

**Electronic Supplementary Information**  
**(ESI)**

**Electrode instead of catalyst and enzyme. A greener  
protocol for the synthesis of new 2-  
hydroxyacetamides derivatives containing  $\gamma$ -lactone  
ring**

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Zeinodini-Meimand

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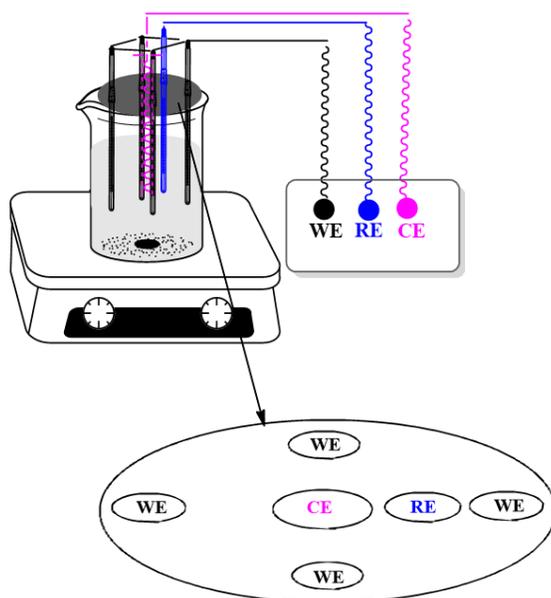
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## Apparatus

Cyclic voltammetry, controlled-potential coulometry and preparative electrolysis were performed using an Autolab model PGSTAT302N potentiostat/galvanostat. The working electrode used in the voltammetry experiments was a glassy carbon disc (1.8 mm<sup>2</sup> area) and platinum wire was used as a counter electrode. The working electrode used in controlled-potential coulometry and macroscale electrolysis was an assembly of four carbon rods (6 mm diameter and 4 cm length), placed as single rods in the edges of a square, and large stainless steely gauze constitutes the counter electrode. The working electrode potentials were measured versus Ag/AgCl (all electrodes from AZAR electrode). The electrolysis was performed in a simple cell (a narrow beaker type cell, 100 ml), equipped with a magnetic stirrer.



Arrangement of the electrodes and cell

## Reagens

3,5-di-*tert*-butylcatechol, *n*-butylamine, *n*-propylamine, ethylamine, methylamine, benzylamine, cyclopentylamine, cyclohexylamine, cycloheptylamine, cyclooctylamine were reagent-grade materials and carbonate salts were of pro-analysis grade, from E. Merck. These chemicals were used without further purification.

## Electrochemical synthesis **9a-17a**: General procedure

### Controlled-potential method

In a typical procedure, four carbon rods as working electrodes, a stainless steely gauze as auxiliary electrode along with an Ag/AgCl reference electrode were immersed into an undivided cell containing a mixture (60 mL) of water (carbonate buffer,  $c = 0.2$  M, pH = 11)/acetonitrile (40/60 v/v). This mixture was pre-electrolyzed at the 0.05 V versus Ag/AgCl, then 1 mmol of 3,5-di-*tert*-butylcatechol and 1 mmol of **9-17** were added to the cell and the mixture was stirred until homogeneity was achieved. The electrolysis was terminated when the decay of the current became more than 95% (within about 3-4 hours). The process was interrupted during the electrolysis and the carbon anode was washed in acetone in order to reactivate it. At the end of electrolysis, after evaporation of acetonitrile, the residue was transferred to a separating funnel and extracted with cyclohexane or *n*-hexane. The extracted portion was recrystallized in *n*-hexane or chloroform. After purification, all products were characterized by: IR,  $^1\text{H}$  NMR,  $^{13}\text{C}$  NMR and MS. Moreover, product **9a** was also characterized by single crystal X-ray diffraction.

### Constant-current method (Galvanostatic method)

A mixture (60 mL) of water (carbonate buffer,  $c = 0.2$  M, pH = 11)/acetonitrile (40/60 v/v) containing 1 mmol of 3,5-di-*tert*-butylcatechol and 1 mmol of **9-17** was electrolyzed in an undivided cell equipped with a carbon anode (an assembly of four rods, with 30 cm<sup>2</sup>) and a large stainless steely gauze cathode at 25 °C under a constant-current density of 1.0 mA cm<sup>-2</sup>. The other steps are similar to those described above in the controlled-potential method.

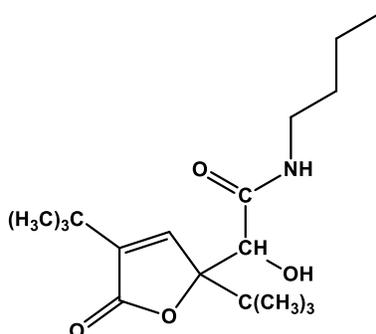
**Table S1.** Electrochemical synthesis of **9a-17a** at constant current condition<sup>a</sup>

Entry	Product	Yields <sup>b</sup> (%)
1	<b>9a</b>	61
2	<b>10a</b>	54
3	<b>11a</b>	62
4	<b>12a</b>	50
5	<b>13a</b>	55
6	<b>14a</b>	60
7	<b>15a</b>	50
8	<b>16a</b>	62
9	<b>17a</b>	70

<sup>a</sup>General procedure: **1** (1 mmol), **9-17** (1 mmol), acetonitrile (24ml), carbonat buffer (36ml), current density 1 mA cm<sup>-2</sup>.  
<sup>b</sup>Yield of isolated product.

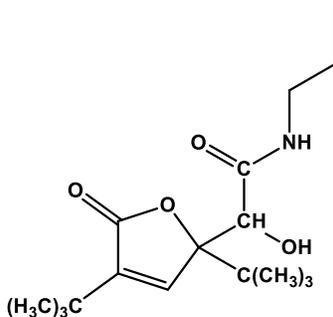
### Characterization of Products

**9a:** *N*-butyl-2-(2,4-di-*tert*-butyl-5-oxo-2,5-dihydrofuran-2-yl)-2-hydroxyacetamide



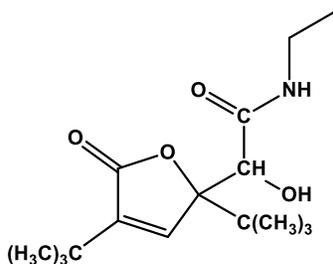
<sup>1</sup>H NMR (300 MHz, acetone-*d*<sub>6</sub>),  $\delta$  (ppm): 0.88 (t, 3H), 1.06 (s, 9H), 1.21 (s, 9H), 1.32 (m, 2H), 1.44(m, 2H), 3.20 (m, 2H), 4.49 (d, 1H), 4.61 (d, 1H), 7.12 (NH, 1H), 7.31 (ring, 1H); <sup>13</sup>C NMR (75 MHz, acetone-*d*<sub>6</sub>),  $\delta$  (ppm): 13.2, 19.8, 25.7, 27.4, 31.3, 31.39, 39.3, 37.7, 39.2, 73.6, 89.3, 142.7, 146.5, 169.9, 170.7; IR (KBr): 3371, 3323, 1741, 1658, 1313 cm<sup>-1</sup>; MS (EI) *m/z* (relative intensity): 326 [M+H<sup>+</sup>] (5), 269 (4), 196 (95), 181 (100), 169 (45), 130 (30), 57(18).

**10a:** *N*-propyl-2-(2,4-di-tert-butyl-5-oxo-2,5-dihydrofuran-2-yl)-2-hydroxyacetamide



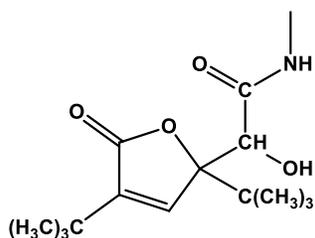
$^1\text{H}$  NMR (300 MHz, acetone- $d_6$ ),  $\delta$  (ppm): 0.86 (t, 3H), 1.05 (s, 9H), 1.20 (s, 9H), 1.46 (t, 2H), 3.07 (m, CH<sub>2</sub>), 4.50 (d, 1H), 4.73 (d, 1H), 7.10 (NH, 1H), 7.32 (ring, 1H);  $^{13}\text{C}$  NMR (75 MHz, acetone- $d_6$ ),  $\delta$  (ppm): 10.9, 22.3, 25.6, 27.4, 31.3, 37.7, 41.2, 73.7, 89.4, 142.7, 146.57, 170.0, 170.8; IR (KBr): 3381, 3327, 1743, 1658, 1315  $\text{cm}^{-1}$ ; MS (EI)  $m/z$  (relative intensity): 313 [ $\text{M}+2\text{H}^+$ ] (3), 312 [ $\text{M}+1\text{H}^+$ ] (10), 196 (55), 181 (63), 169 (42), 116 (42), 57 (100), 43 (98), 41 (90).

**11a:** *N*-ethyl-2-(2,4-di-tert-butyl-5-oxo-2,5-dihydrofuran-2-yl)-2-hydroxyacetamide



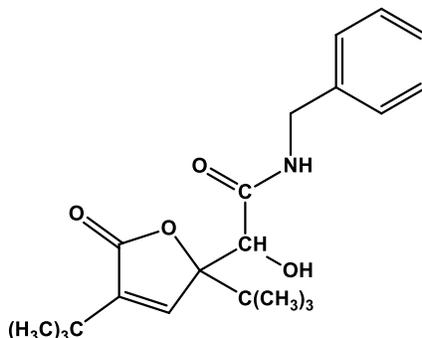
$^1\text{H}$  NMR (300 MHz, acetone- $d_6$ ),  $\delta$  (ppm): 1.07 (s, 12H), 1.21 (s, 9H), 3.19 (t, 2H), 4.50 (q, 2H), 7.13 (NH, 1H), 7.31 (ring, 1H);  $^{13}\text{C}$  NMR (75 MHz, acetone- $d_6$ ),  $\delta$  (ppm): 13.9, 25.6, 27.4, 31.3, 34.2, 37.6, 73.5, 89.2, 142.8, 146.4, 169.7, 170.7; IR (KBr): 3385, 3321, 1741, 1663, 1317  $\text{cm}^{-1}$ ; MS (EI)  $m/z$  (relative intensity): 300 [ $\text{M}+3$ ] (1), 299 [ $\text{M}+2$ ] (4), 298 [ $\text{M}+1$ ] (40), 296 (1), 196 (12), 181(20), 102 (8), 72 (25), 57 (100), 41 (61), 29 (75).

**12a:** *N*-methyl-2-(2,4-di-tert-butyl-5-oxo-2,5-dihydrofuran-2-yl)-2-hydroxyacetamide



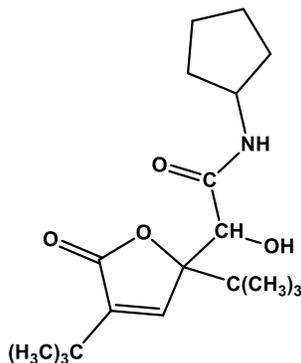
$^1\text{H}$  NMR (300 MHz, acetone- $d_6$ ),  $\delta$  (ppm): 1.07 (s, 9H), 1.21 (s, 9H), 2.70 (d, CH<sub>3</sub>), 4.47 (d, 1H), 4.60 (d, 1H), 7.10 (NH, 1H), 7.30 (ring, 1H);  $^{13}\text{C}$  NMR (75 MHz, acetone- $d_6$ ),  $\delta$  (ppm): 25.5, 25.6, 27.4, 31.2, 37.5, 73.7, 89.3, 142.8, 146.4, 170.4, 170.7; IR (KBr): 3362, 3319, 1740, 1664, 1313  $\text{cm}^{-1}$ ; MS (EI)  $m/z$  (relative intensity): 285 [M+2] (1), 284 [M+1] (6), 196 (39), 181 (65), 169 (55), 125 (32), 57 (100), 41 (54), 29 (63).

**13a:** N-benzyl-2-(2,4-di-tert-butyl-5-oxo-2,5-dihydrofuran-2-yl)-2-hydroxyacetamide



$^1\text{H}$  NMR (300 MHz, acetone- $d_6$ ),  $\delta$  (ppm): 1.01 (s, 9H), 1.22 (s, 9H), 2.97 (s, 2H), 4.28 (d, 1H), 4.40 (d, 1H), 4.60 (d, 1H), 4.74 (d, 1H), 7.27 (m, 6H, aromatic), 7.61 (NH, 1H);  $^{13}\text{C}$  NMR (75 MHz, acetone- $d_6$ ),  $\delta$  (ppm): 170.8, 170.2, 146.5, 142.8, 138.6, 128.29, 127.8, 127.0, 89.4, 74.1, 43.20, 37.7, 31.3, 27.4, 25.7; IR (KBr): 3377, 3292, 3086, 2960, 2872, 1739, 1662, 1315  $\text{cm}^{-1}$ ; MS (EI)  $m/z$  (relative intensity): 361 [M+2] (2), 360 [M+1] (12), 285 (5), 196 (40), 181 (50), 164 (30), 106 (40), 91 (100), 57 (80), 41 (55), 29 (33).

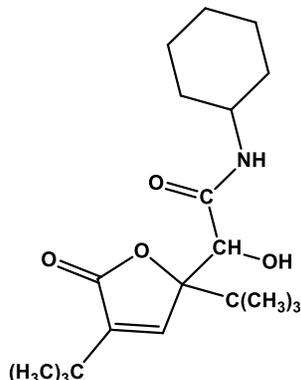
**14a:** N-cyclopentyl-2-(2,4-di-tert-butyl-5-oxo-2,5-dihydrofuran-2-yl)-2-hydroxyacetamide



$^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ ),  $\delta$  (ppm): 1.12 (s, 9H), 1.25 (s, 9H), 1.40 (m, 2H), 1.57 (m, 2H), 1.69 (m, 2H), 1.94 (m, 2H), 4.11 (m, 2H), 4.47 (s, 1H), 5.75 (d, lactone ring, 1H), 7.13 (s, NH, 1H);  $^{13}\text{C}$  NMR (125 MHz,  $\text{CDCl}_3$ ),  $\delta$  (ppm): 24.1, 24.1, 26.6, 28.5, 32.2, 32.9, 33.7, 38.8, 52.4, 73.0, 89.9, 144.6, 146.4, 169.8; IR (KBr): 1636, 1733, 2958, 3309

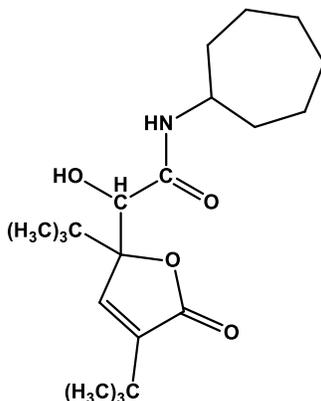
cm<sup>-1</sup>; MS (EI) *m/z* (relative intensity): 339 [M+2] (20), 338 [M+1] (80), 337 [M] (10), 263 (18), 197 (75), 181 (90), 142 (30), 84 (45).

**15a:** *N*-cyclohexyl-2-(2,4-di-tert-butyl-5-oxo-2,5-dihydrofuran-2-yl)-2-hydroxyacetamide



<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>),  $\delta$  (ppm): 1.12 (s, 9H), 1.18 (m, 2H), 1.24 (s, 9H), 1.33 (m, 3H), 1.61 (q, 1H), 1.71 (q, 1H), 1.83, 1.92 (d, 1H), 3.63 (m, 1H), 4.47 (s, 1H), 5.71 (lactone ring, 1H), 7.14 (NH, 1H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>),  $\delta$  (ppm): 171.6, 169.4, 146.5, 144.4, 89.9, 73.0, 50.0, 38.8, 33.3, 32.9, 32.2, 32.0, 28.5, 26.6, 25.7, 25.2; IR (KBr): 3439, 3320, 2959, 2937, 2857, 1753, 1649, 1551 cm<sup>-1</sup>; MS (EI) *m/z* (relative intensity): 352 [M+1] (2), 295 (1), 277 (2), 197 (5), 196 (39), 181 (50), 83 (48), 67 (48), 57(100), 41(77).

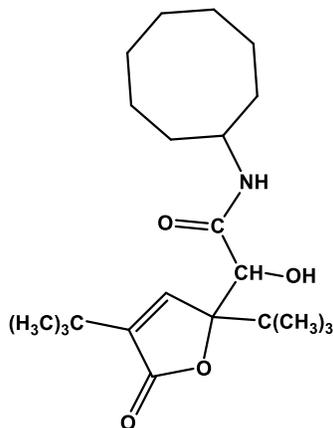
**16a:** *N*-cycloheptyl-2-(2,4-di-tert-butyl-5-oxo-2,5-dihydrofuran-2-yl)-2-hydroxyacetamide



<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>),  $\delta$  (ppm): 7.14 (s, 1H), 5.78 (d, 1H), 4.45 (s, 1H), 4.14 (s, 1H), 3.81 (m, 1H), 1.88 (m, 2H), 1.61 (m, 4H), 1.47 (m, 2H), 1.43 (m, 4H), 1.39 (s, 9H), 1.11 (s, 9H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>),  $\delta$  (ppm): 171.6, 169.1, 146.5, 144.4, 89.9, 73.1, 52.2, 38.8, 35.3, 35.0, 32.2, 28.5, 28.4, 28.4, 26.6, 24.4, 24.4; IR (KBr): 3310, 2957,

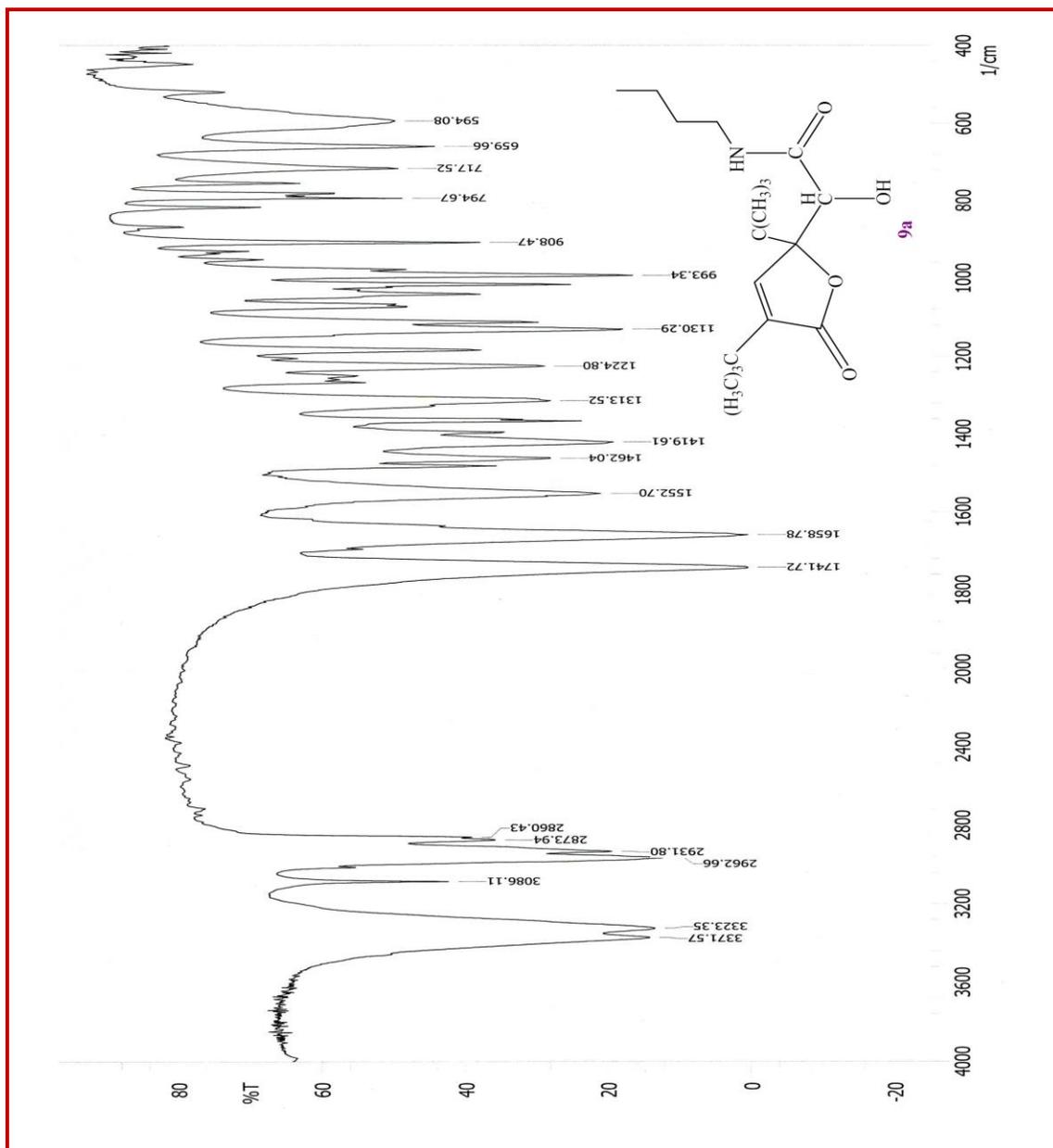
2932, 2866, 1736, 1632  $\text{cm}^{-1}$ ; MS (EI)  $m/z$  (relative intensity): 367 [M+2] (1), 366 [M+1] (2), 269 (0.5), 196 (2), 181 (2), 57 (100), 41 (17).

**17a:** *N*-cyclooctyl-2-(2,4-di-*tert*-butyl-5-oxo-2,5-dihydrofuran-2-yl)-2-hydroxyacetamide

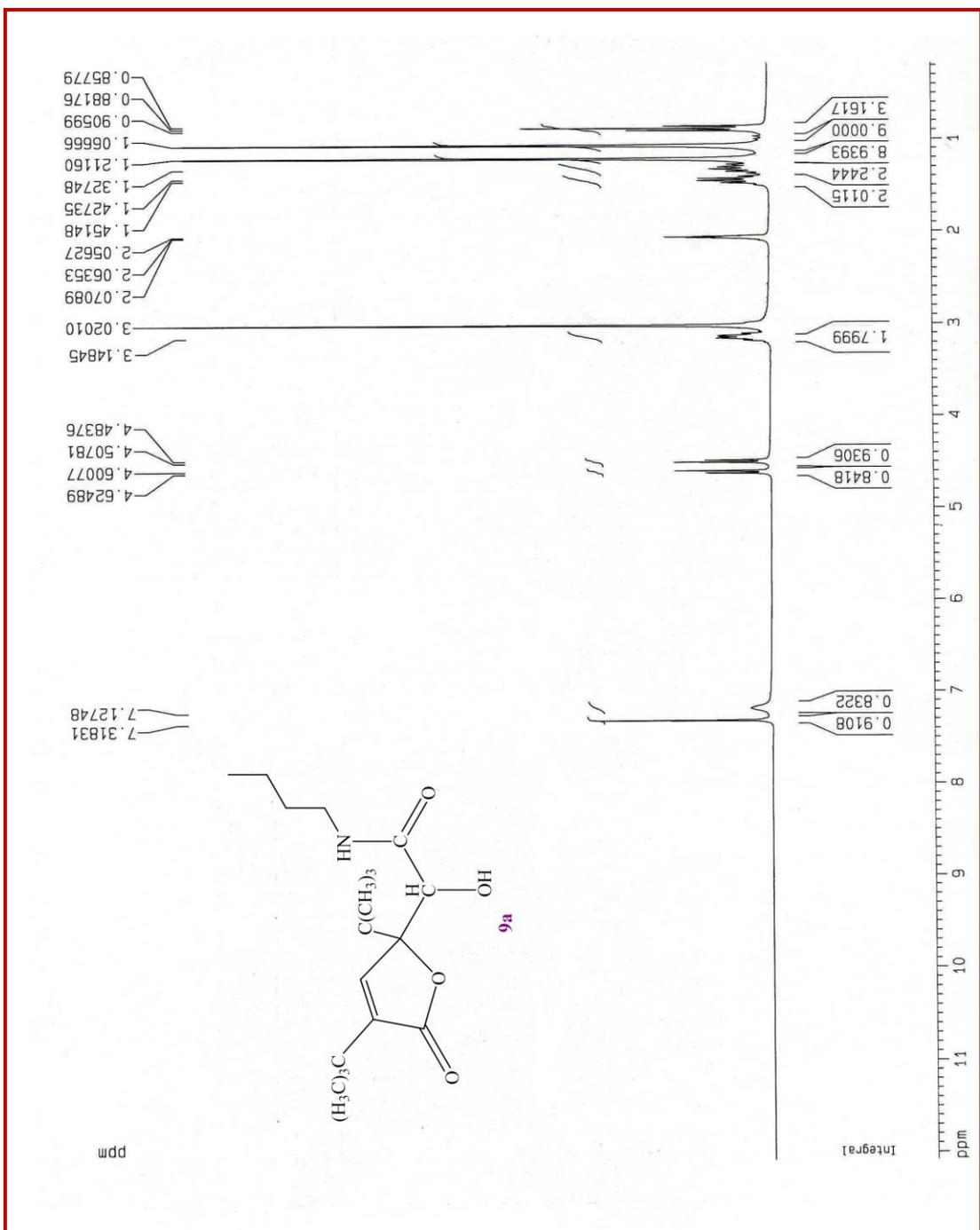


$^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ ),  $\delta$  (ppm): 7.14 (s, 1H), 5.77 (d, 1H), 4.44 (d, 1H), 4.14 (s, 1H), 3.87 (t, 1H), 1.77 (m, 2H), 1.65 (t, 2H), 1.55 (m, 10H), 1.24 (s, 9H), 1.12 (s, 9H);  $^{13}\text{C}$  NMR (125 MHz,  $\text{CDCl}_3$ ),  $\delta$  (ppm): 171.6, 169.1, 146.5, 144.4, 89.9, 73.0, 51.2, 38.9, 32.6, 32.3, 32.2, 28.5, 27.4, 27.3, 26.6, 25.9, 24.0; IR (KBr): 3316, 2926, 1737, 1633, 1533  $\text{cm}^{-1}$ ; MS (EI)  $m/z$  (relative intensity): 382 [M+3] (1), 380 [M+1] (6), 364 (2), 323 (6), 290 (10), 226 (12), 195 (100), 184 (50), 181 (85), 169 (80), 125 (40), 83 (53), 69 (55), 56 (98), 41 (42).

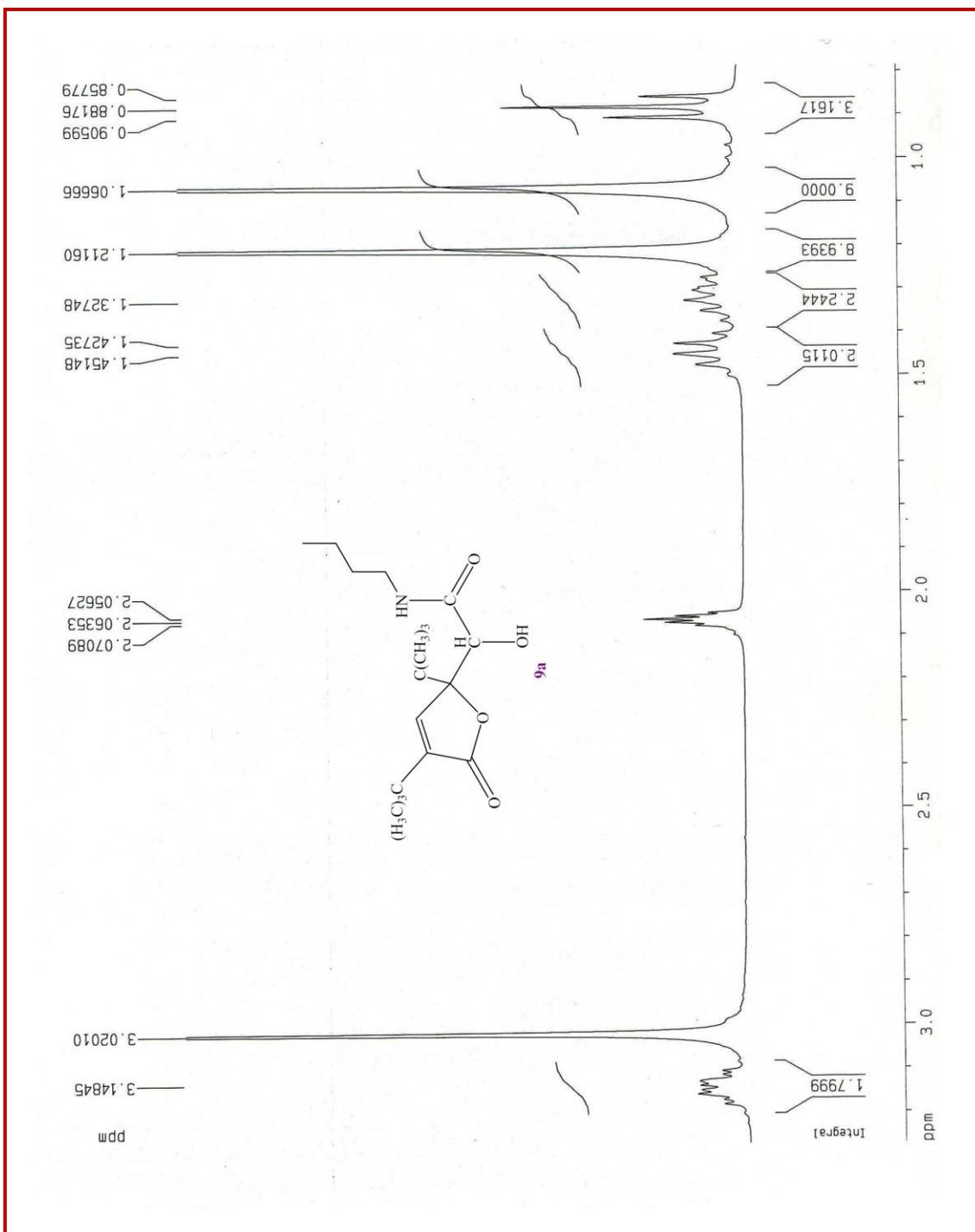
# FTIR, <sup>1</sup>H NMR, <sup>13</sup>C NMR and mass spectra



