

Electronic Supplementary Information (ESI)

A Highly Fluorescent AIE-active Theranostic Agent with Anti-Tumor Activity to Specific Cancer Cells

*Yueyue Zhao,^{ab‡} Ryan T. K. Kwok,^{ab‡} Jacky W. Y. Lam^{ab} and Ben Zhong Tang^{*abc}*

^a HKUST Shenzhen Research Institute, No. 9 Yuexing 1st Road, South Area, Hi-tech Park, Nanshan, Shenzhen 518057, China

^b Department of Chemistry, Hong Kong Branch of Chinese National Engineering Research Center for Tissue Restoration and Reconstruction, Institute for Advanced Study, Division of Biomedical Engineering, Division of Life Science, State Key Laboratory of Molecular Neuroscience, Institute of Molecular Functional Materials, The Hong Kong University of Science and Technology, Clear Water Bay, Kowloon, Hong Kong, China

^c Guangdong Innovative Research Team, SCUT-HKUST Joint Research Laboratory, State Key Laboratory of Luminescent Materials and Devices, South China University of Technology, Guangzhou 510640, China

[‡] These authors contribute equally to the work.

^{*} Corresponding author: tangbenz@ust.hk (B.Z.T)

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Experimental section

Materials

Benzophenone, 4-hydroxybenzophenone, zinc powder, titanium tetrachloride (TiCl_4), potassium carbonate (K_2CO_3), anhydrous dimethylsulfoxide (DMSO), acetone and 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) and other chemicals were all purchased from Sigma-Aldrich and used as received. 2-Bromo-*N,N*-dimethylethanamine hydrobromide was purchased from Annker Organics Co., Ltd. Dulbecco's Modified Eagle Medium (DMEM), modified essential medium (MEM) and LysoTracker red DND-99 (LTR) were the commercial products of Invitrogen. Fetal bovine serum (FBS) and trypsin-EDTA solution were purchased from Life Technologies. Tetrahydrofuran (THF) and dichloromethane (DCM) were distilled from sodium benzophenone ketyl and calcium hydride under nitrogen, respectively. Milli-Q water was supplied by Milli-Q Plus System (Millipore Corporation, United states).

Instruments

^1H NMR and ^{13}C NMR spectra were measured on a Bruker AV 400 NMR spectrometer using CDCl_3 as a solvent and tetramethylsilane (TMS; $\delta = 0$) as an internal reference. Absorption spectra were measured on a Milton Roy Spectronic 3000 Array spectrophotometer. Photoluminescence (PL) spectra were recorded on a Perkin-Elmer LS 55 spectrofluorometer with a xenon discharge lamp excitation. High-resolution mass spectra (HRMS) were recorded on a GCT Premier CAB 048 mass spectrometer system operated in MALDI-TOF mode.

Synthesis of TPE-OH

Into a two-necked round bottom flask equipped a condenser were added a dry THF

solution (40 mL) of benzophenone (1.82 g, 10 mmol), 4-hydroxybenzophenone (1.90 g, 10 mmol), zinc powder (1.60 g, 24 mmol) under nitrogen. The mixture was cooled to -78 °C and TiCl₄ (1.3 mL, 12 mmol) was injected dropwise. The mixture was slowly warmed to room temperature, stirred for 0.5 h, and then heated to reflux overnight. After the mixture was cooled to room temperature, dilute hydrochloric acid was added to quench the reaction. The mixture was extracted with dichloromethane (DCM) three times and the combined organic layer was washed with brine and dried over anhydrous sodium sulfate. After solvent evaporation, the crude product was purified on a silica-gel column using petroleum ether/DCM as eluent. A white solid was obtained. ¹H NMR (400 MHz, CDCl₃), δ (ppm): 7.12–7.07 (m, 9H), 7.07–7.01 (m, 6H), 6.90–6.88 (d, 2H), 6.57–6.55 (d, 2H).

