labelling scheme for $[(bpy)_2Os(Py-BiimzH_2)]^{2+}$ (2), $[(bpy)_2Os(Py-BiimzH)]^+$ (2a) and $[(bpy)_2Os(Py-Biimz)]^0$ (2b) in triplet excited state (using 6-31G(d) and SDD basis sets).



Fig. S45 Schematic drawings of the selective frontier molecular orbitals for $[(bpy)_2Ru(Py-BiimzH_2)]^{2+}$ (1), $[(bpy)_2Ru(Py-BiimzH)]^+$ (1a) and $[(bpy)_2Ru(Py-Biimz)]^0$ (1b) in triplet excited state (using 6-31G(d) and SDD basis sets).



Fig. S46 Schematic drawings of the selective frontier molecular orbitals for $[(bpy)_2Os(Py-BiimzH_2)]^{2+}$ (2), $[(bpy)_2Os(Py-BiimzH)]^+$ (2a) and $[(bpy)_2Os(Py-Biimz)]^0$ (2b) in triplet excited state (using 6-31G(d) and SDD basis sets).

Fig. S47 Calculated single electron transitions for the emissions of T_1 states for $[(bpy)_2Ru(Py-BiimzH_2)]^{2+}$ (1), $[(bpy)_2Ru(Py-BiimzH)]^+$ (1a) and $[(bpy)_2Ru(Py-Biimz)]^0$ (1b) (using 6-31G(d) and SDD basis sets).

Fig. S48 Calculated single electron transitions for the emissions of T_1 states for $[(bpy)_2Os(Py-BiimzH_2)]^{2+}$ (2), $[(bpy)_2Os(Py-BiimzH)]^+$ (2a) and $[(bpy)_2Os(Py-Biimz)]^0$ (2b) (using 6-31G(d) and SDD basis sets).

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