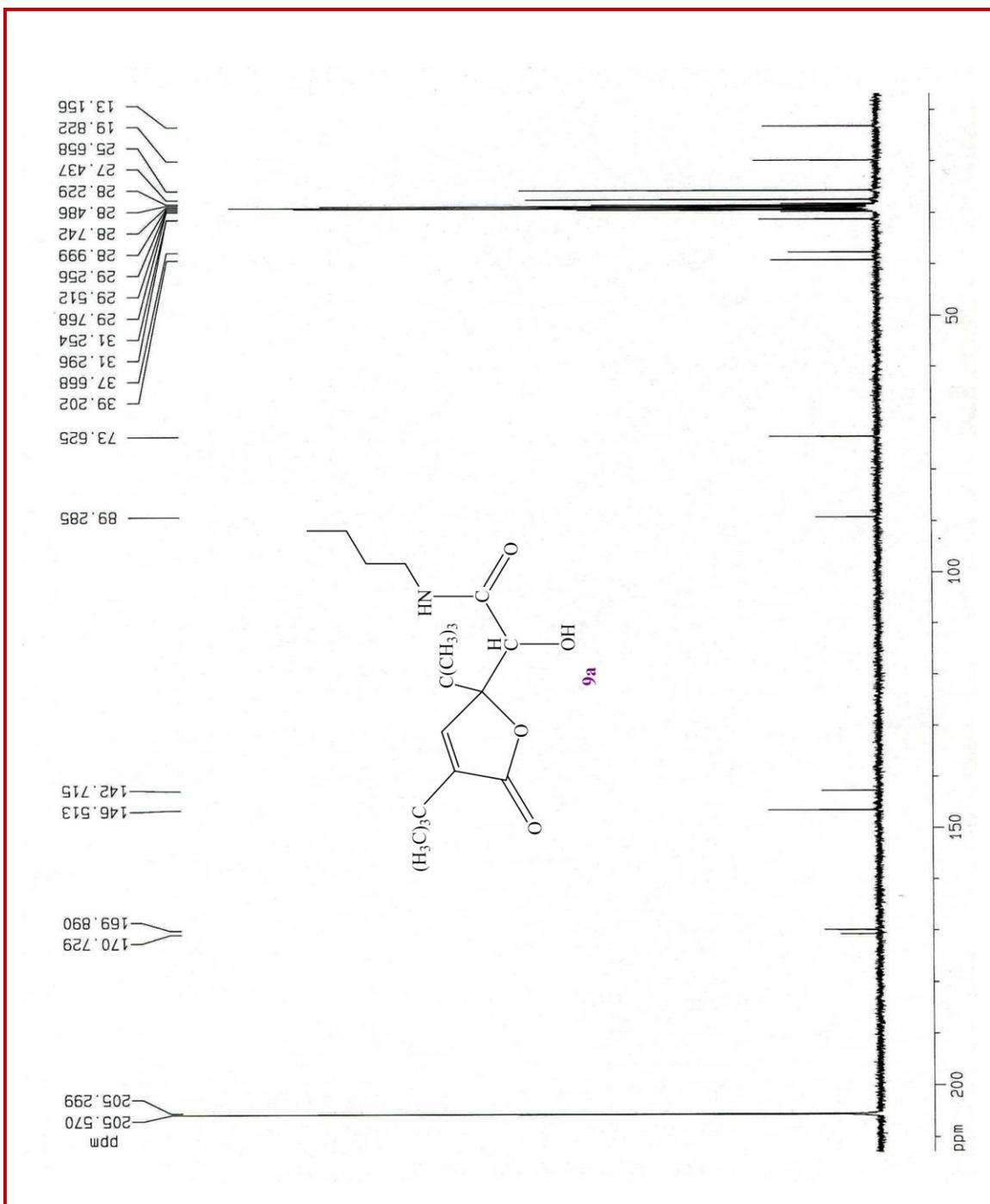
FTIR of 9a



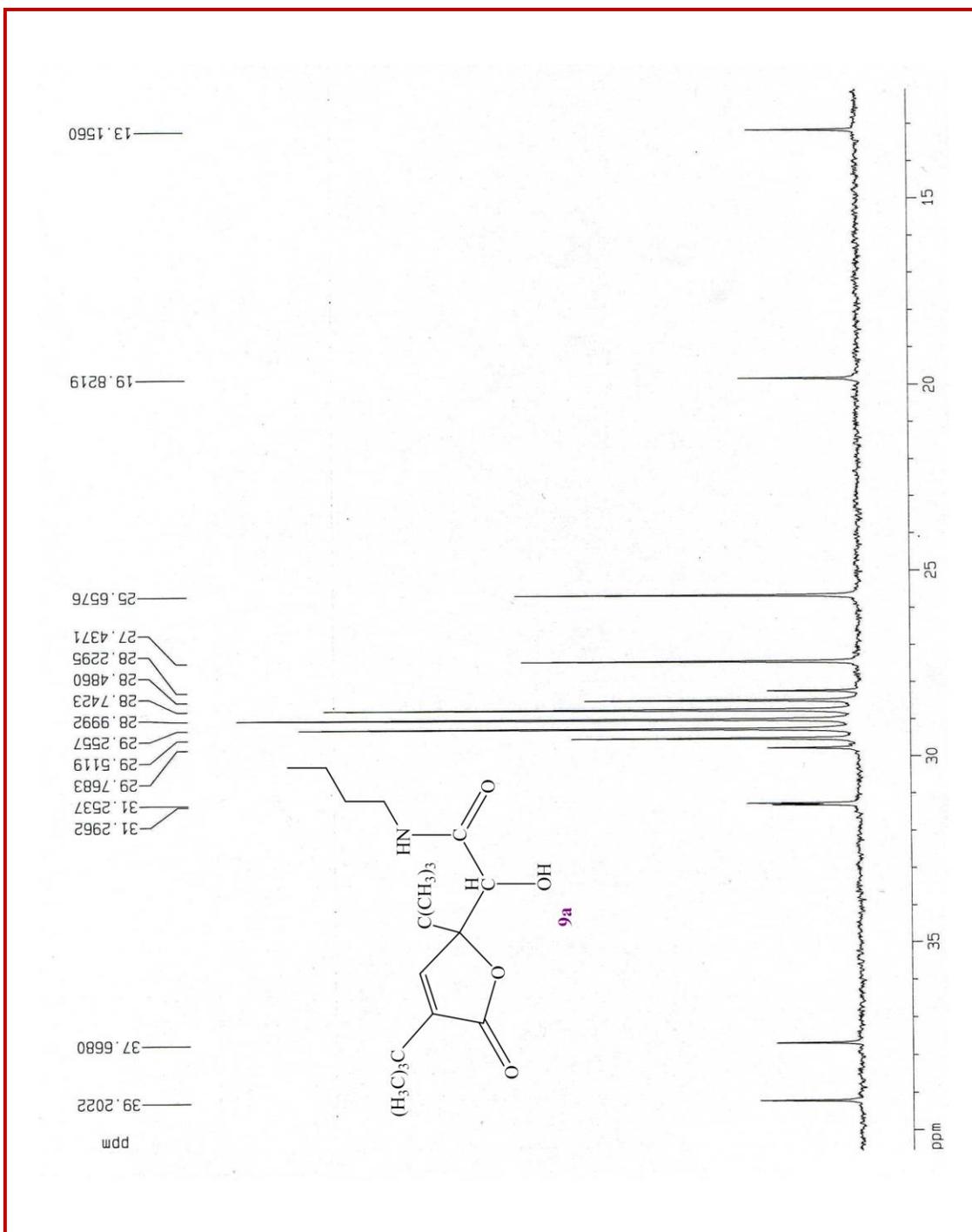
**<sup>1</sup>H NMR of 9a**



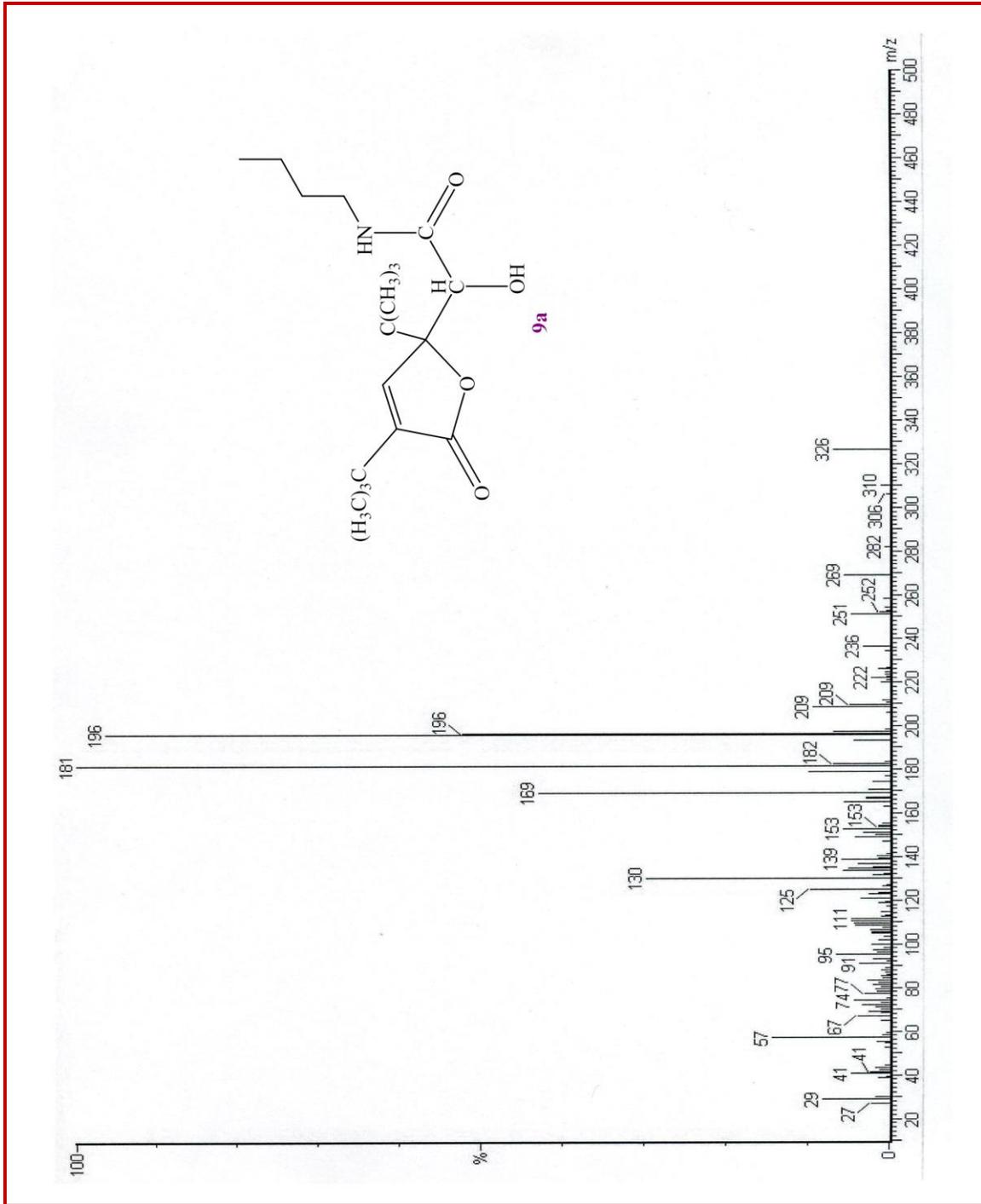
Expanded  $^1\text{H}$  NMR of 9a



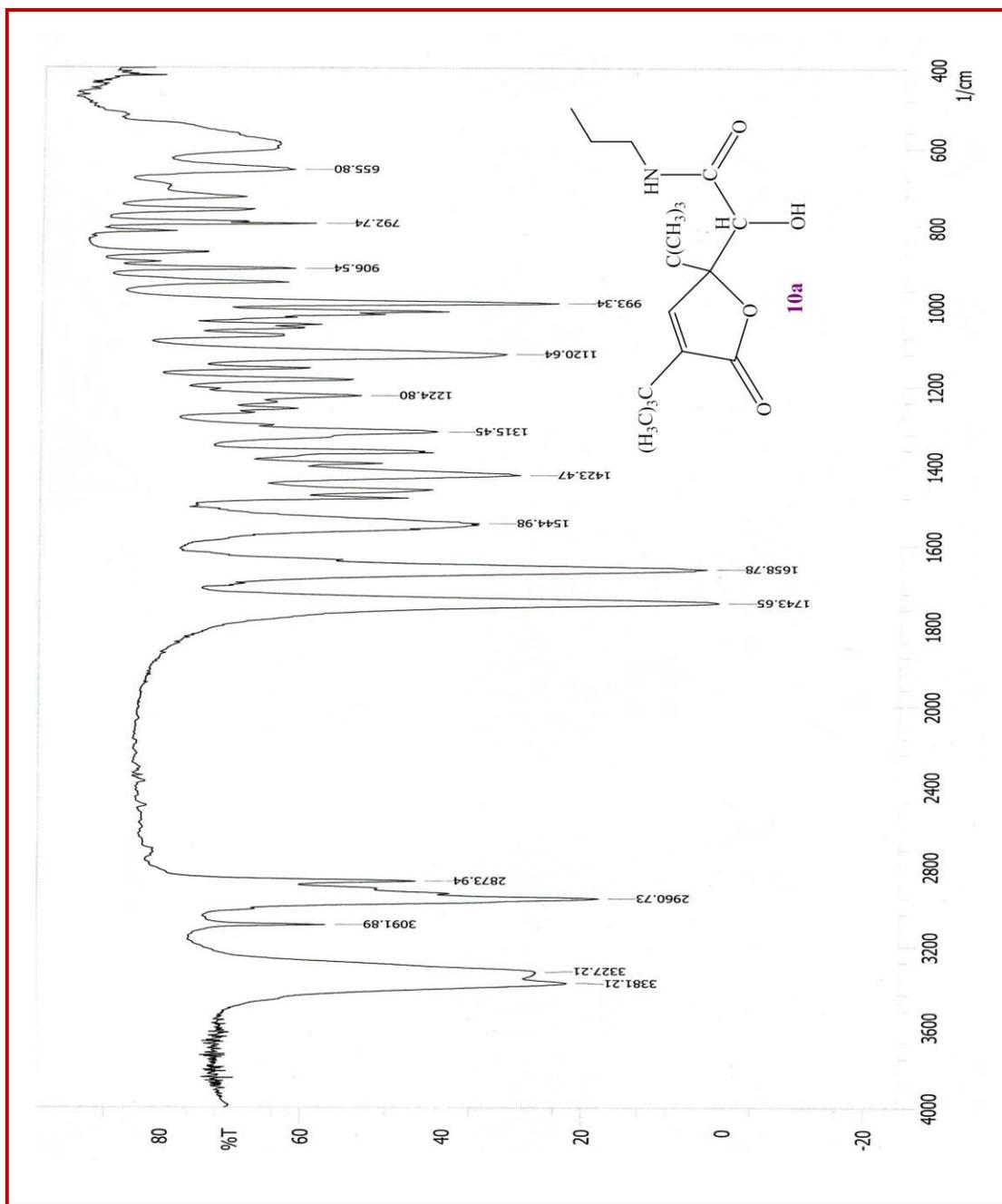
**<sup>13</sup>C NMR of 9a**



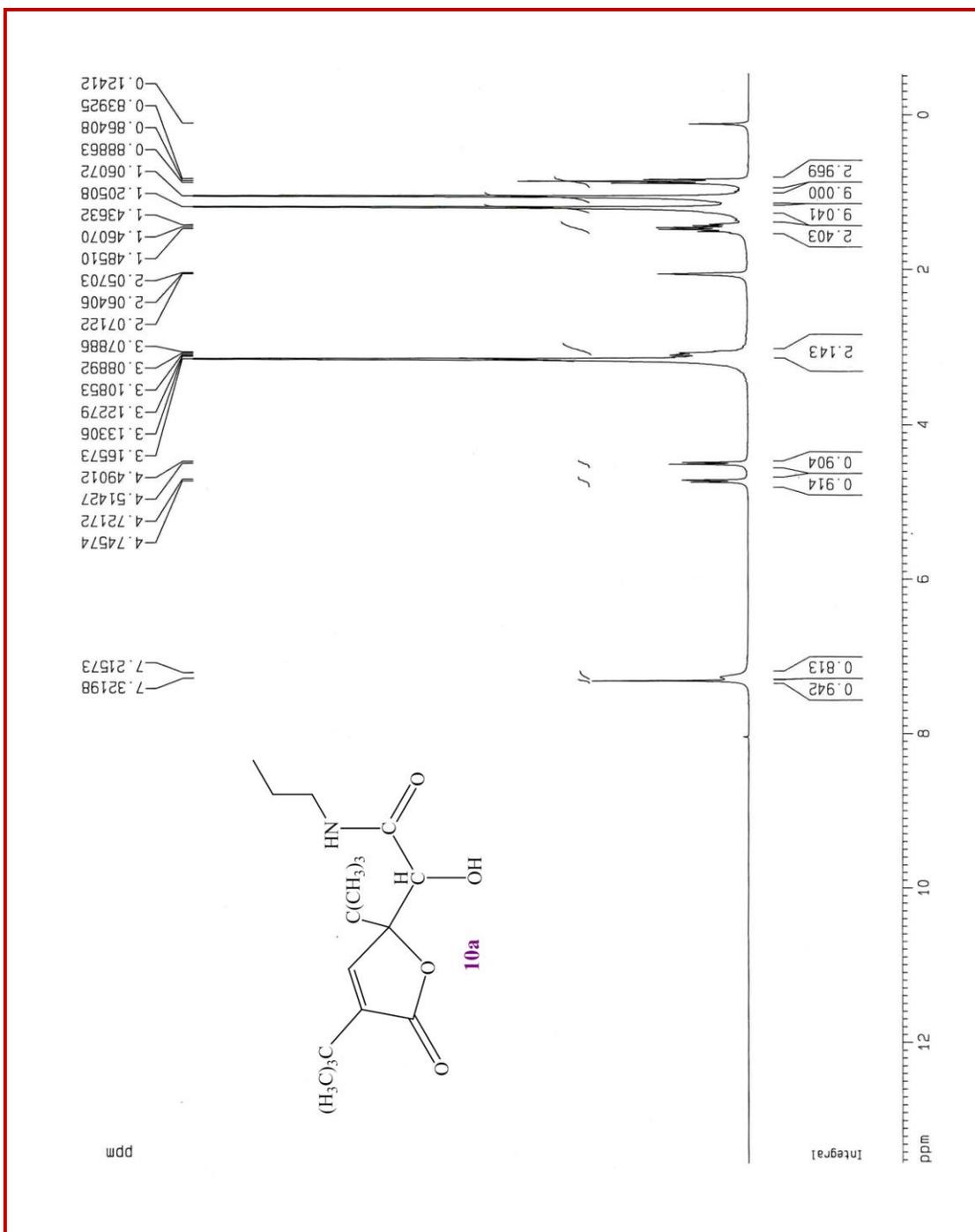
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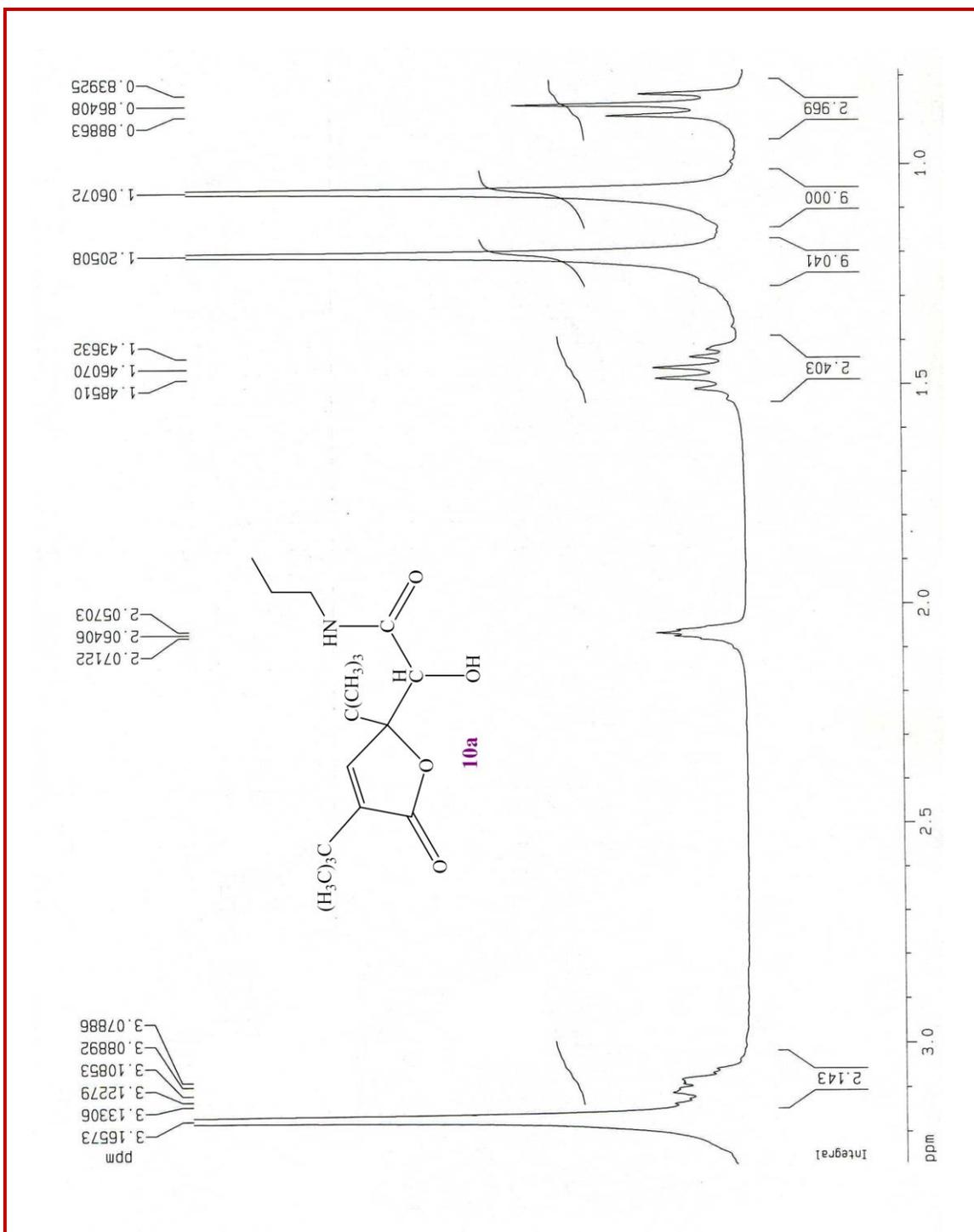


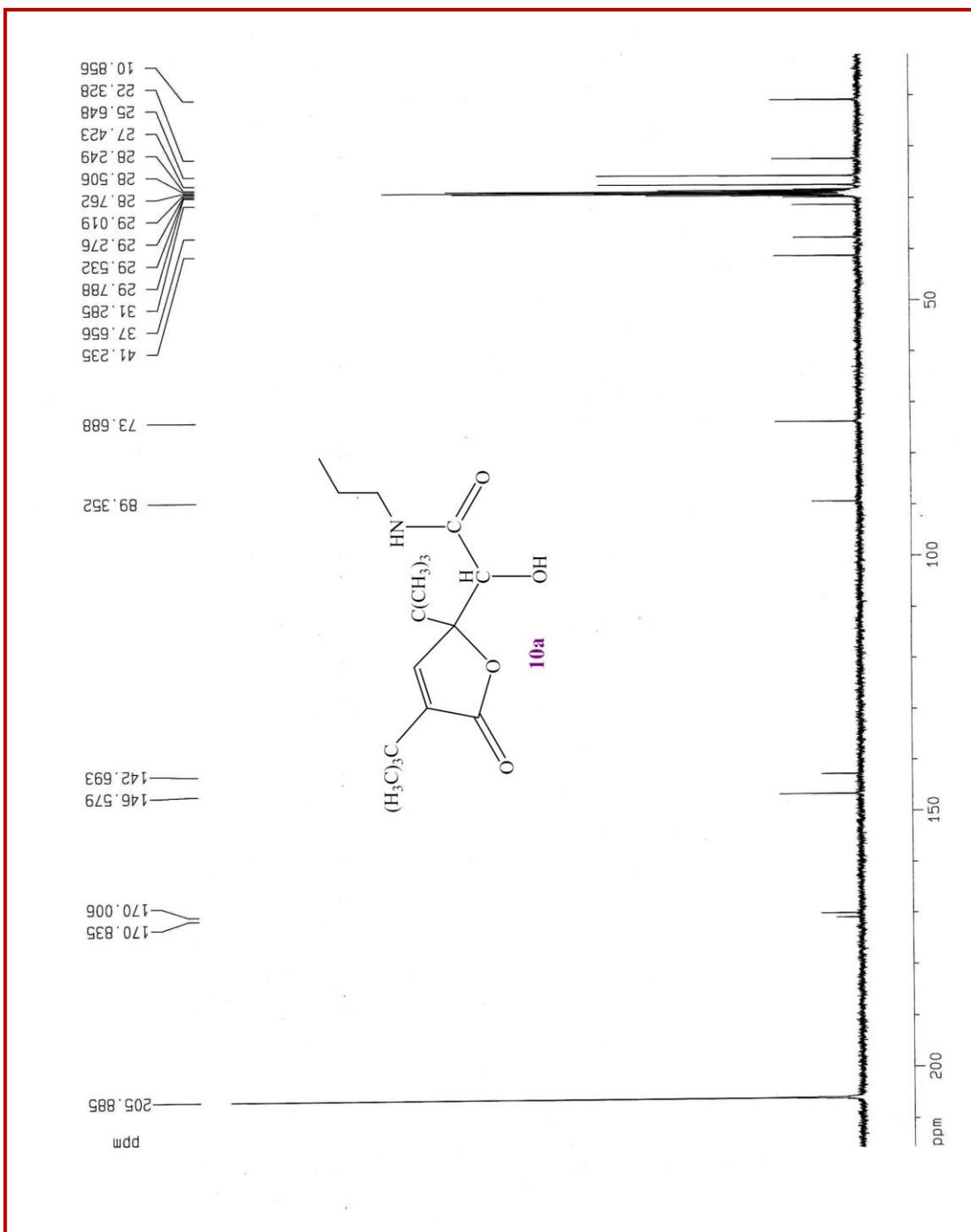
MS of 9a



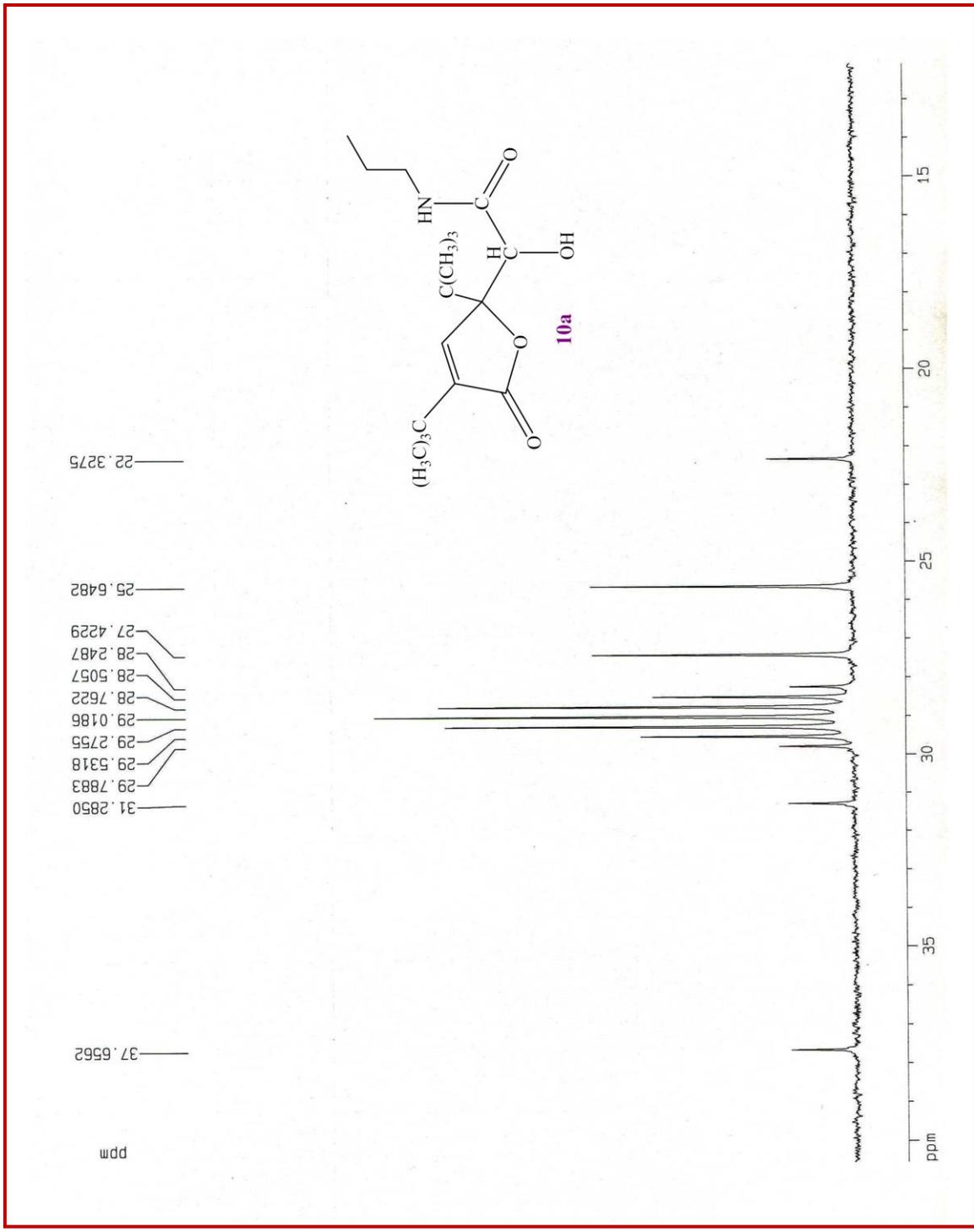
FTIR of 10a



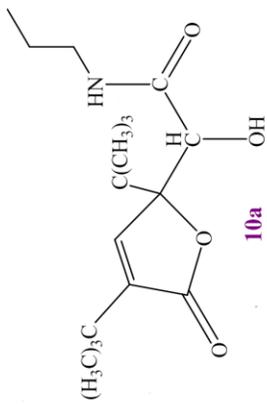
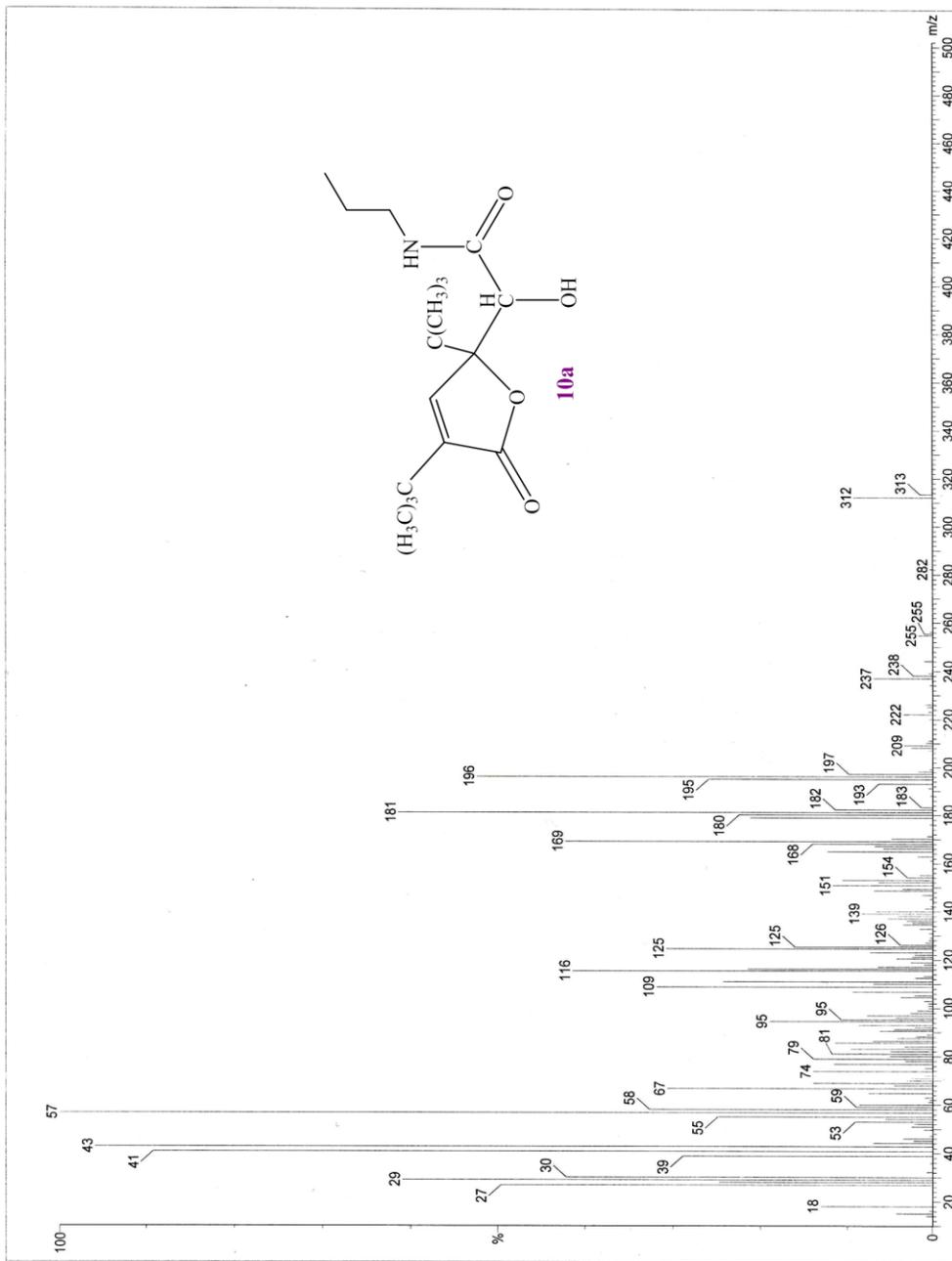




