

Supporting Information

One-pot Total Synthesis of Streptindole, Arsindoline B and Their Congeners Through Tandem Decarboxylative Deaminative Dual-Coupling Reaction of Amino Acids with Indoles

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1. General

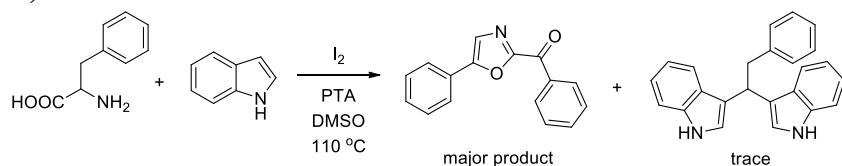
All substrates and reagents were commercially available and used without further purification. TLC analysis was performed using pre-coated glass plates. Column chromatography was performed using silica gel (200–300 mesh). IR spectra were recorded on a Perkin-Elmer PE-983 infrared spectrometer as KBr pellets with absorption in cm^{-1} . ^1H spectra were recorded in CDCl_3 or $\text{DMSO}-d_6$ or CD_3COCD_3 on 400/600 MHz NMR spectrometers and resonances (δ) are given in parts per million relative to tetramethylsilane. Data are reported as follows: chemical shift, multiplicity (s = singlet, d = doublet, t = triplet, m = multiplet q = quadruple), coupling constants (Hz) and integration. ^{13}C spectra were recorded in CDCl_3 or $\text{DMSO}-d_6$ or CD_3COCD_3 on 100/150 MHz NMR spectrometers and resonances (δ) are given in ppm. HRMS were obtained on a Bruker 7-tesla FT-ICR MS equipped with an electrospray source. The X-ray crystal-structure determinations of **3b** and **3g** were obtained on a Bruker SMART APEX CCD system. Melting points were determined using XT-4 apparatus and not corrected. Optical rotation were measured with polarimeter: Atopol IV (an average value of 10 times parallel tests).

2. General procedure for the synthesis of **3** and **4** (**3a** and **4a** as an example)

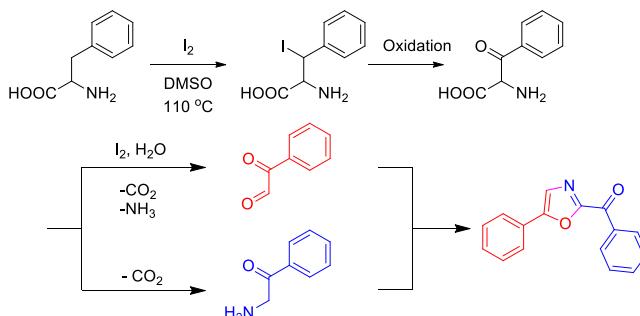
A mixture of 2-amino-2-phenylacetic acid **1a** (0.5 mmol), 1H-indole **2a** (1.0 mmol), alloxan monohydrate (1.0 mmol), phosphotungstic acid 44-hydrate (0.125 mmol) in DMF (1.0 mL) was stirred at 110 °C for 6 hours. Then added 50 mL water and 30 mL saturated brine solution to the mixture, extracted with EtOAc 3 times (3×50 mL). The extract was washed with 10% NaHCO_3 solution, dried over anhydrous Na_2SO_4 and concentrated under reduced pressure. The crude product was purified by column chromatography on silica gel (eluent: petroleum ether/EtOAc=5/1) to afford the product **3a** as yellow solid.

A mixture of 3-acetoxy-2-aminopropanoic acid hydrochloride **1'a** (0.5 mmol), 1H-indole **2a** (1.0 mmol), alloxan monohydrate (1.0 mmol), phosphotungstic acid 44-hydrate (0.125 mmol) in DMF (1.0 mL) was stirred at 110 °C for 6 hours. Then added 50 mL water and 30 mL saturated brine solution to the mixture, extracted with EtOAc 3 times (3×50 mL). The extract was washed with 10% NaHCO_3 solution, dried over anhydrous Na_2SO_4 and concentrated under reduced pressure. The crude product was purified by column chromatography on silica gel (eluent: petroleum ether/EtOAc=2/1) to afford the product **4a** as yellow oil.

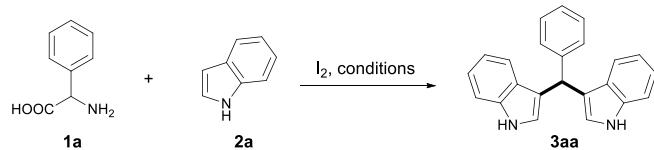
3. The side reaction of benzyl group amino acid in present of I_2 (phenylalanine as an example)



Probable Reaction Pathway:



4. Optimization details

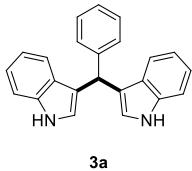


entry	solvent	Additive ^a	Acid	temp (°C)	Time	Yield ^j (%)
1	DMSO	AM	—	100	6h	22
2	DMA	AM	—	100	6h	15
3	DMF	AM	—	100	6h	40
4	CH ₃ CN	AM	—	reflux	6h	11
5	<i>t</i> -BuOH	AM	—	reflux	6h	trace
6	dioxane	AM	—	reflux	6h	trace
7	H ₂ O	AM	—	reflux	6h	0
8	DMF	AM	—	80	6h	trace
9	DMF	AM	—	90	6h	27
10	DMF	AM	—	110	6h	44
11	DMF	AM	—	120	6h	43
12	DMF	AM	—	110	4h	49
13	DMF	AM	TFA (0.25 eq.)	110	4h	75
14	DMF	AM	CF ₃ SO ₃ H (0.25 eq.)	110	4h	73
15	DMF	AM	MeSO ₃ H (0.25 eq.)	110	4h	62
16	DMF	AM	HI (0.25 eq.)	110	4h	67
17	DMF	AM	CuSO ₄ 5H ₂ O (0.25 eq.)	110	4h	27
18	DMF	AM	CuBr ₂ (0.25 eq.)	110	4h	15
19	DMF	AM	CuCl ₂ (0.25 eq.)	110	4h	79
20	DMF	AM	ZnCl ₂ (0.25 eq.)	110	4h	80
21	DMF	AM	AlCl ₃ (0.25 eq.)	110	4h	78
22	DMF	AM	FeCl ₃ (0.25 eq.)	110	4h	74
23	DMF	AM	Ti(i-PrO) ₄ (0.25 eq.)	110	4h	0
24 ^b	DMF	AM0.2	PTA (0.25 eq.)	110	4h	23
25 ^c	DMF	AM0.4	PTA (0.25 eq.)	110	4h	37
27 ^d	DMF	AM2.0	PTA (0.25 eq.)	110	6h	80
28	DMF	AM	PTA (0.25 eq.)	110	4h	86
29	DMF	AM	PTA (1.0 eq.)	110	4h	trace
30	DMF	AM	PTA (0.5 eq.)	110	4h	37

31 ^e	DMF	I ₂	PTA (1.0 eq.)	110	4h	84
32 ^f	DMSO	I ₂	PTA (1.0 eq.)	110	4h	87
33 ^g	DMSO	I ₂	PTA (1.0 eq.)	110	4h	37
34 ^h	DMSO	I ₂	PTA (1.0 eq.)	110	4h	0
35 ⁱ	DMSO	I ₂	PTA (1.0 eq.)	110	4h	trace
36	DMSO	NaClO	PTA (1.0 eq.)	110	4h	0
37	DMF	NaClO	PTA (1.0 eq.)	110	4h	0
38	DMF	IBX	PTA (1.0 eq.)	110	4h	0

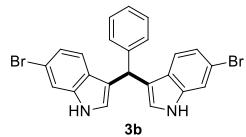
AM = Alloxan Monohydrate; PTA = phosphotungstic acid 44-hydrate; ^eReaction conditions: **1a** (0.5 mmol), **2a** (1.0 mmol), Additive (1.0 mmol. If no otherwise specified), solvent (1.0 mL). ^bAM (0.2 eq.). ^cAM (0.4 eq.). ^dAM (2.0 eq.). ^eI₂ (1.0 eq.). ^fI₂ (0.2 eq.). ^gI₂ (0.2 eq.). for aliphatic amino acid. ^hI₂ (0.2 eq.). for amino acid hydrochloride. ⁱI₂ (0.2 eq.). for aromatic amino acids. ^jisolated yields.

5. Characterization data for compound 3, 4 and 6



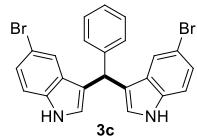
3,3'-(phenylmethylene)bis(1H-indole) (**3a**)

yellow solid; mp = 86-87 °C. ¹H NMR (600 MHz, dmso-*d*₆) δ 10.84 (br, 2H), 7.37 (m, 4H), 7.31 – 7.24 (m, 4H), 7.16 (t, *J* = 7.2 Hz, 1H), 7.04 (t, *J* = 7.2 Hz, 2H), 6.90 – 6.82 (m, 4H), 5.85 (s, 1H). ¹³C NMR (100 MHz, CDCl₃) IR (KBr): 1620.79, 1490.42, 1454.28, 1416.13, 1337.45, 1216.99, 1091.94, 1009.68. HRMS (ESI): m/z [M+Na]⁺ calcd for C₂₃H₁₈N₂Na : 345.1362; found: 345.1365.



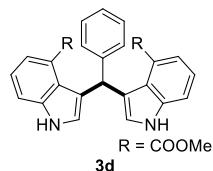
3,3'-(phenylmethylene)bis(6-bromo-1H-indole)

Red solid; mp = 140-142 °C. ¹H NMR (600 MHz, dmso-*d*₆) δ 11.03 (br, 1H), 7.57 (s, 2H), 7.34 (m, 2H), 7.27 (m, 2H), 7.19 (m, 3H), 7.01 (d, *J* = 7.8 Hz, 2H), 6.87 (s, 2H), 5.83 (s, 1H). ¹³C NMR (100 MHz, dmso-*d*₆) δ 144.34, 137.50, 137.34, 128.25, 126.05, 125.60, 124.68, 124.52, 121.18, 120.84, 118.16, 114.10, 113.86. IR (KBr): 1687.23, 1610.42, 1451.65, 1404.27, 1332.34, 1284.79, 1221.54, 1133.97, 1094.93, 1047.51. HRMS (ESI): m/z [M+Na]⁺ calcd for C₂₃H₁₆Br₂N₂Na : 500.9572; found: 500.9573.



3,3'-(phenylmethylene)bis(5-bromo-1H-indole)

yellow solid; mp = 230-232 °C; ¹H NMR (600 MHz, dmso-*d*₆) δ 11.09 (br, 2H), 7.43 (s, 2H), 7.34 (d, *J* = 8.4 Hz, 4H), 7.29 (m, 2H), 7.24 – 7.11 (m, 3H), 6.89 (s, 2H), 5.86 (s, 1H). ¹³C NMR (100 MHz, dmso-*d*₆) δ 144.32, 135.31, 128.42, 128.26, 126.11, 125.30, 123.53, 121.26, 117.70, 113.65, 111.00. IR (KBr): 1598.14, 1558.08, 1455.91, 1416.74, 1333.07, 1216.33, 1098.23. HRMS (ESI): m/z [M+Na]⁺ calcd for C₂₃H₁₆Br₂N₂Na : 500.9572; found: 500.9581



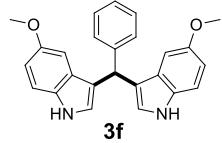
dimethyl 3,3'-(phenylmethylene)bis(1H-indole-4-carboxylate)

White solid; mp = 251-253 °C; ¹H NMR (600 MHz, dmso-*d*₆) δ 11.12 (br, 2H), 7.59 (d, *J* = 7.8Hz, 2H), 7.23 (m, 4H), 7.16 (m, 1H), 7.12 (m, 2H), 6.99 (d, *J* = 7.2 Hz, 2H), 6.39 (s, 1H), 6.32 (s, 2H), 3.38 (s, 6H). ¹³C NMR (100 MHz, dmso-*d*₆) δ 168.56, 145.57, 137.90, 129.02, 127.62, 127.24, 125.53, 124.85, 122.84, 120.73, 119.88, 119.02, 115.33, 51.47, 40.98. IR (KBr): 2948.80, 1704.03, 1613.51, 1495.85, 1433.76, 1344.88, 1280.51, 1243.23, 1198.55, 1143.32, 1036.28. HRMS (ESI): m/z [M+Na]⁺ calcd for C₂₇H₂₂N₂NaO₄ : 461.1472; found: 461.1475.



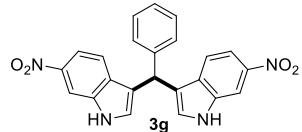
dimethyl 3,3'-(phenylmethylene)bis(1H-indole-6-carboxylate)

White solid; mp = 258-261 °C; ¹H NMR (600 MHz, dmso-*d*₆) δ 11.31 (br, 2H), 8.05 (s, 2H), 7.51 (d, *J* = 8.4 Hz, 2H), 7.36 (d, *J* = 8.4 Hz, 4H), 7.29 (t, *J* = 7.8 Hz, 2H), 7.20 (t, *J* = 7.2 Hz, 1H), 7.11 (d, *J* = 2.4 Hz, 2H), 5.93 (s, 1H), 3.83 (s, 6H). ¹³C NMR (100 MHz, CDCl₃) δ 167.28, 144.29, 135.87, 129.98, 128.80 – 128.39 (m), 127.67, 122.07, 119.03, 118.85, 118.46, 113.64, 51.7, IR (KBr): 1696.87, 1623.33, 1567.29, 1500.36, 1438.20, 1410.83, 1359.07, 1303.66, 1273.72, 1222.66, 1088.27. HRMS (ESI): m/z [M+Na]⁺ calcd for C₂₇H₂₂N₂NaO₄ : 461.1472; found: 461.1475.



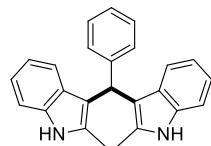
3,3'-(phenylmethylenes)bis(5-methoxy-1H-indole)

Pale red solid; mp = 215-216 °C; ¹H NMR (600 MHz, dmso-*d*₆) δ 10.74 (s, 2H), 7.44 (d, *J* = 7.8 Hz, 2H), 7.36 – 7.26 (m, 4H), 7.19 (t, *J* = 7.2 Hz, 1H), 6.91 (s, 2H), 6.84 (s, 2H), 6.77 (d, *J* = 8.4 Hz, 2H), 5.84 (s, 1H), 3.62 (s, 6H). ¹³C NMR (150 MHz, dmso-*d*₆) δ 152.81, 145.11, 131.95, 128.45, 128.11, 127.17, 125.84, 124.44, 117.85, 112.16, 110.70, 101.58, 55.30, 39.83. IR (KBr): 2937.48, 2829.91, 1622.11, 1584.33, 1485.20, 1450.00, 1331.96, 1291.41, 1258.69, 1208.51, 1171.40, 1127.40, 1095.04, 1024.07. HRMS (ESI): m/z [M+Na]⁺ calcd for C₂₅H₂₂N₂NaO₂ : 405.1573; found: 405.1578.



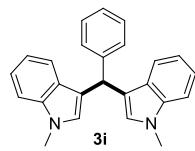
3,3'-(phenylmethylenes)bis(6-nitro-1H-indole)

Yellow solid; mp = 268-270 °C; ¹H NMR (400 MHz, dmso-*d*₆) δ 11.69 (br, 2H), 8.33 (s, 2H), 7.78 (d, *J* = 8.8 Hz, 2H), 7.43 (d, *J* = 8.8 Hz, 2H), 7.35 (s, 2H), 7.31 (s, 4H), 7.21 (s, 1H), 6.02 (s, 1H). ¹³C NMR (100 MHz, dmso-*d*₆) δ 141.93, 134.99, 131.04, 130.7, 128.48, 128.21, 119.17, 118.93, 113.67, 109.36, 108.51. IR (KBr): 1737.65, 1619.40, 1587.25, 1501.51, 1456.64, 1416.07, 1373.16, 1333.60, 1105.12, 1057.28. HRMS (ESI): m/z [M+Na]⁺ calcd for C₂₃H₁₆N₄NaO₄ : 435.1064; found: 435.1070.



3,3'-(phenylmethylenes)bis(2-methyl-1H-indole)

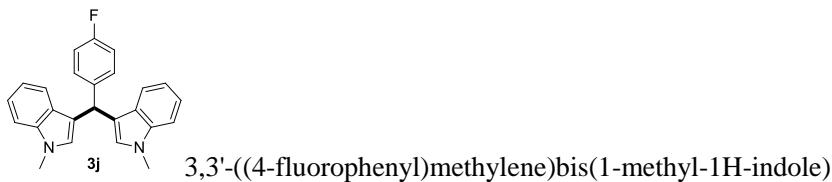
Yellow solid; mp = 257-258 °C; ¹H NMR (600 MHz, dmso-*d*₆) δ 10.75 (s, 2H), 7.27 – 7.22 (m, 2H), 7.22 (m, 1H), 7.20 (m, 4H), 6.88 (t, *J* = 7.8 Hz, 2H), 6.80 (d, *J* = 7.8 Hz, 2H), 6.67 (t, *J* = 7.2 Hz, 2H), 5.92 (s, 1H), 2.06 (s, 6H). ¹³C NMR (100 MHz, dmso-*d*₆) δ 143.93, 134.79, 131.75, 128.40, 128.01, 127.56, 125.41, 119.25, 118.23, 117.65, 111.92, 110.00, 38.38, 11.63. IR (KBr): 2919.23, 1598.01, 1488.07, 1460.10, 1426.35, 1340.51, 1298.34, 1220.93, 1133.17, 1011.65. HRMS (ESI): m/z [M+Na]⁺ calcd for C₂₅H₂₂N₂Na : 373.1675; found: 373.1676.



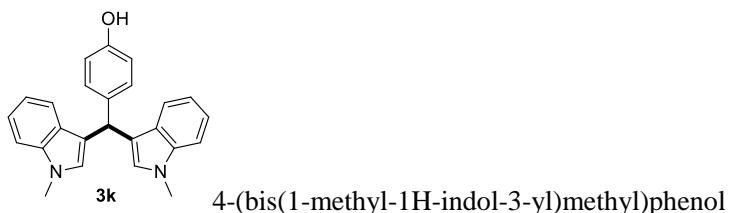
3,3'-(phenylmethylenes)bis(1-methyl-1H-indole)

Pale yellow solid; mp = 200-201 °C; ¹H NMR (600 MHz, CDCl₃) δ 7.40 (d, *J* = 7.8 Hz, 2H), 7.36 (m, 2H), 7.33 – 7.27 (m, 4H), 7.22 (m, 3H), 7.01 (t, *J* = 7.2 Hz, 2H), 6.54 (s, 2H), 5.90 (s, 1H), 3.69 (s, 6H). ¹³C NMR (100 MHz, dmso-*d*₆) δ 144.38, 137.32, 128.64, 128.23, 128.16, 127.37, 125.97, 121.35,

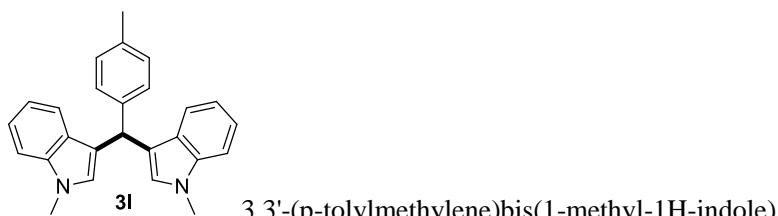
119.98, 118.57, 118.16, 109.02, 40.01, 32.66. IR (KBr): 2933.88, 1615.96, 1549.16, 1474.03, 1424.27, 1368.49, 1328.04, 1227.01, 1200.36, 1152.02, 1119.60, 1055.18, 1010.97. HRMS (ESI): m/z [M+Na]⁺ calcd for C₂₅H₂₂N₂Na : 373.1675; found: 373.1677.



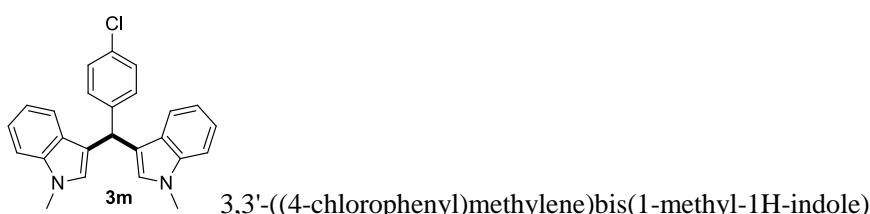
Pale yellow solid; mp = 202-204 °C; ¹H NMR (600 MHz, CDCl₃) δ 7.35 (d, *J* = 7.8 Hz, 2H), 7.28 (m, 4H), 7.20 (m, 3H), 6.97 (m, 4H), 6.50 (s, 2H), 5.86 (s, 1H), 3.66 (s, 6H). ¹³C NMR (100 MHz, CDCl₃) δ 161.27 (*d*, *J* = 242 Hz), 140.06, 137.34, 129.96 (*d*, *J* = 7.4 Hz), 128.17, 127.22, 121.47, 119.90, 118.66, 118.01, 114.88 (*d*, *J* = 21 Hz), 109.09, 39.28, 32.65. IR (KBr): 2935.40, 2845.35, 1894.64, 1599.08, 1550.48, 1501.82, 1472.47, 1424.37, 1368.57, 1328.47, 1218.25, 1153.02, 1119.65, 1091.86, 1056.89, 1010.97. HRMS (ESI): m/z [M+Na]⁺ calcd for C₂₅H₂₁FN₂Na : 391.1581; found: 391.1581.



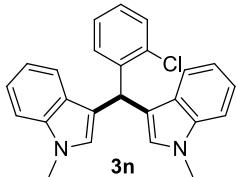
Red solid; mp = 248-249 °C; ¹H NMR (400 MHz, dmso-*d*₆) δ 9.21 (s, 1H), 7.34 (m, 4H), 7.14 (m, 4H), 6.92 (m, 2H), 6.78 (s, 2H), 6.70 (m, 2H), 5.74 (s, 1H), 3.67 (s, 6H). ¹³C NMR (100 MHz, dmso-*d*₆) δ 155.40, 136.98, 135.00, 129.13, 127.77, 126.95, 121.00, 119.35, 118.26, 117.99, 114.88, 109.54, 32.21. IR (KBr): 3457.56, 3107.37, 3050.25, 2933.36, 1669.94, 1604.61, 1545.20, 1508.17, 1474.58, 1370.26, 1330.18, 1289.21, 1254.72, 1231.94, 1196.96, 1159.19, 1129.67, 1053.64, 1011.37. HRMS (ESI): m/z [M+Na]⁺ calcd for C₂₅H₂₂N₂NaO : 389.1624; found: 389.1623.



Red solid; mp = 202-203 °C; ¹H NMR (600 MHz, CDCl₃) δ 7.38 (d, *J* = 7.8 Hz, 2H), 7.26 (m, 2H), 7.22 (m, 2H), 7.19 – 7.15 (m, 2H), 7.06 (d, *J* = 7.8 Hz, 2H), 6.97 (t, *J* = 7.2 Hz, 2H), 6.51 (s, 2H), 5.84 (s, 1H), 3.62 (s, 6H), 2.30 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 141.35, 137.29, 135.29, 128.83, 128.45, 128.13, 127.38, 121.29, 119.99, 118.51, 118.34, 108.97, 39.58, 32.59, 21.08. IR (KBr): 2915.01, 2825.11, 1909.76, 1614.31, 1550.34, 1507.85, 1472.65, 1422.66, 1370.30, 1328.83, 1224.78, 1151.72, 1118.45, 1057.13, 1011.48. HRMS (ESI): m/z [M+Na]⁺ calcd for C₂₆H₂₄N₂Na : 387.1832; found: 387.1831.

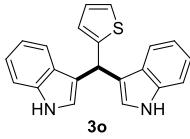


Red solid; mp = 209-212 °C; ¹H NMR (600 MHz, CDCl₃) δ 7.34 (d, *J* = 7.8 Hz, 2H), 7.31 – 7.23 (m, 4H), 7.20 (m, 4H), 6.99 (m, 2H), 6.49 (s, 2H), 5.84 (s, 1H), 3.64 (s, 6H). ¹³C NMR (100 MHz, CDCl₃) δ 142.94, 137.32, 131.57, 129.98 (s), 128.27, 128.20, 127.17, 121.50, 119.85, 118.70, 117.62, 109.11, 39.43, 32.64. IR (KBr): 2930.46, 2862.45, 2818.70, 1902.16, 1614.17, 1586.11, 1547.31, 1480.63, 1329.67, 1253.08, 1224.12, 1200.90, 1152.55, 1127.68, 1085.48, 1058.98, 1011.98. HRMS (ESI): m/z [M+Na]⁺ calcd for C₂₅H₂₁ClN₂Na : 407.1285; found: 407.1287.



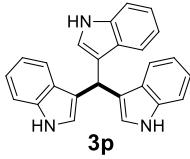
3,3'-(2-chlorophenyl)methylenebis(1-methyl-1H-indole)

White solid; mp = 264-265 °C; ¹H NMR (600 MHz, CDCl₃) δ 7.39 (m, 3H), 7.28 (m, 2H), 7.228 (m, 4H), 7.138 (m, 2H), 6.50 (s, 2H), 6.34 (s, 1H), 3.66 (s, 6H). ¹³C NMR (100 MHz, CDCl₃) δ 141.69, 137.40, 133.83, 130.30, 129.43, 128.38, 127.35, 126.56, 121.47, 119.92, 118.68, 116.83, 109.06, 36.45, 32.68. IR (KBr): 2929.96, 2821.48, 1922.60, 1613.35, 1548.40, 1475.43, 1424.53, 1371.06, 1331.02, 1245.64, 1222.74, 1202.59, 1156.04, 1127.08, 1035.23, 1010.70. HRMS (ESI): m/z [M+Na]⁺ calcd for C₂₅H₂₁ClN₂Na : 407.1285; found: 407.1287.



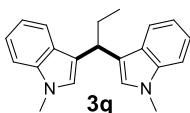
3,3'-(thiophen-2-ylmethylene)bis(1H-indole)

Pale red solid; mp = 171-174 °C; ¹H NMR (600 MHz, CDCl₃) δ 7.90 (s, 2H), 7.47 (d, *J* = 7.8 Hz, 2H), 7.35 (d, *J* = 8.4 Hz, 2H), 7.21 – 7.14 (m, 3H), 7.04 (M, 2H), 6.92 (M, 2H), 6.83 (s, 2H), 6.17 (s, 1H). ¹³C NMR (100 MHz, CDCl₃) δ 148.60, 136.55, 126.73, 126.39, 125.11, 123.58, 123.14, 122.00, 119.74, 119.70, 119.35, 111.08, 77.32, 77.00, 76.68, 35.30. IR (KBr): 1619.36, 1456.24, 1418.50, 1336.37, 1217.33, 1091.23, 1009.90. HRMS (ESI): m/z [M+Na]⁺ calcd for C₂₁H₁₆N₂S : 351.0926; found: 351.0929.



tri(1H-indol-3-yl)methane

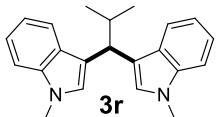
Pale yellow oil; ¹H NMR (400 MHz, CDCl₃) δ 10.73 (br, 3H), 7.41 (d, *J* = 6.8 Hz, 3H), 7.35 (d, *J* = 6.8 Hz, 3H), 7.02 (m, 3H), 6.95 (s, 3H), 6.86 (m, 3H), 6.07 (s, 1H). ¹³C NMR (100 MHz, CDCl₃) δ 136.54, 126.73, 123.16, 120.57, 119.25, 118.24, 117.89, 111.33. IR (KBr): 3258.22, 3052.04, 2922.32, 2588.32, 1618.05, 1485.57, 1455.06, 1338.65, 1240.95, 1216.52, 1225.91, 1093.99, 1022.28, 1003.19. MS: m/z [M]⁺ calcd for C₂₉H₁₉N₃ : 361.16; found: 361.36.



⁶⁰⁰NMR
CDCl₃

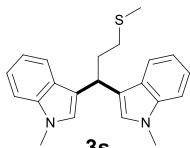
3,3'-(propane-1,1-diyl)bis(1-methyl-1H-indole)

White solid; mp = 88–90 °C; ¹H NMR (600 MHz, CDCl₃) δ 7.77 (m, 2H), 7.42 – 7.36 (m, 2H), 7.33 (s, 2H), 7.18 (m, 2H), 6.97 (s, 2H), 4.52 (m, 1H), 3.78 (s, 6H), 2.37 (m, 2H), 1.21 – 1.06 (m, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 137.14, 127.52, 126.13, 121.14, 119.68, 118.88, 118.29, 109.01, 35.68, 32.48, 29.16, 13.17. IR (KBr): 2957.08, 2924.95, 2866.10, 1915.48, 1879.76, 1761.55, 1612.09, 1585.35, 1549.25, 1476.09, 1423.20, 1371.17, 1327.82, 1250.41, 1228.94, 1195.25, 1156.49, 1132.64, 1085.11, 1052.96, 1011.39. HRMS (ESI): m/z [M+Na]⁺ calcd for C₂₁H₂₂N₂Na : 325.1675; found: 325.1672.



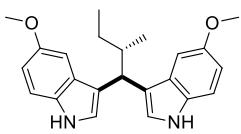
3,3'-(2-methylpropane-1,1-diyl)bis(1-methyl-1H-indole)

White solid; mp = 102–103 °C; ¹H NMR (600 MHz, CDCl₃) δ 7.64 (d, *J* = 7.8 Hz, 2H), 7.24 – 7.18 (m, 2H), 7.15 (t, *J* = 8.4 Hz, 2H), 7.03 (t, *J* = 6.6 Hz, 2H), 6.92 (s, 2H), 4.21 (d, *J* = 9.0 Hz, 1H), 3.67 (m, 6H), 2.69 – 2.51 (m, 1H), 0.97 (d, *J* = 6.6 Hz, 6H). ¹³C NMR (100 MHz, CDCl₃) δ 136.83, 127.93, 128.07, 127.78, 126.37, 121.06, 119.96, 119.7, 118.30, 108.94, 40.94, 33.24, 32.62, 21.90. IR (KBr): 2956.41, 2867.47, 1612.97, 1541.31, 1472.37, 1423.39, 1371.34, 1327.24, 1228.66, 1157.33, 1127.79, 1055.32, 1011.63. HRMS (ESI): m/z [M+Na]⁺ calcd for C₂₂H₂₄N₂Na : 339.1832; found: 339.1832.



3,3'-(3-(methylthio)propane-1,1-diyl)bis(1-methyl-1H-indole)

Yellow oil; ¹H NMR (600 MHz, CDCl₃) δ 7.62 (d, *J* = 7.8 Hz, 2H), 7.24 (d, *J* = 8.4 Hz, 2H), 7.17 (d, *J* = 8.4 Hz, 2H), 7.03 (t, *J* = 7.2 Hz, 2H), 6.85 (s, 2H), 4.65 (s, 1H), 3.66 (s, 6H), 2.55 (t, *J* = 7.2 Hz, 2H), 2.47 (t, *J* = 7.2 Hz, 2H), 2.07 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 137.13, 127.25, 126.20, 121.28, 119.56, 118.46, 118.02, 109.06, 35.30, 32.81, 32.75, 32.52, 15.46. IR (KBr): 2921.08, 1613.80, 1546.10, 1471.08, 1423.00, 1371.83, 1326.62, 1237.90, 1153.88, 1125.06, 1064.22, 1012.40. HRMS (ESI): m/z [M+Na]⁺ calcd for C₂₂H₂₄N₂S : 371.1552; found: 371.1553.



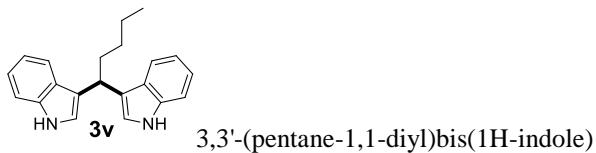
(S)-3,3'-(2-methylbutane-1,1-diyl)bis(5-methoxy-1H-indole)

Red oil; [α]_D +1.83 (c 10mg/ml, MeOH). ¹H NMR (600 MHz, CDCl₃) δ 7.69 (d, *J* = 8.4 Hz, 2H), 7.06 (m, 4H), 6.89 (s, 2H), 6.77 (d, *J* = 8.4 Hz, 2H), 4.23 (d, *J* = 7.8 Hz, 1H), 3.77 (m, 6H), 2.31 (m, 1H), 1.64 – 1.55 (m, 1H), 1.17 – 1.08 (m, 1H), 0.95 (d, *J* = 6.6 Hz, 3H), 0.90 (t, *J* = 7.2 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 153.41, 131.53, 131.40, 128.18, 127.98, 122.78, 122.56, 119.19, 118.64, 111.60, 111.16, 102.02, 55.90, 39.35, 39.14, 27.89, 17.68, 11.97. IR (KBr): 2958.22, 2872.27, 2830.29, 1623.79, 1580.76, 1482.81, 1454.38, 1376.77, 1347.05, 1291.58, 1248.39, 1209.91, 1171.95, 1095.93, 1027.95. MS: m/z [M]⁺ calcd for C₂₃H₂₆N₂O₂ : 362.20; found: 362.37.

