

## Discovery and optimisation of 1-acyl-2-benzylpyrrolidines as potent dual orexin receptor antagonists

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### Electronic supplementary information contents:

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<sup>1</sup>H & <sup>13</sup>C NMR spectra of final compounds

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### Synthetic details for the preparation of compounds:

Commercially available starting materials were used as received without further purification. Flash column chromatography was performed using Biotage SNAP cartridges (10–340 g) and elution was with a Biotage Isolera system. Merck pre-coated thin layer chromatography (TLC) plates were used for TLC analysis. Final compounds were purified to >95% purity (UV and NMR) by reverse phase preparative HPLC using a Waters XBridge column (10 μm, 75 x 30 mm). Conditions: MeCN [eluent A]; water + 0.5% NH<sub>4</sub>OH (25% aq.) [eluent B]; Gradient: 90% B → 5% B over 6.5 min (flow: 75 mL/min). Detection: UV/Vis + MS. Racemates can be separated into their enantiomers by chiral HPLC using a ChiralPaK IC column (5 μm, 250 x 4.6 mm). Conditions: Heptane + 0.05% DEA [eluent A]; EtOH + 0.05% DEA [eluent B]; Isocratic elution with 50% eluent B (flow: 1 mL/min), or a CHIRALCEL OZ-H column (5 μm, 250 x 4.6 mm). Conditions: CO<sub>2</sub> [eluent A]; EtOH + 0.1% DEA [eluent B]; Isocratic elution with 50% eluent B (flow: 4 mL/min).

Mass spectrometry data were recorded by one of the following methods:

#### LC-MS with acidic conditions

**Method A:** Agilent 1100 series with mass spectrometry detection (MS: Finnigan single quadrupole). Column: Zorbax SB-aq (3.5 μm, 4.6 x 50 mm). Conditions:

MeCN [eluent A]; water + 0.04% TFA [eluent B]. Gradient: 95% B → 5% B over 1.5 min (flow: 4.5 mL/min). Detection: UV/Vis + MS.

**Method B:** Agilent 1100 series with mass spectrometry detection (MS: Finnigan single quadrupole). Column: Waters XBridge C18 (2.5 μm, 4.6 x 30 mm). Conditions: MeCN [eluent A]; water + 0.04% TFA [eluent B]. Gradient: 95% B → 5% B over 1.5 min (flow: 4.5 mL/min). Detection: UV/Vis + MS.

#### LC-MS with basic conditions

**Method C:** Agilent 1100 series with mass spectrometry detection (MS: Finnigan single quadrupole). Column: Zorbax Extend C18 (5 μm, 4.6 x 50 mm). Conditions: MeCN [eluent A]; 13 mmol/L NH<sub>3</sub> in water [eluent B]. Gradient: 95% B → 5% B over 1.5 min (flow: 4.5 mL/min). Detection: UV/Vis + MS.

**Method D:** Agilent 1100 series with mass spectrometry detection (MS: Finnigan single quadrupole). Column: Waters XBridge C18 (5 μm, 4.6 x 50 mm). Conditions: MeCN [eluent A]; 13 mmol/L NH<sub>3</sub> in water [eluent B]. Gradient: 95% B → 5% B over 1.5 min (flow: 4.5 mL/min). Detection: UV/Vis + MS.

LC-HRMS parameters were the following: analytical pump Waters Acquity binary, Solvent Manager, MS, SYNAPT G2 MS, source temperature of 150 °C, desolvation temperature of 400 °C, desolvation gas flow of 400 L/h; cone gas flow of 10 L/h, extraction cone of 4 RF; lens 0.1 V; sampling cone 30; capillary 1.5 kV; high resolution mode; gain of 1.0, MS function of 0.2 s per scan, 120–1000 amu in full scan, centroid mode. Lock spray: keucine enkephalin, 2 ng/mL (556.2771 Da), scan time of 0.2 s with interval of 10 s and average of 5 scans; DAD: Acquity UPLC PDA detector. Column was an Acquity UPLC BEH C18 1.7 μm, 2.1 mm x 50 mm from Waters, thermostated in the Acquity UPLC column manager at 60 °C. Eluents were the following: water + 0.05% formic acid; B, acetonitrile + 0.05% formic acid. Gradient was 2–98% B over 3.0 min. Flow was 0.6 mL/min. Detection was at UV 214 nm.

<sup>1</sup>H NMR spectra were recorded on a Bruker (400 or 500 MHz) spectrometer in the indicated deuterated solvent. Chemical shifts are reported in ppm relative to solvent peaks as the internal reference. The substituted pyrrolidine orexin receptor

antagonists described in this manuscript exhibit complex conformations in solution, arising from hindered rotations that are slow on the NMR and LC-MS timescale. The proton spectra typically consist of broad, overlapping multiplets precluding a detailed coupling constant analysis and we feel that pictorial reproductions of the NMR spectra are more informative for comparative purposes; thus, NMR resonances of final compounds will not be listed in numerical format. Instead, we include reproductions of the  $^1\text{H}$  and  $^{13}\text{C}$  NMR spectra at 25 °C of all final compounds. In cases where final compounds appear as a mixture of conformational isomers, visible in their LC-MS spectra, the retention time of the most abundant conformer is given.

clogP values were calculated using ChemBioDraw Ultra 14.0.

Abbreviations used: aq., aqueous; DCM, dichloromethane; DEA, diethylamine; DIPEA, diisopropylethylamine; DMF, dimethylformamide; DMSO, dimethylsulfoxide; er, enantiomeric ratio;  $\text{Et}_2\text{O}$ , diethyl ether; EtOAc, ethyl acetate; EtOH, ethanol; HCl, hydrochloric acid;  $\text{H}_2\text{O}$ , water; HPLC, high performance liquid chromatography; MeCN, acetonitrile; MeOH, methanol; min, minutes;  $\text{NaHCO}_3$ , sodium bicarbonate; NaOH, sodium hydroxide;  $\text{Na}_2\text{SO}_4$ , sodium sulfate;  $^n\text{BuLi}$ , n-butyl-lithium; prep., preparative; py, pyridine; RT, room temperature; Soln., solution; TBME, tert-butyl methyl ether; TBTU, 2-(1*H*-benzotriazole-1-yl)-1,2,3,3-tetramethyluronium tetrafluoroborate; THF, tetrahydrofuran;  $t_{\text{R}}$ , retention time.

### **rac-(2-(3,4-dimethoxybenzyl)piperidin-1-yl)(1-ethyl-1*H*-indol-3-yl)methanone 2**

A prestirred solution of 1-ethyl-1*H*-indole-3-carboxylic acid (17 mg, 0.09 mmol), TBTU (31 mg, 0.1 mmol), and DIPEA (24  $\mu\text{L}$ , 0.14 mmol) in DMF (0.5 mL) was added to a RT solution of 2-(3,4-dimethoxybenzyl)piperidine hydrochloride (purchased from BioBlocks inc.) (25 mg, 0.09 mmol) and DIPEA (24  $\mu\text{L}$ , 0.14 mmol) in DMF (0.5 mL) and the resulting mixture was shaken for 18 h. The reaction mixture was directly purified by prep. HPLC to give the title compound as a yellow oil (26 mg, 69%). LC-MS B:  $t_{\text{R}}$  = 0.92 min;  $[\text{M}+\text{H}]^+$  = 407.21; LC-HRMS:  $t_{\text{R}}$  = 1.32 min;  $[\text{M}+\text{H}]/z$  = 407.2335, found = 407.2340.

**rac-(2-(3,4-dimethoxybenzyl)piperidin-1-yl)(5-methyl-2-(2*H*-1,2,3-triazol-2-yl)phenyl)methanone 4**

The title compound was prepared from 5-methyl-2-(2*H*-1,2,3-triazol-2-yl)benzoic acid (prepared in analogy to the procedure described in WO2008/069997, p28) (19 mg, 0.09 mmol) and 2-(3,4-dimethoxybenzyl)piperidine hydrochloride (purchased from BioBlocks inc.) (25 mg, 0.1 mmol) in analogy to the procedure described for **2**. The product was isolated as a yellow oil (17 mg, 44%). LC-MS D:  $t_R = 0.87$  min;  $[M+H]^+ = 421.22$ ; LC-HRMS:  $t_R = 1.26$  min;  $[M+H]/z = 421.2240$ , found = 421.2243.

**rac-(2-(3,4-dimethoxyphenethyl)piperidin-1-yl)(5-methyl-2-(2*H*-1,2,3-triazol-2-yl)phenyl)methanone 5**

The title compound was prepared from 5-methyl-2-(2*H*-1,2,3-triazol-2-yl)benzoic acid (prepared in analogy to the procedure described in WO2008/069997, p28) (23 mg, 0.11 mmol) and 2-(3,4-dimethoxyphenethyl)piperidine (purchased from UkrOrgSynthesis) (25 mg, 0.1 mmol) in analogy to the procedure described for **2**. The product was isolated as a yellow oil (34 mg, 79%). LC-MS D:  $t_R = 0.95$  min;  $[M+H]^+ = 435.19$ ; LC-HRMS:  $t_R = 1.33$  min;  $[M+H]/z = 435.2396$ , found = 435.2393.

**rac-(2-(3-methoxyphenethyl)piperidin-1-yl)(5-methyl-2-(2*H*-1,2,3-triazol-2-yl)phenyl)methanone 6**

The title compound was prepared from 5-methyl-2-(2*H*-1,2,3-triazol-2-yl)benzoic acid (prepared in analogy to the procedure described in WO2008/069997, p28) (13 mg, 0.06 mmol) and 2-(3-methoxyphenethyl)piperidine (purchased from Matrix Scientific) (13 mg, 0.06 mmol) in analogy to the procedure described for **2**. The product was isolated as a yellow oil (18 mg, 77%). LC-MS D:  $t_R = 1.0$  min;  $[M+H]^+ = 405.29$ ; LC-HRMS:  $t_R = 1.42$  min;  $[M+H]/z = 405.2291$ , found = 405.2298.

**(S)-tetrahydro-3*H*-pyrrolo[1,2-*c*][1,2,3]oxathiazole 1,1-dioxide 29**

A solution of sulfonyl chloride (8 mL, 99 mmol) in DCM (40 mL) was added dropwise over 1.5 h to a well stirred -78°C solution of (S)-(+)-2-(hydroxymethyl)pyrrolidine (10 g, 99 mmol) and pyridine (16 mL, 20 mmol) in DCM (60 mL) under argon. After 3 h the cooling bath was removed and the mixture was allowed to reach 0°C. The reaction mixture was quenched into ice water and transferred to a separating funnel. The layers were separated and the aqueous layer was extracted with DCM (2x). The combined organic extracts were washed with 1M aq. HCl soln., with water, and with

brine. The organic layer was dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and evaporated *in vacuo* to give a yellow solid. This was then re-dissolved in THF (50 mL), filtered through a sintered funnel to remove residual py.HCl and concentrated *in vacuo* to give the title compound as a pale yellow solid (8.8 g, 55%). <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ: 1.81-1.88 (m, 1 H), 1.93-2.09 (m, 2 H), 2.17-2.29 (m, 1 H), 3.26-3.35 (m, 1 H), 3.69-3.78 (m, 1 H), 4.05-4.11 (m, 1 H), 4.25-4.34 (m, 1 H), 4.55-4.61 (m, 1 H).

### **(S)-2-(3,4-dimethoxybenzyl)pyrrolidine 30**

<sup>n</sup>BuLi 1.6 M in hexane (22.8 mL, 36 mmol) was added to a solution of 4-bromoveratrole (5.25 mL, 36 mmol) in THF (40 mL) under Argon at such a rate that the internal temperature remained below -70°C. After a few minutes the mixture became cloudy and a fine white to beige suspension was formed (hard to stir!). After 30 min a solution of (S)-tetrahydro-3*H*-pyrrolo[1,2-*c*][1,2,3]oxathiazole 1,1-dioxide **29** (4.88 g, 30 mmol) in THF (15 mL) was added keeping the internal temperature below -70°C. The cooling bath was removed and the mixture returned slowly to RT where all solids went back into solution. The brown solution was stirred for 1 h at RT and the volatiles were removed *in vacuo*. The residue was dissolved in 2 M aq. HCl (50 mL) and ethanol (50 mL) and heated to 95°C for 40 h. The reaction mixture was cooled to RT, diluted with H<sub>2</sub>O, and the mixture washed with TBME (1x60 mL). This extract was discarded and the aqueous phase was basified with 5 M aq. NaOH and re-extracted with TBME (3x60 mL). The combined organic extracts were dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and evaporated *in vacuo* to give the title compound as a yellow oil (4.43 g, 67%). LC-MS B: t<sub>R</sub> = 0.37 min; er: >99:1; [M+H]<sup>+</sup> = 222.15; <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ: 1.36-1.45 (m, 1 H), 1.67-1.92 (m, 4 H), 2.67-2.76 (m, 2 H), 2.81-2.87 (m, 1 H), 3.03-3.08 (m, 1 H), 3.18-3.25 (m, 1 H), 3.88 (s, 3 H), 3.89 (s, 3 H), 6.75-6.79 (m, 2 H), 6.80-6.83 (m, 1 H).

### **(S)-2-((2,3-dihydrobenzo[*b*][1,4]dioxin-6-yl)methyl)pyrrolidine 31**

The title compound was prepared from 6-bromo-1,4-benzodioxane (2 mL, 15 mmol) and (S)-tetrahydro-3*H*-pyrrolo[1,2-*c*][1,2,3]oxathiazole 1,1-dioxide **29** (2.0 g, 12 mmol) in analogy to the procedure described for **30**. The product was isolated as a brown oil (1.2 g, 45%). LC-MS A: t<sub>R</sub> = 0.51 min; [M+H]<sup>+</sup> = 220.16.

### **(S)-2-(benzo[*d*][1,3]dioxol-5-ylmethyl)pyrrolidine 32**

The title compound was prepared from 4-bromo-1,2-(methylenedioxy)benzene (1.8 mL, 15 mmol) and (*S*)-tetrahydro-3*H*-pyrrolo[1,2-*c*][1,2,3]oxathiazole 1,1-dioxide **29** (2.0 g, 12 mmol) in analogy to the procedure described for **30**. The product was isolated as a brown oil (1.86 g, 74%). LC-MS A:  $t_R = 0.51$  min;  $[M+H]^+ = 206.18$ .

#### **(*S*)-2-(3,5-dimethoxybenzyl)pyrrolidine 33**

The title compound was prepared from 1-bromo-3,5-dimethoxybenzene (3.2 g, 15 mmol) and (*S*)-tetrahydro-3*H*-pyrrolo[1,2-*c*][1,2,3]oxathiazole 1,1-dioxide **29** (2.0 g, 12 mmol) in analogy to the procedure described for **30**. The product was isolated as a brown oil (0.51 g, 19%). LC-MS B:  $t_R = 0.45$  min;  $[M+H]^+ = 222.25$ .

#### **(*S*)-2-(3-chloro-4-methoxybenzyl)pyrrolidine 34**

The title compound was prepared from 4-bromo-2-chloro-1-methoxybenzene (2.2 g, 10 mmol) and (*S*)-tetrahydro-3*H*-pyrrolo[1,2-*c*][1,2,3]oxathiazole 1,1-dioxide **29** (1.34 g, 8.2 mmol) in analogy to the procedure described for **30**. The product was isolated as a yellow oil (0.51 g, 28%). LC-MS A:  $t_R = 0.54$  min;  $[M+H]^+ = 226.2$ .

#### **(*S*)-2-(2-chloro-3-methoxybenzyl)pyrrolidine 35**

The title compound was prepared from 1-chloro-2-methoxybenzene (1.9 g, 13.5 mmol) and (*S*)-tetrahydro-3*H*-pyrrolo[1,2-*c*][1,2,3]oxathiazole 1,1-dioxide **29** (2.0 g, 12 mmol) in analogy to the procedure described for **30**. The product was isolated as an orange oil (1.1 g, 38%). LC-MS A:  $t_R = 0.55$  min;  $[M+H]^+ = 226.15$ .

#### **(*S*)-2-(2-chloro-5-methoxybenzyl)pyrrolidine 36**

The title compound was prepared from 2-bromo-1-chloro-4-methoxybenzene (2.2 g, 10 mmol) and (*S*)-tetrahydro-3*H*-pyrrolo[1,2-*c*][1,2,3]oxathiazole 1,1-dioxide **29** (1.35 g, 8.3 mmol) in analogy to the procedure described for **30**. The product was isolated as a yellow oil (0.6 g, 32%). LC-MS A:  $t_R = 0.57$  min;  $[M+H]^+ = 225.98$ .

#### **(*S*)-2-(3-chloro-4-methylbenzyl)pyrrolidine 37**

The title compound was prepared from 4-bromo-2-chloro-1-methylbenzene (3.0 g, 15 mmol) and (*S*)-tetrahydro-3*H*-pyrrolo[1,2-*c*][1,2,3]oxathiazole 1,1-dioxide **29** (2.0 g, 12 mmol) in analogy to the procedure described for **30**. The product was isolated as a brown oil (1.37 g, 27%). LC-MS A:  $t_R = 0.60$  min;  $[M+H]^+ = 210.11$ .

### **(S)-2-(3,4-dichlorobenzyl)pyrrolidine 38**

The title compound was prepared from 1,2-dichloro-4-iodobenzene (4.1 g, 15 mmol) and (S)-tetrahydro-3H-pyrrolo[1,2-c][1,2,3]oxathiazole 1,1-dioxide **29** (2.0 g, 12 mmol) in analogy to the procedure described for **30**. The product was isolated as a yellow oil (0.44 g, 16%). LC-MS A:  $t_R = 0.59$  min;  $[M+H]^+ = 230.14$ .

### **5-(benzo[d][1,3]dioxol-5-yl)-2-methylthiazole-4-carboxylic acid 39**

**Step 1:** Methyl 5-bromo-2-methylthiazole-4-carboxylate (0.73 g, 3.1 mmol) and 1,3-benzodioxole-5-boronic acid (0.5 g, 3.1 mmol) were dissolved in a mixture of EtOH (20 mL) and toluene (20 mL). Freshly prepared 2 M aq.  $\text{Na}_2\text{CO}_3$  soln. (31 mL) was added and the mixture was degassed by bubbling  $\text{N}_2$  through the mixture for 30 s. Tetrakis(triphenylphosphine) palladium (129 mg, 0.11 mmol) was added quickly and the reaction mixture was heated to reflux for 1 h. The mixture was allowed to cool to RT and some water was added before it was filtered over a celite plug washing with additional water and EtOH. The EtOH was evaporated *in vacuo* and the remaining phases were extracted with DCM (3x). The combined organic extracts were dried over  $\text{Na}_2\text{SO}_4$ , filtered, and evaporated *in vacuo*. The crude product was purified by flash chromatography (eluting with a gradient of 10% to 50% EtOAc in hexane) to give the methyl ester of the title compound as a pale yellow solid (0.55 g, 64%, some transesterification to the ethyl ester was observed). LC-MS B:  $t_R = 0.67$  min;  $[M+H]^+ = 278.14$ .

**Step 2:** The ester from above (0.55 g, 2 mmol) was partitioned between MeOH (30 mL) and 2 M aq. NaOH soln. (15 mL) and heated to a gentle reflux (67°C) overnight. The MeOH was evaporated *in vacuo* and the remaining aqueous layer acidified with 1M aq. HCl before being extracted with DCM (3x). The combined organic extracts were dried over  $\text{Na}_2\text{SO}_4$ , filtered, and evaporated *in vacuo* to give the title compound as a pink solid that was used further without purification (0.46 g, 88%). LC-MS B:  $t_R = 0.58$  min;  $[M+H]^+ = 264.12$ ;  $^1\text{H NMR}$  ( $d_6$ -DMSO)  $\delta$ : 2.65 (s, 3 H), 6.09 (s, 2 H), 6.93-6.99 (m, 2 H), 7.07 (d,  $J = 1.2$  Hz, 1 H), 12.82 (s br, 1 H).

### **5-(2,3-dihydrobenzofuran-5-yl)-2-methylthiazole-4-carboxylic acid 40**

The title compound was prepared from methyl 5-bromo-2-methylthiazole-4-carboxylate (0.48 g, 2 mmol) and (2,3-dihydrobenzofuran-5-yl)boronic acid (0.33 g, 2 mmol) in analogy to the two-step sequence described for **39**. The title compound

was isolated as a white solid (0.29 g, 82%). LC-MS B:  $t_R = 0.59$  min;  $[M+H]^+ = 262.23$ ;  $^1H$  NMR ( $d_6$ -DMSO)  $\delta$ : 2.65 (s, 3 H), 3.17-3.23 (m, 2H), 4.55-4.60 (m, 2 H), 6.78-6.82 (m, 1 H), 7.18-7.22 (m, 1 H), 7.34 (s br, 1 H), 12.75 (s br, 1 H).

**(S)-(2-(3,4-dimethoxybenzyl)pyrrolidin-1-yl)(5-methyl-2-(2H-1,2,3-triazol-2-yl)phenyl)methanone 7**

The title compound was prepared from 5-methyl-2-(2H-1,2,3-triazol-2-yl)benzoic acid (prepared in analogy to the procedure described in WO2008/069997, p28) (184 mg, 0.90 mmol) and (S)-2-(3,4-dimethoxybenzyl)pyrrolidine **30** (200 mg, 0.90 mmol) in analogy to the procedure described for **2**. The product was isolated as a yellow solid (160 mg, 44%). LC-MS D:  $t_R = 0.89$  min;  $[M+H]^+ = 407.23$ ; er: >99:1; LC-HRMS:  $t_R = 1.21$  min;  $[M+H]/z = 407.2083$ , found = 407.2084.

**(S)-(2-(2H-1,2,3-triazol-2-yl)phenyl)(2-(3,4-dimethoxybenzyl)pyrrolidin-1-yl)methanone 8**

The title compound was prepared from 2-(2H-1,2,3-triazol-2-yl)benzoic acid (256 mg, 1.36 mmol) and (S)-2-(3,4-dimethoxybenzyl)pyrrolidine **30** (300 mg, 1.36 mmol) in analogy to the procedure described for **2**. The product was isolated as a cream solid (398 mg, 75%). LC-MS D:  $t_R = 0.85$  min;  $[M+H]^+ = 393.05$ ; LC-HRMS:  $t_R = 1.15$  min;  $[M+H]/z = 393.1927$ , found = 393.1931.

**(S)-(2-(3,4-dimethoxybenzyl)pyrrolidin-1-yl)(5-methyl-2-(1H-pyrazol-1-yl)phenyl)methanone 9**

The title compound was prepared from 5-methyl-2-(1H-pyrazol-1-yl)benzoic acid (prepared in analogy to the procedure described in WO2014/057435, p92) (12 mg, 0.06 mmol) and (S)-2-(3,4-dimethoxybenzyl)pyrrolidine **30** (14 mg, 0.06 mmol) in analogy to the procedure described for **2**. The product was isolated as a white solid (7 mg, 24%). LC-MS D:  $t_R = 0.88$  min;  $[M+H]^+ = 406.17$ ; LC-HRMS:  $t_R = 1.20$  min;  $[M+H]/z = 406.2131$ , found = 406.2136.

**(S)-(2-(3,4-dimethoxybenzyl)pyrrolidin-1-yl)(2-(pyrimidin-2-yl)phenyl)methanone 10**

The title compound was prepared from 2-(pyrimidin-2-yl)benzoic acid (0.91 g, 4.5 mmol) and (S)-2-(3,4-dimethoxybenzyl)pyrrolidine **30** (1.0 g, 4.5 mmol) in analogy to the procedure described for **2**. The product was isolated as a white solid (0.56 g,

31%). LC-MS D:  $t_R = 0.83$  min;  $[M+H]^+ = 404.07$ ; LC-HRMS:  $t_R = 1.13$  min;  $[M+H]/z = 404.1974$ , found = 404.1980.

**(S)-2-(3,4-dimethoxybenzyl)pyrrolidin-1-yl(6-methyl-3-(pyrimidin-2-yl)pyridin-2-yl)methanone 11**

The title compound was prepared from 6-methyl-3-(pyrimidin-2-yl)picolinic acid (prepared in analogy to the procedure described in WO2010/063662, p31) (15 mg, 0.07 mmol) and (S)-2-(3,4-dimethoxybenzyl)pyrrolidine **30** (15 mg, 0.07 mmol) in analogy to the procedure described for **2**. The product was isolated as a white solid (10 mg, 36%). LC-MS D:  $t_R = 0.79$  min;  $[M+H]^+ = 419.13$ ; LC-HRMS:  $t_R = 1.11$  min;  $[M+H]/z = 419.2083$ , found = 419.2082.

**(S)-2-(3,4-dimethoxybenzyl)pyrrolidin-1-yl(3-(*m*-tolyl)pyrazin-2-yl)methanone 12**

The title compound was prepared from 3-(*m*-tolyl)pyrazine-2-carboxylic acid (prepared in analogy to the procedure described in WO2010/38200, p48) (214 mg, 1.0 mmol) and (S)-2-(3,4-dimethoxybenzyl)pyrrolidine **30** (221 mg, 1.0 mmol) in analogy to the procedure described for **2**. The product was isolated as a cream solid (331 mg, 79%). LC-MS D:  $t_R = 0.88$  min;  $[M+H]^+ = 418.16$ ; LC-HRMS:  $t_R = 1.24$  min;  $[M+H]/z = 418.2131$ , found = 418.2135.

**(S)-2-(3,4-dimethoxybenzyl)pyrrolidin-1-yl(2-methyl-5-(*m*-tolyl)thiazol-4-yl)methanone 13**

The title compound was prepared from 2-methyl-5-(*m*-tolyl)thiazole-4-carboxylic acid (211 mg, 0.90 mmol) and (S)-2-(3,4-dimethoxybenzyl)pyrrolidine **30** (200 mg, 0.90 mmol) in analogy to the procedure described for **2**. The product was isolated as a yellow solid (167 mg, 42%). LC-MS D:  $t_R = 0.95$  min;  $[M+H]^+ = 437.2$ ; LC-HRMS:  $t_R = 1.33$  min;  $[M+H]/z = 437.1899$ , found = 437.1900.

**(S)-2-(3,4-dimethoxybenzyl)pyrrolidin-1-yl(2-(3-methyl-1,2,4-oxadiazol-5-yl)phenyl)methanone 14**

The title compound was prepared from 2-(3-methyl-1,2,4-oxadiazol-5-yl)benzoic acid (281 mg, 1.38 mmol) and (S)-2-(3,4-dimethoxybenzyl)pyrrolidine **30** (305 mg, 1.38 mmol) in analogy to the procedure described for **2**. The product was isolated as a yellow solid (403 mg, 72%). LC-MS D:  $t_R = 0.85$  min;  $[M+H]^+ = 408.1$ ; LC-HRMS:  $t_R = 1.16$  min;  $[M+H]/z = 408.1923$ , found = 408.1931.

**(S)-2-((2,3-dihydrobenzo[*b*][1,4]dioxin-6-yl)methyl)pyrrolidin-1-yl)(2-methyl-5-(*m*-tolyl)thiazol-4-yl)methanone 15**

The title compound was prepared from 2-methyl-5-(*m*-tolyl)thiazole-4-carboxylic acid (23 mg, 0.1 mmol) and (S)-2-((2,3-dihydrobenzo[*b*][1,4]dioxin-6-yl)methyl)pyrrolidine **31** (22 mg, 0.1 mmol) in analogy to the procedure described for **2**. The product was isolated as a white solid (20 mg, 47%). LC-MS D:  $t_R = 0.98$  min;  $[M+H]^+ = 435.07$ ; LC-HRMS:  $t_R = 1.37$  min;  $[M+H]/z = 435.1742$ , found = 435.1750.

**(S)-2-(benzo[*d*][1,3]dioxol-5-ylmethyl)pyrrolidin-1-yl)(2-methyl-5-(*m*-tolyl)thiazol-4-yl)methanone 16**

The title compound was prepared from 2-methyl-5-(*m*-tolyl)thiazole-4-carboxylic acid (23 mg, 0.1 mmol) and (S)-2-(benzo[*d*][1,3]dioxol-5-ylmethyl)pyrrolidine **32** (21 mg, 0.1 mmol) in analogy to the procedure described for **2**. The product was isolated as a white solid (29 mg, 69%). LC-MS D:  $t_R = 0.99$  min;  $[M+H]^+ = 421.08$ ; LC-HRMS:  $t_R = 1.39$  min;  $[M+H]/z = 421.1586$ , found = 421.1588.

**(S)-2-(3,5-dimethoxybenzyl)pyrrolidin-1-yl)(2-methyl-5-(*m*-tolyl)thiazol-4-yl)methanone 17**

The title compound was prepared from 2-methyl-5-(*m*-tolyl)thiazole-4-carboxylic acid (211 mg, 0.9 mmol) and (S)-2-(3,5-dimethoxybenzyl)pyrrolidine **33** (200 mg, 0.9 mmol) in analogy to the procedure described for **2**. The product was isolated as a yellow solid (290 mg, 74%). LC-MS D:  $t_R = 1.0$  min;  $[M+H]^+ = 437.11$ ; LC-HRMS:  $t_R = 1.41$  min;  $[M+H]/z = 437.1899$ , found = 437.1899.

**(S)-2-(3-chloro-4-methoxybenzyl)pyrrolidin-1-yl)(2-methyl-5-(*m*-tolyl)thiazol-4-yl)methanone 18**

The title compound was prepared from 2-methyl-5-(*m*-tolyl)thiazole-4-carboxylic acid (200 mg, 0.89 mmol) and (S)-2-(3-chloro-4-methoxybenzyl)pyrrolidine **34** (200 mg, 0.89 mmol) in analogy to the procedure described for **2**. The product was isolated as a yellow solid (290 mg, 74%). LC-MS D:  $t_R = 1.02$  min;  $[M+H]^+ = 441.07$ ; LC-HRMS:  $t_R = 1.46$  min;  $[M+H]/z = 441.1404$ , found = 441.1412.

**(S)-2-(2-chloro-3-methoxybenzyl)pyrrolidin-1-yl)(2-methyl-5-(*m*-tolyl)thiazol-4-yl)methanone 19**

The title compound was prepared from 2-methyl-5-(*m*-tolyl)thiazole-4-carboxylic acid (23 mg, 0.1 mmol) and (S)-2-(2-chloro-3-methoxybenzyl)pyrrolidine **35** (23 mg, 0.1

mmol) in analogy to the procedure described for **2**. The product was isolated as a brown solid (15 mg, 33%). LC-MS D:  $t_R = 0.96$  min;  $[M+H]^+ = 441.16$ ; LC-HRMS:  $t_R = 1.42$  min;  $[M+H]/z = 441.1404$ , found = 441.1410.

**(S)-(2-(2-chloro-5-methoxybenzyl)pyrrolidin-1-yl)(2-methyl-5-(*m*-tolyl)thiazol-4-yl)methanone 20**

The title compound was prepared from 2-methyl-5-(*m*-tolyl)thiazole-4-carboxylic acid (23 mg, 0.1 mmol) and (S)-2-(2-chloro-5-methoxybenzyl)pyrrolidine **36** (23 mg, 0.1 mmol) in analogy to the procedure described for **2**. The product was isolated as a brown solid (36 mg, 81%). LC-MS D:  $t_R = 1.0$  min;  $[M+H]^+ = 441.16$ ; LC-HRMS:  $t_R = 1.49$  min;  $[M+H]/z = 441.1404$ , found = 441.1410.

**(S)-(2-(3-chloro-4-methylbenzyl)pyrrolidin-1-yl)(2-methyl-5-(*m*-tolyl)thiazol-4-yl)methanone 21**

The title compound was prepared from 2-methyl-5-(*m*-tolyl)thiazole-4-carboxylic acid (23 mg, 0.1 mmol) and (S)-2-(3-chloro-4-methylbenzyl)pyrrolidine **37** (21 mg, 0.1 mmol) in analogy to the procedure described for **2**. The product was isolated as a brown solid (24 mg, 57%). LC-MS D:  $t_R = 1.08$  min;  $[M+H]^+ = 425.06$ ; LC-HRMS:  $t_R = 1.59$  min;  $[M+H]/z = 425.1454$ , found = 425.1460.

**(S)-(2-(3,4-dichlorobenzyl)pyrrolidin-1-yl)(2-methyl-5-(*m*-tolyl)thiazol-4-yl)methanone 22**

The title compound was prepared from 2-methyl-5-(*m*-tolyl)thiazole-4-carboxylic acid (35 mg, 0.15 mmol) and (S)-2-(3,4-dichlorobenzyl)pyrrolidine **38** (35 mg, 0.15 mmol) in analogy to the procedure described for **2**. The product was isolated as a yellow solid (53 mg, 79%). LC-MS D:  $t_R = 1.08$  min;  $[M+H]^+ = 445.03$ ; LC-HRMS:  $t_R = 1.59$  min;  $[M+H]/z = 445.0908$ , found = 445.0910.

**(S)-(2-(3,4-dimethoxybenzyl)pyrrolidin-1-yl)(5-(4-fluorophenyl)-2-methylthiazol-4-yl)methanone 23**

The title compound was prepared from 5-(4-fluorophenyl)-2-methylthiazole-4-carboxylic acid (24 mg, 0.1 mmol) and (S)-2-(3,4-dimethoxybenzyl)pyrrolidine **30** (22 mg, 0.1 mmol) in analogy to the procedure described for **2**. The product was isolated as a yellow oil (27 mg, 60%). LC-MS D:  $t_R = 0.92$  min;  $[M+H]^+ = 441.17$ ; LC-HRMS:  $t_R = 1.28$  min;  $[M+H]/z = 441.1648$ , found = 441.1650.

**(S)-(5-(3-chlorophenyl)-2-methylthiazol-4-yl)(2-(3,4-dimethoxybenzyl)pyrrolidin-1-yl)methanone 24**

The title compound was prepared from 5-(3-chlorophenyl)-2-methylthiazole-4-carboxylic acid (prepared in analogy to the procedure described in WO2009/16560, p63) (25 mg, 0.1 mmol) and (S)-2-(3,4-dimethoxybenzyl)pyrrolidine **30** (22 mg, 0.1 mmol) in analogy to the procedure described for **2**. The product was isolated as a yellow oil (32 mg, 71%). LC-MS D:  $t_R = 0.96$  min;  $[M+H]^+ = 457.02$ ; LC-HRMS:  $t_R = 1.36$  min;  $[M+H]/z = 457.1353$ , found = 457.1361.

