

Locked Nucleic Acid (LNA) induced effect on hybridization and fluorescence properties of oligodeoxyribonucleotides modified with nucleobase-functionalized DNA monomers

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ELECTRONIC SUPPLEMENTARY INFORMATION

Table of contents

MALDI-ToF MS of modified 9-mer ONs (Table S1)	S2
Representative thermal denaturation curves (Figure S1)	S3
Absorption spectra for pyrene-modified ONs and duplexes (Figures S2-S5)	S4
Biophysical characterization of 13-mer Y/M -modified ONs	S8
MALDI-ToF MS of modified 13-mer ONs (Table S2)	S8
Thermal denaturation data for 13-mer Y - or M -modified ONs and duplexes (Tables S3 and S4 S6)	S9
Absorption spectra for 13-mer Y -modified ONs and duplexes (Figure S6)	S10
Absorption maxima for 13-mer Y -modified ONs and duplexes (Table S5)	S11
Absorption spectra for 13-mer M -modified ONs and duplexes (Figure S7)	S12
Absorption maxima for 13-mer M -modified ONs and duplexes (Table S6)	S13
Fluorescence emission spectra for 13-mer Y -modified ONs and duplexes (Figure S8)	S14
Fluorescence emission spectra for 13-mer M -modified ONs and duplexes (Figure S9)	S15
NMR spectra for nucleosides	S16

Table S1. MALDI-ToF MS of modified 9-mer ONs.^a

(M+H)				(M+H)			
ON	Sequence	CALC	OBS	ON	Sequence	CALC	OBS
T2	5'-GTG aTa TGC	2809.5	2810.7	A5	5'-GCA tAt CAC	2739.5	2738.2
T3	5'-GT g ATA tGC	2809.5	2810.6	A6	5'-GC a TAT cAC	2752.5	2751.2
W1	5'-GTG AWA TGC	2762.5	2763.6	L4	5'-GCA TLT CAC	2706.5	2706.3
W2	5'-GTG aWa TGC	2818.5	2819.6	L5	5'-GCA tLt CAC	2775.0	2776.0
W3	5'-GT g AWA tGC	2818.5	2819.6	L6	5'-GC a TLT cAC	2761.5	2762.3
X1	5'-GTG AXA TGC	2791.5	2793.0	M4	5'-GCA TMT CAC	2907.7	2906.7
X2	5'-GTG aXa TGC	2847.5	2848.0	M5	5'-GCA tMt CAC	2977.0	2977.0
X3	5'-GT g AXA tGC	2847.5	2848.7	M6	5'-GC a TMT cAC	2964.7	2964.0
Y1	5'-GTG AYA TGC	2964.7	2962.1	N4	5'-GCA TNT CAC	2962.6	2963.3
Y2	5'-GTG aYa TGC	3020.7	3019.6	N5	5'-GCA tNt CAC	3033.6	3033.3
Y3	5'-GT g AYA tGC	3020.7	3020.0	N6	5'-GC a TNT cAC	3018.6	3021.0
Z1	5'-GTG AZA TGC	3019.6	3020.7				
Z2	5'-GTG aZa TGC	3075.5	3076.6				
Z3	5'-GT g AZA tGC	3075.5	3076.6				

^a A/C/G/T = adenin-9-yl/cytosin-1-yl/guanin-9-yl/thymin-1-yl DNA monomers. For structures of monomers **W/X/Y/Z** and **L/M/N**, see Figure 1 in main manuscript. LNA modifications are shown in lower case (“c” = 5-methylcytosine LNA monomer). Modified ONs were obtained in a purity of at least 75%.

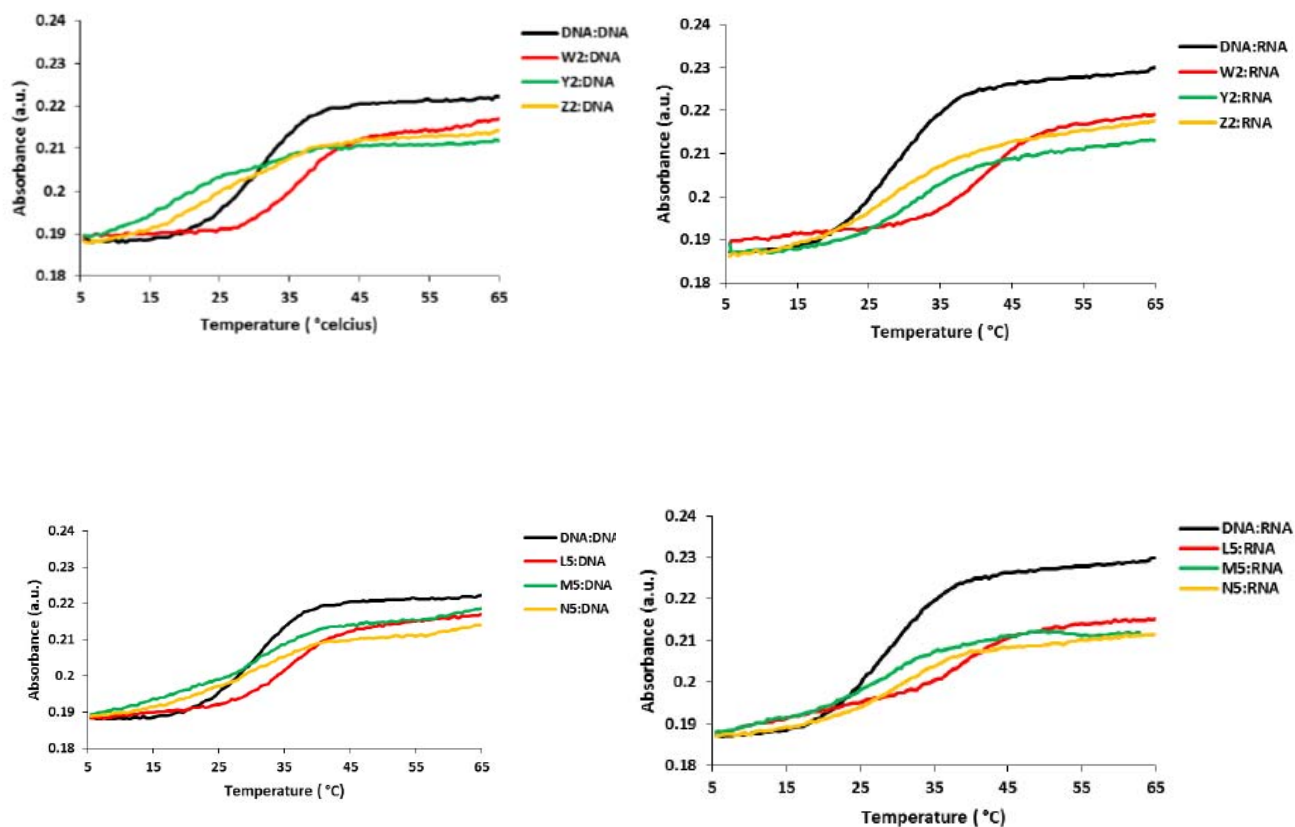


Figure S1. Representative thermal denaturation curves of duplexes between **W2/Y2/Z2/L5/M5/N5** and complementary DNA or RNA targets. For experimental conditions, see Table 1.