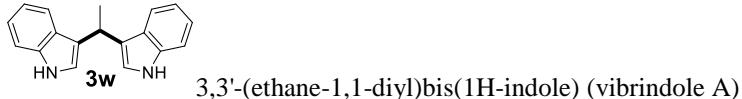


3,3'-(3-methylbutane-1,1-diyl)bis(1H-indole)

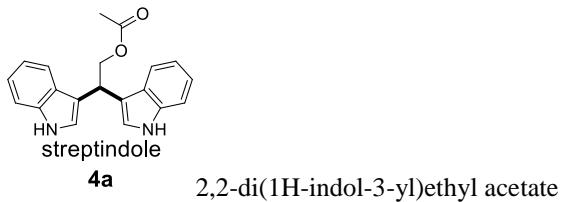
Yellow oil; ^1H NMR (400 MHz, CDCl_3) δ 7.59 (d, $J = 8.0$ Hz, 2H), 7.43 (s, 2H), 7.09 (s, 1H), 7.08 (s, 1H), 7.02 (m, 2H), 6.65 (d, $J = 2.0$ Hz, 2H), 4.53 (t, $J = 7.6$ Hz, 1H), 2.02 (t, $J = 7.2$ Hz, 2H), 1.65 – 1.49 (m, 1H), 0.93 (d, $J = 6.8$ Hz, 6H). ^{13}C NMR (100 MHz, CDCl_3) δ 136.34, 126.88, 121.50, 121.46, 120.09, 119.40, 118.82, 111.14, 77.32, 77.00, 76.68, 45.11, 31.46, 25.80, 22.77. IR (KBr): 2953.91, 2865.73, 1618.06, 1456.93, 1419.00, 1338.93, 1220.38, 1094.36, 1010.92. HRMS (ESI): m/z [M+Na]⁺ calcd for $\text{C}_{21}\text{H}_{22}\text{N}_2\text{Na}$: 325.1675; found: 325.1675.



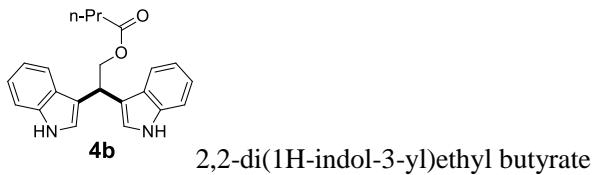
Red oil; ^1H NMR (600 MHz, CDCl_3) δ 7.57 (d, $J = 7.8$ Hz, 2H), 7.50 (s, 2H), 7.15 (d, $J = 7.8$ Hz, 2H), 7.10 (t, $J = 7.8$ Hz, 2H), 7.01 (t, $J = 7.2$ Hz, 2H), 6.75 (s, 2H), 4.41 (t, $J = 7.2$ Hz, 1H), 2.16 (d, $J = 6.6$ Hz, 2H), 1.35 (s, 4H), 0.84 (d, $J = 6.6$ Hz, 3H). ^{13}C NMR (100 MHz, CDCl_3) δ 136.43, 127.05, 121.57, 121.42, 120.34, 119.55, 118.88, 111.08, 35.57, 33.86, 30.47, 22.77, 14.08. IR (KBr): 2954.67, 2926.62, 2857.79, 1617.55, 1521.63, 1486.14, 1455.68, 1418.68, 1337.90, 1219.63, 1151.37, 1093.40, 1010.27. MS: m/z [M]⁺ calcd for $\text{C}_{21}\text{H}_{22}\text{N}_2$: 302.18; found: 302.34.



White solid; mp = 153–155 °C; ^1H NMR (600 MHz, CDCl_3) δ 7.75 (br, 2H), 7.63 (d, $J = 7.8$ Hz, 2H), 7.34 (d, $J = 8.4$ Hz, 2H), 7.22 (t, $J = 7.8$ Hz, 2H), 7.10 (t, $J = 7.2$ Hz, 2H), 6.86 (s, 2H), 4.71 (d, $J = 6.6$ Hz, 1H), 1.84 (d, $J = 7.2$ Hz, 3H). ^{13}C NMR (100 MHz, CDCl_3) δ 136.56, 126.83, 121.69, 121.53, 121.19, 119.66, 118.94, 111.05, 28.11, 21.69. IR (KBr): 2958.13, 2866.18, 2836.91, 1624.23, 1548.86, 1484.13, 1454.83, 1421.09, 1337.75, 1291.37, 1220.61, 1124.66, 1095.80, 1013.70. HRMS (ESI): m/z [M+Na]⁺ calcd for $\text{C}_{18}\text{H}_{16}\text{N}_2\text{Na}$: 283.1206; found: 283.1209.

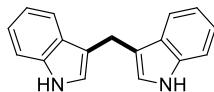


Yellow oil; ^1H NMR (600 MHz, CDCl_3) δ 7.80 (br, 2H), 7.57 (d, $J = 7.8$ Hz, 2H), 7.18 (d, $J = 7.8$ Hz, 2H), 7.11 (t, $J = 7.8$ Hz, 2H), 7.01 (t, $J = 7.2$ Hz, 2H), 6.70 (s, 2H), 4.88 (t, $J = 7.2$ Hz, 1H), 4.67 (d, $J = 6.6$ Hz, 2H), 1.89 (s, 3H). ^{13}C NMR (100 MHz, CDCl_3) δ 171.41, 136.34, 126.90, 122.14, 121.93, 119.40, 116.07, 111.16, 67.42, 33.45, 21.03. IR (KBr): 2924.62, 2853.71, 1721.32, 1631.42, 1457.13, 1422.17, 1384.17, 1263.84, 1097.32, 1038.64. HRMS (ESI): m/z [M+Na]⁺ calcd for $\text{C}_{20}\text{H}_{18}\text{N}_2\text{NaO}$: 341.1260; found: 341.1261.



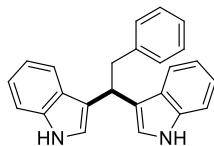
Yellow oil; ^1H NMR (600 MHz, CDCl_3) δ 7.84 (br, 2H), 7.59 (d, $J = 7.8$ Hz, 2H), 7.22 (m, 2H), 7.13 (m, 2H), 7.03 (m, 2H), 6.76 (s, 2H), 4.90 (t, $J = 7.2$ Hz, 1H), 4.70 (d, $J = 7.2$ Hz, 2H), 2.17 (t, $J = 7.2$ Hz, 2H), 1.51 (m, 2H), 0.79 (t, $J = 7.2$ Hz, 3H). ^{13}C NMR (100 MHz, CDCl_3) δ 174.00, 136.35, 126.93,

122.15, 121.89, 119.42, 119.22, 116.11, 111.12, 77.32, 77.00, 76.68, 67.12, 36.17, 33.50, 18.28, 13.50. IR (KBr): 2962.23, 2928.69, 2872.81, 1714.79, 1456.92, 1421.18, 1385.17, 1341.43, 1266.21, 1184.96, 1096.08, 1008.02. HRMS (ESI): m/z [M+Na]⁺ calcd for C₂₂H₂₂N₂NaO₂ : 369.1573; found: 369.1577.



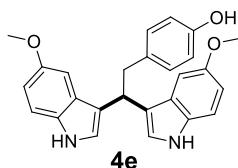
4c di(1H-indol-3-yl)methane

Yellow oil; ¹H NMR (600 MHz, dmso-*d*₆) δ 10.83 (br, 2H), 7.63 (d, *J* = 7.8 Hz, 2H), 7.43 (d, *J* = 8.4 Hz, 2H), 7.22 (s, 2H), 7.12 (m, 2H), 7.01 (m, 2H), 4.24 (s, 2H). ¹³C NMR (150 MHz, dmso-*d*₆) δ 136.54, 136.38, 127.34, 122.87, 122.71, 120.91, 118.83, 118.20, 114.37, 114.33, 111.45, 21.10. IR (KBr): 2921.85, 1620.61, 1488.27, 1455.74, 1422.61, 1341.12, 1221.66, 1091.03, 1006.56. HRMS (ESI): m/z [M+Na]⁺ calcd for C₁₇H₁₄N₂Na : 269.1049; found: 269.1047.



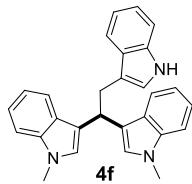
4d 3,3'-(2-phenylethane-1,1-diyl)bis(1H-indole)

White solid; mp = 76-78°C; ¹H NMR (600 MHz, CDCl₃) δ 7.58 (d, *J* = 7.8 Hz, 2H), 7.15 – 7.08 (m, 10H), 7.05 (m, 3H), 6.61 (m, 2H), 4.76 (t, *J* = 7.8 Hz, 1H), 3.50 (d, *J* = 7.2 Hz, 2H). ¹³C NMR (150 MHz, CDCl₃) δ 141.17, 136.26, 128.85, 127.89, 126.68, 125.63, 121.99, 121.54, 119.46, 118.91, 118.88, 111.11, 41.57, 36.02. IR (KBr): 3416, 3054, 1631, 1492, 1455, 1417, 1340, 1244, 1094, 1011. MS: m/z [M]⁺ calcd for C₂₄H₂₀N₂ : 336.16; found: 336.22.



4e 4-(2,2-bis(5-methoxy-1H-indol-3-yl)ethyl)phenol

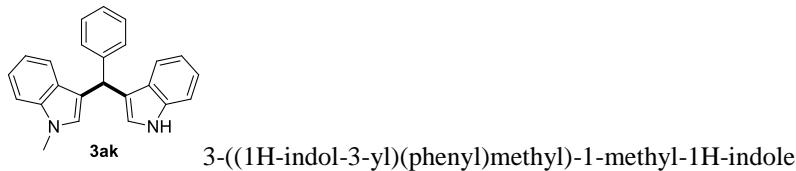
Yellow oil; ¹H NMR (600 MHz, dmso-*d*₆) δ 10.54 (br, 2H), 9.03 (s, 1H), 7.17 (m, 4H), 6.97 (m, 4H), 6.65 (m, 2H), 6.54 (d, *J* = 7.8 Hz, 2H), 4.56 (t, *J* = 7.8 Hz, 1H), 3.67 (s, 6H), 3.35 (d, *J* = 7.8 Hz, 2H). ¹³C NMR (75 MHz, CDCl₃) δ 153.60, 153.46, 133.47, 131.78, 130.12, 127.43, 122.79, 119.15, 114.80, 111.64, 111.55, 101.99, 55.90, 40.71, 36.53. IR (KBr): 3413.36, 2933.77, 2831.05, 1619.97, 1582.11, 1513.25, 1482.86, 1446.52, 1350.50, 1290.68, 1211.98, 1170.18, 1097.02, 1058.26, 1025.49. HRMS (ESI): m/z [M+Na]⁺ calcd for C₂₆H₂₄N₂O₃Na: 435.1679; found: 435.1687.



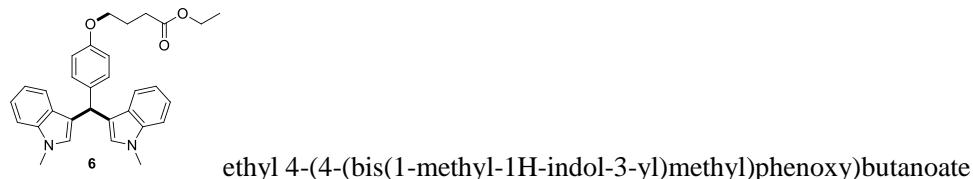
4f 3,3'-(2-(1H-indol-3-yl)ethane-1,1-diyl)bis(1-methyl-1H-indole)

¹H NMR (600 MHz, acetone-*d*₆) δ 9.75 (br, 1H), 7.65 (d, *J* = 7.8 Hz, 1H), 7.63 (d, *J* = 8.4 Hz, 2H), 7.31 (d, *J* = 7.8 Hz, 1H), 7.28 (d, *J* = 8.4 Hz, 2H), 7.10 (m, 3H), 7.09 – 7.04 (m, 2H), 7.01 (m, 1H), 6.94 (m, 2H), 6.90 (s, 1H), 5.00 (t, *J* = 7.2 Hz, 1H), 3.85 – 3.55 (m, 8H). ¹³C NMR (100 MHz, CDCl₃) δ 137.23, 135.88, 127.88, 127.50, 126.62, 122.14, 121.60, 121.22, 121.21, 119.76, 119.03, 118.88,

118.77, 118.45, 115.37, 110.91, 109.06, 34.27, 32.60, 31.92. IR (KBr): 3050.19, 2927.13, 1616.00, 1547.46, 1475.55, 1421.04, 1372.09, 1328.48, 1244.95, 1154.10, 1125.57, 1012.44. HRMS (ESI): m/z [M+Na]⁺ calcd for C₂₈H₂₅N₂Na: 426.1941; found: 426.1952.



Red oil; ¹H NMR (600 MHz, CDCl₃) δ 7.63 (br, 1H), 7.36 (m, 2H), 7.31 (m, 2H), 7.27 – 7.08 (m, 7H), 6.96 (m, 2H), 6.51 (s, 1H), 6.45 (s, 1H), 5.85 (s, 1H), 3.55 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 144.15, 137.24, 136.36, 128.55, 128.08, 127.31, 126.81, 125.96, 123.50, 121.64, 121.34, 119.89, 119.66, 119.25, 119.00, 118.57, 117.90, 110.99, 109.05, 40.00, 32.19. IR (KBr): 2929.97, 1609.63, 1546.59, 1455.04, 1418.74, 1369.65, 1330.07, 1221.13, 1153.63, 1123.54, 1091.74, 1048.21, 1010.29. HRMS (ESI): m/z [M+Na]⁺ calcd for C₂₄H₂₀N₂Na: 359.1519; found: 359.1518.



Red oil; ¹H NMR (600 MHz, CDCl₃) δ 7.37 (d, *J* = 7.8 Hz, 2H), 7.28 (d, *J* = 8.4 Hz, 2H), 7.22 (m, 2H), 7.19 (t, *J* = 7.2 Hz, 2H), 6.98 (t, *J* = 7.2 Hz, 2H), 6.79 (d, *J* = 8.4 Hz, 2H), 6.51 (s, 2H), 5.82 (s, 1H), 4.13 (m, 2H), 3.96 (t, *J* = 6.6 Hz, 2H), 3.66 (s, 6H), 2.50 (t, *J* = 7.2 Hz, 2H), 2.083 (m, 2H), 1.25 (t, *J* = 7.2 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 173.16, 157.02, 137.32, 136.64, 129.44, 128.07, 127.35, 121.28, 119.96, 118.48, 114.00, 108.94, 100.76, 39.15, 32.50, 30.80, 24.67, 14.16. IR (KBr): 2933.26, 1731.78, 1609.57, 1507.87, 1470.47, 1422.45, 1371.27, 1327.34, 1244.44, 1175.10, 1121.39, 1046.63. HRMS (ESI): m/z [M+Na]⁺ calcd for C₃₁H₃₂N₂NaO: 503.2305; found: 503.2308.

4. 3,3'-(phenylmethylene)bis(1-methyl-1H-indole)

X-ray structure determination was obtained via slow evaporation of compound 3i in CHCl₃/MeOH (10:1) at room temperature.

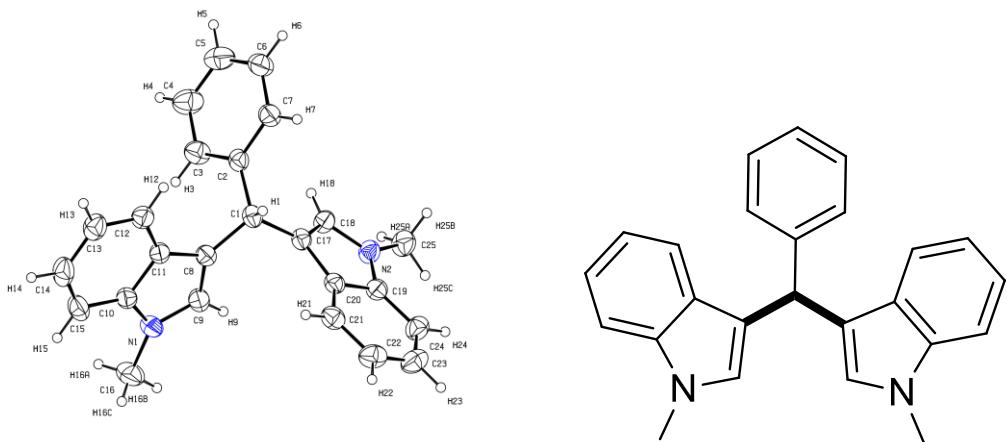
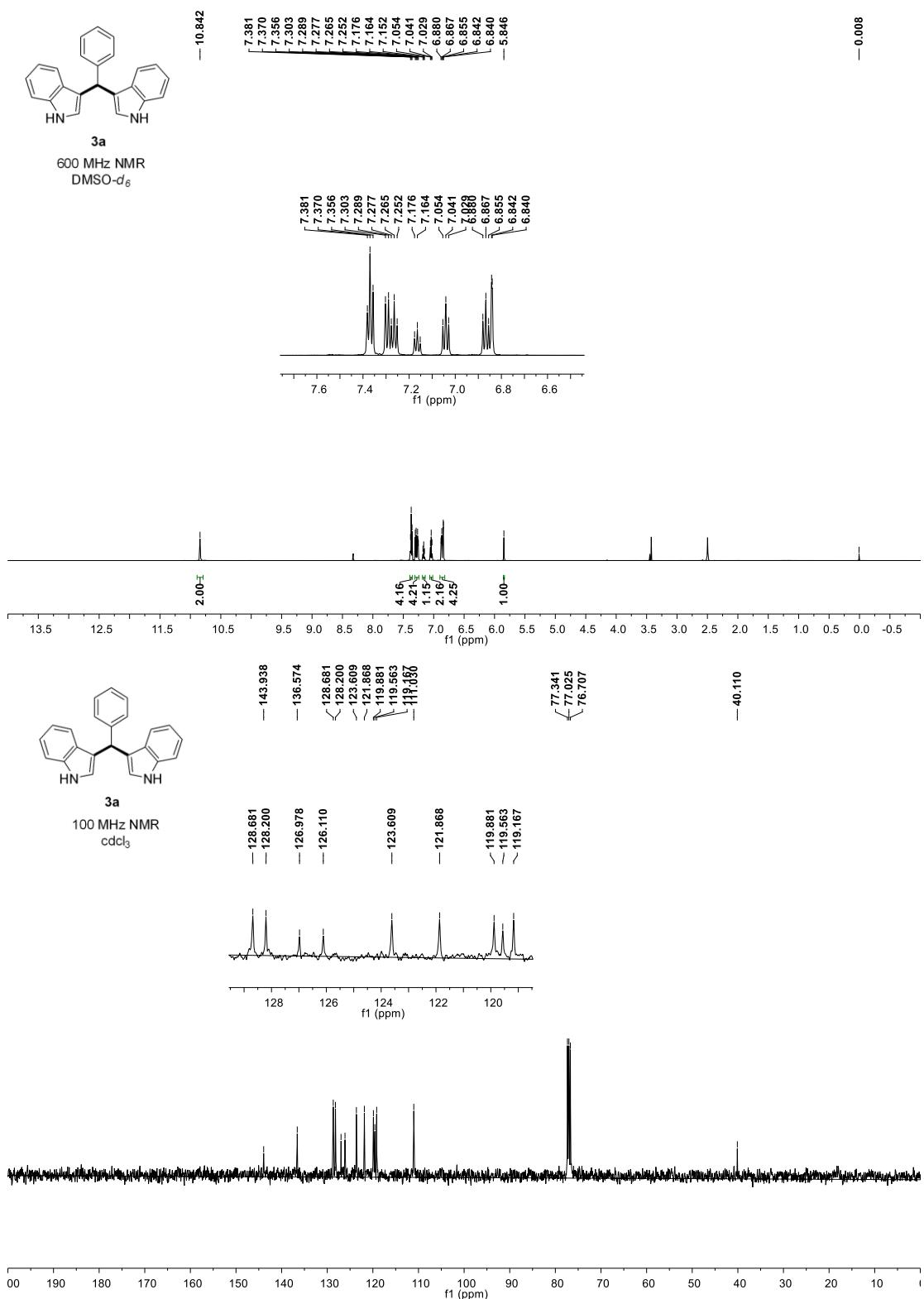


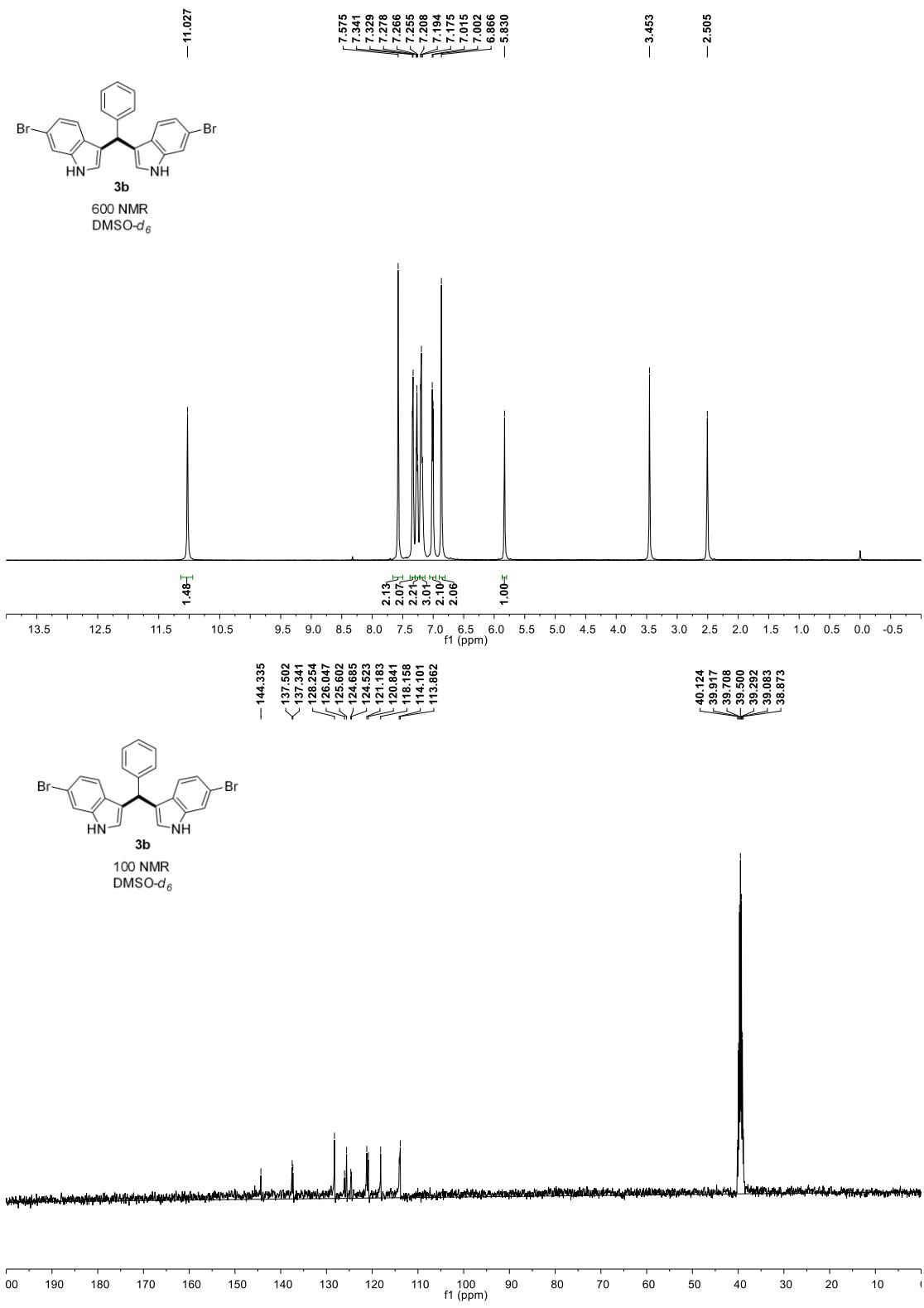
Figure S1 X-ray crystal structure of **3i**; ellipsoids depicted at the 30% probability level.

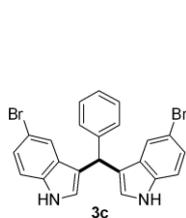
Table S1. Crystal data and structure refinement for compound **3i** (CCDC: 1030461)

Empirical formula	C ₂₅ H ₂₂ N ₂		Absorption coefficient	0.070 mm ⁻¹
Formula weight	350.45		F(000)	744
Temperature	298(2)		Crystal size	0.15 x 0.12 x 0.10 mm ³
Wavelength	0.71073		Reflections collected	12169
Crystal system	Monoclinic		Independent reflections	3388 [R(int) = 0.0446]
Space group	P 21/c		Max. and min. transmission	0.992 and 0.993
Unit cell dimensions	a = 9.4408(17) Å	α= 90.00(4) °	Refinement method	Full-matrix least-squares on F ²
	b = 25.506(5) Å	β= 95.641(4) °	Goodness-of-fit on F ²	1.014
	c = 8.0660(15) Å	γ = 90.00(4) °	Final R indices [I>2sigma(I)]	R1 = 0.0476, wR2 = 0.1148
Volume	558.7(2) Å ³		R indices (all data)	R1 = 0.0871, wR2 = 0.1367
Z	4		Largest diff. peak and hole	0.180 and -0.162e. ⁻³
Density (calculated)	1.204 Mg/m ³			

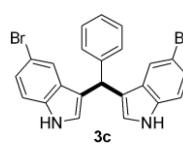
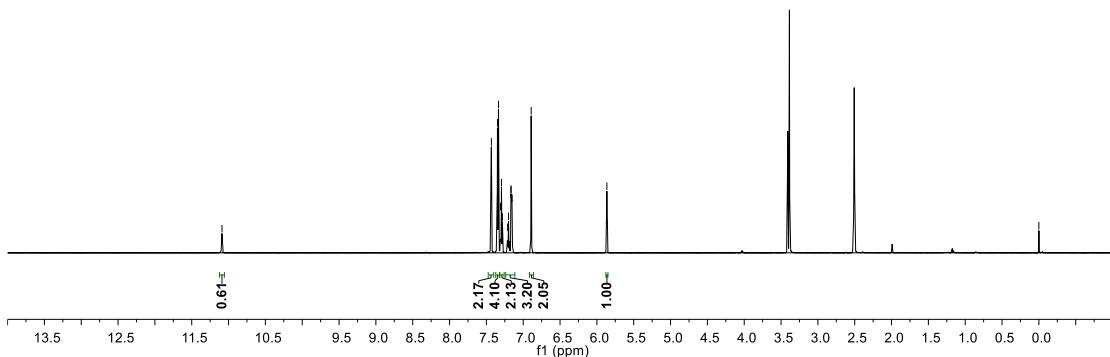
5. Copies of ^1H NMR, ^{13}C NMR and HRMS/MS Spectra



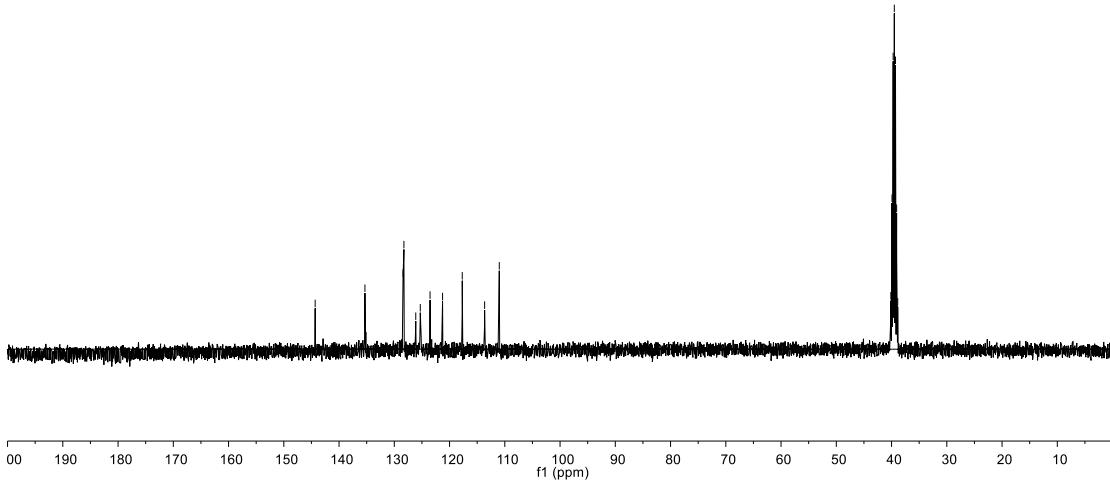


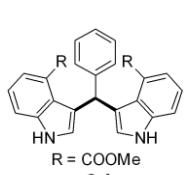


600 NMR
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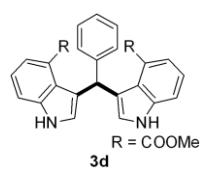
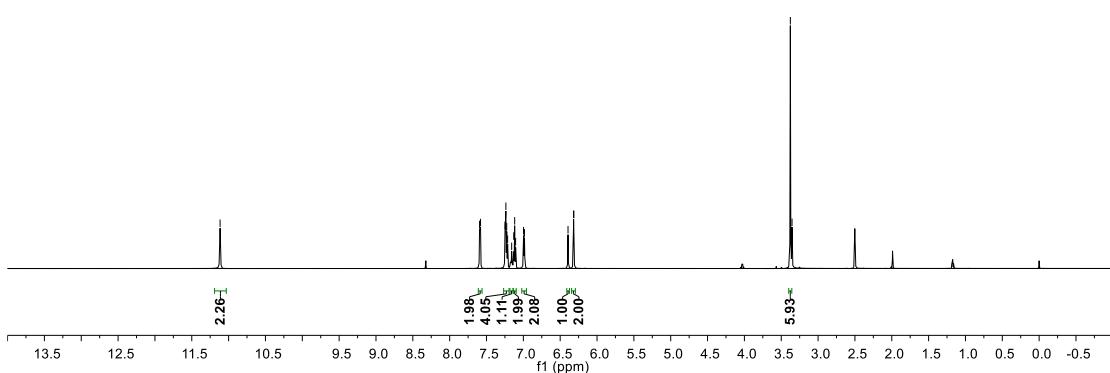


