#### **ELECTRONIC SUPPORTING INFORMATION**

#### Low generation Polyamine Dendrimers Bearing Flexible Tetraethylene Glycol as nanocarriers for plasmids and siRNA

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Figure S1. <sup>1</sup>H NMR spectrum of compound 9 (CDCl<sub>3</sub>, 300 MHz).



Figure S2.<sup>13</sup>C NMR spectrum of compound 9 (CDCl<sub>3</sub>, 75 MHz).



Figure S3. COSY spectrum of compound 9.



**Figure S4.** HRMS analysis (ESI<sup>+</sup>) for compound **9**.



Figure S5. IR spectrum of compound 9.





**Figure S6.** <sup>1</sup>H NMR spectrum of compound **3** (D<sub>2</sub>O, 300 MHz).



**Figure S7.** <sup>13</sup>C NMR spectrum of compound **3** ( $D_2O$ , 151 MHz).



Figure S8. COSY spectrum of compound 3







**Figure S10.** HRMS analysis (ESI<sup>+</sup>) for compound **3**.



Figure S11. IR spectrum of compound 3.





Figure S13. <sup>13</sup>C NMR spectrum of compound 11 (CDCl<sub>3</sub>, 75 MHz).



Figure S14. COSY spectrum of compound 11.



Figure S15. MALDI-TOF trace for compound 11.



Figure S16. MALDI-TOF trace for compound 11.



Figure S17. IR spectrum of compound 11



Figure S18. <sup>1</sup>H NMR spectrum of compound 4 (D<sub>2</sub>O, 300 MHz).



Figure S19. <sup>13</sup>C NMR spectrum of compound 4 (D<sub>2</sub>O, 75 MHz).



Figure S20. COSY spectrum of compound 4.



Figure S21. MALDI-TOF trace for compound 4 (DHB matrix).



Figure S22. Expanded MALDI-TOF trace for compound 4.



Figure S23. IR spectrum of compound 4.



Figure S24. <sup>1</sup>H NMR spectrum of compound 13 (CDCl<sub>3</sub>, 300 MHz).



Figure S25. <sup>13</sup>C NMR spectrum of compound 13 (CDCl<sub>3</sub>, 151 MHz).



Figure S26. COSY spectrum of compound 13.



Figure S27. MALDI-TOF trace for glycodendrimer 13.



Figure S28. IR spectrum of compound 13.



**Figure S29.**<sup>1</sup>H NMR spectrum of glycodendrimer **5** (D<sub>2</sub>O, 300 MHz).



Figure S30. <sup>13</sup>C NMR spectrum of glycodendrimer 5 (D<sub>2</sub>O, 151 MHz).



Figure S31. COSY spectrum of glycodendrimer 5.



Figure S32. ESI spectrum of compound 5.



Figure S33. IR spectrum of compound 5.





**Figure S34.** <sup>1</sup>H NMR spectrum of compound **15** (CDCl<sub>3</sub>, 300 MHz).

Rishi-B2-046-P31 Rishi-B2-046-P31 phosphore CDCl3 C:\Bruker\TOPSPIN Roy





Figure S35.<sup>31</sup>P NMRspectrum of compound 15 (122 MHz, CDCl<sub>3</sub>).



Figure S36. <sup>13</sup>C NMR spectrum of compound 15 (CDCl<sub>3</sub>, 75 MHz).



Figure S37. COSY spectrum of compound 15.



Figure S38. MALDI-TOF spectrum of compound 15 (DHB matrix).



Figure S39. IR spectrum of compound 15.





Figure S40.<sup>1</sup>H NMR spectrum of glycodendrimer 6 (D<sub>2</sub>O, 300 MHz)



**Figure S41.**<sup>13</sup>C NMR spectrum of glycodendrimer **6** (D<sub>2</sub>O, 151 MHz).



Figure S42. COSY spectrum of glycodendrimer 6.



Figure S43.<sup>31</sup>P NMRspectrum of compound 6 (122 MHz, CD<sub>3</sub>OD).



Figure S44. HR ESI spectrum of compound 6



Figure S45. IR spectrum of glycodendrimer 6.





Figure S46.<sup>1</sup>H NMR spectrum of glycodendrimer 6 (D<sub>2</sub>O, 600 MHz).



Figure S47.<sup>13</sup>C NMR spectrum of glycodendrimer 16 (D<sub>2</sub>O, 150 MHz).



Figure S48. IR spectrum of glycodendrimer 16.







