

Supporting information

For

Development of novel bis-naphthalimide derivatives and their anticancer properties

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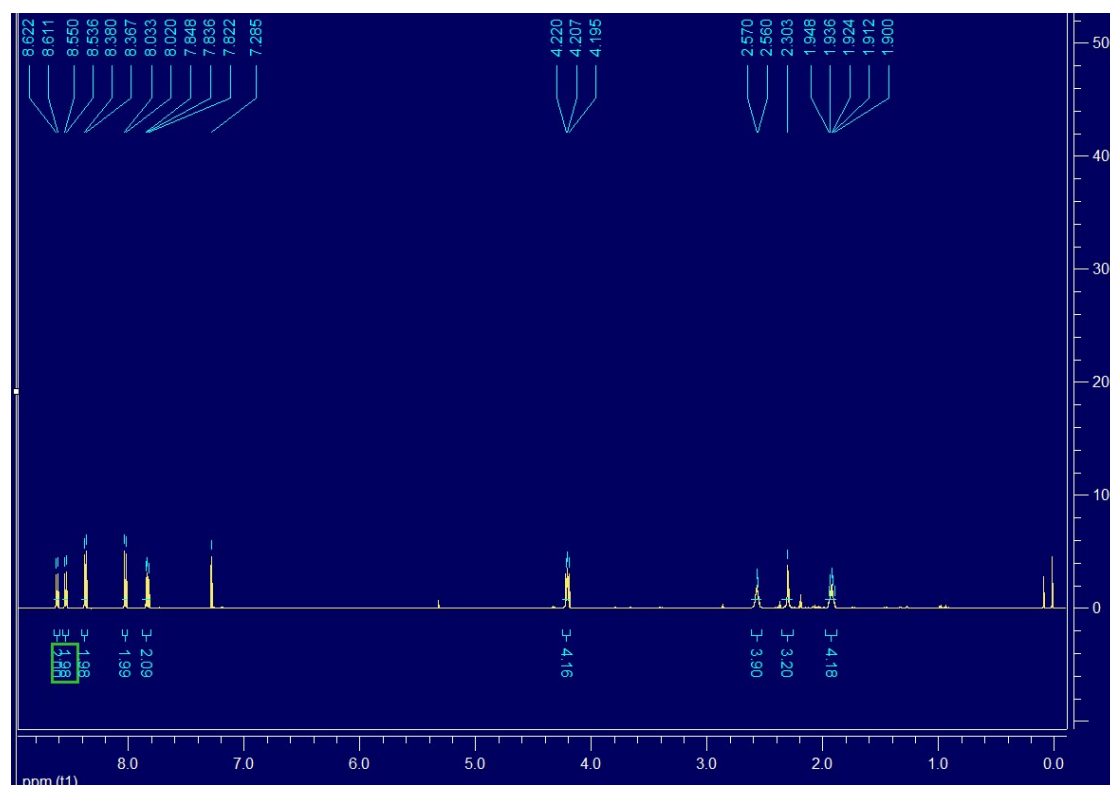


Fig. S1 ¹H NMR (600 MHz, CDCl₃) of compound M1.

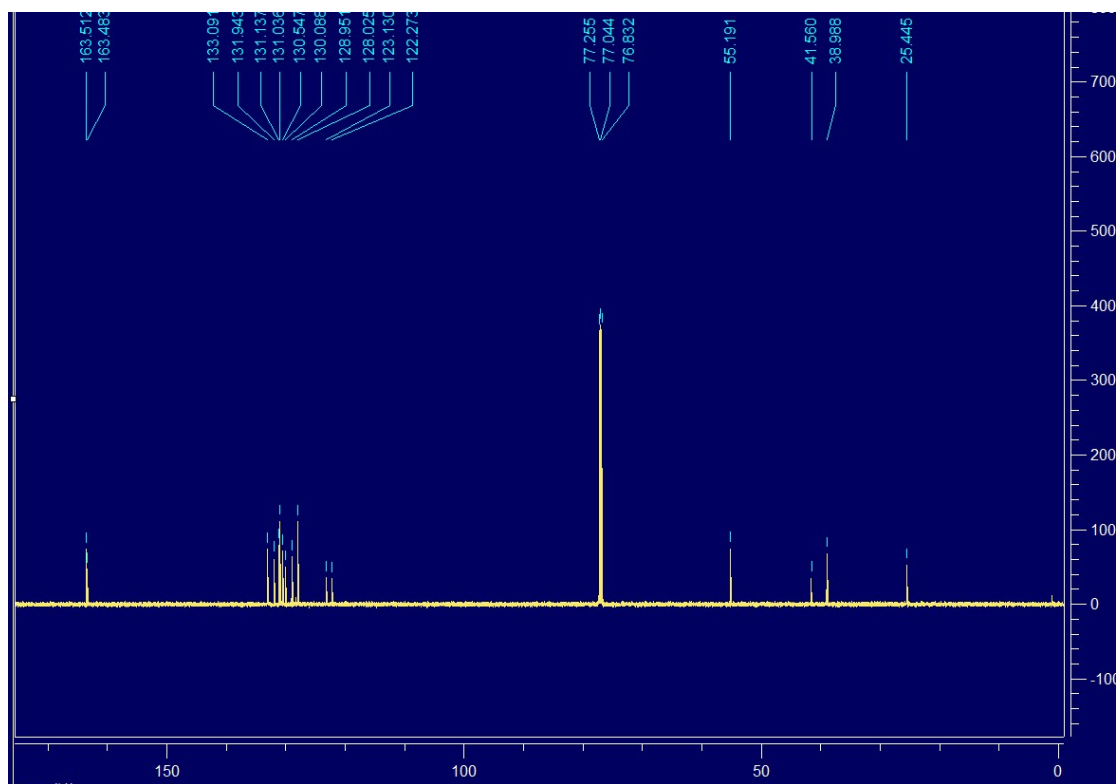


Fig. S2 ^{13}C NMR (150 MHz, CDCl_3) of compound **M1**.

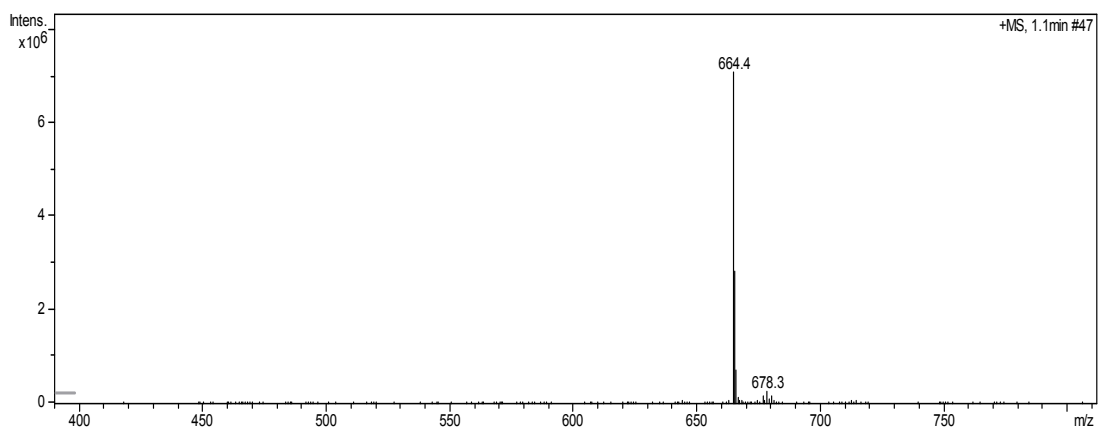


Fig. S3 MS (ESI) of compound **M1**.