100 NMR
DMSO-*d*₆

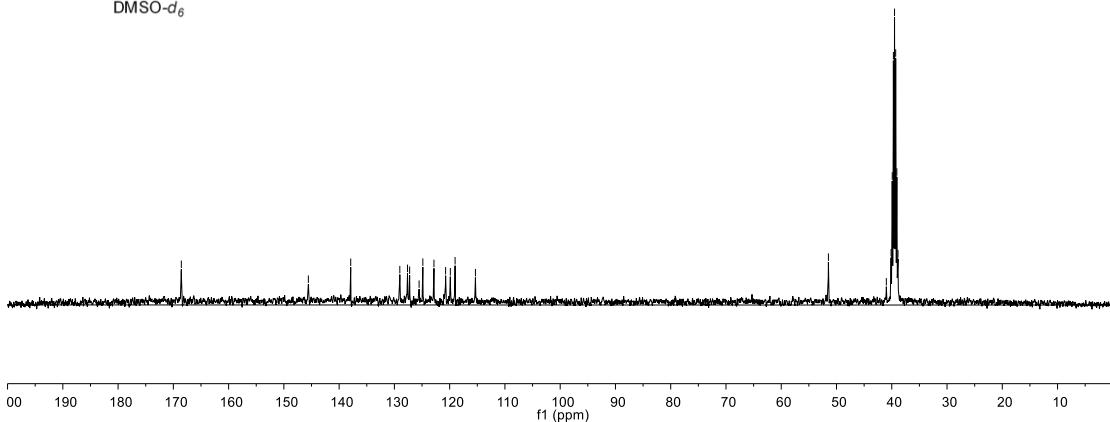


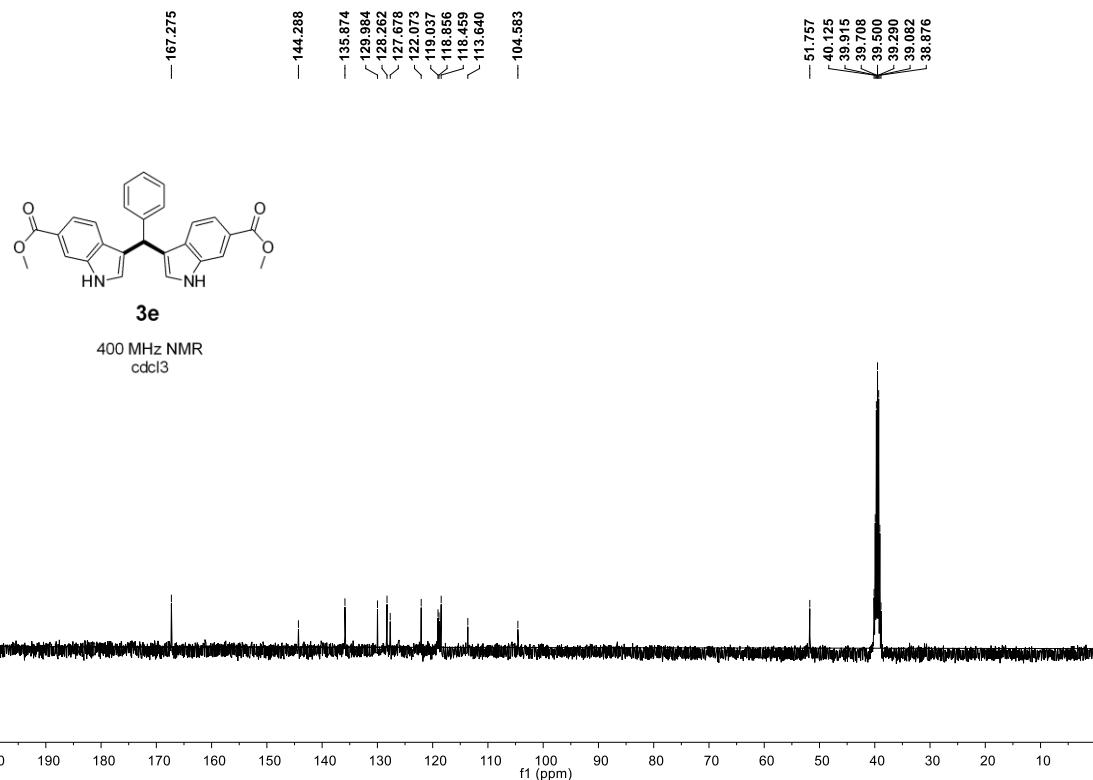
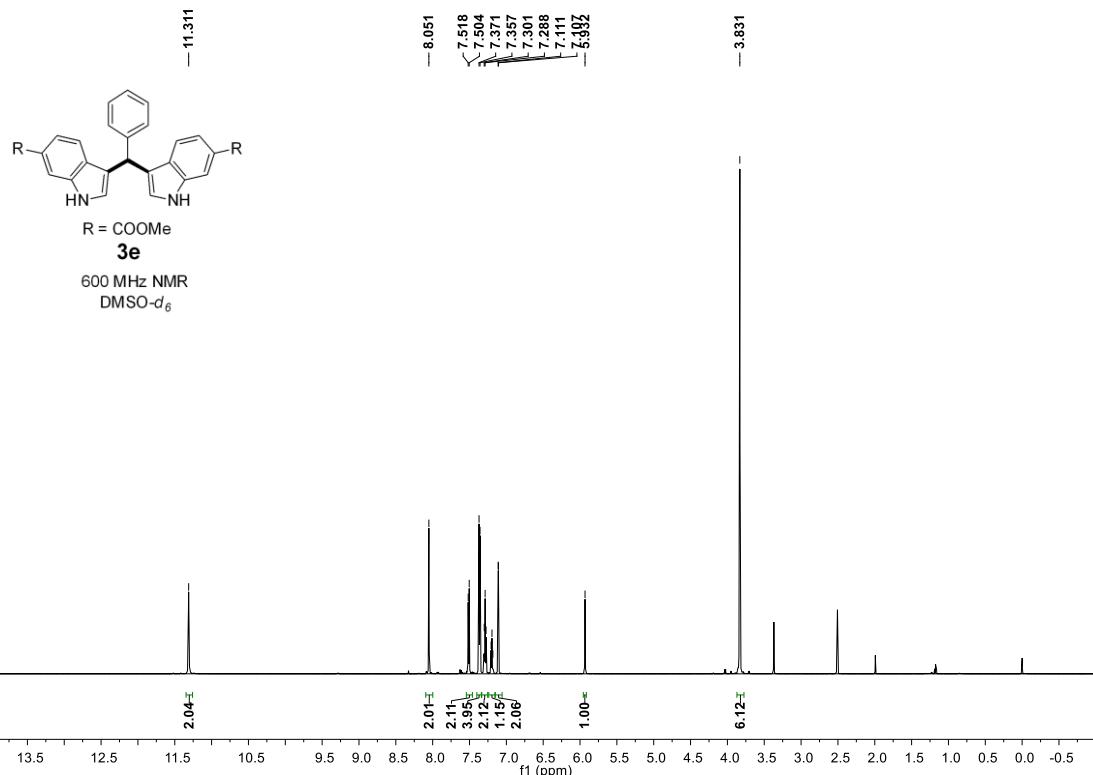


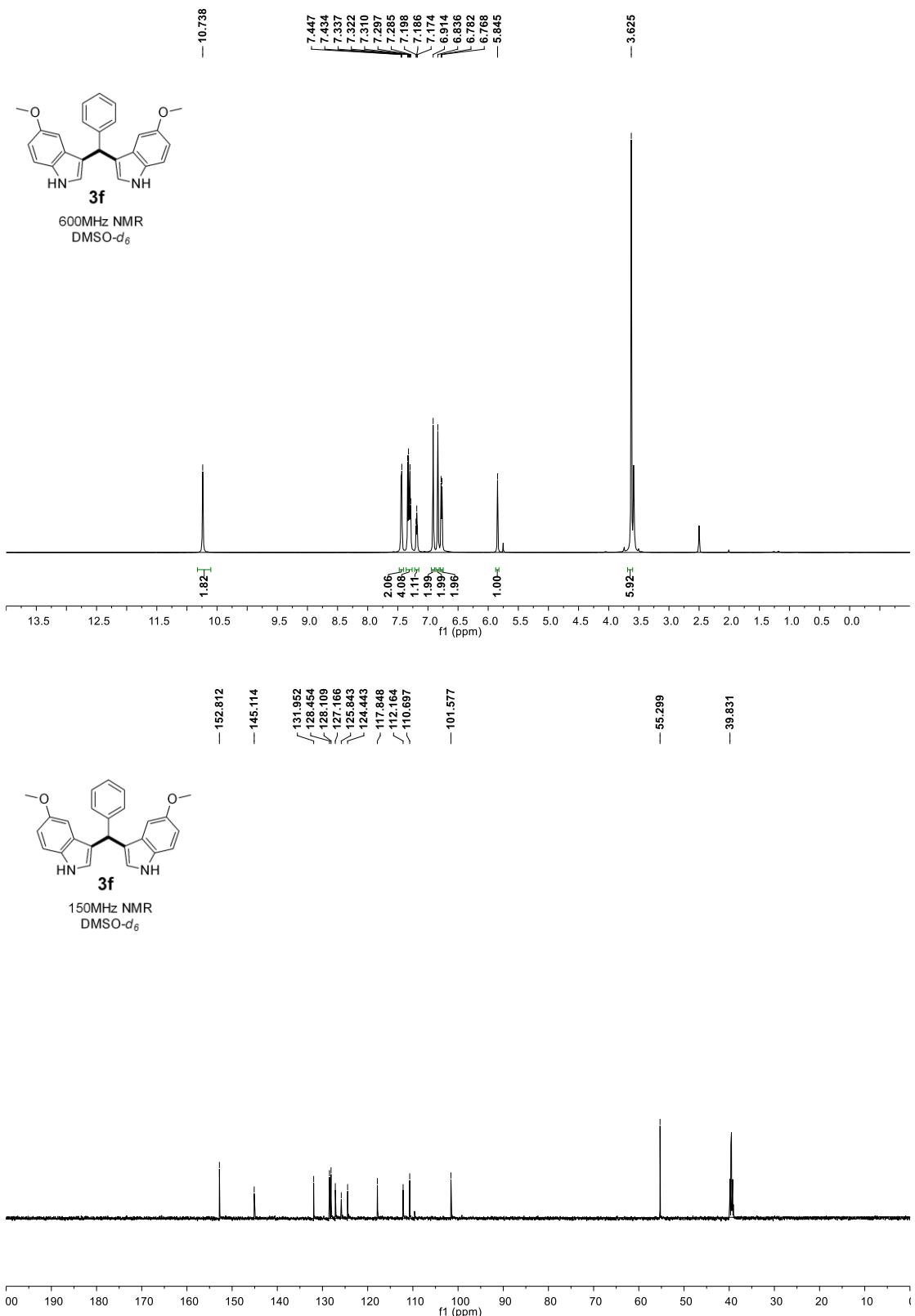
600 NMR
DMSO-*d*₆

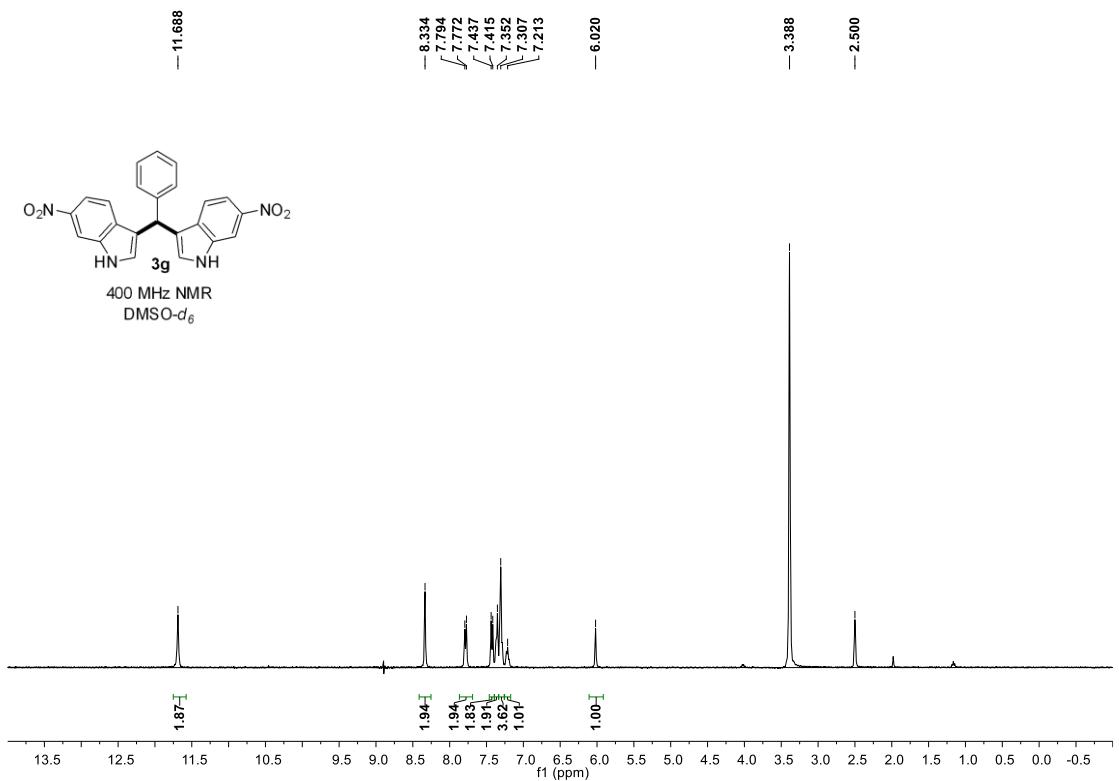


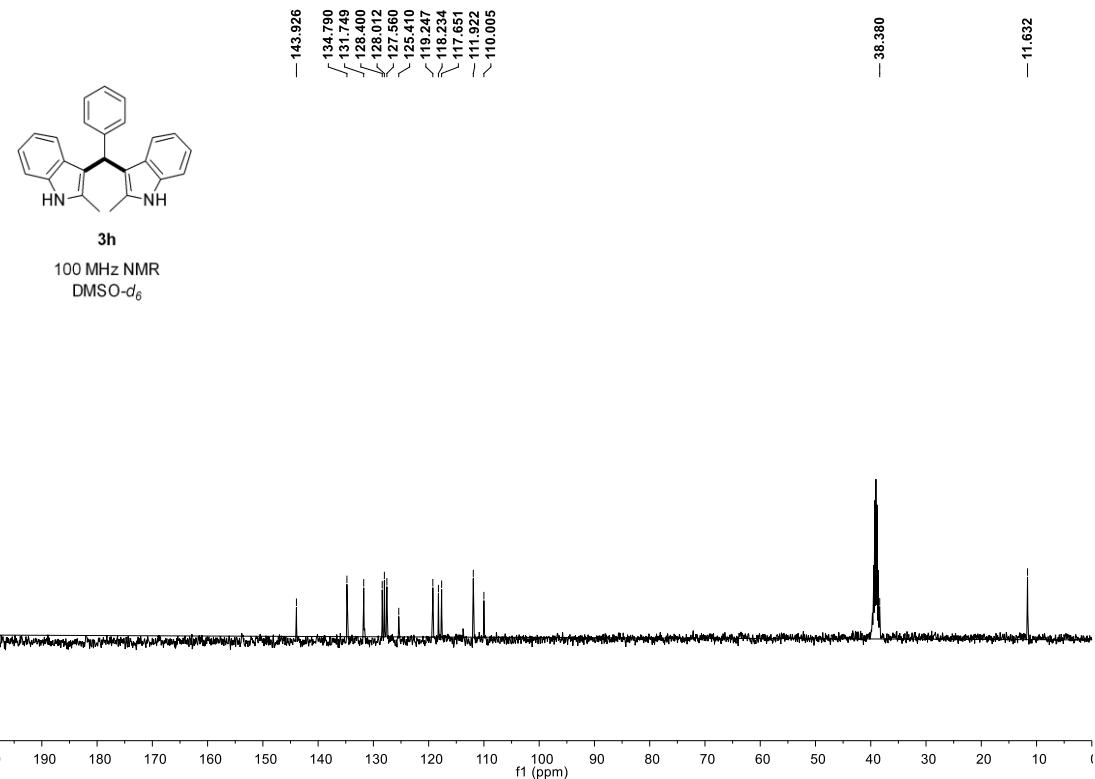
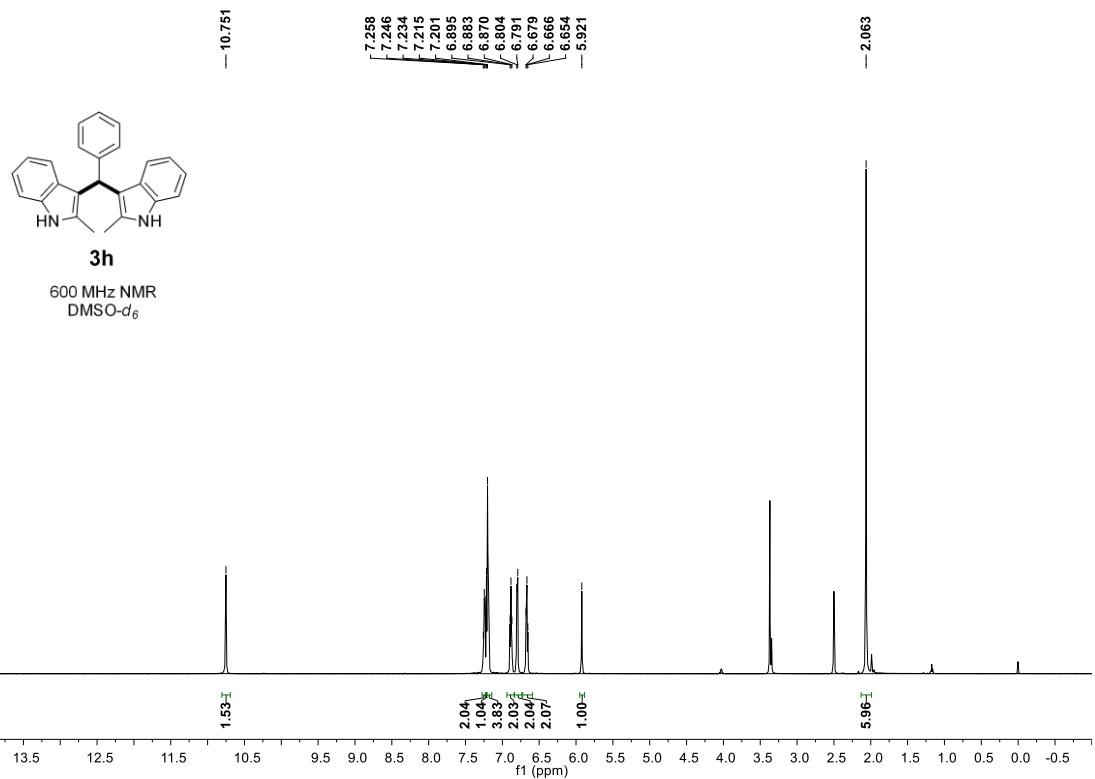
100 NMR
DMSO-*d*₆

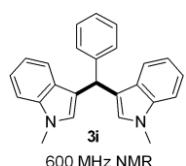




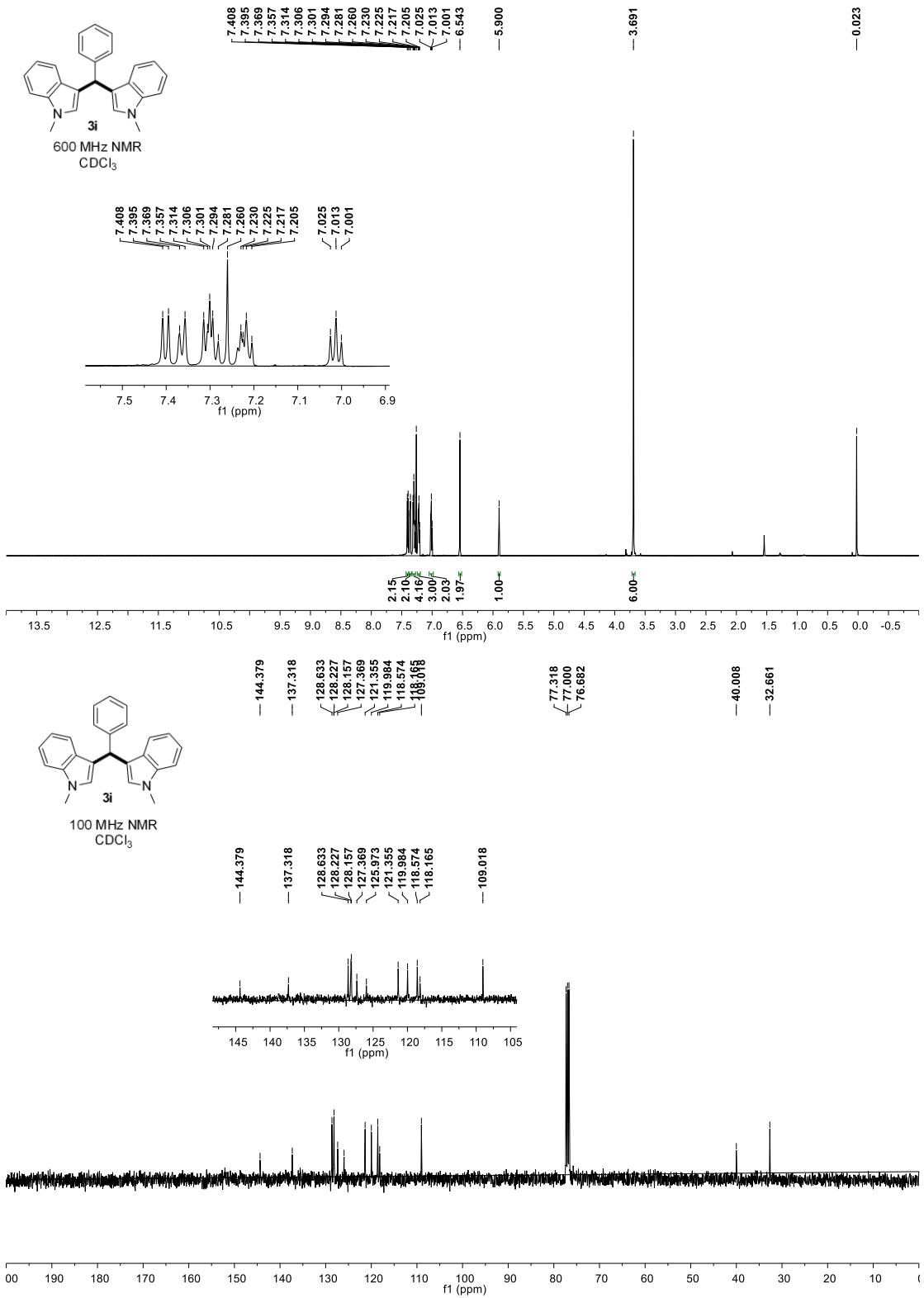


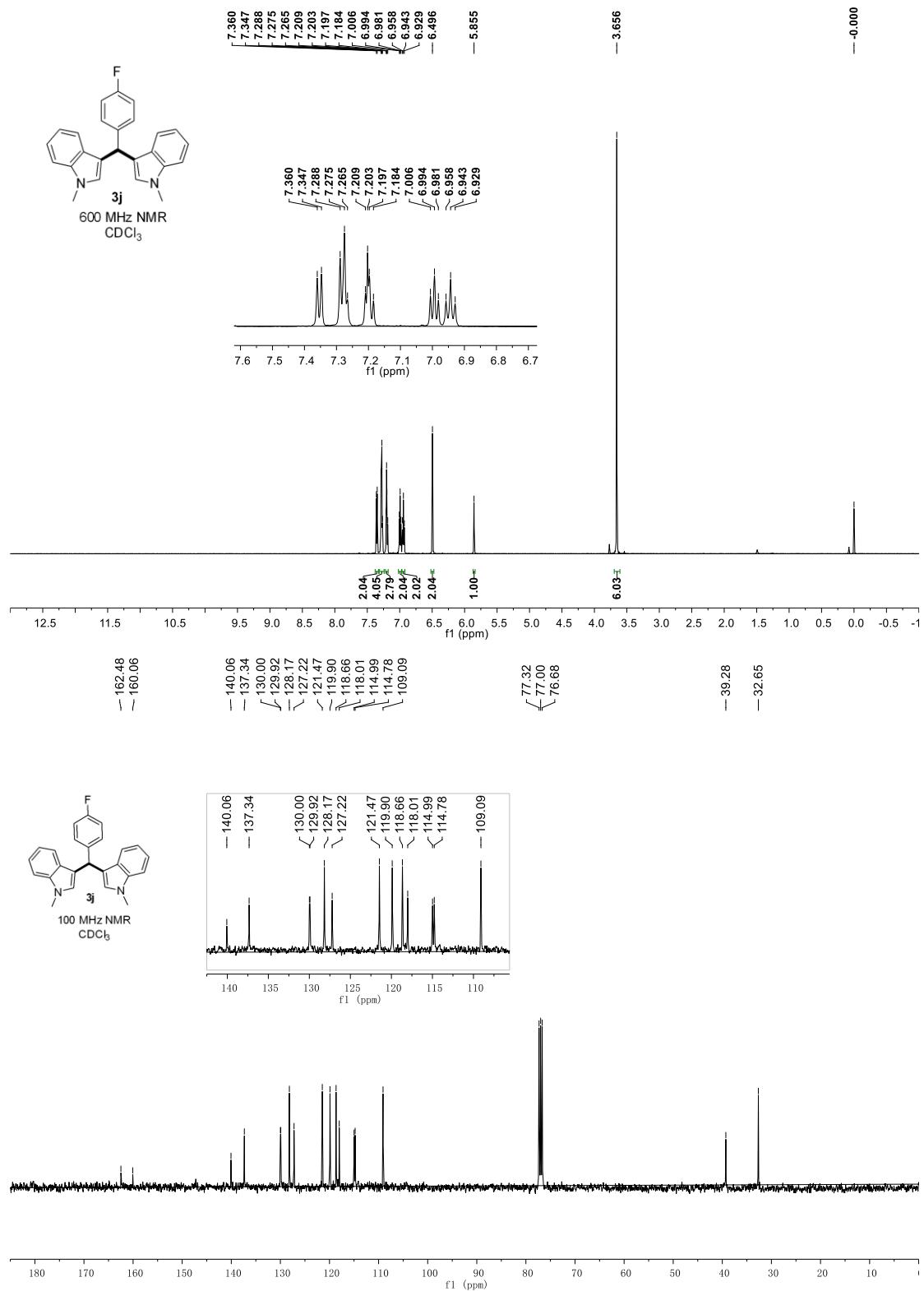


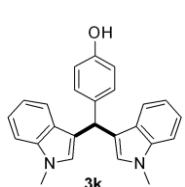




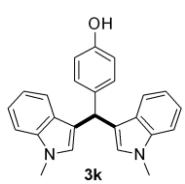
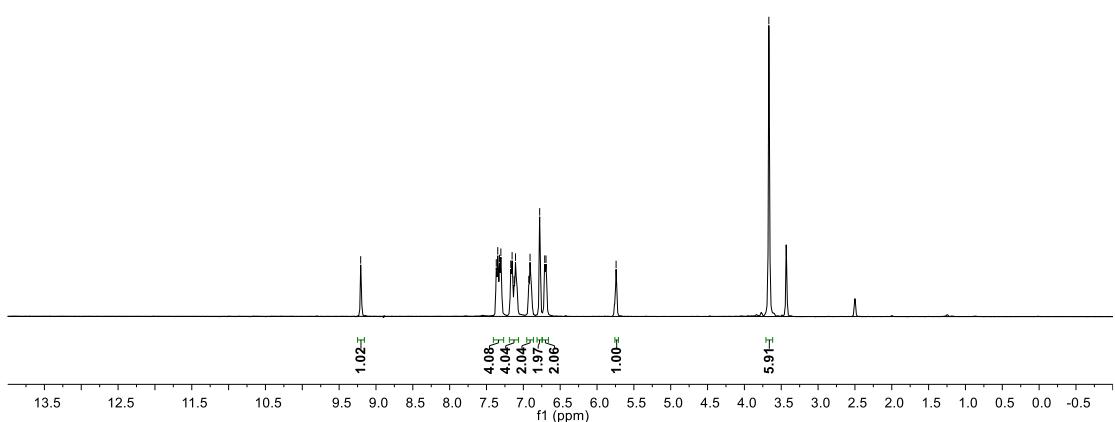
600 MHz NMR
 CDCl_3