Figure S50. ExpandedHR ESI spectrum of compound 16



Figure S51. Cytotoxicity of polyamine dendrimers G0-4 (1), G0-6 (2), G1-12 (3), G1-18 (4) and G1-30 (5 and 6, dipentaerythritol and aromatic phosphazene cores) in A) U251N human glioblatoma, B) MCF-7 human breast carcinoma, and C) HepG2 human liver adenocarcinoma cells. Cells were treated with increasing concentrations of dendrimers for 24 hrs, after which the mitochondrial metabolic activity was measured by the MTT assay. Shown are mean percentage values ± SEM as compared to the untreated control from three independent experiments.



**Figure S52.** Cytotoxicity of polyamine dendrimers G1-12 (**3**), G1-18 (**4**) and G1-30 (**5**, dipentaerythritol core) in U251N human glioblastoma cells. Cells were treated with increasing concentrations of dendrimers for 24 hrs, after which cell nuclei were labeled with Hoechst 33342 and counted. Shown are mean percentage values  $\pm$  SEM as compared to the untreated control from three independent experiments.



# Figure S53. Transfection negative controls of eGFP plasmid DNA alone and dendrimer vectors alone in HEK 293 human embryonic kidney cells

Fluorescence micrographs of HEK 293 human embryonic kidney cells incubated with (A) eGFP plasmid DNA (0.5  $\mu$ g/well), (B) G1-12-NH<sub>2</sub> HCl (**3**) (2.9  $\mu$ M) or (C) G1-18-NH<sub>2</sub> HCl (**4**) (1.97  $\mu$ M) for 24h in serum deprived media. Plasmid DNA and dendrimer concentrations correspond to that of transfections at a charge ratio of 5 (dendrimer to plasmid DNA). No significant eGFP fluorescence was detected. Cell nuclei are labeled with Hoechst 33342 (blue).



# Figure S54. Transfection negative controls of Cy3-labelled anti-HSP70 siRNA alone and dendrimer vectors alone in U251N glioblastoma cells

Fluorescence micrographs of human U251N glioblastoma cells incubated with (A) Cy3-labeled anti-HSP70 siRNA (10 nM), (B) G1-12-NH<sub>2</sub> HCl (**3**) (1.18  $\mu$ M) or (C) G1-18-NH<sub>2</sub> HCl (**4**) (0.78  $\mu$ M) for 24h in serum deprived media. siRNA and dendrimer concentrations correspond to that of transfections at a charge ratio of 1 (dendrimer to siRNA). No significant Cy3 fluorescence was detected. Cell nuclei are labeled with Hoechst 33342 (blue).



# Figure S55. Down-regulation of heat-shock protein 70 (HSP70) protein expression in U251N glioblastoma cells using small interfering RNA (siRNA) delivered with G1-12 and G1-18 dendrimers.

The expression of HSP70 in control and transfected cells was verified by immunoblotting.  $\beta$ -actin served as loading control. Cells were treated with siRNA (10 nM) alone, siRNA complexed with dendrimers G1-12 (3) and G1-18 (4) at a charge ratio of 1 to 20 (siRNA to dendrimer), or dendrimers alone for 48h. Shown are representative immunoblots and mean percentage HSP70 expression (normalized to the loading control) as compared to the untreated control  $\pm$  S.D. from two independent experiments. (\*p<0.05)



Figure S56. Mitochondria metabolic (MTT) activity of HEK 293 human embryonic kidney cells and U251N human glioblastoma cells in response to transfection of eGFP plasmid DNA (pDNA) and heat-shock protein 70 (HSP70) small interfering RNA (siRNA), respectively, delivered with G1-12 (3) and G1-18 (4) dendrimers for 24h.

(A) HEK 293 cells were transfected with eGFP plasmid DNA (0.5  $\mu$ g/well) using G1-12 (3) or G1-18 (4) polyamine dendrimers at different charge ratios (dendrimer to pDNA) for 24h.

Lipofectamine 2000© was included as a commercial comparator, and was used as recommended by the manufacturer. Mitochondrial metabolic activity was measured by the MTT assay. Shown are mean percentage values  $\pm$  S.D. as compared to untreated controls. Each treatment was included in triplicate. (\*p<0.01)

(B) U251N cells were transfected with HSP70 siRNA (10 nM) using G1-12 or G1-18 polyamine dendrimers at different charge ratios (dendrimer to siRNA) for 24h. Lipofectamine 2000© was included as a commercial comparator, and was used as recommended by the manufacturer. Mitochondrial metabolic activity was measured by the MTT assay. Shown are mean percentage values  $\pm$  S.D. as compared to untreated controls. Each treatment was included in triplicate. (\*p<0.01)