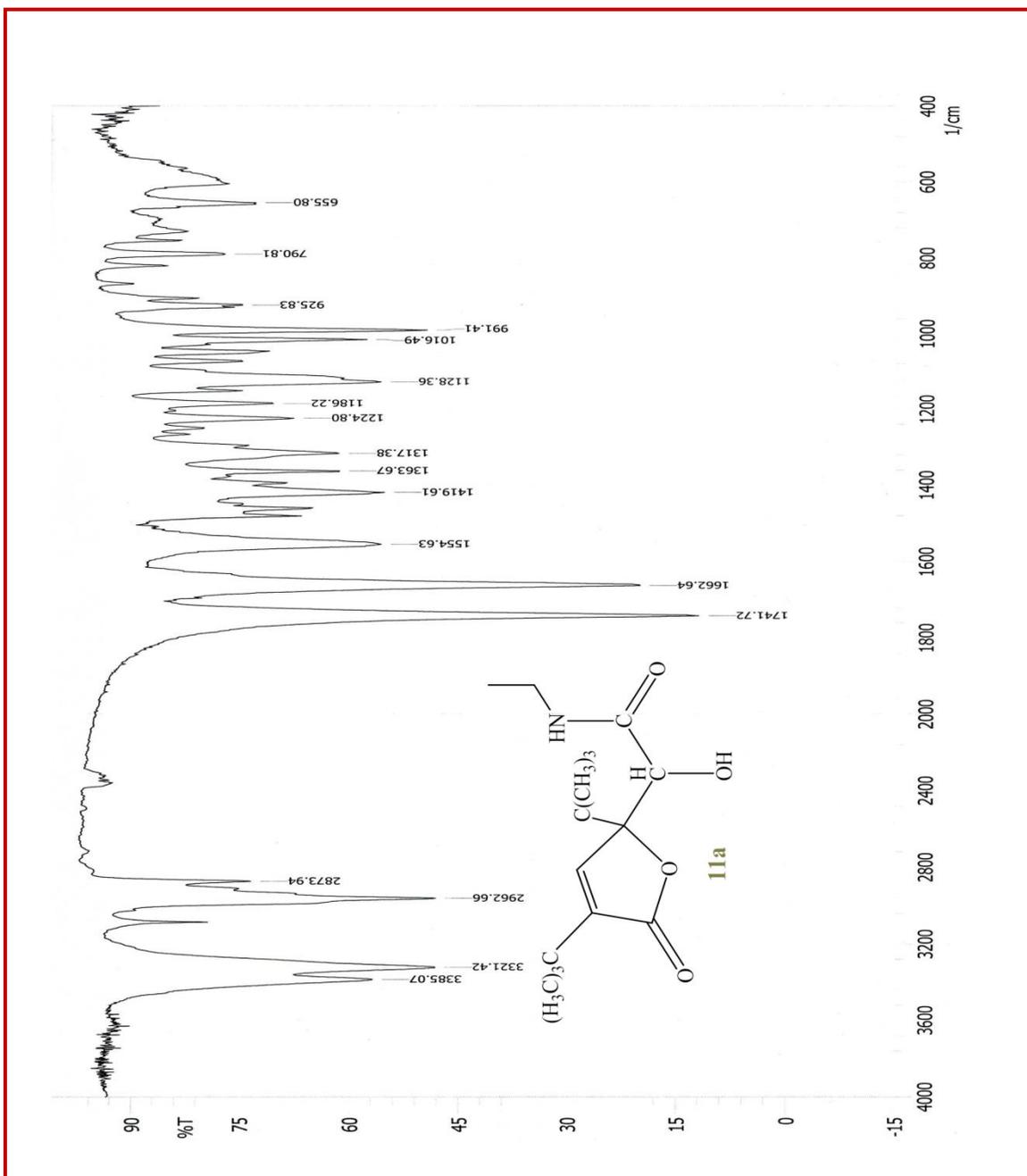
**<sup>13</sup>C NMR of 10a**



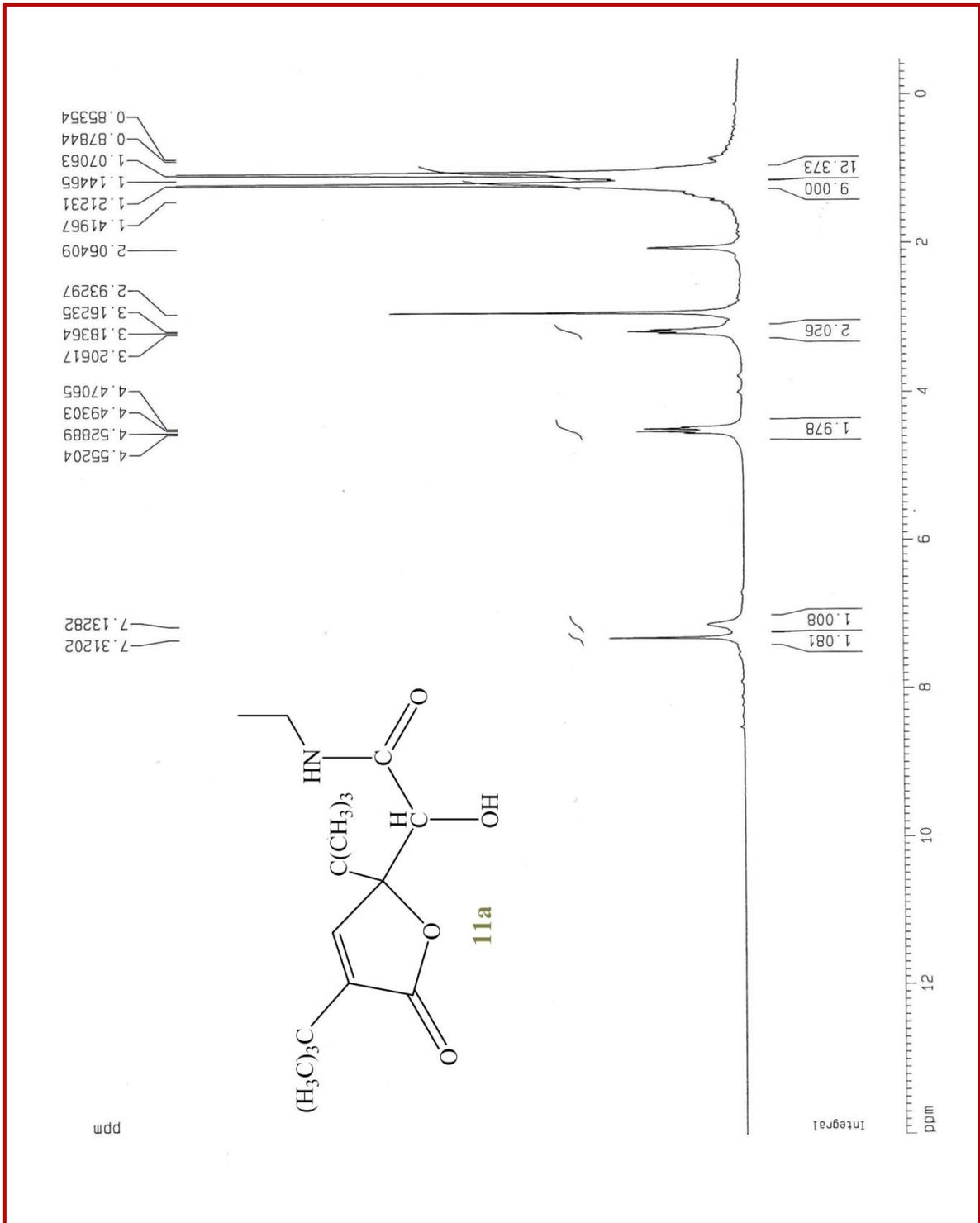
Expanded  $^{13}\text{C}$  NMR of 10a



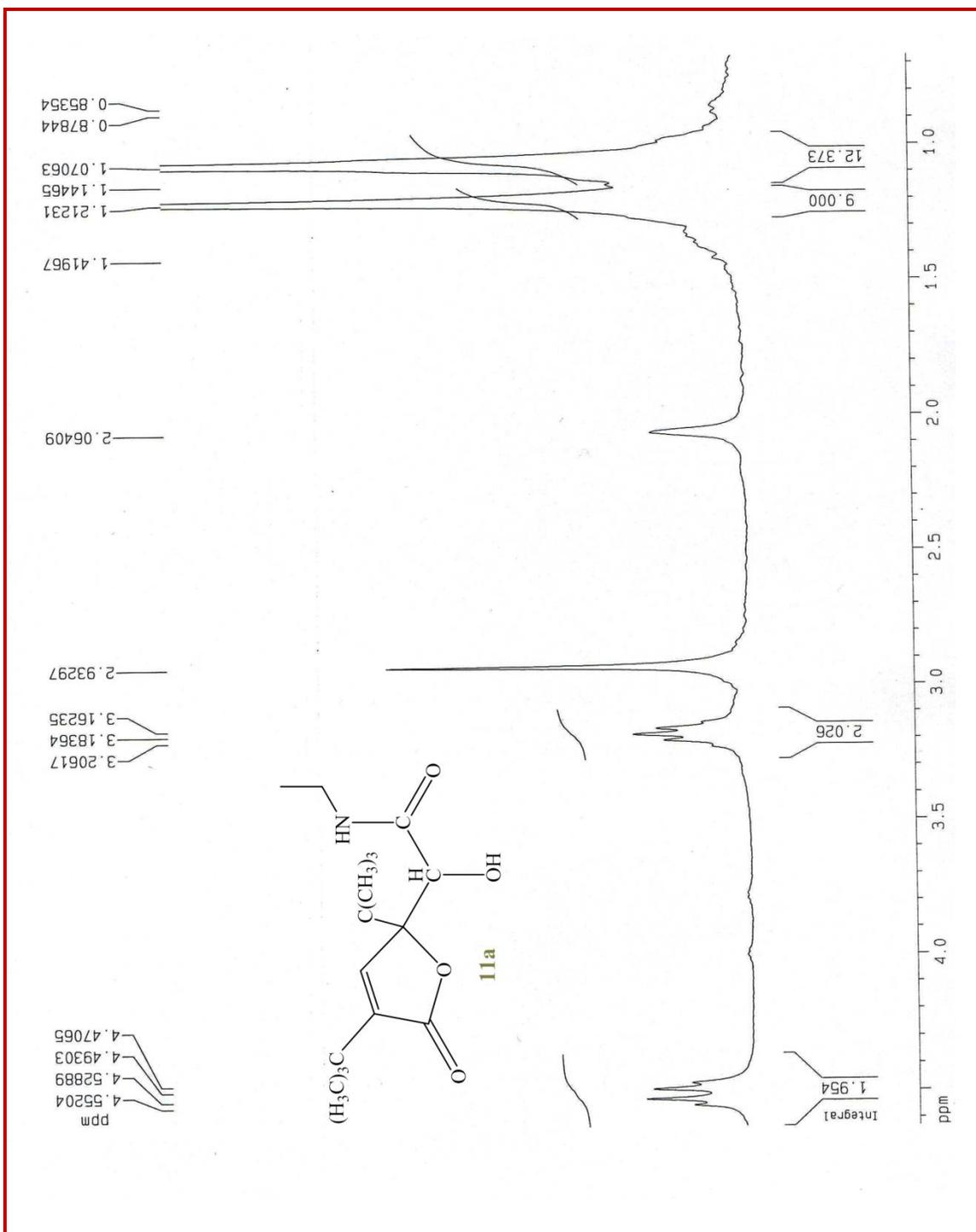
MS of 10a



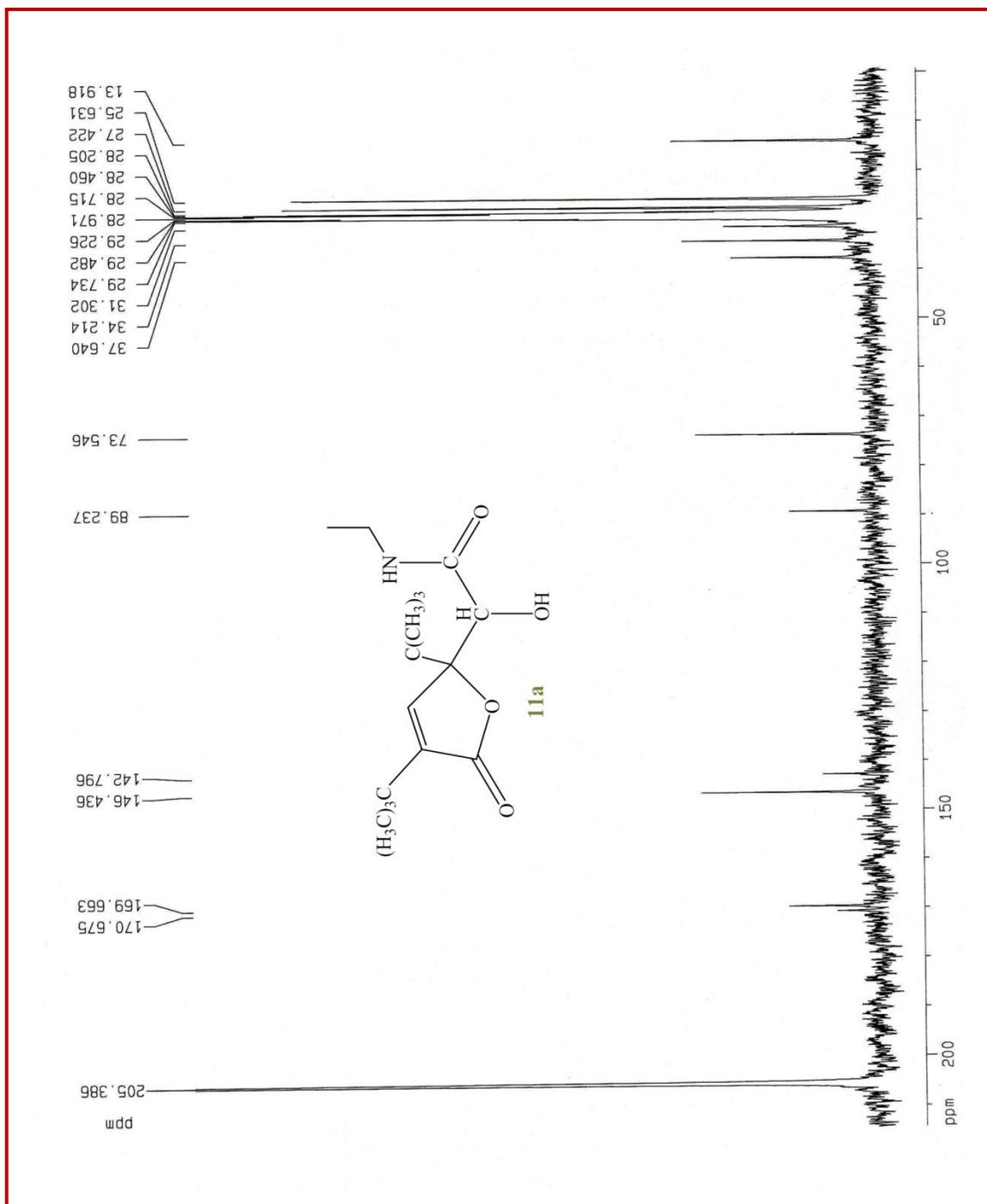
IR of 11a



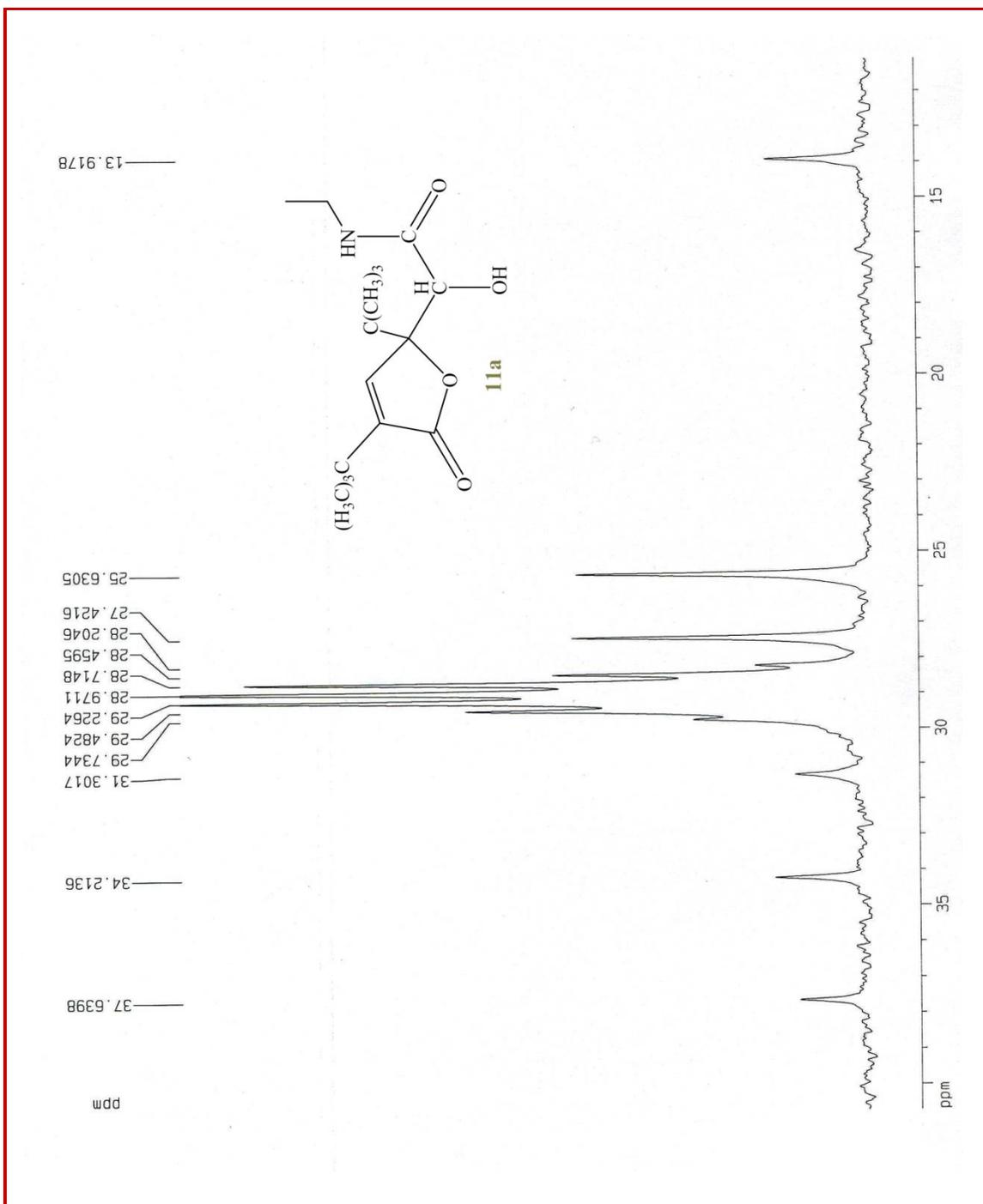
**<sup>1</sup>H NMR of 11a**



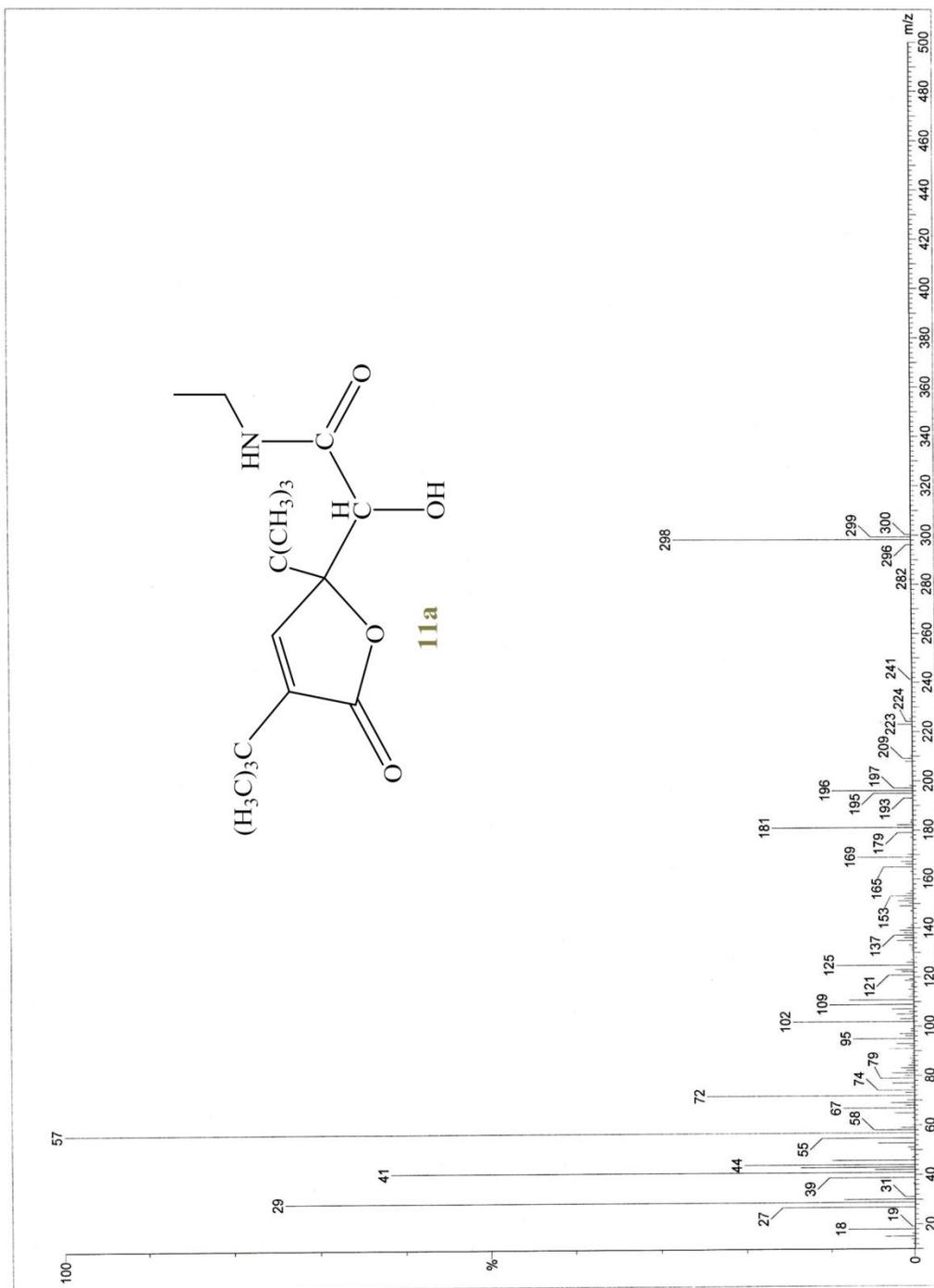
Expanded  $^1\text{H}$  NMR of 11a



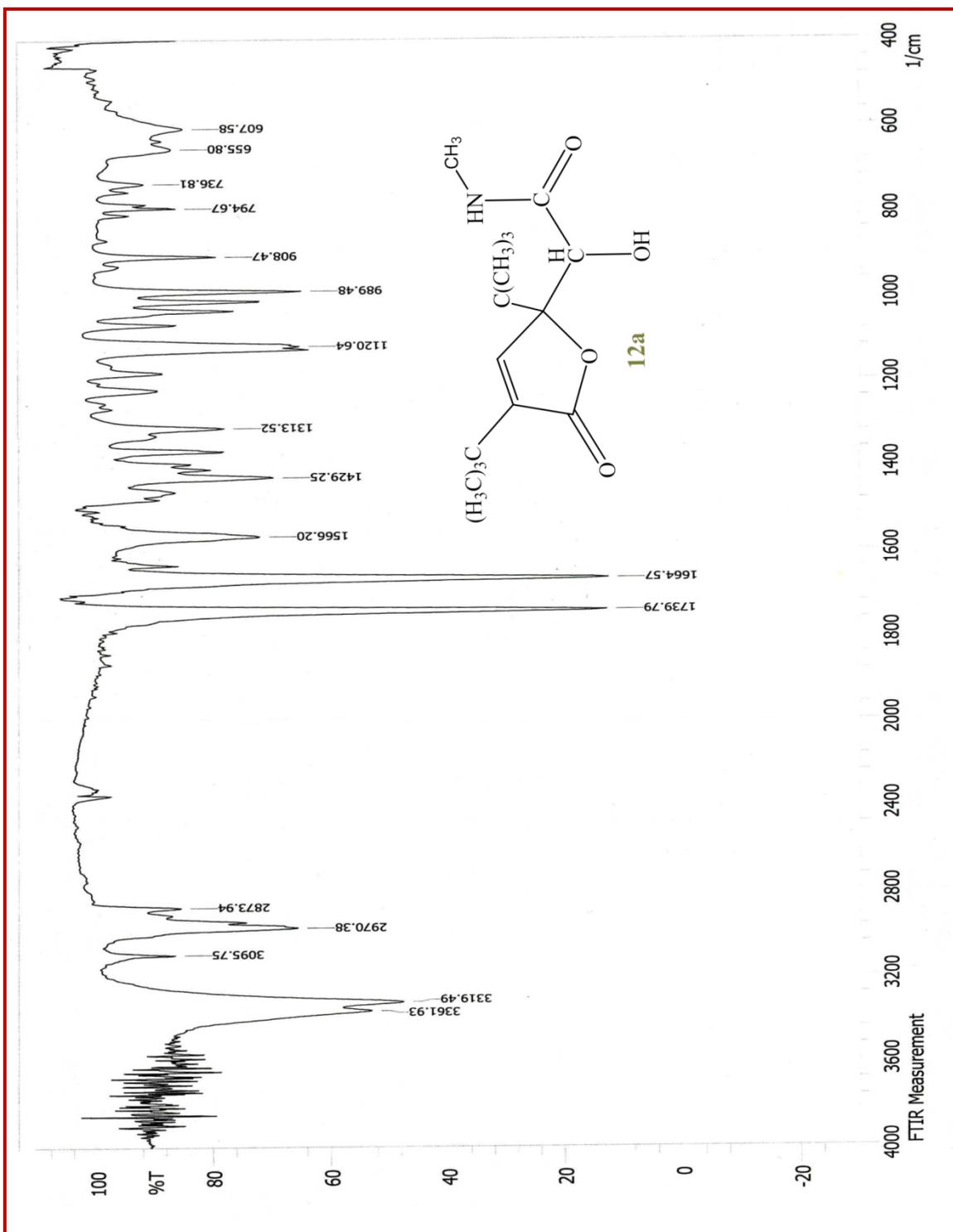
$^{13}\text{C}$  NMR of 11a



