

1 [Supporting Information]

2 Surface Engineering Superparamagnetic
3 Nanoparticles for Aqueous Applications: Design and
4 Characterization of Tailored Organic Bilayers

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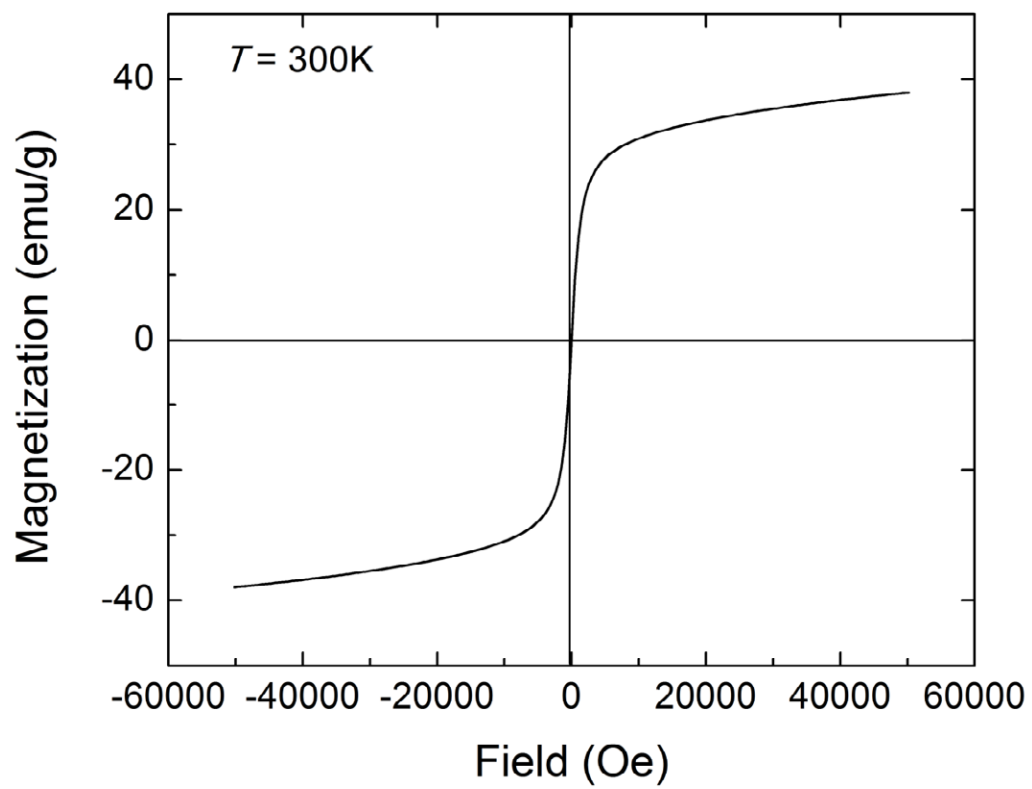
