

## Supporting Information

# Rapid and Scalable Synthesis of Innovative Unnatural $\alpha$ , $\beta$ or $\gamma$ -Amino Acids Functionalized with Tertiary Amines on their Side-chain

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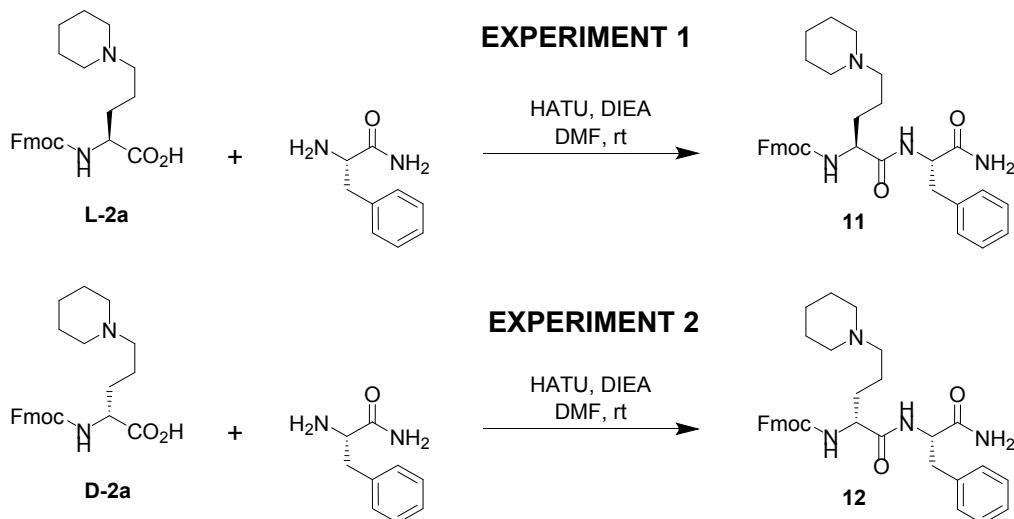
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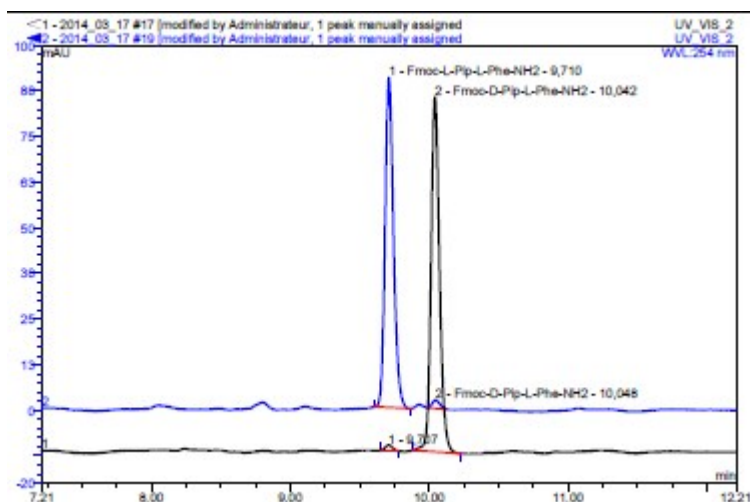
## The ruthenium-catalyzed reduction does not affect the chirality of the amino acid

We evaluate whether this ruthenium-catalyzed reduction could affect the chirality of the amino acid. To answer this issue, both enantiomers **L-2a** and **D-2a** were coupled to the commercially available H-L-Phe-NH<sub>2</sub> using HATU as coupling reagent. In both case, HATU was the last reagent to be added.



After 1 hour, each reaction mixture was analyzed by RP-HPLC-UV without any treatment (Figure 1).

Analytical RP-HPLC-DAD was performed with a Dionex Ultimate3000 with a Waters XSelect CSH™ column (150 x 4.6 mm; 5 μm) using the following parameters : Solvent system : A (MeOH) and B (0.05% TFA in H<sub>2</sub>O); Method : Linear gradient from t = 0 min (80%B) to t = 12 min (0% B); Flow rate : 1 mL/min; Column temperature: 30°C; The ratio of product was determined by integration of UV spectra recorded at 254 nm.



Both diastereoisomers were easily separated: Experiment 1 (blue) led to 99% of compound **11** with 1% of the epimere **12**, while experiment 2 (black) led to 99% of compound **12** with 1% of the epimere **11**. Interestingly, we observed that when compound **2a** was pre-activated with the coupling reagent HATU for one or two minutes before adding phenylalanine, the epimerization ratio increased up to 15%. This observation was also true with other coupling reagents such as BOP or COMU. Consequently, we think that the trace of epimerization we observed in experiments 1 and 2 is more likely due to the coupling reaction, and not to the ruthenium-catalyzed reduction.

*The ruthenium-catalyzed reduction described in this work does not affect the chirality of the resulting basic non-natural amino acids. However, these new amino acids are quite sensitive to epimerization during coupling reaction, and pre-activation has to be avoided.*

## Experimental section:

### Materials and apparatus:

Chemical reagents and anhydrous solvents were obtained from commercial sources and used as-is. Analytical thin-layer chromatography was performed using silica gel plates Merck 60F254 and plates were visualized by exposure to ultraviolet light. When required, compounds were purified using Armen Spot flash chromatography on silica gel Merck 60 (particle size 0.040-0.063mm), or on reversed-phase column (AIT Simply Connect C18 columns – 35g - 0.040-0.063mm). Optical rotations were determined with a Jasco P-2000 polarimeter. <sup>1</sup>H and <sup>13</sup>C NMR spectra were recorded on Bruker Advance spectrometer operating at 400 MHz or 500 MHz for proton and 100 MHz or 125 MHz for carbon. All chemical shift values and coupling constants J are quoted in ppm and in Hz, respectively. Traces of ruthenium were quantified by ICP-AES (Analytical platform ECPM, Strasbourg).

### Abbreviations:

HATU: 1-[Bis(dimethylamino)methylene]-1H-1,2,3-triazolo[4,5-b]pyridinium 3-oxid hexafluorophosphate; BOP: (Benzotriazol-1-yloxy)tris(dimethylamino)phosphonium hexafluorophosphate. COMU: (1-Cyano-2-ethoxy-2-oxoethylideneaminoxy)dimethylamino-morpholino-carbenium hexafluorophosphate ; TMDS: Tetramethyldisiloxane; Ru<sub>3</sub>(CO)<sub>12</sub>: dodecakis(methanidyldyneoxidanium)triruthenium; TFA: Trifluoroacetic acid;

### General procedure for the synthesis of amides 1:

To a solution of commercially available Fmoc-Glu(OH)-OtBu (2.35 mmol, 1 equiv.) in DMF (20 mL) were added DIEA (3.53 mmol, 1.5 equiv.), amine (2.4 mmol, 1.02 equiv.), and HATU (2.47 mmol, 1.05 equiv.). The reaction mixture was stirred at room temperature for 3 hours, and the mixture was concentrated under reduced pressure. The resulting mixture was solubilized in EtOAc, and treated with aqueous 1N HCl, aqueous saturated NaHCO<sub>3</sub>, and aqueous saturated NaCl. The organic layer was dried (MgSO<sub>4</sub>), filtered, and concentrated to afford compounds **1** without further purification steps, excepted for compounds **1c**, **1i**, and **1j** which required a purification on silica gel.

**tert-butyl (2S)-5-(piperidin-1-yl)-2-[[9H-fluoren-9-ylmethoxy]carbonyl]amino}-5-oxopentanoate (L-1a).** 99% yield; <sup>1</sup>H NMR (500 MHz, DMSO-d<sub>6</sub>): δ (ppm) = 7.86 (d, *J*=7.6 Hz, 2H), 7.62-7.75 (m, 3H), 7.39 (t, *J*=7.5 Hz, 2H), 7.30 (t, *J*=7.4 Hz, 2H), 4.34-4.14 (m, 3H), 3.99-3.92 (m, 1H), 3.39 (br d, *J*=4.3 Hz, 2H), 3.36-3.28 (m, 3H), 2.49 (d, *J*=9.5 Hz, 1H), 2.41-2.28 (m, 2H), 1.98-1.87 (m, 1H), 1.87-1.70 (m, 1H), 1.58-1.46 (m, 2H), 1.41 (br s, 3H), 1.37 (s, 9H); <sup>13</sup>C NMR (125 MHz, DMSO-d<sub>6</sub>): δ (ppm) = 171.5, 169.3, 156.1, 143.8, 140.7, 127.6, 127.0, 125.2, 120.1, 80.5, 65.6, 54.0, 46.7, 45.7, 41.9, 28.7, 27.6, 26.5, 26.0, 25.3, 24.0.

**tert-butyl (2S)-5-(azepan-1-yl)-2-[[9H-fluoren-9-ylmethoxy]carbonyl]amino}-5-oxopentanoate (1b).** 99% Yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ (ppm) = 7.67 (d, *J*= 7.4 Hz, 2H), 7.53 (t, *J*= 7.3 Hz, 2H), 7.31 (t, *J*= 7.5 Hz, 2H), 7.22 (t, *J*= 7.3 Hz, 2H), 5.79 (d, *J*= 7.6 Hz, 1H), 4.33-4.12 (m, 4H), 3.45 (t, *J*= 5.9, 2H), 3.30 (t, *J*= 5.9, 2H), 2.44-2.26 (m, 2H), 2.21-2.11 (m, 1H), 2.04-1.92 (m, 1H), 1.65-1.55 (m, 4H), 1.48-1.43 (m, 4H), 1.39 (s, 9H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): δ (ppm) = 171.7, 171.5, 156.3, 144.1, 141.4, 127.8, 127.1, 125.4, 120.0, 82.2, 67.1, 54.6, 47.9, 47.3, 46.3 29.5, 29.1, 28.2, 27.8, 27.7, 27.2, 26.9.

**tert-butyl (2S)-2-[[9H-fluoren-9-ylmethoxy]carbonyl]amino}-5-[(2R)-2-methylpyrrolidin-1-yl]-5-oxopentanoate (1c).** 77% yield after column chromatography (EtOAc – Heptane, 40/60); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ (ppm) = 7.68 (d, *J* = 7.7 Hz, 2H), 7.54 (*t*, *J* = 6.8 Hz, 2H), 7.32 (*t*, *J* = 7.3 Hz, 2H), 7.23 (*t*, *J* = 7.3 Hz, 2H), 5.89-5.75 (m, 1H), 4.36-4.29 (m, 1H), 4.25-4.18 (m, 1H), 4.16-4.11 (m, 2H), 3.43-3.24 (m, 2H), 2.39-2.10 (m, 3H), 2.04-1.75 (m, 4H), 1.60-1.42 (m, 2H), 1.40 (s, 9H), 1.12-1.07 (m, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): δ (ppm) = 171.6, 170.5, 156.4, 144.2, , 141.5, 127.8, 127.2, 125.4, 120.1, 82.3, 67.2, 54.8, 53.3, 47.4, 47.0, , 32.0, 31.3, 27.3, 24.0, 22.0, 19.7.

**tert-butyl(2S)-2-[[9H-fluoren-9-ylmethoxy]carbonyl]amino}-5-oxo-5-[(1S,5R)-1,3,3-trimethyl-6-azabicyclo[3.2.1]octan-6-yl]pentanoate (1d).** 95 % yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ (ppm) = 7.69 (d, *J* = 7.6 Hz, 2H), 7.54 (*t*, *J* = 7.6 Hz, 2H), 7.32 (*t*, *J* = 7.5 Hz, 2H), 7.24 (*t*, *J* = 7.5 Hz, 2H), 5.89-5.73 (m, 1H), 4.43-3.97 (m, 5H), 3.40-3.19 (m, 1H), 3.00-2.95 (m, 1H), 2.46-1.89 (m, 4H), 1.69-1.49 (m, 2H), 1.46-1.18 (m, 12 H), 0.99-0.97 (m, 3H), 0.88-0.75 (m, 7H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): δ (ppm) = 171.5, 169.6, 158.7, 144.0, 141.4, 127.8, 127.2, 125.5, , 120.1, 82.3, 67.2, 57.3, 56.555.2, 54.7, 52.0, 51.5, 47.3, 44.5, 43.4, 41.4, 40.3, 38.2, 36.6, 31.7, 28.2, 25.2, .

**tert-butyl(2S)-2-[[9H-fluoren-9-ylmethoxy]carbonyl]amino}-5-(4-methylpiperazin-1-yl)-5-oxopentanoate (1e).** 98% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ (ppm) = 7.73 (d, *J* = 7.6 Hz, 2H), 7.58 (*t*, *J* = 7.6 Hz, 2H), 7.37 (*t*, *J* = 7.5 Hz, 2H), 7.28 (*t*, *J* = 7.3 Hz, 2H), 5.68 (d, *J* = 7.5 Hz, 1H), 4.35-4.13 (m, 4H), 3.69-3.51 (m, 2H), 3.40-3.32 (m, 2H), 2.40-2.11 (m, 10 H), 1.97-1.90 (m, 1H), 1.4 (s, 9H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): δ (ppm) = 171.4, 170.5, 156.3, 144.1, 141.4, 127.8, 127.2, 125.3, 120.1, 82.4, 67.2, 55.1, 54.8, 54.4, 47.3, 46.1, 45.4, 41.8, 29.4, 29.1.

**tert-butyl(2S)-2-[[9H-fluoren-9-ylmethoxy]carbonyl]amino}-5-oxo-5-[4-(pyrimidin-2-yl)piperazin-1-yl]pentanoate (1f).** 85% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ (ppm) = 8.25 (d, *J* = 4.8 Hz, 2H), 7.69 (d, *J* = 7.3 Hz, 2H), 7.55-7.52 (m, 2H), 7.32 (*t*, *J* = 7.3 Hz, 2H), 7.24 (*t*, *J* = 7.3 Hz, 2H), 6.47 (*t*, *J* = 4.8 Hz, 1H), 5.59 (d, *J* = 7.9 Hz, 1H), 4.34-4.13 (m, 4H), 3.79-3.73 (m, 4H), 3.66-3.57 (m, 2H), 3.46-3.38 (m, 2H), 2.48-2.31 (m, 2H), 2.24-2.16 (m, 1H), 2.00-1.94 (m, 1H), 1.41 (s, 9H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): δ (ppm) = 171.4, 170.8, 161.5, 157.9, 156.3, 143.9, 141.5, 127.9, 127.3, 125.4, 120.1, 110.6, 82.5, 67.2, 54.4, 47.3, 45.4, 43.7, 41.7, 29.5, 28.2.

**tert-butyl(2S)-2-[[9H-fluoren-9-ylmethoxy]carbonyl]amino}-5-oxo-5-[4-(piperidin-1-yl)piperidin-1-yl]pentanoate (1g).** 90% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ (ppm) = 7.69 (d, *J* = 7.6 Hz, 2H), 7.56-7.51 (m, 2H), 7.32 (*t*, *J* = 7.6 Hz, 2H), 7.24 (*t*, *J* = 7.3 Hz, 2H), 5.64 (m, 1H), 4.34-4.13 (m, 4H), 3.31-2.71 (m, 5H), 2.65-2.14 (m, 4H), 2.11-1.69 (m, 6H), 1.65-1.35 (m, 12H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): δ (ppm) = 171.5, 170.2, 156.4, 144.2, 141.4, 127.9, 127.2, 125.4, 120.2, 82.4, 67.2, 62.8, 54.6, 50.3, 47.3, 45.3, 41.8, 28.8, 28.2, 27.7, 26.3, 24.7.

**Ethyl-1-[(4S)-5-(tert-butoxy)-4-[[9H-fluoren-9-ylmethoxy]carbonyl]amino}-5-oxopentanoyl]piperidine-4-carboxylate (1h).** 83% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ (ppm) = 7.74 (d, *J* = 7.5 Hz, 2H), 7.54 (*t*, *J* = 7.4 Hz, 2H), 7.38 (*t*, *J* = 7.4 Hz, 2H), 7.29 (*t*, *J* = 7.4 Hz, 2H), 5.63-5.59 (m, 1H), 4.35-4.13 (m, 5H), 4.11 (q, *J* = 6.9 Hz, 2H), 3.72-3.66 (m, 1H), 3.06-2.95 (m, 1H), 2.79-2.69 (m, 1H), 2.51-2.31 (m, 3H), 2.14-2.27 (m, 1H), 2.06-1.83 (m, 3H), 1.65-1.54 (m, 2H), 1.45 (s, 9H), 1.22 (*t*, *J* = 6.9 Hz, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): δ (ppm) = 174.3, 171.5, 170.4, 156.3, 144.2, 127.9, 127.3, 125.4, 120.2, 82.5, 67.2, 60.8, 54.5, 47.3, 44.9, 41.4, 41.2, 29.6, 28.6, 28.2, 28.0, 14.4.

**tert-butyl(2S)-2-[[9H-fluoren-9-ylmethoxy]carbonyl]amino}-5-(4-hydroxypiperidin-1-yl)-5-oxopentanoate (1i).** 70% yield after column chromatography (DCM – MeOH, 96/4); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ (ppm) = 7.68 (d, *J* = 7.5 Hz, 2H), 7.53 (*t*, *J* = 7.3 Hz, 2H), 7.31 (*t*, *J* = 7.3 Hz, 2H),

7.22 (*t*, *J* = 7.5 Hz, 2H), 5.72 (m, 1H), 4.33-4.12 (m, 4H), 4.03-3.93 (m, 1H), 3.81 (m, 1H), 3.64-3.56 (m, 1H), 3.02-3.24 (m, 2H), 2.43-1.89 (m, 5H), 1.70-1.81 (m, 2H), 1.31-1.52 (m, 11H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): δ (ppm) = 171.5, 170.4, 156.3, 144.1, 141.4, 127.9, 127.3, 127.2, 125.4, 120.1, 82.4, 67.2, 67.1, 54.5, 47.3, 42.9, 39.3, 34.6, 34.0, 29.5, 28.2, 28.1.

**tert-butyl(2S)-5-(4-benzylpiperidin-1-yl)-2-[(9H-fluoren-9-ylmethoxy)carbonyl]amino}-5-oxopentanoate (1j).** 70% yield after column chromatography (EtOAc – Heptane, 5/95 to 30/70); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ (ppm) = 7.69 (d, *J* = 7.6 Hz, 2H), 7.53 (*t*, *J* = 7.3 Hz, 2H), 7.32 (*t*, *J* = 7.4 Hz, 2H), 7.25-7.18 (m, 4H), 7.14-7.09 (m, 1H), 7.04-7.01 (m, 2H), 5.66 (d, *J* = 7.1 Hz, 1H), 4.53 (d, *J* = 11.8 Hz, 1H), 4.34-4.13 (m, 4H), 3.69 (d, *J* = 12.5 Hz, 1H), 2.83 (*t*, *J* = 12.5 Hz, 1H), 2.5-2.25 (m, 5H), 2.18-2.09 (m, 1H), 1.98-1.89 (m, 1H), 1.70-1.58 (m, 4H), 1.39 (s, 9H), 1.27-1.16 (m, 1H), 1.13-0.98 (m, 2H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): δ (ppm) = 171.5, 170.3, 156.3, 144.0, 141.5, 140.1, 129.2, 128.5, 127.9, 127.3, 126.3, 125.4, 120.2, 82.4, 67.2, 54.6, 47.4, 46.0, 43.1, 42.4, 38.4, 32.7, 31.9, 29.6, 28.2, 28.1.

**tert-butyl(2S)-2-[(9H-fluoren-9-ylmethoxy)carbonyl]amino}-5-oxo-5-(1,2,3,4-tetrahydroisoquinolin-2-yl)pentanoate (1k).** 98% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ (ppm) = 7.73 (d, *J* = 7.4 Hz, 2H), 7.54-7.52 (m, 2H), 7.37 (*t*, *J* = 7.4 Hz, 2H), 7.28 (*t*, *J* = 7.3 Hz, 2H), 7.11-6.98 (m, 4H), 5.63 (m, 1H), 4.65 (s, 1H), 4.50 (s, 1H), 4.40-4.10 (m, 4H), 3.81 (*t*, *J* = 5.6 Hz, 1H), 3.61 (*t*, *J* = 5.4 Hz, 1H), 2.81-2.77 (m, 2H), 2.48-2.35 (m, 2H), 2.24-2.14 (m, 1H), 2.08-1.94 (m, 1H), 1.40 (s, 9H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): δ (ppm) = 171.5, 171.1, 156.3, 144.1, 144.0, 141.5, 135.3, 134.2, 133.6, 132.5, 129.1, 128.4, 127.9, 127.3, 127.2, 126.9, 126.6, 126.3, 125.4, 125.3, 120.2, 82.5, 67.2, 54.6, 47.40, 44.5, 43.4, 40.1, 30.0, 29.6, 28.7, 28.2, 28.1.