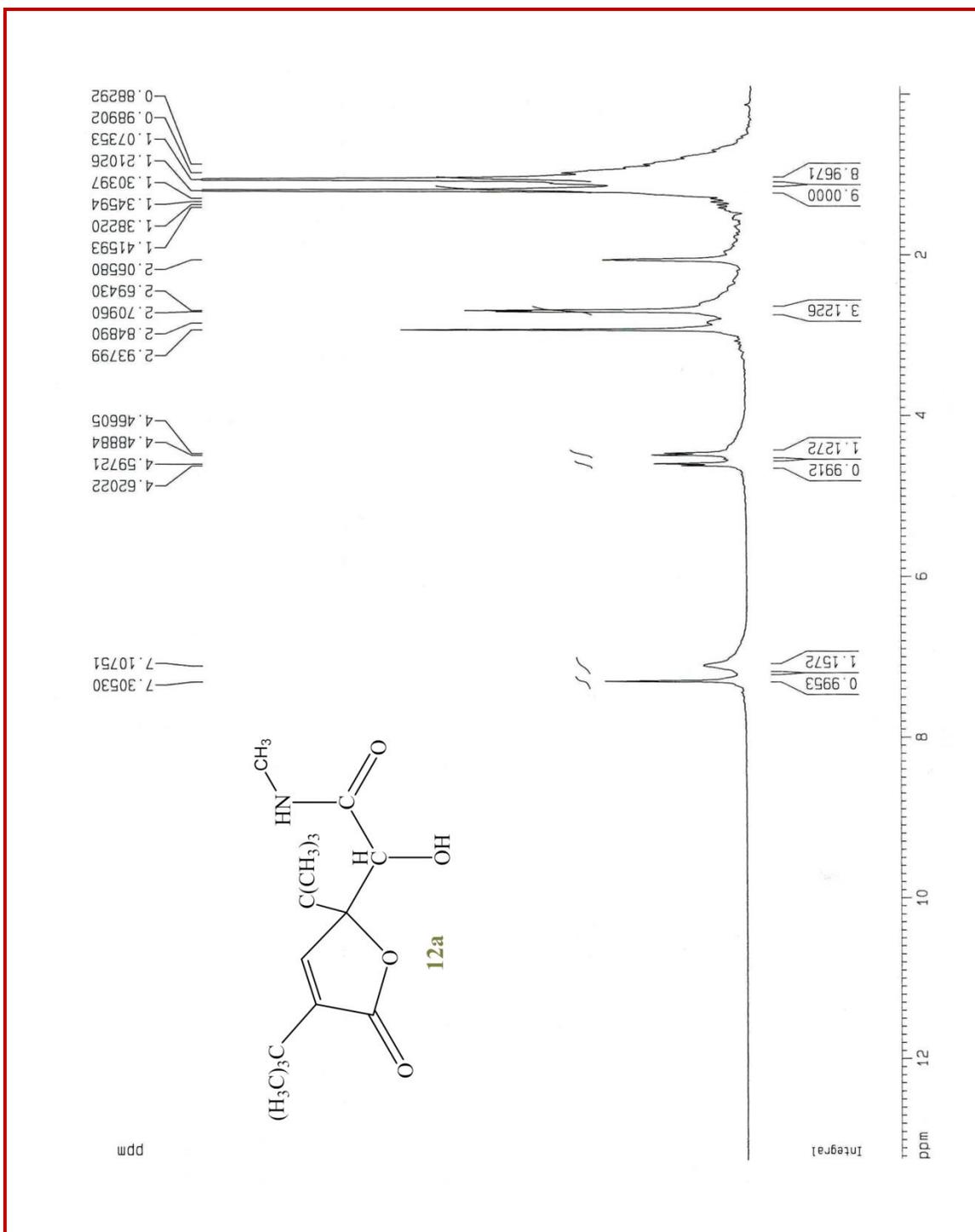
Expanded  $^{13}\text{C}$  NMR of 11a

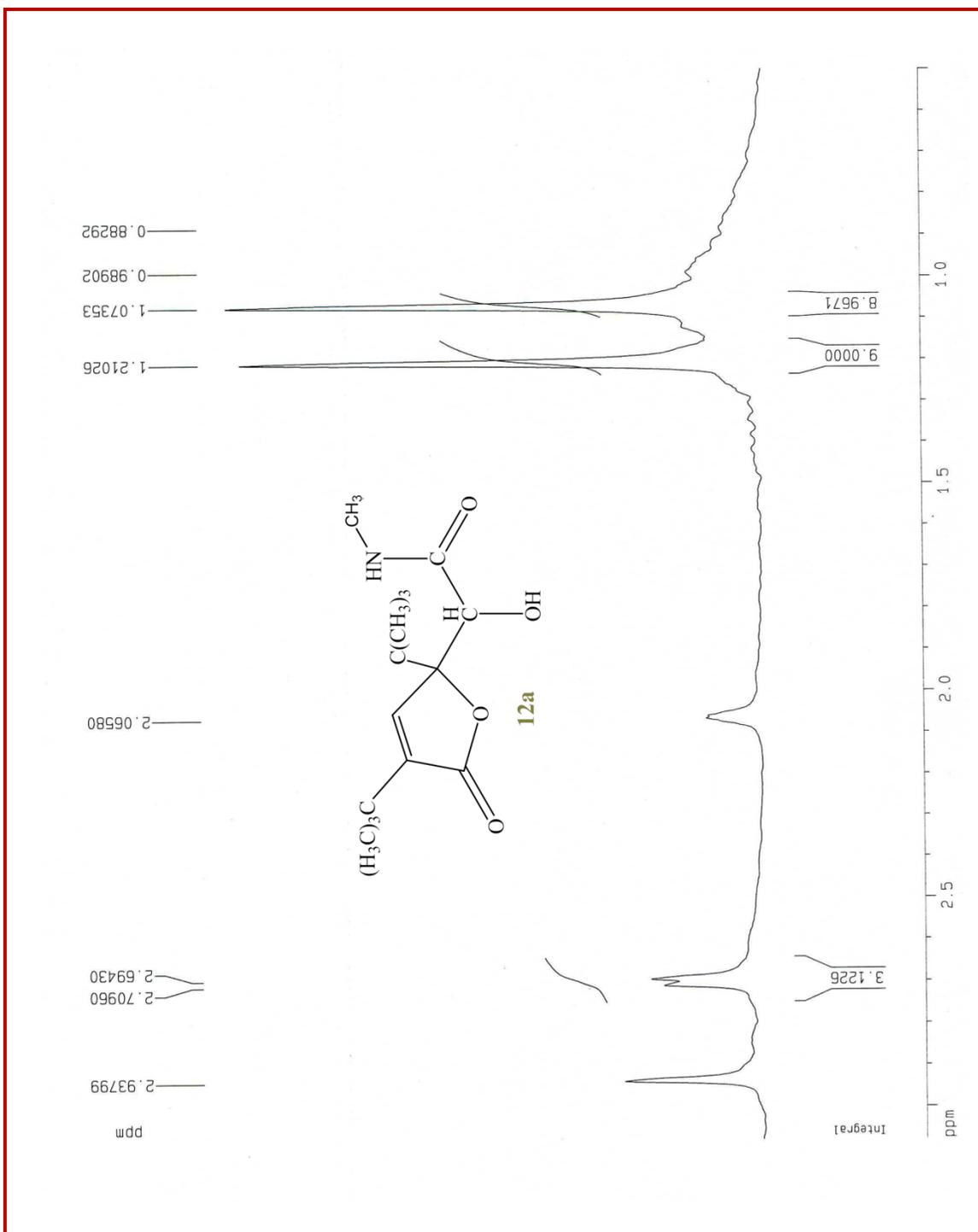


MS of 11a

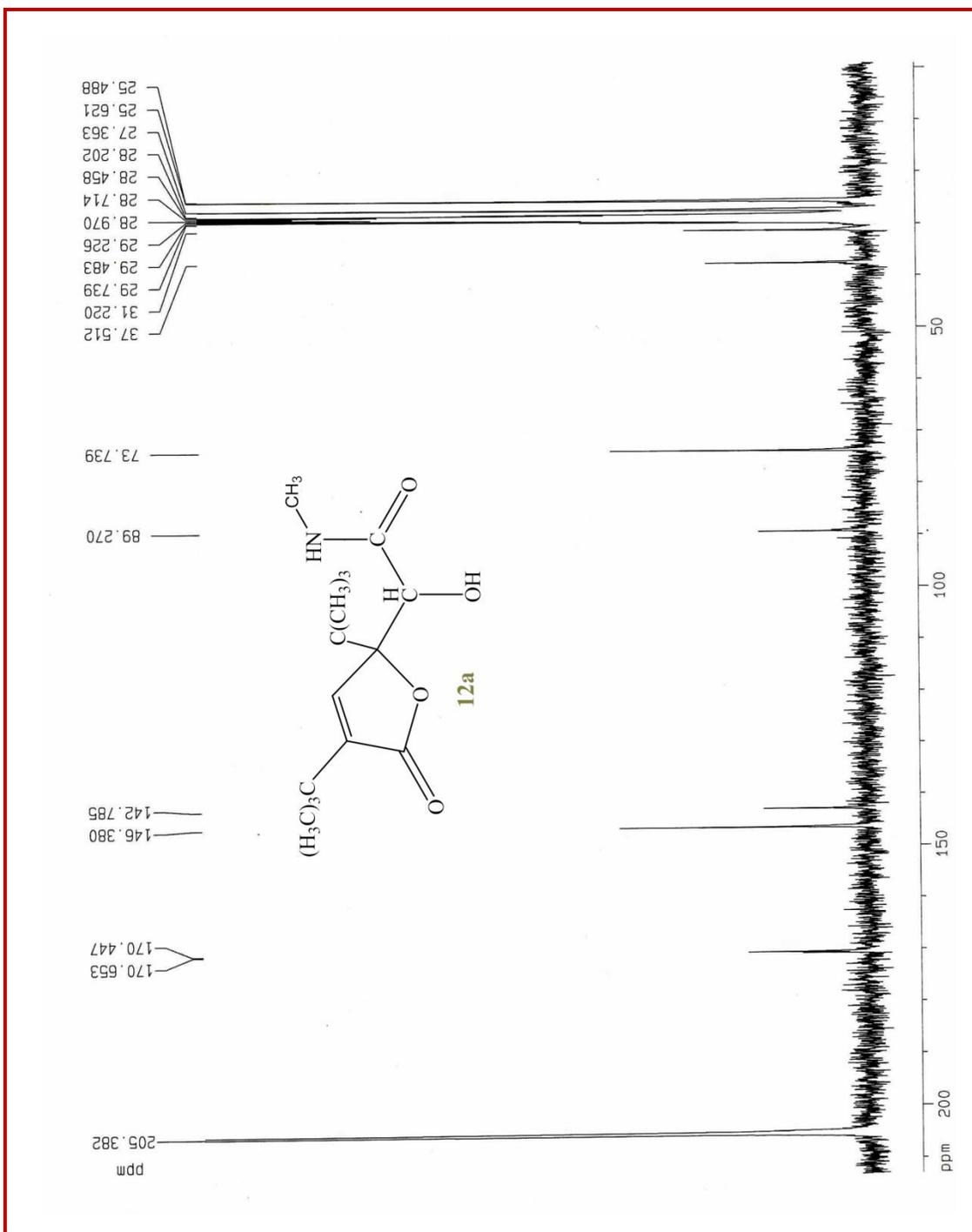


IR of 12a

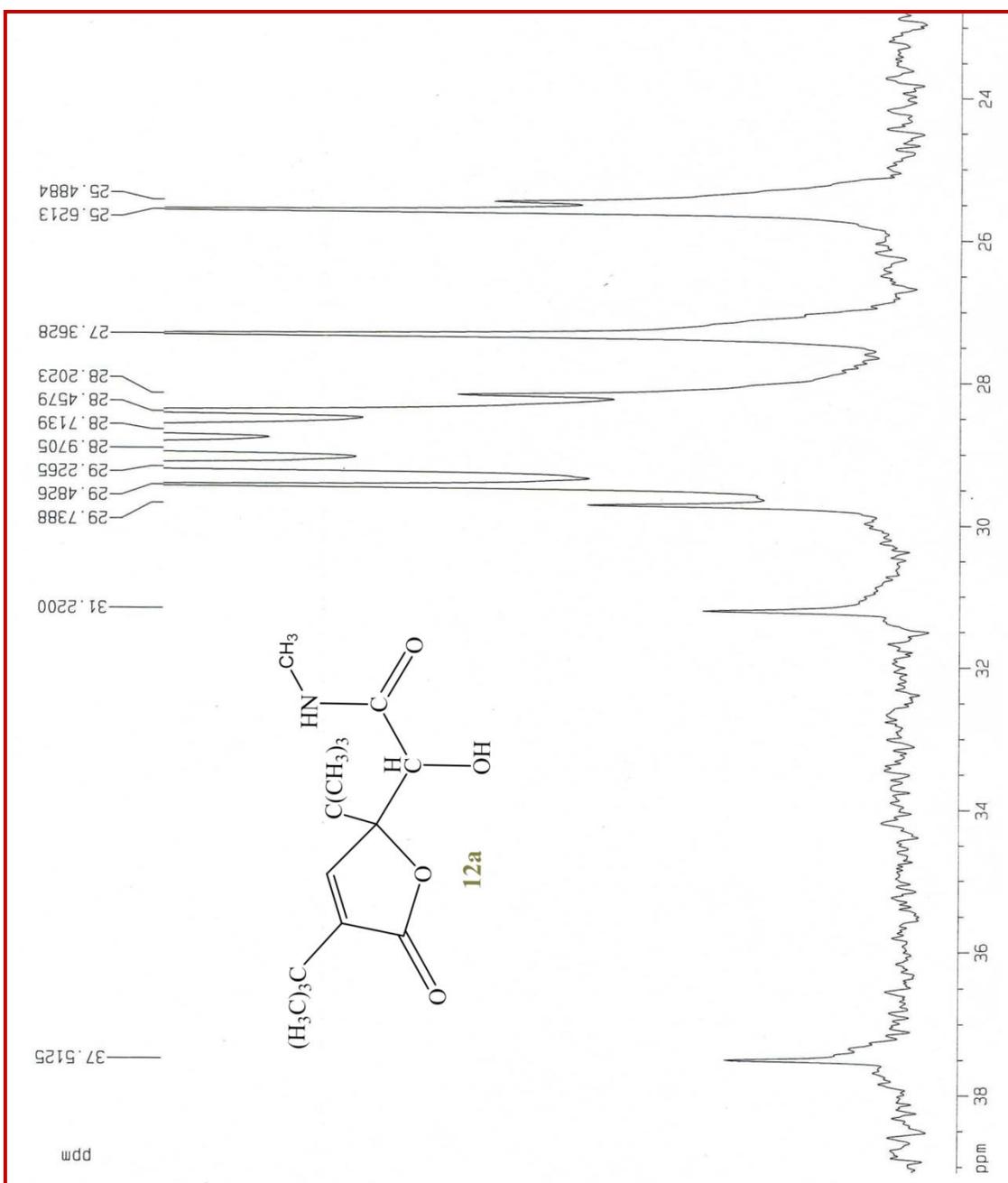




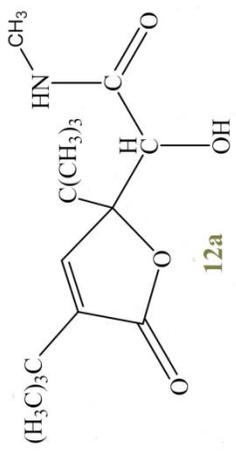
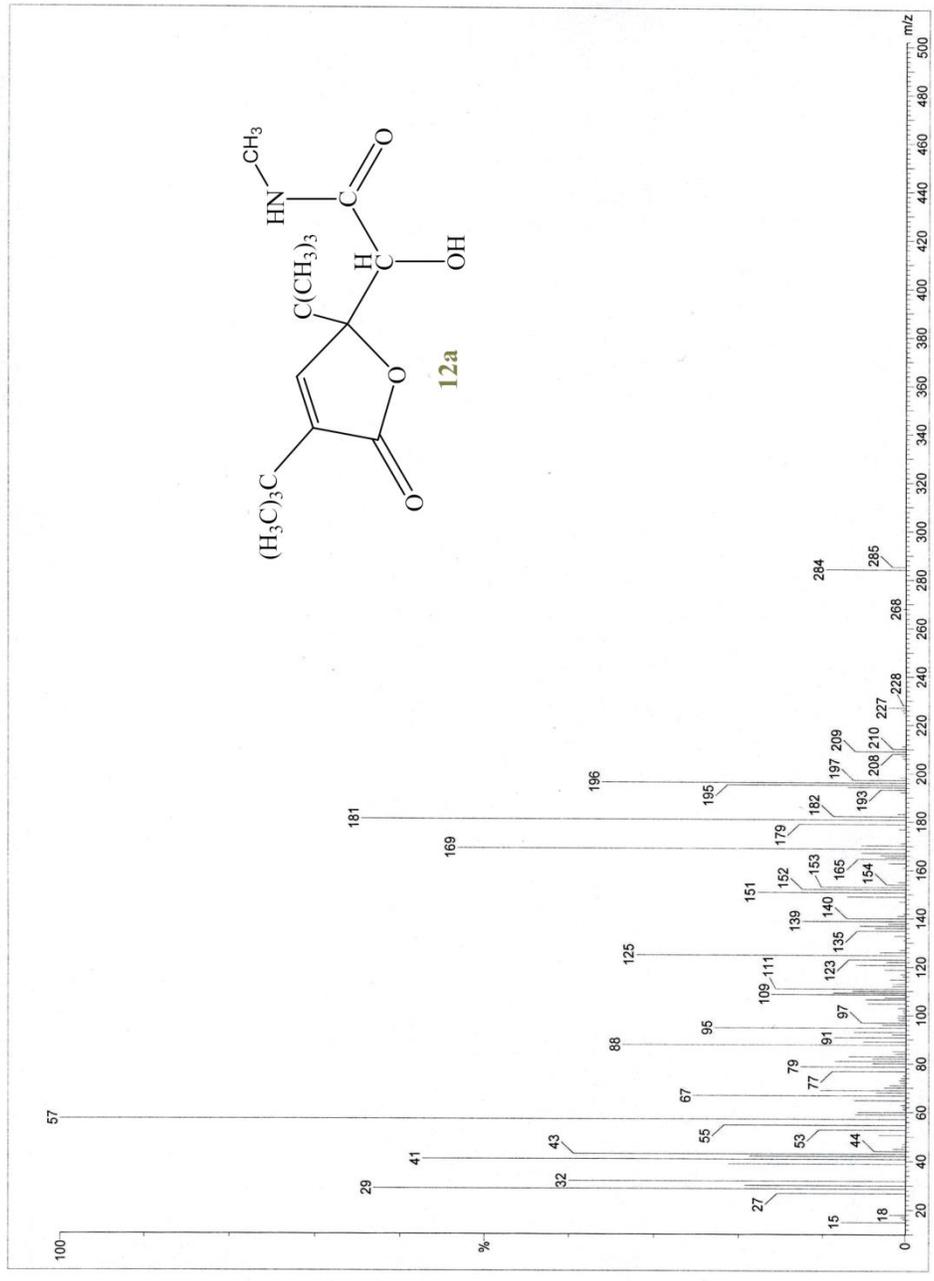
Expanded <sup>1</sup>H NMR of 12a



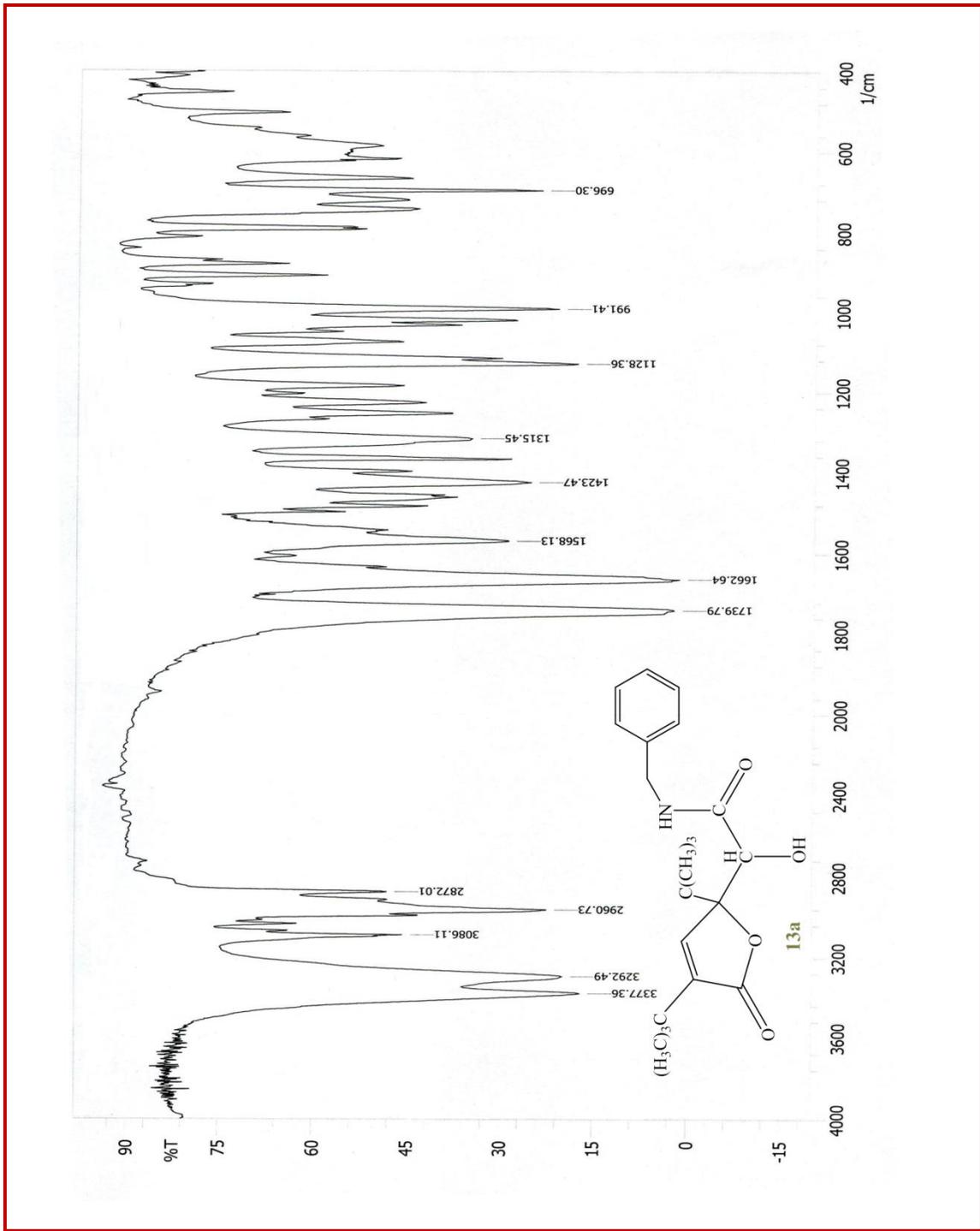
$^{13}\text{C}$  NMR of **12a**



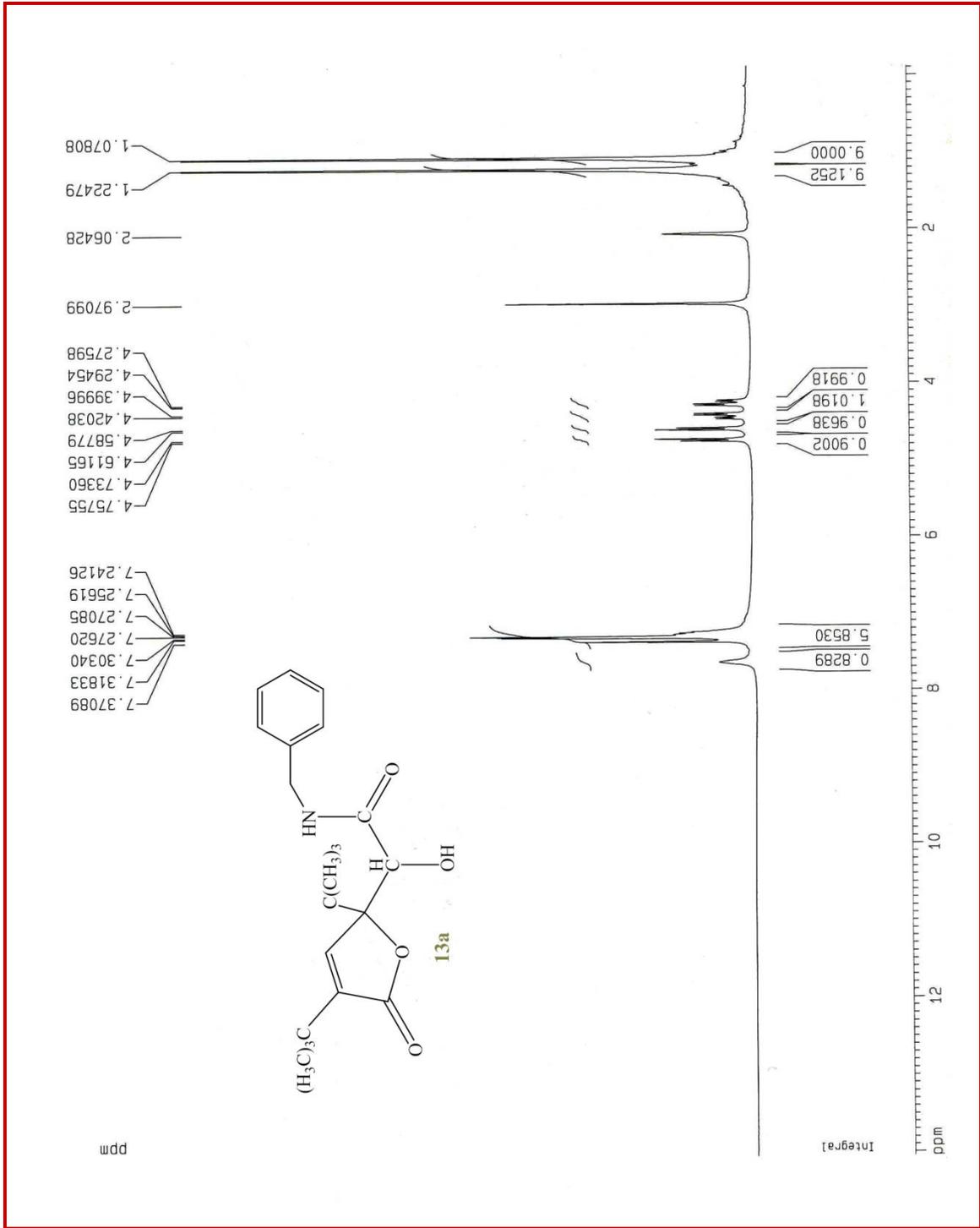
Expanded  $^{13}\text{C}$  NMR of **12a**