Synthesis of TPE-TMX

Into a two-necked round bottom flask equipped a condenser were added an acetone solution (15 mL) of 2-bromo-*N,N*-dimethylethanamine hydrobromide (1.74 g, 7.5 mmol), TPE-OH (1.74 g, 5 mmol), potassium carbonate (3.45 g, 25 mmol). Afterwards, the mixture was heated to 85 °C overnight. After the mixture cooled to room temperature, the mixture was extracted with DCM three times and the combined organic layer was washed with brine and dried over anhydrous sodium sulfate. After solvent evaporation, the crude product was purified on an aluminum oxide-gel column to afford the desirable white solid. ¹H NMR (400 MHz, CDCl₃), δ (ppm): 7.14–7.10 (m, 9H), 7.07–7.03 (m, 6H), 6.96–6.94 (d, 2H, *J* = 8.4 Hz), 4.06–4.03 (d, 2H, *J* = 4.5 Hz), 2.78–2.76 (d, 2H, *J* = 4.5 Hz), 2.39 (m, 6H). ¹³C NMR (100 MHz, CDCl₃), δ (TMS, ppm): 157.2, 144.0, 143.8, 140.5, 140.2, 136.3, 132.6, 131.4, 131.4, 131.3, 127.7, 127.6, 126.4, 126.3, 113.7, 77.4, 77.1, 76.8, 65.5, 58.2, 45.8. MS (MALDI-TOF): *m/z* 418.2 (M⁺, calcd. 419.2).

Cell culture

MCF-7, MDA-MB-231, HeLa and COS-7 cell lines were provided by American Type Culture Collection. The HeLa cells were cultured in MEM while MCF-7, MDA-MB-231 and COS-7 cells were cultured in DMEM at 37 °C in a humidified incubator with 5% CO₂. Both culture mediums contained 10% heat-inactivated FBS 100 U mL⁻¹ penicillin and 100 µg mL⁻¹ streptomycin. Before experiment, the cells were pre-cultured until confluence was reached.

Cell imaging

Cells were grown overnight on a 35 mm petri dish with a cover slip. The live cells were incubated with the aqueous solution of TPE-TMX (2 µM) for 24 h followed by LTR (50 nM) for 15 min. The dye-labelled cells were then mounted in standard mounting media and imaged by fluorescence microscopy (BX 41 Microscope). Conditions: for TPE-TMX, excitation wavelength: 330–385 nm, dichroic mirror: 400 nm and emission long pass filter: 420 nm; for LTR, excitation filter: 540–580 nm, dichroic mirror: 600 nm and emission long pass filter: 610 nm.

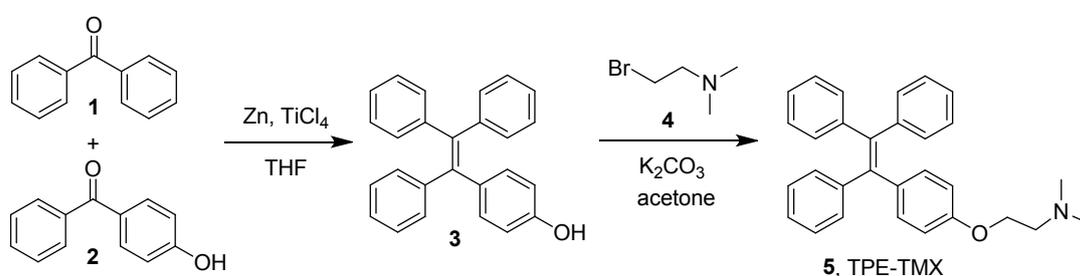
Photostability test

The dye-labelled MCF-7 cells were imaged by confocal microscope (Zeiss laser scanning confocal microscope LSM7 DUO). Conditions: excitation wavelength: 405 nm and emission filter: 500–750 nm (TPE-TMX); excitation wavelength: 561 nm and emission filter: 580–750 nm (LTR). Laser power was unified as 0.1 mW.

Cytotoxicity study

MTT assays were used to evaluate the cytotoxicity of TPE-TMX to different cell lines. For each cell line, the cells were seeded in 96-well plates (Costar, IL, USA) at a density of 5×10^3 cells/well. After 24 h incubation, the cells were exposed to a series

of doses of TPE-TMX (0–10 μM) in culture medium at 37 $^{\circ}\text{C}$. After 24 h incubation, 10 μL of freshly prepared MTT solution (5 mg/mL in PBS) was added into each well. After 4 h incubation, 100 μL of solubilizing solution containing 10% SDS and 0.01 M HCl was added to dissolve the purple crystals. After 8 h incubation, the absorbance of MTT at 595 nm was monitored using a Perkin-Elmer Victor plate reader. Cell viability was expressed by the ratio of absorbance of the cells incubated with TPE-TMX to that of cells incubated with culture medium only. Each of the experiments was performed at least three times.