**(S)-(2-(3,4-dimethoxybenzyl)pyrrolidin-1-yl)(5-(3-methoxyphenyl)-2-methylthiazol-4-yl)methanone 25**

The title compound was prepared from 5-(3-methoxyphenyl)-2-methylthiazole-4-carboxylic acid (prepared in analogy to the procedure described in WO2010/044054, p82) (25 mg, 0.1 mmol) and (S)-2-(3,4-dimethoxybenzyl)pyrrolidine **30** (22 mg, 0.1 mmol) in analogy to the procedure described for **2**. The product was isolated as a cream solid (28 mg, 62%). LC-MS D:  $t_R = 0.92$  min;  $[M+H]^+ = 453.12$ ; LC-HRMS:  $t_R = 1.26$  min;  $[M+H]/z = 453.1848$ , found = 453.1858.

**(S)-(2-(3,4-dimethoxybenzyl)pyrrolidin-1-yl)(5-(4-methoxyphenyl)-2-methylthiazol-4-yl)methanone 26**

The title compound was prepared from 5-(4-methoxyphenyl)-2-methylthiazole-4-carboxylic acid (prepared in analogy to the procedure described in WO2010/044054, p82) (25 mg, 0.1 mmol) and (S)-2-(3,4-dimethoxybenzyl)pyrrolidine **30** (22 mg, 0.1 mmol) in analogy to the procedure described for **2**. The product was isolated as a cream solid (8 mg, 17%). LC-MS D:  $t_R = 0.91$  min;  $[M+H]^+ = 452.99$ ; LC-HRMS:  $t_R = 1.25$  min;  $[M+H]/z = 453.1848$ , found = 453.1856.

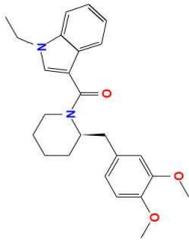
**(S)-(5-(benzo[d][1,3]dioxol-5-yl)-2-methylthiazol-4-yl)(2-(3,4-dimethoxybenzyl)pyrrolidin-1-yl)methanone 27**

The title compound was prepared from 5-(benzo[d][1,3]dioxol-5-yl)-2-methylthiazole-4-carboxylic acid **39** (238 mg, 0.9 mmol) and (S)-2-(3,4-dimethoxybenzyl)pyrrolidine **30** (200 mg, 0.9 mmol) in analogy to the procedure described for **2**. The product was isolated as a white solid (252 mg, 60%). LC-MS D:  $t_R = 0.89$  min;  $[M+H]^+ = 466.99$ ; LC-HRMS:  $t_R = 1.23$  min;  $[M+H]/z = 467.1641$ , found = 467.1652.

**(S)-(5-(2,3-dihydrobenzofuran-5-yl)-2-methylthiazol-4-yl)(2-(3,4-dimethoxybenzyl)pyrrolidin-1-yl)methanone 28**

The title compound was prepared from 5-(2,3-dihydrobenzofuran-5-yl)-2-methylthiazole-4-carboxylic acid **40** (26 mg, 0.1 mmol) and (S)-2-(3,4-dimethoxybenzyl)pyrrolidine **30** (22 mg, 0.1 mmol) in analogy to the procedure described for **2**. The product was isolated as a yellow oil (33 mg, 71%). LC-MS D:  $t_R$  = 0.90 min;  $[M+H]^+$  = 464.98; LC-HRMS:  $t_R$  = 1.25 min;  $[M+H]/z$  = 465.1848, found = 465.1855.

**$^1H$  &  $^{13}C$  NMR spectra of final compounds:**



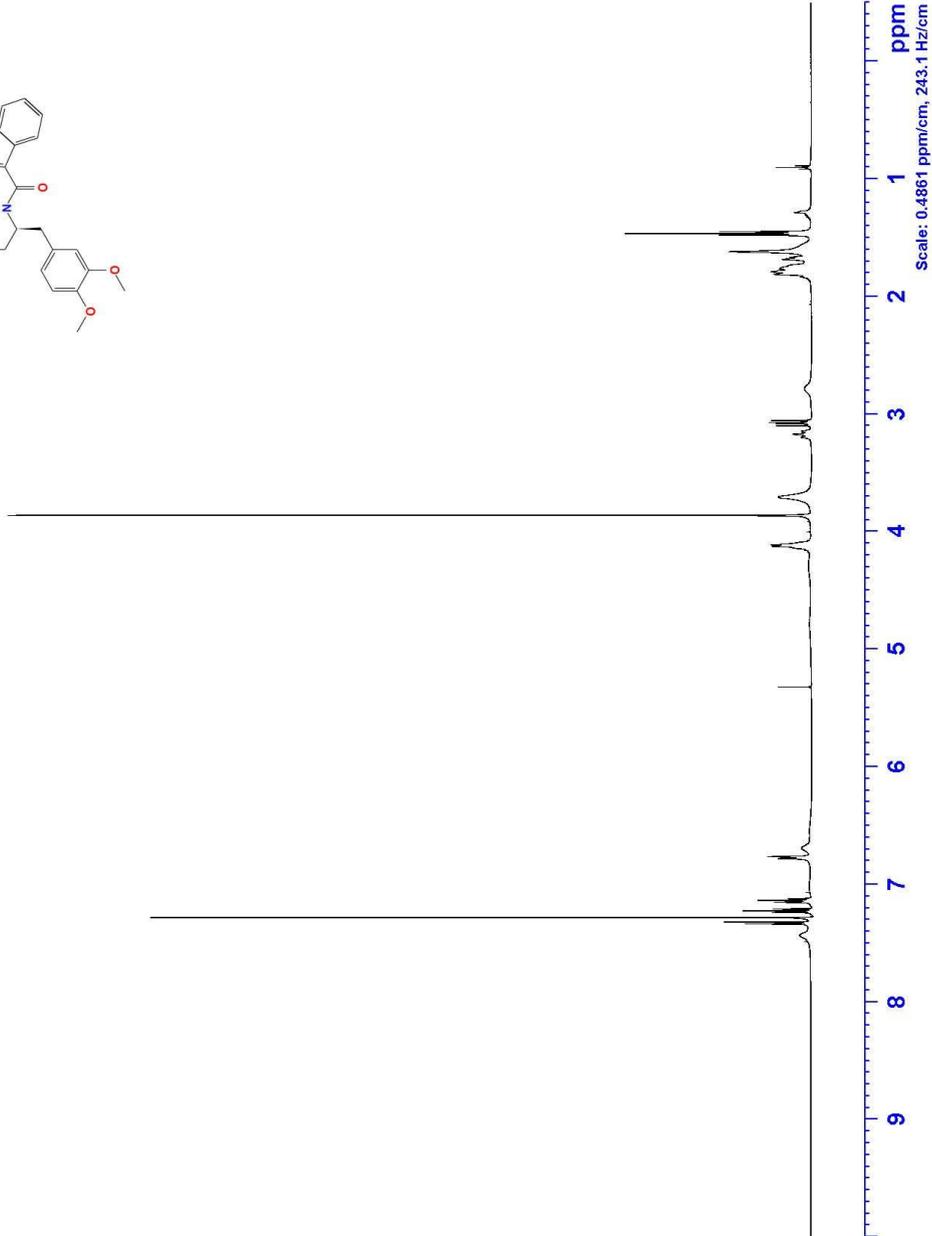
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PROCNO   1

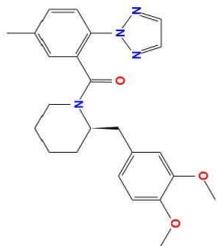
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SOLVENT  CDCl3
NS       16
DS       2
SWH      10000.000 Hz
FIDRES   0.152888 Hz
AQ       3.276793 sec
RG       768.000
AQ       50.000 USEC
DE       40.00 USEC
TE       298.0 K
D1       1.0000000 sec
TD0      1

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SF01    500.1300885 MHz
NUC1     1H
P1       9.25 usec
ELW1    13.0000000 W

F2 - Processing parameters
SI       65536
WDW      EM
SSB      0
GB       0
PC       1.00
  
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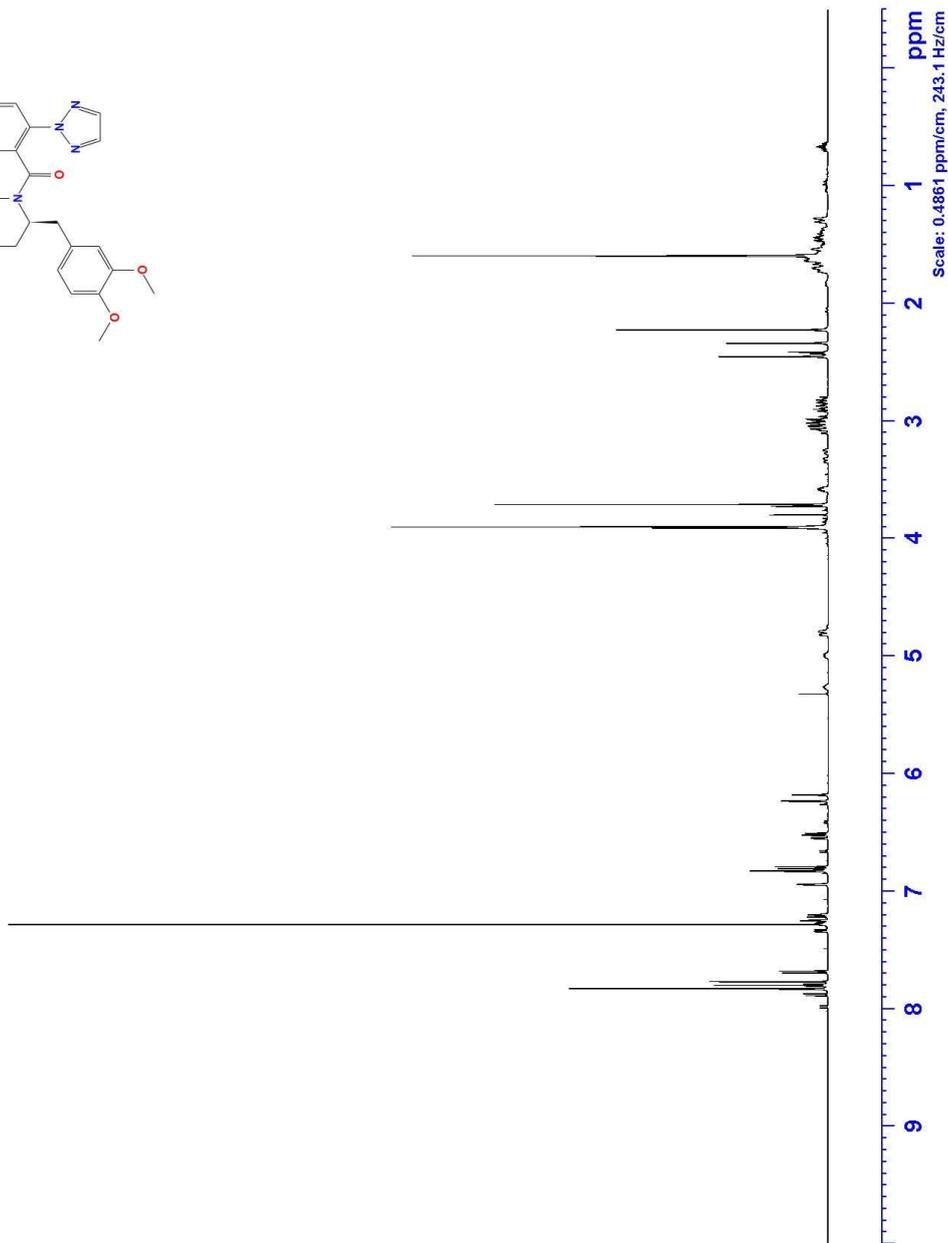


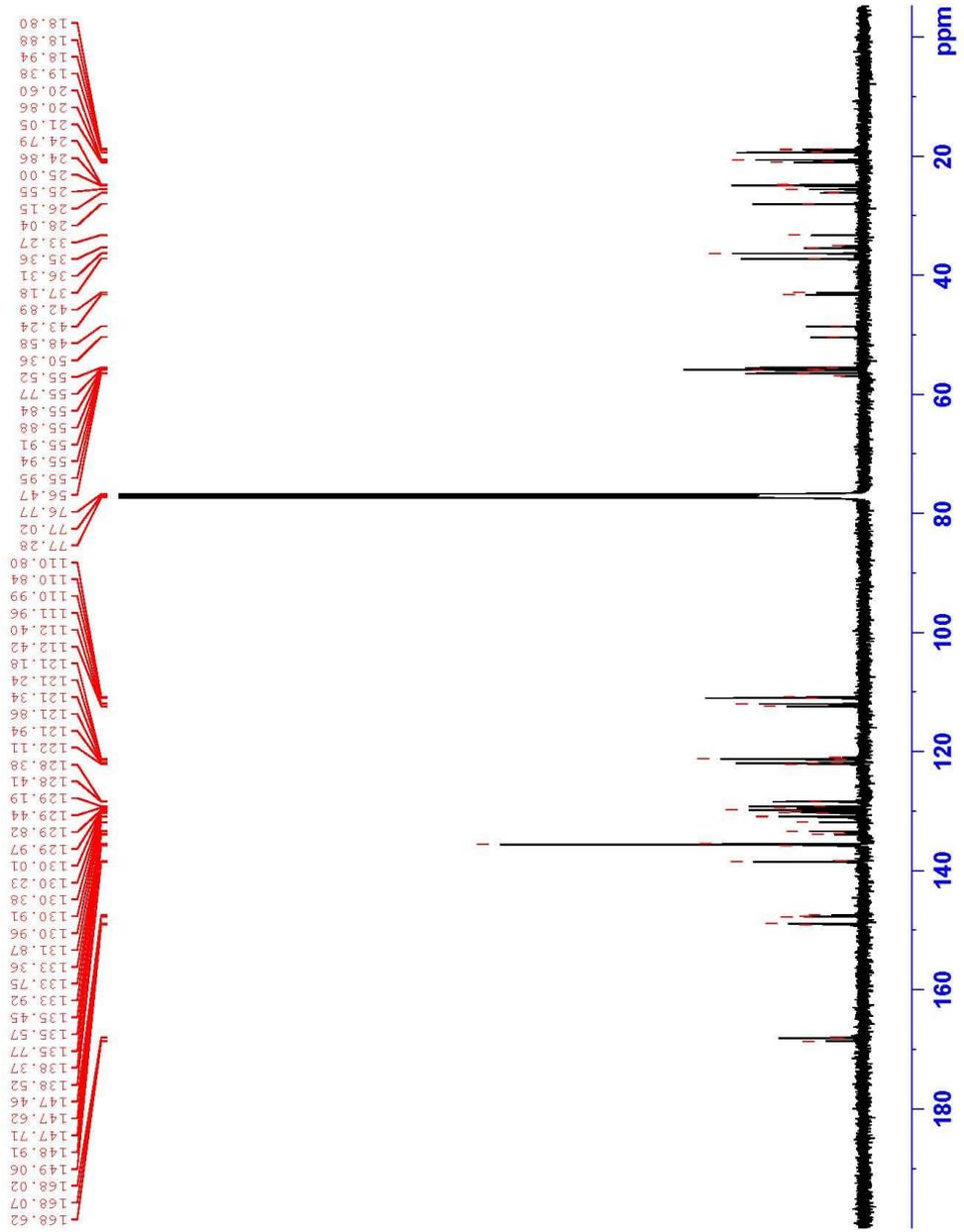
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 EXPNO 10  
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 Time 20.35  
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 PULPROG zgpg30  
 TD 65536  
 CHAN13  
 SOLVENT  
 NS 16  
 DS 2  
 SWH 10000.000 Hz  
 SRH 0.152888 Hz  
 FIDRES 3.276793 SEC  
 AQ 76.000 SEC  
 RG 409.6  
 DW 50.000 USEC  
 DE 40.000 USEC  
 TE 298.0 K  
 D1 1.00000000 sec  
 TDO 1

==== CHANNEL f1 =====  
 SF01 500.1330885 MHz  
 NUC1 1H  
 P1 9.25 usec  
 ELW1 13.0000000 W

F2 - Processing parameters  
 SI 65536  
 WDW EM  
 SSB 0  
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 PC 1.00





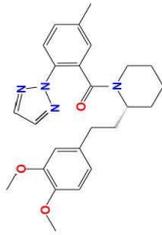
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PROCNO   1

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PULPROG zgpg30
TD        65536
SOLVENT  CDCl3
NS        256
DS        4
SSB       31512.605 Hz
AQ        0.480844 Hz
RG        1.0298378 sec
DE        10.000000 Hz
TE        60.46 usec
TE        298.0 K
D1        3.0000000 sec
D11       0.0300000 sec
TD0       1

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NUC1      13C
P1        11.00 usec
PL1       18.0000000 N

===== CHANNEL f2 =====
SFO2     500.1320005 MHz
NUC2     1H
P2        19.00 usec
PL2       0.0000000 N
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SFO2     13.0000000 MHz
NUC2     13C
P2        13.00 usec
PL2       0.1738000 N
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SFO2     0.1112500 MHz
NUC2     1H
P2        0.1112500 N
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SSB       0
LB        1.00 Hz
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PC        1.40
  
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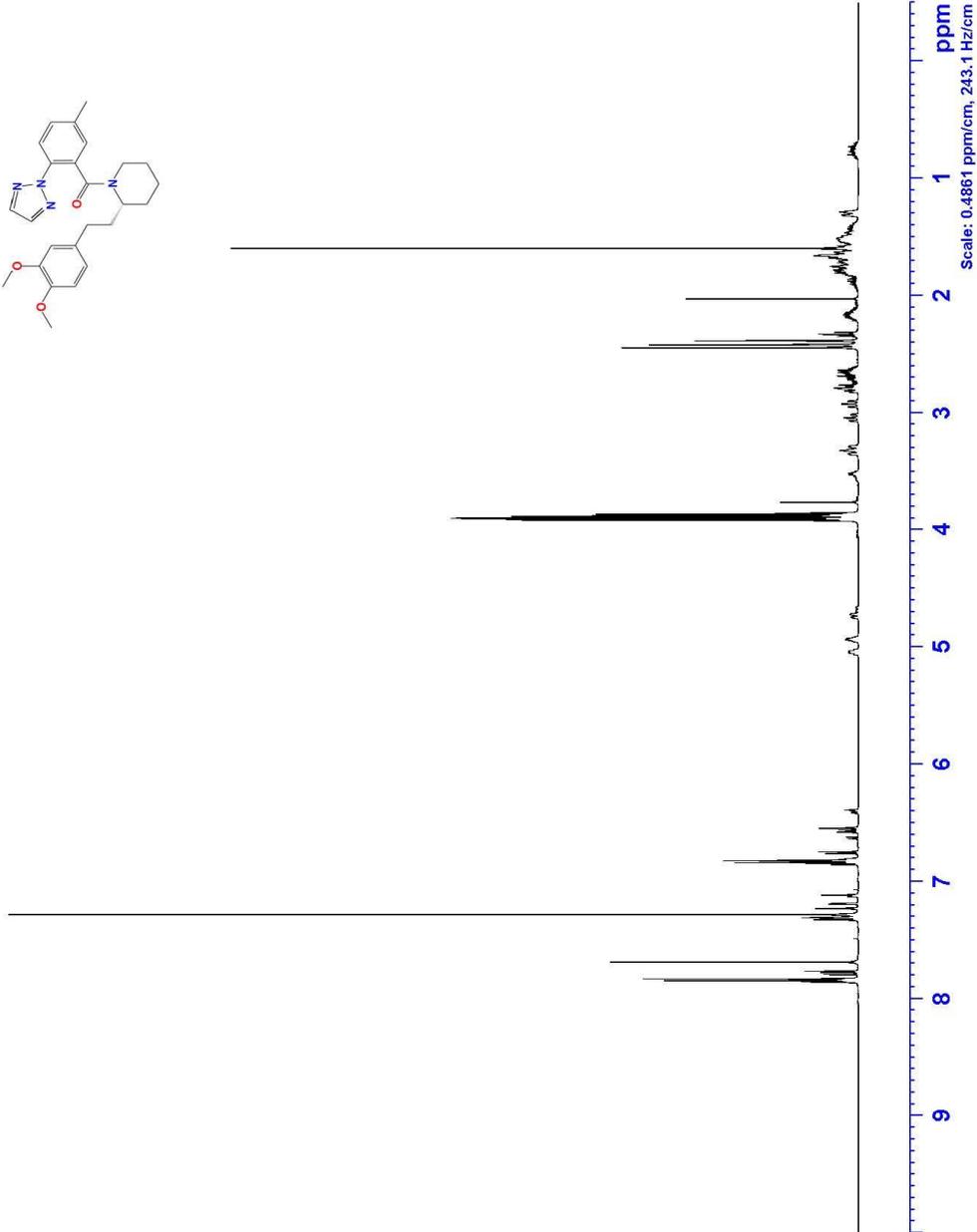
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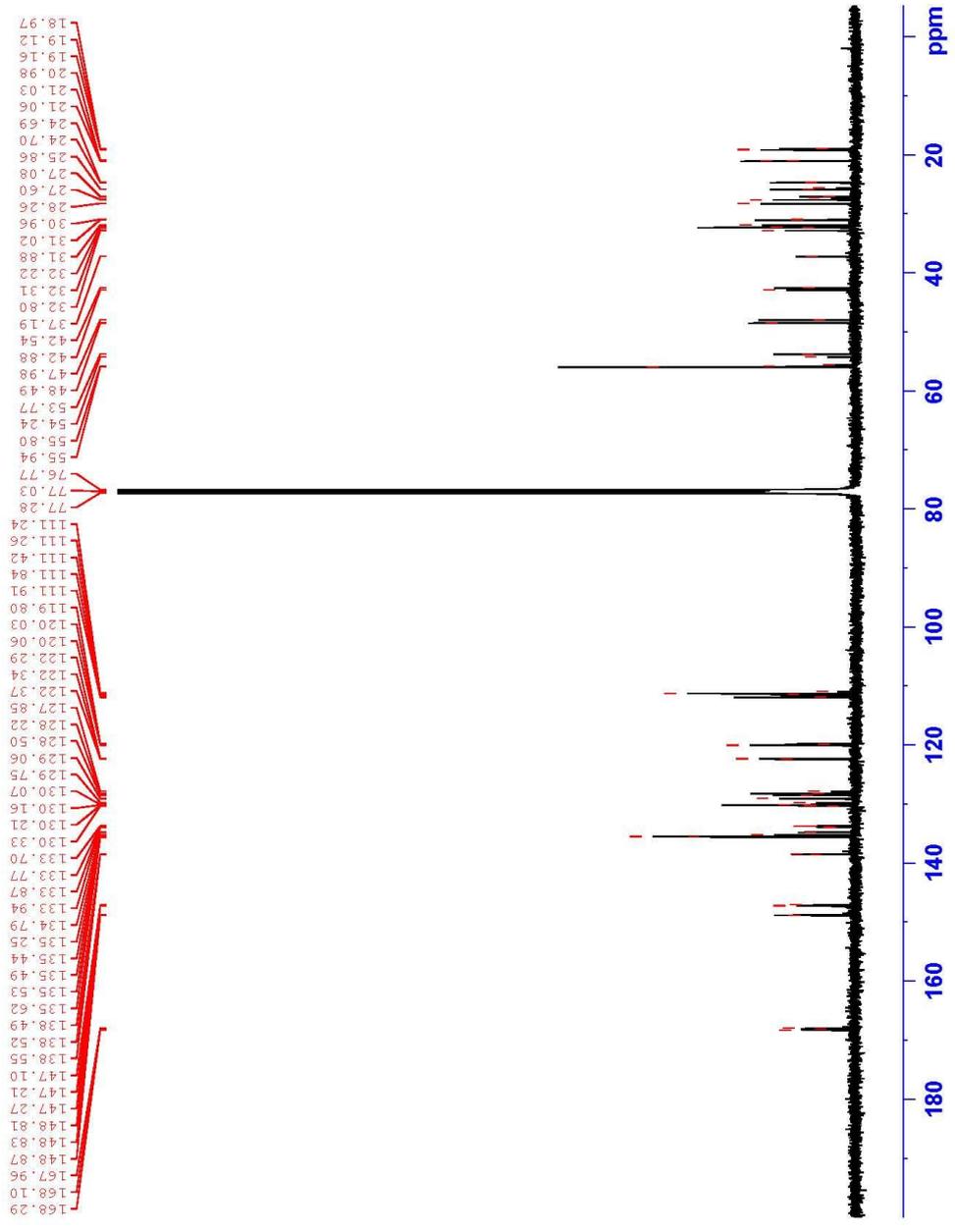
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EXPNO        10
PROCNO       1

F2 - Acquisition Parameters
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Time         20.53
INSTRUM      SPECT
PROBHD       5 mm CPDCH 13C
PULPROG      zgpg30
TD           65536
SOLVENT      CDCl3
NS           16
DS           2
SWH           10000.000 Hz
FIDRES       0.152888 Hz
AQ           3.276793 sec
RG           768.000
AQ           3.276793 sec
RG           768.000
DW           50.000 USEC
DE           40.000 USEC
TE           298.0 K
D1           1.00000000 sec
TD0          1

===== CHANNEL f1 =====
SF01         500.1330885 MHz
NUC1         1H
P1           9.25 usec
ELW1         13.00000000 W

F2 - Processing parameters
SI           65536
WDW          EM
SSB          0
GB           0
PC           1.00
  
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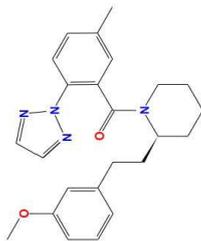
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 Compound: 2  
 PROCNO: 1

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 PROBD: 5 mm CPDCH 13C  
 P1: 11.00 usec  
 SFO2: 500.1320005 MHz  
 SOLVENT: CDCl3  
 NS: 256  
 DS: 4  
 SSB: 31512.605 Hz  
 FIDRES: 0.480844 Hz  
 AQ: 1.0398378 sec  
 RG: 1000  
 IN: 15.667 usec  
 DE: 60.46 usec  
 TE: 298.0 K  
 D1: 3.0000000 sec  
 DI: 0.0300000 sec  
 TD0: 1

===== CHANNEL f1 =====  
 NUC1: 13C  
 P1: 11.00 usec  
 PLW1: 18.00000000 N

===== CHANNEL f2 =====  
 SFO2: 500.1320005 MHz  
 NUC2: 1H  
 P2: 11.00 usec  
 PLW2: 18.00000000 N  
 SFO1: 125.7611000 MHz  
 NUC1: 13C  
 P1: 11.00 usec  
 PLW1: 18.00000000 N

F2 - Processing parameters  
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 SF: 125.7577890 MHz  
 DS: 4  
 SSB: 0  
 LB: 0 1.00 Hz  
 GB: 0  
 PC: 1.40

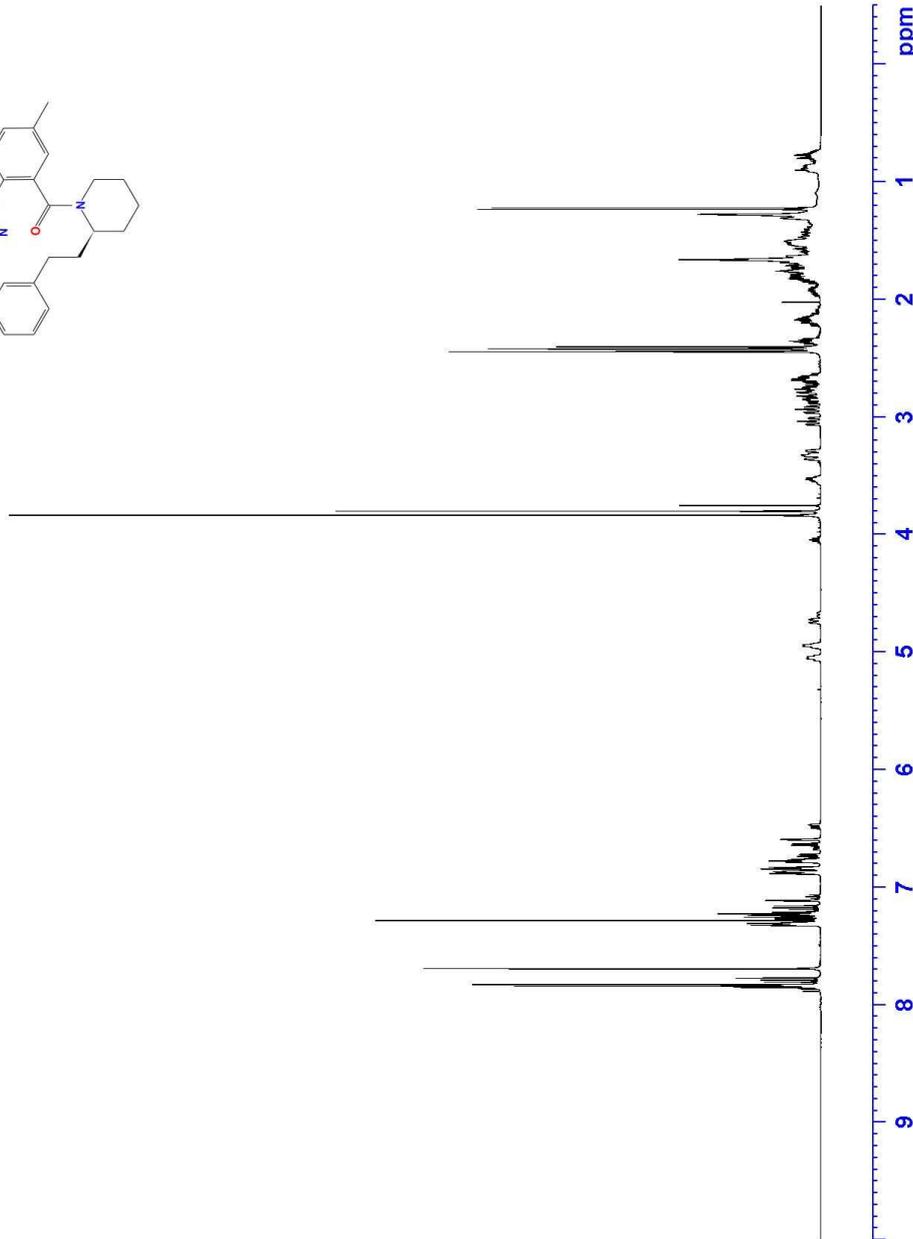


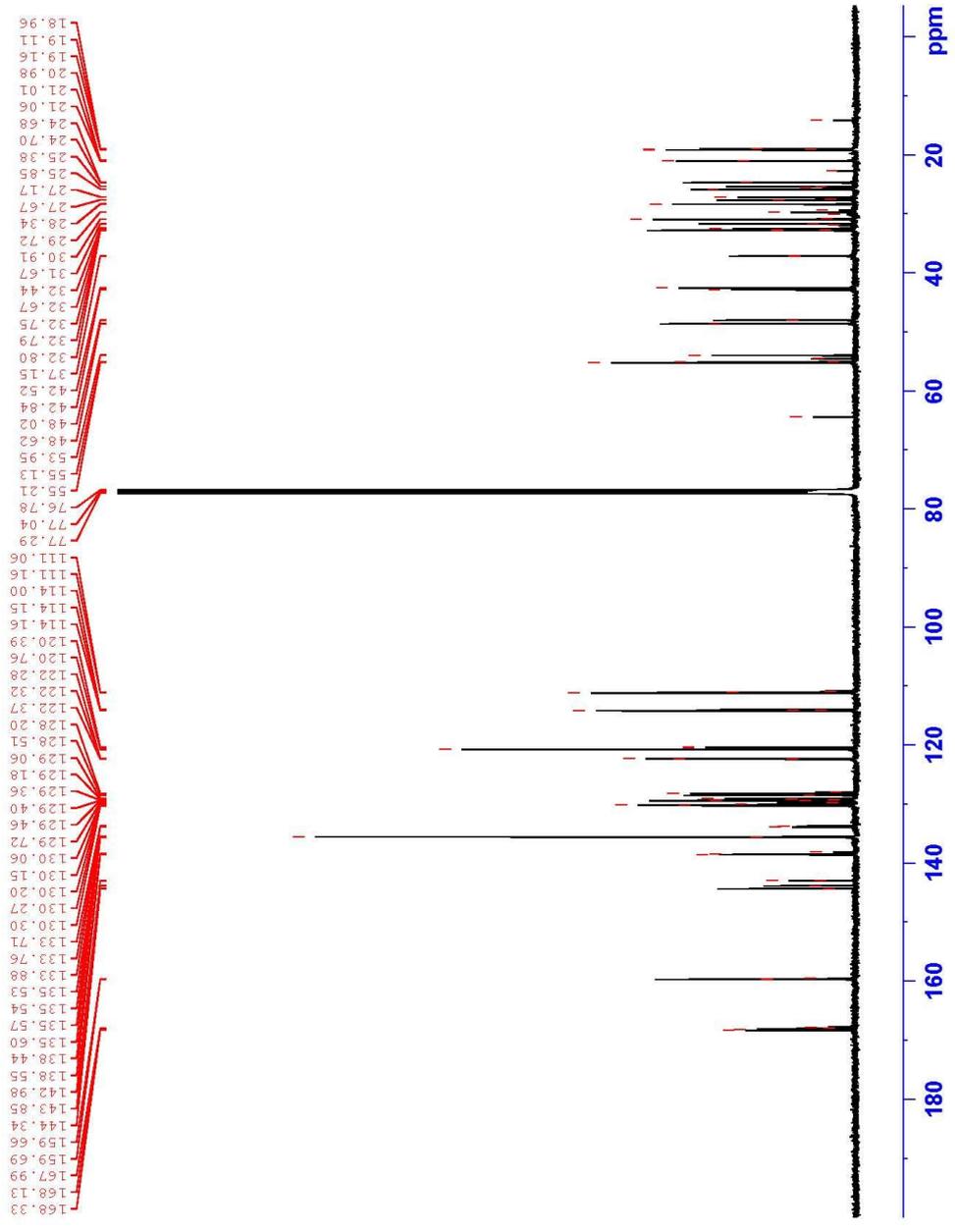
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Current Data Parameters
NAME      Compound 6
EXPNO    10
PROCNO   1

F2 - Acquisition Parameters
Date_    20141113
Time     0.33
INSTRUM  spect
PROBHD   5 mm CPDCH 13C
PULPROG  zgpg30
TD       65536
SOLVENT  CDCl3
NS       16
DS       2
SWH      10000.000 Hz
FIDRES   0.152888 Hz
AQ       3.276797 sec
RG       34.601
WDW      50.000 usec
DE       40.000 usec
TE       298.0 K
D1       1.00000000 sec
TD0      1