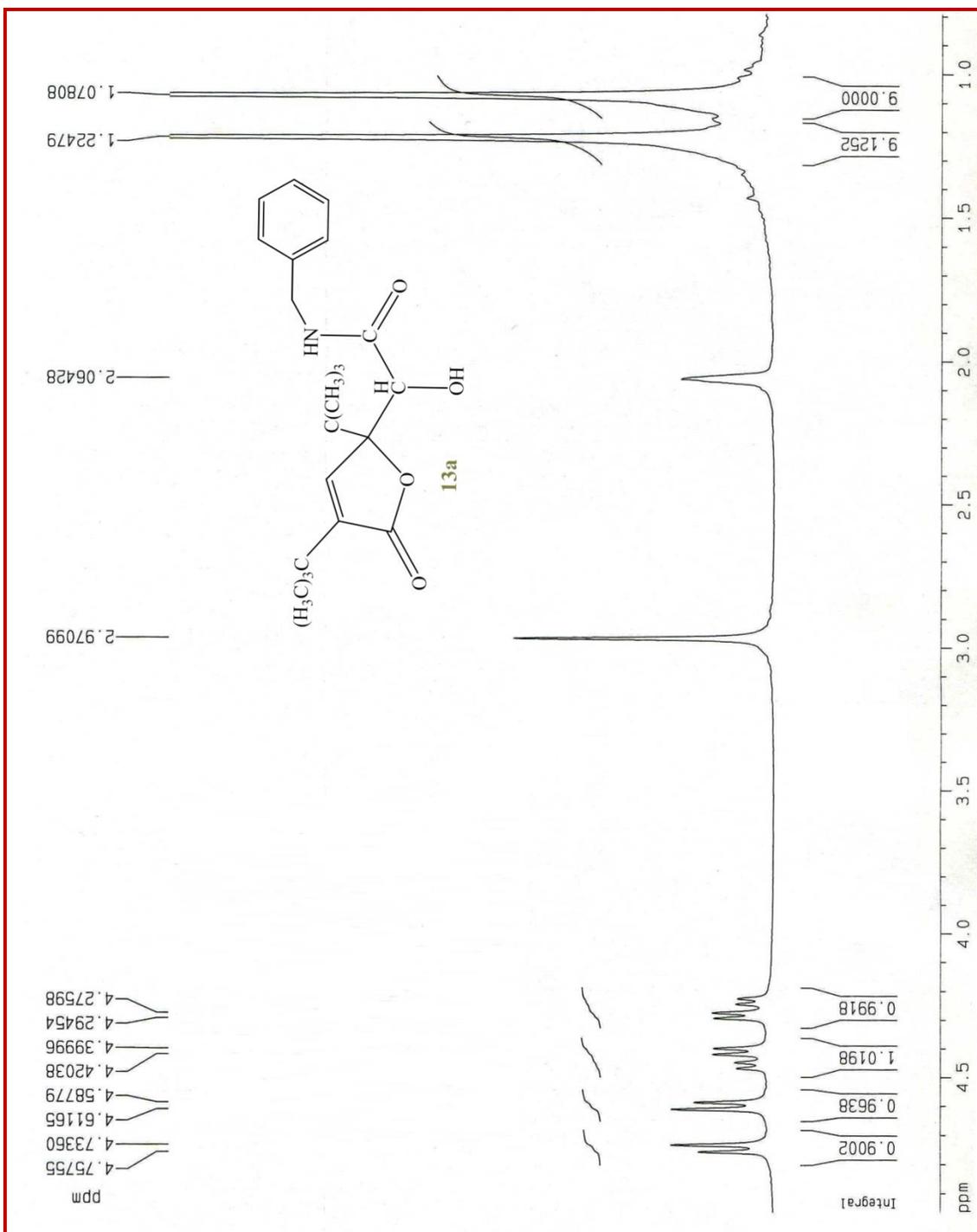
MS of 12a



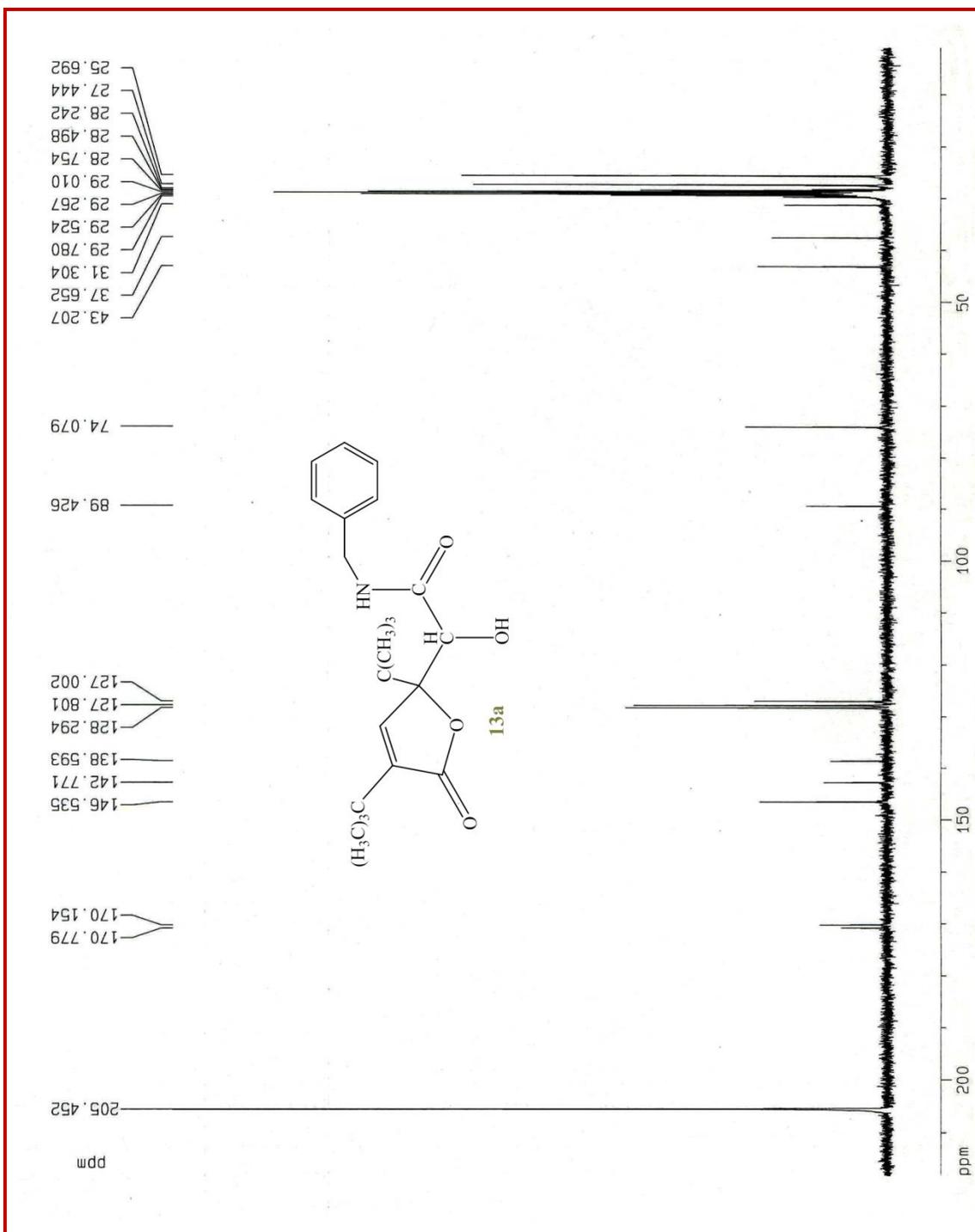
IR of 13a



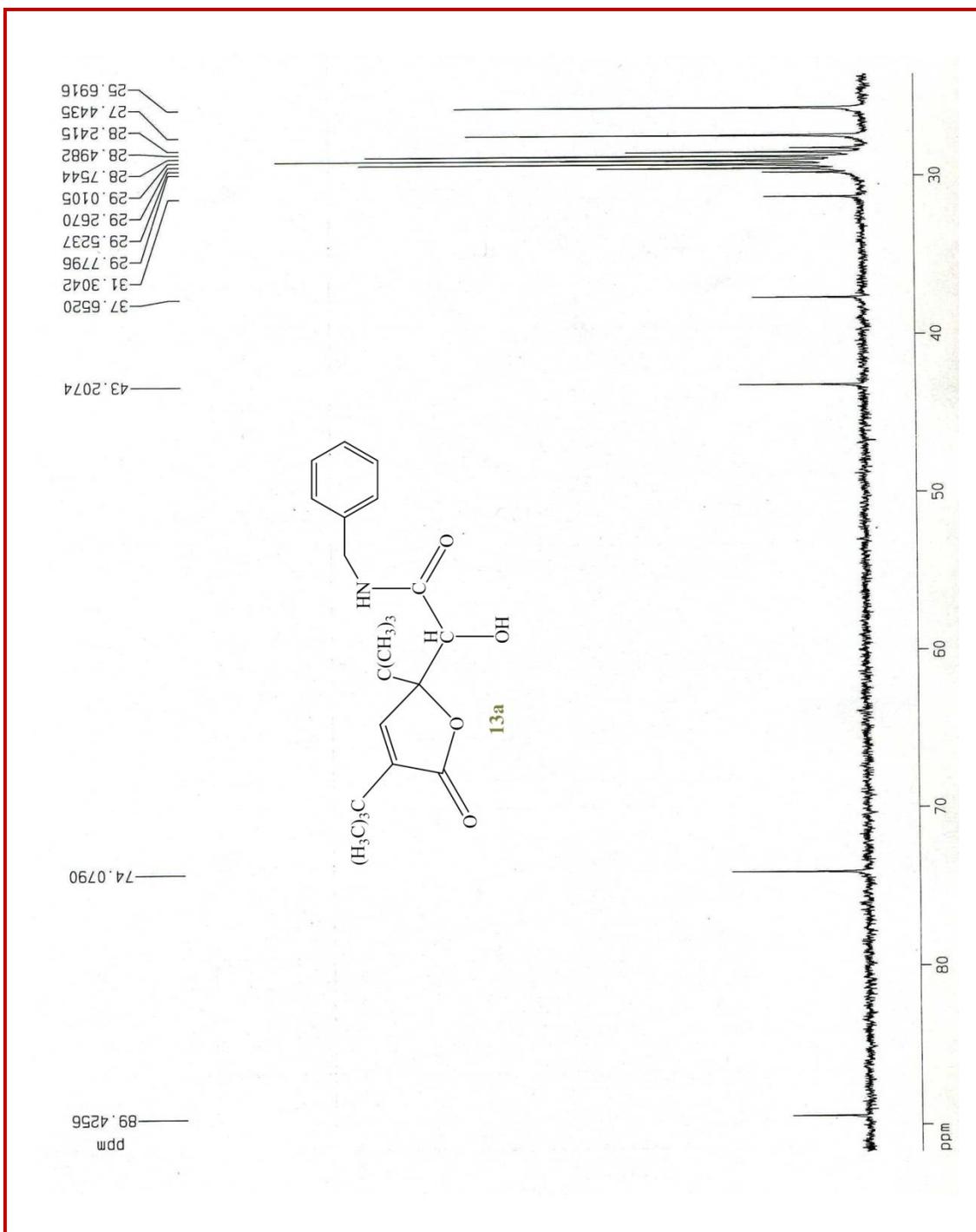
**<sup>1</sup>H NMR of 13a**



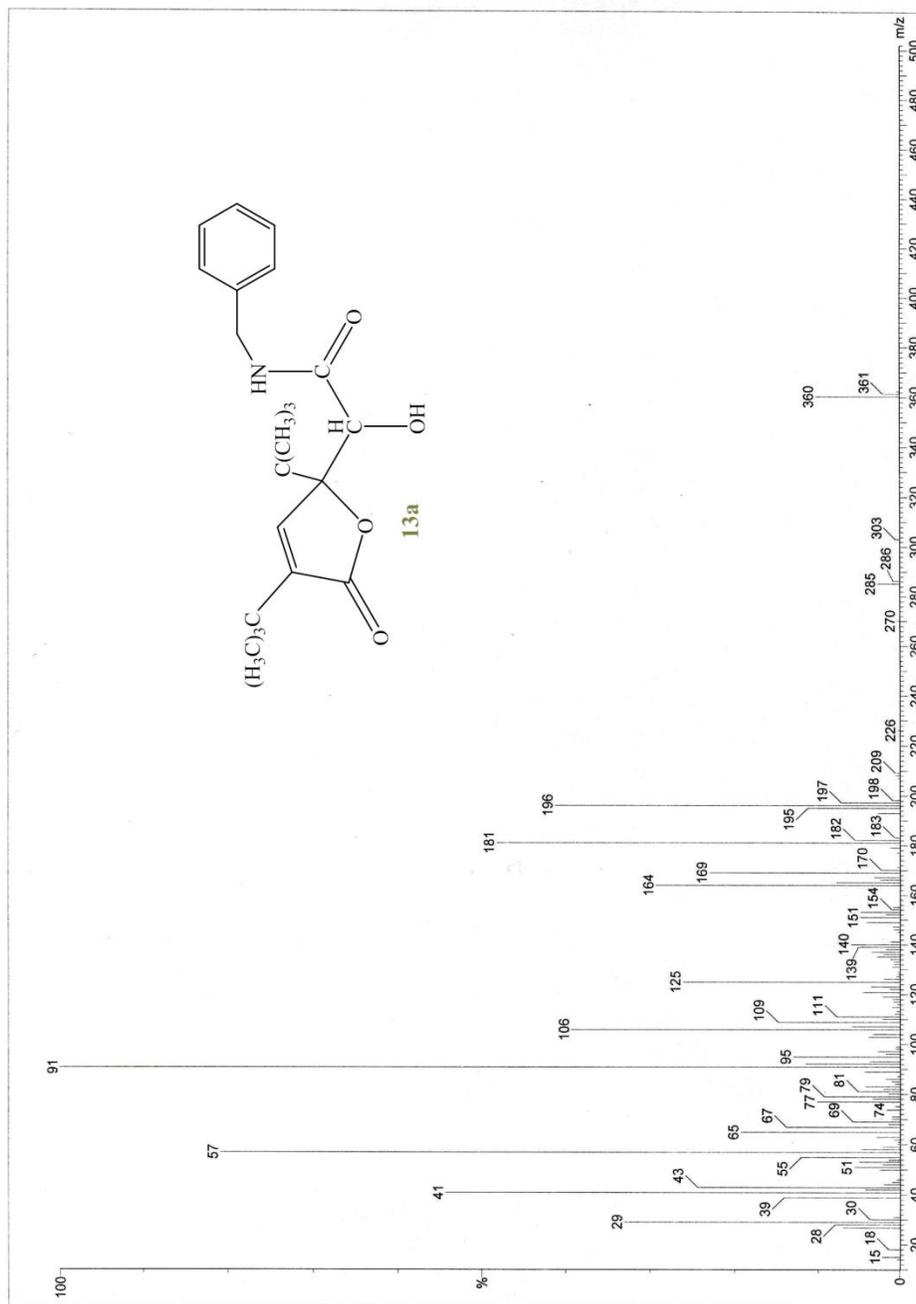
Expanded <sup>1</sup>H NMR of 13a



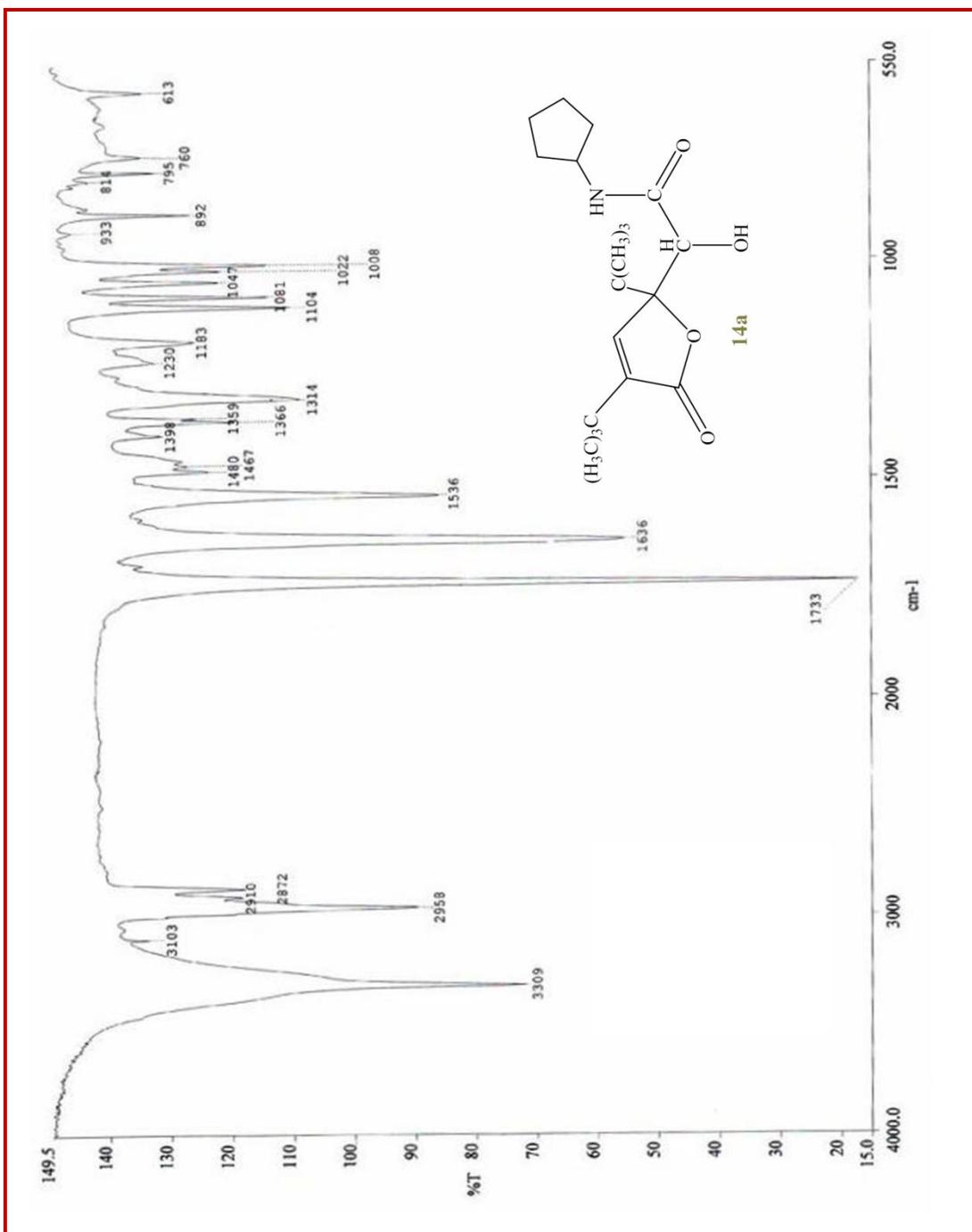
<sup>13</sup>C NMR of 13a



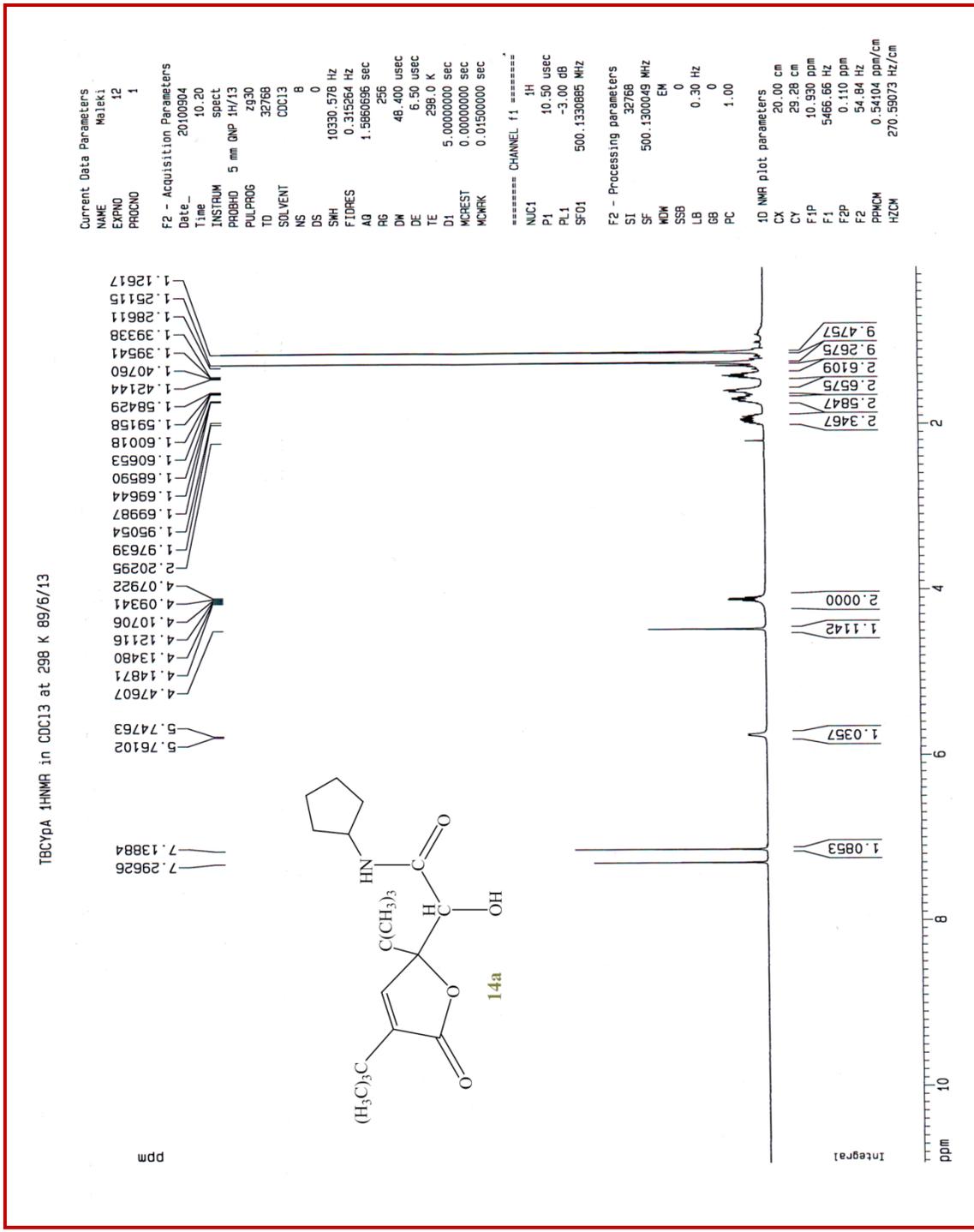
Expanded  $^{13}\text{C}$  NMR of 13a



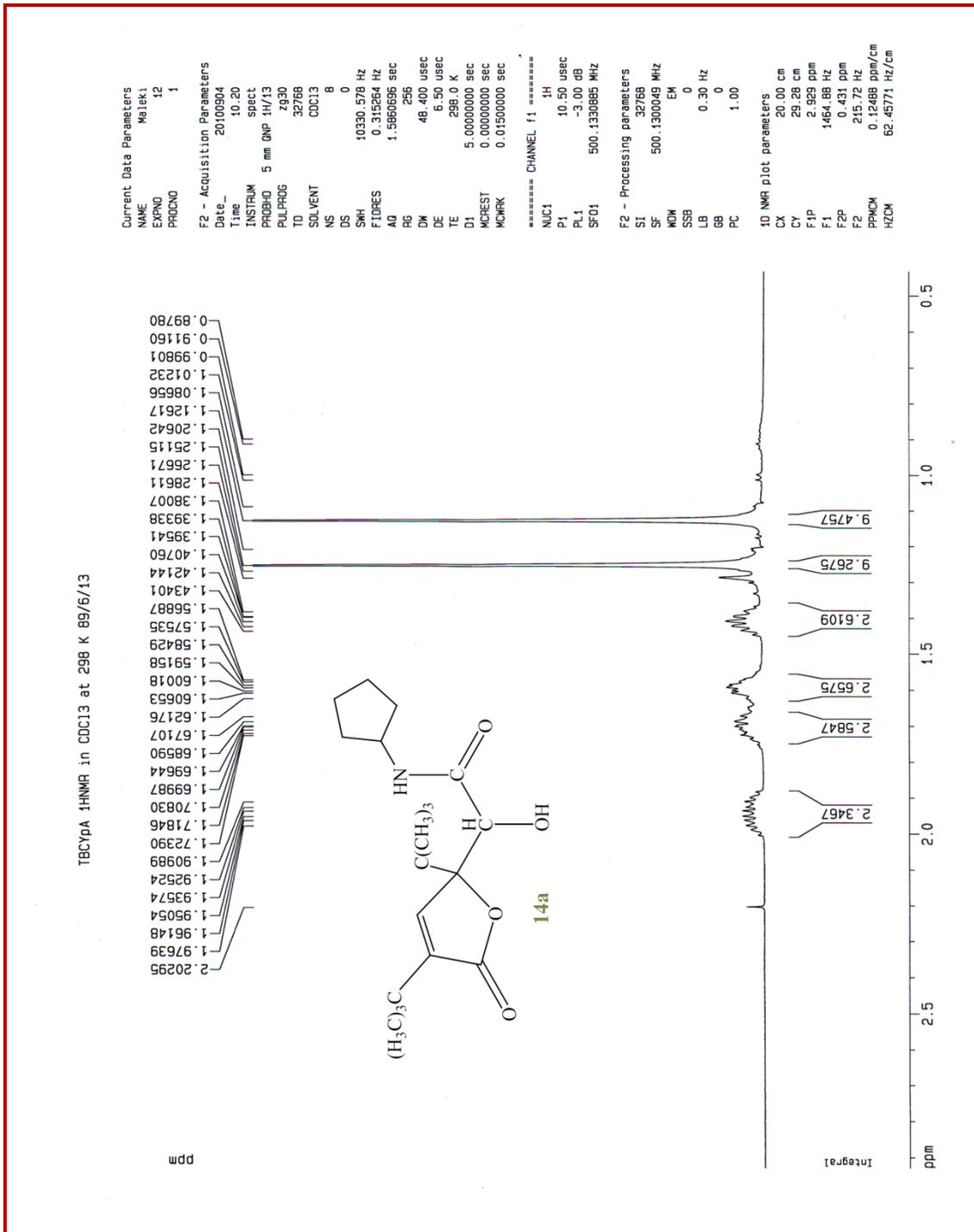
MS of 13a



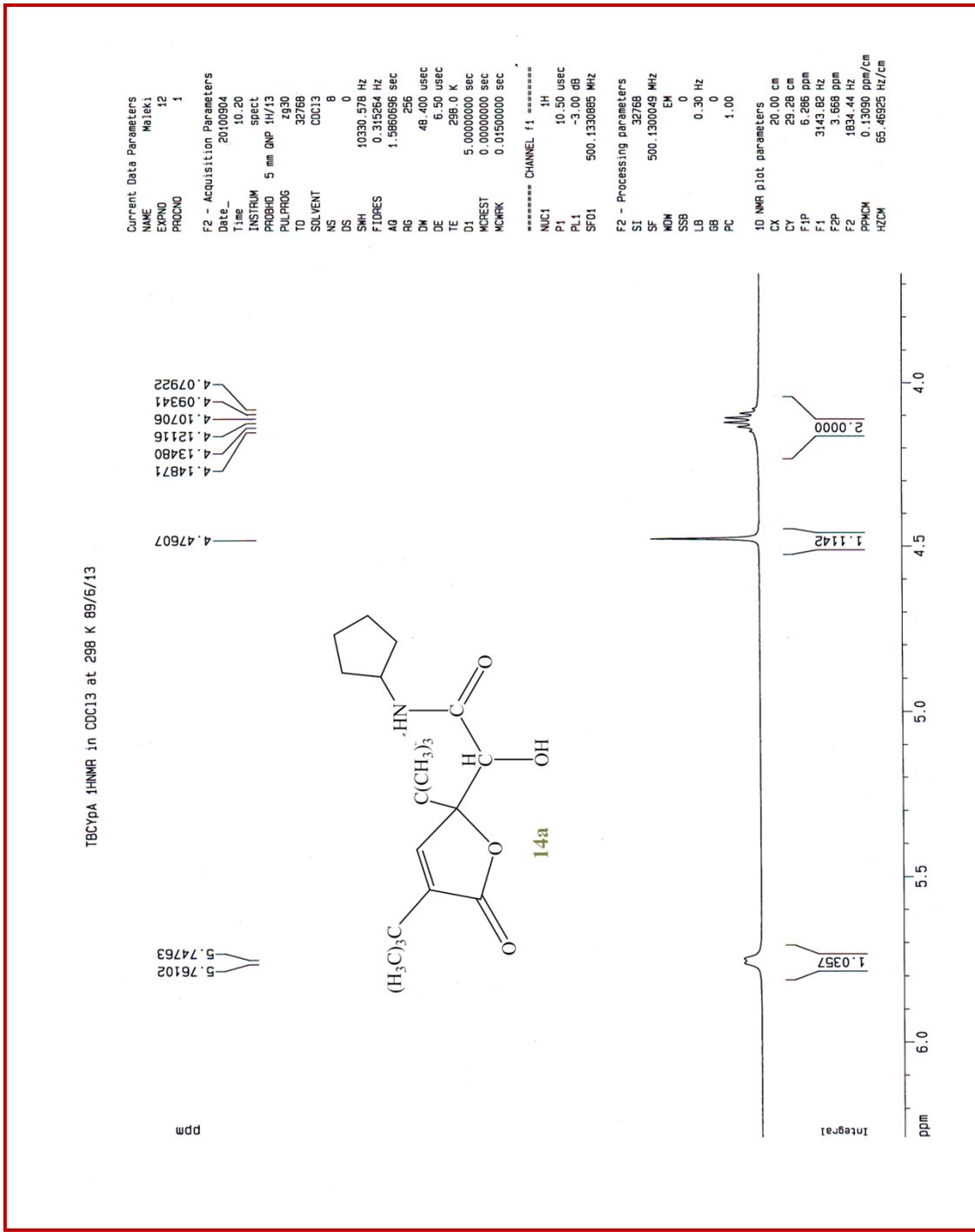
IR of 14a



**<sup>1</sup>H NMR of 14a**

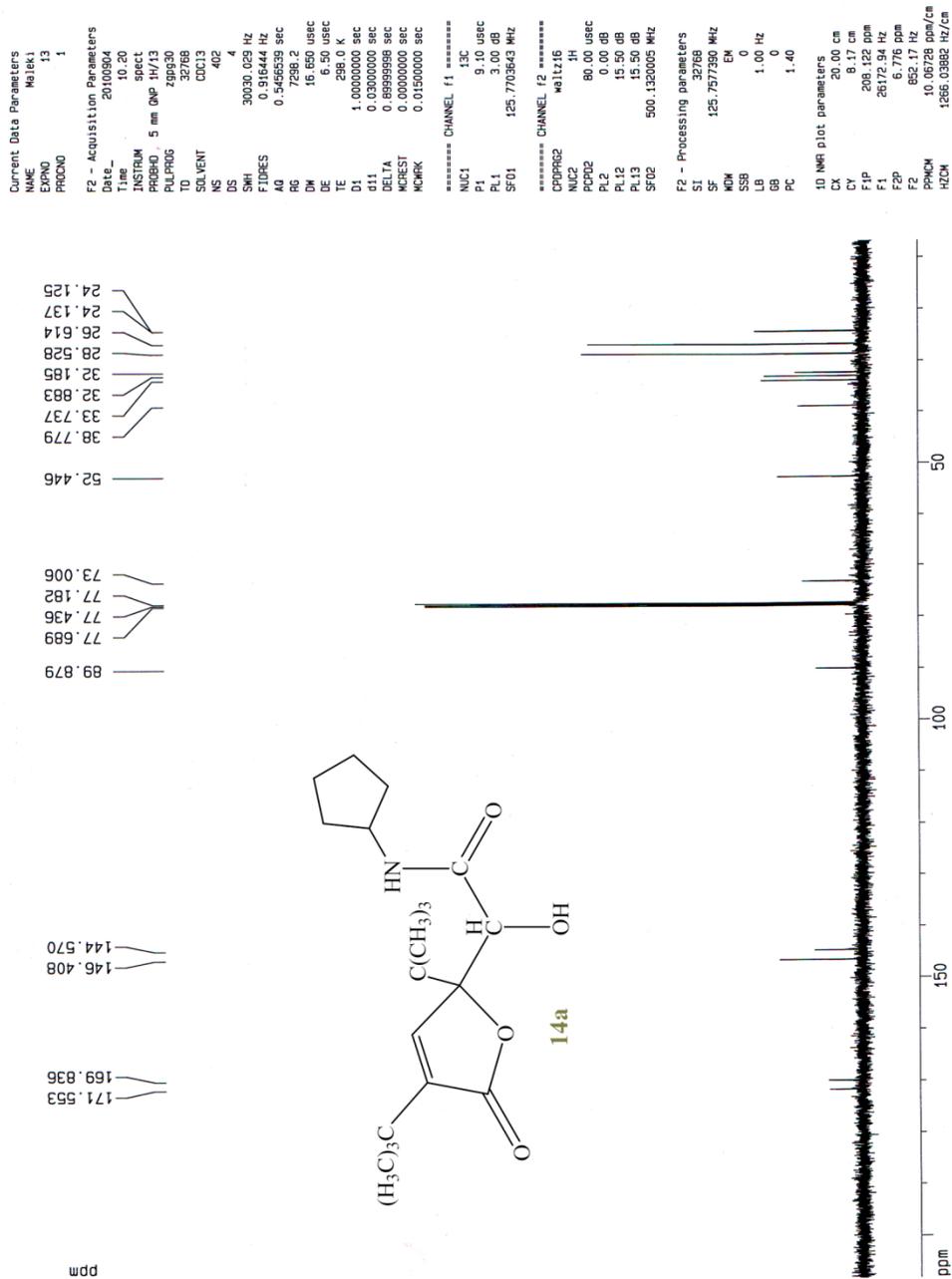


Expanded <sup>1</sup>H NMR of 14a

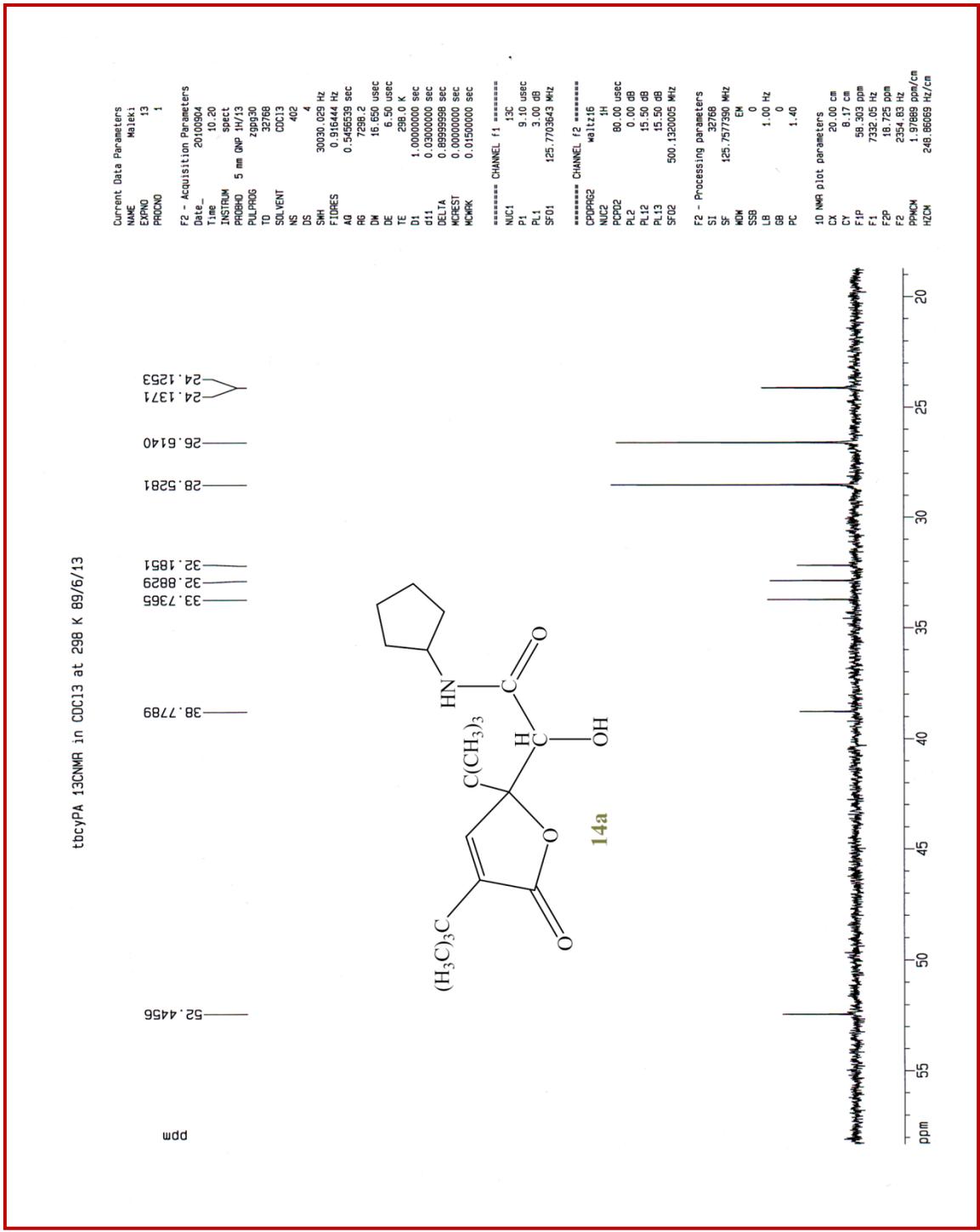


Expanded <sup>1</sup>H NMR of 14a

tbcyPA 13CNMR in CDCl3 at 298 K 89/6/13

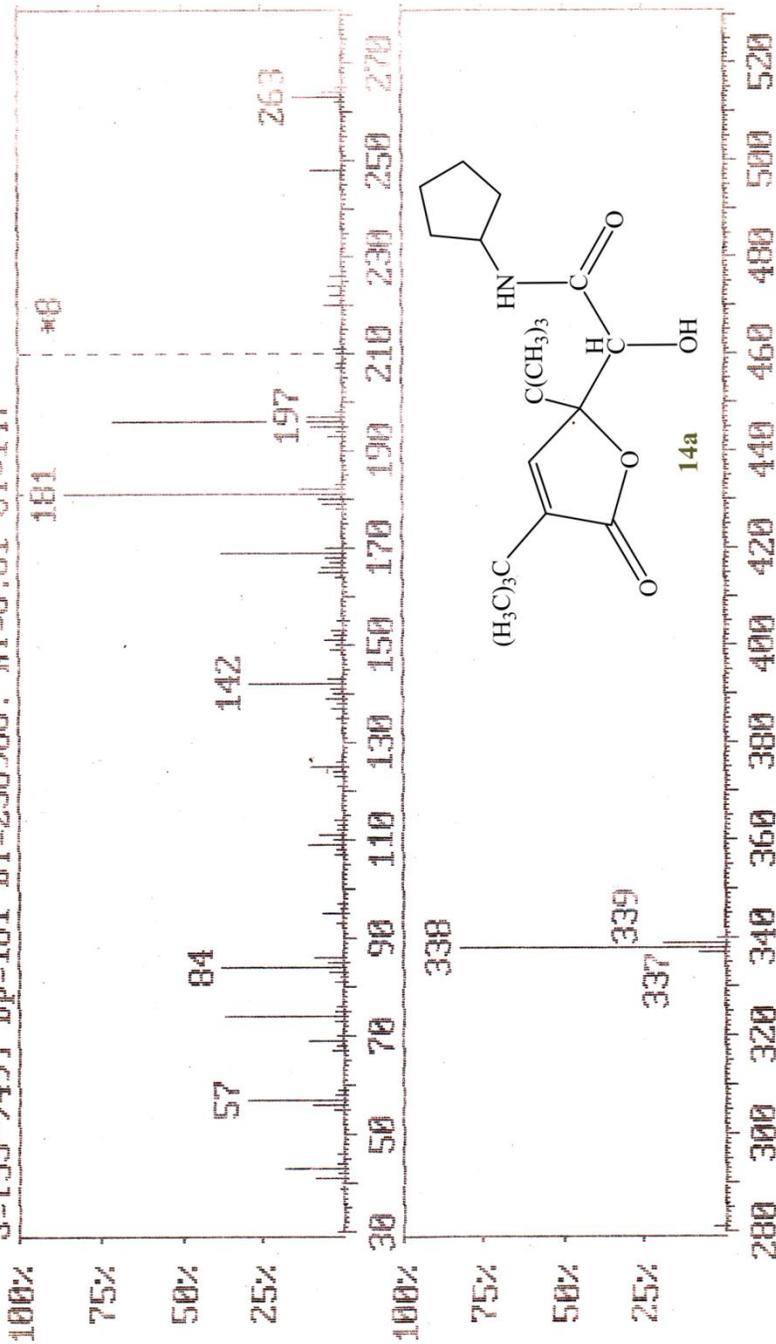


### <sup>13</sup>C NMR of 14a



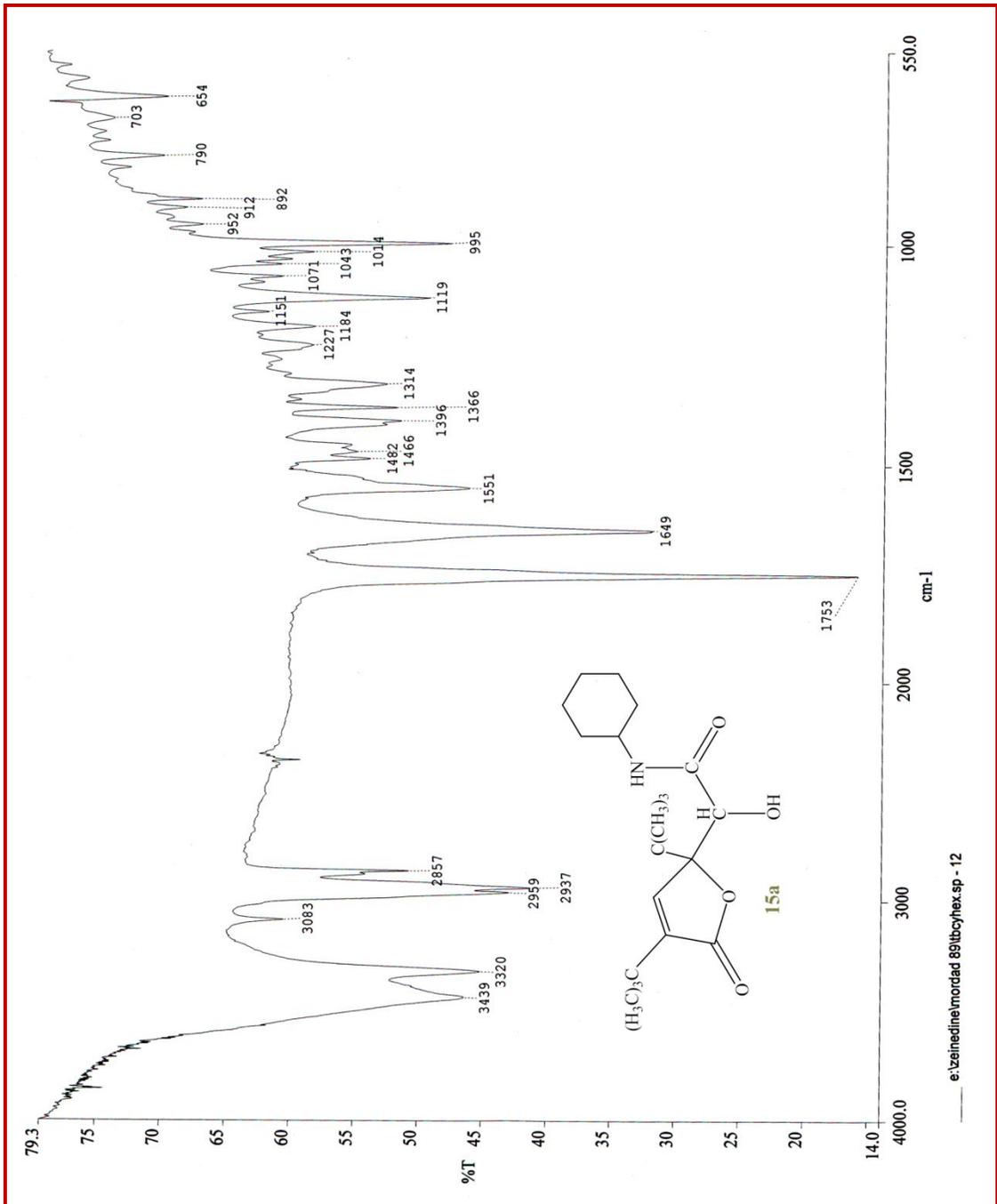
Expanded <sup>13</sup>C NMR of 14a

DI/ZEINOLDINI-TBCPAN/89.03.30  
 File : DI\_83.X10 Date 9/ 9/10 Time 02:12:16  
 S=[35->49] Bp=181 Bi=238960. RT=0.81 CT=147



SB=30 SE=340 DB=30 DE=510 N=0 Z=2 T=0.0 Fact[210->530] \*8  
 S List > S=[35->49] B=0 Pos=3 Tot=3

MS of 14a



FTIR of 15a

tucyhex <sup>1</sup>HNMR in CDCl<sub>3</sub> at 298 K 89/5/13

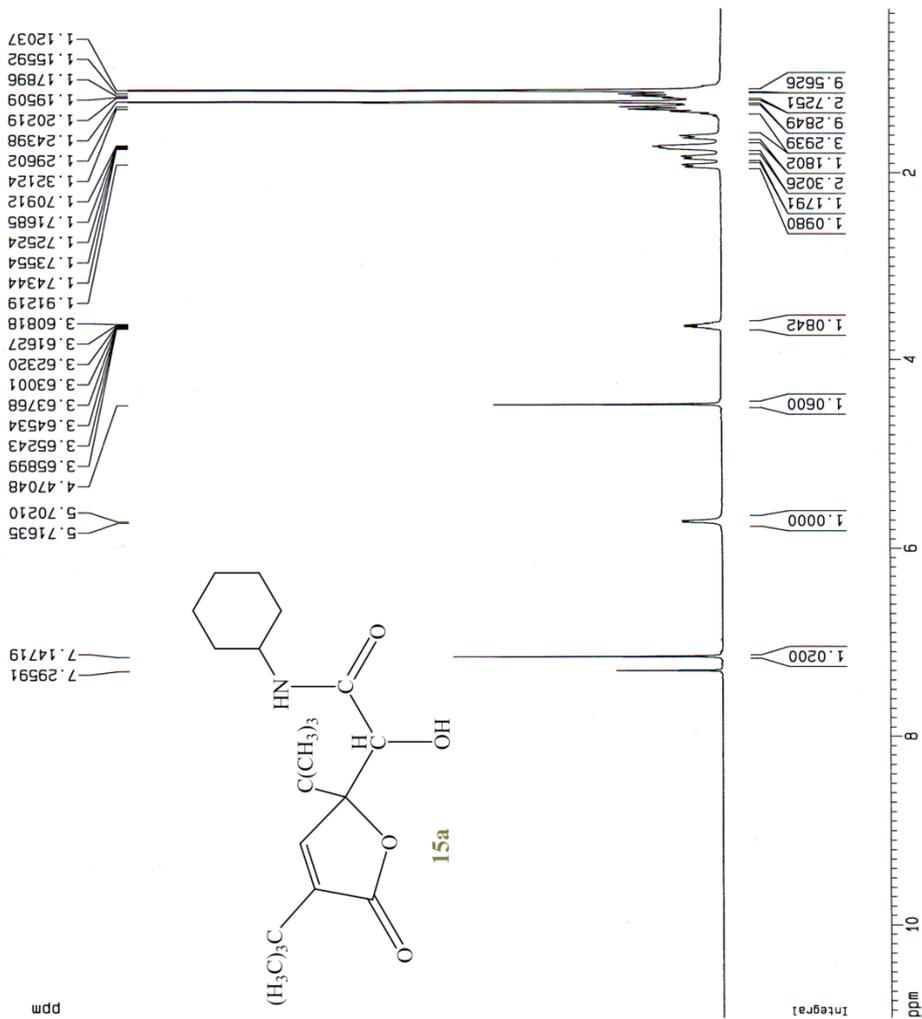
Current Data Parameters  
NAME Melik1  
EXPNO 10  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20100904  
Time 9.58  
INSTRUM spect  
PROBHD 5 mm QNP 1H/13  
PULPROG zg30  
TD 32768  
SOLVENT CDCl<sub>3</sub>  
NS 8  
DS 0  
SWH 10330.578 Hz  
FIDRES 0.315264 Hz  
AQ 1.3960696 sec  
RG 181  
DM 48.400 usec  
DE 6.50 usec  
TE 298.0 K  
D1 5.0000000 sec  
ICREST 0.0000000 sec  
MCWK 0.0150000 sec