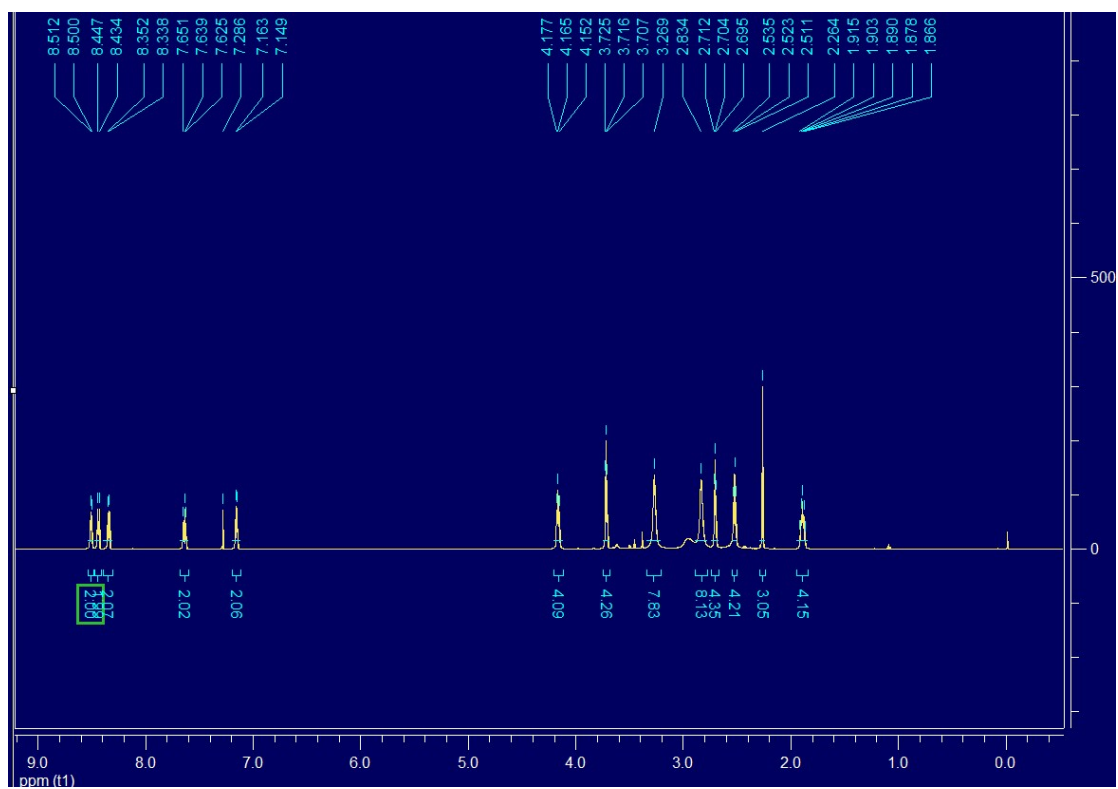


Fig. S4 ^1H NMR (600 MHz, CDCl_3) of compound **N1**.

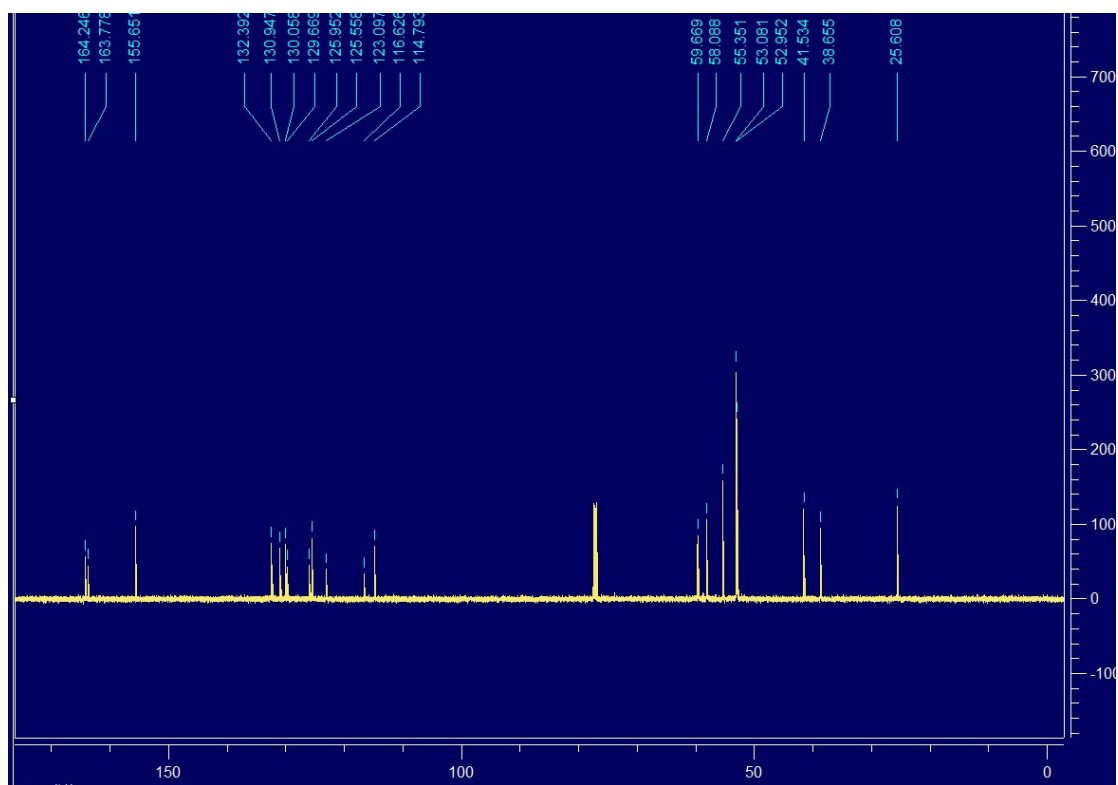


Fig. S5 ^{13}C NMR (150 MHz, CDCl_3) of compound **N1**.

Mass Spectrum SmartFormula Report

Analysis Info

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 Method 20130927
 Sample Name
 Comment

