Electronic Supplementary Information (ESI)

Bone-repair properties of biodegradable hydroxyapatite nano-rods superstructures

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Single roughness depth $(R_{Z,i})$; the maximum roughness depth $(R_{Z,max})$ and the mean roughness depth ($R_{Z,mean}$) parameters. Examples of $R_{z,i}$ are indicated in materials roughness profiles.





R _{z, mean}	SD	Minimum	Median	R _{z, max}
56.7	14.4	27.3	58.5	85

Figure 2 ESI: MII surface roughness profile.

R_{z, mean} 42.9



91.5



Figure 3 ESI: MIII surface roughness profile.

Figure 4 ESI: MIV surface roughness profile.



71.7

Figure 5 ESI: Nano-hydroxyapatite superstructures degradation in physiological fluid conditions (phosphate buffer saline, pH = 7.4). At 36°C all materials exhibited a weight loss minor than 2.5%; while at 25°C no degradation can be recorded.



Figure 6 ESI: Optical microphotographs showing MSCs adhesion on the nano-HAp powder coatings (MI, MII, MIII and MIV) with different nanoparticles amounts. At high amount of nano-HAp ($3100 \ \mu g/cm^2$) non-adherent cells are observed.



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Figure 7 ESI: Optical microphotographs showing MSCs morphology after adhesion on Co-I; MI/ Co-I coatings and non-coated glass slide used as positive control (C+). ⁽¹⁾ Co-I ($31 \mu g / cm^2$); ⁽²⁾ HAp (MI) 7200 $\mu g/cm^2$. **Slide A:** A1, A2 cell adhesion on Co-I coating; A4, A5 example area with high mineral concentration and no visible adherent cells. **Slide B:** adherent cells on C+. **Slide C:** C1, C5 adherent cells on Co-I coatings; C2, C6 example area with high mineral concentration and no visible adherent cells; C3, C7 adherent cells; C3, C7 adherent cells; C3, C7 adherent cells; C3, C7 adherent cells; C3, C6 example area with high mineral concentration and no visible adherent cells; C3, C7 adherent cells; C3, C6 example area with high mineral concentration and no visible adherent cells; C3, C7 adherent cells; C3, C6 example area with high mineral concentration and no visible adherent cells; C3, C7 adherent cells; C3, C6 example area with high mineral concentration and no visible adherent cells; C3, C7 adherent cells; C3, C6 example area with high mineral concentration and no visible adherent cells; C3, C7 adherent cells; C3, C6 example area with high mineral concentration and no visible adherent cells; C3, C7 adherent cells; C3, C6 example area with high mineral concentration and no visible adherent cells; C3, C7 adherent cells; C3, C6 example area with high mineral concentration and no visible adherent cells; C3, C6 example area with high mineral concentration and no visible adherent cells; C3, C6 example area with high mineral concentration and no visible adherent cells; C4, C6 example area with high mineral concentration and no visible adherent cells; C4, C6 example area with high mineral concentration and no visible adherent cells; C4, C6 example area with high mineral concentration and no visible adherent cells; C4, C6 example area with high mineral concentration and no visible adherent cells; C4, C6 example area with high mineral concentration and no visible adherent cells; C4, C6 example area w









MI / Co I Coating

Figure 8 ESI: Laser scanning confocal microphotographs showing MSCs α -SMA expression using large amounts of hydroxyapatite MI (7200 μ g/cm²) / Co I (31 μ g/cm²) coatings to test cytotoxicity.



Figure 9 ESI: Figure 10 (a) supplementary information. Immunofluorescence assay: confocal microphotographs $10\times$. The bar graph shows the number of nucleus counted by Image J software in each replicate slide. Non-coated glass slide was used as positive control (C+).



Figure 10 ESI: Image analysis for α-SMA expression (monochromatic image, green laser 488nm), the mean green pixel intensity was determined from the mean brightness value of all green pixels analyzed per image. Supplementary information of methodology section 2.7.3: "*MSC actin-based spreading on Collagen type I without and with nano-Hap /: Immunofluorescence confocal microscopy*".