**tert-butyl (2S)-5-(2,3-dihydro-1H-indol-1-yl)-2-[(9H-fluoren-9-ylmethoxy)carbonyl]amino}-5-oxopentanoate (1l).** 98% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ (ppm) = 8.17 (d, *J* = 8.0 Hz, 1H), 7.67 (d, *J* = 7.3 Hz, 2H), 7.50 (d, *J* = 7.3 Hz, 2H), 7.30 (*t*, *J* = 7.4 Hz, 2H), 7.21-7.16 (m, 2H), 7.13-7.09 (m, 2H), 6.93 (*t*, *J* = 7.3 Hz, 1H), 5.67 (d, *J* = 7.5 Hz, 1H), 4.28-3.89 (m, 6H), 3.09 (*t*, *J* = 8.4 Hz, 2H), 2.52-2.01 (m, 4H), 1.41 (s, 9H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): δ (ppm) = 171.4, 170.4, 156.4, 144.1, 144.0, 143.1, 141.5, 131.2, 127.9, 127.8, 127.2, 125.4, 124.7, 123.9, 120.1, 117.2, 82.5, 67.2, 54.5, 48.1, 47.3, 32.3, 28.2, 28.1, 27.5.

**tert-butyl (2S)-5-(1H-1,3-benzodiazol-1-yl)-2-[(9H-fluoren-9-ylmethoxy)carbonyl]amino}-5-oxopentanoate (1m).** 98% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ (ppm) = 8.54-8.24 (m, 1H), 8.14-8.04 (m, 1H), 7.74-7.39 (m, 5H), 7.35-7.05 (m, 6H), 5.63-5.51 (d, *J* = 8.1 Hz, 1H), 4.36-4.21 (m, 3H), 4.06-3.96 (m, 1H), 3.19-2.65 (m, 2H), 2.45-2.25 (m, 1H), 2.15-1.95 (m, 1H), 1.37 (s, 9H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): δ (ppm) = 170.9, 169.6, 156.3, 151.8, 143.8, 141.4, 141.1, 131.5, 129.6, 127.8, 127.1, 126.1, 125.2, 125.1, 120.6, 120.1, 115.6, 83.0, 67.0, 53.7, 47.2, 32.1, 28.1, 27.4.

**tert-butyl(2S)-2-[(9H-fluoren-9-ylmethoxy)carbonyl]amino}-4-[methyl(phenyl)carbamoyl]butanoate (1n).** 97% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ (ppm) = 7.69 (d, *J* = 7.4 Hz, 2H), 7.52-7.49 (m, 2H), 7.34-7.18 (m, 7H), 7.07 (d, *J* = 7.5 Hz, 2H), 5.53 (d, *J* = 7.4 Hz, 1H), 4.29-4.19 (m, 2H), 4.13-4.02 (m, 2H), 3.18 (s, 3H), 2.09-1.87 (m, 4H), 1.32 (s, 9H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): δ (ppm) = 172.2, 171.4, 156.1, 144.1, 143.9, 141.4, 130.0, 128.1, 127.9, 127.4, 127.2, 125.4, 120.1, 82.2, 67.2, 54.4, 47.3, 37.6, 30.6, 28.2, 28.1.

**tert-butyl (2S)-2-[(9H-fluoren-9-ylmethoxy)carbonyl]amino}-4-[methyl(2-phenylethyl)carbamoyl] butanoate (1o).** 94% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ (ppm) = 7.74 (*dd*, *J* = 7.5, 3.4 Hz, 2H), 7.55-7.52 (m, 2H), 7.34-7.29 (m, 2H), 7.25-7.11 (m, 6H), 7.03 (d, *J* = 7.0 Hz, 1H), 5.58 (*dd*, *J* = 76.7, 8.0 Hz, 1H), 4.46-4.10 (m, 4H), 3.63-3.39 (m, 2H), 3.01-2.71 (m, 5H), 2.49-

1.74 (m, 4H), 1.55-1.39 (m, 9H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): δ (ppm) = 172.0, 171.5, 156.4, 144.2, 141.5, 139.5, 138.3, 129.0, 128.9, 127.9, 127.3, 127.0, 126.6, 125.4, 120.2, 82.4, 67.2, 54.6, 51.8, 50.5, 47.4, 36.3, 34.9, 34.0, 33.9, 30.0, 29.1, 28.2, 28.0, 27.8.

**tert-butyl-(2S)-4-[butyl(methyl)carbamoyl]-2-[(9H-fluoren-9-**

**ylmethoxy)carbonyl]amino}butanoate (1p).** 95% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ (ppm) = 7.72 (d, *J*= 7.7 Hz, 2H), 7.59 (t, *J*= 7.4 Hz, 2H), 7.36 (t, *J*= 7.3 Hz, 2H), 7.4 (t, *J*= 7.4 Hz, 2H), 5.91-5.78 (m, 1H), 4.41-4.09 (m, 4H), 3.34 (t, *J*= 7.5 Hz, 1H), 3.19 (t, *J*=7.5 Hz, 1H), 2.89 (s, 3H), 2.44 (m, 2H), 2.19 (m, 1H), 2.10-1.92 (m, 1H), 1.47-1.42 (m, 2H), 1.39 (s, 9H), 1.32-1.21 (m, 2H), 0.95-0.82 (m, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): δ (ppm) = 171.8, 171.5, 156.3, 144.1, 141.4, 127.8, 127.2, 125.4, 120.1, 82.2, 67.1, 54.6, 47.8, 47.3, 33.6, 29.9, 29.5, 28.1, 27.7, 20.2, , 13.9.

**tert-butyl(2S)-4-[(2-cyanoethyl)(methyl)carbamoyl]-2-[(9H-fluoren-9-**

**ylmethoxy)carbonyl]amino} butanoate (1q).** 91% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ (ppm) = 7.65-7.20 (m, 8H), 5.67 (s, 1H), 4.28-4.12 (m, 4H), 3.47-3.35 (m, 2H), 2.95-2.66 (m, 3H), 2.49-1.87 (m, 6H), 1.37 (9H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): δ (ppm) = 172.5, 171.3, 156.2, 144.0, 141.3, 127.7, 127.1, 125.2, 120.0, 118.4, 82.3, 67.0, 54.1, 47.2, 45.0, 36.8, 29.5, 28.0, 27.6, 16.2.

**tert-butyl**

**(2S)-4-{N-benzyl-N'-[(tert-butoxy)carbonyl]hydrazinecarbonyl}-2-[(9H-fluoren-9-ylmethoxy)carbonyl]amino}butanoate (1r).** 65% yield after column chromatography; <sup>1</sup>H NMR (400MHz, CDCl<sub>3</sub>): δ (ppm) = 7.76 (d, *J*=7.5 Hz, 2H), 7.58 (d, *J*=7.3 Hz, 2H), 7.36 (t, *J*=6.9 Hz, 2H), 7.32-7.18 (m, 7H), 5.70-5.50 (m, 1H), 5.49-5.22 (m, 1H), 4.42-3.92 (m, 5H), 2.72-1.79 (m, 4H), 1.45 (s, 9H), 1.42-1.32 (m, 9H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): δ (ppm) = 174.9, 171.5, 144.3, 141.2, 135.7, 129.2, 128.9, 128.1, 127.9, 127.2, 125.3, 120.6, 82.5, 67.3, 47.3, 28.3, 28.1

**(2S)-2-[(9H-fluoren-9-ylmethoxy)carbonyl]amino}-4-oxo-4-(piperidin-1-yl)butanoic acid (3).**

Same procedure than for compounds **1**, starting from commercially available Fmoc-L-Asp-OtBu. 69% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ (ppm) = 7.73(d, *J*=7.5 Hz, 2H), 7.60 (dd, *J*=7.3, 3.5 Hz, 2H), 7.37 (t, *J*=7.5, 2H), 7.28 (br tt, *J*=7.5, 1.0 Hz, 2H), 6.14 (br d, *J*=9.0 Hz, 1H), 4.55-4.47 (m, 1H), 4.44 (dd, *J*=9.9, 6.7 Hz, 1H), 4.29-4.16 (m, 2H), 3.59 (br s, 1H), 3.49 (br dd, *J*=7.0, 4.4 Hz, 1H), 3.44-3.26 (m, 2H), 3.10 (dd, *J*=16.6, 4.0 Hz, 1H), 2.72 (dd, *J*=16.6, 3.9 Hz, 1H), 1.70-1.59 (m, 4H), 1.59-1.48 (m, 4H), 1.48 (s, 9H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): δ (ppm) = 170.3, 168.3, 156.4, 144.0, 143.9, 141.3, 127.6, 127.0, 125.3, 120.0, 82.0, 67.3, 51.4, 47.4, 46.8, 42.9, 35.8, 28.1, 26.6, 25.8, 24.6.

**tert-butyl (3S)-3-[(9H-fluoren-9-ylmethoxy)carbonyl]amino}-4-oxo-4-(piperidin-1-yl)butanoate**

**(5).** Same procedure than for compounds **1**, starting from commercially available Fmoc-L-Asp(OtBu)-OH. 91% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ (ppm) = 7.74 (d, *J*=7.7 Hz, 2H), 7.58 (t, *J*=1.0 Hz, 2H), 7.37 (t, *J*=1.0 Hz, 2H), 7.32-7.26 (m, 2H), 5.83 (d, *J*=8.2 Hz, 1H), 4.72 (br td, *J*=3.6 Hz, 1H), 4.33 (d, *J*=7.2 Hz, 2H), 4.19 (br t, *J*=7.0 Hz, 1H), 3.64-3.42 (m, 4H), 2.40-2.22 (m, 1H), 2.06-1.94 (m, 1H), 1.79-1.68 (m, *J*=7.9 Hz, 1H), 1.68-1.49 (m, 7H), 1.43 (s, 9H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): δ (ppm) = 172.1, 169.5, 156.1, 144.0, 141.3, 127.6, 127.0, 125.2, 120.1, 80.5, 67.2, 50.1, 47.4, 46.8, 43.5, 30.9, 28.6, 28.3, 26.7, 25.7, 24.6.

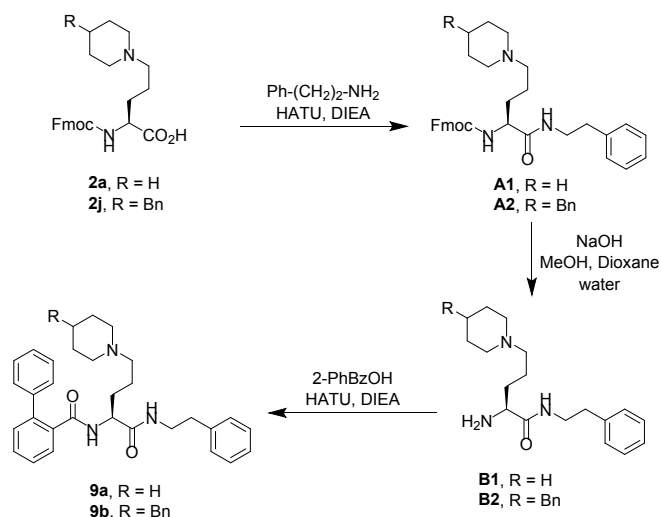
**tert-butyl**

**(4S)-4-[(9H-fluoren-9-ylmethoxy)carbonyl]amino}-5-oxo-5-(piperidin-1-yl)pentanoate (6).**

Same procedure than for compounds **1**, starting from commercially available Fmoc-L-GluO(tBu)-OH. 95% yield; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ (ppm) = 7.73 (d, *J*=7.5 Hz, 2H), 7.56 (br d, *J*=7.4 Hz, 2H), 7.37 (t, *J*=7.4 Hz, 2H), 7.28 (t, *J*=7.4 Hz, 2H), 5.78 (br d, *J*=9.2 Hz, 1H), 5.05-4.95 (m, 1H), 4.34 (d, *J*=7.2

Hz, 2H), 4.19 (t,  $J=7.1$  Hz, 1H), 3.64-3.40 (m, 4H), 2.68 (dd,  $J=15.4, 6.0$  Hz, 1H), 2.49 (dd,  $J=15.4, 6.5$  Hz, 1H), 1.68-1.48 (m, 6H), 1.42 (s, 9H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ ):  $\delta$  (ppm) = 169.7, 168.9, 155.8, 144.0, 141.5, 127.9, 127.2, 125.3, 120.1, 81.5, 67.3, 47.9, 47.3, 46.9, 43.5, 39.4, 28.1, 26.6, 25.7, 24.7.

### Synthesis of compounds 9a and 9b



To a solution of **2** (1.96 mmol, 1 equiv.) in DMF (10 mL) were added DIEA (4.9 mmol, 2.5 equiv.), phenethylamine (2.05 mmol, 1.05 equiv.), and HATU (2.05 mmol, 1.05 equiv.). The reaction mixture was stirred at room temperature for 1 hour, and the mixture was concentrated under reduced pressure. The resulting mixture was solubilized in EtOAc, and treated with aqueous 1N HCl, aqueous saturated  $\text{NaHCO}_3$ , and aqueous saturated NaCl. The organic layer was dried ( $\text{MgSO}_4$ ), filtered, and concentrated to afford the expected amide **A** without further purification steps.

**A1** : R = H; 91% yield;  $^1\text{H}$  NMR (400 MHz,  $\text{MeOD-d}_4$ )  $\delta$  (ppm) : 7.81 (d,  $J = 7.5$  Hz, 2H ), 7.66 (m, 2H ), 7.40 (dd,  $J = 7.5, 7.5$  Hz, 2H), 7.31 (dd,  $J = 7.5, 7.5$  Hz, 2H), 7.28 – 7.15 (m, 5H), 4.41 (m, 2H), 4.21 (m, 1H), 3.97 (m, 1H), 3.41 (m, 2H), 2.79 (m, 2H), 2.72 – 2.38 (m, 6H), 1.83 – 1.51 (m, 10H).

**A2** : R = Bn; 87% yield;  $^1\text{H}$  NMR (400 MHz,  $\text{MeOD-d}_4$ )  $\delta$  (ppm) : 7.78 (d,  $J = 7.5$  Hz, 2H ), 7.65 (m, 2H ), 7.38 (dd,  $J = 7.5, 7.5$  Hz, 2H), 7.32 – 7.09 (m, 12H), 4.39 (m, 2H), 4.19 (m, 1H), 4.01 (m, 1H), 3.41 (m, 2H), 2.92 (m, 2H), 2.77 (m, 3H), 2.51 (m, 3H), 2.39 (m, 2H), 1.77 – 1.44 (m, 7H), 1.36 – 1.22 (m, 2H).

To a solution of amide **A** (0.97 mmol, 1 equiv.) in a mixture of methanol (72 mL) and dioxane (19 mL) was added NaOH (4M in water, 4.8 mL, 20 equiv.). The reaction mixture was stirred at room temperature for 45 minutes, and the mixture was concentrated under reduced pressure. Water was added to the resulting mixture to afford a white solid. The precipitate was solubilized in dichloromethane, and the resulting organic layer was dried ( $\text{MgSO}_4$ ), filtered and evaporated to afford an oil. HCl 2N in ether (20 mL) was added to afford the amine **B** as a powder (> 80% yield), which was directly used in the next step without further purification steps.

To a solution of 2-biphenylcarboxylic acid (0.19 mmol, 1.05 equiv.) in DMF (1 mL) were added DIEA (0.61 mmol, 2.5 equiv.), **B** (0.176 mmol, 1 equiv.) and HATU (0.19 mmol, 1.05 equiv.). The reaction mixture was stirred at room temperature for 1 hour, and the mixture was concentrated under reduced pressure. The resulting mixture was purified by column chromatography using a reversed-phase C18 column (eluent: MeOH – water containing 0.05% TFA). Compounds **9** were obtained as

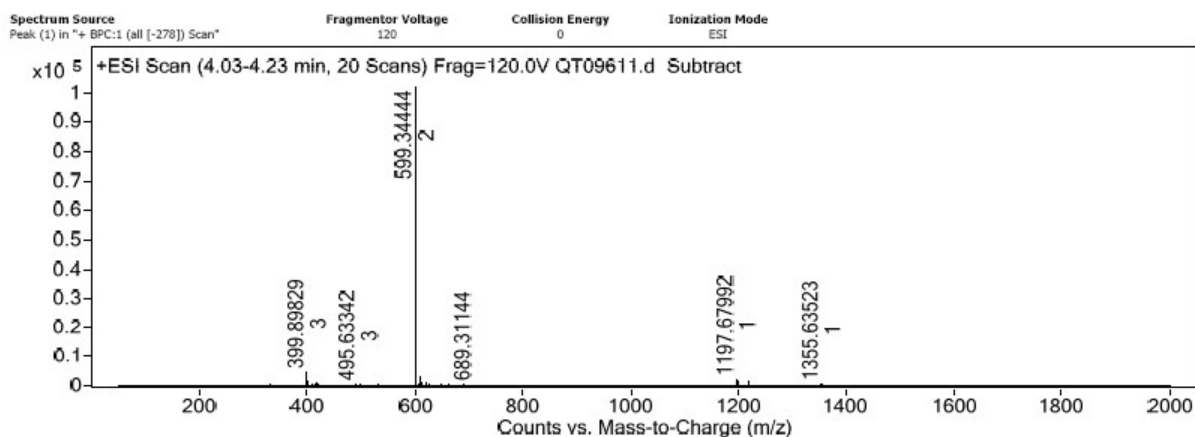
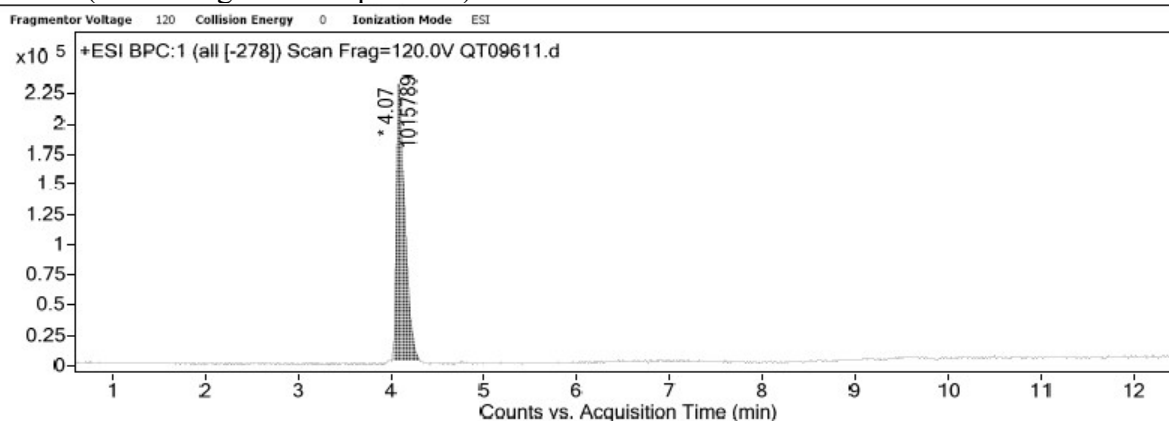
trifluoroacetic acid salts (**9a** : 72% yield; **9b** : 34% yield); Analytical data were in full agreement with those previously published (Bihel et al. *ACS Chem Neurosci* **2015**. DOI: 10.1021/cn500219h).



## Synthesis of peptide 10

The peptide chain elongation was carried out manually using a Rink-Amide resin. Typically the coupling was carried out using a solution of the Fmoc-amino acid (3 equiv.), HATU (3.8 equiv.) and DIEA (8 equiv.) in DMF. 1-1.5 h reaction is enough for most amino acids. Non-natural amino acid **2j** was used to replace Arg. In this case, it was important to add HATU as the last reagent to avoid any epimerization. After the coupling reaction the resin was washed with DMF and DCM. The Fmoc group was removed by treatment with 20% piperidine in DMF (twice: 5 min, 10 min). Ninhydrin test may be carried out to check the coupling efficiency. After assembly of peptide chains, the N-terminal Fmoc group was removed. After washing the resin the cleavage/deprotection step was carried out by treating the resin with cleavage cocktail (TFA/TIPS/thiophenol/, 92:3:5) for 1 h. Then the solution was collected and concentrated under reduced pressure. After precipitation with cold Et<sub>2</sub>O and centrifugation the crude peptides can be obtained. Analysis and purification was carried out using HPLC and MS. After lyophilisation the desired peptide **10** was obtained. HRMS (ESI+) calcd. for C<sub>65</sub>H<sub>88</sub>N<sub>12</sub>O<sub>10</sub> [M + 2H]<sup>+</sup> 599.3444, found 599.3444.

