

A Lactate-Derived Chiral Aldehyde for Determination of the Enantiopurity of Primary Amines

Rachel M. Lanigan, Samantha M. Gibson, Laure Benhamou, Abil E. Aliev and Tom
D. Sheppard*

*Department of Chemistry, University College London,
Christopher Ingold Laboratories, 20 Gordon St, London, WC1H 0AJ, UK*

tom.sheppard@ucl.ac.uk

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General Experimental Information

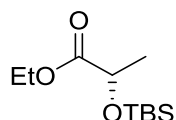
All solvents and chemicals were used as supplied unless otherwise indicated. All resins were washed with CH_2Cl_2 and dried under vacuum prior to use. Column chromatography was carried out using silica gel and analytical thin layer chromatography was carried out using aluminium-backed silica plates. Components were visualized using combinations of UV (254 nm) and potassium permanganate. $[\alpha]_D$ values are given in $10^{-1} \text{ deg cm}^2 \text{ g}^{-1}$, concentration (c) in g per 100 mL. ^1H NMR spectra were recorded at 300, 400, 500 or 600 MHz in the stated solvent using residual protic solvent CDCl_3 ($\delta = 7.26$ ppm, s), DMSO ($\delta = 2.56$ ppm, qn) or MeOD ($\delta = 4.87$, s and 3.31, quintet) as the internal standard. Chemical shifts are quoted in ppm using the following abbreviations: s, singlet; d, doublet; t, triplet; q, quartet; qn, quintet; m, multiplet; br, broad or a combination of these. The coupling constants (J) are measured in Hertz. ^{13}C NMR spectra were recorded at 150 MHz on a Bruker 600 MHz machine with cryoprobe in the stated solvent using the central reference of CDCl_3 ($\delta = 77.0$ ppm, t), DMSO ($\delta = 39.52$ ppm, septet) or MeOD ($\delta = 49.15$ ppm, septet) as the internal standard. Chemical shifts are reported to the nearest 0.1 ppm. Mass Spectrometry data were collected on either TOF or magnetic sector analysers. The ionization method is reported in the experimental data.

The enantiopurity of the commercially available lactates was determined prior to use via formation of the corresponding Mosher's esters as follows: MTPA (40 mg, 0.17 mmol), EDCl.HCl (33 mg, 0.17 mmol) and DMAP (23 mg, 0.19 mmol) were added to a solution of lactate (~ 0.06 mmol) in CH_2Cl_2 (1 mL). The reaction was stirred overnight, and then concentrated *in vacuo*. The crude mixture was dissolved in MeOD and analysed by ^1H NMR. The chemical shifts of the methyl groups at 1.54 and 1.47 ppm were integrated to determine the enantiomeric ratio. Samples of (*R*)-methyl lactate were found to consistently have high enantiopurity ($\geq 99:1$), whereas the enantiopurity of (*S*)-ethyl lactate was more variable. Typically, most commercial sources of (*S*)-ethyl lactate were found to be not quite enantiopure (97:3 er) with the exception of (*S*)-ethyl lactate (99%) purchased from Alfa Aesar (catalogue number A10900) which was determined to be $>99:1$ er. In all cases, the enantiopurity of a batch of aldehyde **4** was found to be identical to the enantiopurity of the lactate ester **6** from which it was prepared.

Samples of the aldehydes **4** were stored in the freezer under argon. Although no drop in enantiopurity was observed over the course of 1 month, small amounts of decomposition were observed. It is recommended that aldehyde that has been stored for longer periods is not used for analysis, without first checking both the quality and enantiopurity of the sample.

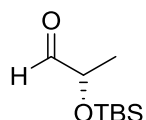
Preparation of Chiral Aldehydes

Ethyl (S)-2-((tert-butyldimethylsilyl)oxy)propanoate (S)-6a



tert-Butylchlorodimethylsilane (16.0 g, 0.11 mol, 1.2 equiv.), was added to a stirring solution of *S*-ethyl lactate (10 mL, 0.09 mol, 1 equiv.) and imidazole (9.0 g, 0.13 mol, 1.4 equiv.) in DMF (88 mL). The solution was stirred for 30 min at rt, water (150 mL) was added and the reaction mixture extracted with Et₂O (2 × 150 mL). The combined organic layers were washed with brine (150 mL), dried over MgSO₄ and concentrated under reduced pressure to give **80** as a colourless oil (20.49 g, 0.088 mol, 98%); [α]_D²³ -21.9 (*c* = 1.33, CHCl₃) [lit -28.9 (*c* = 1.26, CHCl₃, 23 °C)]¹; δ_{H} (400 MHz, CDCl₃) 0.07 (s, 3H, SiCH₃), 0.11 (s, 3H, SiCH₃), 0.90 (s, 9H, SiC(CH₃)₃), 1.29 (t, *J* 7.1, CH₂CH₃), 1.40 (d, *J* 6.8, 3H, CH₃CH), 4.17-4.21 (2H, m, CH₂CH₃), 4.31 (q, *J* 6.8, 1H, CHCH₃); δ_{C} (100 MHz, CDCl₃) -5.3, -5.0, 14.2, 18.3, 21.3, 25.7, 60.7, 68.5, 174.1.

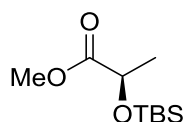
(S)-2-((tert-Butyldimethylsilyl)oxy)propanal (S)-4



Diisobutylaluminium hydride (44 mL, 1.1 M in cyclohexane, 48.4 mmol, 1.6 equiv.) was added dropwise (20 mL/h) to a stirring solution of ester **80** (7 g, 30.1 mmol, 1 equiv.) in Et₂O (200 mL) at -100 °C. After completion of addition, the reaction was stirred for 10 min at -78 °C then quenched by dropwise addition of MeOH (2 mL) and H₂O (5 mL). After warming to rt and stirring for 1.5 h Na₂SO₄ and MgSO₄ were added and the suspension stirred for 15 min, then filtered through a short plug of celite and silica eluting with Et₂O. The filtrate was concentrated under reduced pressure and the residue distilled to give aldehyde (S)-**81** as a colourless liquid (4.56 g, 24.2 mmol, 80%); bp 79-81 °C (19 Torr) [Lit. 67-68 °C (14 Torr)]²;

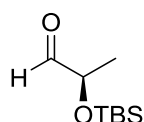
$[\alpha]_D^{23} -12.3$ ($c = 1.24$, CHCl_3) [Lit -11.1 , ($c = 2.7$, CHCl_3 , 22°C)]²; δ_{H} (600 MHz, CDCl_3) 0.08 (s, 3H, SiCH_3), 0.09 (s, 3H, SiCH_3), 0.91 (s, 9H, $\text{SiC}(\text{CH}_3)_3$), 1.27 (d, 3H, J 6.9, CHCH_3), 4.08 (qd, 1H, J 1.2, 6.9, CHCH_3), 9.60 (d, 1H, J 1.2, CHO); δ_{C} (150 MHz, CDCl_3) -4.7, -4.6, 18.3, 18.6, 25.8, 73.9, 204.4.

Methyl (*R*)-2-((*tert*-butyldimethylsilyl)oxy)propanoate (*R*)-6b



tert-Butylchlorodimethylsilane (18.0 g, 0.12 mol, 1.2 equiv.), was added to a stirring solution of *R*-methyl lactate (10 mL, 0.10 mol, 1 equiv.) and imidazole (10.0 g, 0.15 mol, 1.5 equiv.) in DMF (90 mL). The solution was stirred for 30 min at rt, water (150 mL) was added and the reaction mixture extracted with Et_2O (2×150 mL). The combined organic layers were washed with brine (150 mL), dried over MgSO_4 and concentrated under reduced pressure to give the ester as a colourless oil (17.85 g, 81.7 mmol, 82%); $[\alpha]_D^{24} +26.5$ ($c = 2.44$, CHCl_3) [lit $+28.8$ ($c = 2.35$, CHCl_3 , 24°C)]³; δ_{H} (600 MHz, CDCl_3) 0.05 (s, 3H, SiCH_3), 0.08 (s, 3H, SiCH_3), 0.88 (s, 9H, $\text{SiC}(\text{CH}_3)_3$), 1.38 (d, J 6.8, 3H, CH_3CH), 3.70 (s, 3H, CH_3O), 4.31 (q, J 6.8, 1H, CHCH_3); δ_{C} (150 MHz, CDCl_3) -5.2, -4.9, 18.4, 21.5, 25.8, 52.0, 68.5, 174.1.

(*R*)-2-((*tert*-Butyldimethylsilyl)oxy)propanal (*R*)-4



Diisobutylaluminium hydride (47 mL, 1.1 M in cyclohexane, 51.7 mmol, 1.6 equiv.) was added dropwise (20 mL/h) to a stirring solution of methyl (*R*)-2-((*tert*-butyldimethylsilyl)oxy)propanoate (7 g, 32.1 mmol, 1 equiv.) in Et_2O (200 mL) at -100°C . After completion of addition, the reaction was stirred for 10 min at -78°C then quenched by dropwise addition of MeOH (2 mL) and H_2O (5 mL). After warming to rt and stirring for 1.5 h Na_2SO_4 and MgSO_4 were added and the suspension stirred for 15 min, then filtered through a short plug of celite and silica eluting with Et_2O . The filtrate was concentrated under reduced pressure and the residue distilled to give aldehyde (*R*)-**81** as a colourless liquid (4.35 g, 23.1 mmol, 72%); bp $83-86^\circ\text{C}$ (28 Torr); $[\alpha]_D^{24} +15.3$ ($c = 1.59$, CHCl_3) [Lit. $+12.8$ ($c = 1.59$, CHCl_3 , 24°C)]³; δ_{H} (600 MHz, CDCl_3) 0.08 (s, 3H, SiCH_3), 0.09 (s, 3H, SiCH_3), 0.91 (s, 9H,

SiC(CH₃)₃), 1.27 (d, 3H, *J* 6.9, CHCH₃), 4.08 (qd, 1H, *J* 1.0, 6.8, CHCH₃), 9.60 (d, 1H, *J* 1.0, CHO); δ_C (150 MHz, CDCl₃) -4.72, -4.66, 18.3, 18.6, 25.8, 73.9, 204.4.

Determination of the Enantiopurity of Aldehyde 4

A solution of phenylalanine methyl ester (54 mg, 0.30 mmol) in C₆D₆ (0.7 mL) was added to the aldehyde (45 mg, 0.24 mmol). The solution was mixed and transferred to an NMR tube and the enantiopurity was determined via ¹³C NMR as below.

A sample of (*R*)-**4** was analysed via this method after 0, 14, and 31 days (see pages 9-10 for NMR spectra). No drop in enantiopurity (99:1) was observed over this time period, but some impurities were observed in the ¹H NMR of **4** after storing for 14 and 30 days (see page 11 for NMR spectra).

General Procedure for Analysis of Chiral Primary Amines

A chiral amine of unknown enantiopurity was weighed into two separate vials (~0.2 mmol in each, 1 equiv.). A solution of (*S*)-**4** (~0.24 mmol, 1.2 equiv.) in MeOD-*d*₄ or another appropriate NMR solvent (~0.7 mL) was added to one sample, and a similar solution of (*R*)-**4** was added to the other sample. The resulting solutions of imine were then transferred to NMR tubes and analysed via a 256 scan ¹³C NMR (600 MHz NMR with cryoprobe; experiment time ~15 minutes). The enantiopurity was determined by integration of the imine carbon signals (ca 160-170 ppm) in each sample. The average enantiomeric ratio from the two samples was taken as the enantiopurity of the sample.

tert-Butyl (1-oxo-1-(propylamino)propan-2-yl)carbamate (7)

Propylamine (82 μ L, 1.0 mmol) and B(OCH₂CF₃)₃ (616 mg, 2.0 mmol) were added to a solution of ($\pm$)-Boc-alanine or Boc-L-alanine (188 mg, 1 mmol) in acetonitrile (2 mL). The reaction was stirred at 80 °C overnight. Dichloromethane (3 mL) and H₂O (1 mL) were added to the reaction mixture. Amberlyst A-26(OH), Amberlyst 15 and Amberlite IRA743 were added and the mixture stirred for 30 min. MgSO₄ was added and the mixture was filtered. The solution was concentrated *in vacuo* to give the Boc protected amide as a white solid (228 mg, 0.99 mmol, 99%); ¹H NMR (600 MHz, CDCl₃) δ 6.74 (1H, br s, NH), 5.43 (1H, br d, *J* = 6.9, NH), 4.16 (1H, br s, CHCH₃), 3.14 (2H, q, *J* = 6.9, NHCH₂), 1.46 (2H, sext, *J* = 7.4, CH₂CH₃), 1.39 (9H, s, tBu), 1.29 (3H, d, *J* = 6.9, CHCH₃), 0.86 (3H, t, *J* = 7.4, CH₂CH₃); ¹³C

NMR (150 MHz, CDCl₃) δ 173.0, 155.7, 79.9, 50.1, 41.2, 28.4, 22.8, 18.8, 11.4; LRMS (ES+) 231 [M+H]⁺; mp 82-85 °C (CH₂Cl₂).

The Boc protected amide was deprotected as described below, and then the diastereomeric imines were prepared and analysed via the procedure described above in a range of NMR solvents.

Preparation and analysis of unknown samples (Table 3)

Sample preparation

Samples of ‘unknown’ enantiopurity were prepared by weighing appropriate amounts of the individual enantiomeric amines into an appropriately size volumetric flask, and preparing a solution in MeOH of approximately 0.4 M concentration.

Each researcher prepared three different samples, and each sample was then analysed independently by two other researchers as described below.

Analysis of ‘Unknown’ Samples

Two separate 0.5 mL portions of the unknown solution were concentrated into pre-weighed vials to leave ~0.2 mmol of amine in each. A solution of (*S*)-**4** (1.2 equiv) in MeOD-*d*₄ (~0.7 mL) was added to one vial and a solution of (*R*)-**4** (1.2 equiv) in MeOD-*d*₄ (~0.7 mL) was added to the other. Both samples were analysed by ¹³C NMR as described above.

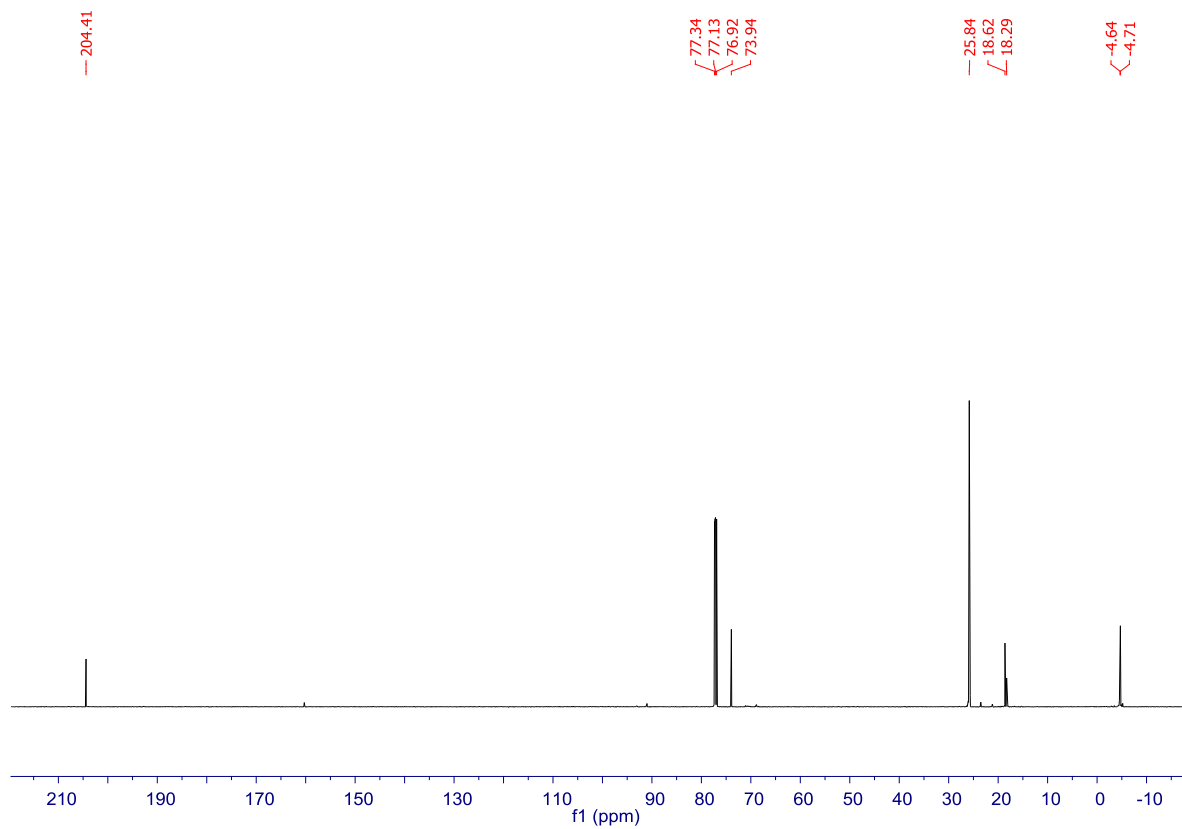
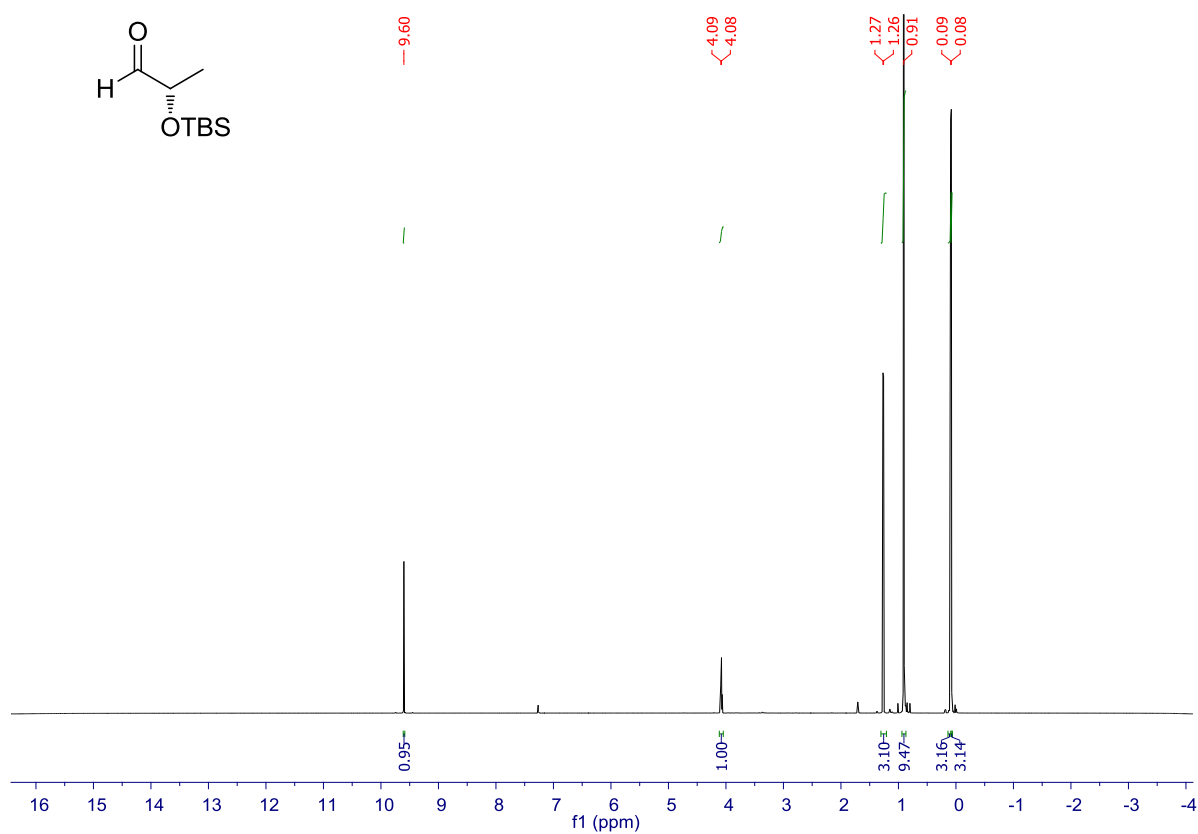
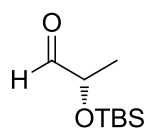
Preparation of Amides 25