Scheme S1 Synthetic route of TPE-TMX.

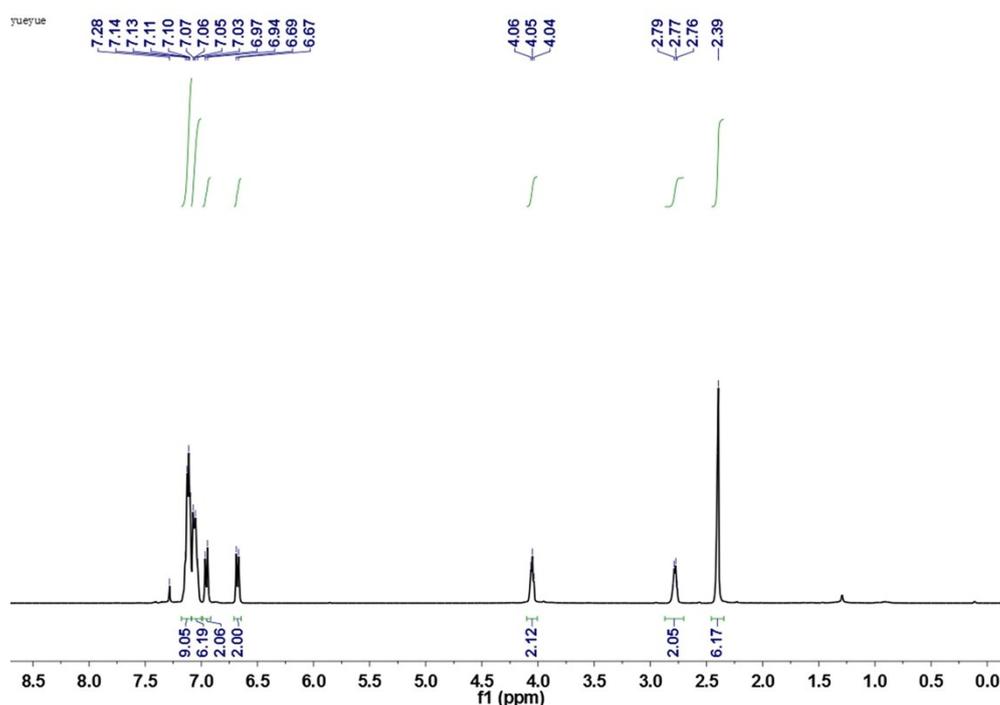


Fig. S1 ^1H NMR spectrum of TPE-TMX in CDCl_3 .

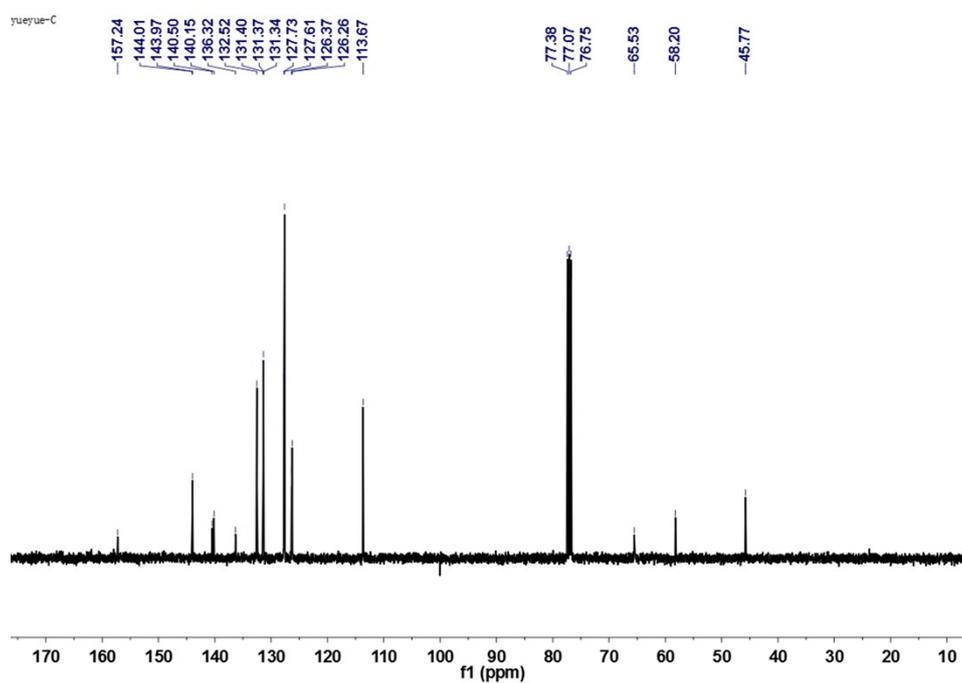


Fig. S2 ^{13}C NMR spectrum of TPE-TMX in CDCl_3 .