Acquisition Date 2013/10/12 15:11:45

Operator
 Instrument apex-Ultra

Acquisition Parameter

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Averaged Scans	2	No. of Cell Fills	1	Laser Power	51.0 %
Broadband Low Mass	100.3 m/z	End Plate	3500.0 V	MALDI Plate	300.0 V
Broadband High Mass	800.0 m/z	Capillary Entrance	4000.0 V	Imaging Spot Diameter	2000.0 μ m
Acquisition Mode	Single MS	Skimmer 1	20.0 V		
Pulse Program	basic	Drying Gas Temperature	180.0 $^{\circ}$ C	Calibration Date	Tue Oct 8 03:08:56 2013
Source Accumulation	0.0 sec	Drying Gas Flow Rate	4.0 L/min	Data Acquisition Size	131072
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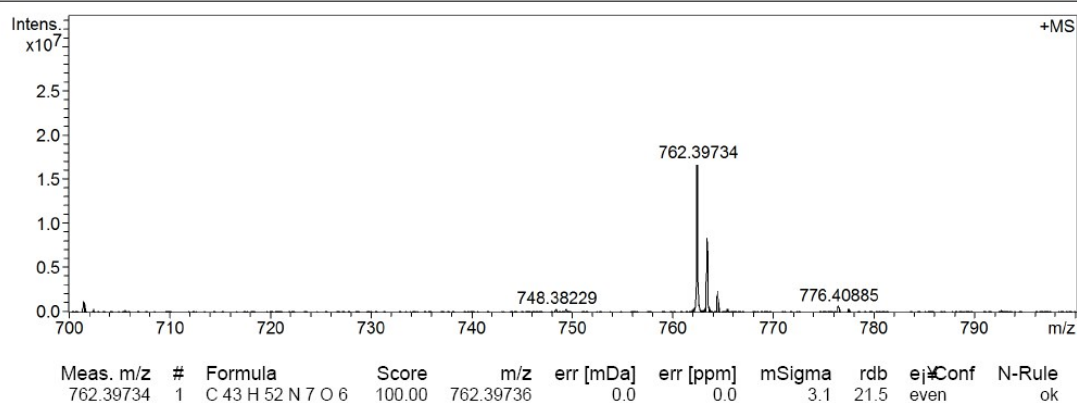


Fig. S6 HRMS (ESI) of compound N1.

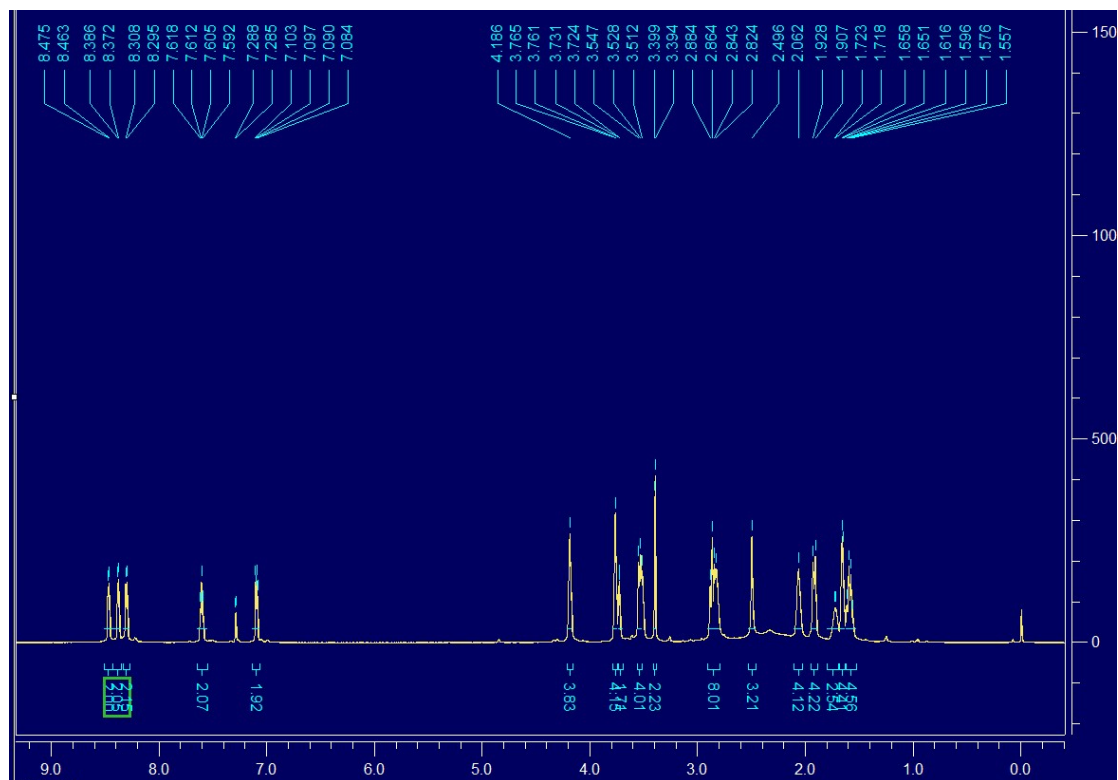


Fig. S7 ^1H NMR (600 MHz, CDCl_3) of compound N2.

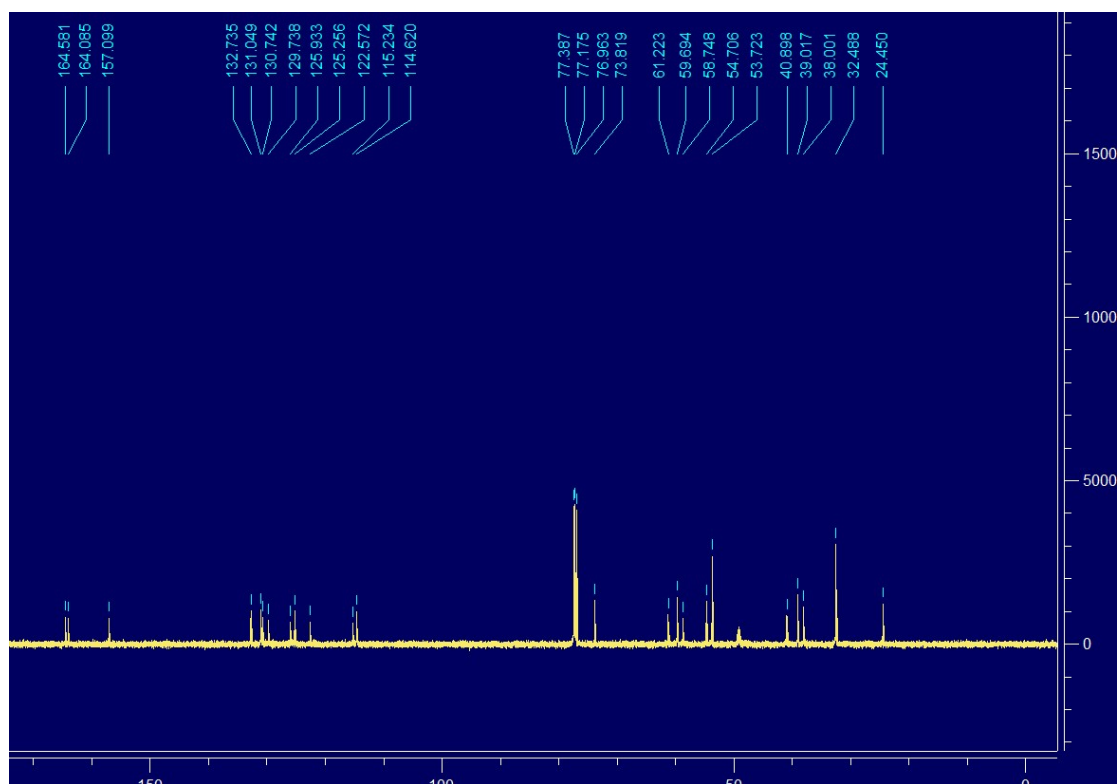


Fig. S8 ^{13}C NMR (150 MHz, CDCl_3) of compound **N2**.