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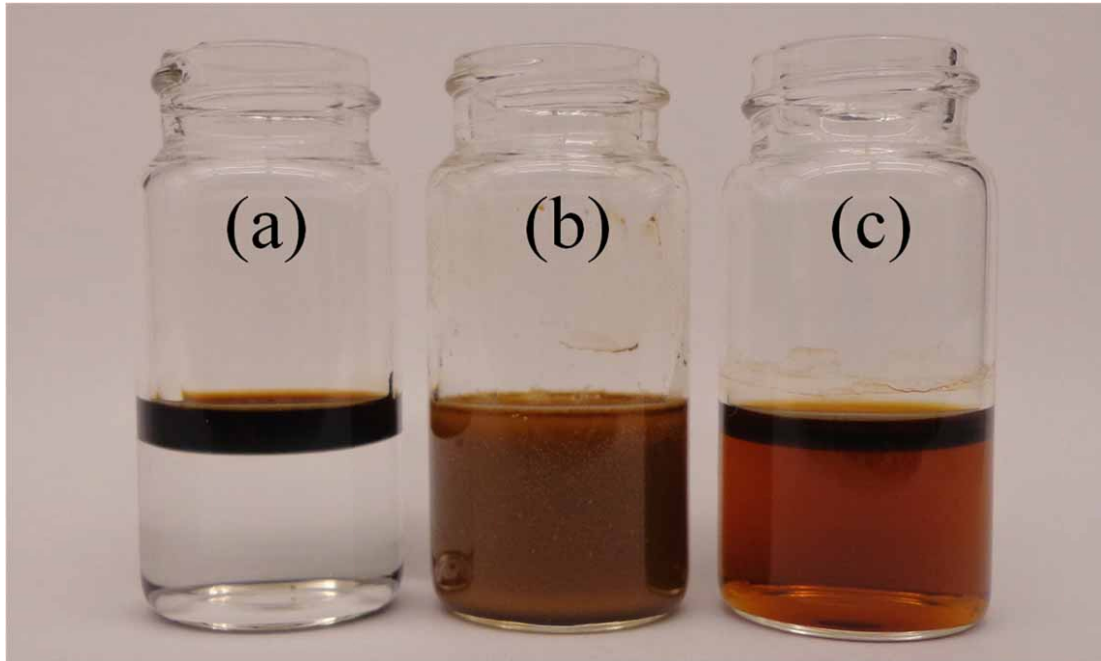
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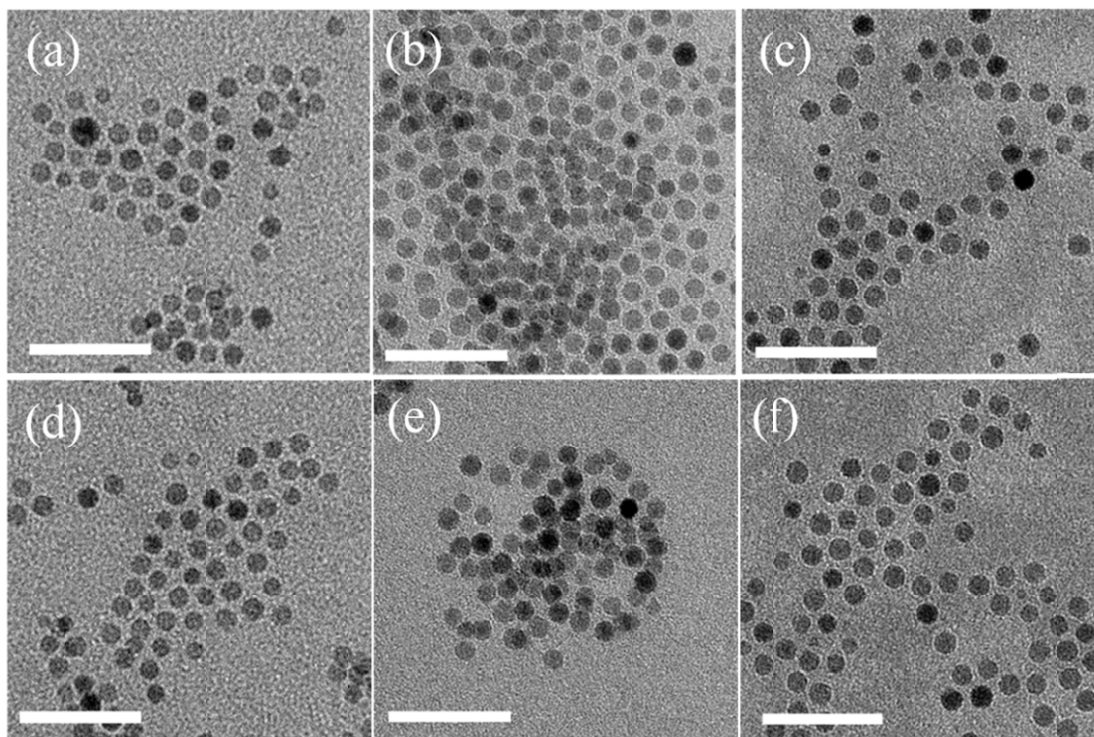
19 **Figure S1.** Hysteresis loops of 8 nm iron oxide NPs at 300 K.