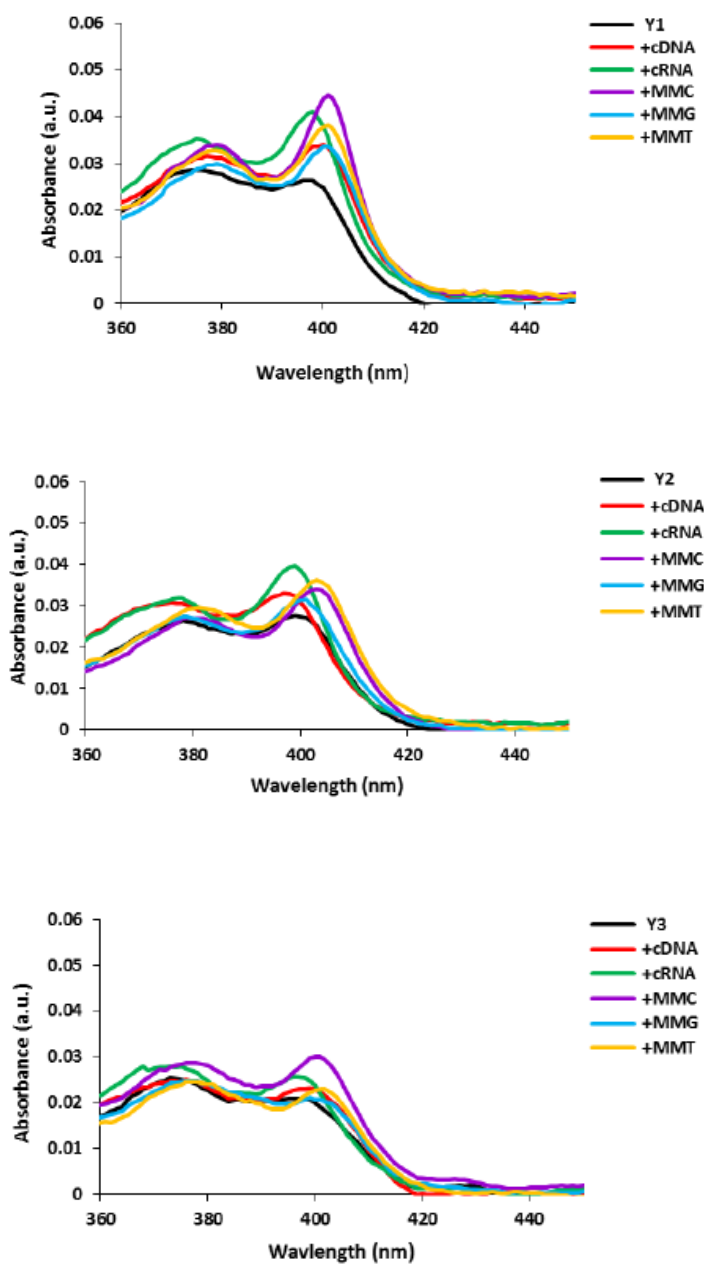


Figure S2. Absorption spectra of single-stranded **Y1-Y3** and the corresponding duplexes with complementary DNA/RNA (cDNA/cRNA) or centrally mismatched DNA targets (mismatched nucleoside is specified) - for sequences of matched/mismatched targets, see footnote of Table 5. Spectra were recorded in T_m buffer at $T = 5^\circ\text{C}$ using each strand at $1\ \mu\text{M}$ concentration.

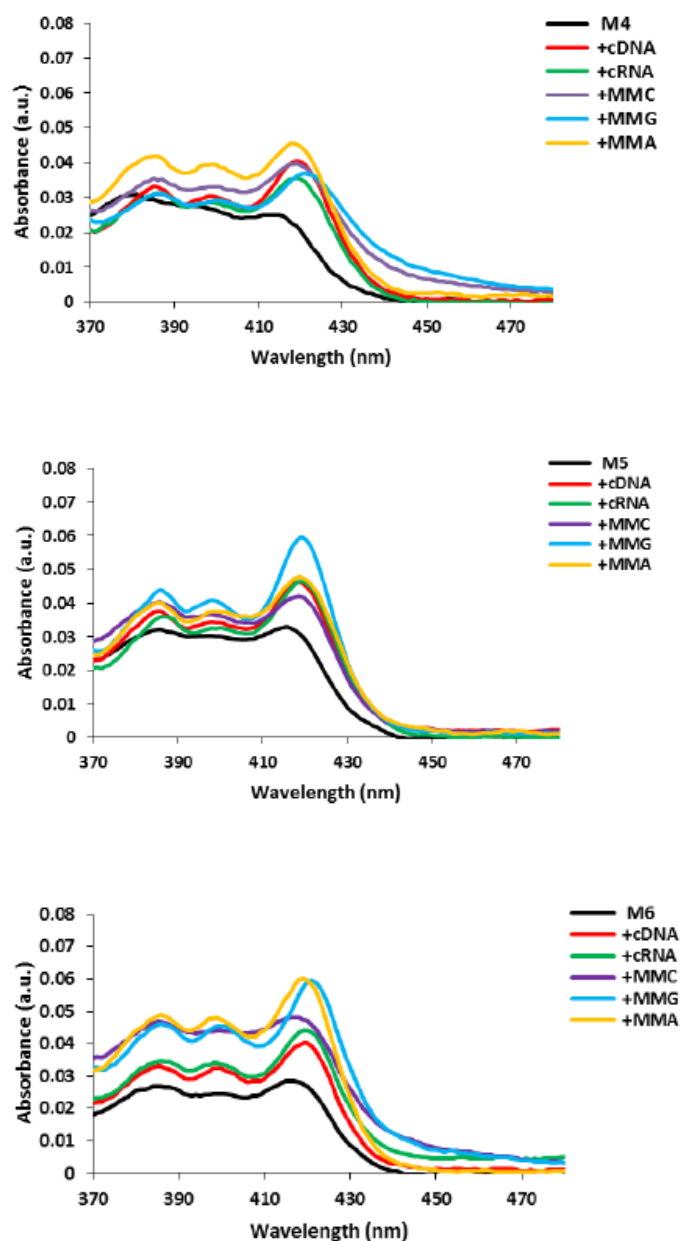


Figure S3. Absorption spectra of single-stranded **M4-M6** and the corresponding duplexes with complementary DNA/RNA or centrally mismatched DNA targets (mismatched nucleoside is specified) - for sequences of matched/mismatched targets, see footnote of Table 5. For experimental conditions, see Figure S2.

