

1 **Supplemental Information for:**

2 **Deformability based Sorting of Red Blood Cells Improves Diagnostic Sensitivity**  
 3 **for Malaria Caused by Plasmodium Falciparum**

4 Quan Guo<sup>a</sup>, Simon P. Duffy<sup>a</sup>, Kerryn Matthews<sup>a</sup>, Xiaoyan Deng<sup>a</sup>, Aline T. Santoso<sup>a</sup>, Emel Islamzada<sup>a</sup> and  
 5 Hongshen Ma<sup>a,b,c</sup>

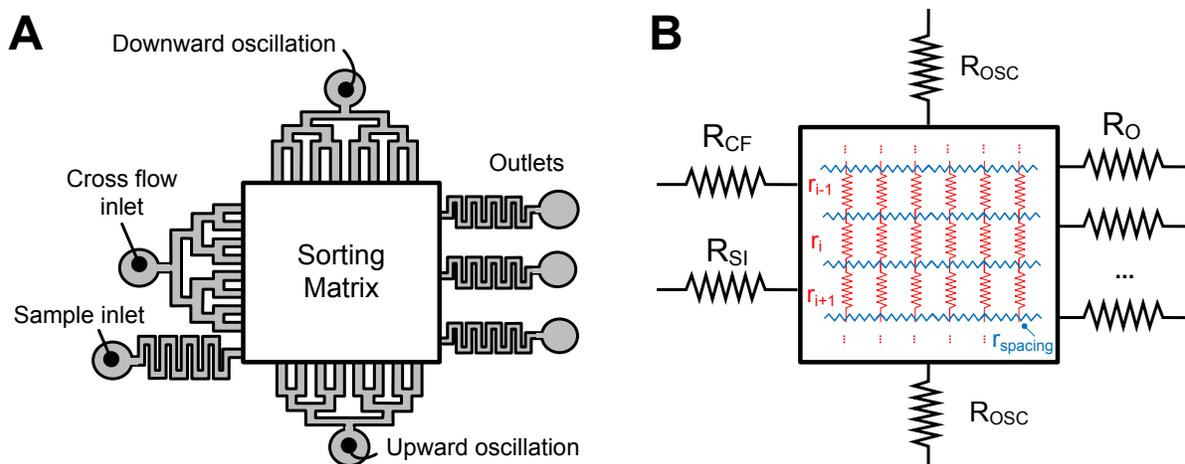
6 *a Department of Mechanical Engineering, University of British Columbia, 2054-6250 Applied Science Lane,*  
 7 *Vancouver, BC, Canada V6T 1Z4*

8 *b Department of Urologic Science, University of British Columbia, Vancouver, BC, Canada*

9 *c Vancouver Prostate Centre, Vancouver General Hospital, Vancouver, BC, Canada*

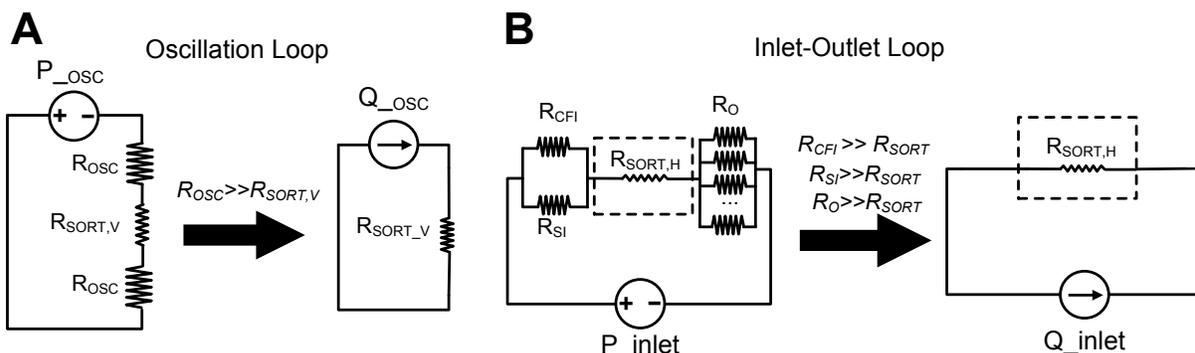
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11 **SUPPLEMENTAL FIGURES AND TABLES:**