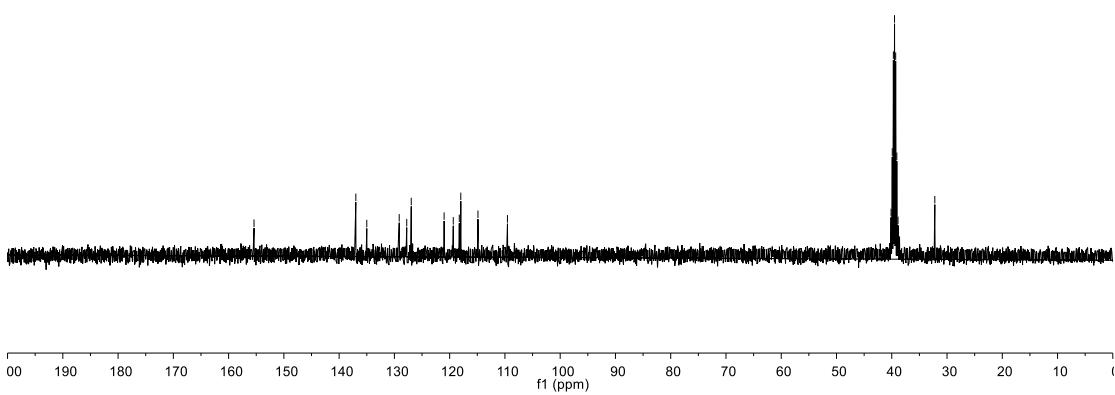


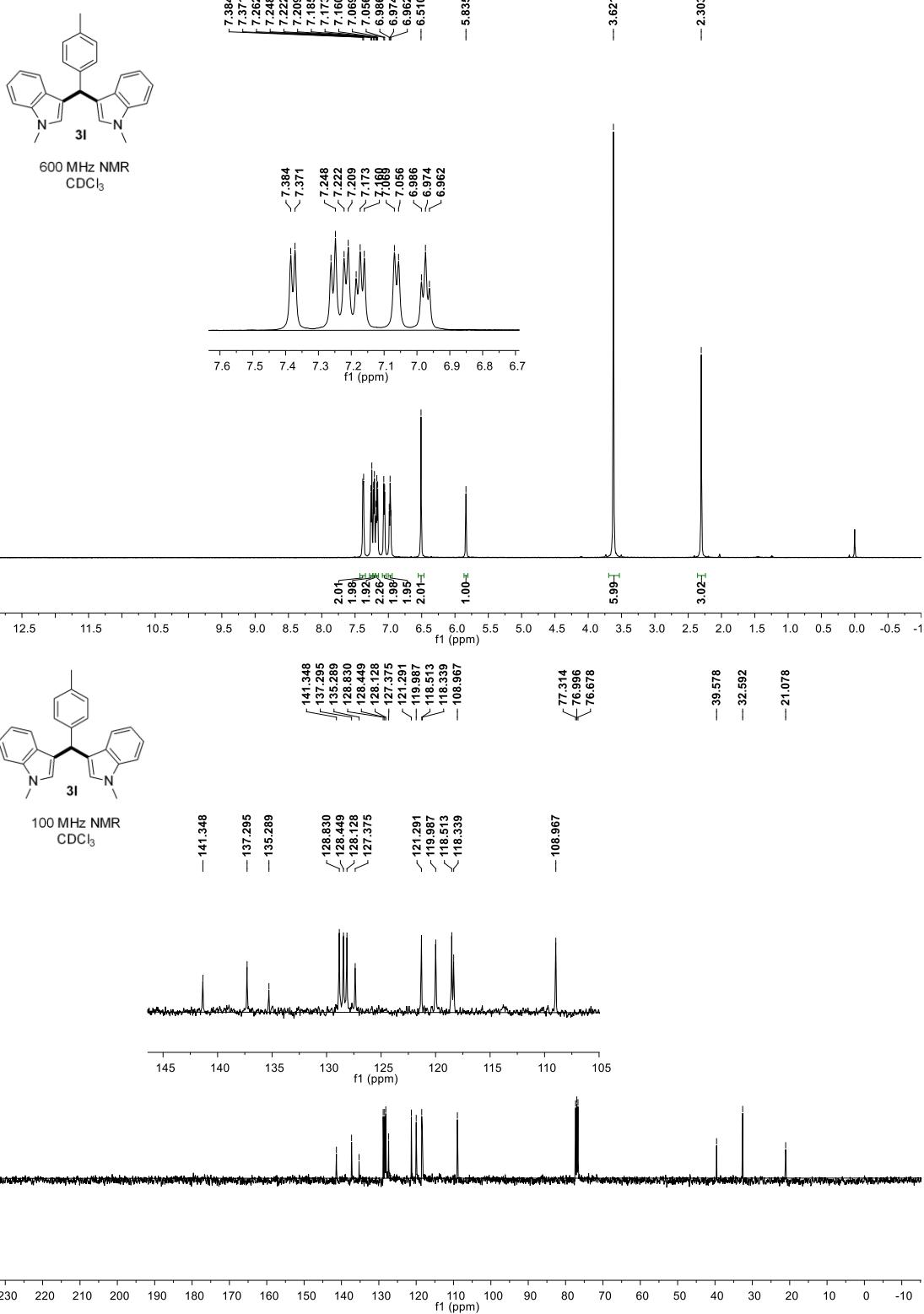


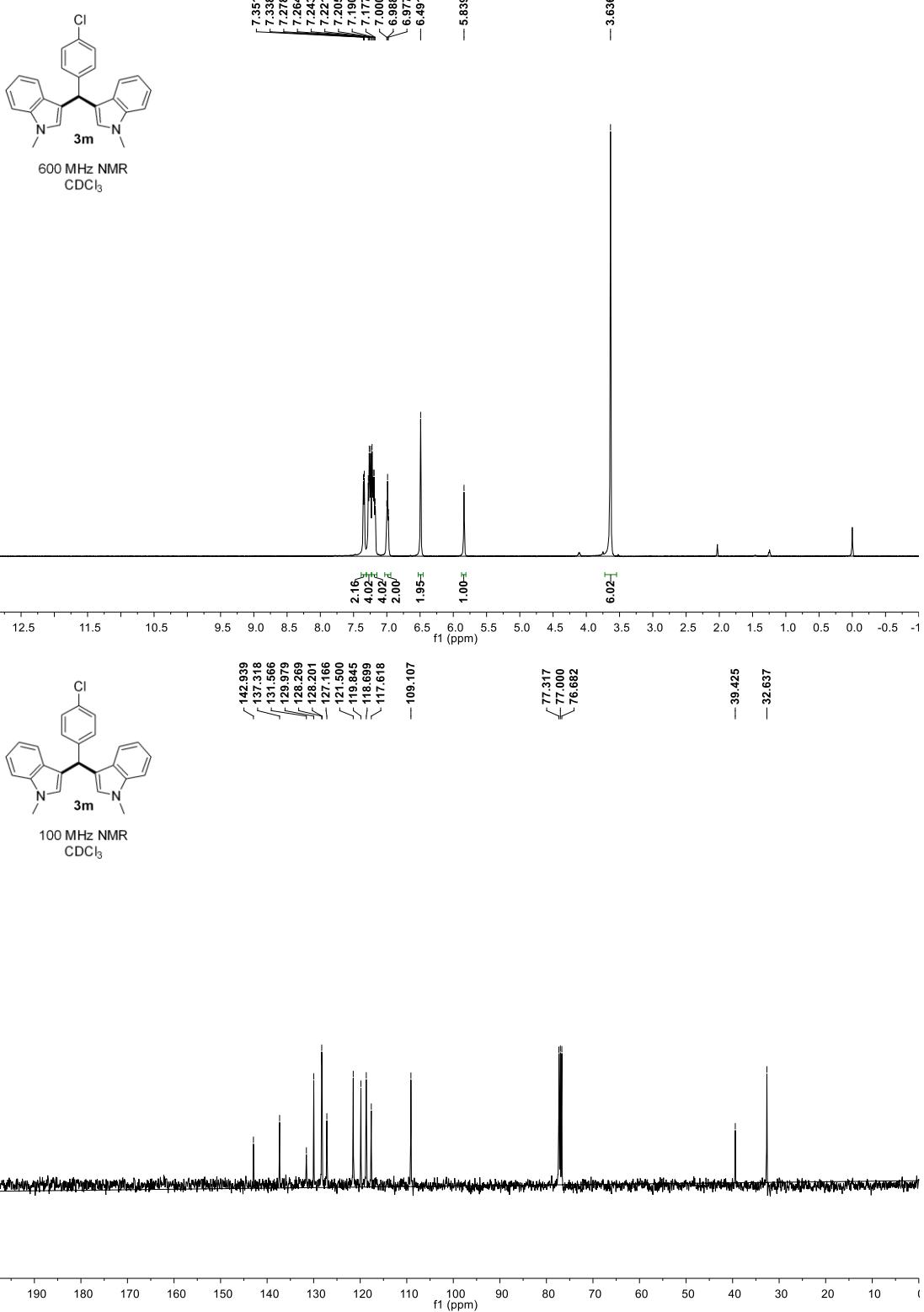
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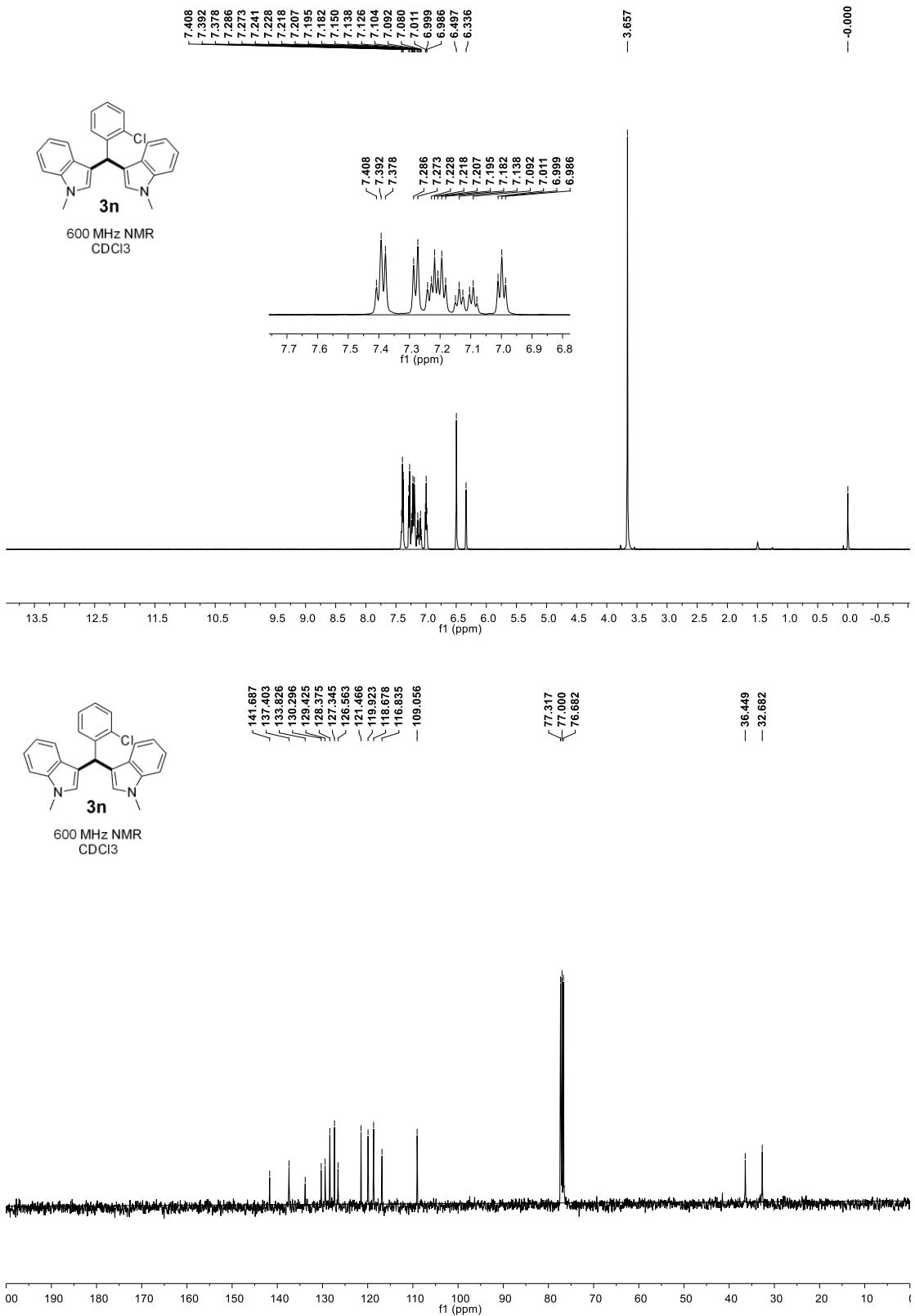


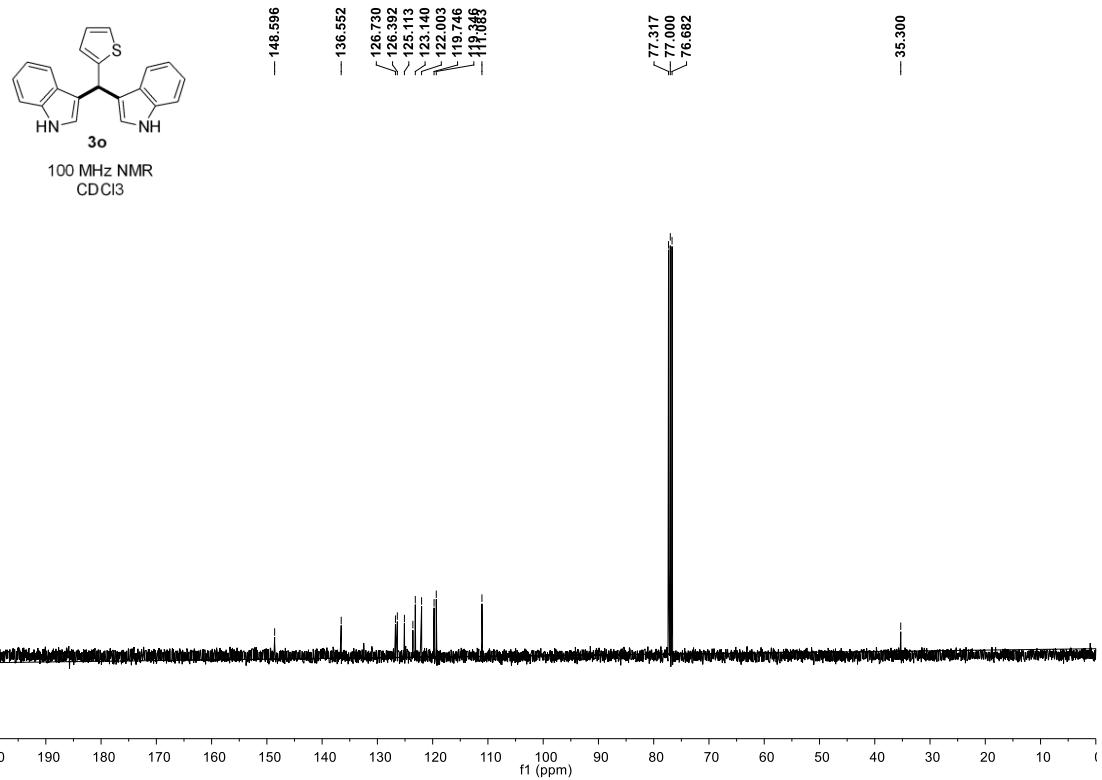
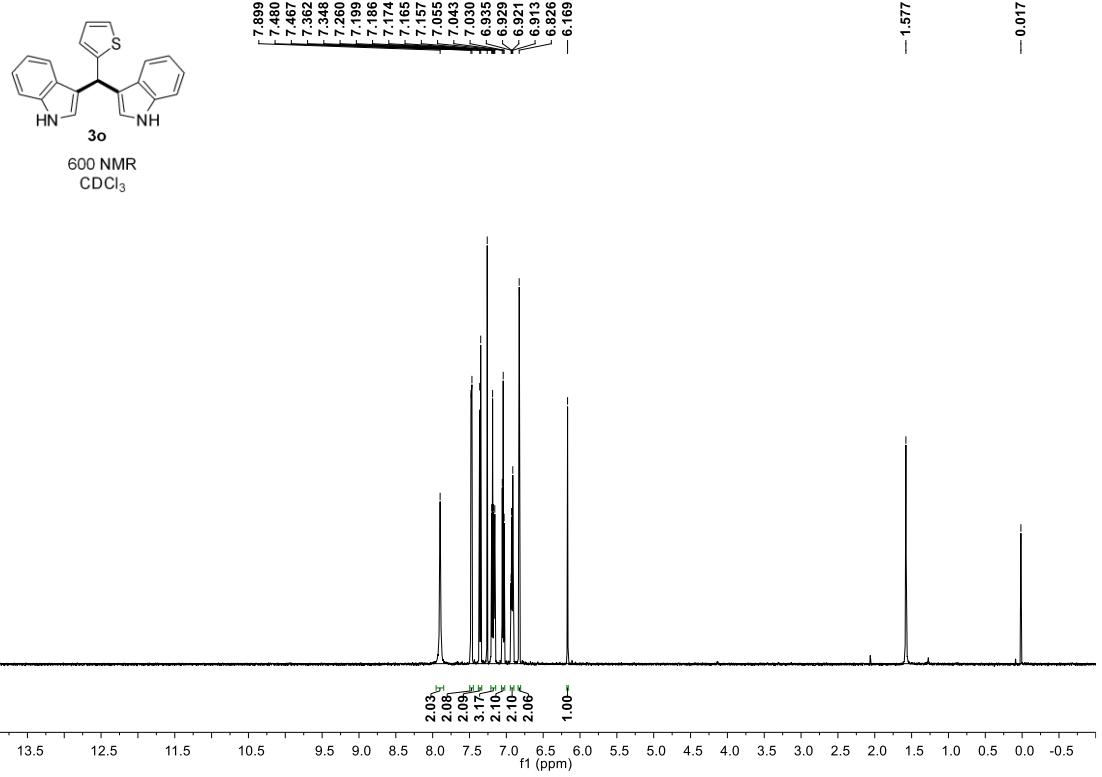
100 MHz NMR
DMSO-*d*₆

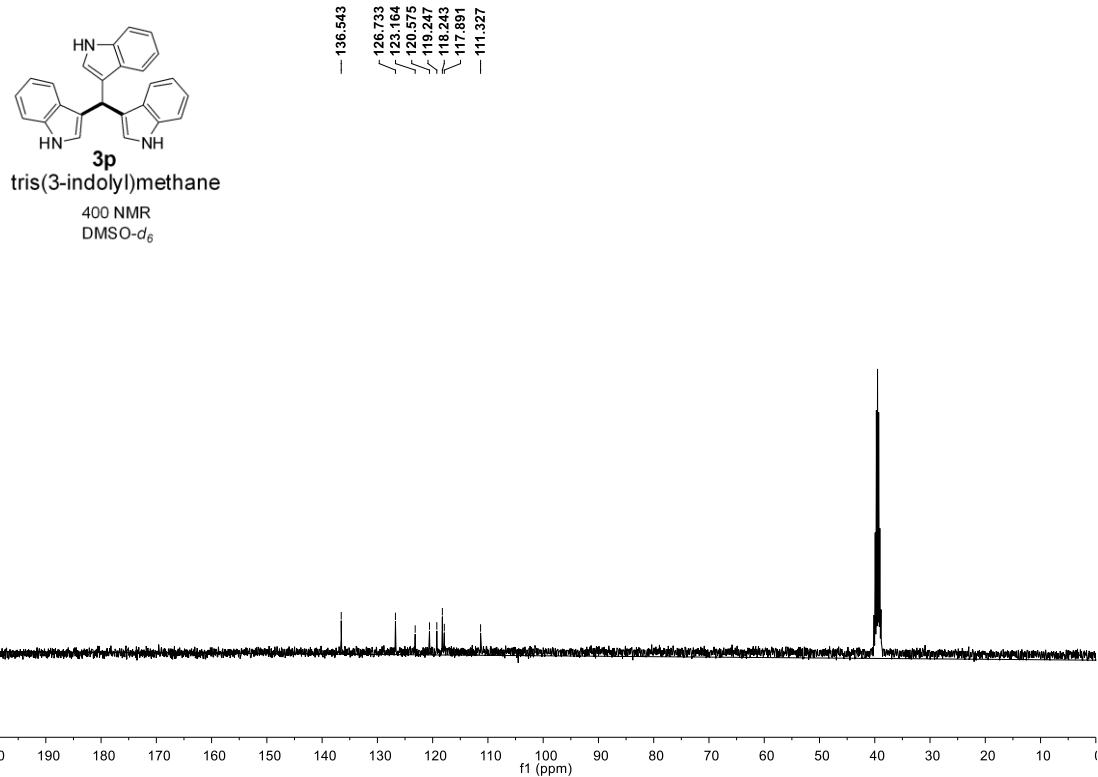
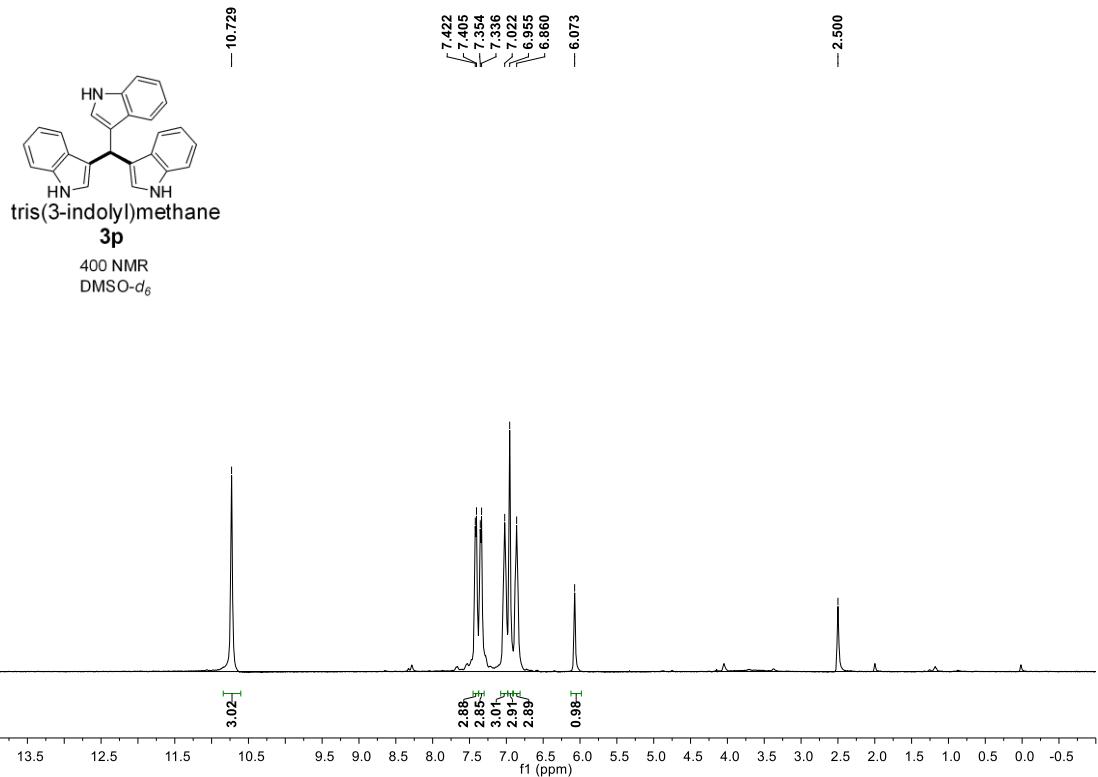


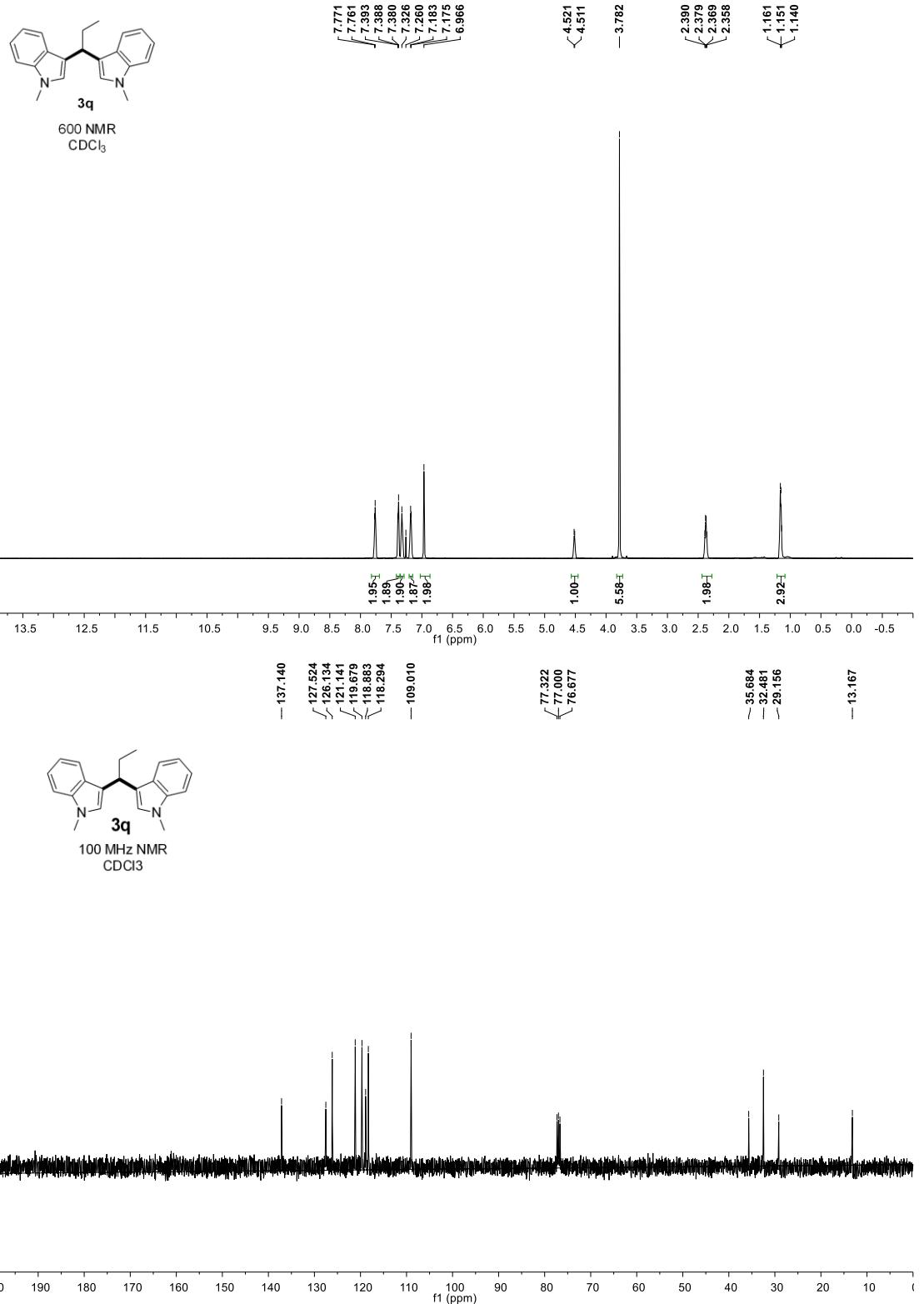


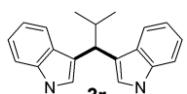




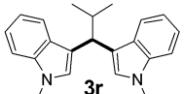
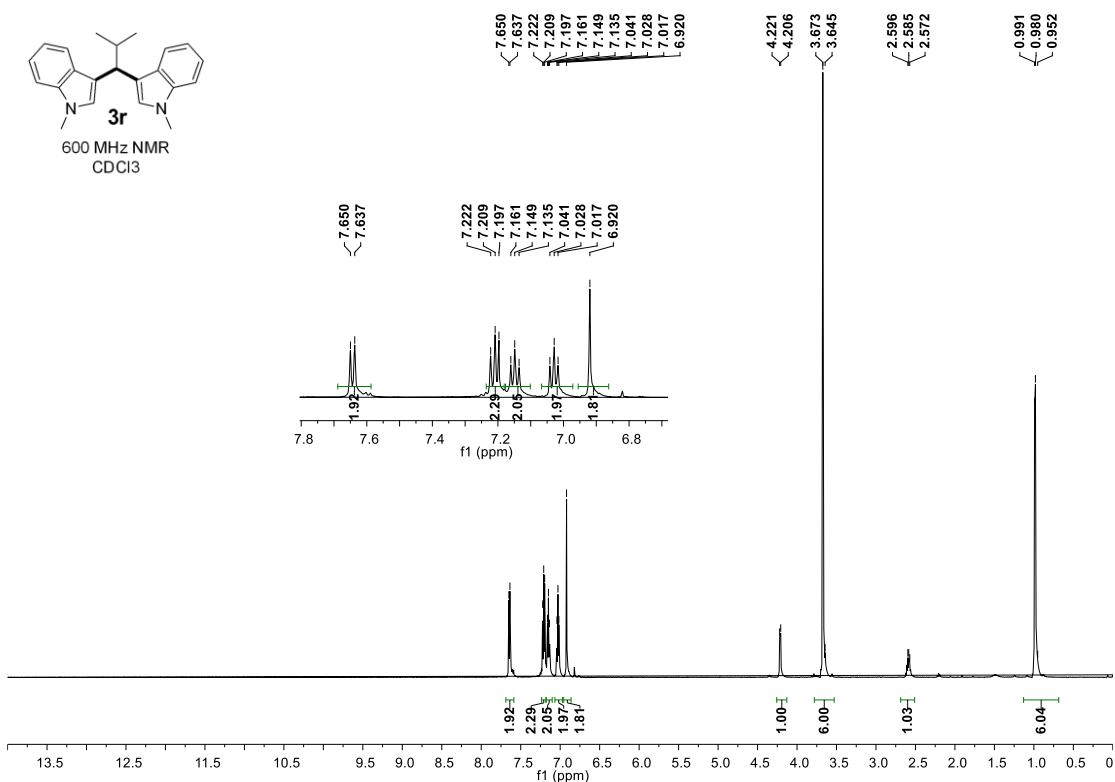




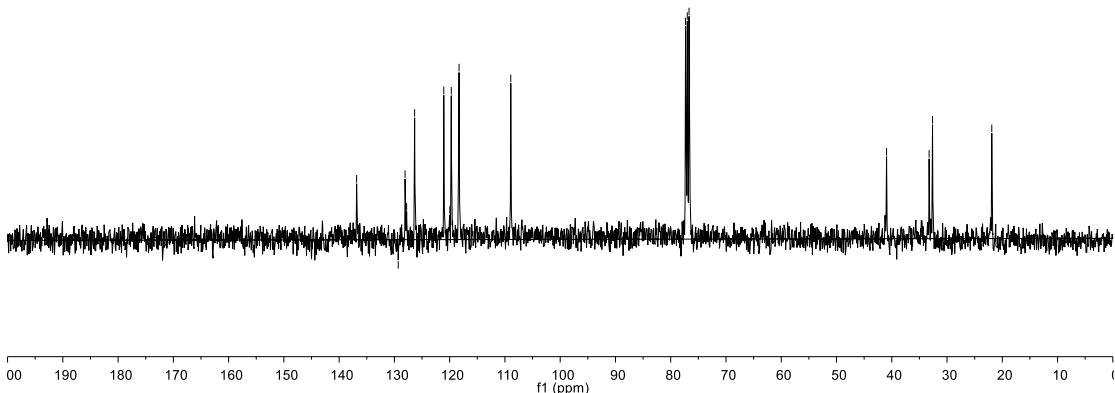


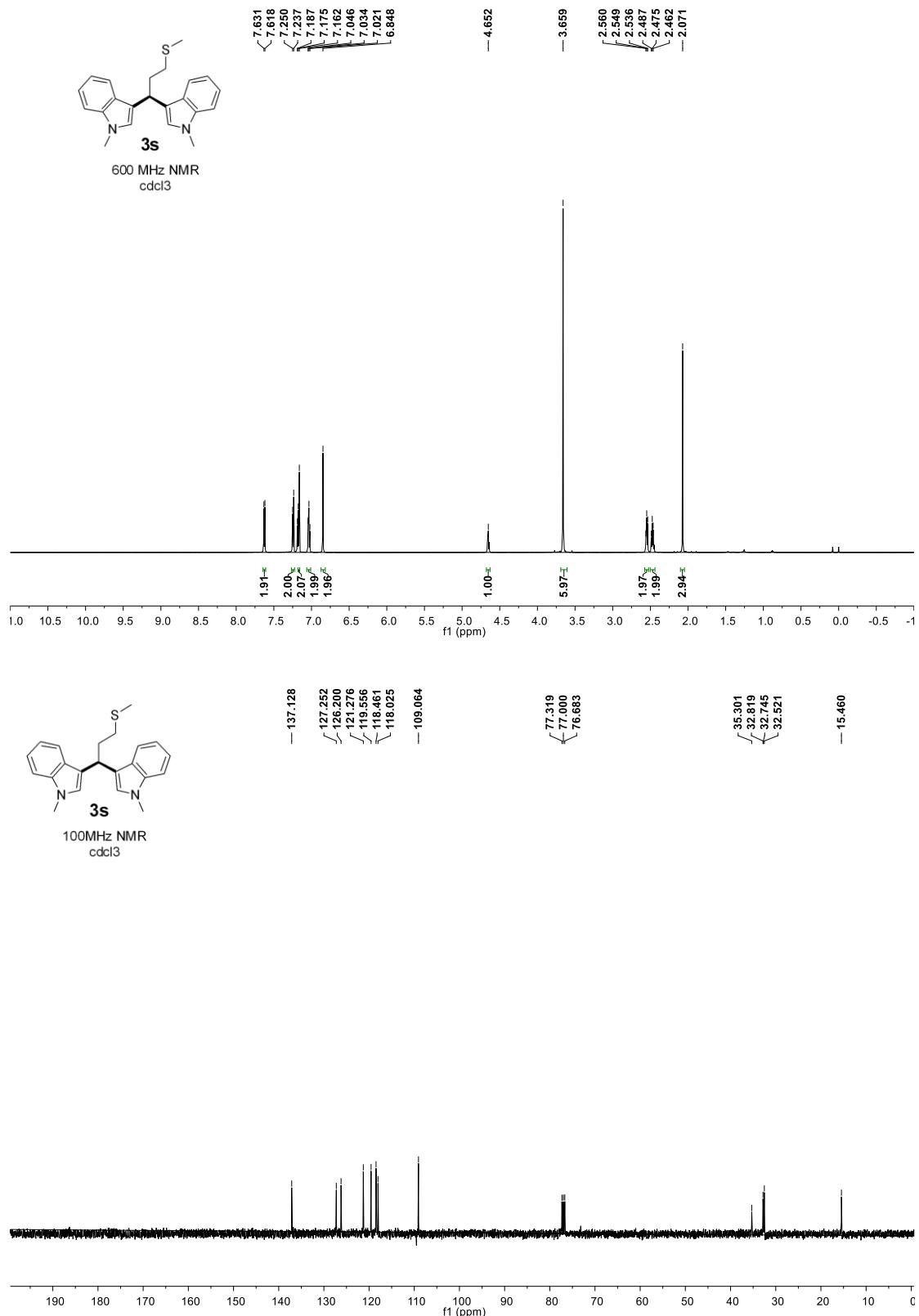


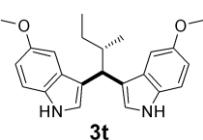
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CDCl₃



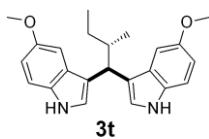
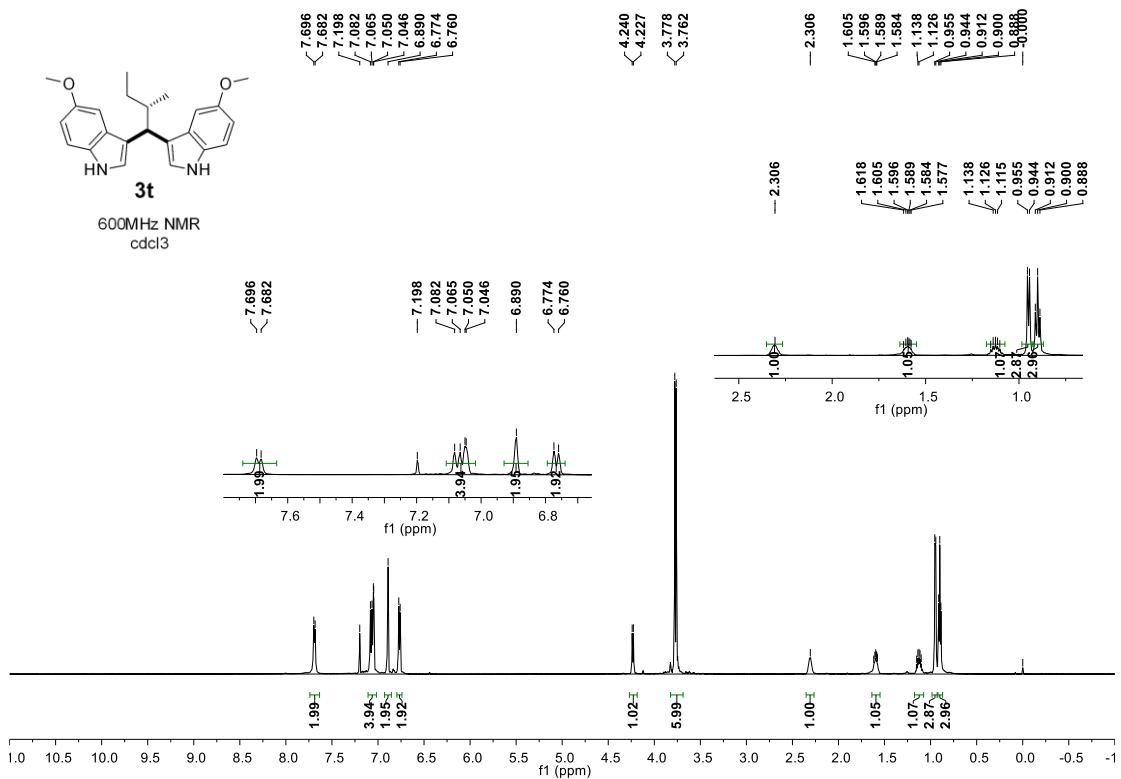
100 MHz NMR
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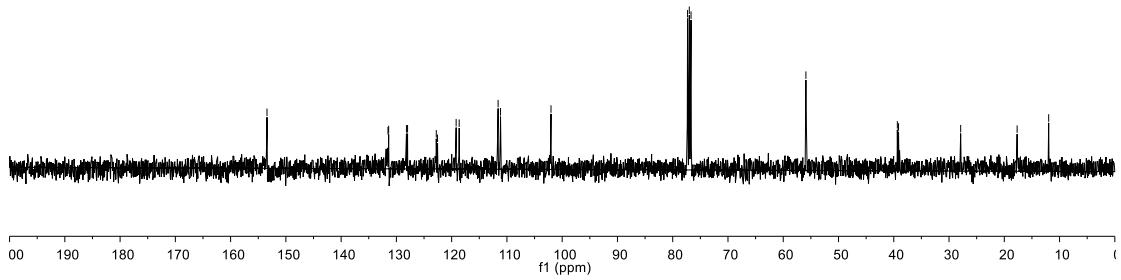