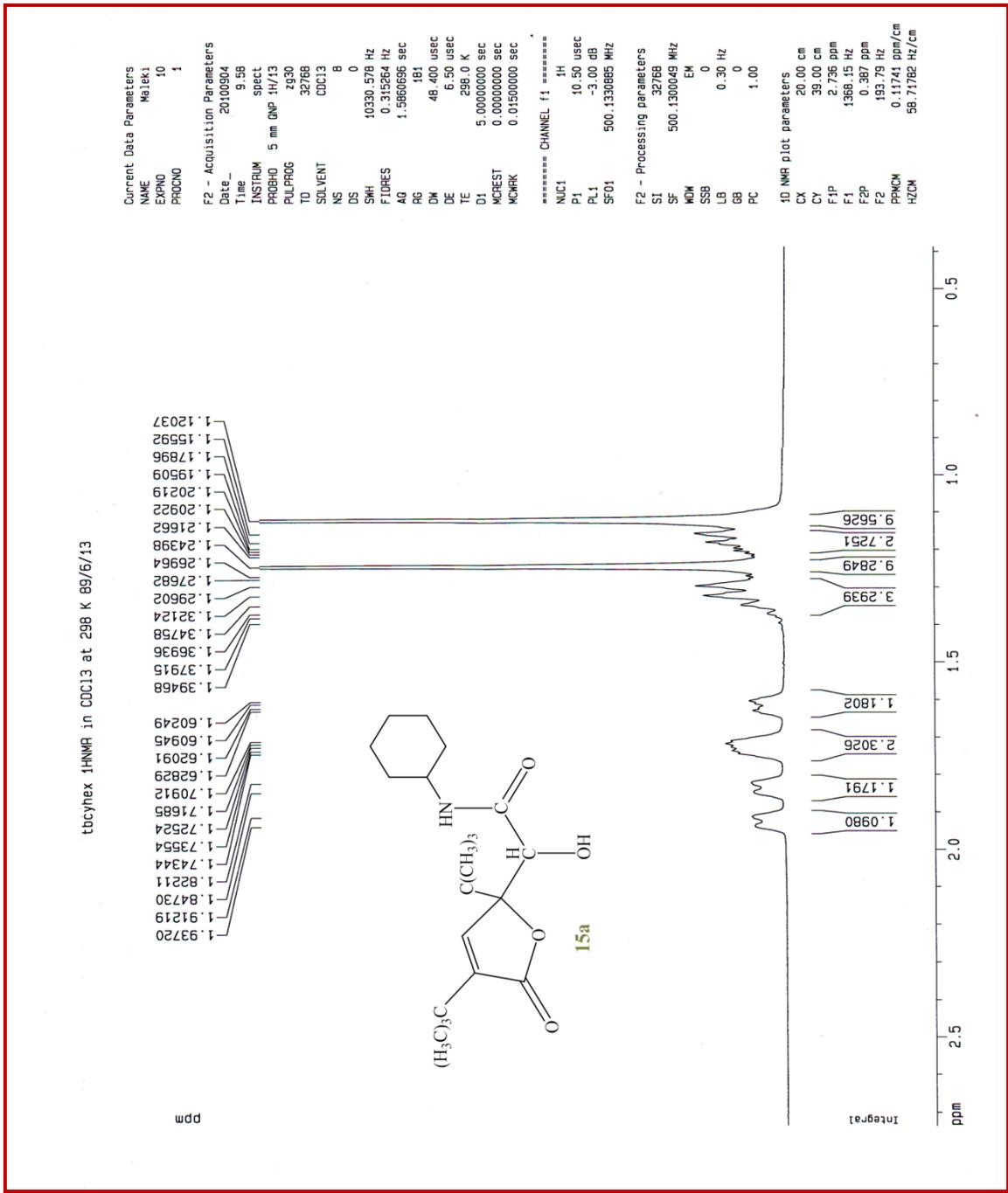
\*\*\*\*\* CHANNEL f1 \*\*\*\*\*  
NUC1 1H  
P1 10.50 usec  
PL1 -3.00 dB  
SFO1 500.1330985 MHz

F2 - Processing parameters  
SI 32768  
SF 500.1300049 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00

1D NMR plot parameters  
CX 20.00 cm  
CY 39.00 cm  
F1P 10.977 ppm  
F2P 0.250 ppm  
F2 125.28 Hz  
PPHM 0.53635 ppm/cm  
HZCM 268.24286 Hz/cm

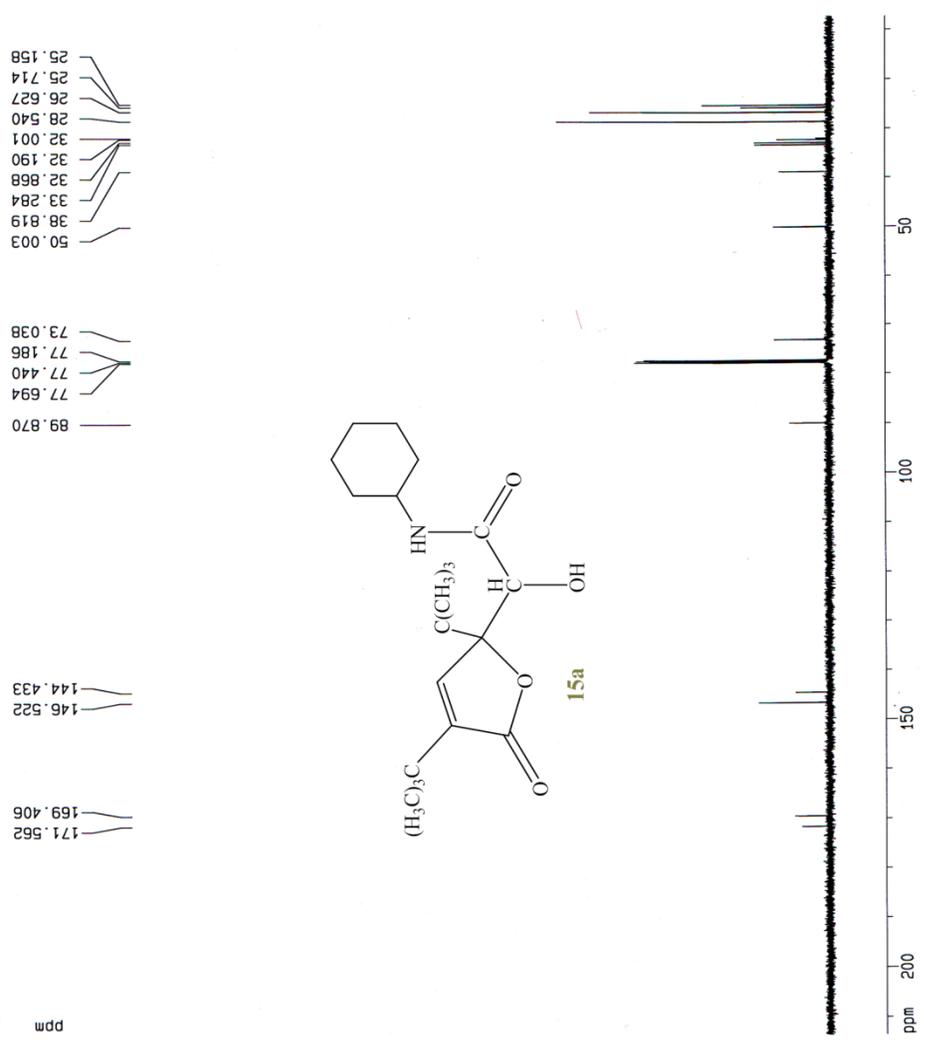


<sup>1</sup>HNMR of 15a



Expanded <sup>1</sup>H NMR of 15a

tbcyhex-13CNMR in CDCl3 at 298 K 89/6/13



```

Current Data Parameters
NAME      Malek1
EXPNO    11
PROCNO   1

F2 - Acquisition Parameters
Date_    20100904
Time     9.58
INSTRUM spect
PROBHD   5 mm QNP 1H/13
PULPROG zgpg30
TD        32768
SOLVENT  CDCl3
NS        276
DS        4
SWH      30030.029 Hz
FIDRES   0.046474 Hz
AQ        0.548626 sec
RG        61.952
DM        16.650 usec
DE        6.50 usec
TE        298.0 K
D1        1.0000000 sec
d11       0.0300000 sec
DELTA    0.8898988 sec
ICREST   0.0000000 sec
MCWK     0.0150000 sec

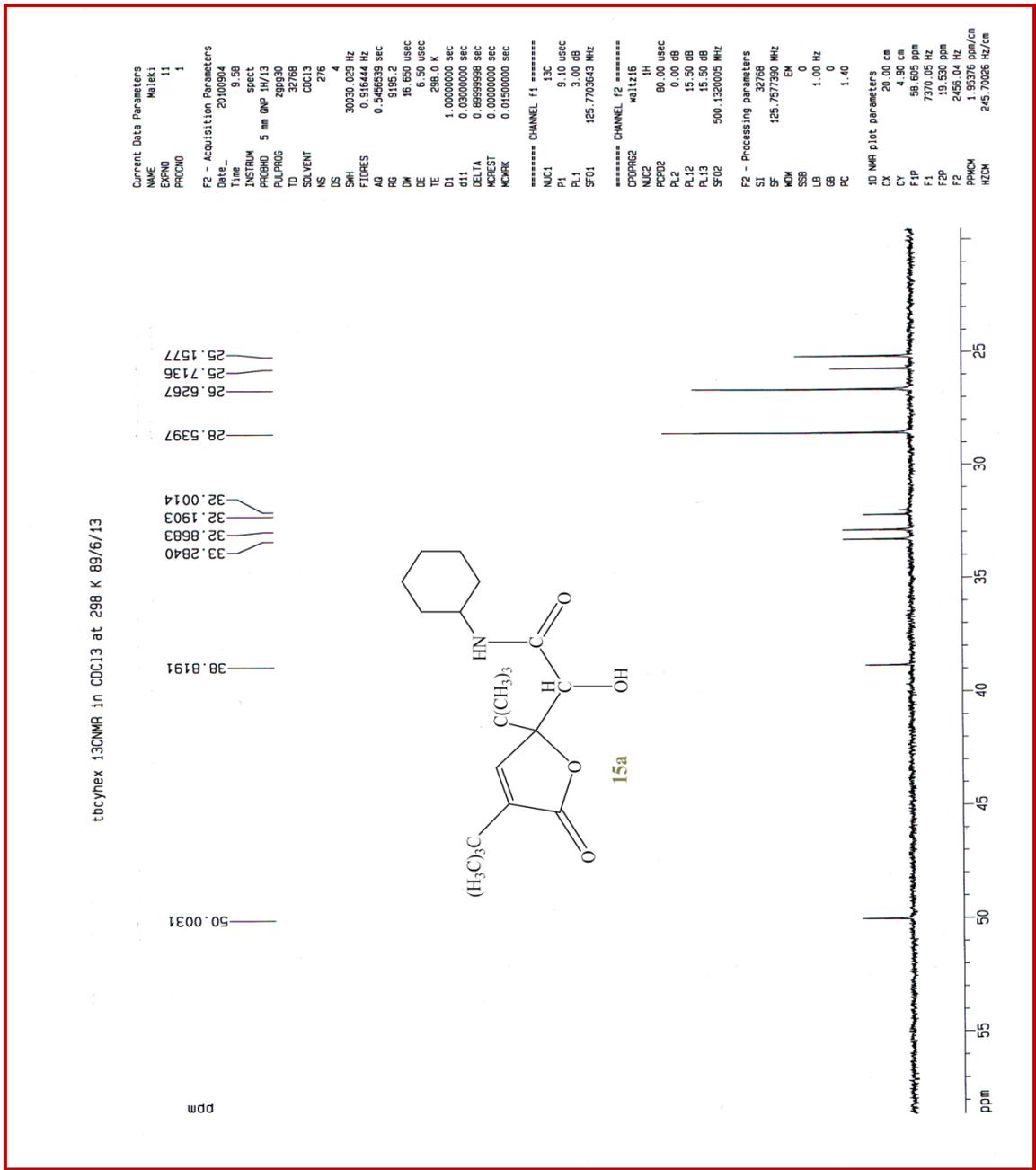
***** CHANNEL f1 *****
NUC1      13C
P1        9.10 usec
PL1       3.00 dB
SF01     125.7703643 MHz

***** CHANNEL f2 *****
CPDPRG2  waltz16
NUC2      1H
P2        60.00 usec
PL2       0.00 dB
PL12     15.50 dB
PL13     15.50 dB
SF02     500.1300005 MHz

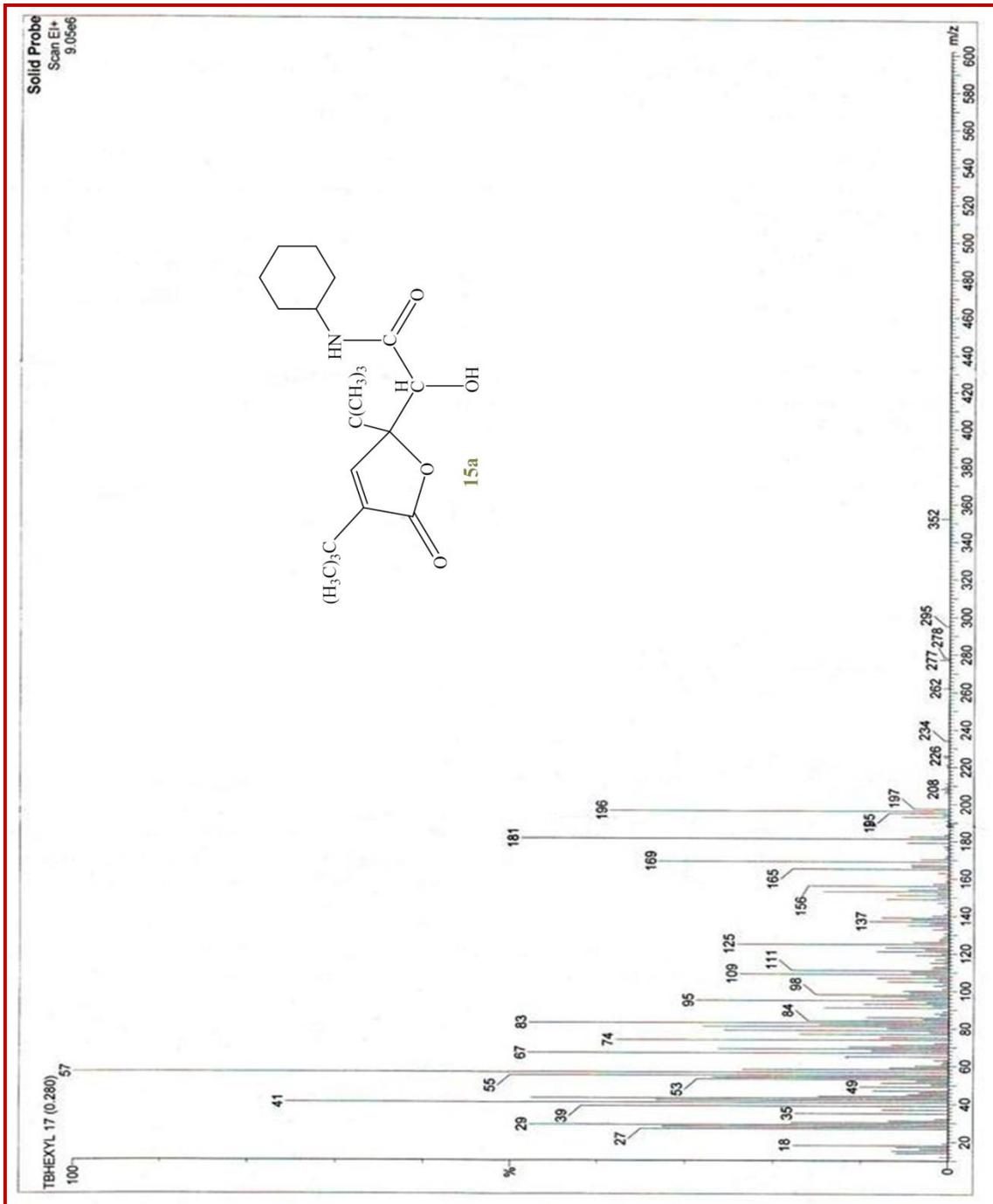
F2 - Processing parameters
SI        32768
SF        125.7577390 MHz
MGM       EM
SSB       0
LB        1.00 Hz
GB        0
PC        1.40

1D NMR plot parameters
CY        20.00 cm
CX        4.00 cm
FIP       213.549 ppm
F1        26865.45 Hz
F2        7.316 ppm
FREQM     10.31150 ppm/cm
HZCM      1296.75122 Hz/cm
  