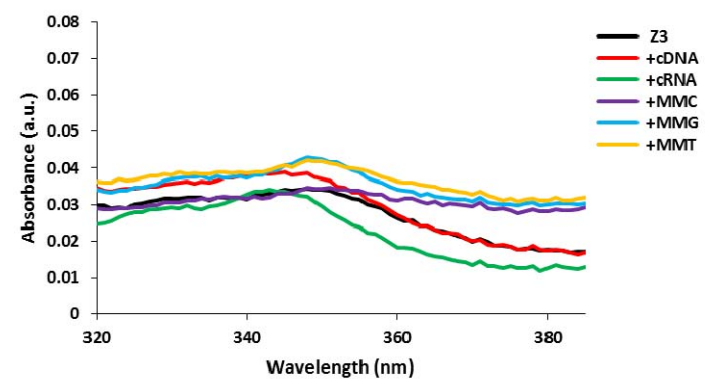
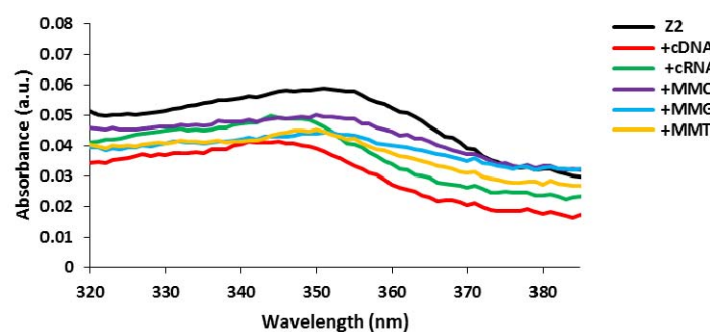
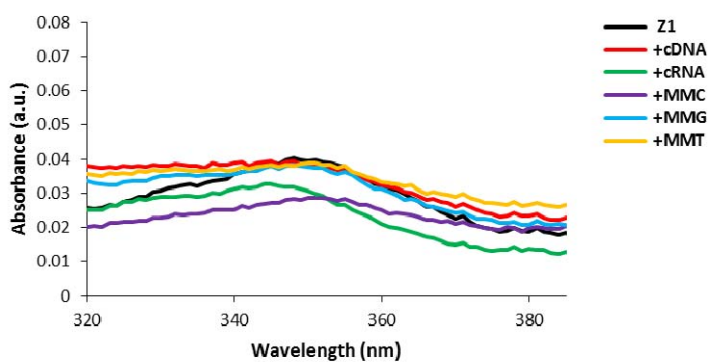


Figure S4. Absorption spectra of single-stranded **Z1-Z3** and the corresponding duplexes with complementary DNA/RNA or centrally mismatched DNA targets (mismatched nucleoside is specified) - for sequences of matched/mismatched targets, see footnote of Table 5. For experimental conditions, see Figure S2.

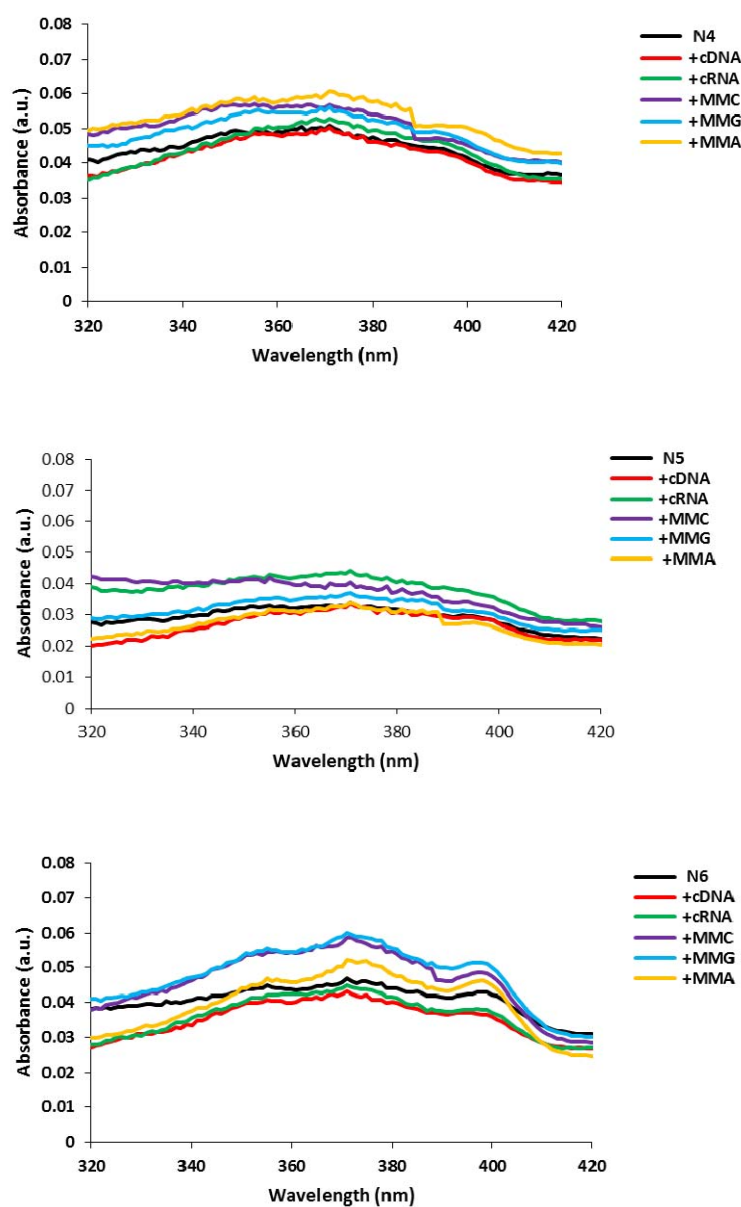


Figure S5. Absorption spectra of single-stranded **N4-N6** and the corresponding duplexes with complementary DNA/RNA or centrally mismatched DNA targets (mismatched nucleoside is specified) - for sequences of matched/mismatched targets, see footnote of Table 5. For experimental conditions, see Figure S2.

Biophysical characterization of 13-mer Y/M-modified ONs. Systematic studies of 13-mer ONs, in which the C5- or C8-ethynylpyrene functionalized monomers **Y** or **M** are directly flanked by LNA nucleotides, were performed (Table S2). The observed trends mirror those for the 9-mer ONs (see main manuscript), with the following exceptions: i) **M11-M14** exhibit lower-than-anticipated cDNA (Table S4), ii) **Y10** displays minimal blue-shifts in pyrene absorption maxima upon cDNA hybridization (Table S5), iii) **M13** displays blue-shifts in pyrene absorption maxima upon hybridization with matched/mismatched DNA targets; this may be due to a structured state of the single-stranded probe (note the high absolute λ_{max} value for **M13**, Table S5), and iv) hybridization of **M11-M14** to cDNA targets does not result in increased emission.

Table S2. MALDI-ToF MS of modified 13-mer ONs.^a

ON	Sequence	(M+H)	
		CALC	OBS
Y7	5'-CG CAA aYa AAC GC	4185.0	4184.0
Y8	5'-CG CAA cYc AAC GC	4232.7	4233.0
Y9	5'-CG CAA gYg AAC GC	4180.7	4182.0
Y10	5'-CG CAA tYt AAC GC	4200.0	4202.0
M11	3'-GCGTT aMa TTGCG	4249.7	4250.6
M12	3'-GCGTT cMc TTGCG	4299.7	4300.0
M13	3'-GCGTT gMg TTGCG	4247.7	4248.0
M14	3'-GCGTT tMt TTGCG	4267.8	4268.5

^a A/C/G/T = adenin-9-yl/cytosin-1-yl/guanin-9-yl/thymin-1-yl DNA monomers. LNA modifications are shown in lower case (**c** = 5-methylcytosine LNA). For structures of monomers **Y** and **M**, see Figure 1 in main manuscript.

Table S3. Thermal denaturation data for duplexes between **Y**-modified ONs and complementary or singly mismatched DNA targets.