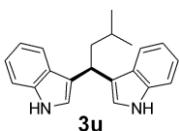


600MHz NMR
cdcl3

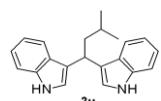
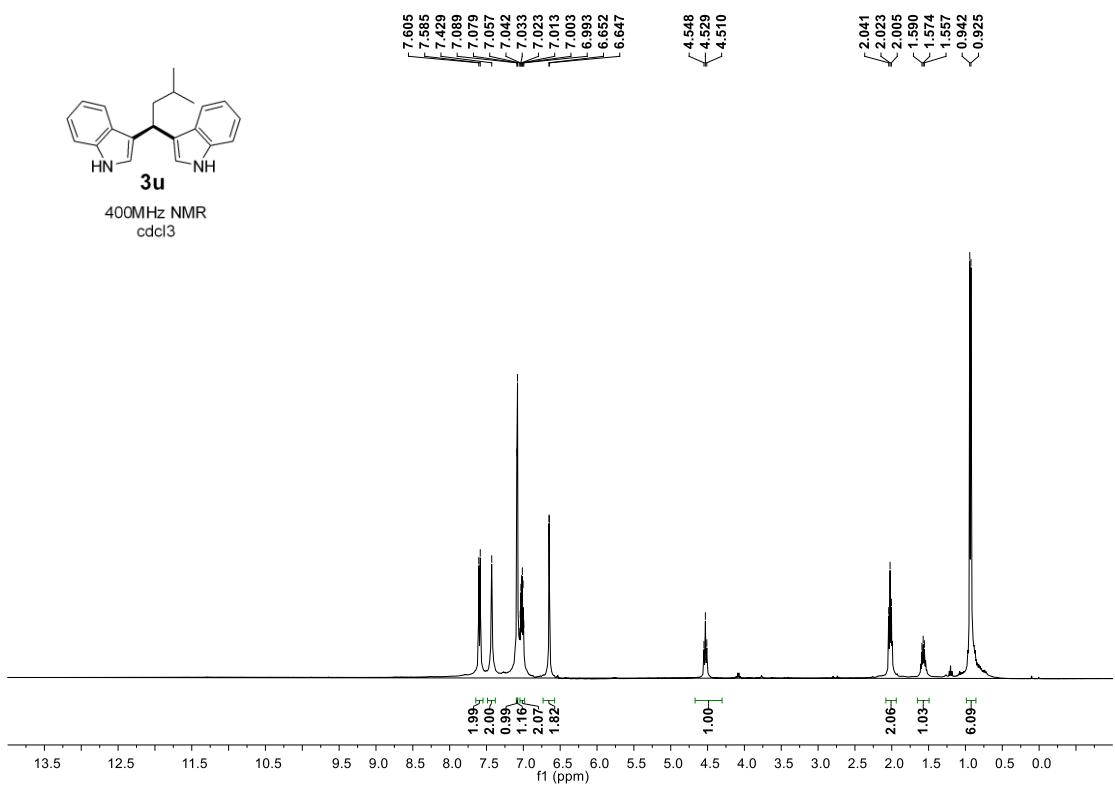


100MHz NMR
cdcl3

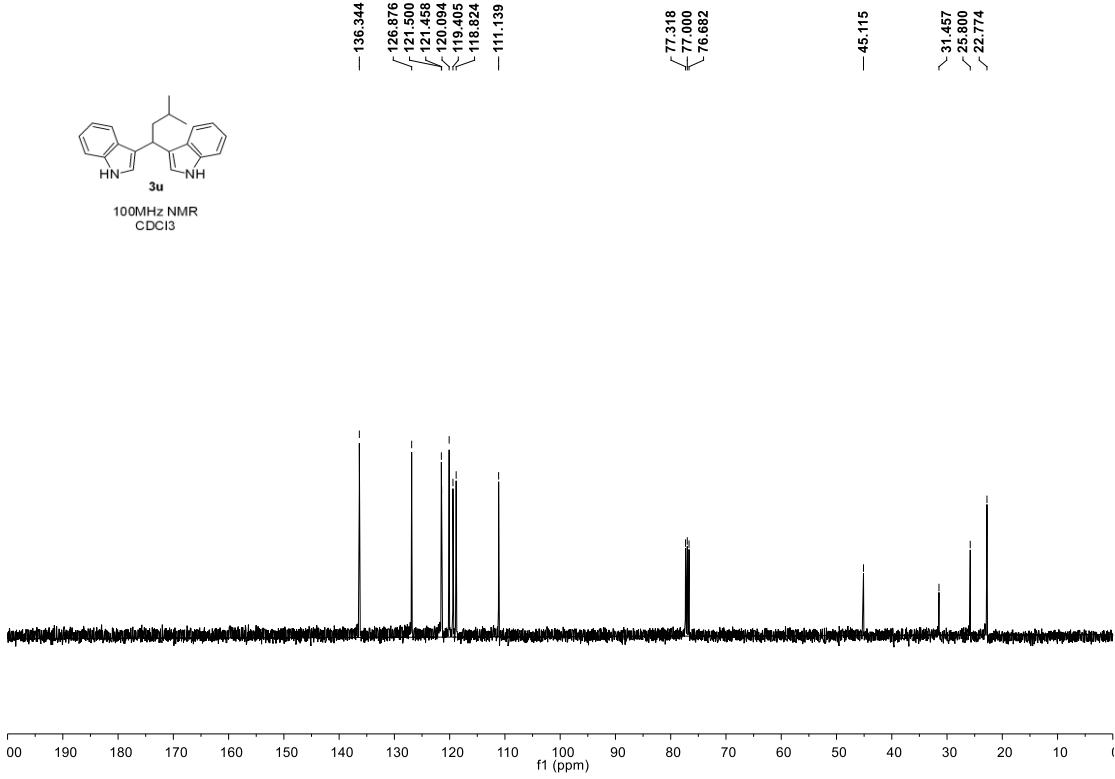


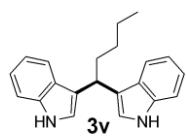


400MHz NMR
cdcl₃

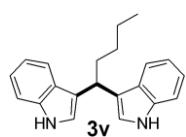
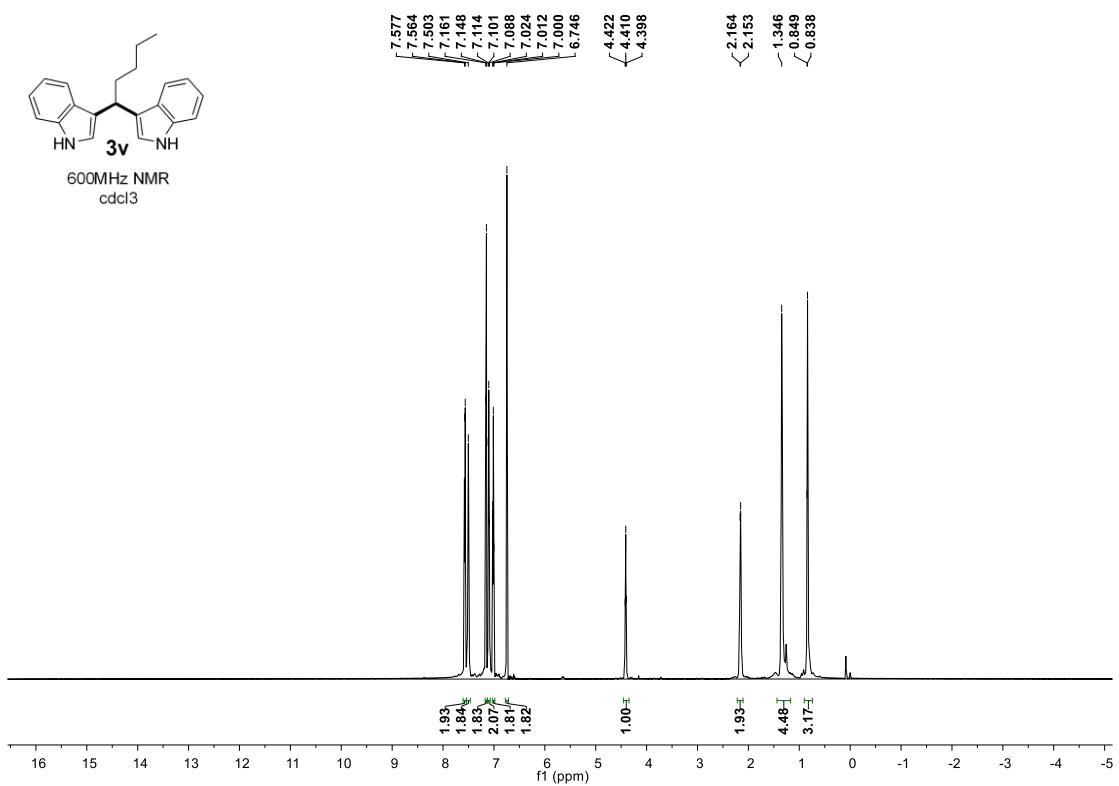


100MHz NMR
CDCl₃

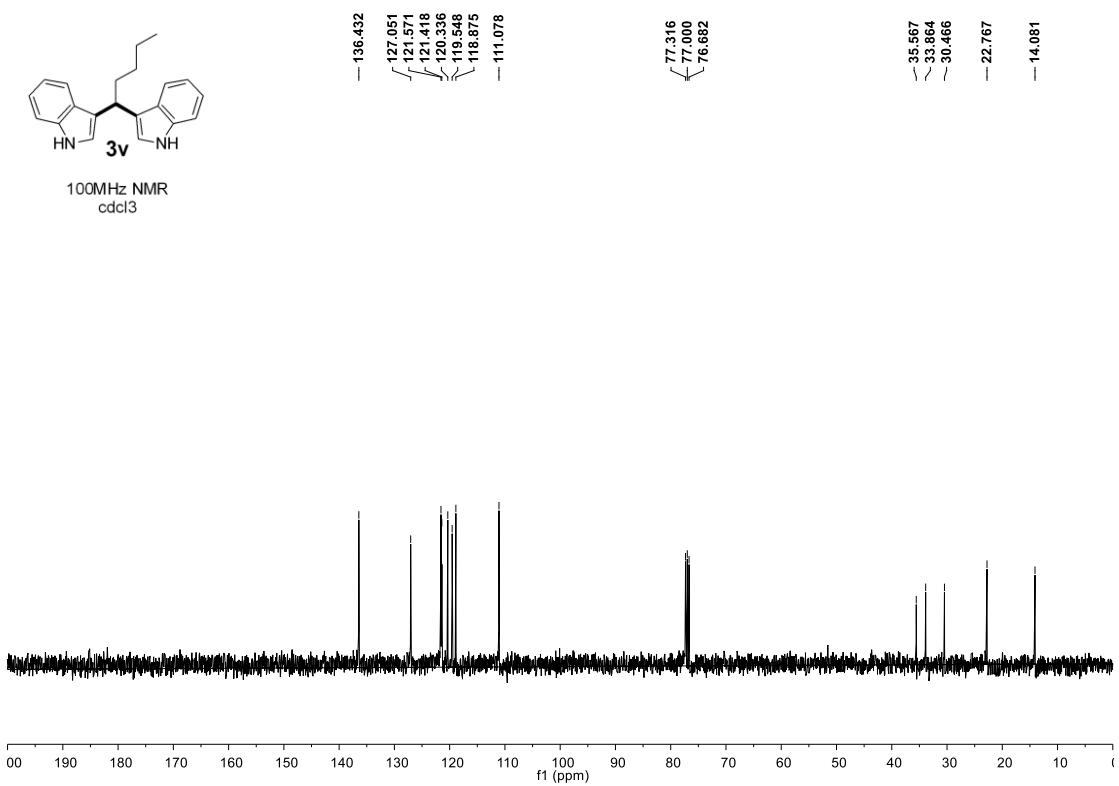


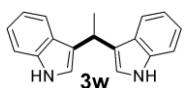


600MHz NMR
cdcl3

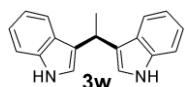
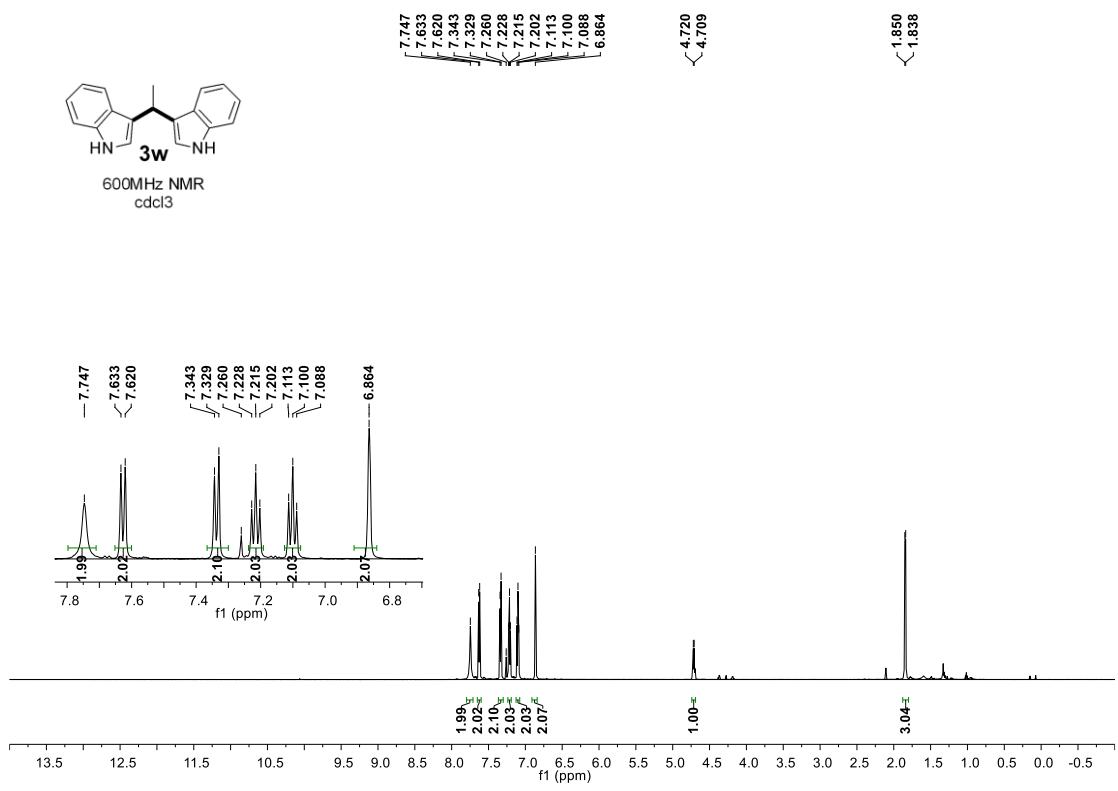


100MHz NMR
cdcl3

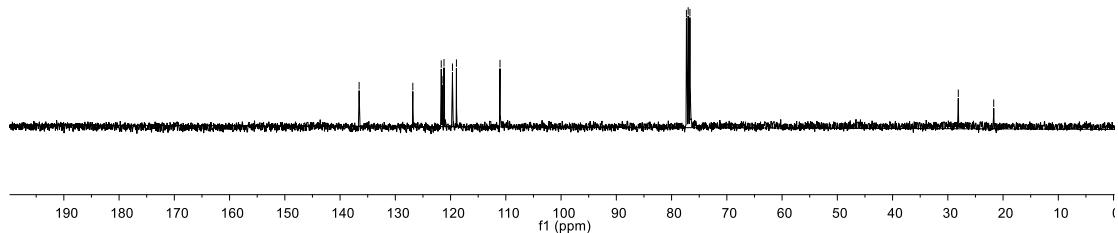




600MHz NMR
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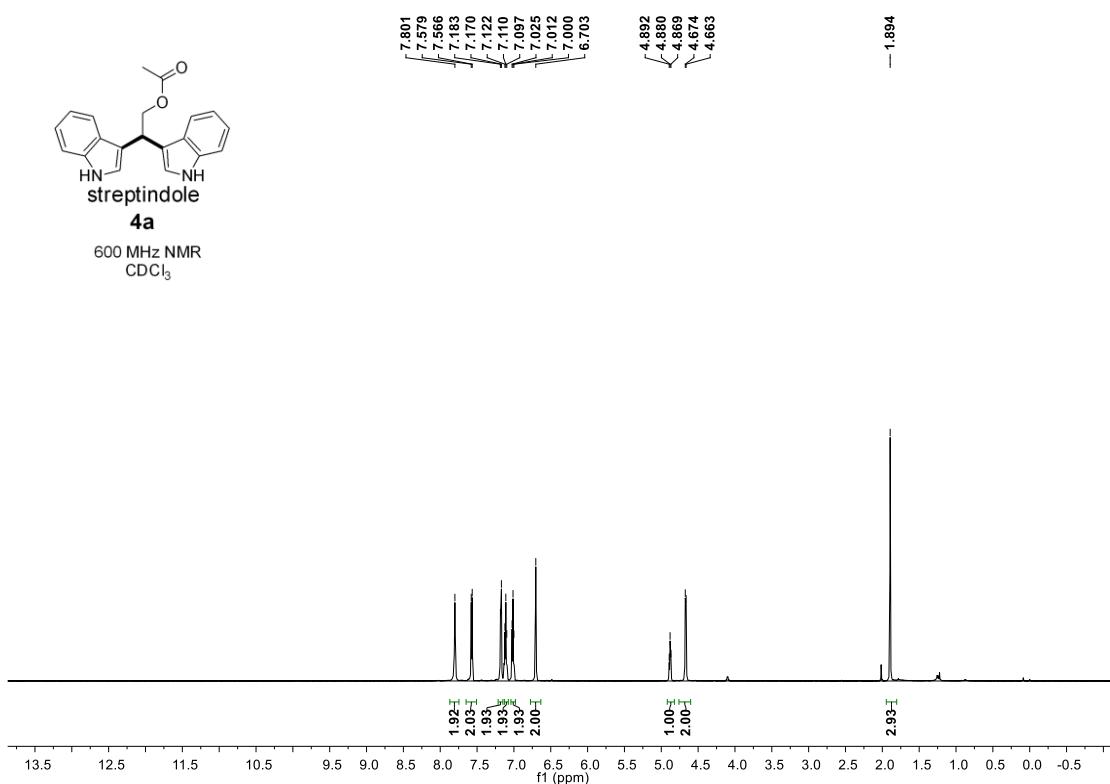


100MHz NMR
cdcl3

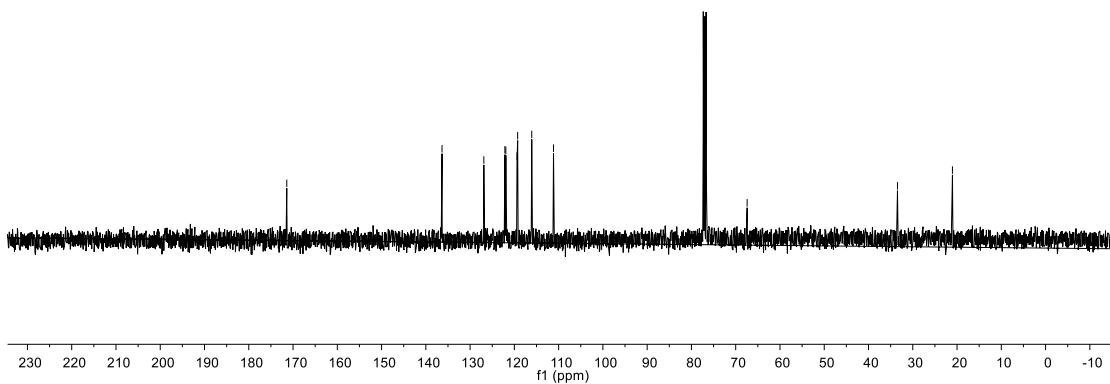




600 MHz NMR
 CDCl_3

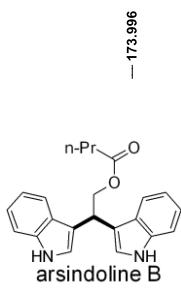
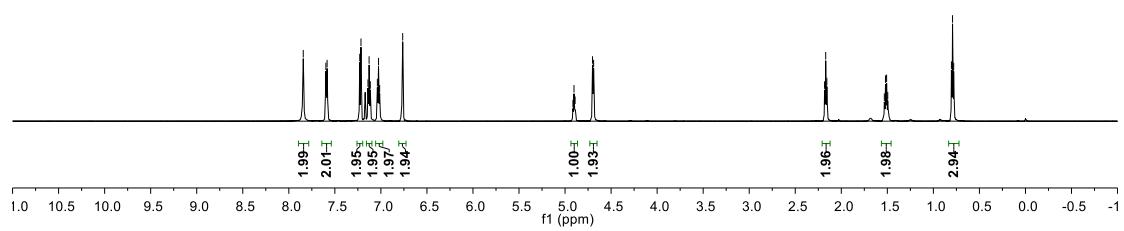


100 MHz NMR
 CDCl_3

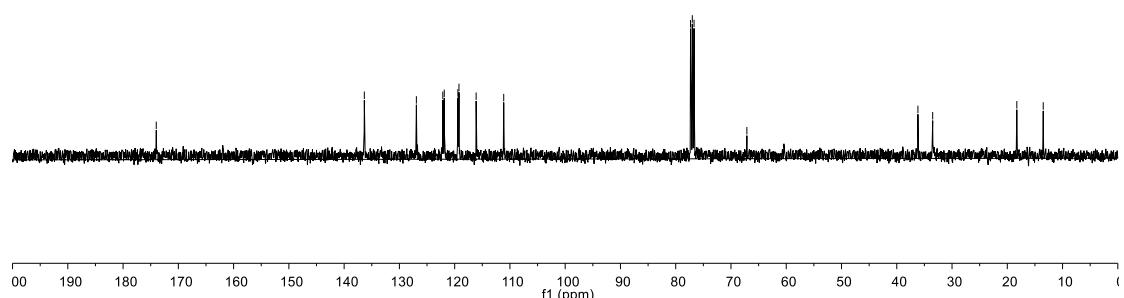


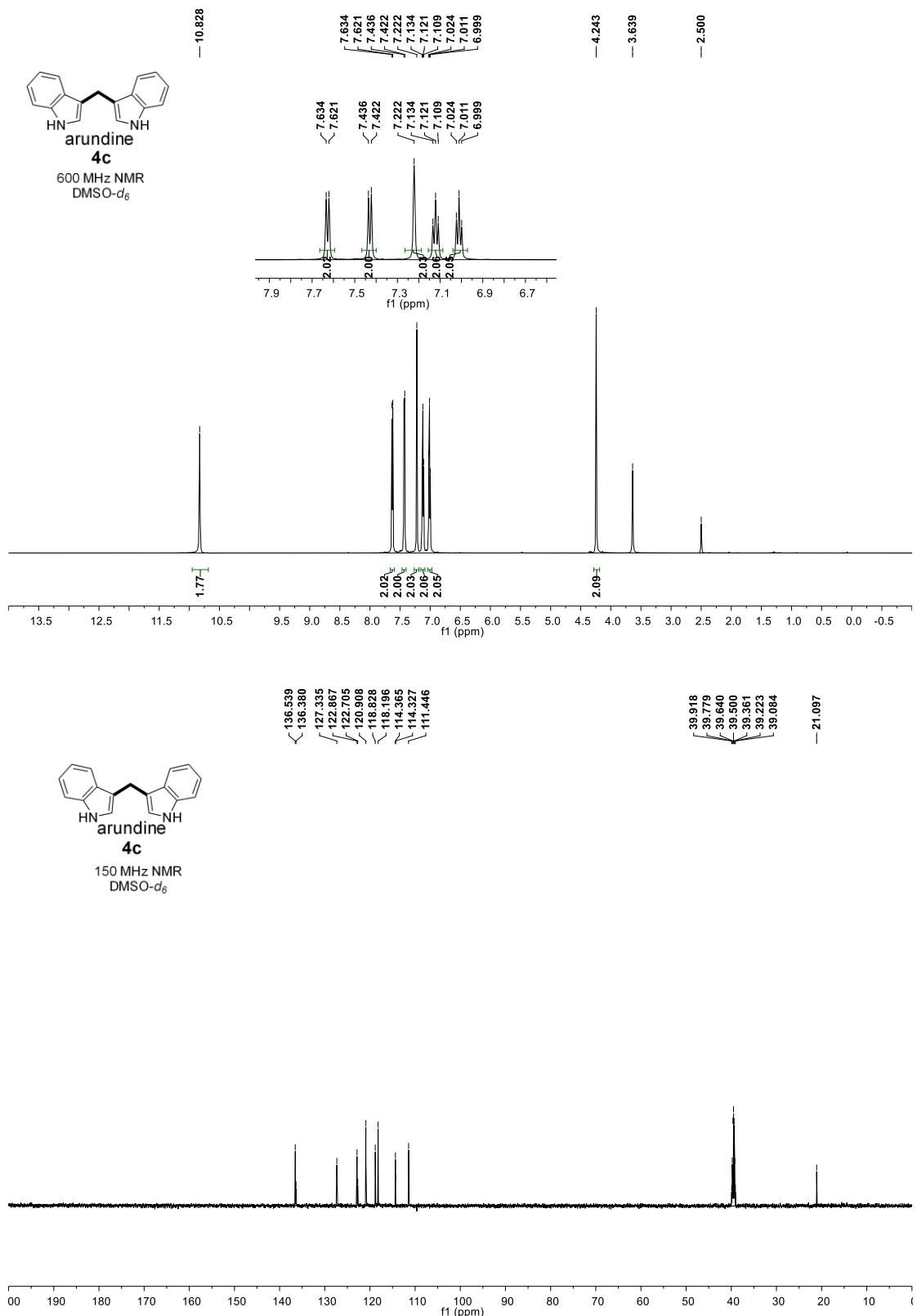


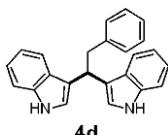
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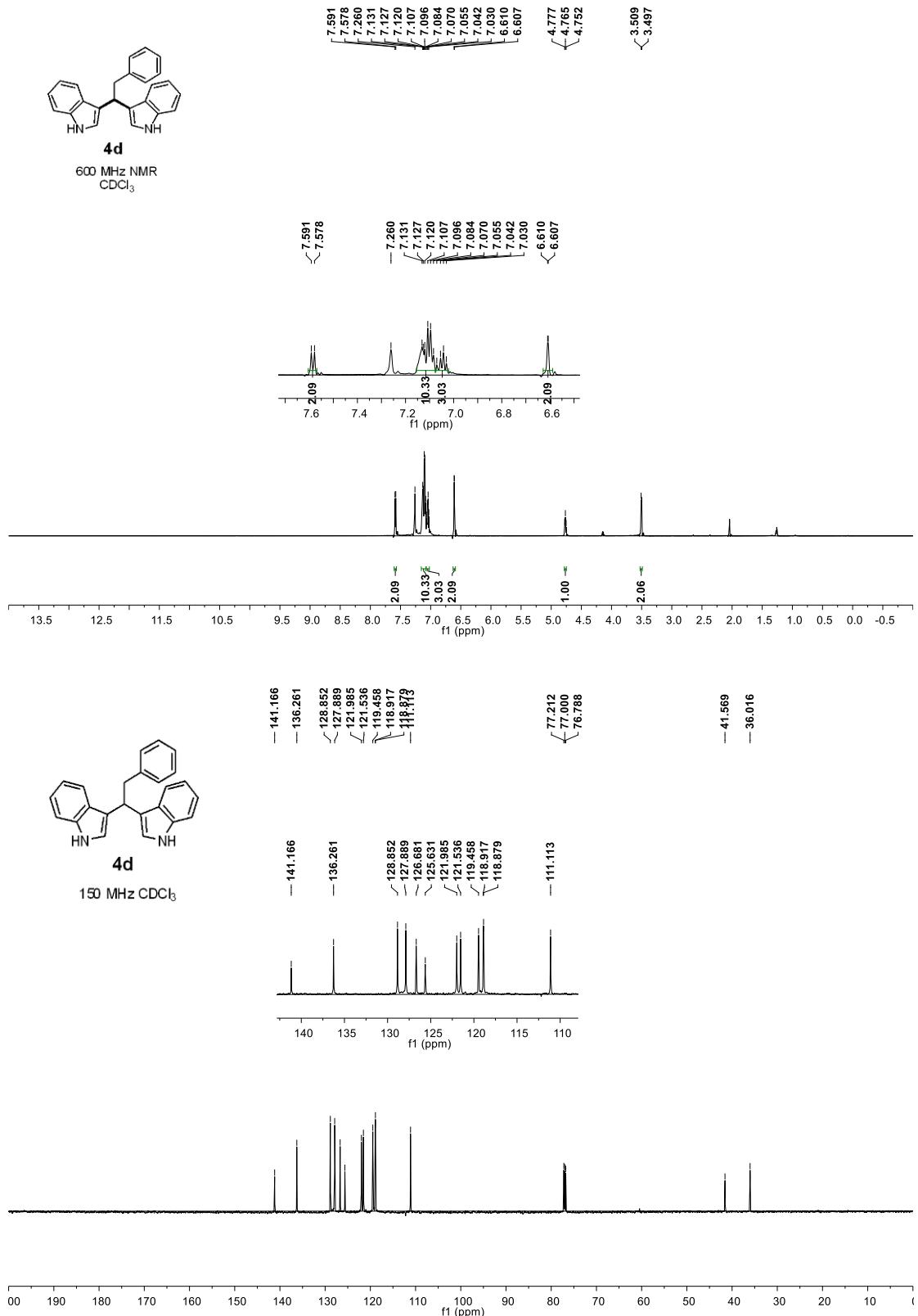
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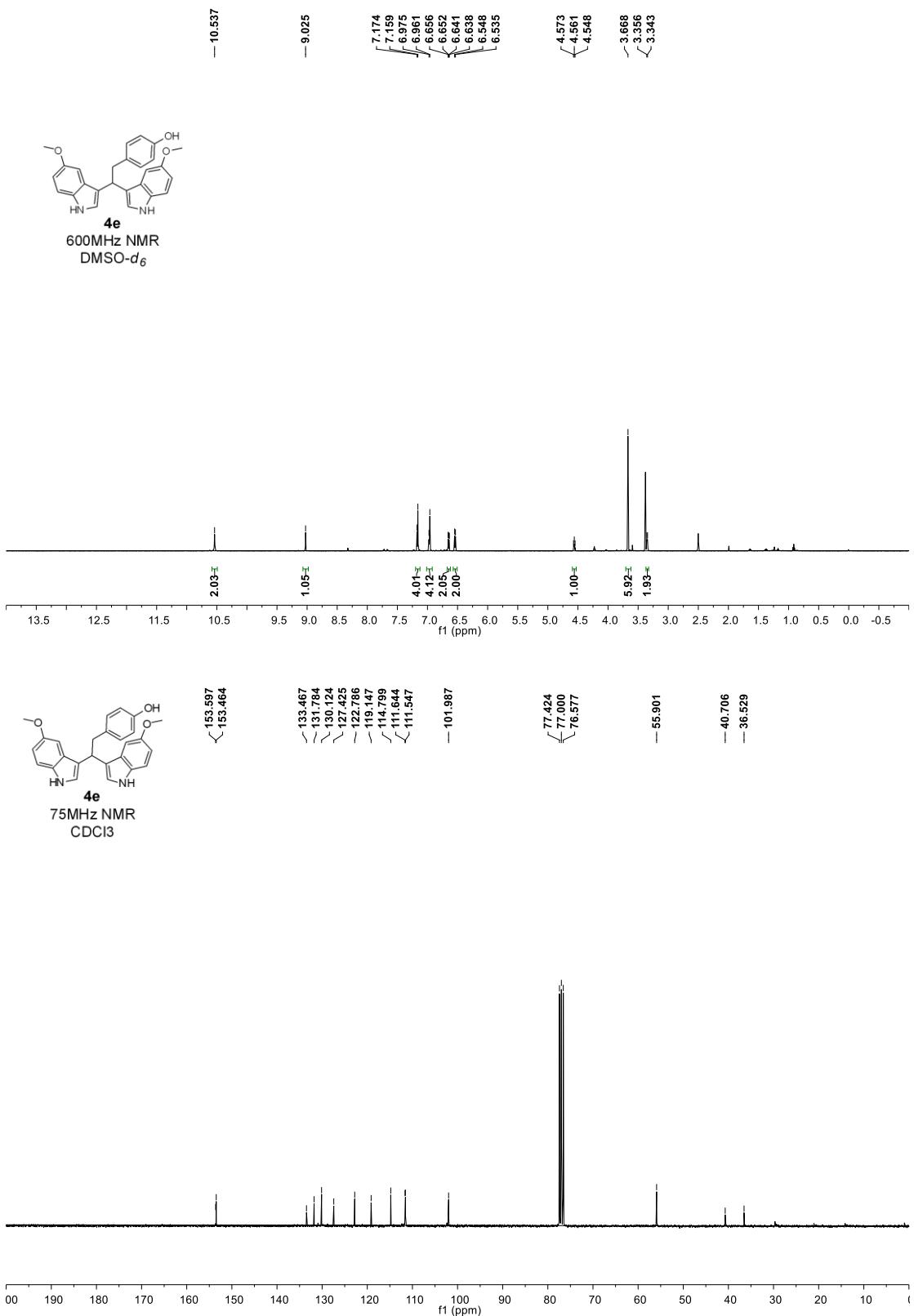