## HRMS (chromatogram and spectrum)



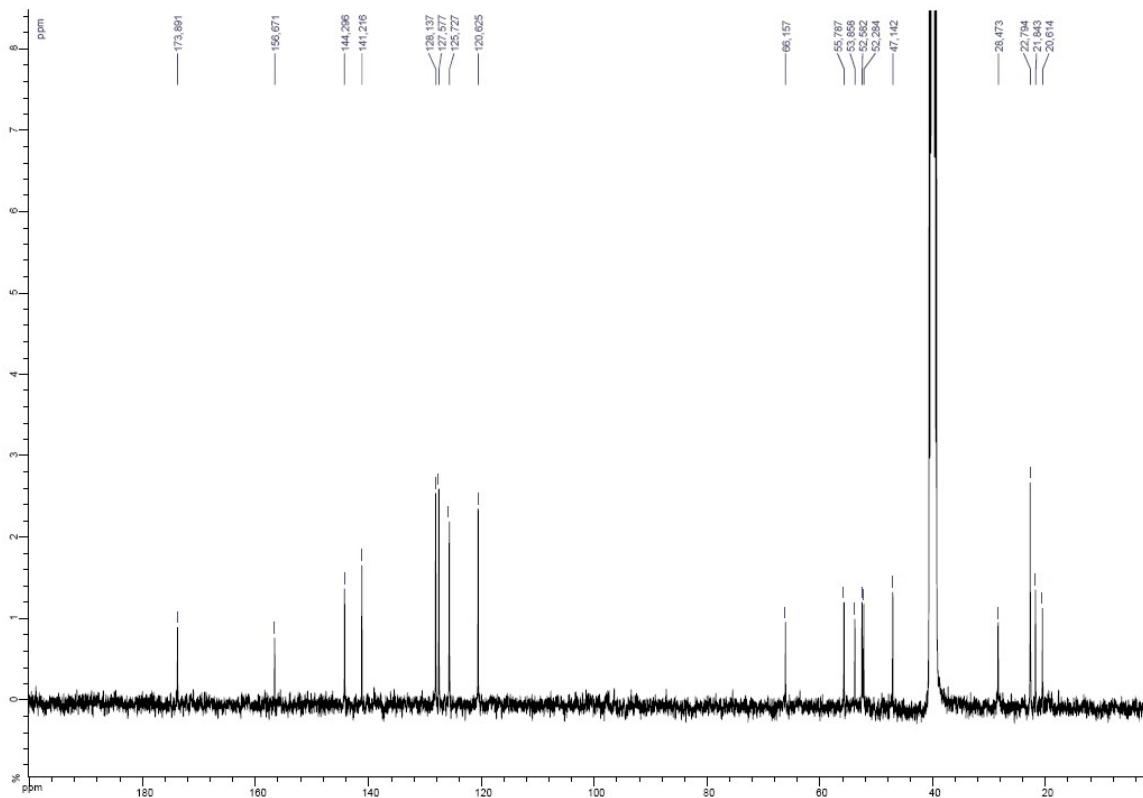
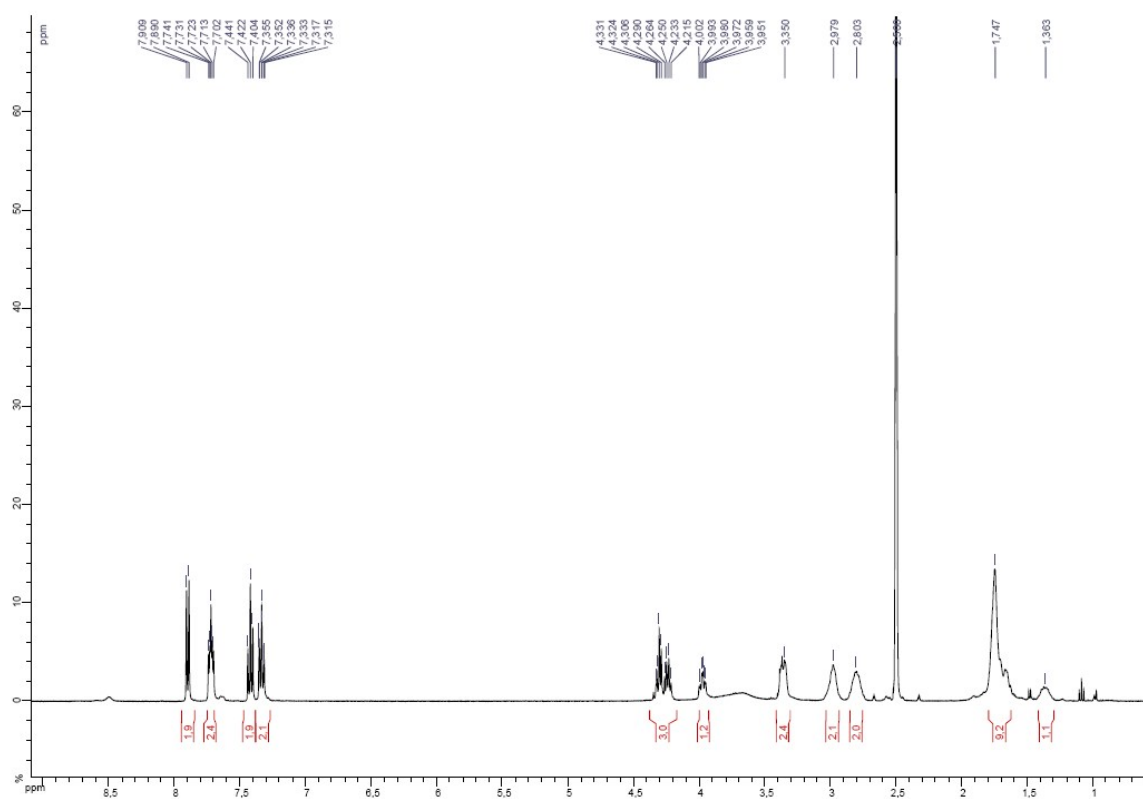
### **NPPFR1 and NPPFR2 receptor binding assays**

Competition experiments with membranes from CHO cells stably expressing hNPPFR1 and hNPPFR2 were performed essentially as described [1]. Briefly, hNPPFR1 or hNPPFR2 membranes (5 to 10 µg of proteins) were incubated (1 hr at 25°C; 0.25 mL total volume) with 0.015 nM [D-Tyr<sup>1</sup>[<sup>125</sup>I], N-MePhe<sup>3</sup>]-NPPF (Hartmann Analytic GmbH) in 50 mM HEPES (pH 7.4), 1 mM CaCl<sub>2</sub>, 1 mM MgCl<sub>2</sub> and 0.1% bovine serum albumin. Non-specific binding was determined in the presence of 10 µM RFRP-3. Incubation was terminated by rapid filtration through a 96-well GF/B unifilter apparatus (Perkin Elmer Life and Analytical Sciences, Courtaboeuf, France). Unifilters were washed three times with binding buffer, then dried for 1 h at 65°C. After addition of 40 µL scintillation cocktail (Microscint-O, Perkin Elmer) per well, bound radioactivity was determined on a TopCount scintillation counter (Perkin Elmer).

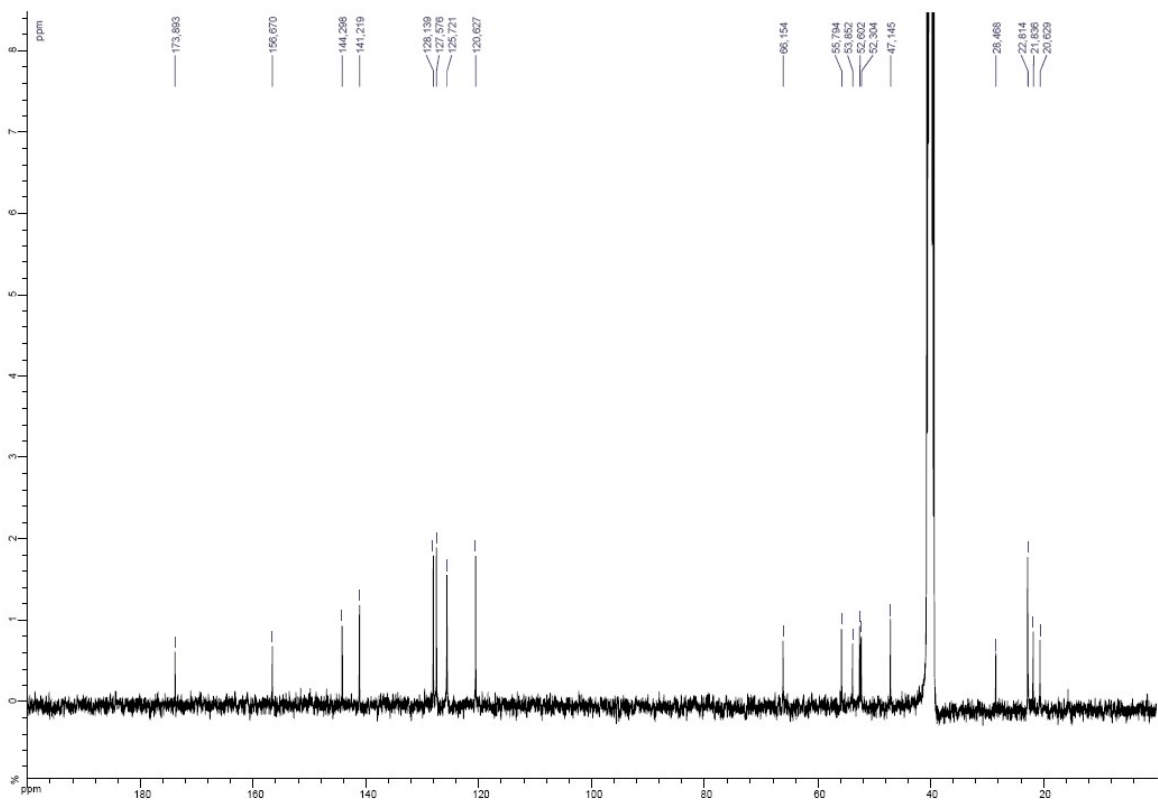
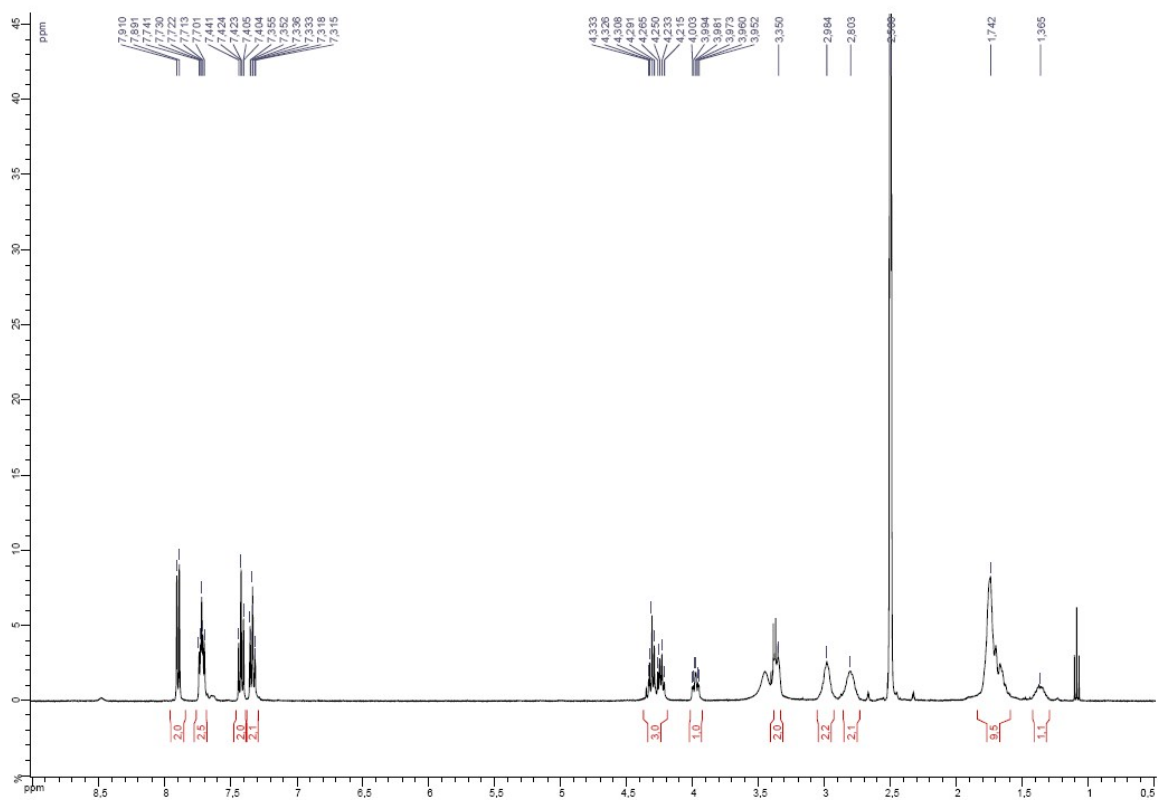
### **Reference**

1. Elhabazi, K., J.P. Humbert, I. Bertin, M. Schmitt, F. Bihel, J.J. Bourguignon, B. Bucher, J.A. Becker, T. Sorg, H. Meziane, B. Petit-Demouliere, B. Ilien, and F. Simonin, *Endogenous mammalian RF-amide peptides, including PrRP, kisspeptin and 26RFa, modulate nociception and morphine analgesia via NPPF receptors*. *Neuropharmacology*, 2013. 75C, 164-171.

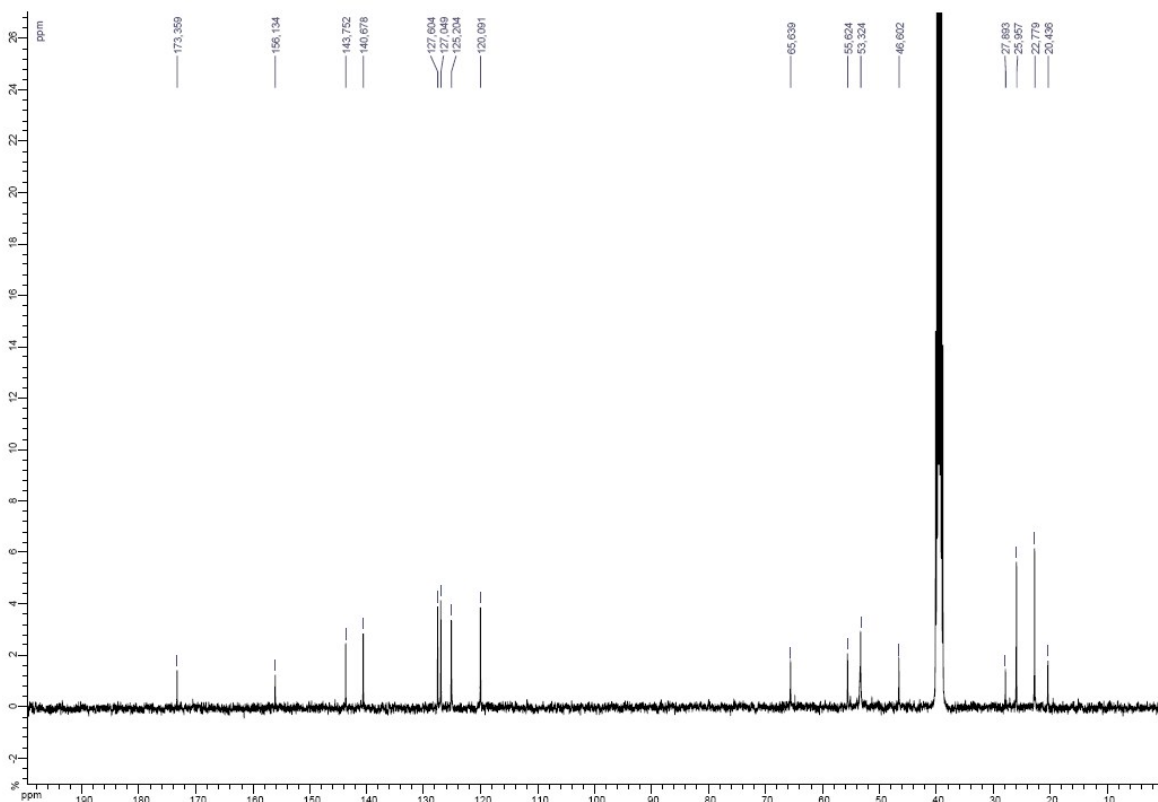
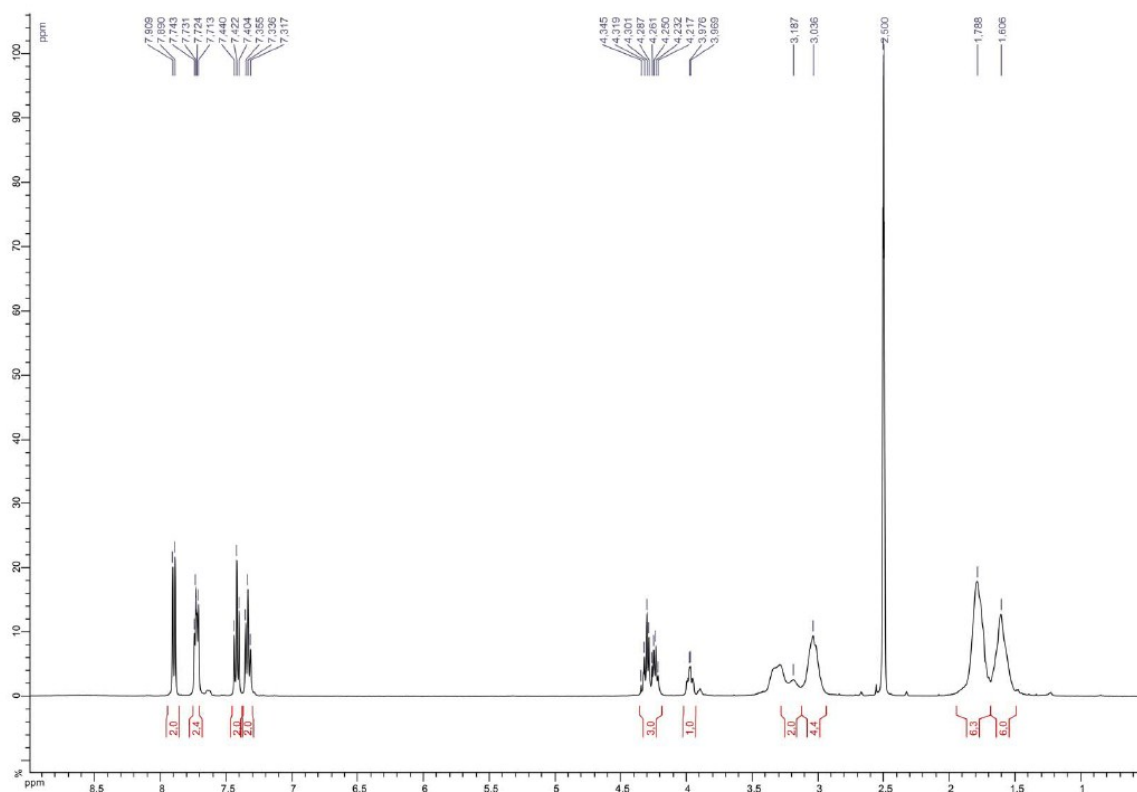
# Compound L-2a