Mass Spectrum SmartFormula Report

Analysis Info

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 Method 20130927
 Sample Name
 Comment

Acquisition Date 2013/10/12 15:25:14

Operator
 Instrument apex-Ultra

Acquisition Parameter

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Broadband High Mass	800.0 m/z	Capillary Entrance	4000.0 V	Imaging Spot Diameter	2000.0 μm
Acquisition Mode	Single MS	Skimmer 1	20.0 V	Calibration Date	Tue Oct 8 03:08:56 2013
Pulse Program	basic	Drying Gas Temperature	180.0 $^{\circ}\text{C}$	Data Acquisition Size	131072
Source Accumulation	0.0 sec	Drying Gas Flow Rate	4.0 L/min	Apodization	Sine-Bell Multiplication
Ion Accumulation Time	0.0 sec	Nebulizer Gas Flow Rate	1.0 L/min		
Flight Time to Acq. Cell	0.0 sec				

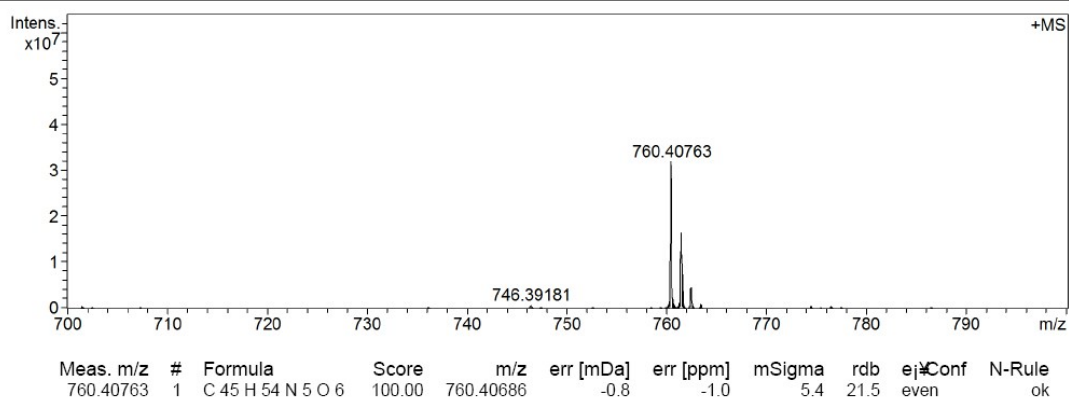


Fig. S9 HRMS (ESI) of compound **N2**.



Mass Spectrum SmartFormula Report

Analysis Info

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 Method 20130927
 Sample Name
 Comment

Acquisition Date 2013/10/12 15:26:19

Operator
 Instrument apex-Ultra

Acquisition Parameter

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Broadband Low Mass	100.3 m/z	End Plate	3500.0 V	MALDI Plate	300.0 V
Broadband High Mass	800.0 m/z	Capillary Entrance	4000.0 V	Imaging Spot Diameter	2000.0 μ m
Acquisition Mode	Single MS	Skimmer 1	20.0 V		
Pulse Program	basic	Drying Gas Temperature	180.0 $^{\circ}$ C	Calibration Date	Tue Oct 8 03:08:56 2013
Source Accumulation	0.0 sec	Drying Gas Flow Rate	4.0 L/min	Data Acquisition Size	131072
Ion Accumulation Time	0.0 sec	Nebulizer Gas Flow Rate	1.0 L/min	Apodization	Sine-Bell Multiplication
Flight Time to Acq. Cell	0.0 sec				

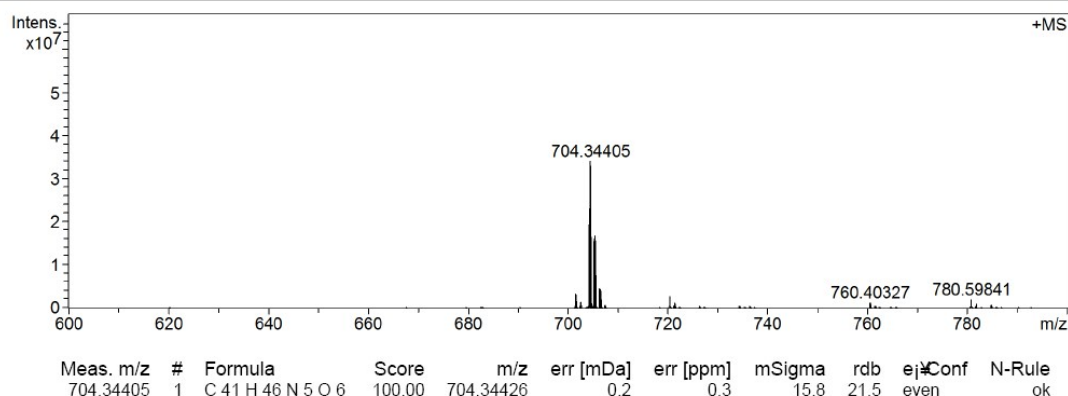


Fig. S12 HRMS (ESI) of compound N3.

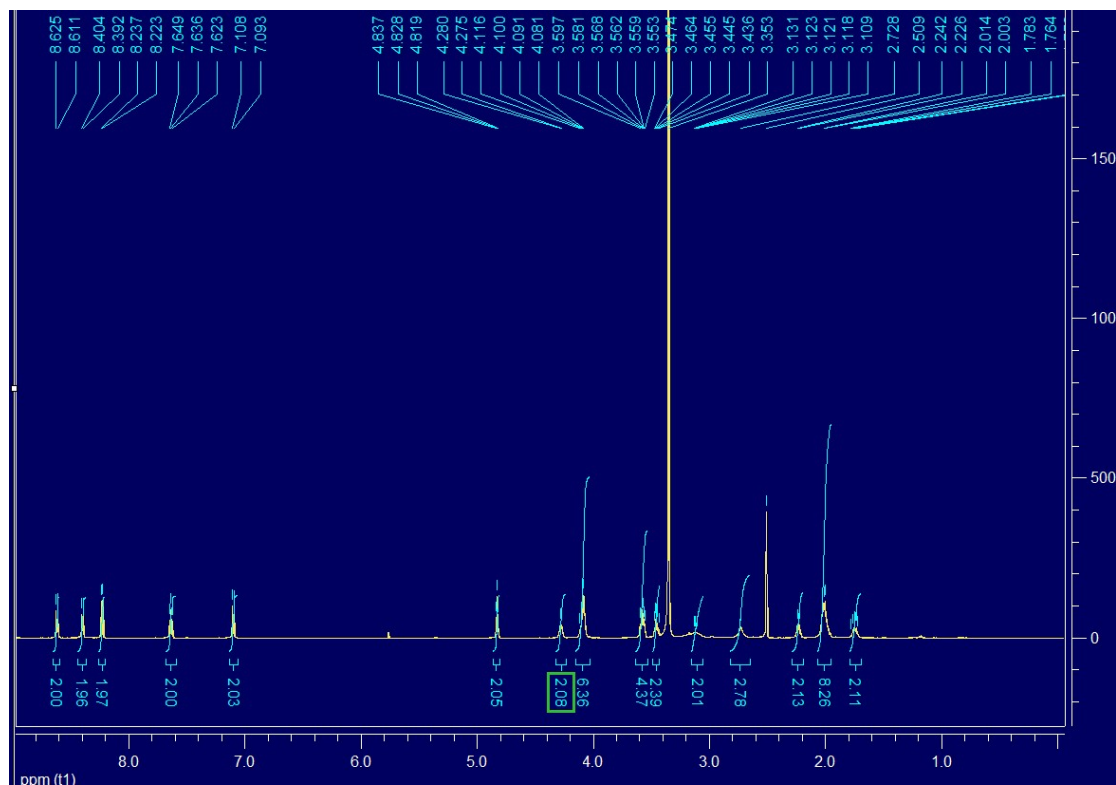


Fig. S13 ^1H NMR (600 MHz, $\text{DMSO-d}_6 + \text{D}_2\text{O}$) of compound N4.