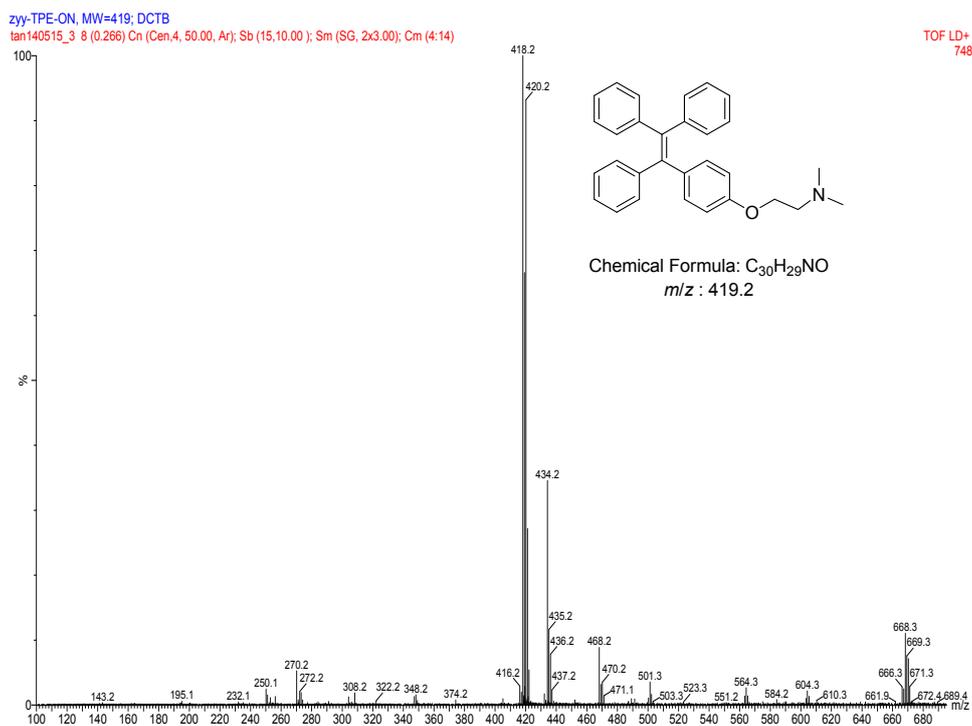


Fig. S3 Mass spectrum of TPE-TMX.