Boc-protected amides **25a-25c** were prepared as described previously.⁴

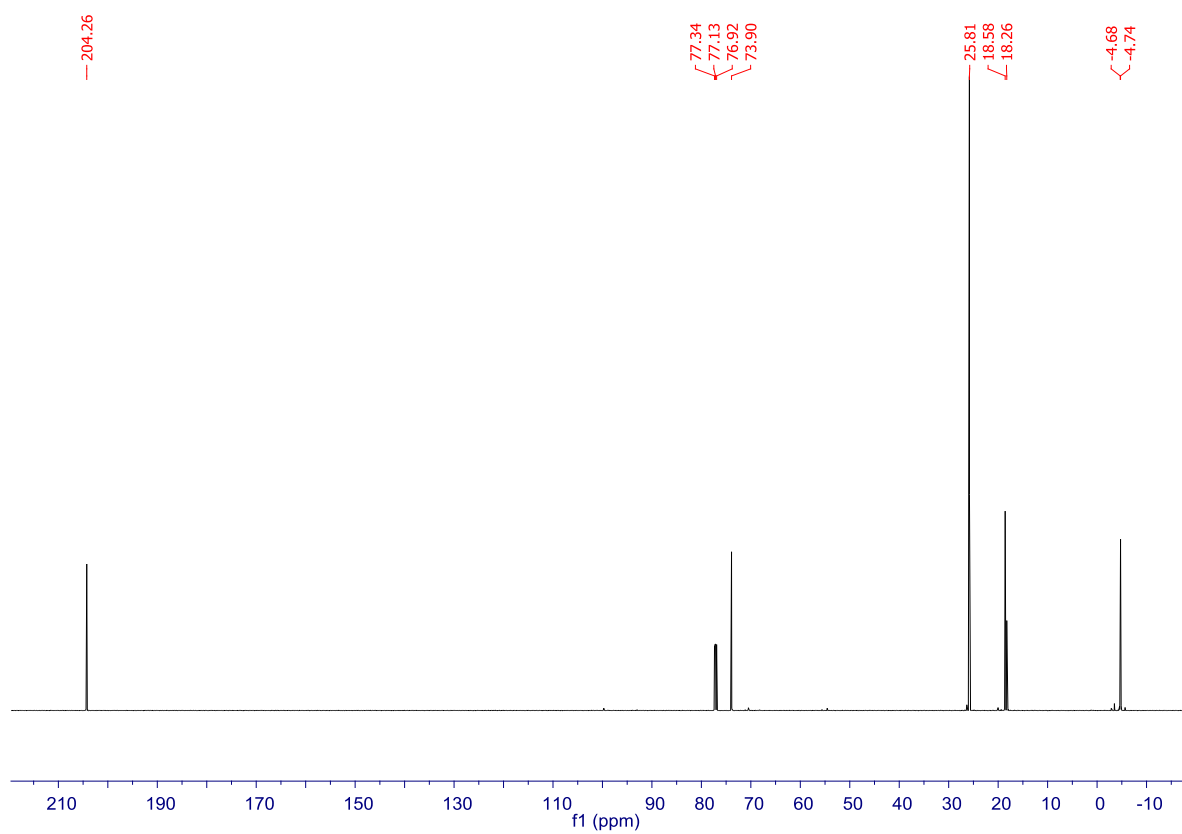
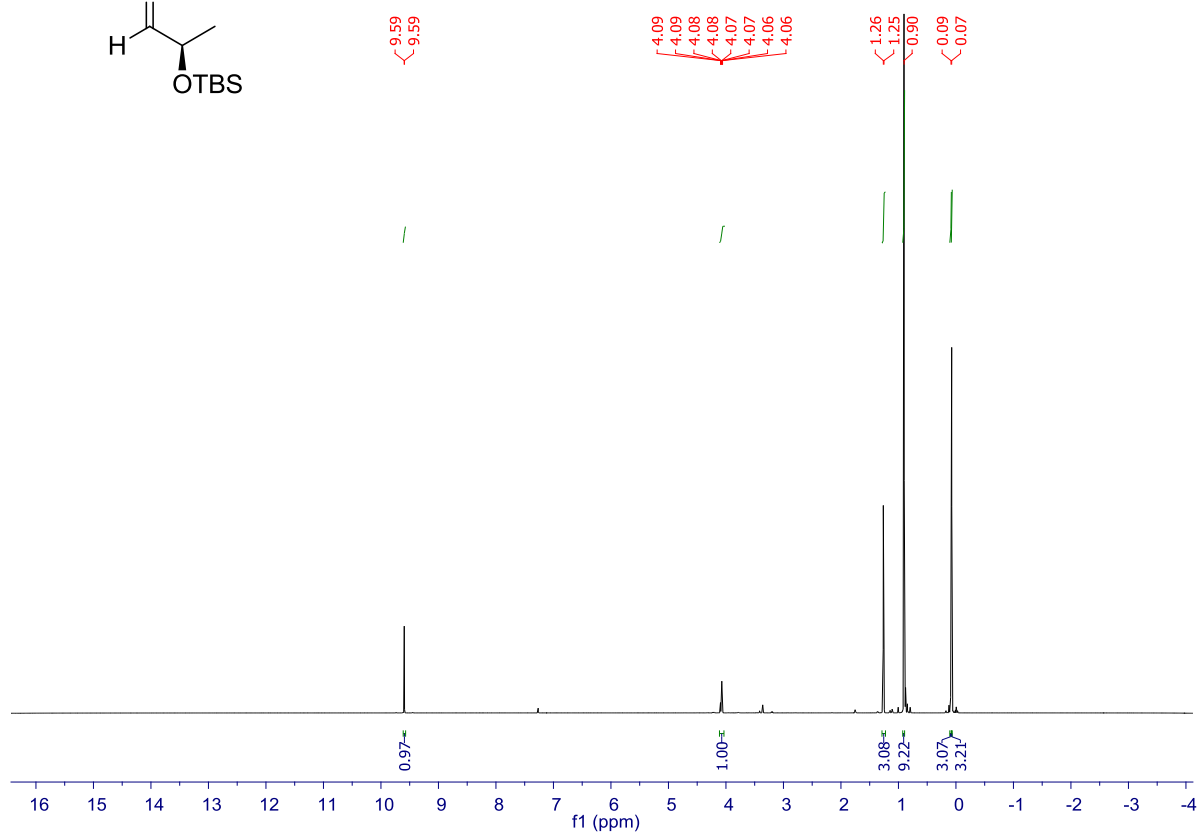
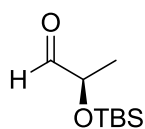
General Procedure for Boc Deprotection and Analysis

The Boc-protected amino acid amide (~0.25 mmol) was diluted in a 1:1 solution of TFA and CH₂Cl₂ (5 mL, 0.05 M) and stirred at rt for 1 h. The reaction mixture was concentrated in vacuo to give the TFA salt. The salt was redissolved in CH₂Cl₂ and stirred with Amberlite A26(OH) (~50 mg) at rt for 30 min. The resin was then removed by filtration and rinsed three times with CH₂Cl₂ (3 × 10 mL). The filtrate was concentrated under reduced pressure to give the free amino acid amide and the enantiopurity was determined as described above.

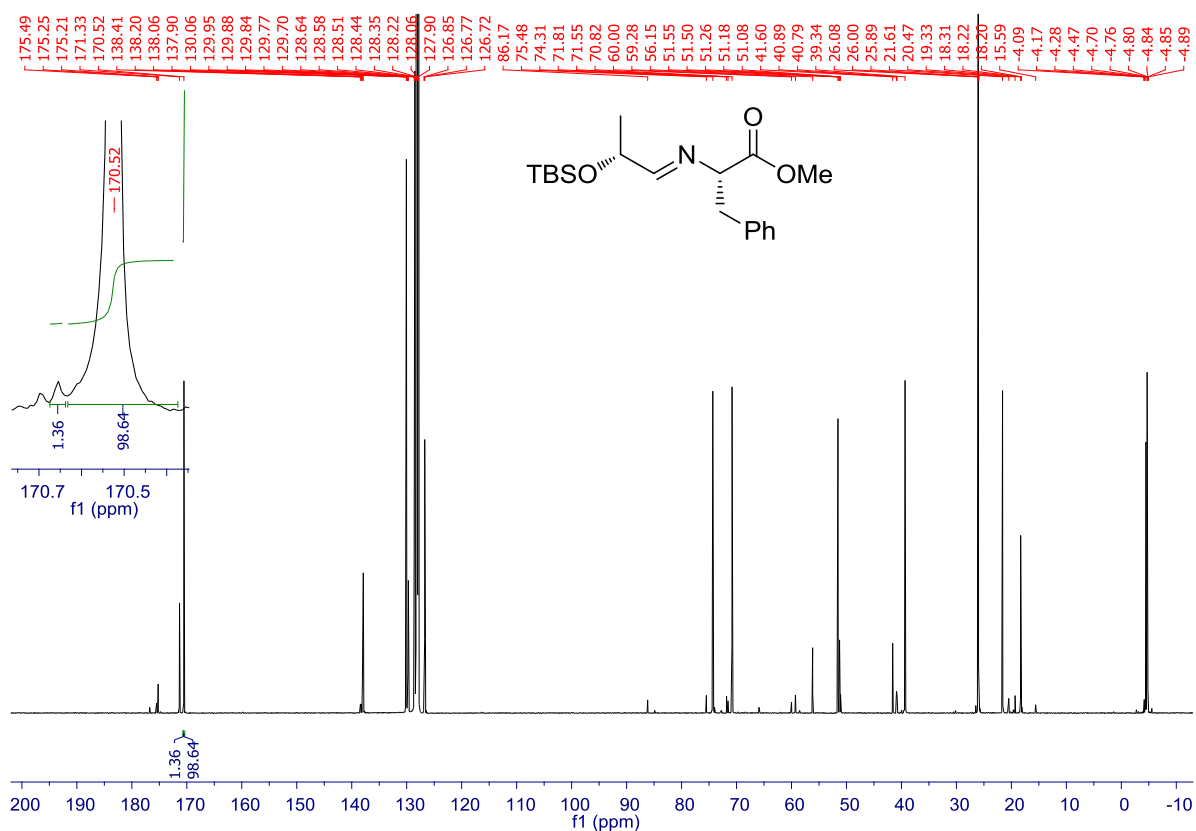
(S)-2-((*tert*-Butyldimethylsilyl)oxy)propanal (S)-4



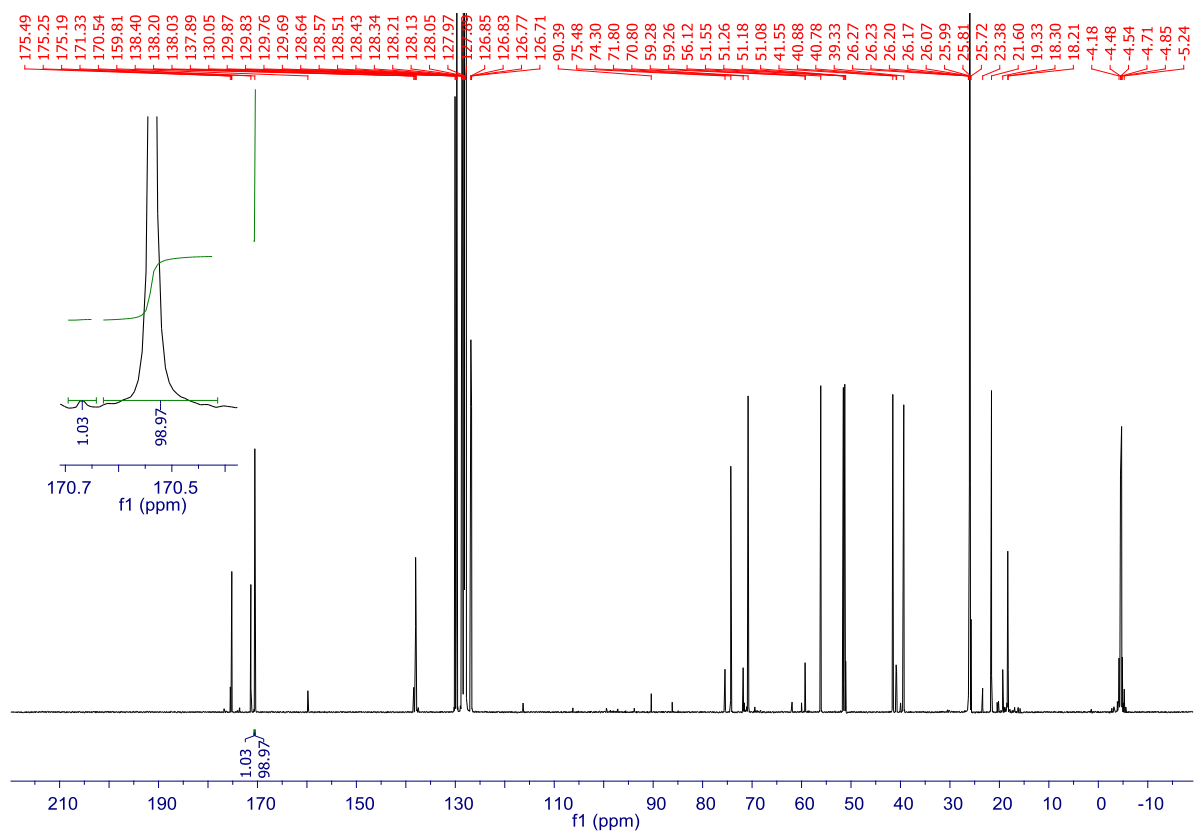
(R)-2-((*tert*-Butyldimethylsilyl)oxy)propanal (R)-4



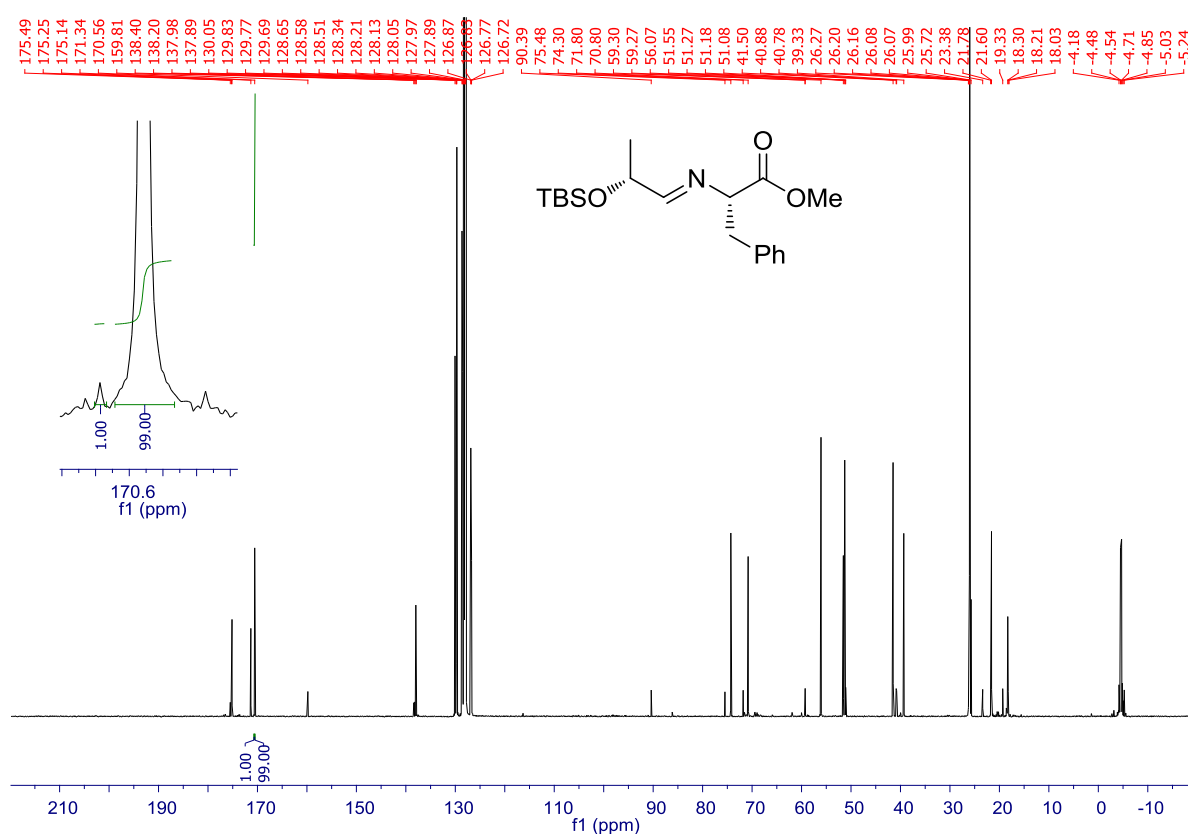
(R)-4 and H-Phe-OMe, C₆D₆ 0 days



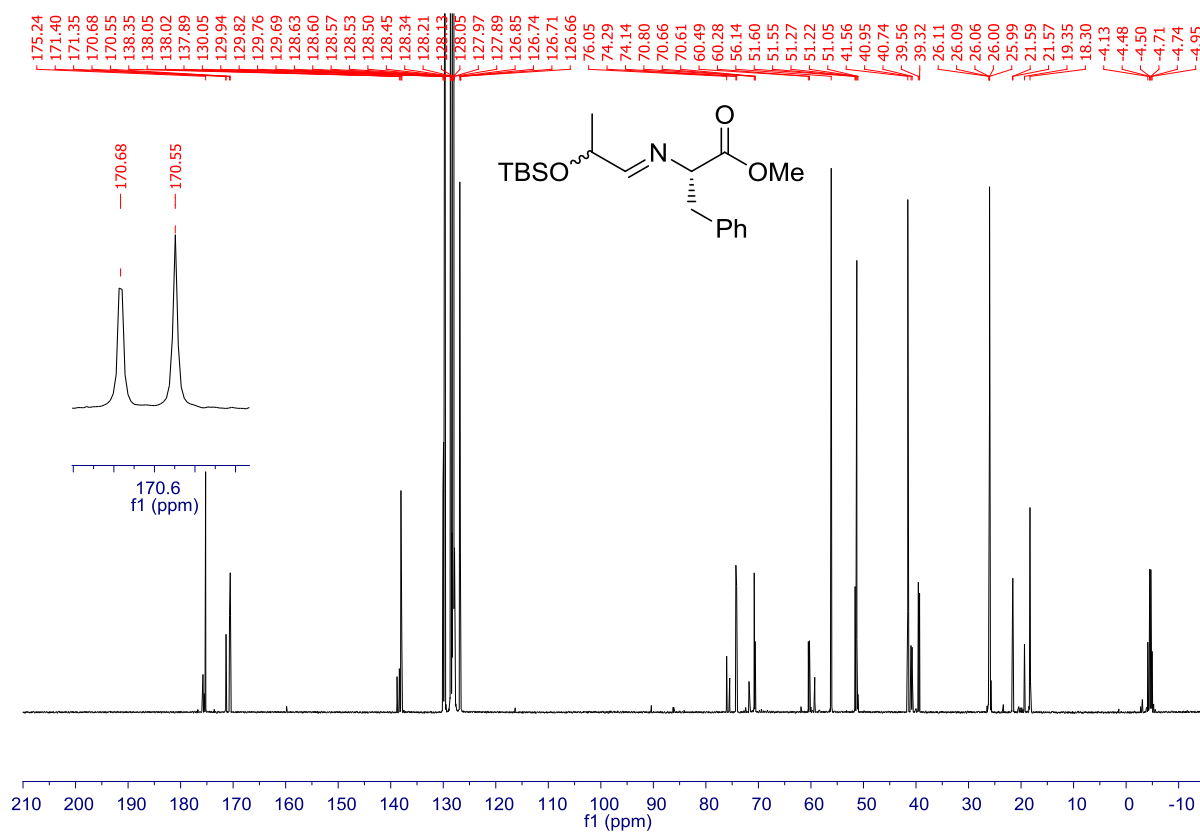
14 days



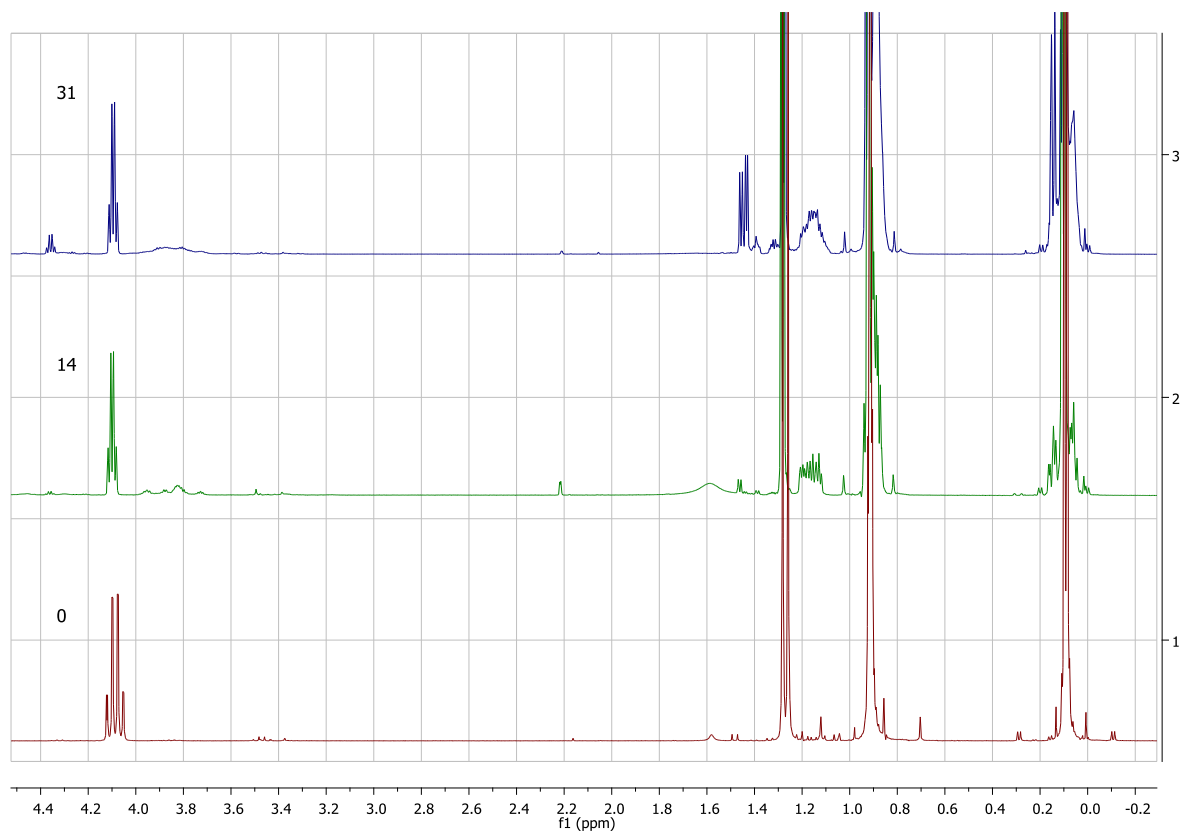
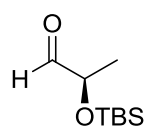
31 days