===== CHANNEL f1 =====
SFO1     500.1300885 MHz
NUC1     1H
P1       9.25 usec
ELW1    13.00000000 W
F2 - Processing parameters
SI       65536
WDW      0
SSB      0
GB       0
PC       0
Scale: 0.4861 ppm/cm, 243.1 Hz/cm
  
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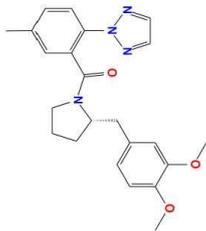
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Name      Compound
EXPNO    2
PROCNO   1

F2 - Acquisition Parameters
Time     20.10
Date_    0.48
INSTRUM  spect
PROBHD   5 mm CPDCH 13C
PULPROG  zgpg30
TD       65536
SOLVENT  CDCl3
NS       256
DS       4
SFO      31512.605 Hz
AQ       0.480844 Hz
FIDRES   1.0298378 sec
RG       10.000000
DE       15.667 usec
TE       60.46 usec
TE       298.0 K
D1       2.0000000 sec
D11      0.0300000 sec
TD0      1

===== CHANNEL f1 =====
NUC1      13C
P1       11.00 usec
PL1      18.00000000 N

===== CHANNEL f2 =====
SFO2     500.1320005 MHz
NUC2     1H
P2       11.00 usec
PL2      0.00000000 N
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PROC2    waltz16
NUC2     13C
PL2      13.00000000 N
PLM2     0.17380001 N
PLM3     0.11125000 N

F2 - Processing parameters
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SSB      0
LB       1.00 Hz
GB       0
PC       1.40
  
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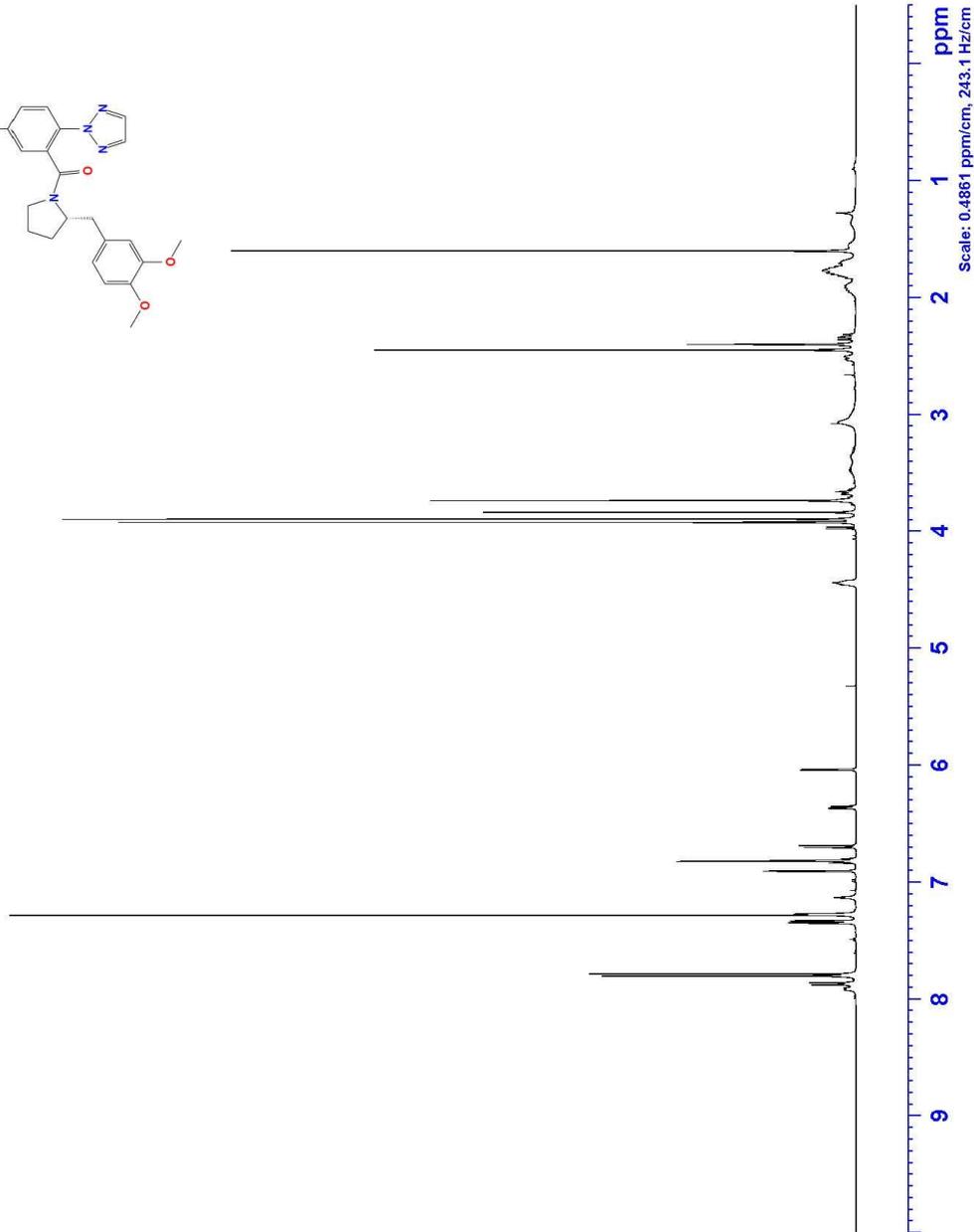


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Current Data Parameters
NAME          Compound 7
EXPNO        10
PROCNO       1

F2 - Acquisition Parameters
Date_        20141117
Time         20.17
INSTRUM     spect
PROBHD      5 mm CPDCH 13C
PULPROG     zgpg30
TD          65536
SOLVENT     CDCl3
NS          16
DS          2
SWH         10000.000 Hz
FIDRES     0.152888 Hz
AQ         3.276793 sec
RG         327.68
WDW         50.000 usec
DE         40.000 usec
TE         298.0 K
D1         1.00000000 sec
TD0        1

===== CHANNEL f1 =====
SF01      500.1300885 MHz
NUC1       1H
P1         9.25 usec
ELW1      13.00000000 W
F2 - Processing parameters
SI         65536
WDW        EM
SSB        0
GB         0
PC         1.00
  
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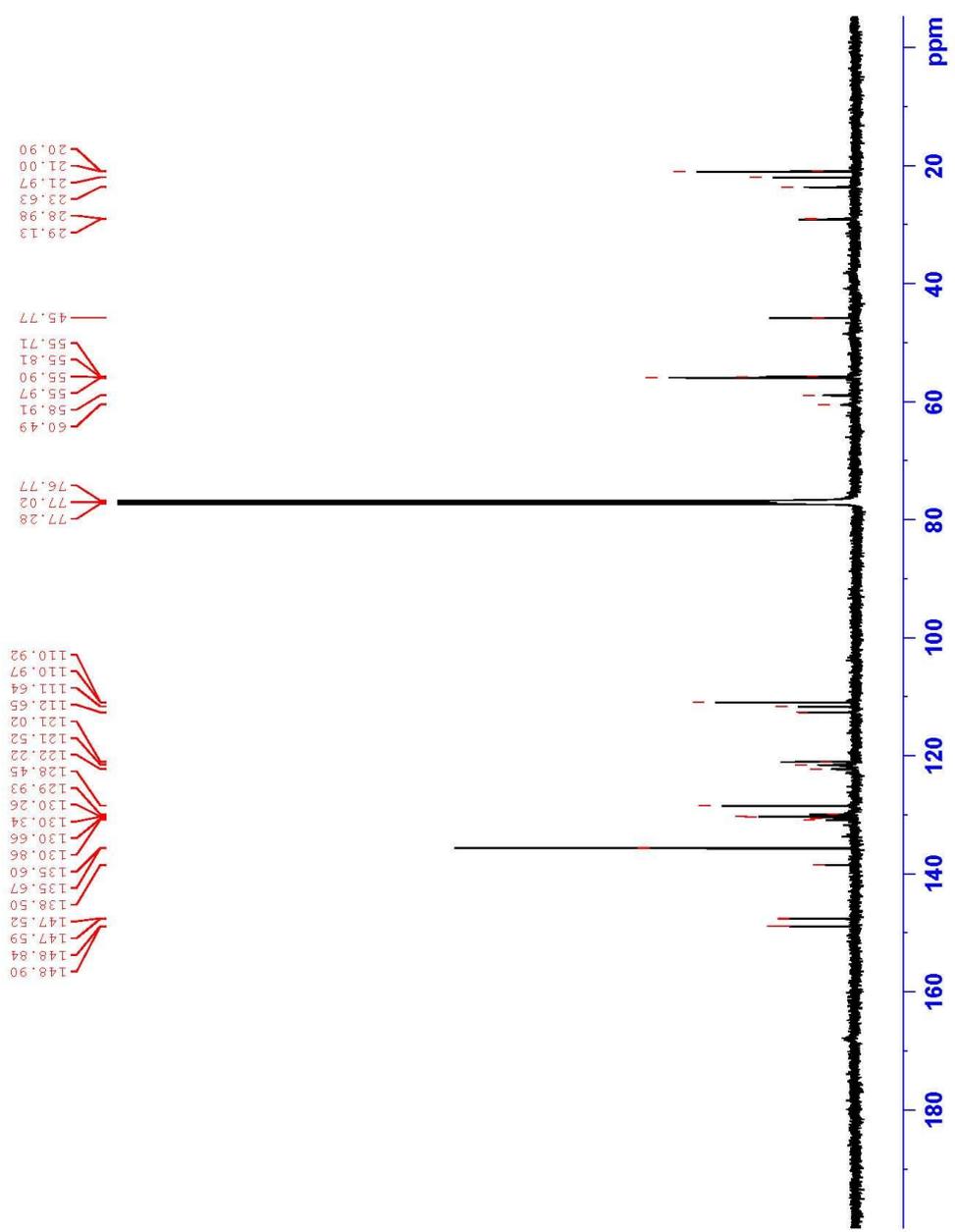
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Name          Compound
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PROCNO       1

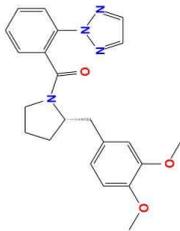
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INSTRUM      spect
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PULPROG      zgpg30
TD           65536
SOLVENT      CDCl3
NS           256
DS           4
SWH           31512.605 Hz
FIDRES       0.480844 Hz
AQ           1.0298378 sec
RG           106.667
DE           15.667 usec
TE           60.46 usec
TD0          258.0 K
D1           2.0000000 sec
D11          0.0300000 sec
TD0          1

----- CHANNEL f1 -----
NUC1         13C
P1           11.00 usec
PL1         18.00000000 N

----- CHANNEL f2 -----
SFO2         500.1320005 MHz
NUC2         1H
P2           12.00 usec
PL2         0.00000000 N
SFO1         125.7678900 MHz
NUC1         13C
P1           11.00 usec
PL1         18.00000000 N

----- Processing parameters -----
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SF           125.7678900 MHz
WDW          EM
SSB          0
LB           1.00 Hz
GB           0
PC           1.40
  
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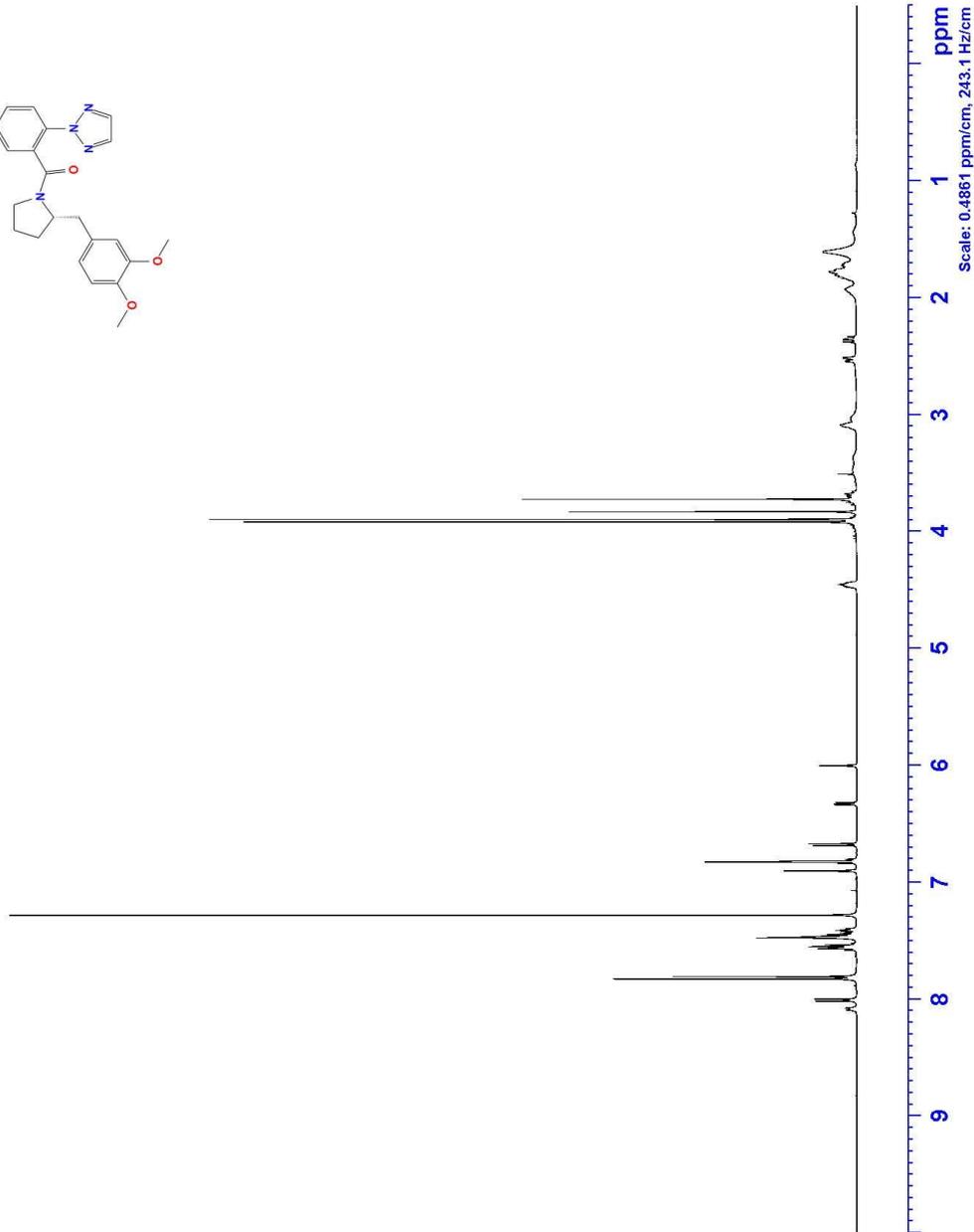
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Current Data Parameters
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EXPNO    10
PROCNO   1

F2 - Acquisition Parameters
Date_    20141111
Time     0.15
INSTRUM  SPECT
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PULPROG  zg30
TD       65536
SOLVENT  CDCl3
NS       16
DS       2
SWH      10000.000 Hz
FIDRES   0.152888 Hz
AQ       3.276793 sec
RG       768.000
AQ       50.000 usec
DE       40.000 usec
TE       298.0 K
D1       1.00000000 sec
TD0      1

===== CHANNEL f1 =====
SF01     500.1330885 MHz
NUC1      1H
P1       9.25 usec
ELW1     13.00000000 W

F2 - Processing parameters
SI       65536
WDW      EM
SSB      0
GB       0
PC       1.00
  
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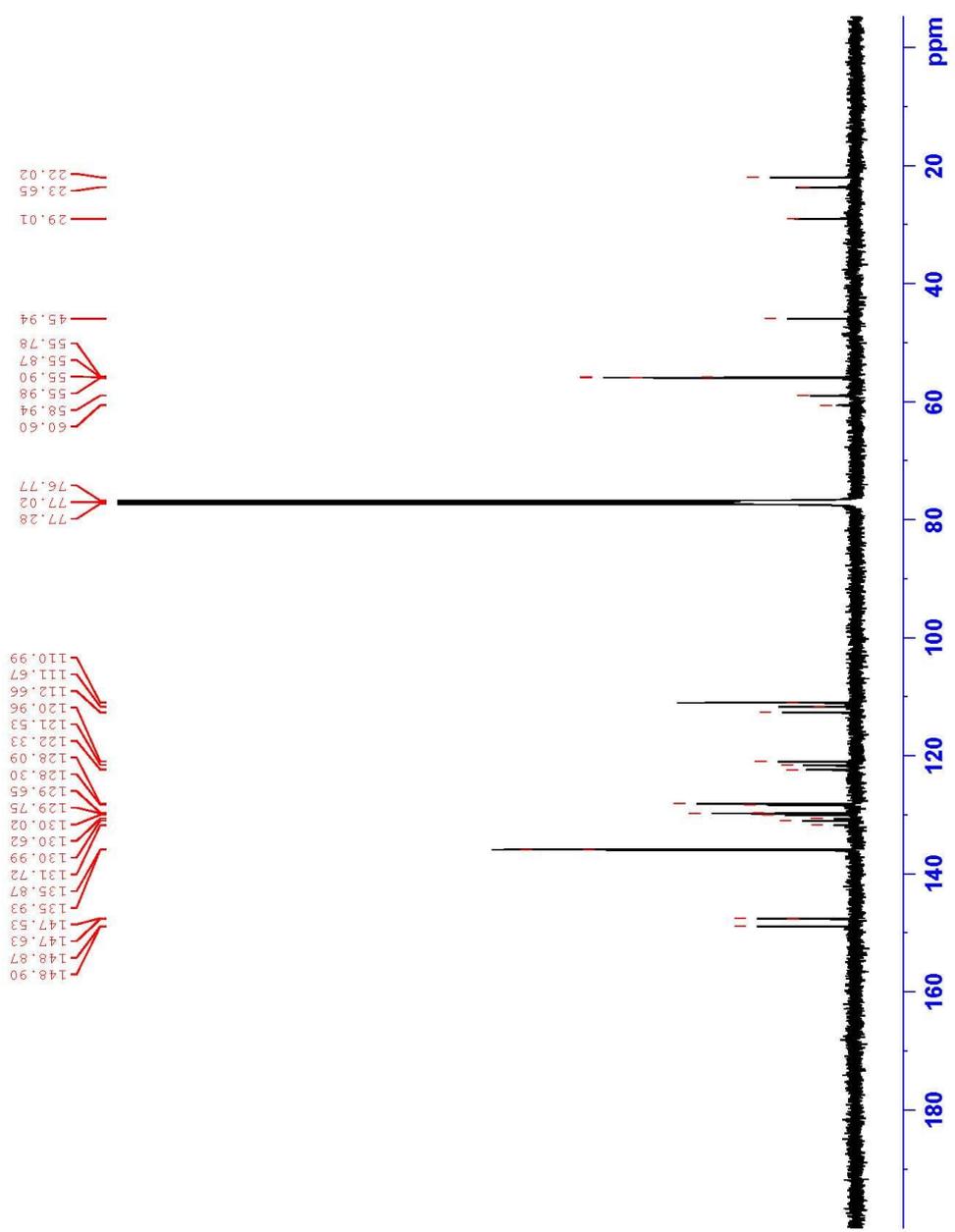
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EXPNO    2
PROCNO   1

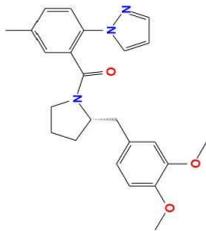
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Date_    0.29
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TD        65536
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NS        256
DS        4
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FIDRES   0.480844 Hz
AQ        1.0298378 sec
RG         10.000
DE         15.697 usec
TE         60.46 usec
TD0       258.0 K
D1         2.0000000 sec
D11        0.0300000 sec
TD0        1

===== CHANNEL f1 =====
NUC1      13C
P1        11.00 usec
PL1       18.00000000 N

===== CHANNEL f2 =====
SFO2     500.1320005 MHz
NUC2      1H
P2        13.00000000 usec
PL2       0.00000000 N
PLM2     0.17380001 N
PLM3     0.11125000 N

F2 - Processing parameters
SI        32768
SF        125.7577890 MHz
WDW       EM
SSB       0
LB        1.00 Hz
GB        0
PC        1.40
  
```



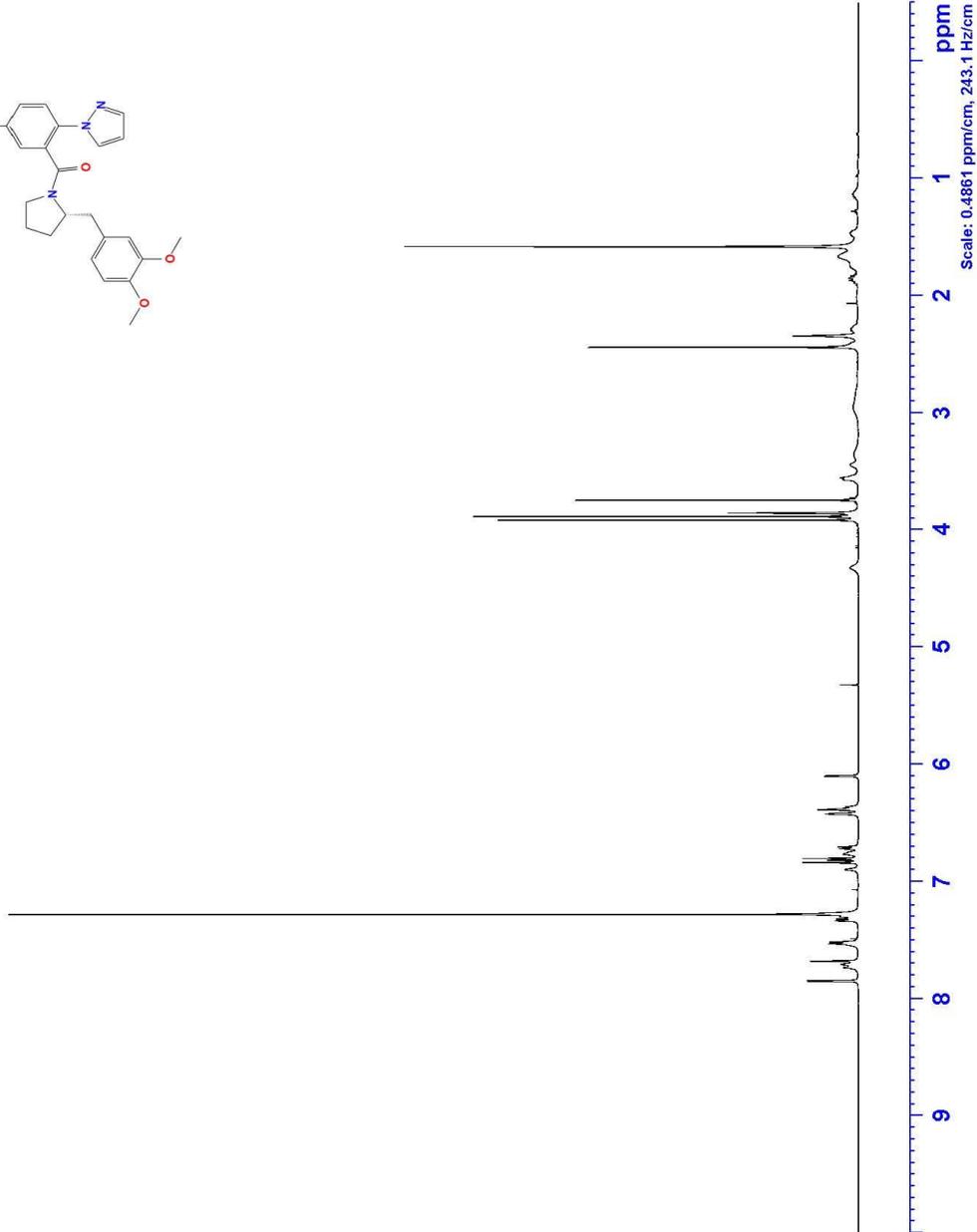


Current Data Parameters  
 NAME Compound 9  
 EXPNO 10  
 PROCNO 1

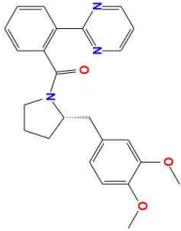
F2 - Acquisition Parameters  
 Date\_ 20110313  
 Time 15:33  
 INSTRUM SPECT  
 PROBHD 5 mm CPDCH 13C  
 PULPROG zgpg30  
 TD 65536  
 CHAN13  
 SOLVENT  
 NS 16  
 DS 2  
 SWH 10000.000 Hz  
 SRH 0.152888 Hz  
 FIDRES 3.276793 SEC  
 AQ 76.000000  
 RG 409.600000  
 DW 50.000 USEC  
 DE 40.000 USEC  
 TE 298.0 K  
 D1 1.00000000 sec  
 TDO 1

==== CHANNEL f1 =====  
 SF01 500.1330085 MHz  
 NUC1 1H  
 P1 9.25 usec  
 ELW1 13.0000000 W

F2 - Processing parameters  
 SI 65536  
 SF 500.1330000 MHz  
 WDM 0  
 SSB 0  
 GB 0  
 PC 0 1.00





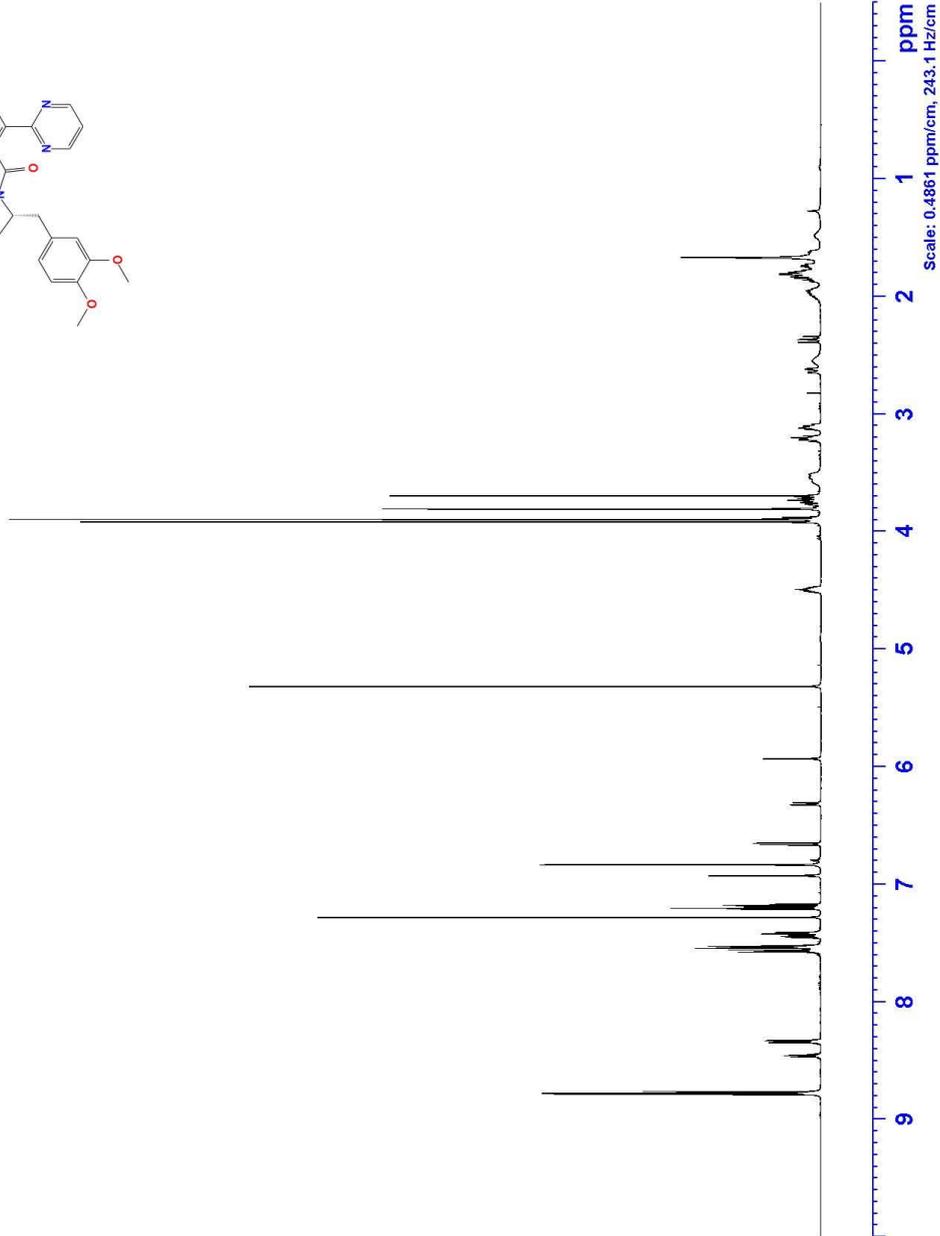


Current Data Parameters  
 NAME Compound 10  
 EXPNO 10  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20111117  
 Time 19:22  
 INSTRUM SPECT  
 PROBHD 5 mm CPDCH 13C  
 PULPROG zgpg30  
 TD 65536  
 CHAN13  
 SOLVENT  
 NS 16  
 DS 2  
 SWH 10000.000 Hz  
 SRH 0.152888 Hz  
 FIDRES 3.276797 SEC  
 AQ 14.601  
 RG 409  
 DW 50.000 USEC  
 DE 40.000 USEC  
 TE 298.0 K  
 D1 1.00000000 sec  
 TDO 1

==== CHANNEL f1 =====  
 SF01 500.1330885 MHz  
 NUC1 1H  
 P1 9.25 USEC  
 ELW1 13.0000000 W

F2 - Processing parameters  
 SI 65536  
 WDW EM  
 SSB 0  
 GB 0  
 EC 0 1.00



```

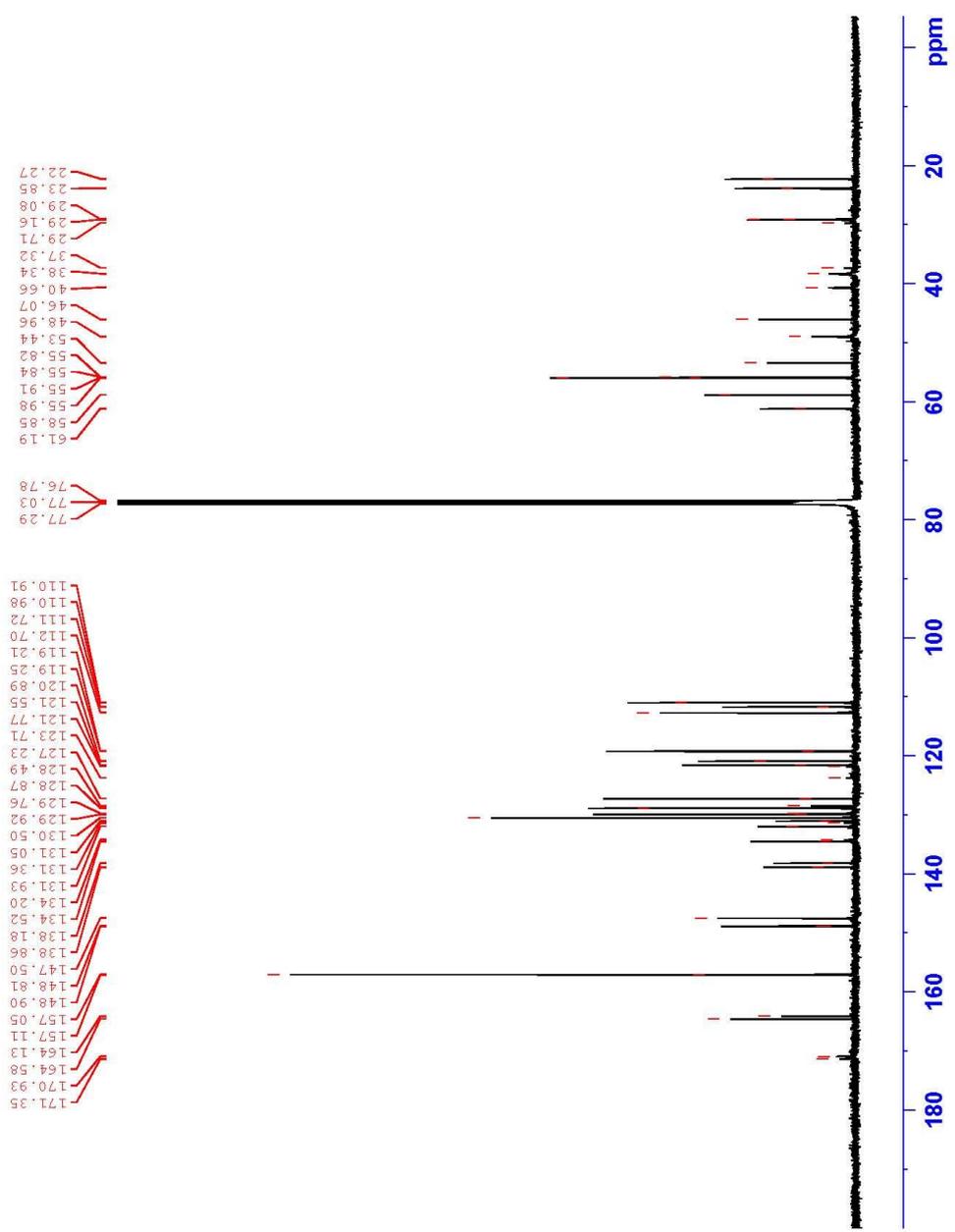
Current Data Parameters
Name      Compound 10
EXPNO    1
PROCNO   1

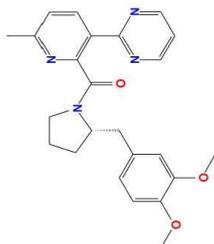
F2 - Acquisition Parameters
Time     20.18
Date_    19.36
INSTRUM  spect
PROBHD   5 mm CPDCH 13C
PULPROG  zgpg30
TD        65536
SOLVENT  CDCl3
NS        256
DS        4
SFO1     31512.605 Hz
AQ        0.480844 Hz
RG         1.0298378 sec
DE         10.000000 sec
TE         60.46 usec
TE        298.0 K
D1         2.0000000 sec
D11        0.0300000 sec
TD0        1

----- CHANNEL f1 -----
NUC1      13C
P1         11.00 usec
PL1        18.00000000 N

----- CHANNEL f2 -----
SFO2     500.1320005 MHz
NUC2      1H
P2         12.00 usec
PL2        0.00000000 N
SFO3     13.00000000 MHz
NUC3      13C
P3         12.00 usec
PL3        0.00000000 N
SFO4     0.17380001 MHz
NUC4      1H
P4         12.00 usec
PL4        0.11125000 N