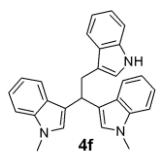




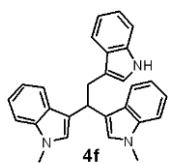
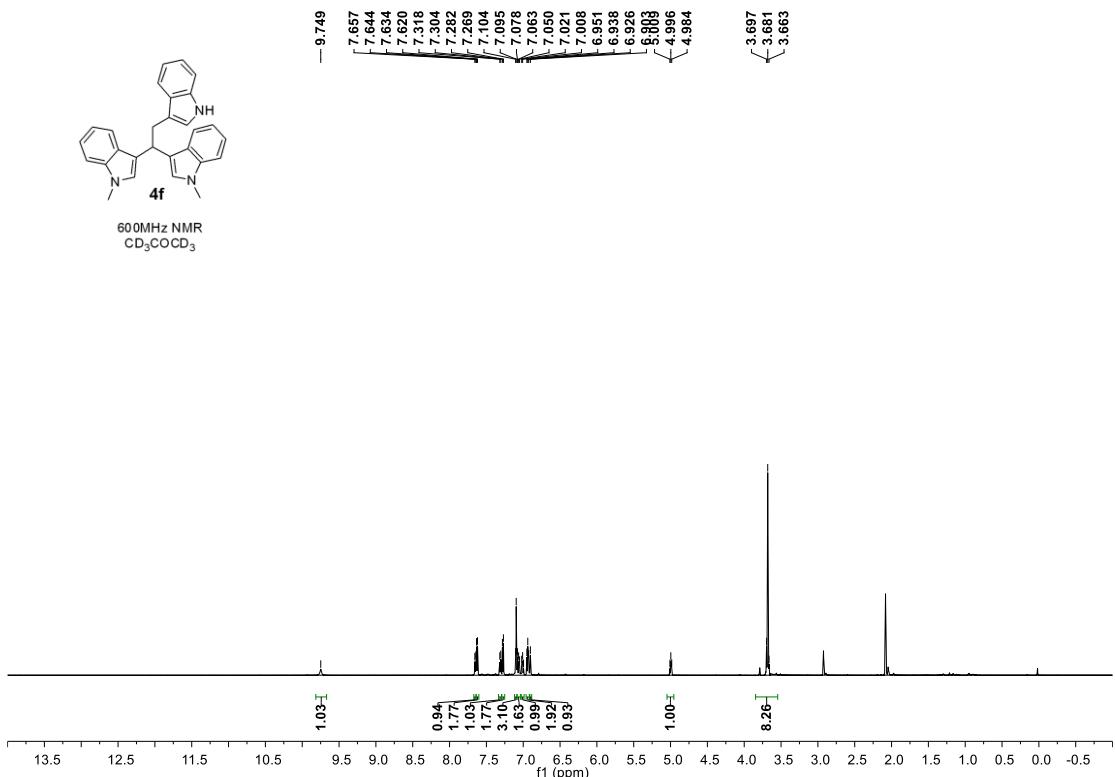
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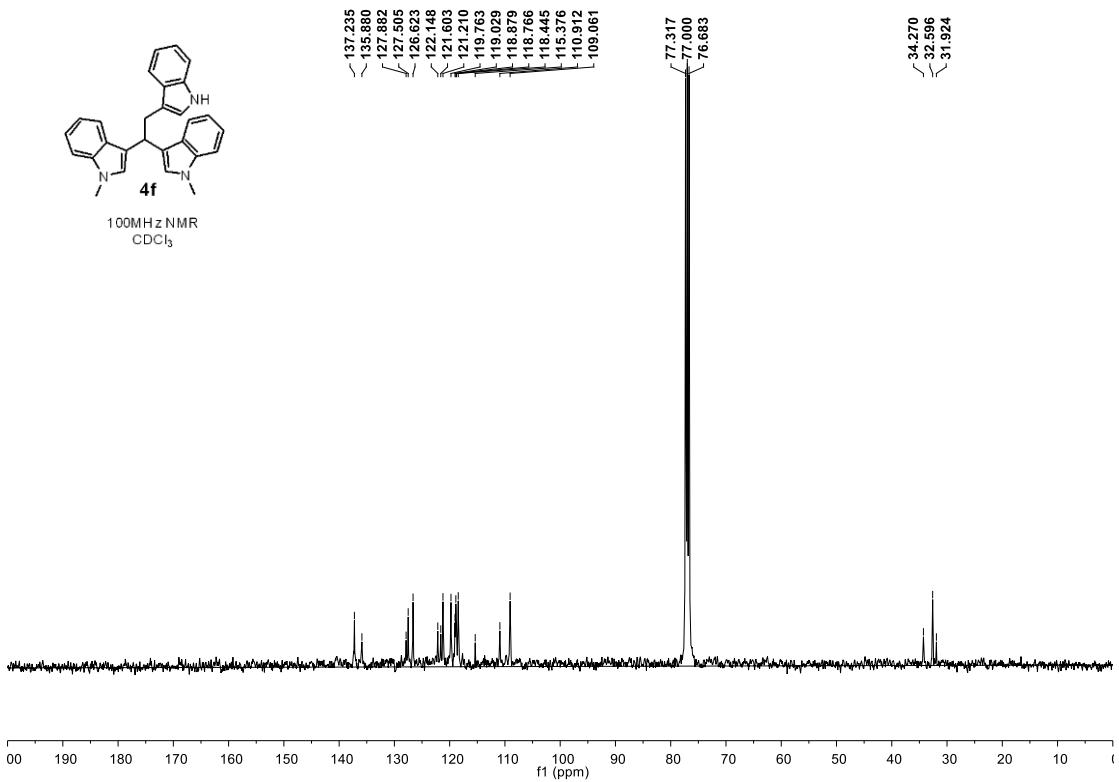


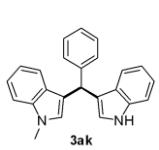


600MHz NMR
 CD_3COCD_3

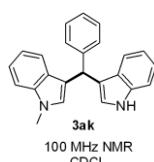
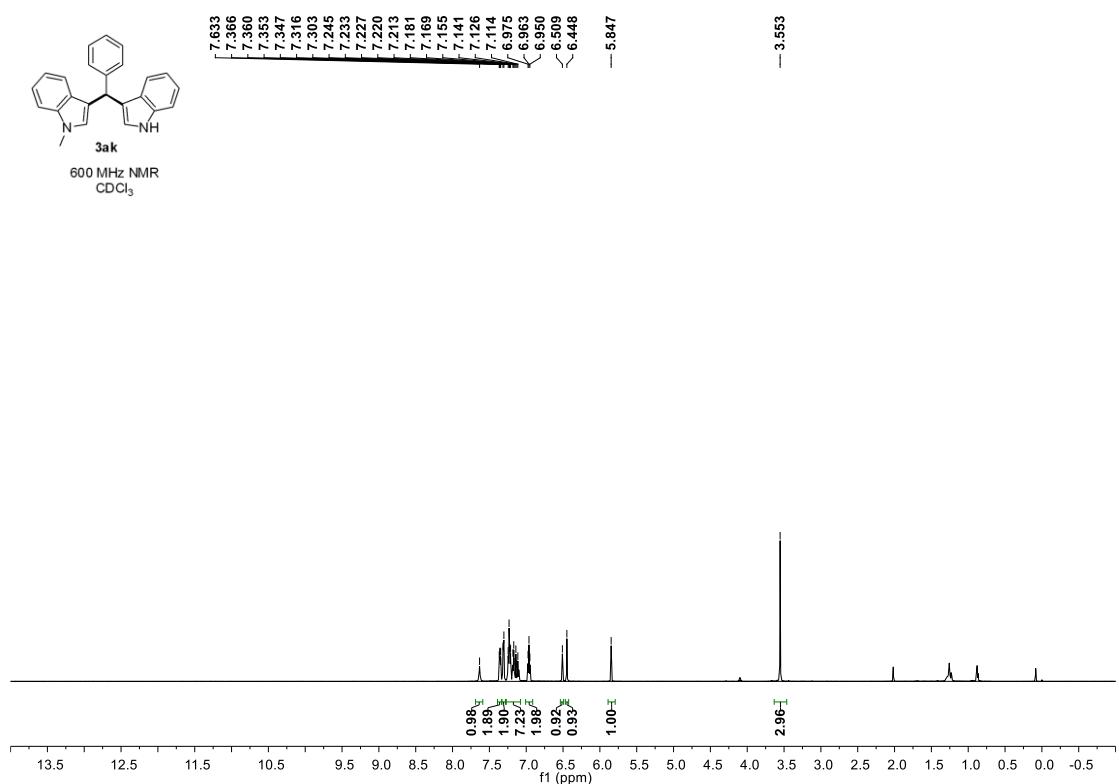


100MHz NMR
 CDCl_3

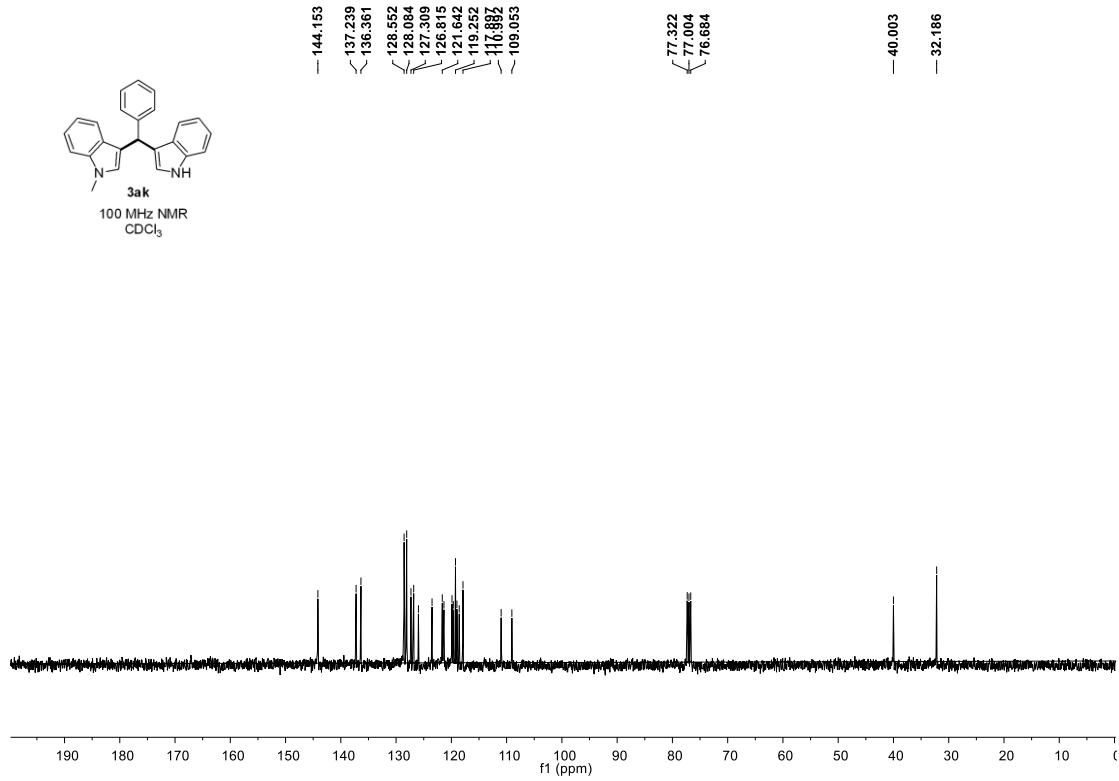


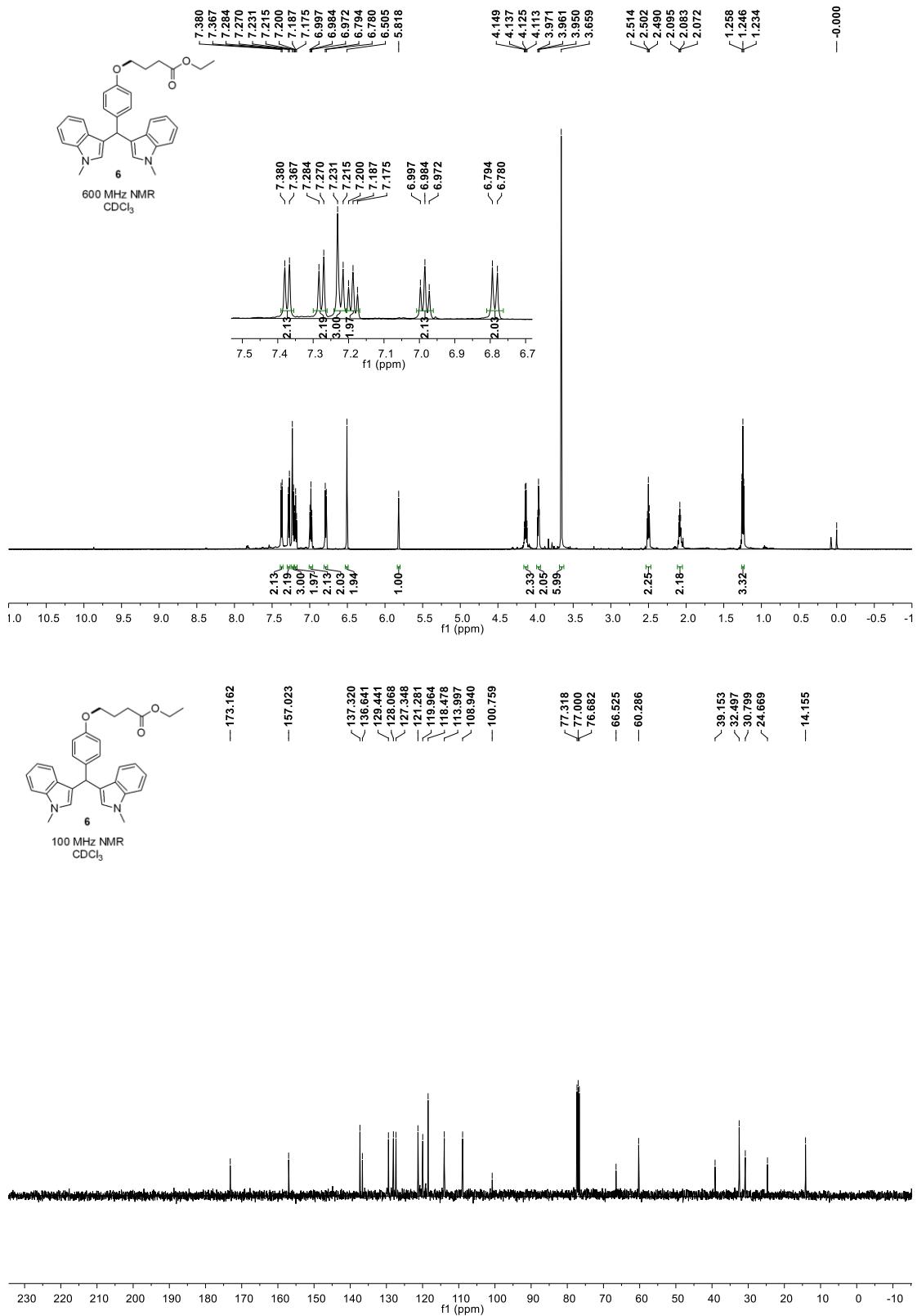


600 MHz NMR
 CDCl_3



100 MHz NMR
 CDCl_3





Mass Spectrum SmartFormula Report

Analysis Info

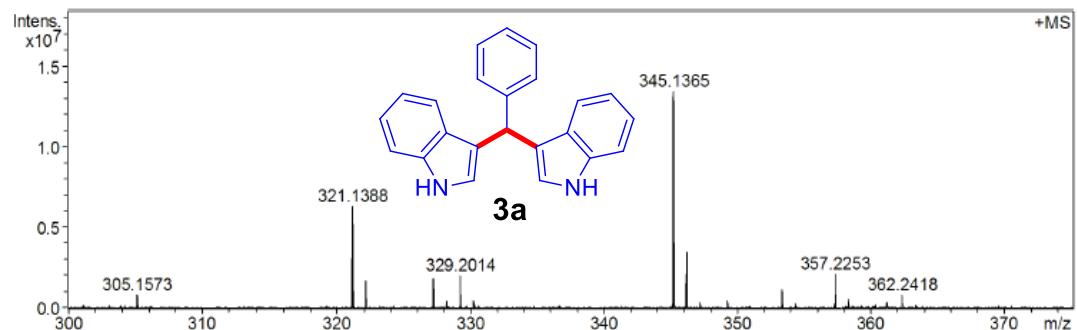
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 Sample Name Student_4-6
 Comment

Acquisition Date 2014-7-2 17:03:20

 Operator
 Instrument apex-Ultra

Acquisition Parameter

Polarity	Positive	Source	ESI	No. of Laser Shots	20
Averaged Scans	2	No. of Cell Fills	1	Laser Power	51.0 %
Broadband Low Mass	100.3 m/z	End Plate	3500.0 V	MALDI Plate	300.0 V
Broadband High Mass	1200.0 m/z	Capillary Entrance	4000.0 V	Imaging Spot Diameter	2000.0 μ m
Acquisition Mode	Single MS	Skimmer 1	20.0 V		
Pulse Program	basic	Drying Gas Temperature	180.0 °C	Calibration Date	Wed Jun 18 07:18:23
Source Accumulation	0.0 sec	Drying Gas Flow Rate	4.0 L/min	Data Acquisition Size	281072
Ion Accumulation Time	0.1 sec	Nebulizer Gas Flow Rate	1.0 L/min	Apodization	Sine-Bell Multiplication
Flight Time to Acq. Cell	0.0 sec				



Meas . m/z	#	Form ula	m/z	err [ppm]	Mean err [ppm]	rdb	N-Rule	ej‡ Conf	mSig ma	Std I	Std Mean m/z	Std VarN orm	Std m/z Diff	Std Com b Dev
345.1 365	1	C 23 H 18 N 2 Na	345.1 362	-0.8	-0.7	15.5	ok	even	3.81	0.005 4	0.000 3	0.002 4	0.000 3	0.842 7

Mass Spectrum SmartFormula Report

Analysis Info

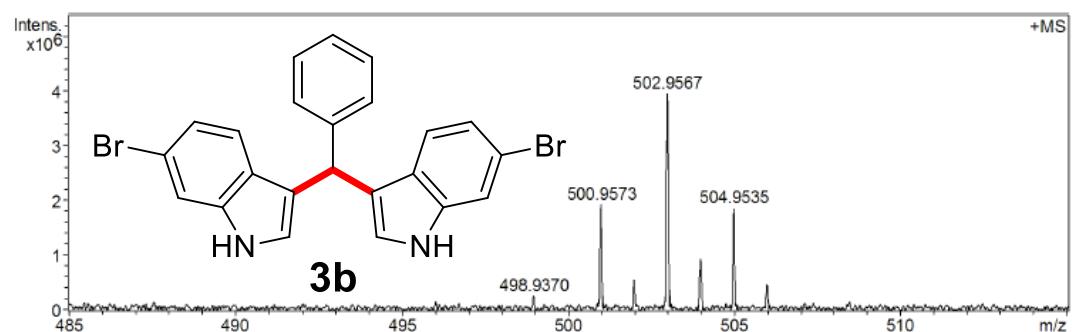
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 Sample Name Student_4-6
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 Operator
 Instrument apex-Ultra

Acquisition Parameter

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Broadband Low Mass	100.3 m/z	End Plate	3500.0 V	MALDI Plate	300.0 V
Broadband High Mass	1200.0 m/z	Capillary Entrance	4000.0 V	Imaging Spot Diameter	2000.0 μ m
Acquisition Mode	Single MS	Skimmer 1	20.0 V		
Pulse Program	basic	Drying Gas Temperature	180.0 °C	Calibration Date	Wed Jun 18 07:18:23
Source Accumulation	0.0 sec	Drying Gas Flow Rate	4.0 L/min	Data Acquisition Size	201072
Ion Accumulation Time	0.1 sec	Nebulizer Gas Flow Rate	1.0 L/min	Apodization	Sine-Bell Multiplication
Flight Time to Acq. Cell	0.0 sec				



Meas . m/z	#	Form ula	m/z	err [ppm]	Mean err [ppm]	rdb	N-Rule	eJ Conf	mSig ma	Std I	Std Mean m/z	Std VarN orm	Std m/z Diff	Std Com b Dev
500.9 573	1	C 23 H 16 Br 2 N 2 Na	500.9 572	-0.2	-0.7	15.5	ok	even	18.27	0.017 0	0.001 1	0.005 9	0.001 8	0.842 7

Mass Spectrum SmartFormula Report

Analysis Info

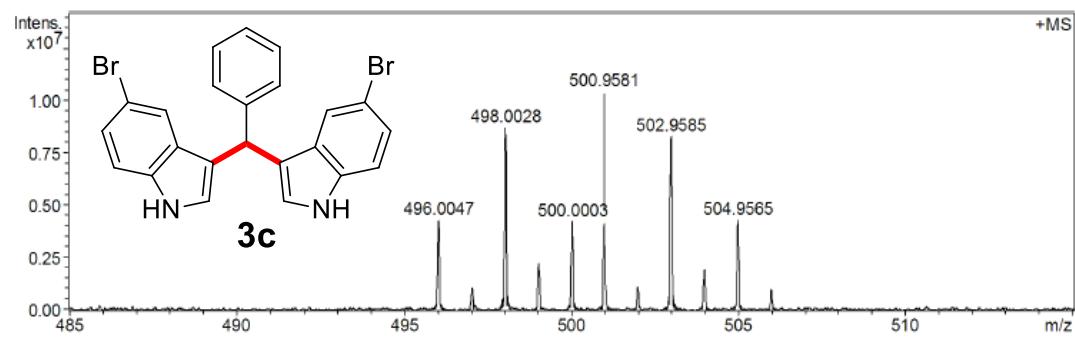
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 Sample Name Student_4-6
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Acquisition Date 2014-7-2 17:05:12

 Operator
 Instrument apex-Ultra

Acquisition Parameter

Polarity	Positive	Source	ESI	No. of Laser Shots	20
Averaged Scans	2	No. of Cell Fills	1	Laser Power	51.0 %
Broadband Low Mass	100.3 m/z	End Plate	3500.0 V	MALDI Plate	300.0 V
Broadband High Mass	1200.0 m/z	Capillary Entrance	4000.0 V	Imaging Spot Diameter	2000.0 μ m
Acquisition Mode	Single MS	Skimmer 1	20.0 V		
Pulse Program	basic	Drying Gas Temperature	180.0 °C	Calibration Date	Wed Jun 18 07:18:23
Source Accumulation	0.0 sec	Drying Gas Flow Rate	4.0 L/min	Data Acquisition Size	281072
Ion Accumulation Time	0.1 sec	Nebulizer Gas Flow Rate	1.0 L/min	Apodization	Sine-Bell Multiplication
Flight Time to Acq. Cell	0.0 sec				



Meas . m/z	#	Form ular	m/z	err [ppm]	Mean err [ppm]	rdb	N-Rule	ej% Conf	mSig ma	Std I m/z	Std Mean m/z	Std VarN orm	Std m/z Diff	Std Com b Dev
500.9581	1	C 23 H 16 Br 2 N 2 Na	500.9572	-1.6	-5.2	15.5	ok	even	9.75	0.0103	0.0028	0.0045	0.0012	0.8427

Mass Spectrum SmartFormula Report

Analysis Info

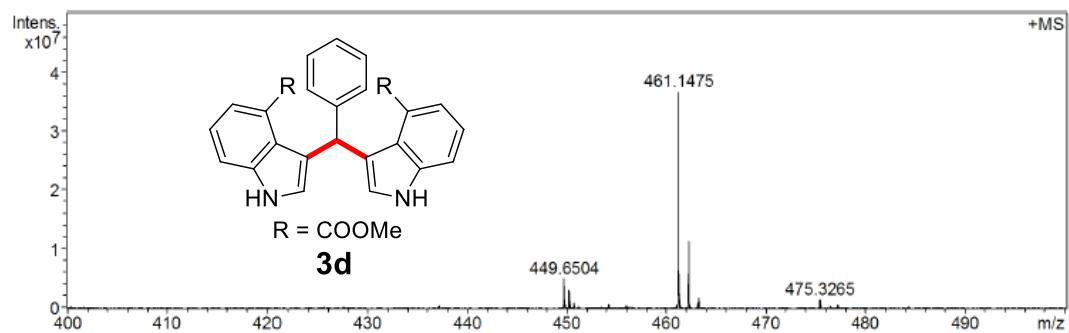
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 Method Student_LCms-140618
 Sample Name Student_4-6
 Comment

Acquisition Date 2014-7-2 17:07:41

 Operator
 Instrument apex-Ultra

Acquisition Parameter

Polarity	Positive	Source	ESI	No. of Laser Shots	20
Averaged Scans	2	No. of Cell Fills	1	Laser Power	51.0 %
Broadband Low Mass	100.3 m/z	End Plate	3500.0 V	MALDI Plate	300.0 V
Broadband High Mass	1200.0 m/z	Capillary Entrance	4000.0 V	Imaging Spot Diameter	2000.0 μm
Acquisition Mode	Single MS	Skimmer 1	20.0 V		
Pulse Program	basic	Drying Gas Temperature	180.0 °C	Calibration Date	Wed Jun 18 07:18:23
Source Accumulation	0.0 sec	Drying Gas Flow Rate	4.0 L/min	Data Acquisition Size	201072
Ion Accumulation Time	0.1 sec	Nebulizer Gas Flow Rate	1.0 L/min	Apodization	Sine-Bell Multiplication
Flight Time to Acq. Cell	0.0 sec				



Meas . m/z	#	Form ula	m/z	err [ppm]	Mean err [ppm]	rdb	N-Rule	e/Conf	mSig ma	Std I	Std I	Std I	Std m/z	Std VarN orm	Std m/z Diff	Std Com b Dev
461.1 475	1	C 27 H 22 N 2 Na O 4	461.1 472	-0.7	-0.7	17.5	ok	even	5.19	0.007 4	0.000 4	0.003 1	0.000 1	0.842 7		

Mass Spectrum SmartFormula Report

Analysis Info

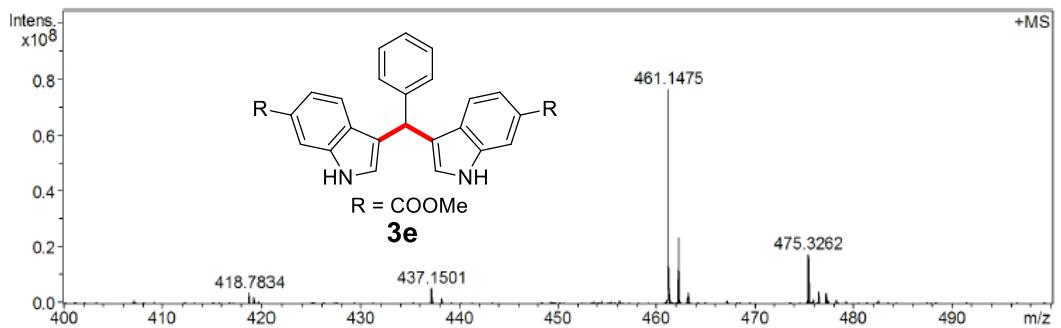
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 Sample Name Student_4-6
 Comment

Acquisition Date 2014-7-2 17:08:27

 Operator
 Instrument apex-Ultra

Acquisition Parameter

Polarity	Positive	Source	ESI	No. of Laser Shots	20
Averaged Scans	2	No. of Cell Fills	1	Laser Power	51.0 %
Broadband Low Mass	100.3 m/z	End Plate	3500.0 V	MALDI Plate	300.0 V
Broadband High Mass	1200.0 m/z	Capillary Entrance	4000.0 V	Imaging Spot Diameter	2000.0 μ m
Acquisition Mode	Single MS	Skimmer 1	20.0 V		
Pulse Program	basic	Drying Gas Temperature	180.0 °C	Calibration Date	Wed Jun 18 07:18:23
Source Accumulation	0.0 sec	Drying Gas Flow Rate	4.0 L/min	Data Acquisition Size	201072
Ion Accumulation Time	0.1 sec	Nebulizer Gas Flow Rate	1.0 L/min	Apodization	Sine-Bell Multiplication
Flight Time to Acq. Cell	0.0 sec				



Meas . m/z	#	Form u la	m/z	err [ppm]	Mean err [ppm]	rdb	N-Rul e	e‡ Conf	mSig ma	Std I	Std I	Std m/z	Std VarN orm	Std m/z Diff	Std Com b Dev
461.1															
475	1	C 27 H 22 N 2 Na O 4	461.1 472	-0.7	-0.6	17.5	ok	even	3.20	0.004 7	0.000 3	0.001 7	0.000 3	0.842 7	

Mass Spectrum SmartFormula Report

Analysis Info

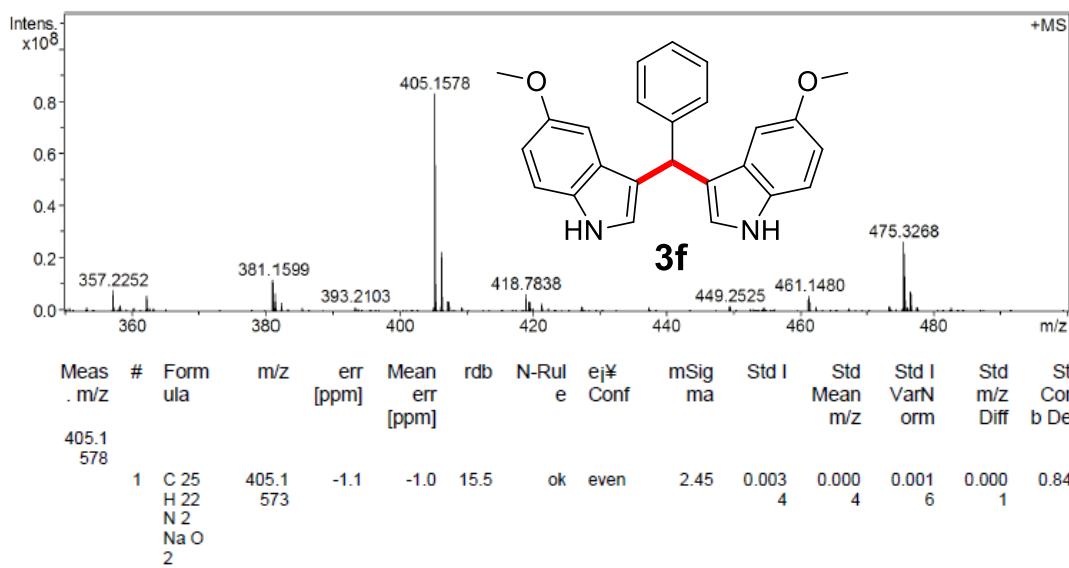
Analysis Name H:\20140702-1\JC\jc564_000002.d
 Method Student_LCms-140618
 Sample Name Student_4-6
 Comment

Acquisition Date 2014-7-2 17:09:15

Operator Instrument apex-Ultra

Acquisition Parameter

Polarity	Positive	Source	ESI	No. of Laser Shots	20
Averaged Scans	2	No. of Cell Fills	1	Laser Power	51.0 %
Broadband Low Mass	100.3 m/z	End Plate	3500.0 V	MALDI Plate	300.0 V
Broadband High Mass	1200.0 m/z	Capillary Entrance	4000.0 V	Imaging Spot Diameter	2000.0 μm
Acquisition Mode	Single MS	Skimmer 1	20.0 V	Calibration Date	Wed Jun 18 07:18:23
Pulse Program	basic	Drying Gas Temperature	180.0 °C	Data Acquisition Size	201972
Source Accumulation	0.0 sec	Drying Gas Flow Rate	4.0 L/min	Apodization	Sine-Bell Multiplication
Ion Accumulation Time	0.1 sec	Nebulizer Gas Flow Rate	1.0 L/min		
Flight Time to Acq. Cell	0.0 sec				



Mass Spectrum SmartFormula Report

Analysis Info

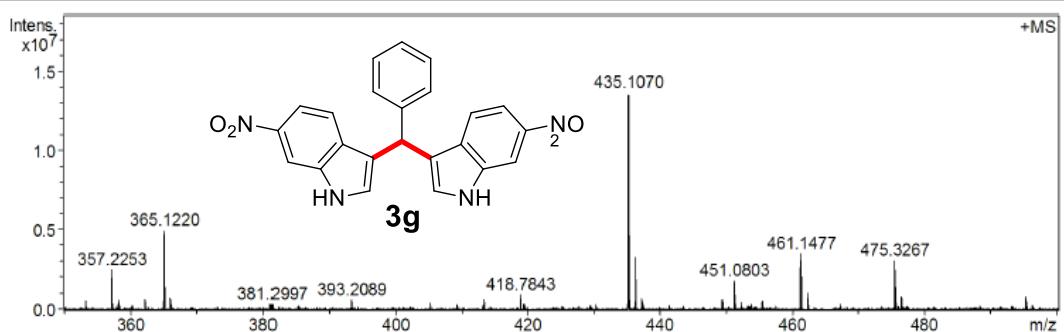
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 Method Student_LCms-140618
 Sample Name Student_4-6
 Comment