```

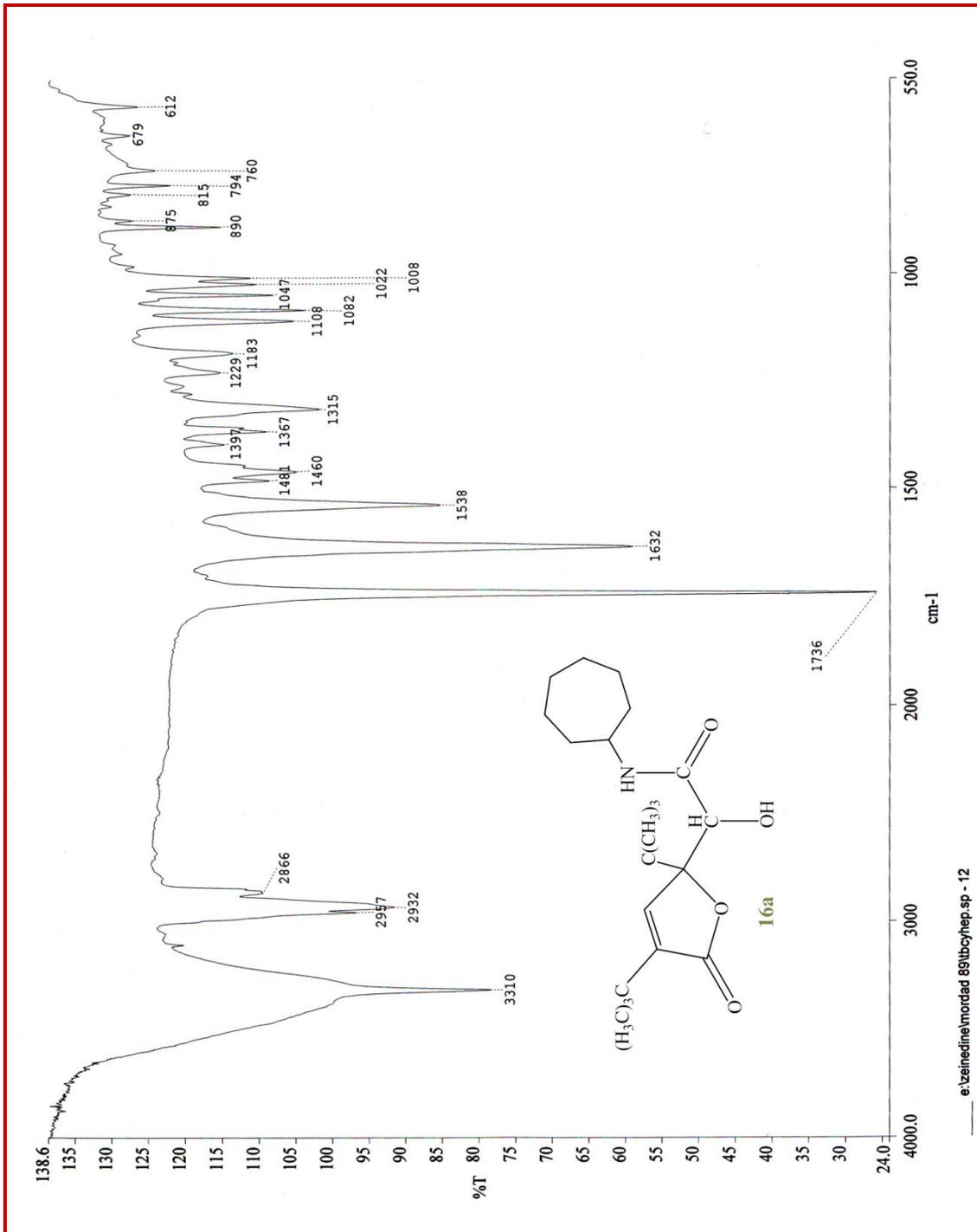
### <sup>13</sup>C NMR of 15a



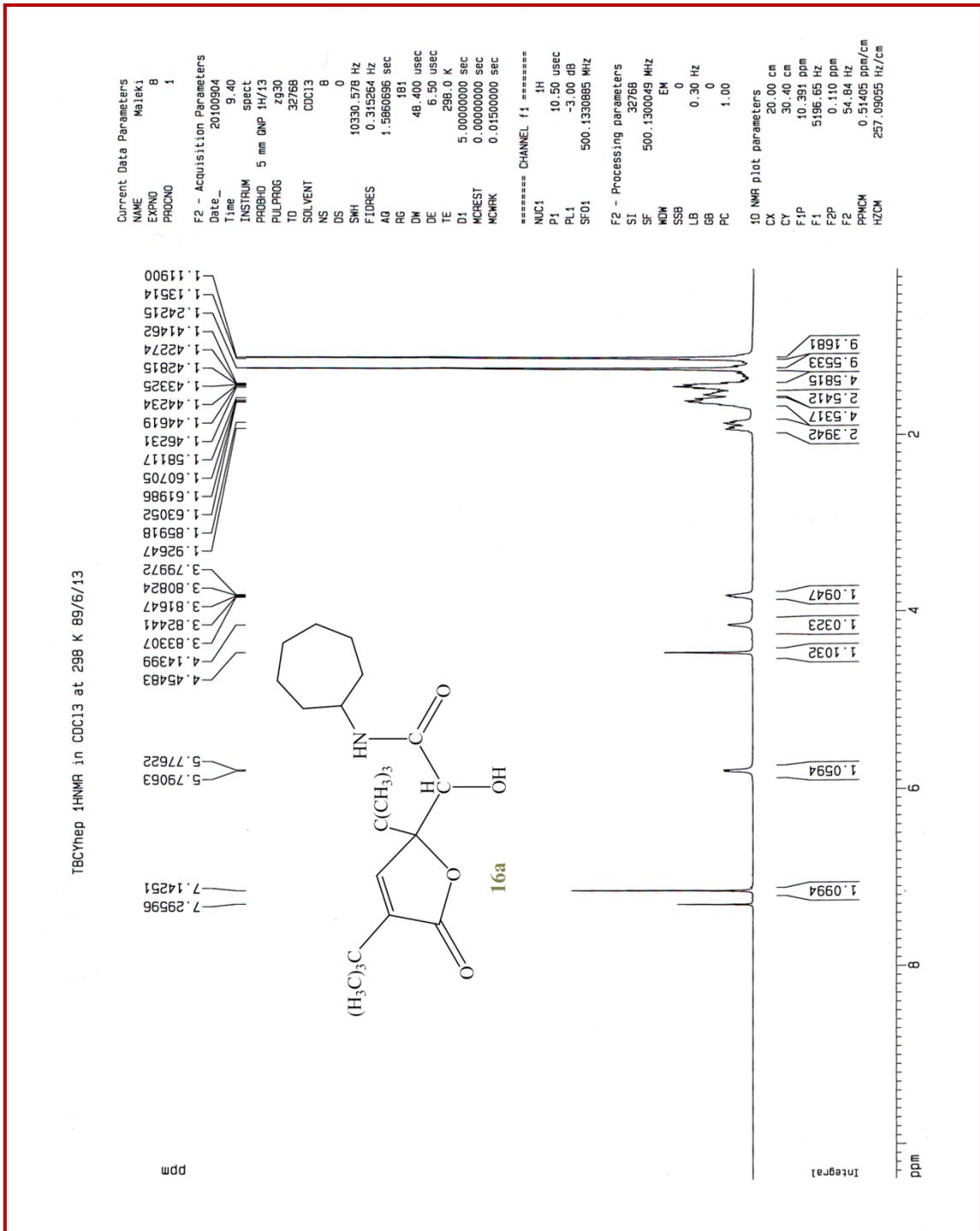
Expanded <sup>13</sup>C NMR of 15a



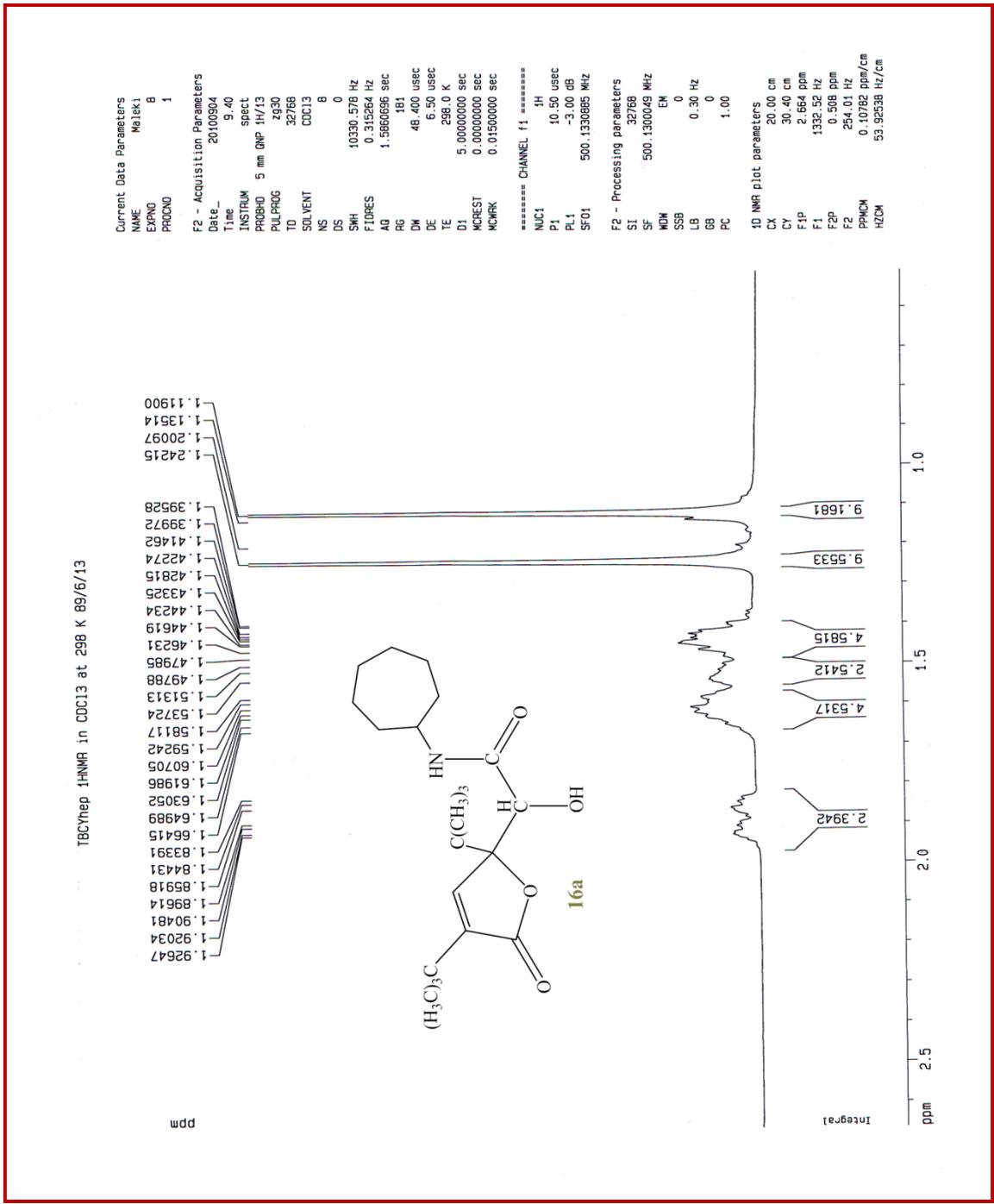
MS of 15a



IR of 16a

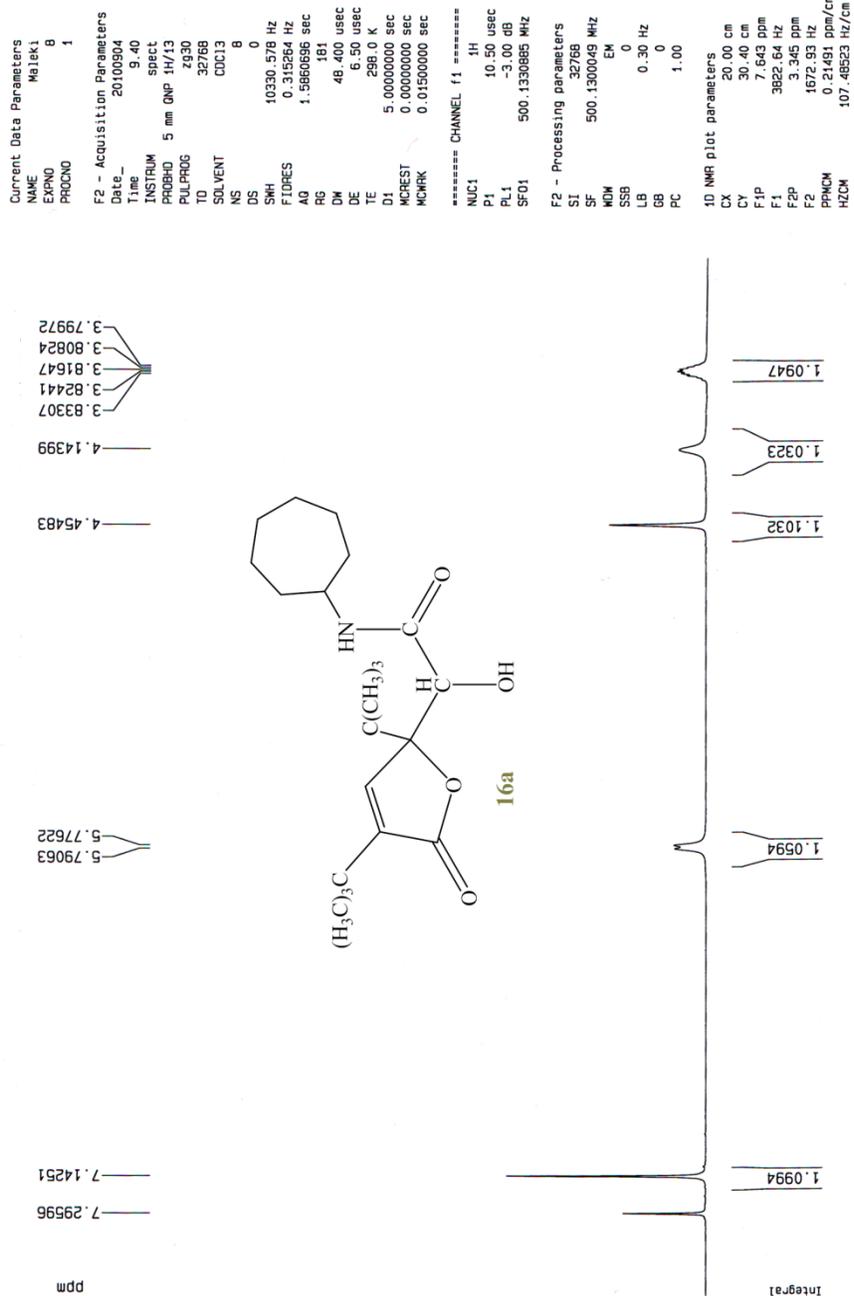


Expanded <sup>1</sup>H NMR of 16a

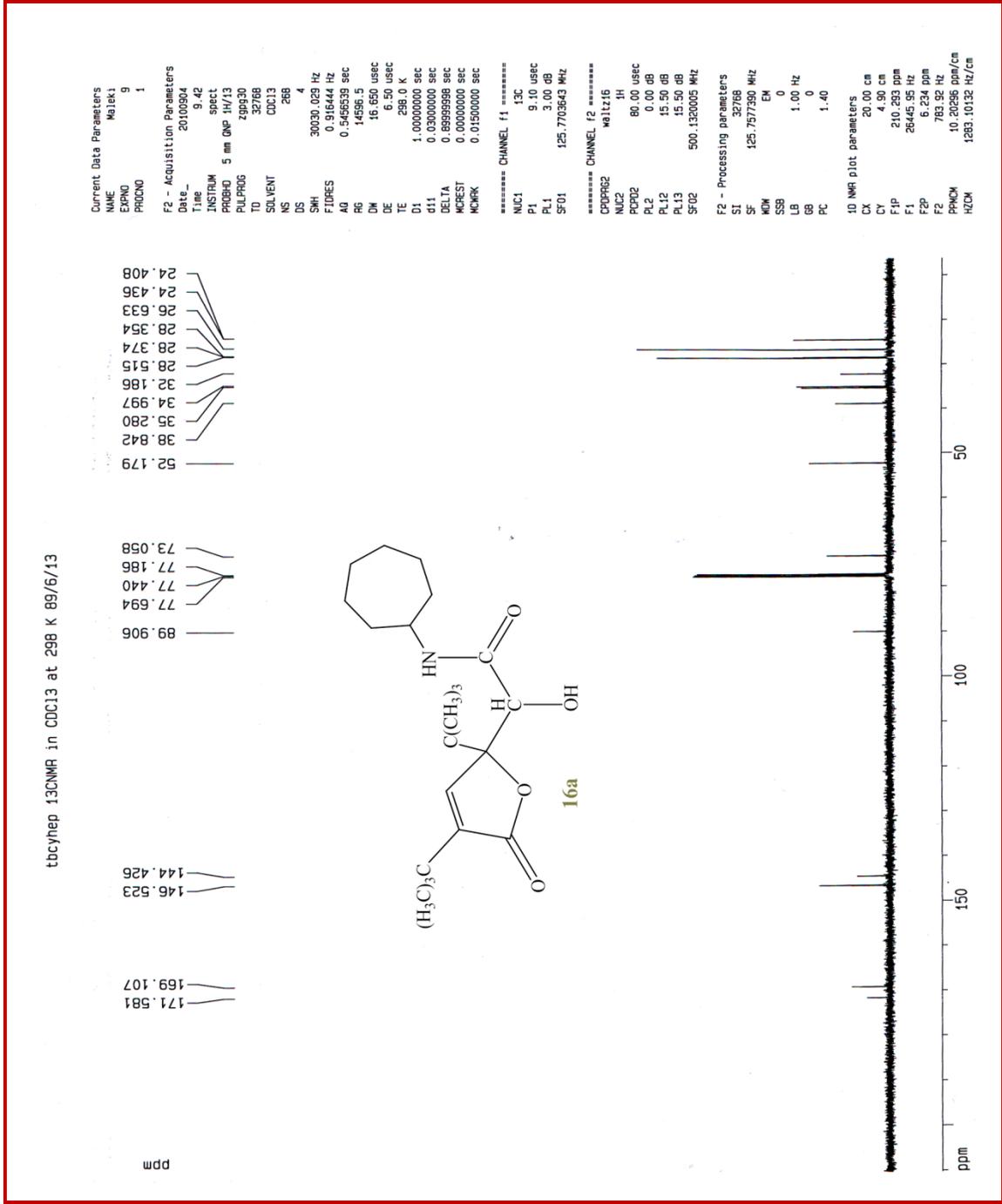


Expanded <sup>1</sup>H NMR of 16a

TBCYhep 1HNMR in CDCl3 at 298 K 89/6/13



Expanded <sup>1</sup>H NMR of 16a



<sup>13</sup>C NMR of 16a

tbodyhep <sup>13</sup>C NMR in CDCl<sub>3</sub> at 298 K 89/6/13

```

Current Data Parameters
NAME      Maleki
EXPNO     9
PROCNO    1

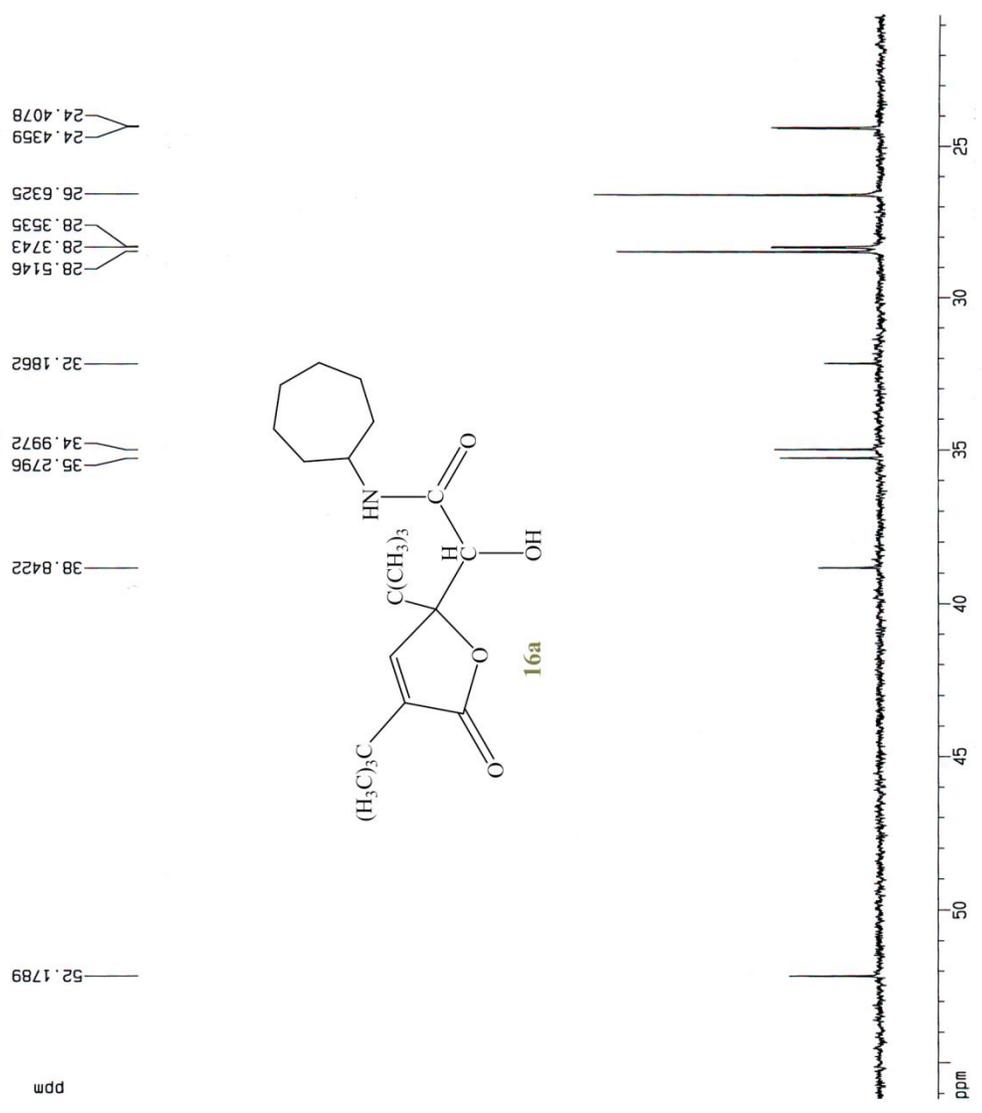
F2 - Acquisition Parameters
Date_     20100904
Time      9.42
INSTRUM   spect
PROBHD    5 mm QNP 1H/13
PULPROG   zgpg30
TD         32768
SOLVENT   CDCl3
NS         268
DS         4
SWH        30030.029 Hz
FIDRES     0.016444 Hz
AQ          0.546659 sec
RG          1496.5
DM          16.650 usec
DE          6.50 usec
TE          298.0 K
D1          1.0000000 sec
d11        0.0300000 sec
DELTA      0.8999998 sec
MORREST    0.0000000 sec
MORMR      0.0150000 sec

===== CHANNEL f1 =====
NUC1       13C
P1          9.10 usec
PL1         3.00 dB
SFO1       125.7703643 MHz

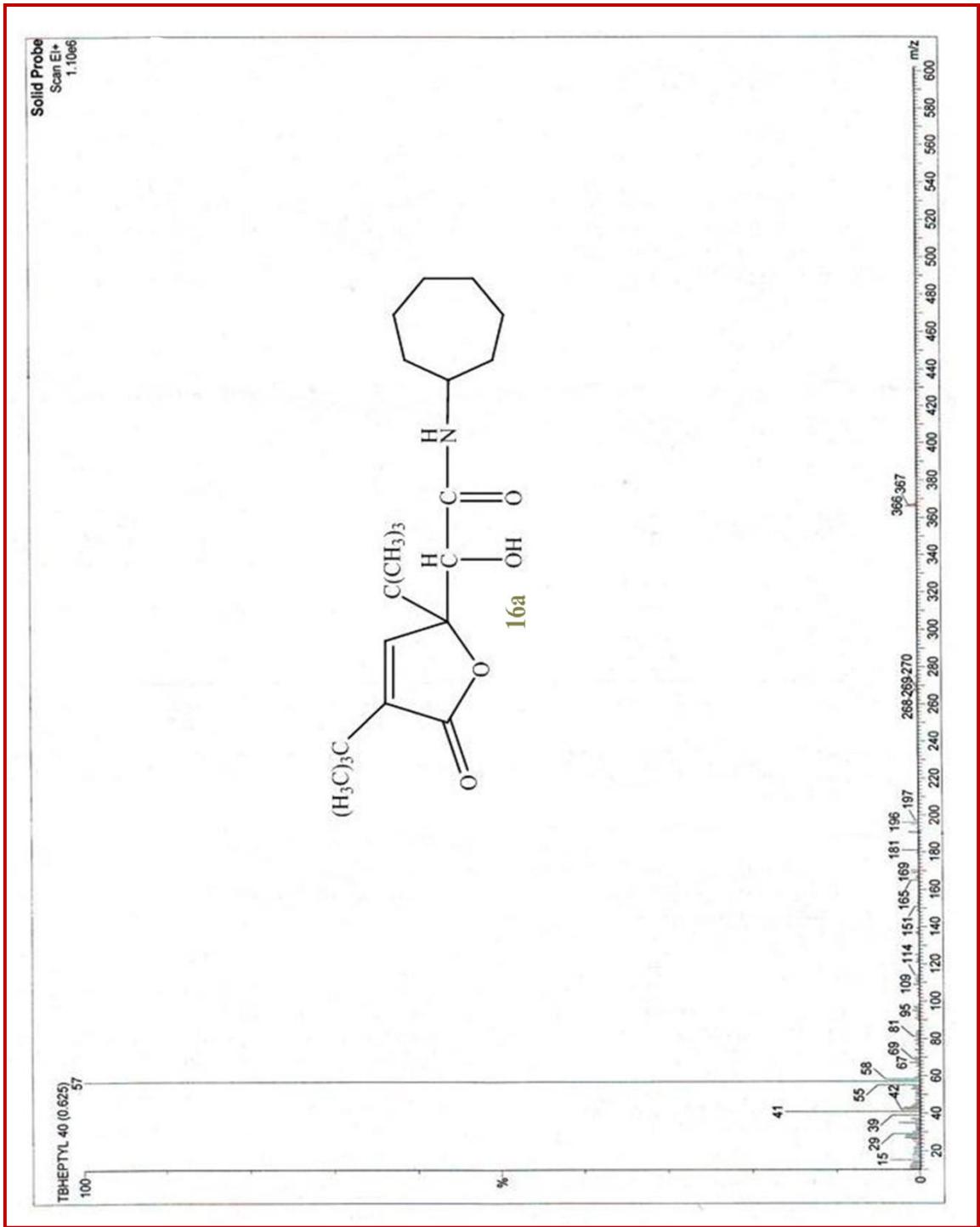
===== CHANNEL f2 =====
CPDPRG2    waltz16
NUC2        1H
PCPD2       80.00 usec
PL2         0.00 dB
PL12        15.50 dB
PL13        15.50 dB
SFO2        500.1320005 MHz

F2 - Processing parameters
SI          32768
SF          125.7577890 MHz
WDW         EM
SSB         0
LB          1.00 Hz
GB          0
PC          1.40

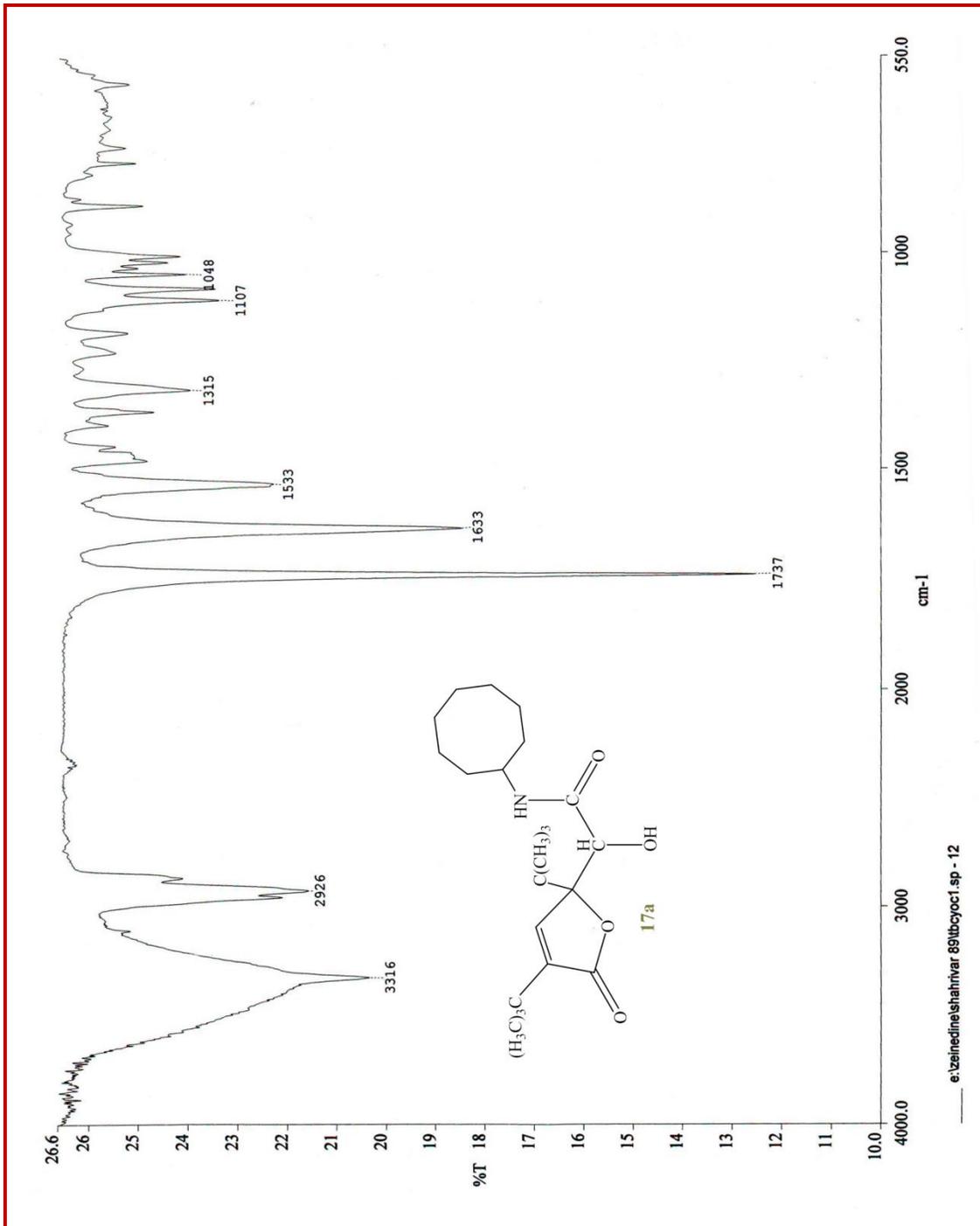
ID NMR plot parameters
CA          20.00 cm
CT          4.90 cm
F1P         56.175 ppm
F1          7064.48 Hz
F2P         20.674 ppm
F2          2599.91 Hz
PPHCK       1.77506 ppm/cm
HZCM        223.22813 Hz/cm
    
```