F2 - Processing parameters
SI        32768
SF        125.7577890 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         1.40
  
```





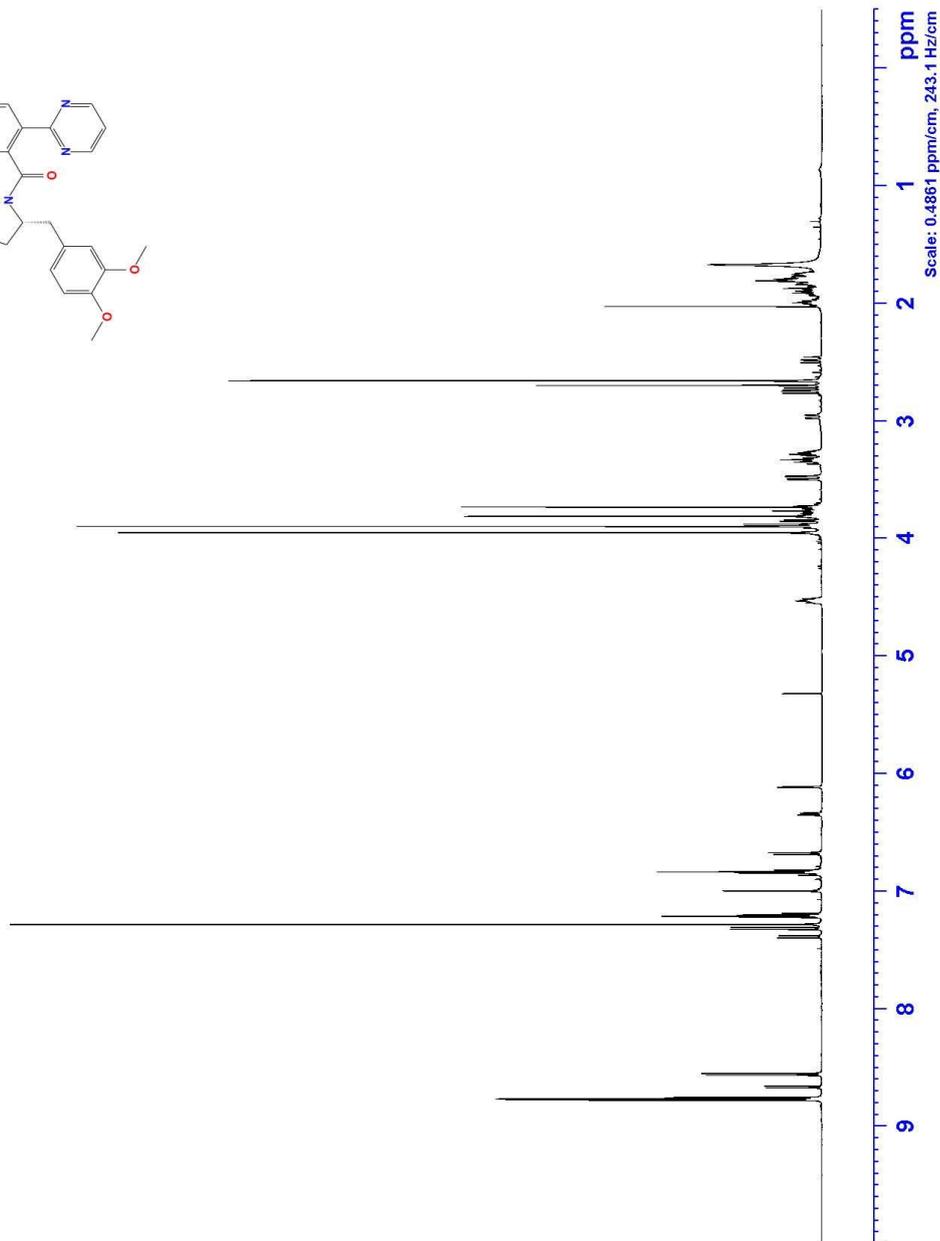
```

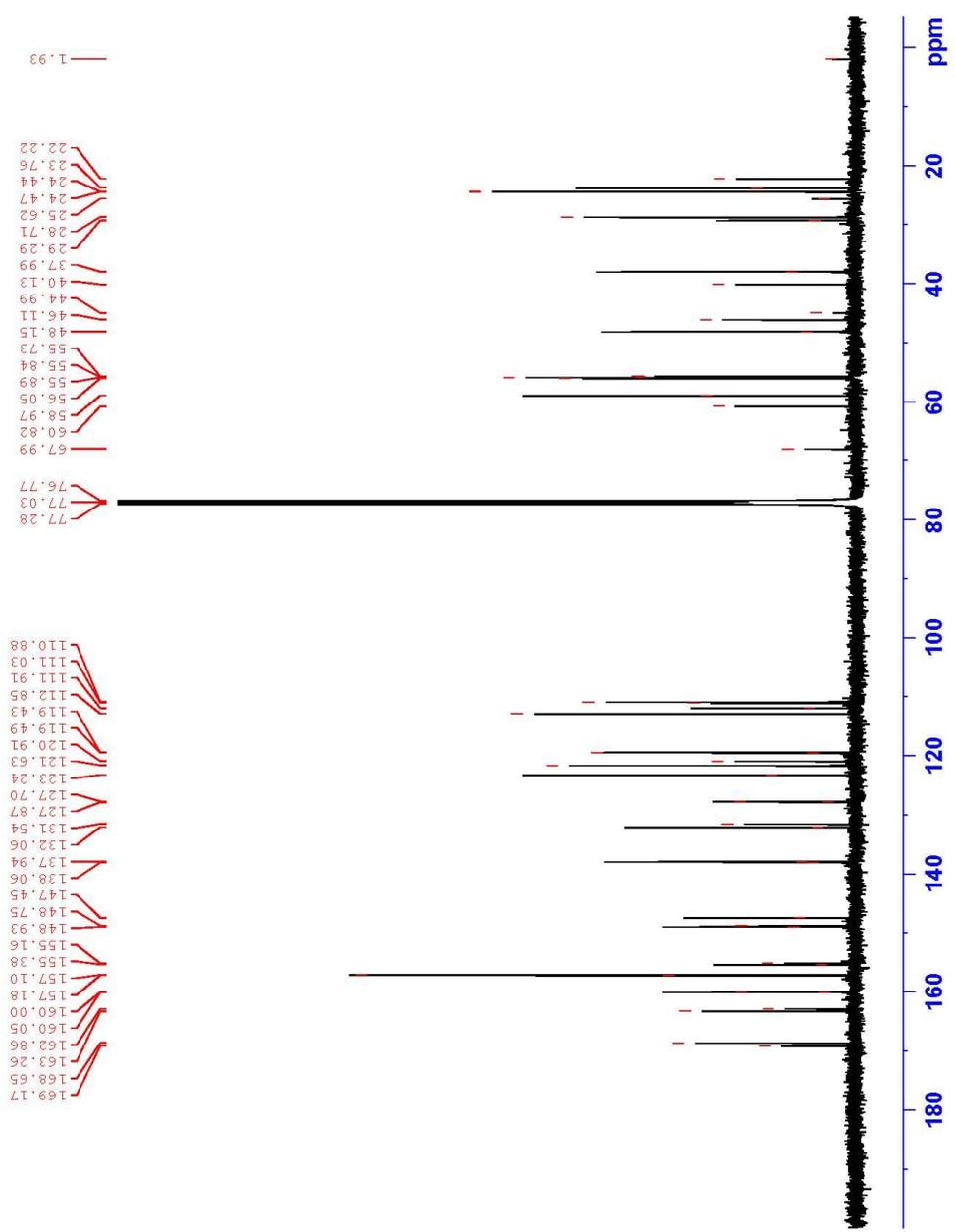
Current Data Parameters
NAME      Compound 11
EXPNO    10
PROCNO   1

F2 - Acquisition Parameters
Date_    20141117
Time     23.02
INSTRUM SPECT
PROBHD   5 mm CPDCH 13C
PULPROG zgpg30
TD       65536
SOLVENT  CDCl3
NS       16
DS       2
SWH      10000.000 Hz
FIDRES   0.152888 Hz
AQ       3.276793 sec
RG       768.000
AQ       50.000 usec
DE       40.000 usec
TE       298.0 K
D1       1.00000000 sec
TD0      1

===== CHANNEL f1 =====
SF01    500.1300885 MHz
NUC1     13C
P1       9.25 usec
ELW1    13.00000000 W

F2 - Processing parameters
SI       65536
WDW      EM
SSB      0
GB       0
PC       1.00
  
```





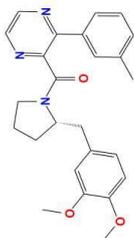
Current Data Parameters  
 Name: Compound\_11  
 EXPNO: 1  
 PROCNO: 1

F2 - Acquisition Parameters  
 Date\_ Time: 2016.02.16  
 Time: 23.16  
 INSTRUM: spect  
 PROBD: 5 mm CPDCH 13C  
 PULPROG: zgpg30  
 TD: 65536  
 SOLVENT: CDCl3  
 NS: 256  
 DS: 4  
 SSB: 31512.605 Hz  
 AQ: 0.480844 Hz  
 FIDRES: 1.0298378 sec  
 RG: 106.667  
 INEG: 15.667  
 DE: 60.46 usec  
 TE: 298.0 K  
 D1: 2.0000000 sec  
 D11: 0.0300000 sec  
 TD0: 1

===== CHANNEL f1 =====  
 NUC1: 13C  
 NUC1: 125.772000 MHz  
 P1: 11.00 usec  
 PL1: 18.00000000 N  
 PLO1: 18.00000000 N

===== CHANNEL f2 =====  
 SFO2: 500.1320005 MHz  
 NUC2: 1H  
 NUC2: 500.1320005 MHz  
 P2: 12.00 usec  
 PL2: 0.00000000 N  
 PLO2: 0.00000000 N  
 P2: 13.00000000 N  
 PLM2: 0.17380001 N  
 PLO3: 0.11125000 N

F2 - Processing parameters  
 SI: 32768  
 SF: 500.1320005 MHz  
 DS: 4  
 SSB: 0  
 LB: 1.00 Hz  
 GB: 0  
 PC: 1.40



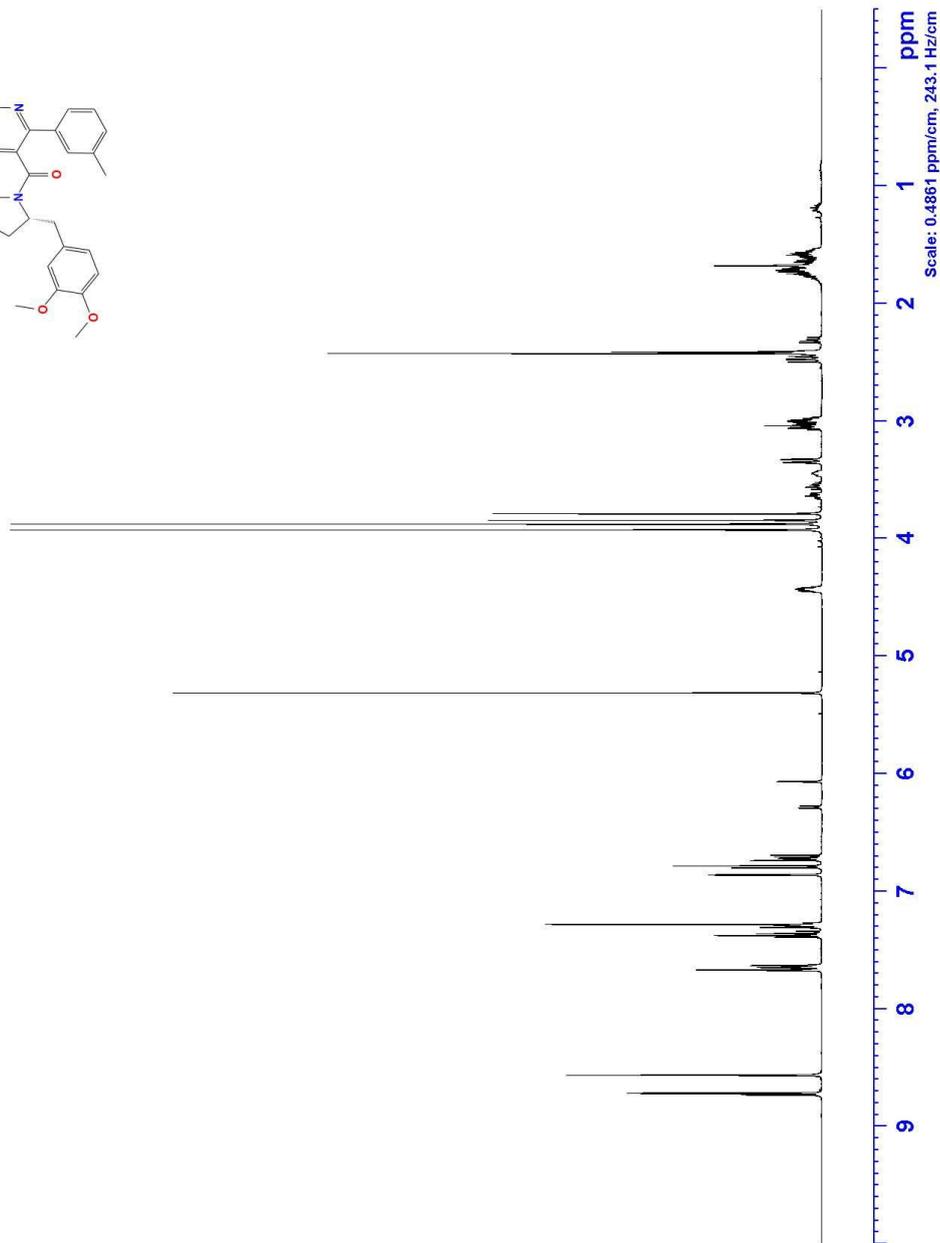
```

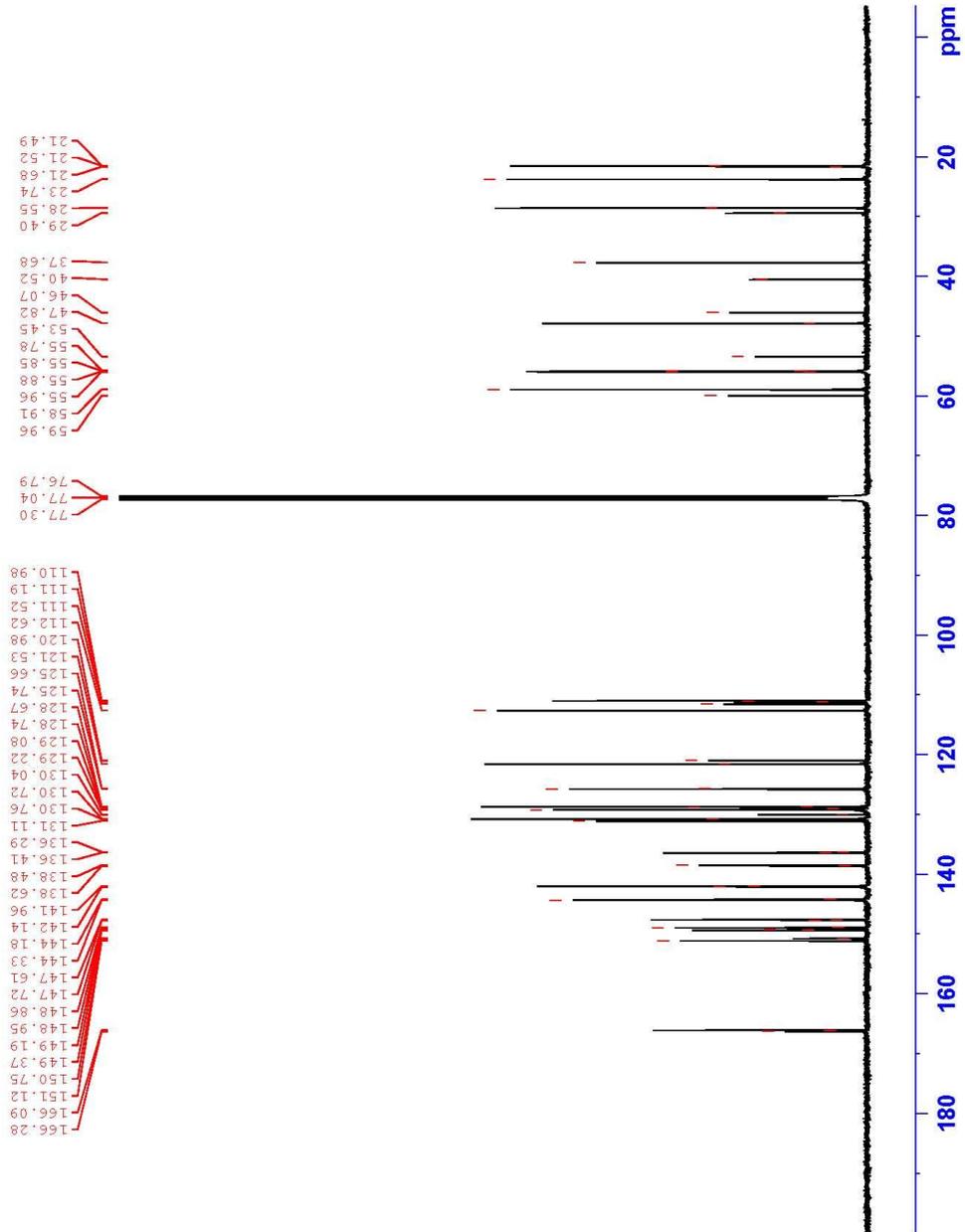
Current Data Parameters
NAME      Compound 12
EXPNO    10
PROCNO   1

F2 - Acquisition Parameters
Date_    20111117
Time     23:57
INSTRUM  spect
PROBHD   5 mm CPDCH 13C
PULPROG  zgpg30
TD       65536
SOLVENT  CDCl3
NS       16
DS       2
SWH      10000.000 Hz
FIDRES   0.152688 Hz
AQ       3.276797 sec
RG       327.5
AQ       14.601000 sec
DE       50.000 USSEC
TE       400.00 USSEC
TB       298.0 K
D1       1.00000000 sec
TD0      1

===== CHANNEL f1 =====
SF01     500.1300885 MHz
NUC1      1H
P1       9.25 usec
ELW1     13.00000000 W

F2 - Processing parameters
SI       65536
WDW      EM
SSB      0
GB       0
PC       1.00
  
```





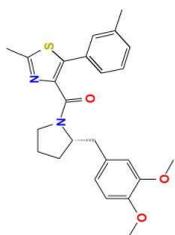
Current Data Parameters  
 Name: Compound 12  
 EXPNO: 1  
 PROCNO: 1

F2 - Acquisition Parameters  
 Date\_ :  
 Time: 2010.01.11  
 INSTRUM: spect  
 PROBD: 5 mm CPDCH 13C  
 PULPROG: zgpg30  
 TD: 65536  
 SOLVENT: CDCl3  
 NS: 256  
 DS: 4  
 SSBH: 31512.605 Hz  
 FIDRES: 0.480844 Hz  
 AQ: 1.0298378 sec  
 RG: 106.600  
 RW: 15.667 usec  
 DE: 60.46 usec  
 TE: 298.0 K  
 D1: 2.0000000 sec  
 D11: 0.0300000 sec  
 TD0: 1

----- CHANNEL f1 -----  
 NUC1: 13C  
 P1: 11.00 usec  
 PL1: 18.00000000 N

----- CHANNEL f2 -----  
 SFO2: 500.1320005 MHz  
 NUC2: 13C  
 P2: 11.00 usec  
 PL2: 18.00000000 N  
 SFO2: 500.1320005 MHz  
 NUC2: 1H  
 P2: 11.00 usec  
 PL2: 18.00000000 N  
 SFO2: 13.00000000 MHz  
 NUC2: 13C  
 P2: 11.00 usec  
 PL2: 18.00000000 N

F2 - Processing parameters  
 SI: 32768  
 SF: 125.7577890 MHz  
 WDW: EM  
 SSB: 0  
 LB: 1.00 Hz  
 GB: 0  
 PC: 1.40

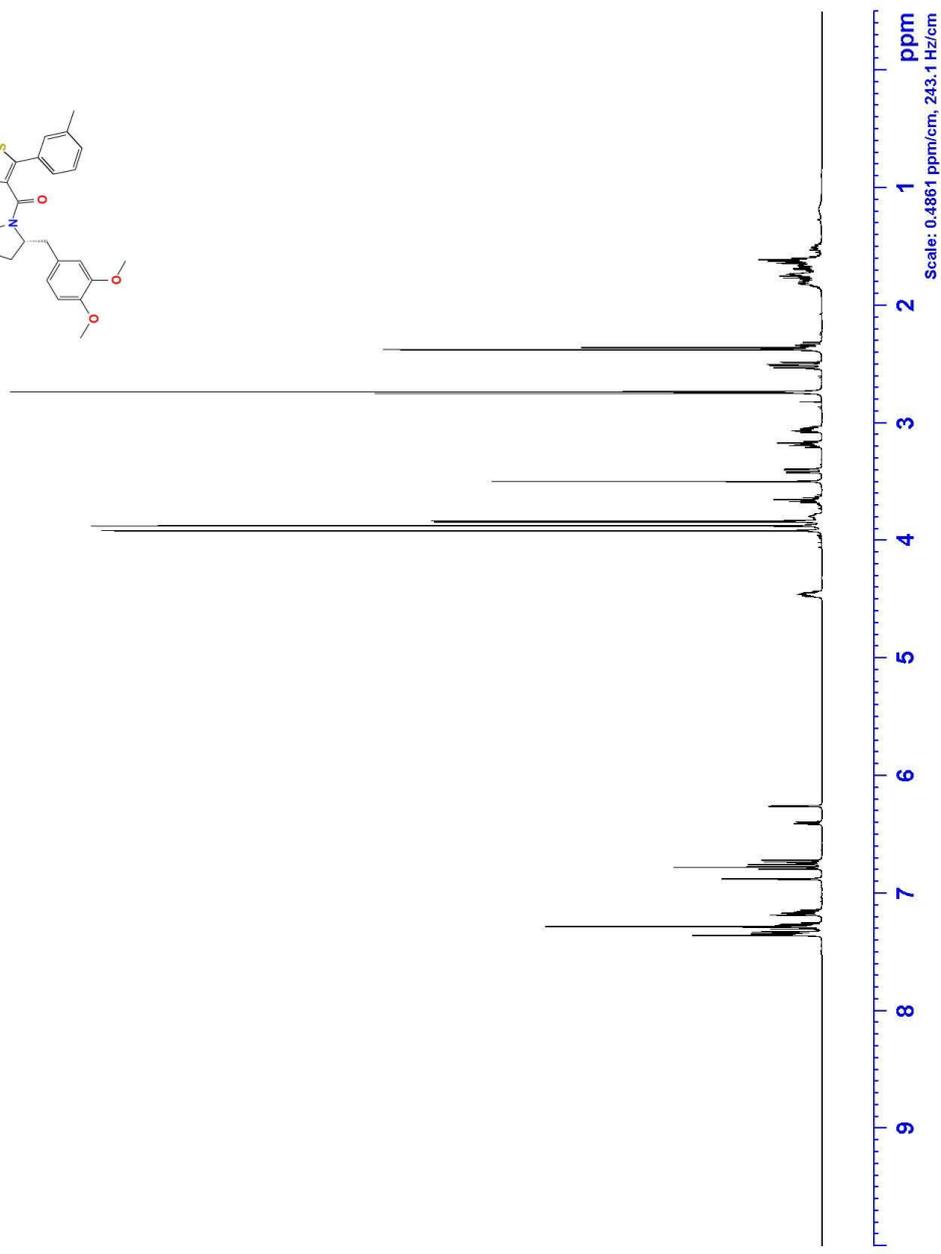


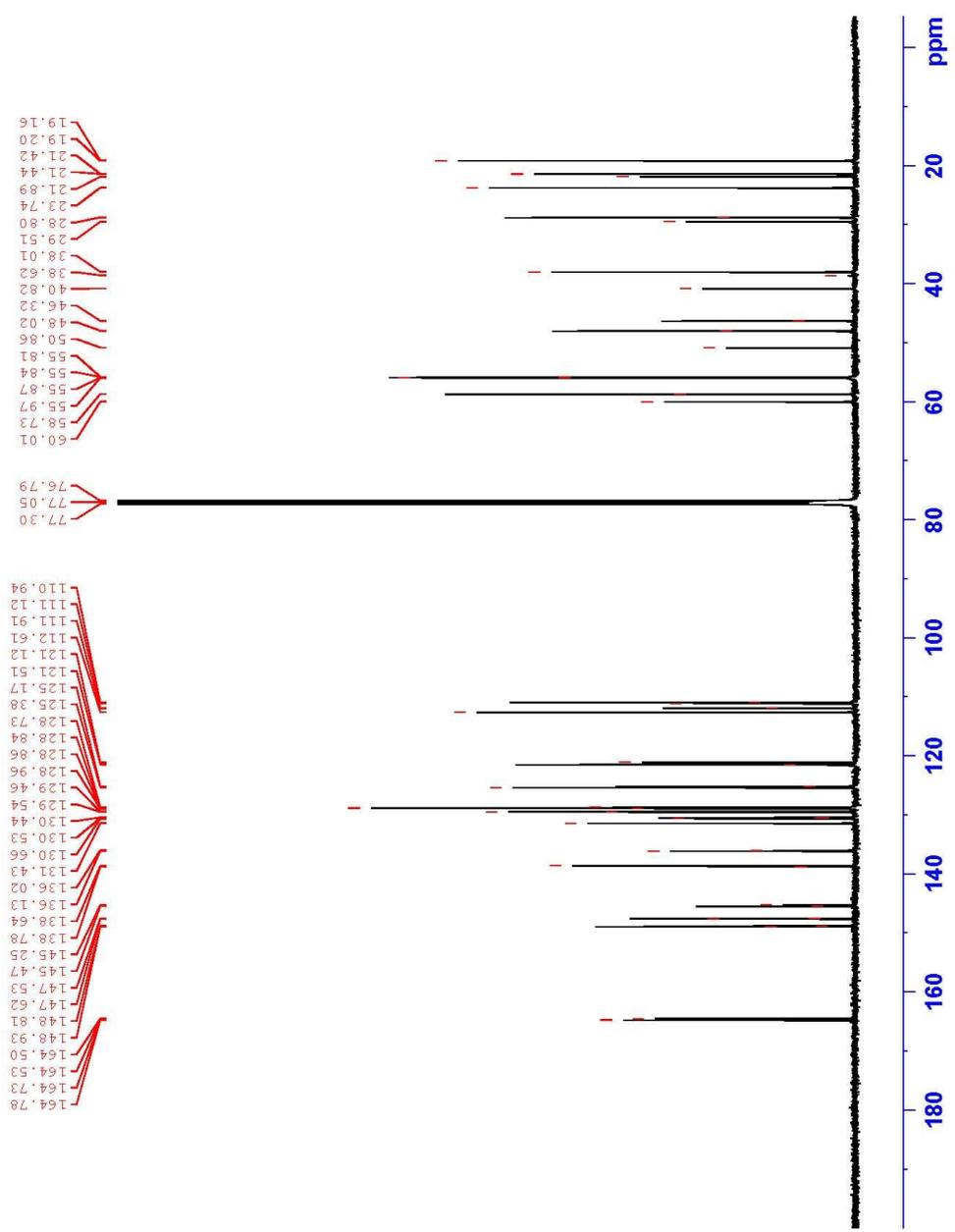
Current Data Parameters  
 NAME Compound 13  
 EXPNO 10  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20110818  
 Time 23:38  
 INSTRUM spect  
 PROBHD 5 mm CPDCH 13C  
 PULPROG zgpg30  
 TD 65536  
 CHAN13  
 SOLVENT  
 NS 16  
 DS 2  
 SWH 10000.000 Hz  
 SFR 0.152888 Hz  
 AQ 3.276797 sec  
 FWHM 14.601 Hz  
 DQ 50.000 usec  
 DE 40.000 usec  
 TE 298.0 K  
 D1 1.00000000 sec  
 TDO 1

==== CHANNEL f1 =====  
 SF01 500.1330885 MHz  
 NUC1 1H  
 P1 9.25 usec  
 ELW1 13.0000000 W

F2 - Processing parameters  
 SI 65536  
 WDW EM  
 SSB 0  
 GB 0  
 PC 0 1.00





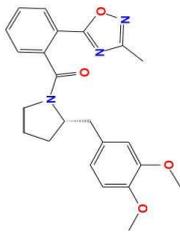
Current Data Parameters  
 Name: 13  
 Compound: 13  
 EXPNO: 1  
 PROCNO: 1

F2 - Acquisition Parameters  
 Date\_ :  
 Time: 23.53  
 INSTRUM: spect  
 PROBD: 5 mm CPDCH 13C  
 PULPROG: zgpg30  
 TD: 65536  
 SOLVENT: CDCl3  
 NS: 256  
 DS: 4  
 SSB: 31512.605 Hz  
 FIDRES: 0.480844 Hz  
 AQ: 1.0398378 sec  
 RG: 1066  
 INEG: 15.667 usec  
 DE: 60.46 usec  
 TE: 298.0 K  
 D1: 2.0000000 sec  
 D11: 0.0300000 sec  
 TD0: 1

----- CHANNEL f1 -----  
 NUC1: 13C  
 P1: 11.00 usec  
 PL1: 18.00000000 N

----- CHANNEL f2 -----  
 SFO2: 500.1320005 MHz  
 NUC2: 1H  
 P2: 19.00 usec  
 PL2: 0.00000000 N  
 PL12: 13.00000000 N  
 PL13: 0.17380001 N  
 PL14: 0.11125000 N

F2 - Processing parameters  
 SI: 32768  
 SF: 125.7577890 MHz  
 EQ: BR  
 SSB: 0  
 LB: 1.00 Hz  
 GB: 0  
 PC: 1.40

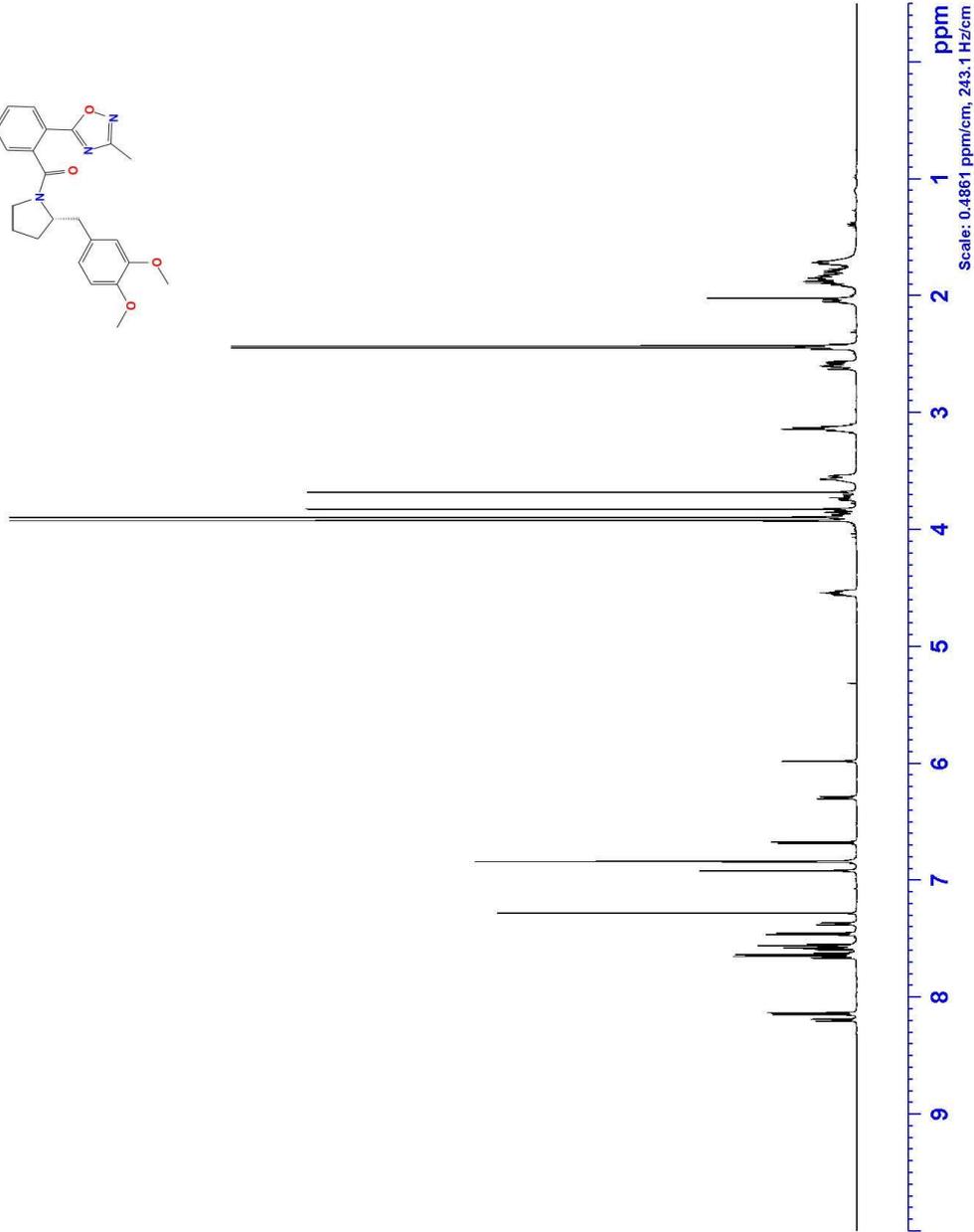


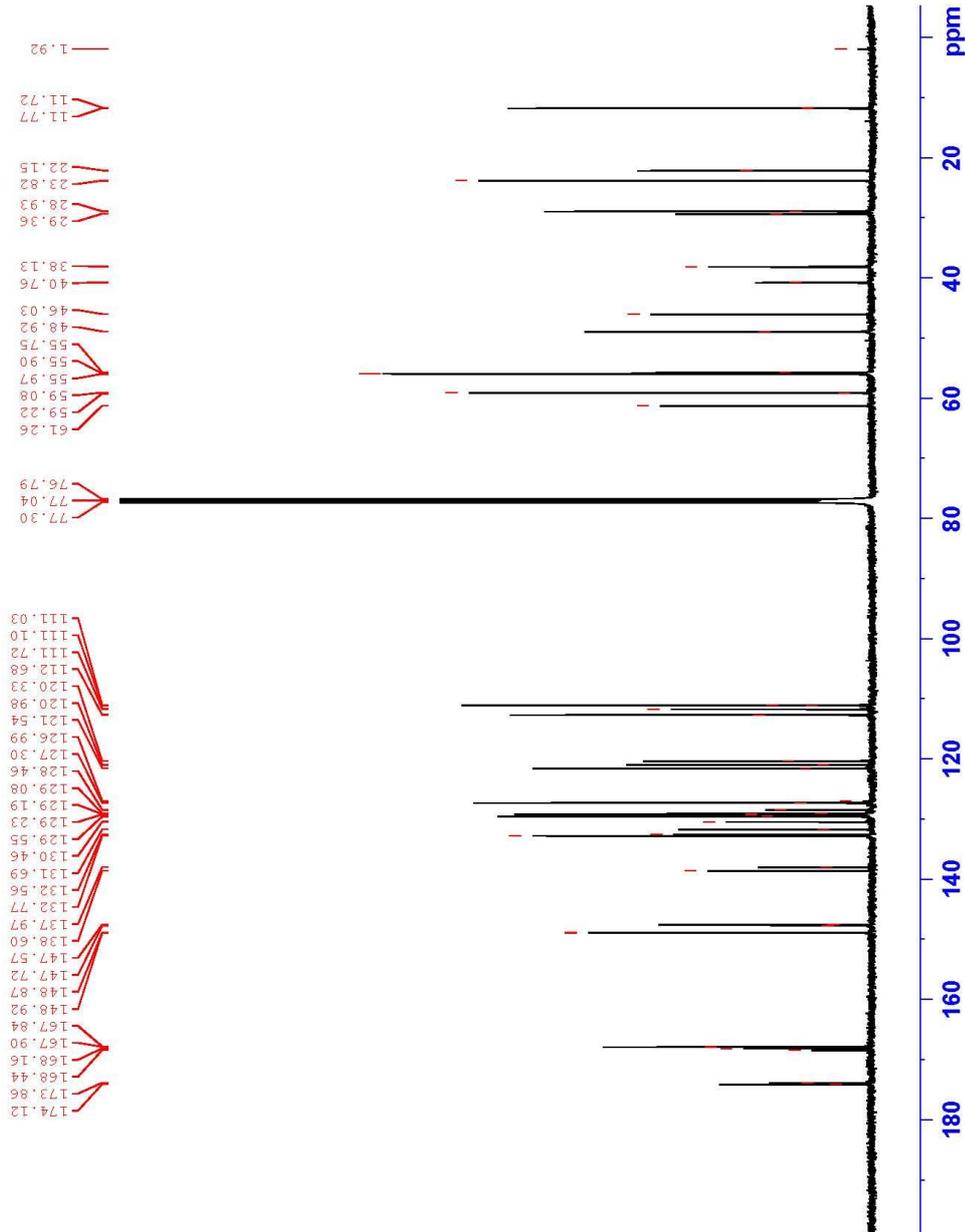
Current Data Parameters  
 NAME Compound 14  
 EXPNO 10  
 PROCNO 1

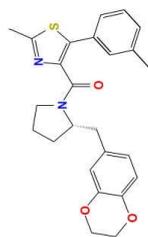
F2 - Acquisition Parameters  
 Date\_ 20141114  
 Time 19:40  
 INSTRUM SPECT  
 PROBHD 5 mm CPDCH 13C  
 PULPROG zgpg30  
 TD 65536  
 CHAN13  
 SOLVENT  
 NS 16  
 DS 2  
 SWH 10000.000 Hz  
 SRH 0.152888 Hz  
 FIDRES 3.276797 SEC  
 AQ 14.601 USEC  
 RG 409.6  
 DW 50.000 USEC  
 DE 40.00 USEC  
 TE 298.0 K  
 D1 1.0000000 sec  
 TDO 1

==== CHANNEL f1 =====  
 SF01 500.1300885 MHz  
 NUC1 1H  
 P1 9.25 USEC  
 ELW1 13.0000000 W

F2 - Processing parameters  
 SI 65536  
 WDW EM  
 SSB 0  
 GB 0  
 PC 0 1.00





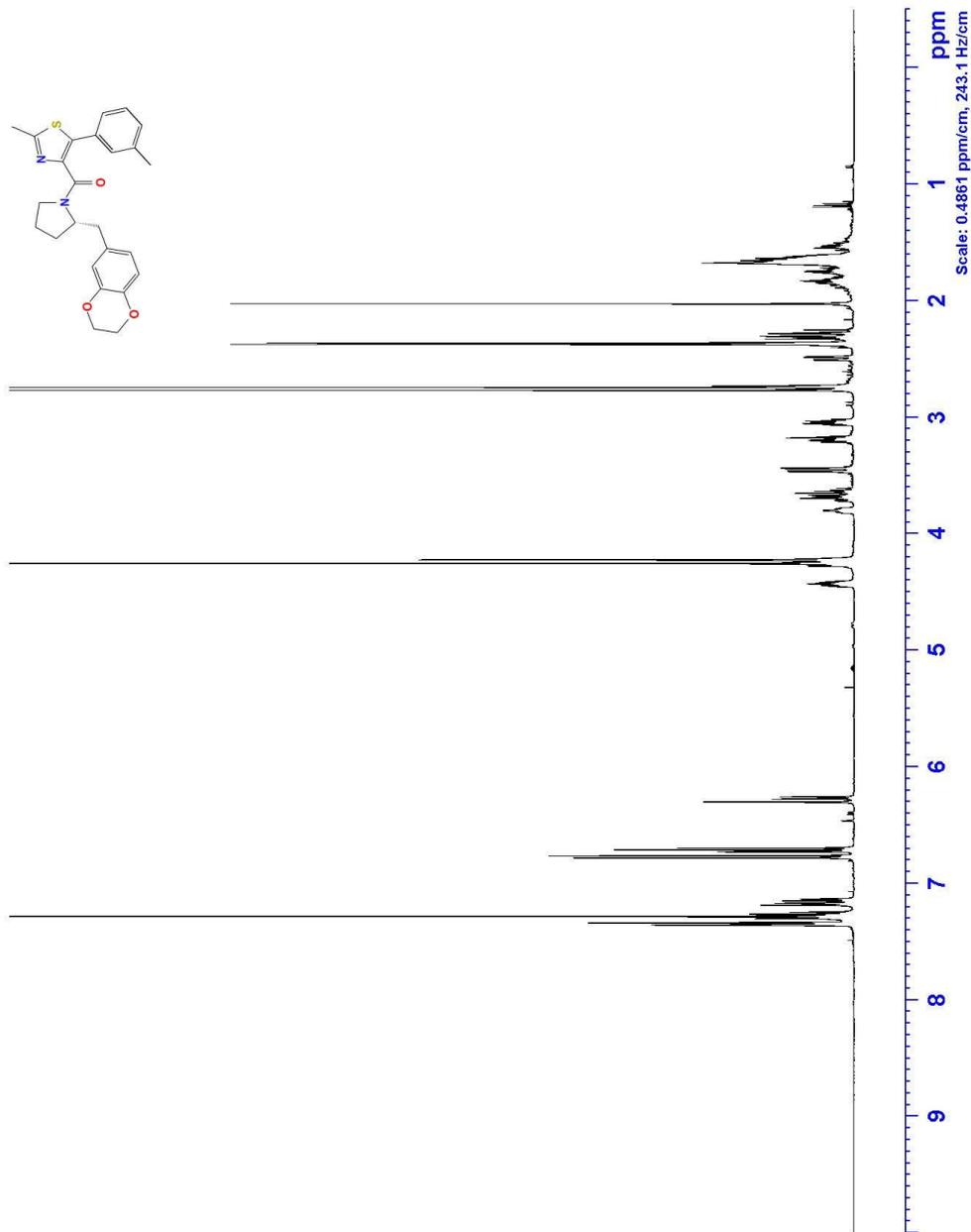


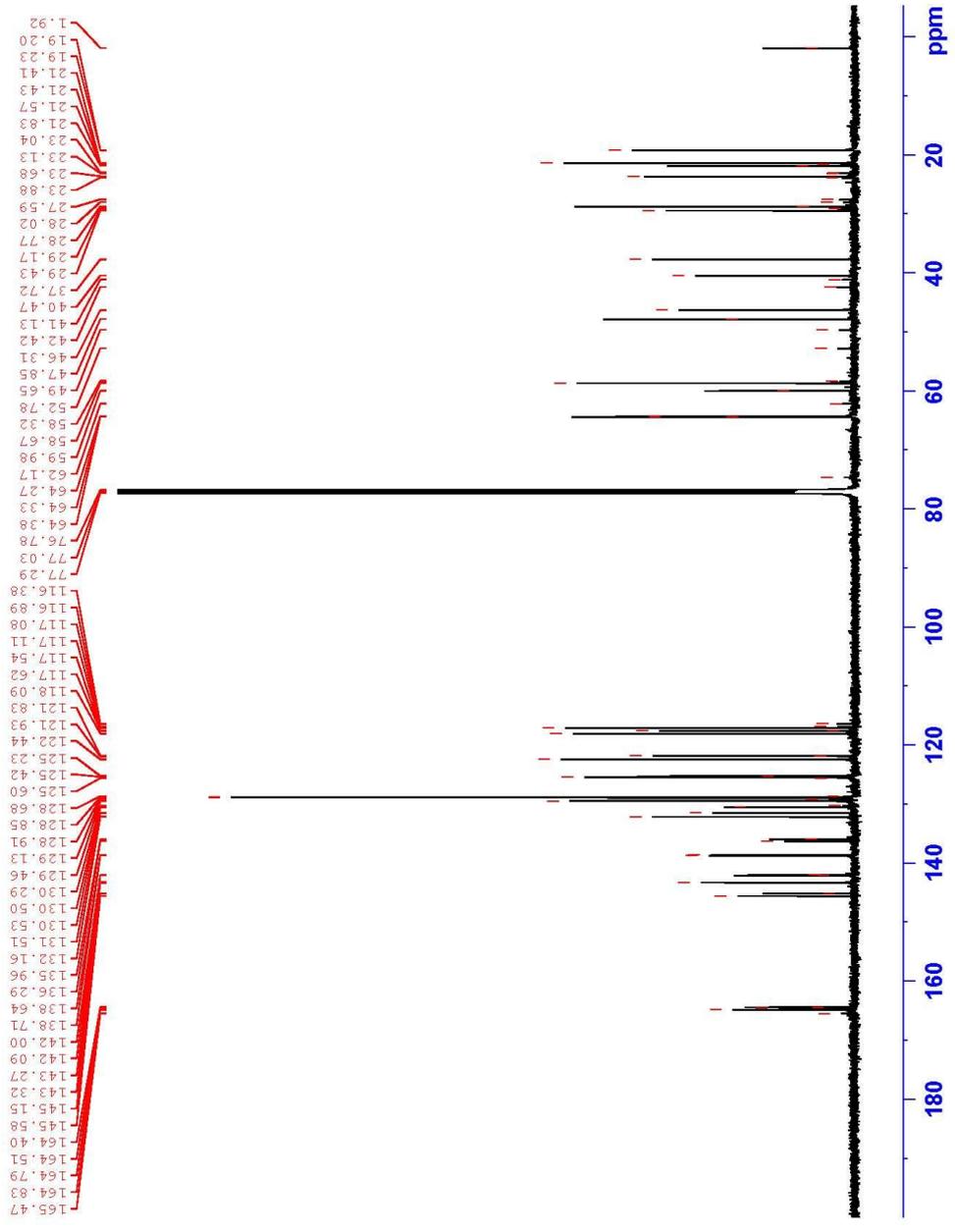
```