Acquisition Date 2014-7-2 17:10:09

 Operator
 Instrument apex-Ultra

Acquisition Parameter

Polarity	Positive	Source	ESI	No. of Laser Shots	20
Averaged Scans	2	No. of Cell Fills	1	Laser Power	51.0 %
Broadband Low Mass	100.3 m/z	End Plate	3500.0 V	MALDI Plate	300.0 V
Broadband High Mass	1200.0 m/z	Capillary Entrance	4000.0 V	Imaging Spot Diameter	2000.0 μm
Acquisition Mode	Single MS	Skimmer 1	20.0 V		
Pulse Program	basic	Drying Gas Temperature	180.0 °C	Calibration Date	Wed Jun 18 07:18:23
Source Accumulation	0.0 sec	Drying Gas Flow Rate	4.0 L/min	Data Acquisition Size	201072
Ion Accumulation Time	0.1 sec	Nebulizer Gas Flow Rate	1.0 L/min	Apodization	Sine-Bell Multiplication
Flight Time to Acq. Cell	0.0 sec				



Meas. m/z	#	Form ula	m/z	err [ppm]	Mean err [ppm]	rdb	N-Rul e	ej¥ Conf	mSig ma	Std I m/z	Std I Mean m/z	Std I VarN orm	Std m/z Diff	Std Com b Dev
435.1 070	1	C 23 H 16 N 4 Na O 4	435.1 064	-1.5	-1.6	17.5	ok	even	12.84	0.016 9	0.000 7	0.009 8	0.000 3	0.842 7

Mass Spectrum SmartFormula Report

Analysis Info

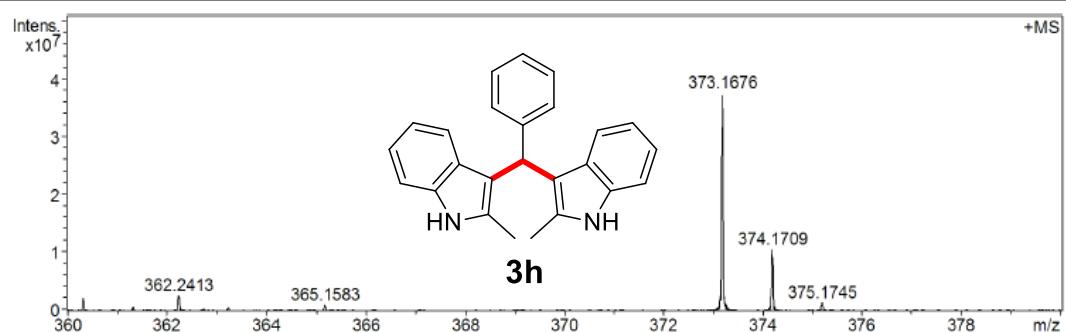
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 Sample Name Student_4-6
 Comment

Acquisition Date 2014-7-2 17:11:05

 Operator
 Instrument apex-Ultra

Acquisition Parameter

Polarity	Positive	Source	ESI	No. of Laser Shots	20
Averaged Scans	2	No. of Cell Fills	1	Laser Power	51.0 %
Broadband Low Mass	100.3 m/z	End Plate	3500.0 V	MALDI Plate	300.0 V
Broadband High Mass	1200.0 m/z	Capillary Entrance	4000.0 V	Imaging Spot Diameter	2000.0 μm
Acquisition Mode	Single MS	Skimmer 1	20.0 V		
Pulse Program	basic	Drying Gas Temperature	180.0 °C	Calibration Date	Wed Jun 18 07:18:23
Source Accumulation	0.0 sec	Drying Gas Flow Rate	4.0 L/min	Data Acquisition Size	281872
Ion Accumulation Time	0.1 sec	Nebulizer Gas Flow Rate	1.0 L/min	Apodization	Sine-Bell Multiplication
Flight Time to Acq. Cell	0.0 sec				



Meas . m/z	#	Form ula	m/z	err [ppm]	Mean err [ppm]	rdb	N-Rule	ej‡ Conf	mSig ma	Std I	Std I	Std I	Std m/z	Std VarN orm	Std m/z Diff	Std Com b Dev
373.1 676	1	C 25 H 22 N 2 Na	373.1 675	-0.2	-0.2	15.5	ok	even	1.79	0.002 5	0.000 1	0.001 1	0.000 0	0.842 7		

Mass Spectrum SmartFormula Report

Analysis Info

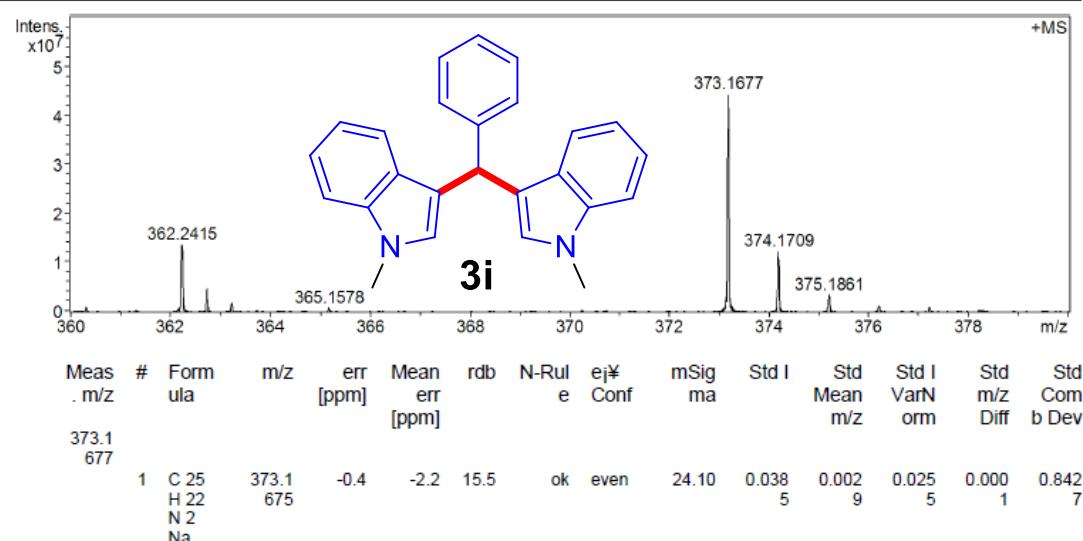
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 Method Student_LCms-140618
 Sample Name Student_4-6
 Comment

Acquisition Date 2014-7-2 17:11:55

Operator
 Instrument apex-Ultra

Acquisition Parameter

Polarity	Positive	Source	ESI	No. of Laser Shots	20
Averaged Scans	2	No. of Cell Fills	1	Laser Power	51.0 %
Broadband Low Mass	100.3 m/z	End Plate	3500.0 V	MALDI Plate	300.0 V
Broadband High Mass	1200.0 m/z	Capillary Entrance	4000.0 V	Imaging Spot Diameter	2000.0 μ m
Acquisition Mode	Single MS	Skimmer 1	20.0 V		
Pulse Program	basic	Drying Gas Temperature	180.0 °C	Calibration Date	Wed Jun 18 07:18:23
Source Accumulation	0.0 sec	Drying Gas Flow Rate	4.0 L/min	Data Acquisition Size	281072
Ion Accumulation Time	0.1 sec	Nebulizer Gas Flow Rate	1.0 L/min	Apodization	Sine-Bell Multiplication
Flight Time to Acq. Cell	0.0 sec				



Mass Spectrum SmartFormula Report

Analysis Info

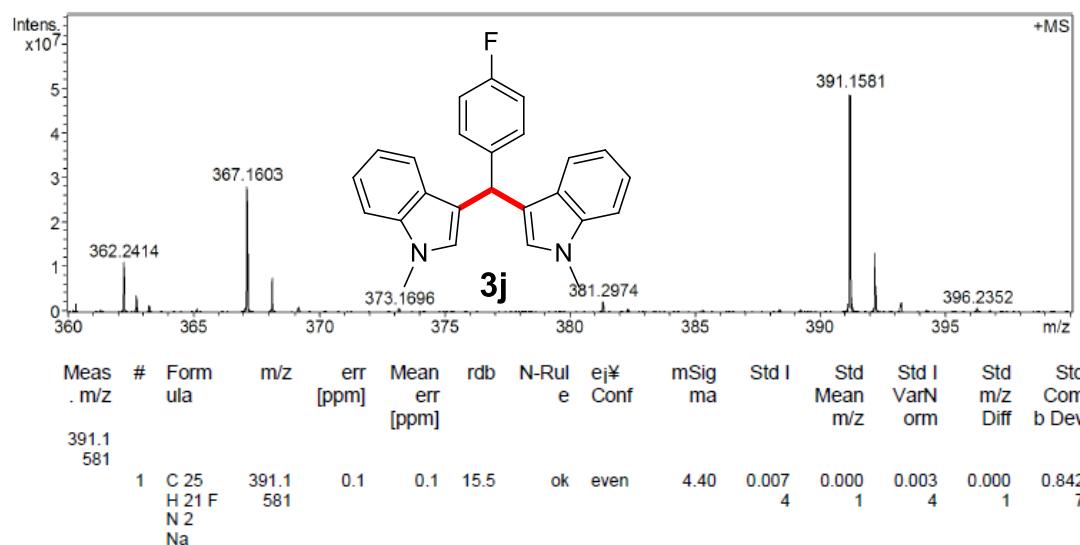
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 Method Student_LCms-140618
 Sample Name Student_4-6
 Comment

Acquisition Date 2014-7-2 17:12:46

 Operator
 Instrument apex-Ultra

Acquisition Parameter

Polarity	Positive	Source	ESI	No. of Laser Shots	20
Averaged Scans	2	No. of Cell Fills	1	Laser Power	51.0 %
Broadband Low Mass	100.3 m/z	End Plate	3500.0 V	MALDI Plate	300.0 V
Broadband High Mass	1200.0 m/z	Capillary Entrance	4000.0 V	Imaging Spot Diameter	2000.0 μm
Acquisition Mode	Single MS	Skimmer 1	20.0 V		
Pulse Program	basic	Drying Gas Temperature	180.0 °C	Calibration Date	Wed Jun 18 07:18:23
Source Accumulation	0.0 sec	Drying Gas Flow Rate	4.0 L/min	Data Acquisition Size	281972
Ion Accumulation Time	0.1 sec	Nebulizer Gas Flow Rate	1.0 L/min	Apodization	Sine-Bell Multiplication
Flight Time to Acq. Cell	0.0 sec				



Mass Spectrum SmartFormula Report

Analysis Info

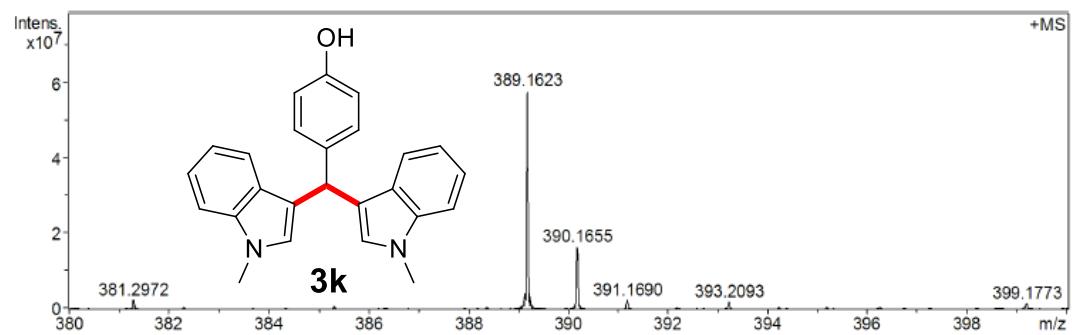
Analysis Name H:\20140702-1\JC\jc523_000002.d
 Method Student_LCms-140618
 Sample Name Student_4-6
 Comment

Acquisition Date 2014-7-2 17:13:57

Operator
 Instrument apex-Ultra

Acquisition Parameter

Polarity	Positive	Source	ESI	No. of Laser Shots	20
Averaged Scans	2	No. of Cell Fills	1	Laser Power	51.0 %
Broadband Low Mass	100.3 m/z	End Plate	3500.0 V	MALDI Plate	300.0 V
Broadband High Mass	1200.0 m/z	Capillary Entrance	4000.0 V	Imaging Spot Diameter	2000.0 μ m
Acquisition Mode	Single MS	Skimmer 1	20.0 V		
Pulse Program	basic	Drying Gas Temperature	180.0 °C	Calibration Date	Wed Jun 18 07:18:23
Source Accumulation	0.0 sec	Drying Gas Flow Rate	4.0 L/min	Data Acquisition Size	201072
Ion Accumulation Time	0.1 sec	Nebulizer Gas Flow Rate	1.0 L/min	Apodization	Sine-Bell Multiplication
Flight Time to Acq. Cell	0.0 sec				



Meas . m/z	#	Form u la	m/z	err [ppm]	Mean err [ppm]	rdb	N-Rule	eJ% Conf	mSig ma	Std I	Std I	Std m/z	Std VarN orm	Std m/z Diff	Std Com b Dev
389.1 623	1	C 25 H 22 N 2 Na O	389.1 624	0.3	0.4	15.5	ok	even	4.55	0.007 1	0.000 2	0.003 1	0.000 1	0.842 7	

Mass Spectrum SmartFormula Report

Analysis Info

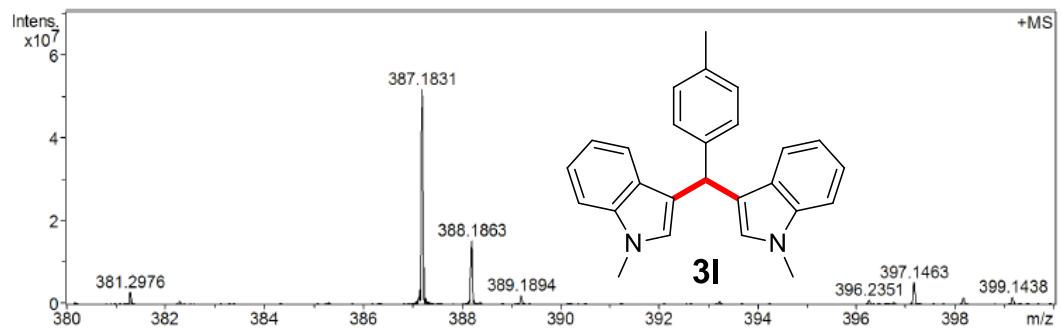
Analysis Name H:\20140702-1\JC\jc525_000002.d
 Method Student_LCms-140618
 Sample Name Student_4-6
 Comment

Acquisition Date 2014-7-2 17:15:08

 Operator
 Instrument apex-Ultra

Acquisition Parameter

Polarity	Positive	Source	ESI	No. of Laser Shots	20
Averaged Scans	2	No. of Cell Fills	1	Laser Power	51.0 %
Broadband Low Mass	100.3 m/z	End Plate	3500.0 V	MALDI Plate	300.0 V
Broadband High Mass	1200.0 m/z	Capillary Entrance	4000.0 V	Imaging Spot Diameter	2000.0 μ m
Acquisition Mode	Single MS	Skimmer 1	20.0 V		
Pulse Program	basic	Drying Gas Temperature	180.0 °C	Calibration Date	Wed Jun 18 07:18:23
Source Accumulation	0.0 sec	Drying Gas Flow Rate	4.0 L/min	Data Acquisition Size	201072
Ion Accumulation Time	0.1 sec	Nebulizer Gas Flow Rate	1.0 L/min	Apodization	Sine-Bell Multiplication
Flight Time to Acq. Cell	0.0 sec				



Meas . m/z	#	Form ula	m/z	err [ppm]	Mean err [ppm]	rdb	N-Rul e	ej‡ Conf	mSig ma	Std I	Std I	Std I	Std m/z	Std VarN orm	Std m/z Diff	Std Com b Dev
387.1 831	1	C 26 H 24 N 2 Na	387.1 832	0.2	0.3	15.5	ok	even	3.95	0.005 2	0.000 1	0.002 5	0.000 1	0.842 7		

Mass Spectrum SmartFormula Report

Analysis Info

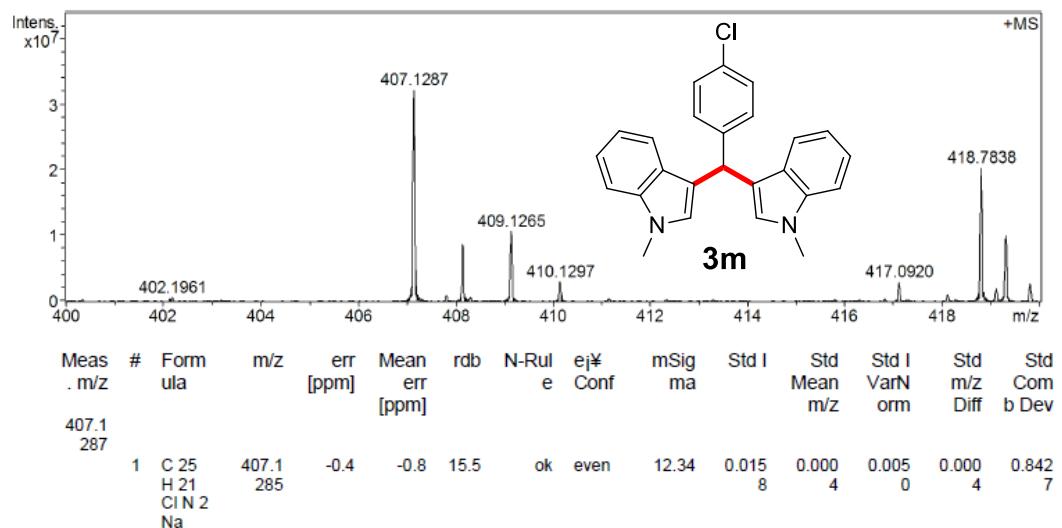
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 Method Student_LCms-140618
 Sample Name Student_4-6
 Comment

Acquisition Date 2014-7-2 17:16:24

 Operator
 Instrument apex-Ultra

Acquisition Parameter

Polarity	Positive	Source	ESI	No. of Laser Shots	20
Averaged Scans	2	No. of Cell Fills	1	Laser Power	51.0 %
Broadband Low Mass	100.3 m/z	End Plate	3500.0 V	MALDI Plate	300.0 V
Broadband High Mass	1200.0 m/z	Capillary Entrance	4000.0 V	Imaging Spot Diameter	2000.0 μ m
Acquisition Mode	Single MS	Skimmer 1	20.0 V		
Pulse Program	basic	Drying Gas Temperature	180.0 °C	Calibration Date	Wed Jun 18 07:18:23
Source Accumulation	0.0 sec	Drying Gas Flow Rate	4.0 L/min	Data Acquisition Size	281072
Ion Accumulation Time	0.1 sec	Nebulizer Gas Flow Rate	1.0 L/min	Apodization	Sine-Bell Multiplication
Flight Time to Acq. Cell	0.0 sec				



Mass Spectrum SmartFormula Report

Analysis Info

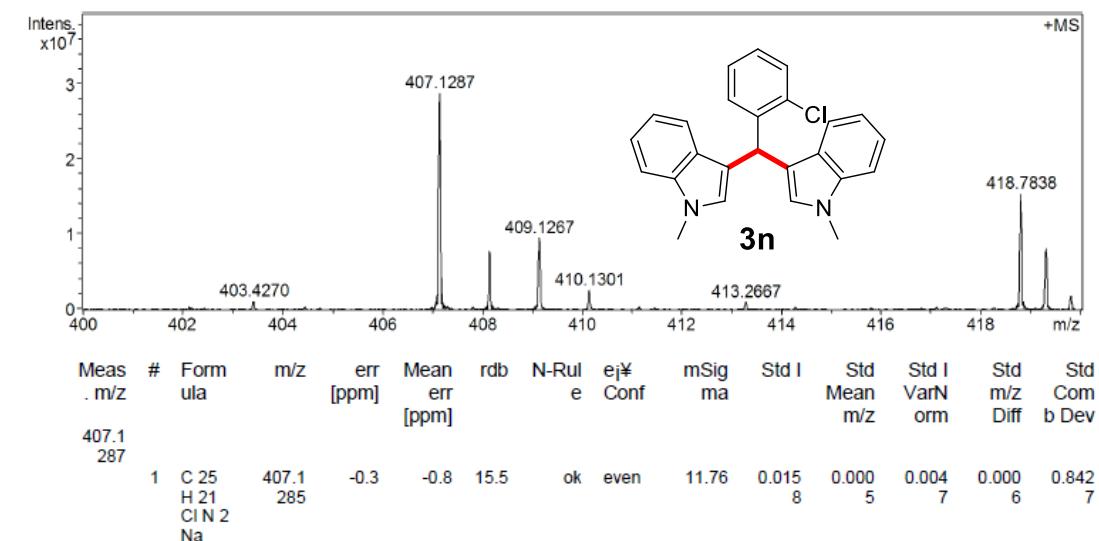
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 Sample Name Student_4-6
 Comment

Acquisition Date 2014-7-2 17:17:21

 Operator
 Instrument apex-Ultra

Acquisition Parameter

Polarity	Positive	Source	ESI	No. of Laser Shots	20
Averaged Scans	2	No. of Cell Fills	1	Laser Power	51.0 %
Broadband Low Mass	100.3 m/z	End Plate	3500.0 V	MALDI Plate	300.0 V
Broadband High Mass	1200.0 m/z	Capillary Entrance	4000.0 V	Imaging Spot Diameter	2000.0 μ m
Acquisition Mode	Single MS	Skimmer 1	20.0 V	Calibration Date	Wed Jun 18 07:18:23
Pulse Program	basic	Drying Gas Temperature	180.0 °C	Data Acquisition Size	201072
Source Accumulation	0.0 sec	Drying Gas Flow Rate	4.0 L/min	Apodization	Sine-Bell Multiplication
Ion Accumulation Time	0.1 sec	Nebulizer Gas Flow Rate	1.0 L/min		
Flight Time to Acq. Cell	0.0 sec				



Mass Spectrum SmartFormula Report

Analysis Info

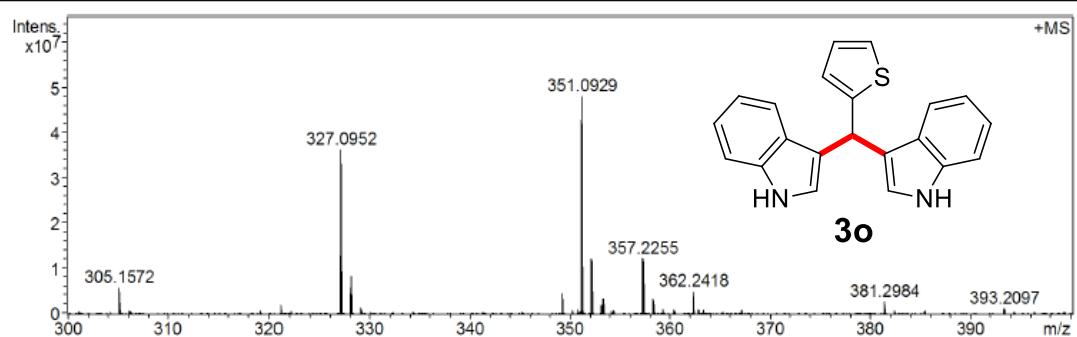
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 Sample Name Student_4-6
 Comment

Acquisition Date 2014-7-2 17:05:57

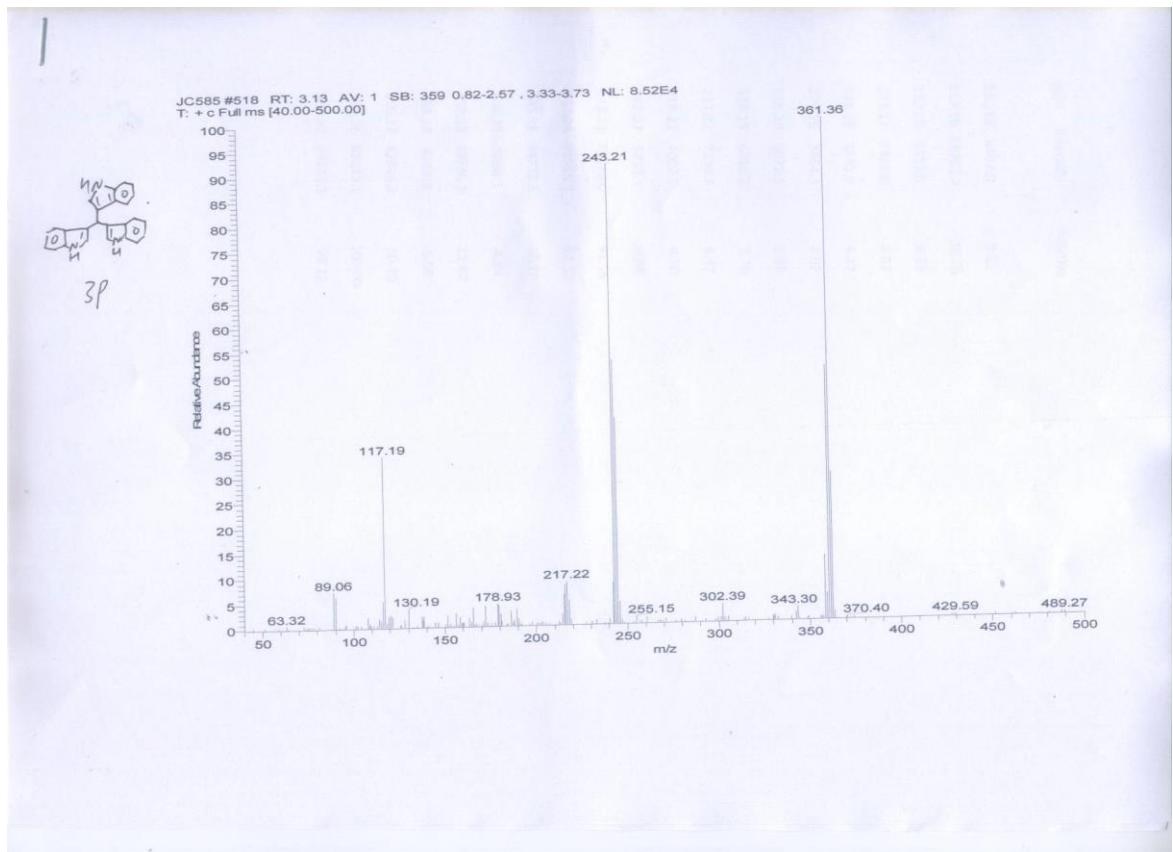
 Operator
 Instrument apex-Ultra

Acquisition Parameter

Polarity	Positive	Source	ESI	No. of Laser Shots	20
Averaged Scans	2	No. of Cell Fills	1	Laser Power	51.0 %
Broadband Low Mass	100.3 m/z	End Plate	3500.0 V	MALDI Plate	300.0 V
Broadband High Mass	1200.0 m/z	Capillary Entrance	4000.0 V	Imaging Spot Diameter	2000.0 μ m
Acquisition Mode	Single MS	Skimmer 1	20.0 V		
Pulse Program	basic	Drying Gas Temperature	180.0 °C	Calibration Date	Wed Jun 18 07:18:23
Source Accumulation	0.0 sec	Drying Gas Flow Rate	4.0 L/min	Data Acquisition Size	201072
Ion Accumulation Time	0.1 sec	Nebulizer Gas Flow Rate	1.0 L/min	Apodization	Sine-Bell Multiplication
Flight Time to Acq. Cell	0.0 sec				



Meas . m/z	#	Form ula	m/z	err [ppm]	Mean err [ppm]	rdb	N-Rul e	ej% Conf	mSig ma	Std I Std m/z	Std I Mean m/z	Std I VarN orm	Std m/z Diff	Std Com b Dev
351.0 929	1	C 21 H 16 N 2 Na S	351.0 926	-0.8	-0.6	14.5	ok	even	17.51	0.030 7	0.000 3	0.014 1	0.000 4	0.842 7



Mass Spectrum SmartFormula Report

Analysis Info

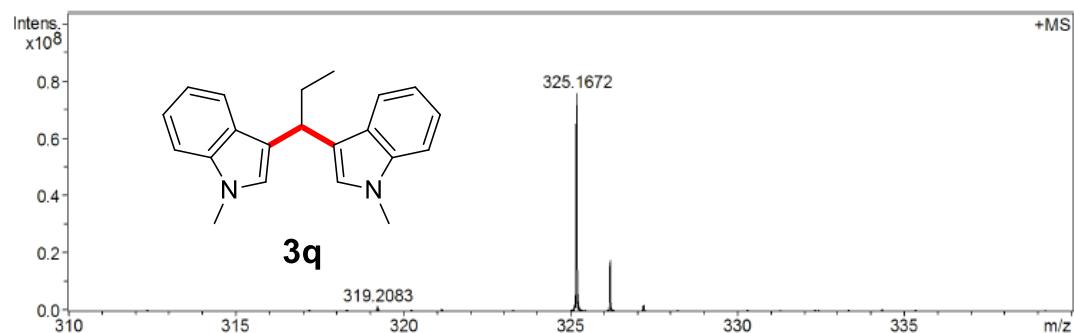
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 Method Student_LCms-140618
 Sample Name Student_4-6
 Comment

Acquisition Date 2014-7-2 17:45:35

 Operator
 Instrument apex-Ultra

Acquisition Parameter

Polarity	Positive	Source	ESI	No. of Laser Shots	20
Averaged Scans	2	No. of Cell Fills	1	Laser Power	51.0 %
Broadband Low Mass	100.3 m/z	End Plate	3500.0 V	MALDI Plate	300.0 V
Broadband High Mass	1200.0 m/z	Capillary Entrance	4000.0 V	Imaging Spot Diameter	2000.0 μ m
Acquisition Mode	Single MS	Skimmer 1	20.0 V		
Pulse Program	basic	Drying Gas Temperature	180.0 °C	Calibration Date	Wed Jun 18 07:18:23
Source Accumulation	0.0 sec	Drying Gas Flow Rate	4.0 L/min	Data Acquisition Size	281072
Ion Accumulation Time	0.1 sec	Nebulizer Gas Flow Rate	1.0 L/min	Apodization	Sine-Bell Multiplication
Flight Time to Acq. Cell	0.0 sec				



Meas . m/z	#	Form u la	m/z	err [ppm]	Mean err [ppm]	rdb	N-Rule	ej% Conf	mSig ma	Std I m/z	Std Mean m/z	Std VarN orm	Std m/z Diff	Std Com b Dev
325.1 672	1	C 21 H 22 N 2 Na	325.1 675	1.0	1.0	11.5	ok	even	1.85	0.003 4	0.000 3	0.001 6	0.000 0	0.842 7

Mass Spectrum SmartFormula Report

Analysis Info

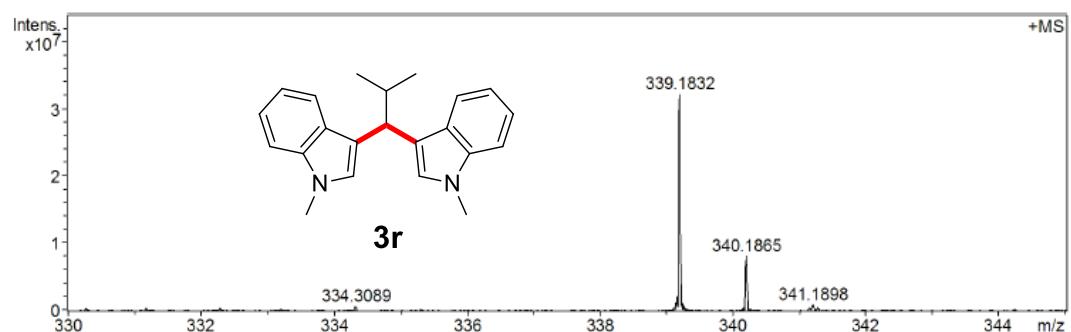
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 Sample Name Student_4-6
 Comment

Acquisition Date 2014-7-2 17:46:34

 Operator
 Instrument apex-Ultra

Acquisition Parameter

Polarity	Positive	Source	ESI	No. of Laser Shots	20
Averaged Scans	2	No. of Cell Fills	1	Laser Power	51.0 %
Broadband Low Mass	100.3 m/z	End Plate	3500.0 V	MALDI Plate	300.0 V
Broadband High Mass	1200.0 m/z	Capillary Entrance	4000.0 V	Imaging Spot Diameter	2000.0 μm
Acquisition Mode	Single MS	Skimmer 1	20.0 V		
Pulse Program	basic	Drying Gas Temperature	180.0 °C	Calibration Date	Wed Jun 18 07:18:23
Source Accumulation	0.0 sec	Drying Gas Flow Rate	4.0 L/min	Data Acquisition Size	201072
Ion Accumulation Time	0.1 sec	Nebulizer Gas Flow Rate	1.0 L/min	Apodization	Sine-Bell Multiplication
Flight Time to Acq. Cell	0.0 sec				



Meas . m/z	#	Form ula	m/z	err [ppm]	Mean err [ppm]	rdb	N-Rule	eJ% Conf	mSig ma	Std I	Std I	Std I	Std m/z Diff	Std Com b Dev
339.1 832	1	C 22 H 24 N 2 Na	339.1 832	-0.1	-0.1	11.5	ok	even	4.22	0.006 2	0.000 0	0.002 7	0.000 0	0.842 7