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21 **Figure S2.** Bilayer phase transfer process at different stages: (a) at beginning, (b) just after

22 sonication and (c) after 1 day settlement.



23
24 **Figure S3.** Representative TEM micrographs of (a) RA-NPs, (b) SA-NPs, (c) SDP-NPs, (d)
25 SDS-NPs, (e) C₁₂TAB-NPs, and (f) EMPIGEN-NPs in water. All scale bars are 50 nm.

26 **Table S1.** Detailed hydrodynamic diameter and transfer yield as functions of sonication
 27 amplitude, sonication time and surfactant concentration tested for ricinoleic acid (RA) coated
 28 NPs.

Dependent Variable	Time Tested (min)	Conc. Tested (mM)	Amplitude Tested	Hydrodynamic Diameter (nm)	Transfer Yield (%)
Amplitude	4	10	Dependent Variable		
50%				20.4 ± 6.6	47.3 ± 1.3
60%				23.7 ± 2.1	72.0 ± 3.1
70%				22.4 ± 0.5	81.2 ± 2.2
80%				21.3 ± 0.6	84.1 ± 1.9
Time (min)	Dependent Variable	10	70%		
2				23.6 ± 0.9	53.7 ± 2.5
4				21.8 ± 1.8	81.2 ± 2.2
6				20.9 ± 1.7	75.9 ± 2.0
8				23.1 ± 3.3	77.3 ± 1.0
Conc. (mM)	5	Dependent Variable	70%		
2.5				24.3 ± 0.7	62.5 ± 1.5
5				24.2 ± 0.8	78.6 ± 2.2
10				21.8 ± 1.2	83.5 ± 1.6
15				21.3 ± 1.8	85.1 ± 3.5
20				22.2 ± 5.5	84.3 ± 1.5

29

30 **Table S2.** Detailed hydrodynamic diameter and transfer yield as functions of sonication
 31 amplitude, sonication time and surfactant concentration tested for elaidic acid (EA) coated NPs.

Dependent Variable	Time Tested (min)	Conc. Tested (uL)	Amplitude Tested	Hydrodynamic Diameter (nm)	Transfer Yield (%)
Amplitude	4	50	Dependent Variable		
50%				45.4 ± 2.9	13.4 ± 1.5
60%				41.2 ± 3.0	19.3 ± 2.3
70%				24.7 ± 5.7	26.2 ± 1.1
80%				22.7 ± 5.8	25.7 ± 3.2
Time (min)	Dependent Variable	50	75%		
2				31.4 ± 4.6	16.5 ± 1.7
4				18.5 ± 4.3	26.6 ± 2.2
6				20.5 ± 4.1	25.2 ± 1.4
8				19.8 ± 2.7	21.3 ± 0.5
Conc. (uL)	6	Dependent Variable	75%		
12.5				39.4 ± 10.5	17.7 ± 0.4
25				30.4 ± 5.2	21.1 ± 1.8
50				19.5 ± 5.5	25.8 ± 2.1
75				25.2 ± 4.2	24.1 ± 1.4
100				32.6 ± 2.7	16.0 ± 0.9

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33 **Table S3.** Detailed hydrodynamic diameter and transfer yield as functions of sonication
 34 amplitude, sonication time and surfactant concentration tested for stearic acid (SA) coated NPs.

Dependent Variable	Time Tested (min)	Conc. Tested (mM)	Amplitude Tested	Hydrodynamic Diameter (nm)	Transfer Yield (%)
Amplitude	4	10	Dependent Variable		
50%				14.9 ± 3.4	41.7 ± 3.2
60%				21.4 ± 2.5	65.1 ± 2.0
70%				18.6 ± 1.4	65.2 ± 1.7
80%				17.1 ± 2.1	71.5 ± 3.5
Time (min)	Dependent Variable	10	70%		
2				25.0 ± 10.4	47.4 ± 2.7
4				26.4 ± 8.0	63.3 ± 1.6
6				23.0 ± 3.5	75.4 ± 2.5
Conc. (mM)	5	Dependent Variable	70%		
2.5				17.7 ± 6.2	37.4 ± 0.3
5				33.5 ± 3.8	51.3 ± 2.4
10				20.8 ± 2.6	73.3 ± 4.0

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36 **Table S4.** Detailed hydrodynamic diameter and transfer yield as functions of sonication
 37 amplitude, sonication time and surfactant concentration tested for palmitic acid (PA) coated NPs.