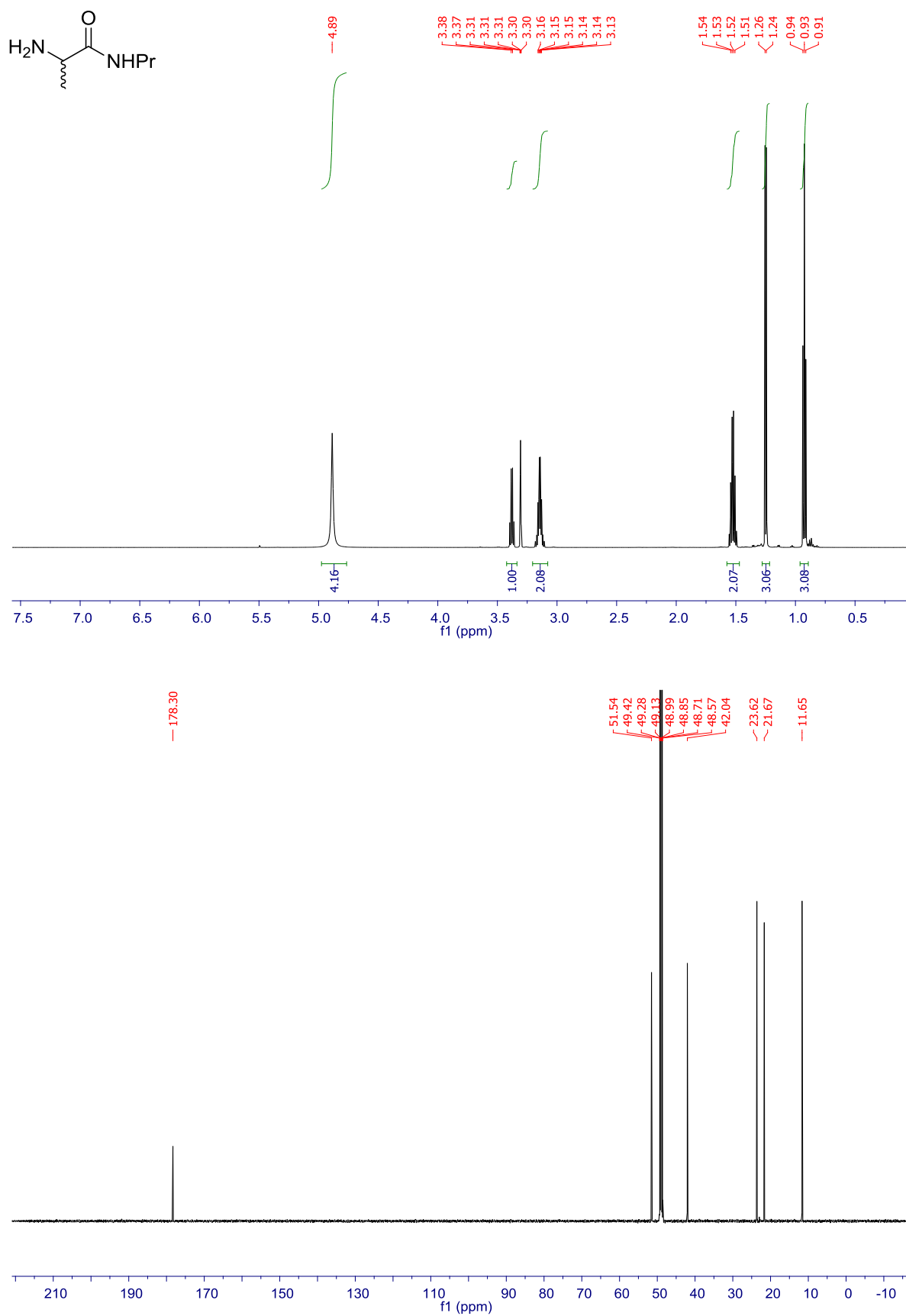
Mixture of (R)-4 and (S)-4 with H-Phe-OMe



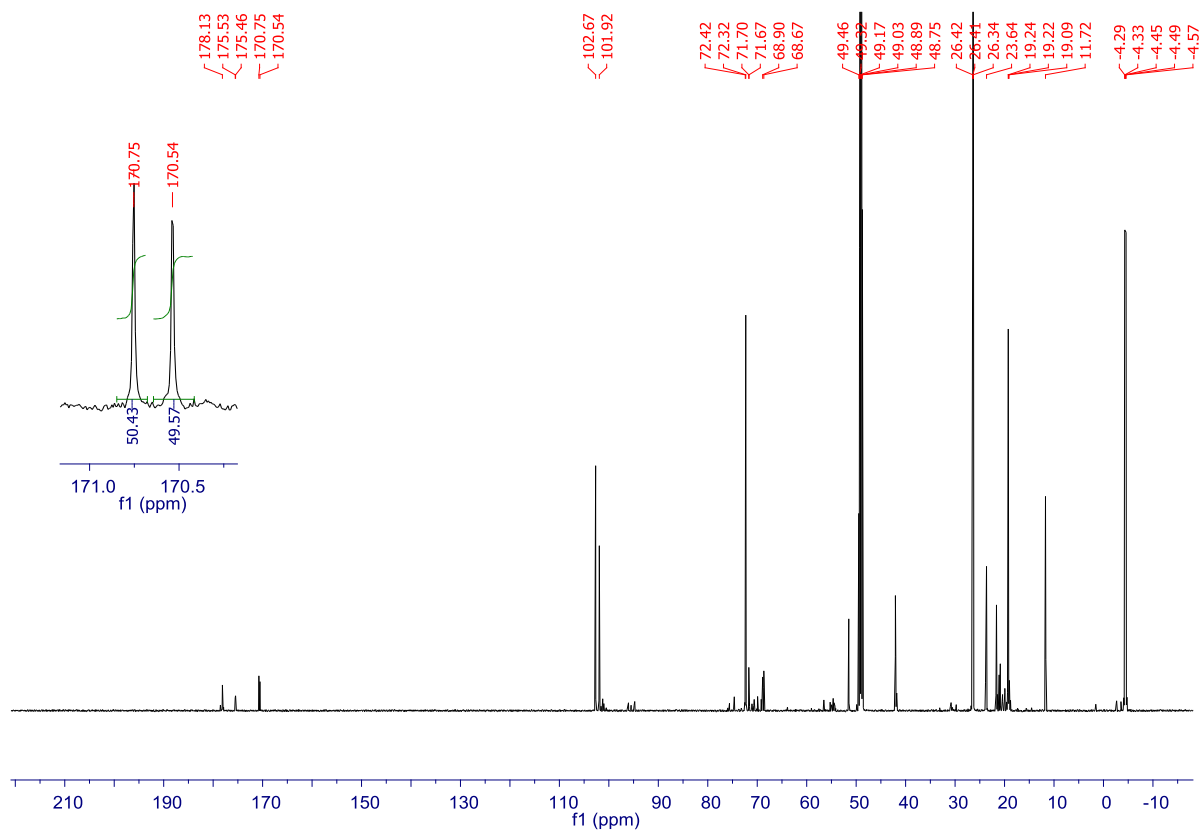
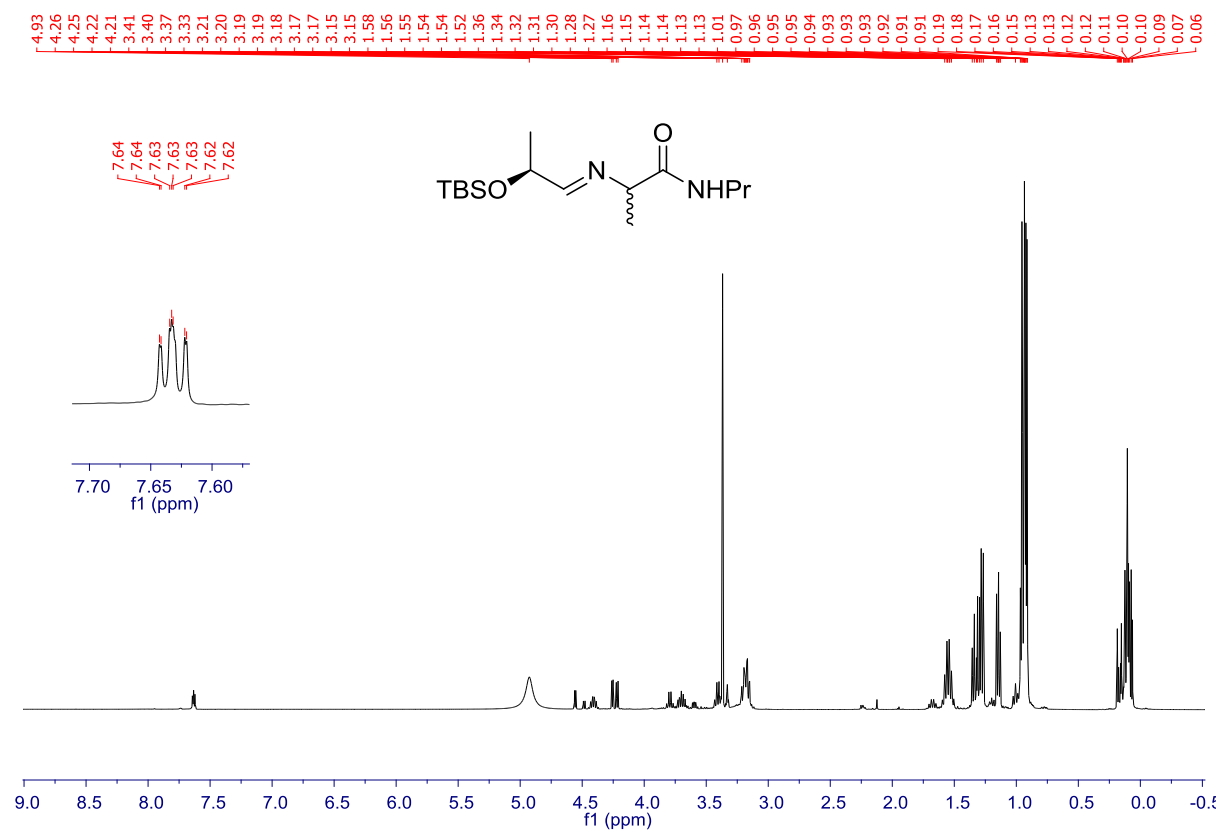
^1H NMR of (*R*)-4 after 0, 14 and 31 days



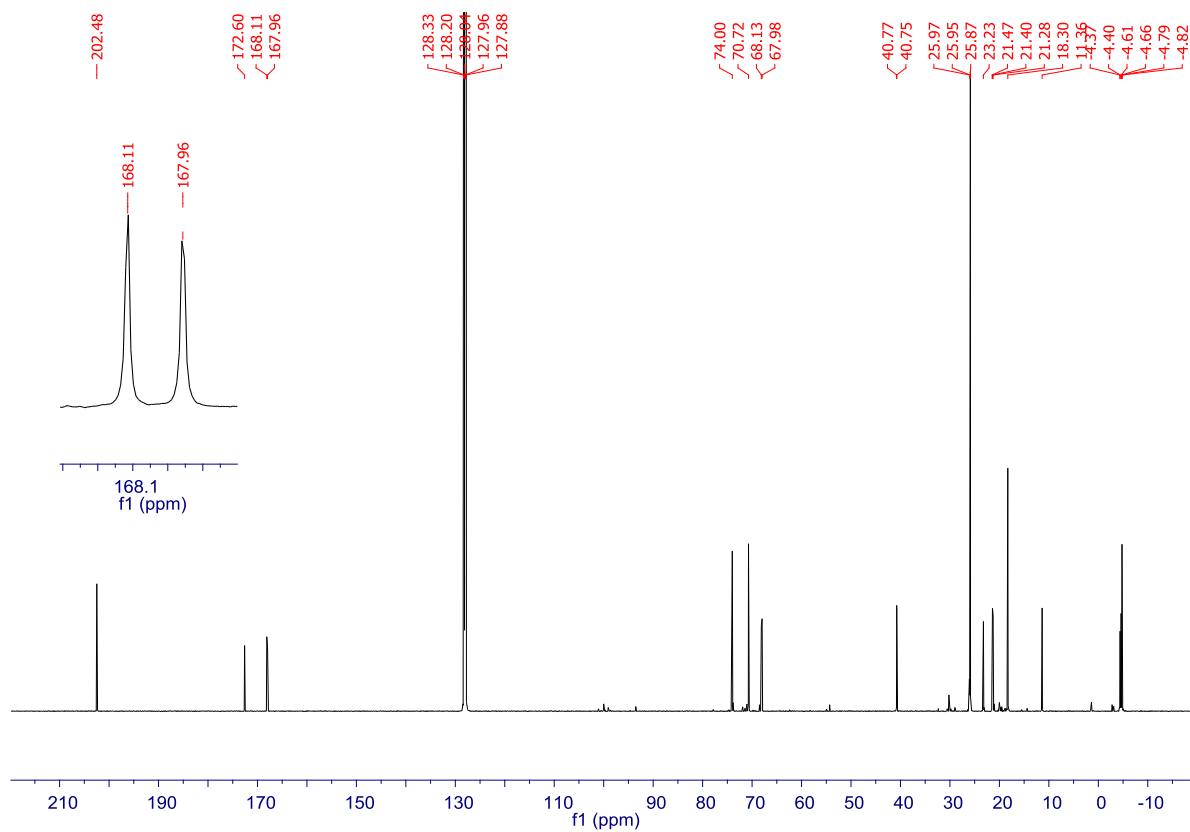
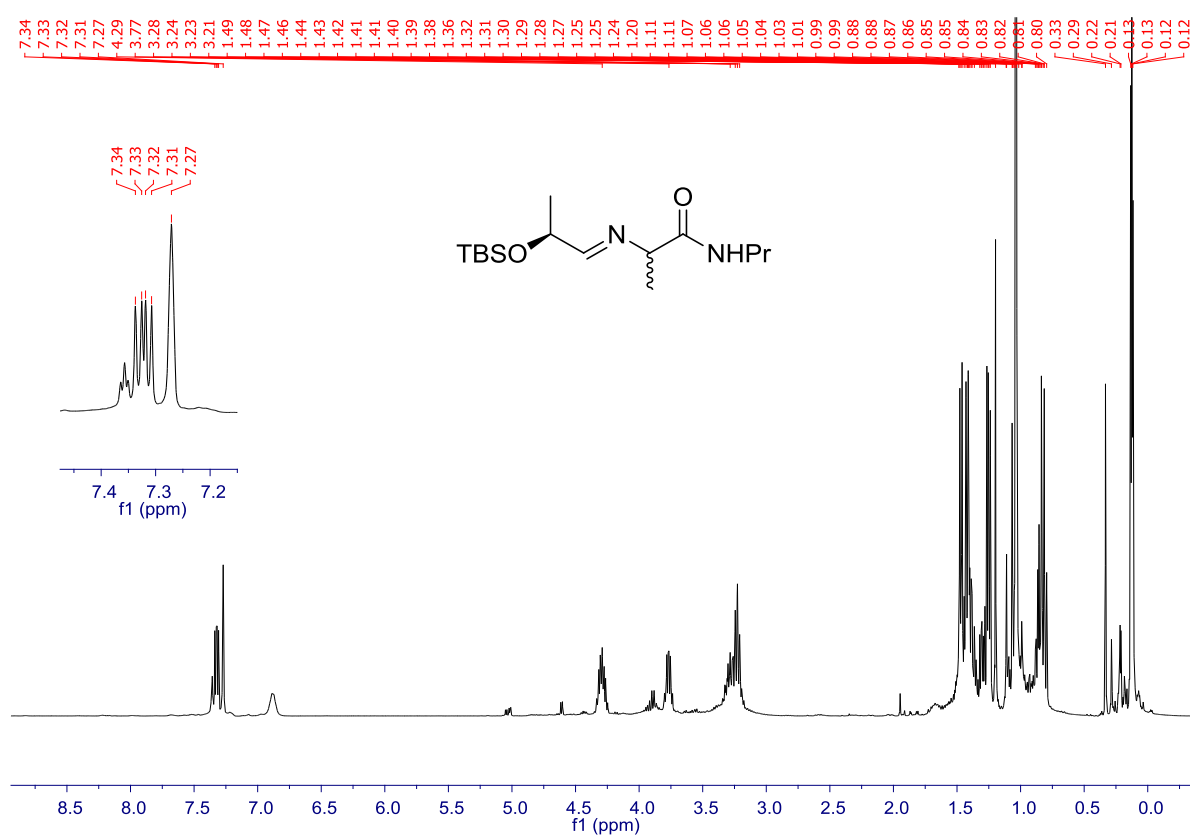
***tert*-butyl (1-oxo-1-(propylamino)propan-2-yl)carbamate (7) (*d*₄-MeOH)**



