# LabDisk for SAXS: A centrifugal microfluidic sample preparation platform for small-angle X-ray scattering

# Supplementary information

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## Processing device for the LabDisk for SAXS

ESI Fig. 1: The processing device for the LabDisk for SAXS.

#### LabDisk for SAXS in P12 beamline



ESI Fig. 2: LabDisk for SAXS in P12 beamline (PETRA III, DESY) at EMBL Hamburg.

## Liquid loss due to cornerflow



ESI Fig. 3: Liquid loss due to cornerflow in the first and last aliquoting fingers of the LabDisk for SAXS. Displayed is an older version of the LabDisk for SAXS fabricated as a milled disk in PMMA and sealed with adhesive film. After long holding times liquid loss is clearly visible in the first and last aliquoting finger.

## Buffer subtracted curves collected with the LabDisk for SAXS



ESI Fig. 4: Buffer subtracted curves collected with the LabDisk for SAXS: the curves are offset for clarity. Protein concentration: 1.8 mg/ml (black), 3.7 mg/ml (red), 5.5 mg/ml (red), 11 mg/ml (yellow). NaCl concentration (from bottom to top): 0 mM, 83 mM, 166 mM, 250 mM, 333 mM.

#### **Dead volumes**

ESI Table 1 shows which amount of protein volume is used for fluidic processing and which amount of protein volume ends up in the actual read-out chambers. Reduction of total sample volume without change to the read-out chambers could be acchieved by reducing the pipetting tolerance, reducing the size of the feeding channel or reducing the excess volume in the read-out chambers.

Another way to reduce total sample volume would be to shrink the size of all volumes. However, this would include reduction of the size of the read-out chambers. In the current design the read-out chamber in the frontside foil has an aspect ratio of 1. Reducing the diameter of the read-out chamber would lead to an aspect ratio larger than 1, which is not recommended for micro-thermoforming. Reducing the depth of the read-out chamber would decrease the height of the measured liquid column, 860  $\mu$ m in the current design, and consequently reduce the data quality.

ESI Table 1: Calculation of the protein volume used for SAXS analysis. Out of the total input protein volume of 2.5  $\mu$ l, 680 nl are used for the actual SAXS analysis. Additional liquid volume is required to tolerate pipetting errors, to ensure proper aliquoting and to ensure complete filling of the read-out chambers.

Туре	Dead volume	Remaining protein volume
Total input protein volume	-	2.5 μl
Extra volume included for	500 nl	2.0 µl
pipetting tolerance		
Extra volume required to ensure	~680 nl	1.32 µl
proper aliquoting (limitted by the		
size of the feeding channel)		
Excess volume included to ensure	49%	<u>680 nl</u>
complete filling of the read-out	(123 nl per read-out chamber)	
chambers		

## Calculation of $\chi^2$

 $\chi^2$  can be calculated from eqn (1), where n is the number of data points and  $\sigma$  is the

experimental error.<sup>1</sup>

$$\chi^{2} = \frac{1}{n-1} \sum_{k=1}^{n} \left[ \frac{I_{exp}(q_{k}) - I_{calc}(q_{k})}{\sigma(I_{exp}(q_{k}))} \right]^{2}$$

#### **References**

1 K. Pearson, *The London, Edinburgh, and Dublin Philosophical Magazine and Journal of Science*, 1900, **50**, 157–175.