ON	Sequence	B =	$T_m/^{\circ}\text{C}$	$\Delta T_m/^{\circ}\text{C}$		
			A	C	G	T
Y7	5'-CG CAA aYa AAC GC 3'-GC GTT TBT TTG CG		45.0	0.0	+1.0	+1.0
Y8	5'-CG CAA cYc AAC GC 3'-GC GTT GBG TTG CG		57.0	-5.0	-2.0	-6.0
Y9	5'-CG CAA gYg AAC GC 3'-GC GTT CBC TTG CG		49.0	0.0	-8.0	-3.0
Y10	5'-CG CAA tYt AAC GC 3'-GC GTT ABA TTG CG		45.5	-1.0	-1.0	-3.0

^aFor experimental conditions, see Table 1 in main manuscript. T_m 's of the duplexes between cDNA and the unmodified control ONs for **Y7-Y10** are 48.5 °C, 55.5 °C, 55.0 °C and 48.5 °C, respectively. ΔT_m = change in T_m 's relative to fully matched duplex (**B** = A).

Table S4. Thermal denaturation data for duplexes between **M**-modified ONs and complementary or singly mismatched DNA targets.

ON	Sequence	B =	$T_m/^{\circ}\text{C}$	$\Delta T_m/^{\circ}\text{C}$		
			T	A	C	G
M11	5'-CGCAA TBT AACGC 3'-GCGTT aMa TTGCG		41.0	-0.5	+2.5	-0.5
M12	5'-CGCAA GBG AACGC 3'-GCGTT cMc TTGCG		54.0	-1.5	-2.5	-1.5
M13	5'-CGCAA CBC AACGC 3'-GCGTT gMg TTGCG		47.0	+1.5	+2.5	-3.5
M14	5'-CGCAA ABA AACGC 3'-GCGTT tMt TTGCG		42.0	-1.5	-0.5	+1.5

^aFor experimental conditions, see Table 1 in main manuscript. T_m 's of the duplexes between cDNA and the unmodified control ONs for **M11-M14** are 48.5 °C, 55.0 °C, 55.5 °C and 48.5 °C, respectively. ΔT_m = change in T_m 's relative to fully matched duplex (**B** = T).

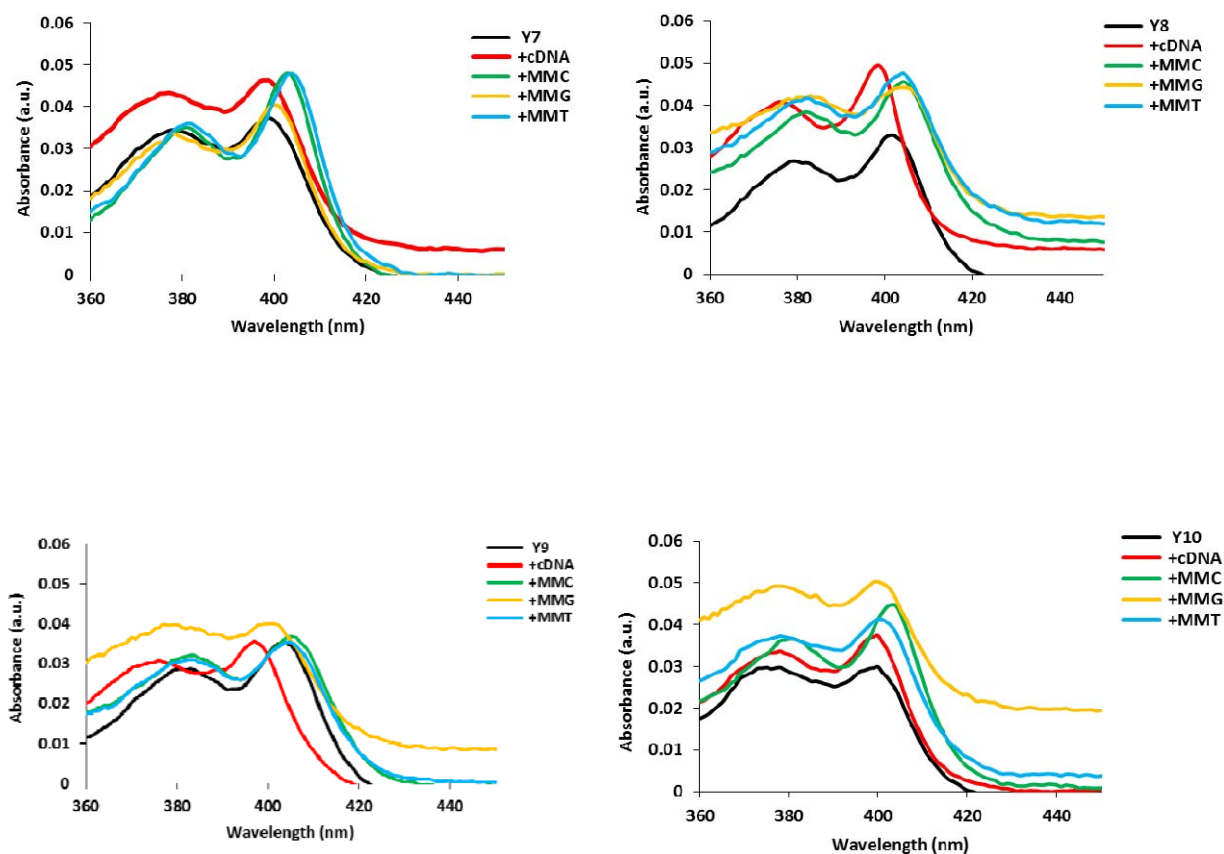


Figure S6. Absorption spectra of 13-mer Y-modified ONs and the corresponding duplexes with complementary or centrally mismatched DNA targets (nucleoside opposite of monomer Y is specified) – for sequences see Table S3. Spectra were recorded in T_m buffer at $T = 5^\circ\text{C}$ using each strand at $1\ \mu\text{M}$ concentration.

Table S5. Absorption maxima of 13-mer **Y**-modified ONs in the presence or absence of complementary or singly mismatched DNA targets (nucleoside opposite of monomer **Y** is specified).

ON	Sequence	$\lambda_{\max}/\text{nm} (\Delta\lambda_{\max})$				
		SSP	B = A	B = C	B = G	B = T
Y7	5'-CG CAA aYa AAC GC 3'-GC GTT TBT TTG CG	399	-1	+4	+1	+5
Y8	5'-CG CAA cYc AAC GC 3'-GC GTT GBG TTG CG	402	-4	+2	+2	+2
Y9	5'-CG CAA gYg AAC GC 3'-GC GTT CBC TTG CG	403	-6	+2	-1	+1
Y10	5'-CG CAA tYt AAC GC 3'-GC GTT ABA TTG CG	400	+0	+4	± 0	± 0

^a Recorded in T_m buffer at $T = 5^\circ\text{C}$ using $1.0\ \mu\text{M}$ of each strand. SSP = single-stranded probe.