Dependent Variable	Time Tested (min)	Conc. Tested (mM)	Amplitude Tested	Hydrodynamic Diameter (nm)	Transfer Yield (%)
Amplitude	4	10	Dependent Variable		
50%				24.7 ± 4.8	35.2 ± 1.3
60%				30.6 ± 2.6	61.0 ± 2.4
70%				26.2 ± 0.7	62.7 ± 5.1
80%				27.4 ± 3.3	71.4 ± 3.7
Time (min)	Dependent Variable	10	65%		
2				29.2 ± 3.5	57.3 ± 2.5
4				31.9 ± 3.9	63.3 ± 0.8
6				20.7 ± 7.4	67.4 ± 3.1
8				23.2 ± 2.8	67.2 ± 2.3
Conc. (mM)	6	Dependent Variable	65%		
2.5				31.2 ± 1.2	57.1 ± 3.3
5				33.5 ± 3.7	66.3 ± 3.4
10				33.2 ± 7.7	68.4 ± 1.7
15				30.8 ± 1.4	68.6 ± 3.2
20				23.7±11.1	70.7 ± 1.9

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39 **Table S5.** Detailed hydrodynamic diameter and transfer yield as functions of sonication
 40 amplitude, sonication time and surfactant concentration tested for myristic acid (MA) coated
 41 NPs.

Dependent Variable	Time Tested (min)	Conc. Tested (mM)	Amplitude Tested	Hydrodynamic Diameter (nm)	Transfer Yield (%)
Amplitude	4	10	Dependent Variable		
50%				24.3 ± 7.7	70.5 ± 2.3
60%				24.0 ± 2.7	81.0 ± 3.6
70%				24.3 ± 0.3	82.1 ± 3.9
80%				22.8 ± 0.6	80.0 ± 1.2
Time (min)	Dependent Variable	10	65%		
1				26.7 ± 1.1	73.3 ± 1.2
2				24.2 ± 1.8	82.5 ± 5.6
4				21.8 ± 1.0	83.9 ± 1.9
6				21.4 ± 0.3	80.4 ± 2.8
Conc. (mM)	4	Dependent Variable	65%		
2.5				25.4 ± 2.8	60.4 ± 2.1
5				24.2 ± 2.0	67.5 ± 1.8
10				21.5 ± 1.9	74.1 ± 2.3
15				20.9 ± 1.4	77.3 ± 2.7
20				21.1 ± 1.1	75.5 ± 4.4

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43 **Table S6.** Detailed hydrodynamic diameter and transfer yield as functions of sonication
 44 amplitude, sonication time and surfactant concentration tested for lauric acid (LA) coated NPs.

Dependent Variable	Time Tested (min)	Conc. Tested (mM)	Amplitude Tested	Hydrodynamic Diameter (nm)	Transfer Yield (%)
Amplitude	3	5	Dependent Variable		
50%				23.2 ± 10.4	58.4 ± 0.8
60%				25.7 ± 8.6	61.5 ± 3.6
70%				26.5 ± 2.1	77.4 ± 5.4
80%				25.6 ± 2.7	84.5 ± 3.6
90%				24.3 ± 0.5	87.3 ± 4.1
Time (min)	Dependent Variable	5	75%		
1				37.6 ± 4.8	80.3 ± 4.1
2				31.6 ± 2.6	86.5 ± 2.6
3				20.8 ± 4.0	88.2 ± 3.3
4				28.0 ± 1.7	84.2 ± 3.9
5				22.4 ± 9.0	88.5 ± 2.6
Conc. (mM)	3	Dependent Variable	75%		
1.25				34.5 ± 0.7	70.1 ± 2.8
2.5				31.6 ± 2.5	74.5 ± 1.7
5				26.9 ± 1.7	85.1 ± 4.4
7.5				25.2 ± 1.7	88.8 ± 2.6
10				19.2 ± 0.4	87.6 ± 1.9

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46 **Table S7.** Detailed hydrodynamic diameter and transfer yield as functions of sonication
 47 amplitude, sonication time and surfactant concentration tested for decanoic acid (DA) coated
 48 NPs.