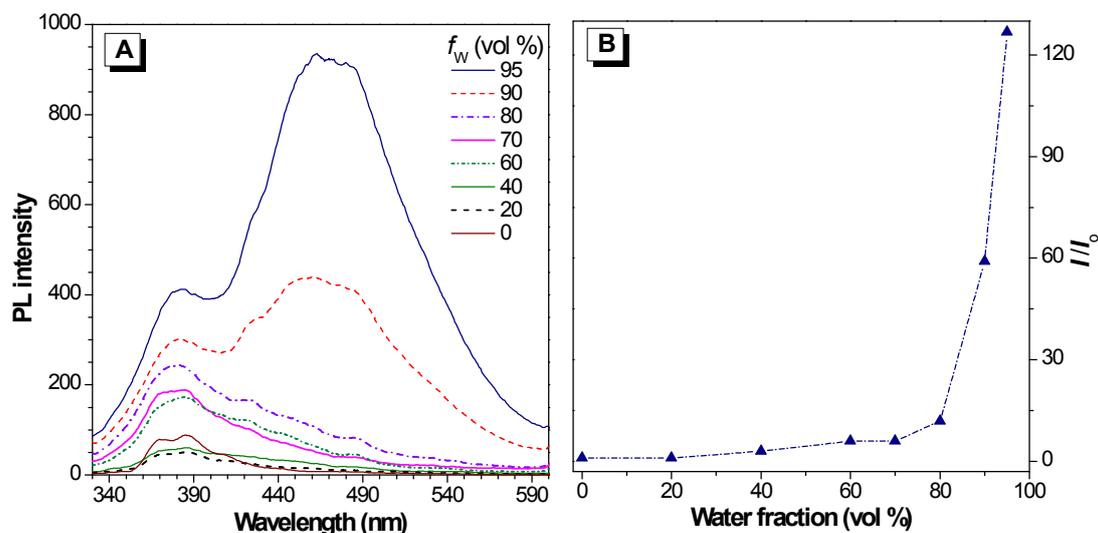


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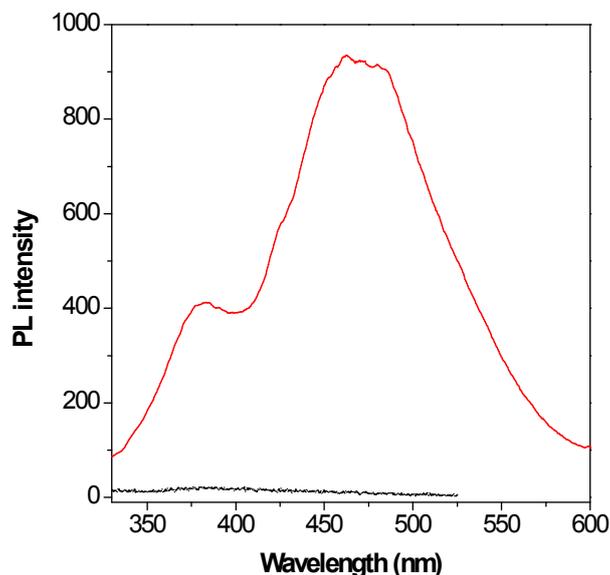


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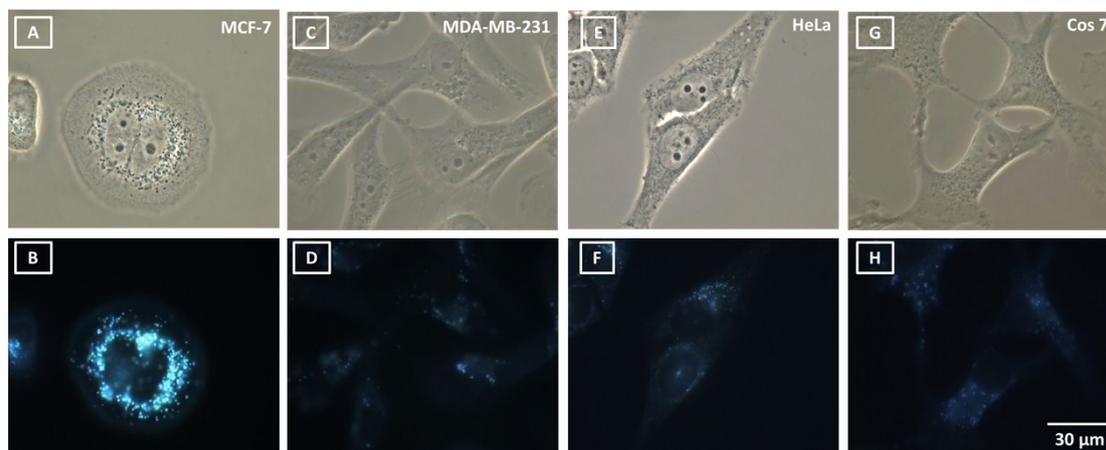