Current Data Parameters
NAME      Compound 15
EXPNO    10
PROCNO   1

F2 - Acquisition Parameters
Date_    20111118
Time     17.56
INSTRUM  spect
PROBHD   5 mm CPDCH 13C
PULPROG  zgpg30
TD       65536
SOLVENT  CDCl3
NS       16
DS       2
SWH      10000.000 Hz
FIDRES   0.152888 Hz
AQ       3.276797 sec
RG       327.5
WDW      EM
SSB      0
GB       0
DE       40.00 USSEC
TE       298.0 K
D1       1.00000000 sec
TD0      1

===== CHANNEL f1 =====
SF01    500.1300885 MHz
NUC1     13C
P1       9.25 usec
ELW1    13.00000000 W
F2 - Processing parameters
SI       65536
WDW      EM
SSB      0
GB       0
DE       40.30 Hz
TE       0
PC       1.00
  
```





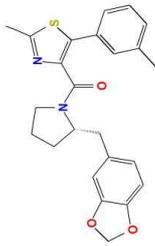
Current Data Parameters  
 Name: 15  
 Compound: 15  
 PROCNO: 1

F2 - Acquisition Parameters  
 Date\_ Time: 2016.11.18.11  
 INSTRUM: spect  
 PROBD: 5 mm CPDCH 13C  
 PULPROG: zgpg30  
 TD: 65536  
 SOLVENT: CDCl3  
 NS: 256  
 DS: 4  
 SSB: 31512.605 Hz  
 FIDRES: 0.480844 Hz  
 AQ: 1.039878 sec  
 RG: 1000  
 INEG: 15.667 usec  
 DE: 60.46 usec  
 TE: 298.0 K  
 D1: 2.0000000 sec  
 D11: 0.0300000 sec  
 TD0: 1

===== CHANNEL f1 =====  
 NUC1: 13C  
 P1: 11.00 usec  
 PL1: 18.00000000 N

===== CHANNEL f2 =====  
 SFO2: 500.1320005 MHz  
 NUC2: 1H  
 P2: 19.00 usec  
 PL2: 0.00000000 N  
 SFO1: 125.7611540 MHz  
 NUC1: 13C  
 P1: 11.00 usec  
 PL1: 18.00000000 N

F2 - Processing parameters  
 SI: 32768  
 SF: 125.7577890 MHz  
 DS: 4  
 SSB: 0  
 LB: 1.00 Hz  
 GB: 0  
 PC: 1.40



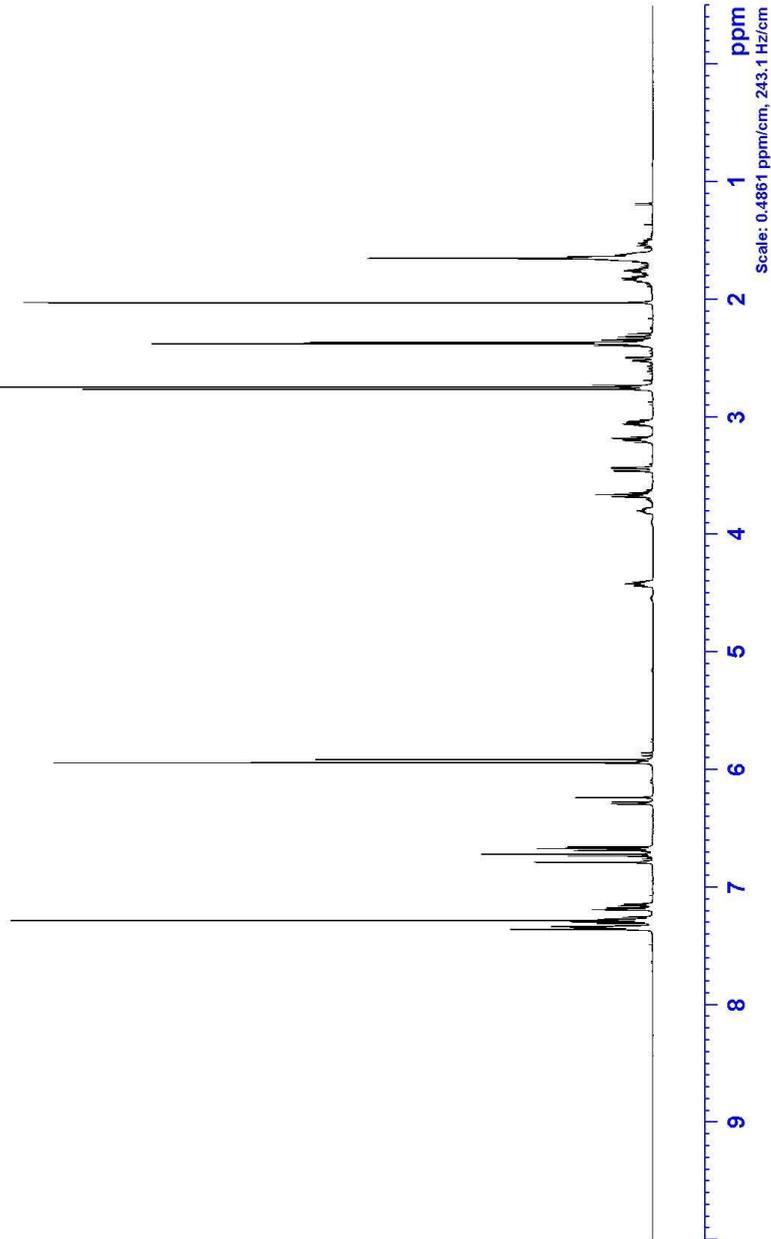
```

Current Data Parameters
NAME      Compound 16
EXPNO    10
PROCNO   1

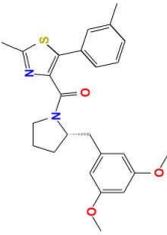
F2 - Acquisition Parameters
Date_    20111111
Time     16.14
INSTRUM  SPECT
PROBHD   5 mm CPDCH 13C
PULPROG  zgpg30
TD       65536
SOLVENT  CDCl3
NS       16
DS       2
SWH      10000.000 Hz
FIDRES   0.152888 Hz
AQ       3.276797 sec
RG       327.5
AQ       50.000 USSEC
DE       40.00 USSEC
TE       298.0 K
D1       1.00000000 sec
TD0      1

===== CHANNEL f1 =====
SF01    500.1330885 MHz
NUC1     13C
P1       9.25 usec
ELW1    13.00000000 W

F2 - Processing parameters
SI       65536
WDW      EM
SSB      0
GB       0
PC       1.00
  
```







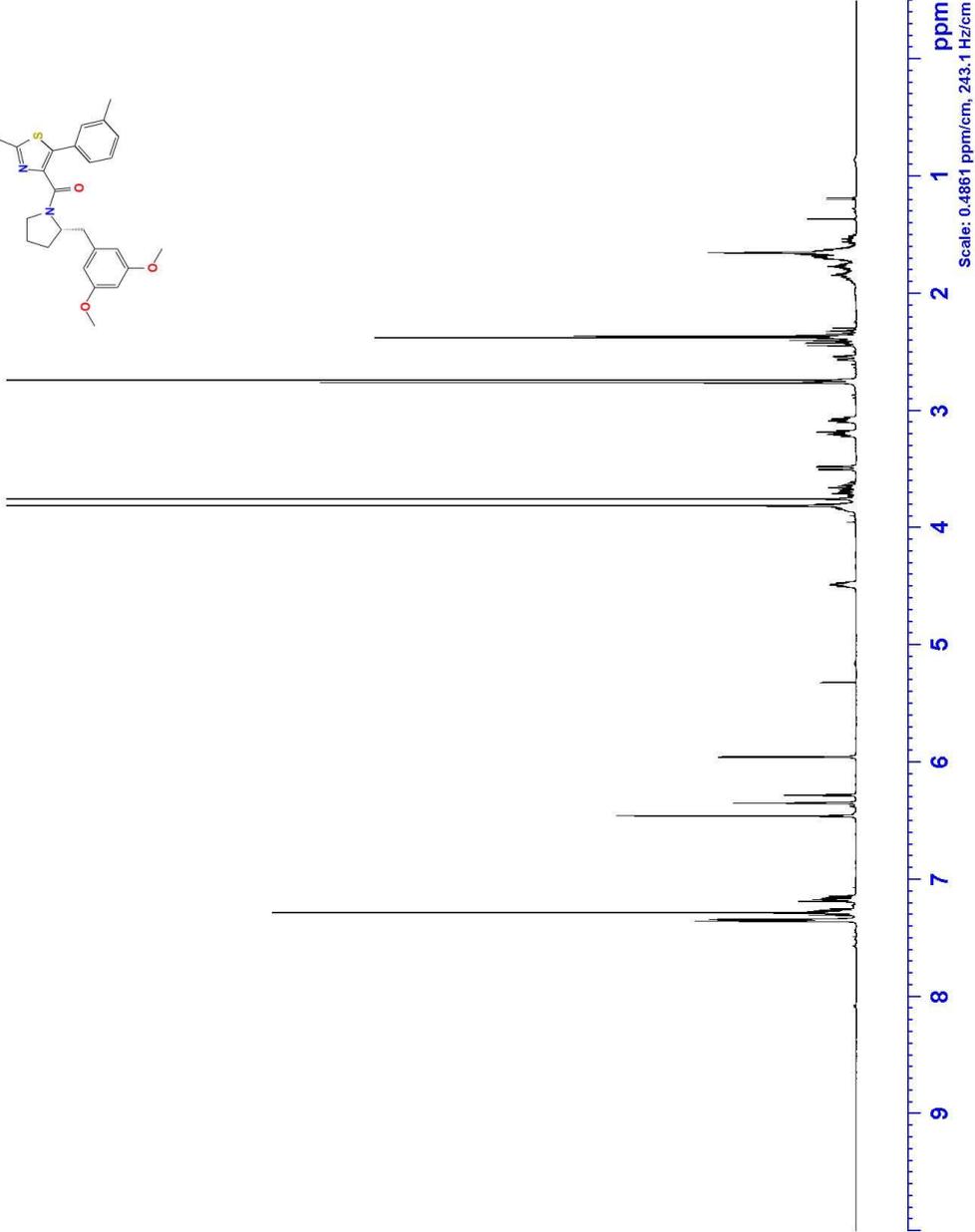
```

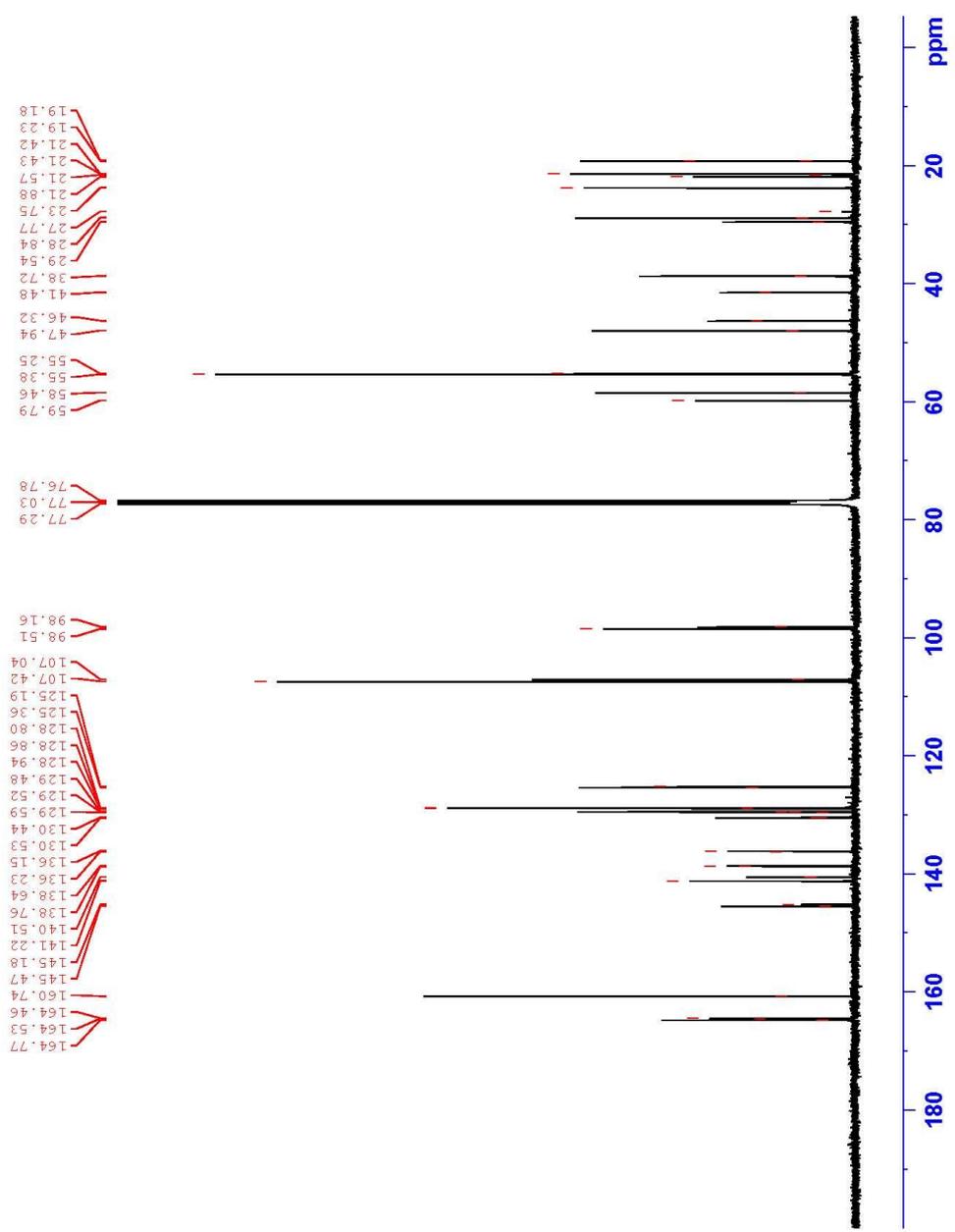
Current Data Parameters
NAME      Compound 17
EXPNO    10
PROCNO   1

F2 - Acquisition Parameters
Date_    20110818
Time     19:58
INSTRUM  spect
PROBHD   5 mm CPDCH 13C
PULPROG  zgpg30
TD       65536
SOLVENT  CDCl3
NS       16
DS       2
SWH      10000.000 Hz
FIDRES   0.152888 Hz
AQ       3.276797 sec
RG       346.000
DW       50.000 usec
DE       40.000 usec
TE       298.0 K
D1       1.00000000 sec
TD0      1

===== CHANNEL f1 =====
SF01    500.1330885 MHz
NUC1     1H
P1       9.25 usec
ELW1    13.00000000 W

F2 - Processing parameters
SI      65536
WDW     EM
SSB     0
GB      0
PC      1.00
  
```





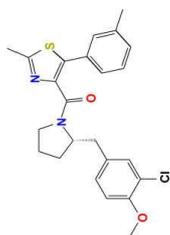
Current Data Parameters  
 Name: 17  
 Compound: 17  
 EXPNO: 1  
 PROCNO: 1

F2 - Acquisition Parameters  
 Date\_ : 2013  
 Time: 20.13  
 INSTRUM: spect  
 PROBD: 5 mm CPDCH 13C  
 PULPROG: zgpg30  
 TD: 65536  
 SOLVENT: CDCl3  
 NS: 256  
 DS: 4  
 SSB: 31512.605 Hz  
 FIDRES: 0.480844 Hz  
 AQ: 1.0298378 sec  
 RG: 1000  
 INEG: 15.667 usec  
 DE: 60.46 usec  
 TE: 298.0 K  
 D1: 3.0000000 sec  
 D11: 0.0300000 sec  
 TD0: 1

----- CHANNEL f1 -----  
 NUC1: 13C  
 P1: 11.00 usec  
 PL1: 18.00000000 N

----- CHANNEL f2 -----  
 SFO2: 500.1320005 MHz  
 NUC2: 1H  
 P2: 19.00 usec  
 PL2: 0.00000000 N  
 PL12: 13.00000000 N  
 PL13: 0.17380001 N  
 PL14: 0.11125000 N

F2 - Processing parameters  
 SI: 32768  
 SF: 125.7577890 MHz  
 EQ: EN  
 SSB: 0  
 LB: 1.00 Hz  
 GB: 0  
 PC: 1.40

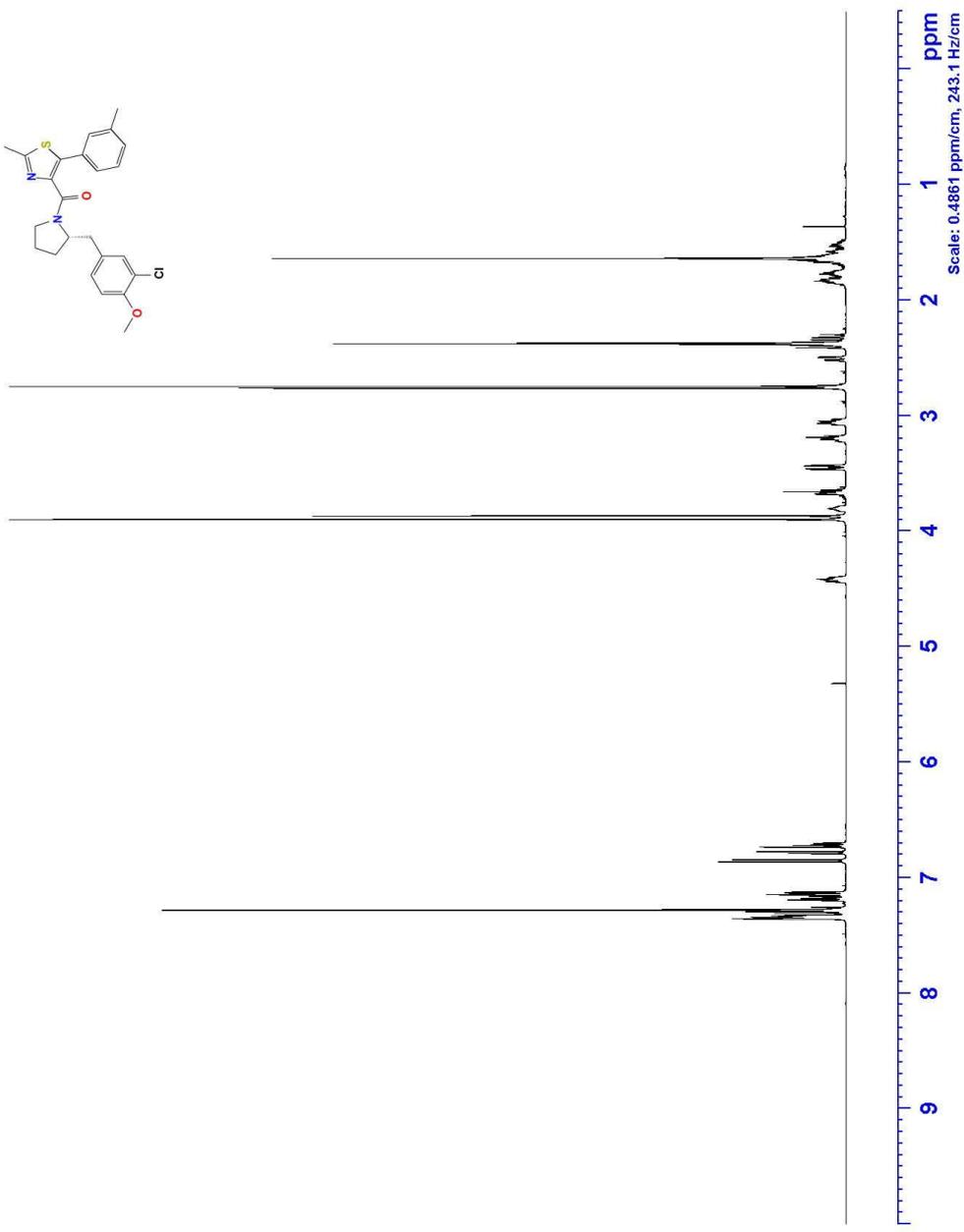


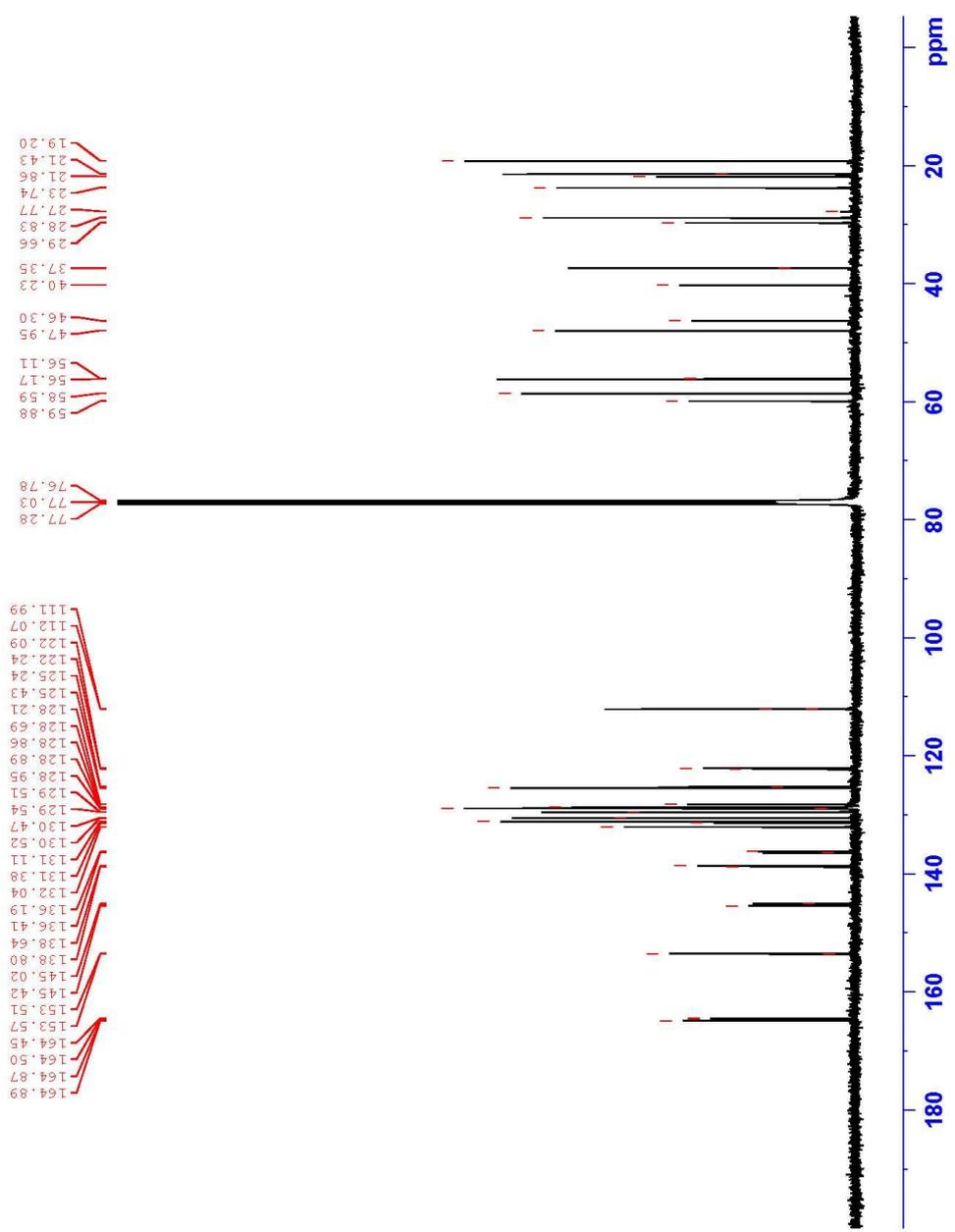
Current Data Parameters  
 NAME Compound 18  
 EXPNO 10  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20111114  
 Time 21:48  
 INSTRUM SPECT  
 PROBHD 5 mm CPDCH 13C  
 PULPROG zgpg30  
 TD 65536  
 CHAN13  
 SOLVENT  
 NS 16  
 DS 2  
 SWH 10000.000 Hz  
 FIDRES 0.152888 Hz  
 AQ 3.276797 sec  
 RG 4096  
 DW 50.000 usec  
 DE 40.000 usec  
 TE 298.0 K  
 D1 1.00000000 sec  
 TDO 1

==== CHANNEL f1 =====  
 SF01 500.1300885 MHz  
 NUC1 1H  
 P1 9.25 usec  
 ELW1 13.0000000 W

F2 - Processing parameters  
 SI 65536  
 WDW EM  
 SSB 0  
 GB 0  
 PC 0 1.00





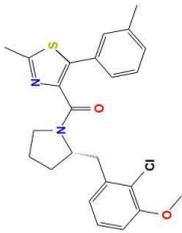
```