Mass Spectrum SmartFormula Report

Analysis Info

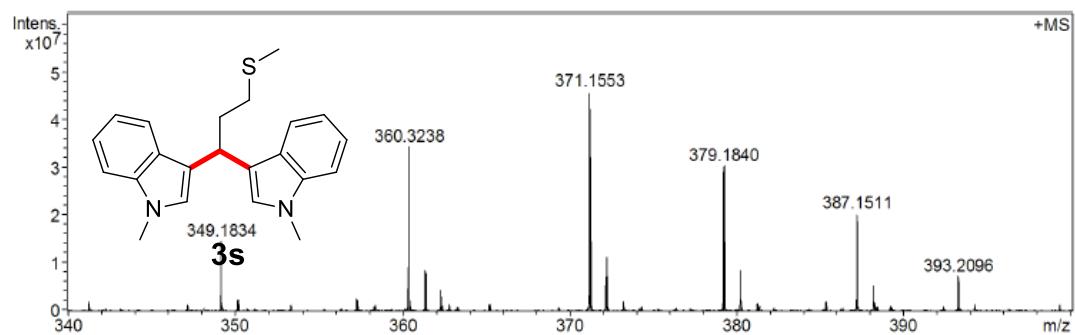
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 Sample Name Student_4-6
 Comment

Acquisition Date 2014-7-2 17:47:25

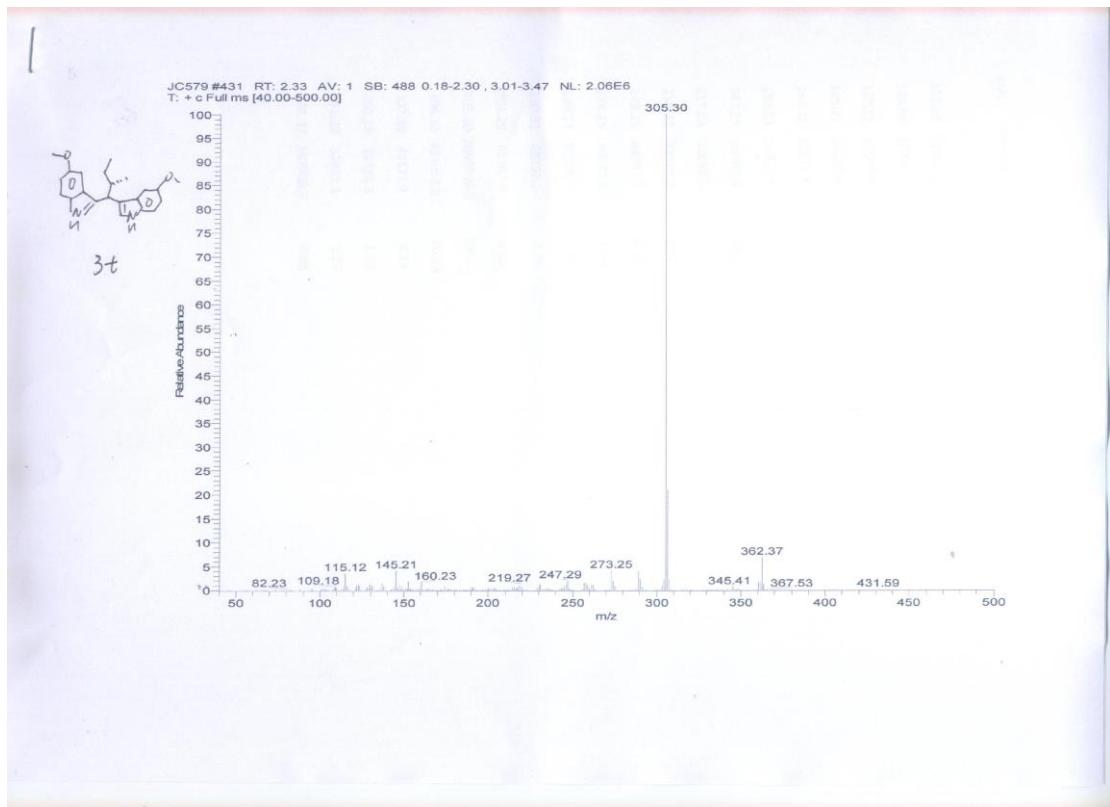
 Operator
 Instrument apex-Ultra

Acquisition Parameter

Polarity	Positive	Source	ESI	No. of Laser Shots	20
Averaged Scans	2	No. of Cell Fills	1	Laser Power	51.0 %
Broadband Low Mass	100.3 m/z	End Plate	3500.0 V	MALDI Plate	300.0 V
Broadband High Mass	1200.0 m/z	Capillary Entrance	4000.0 V	Imaging Spot Diameter	2000.0 μ m
Acquisition Mode	Single MS	Skimmer 1	20.0 V		
Pulse Program	basic	Drying Gas Temperature	180.0 °C	Calibration Date	Wed Jun 18 07:18:23
Source Accumulation	0.0 sec	Drying Gas Flow Rate	4.0 L/min	Data Acquisition Size	281072
Ion Accumulation Time	0.1 sec	Nebulizer Gas Flow Rate	1.0 L/min	Apodization	Sine-Bell Multiplication
Flight Time to Acq. Cell	0.0 sec				



Meas . m/z	#	Form u la	m/z	err [ppm]	Mean err [ppm]	rdb	N-Rul e	ej% Conf	mSig ma	Std I	Std Mean m/z	Std VarN orm	Std m/z Diff	Std Com b Dev
371.1553	1	C 22 H 24 N 2 Na S	371.1552	-0.1	-0.2	11.5	ok	even	17.03	0.0338	0.0003	0.0139	0.0010	0.8427



Mass Spectrum SmartFormula Report

Analysis Info

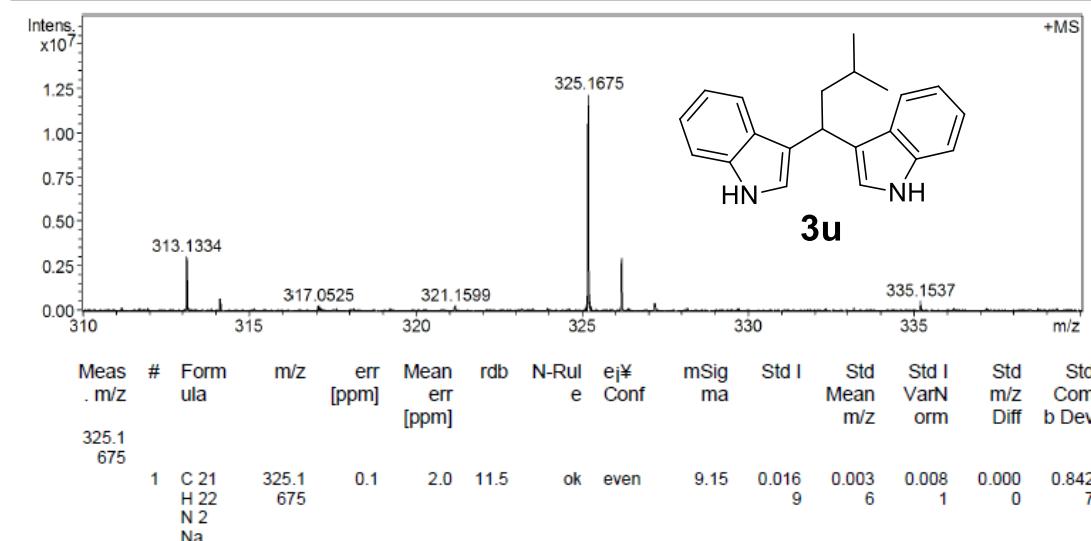
Analysis Name H:\20140702-1\JCjc550_000002.d
 Method Student_LCms-140618
 Sample Name Student_4-6
 Comment

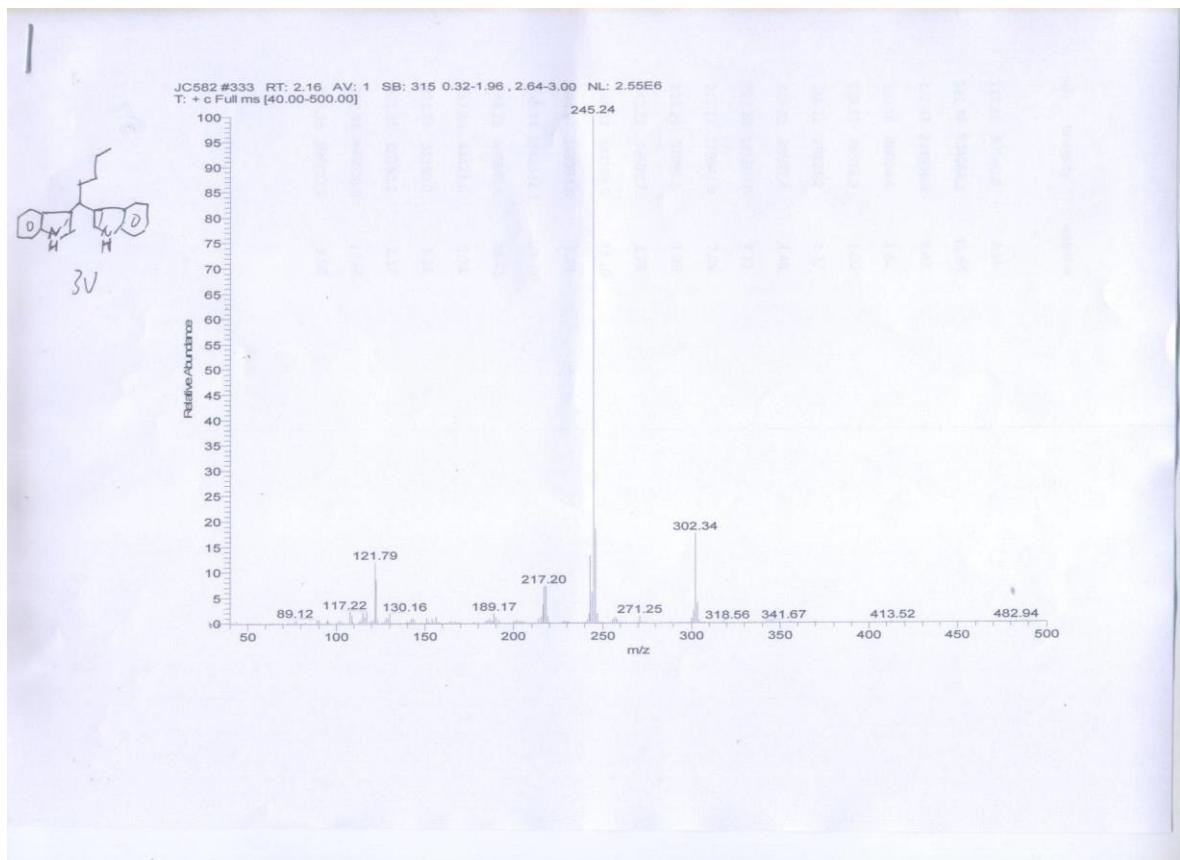
Acquisition Date 2014-7-2 17:51:30

 Operator
 Instrument apex-Ultra

Acquisition Parameter

Polarity	Positive	Source	ESI	No. of Laser Shots	20
Averaged Scans	2	No. of Cell Fills	1	Laser Power	51.0 %
Broadband Low Mass	100.3 m/z	End Plate	3500.0 V	MALDI Plate	300.0 V
Broadband High Mass	1200.0 m/z	Capillary Entrance	4000.0 V	Imaging Spot Diameter	2000.0 μ m
Acquisition Mode	Single MS	Skimmer 1	20.0 V		
Pulse Program	basic	Drying Gas Temperature	180.0 °C	Calibration Date	Wed Jun 18 07:18:23
Source Accumulation	0.0 sec	Drying Gas Flow Rate	4.0 L/min	Data Acquisition Size	201072
Ion Accumulation Time	0.1 sec	Nebulizer Gas Flow Rate	1.0 L/min	Apodization	Sine-Bell Multiplication
Flight Time to Acq. Cell	0.0 sec				





Mass Spectrum SmartFormula Report

Analysis Info

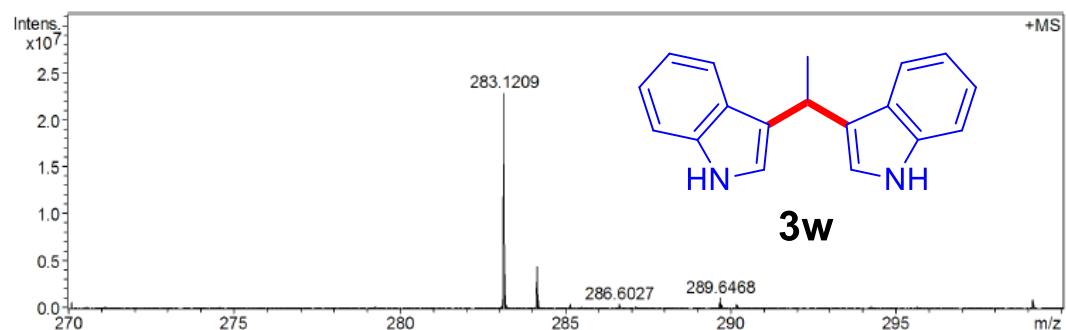
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 Method: Student_LCms-140618
 Sample Name: Student_4-6
 Comment:

Acquisition Date: 2014-7-2 17:53:02

Operator: Instrument: apex-Ultra

Acquisition Parameter

Polarity	Positive	Source	ESI	No. of Laser Shots	20
Averaged Scans	2	No. of Cell Fills	1	Laser Power	51.0 %
Broadband Low Mass	100.3 m/z	End Plate	3500.0 V	MALDI Plate	300.0 V
Broadband High Mass	1200.0 m/z	Capillary Entrance	4000.0 V	Imaging Spot Diameter	2000.0 μm
Acquisition Mode	Single MS	Skimmer 1	20.0 V		
Pulse Program	basic	Drying Gas Temperature	180.0 °C	Calibration Date	Wed Jun 18 07:18:23
Source Accumulation	0.0 sec	Drying Gas Flow Rate	4.0 L/min	Data Acquisition Size	281072
Ion Accumulation Time	0.1 sec	Nebulizer Gas Flow Rate	1.0 L/min	Apodization	Sine-Bell Multiplication
Flight Time to Acq. Cell	0.0 sec				



Meas. m/z	#	Form ula	m/z	err [ppm]	Mean err [ppm]	rdb	N-Rule	eJ \pm Conf	mSig ma	Std I m/z	Std Mean m/z	Std I VarN orm	Std m/z Diff	Std Com b Dev
283.1 209	1	C 18 H 16 N 2 Na	283.1 206	-1.3	-1.5	11.5	ok	even	4.50	0.008 6	0.000 4	0.003 6	0.000 2	0.842 7

Mass Spectrum SmartFormula Report

Analysis Info

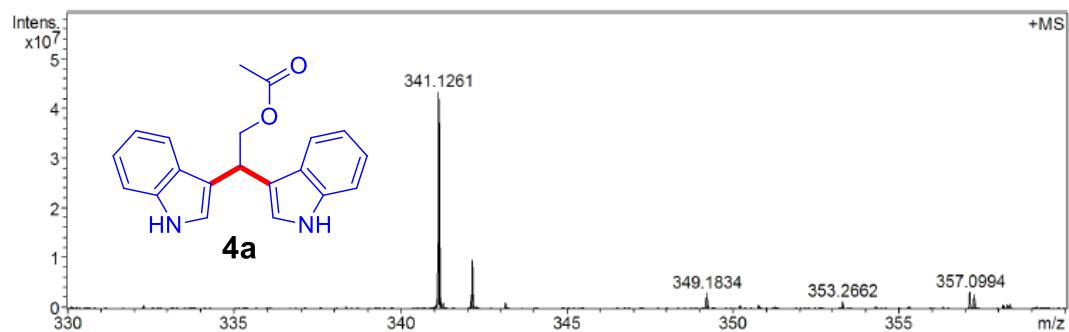
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 Method Student_LCms-140618
 Sample Name Student_4-6
 Comment

Acquisition Date 2014-7-2 17:53:55

 Operator
 Instrument apex-Ultra

Acquisition Parameter

Polarity	Positive	Source	ESI	No. of Laser Shots	20
Averaged Scans	2	No. of Cell Fills	1	Laser Power	51.0 %
Broadband Low Mass	100.3 m/z	End Plate	3500.0 V	MALDI Plate	300.0 V
Broadband High Mass	1200.0 m/z	Capillary Entrance	4000.0 V	Imaging Spot Diameter	2000.0 μm
Acquisition Mode	Single MS	Skimmer 1	20.0 V		
Pulse Program	basic	Drying Gas Temperature	180.0 °C	Calibration Date	Wed Jun 18 07:18:23
Source Accumulation	0.0 sec	Drying Gas Flow Rate	4.0 L/min	Data Acquisition Size	201072
Ion Accumulation Time	0.1 sec	Nebulizer Gas Flow Rate	1.0 L/min	Apodization	Sine-Bell Multiplication
Flight Time to Acq. Cell	0.0 sec				



Meas . m/z	#	Form ula	m/z	err [ppm]	Mean err [ppm]	rdb	N-Rule	eJ% Conf	mSig ma	Std I m/z	Std Mean m/z	Std VarN orm	Std m/z Diff	Std Com b Dev
341.1261	1	C ₂₀ H ₁₈ N ₂ O ₂	341.1260	-0.2	-0.1	12.5	ok	even	2.33	0.0045	0.0001	0.0026	0.0000	0.8427

Mass Spectrum SmartFormula Report

Analysis Info

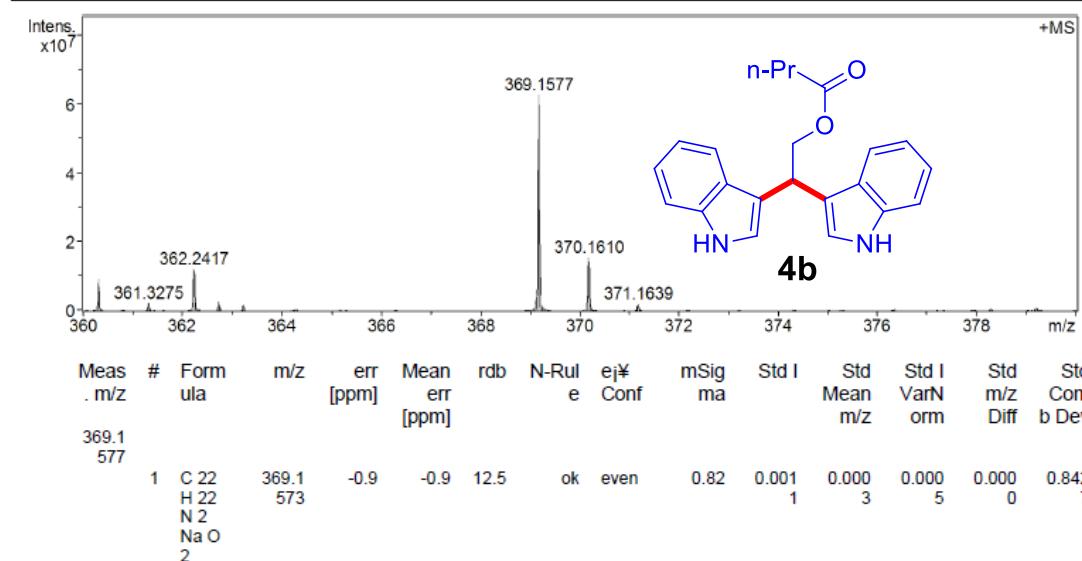
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 Sample Name Student_4-6
 Comment

Acquisition Date 2014-7-2 17:54:43

 Operator
 Instrument apex-Ultra

Acquisition Parameter

Polarity	Positive	Source	ESI	No. of Laser Shots	20
Averaged Scans	2	No. of Cell Fills	1	Laser Power	51.0 %
Broadband Low Mass	100.3 m/z	End Plate	3500.0 V	MALDI Plate	300.0 V
Broadband High Mass	1200.0 m/z	Capillary Entrance	4000.0 V	Imaging Spot Diameter	2000.0 μm
Acquisition Mode	Single MS	Skimmer 1	20.0 V	Calibration Date	Wed Jun 18 07:18:23
Pulse Program	basic	Drying Gas Temperature	180.0 °C	Data Acquisition Size	201072
Source Accumulation	0.0 sec	Drying Gas Flow Rate	4.0 L/min	Apodization	Sine-Bell Multiplication
Ion Accumulation Time	0.1 sec	Nebulizer Gas Flow Rate	1.0 L/min		
Flight Time to Acq. Cell	0.0 sec				



Mass Spectrum SmartFormula Report

Analysis Info

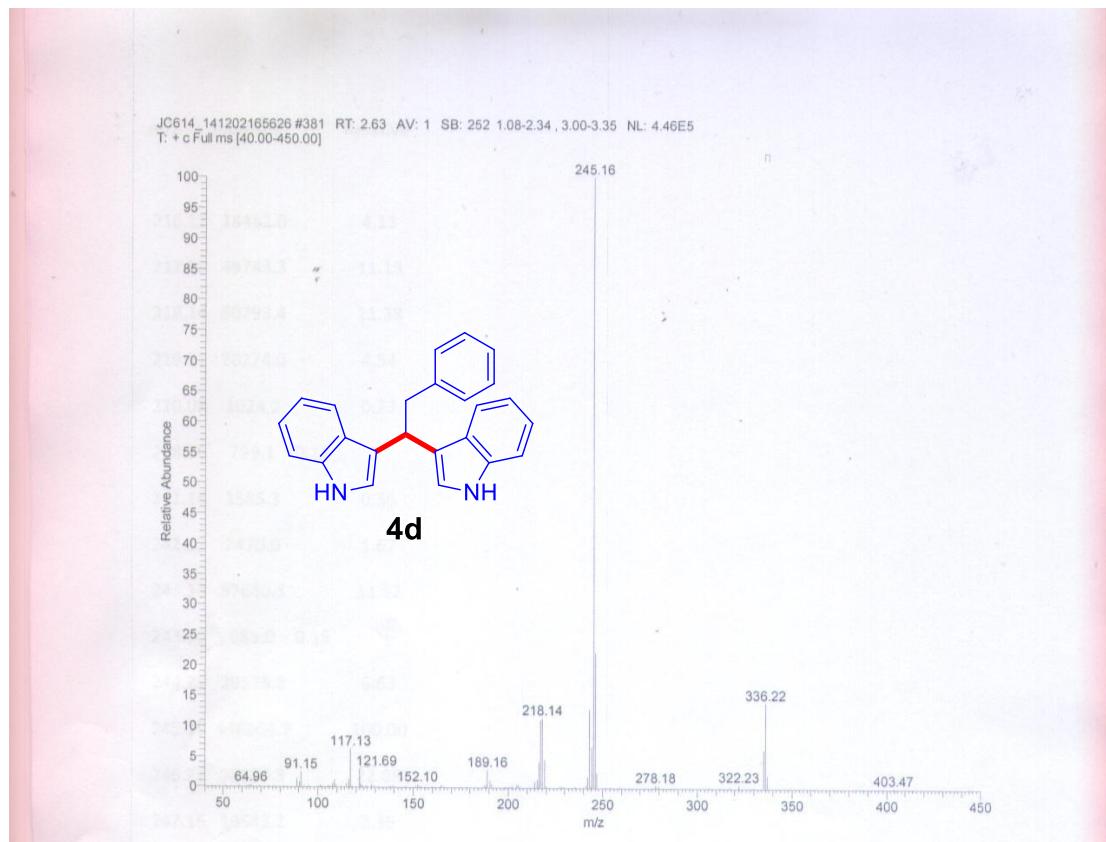
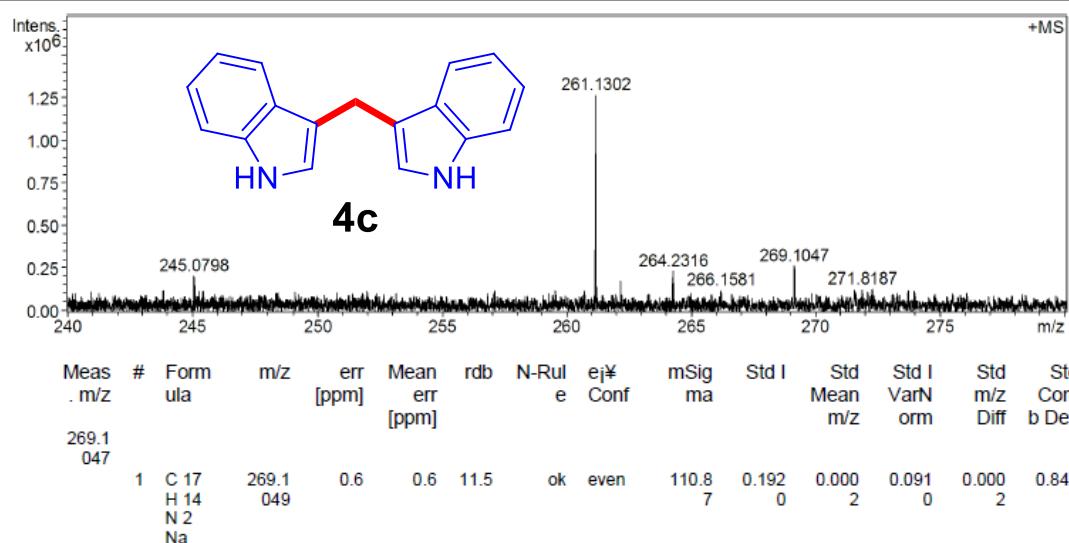
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 Method Student_LCms-140618
 Sample Name Student_4-6
 Comment

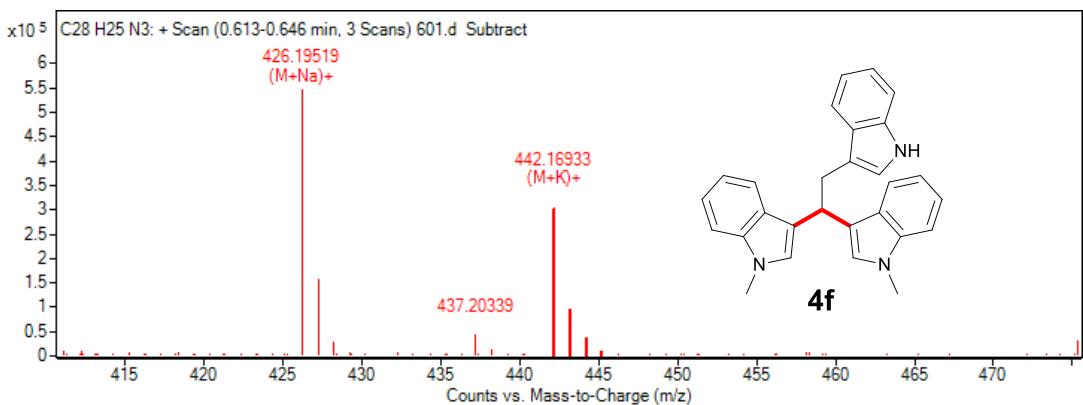
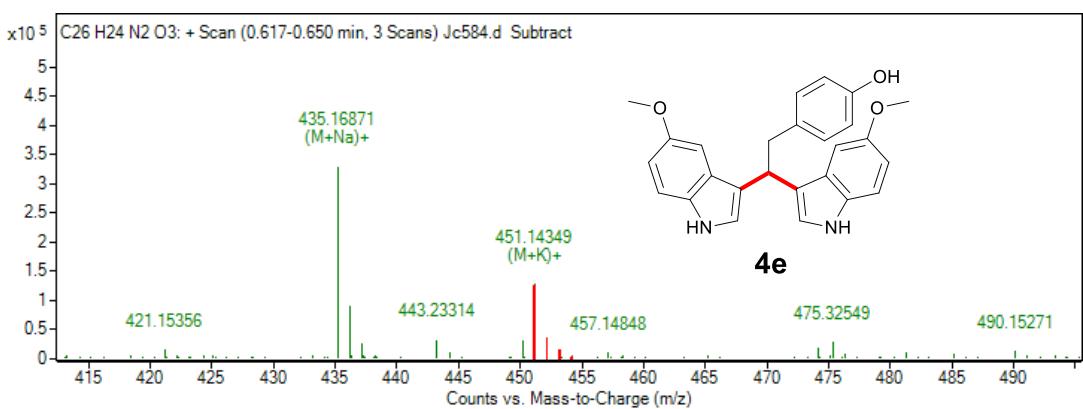
Acquisition Date 2014-7-2 17:48:59

 Operator
 Instrument apex-Ultra

Acquisition Parameter

Polarity	Positive	Source	ESI	No. of Laser Shots	20
Averaged Scans	2	No. of Cell Fills	1	Laser Power	51.0 %
Broadband Low Mass	100.3 m/z	End Plate	3500.0 V	MALDI Plate	300.0 V
Broadband High Mass	1200.0 m/z	Capillary Entrance	4000.0 V	Imaging Spot Diameter	2000.0 μ m
Acquisition Mode	Single MS	Skimmer 1	20.0 V		
Pulse Program	basic	Drying Gas Temperature	180.0 °C	Calibration Date	Wed Jun 18 07:18:23
Source Accumulation	0.0 sec	Drying Gas Flow Rate	4.0 L/min	Data Acquisition Size	201072
Ion Accumulation Time	0.1 sec	Nebulizer Gas Flow Rate	1.0 L/min	Apodization	Sine-Bell Multiplication
Flight Time to Acq. Cell	0.0 sec				





Mass Spectrum SmartFormula Report

Analysis Info

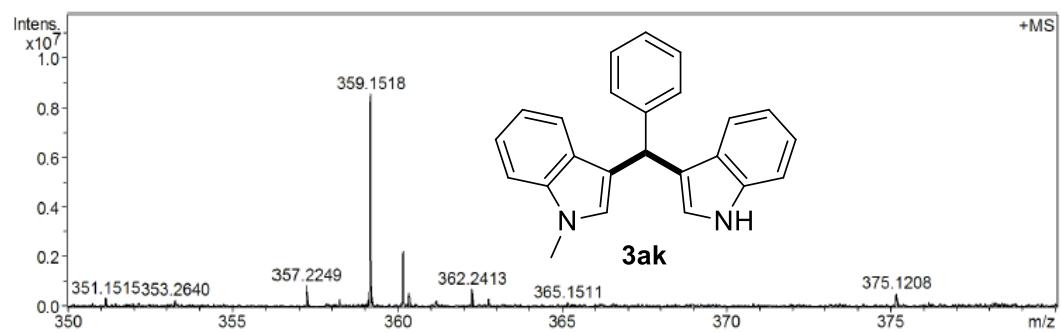
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 Method Student_LCms-140618
 Sample Name Student_4-6
 Comment

Acquisition Date 2014-7-2 17:58:08

 Operator
 Instrument apex-Ultra

Acquisition Parameter

Polarity	Positive	Source	ESI	No. of Laser Shots	20
Averaged Scans	2	No. of Cell Fills	1	Laser Power	51.0 %
Broadband Low Mass	100.3 m/z	End Plate	3500.0 V	MALDI Plate	300.0 V
Broadband High Mass	1200.0 m/z	Capillary Entrance	4000.0 V	Imaging Spot Diameter	2000.0 μm
Acquisition Mode	Single MS	Skimmer 1	20.0 V		
Pulse Program	basic	Drying Gas Temperature	180.0 °C	Calibration Date	Wed Jun 18 07:18:23
Source Accumulation	0.0 sec	Drying Gas Flow Rate	4.0 L/min	Data Acquisition Size	201072
Ion Accumulation Time	0.0 sec	Nebulizer Gas Flow Rate	1.0 L/min	Apodization	Sine-Bell Multiplication
Flight Time to Acq. Cell	0.0 sec				



Meas . m/z	#	Form ula	m/z	err [ppm]	Mean err [ppm]	rdb	N-Rule	ej Conf	mSig ma	Std I	Std Mean m/z	Std I VarN orm	Std m/z Diff	Std Com b Dev
359.1518	1	C 24 H 20 N 2 Na	359.1519	0.2	0.3	15.5	ok	even	4.93	0.0086	0.0002	0.0040	0.0001	0.8427

Mass Spectrum SmartFormula Report

Analysis Info

Analysis Name H:\20140702-1\JC\jc581_000002.d
 Method Student_LCms-140618
 Sample Name Student_4-6
 Comment

Acquisition Date 2014-7-2 17:56:46

 Operator
 Instrument apex-Ultra

Acquisition Parameter

Polarity	Positive	Source	ESI	No. of Laser Shots	20
Averaged Scans	2	No. of Cell Fills	1	Laser Power	51.0 %
Broadband Low Mass	100.3 m/z	End Plate	3500.0 V	MALDI Plate	300.0 V
Broadband High Mass	1200.0 m/z	Capillary Entrance	4000.0 V	Imaging Spot Diameter	2000.0 μm
Acquisition Mode	Single MS	Skimmer 1	20.0 V		
Pulse Program	basic	Drying Gas Temperature	180.0 °C	Calibration Date	Wed Jun 18 07:18:23
Source Accumulation	0.0 sec	Drying Gas Flow Rate	4.0 L/min	Data Acquisition Size	201072
Ion Accumulation Time	0.1 sec	Nebulizer Gas Flow Rate	1.0 L/min	Apodization	Sine-Bell Multiplication
Flight Time to Acq. Cell	0.0 sec				