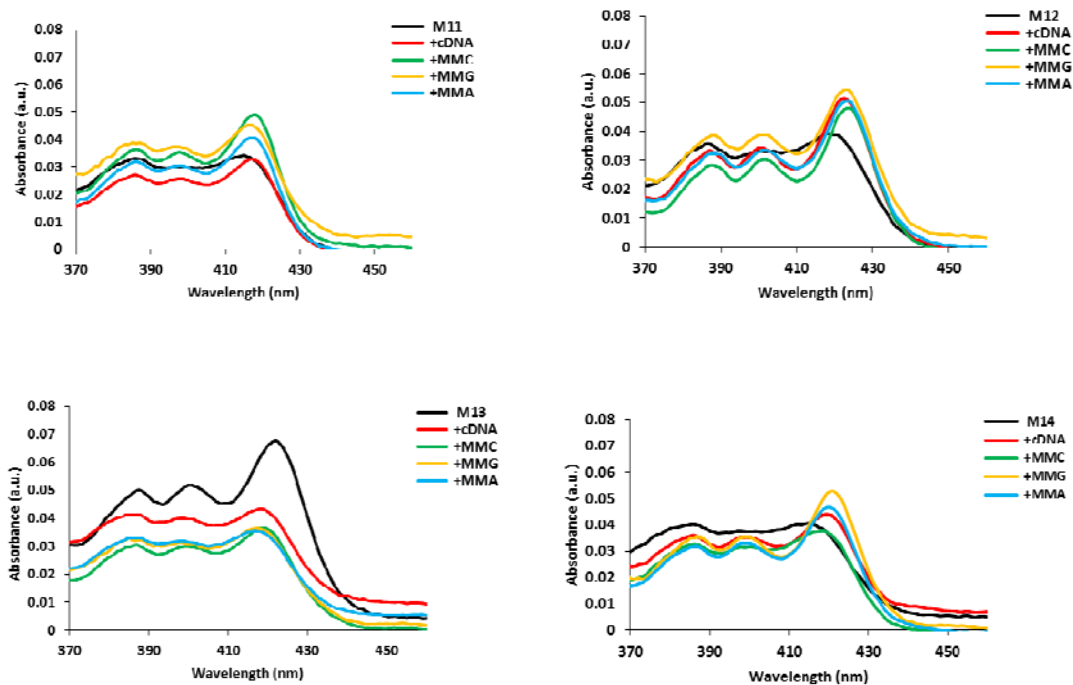


Figure S7. Absorption spectra of 13-mer **M**-modified ONs and the corresponding duplexes with complementary or centrally mismatched DNA targets (nucleoside opposite of monomer **M** is specified) – for sequences see Table S4. Spectra were recorded in T_m buffer at $T = 5\text{ }^{\circ}\text{C}$ using each strand at $1\text{ }\mu\text{M}$ concentration.

Table S6. Absorption maxima of 13-mer **M**-modified ONs in the absence (SSP) or presence of complementary or singly mismatched DNA targets (nucleoside opposite of monomer **M** is specified).^a

ON	Sequence	$\lambda_{\max}/\text{nm} (\Delta\lambda_{\max})$				
		SSP	B = T	B = C	B = G	B = A
M11	3'-GCGTT aMa TTGCG 5'-CGCAA TBT AACGC	415	+2	+3	+2	+2
M12	3'-GCGTT cMc TTGCG 5'-CGCAA GBG AACGC	418	+4	+5	+5	+5
M13	3'-GCGTT gMg TTGCG 5'-CGCAA CBC AACGC	422	-4	-3	-5	-5
M14	3'-GCGTT tMt TTGCG 5'-CGCAA ABA AACGC	415	+4	+3	+3	+5

^[a] Recorded in T_m buffer at $T = 5\text{ }^{\circ}\text{C}$ using $1.0\text{ }\mu\text{M}$ of each strand. SSP = single-stranded probe.

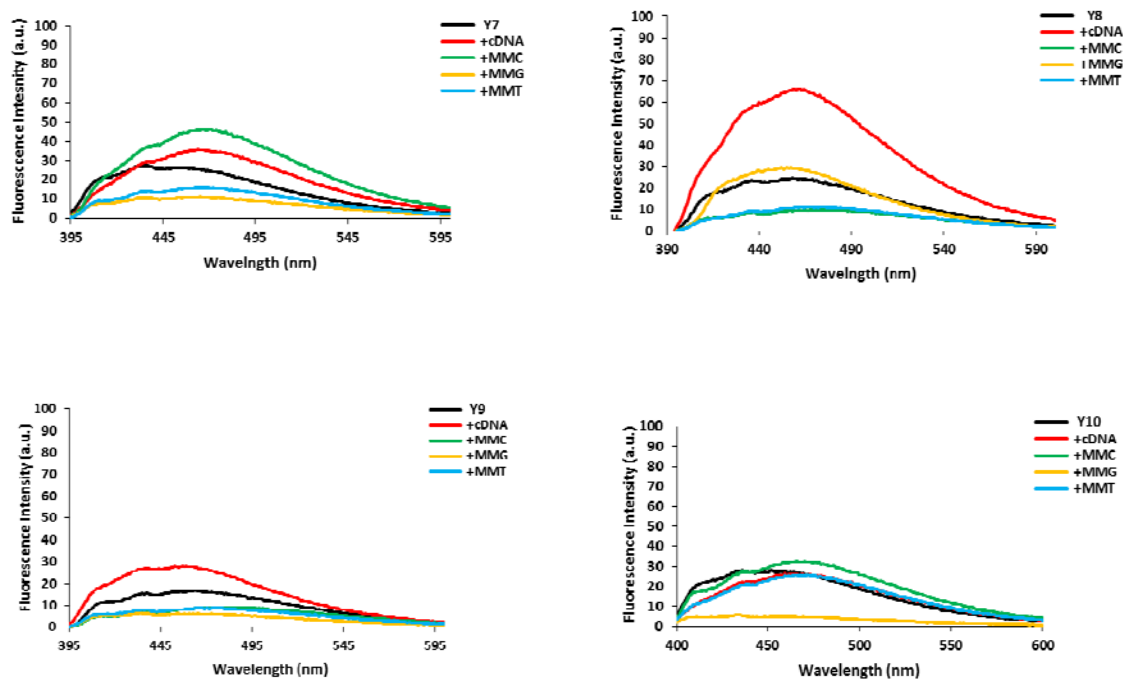


Figure S8. Steady-state fluorescence emission spectra of 13-mer **Y**-modified ONs in the presence or absence of complementary or centrally mismatched DNA targets (mismatched nucleoside opposite of monomer **Y** is specified) – for sequences see Table S3. Spectra were recorded in T_m buffer at $T = 5\text{ }^{\circ}\text{C}$ using each strand at $1\text{ }\mu\text{M}$ concentration and $\lambda_{\text{ex}} = 380\text{ nm}$.

