

## Electronic Supporting Information (ESI)

### **Discovery of 4-acetyl-3-(4-fluorophenyl)-1-(p-tolyl)-5-methylpyrrole as a dual inhibitor of human P-glycoprotein and *Staphylococcus aureus* Nor A efflux pump<sup>□</sup>**

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## CONTENTS

Content	Page no.
<b>S1.</b> General procedure for synthesis of pyrroles <b>5a-q</b>	S2
<b>S2.</b> Spectral data of representative compounds	S3
<b>S3.</b> <sup>1</sup> H, <sup>13</sup> C, DEPT135, HSQC, and HMBC spectra scans of compound <b>5q</b>	S4
<b>S4.</b> <sup>1</sup> H, <sup>13</sup> C, DEPT135, <sup>19</sup> F and HMBC NMR spectra scans of compound <b>5i</b>	S7
<b>S5.</b> HMBC correlations and structure assignments for compound <b>5q</b>	S10
<b>S6.</b> Pharmacokinetic parameters of compound <b>5i</b>	S12

## S1. General procedure for synthesis of pyrroles 5a-q.

To the stirred solution of aniline (**2a**, 0.132 g, 1.42 mmol), benzaldehyde (**3a**, 0.1 g, 0.94 mmol), and acetylacetone (**4a**, 0.094 g, 0.94 mmol) in nitromethane (**5a**, 1.1 ml, 20 mmol) was added 10 mol% montmorillonite clay K10 or clay KSF catalyst. The mixture was refluxed for 5-8 h and then cooled to room temperature. The excess solvent was removed under vacuum, and the residue was purified by silica gel (#100-200) column chromatography using EtOAc: n-hexane (95:5) to get pyrroles **7a-q** in 68-88% yield. Pyrroles **1a-1d** and **1m** were characterized by comparison of their spectral data with literature values.<sup>1</sup>

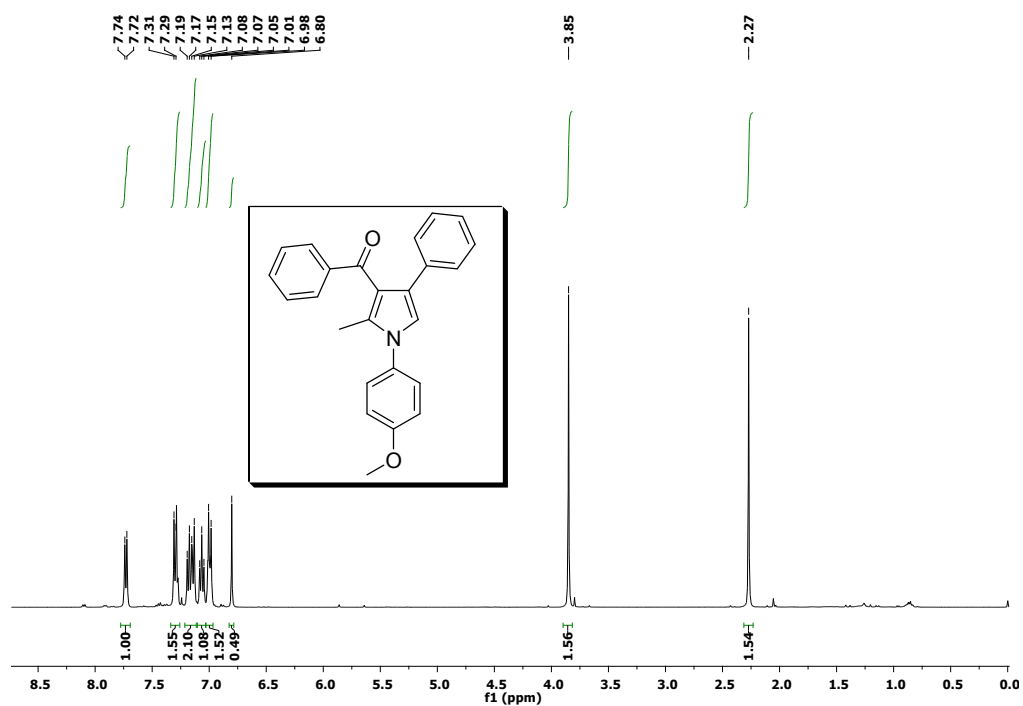
## S2. Spectral data of representative compounds:

**1-(4-(4-Fluorophenyl)-2-methyl-1-p-tolyl-1H-pyrrol-3-yl)ethanone (5i):** Yellow Solid; m.p. 109-110 °C; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz): δ 7.33 (t, *J* = 8.0 Hz, 2H), 7.29 (d, *J* = 8.0 Hz, 2H), 7.21 (d, *J* = 8.0 Hz, 2H), 7.06 (t, *J* = 8.0 Hz, 2H), 6.61 (s, 1H), 2.42 (s, 3H), 2.39 (s, 3H), 2.07 (s, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz): δ 197.2, 163.1 (d, <sup>1</sup>*J*<sub>CF</sub> = 230 Hz), 138.2, 136.1, 135.6, 132.1, 130.8, 130.0, 126.1, 125.1, 122.4, 120.8, 115.3 (d, <sup>2</sup>*J*<sub>CF</sub> = 53.8 Hz), 31.1, 21.1, 13.0; <sup>19</sup>F NMR (376.5 MHz, CDCl<sub>3</sub>): δ -116.0, -116.0 (m, 1F); IR (CHCl<sub>3</sub>): ν<sub>max</sub> 2918, 1650, 1553, 1418, 1384, 1275, 1220, 1041 cm<sup>-1</sup>; ESI-MS: *m/z* 308.00 [M+H]<sup>+</sup>; HR-ESIMS: *m/z* 308.1449 calcd for C<sub>20</sub>H<sub>18</sub>FNO+H<sup>+</sup> (308.1445).

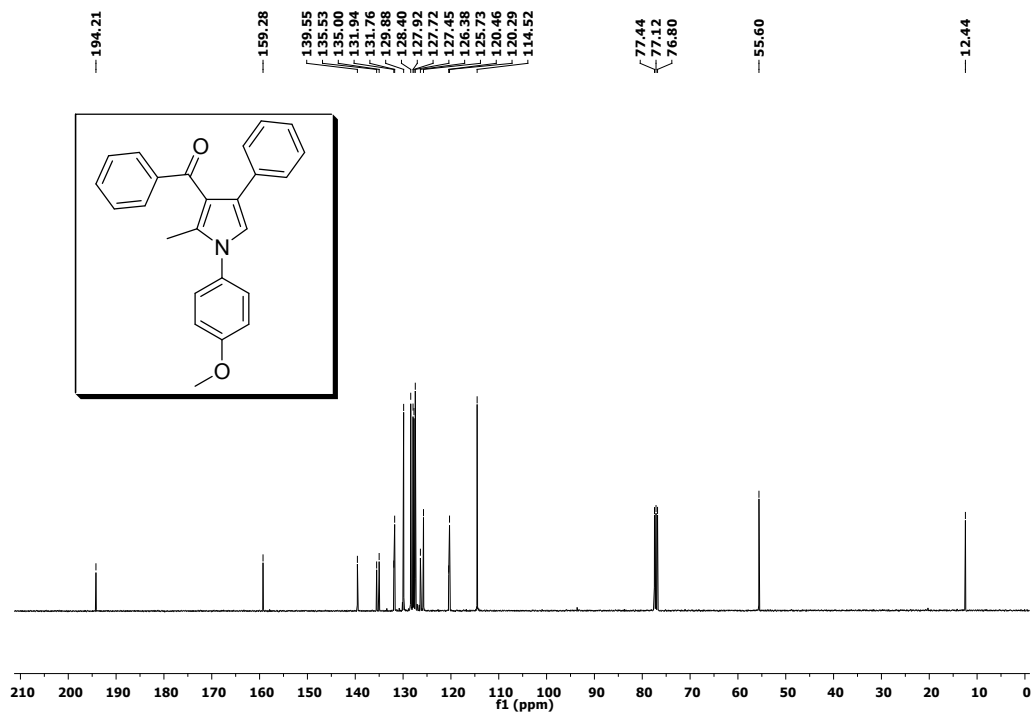
**(1-(4-Methoxyphenyl)-2-methyl-4-phenyl-1H-pyrrol-3-yl)(phenyl)methanone (5q):** Brown sticky solid; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz): δ 7.74 (d, *J* = 8.0 Hz, 2H), 7.31 (d, *J* = 8.0 Hz, 3H), 7.19-7.13 (m, 4H), 7.07 (t, *J* = 8.0 Hz, 2H), 7.01 (d, *J* = 12.0 Hz, 3H), 6.80 (s, 1H), 3.85 (s, 3H), 2.27 (s, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz): δ 194.2, 159.3, 157.9, 139.6, 135.5, 135.0, 131.9, 131.8, 129.9, 128.4, 127.9, 127.7, 127.5, 126.4, 125.7, 120.5, 120.3, 114.5, 55.6, 12.5; IR (CHCl<sub>3</sub>): ν<sub>max</sub> 2905, 1720, 1616, 1601, 1574, 1320, 1251 cm<sup>-1</sup>; ESI-MS: *m/z* 368.00 [M+H]<sup>+</sup>; HR-ESIMS: *m/z* 368.1648 calcd for C<sub>25</sub>H<sub>21</sub>NO<sub>2</sub>+H<sup>+</sup> (368.1645).