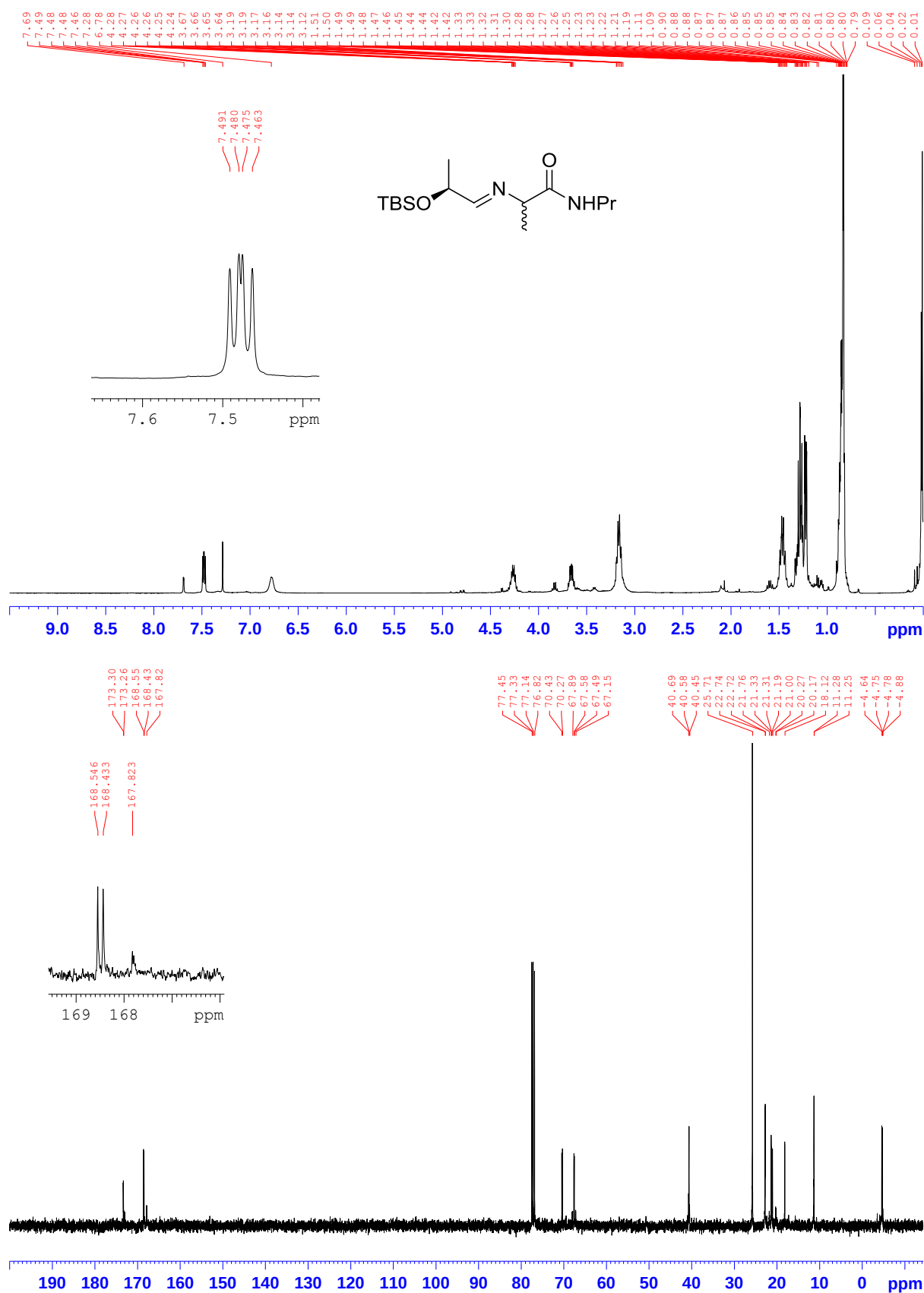
(±)-7 and (S)-4, *d*₄-MeOH



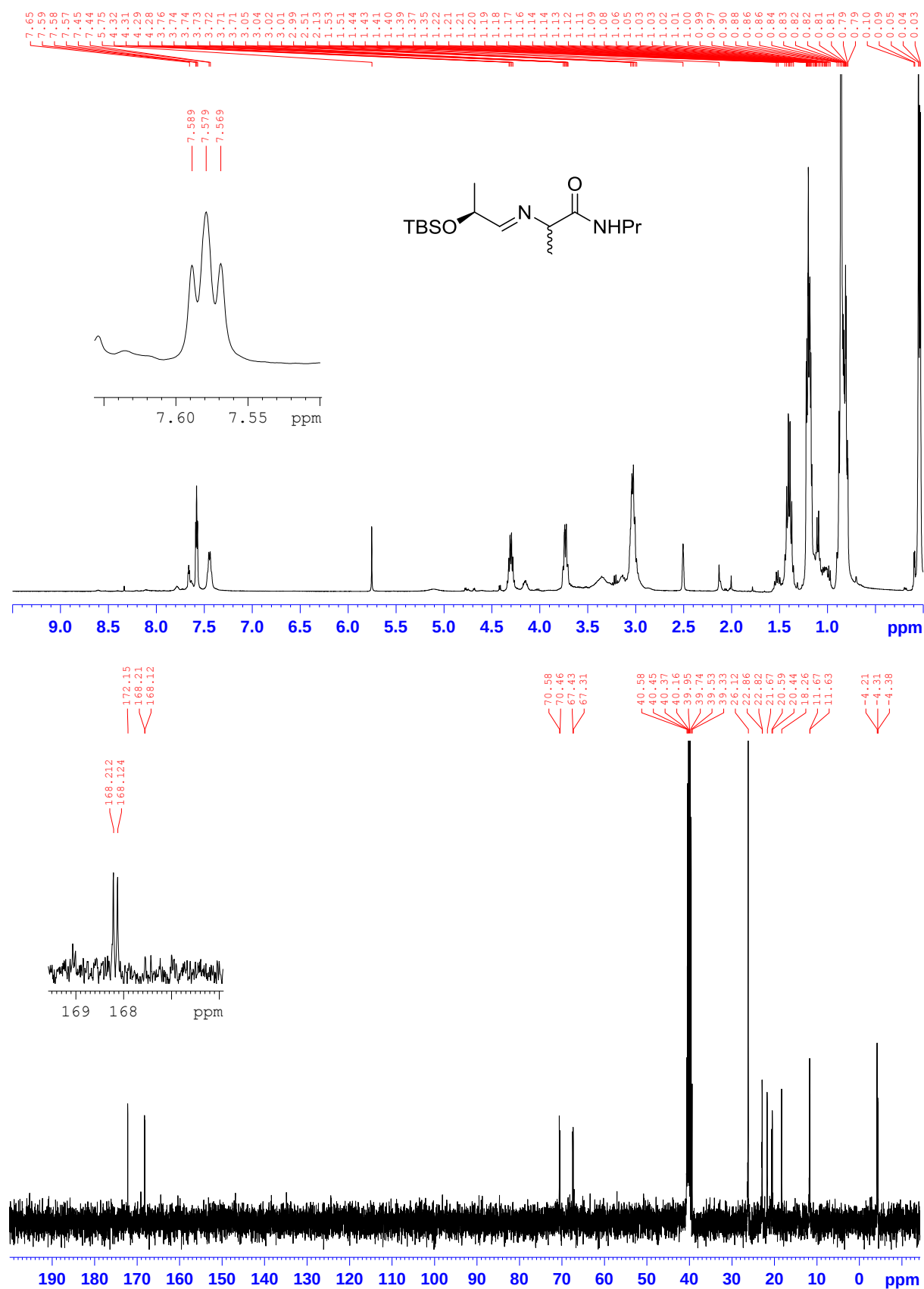
(±)-7 and (S)-4, C₆D₆



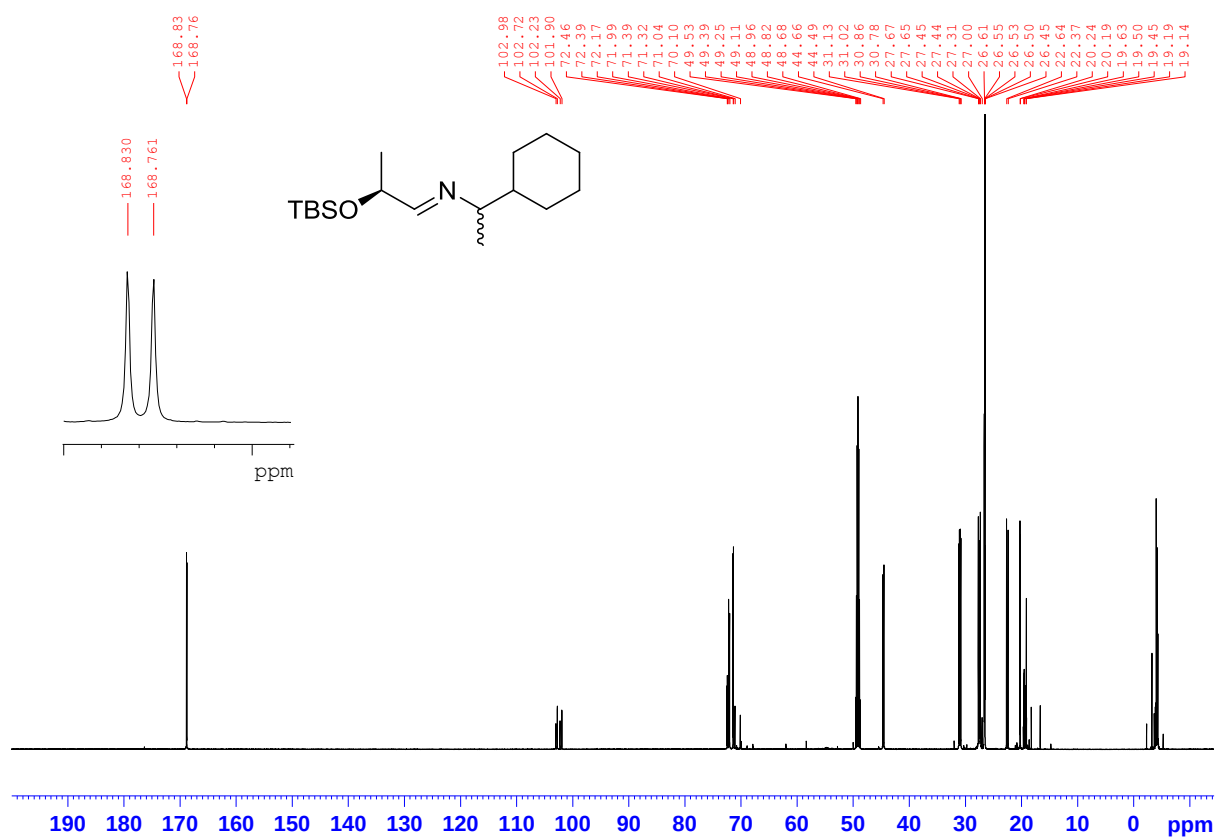
(±)-7 and (S)-4, CDCl₃



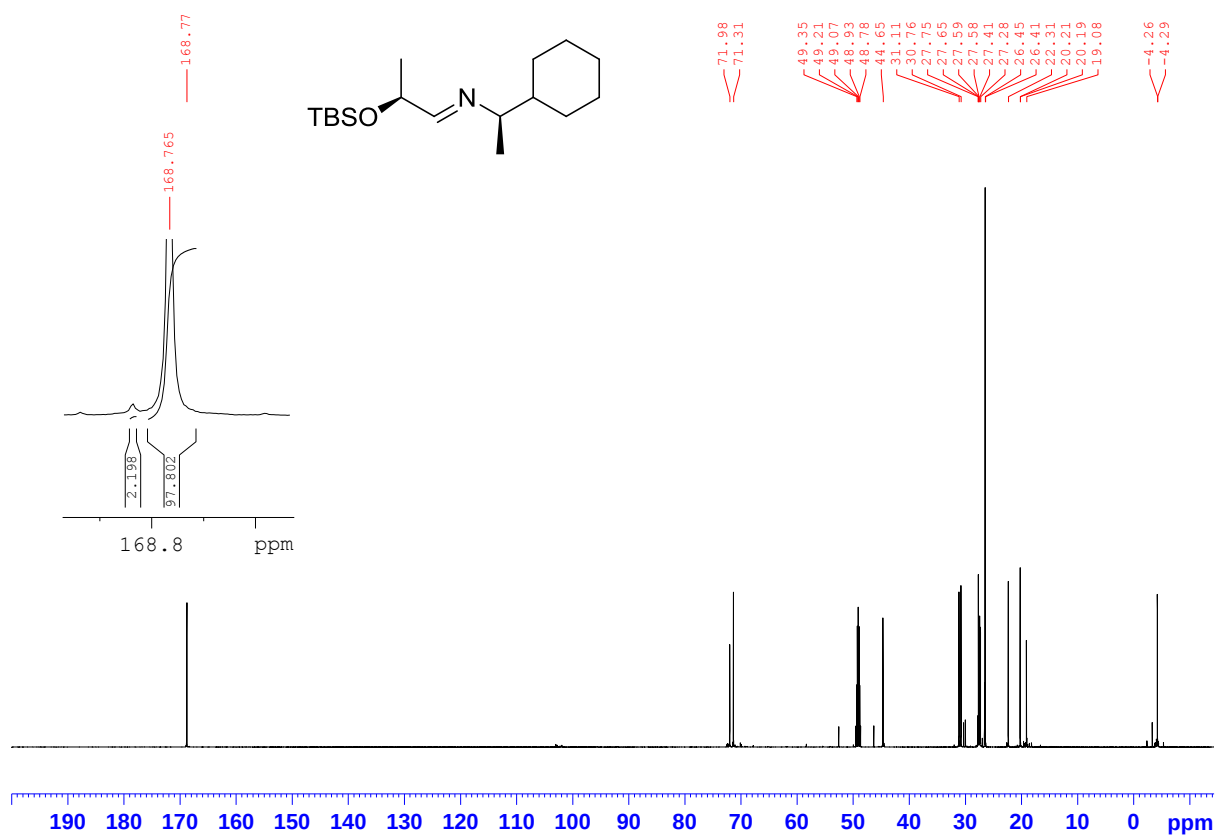
(±)-7 and (S)-4, DMSO-*d*₆



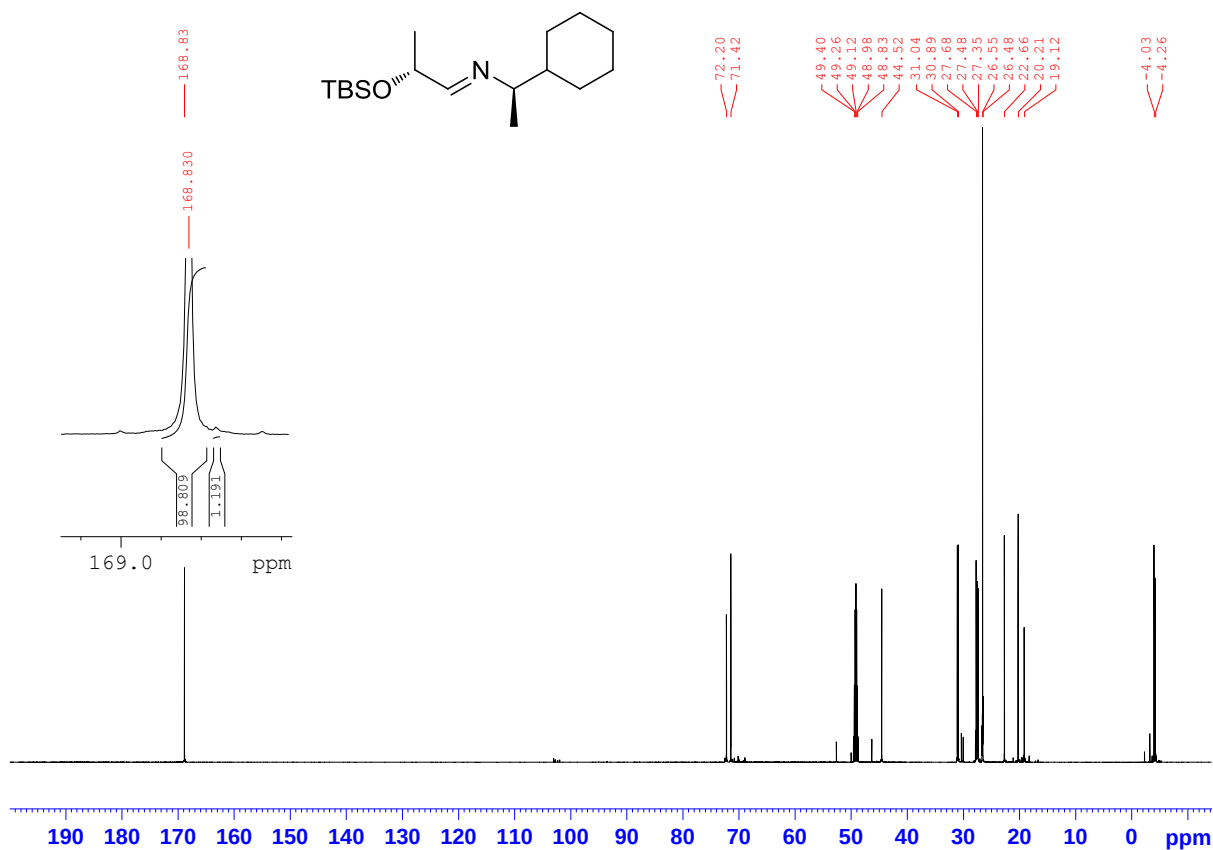
(±)-1-cyclohexylethylamine-(9) and (S)-4, *d*₄-MeOH



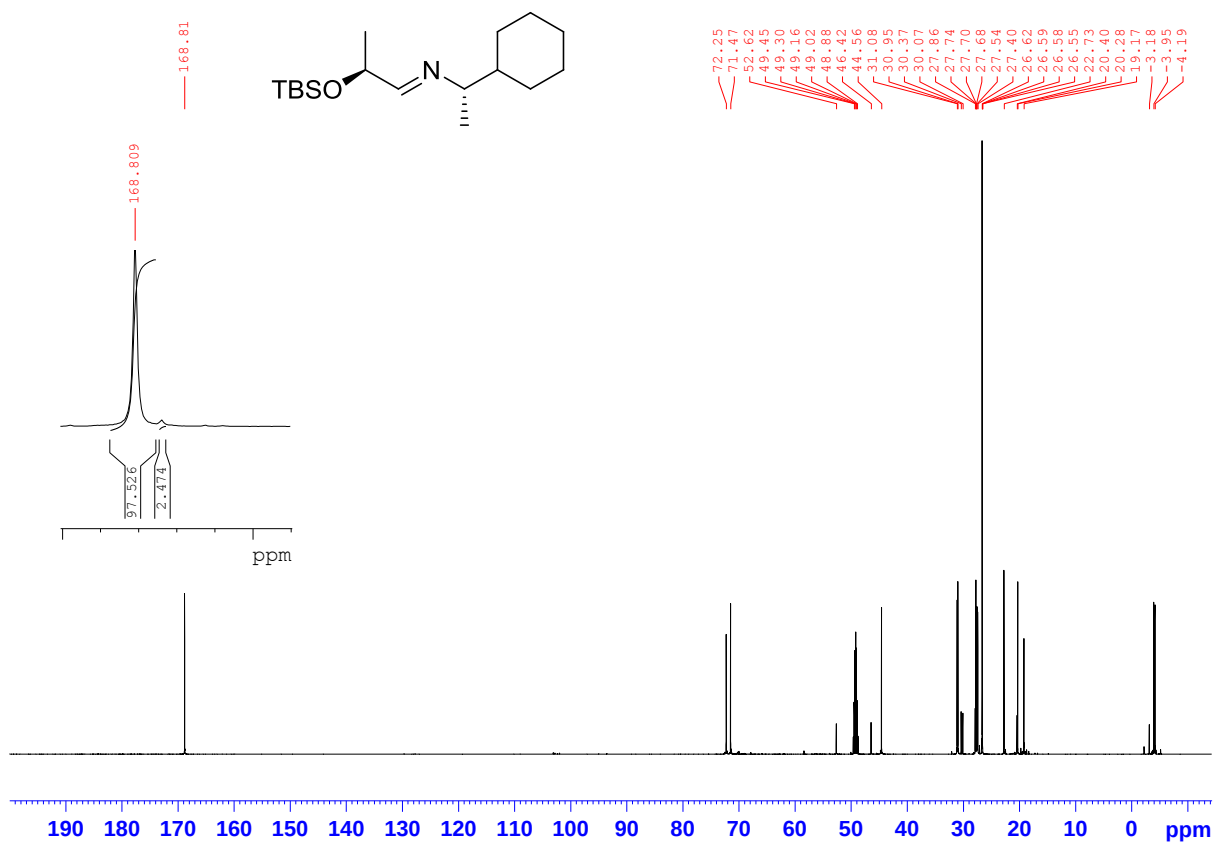
(R)-1-cyclohexylethylamine-(9) and (S)-4, *d*₄-MeOH



(R)-1-cyclohexylethylamine-(9) and (R)-4, *d*₄-MeOH



(S)-1-cyclohexylethylamine-(9) and (S)-4, *d*₄-MeOH



Chemical structure: CC1(C)N(C1)C(C)C2CCCCC2

¹H NMR spectrum (top):

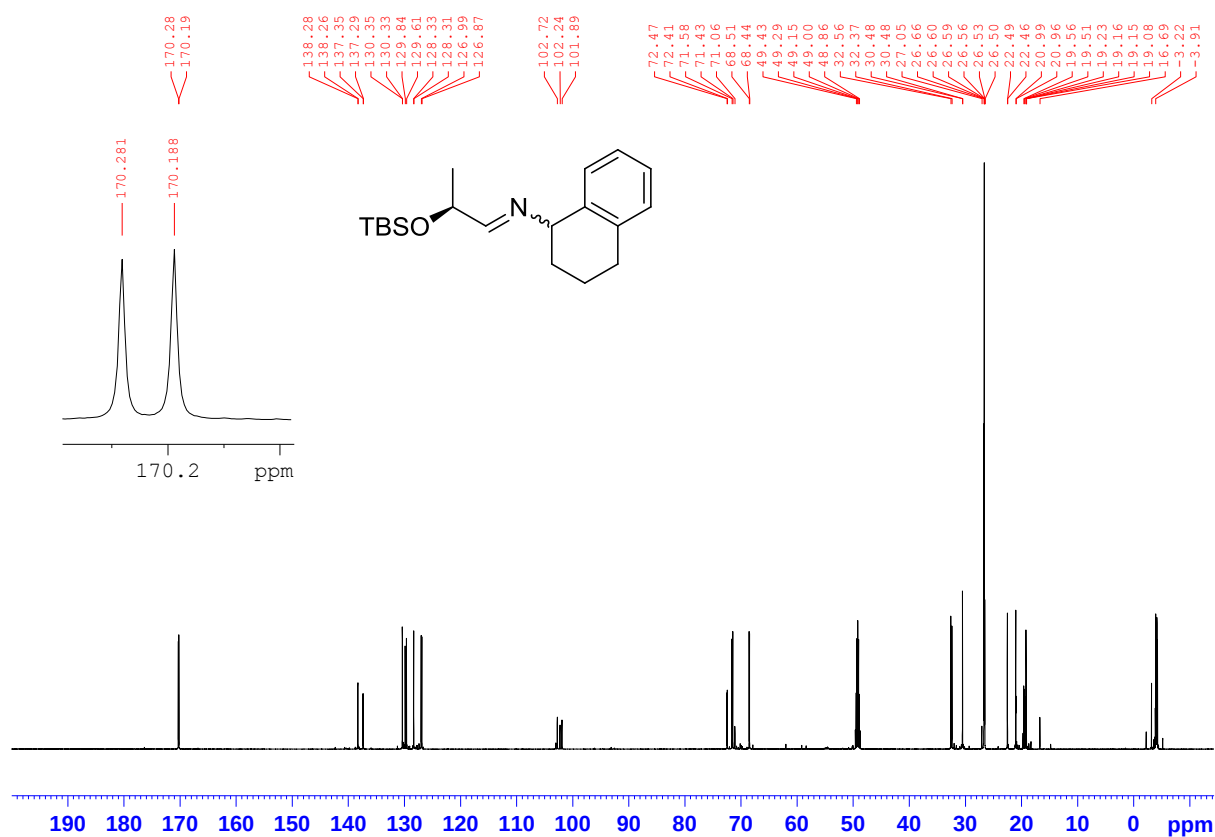
- 7.213, 7.145
- 49.51, 49.37, 49.23, 49.08, 48.94, 44.79, 31.24, 30.32, 30.12, 29.12, 27.83, 27.75, 27.74, 27.59, 27.46, 26.68, 26.64, 22.54, 20.45, 20.42, 19.25, -4.02, -4.05

¹³C NMR spectrum (bottom):

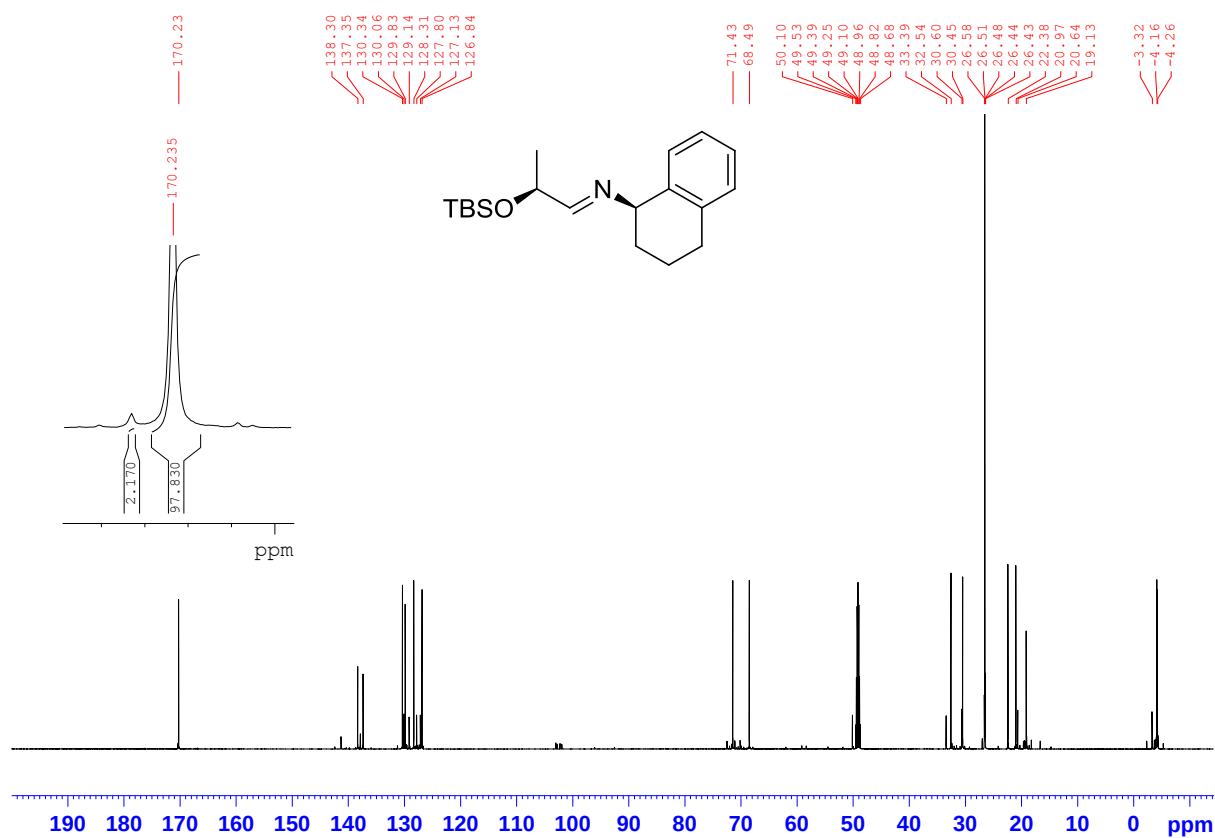
- 168.74

1-cyclohexylethylamine (9)	Aldehyde (4)	<i>S</i>	<i>R</i>
<i>R</i>		2:98	99:1
<i>S</i>		98:2	1:99