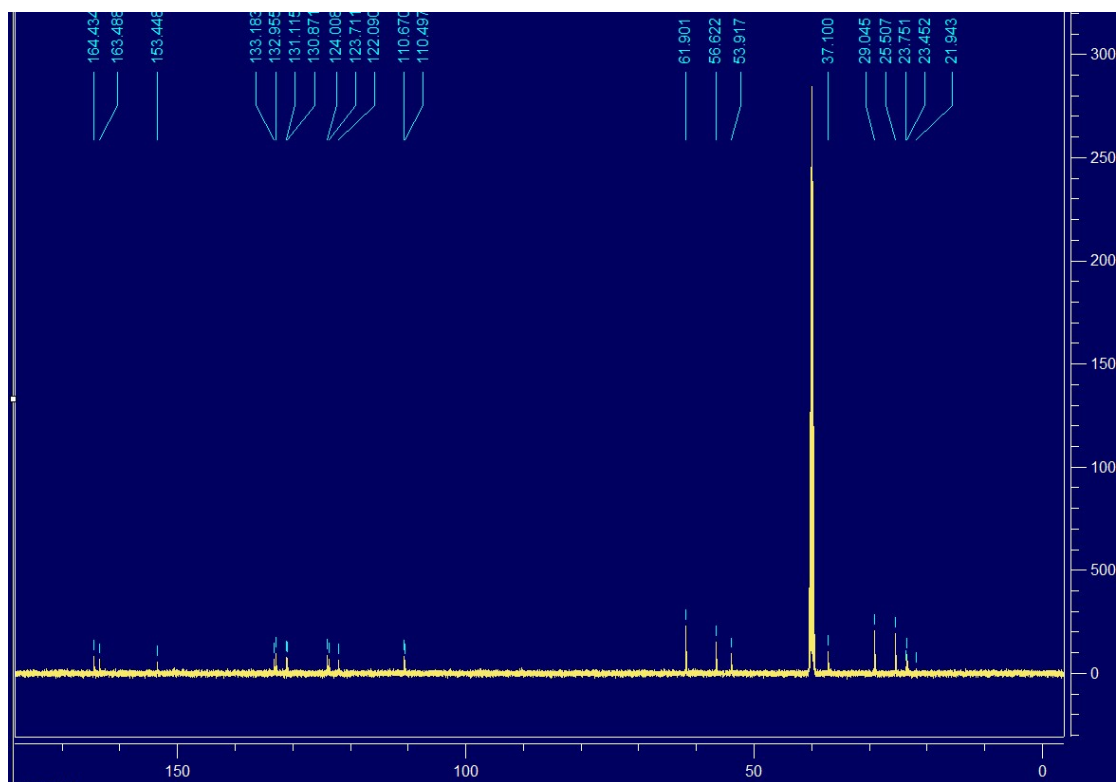


Fig. S14 ^{13}C NMR (150 MHz, $\text{DMSO-d}_6 + \text{D}_2\text{O}$) of compound **N4**.

Mass Spectrum SmartFormula Report

Analysis Info

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 Method 20130927
 Sample Name
 Comment

Acquisition Date 2013/10/12 15:27:36

Operator
 Instrument apex-Ultra

Acquisition Parameter

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Averaged Scans	2	No. of Cell Fills	1	Laser Power	51.0 %
Broadband Low Mass	100.3 m/z	End Plate	3500.0 V	MALDI Plate	300.0 V
Broadband High Mass	800.0 m/z	Capillary Entrance	4000.0 V	Imaging Spot Diameter	2000.0 μm
Acquisition Mode	Single MS	Skimmer 1	20.0 V		
Pulse Program	basic	Drying Gas Temperature	180.0 $^{\circ}\text{C}$	Calibration Date	Tue Oct 8 03:08:56 2013
Source Accumulation	0.0 sec	Drying Gas Flow Rate	4.0 L/min	Data Acquisition Size	131072
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Flight Time to Acq. Cell	0.0 sec				

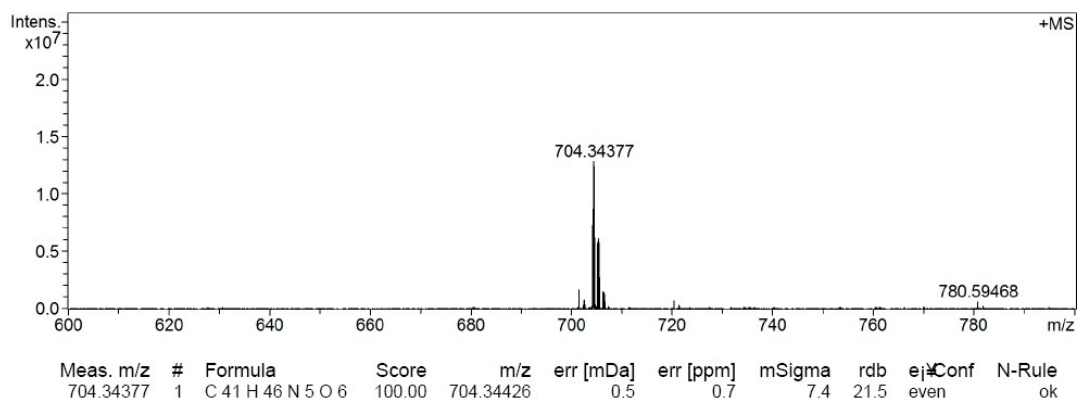


Fig. S15 HRMS (ESI) of compound **N4**.