### S3. $^1\text{H}$ , $^{13}\text{C}$ , DEPT135 and HMBC NMR spectra scans of compound 5q

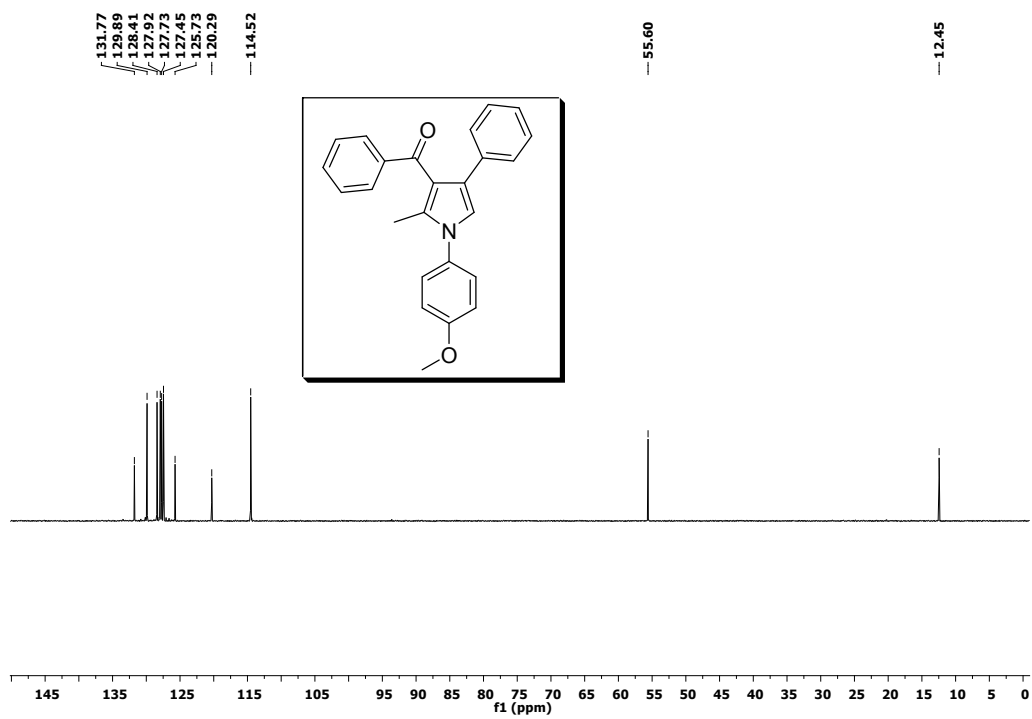
#### $^1\text{H}$ NMR



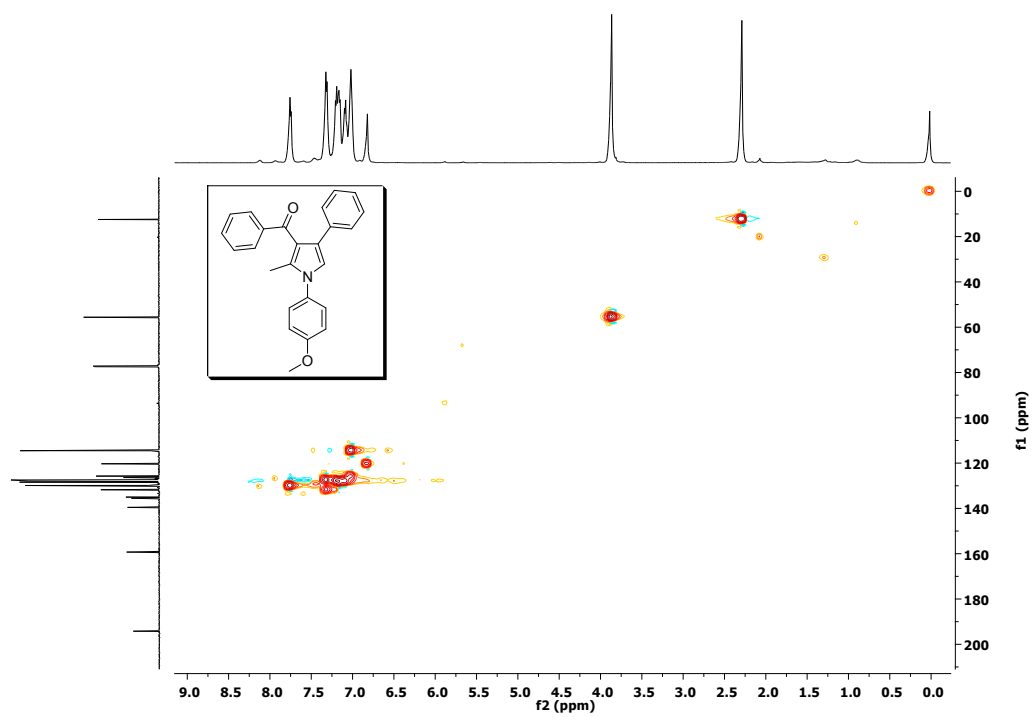
#### $^{13}\text{C}$ NMR



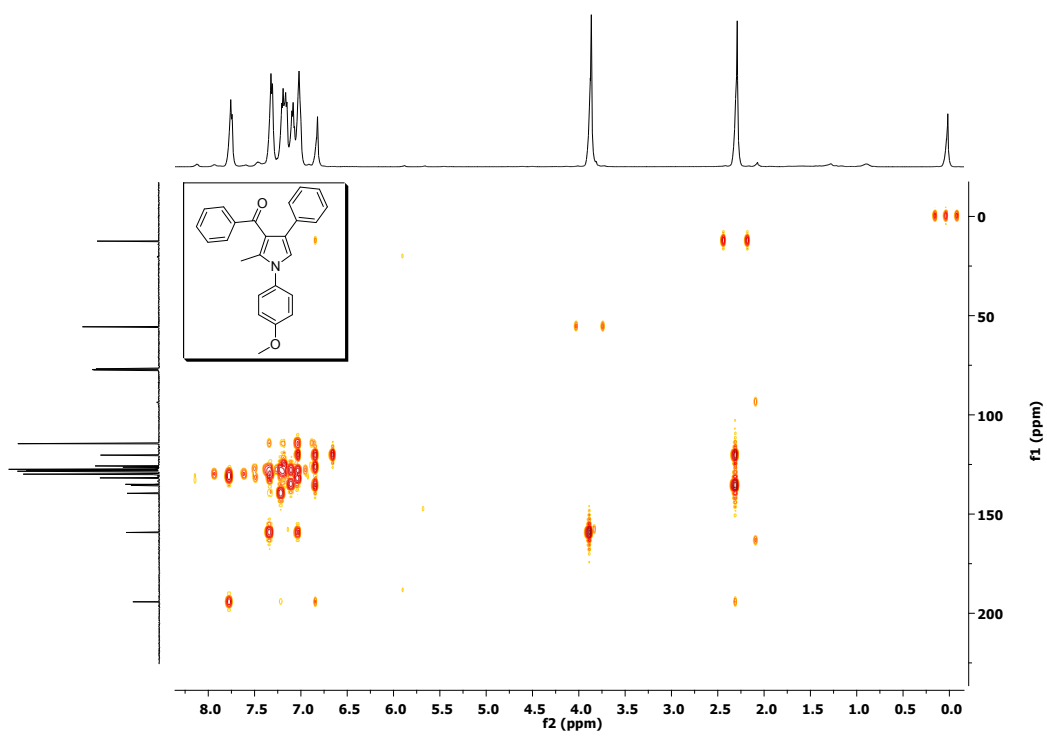
# DEPT



# HSQC

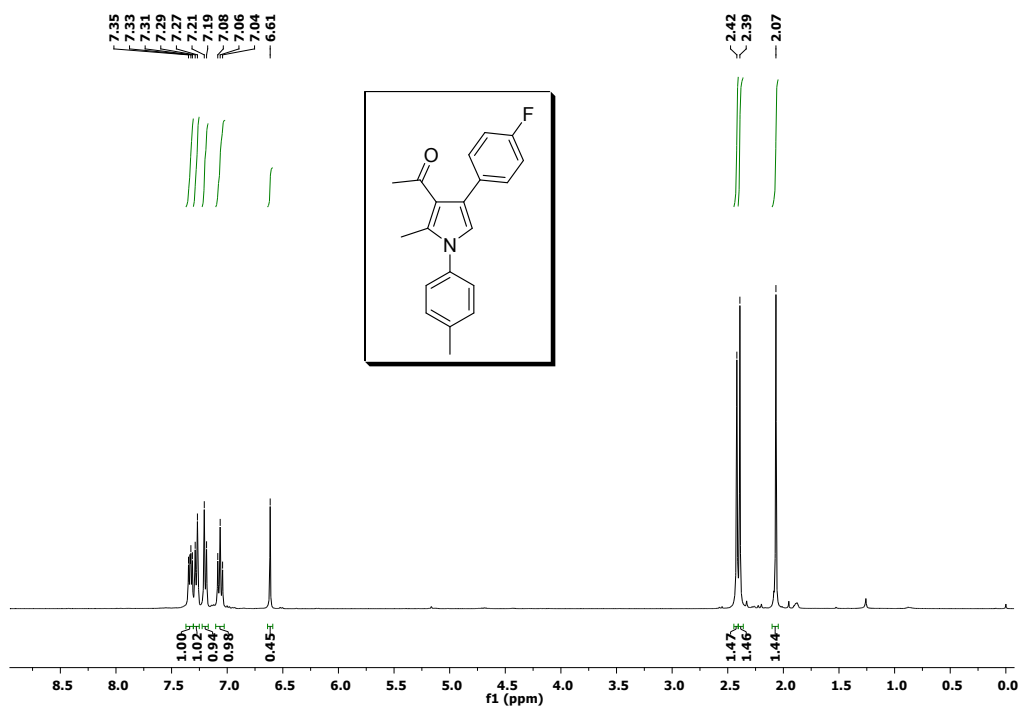


# HMBC

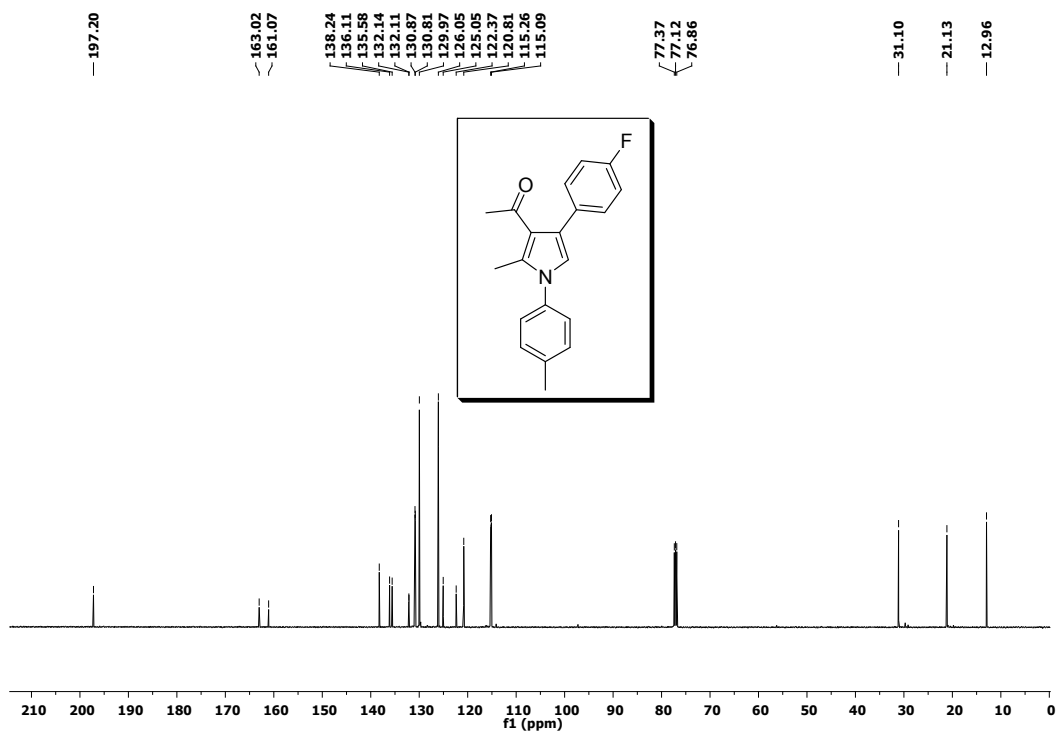


# S4. <sup>1</sup>H, <sup>13</sup>C, DEPT135 and HMBC NMR spectra scans of compound 5i

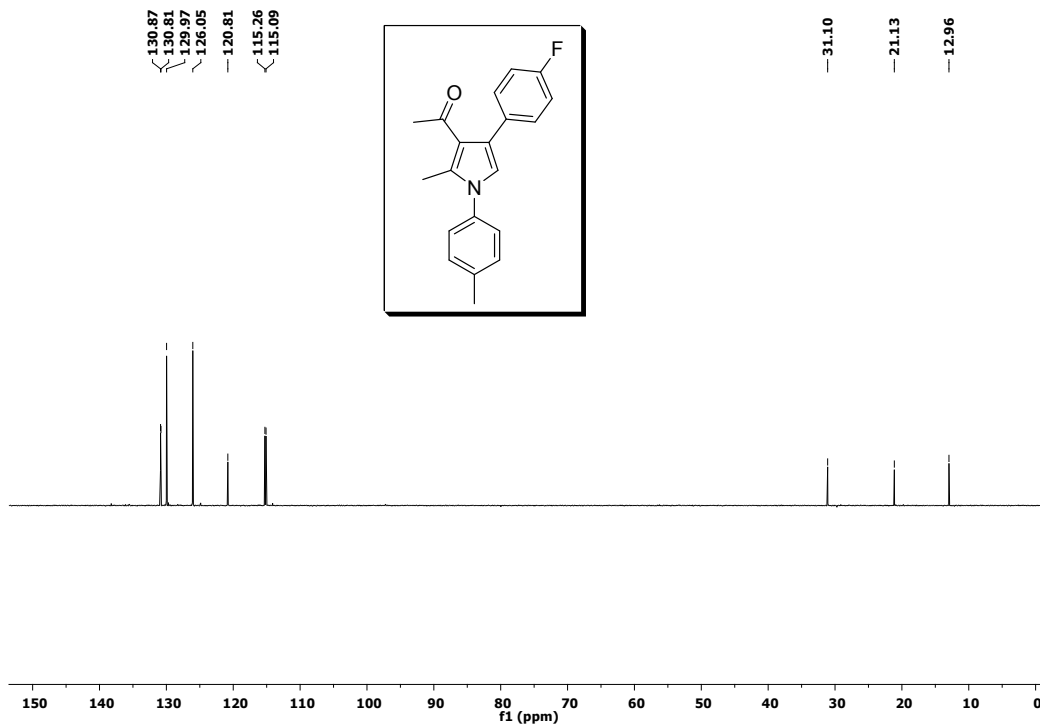
