Supporting Information

Rational Synthesis of Pd Nanoparticles-Embedded Reduced Graphene Oxide Frameworks with Enhanced Selective Catalysis in Water

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Figure S1. TEM (A) and SEM (B) images of GO-GOOH.



Figure S2. (A~D) TEM images of GOF; (E) the typical EDX pattern of GOF.



Figure S3. (A, B) TEM images, (C) the typical EDX pattern, and (D~F) SEM images of Pd²⁺-GOF.



Figure S4. The typical EDX pattern of Pd-rGOF.



Figure S5. (A) TEM images and (B) SEM images of Pd-rGOF after 10 th cycles.



Figure S6. (A) XPS spectrum wide scan, (B) XRD pattern, and (C) Raman spectrum of Pd-rGOF of the 3D Pd-rGOF after 10 th cycles. Inset of D shows the pore size distribution of the 3D Pd-rGOF.

$\begin{array}{c} 0 & 0 \\ 0 & 0 \\ 0 & 0 \\ 0 \\ 0 \\ \end{array} + \\ \begin{array}{c} Catalyst \\ PPh_3 \\ \end{array} + \\ \begin{array}{c} 0 & 0 \\ 0 \\ 0 \\ \end{array} + \\ \begin{array}{c} 0 & 0 \\ 0 \\ 0 \\ \end{array} + \\ \begin{array}{c} 0 & 0 \\ 0 \\ 0 \\ \end{array} + \\ \begin{array}{c} 0 & 0 \\ 0 \\ 0 \\ \end{array} + \\ \begin{array}{c} 0 & 0 \\ 0 \\ 0 \\ \end{array} + \\ \begin{array}{c} 0 & 0 \\ 0 \\ 0 \\ \end{array} + \\ \begin{array}{c} 0 & 0 \\ 0 \\ 0 \\ \end{array} + \\ \begin{array}{c} 0 & 0 \\ 0 \\ 0 \\ \end{array} + \\ \begin{array}{c} 0 & 0 \\ 0 \\ 0 \\ \end{array} + \\ \begin{array}{c} 0 & 0 \\ 0 \\ 0 \\ \end{array} + \\ \begin{array}{c} 0 & 0 \\ 0 \\ 0 \\ 0 \\ \end{array} + \\ \begin{array}{c} 0 & 0 \\ 0 \\ 0 \\ 0 \\ \end{array} + \\ \begin{array}{c} 0 & 0 \\ 0 \\ 0 \\ 0 \\ \end{array} + \\ \begin{array}{c} 0 & 0 \\ 0 \\ 0 \\ 0 \\ \end{array} + \\ \begin{array}{c} 0 & 0 \\ 0 \\ 0 \\ 0 \\ 0 \\ \end{array} + \\ \begin{array}{c} 0 & 0 \\ 0 \\ 0 \\ 0 \\ 0 \\ \end{array} + \\ \begin{array}{c} 0 & 0 \\ 0 \\ 0 \\ 0 \\ 0 \\ 0 \\ \end{array} + \\ \begin{array}{c} 0 & 0 \\ 0 \\ 0 \\ 0 \\ 0 \\ 0 \\ 0 \\ 0 \\ 0 \\$								
1		2	"	3	4			
Entry	$Pd_{(\mu mol)}$	PPh _{3(µmol)}	T(°C)	Yield $(\%)^b$	3 : 4 (%) ^b			
1	1.4	0	100	21	100:0			
2	1.4	3.5	100	38	75:25			
3	1.4	7.0	100	>99	0:100			
4	1.4	10.5	100	63	70:30			
5	0	7.0	100	0	-			
6	1.4	7.0	90	>99	20:80			
7	1.4	7.0	80	70	80:20			
8	1.4	7.0	70	50	80:20			
9^c	1.4	0	70	63	100:0			
10^d	1.4	7.0	70	70	80:20			
11^e	1.4	7.0	100	0	-			

Table S1 The optimal condition choice of Tsuji-Trost reaction using Pd/TETA/CGO catalyst in H_2O and air.^{*a*}

^a Reaction condition: 1(2.0 mmol), 2 (5.0 mmol), H₂O (3.0 mL), 0.5 h.

^b Yields of isolated products and determined by ¹H-NMR, ¹³C-NMR. Tetramethylsilane was used as an internal standard. ^{c, d} K₂CO₃ (2.5 mmol) was added. ^e **2** was blank.