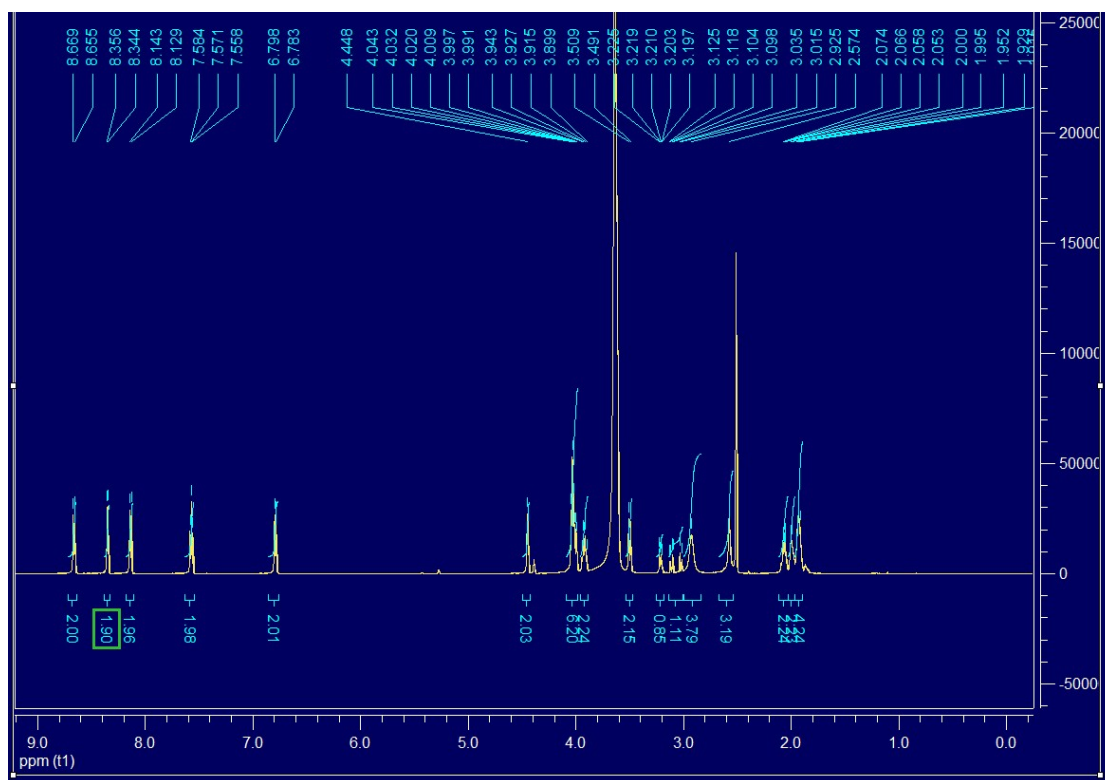


Fig. S16 ^1H NMR (600 MHz, $\text{DMSO-d}_6 + \text{D}_2\text{O}$) of compound **N5**.

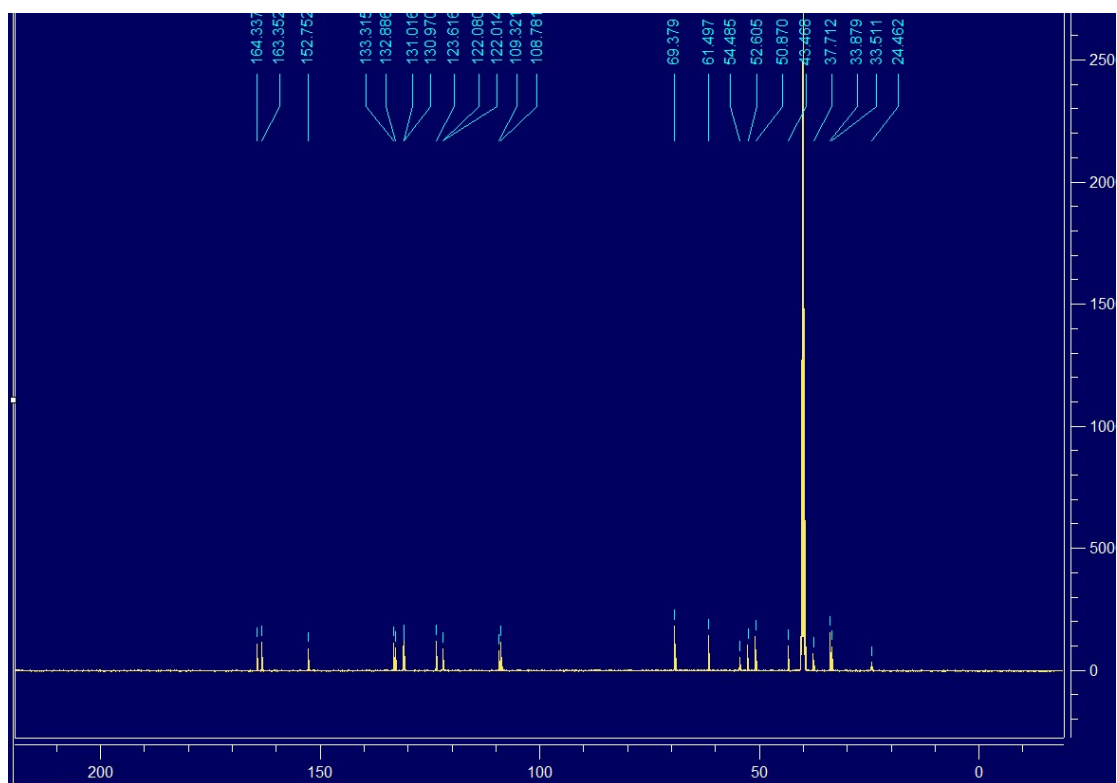


Fig. S17 ^{13}C NMR (150 MHz, $\text{DMSO-d}_6 + \text{D}_2\text{O}$) of compound **N5**.

Mass Spectrum SmartFormula Report

Analysis Info

Analysis Name D:\Data\20131012\Q10-ESI-POS-MSMS_000002.d
 Method 20130927
 Sample Name
 Comment

Acquisition Date 2013/10/12 15:29:51

Operator
 Instrument apex-Ultra

Acquisition Parameter

Polarity	Positive	Source	ESI	No. of Laser Shots	20
Averaged Scans	2	No. of Cell Fills	1	Laser Power	51.0 %
Broadband Low Mass	100.3 m/z	End Plate	3500.0 V	MALDI Plate	300.0 V
Broadband High Mass	800.0 m/z	Capillary Entrance	4000.0 V	Imaging Spot Diameter	2000.0 μ m
Acquisition Mode	Single MS	Skimmer 1	20.0 V		
Pulse Program	basic	Drying Gas Temperature	180.0 $^{\circ}$ C	Calibration Date	Tue Oct 8 03:08:56 2013
Source Accumulation	0.0 sec	Drying Gas Flow Rate	4.0 L/min	Data Acquisition Size	131072
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Flight Time to Acq. Cell	0.0 sec				

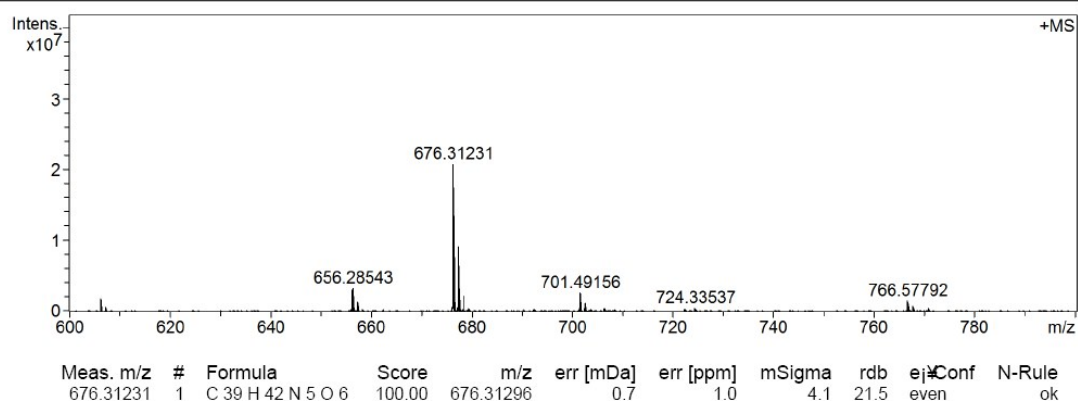


Fig. S18 HRMS (ESI) of compound N5.