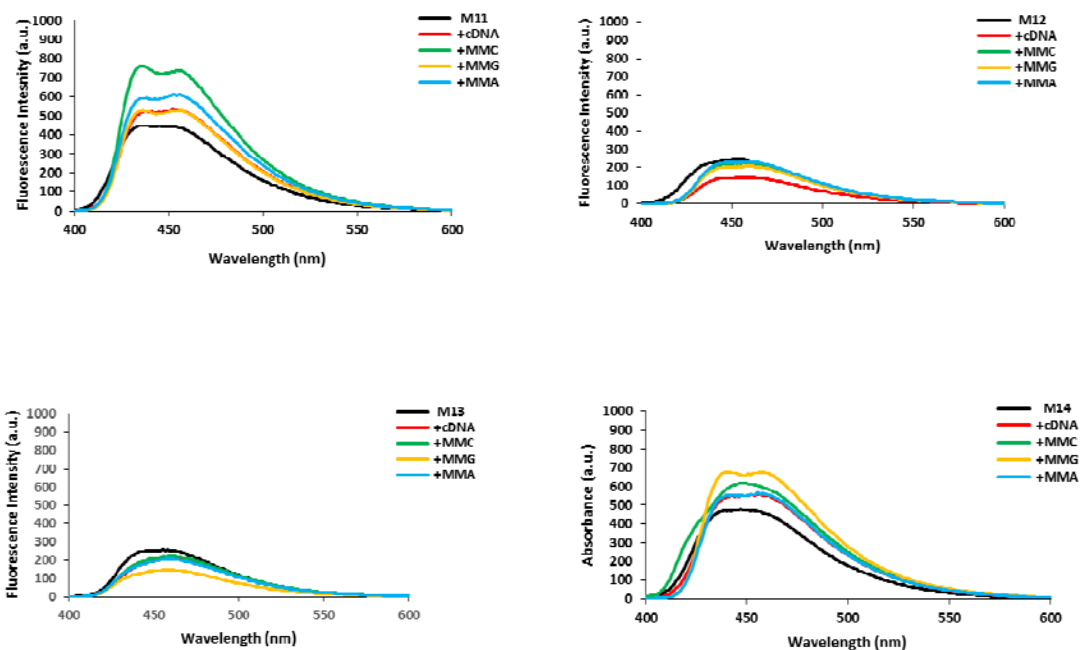
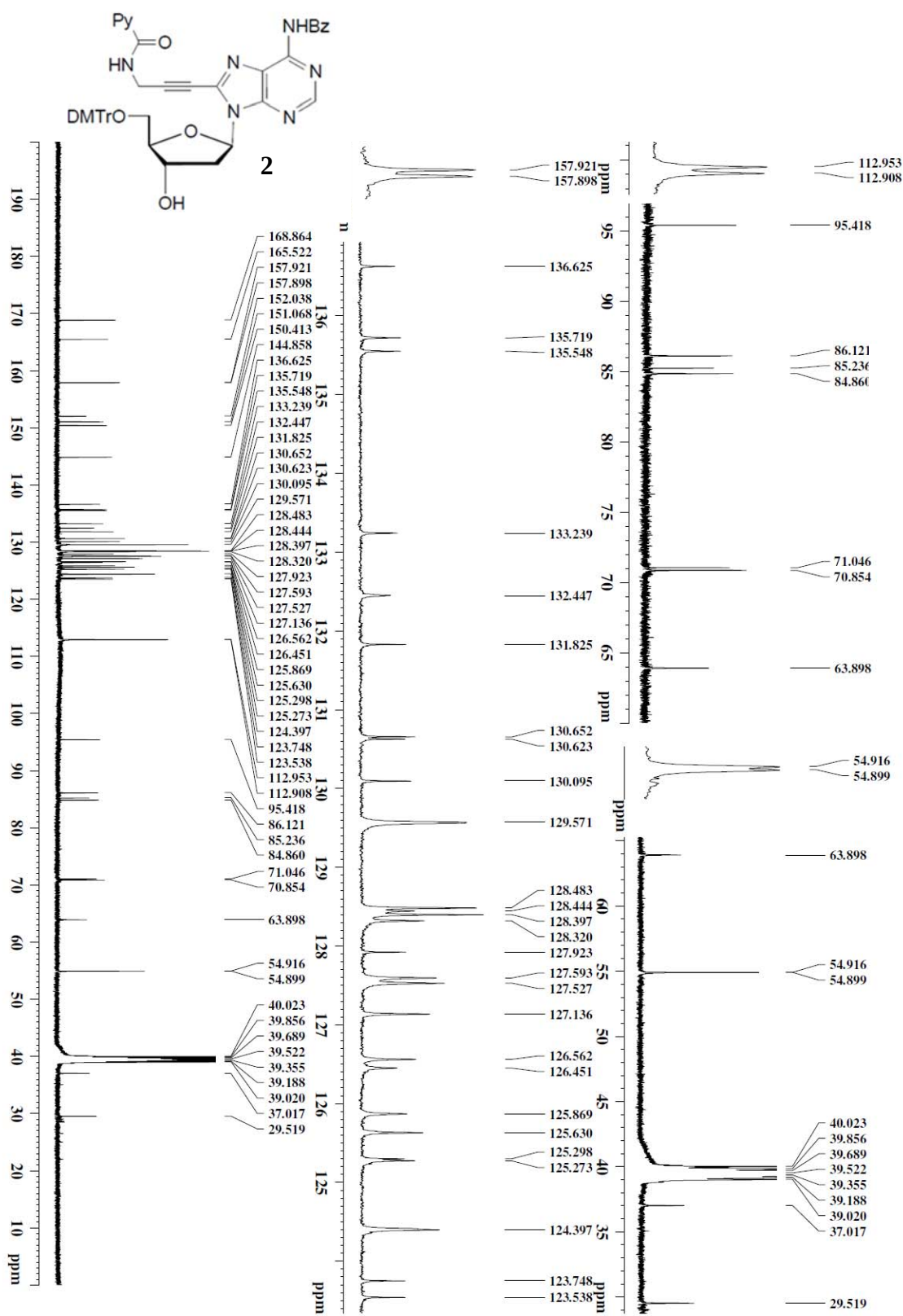
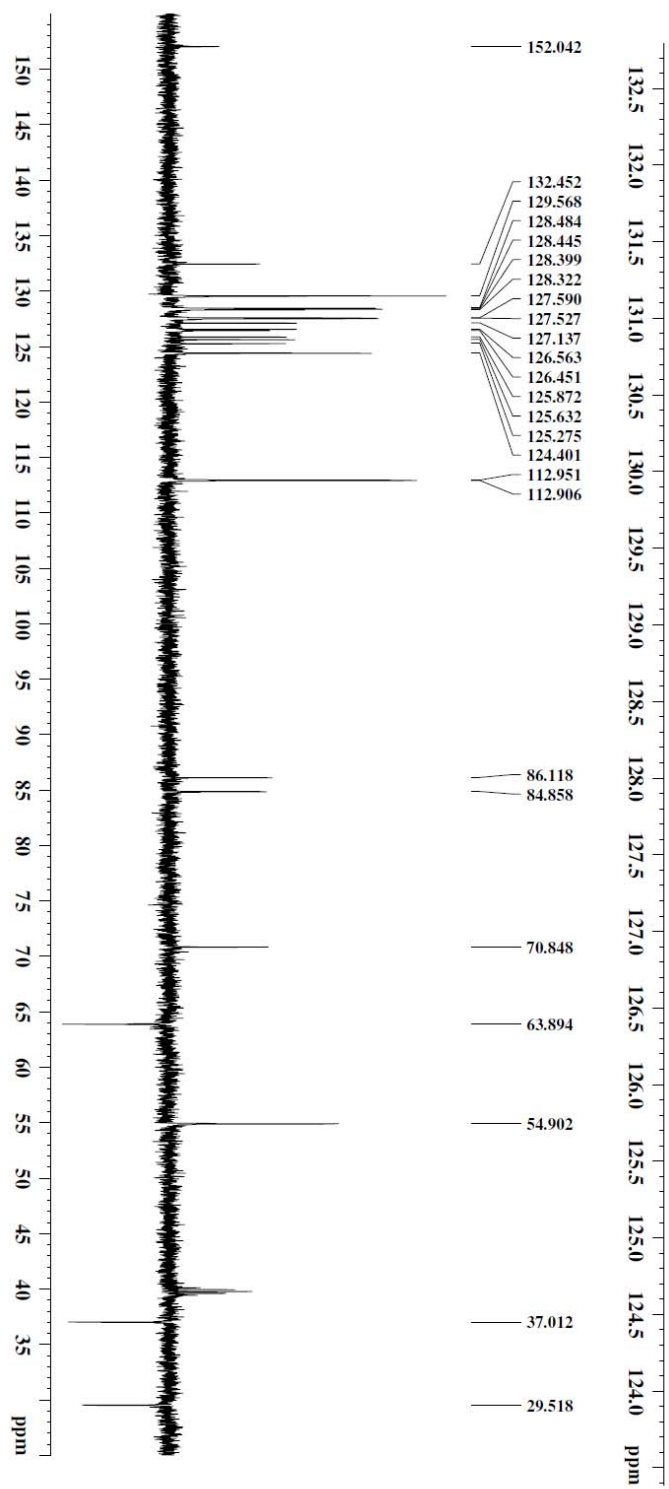
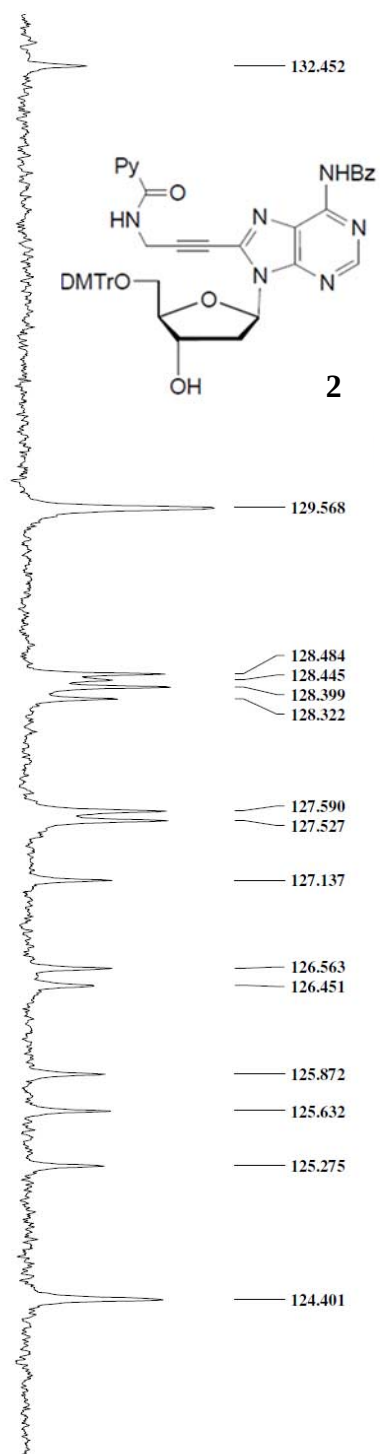
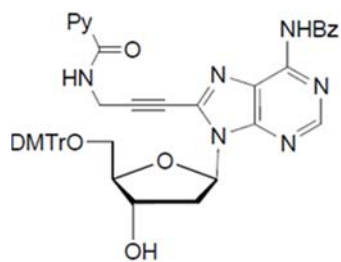