Dependent Variable	Time Tested (min)	Conc. Tested (mM)	Amplitude Tested	Hydrodynamic Diameter (nm)	Transfer Yield (%)
Amplitude	4	10	Dependent Variable		
50%				24.4 ± 2.5	26.7 ± 3.1
60%				27.5 ± 1.5	30.8 ± 5.0
70%				26.1 ± 2.3	40.6 ± 2.4
80%				28.4 ± 0.6	59.3 ± 4.2
Time (min)	Dependent Variable	10	75%		
2				28.4 ± 0.4	55.4 ± 2.0
4				26.5 ± 0.6	54.3 ± 2.4
6				25.1 ± 0.7	55.0 ± 1.8
8				25.4 ± 0.8	59.3 ± 3.6
Conc. (mM)	4	Dependent Variable	75%		
2.5				30.4 ± 8.9	38.4 ± 0.7
5				28.9 ± 0.7	40.4 ± 2.1
10				28.5 ± 0.6	46.4 ± 1.1
15				27.4 ± 1.9	45.6 ± 1.6
20				28.9 ± 0.3	53.4 ± 3.0

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50 **Table S8.** Detailed hydrodynamic diameter and transfer yield as functions of sonication
 51 amplitude, sonication time and surfactant concentration tested for sodium monododecyl
 52 phosphate (SDP) coated NPs.

Dependent Variable	Time Tested (min)	Conc. Tested (mM)	Amplitude Tested	Hydrodynamic Diameter (nm)	Transfer Yield (%)
Amplitude	3	5	Dependent Variable		
50%				44.5 ± 0.5	51.2 ± 3.4
60%				41.6 ± 2.4	67.4 ± 3.9
70%				39.1 ± 2.9	85.6 ± 4.6
80%				36.2 ± 0.6	93.1 ± 4.9
90%				39.0 ± 1.0	91.4 ± 5.5
Time (min)	Dependent Variable	5	70%		
1				53.2 ± 3.2	67.4 ± 1.3
2				43.7 ± 1.6	79.3 ± 5.7
3				38.7 ± 0.8	90.9 ± 4.7
4				39.0 ± 1.0	92.7 ± 3.2
5				38.4 ± 2.5	95.2 ± 1.9
Conc. (mM)	3	Dependent Variable	70%		
1.25				54.9 ± 3.3	66.9 ± 1.1
2.5				43.1 ± 2.3	77.3 ± 2.2
5				40.6 ± 1.2	87.6 ± 3.9
7.5				34.3 ± 0.2	85.2 ± 6.7
10				23.4 ± 3.5	90.1 ± 2.3

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54 **Table S9.** Detailed hydrodynamic diameter and transfer yield as functions of sonication
 55 amplitude, sonication time and surfactant concentration tested for sodium monododecyl sulfate
 56 (SDS) coated NPs.

Dependent Variable	Time Tested (min)	Conc. Tested (mM)	Amplitude Tested	Hydrodynamic Diameter (nm)	Transfer Yield (%)
Amplitude	5	5	Dependent Variable		
50%				20.9 ± 3.0	40.8 ± 0.6
60%				20.4 ± 1.1	50.4 ± 1.2
70%				19.0 ± 1.0	64.3 ± 3.6
80%				20.2 ± 1.6	72.3 ± 7.4
90%				20.3 ± 0.1	70.7 ± 0.9
Time (min)	Dependent Variable	5	70%		
1				22.6 ± 10.4	55.2 ± 2.0
2				20.9 ± 2.6	59.7 ± 1.0
3				21.9 ± 0.2	73.6 ± 4.0
4				22.1 ± 1.3	79.1 ± 0.7
5				22.2 ± 1.5	74.1 ± 1.8
Conc. (mM)	3	Dependent Variable	70%		
1.0				28.1 ± 1.0	57.5 ± 2.7
2.5				25.0 ± 0.5	76.2 ± 2.1
5				20.6 ± 0.7	75.9 ± 3.2
7.5				19.7 ± 0.3	78.2 ± 3.1
10				16.6 ± 0.7	76.0 ± 1.8

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58 **Table S10.** Detailed hydrodynamic diameter and transfer yield as functions of sonication
 59 amplitude, sonication time and surfactant concentration tested for sodium
 60 dodecylbenzenesulfonate (SDBS) coated NPs.