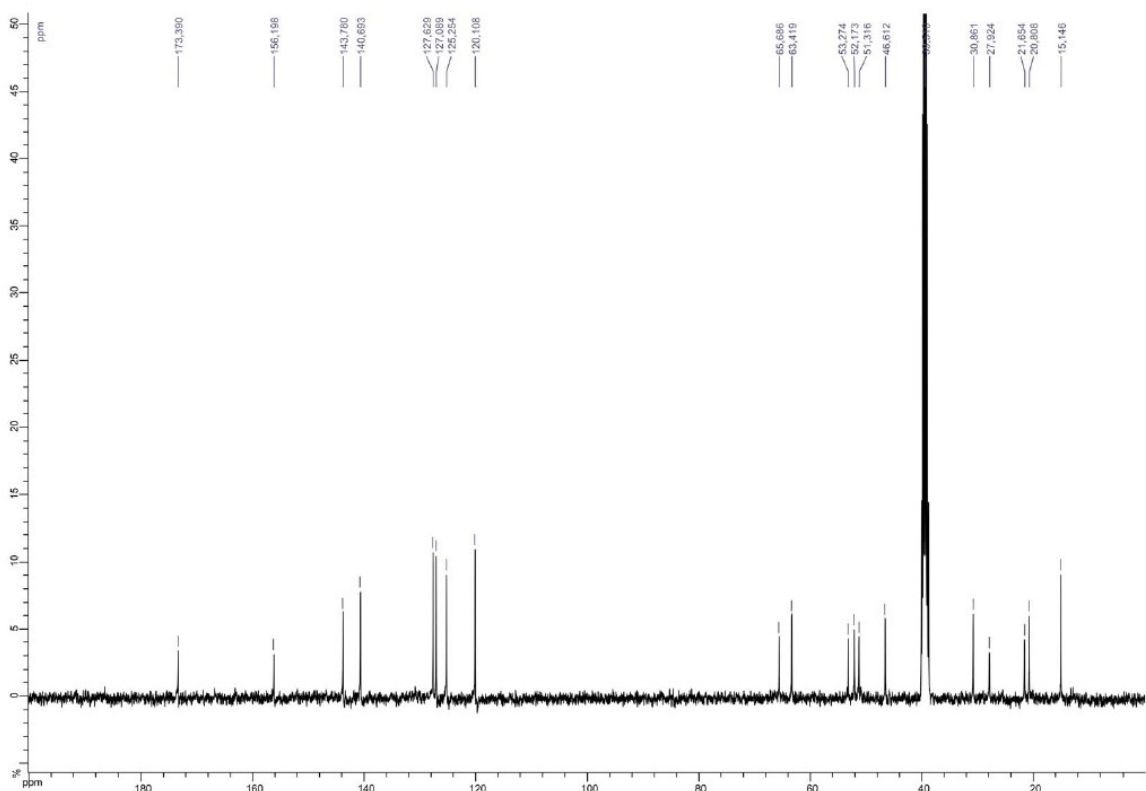
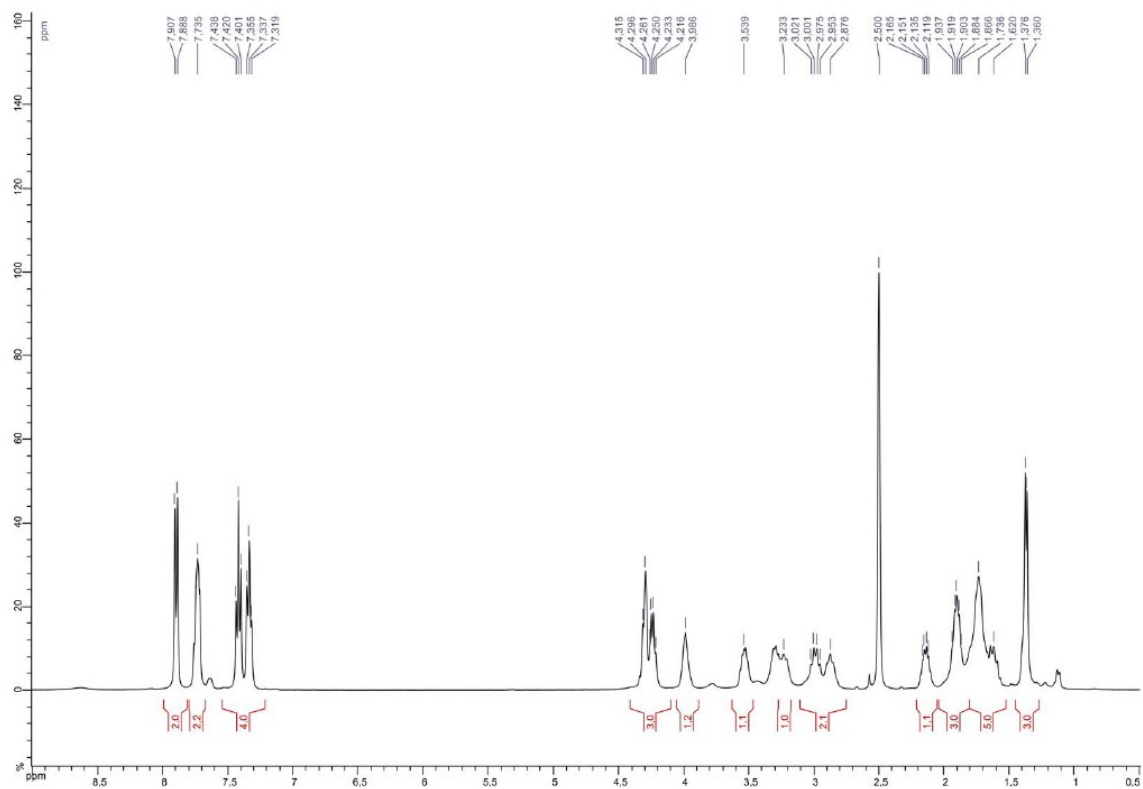
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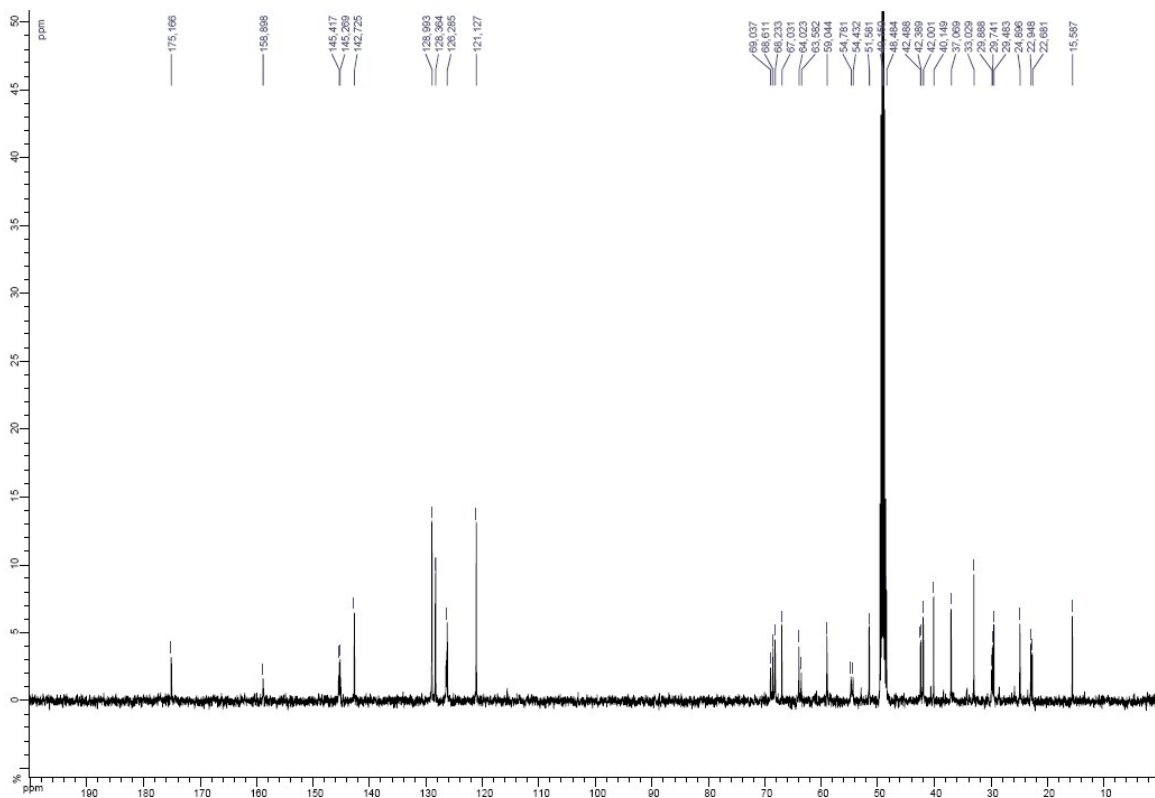
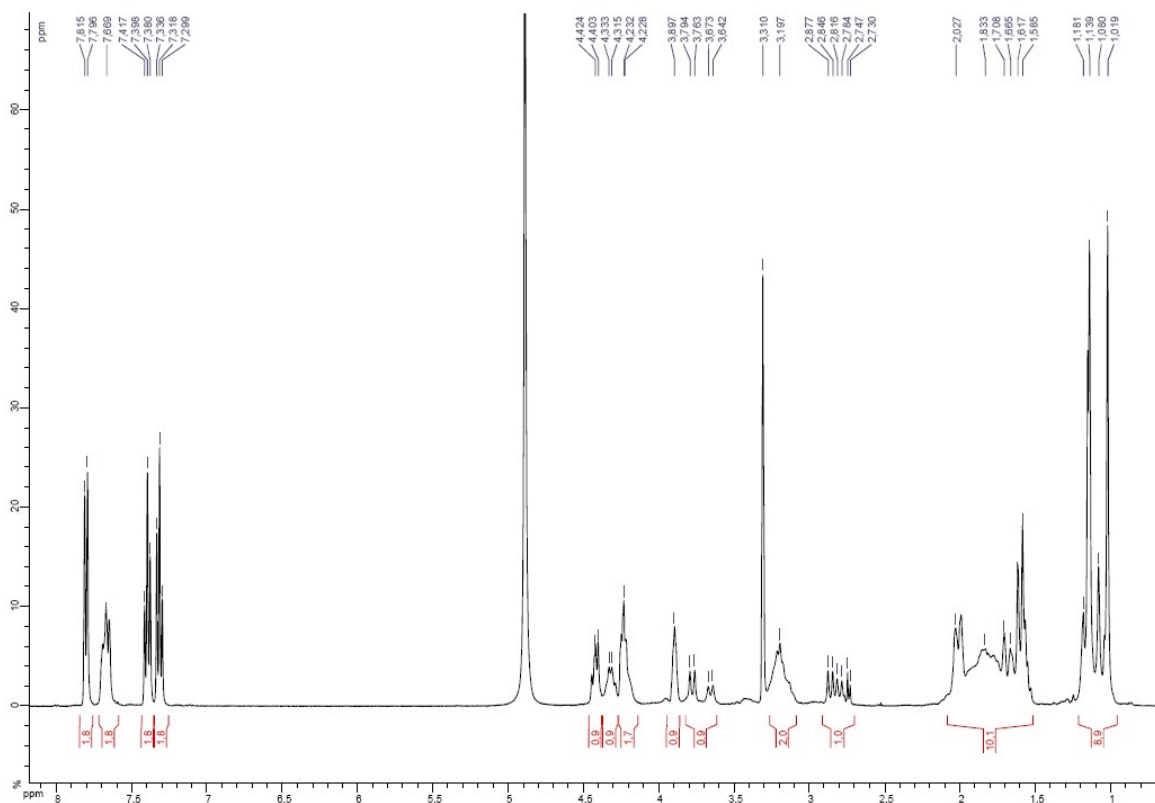
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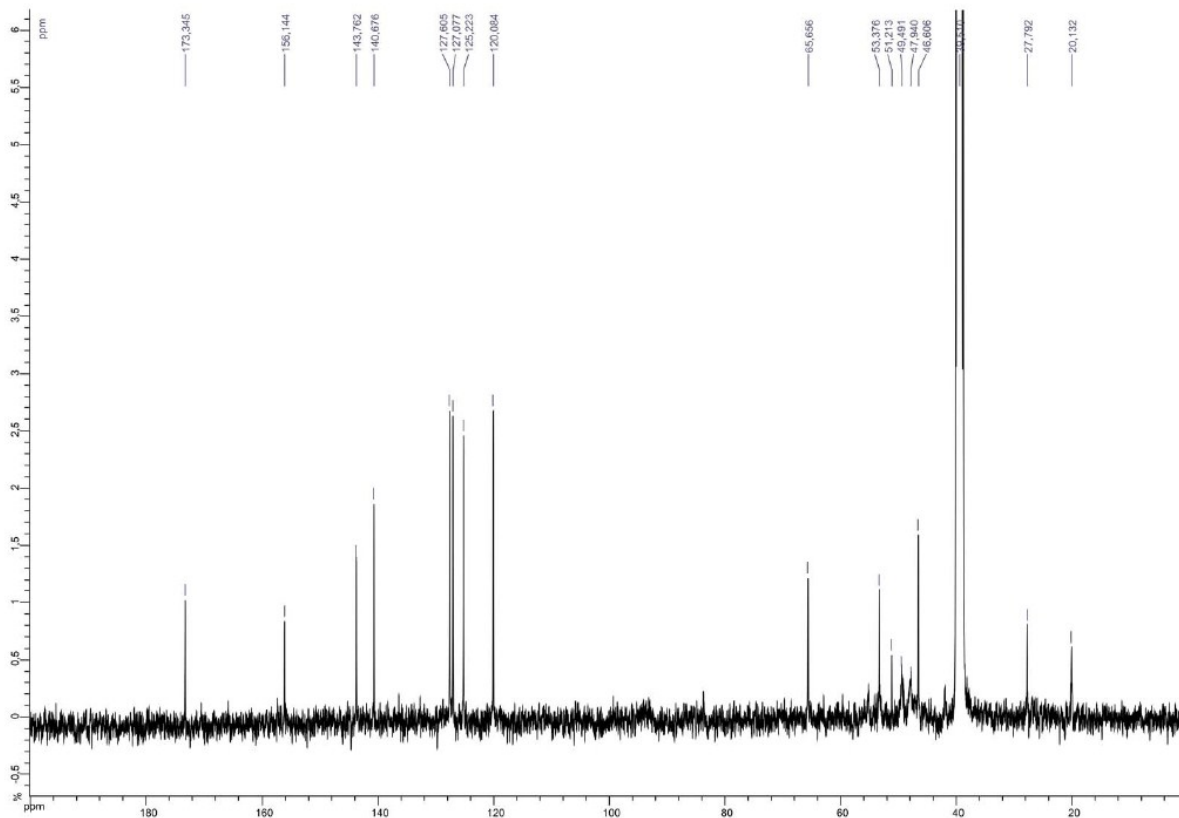
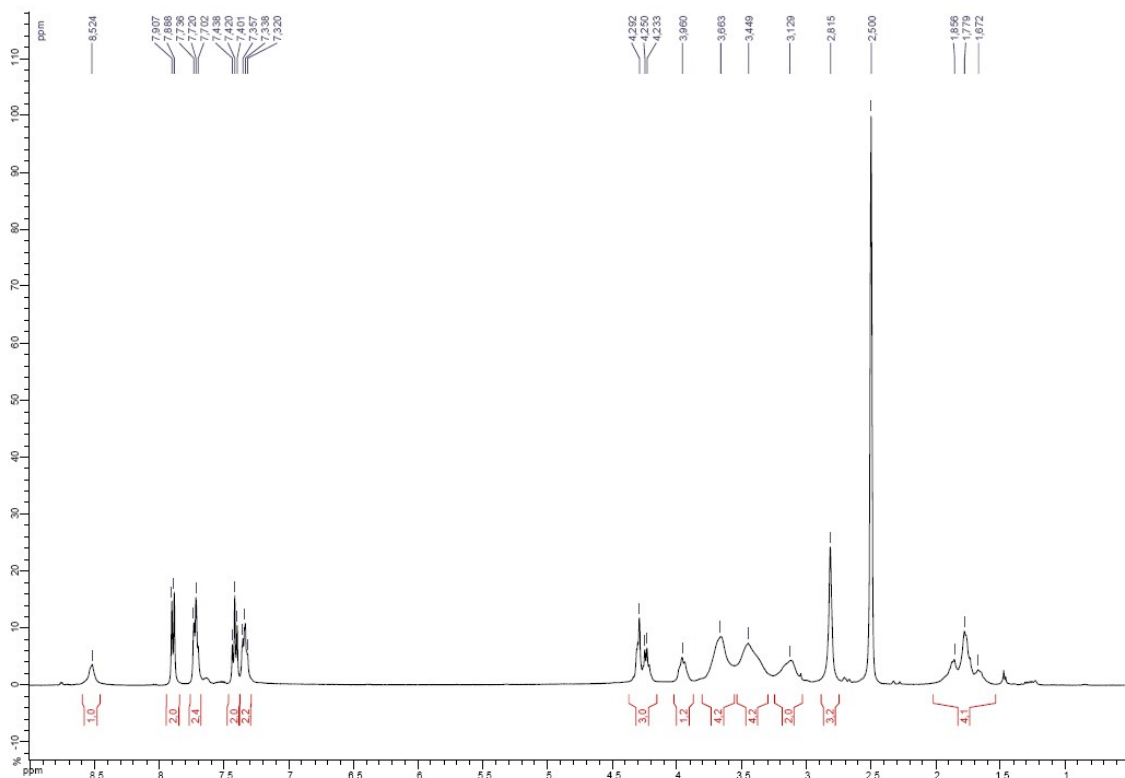
# Compound 2c



# Compound 2d

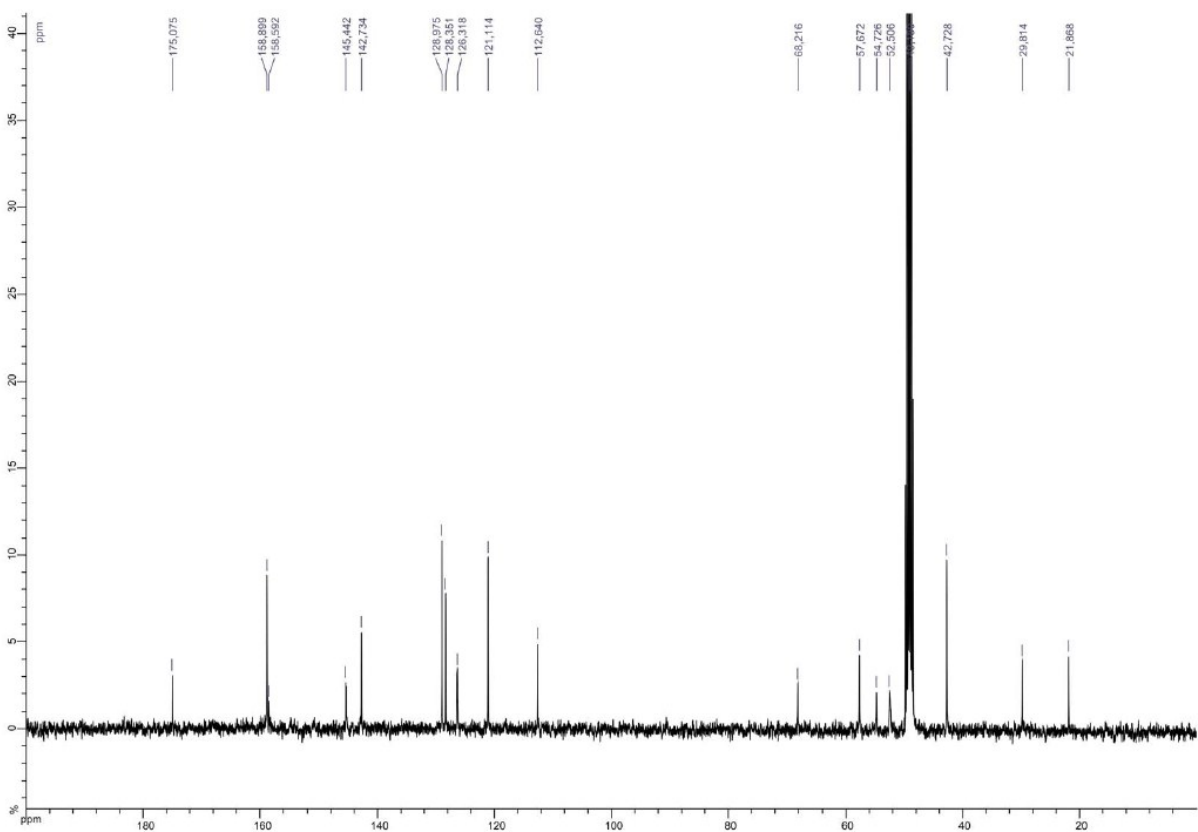
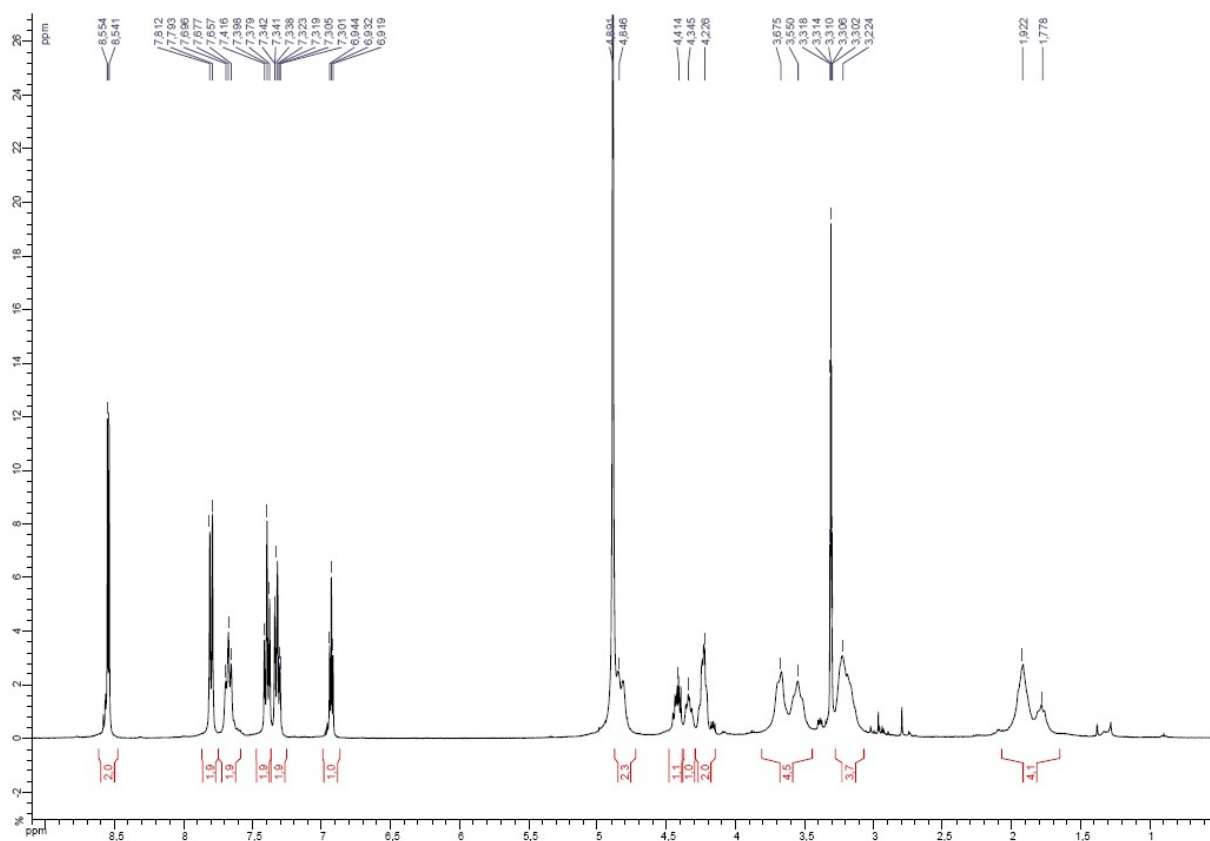