Current Data Parameters
=====
Name      Compound 18
EXPNO    1
PROCNO   1

F2 - Acquisition Parameters
=====
Time          20.00
Date_         22.03
INSTRUM      spect
PROBHD       5 mm CPDCH 13C
PULPROG      zgpg30
TD           65536
SOLVENT      CDCl3
NS           256
DS           4
SWH          31512.605 Hz
FIDRES       0.480844 Hz
AQ           1.0298378 sec
RG           106.67
DE           15.667 usec
TE           60.46 usec
TE          298.0 K
D1           2.0000000 sec
d11          0.0300000 sec
TD0          1

===== CHANNEL f1 =====
NUC1         13C
P1           11.00 usec
PL1         18.0000000 N

===== CHANNEL f2 =====
SFO2         500.1320005 MHz
NUC2         1H
P2           19.00 usec
PL2         0.0000000 N
=====
SFO1         125.7620000 MHz
=====
SFO3         13.0000000 MHz
=====
SFO4         0.17380001 MHz
=====
SFO5         0.11125000 MHz
=====
F2 - Processing parameters
=====
SI           32768
SF           125.7577890 MHz
WDW          EM
SSB          0
LB           1.00 Hz
GB           0
PC           1.40
  
```

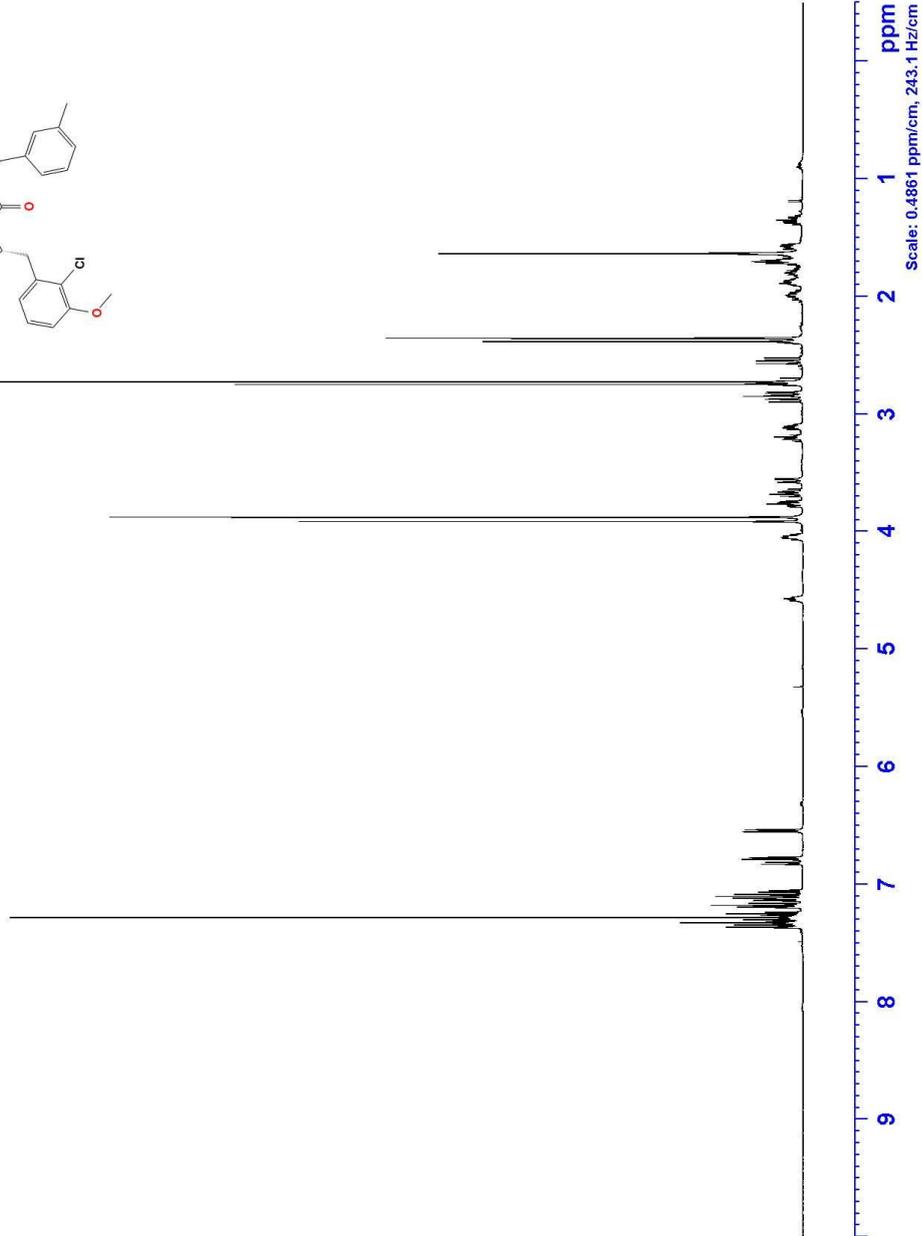


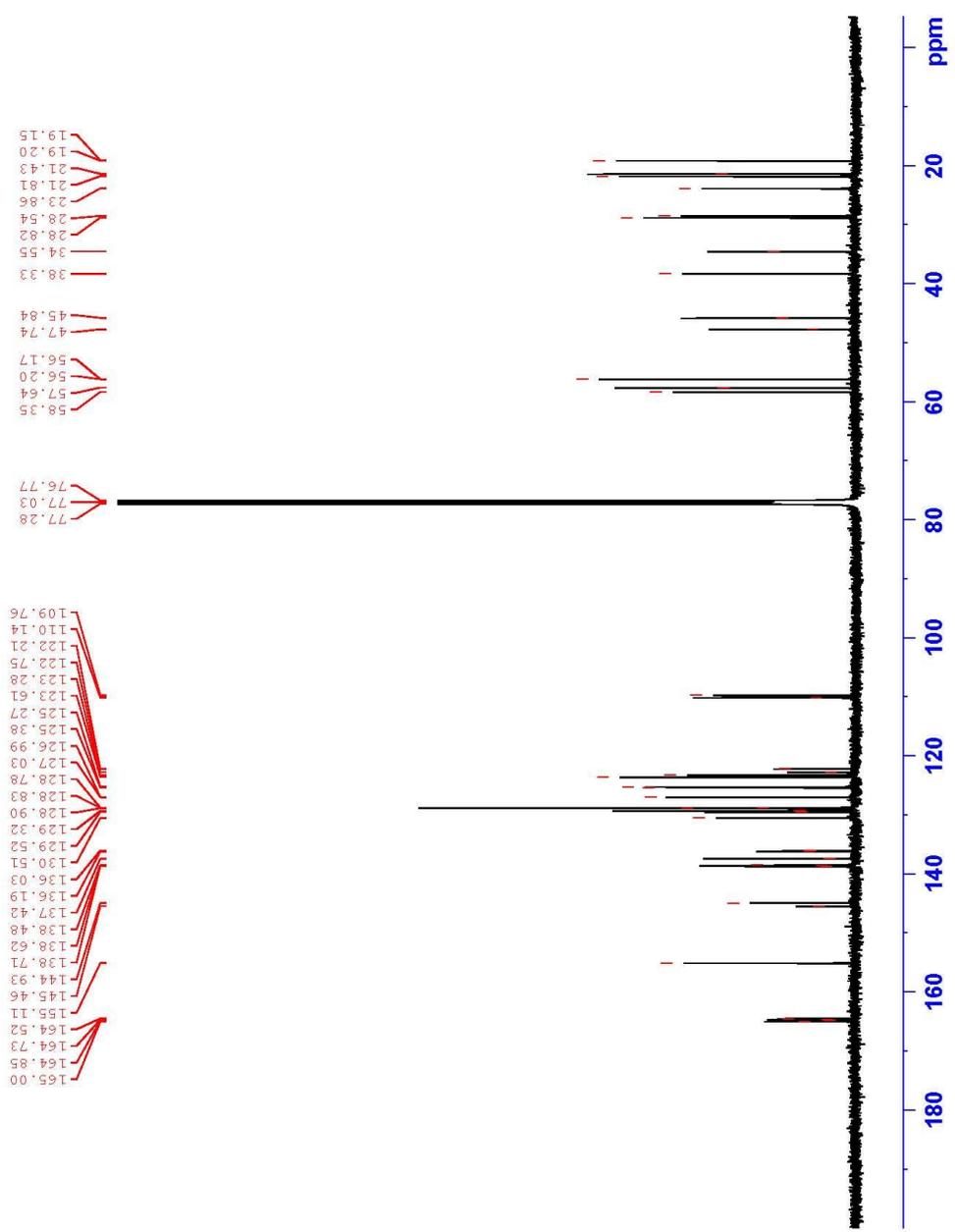
Current Data Parameters  
 NAME Compound 19  
 EXPNO 10  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20141118  
 Time 1.38  
 INSTRUM SPECT  
 PROBHD 5 mm CPDCH 13C  
 PULPROG zgpg30  
 TD 65536  
 CHAN13  
 SOLVENT  
 NS 16  
 DS 2  
 SWH 10000.000 Hz  
 FIDRES 0.152688 Hz  
 AQ 3.276793 sec  
 RG 400.000  
 DW 50.000 USEC  
 DE 40.000 USEC  
 TE 298.0 K  
 D1 1.00000000 sec  
 TDO 1

==== CHANNEL f1 =====  
 SF01 500.1330085 MHz  
 NUC1 1H  
 P1 9.25 usec  
 ELW1 13.0000000 W

F2 - Processing parameters  
 SI 65536  
 WDW EM  
 SSB 0  
 GB 0  
 EC 0 1.00





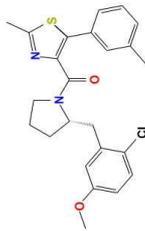
Current Data Parameters  
 Name: 13C  
 Compound: 15  
 EXPNO: 2  
 PROCNO: 1

F2 - Acquisition Parameters  
 Date\_ Time: 20160811 14.43  
 INSTRUM: spect  
 PROBD: 5 mm CPDCH 13C  
 PULPROG: zgpg30  
 TD: 65536  
 SOLVENT: CDCl3  
 NS: 256  
 DS: 4  
 SSB: 31512.605 Hz  
 AQ: 0.480844 Hz  
 FIDRES: 1.0298378 sec  
 RG: 106.667 Hz  
 RW: 15.667 Hz  
 DE: 60.46 usec  
 TE: 298.0 K  
 D1: 2.0000000 sec  
 D11: 0.0300000 sec  
 TD0: 1

===== CHANNEL f1 =====  
 NUC1: 13C  
 P1: 11.00 usec  
 PL1: 18.0000000 N

===== CHANNEL f2 =====  
 SFO2: 500.1320005 MHz  
 NUC2: 13C  
 P2: 11.00 usec  
 PL2: 18.0000000 N  
 SFO1: 125.7611250 MHz  
 NUC1: 1H  
 P1: 11.00 usec  
 PL1: 18.0000000 N

F2 - Processing parameters  
 SI: 32768  
 SF: 125.7577890 MHz  
 EQ: EN  
 SSB: 0  
 LB: 0 1.00 Hz  
 GB: 0  
 PC: 1.40

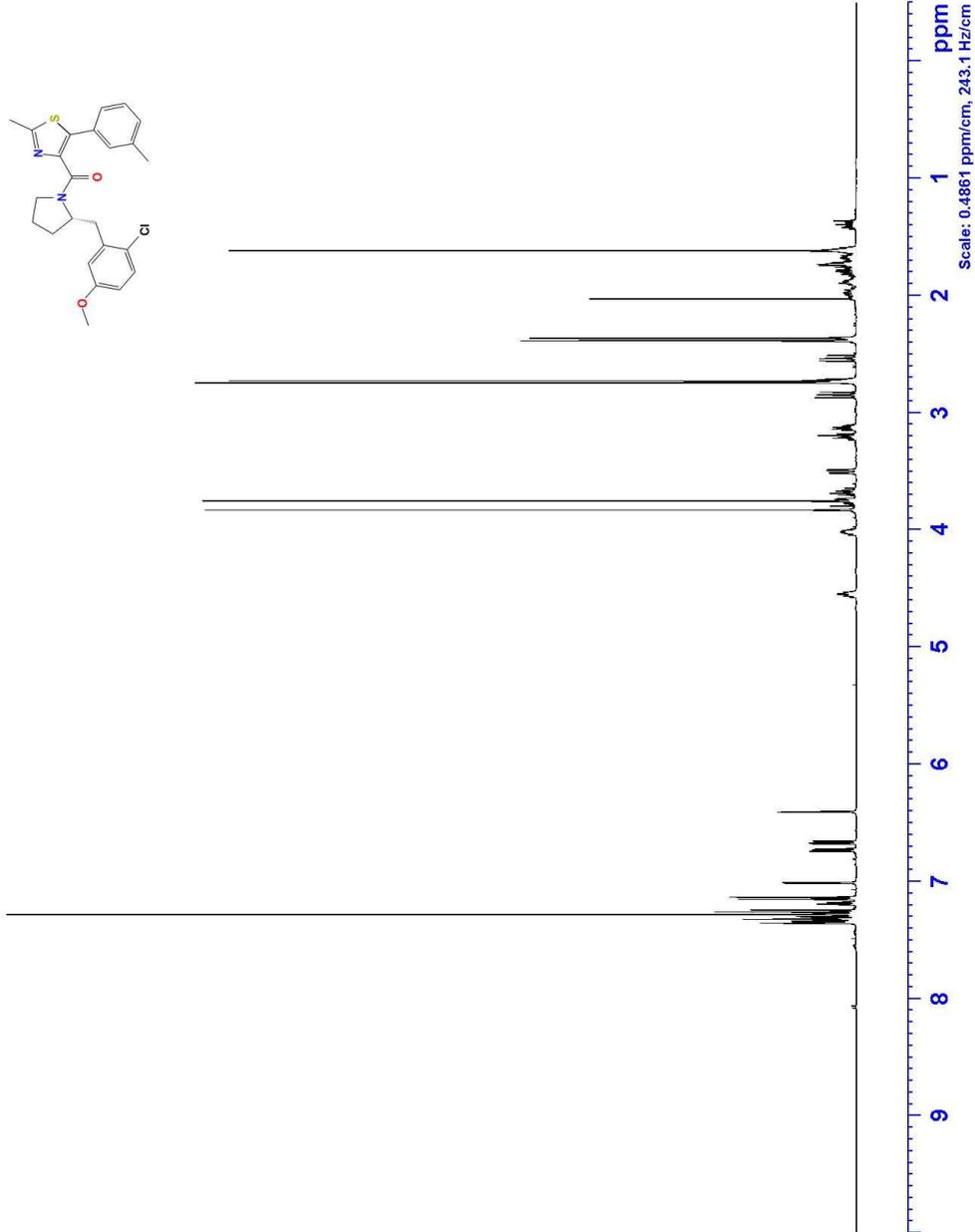


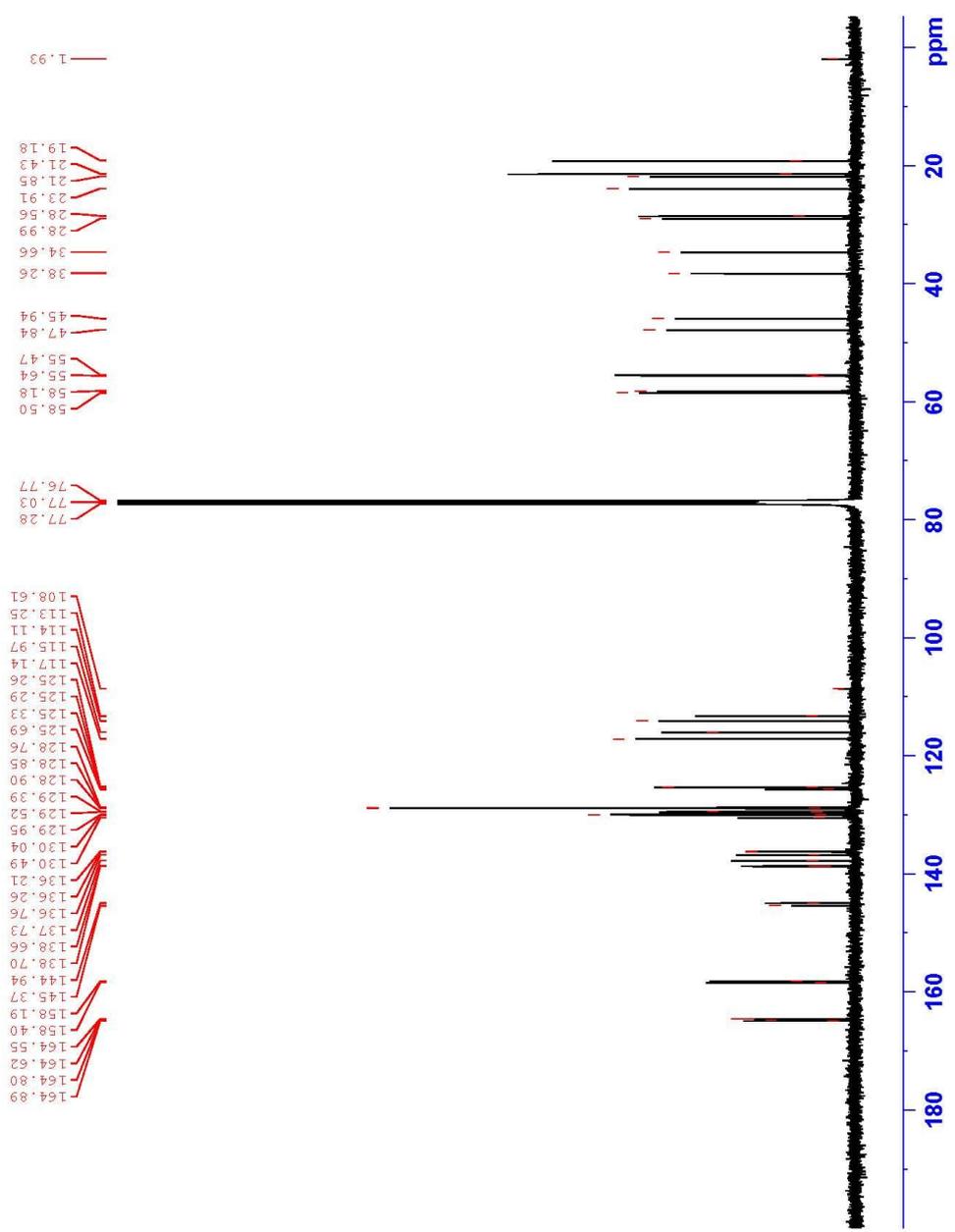
Current Data Parameters  
 NAME Compound 20  
 EXPNO 10  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20141118  
 Time 21:30  
 INSTRUM spect  
 PROBHD 5 mm CPDCH 13C  
 PULPROG zgpg30  
 TD 65536  
 CHAN13  
 SOLVENT  
 NS 16  
 DS 2  
 SWH 10000.000 Hz  
 FIDRES 0.152888 Hz  
 AQ 3.276793 sec  
 RG 409.6  
 DW 50.000 usec  
 DE 40.000 usec  
 TE 298.0 K  
 D1 1.00000000 sec  
 TDO 1

==== CHANNEL f1 =====  
 SF01 500.1300885 MHz  
 NUC1 1H  
 P1 9.25 usec  
 ELW1 13.0000000 W

F2 - Processing parameters  
 SI 65536  
 WDW EM  
 SSB 0  
 GB 0  
 PC 0 1.00





Current Data Parameters  
 Name: Compound 20  
 EXPNO: 2  
 PROCNO: 1

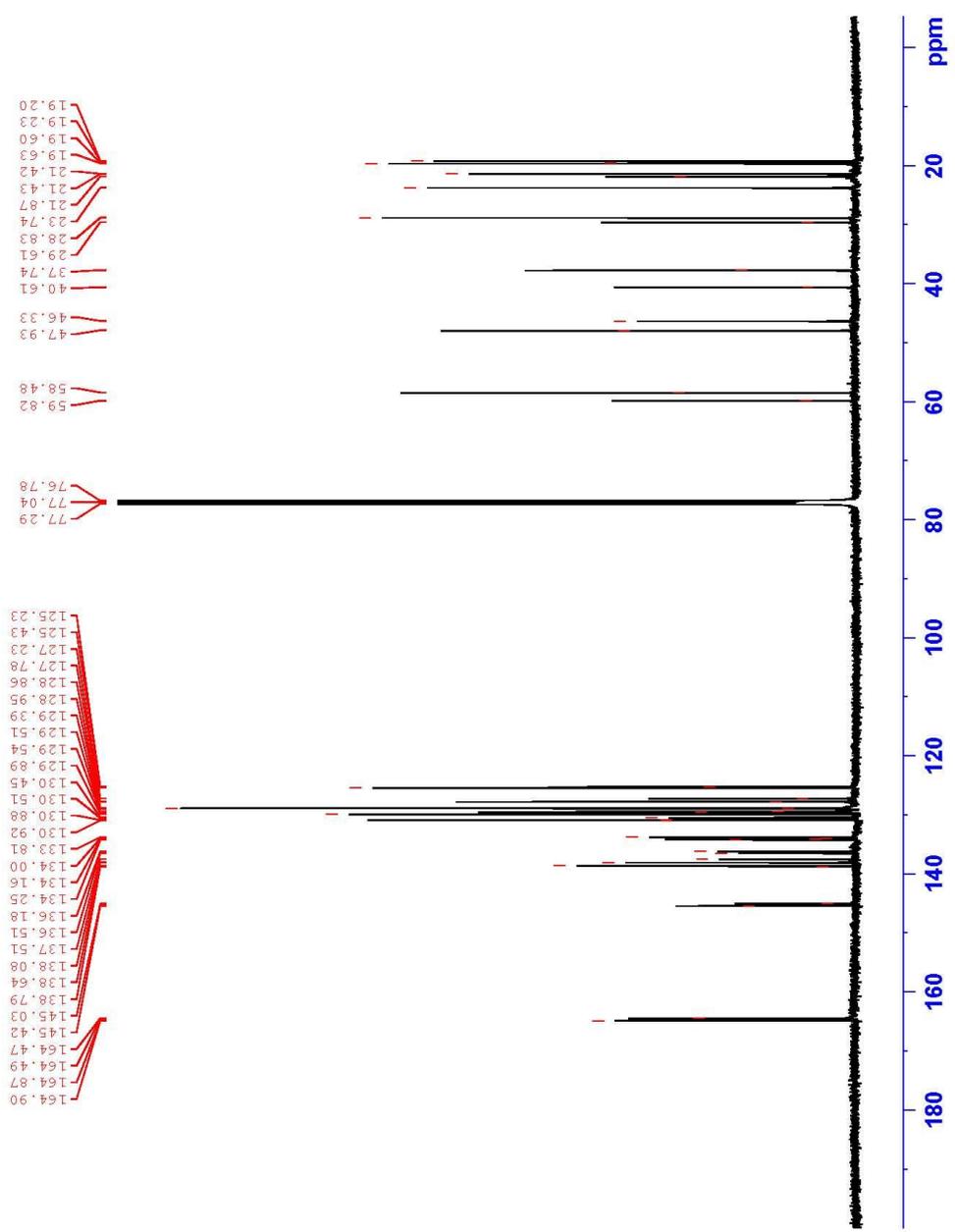
F2 - Acquisition Parameters  
 Date\_Time: 20111121 21:44  
 INSTRUM: spect  
 PROBD: 5 mm CPDCH 13C  
 PULPROG: zgpg30  
 TD: 65536  
 SOLVENT: CDCl3  
 NS: 256  
 DS: 4  
 SSB: 31512.605 Hz  
 AQ: 0.480844 Hz  
 FIDRES: 1.0298378 sec  
 RG: 106.667  
 RW: 15.667 usec  
 DE: 60.46 usec  
 TE: 298.0 K  
 D1: 2.0000000 sec  
 D11: 0.0300000 sec  
 TD0: 1

===== CHANNEL f1 =====  
 NUC1: 13C  
 P1: 11.00 usec  
 PL1: 18.0000000 N

===== CHANNEL f2 =====  
 SFO2: 500.1320005 MHz  
 NUC2: 1H  
 P2: 19.00 usec  
 PL2: 0.0000000 N  
 PLM2: 13.0000000 N  
 PLM3: 0.1738001 N  
 PLM4: 0.1112500 N

F2 - Processing parameters  
 SI: 32768  
 SF: 125.7577890 MHz  
 EQ: EN  
 SSB: 0  
 LB: 1.00 Hz  
 GB: 0  
 PC: 1.40





```

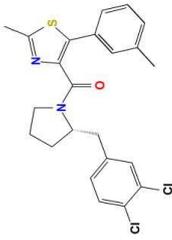
Current Data Parameters
Name      Compound 21
EXPNO    1
PROCNO   1

F2 - Acquisition Parameters
Date_    20160806
Time     1.06
INSTRUM  spect
PROBHD   5 mm CPDCH 13C
PULPROG  zgpg30
TD       65536
SOLVENT  CDCl3
NS       256
DS       4
SSBH     31512.605 Hz
AQ       0.480844 Hz
FIDRES   1.0298378 sec
RG       1000
DE       15.667 usec
TE       60.46 usec
TE       298.0 K
D1       2.0000000 sec
d11      0.0300000 sec
TD0      1

----- CHANNEL f1 -----
NUC1     13C
P1       11.00 usec
PL1      18.00000000 N

----- CHANNEL f2 -----
SFO2     500.1320005 MHz
NUC2     1H
P2       19.00 usec
PL2      0.00000000 N
PCPD2    30.00 usec
PLM2     13.00000000 N
PLM3     0.17380001 N
PLM4     0.11125000 N

F2 - Processing parameters
SI       32768
SF       125.7577890 MHz
WDW      EM
SSB      0
LB       1.00 Hz
GB       0
PC       1.40
  
```



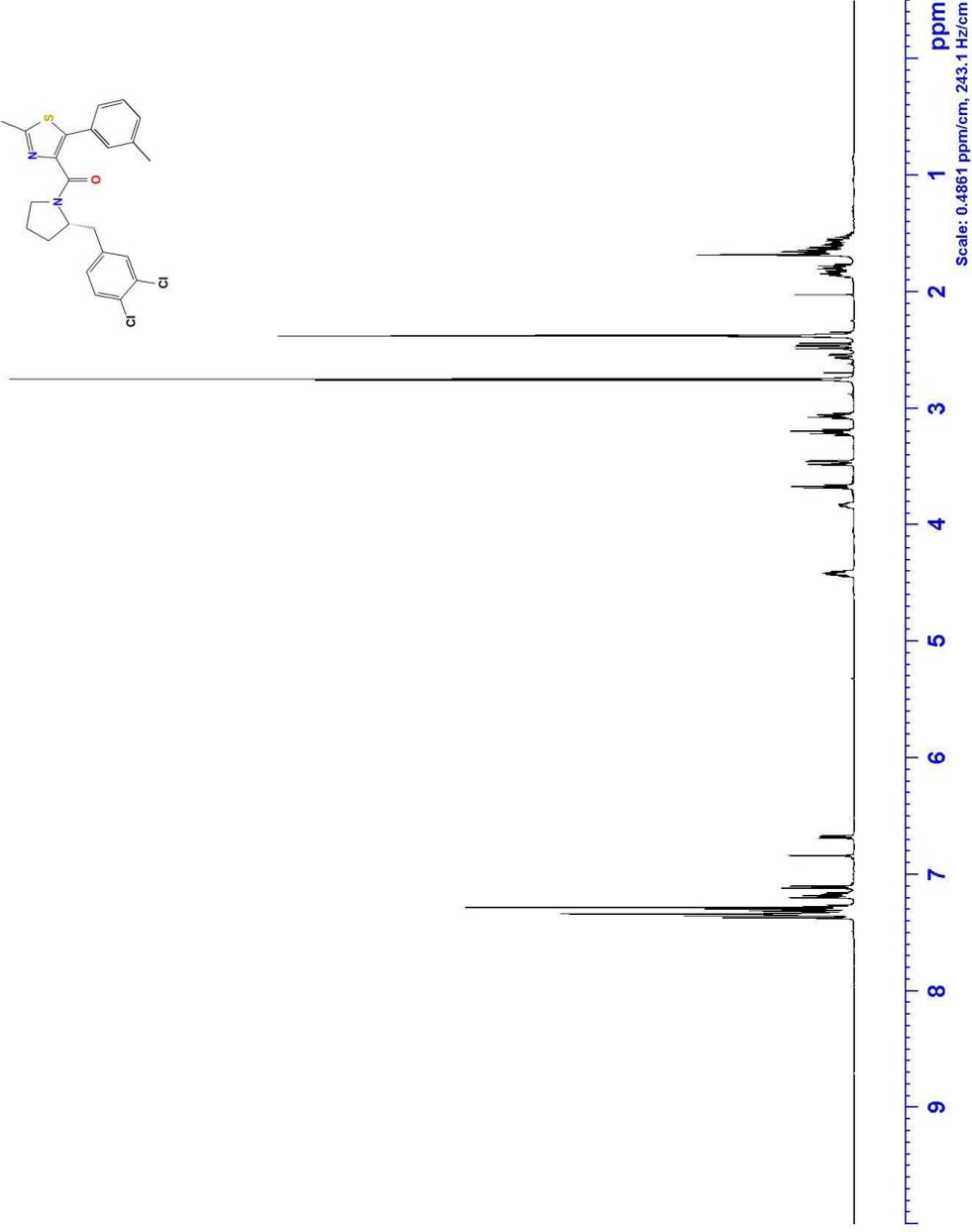
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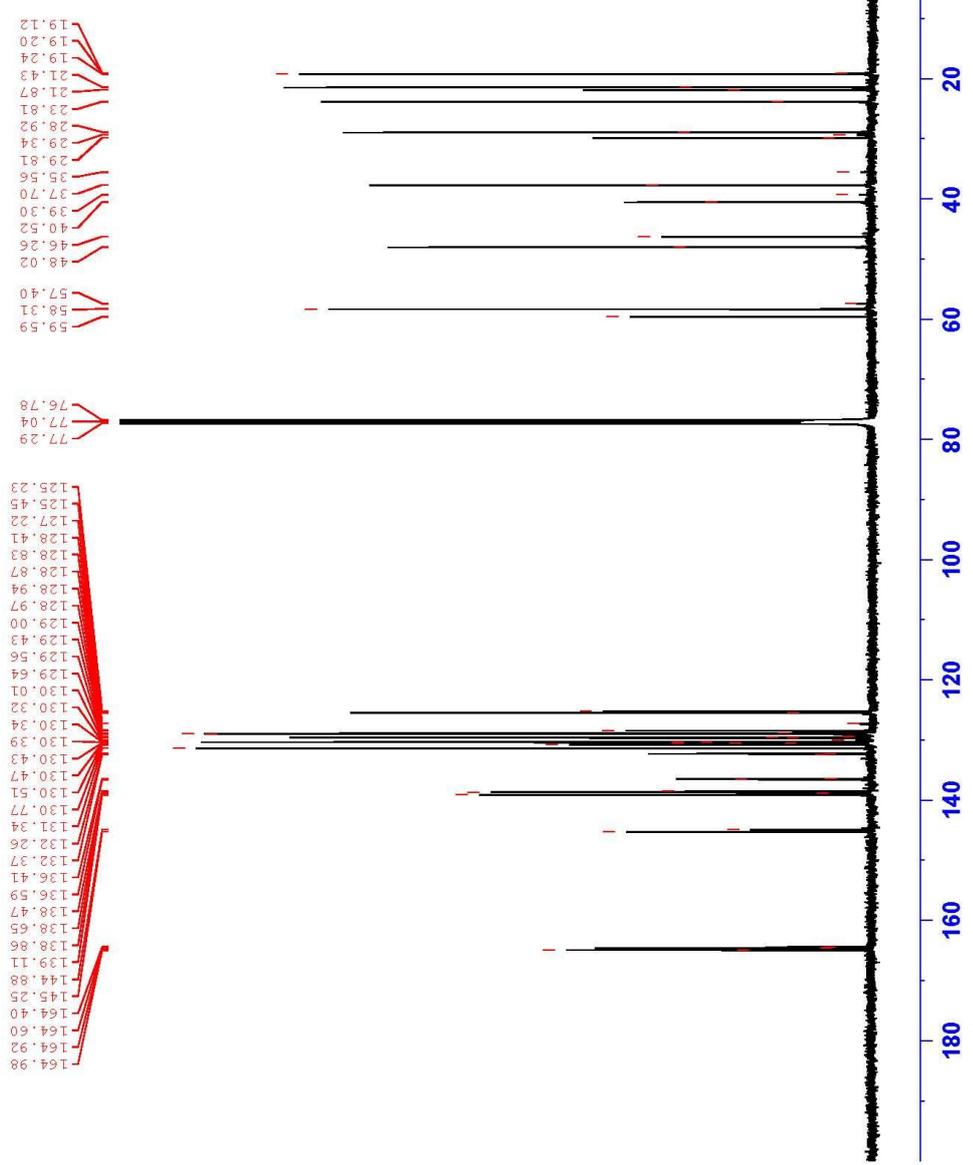
Current Data Parameters
NAME      Compound 22
EXPNO    10
PROCNO   1

F2 - Acquisition Parameters
Date_    20141111
Time     11.10
INSTRUM  spect
PROBHD   5 mm CPDCH 13C
PULPROG  zgpg30
TD       65536
SOLVENT  CDCl3
NS       16
DS       2
SWH      10000.000 Hz
FIDRES   0.152888 Hz
AQ       3.276797 sec
RG       14.601
DW       50.000 usec
DE       40.000 usec
TE       298.0 K
D1       1.00000000 sec
TD0      1

===== CHANNEL f1 =====
SF01    500.1300885 MHz
NUC1     13C
P1       9.25 usec
ELW1    13.00000000 W

F2 - Processing parameters
SI      65536
WDW     EM
SSB     0
GB      0
PC      1.00
  