Dependent Variable	Time Tested (min)	Conc. Tested (mM)	Amplitude Tested	Hydrodynamic Diameter (nm)	Transfer Yield (%)
Amplitude	4	10	Dependent Variable		
50%				16.4 ± 3.1	36.3 ± 0.7
60%				15.0 ± 0.6	58.4 ± 2.2
70%				17.1 ± 1.5	63.3 ± 0.4
80%				16.2 ± 1.9	66.6 ± 1.7
Time (min)	Dependent Variable	10	70%		
2				21.0 ± 2.7	53.0 ± 1.7
4				16.6 ± 1.9	67.5 ± 1.6
6				15.1 ± 3.0	68.4 ± 1.6
Conc. (mM)	5	Dependent Variable	70%		
5				18.9 ± 0.3	45.5 ± 1.4
10				17.2 ± 0.8	62.8 ± 1.0
20				15.2 ± 4.1	66.1 ± 1.4
30				18.2 ± 3.4	64.9 ± 5.5
40				17.9 ± 0.3	64.5 ± 2.3

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62 **Table S11.** Detailed hydrodynamic diameter and transfer yield as functions of sonication
 63 amplitude, sonication time and surfactant concentration tested for dodecyltrimethylammonium
 64 bromide (C₁₂TAB) coated NPs.

Dependent Variable	Time Tested (min)	Conc. Tested (mM)	Amplitude Tested	Hydrodynamic Diameter (nm)	Transfer Yield (%)
Amplitude	4	10	Dependent Variable		
50%				22.3 ± 4.1	28.7 ± 3.4
60%				19.7 ± 1.7	33.8 ± 2.2
70%				21.3 ± 1.3	35.5 ± 5.2
80%				21.5 ± 0.8	42.8 ± 2.3
Time (min)	Dependent Variable	10	75%		
2				19.6 ± 1.0	38.5 ± 0.7
4				20.7 ± 1.0	45.1 ± 1.9
6				20.3 ± 2.3	47.4 ± 1.4
Conc. (mM)	4	Dependent Variable	75%		
5				34.5 ± 0.4	30.2 ± 2.9
10				20.7 ± 1.0	44.6 ± 1.6
20				36.1 ± 0.6	41.1 ± 3.5
30				43.8 ± 11.5	42.6 ± 2.2
40				59.2 ± 3.4	40.4 ± 4.5

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66 **Table S12.** Detailed hydrodynamic diameter and transfer yield as functions of sonication
 67 amplitude, sonication time and surfactant concentration tested for N,N-Dimethyl-N-
 68 dodecylglycine betaine (EMPIGEN) coated NPs.

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Dependent Variable	Time Tested (min)	Conc. Tested (mM)	Amplitude Tested	Hydrodynamic Diameter (nm)	Transfer Yield (%)
Amplitude	5	20	Dependent Variable		
50%				29.0 ± 9.8	30.2 ± 2.7
60%				36.9 ± 8.1	38.4 ± 1.2
70%				36.0 ± 6.9	42.3 ± 3.3
80%				33.1 ± 13.7	46.3 ± 4.5
90%				38.3 ± 1.9	46.1 ± 2.3
Time (min)	Dependent Variable	20	70%		
2				36.1 ± 0.9	46.4 ± 3.5
4				33.8 ± 9.9	42.4 ± 1.2
5				30.8 ± 7.4	50.5 ± 2.3
6				30.0 ± 0.9	55.8 ± 3.6
8				32.1 ± 0.7	52.8 ± 2.8
Conc. (mM)	6	Dependent Variable	70%		
10				56.8 ± 6.6	37.1 ± 1.3
20				31.5 ± 2.9	46.4 ± 2.6
30				27.9 ± 3.5	54.9 ± 1.9
40				17.3 ± 1.5	65.4 ± 4.3
50				21.6 ± 1.0	67.9 ± 3.1

70 **Table S13.** Total organic carbon (TOC) concentrations of bilayer coated iron oxide NPs (50
 71 ppm Fe).

Outer layer	TOC (ppm)	Outer layer density (mol/mol Fe ₃ O ₄) ^a	Outer layer density (mol/mol # NPs) ^a	Out layer density (10 ⁻⁵ mol/m ²) ^a
Oleic acid (OA)	102.8 ± 0.3	1.32	4953	3.99
Ricinoleic acid (RA)	100.1 ± 0.9	1.28	4792	3.86
Elaidic acid (EA)	85.0 ± 0.2	1.04	3917	3.16
Stearic acid (SA)	99.8 ± 0.9	1.27	4777	3.85
Palmitic acid (PA)	99.4 ± 0.2	1.43	5350	4.31
Myristic acid (MA)	94.3 ± 0.5	1.53	5730	4.62
Lauric acid (LA)	87.2 ± 0.5	1.62	6063	4.89
Decanoic acid (DA)	81.3 ± 0.3	1.77	6655	5.36
SDP	86.7 ± 0.5	1.60	6016	4.85
SDS	88.8 ± 0.4	1.65	6207	5.00
SDBS	100.5 ± 1.9	1.28	4814	3.88
C ₁₂ TAB	95.0 ± 0.1	1.44	5395	4.35
EMPIGEN	98.3 ± 1.6	1.41	5276	4.25