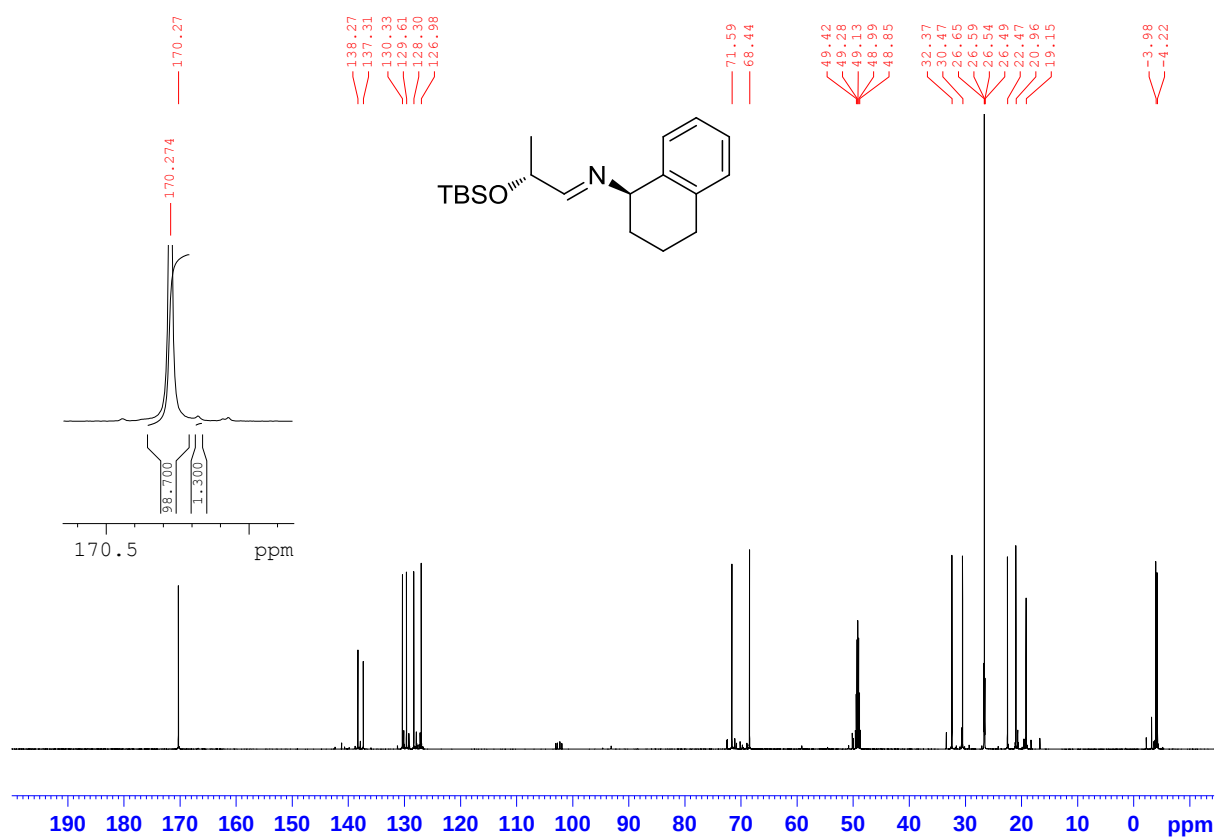
(±)-tetrahydro-1-naphthylamine-(10) and (S)-4, d₄-MeOH



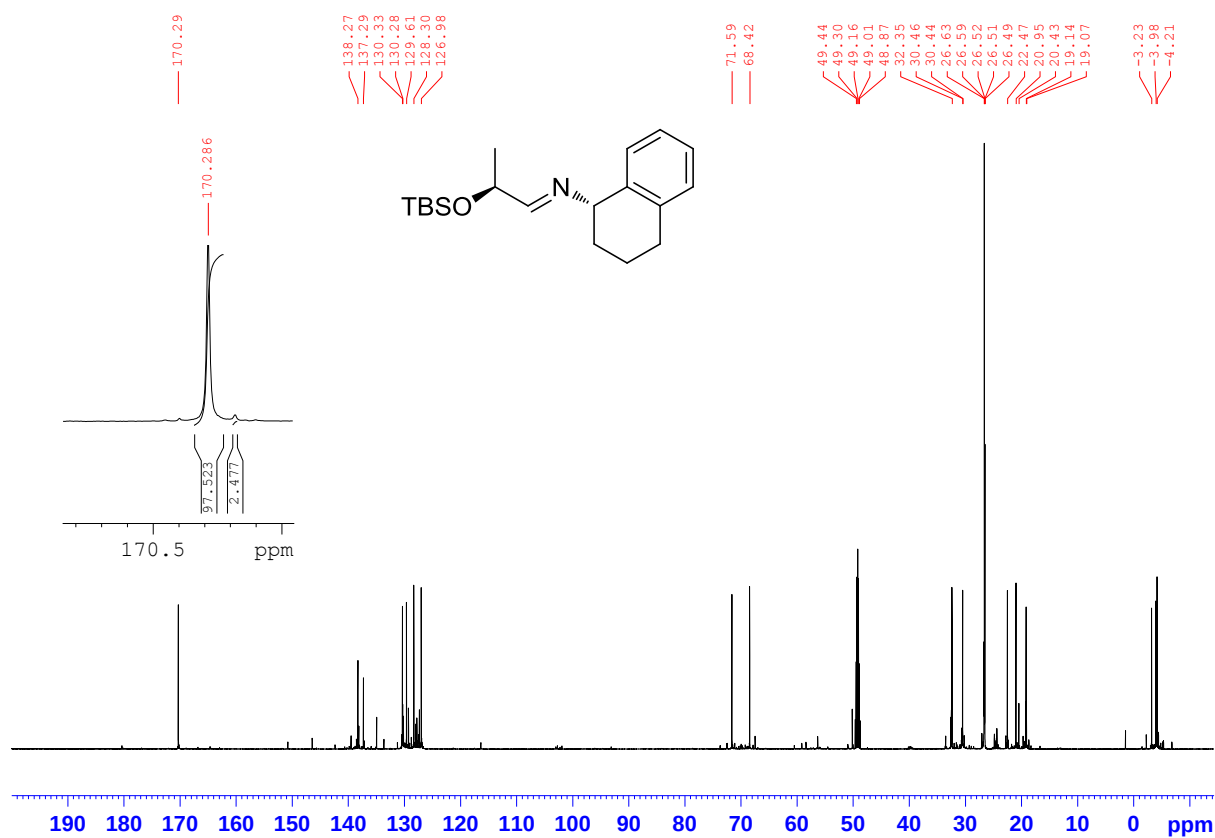
(R)-Tetrahydro-1-naphthylamine-(10) and (S)-4, d₄-MeOH



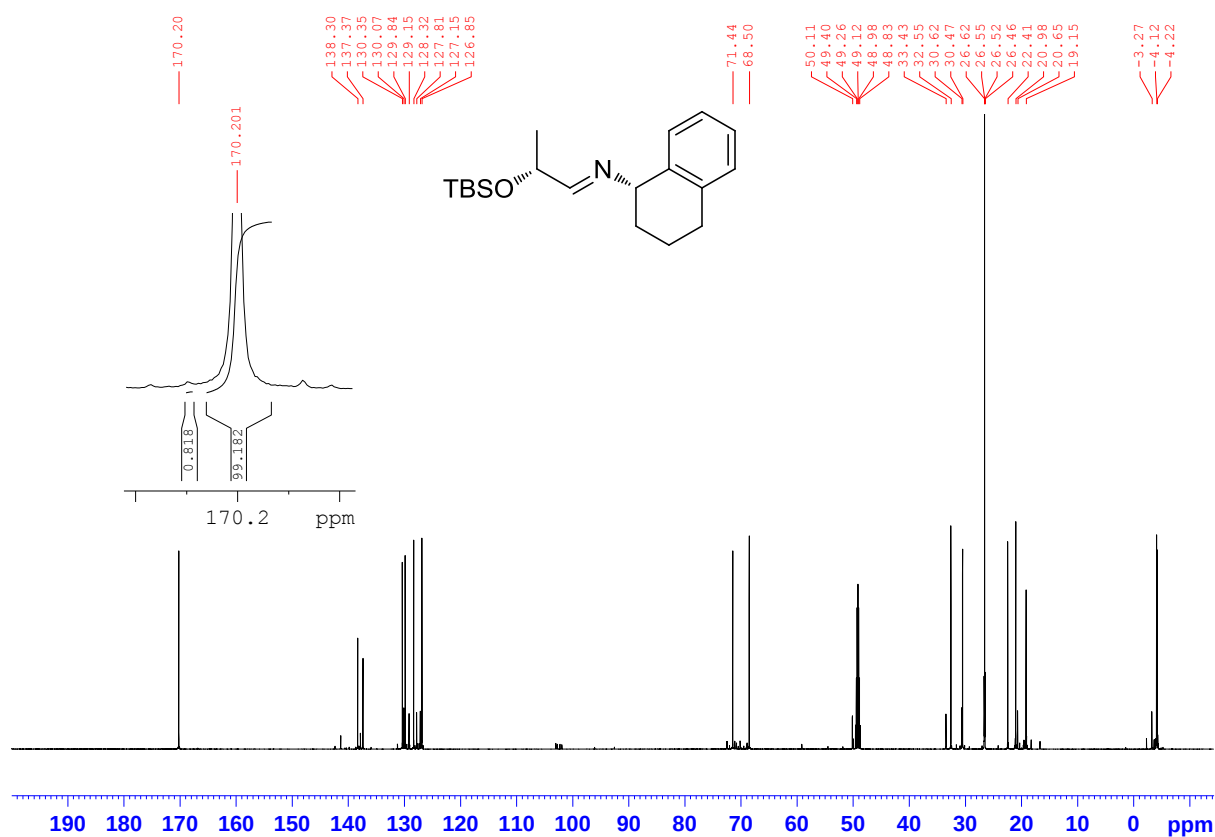
(R)-tetrahydro-1-naphthylamine-(10) and (R)-4, *d*₄-MeOH



(S)-tetrahydro-1-naphthylamine-(10) and (S)-4, *d*₄-MeOH



(S)-tetrahydro-1-naphthylamine-(10) and (R)-4, *d*₄-MeOH



1,2,3,4-tetrahydro-1-naphthylamine (10)	Aldehyde (4)	<i>S</i>	<i>R</i>
<i>R</i>		2:98	99:1
<i>S</i>		98:2	1:99

Chemical structure: CC(C)(O[Si](C)(C)C(C)(C)C)C=Cc1ccccc1

¹³C NMR spectrum (CDCl₃) showing peaks (ppm):

- 169.156
- 169.017
- 169.16
- 169.02
- 145.09
- 144.98
- 129.62
- 129.60
- 129.59
- 129.54
- 129.50
- 128.30
- 128.25
- 128.18
- 128.15
- 128.10
- 127.75
- 127.64
- 102.69
- 102.20
- 101.91
- 72.44
- 72.36
- 71.53
- 71.04
- 70.34
- 70.12
- 70.03
- 69.95
- 49.52
- 49.38
- 49.24
- 49.10
- 48.96
- 48.82
- 48.61
- 48.56
- 48.54
- 48.47
- 46.43
- 46.40
- 46.38
- 45.61
- 44.01
- 23.92
- 22.14
- 22.12
- 19.92
- 19.39
- 19.36
- 19.16
- 19.15
- 19.10
- 19.08
- 19.06
- 19.05
- 19.01
- 18.17

Inset peaks (ppm):

- 169.156
- 169.017

Chemical structure: CC(C)C(=N[C@@H](C)Cc1ccccc1)C(=O)N1CCCC1

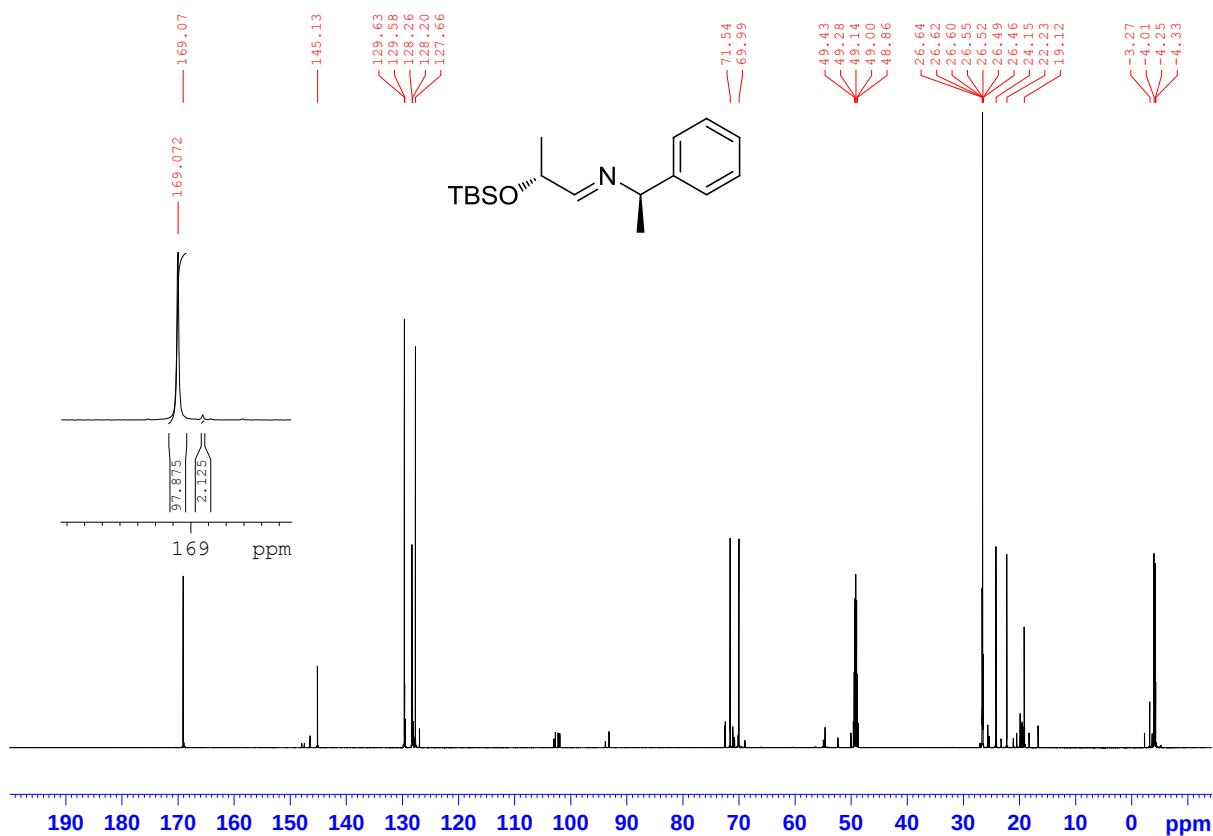
¹³C NMR spectrum (CDCl₃) showing peaks (ppm):

- 168.95, 168.946 (Carbonyl)
- 145.00
- 129.61, 129.52, 128.32, 128.17, 128.13, 128.04, 127.77, 126.92 (Aromatic)
- 71.54, 70.33, 70.06 (Solvent)
- 49.56, 49.42, 49.27, 49.13, 48.99, 48.85, 48.71, 48.61, 46.58, 46.54, 46.50, 46.47, 46.45, 37.70, 32.00, 32.18, 30.01, 19.08, 19.04 (Aliphatic)
- 3.29, -4.18, -4.27, -4.32, -4.35 (Methyls)

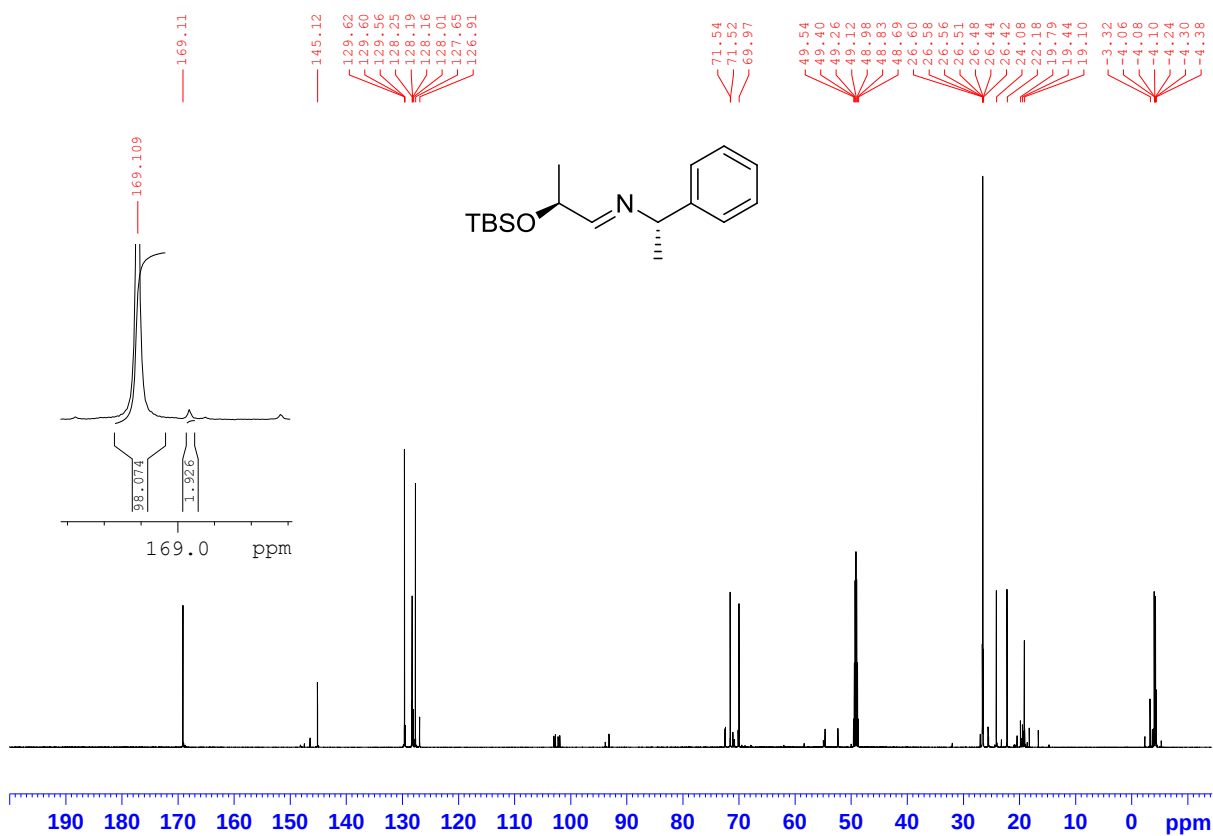
Zoomed-in view of the carbonyl region (ppm):

- 169.057 (Integration: 3.057)
- 168.943 (Integration: 96.943)

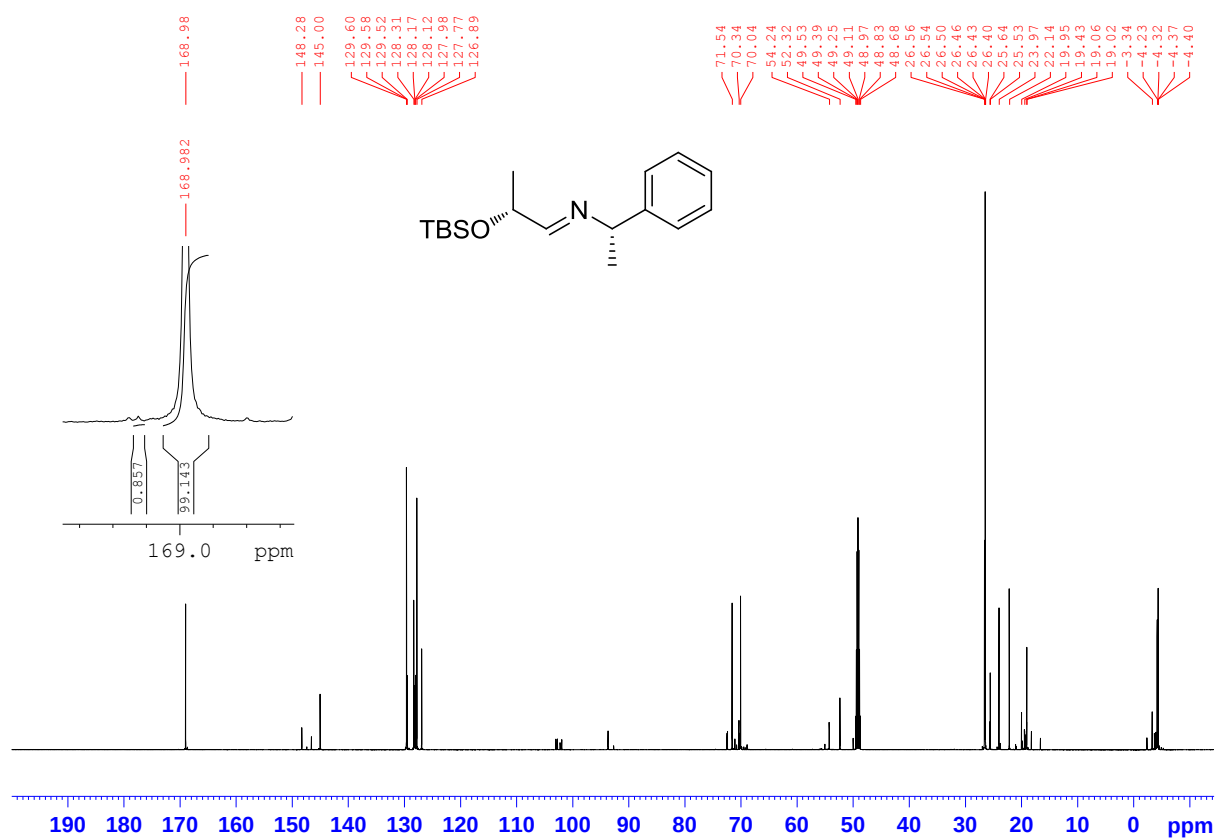
(R)- α -methylbenzylamine-(11) and (R)-4, d_4 -MeOH