```





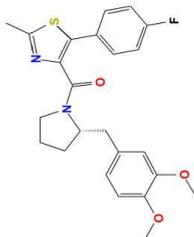
Current Data Parameters  
 Name: Compound 22  
 EXPNO: 1  
 PROCNO: 1

F2 - Acquisition Parameters  
 Date\_ Time: 20180816 1:24  
 INSTRUM: spect  
 PROBD: 5 mm CPDCH 13C  
 PULPROG: zgpg30  
 TD: 65536  
 SOLVENT: CDCl3  
 NS: 256  
 DS: 4  
 SSB: 31512.605 Hz  
 FIDRES: 0.480844 Hz  
 AQ: 1.0298378 sec  
 RG: 106.667  
 IN: 15.667  
 DE: 60.46 usec  
 TE: 298.0 K  
 D1: 2.0000000 sec  
 D11: 0.0300000 sec  
 TD0: 1

===== CHANNEL f1 =====  
 NUC1: 13C  
 P1: 11.00 usec  
 PL1: 18.00000000 N

===== CHANNEL f2 =====  
 SFO2: 500.1320005 MHz  
 NUC2: 1H  
 P2: 19.00 usec  
 PL2: 0.00000000 N  
 PLM2: 13.00000000 N  
 PLM3: 0.17380001 N  
 PLM13: 0.11125000 N

F2 - Processing parameters  
 SI: 32768  
 SF: 125.7577890 MHz  
 SSB: 0  
 LB: 1.00 Hz  
 GB: 0  
 PC: 1.40



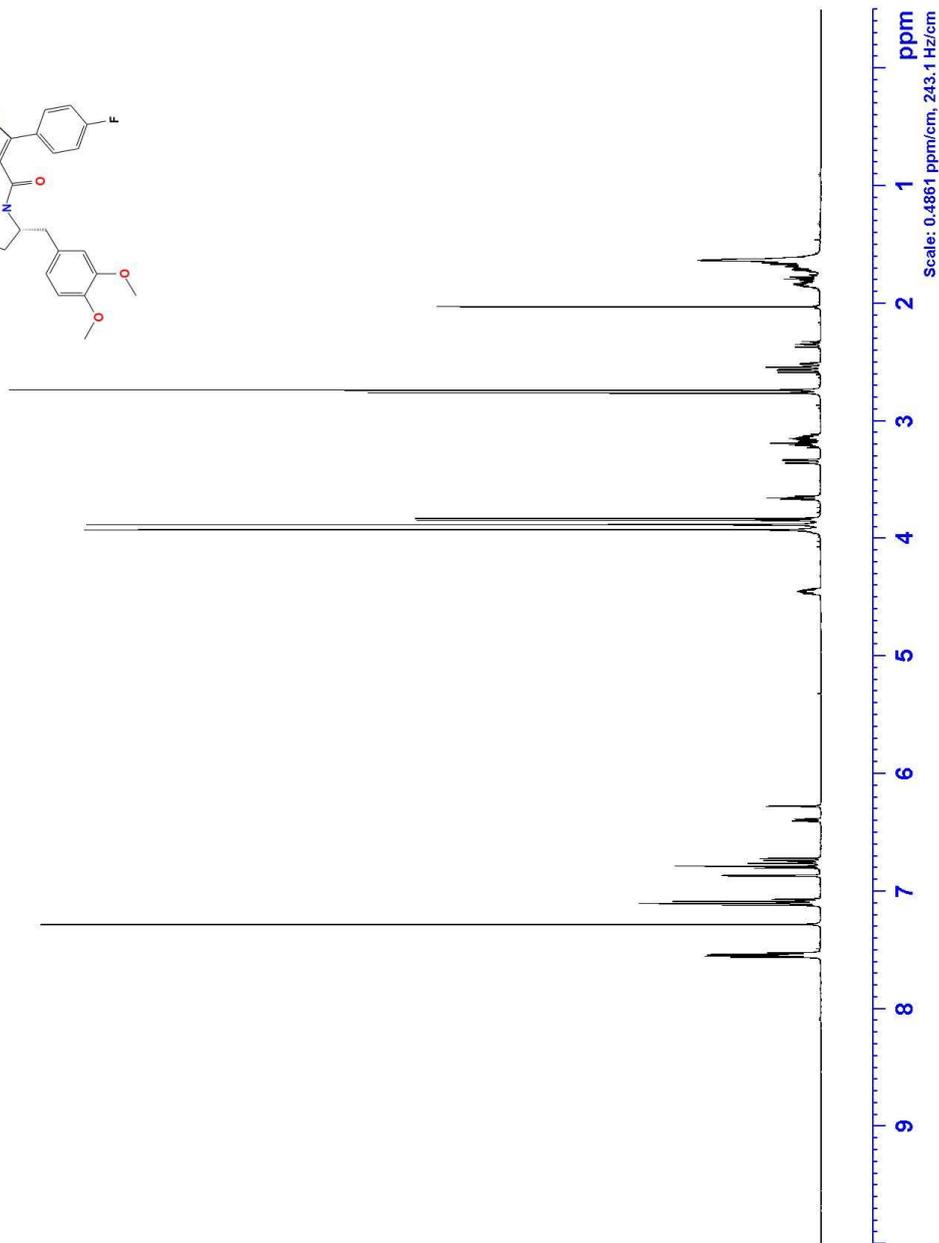
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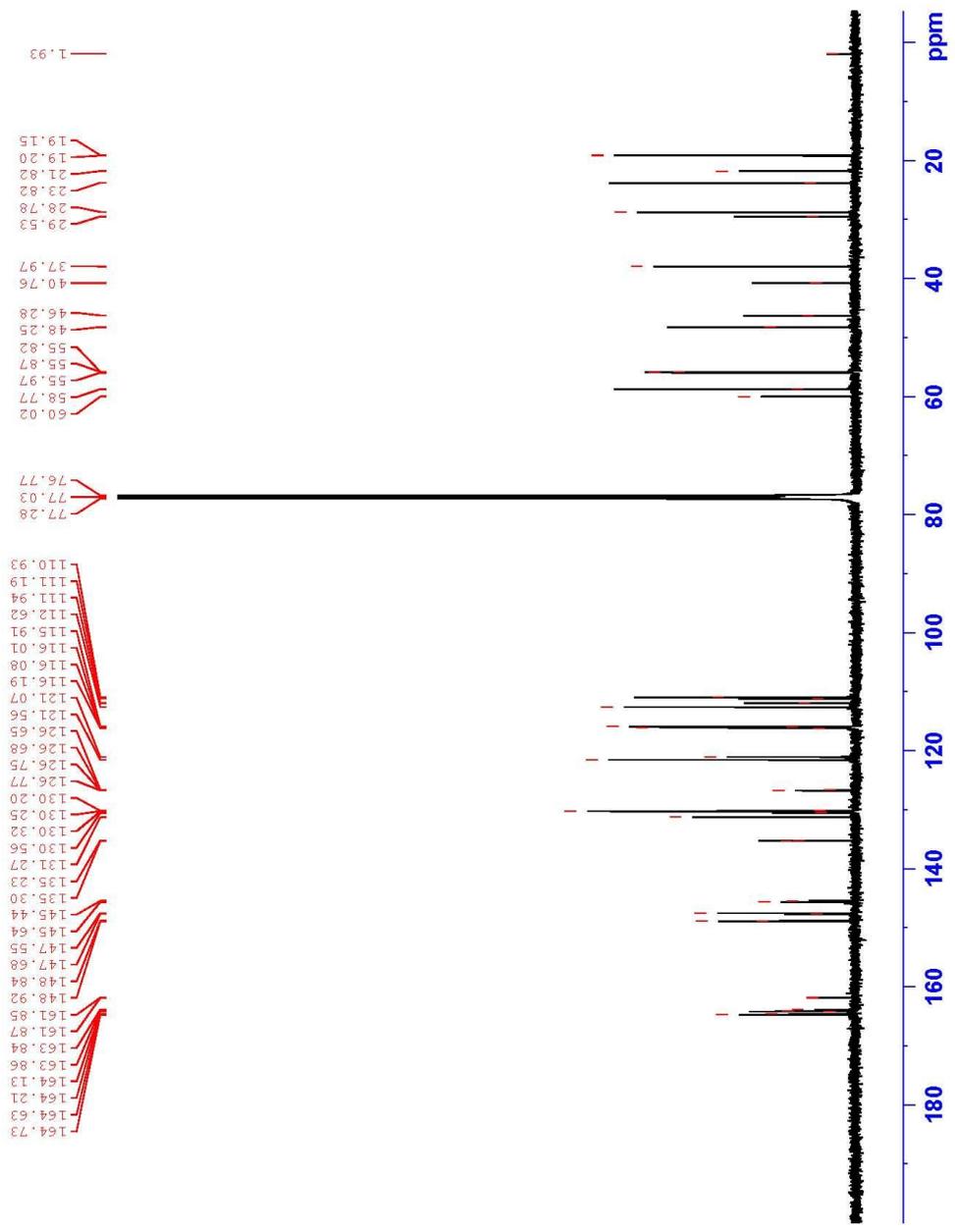
Current Data Parameters
NAME          Compound 23
EXPNO        10
PROCNO       1

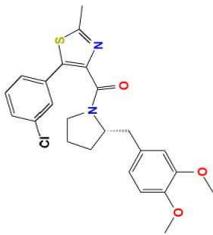
F2 - Acquisition Parameters
Date_         20141117
Time         21.12
INSTRUM      SPECT
PROBHD       5 mm CPDCH 13C
PULPROG      zgpg30
TD           65536
SOLVENT      CDCl3
NS           16
DS           2
SWH           10000.000 Hz
FIDRES       0.152888 Hz
AQ           3.276793 sec
RG           768.000
WDW           EM
SSB           0
DE           40.000 USSEC
TE           298.0 K
D1           1.00000000 sec
TD0          1

===== CHANNEL f1 =====
SF01        500.1330885 MHz
NUC1         1H
P1           9.25 usec
ELW1        13.00000000 W

F2 - Processing parameters
SI           65536
WDW          EM
SSB          0
GB          0
PC          1.00
  
```





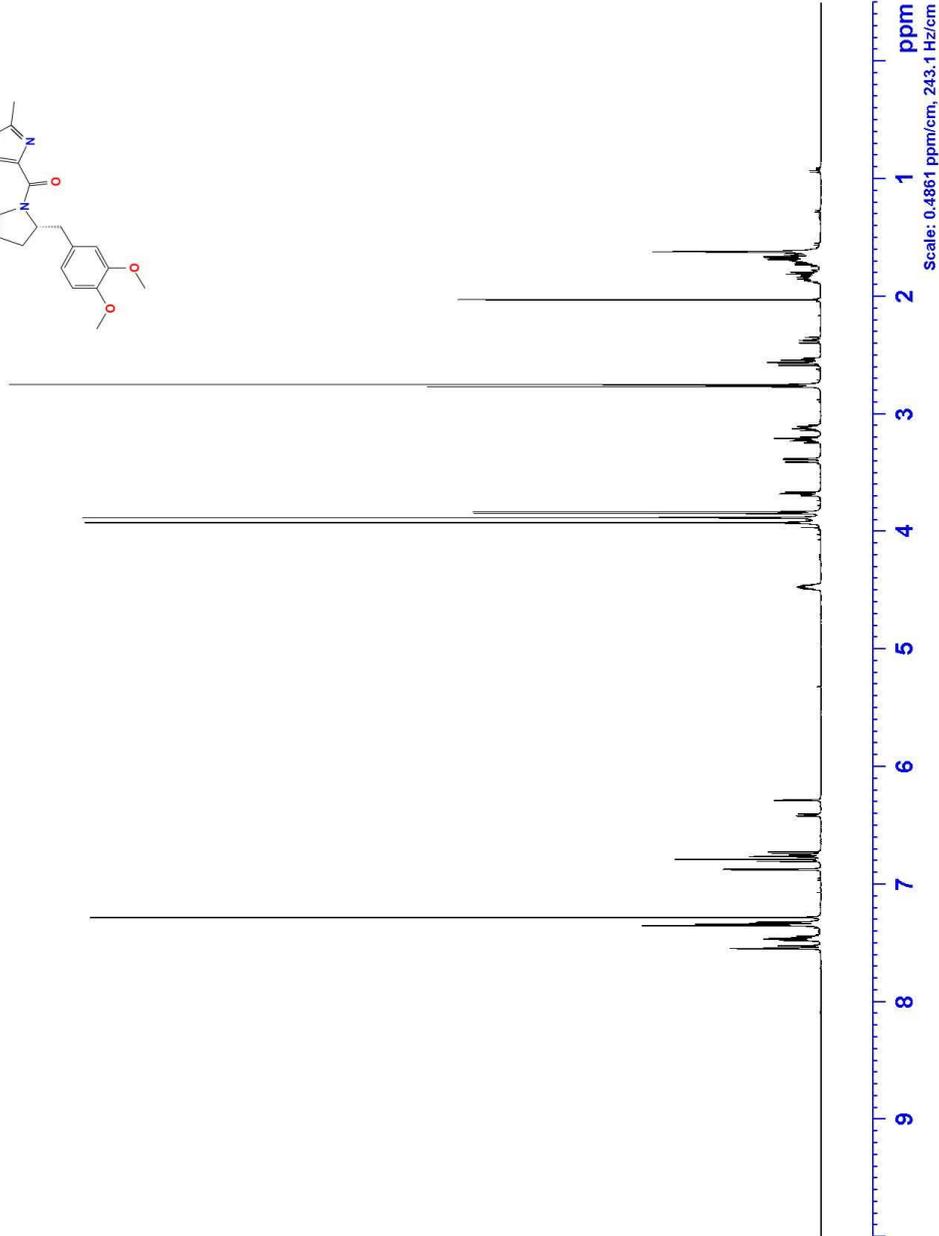


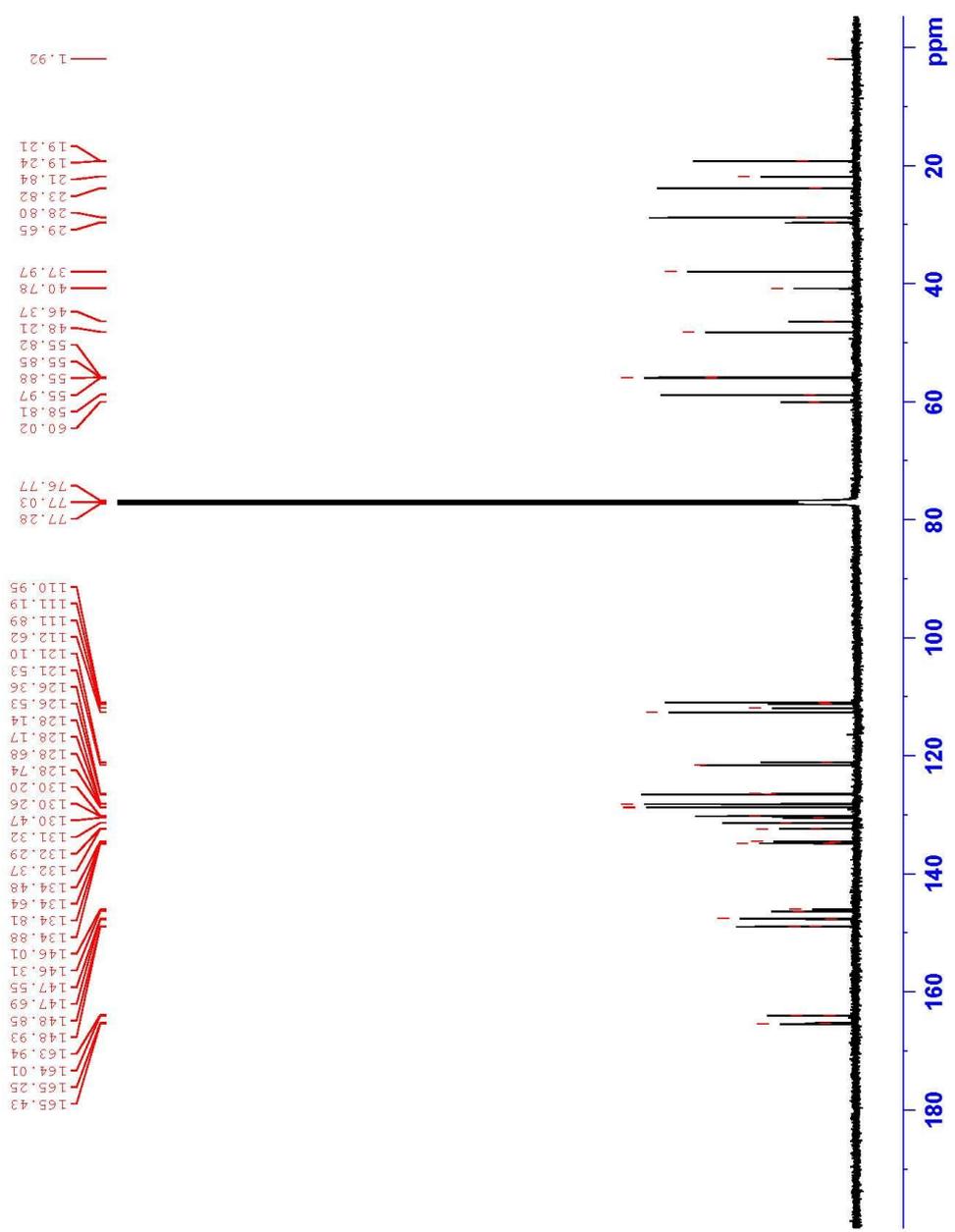
Current Data Parameters  
 NAME Compound 24  
 EXPNO 10  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20141111  
 Time 17.01  
 INSTRUM SPECT  
 PROBHD 5 mm CPDCH 13C  
 PULPROG zgpg30  
 TD 65536  
 CHAN13  
 SOLVENT  
 NS 16  
 DS 2  
 SWH 10000.000 Hz  
 SFR 0.152888 Hz  
 AQ 3.276793 sec  
 RG 409.6  
 DW 50.000 USEC  
 DE 40.000 USEC  
 TE 298.0 K  
 D1 1.00000000 sec  
 TDO 1

==== CHANNEL f1 =====  
 SF01 500.1330885 MHz  
 NUC1 1H  
 P1 9.25 usec  
 ELW1 13.0000000 W

F2 - Processing parameters  
 SI 65536  
 WDW EM  
 SSB 0  
 GB 0  
 PC 0 1.00





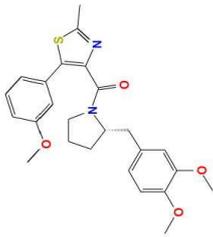
Current Data Parameters  
 Name: Compound\_24  
 EXPNO: 4  
 PROCNO: 1

F2 - Acquisition Parameters  
 Date\_ Time: 20180808 8:53  
 INSTRUM: spect  
 PROBD: 5 mm CPDCH 13C  
 PULPROG: zgpg30  
 TD: 65536  
 SOLVENT: CDCl3  
 NS: 256  
 DS: 4  
 SSBH: 31512.605 Hz  
 FIDRES: 0.480844 Hz  
 AQ: 1.0298378 sec  
 RG: 106.667  
 RW: 15.667 usec  
 DE: 60.46 usec  
 TE: 298.0 K  
 D1: 2.0000000 sec  
 D11: 0.0300000 sec  
 TD0: 1

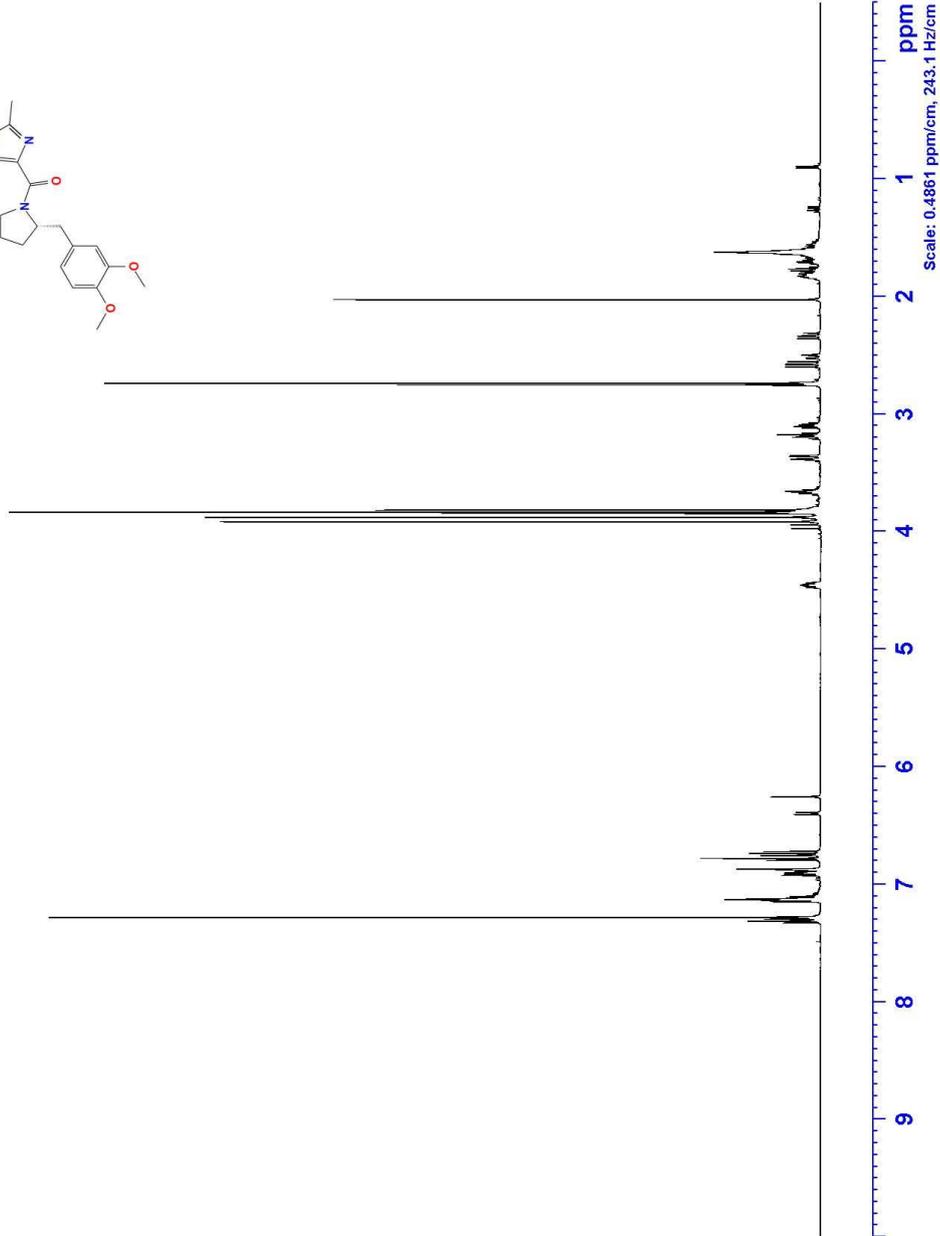
===== CHANNEL f1 =====  
 NUC1: 13C  
 P1: 11.00 usec  
 PL1: 18.00000000 N

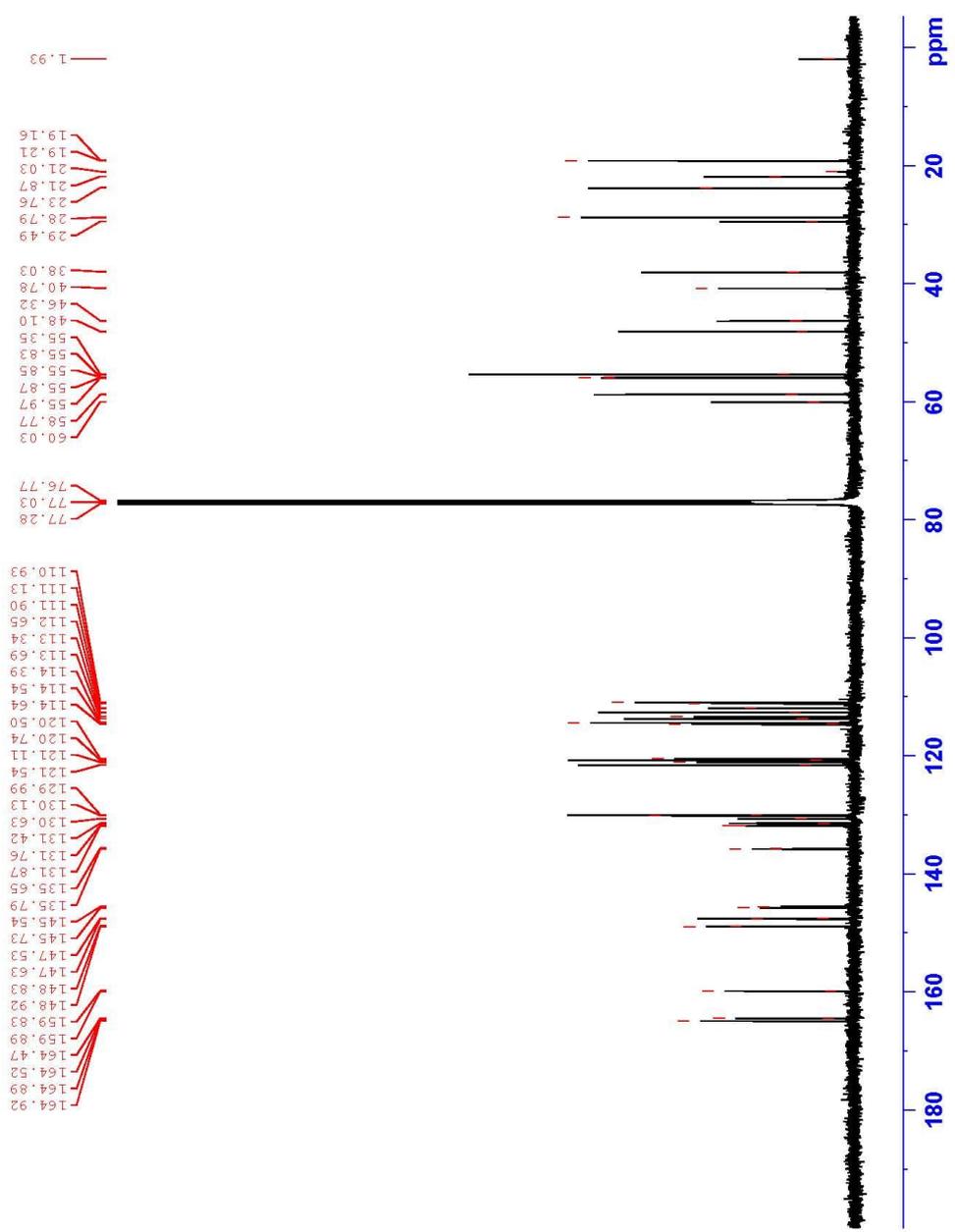
===== CHANNEL f2 =====  
 SFO2: 500.1320005 MHz  
 NUC2: 1H  
 P2: 19.00 usec  
 PL2: 0.00000000 N  
 SFO1: 125.7611000 MHz  
 NUC1: 13C  
 P1: 11.00 usec  
 PL1: 18.00000000 N

F2 - Processing parameters  
 SI: 32768  
 SF: 125.7577890 MHz  
 EQ: BR  
 SSB: 0  
 LB: 1.00 Hz  
 GB: 0  
 PC: 1.40



Current Data Parameters  
 NAME Compound 25  
 EXPNO 10  
 PROCNO 1  
 F2 - Acquisition Parameters  
 Date\_ 20111114  
 Time 22.44  
 INSTRUM SPECT  
 PROBHD 5 mm CPDCH 13C  
 PULPROG zgpg30  
 TD 65536  
 CHAN13  
 SOLVENT  
 NS 16  
 DS 2  
 SWH 10000.000 Hz  
 SRH 0.152888 Hz  
 FIDRES 3.276793 SEC  
 AQ 76.000000 SEC  
 RG 655.36  
 DW 50.000 USEC  
 DE 40.000 USEC  
 TE 298.0 K  
 D1 1.00000000 sec  
 TDO 1  
 ===== CHANNEL f1 =====  
 SF01 500.1300885 MHz  
 NUC1 1H  
 P1 9.25 usec  
 ELW1 13.0000000 W  
 F2 - Processing parameters  
 SI 65536  
 WDW EM  
 SSB 0  
 GB 0  
 PC 0 1.00





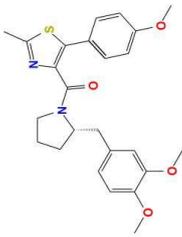
Current Data Parameters  
 Name: Compound 25  
 EXPNO: 1  
 PROCNO: 1

F2 - Acquisition Parameters  
 Date\_ Time: 2010-02-22 22:58  
 INSTRUM: spect  
 PROBD: 5 mm CPDCH 13C  
 PULPROG: zgpg30  
 TD: 65536  
 SOLVENT: CDCl3  
 NS: 256  
 DS: 4  
 SSB: 31512.605 Hz  
 FIDRES: 0.480844 Hz  
 AQ: 1.0298378 sec  
 RG: 1000  
 INEG: 15.667 usec  
 DE: 60.46 usec  
 TE: 298.0 K  
 D1: 2.0000000 sec  
 D11: 0.0300000 sec  
 TD0: 1

----- CHANNEL f1 -----  
 NUC1: 13C  
 P1: 11.00 usec  
 PL1: 18.00000000 N

----- CHANNEL f2 -----  
 SFO2: 500.1320005 MHz  
 NUC2: 13C  
 P2: 11.00 usec  
 PL2: 18.00000000 N  
 WALTZ16: 1H  
 WALTZ16: 40.00 usec  
 WALTZ16: 40.00 usec  
 FLM2: 13.00000000 N  
 FLM12: 0.17380001 N  
 FLM13: 0.11125000 N

F2 - Processing parameters  
 SI: 32768  
 SF: 125.7577890 MHz  
 WID: 6553.6  
 SSB: 0  
 LB: 1.00 Hz  
 GB: 0  
 PC: 1.40



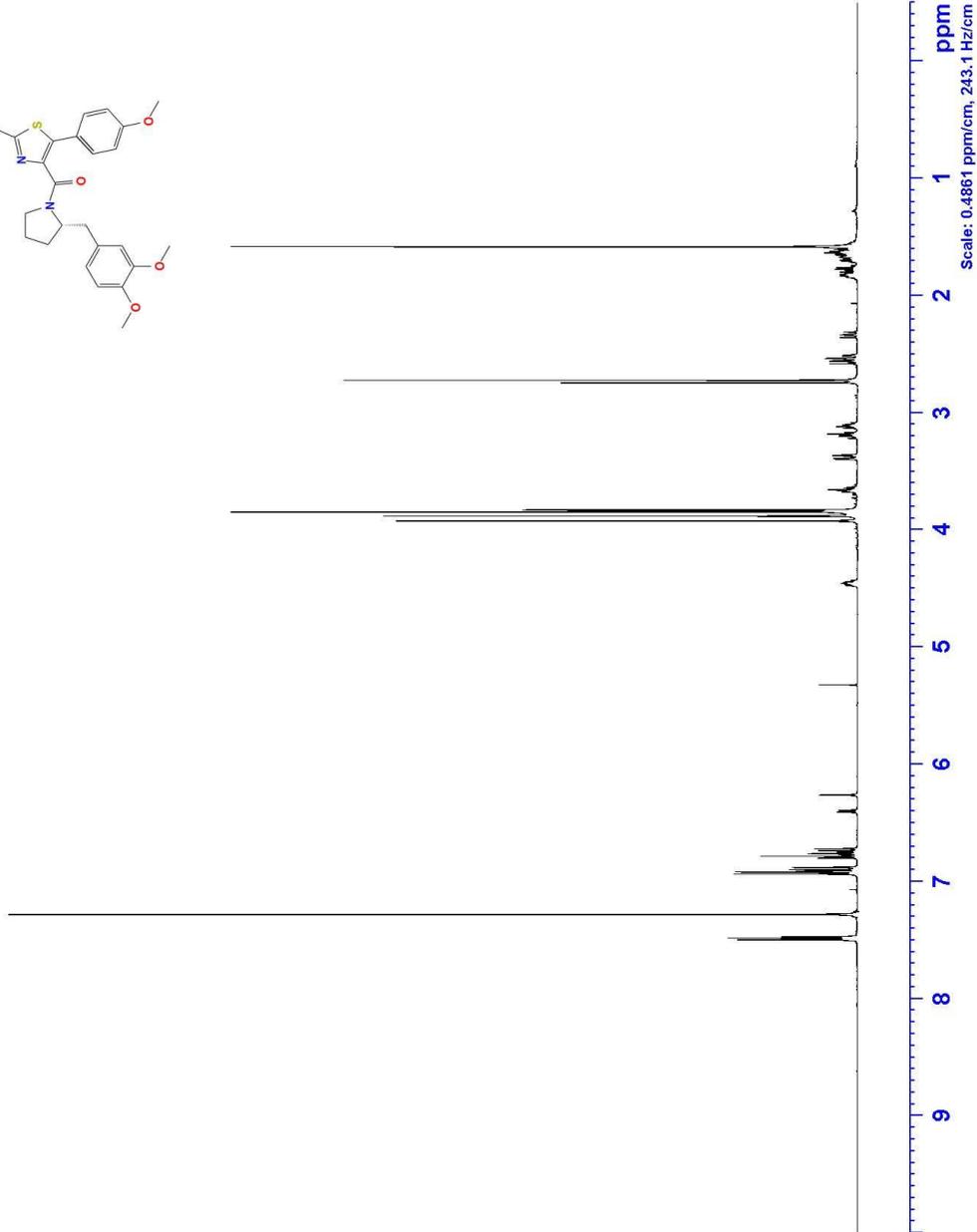
```