## <sup>1</sup>H NMR



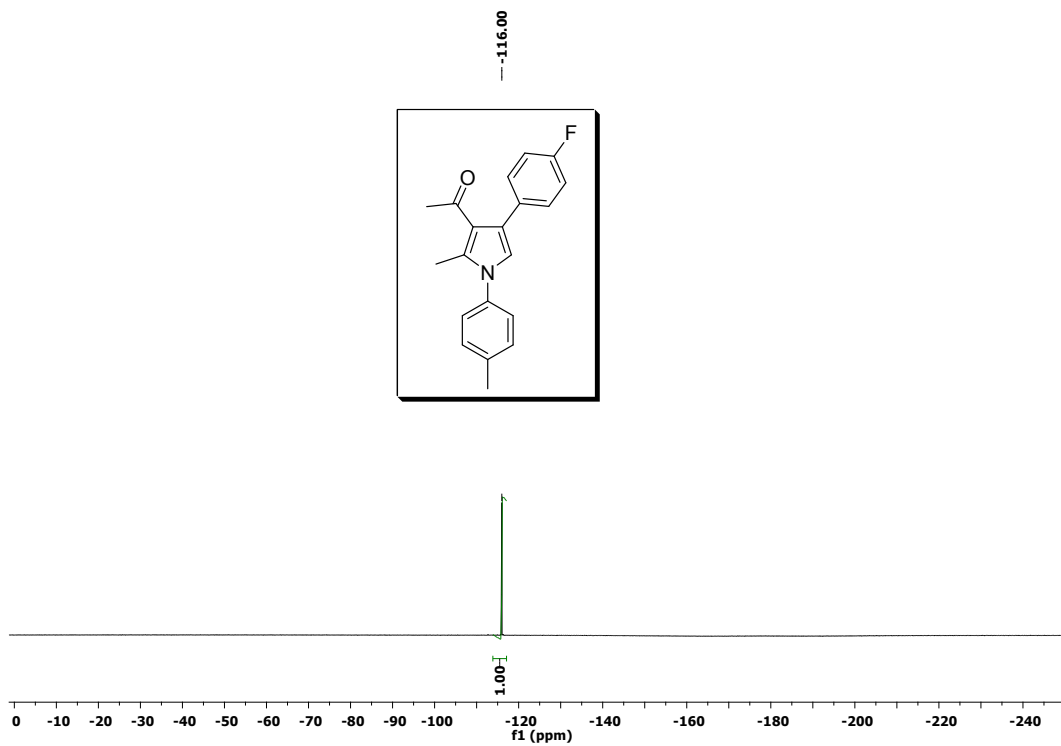
## <sup>13</sup>C NMR



# DEPT

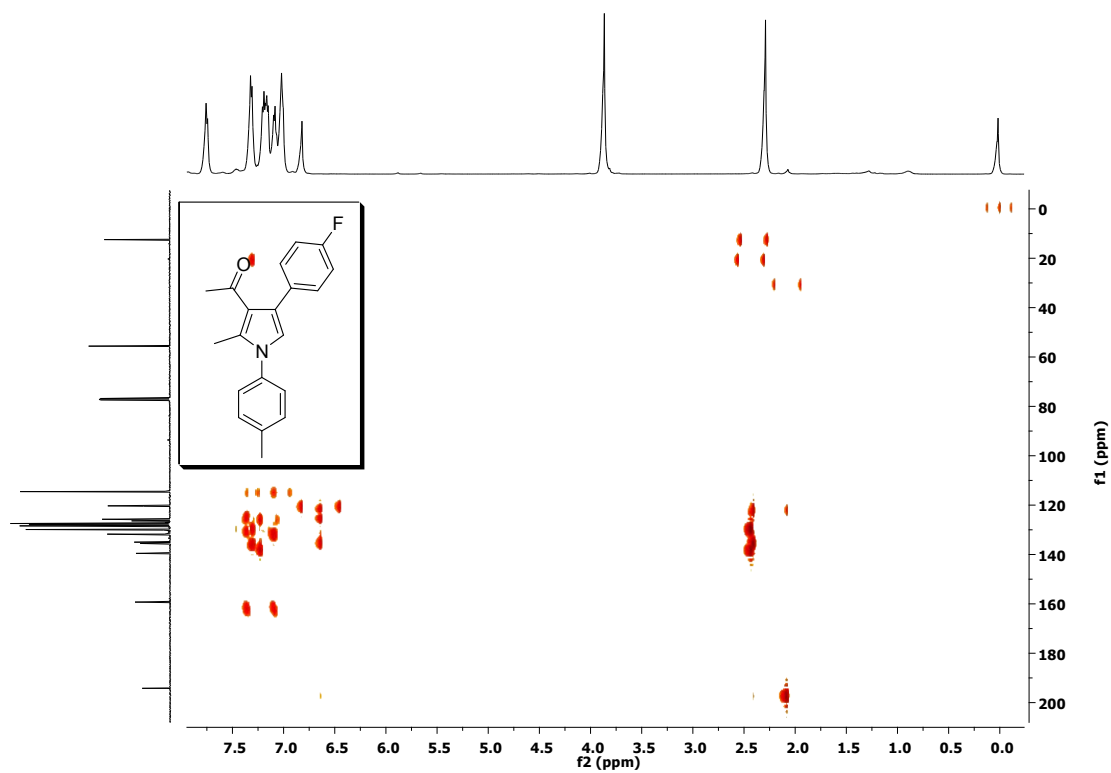


# <sup>19</sup>F NMR



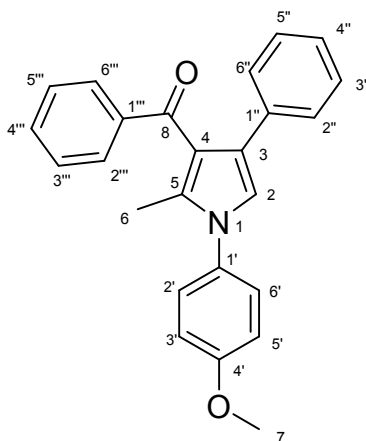


# HMBC



## S5. HMBC correlations and structure assignments for compound 5q

Compound numbering:



Key correlations and assignments:

<p>HMBC</p>		
<p><b>Key HMBC correlations</b></p>	<p><b><sup>13</sup>C NMR assignments</b></p>	<p><b><sup>1</sup>H NMR assignments</b></p>

Assignment table:

position	5q		Key HMBC correlations