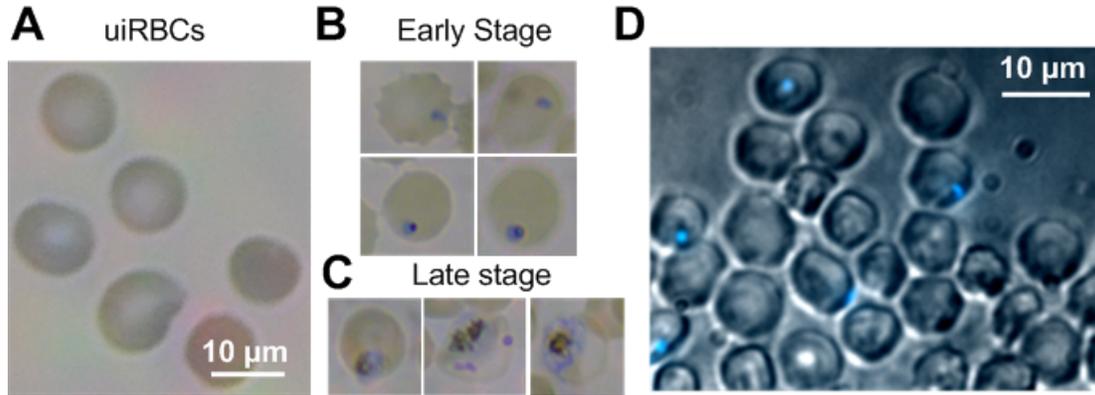
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13 **Supplemental Figure 1.** Schematic illustration of the microfluidic ratchet sorting device (A) and its  
 14 equivalent hydrodynamic resistance model (B)



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16 **Supplemental Figure 2.** Design principle of the microfluidic ratchet sorting device explained using the  
 17 system hydrodynamic resistance modeling. (A) Equivalent hydrodynamic resistance model of the  
 18 oscillation flow loop. (B) Equivalent hydrodynamic resistance model of the inlet-outlet flow loop.



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20 **Supplemental Figure 3.** Giemsa stain light microscopy for determining pre-sorted sample parasitemia  
 21 and Hoechst DNA stain of parasitized RBCs at outlets after sorting. Giemsa stain microscopy showing (A)  
 22 uiRBCs as well as *Pf*-iRBCs at (B) early (0-24 hours) and (C) late stages (24-48 hours). (D) Hoechst 33342  
 23 stain of parasites within the infected RBCs, illuminating parasite DNA under fluorescence microscope.

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25 **Supplemental Table 1. Summary of hydrodynamic resistance of ratchet funnel**

Pore size ( $\mu\text{m}$ )	1.5	1.75	2	2.25	2.5	3	3.5	6	7.5
Hydrodynamic resistance ( $\times 10^{14} \text{ Pa s/m}^3$ )	64.6	46.3	34.9	27.5	22.5	16.1	14.3	12.4	5.07

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27 **Supplemental Table 2. Summary of hydrodynamic resistance of various components of the device**

Components	Hydrodynamic resistance ( $\times 10^{14} \text{ Pa s/m}^3$ )
Cross flow inlet ( $R_{CF}$ )	150
Sample inlet ( $R_{SI}$ )	120
Oscillation inlet ( $R_{OSC}$ )	100
Single outlet channel ( $R_O$ )	1200
Sorting region (vertical direction: $R_{SORT,V}$ )	1.65
Sorting region (horizontal direction: $R_{SORT,H}$ )	20

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