Table S2 Tsuji-Trost reaction of various 1,3-dicarbonyl compounds with allyl ethyl carbonate in H_2O and air.^{*a*}

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Entry	1,3-Dicarbonyl	Catalyst	Time (h)	Mono-:Di-	Yield $(\%)^b$
1	0 0	Pd-GOF	0.5	0:100	>99
2		Pd/C	0.5	35:65	60
3	0 0	Pd-GOF	0.5	0:100	>99
4		Pd/C	0.5	20:80	61
5		Pd-GOF	1.0	0:100	>99
6	\sim_0	Pd/C	1.0	45:55	58
7	0 0	Pd-GOF	1.0	0:100	>99
8	\sim	Pd/C	1.0	41:59	67
9		Pd-GOF	1.5	0:100	90
10		Pd/C	1.5	0:100	64
11	0 0	Pd-GOF	1.0	100:0	90
12	$\sim 0^{-1}$	Pd/C	1.0	100:0	40
13	0	Pd-GOF	1.5	100:0	90
14	$\langle \mathcal{A}_{\mathbf{o}} \rangle$	Pd/C	1.5	100:0	60
15	0 0	Pd-GOF	2.0	100:0	60
16	$\sim_0 \sim \sim_0 \sim$	Pd/C	2.0	100:0	35

^{*a*} Reaction condition: Pd-rGOF (Pd: 1.4 μmol), Pd/C (Pd: 1.4 μmol), PPh₃ (7.0 μmol)), **1** (2.0 mmol), **2** (5.0 mmol), H₂O (3.0 mL), 100 °C.

^b Yields of isolated products and determined by ¹H-NMR and ¹³C-NMR.

1. Methyl 2-acetylpent-4-enoate

¹H NMR (400 MHz, CDCl₃) δ 2.18 (s, 3H), 2.51-2.56 (m, 2H), 3.50 (t, 1H), 3.68 (s, 3H), 4.97-5.07 (m, 2H), 5.63-5.73 (m, 1H) ¹³C NMR (100 MHz, CDCl₃) δ 29.3, 32.3, 52.5, 59.1, 117.6, 134.2, 169.8, 202.9 MS (EI) m/z (%): 113 (M⁺-CH₃CO), 97, 81, 71, 55, 43 (100)

Methyl 2-acetyl-2-allylpent-4-enoate



¹**H NMR** (400 MHz, CDCl₃) δ 2.14 (s, 3H), 2.56-2.67 (m, 4H), 3.74 (s, 3H), 5.09-5.14 (m, 4H), 5.54-5.65 (m, 2H)

¹³C NMR (100 MHz, CDCl₃) δ 27.0, 36.0, 52.4, 63.4, 119.3, 132.2, 171.9, 202.9 MS (EI) m/z (%): 154 (M⁺-CH₃CO), 149, 137, 123, 111, 95, 81, 71, 57, 43 (100)

2. Ethyl 2-acetylpent-4-enoate

¹**H** NMR (400 MHz, CDCl₃) δ 1.22 (t, J = 7.1Hz, 3H), 2.18 (s, 3H), 2.54 (quint, 2H), 3.46 (t, J = 7.4Hz, 1H), 4.15 (q, J = 7.1Hz, 2H), 4.97-5.06 (m, 2H), 5.63-5.74 (m, 1H) ¹³C NMR (100 MHz, CDCl₃) δ 14.0, 29.0, 32.0, 59.1, 61.3, 117.3, 134.1, 169.1, 202.3 MS (EI) m/z (%): 127 (M⁺ -CH₃CO), 99, 81, 55, 43 (100)

Ethyl 2-acetyl-2-allylpent-4-enoate

¹**H** NMR (400 MHz, CDCl₃) δ 1.27 (t, J = 7.1Hz, 3H), 2.14 (s, 3H), 2.61 (quint, 4H), 4.20 (q, J = 7.1Hz, 2H), 5.08-5.13 (m, 4H), 5.55-5.65 (m, 2H) ¹³**C** NMR (100 MHz, CDCl₃) δ 14.2, 26.9, 35.9, 61.4, 63.2, 118.2, 132.2, 172.1, 203.8 MS (EI) m/z (%): 168 (M⁺ -CH₃CO), 123, 95, 79, 67, 43 (100)

3. Isopropyl 2-acetylpent-4-enoate

¹H NMR (400 MHz, CDCl₃) δ 1.25 (d, 6H), 2.24 (s, 3H), 2.59 (quint, 2H), 2.47-2.51 (m, 1H), 5.04-5.13 (m, 3H), 5.70-5.80 (m, 1H)
¹³C NMR (100 MHz, CDCl₃) δ 21.8, 29.1, 32.2, 59.5, 69.2, 117.5, 134.3, 168.9, 202.4
MS (EI) m/z (%): 141 (M⁺-CH₃CO), 124, 99, 92, 82, 71, 57, 43 (100)

Isopropyl 2-acetyl-2-allylpent-4-enoate

¹H NMR (400 MHz, CDCl₃) δ 1.25 (d, 6H), 2.13 (s, 3H), 2.55-2.67 (m, 4H), 5.05-5.13 (m, 5H), 5.55-5.65 (m, 2H)
¹³C NMR (100 MHz, CDCl₃) δ 21.7, 26.9, 35.9, 63.1, 69.1, 119.3, 132.3, 171.6, 204.0
MS (EI) m/z (%): 180 (M⁺-CH₃CO), 178, 169, 149, 123, 95, 79, 57, 44 (100)