(S)- α -methylbenzylamine-(11) and (S)-4, d_4 -MeOH

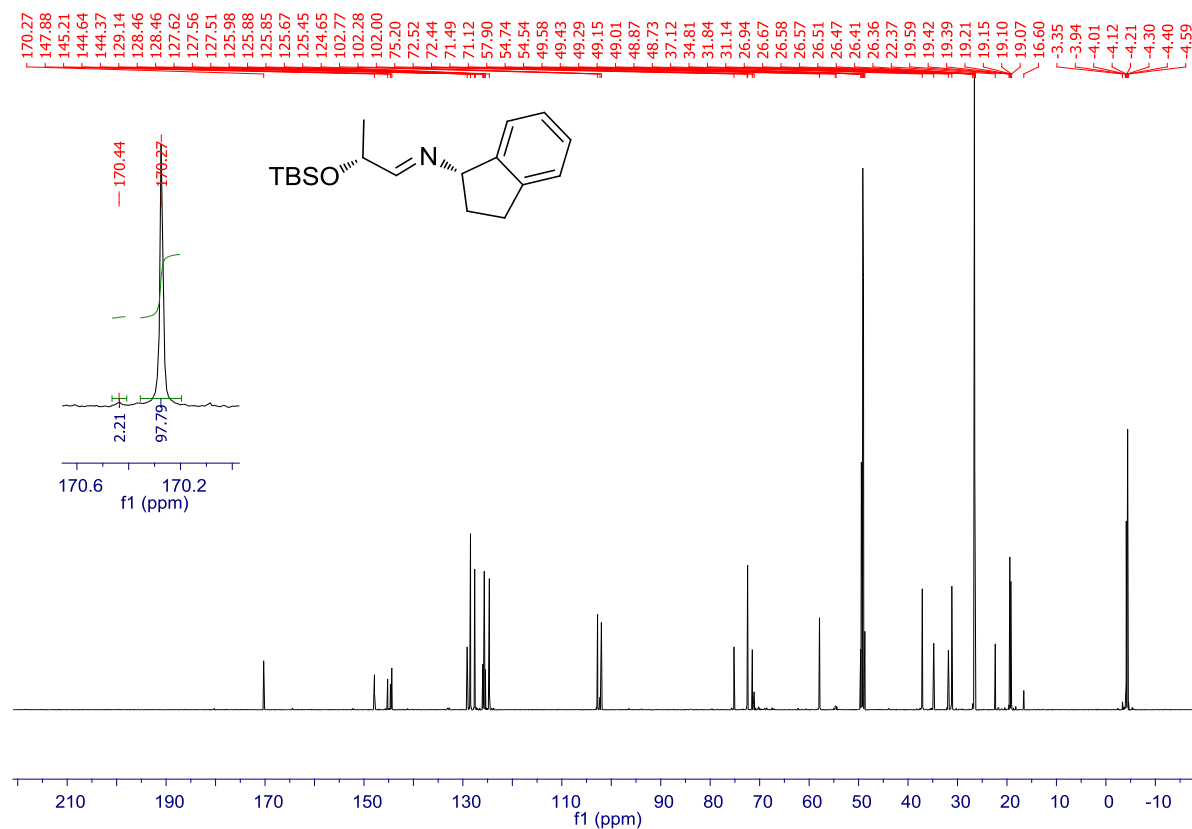


(S)- α -methylbenzylamine-(11) and (R)-4, d_4 -MeOH

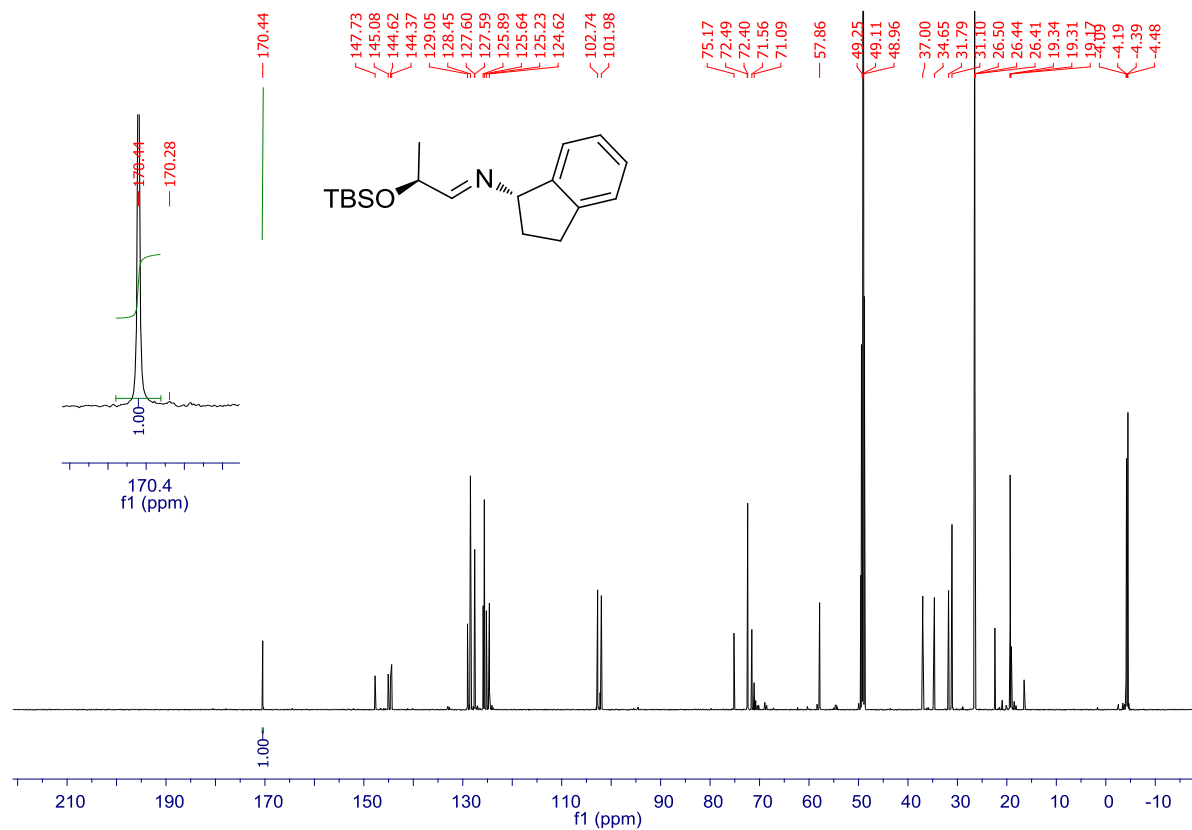


α -methylbenzylamine (11)	Aldehyde (4)	S	R
R		3:97	98:2
S		98:2	1:99

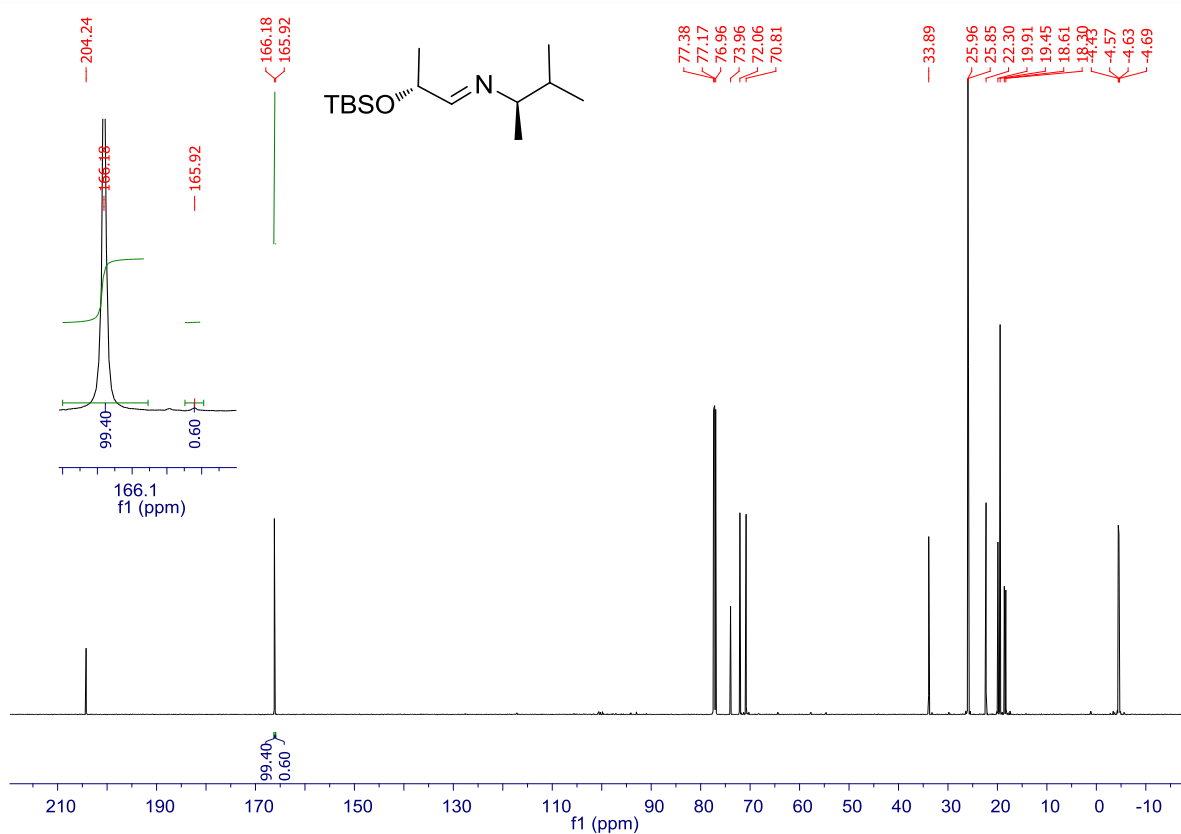
(S)-(+)-1-Aminoindan-(12) and (R)-4, d_4 -MeOH



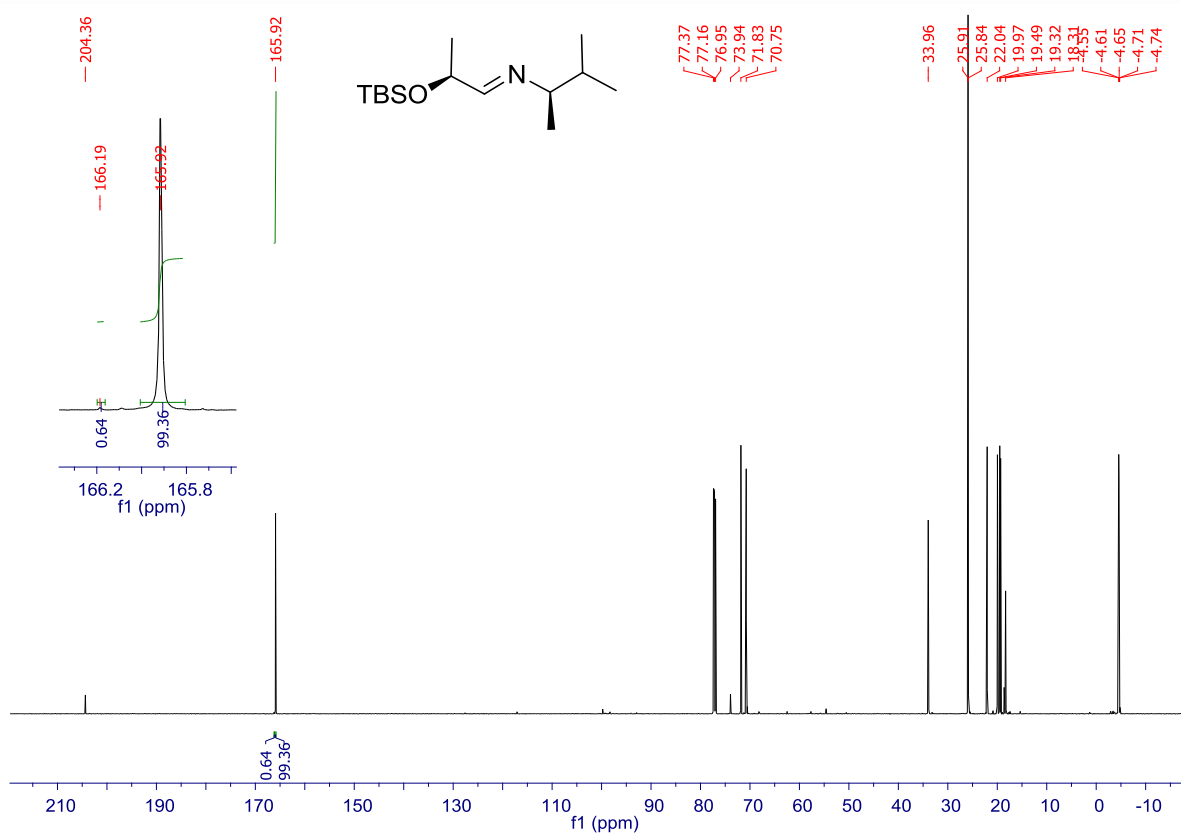
(S)-(+)-1-Aminoindan-(12) and (S)-4, d_4 -MeOH



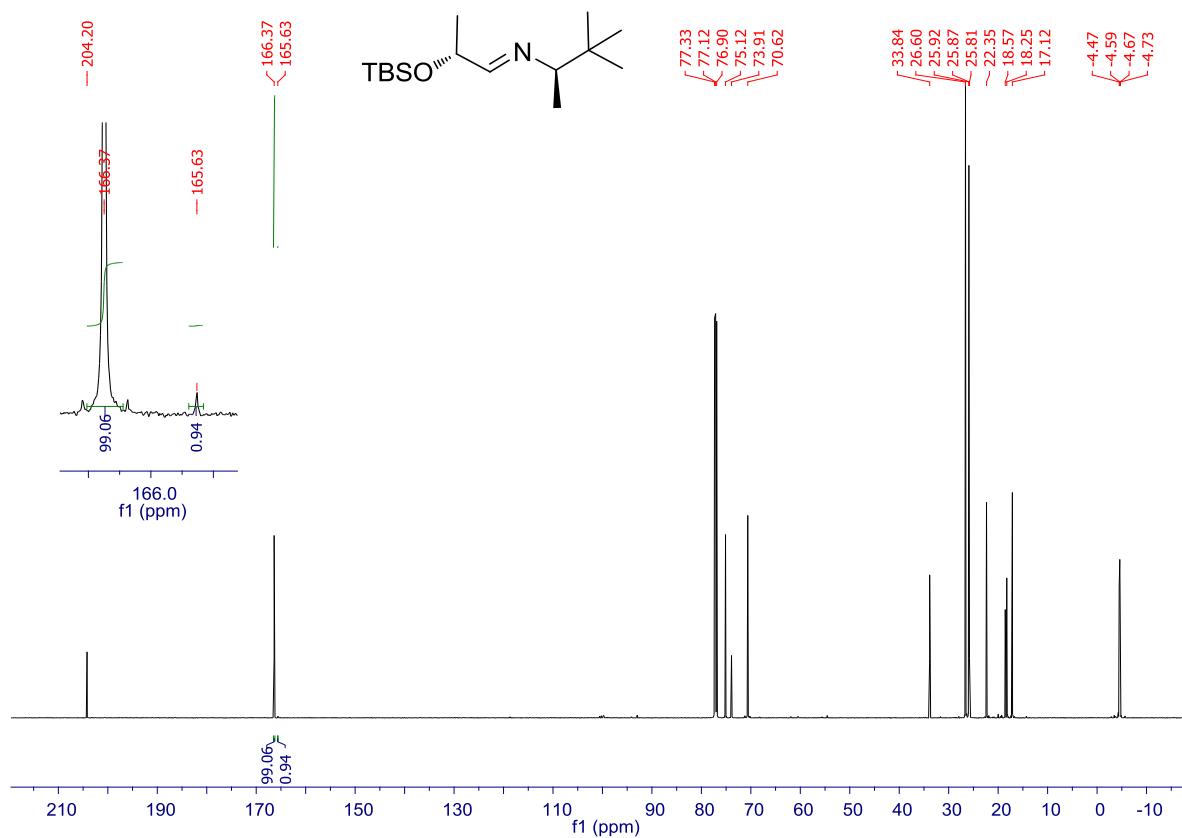
(R)-(-)-2-Amino-3-methylbutane-(13) and (R)-4, CDCl₃



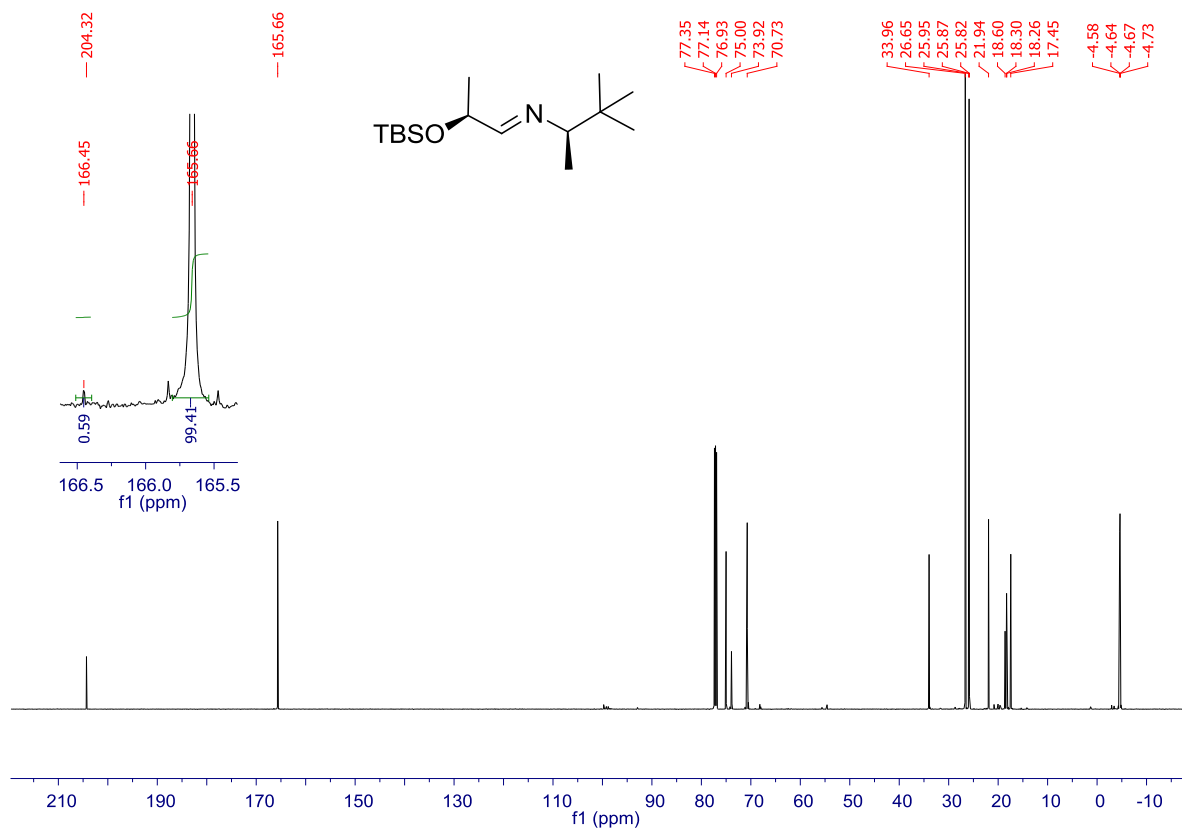
(R)-(-)-2-Amino-3-methylbutane-(13) and (S)-4, CDCl₃



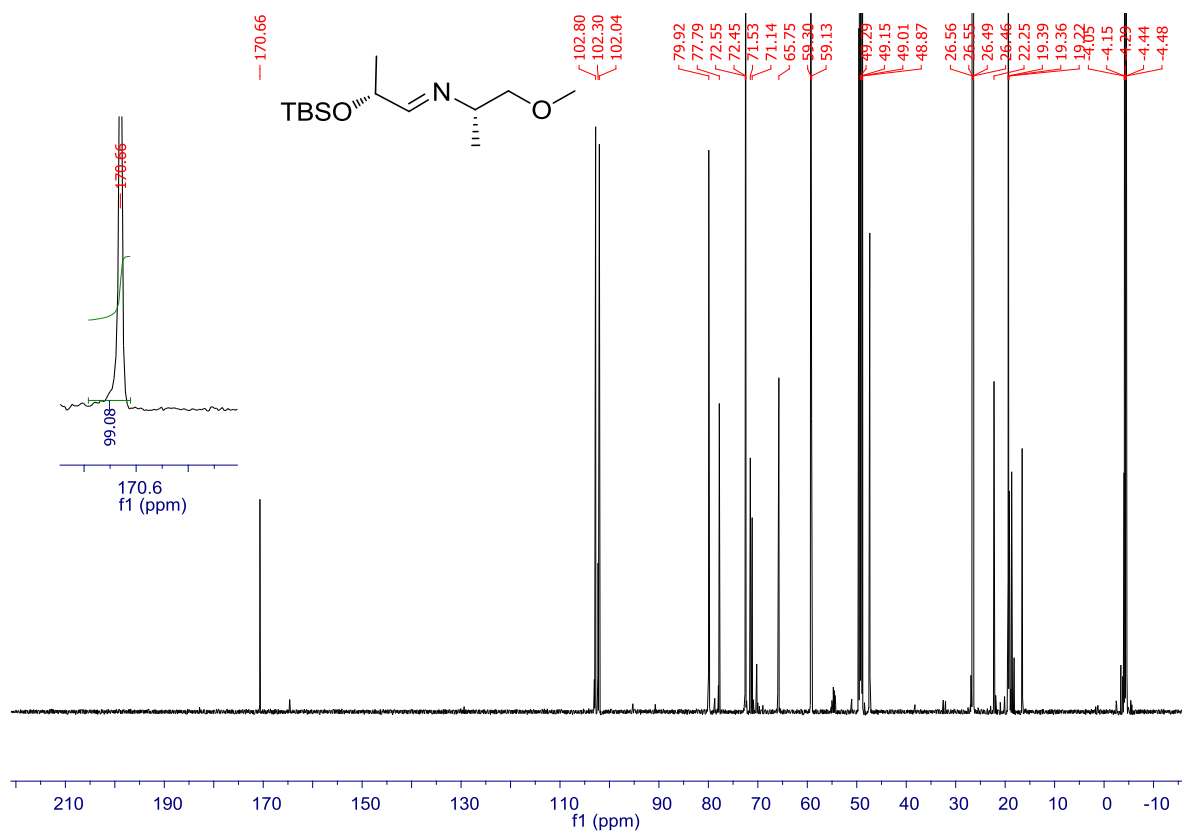
(R)-(-)-3,3-Dimethyl-2-butylamine-(14) and (R)-4, CDCl₃