	$\delta_C$	$\delta_H$ (J)	
2	120.3, CH	6.80, s	C <sub>3</sub> (139.5), C <sub>5</sub> (120.46), C <sub>8</sub> (194.2), C <sub>6'</sub> (125.73)
3	139.5, C		
4	126.4, C		
5	120.5, C		
6	12.4, CH <sub>3</sub>	2.27, s	C <sub>5</sub> (120.46), C <sub>1'</sub> (135.53)

7	55.6, CH <sub>3</sub>	3.85 s	
8	194.2, CO		
1'	135.5, C		
2', 6'	125.7, CH	7.10-7.08, m	C <sub>1</sub> ' (135.53)
3', 5'	114.5, CH	7.02, d (12)	C <sub>4</sub> ' (159.3), C <sub>3</sub> ' (114.52)
4'	159.3, C		
1''	135.0, C		
2'', 6''	127.7, CH	7.19-7.13, m	C <sub>3</sub> (139.5), C <sub>4</sub> '' (128.41)
3'', 5''	127.9, CH	7.19-7.13, m	
4''	128.4, CH	7.08, t, (12)	C <sub>3</sub> '' (127.92), C <sub>2</sub> '' (127.72)
1'''	131.9, C		
2''', 6'''	129.9, CH	7.74, d, (8)	C <sub>6</sub> ''' (129.89), C <sub>1</sub> ''' (131.94), C <sub>8</sub> (194.2), C <sub>5</sub> ''' (131.76), C <sub>4</sub> ''' (127.45)
3''', 5'''	131.8, CH	7.31, d, (8)	
4'''	127.5, CH	7.31, d, (8)	

**S6. Pharmacokinetic parameters of compound 5i.