Natural Nitric Oxide (NO) inhibitors from the rhizomes of Curcuma phaeocaulis

Supplementary Information

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Figure S2. ¹H NMR spectra of phasalvione (1)





Figure S3. ¹H-¹H COSY spectra of phasalvione (1)







Figure S5. HMBC spectra of phasalvione (1)



Figure S6. NOESY spectra of phasalvione (1)



Figure S7. Rh₂(OCOCF₃)₄-induced CD spectra of phasalvione (1)

Bio-Kine Software V4.62 Date : 2012-12-28 Time : 17:22:52

COMMENTS :

File name : d:US#Nje-16.bka_1 Savitzky-Golay Smooth of sav-golay Window Points=15 Polynomial Order=3 Derivative=0

Figure S8. HRESIMS spectra of phasalvione (1)







Figure S10. ¹H NMR spectra of phaeocaudione (2)





Figure S11. ¹H-¹H COSY spectra of phaeocaudione (2)



Figure S12. HSQC spectra of phaeocaudione (2)



Figure S13. HMBC spectra of phaeocaudione (2)



Figure S14. NOESY spectra of phaeocaudione (2)

0.00 Hz 0



Figure S15. Rh₂(OCOCF₃)₄-induced CD spectra of phaeocaudione (2)

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Figure S16. HRESIMS spectra of phaeocaudione (2)

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Figure S17. ¹³C NMR spectra of phaeocauone (3)





Figure S18. ¹H NMR spectra of phaeocauone (3)







Figure S21. NOESY spectra of phaeocauone (3)

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Figure S22. HRESIMS spectra of phaeocauone (3)

Mass Spectrum Molecular Formula Report







Figure S24. ¹H NMR spectra of 3-methyl-4-(3-oxobutyl)-benzoic acid (4)





Figure S25. HSQC spectra of 3-methyl-4-(3-oxobutyl)-benzoic acid (4)



Figure S26. HMBC spectra of 3-methyl-4-(3-oxobutyl)-benzoic acid (4)

Figure S27. HRESIMS spectra of 3-methyl-4-(3-oxobutyl)-benzoic acid (4)

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Figure S28. ¹³C NMR spectra of 8β (H)-elema-1,3,7(11)-trien-8,12-lactam (5)



Figure S29. ¹H NMR spectra of 8β (H)-elema-1,3,7(11)-trien-8,12-lactam (5)





Figure S30. HSQC spectra of 8β (H)-elema-1,3,7(11)-trien-8,12-lactam (5)



Figure S31. HMBC spectra of 8β (H)-elema-1,3,7(11)-trien-8,12-lactam (5)



Figure S32. NOESY spectra of 8β (H)-elema-1,3,7(11)-trien-8,12-lactam (5)

Figure S33. HRESIMS spectra of 8β (H)-elema-1,3,7(11)-trien-8,12lactam (5) Qualitative Analysis

		Report		
Sample Type Instrument Name Acq Method IRM Calibration Status Comment	Semple Instrument 1 HR(+).m Success	Position User Name Acquired Time DA Method	19 P1-P1 S/17/2014 10:30:24 AM 1.m	
Sample Group Into. User Chromatograms User Spectra				
Fragmentor Voltage 175	Collision Energy 0	Ionization Mode ESI		
x10 4 +ESI Scan (0.164 5 2 4 3 2 1	min) Frag=175.0V 19 32.1704 233.1716	.d		

Formula Calculator Results										
Formula	Best	Hass	Tgt Mass	Diff (ppm)	Ion Species	Score				
C15 H21 N O	TRUE	231.1631	231,1623	-3.45	C15 H22 N O	91.63				

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Figure S35. ¹H NMR spectra of 8β -methoxy-isogerma furenolide (6)

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Figure S36. HSQC spectra of 8β -methoxy-isogerma furenolide (6)



Figure S37. HMBC spectra of 8β -methoxy-isogerma furenolide (6)



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Figure S38. NOESY spectra of 8β -methoxy-isogerma furenolide (6)

Figure S39. HRESIMS spectra of 8β -methoxy-isogerma furenolide (6)

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Figure S40. ¹³C NMR spectra of phaeusmane I (7)













Figure S43. HMBC spectra of phaeusmane I (7)



Figure S44. NOESY spectra of phaeusmane I (7)

Figure S45. HRESIMS spectra of phaeusmane I (7) Qualitative Analysis

Report Sample Type Sample Position P1-52 Instrument Name Instrument 1 User Name Acquired Time 5/17/2014 10:32:23 AM Acg Method HR(+).m **IRM Calibration Status** DA Hethod Lm Success Comment Sample Group Info. **User Chromatograms User Spectra** Fragmentor Voltage **Collision Energy Jonization Mode** 651 125 0 +ESI 8can (0.173 min) Frag=175.0V 20.d x10.8 266.]767 2.5 $\mathbf{2}$ 1.51 267.1789 0.50 205 206 207 Counts vs. Mass-to-Charge (m/z) 262 263 264269 270 268

Formula Calculator Results										
Formula	Best	Mass	Tgt Mass	Diff (ppm)	Ion Species	Score				
C15 H23 N 03	TRUE	265.1684	265.1676	-2.45	C15 H24 N C5	95.22				

---- End Of Report ----





Figure S46. ¹³C NMR spectra of phaeoheptanoxide (8)

Figure S47. ¹H NMR spectra of phaeoheptanoxide (8)









Figure S51. HRESIMS spectra of phaeoheptanoxide (8)

Mass Spectrum Molecular Formula Report

Analysis Info Acquisition Date 12/31/2012 10:01:16 AM Analysis Name D3Data/20121231UE-61.d Method. LIU 250-550POS.m Bruker Customer Operator. Instrument / Ser# micrOTOF-Q 125 Sample Name JE-61 Comment. Acquisition Parameter 0.3 Bar 180 °C Source Type 051 Ion Polarity Positive Set Nebulkter 4500 V Set Dry Heater Fecus Not active. Set Capillary Set End Plate Offset Scan Begin 50 mitz -500 V Set Dry Gas 4.0 Umin Scan End 3000 m/z. Set Collsion Cell RF 500.0 Vpp Set Divert Valve Source Generate Molecular Formula Parameter Formula, min. C19H22O5Na Formula, max. Measured mit: 353,136 Tolerance 5 Charge ppm **Check Valence** Minimum. 0 Ö, no. Maximum Nirogen Rule Electron Configuration 56ft np. Filter H/C Ratio Minimum. Maximum 3 ne. 0 Estimate Carbon yes intens. +MS, 2.7min #198 x10⁴ 6 353,1360 4 2° 0+ 330 335 340 345 350 355 360 365 m/z Sum Formula Sigma Err [som] Mean Err [pprn] Err [mDa] rdb. N Rule mbr. 67 -0.07 C 19H 22 Na 1 O 5 0.123 353, 1359 8.50 -0.200.25 68 even

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Page 1 of 1

Figure S52. Chiral HPLC analytical chromatograms for compounds 5 and 6



(Daicel, Chiralpak AD-RH, 5 µm, 150 × 4.6 mm; MeCN/H2O 45:55; flow rate 1.0 ml/min; 220 nm)

Figure S53. Chiral HPLC analytical chromatogram for compound 7