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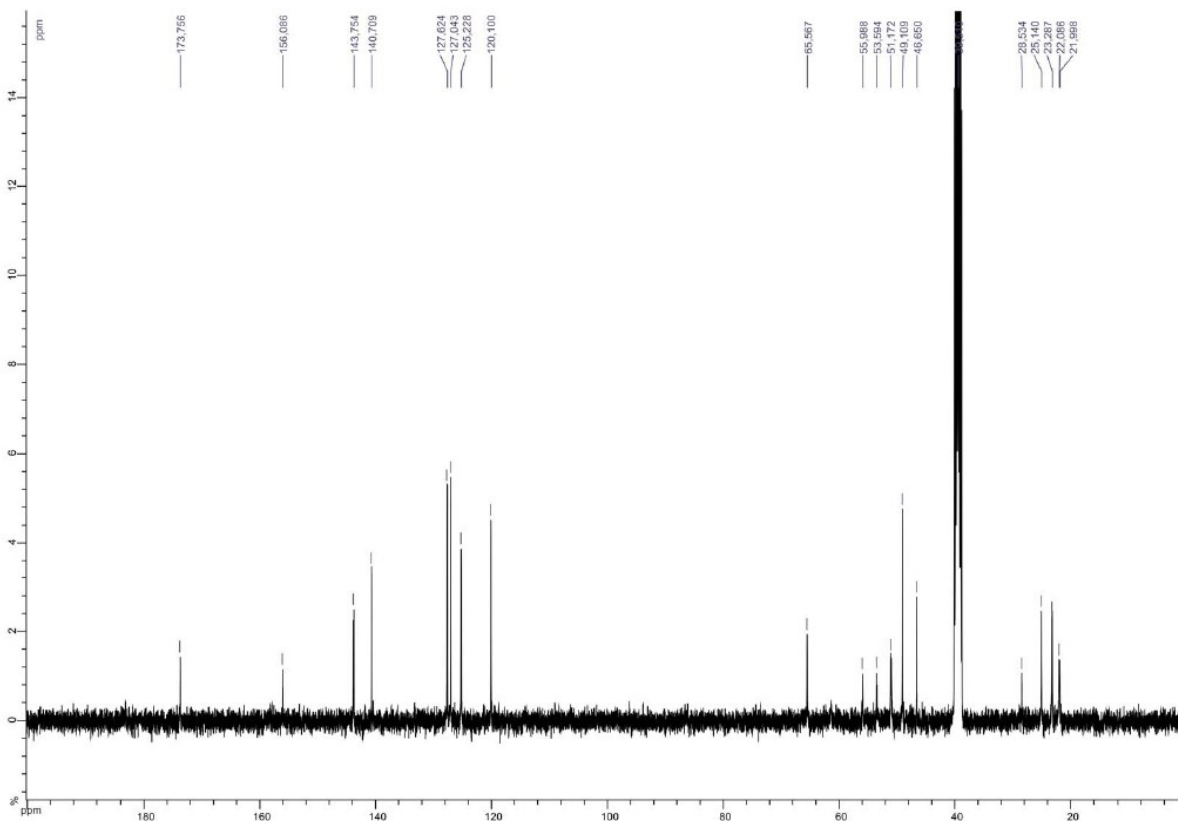
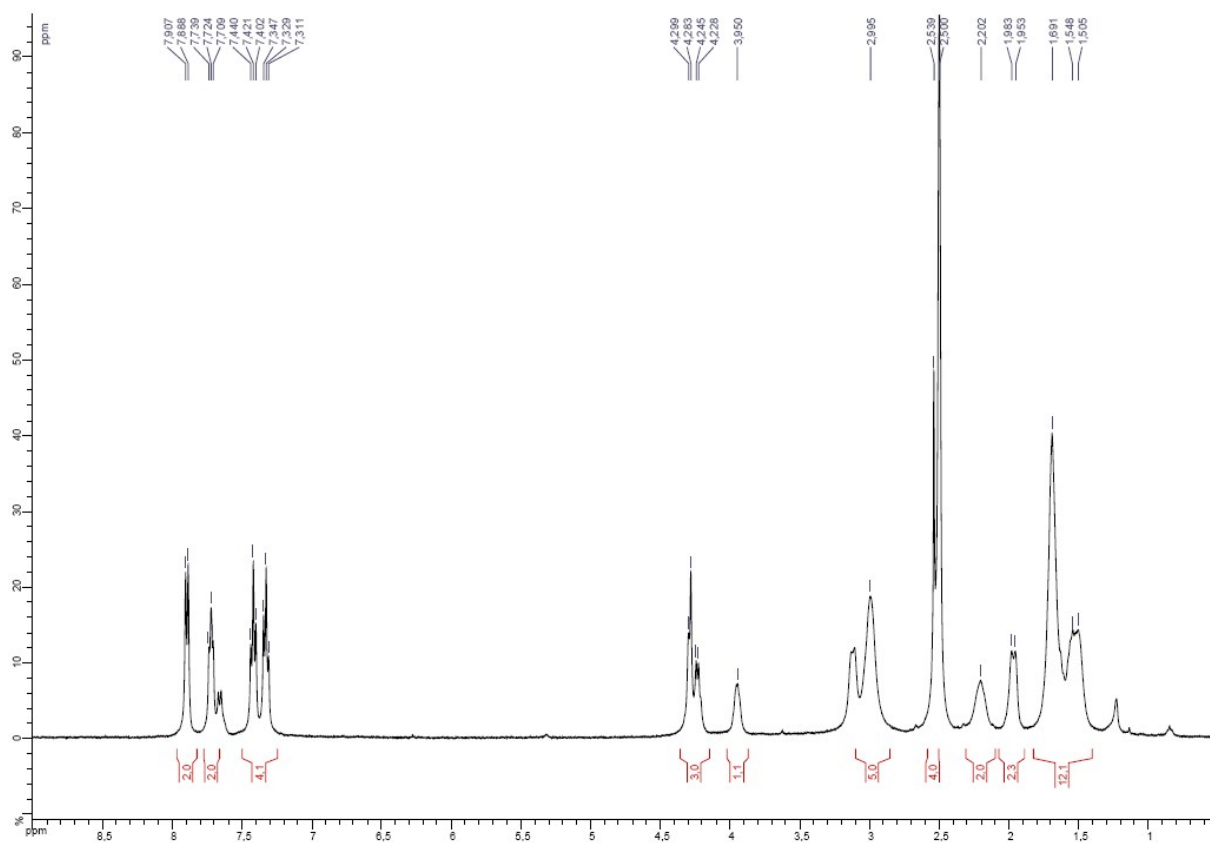