** The pharmacokinetic study of compound **5i** was carried out in BALB/c male mice of age 4-6 weeks, by administering compound orally and IV formulation at dose of 10 and 1 mg/kg, respectively. Plasma samples were collected at appropriate time points between the range of 0 hours to 24 hours (0.25, 0.5, 1, 2, 4, 8, 10 and 24 h time intervals) and analyzed by LC-MS-MS. Mean plasma concentration was calculated and data was further analyzed for PK parameters evaluation using WinNonlin 5.3 software package.

The pharmacokinetic parameters are listed in Table S1.

**Table S1.** Pharmacokinetic parameters of compound **5i** in BALB/c mice

Parameter	Unit	IV (1 mg/kg)	PO (10 mg/kg)
$t_{1/2, \beta}$	(h)	1.31	0.82
$C_{\max}$	(ng/mL)	158	15.9
$C_0$	(ng/mL)	216	nd
$AUC_{0-t}$	(ng·h/mL)	75.4	14.1
$AUC_{0-\infty}$	(ng·h/mL)	79.5	17.8
CL	(mL/min/kg)	210	nd
$V_d$	(L/kg)	23.8	nd
$V_{dss}$	(L/kg)	11.4	nd
$T_{\text{last}}$	(h)	4.00	nd
Time points considered for $t_{1/2, \beta}$ calculation		1-4 h	0.5-2 h
Bioavailability	F (%)	-	2.25

## References

1. S. Maiti, S. Biswas and U. Jana, *J. Org. Chem.*, 2010, **75**, 1674-1683.