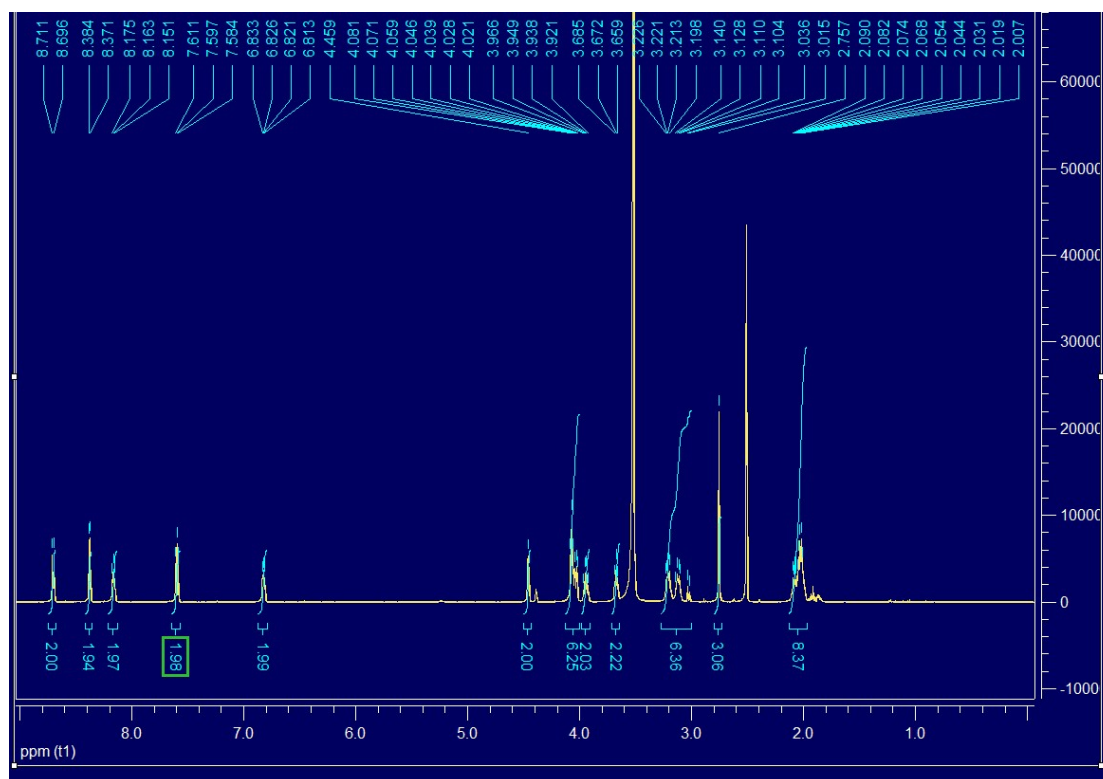


Fig. S19 ^1H NMR (600 MHz, DMSO- d_6 + D_2O) of compound N6.

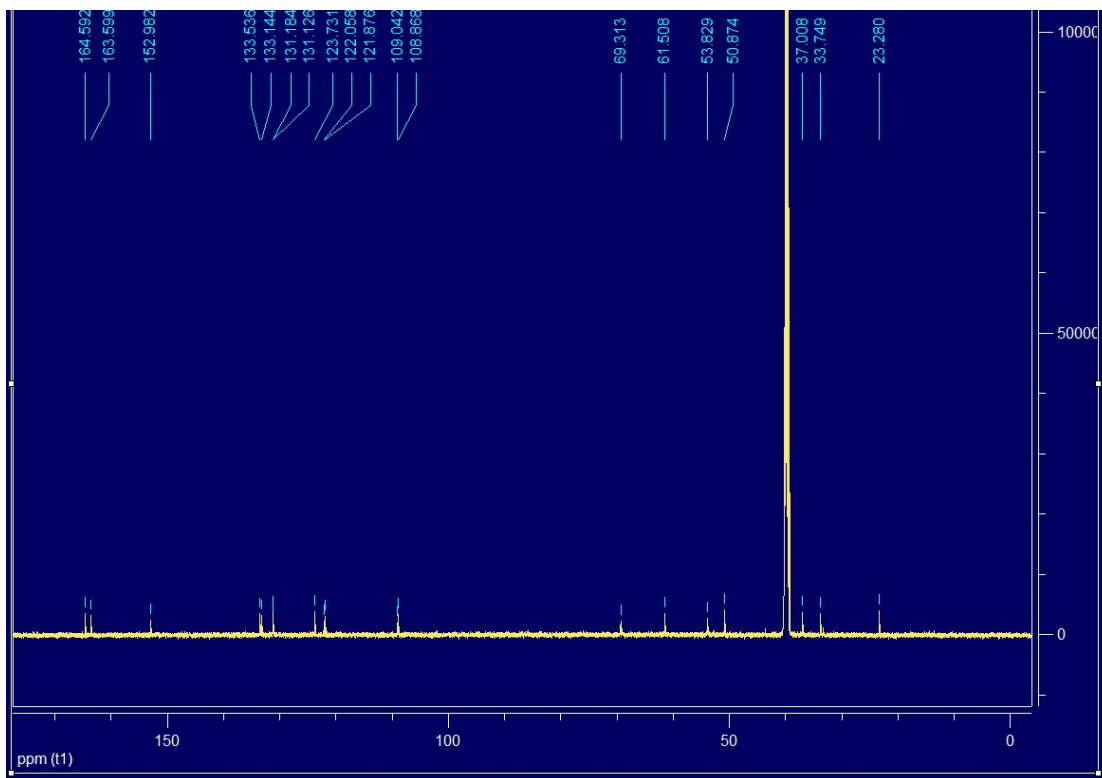


Fig. S20 ^1H NMR (600 MHz, $\text{DMSO-d}_6 + \text{D}_2\text{O}$) of compound **N6**.

Mass Spectrum SmartFormula Report

Analysis Info

Analysis Name D:\Data\20131012\Q9-ESI-POS-MSMS_000002.d
 Method 20130927
 Sample Name
 Comment

Acquisition Date 2013/10/12 15:28:41

Operator
 Instrument apex-Ultra

Acquisition Parameter

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Averaged Scans	2	No. of Cell Fills	1	Laser Power	51.0 %
Broadband Low Mass	100.3 m/z	End Plate	3500.0 V	MALDI Plate	300.0 V
Broadband High Mass	800.0 m/z	Capillary Entrance	4000.0 V	Imaging Spot Diameter	2000.0 μm
Acquisition Mode	Single MS	Skimmer 1	20.0 V	Calibration Date	Tue Oct 8 03:08:56 2013
Pulse Program	basic	Drying Gas Temperature	180.0 $^{\circ}\text{C}$	Data Acquisition Size	131072
Source Accumulation	0.0 sec	Drying Gas Flow Rate	4.0 L/min	Apodization	Sine-Bell Multiplication
Ion Accumulation Time	0.0 sec	Nebulizer Gas Flow Rate	1.0 L/min		
Flight Time to Acq. Cell	0.0 sec				

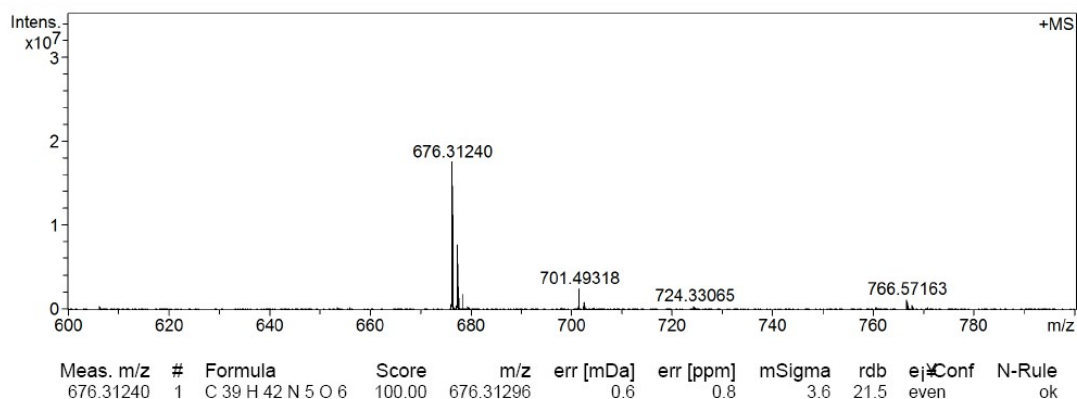


Fig. S21 HRMS (ESI) of compound **N6**.