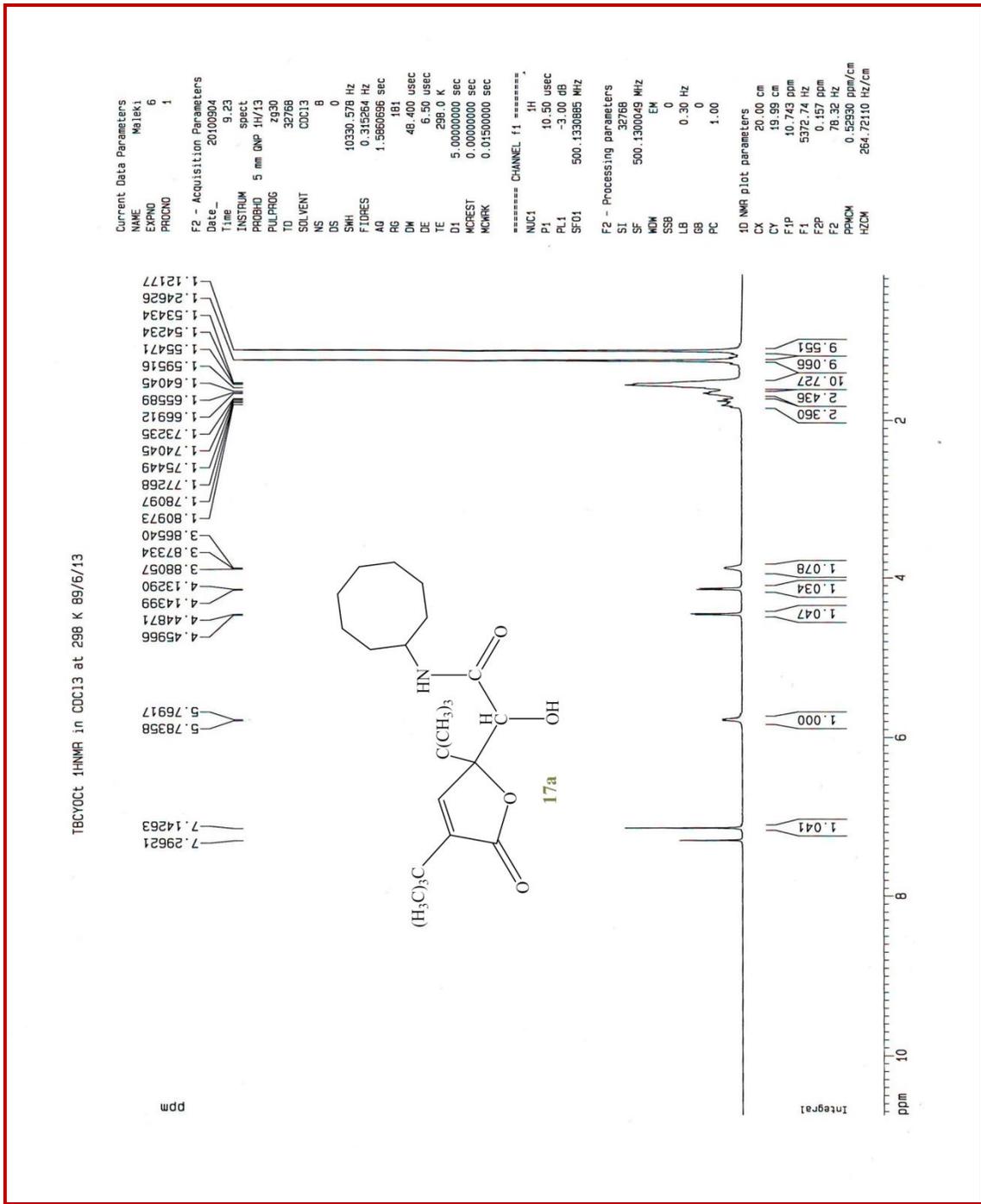
Expanded <sup>13</sup>C NMR of 16a



MS of 16a



FTIR of 17a



<sup>1</sup>H NMR of 17a

TBCYOct 1HNMR in CDCl3 at 298 K 89/6/13

Current Data Parameters  
NAME Maleki  
EXPNO 6  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20100904  
Time 9.23

INSTRUM spect  
PROBHD 5 mm QNP 1H/13  
PULPROG zg30  
TD 32768

SOLVENT CDCl3  
NS 8  
DS 0

SWH 10330.578 Hz  
FIDRES 0.315264 Hz  
AQ 1.5860596 sec

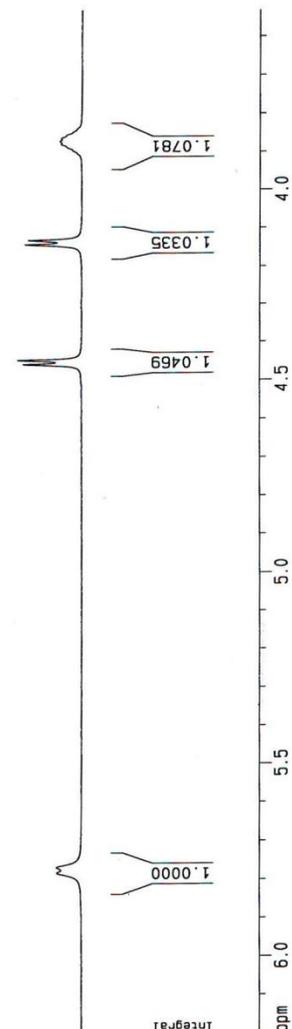
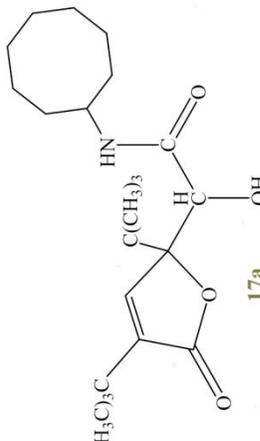
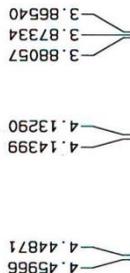
RG 181  
DM 48.400 usec  
DE 6.50 usec  
TE 298.0 K

D1 5.0000000 sec  
MCRET 0.0000000 sec  
MCHK 0.01500000 sec

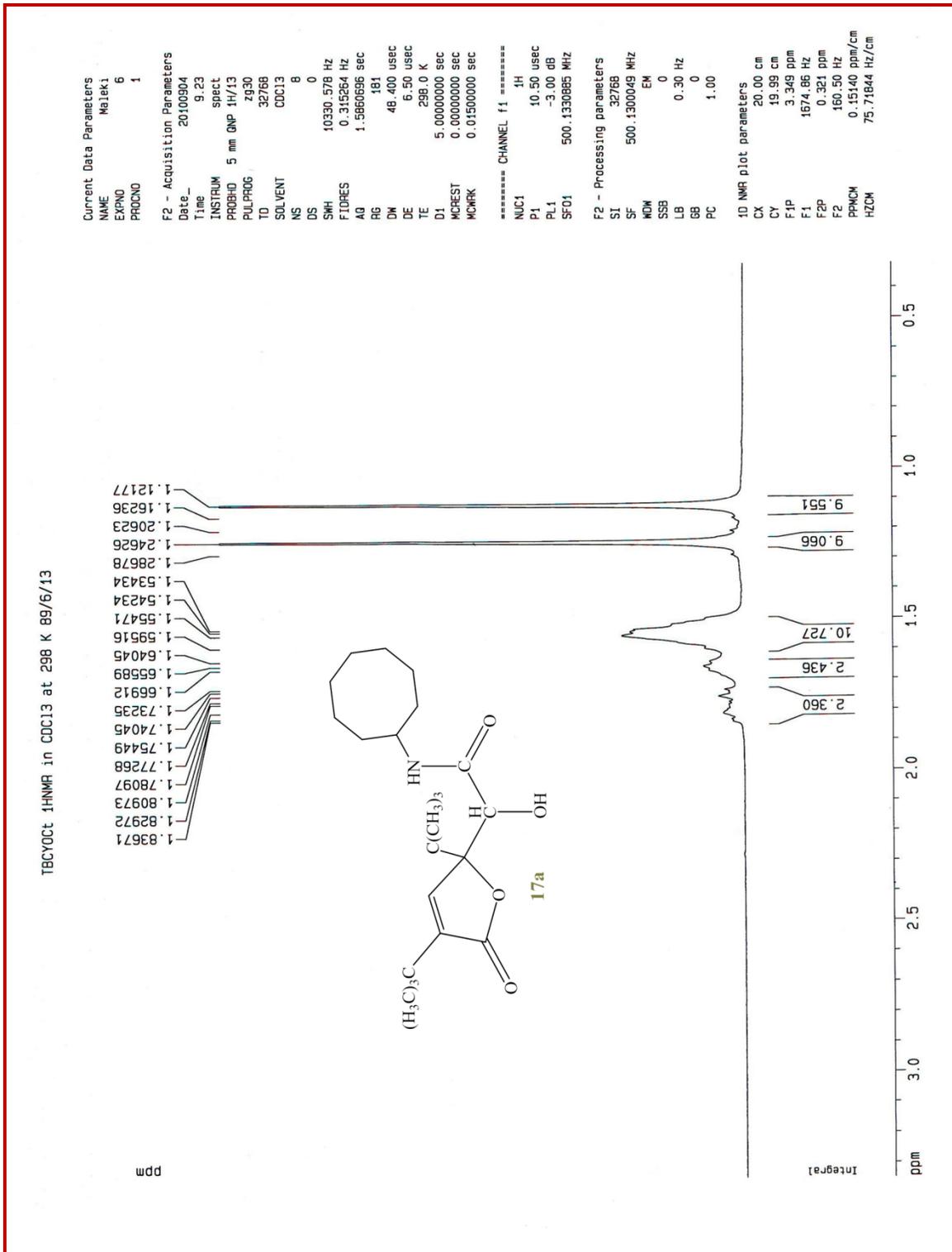
\*\*\*\*\* CHANNEL f1 \*\*\*\*\*  
NUC1 1H  
P1 10.50 usec  
PL1 -3.00 dB  
SFO1 500.1330985 MHz

F2 - Processing parameters  
SI 32768  
SF 500.1300049 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00

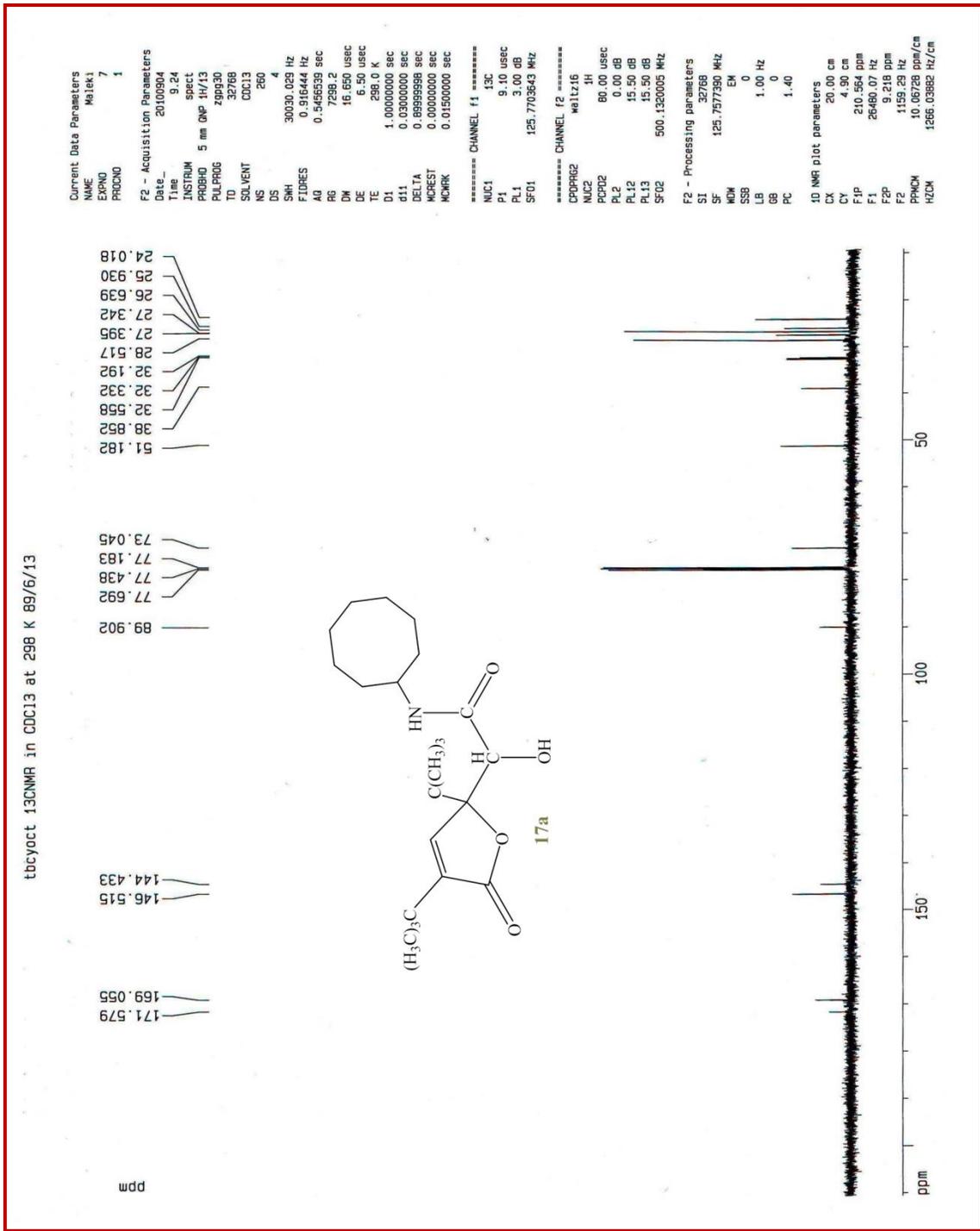
1D NMR plot parameters  
CX 20.00 cm  
CY 19.99 cm  
F1P 6.201 ppm  
F1 3101.19 Hz  
F2P 3.532 ppm  
F2 1766.43 Hz  
PPMCM 0.13344 ppm/cm  
HZCM 66.73786 Hz/cm



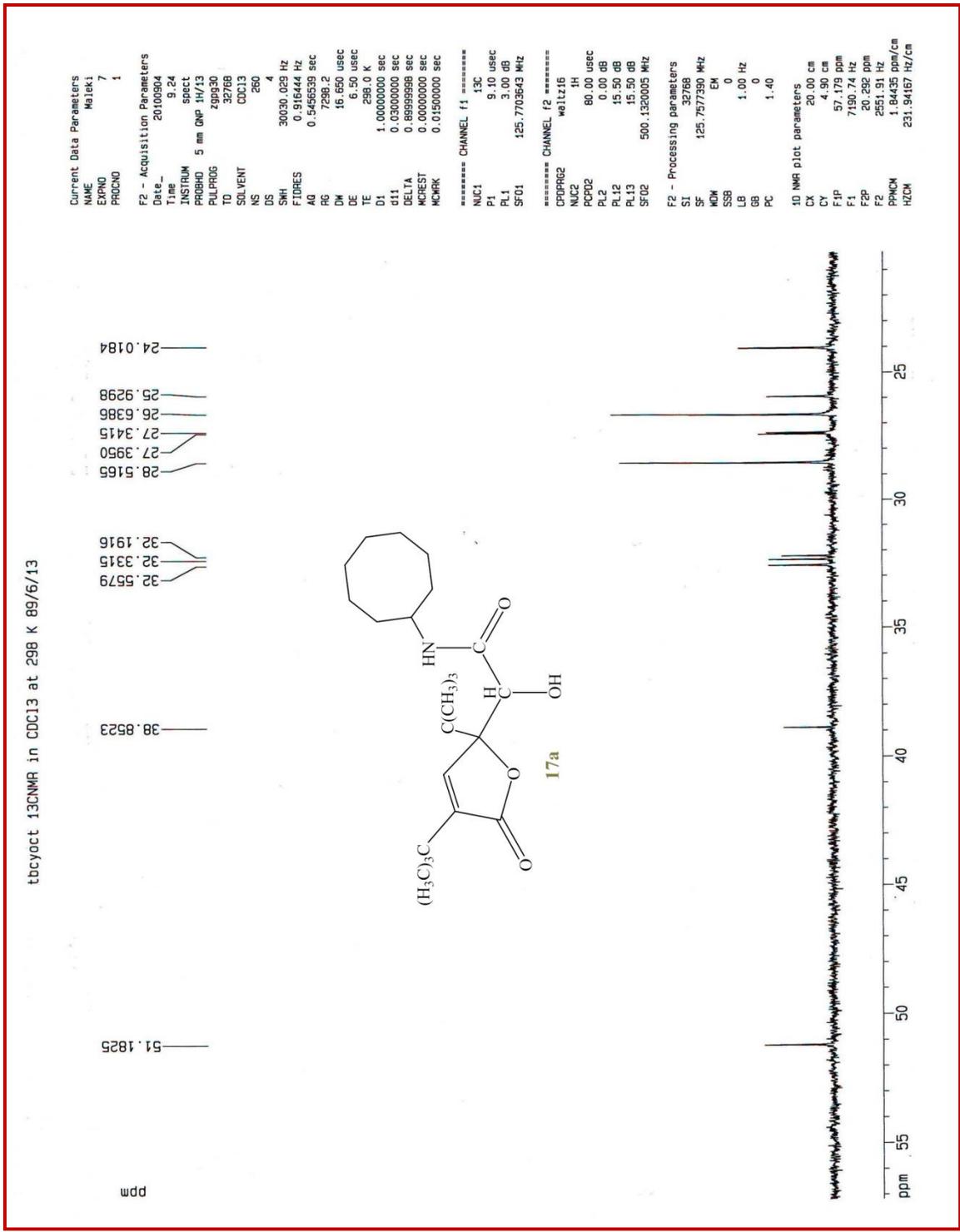
Expanded <sup>1</sup>H NMR of 17a



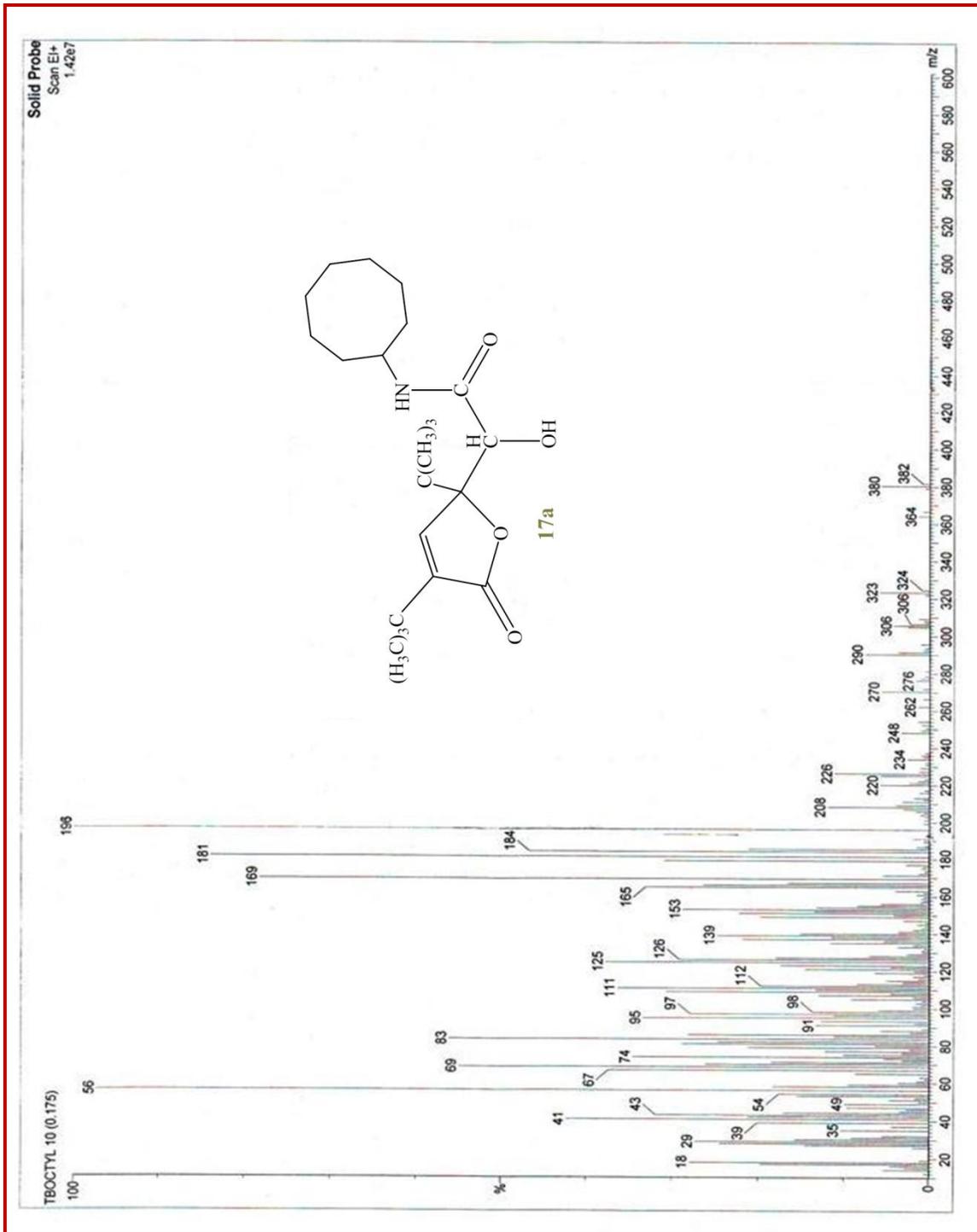
Expanded <sup>1</sup>H NMR of 17a



<sup>13</sup>C NMR of 17a



Expanded <sup>13</sup>C NMR of 17a



MS of 17a

## VI. Crystallography of 9a

Table S2. Selected bond lengths (Å) and bond Angles (°) of compound 9a

<i>Bond lengths</i>	
C(2)–C(7)	1.326(3)
C(7)–C(8)	1.498(2)
C(8)–O(2)	1.459(2)
C(1)–O(2)	1.353(2)
C(1)–O(1)	1.205(2)
C(13)–C(14)	1.520(3)
C(8)–C(13)	1.565(3)
C(14)–N(1)	1.330(3)
C(14)–O(4)	1.233(2)
C(15)–N(1)	1.459(3)
<i>Bond angles</i>	
O(3)–C(13)–C(8)	112.89(16)
O(3)–C(13)–C(14)	109.89(16)
C(14)–C(13)–C(8)	110.50(16)
O(4)–C(14)–N(1)	122.7(2)
O(4)–C(14)–C(13)	120.16(19)
N(1)–C(14)–C(13)	117.11(17)
N(1)–C(15)–C(16)	112.5(2)
C(14)–N(1)–C(15)	122.57(19)
C(1)–O(2)–C(8)	109.69(13)
O(1)–C(1)–C(2)	130.32(19)

Table S3: Crystal data and structure refinement for compound **9a**

Empirical formula	C <sub>18</sub> H <sub>31</sub> NO <sub>4</sub>
Formula weight	326
Temperature (K)	298(2)
Wavelength (Å)	0.71073
Crystal system	Triclinic
Space group	P -1
a (Å)	9.384(2)
b (Å)	10.484(2)
c (Å)	11.281(2)
$\alpha$ (°)	102.829(17)
$\beta$ (°)	94.191(18)
$\gamma$ (°)	113.456(18)
Volume (Å <sup>3</sup> )	976.4(4)
Z	2
Density(Mg/m <sup>3</sup> )	1.107
Crystal size (mm <sup>3</sup> )	0.50×0.17×0.15
Absorption coefficient (mm <sup>-1</sup> )	0.077
$\theta$ range for data collection (°)	1.88 to 29.31
Index ranges	-10 ≤ <i>h</i> ≤ 12 -14 ≤ <i>k</i> ≤ 14 -15 ≤ <i>l</i> ≤ 15
Reflections collected	11380
Independent reflections	5246 [R <sub>int</sub> = 0.0611]
Absorption correction	None
Max. and min. transmission	0.5263 and 0.2964
Refinement method	Full-matrix least-squares on $F^2$
Goodness-of-fit on $F^2$	1.127
Final <i>R</i> indices [ <i>I</i> > 2σ( <i>I</i> )]	<i>R</i> <sub>1</sub> = 0.0744, <i>wR</i> <sub>2</sub> = 0.1880
<i>R</i> indices (all data)	<i>R</i> <sub>1</sub> = 0.1106, <i>wR</i> <sub>2</sub> = 0.2092
Largest diff. peak and hole (e.Å <sup>-3</sup> )	0.401 and -0.392

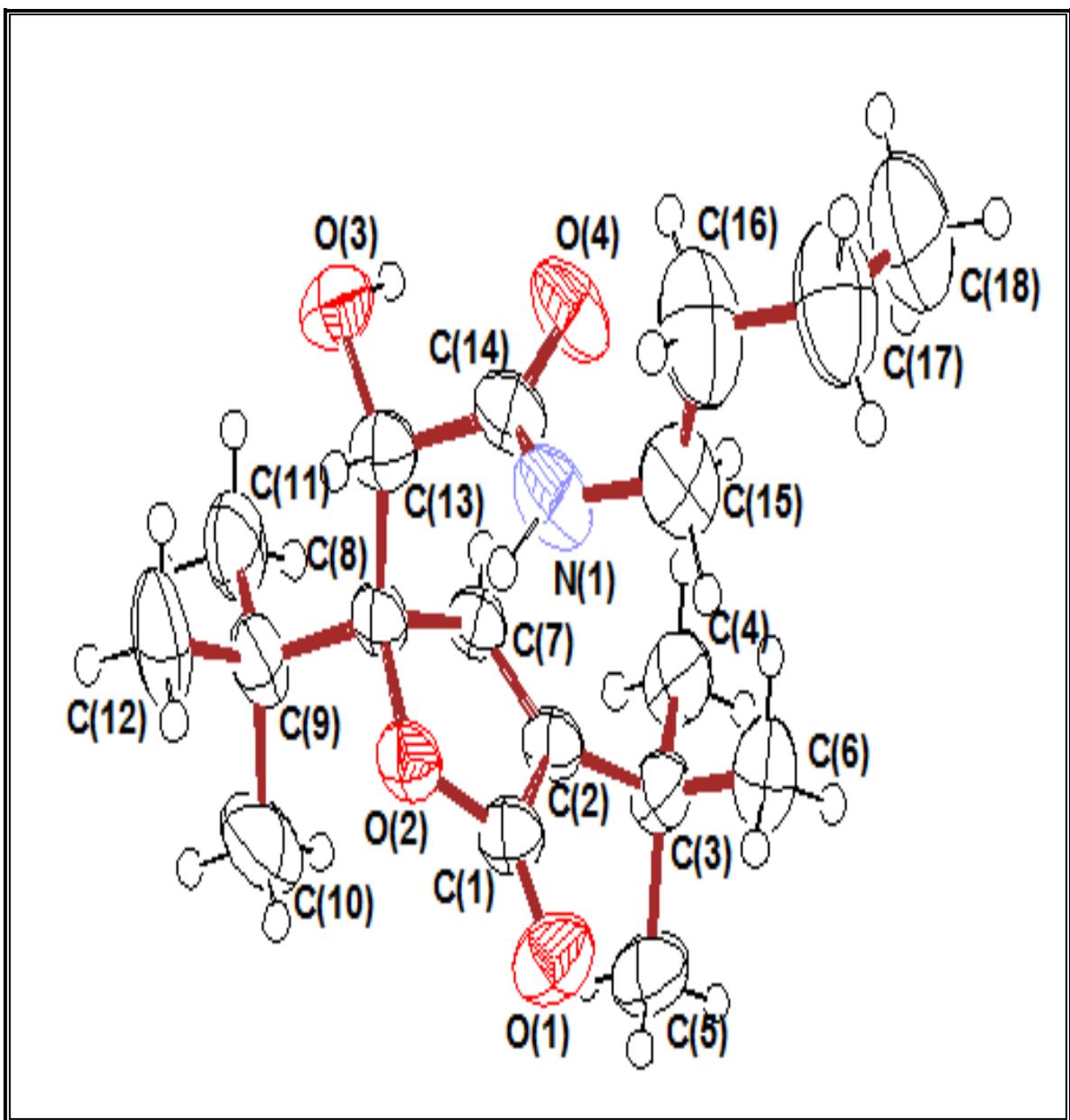
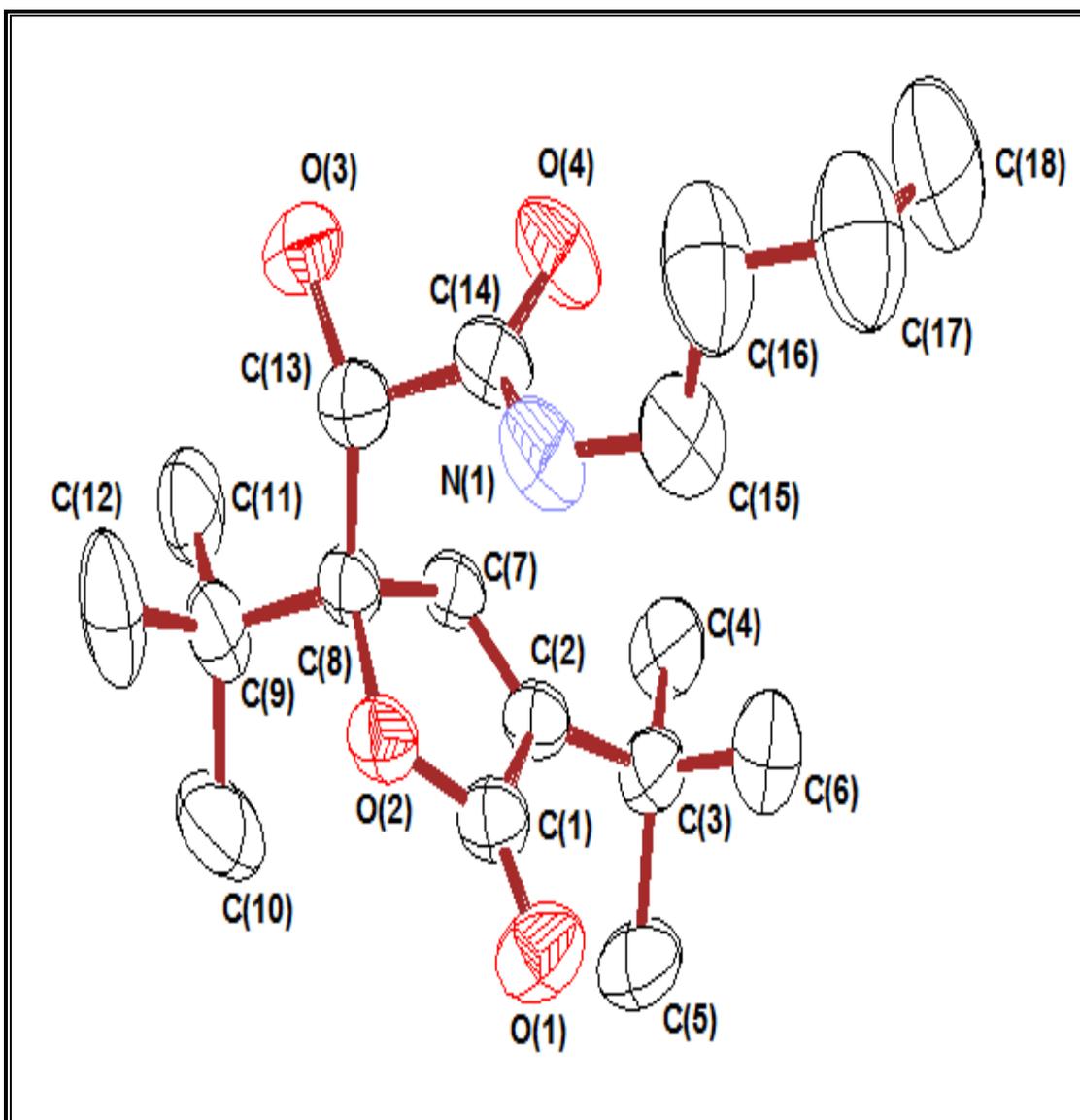


Figure S1. X-ray crystal structure of compound **9a**.



**Figure S2.** X-ray crystal structure of compound **9a**. The hydrogen atoms are omitted for the reason of clarity.