Figure S9. Steady-state fluorescence emission spectra of 13-mer **M**-modified ONs in the presence or absence of complementary or centrally mismatched DNA targets (mismatched nucleoside opposite of monomer **M** is specified) – for sequences see Table S4. Spectra were recorded in T_m buffer at $T = 5\text{ }^{\circ}\text{C}$ using each strand at $1\text{ }\mu\text{M}$ concentration, $\lambda_{\text{ex}} = 385\text{ nm}$.

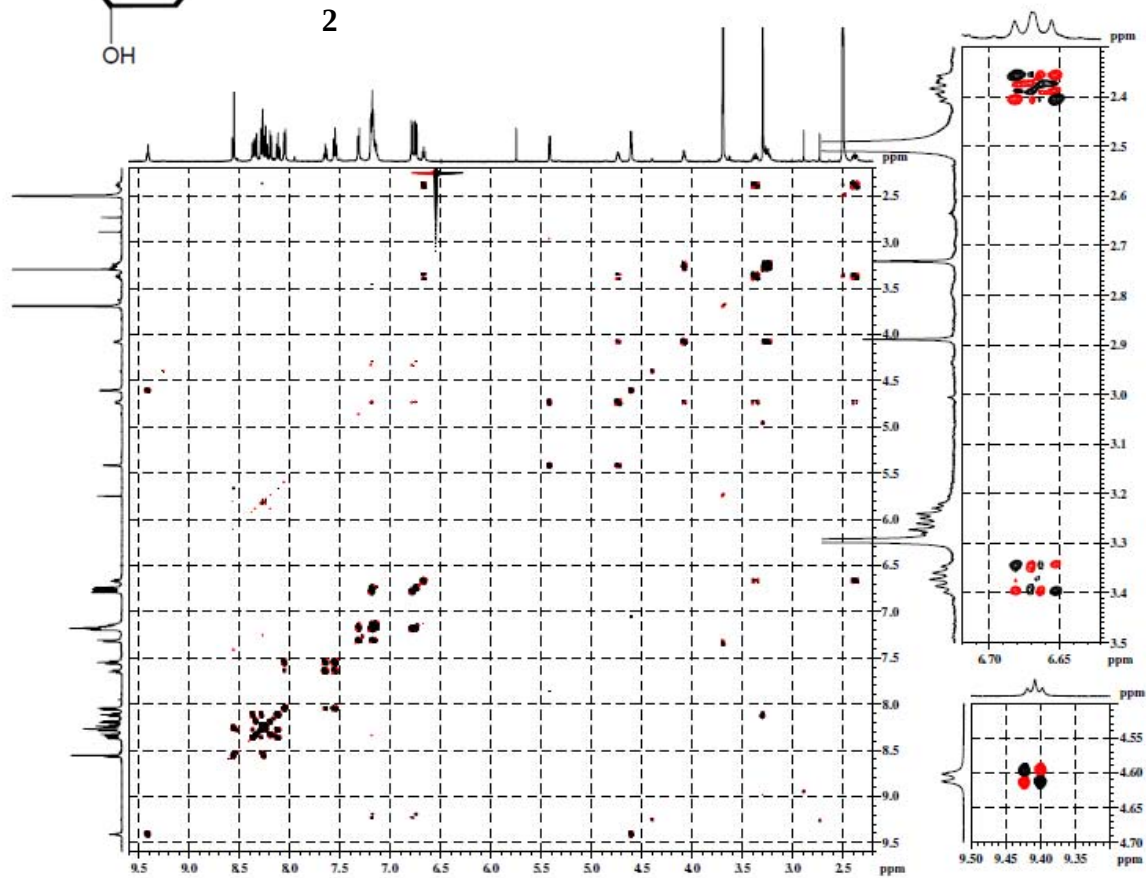


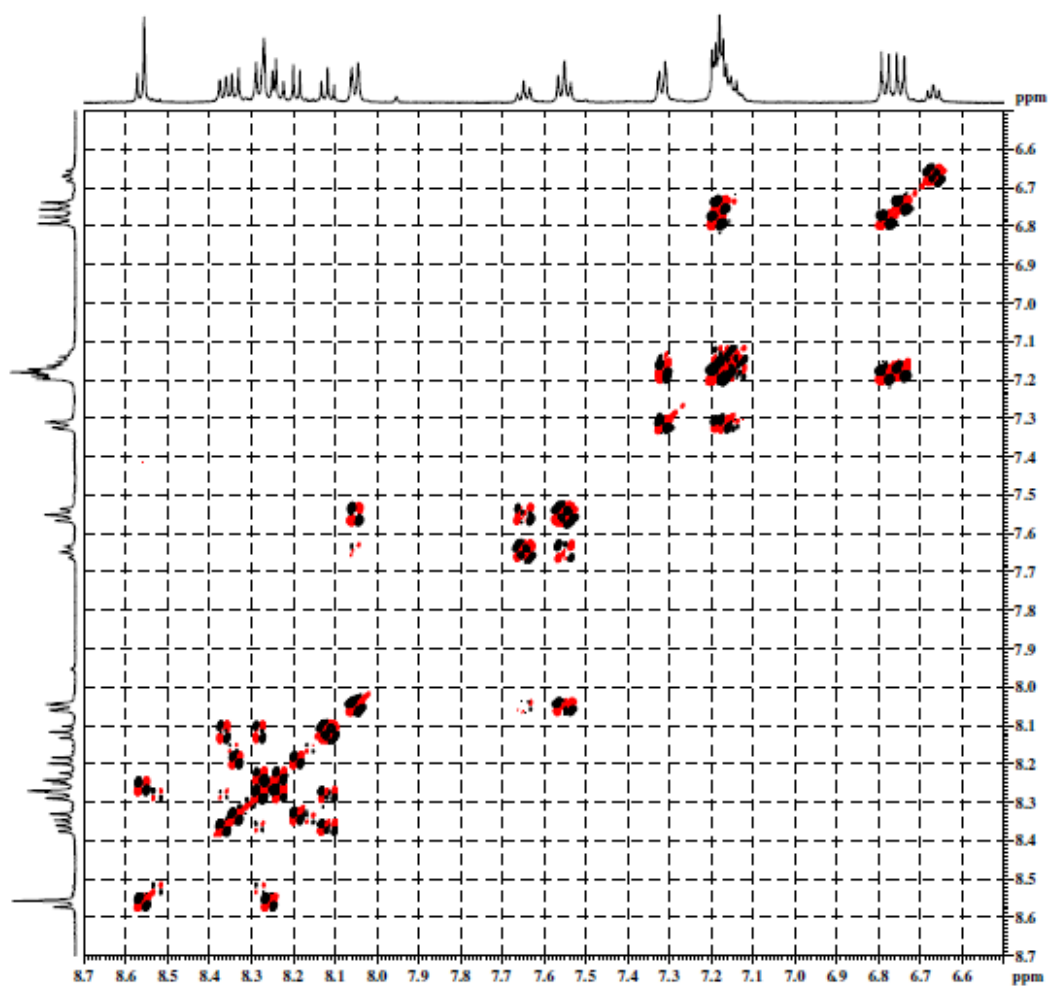
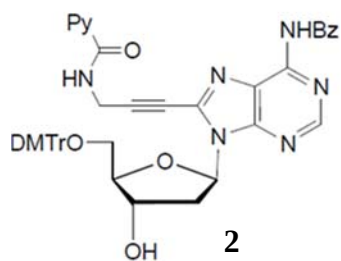


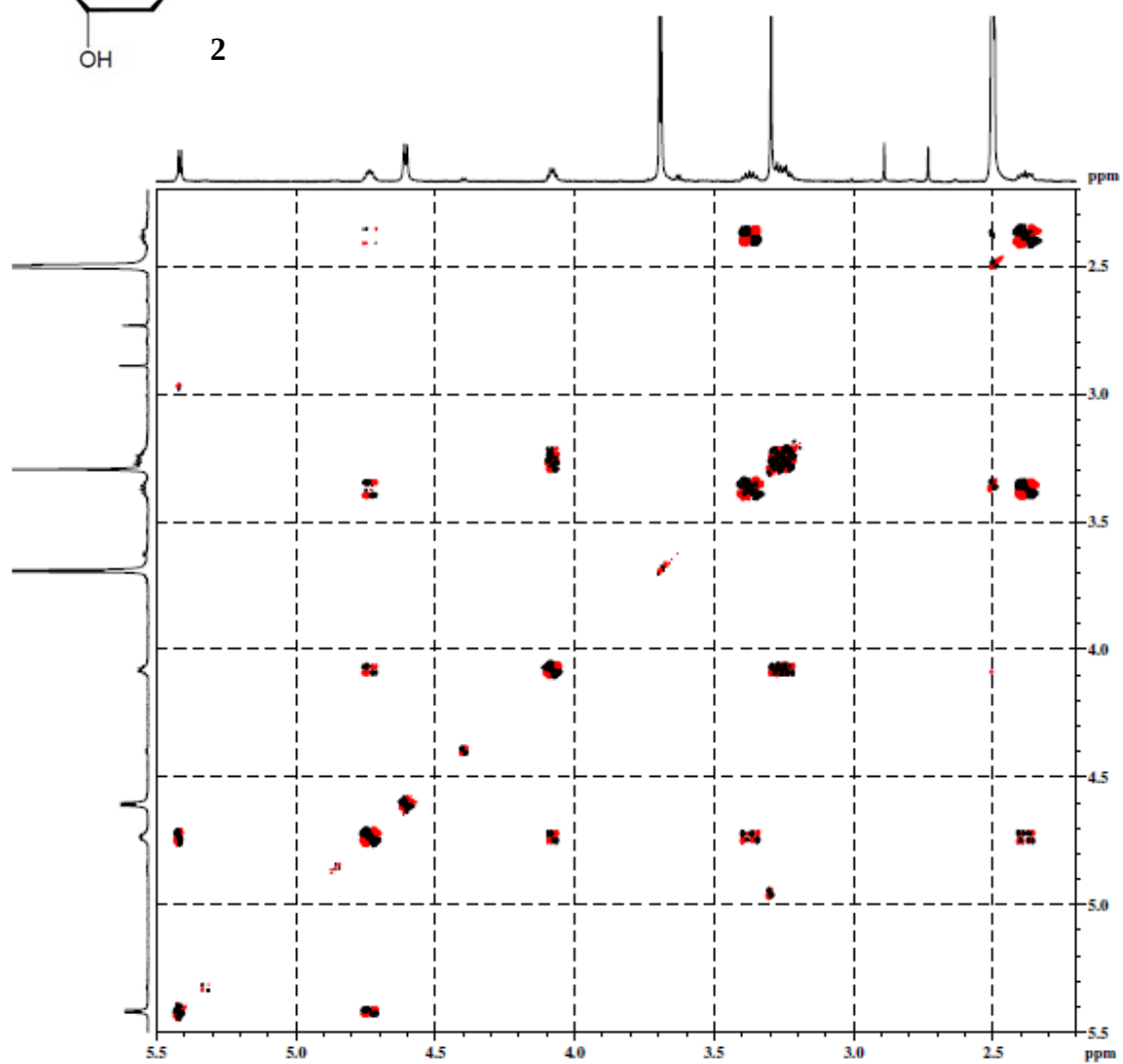
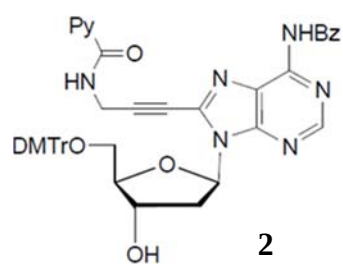


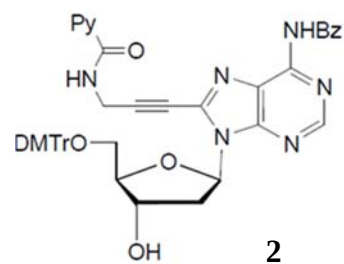


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