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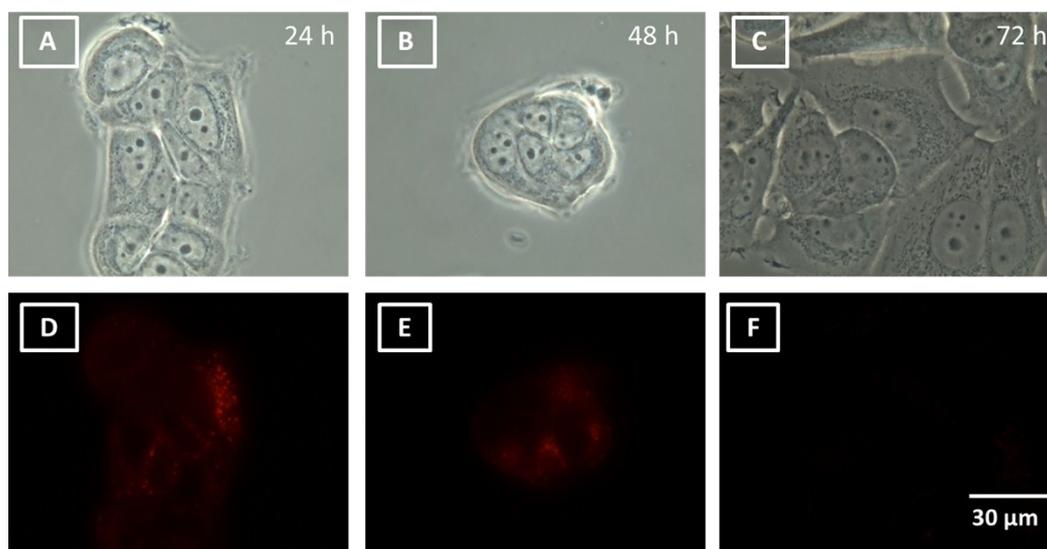


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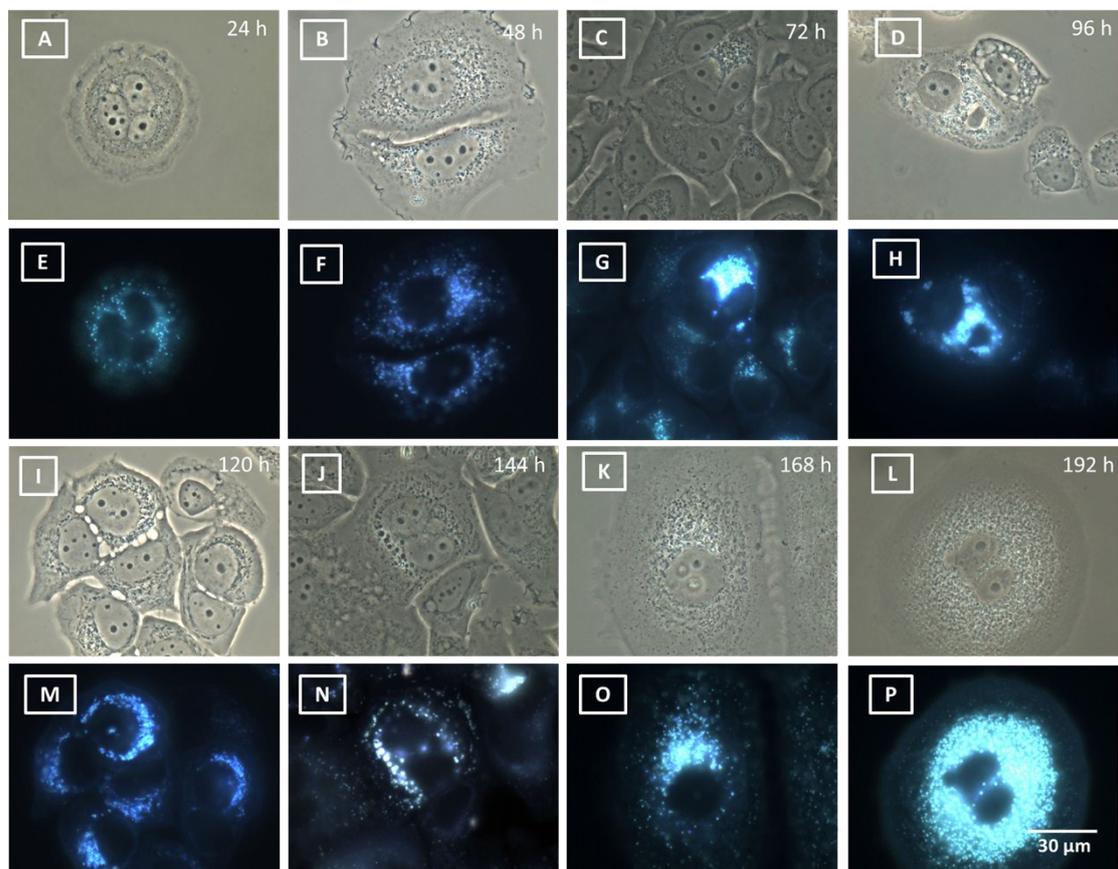


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