72 ^a Assumes the surface area per oleic acid molecule (*A*) to be 0.2 nm²,¹ the radius of IONPs (*r*)
 73 to be 8.1/2=4.05 nm, the density of iron oxide (ρ) is 5.17 g/cm³. The base oleic acid layer is
 74 calculated to be 23.2 mg per 50 mg Fe, which is equal to 0.28 mmol oleic acid/mmol Fe₃O₄.

$$\begin{aligned}
75 \quad \text{Base OA layer} &= \frac{(\text{base OA layer weight on each NP}) * \text{Fe concentration}}{\text{each NP weight (as Fe)}} = \\
76 \quad &\frac{(\# \text{ of OA molecule on each NP}) * (\text{molecular weight of OA}) / N_A}{(\text{volume of each NP}) * (\text{density of iron oxide}) * (\text{percentage of Fe in each NP})} \text{Fe concentration} = \\
77 \quad &\frac{(4\pi r^2 / A) * (M_w \text{ of OA}) / N_A}{(4\pi r^3 / 3) * \rho_{Fe_3O_4} * (3 * M_w \text{ of Fe} / M_w \text{ of Fe}_3O_4)} \text{Fe concentration} \quad (1)
\end{aligned}$$

$$78 \quad \text{The C content in the base OA layer} = \frac{18 * (M_w \text{ of C})}{M_w \text{ of OA}} \text{base OA layer content} \quad (2)$$

79 The C concentration in the base OA layer was calculated to be 17.74 mg per 50 mg Fe. By
80 subtracting the C concentration from the total carbon concentration, we can derive the C
81 concentration in the second layer and thus the mol concentration of second layer.

$$82 \quad \text{The mol concentration of } Fe_3O_4 = \frac{\text{Fe concentration}}{3 * (M_w \text{ of Fe})} = 0.298 \text{ mmol as in 50 ppm Fe} \quad (3)$$

$$83 \quad \text{Outer layer density} \left(\frac{\text{mol}}{\text{mol } Fe_3O_4} \right) = \frac{\text{mol concentration of second layer}}{\text{mol concentration of } Fe_3O_4} \quad (4)$$

$$84 \quad \text{Outer layer density} \left(\frac{\text{mol}}{\text{mol \# NPs}} \right) = \frac{\text{mol concentration of second layer}}{\text{mol \# NPs}} =$$

$$85 \quad \frac{\text{mol concentration of second layer}}{(\text{total weight as } Fe_3O_4 / \text{each NP weight} * N_A)} \quad (5)$$

$$86 \quad \text{Outer layer density} \left(\frac{\text{mol}}{m^2} \right) = \frac{\text{mol concentration of second layer}}{\text{mol \# NPs}} =$$

$$87 \quad \frac{\text{mol concentration of second layer}}{(\text{surface area of each NP}) * (\text{total weight as } Fe_3O_4) / (\text{each NP weight})} \quad (6)$$

88 **Table S14.** Summary of CCC values for bilayer coated iron oxide NPs.

Outer layer	Critical Coagulation Concentration	
	NaCl (mM)	CaCl ₂ (mM)
Oleic acid (OA)	710	10.6
Ricinoleic acid (RA)	746	10.8
Elaidic acid (EA)	260	7.4
Stearic acid (SA)	452	9.3
Palmitic acid (PA)	257	5.3
Myristic acid (MA)	94	3.9
Lauric acid (LA)	16	0.5
Decanoic acid (DA)	27	1.6
SDP	250	3.6
SDS	45	1.4
SDBS	46	6.0
C ₁₂ TAB	555	11.1
EMPIGEN	766	11.3

89

90 **References**

- 91 1. A. G. Roca, M. P. Morales, K. O'Grady and C. J. Serna, *Nanotechnology*, 2006, **17**,
92 2783.