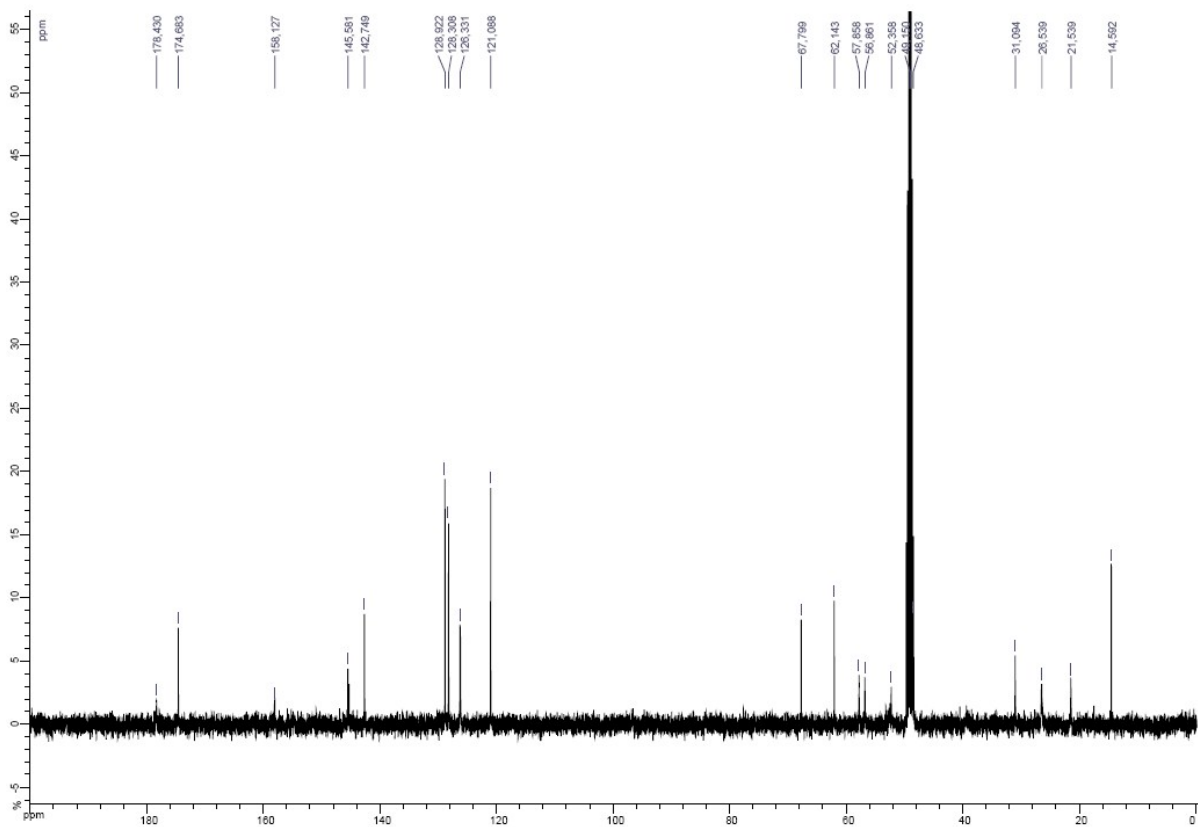
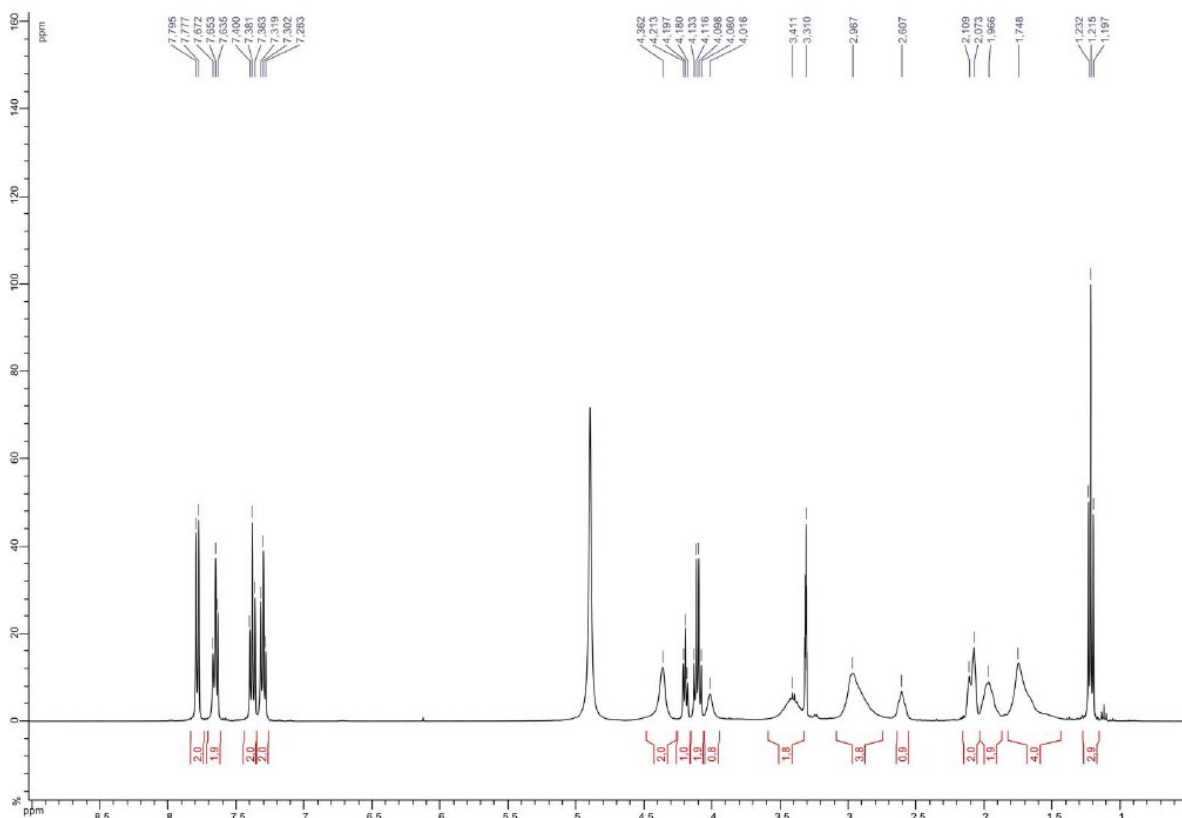
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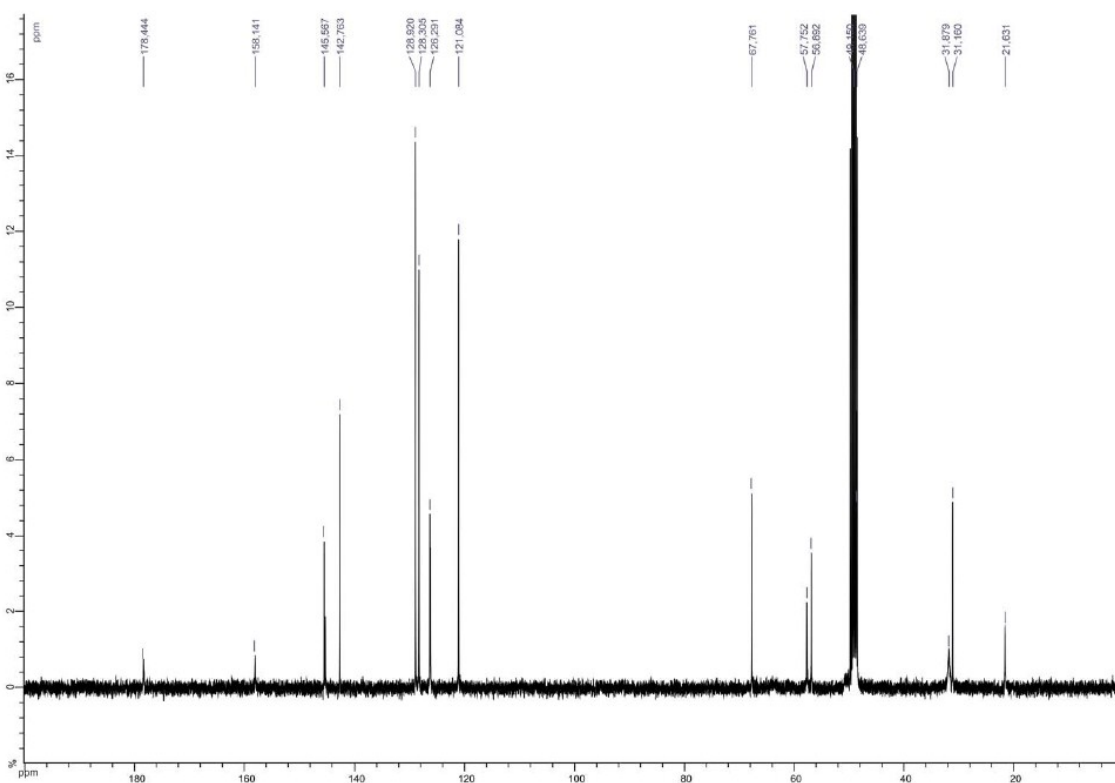
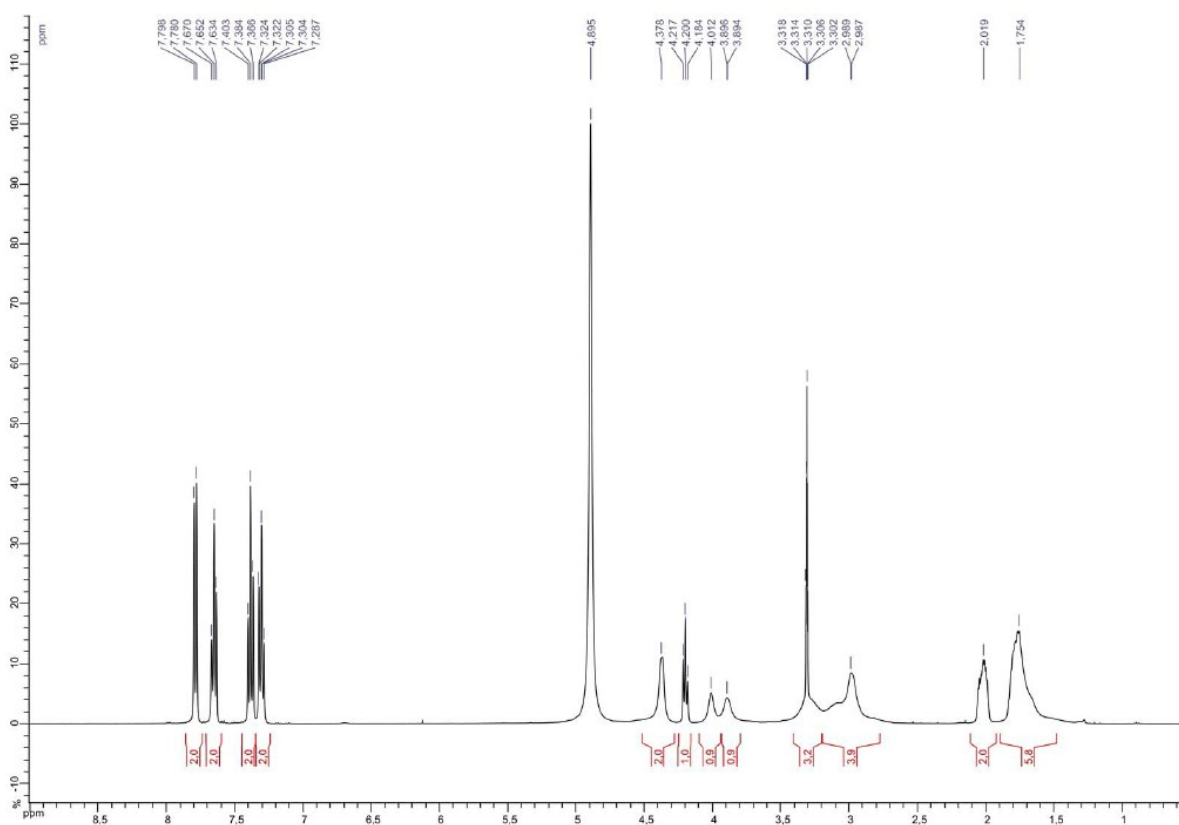
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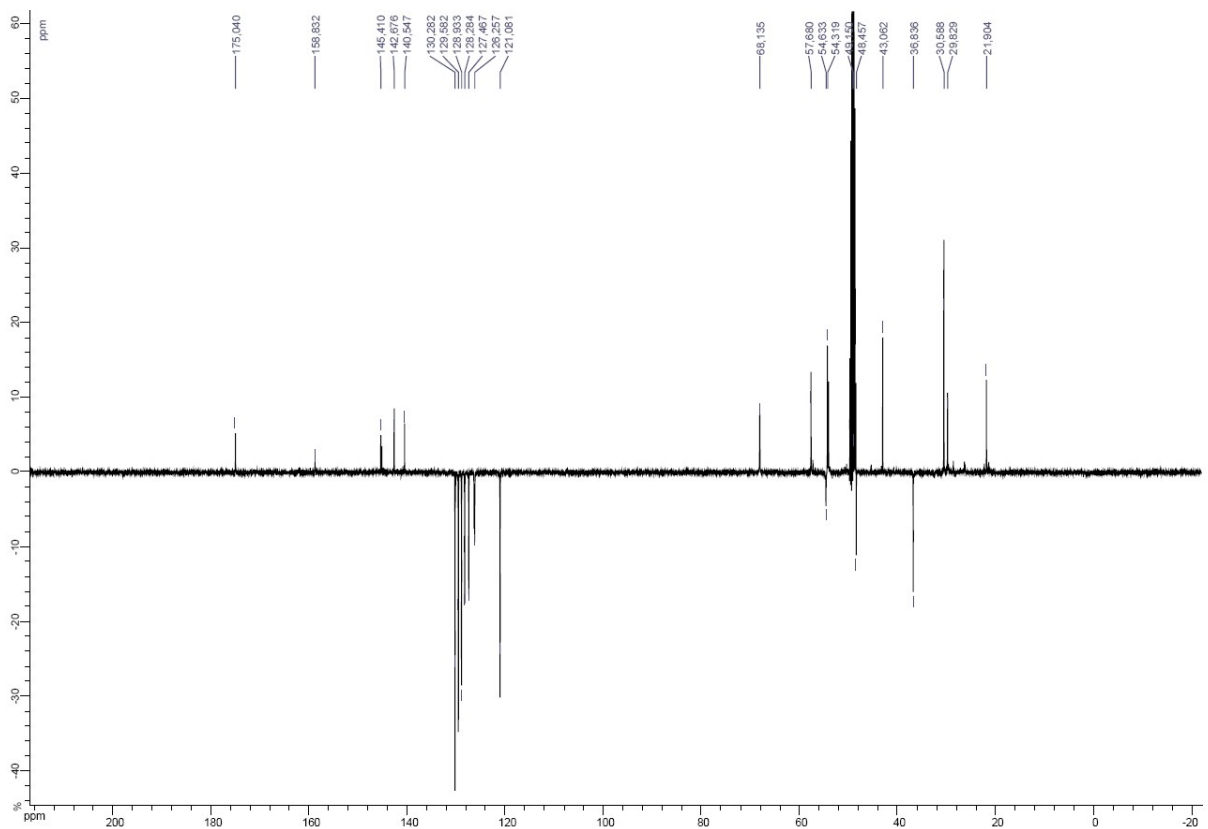
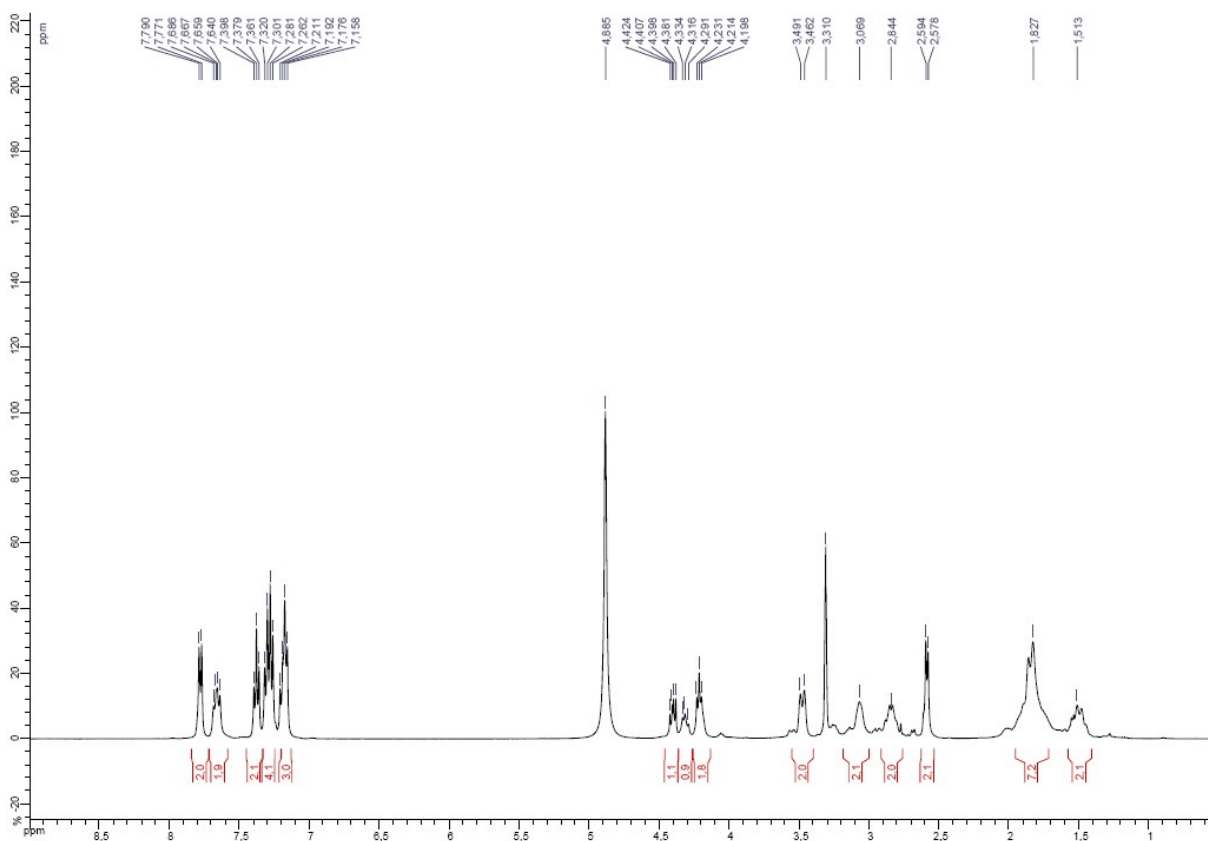
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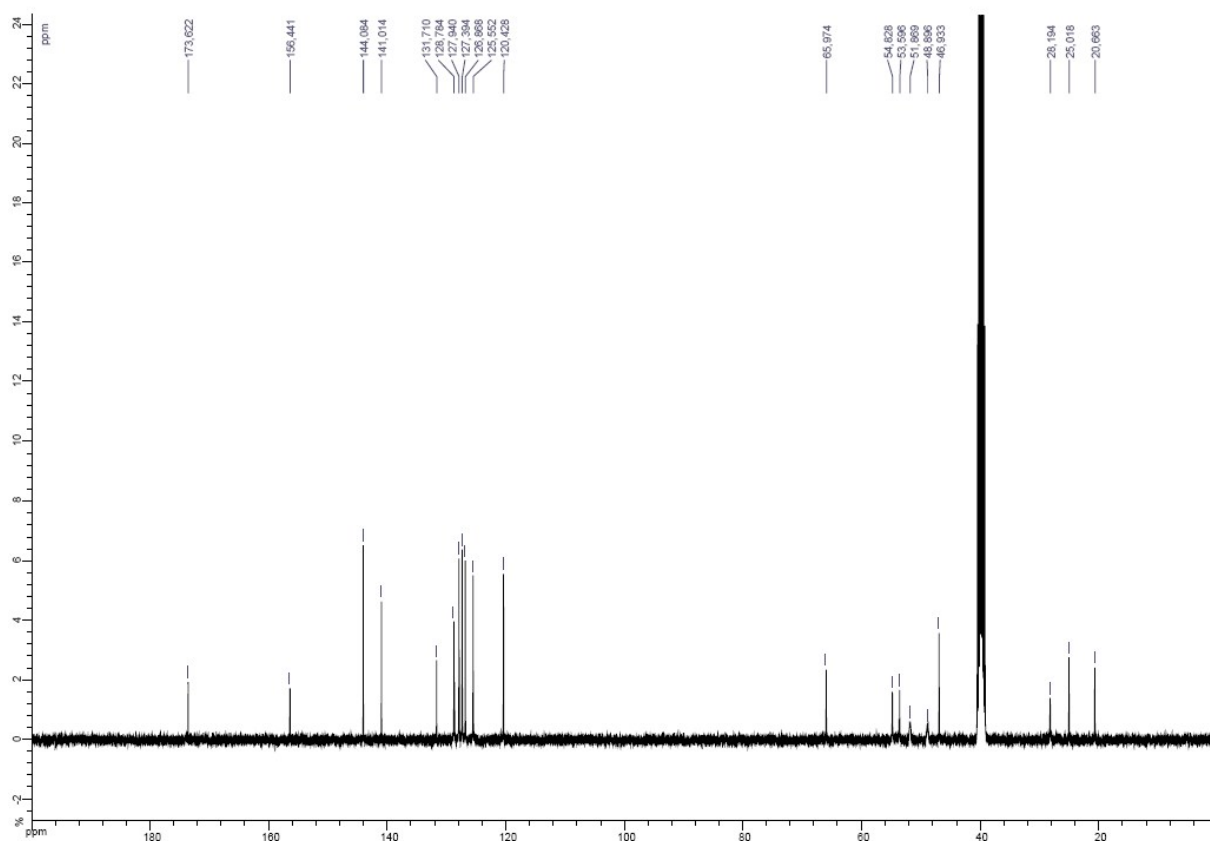
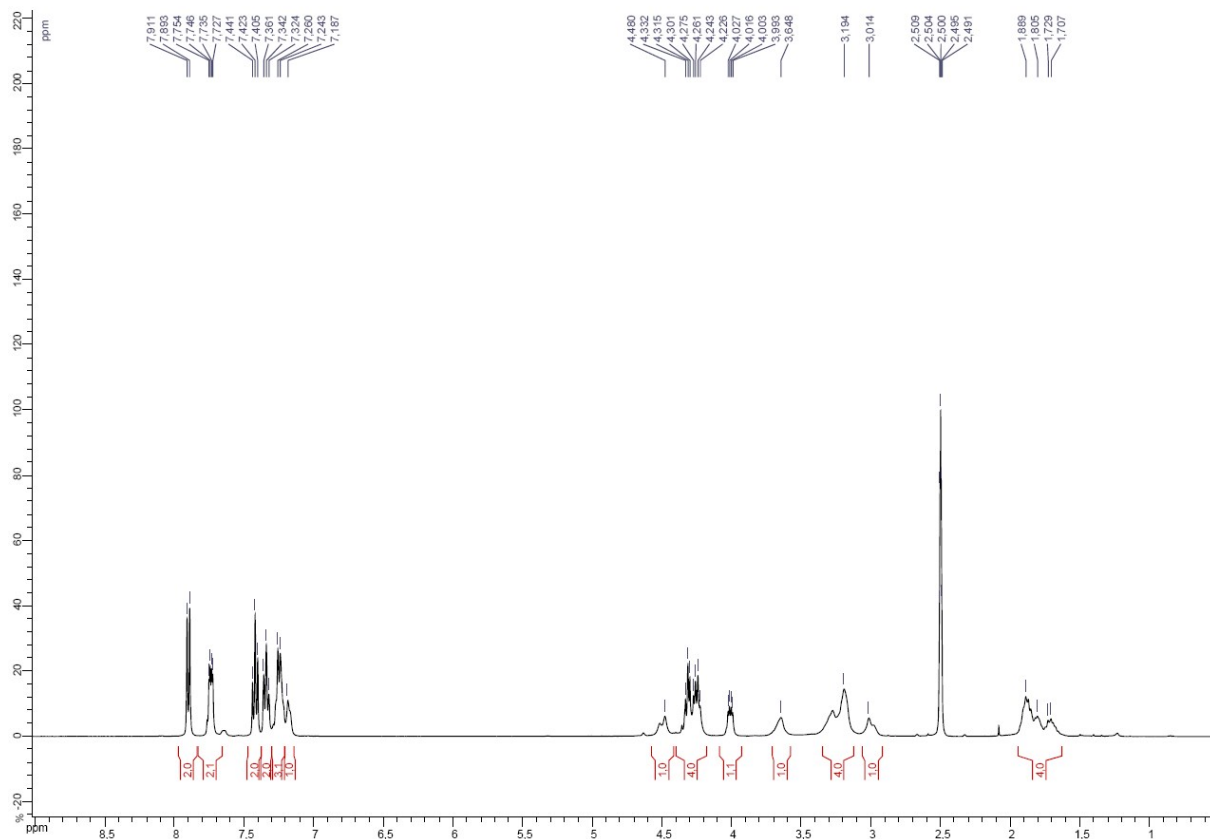
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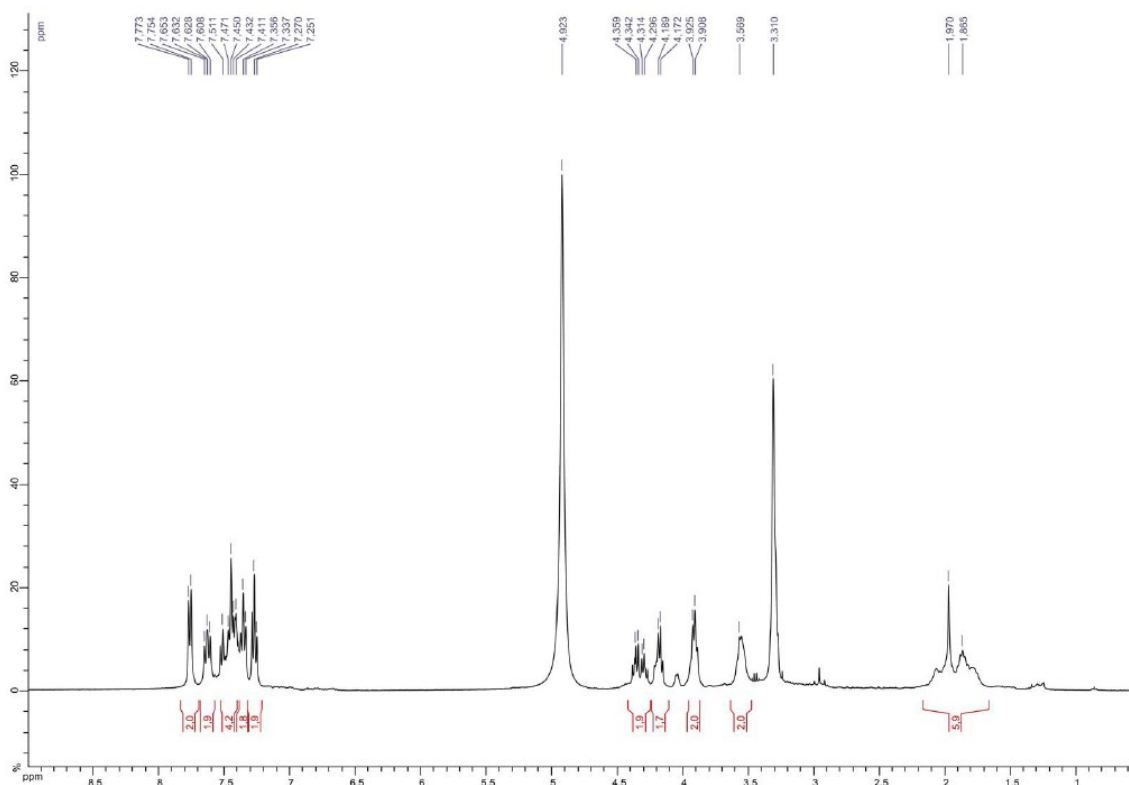
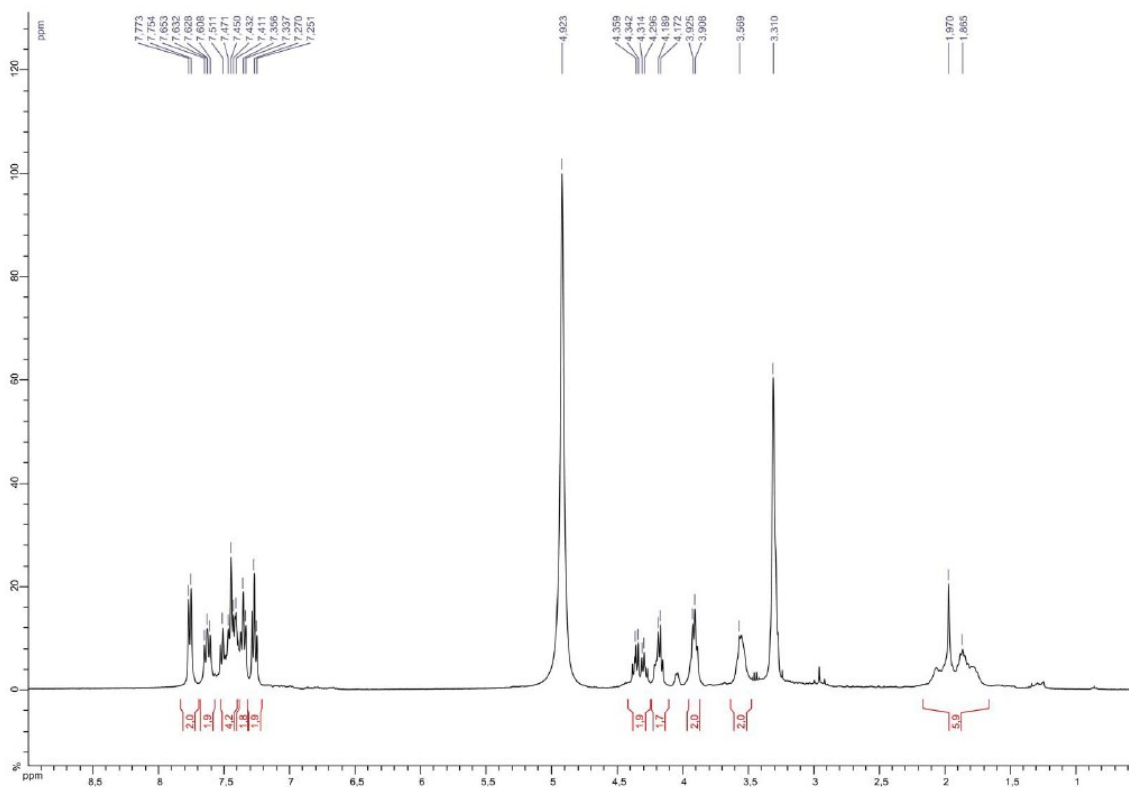
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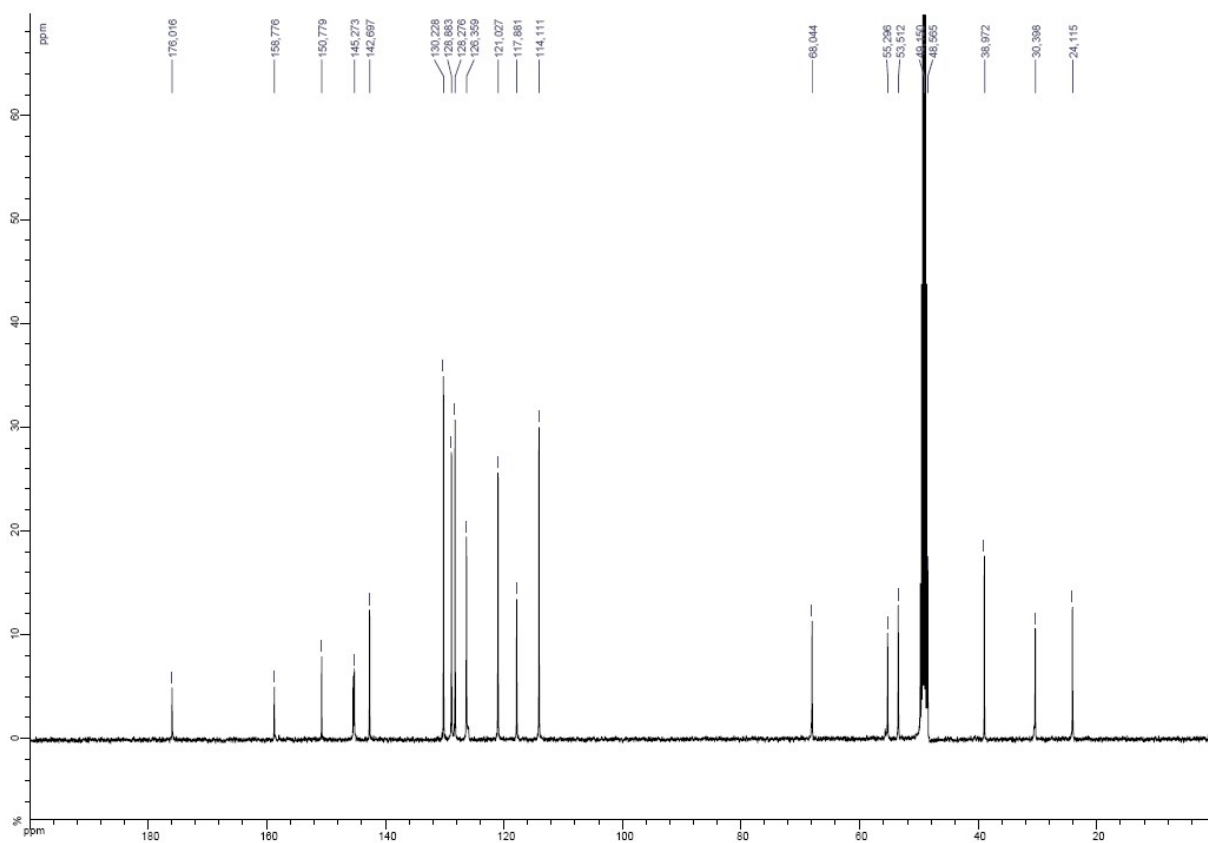
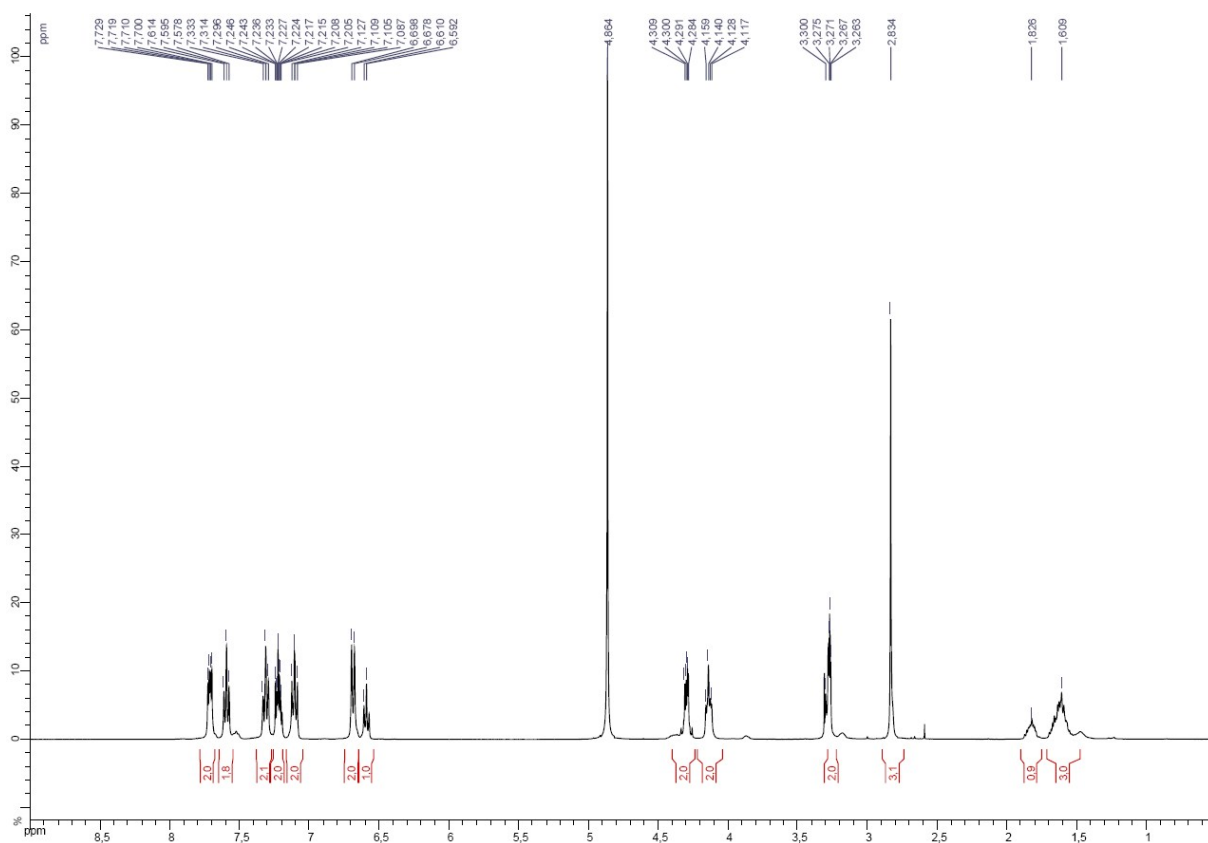
# Compound 2k