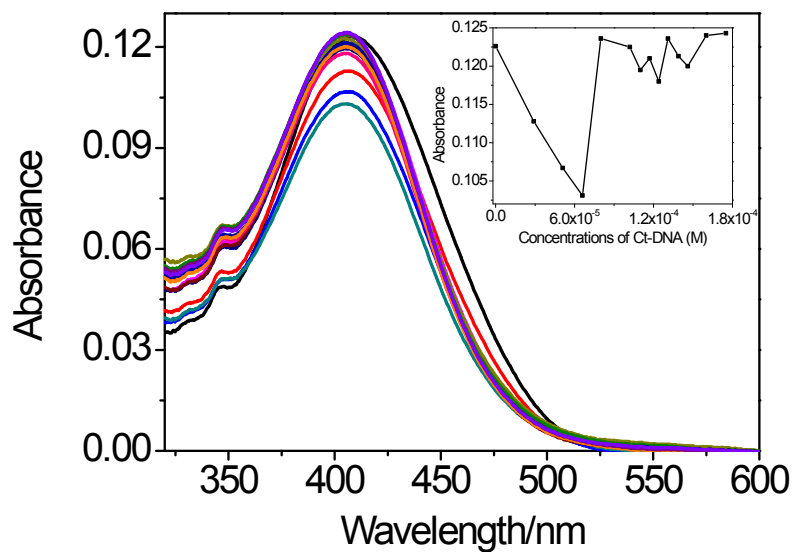


Fig. S22 UV-Vis spectra of compound NI1 (1 × 10⁻⁵ M) binding with Ct-DNA in phosphate buffer (10 mM, pH 7.4, 50 mM NaCl).

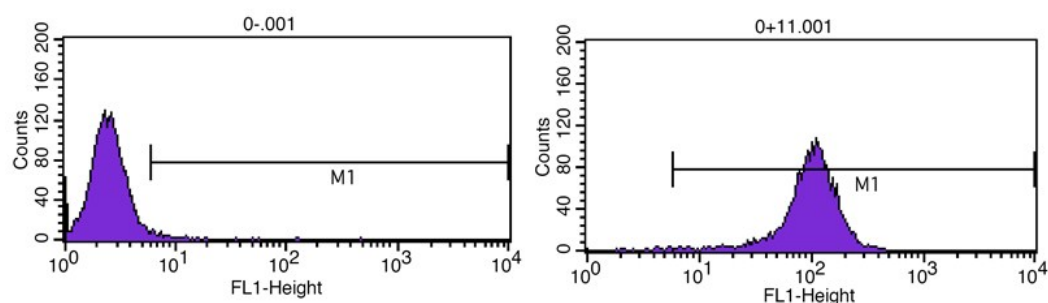


Fig. S23 Compound NI1 induced lysosomal membrane permeabilization (LMP). Hela cells were stained with Annexin V-TITC. Numbers indicate the percentage of cells with Annexin V-TITC fluorescence.

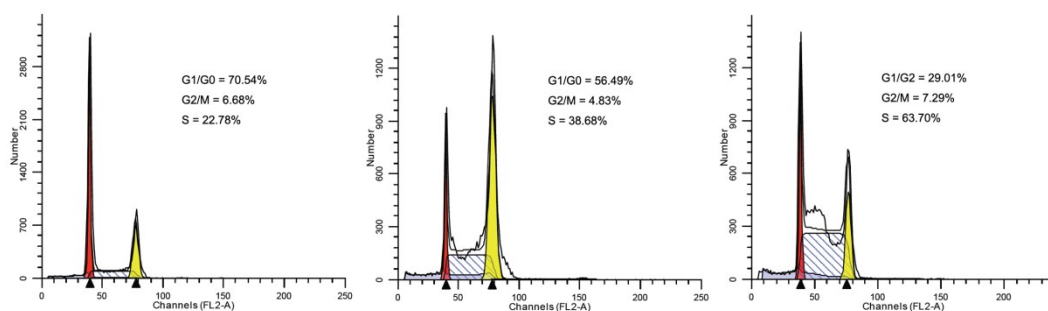


Fig. S24 Inhibition of cell cycle process in Hela cell treated with Amonafide for 48 h. Cell cycle distribution was analyzed by flow cytometry.

Table S1. Quantitative determination of hematotoxic effects of compound **NI1** on the numbers of platelets (PLT), red blood cells (RBC) and white blood cells (WBC) in healthy mice when administered i.v. with different doses (5 mg/kg, 10 mg/kg and 20 mg/kg). Analyses were performed at 24 h, 48 h and 96 h following administration.

	Doses (mg/kg)	PLT ($10^9/\mu\text{L}$)	RBC ($10^{12}/\mu\text{L}$)	WBC ($10^9/\mu\text{L}$)
Control		296.75±9.18	6.24±0.18	3.62±0.04
24 h	5	284.25±4.92*	6.34±0.05	3.48±0.24
	10	288.75±4.65	5.99±0.13	3.35±0.09
	20	299±6.48	5.8±0.46*	3.71±0.28
48 h	5	343.75±7.14*	6.1±0.25	3.71±0.28
	10	259.5±2.38*	6.19±0.96	2.03±0.09*
	20	407.75±23.16*	6.8±0.3	2.8±0.12*
96 h	5	343.5±6.56*	6.27±0.1	3.05±0.04*
	10	384.75±9.74*	6.2±1.4	3.14±0.17*
	20	364.5±6.61*	6.18±0.63	2.43±0.09*

Statistical significance (control versus treatment) was assessed using the Mean-Whitney U-test:

* = $p < 0.05$.

Table S2. Quantitative determination of cardiotoxic effects of compound **NI1** on the levels of aspartate transaminase (AST), creatine kinase (CK) and lactate dehydrogenase (LDH) in healthy mice when administered i.v. with different doses (5 mg/kg, 10 mg/kg and 20 mg/kg). Analyses were performed at 24 h, 48 h and 96 h following administration.

	Doses (mg/kg)	AST (U/L)	LDH (U/L)	CK (U/L)
Control		159.83±9.82	861.5±90.78	1342.5±60.64
24 h	5	175.13±5.91	958.80±9.92	1628.2±111.51
	10	311.9±6.87*	1778.2±72.29*	1832.0±109.12*
	20	356.87±38.79*	1805.9±87.32*	1799.9±305.68
48 h	5	123.97±7.20*	1109.3±59.21*	1292.1±60.21
	10	139.73±8.93*	985.3±78.63	1196.7±37.4
	20	146.8±9.44	947.63±44.87	2035.3±45.66*
96 h	5	146.33±3.22	1146.2±40.06*	1648.8±51.51*
	10	163.37±15.2	1213.5±104.36*	2155.3±63.85*
	20	166.5±11.21	999.13±70.76	1483.7±58.42*

Statistical significance (control versus treatment) was assessed using the Mean-Whitney U-test:

* = $p < 0.05$.