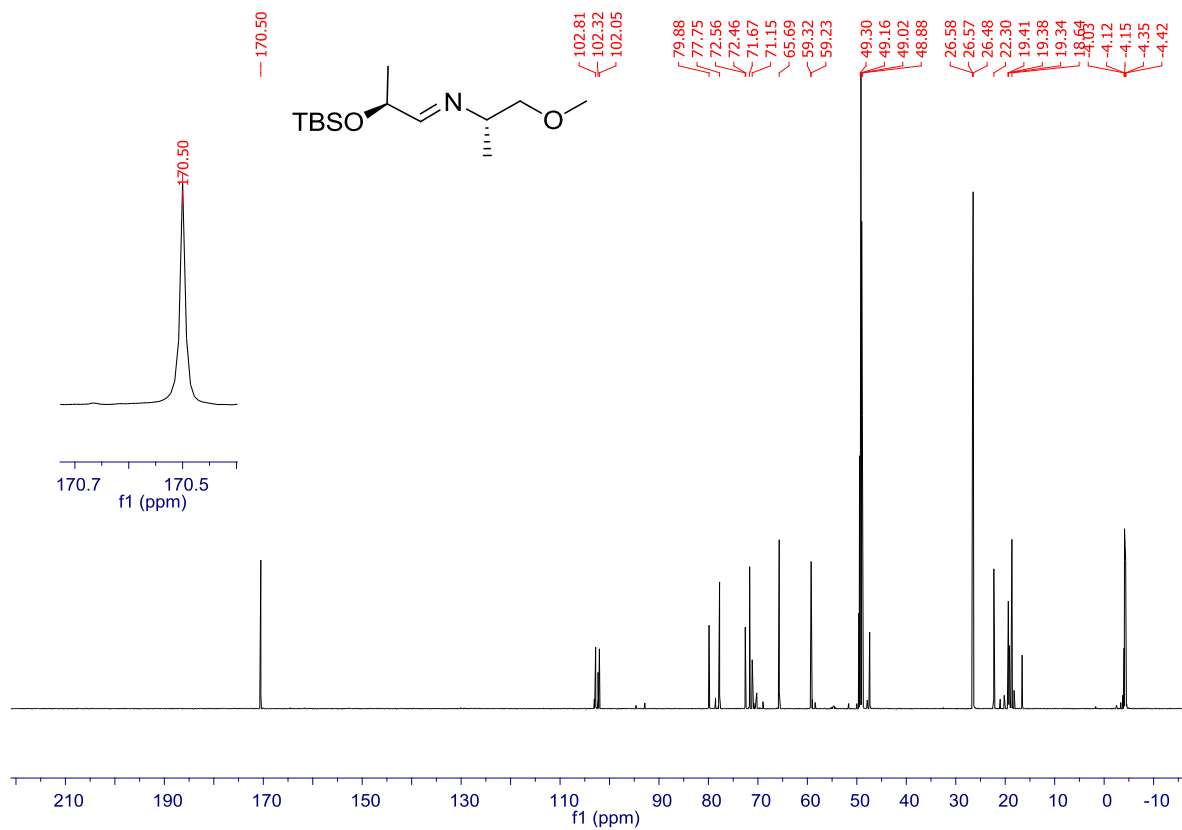
(R)-(-)-3,3-Dimethyl-2-butylamine-(14) and (S)-4, CDCl₃



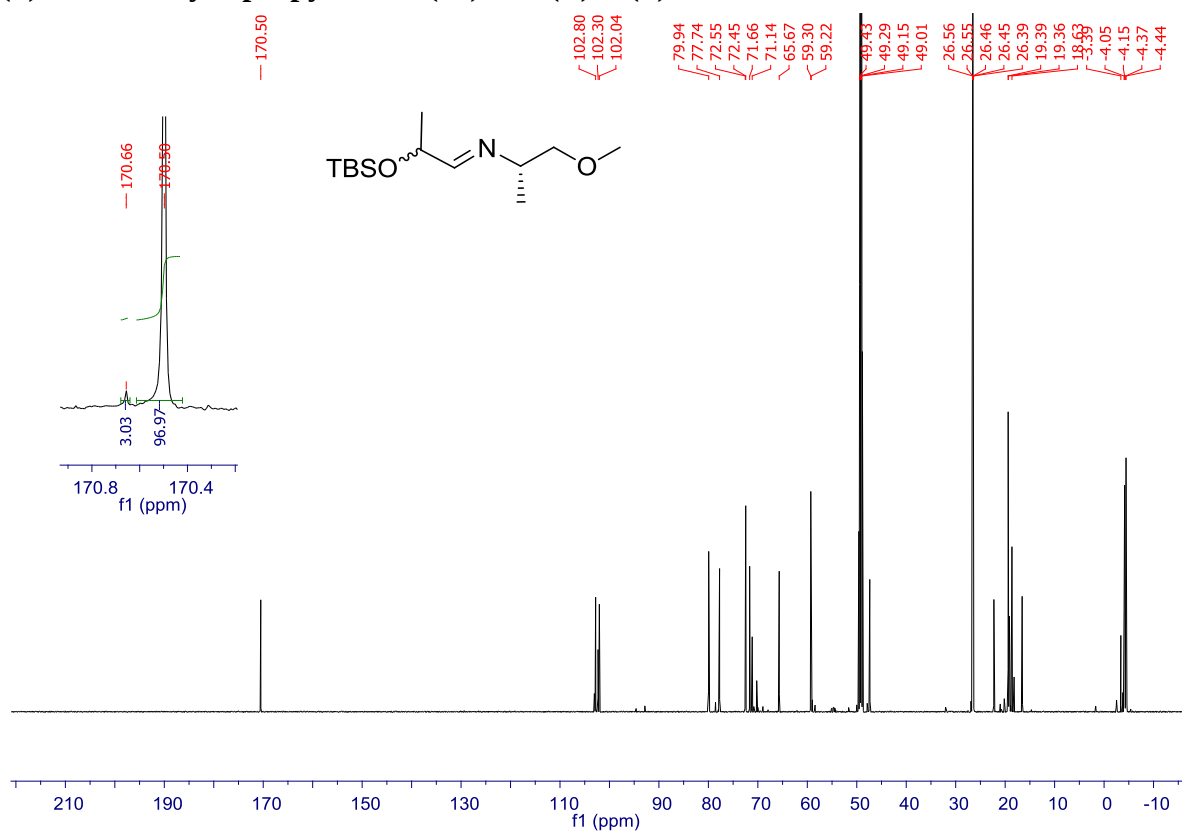
(S)-1-Methoxy-2-propylamine-(15) and (R)-4, d_4 -MeOH



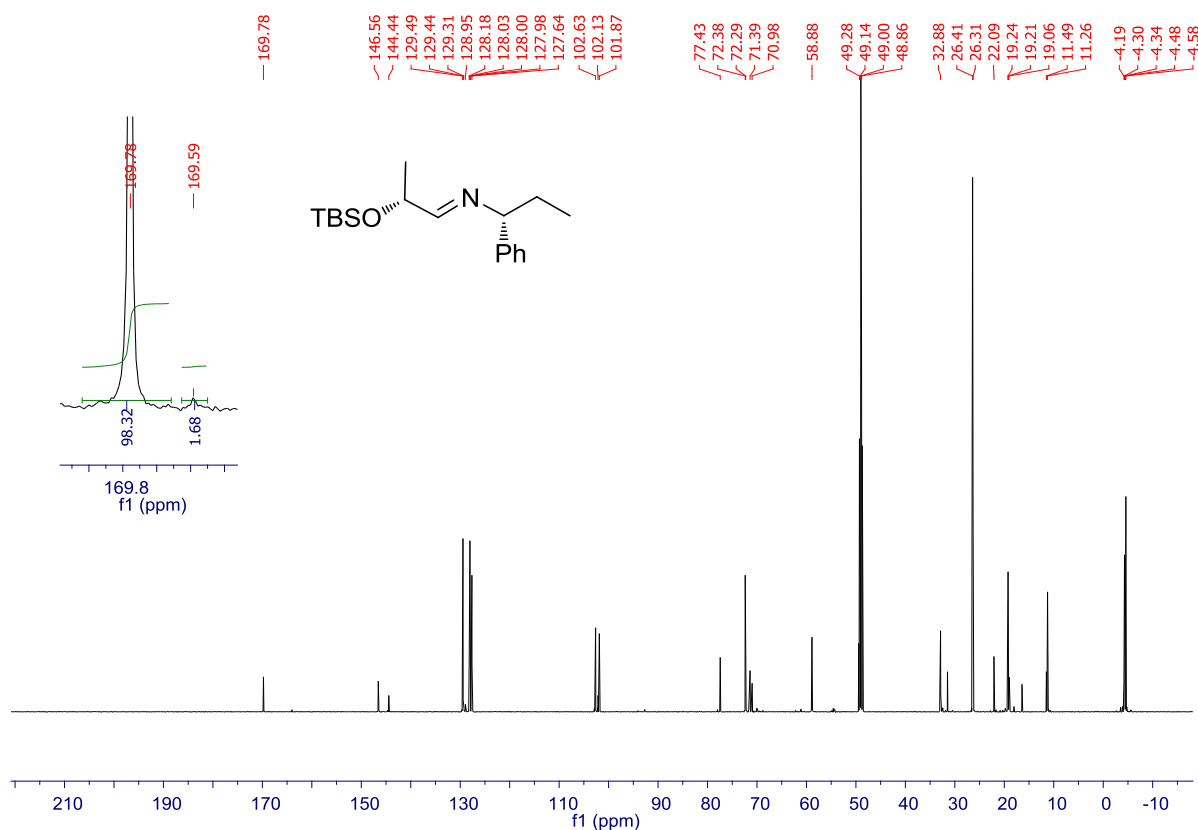
(S)- 1-Methoxy-2-propylamine-(15) and (S)-4, d_4 -MeOH



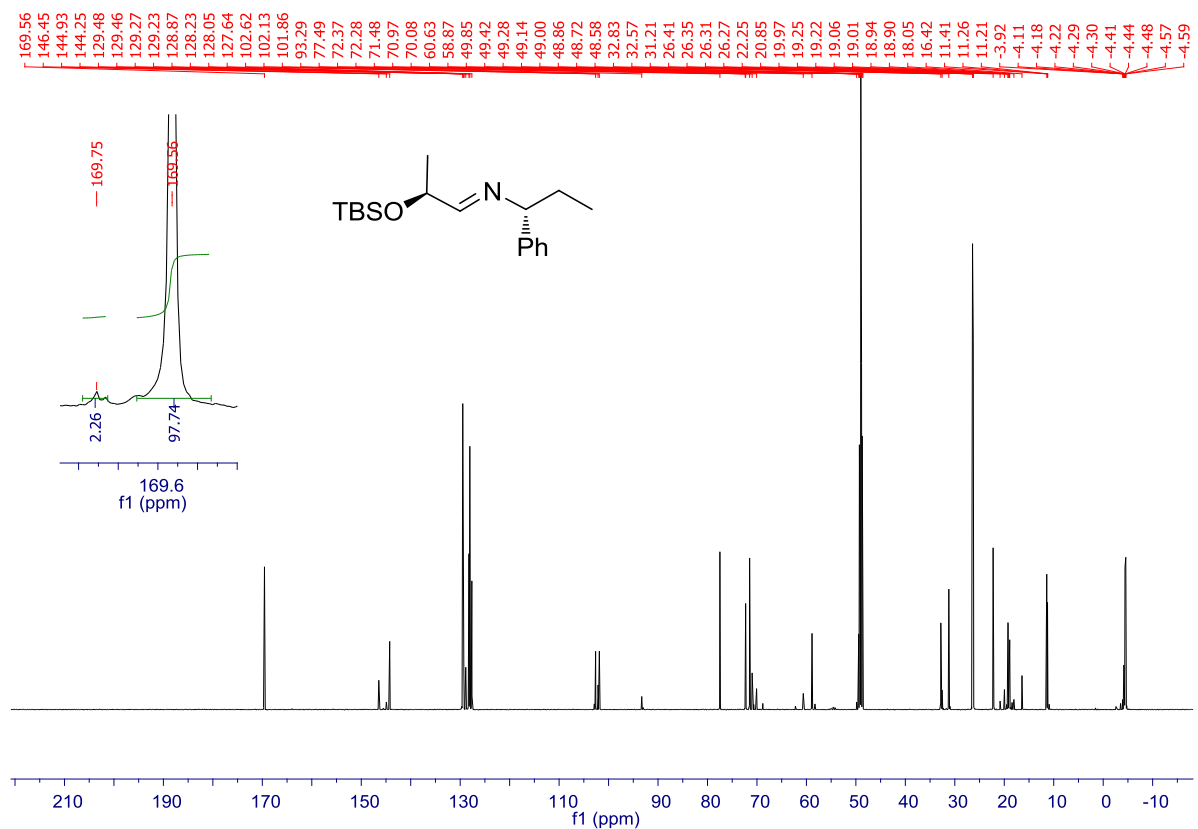
(S)- 1-Methoxy-2-propylamine-(15) and (R)-4:(S)-4 3:97



(R)-(+)- α -Ethylbenzylamine-(16) and (R)-4, d_4 -MeOH



(R)-(+)- α -Ethylbenzylamine-(16) and (S)-4, d_4 -MeOH



Chemical structure of compound 10 is shown above the spectra. The structure is a bicyclic system with an imine group and an OTBS group.

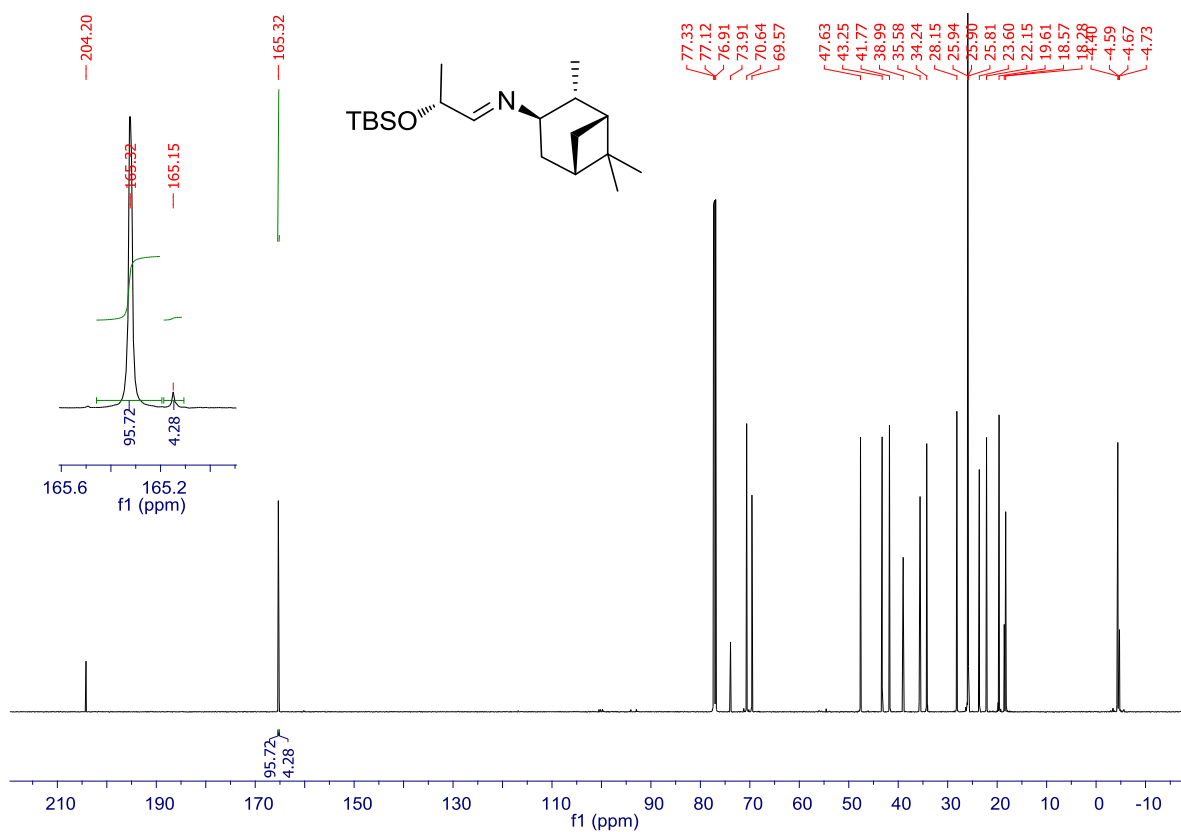
¹H NMR spectrum (bottom) is recorded in CDCl₃. The x-axis is labeled f1 (ppm) and ranges from 0 to 10. The spectrum shows several peaks, with the following chemical shifts (ppm) labeled in red:

- 72.54, 72.45, 71.53, 71.13, 67.66
- 49.43, 49.29, 49.15, 49.01, 48.87, 45.12, 43.30, 34.46, 28.70, 27.29, 26.58, 26.50, 24.18, 21.39, 4.03, 4.12, 4.13, 4.16, 4.31, 4.41
- 171.05, 170.94

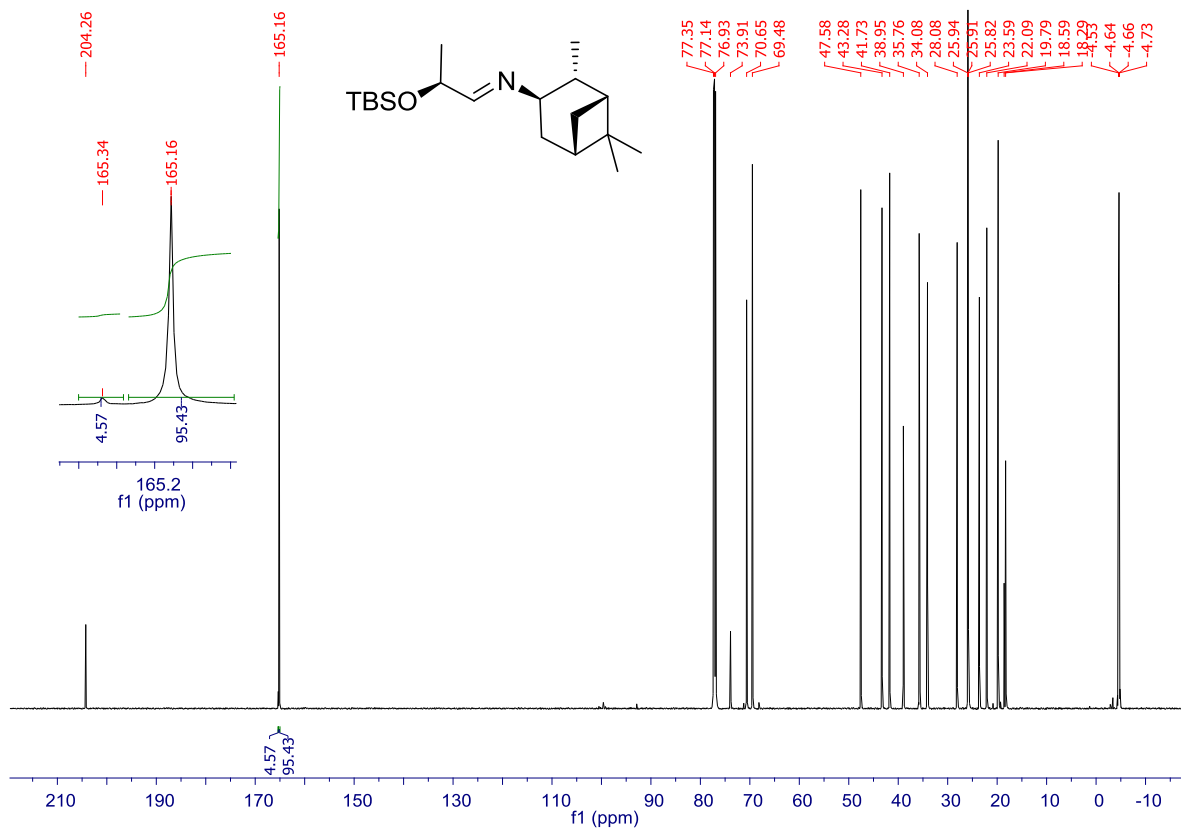
¹³C NMR spectrum (top) is recorded in CDCl₃. The x-axis is labeled f1 (ppm) and ranges from 0 to 210. The spectrum shows several peaks, with the following chemical shifts (ppm) labeled in red:

- 102.79, 102.29, 102.03
- 49.43, 49.29, 49.15, 49.01, 48.87, 45.12, 43.30, 34.46, 28.70, 27.29, 26.58, 26.50, 24.18, 21.39, 4.03, 4.12, 4.13, 4.16, 4.31, 4.41
- 171.05, 170.94

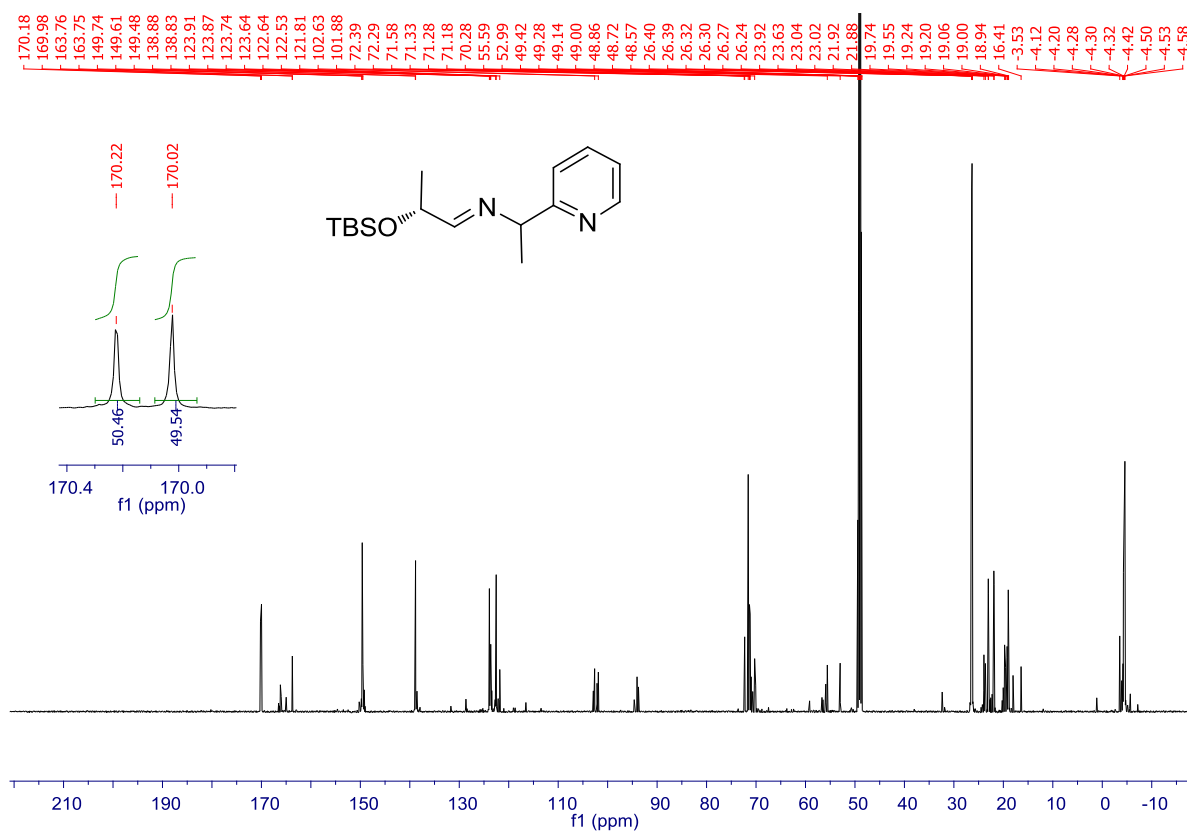
(1*R*,2*R*,3*R*,5*S*)-(-)-Isopinocampheylamine-(18) and (*R*)-4, CDCl₃