# Compound 2l

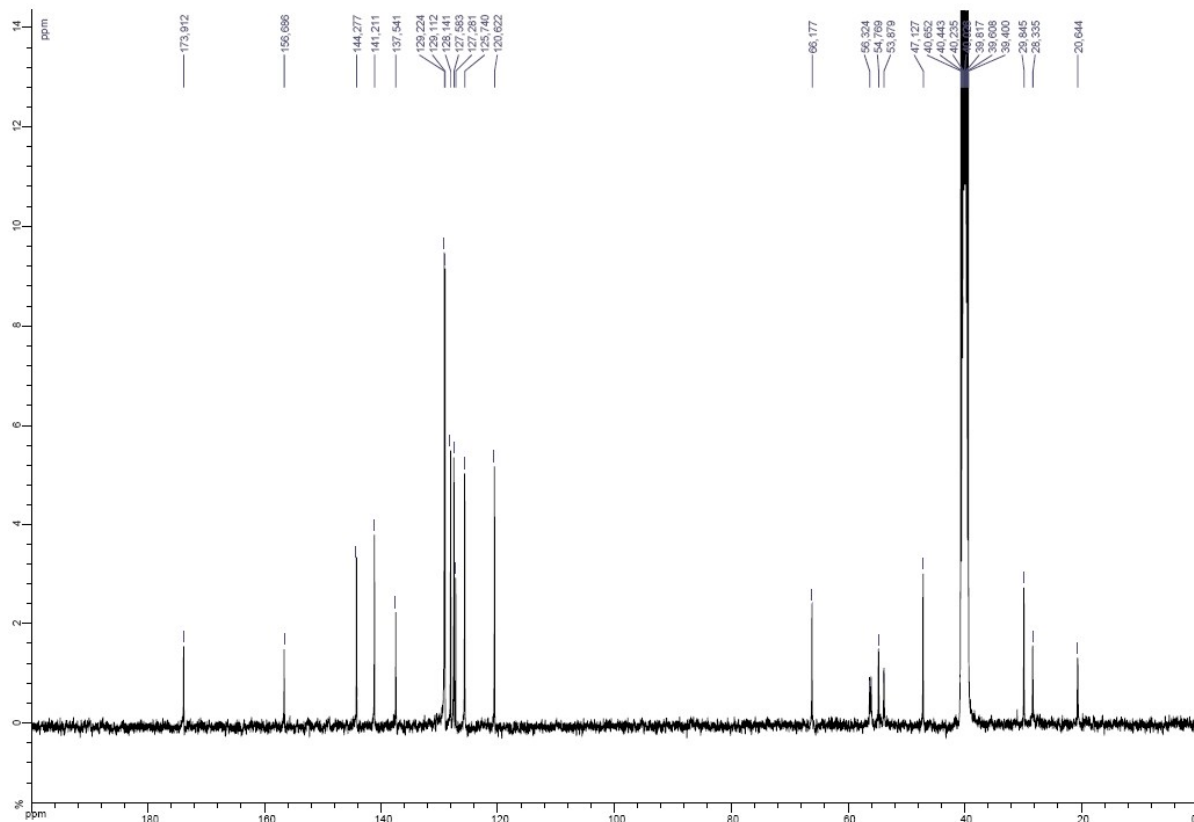
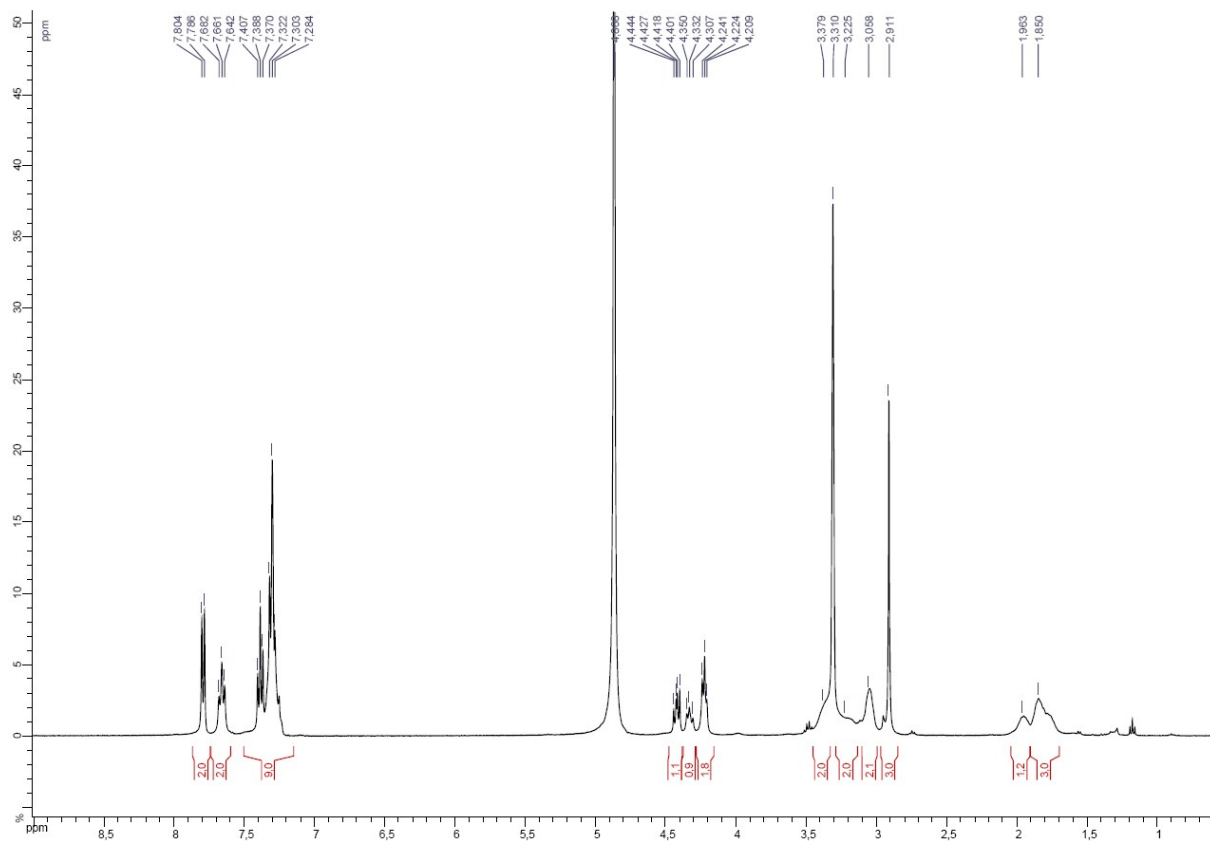


# Compound 2n

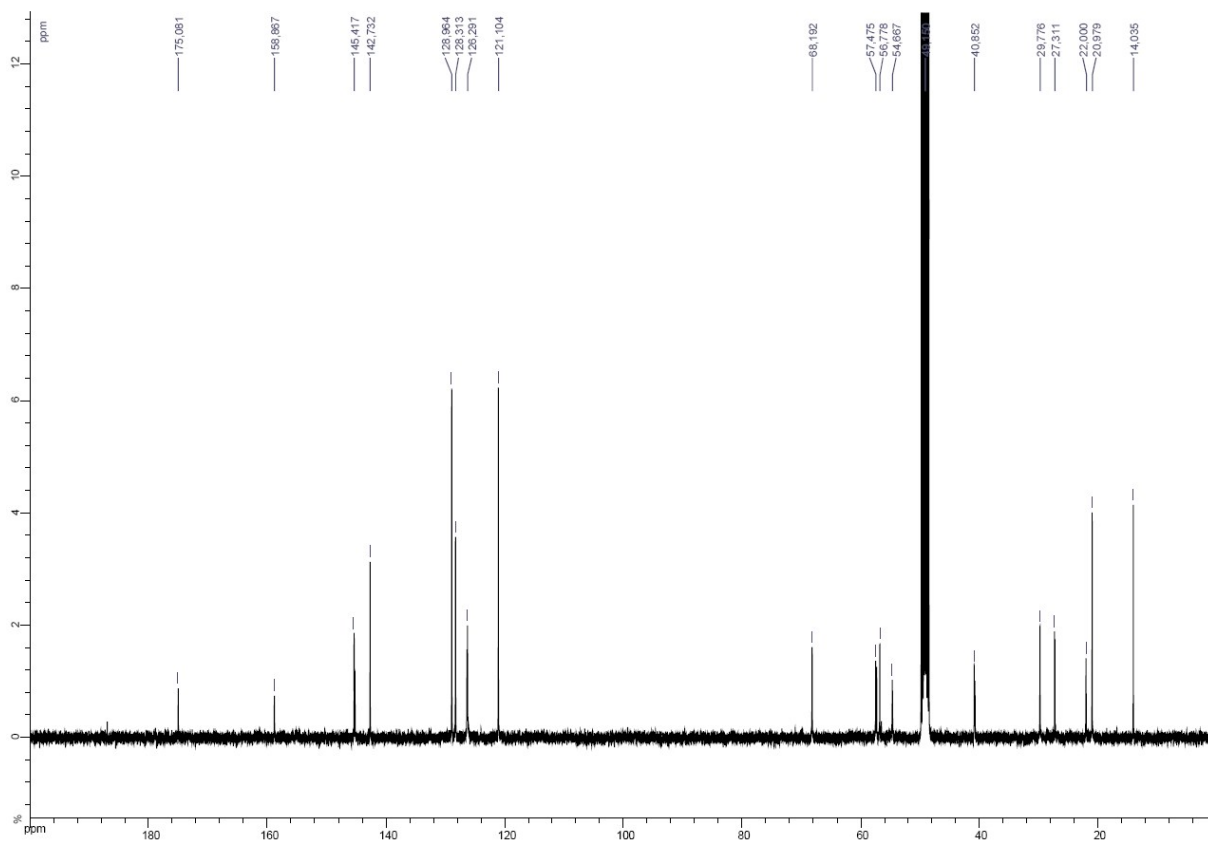
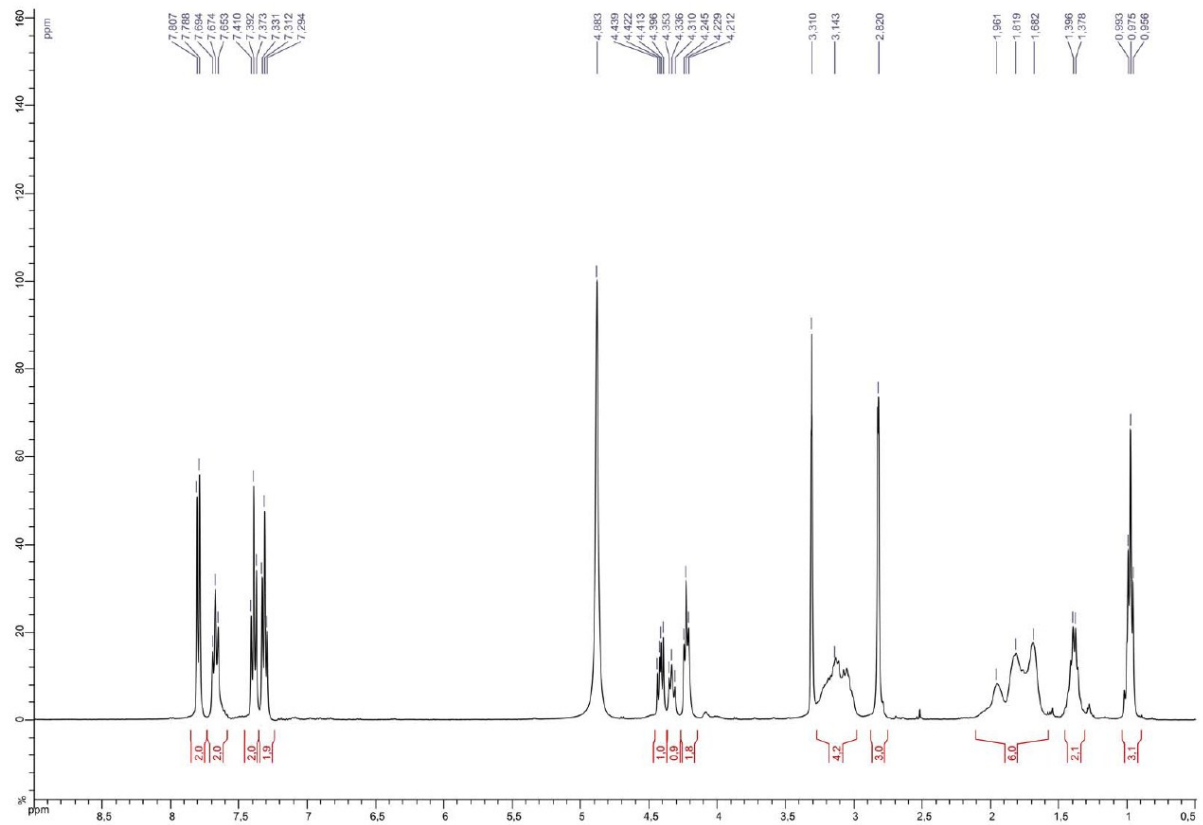