4. Isobutyl 2-acetylpent-4-enoate

¹H NMR (400 MHz, CDCl₃) δ 0.93 (d, 6H), 1.90-2.00 (m, 1H), 2.25 (s, 3H), 2.55-2.66 (m, 2H), 3.55 (t, 2H), 3.92 (d, 2H), 5.04-5.13 (m, 2H), 5.70-5.80 (m, 1H) ¹³C NMR (100 MHz, CDCl₃) δ 19.1, 27.8, 29.3, 32.2, 59.3, 71.6, 117.6, 134.3, 169.4, 202.5 MS (EI) m/z (%):155 (M⁺-CH₃CO), 142, 124, 109, 99, 82, 57, 43 (100)

Isobutyl 2-acetyl-2-allylpent-4-enoate

¹**H** NMR (400 MHz, CDCl₃) δ 0.93 (d, 6H), 1.88-1.98(m, 1H), 2.14 (s, 3H), 2.56-2.69 (m, 4H), 3.90 (d, 2H), 5.08-5.13 (m, 4H), 5.54-5.65 (m, 2H)

¹³C NMR (100 MHz, CDCl₃) δ 19.3, 26.9, 27.6, 36.0, 63.4, 71.7, 119.3, 132.1, 173.3, 203.8 MS (EI) m/z (%): 141 (M⁺-CH₃CO), 123, 95, 79, 57, 43 (100)

5. Ethyl 2-allyl-2-benzoylpent-4-enoate

¹**H NMR** (400 MHz, CDCl₃) δ 1.08 (t, J = 7.1Hz, 3H), 2.81 (m, 4H), 4.13 (q, J = 7.1Hz, 2H), 5.00-5.09 (m, 4H), 5.53-5.64 (m, 2H), 7.40-7.54 (m, 3H), 7.83-7.85 (m, 2H) ¹³**C NMR** (100 MHz, CDCl₃) δ 13.9, 37.2, 60.6, 61.5, 119.1, 128.5, 128.6, 132.0, 132.8, 136.0, 172.7, 196.2 **MS** (EI) m/z (%): 272 (M⁺), 198, 159, 105 (100), 77

6. Ethyl 2-acetyl-2-methylpent-4-enoate

¹**H** NMR (400 MHz, CDCl₃) δ 1.20 (t, J = 7.1Hz, 3H), 1.26 (s, 3H), 2.08 (s, 3H), 2.41-2.60 (m, 2H), 4.13(q, J = 7.1Hz, 2H), 4.97-5.09 (m, 2H), 5.53-5.64 (m, 1H) ¹³C NMR (100 MHz, CDCl₃) δ 14.1, 18.9, 26.2, 39.4, 59.4, 61.4, 119.0, 132.7, 172.5, 204.8 MS (EI) m/z (%): 142, 114, 97, 69, 43 (100) 7. 2-allyl-2-methylcyclopentane-1, 3-dione

¹H NMR (400 MHz, CDCl₃) δ 1.11 (s, 3H), 2.33-2.05 (m, 2H), 2.65-2.82 (m, 4H), 5.04-5.08 (m, 2H), 5.54-5.65 (m, 1H) ¹³C NMR (100 MHz, CDCl₃) δ 19.2, 36.8, 40.4, 57.1, 120.2, 131.9, 289.1 MS (EI) m/z (%): 152 (M⁺), 124, 111, 97 (100), 55, 41

8. Table 2 entry 21 (Mono-)

¹**H** NMR (400 MHz, CDCl₃) δ 1.19 (t, 6H), 2.56 (m, J = 7.2Hz, 2H), 3.35 (t, J = 7.5Hz, 1H), 4.12 (quint, J = 3.9Hz, 4H), 4.93-5.10 (m, 2H), 5.66-5.76 (m, 1H) ¹³**C** NMR (100 MHz, CDCl₃) δ 14.1, 32.8, 51.6, 61.3, 117.4, 134.1, 168.9 MS (EI) m/z (%): 155 (M⁺-OCH₂CH₃), 149, 127, 109, 98 (100), 81, 67, 55, 44

Copies of ¹H-NMR, ¹³C-NMR

Methyl 2-acetylpent-4-enoate







Ethyl 2-acetylpent-4-enoate







Isopropyl 2-acetylpent-4-enoate







Isobutyl 2-acetylpent-4-enoate











Ethyl 2-allyl-2-benzoylpent-4-enoate



2-allyl-2-methylcyclopentane-1, 3-dione







110 100 f1 (ppm)