Current Data Parameters
NAME          Compound 26
EXPNO        10
PROCNO       1

F2 - Acquisition Parameters
Date_         20141115
Time         15.15
INSTRUM      SPECT
PROBHD       5 mm CPDCH 13C
PULPROG      zgpg30
TD           65536
SOLVENT      CDCl3
NS           16
DS           2
SWH           10000.000 Hz
FIDRES       0.152888 Hz
AQ           3.276793 sec
RG           768.000
WDW           EM
SSB           0
DE           40.00 USSEC
TE           298.0 K
D1           1.00000000 sec
TD0          1

===== CHANNEL f1 =====
SF01        500.1300885 MHz
NUC1         13C
P1           9.25 usec
ELW1        13.00000000 W

F2 - Processing parameters
SI           65536
WDW         EM
SSB         0
GB         0
PC         1.00
  
```



```

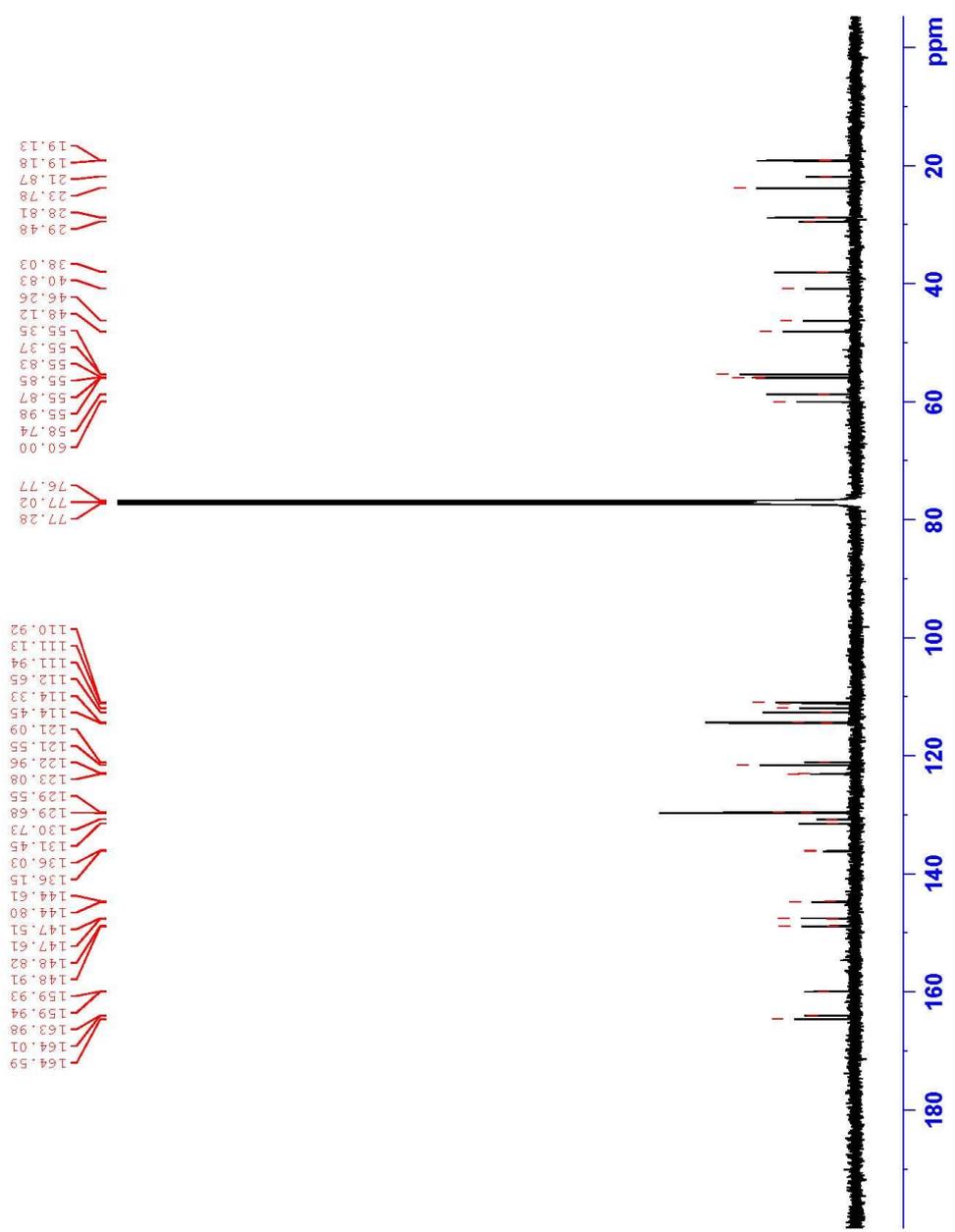
Current Data Parameters
Name          Compound 26
EXPNO        1
PROCNO       1

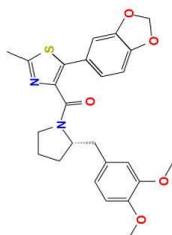
F2 - Acquisition Parameters
Time         2015.15.29
INSTRUM      spect
PROBHD       5 mm CPDCH 13C
PULPROG      zgpg30
TD           65536
SOLVENT      CDCl3
NS           256
DS           4
SWH           31512.605 Hz
FIDRES       0.480844 Hz
AQ           1.0298378 sec
RG           106.667
DE           15.667 usec
TE           60.46 usec
TE           298.0 K
D1           2.0000000 sec
D11          0.0300000 sec
TD0          1

----- CHANNEL f1 -----
NUC1         13C
P1           11.00 usec
PL1         18.00000000 N

----- CHANNEL f2 -----
SFO2         500.1320005 MHz
NUC2         1H
P2           12.00 usec
PL2         0.00000000 N
SFO1         125.7620000 MHz
NUC1         13C
P1           11.00 usec
PL1         18.00000000 N

----- Processing parameters -----
SI           32768
SF           125.7577890 MHz
WDW          EM
SSB          0
LB           1.00 Hz
GB           0
PC           1.40
  
```



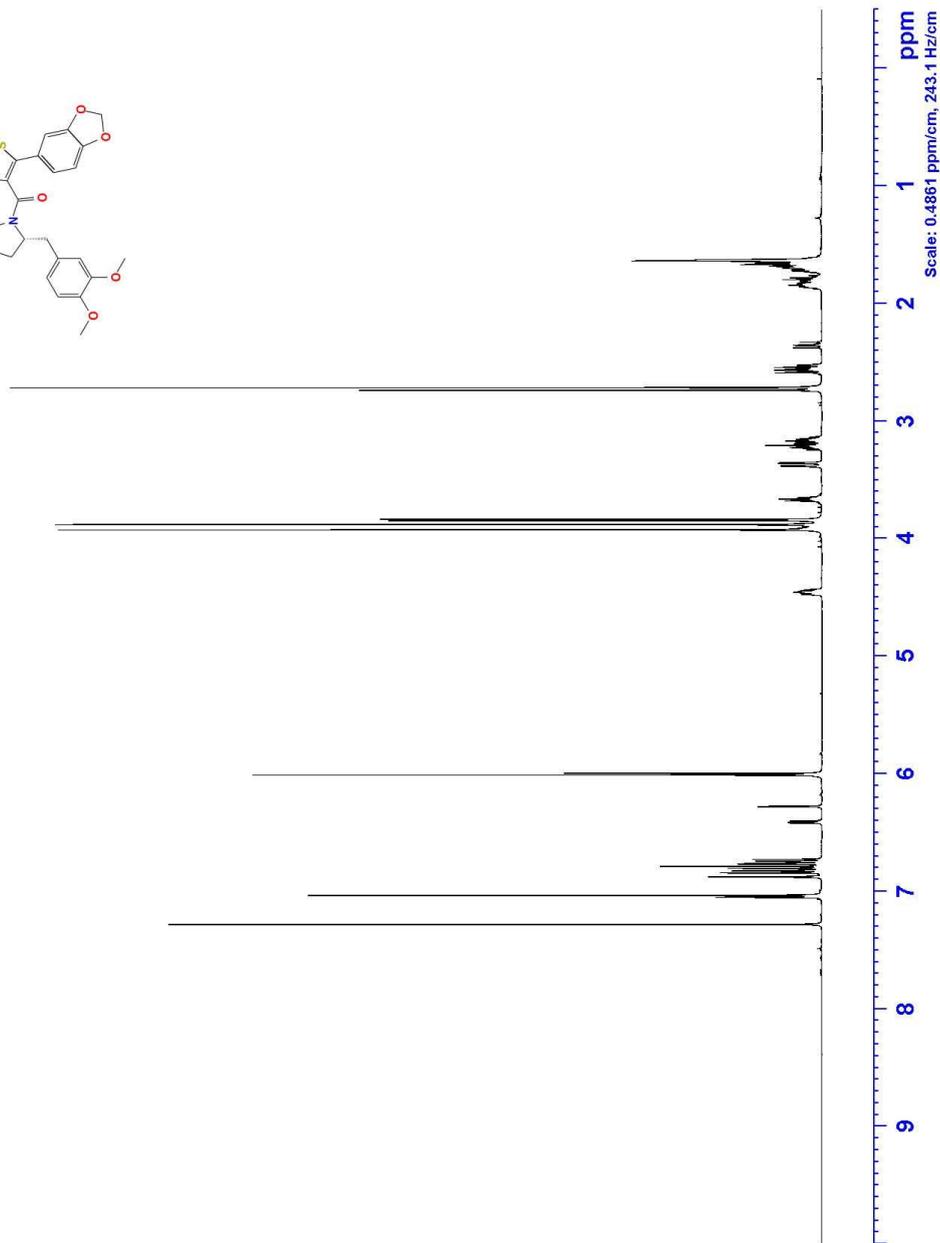


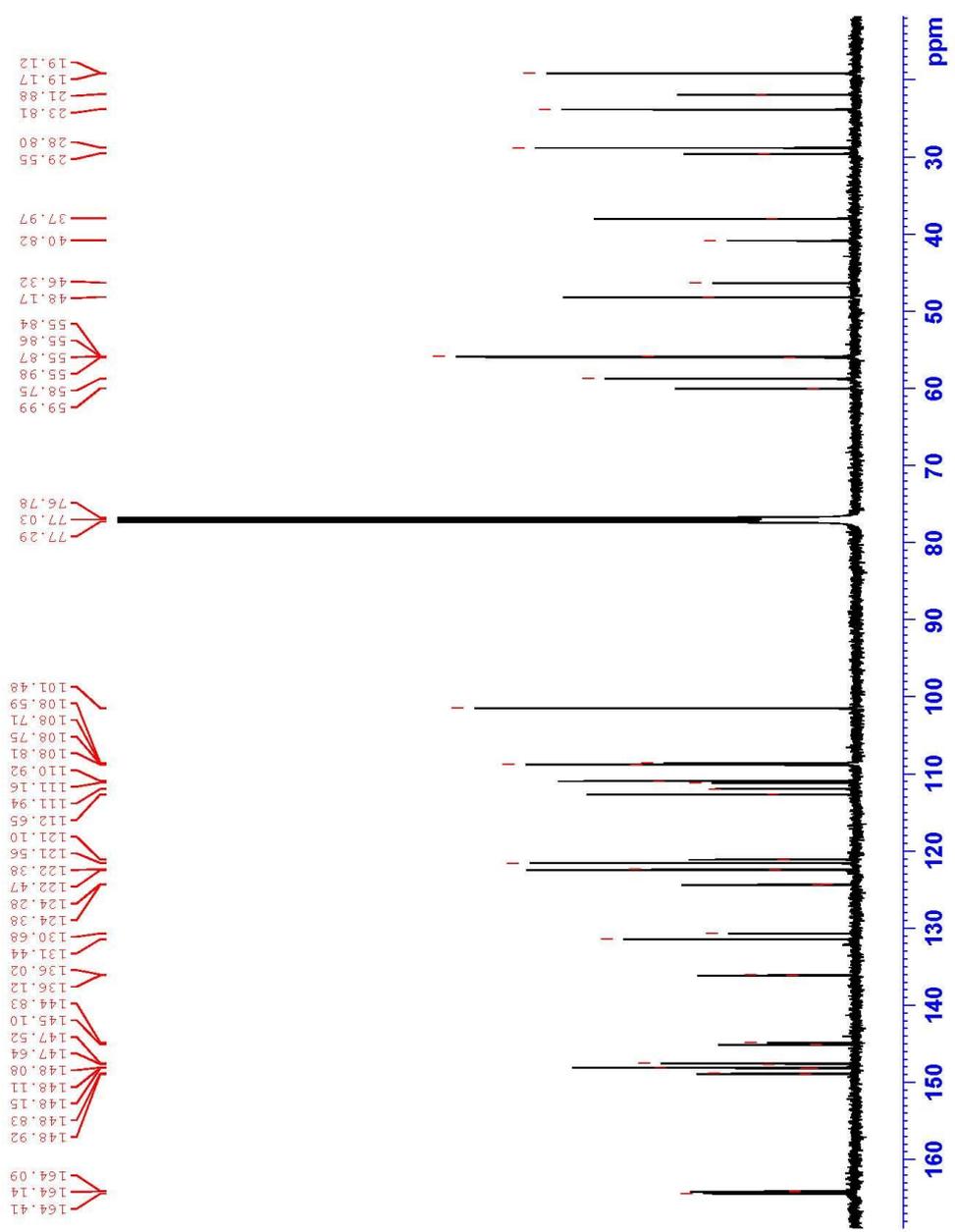
Current Data Parameters  
 NAME Compound 27  
 EXPNO 10  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20111111  
 Time 16.43  
 INSTRUM SPECT  
 PROBHD 5 mm CPDCH 13C  
 PULPROG zgpg30  
 TD 65536  
 CHAN13  
 SOLVENT  
 NS 16  
 DS 2  
 SWH 10000.000 Hz  
 SRH 0.152888 Hz  
 FIDRES 3.276797 SEC  
 AQ 14.601  
 RG 409.600  
 DW 50.000 USEC  
 DE 40.000 USEC  
 TE 298.0 K  
 D1 1.00000000 sec  
 TDO 1

===== CHANNEL f1 =====  
 SF01 500.1330885 MHz  
 NUC1 1H  
 P1 9.25 USEC  
 ELW1 13.0000000 W

F2 - Processing parameters  
 SI 65536  
 WDW EM  
 SSB 0  
 GB 0  
 PC 1.00





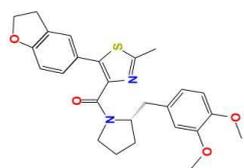
Current Data Parameters  
 ExpNO 27  
 Compound 41  
 PROCNO 1

F2 - Acquisition Parameters  
 Time 16.57  
 Date\_ 20160716  
 INSTRUM spect  
 PROBD 5 mm CPDCH 13C  
 PULPROG zgpg30  
 TD 65536  
 SOLVENT CDCl3  
 NS 256  
 DS 4  
 SSB 31512.605 Hz  
 FIDRES 0.480844 Hz  
 AQ 1.0298378 sec  
 RG 1024  
 RW 15.687 usec  
 DE 60.46 usec  
 TE 298.0 K  
 D1 2.0000000 sec  
 D11 0.0300000 sec  
 TD0 1

----- CHANNEL f1 -----  
 NUC1 133.726133 MHz  
 P1 11.00 usec  
 PL1 18.00000000 N

----- CHANNEL f2 -----  
 SFO2 500.1320005 MHz  
 NUC2 1H  
 P2 19.00 usec  
 PL2 18.00000000 N  
 SFO2 13.00000000 MHz  
 NUC2 133.726133 MHz  
 P1 11.00 usec  
 PL1 18.00000000 N

F2 - Processing parameters  
 SI 32768  
 SF 500.1320005 MHz  
 DS 4  
 SSB 0  
 LB 1.00 Hz  
 GB 0  
 PC 1.40

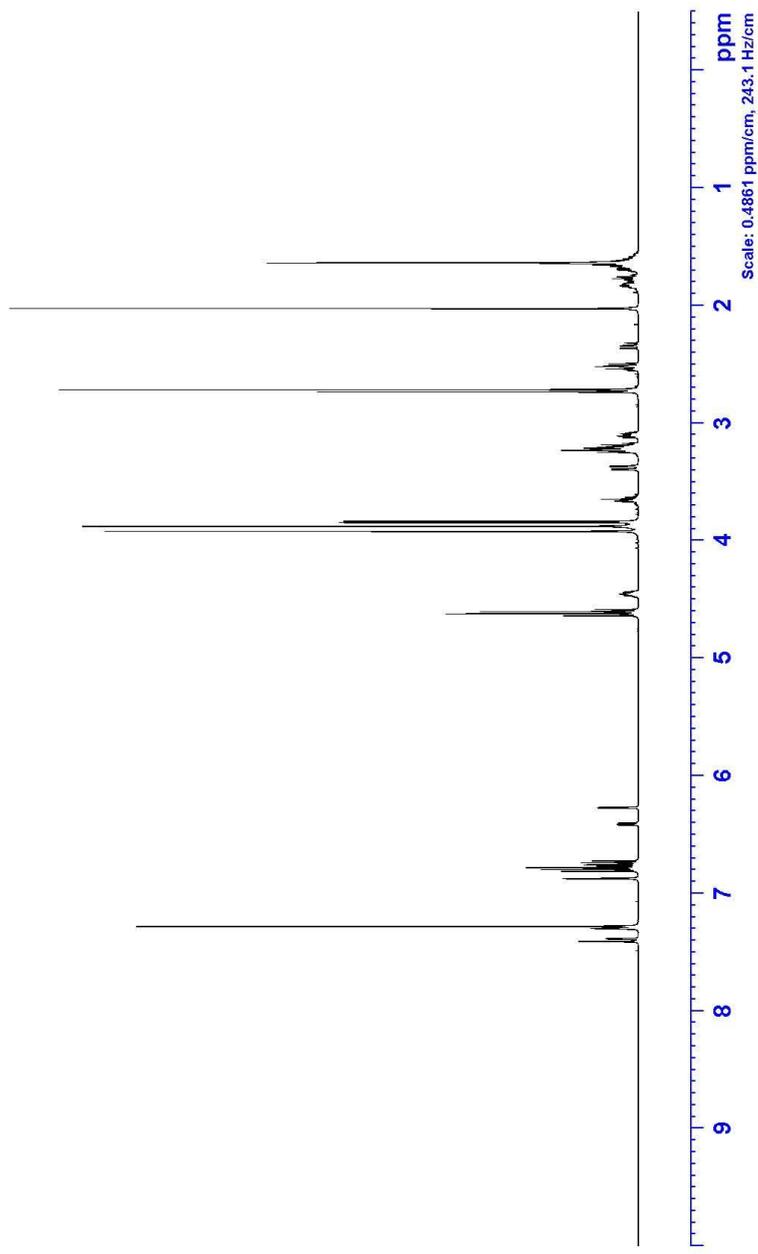


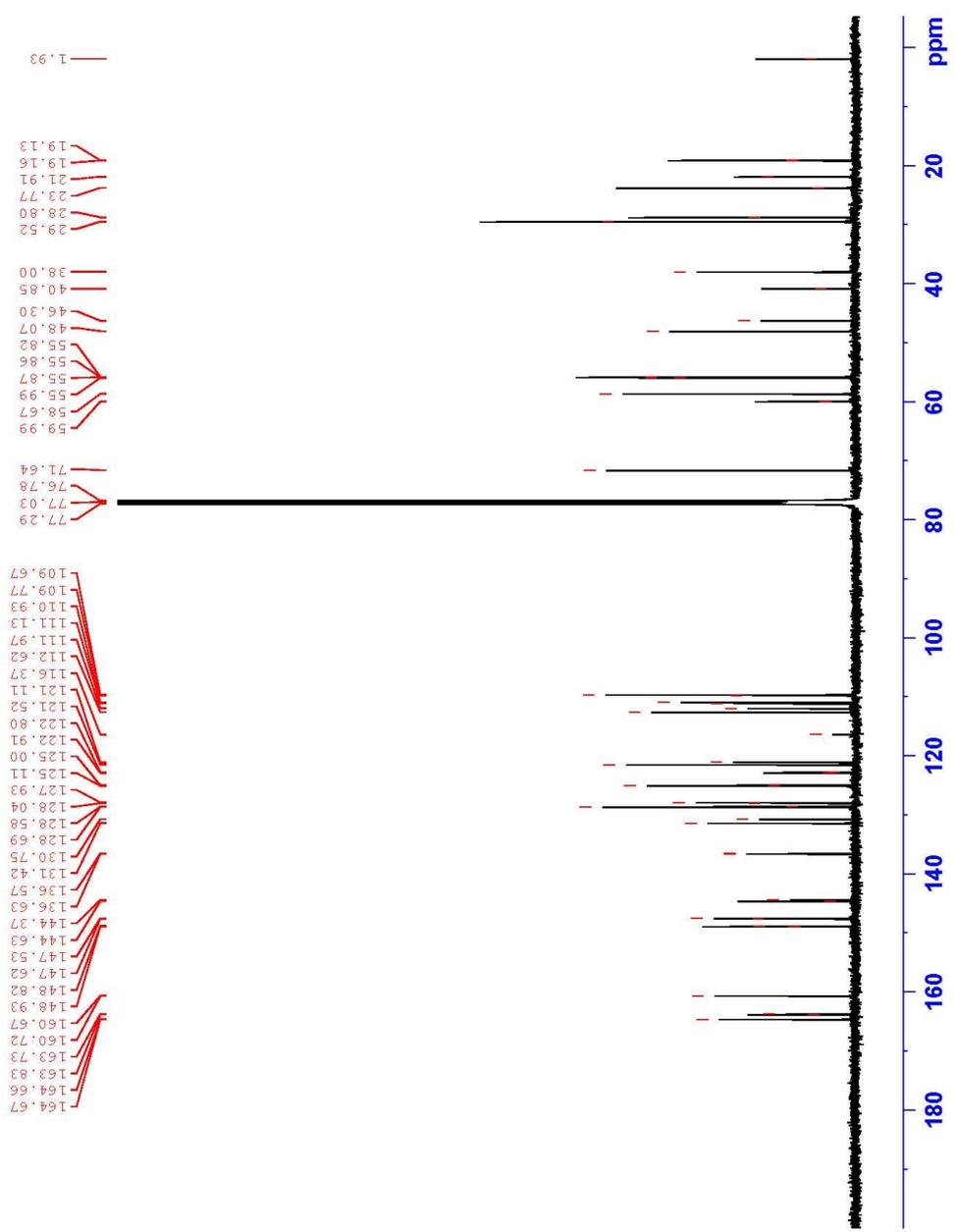
Current Data Parameters  
 NAME Compound 28  
 EXPNO 10  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20111111  
 Time 17:38  
 INSTRUM spect  
 PROBHD 5 mm CPDCH 13C  
 PULPROG zgpg30  
 TD 65536  
 CHAN13  
 SOLVENT  
 NS 16  
 DS 2  
 SWH 10000.000 Hz  
 FIDRES 0.152888 Hz  
 AQ 3.276792 sec  
 RG 409.52  
 DW 50.000 usec  
 DE 40.000 usec  
 TE 298.0 K  
 D1 1.00000000 sec  
 TDO 1

==== CHANNEL f1 =====  
 SF01 500.1300885 MHz  
 NUC1 1H  
 P1 9.25 usec  
 ELW1 13.0000000 W

F2 - Processing parameters  
 SI 65536  
 WDW EM  
 SSB 0  
 GB 0  
 PC 0 1.00





Current Data Parameters  
 Name Compound 28  
 EXPNO 1  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ Time 2017.07.17 17:52  
 INSTRUM spect  
 PROBD 5 mm CPDCH 13C  
 PULPROG zgpg30  
 TD 65536  
 SOLVENT CDCl3  
 NS 256  
 DS 4  
 SSB 31512.605 Hz  
 FIDRES 0.480844 Hz  
 AQ 1.0298378 sec  
 RG 1060  
 RW 15.667 usec  
 DE 60.46 usec  
 TE 298.0 K  
 D1 2.0000000 sec  
 D11 0.0300000 sec  
 TD0 1

----- CHANNEL f1 -----  
 NUC1 13C  
 P1 11.00 usec  
 PL1 18.0000000 N

----- CHANNEL f2 -----  
 SFO2 500.1320005 MHz  
 NUC2 1H  
 P2 19.00 usec  
 PL2 13.0000000 N  
 PLM2 0.1738001 N  
 PLM3 0.1112500 N

F2 - Processing parameters  
 SI 32768  
 SF 125.7577890 MHz  
 EQ  
 SSB 0  
 LB 0 1.00 Hz  
 GB 0  
 PC 1.40

## Biological protocols:

### Intracellular calcium release assays

Chinese hamster ovary (CHO) cells expressing the human or rat orexin-1 receptor or orexin-2 receptor, respectively, were grown in culture medium (Ham F-12 with L-Glutamine) containing 300 µg/mL G418, 100 U/mL penicillin, 100 µg/mL streptomycin and 10% heat inactivated fetal calf serum (FCS). The cells were seeded at 20,000 cells / well into 384-well black clear bottom sterile plates (Greiner). The seeded plates were incubated overnight at 37°C in 5% CO<sub>2</sub>.

Orexin-A as an agonist was prepared as 1 mM stock solution in MeOH: water (1:1), diluted in Hanks balanced salt solution (HBSS) containing 0.1% bovine serum albumin (BSA), NaHCO<sub>3</sub>: 0.375 g/L and 20 mM HEPES for use in the assay at a final concentration of 5 nM (EC<sub>70</sub>).

Antagonists were prepared as 10 mM stock solution in DMSO, then diluted in 384-well plates using DMSO followed by a transfer of the dilutions into HBSS containing 0.1% bovine serum albumin (BSA), NaHCO<sub>3</sub>: 0.375 g/L and 20 mM HEPES. On the day of the assay, 50 µL of staining buffer (HBSS containing 1% FCS, 20 mM HEPES, NaHCO<sub>3</sub>: 0.375 g/L, 5 mM probenecid (Sigma) and 3 µM of the fluorescent calcium indicator fluo-4 AM (1 mM stock solution in DMSO, containing 10% pluronic) was added to each well. The 384-well cell-plates were incubated for 50 min at 37°C in 5% CO<sub>2</sub> followed by equilibration at RT for 30 - 120 min before measurement.

Within the Fluorescent Imaging Plate Reader (FLIPR Tetra, Molecular Devices), antagonists were added to the plate in a volume of 10 µL/well, incubated for 120 min, and finally 10 µL/well of agonist was added. Fluorescence was measured for each well at 1 second intervals, and the height of each fluorescence peak was compared to the height of the fluorescence peak induced by an EC<sub>70</sub> of orexin A with vehicle in place of antagonist. The IC<sub>50</sub> value (the concentration of compound needed to inhibit 50% of the agonistic response) was determined using the proprietary IC<sub>50</sub>witch software.

For the determination of apparent K<sub>b</sub> values, a dilution series of **27** was incubated with the recombinant OX receptor expressing cells for 120 min and then stimulated with a dilution series of orexin-A (OxA) to obtain a set of agonist concentration-response curves in the presence of different fixed antagonist concentrations. The obtained concentration-response curves demonstrated insurmountable antagonism.

To calculate the apparent  $K_b$  values, the  $EC_{50}$  values and Hill slopes for OxA were calculated using the proprietary  $IC_{50}$  Witch software (settings=curve-intrinsic minima and maxima were used). Then, the  $IC_{50}$  values of the antagonists at approximate  $EC_{50-70}$  of OxA (1.6 nM for human  $OX_1$ , human  $OX_2$ , rat  $OX_2$  and 8 nM for rat  $OX_1$ ) were determined using the  $IC_{50}$ witch software (settings=curve-intrinsic minima and maxima were used). From this  $IC_{50}$  value, the on-day OxA  $EC_{50}$  value and the slope (n) of the OxA CRC, the apparent  $K_b$  was calculated using the generalised Cheng-Prusoff equation:<sup>1, 2</sup>

$$K_b = \frac{IC_{50}}{(2 + ([OxA_{stim}]/EC_{50OxA})^n)^{1/n} - 1}$$

For the determination of receptor occupancy half-life ( $ROt_{1/2}$ ) values of **27**, cells were supplemented with 10  $\mu$ L of antagonist dilution series and incubated at RT for 120 min. Then, cells were either stimulated in the FLIPR by the addition of 10  $\mu$ L of 7x concentrated OxA (final assay concentration  $EC_{50}$ –  $EC_{70}$ ), or cells were washed twice with 50  $\mu$ L/well assay buffer (HBSS containing 0.1% BSA, 20 mM HEPES, 0.375 g/L  $NaHCO_3$ , 2.5 mM probenecid, pH 7.4). After 5, 15, 20 and 30 min of incubation at RT, cells were stimulated with an  $EC_{50}$ – $EC_{70}$  of OxA by the addition of 10  $\mu$ L of a 7x concentrated OxA stock in assay buffer. Calcium release was monitored for 3 min. For the estimation of  $ROt_{1/2}$ , the apparent  $K_b$  values were calculated via the generalised Cheng-Prusoff equation using on-plate generated  $EC_{50}$  values of OxA, and the earliest time point with a  $K_b$  value significantly shifted versus the non-washed control  $K_b$  value ( $K_{b0min}$ ) was used. Significant change from  $K_{b0min}$  ( $p < 0.05$ ) was determined using the one-way ANOVA test including Dunnett's post test using GraphPadPrism software and is indicated with an asterisk in Fig. **2**. An approximate  $ROt_{1/2}$  was then calculated assuming first order dissociation kinetics:  $t_{1/2} = t_x / [\log_2 (K_{bxmin}/K_{b0min})]$  with x= time point of first significant change in  $K_b$  after wash-out.

**Table 1** Inhibitory potency of **27** on  $OX_1$  and  $OX_2$ -mediated intracellular  $Ca^{2+}$  release

<b>Test system</b>	Recombinant CHO cells expressing human or rat $OX_1$ or $OX_2$ .
<b>Assay</b>	Inhibition of orexin-A-induced calcium mobilization, measured as increase in fluorescence of calcium-sensitive dye

Receptor	apparent $K_b$ [ $\sigma_g$ ]	
	OX <sub>1</sub>	OX <sub>2</sub>
Human	5.3 nM [1.3]	1.4 nM [1.8]
Rat	7.3 nM [1.7]	1.7 nM [1.7]

Apparent  $K_b$  values were calculated from  $IC_{50}$  values at 1.6 nM OxA (human OX<sub>1</sub>, human OX<sub>2</sub>, rat OX<sub>2</sub>) or 8 nM OxA (rat OX<sub>1</sub>) via the generalised Cheng-Prusoff equation. Apparent  $K_b$  values are expressed as the geometric means of  $n = 4$  values determined from three independent experiments. The geometric standard deviation  $\sigma_g$  is displayed in brackets. CHO, Chinese hamster ovary;  $IC_{50}$ , concentration that causes 50% inhibition;  $K_b$ , equilibrium dissociation constant of a ligand determined by means of a functional assay; OX<sub>1</sub>, orexin receptor 1; OX<sub>2</sub>, orexin receptor 2; OxA, orexin-A.

### Pharmacology protocols:

Experiments were conducted on male, adult Wistar (RccHan:WIST; Harlan, Horst, The Netherlands) rats, which were maintained under standard lab conditions (temperature  $20 \pm 2^\circ\text{C}$ , relative humidity 55–70%, food and water *ad libitum*) under a regular 12 h light–dark cycle (lights on 06:00). After arrival rats were allowed at least one week of habituation to Actelion’s animal facility before experiments commenced. Experimental procedures were approved by the Basel-Landschaft Veterinary Office and strictly adhered to Swiss federal regulations on animal experimentation. Unless noted otherwise, rats were socially housed in groups of four in standard plastic rodent cages, and all tests were conducted during the light phase (08:00 to 18:00) under illumination of  $> 600$  lx, where not otherwise specified.

### For telemetric transmitter implantation (EEG/EMG monitoring)

Rats were equipped with telemetric transmitters (TL11M2-F20-EET; Data Science International, St Paul, MN, USA) that allowed the noninvasive detection of electroencephalograms (EEG), electromyograms (EMG) and activity via signal transmission to a receiver. The surgical transmitter implantation was performed under aseptic conditions. The rat was placed and secured in a stereotaxic apparatus. The body of the transmitter was placed subcutaneously along the dorsal flank of the rat with the leads routed subcutaneously to an incision accessing the cranium. For EEG recordings, two trepanations were placed in the skull, 2 mm from either side of the midline and 2 mm anterior to the lambda suture for placement of one differential

pair of electrodes. Two other superficial trepanations were drilled for screws as support for cementing the electrodes. The EMG leads were inserted in either side of the muscles of the neck and sutured into place.

### **Sleep/wake cycle evaluation**

Sleep/wake cycles were evaluated via radiotelemetry technology in rats. EEG, EMG and home cage activity were recorded from singly housed Wistar rats while under free moving conditions in their home cages. At the start of the experiment rats were placed together with their home cages in ventilated sound attenuating boxes, on a regular 12 h light/dark cycle for 3 days of acclimation before recordings started. Experiments were done in a crossover design, i.e. animals were alternatively treated with drug and vehicle. Recordings started by 24 h baseline (preceding the treatment), the 12 h night-period following the treatment, 36 h of recovery (wash out period) followed by the crossover. Oral administrations occurred at the transition from the day to the night phase (17:45 to 18:00). Sleep and wake stages were evaluated automatically using the Somnologica Science software (Medcare, Embla, USA). The recording is divided into user definable (10 s) contiguous epochs. The scoring is based on frequency estimation for EEG and amplitude discrimination for the EMG and the locomotor activity. Using these measurements, the software determines the probabilities that the EEG and EMG components within each epoch best represent waking, quiet waking, non-REM sleeping or REM sleeping. Essentially, wake consists of low-amplitude EEG activity with relatively greater power in the higher frequency bands such as alpha, from (10–13 Hz), accompanied by moderate to high-level EMG activity. The locomotor activity and the amplitude of the EMG allow the differentiation between wake and quiet wake. Non-REM sleep is defined by high amplitude EEG activity with greater power in the delta frequency band (0.5–5 Hz), and by low EMG activity. REM sleep is characterized by low amplitude EEG activity focused in the theta frequency band (6–9 Hz). There is no EMG activity present during REM sleep.

1. T. R. Miller, D. G. Witte, L. M. Ireland, C. H. Kang, J. M. Roch, J. N. Masters, T. A. Esbenshade and A. A. Hancock, *J. Biomol. Screening*, 1999, **4**, 249-258.
2. Y. Cheng and W. H. Prusoff, *Biochem Pharmacol*, 1973, **22**, 3099-3108.