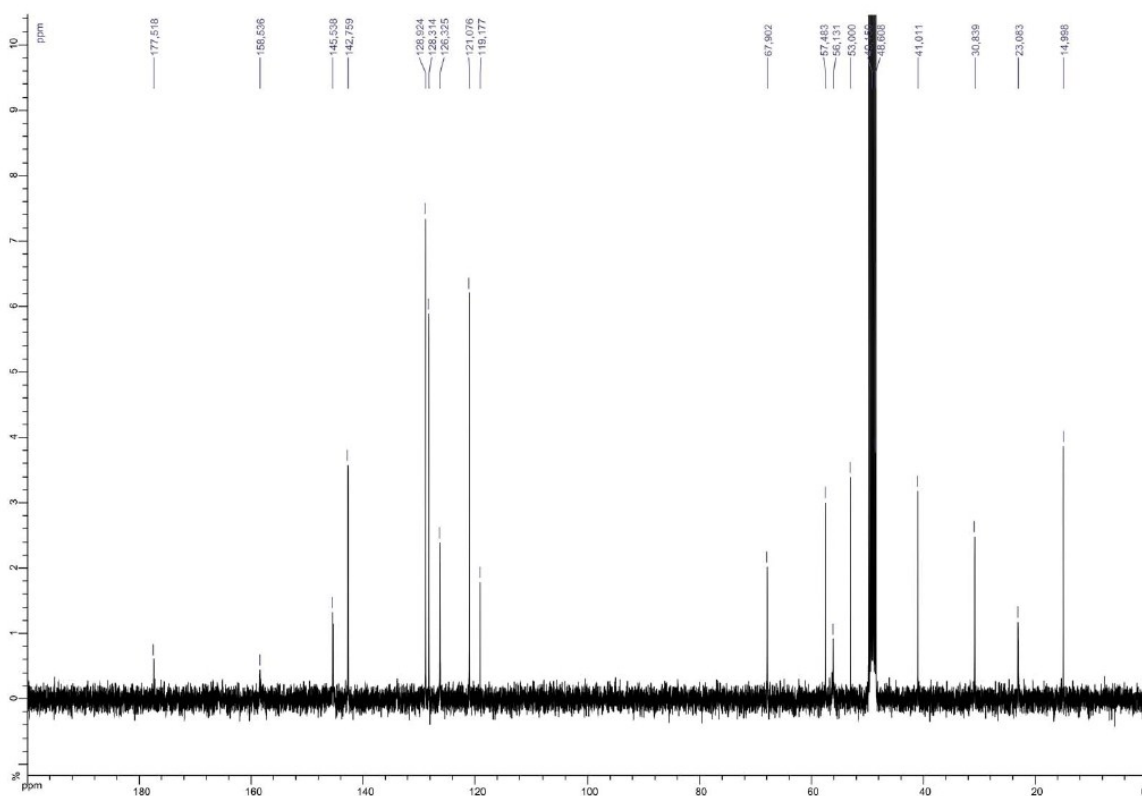
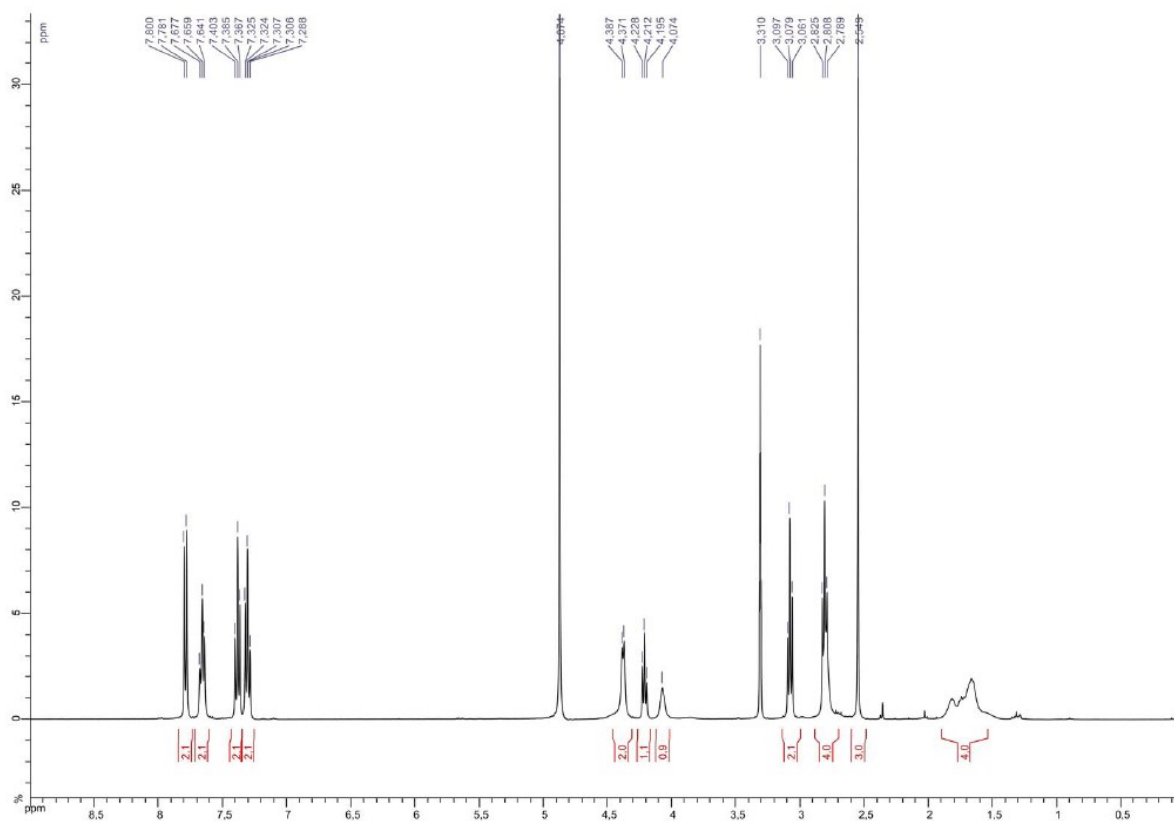
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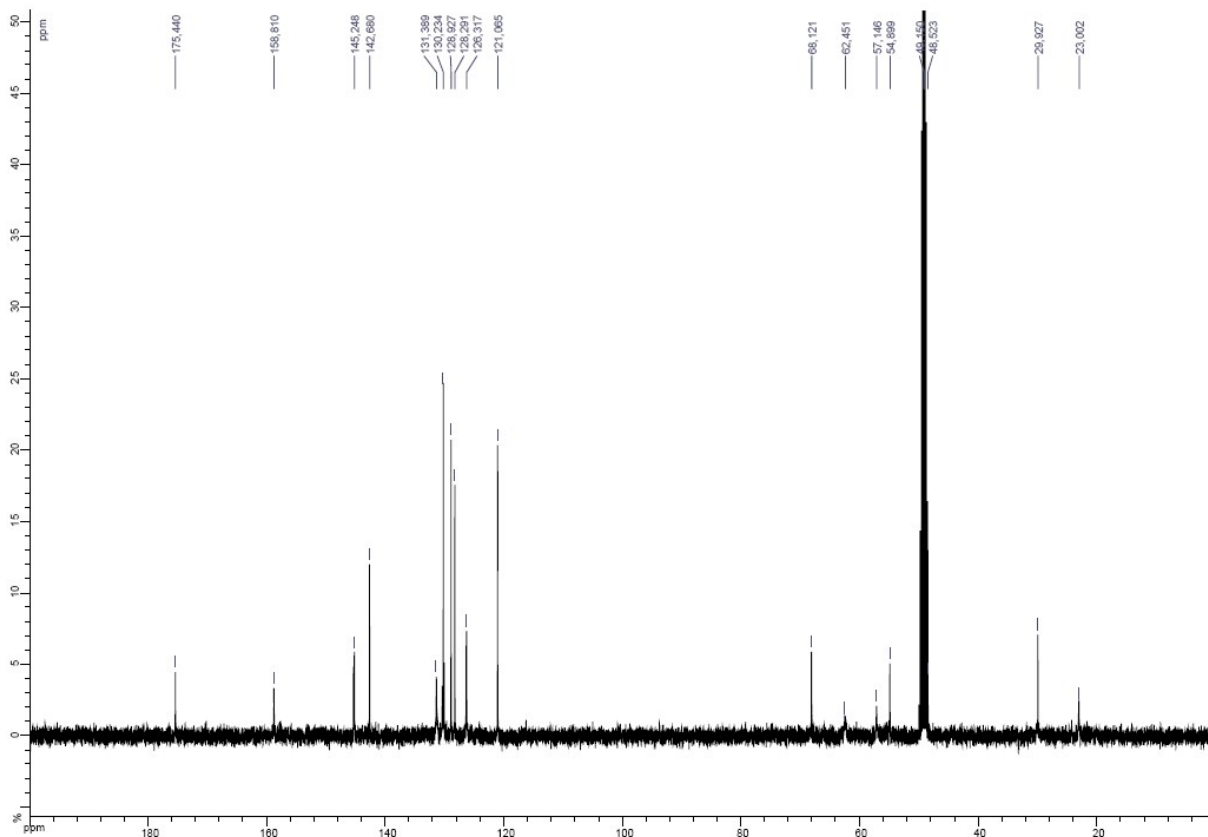
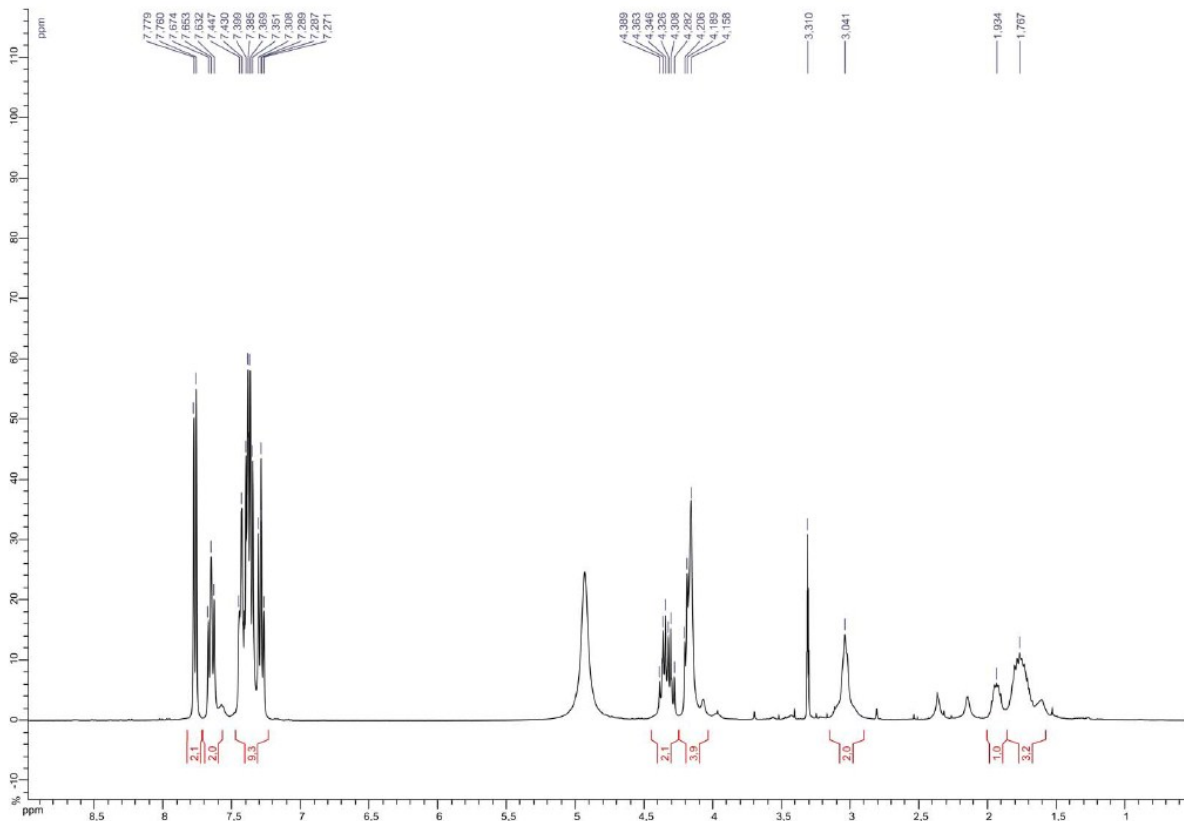
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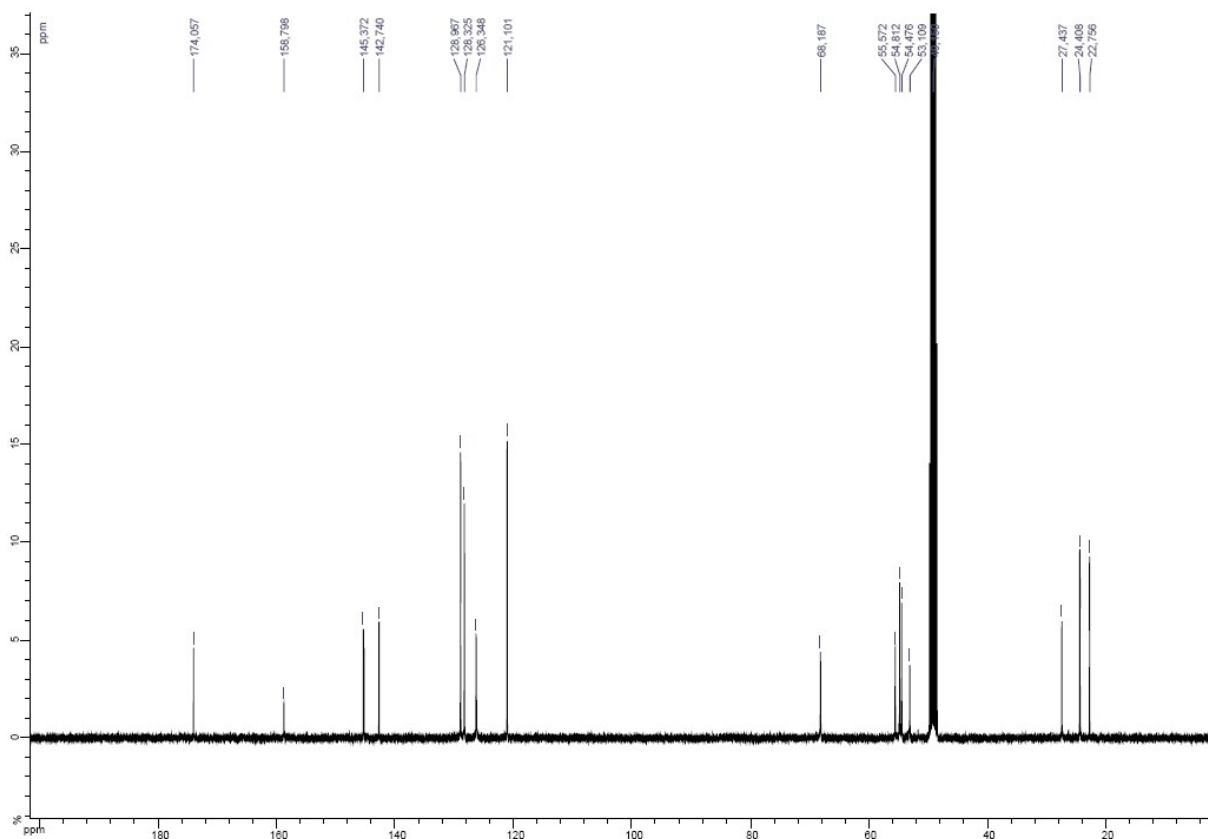
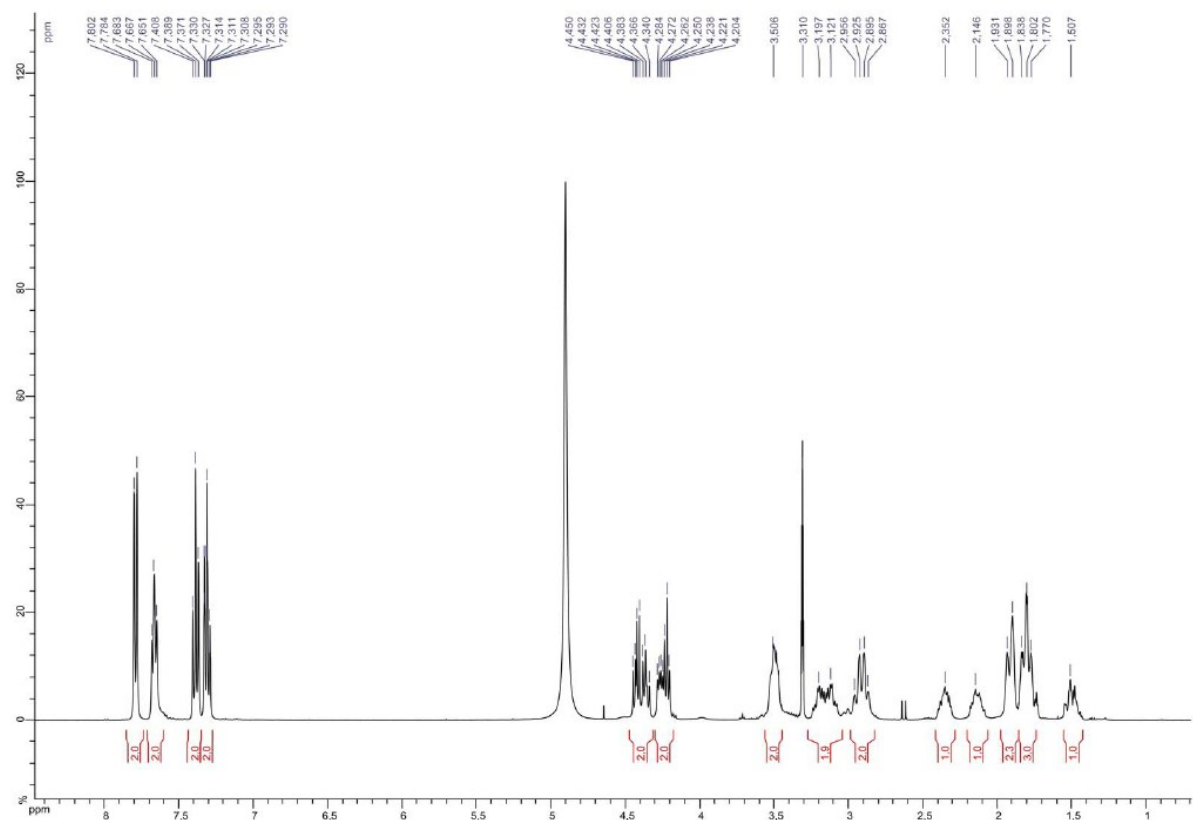
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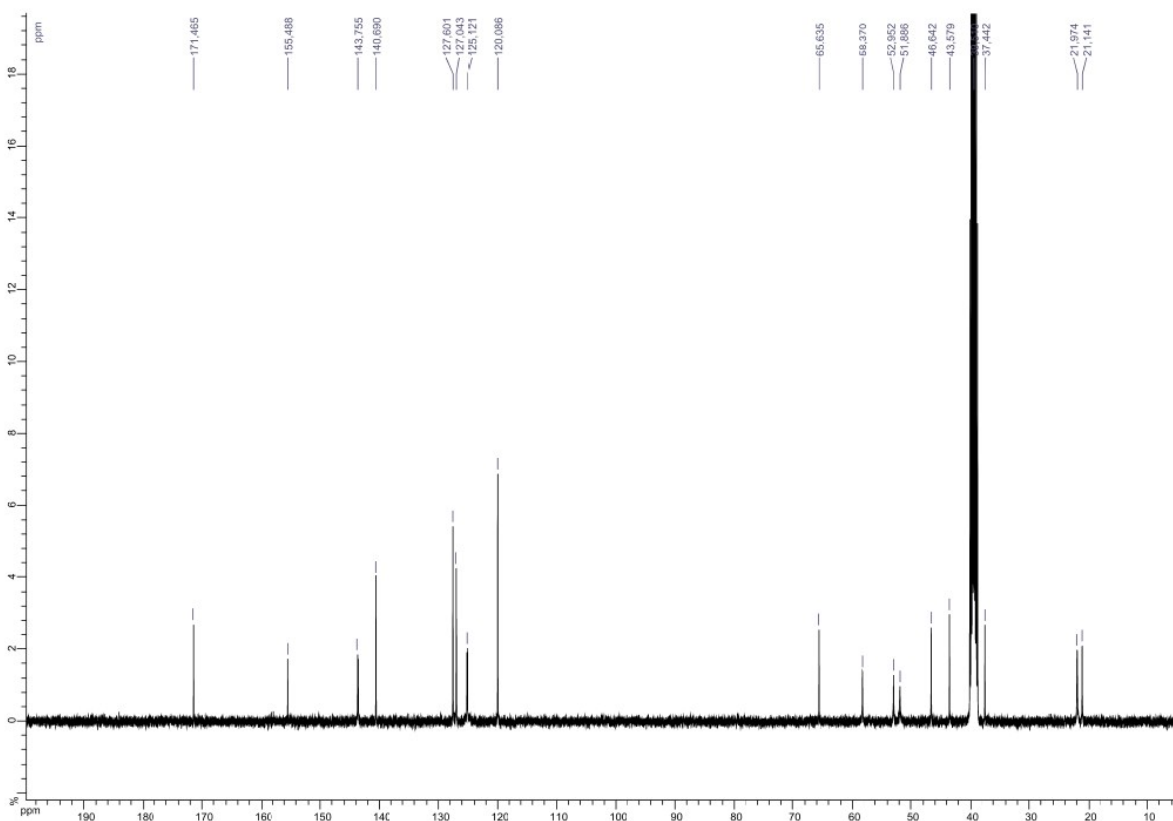
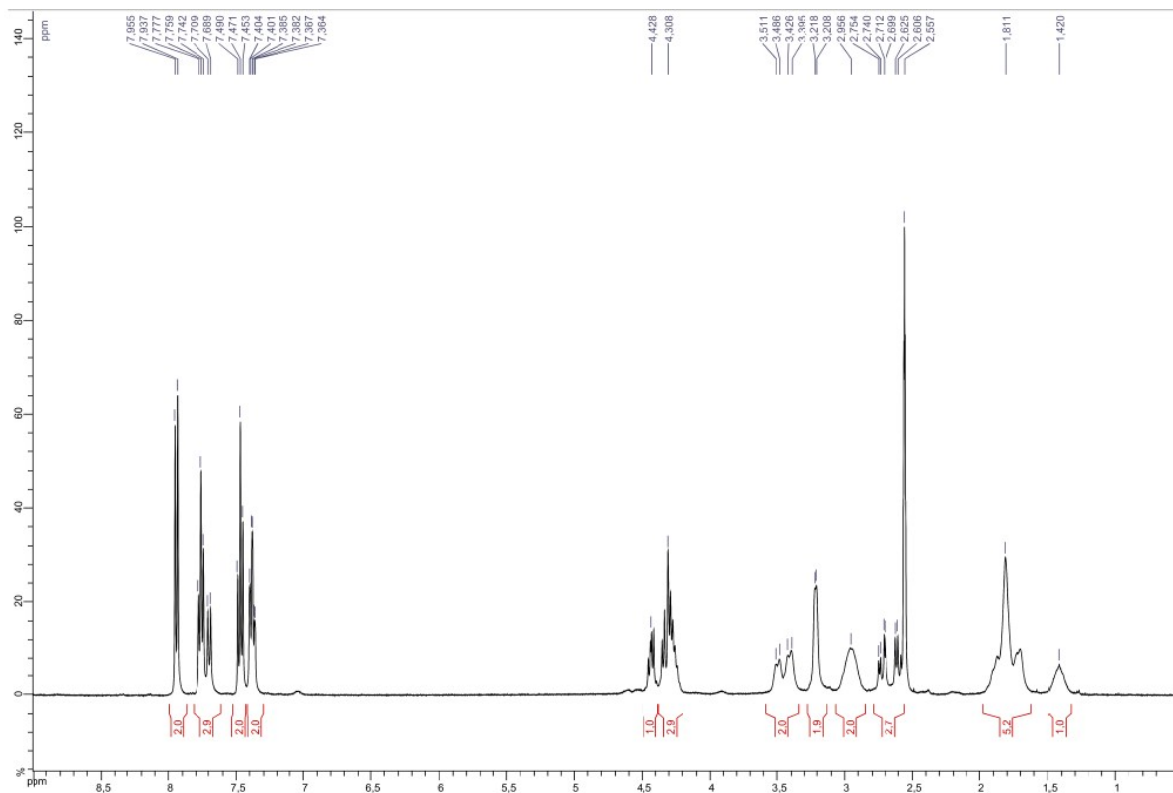
# Compound 2r



# Compound 4



# Compound 7



# Compound 8