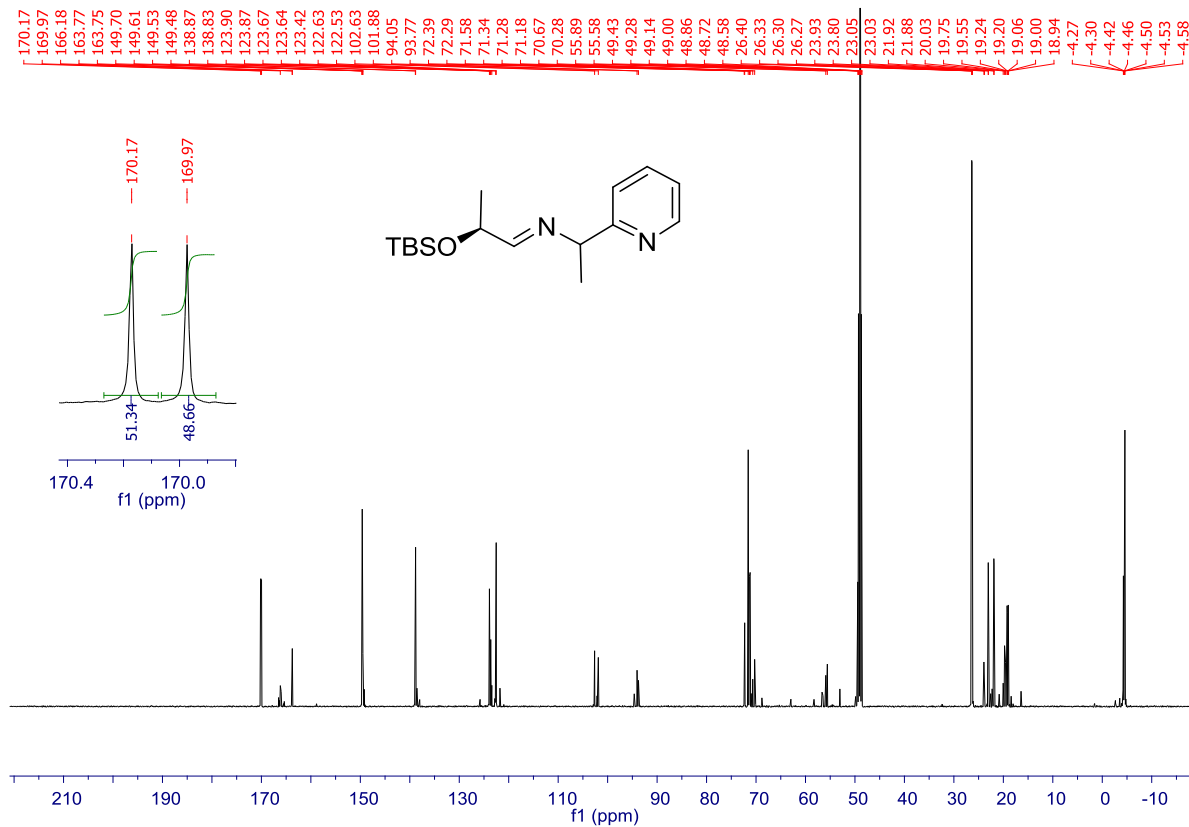
(1*R*,2*R*,3*R*,5*S*)-(-)-Isopinocampheylamine-(18) and (*S*)-4, CDCl₃



(±)-2-(1-Aminoethyl)pyridine-(19) and (R)-4, d₄-MeOH



(±)-2-(1-Aminoethyl)pyridine-(19) and (S)-4, d₄-MeOH



[illegible]

Chemical structure: CC(C)(C)S(=O)(=O)C=C[C@H](C)C(=O)C1(C)C(C)C(C)C1

¹³C NMR spectrum (ppm):

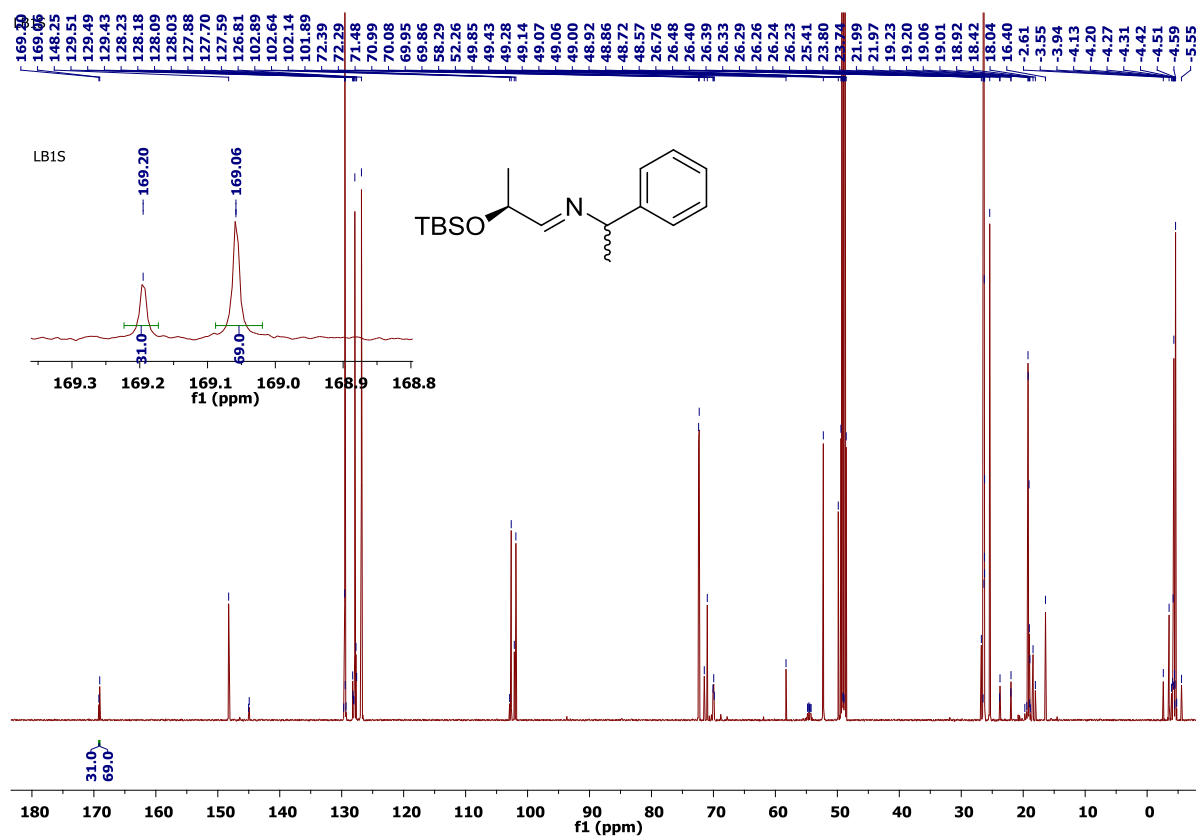
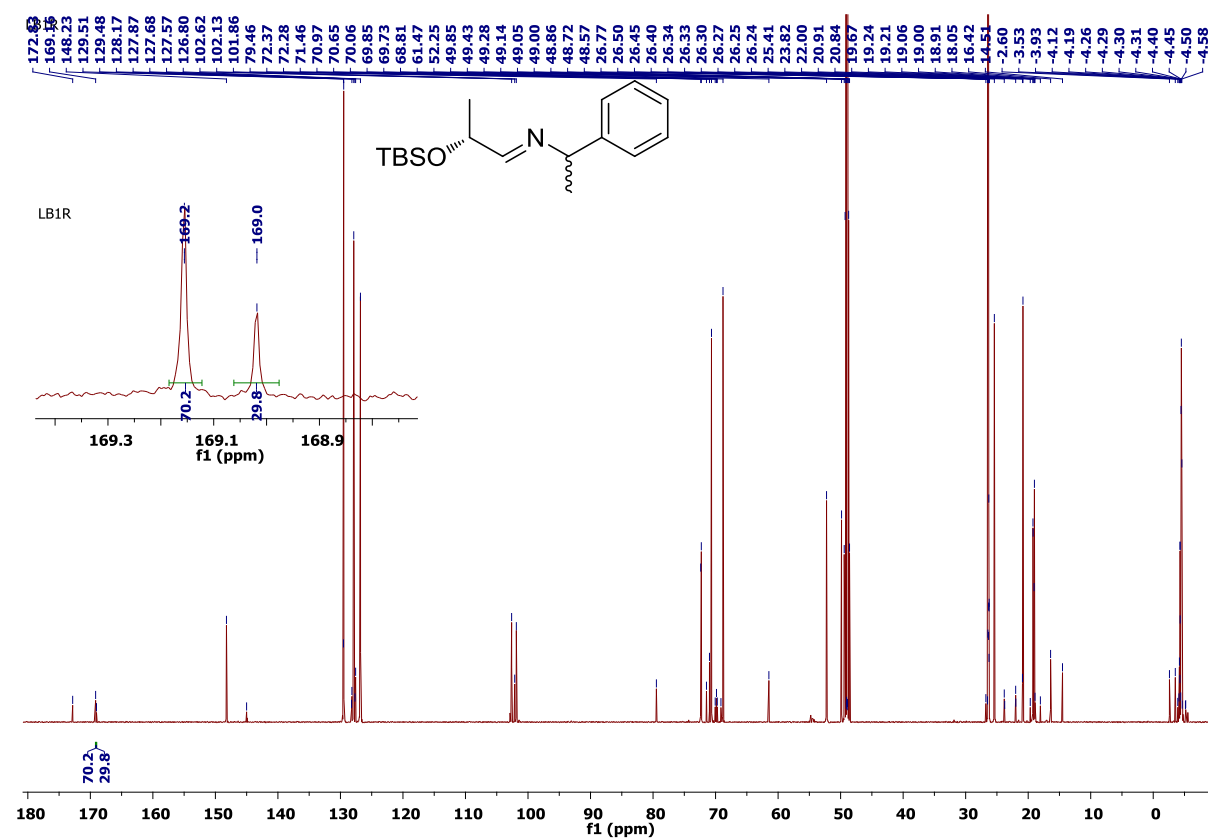
- 202.42
- 163.50
- 163.46
- 128.32
- 128.20
- 128.11
- 128.03
- 127.95
- 127.87
- 74.00
- 73.35
- 73.32
- 70.95
- 70.93
- 44.63
- 44.60
- 39.92
- 39.75
- 36.61
- 36.59
- 35.83
- 35.81
- 29.44
- 29.41
- 26.98
- 26.08
- 25.87
- 21.81
- 18.38
- 18.20
- 4.23
- 4.52
- 4.78
- 4.82

Inset: 163.50 f1 (ppm)

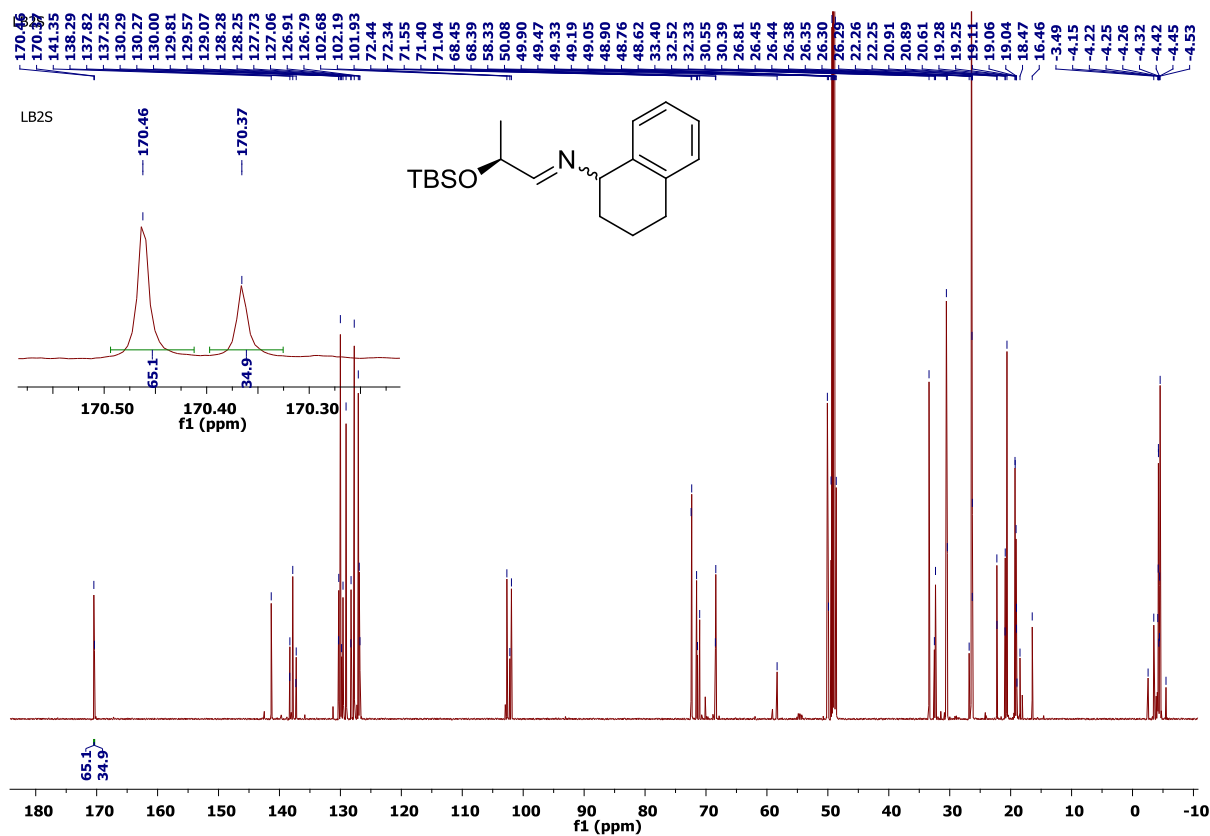
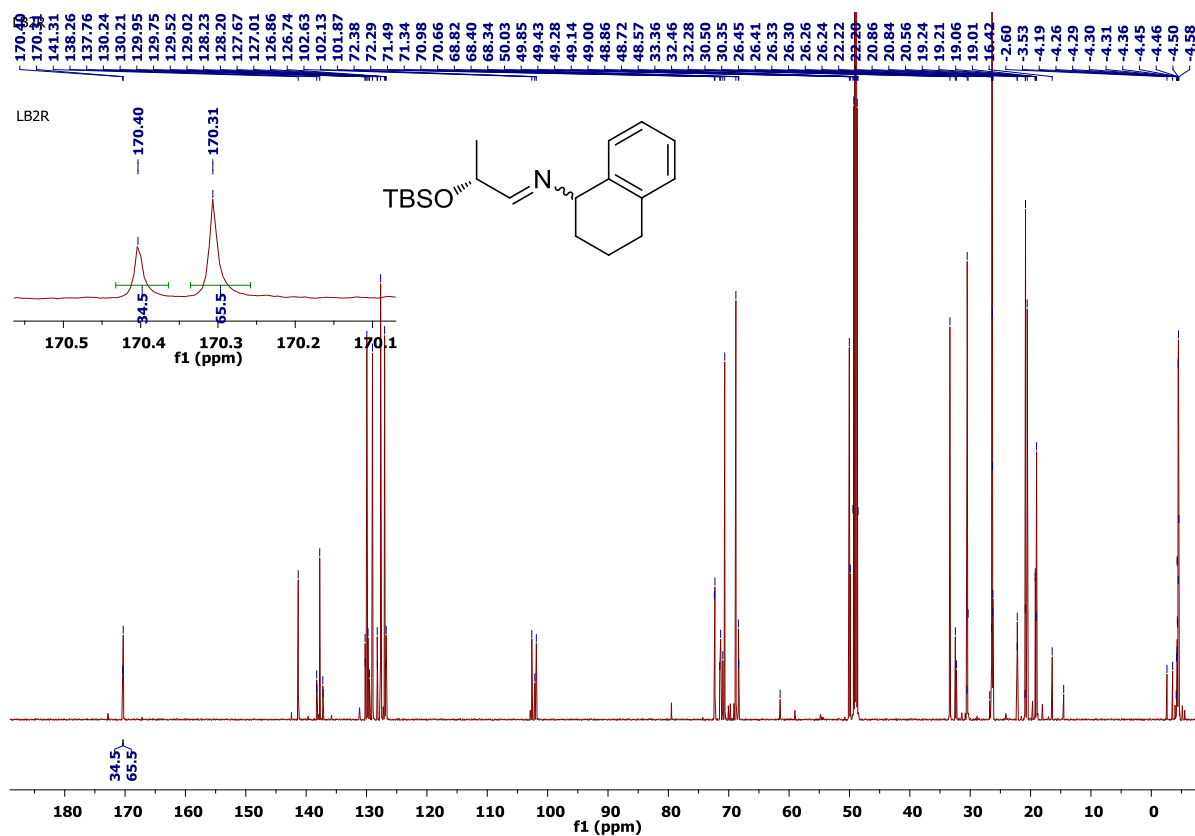
Analysis of ‘Unknown’ Samples (Table 3)

Sample	Prepared by	Actual er <i>R:S</i>	Researcher 1		Researcher 2	
			(S)-4	(R)-4	(S)-4	(R)-4
1	<i>RML</i>	69:31	<i>LB</i>		<i>SMG</i>	
			69:31	69:31	70:30	68:32
2	<i>RML</i>	34:66	<i>LB</i>		<i>SMG</i>	
			35:65	35:65	35:65	35:65
3	<i>RML</i>	77:23	<i>LB</i>		<i>SMG</i>	
			78:22	78:22	77:23	79:21
4	<i>LB</i>	6:94	<i>RML</i>		<i>SMG</i>	
			9:91	8:92	10:90	8:92
5	<i>LB</i>	86:14	<i>RML</i>		<i>SMG</i>	
			86:14	87:13	85:15	85:15
6	<i>LB</i>	24:76	<i>RML</i>		<i>SMG</i>	
			24:76	25:75	25:75	25:75
7	<i>SMG</i>	89:11	<i>RML</i>		<i>LB</i>	
			88:12	87:13	87:13	88:12
8	<i>SMG</i>	53:47	<i>RML</i>		<i>LB</i>	
			52:48	53:47	52:48	54:46
9	<i>SMG</i>	16:84	<i>RML</i>		<i>LB</i>	
			18:82	17:83	17:83	17:83

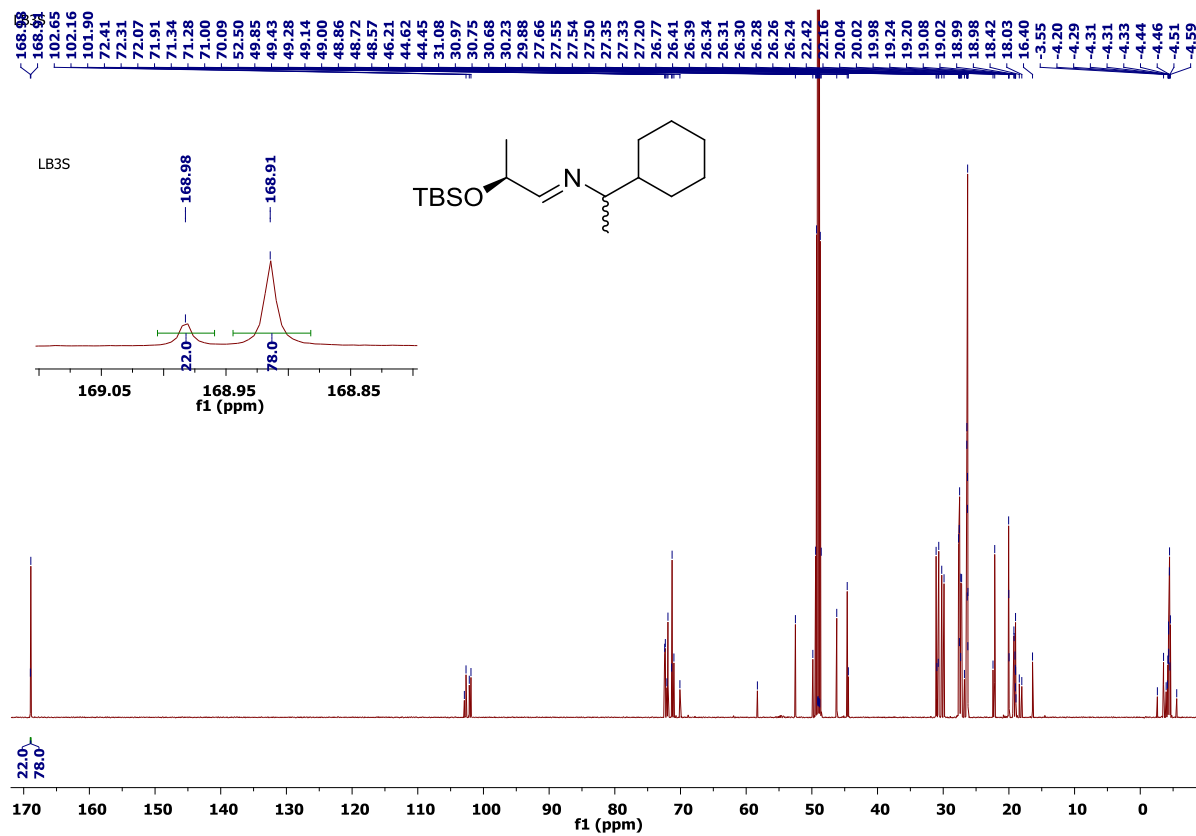
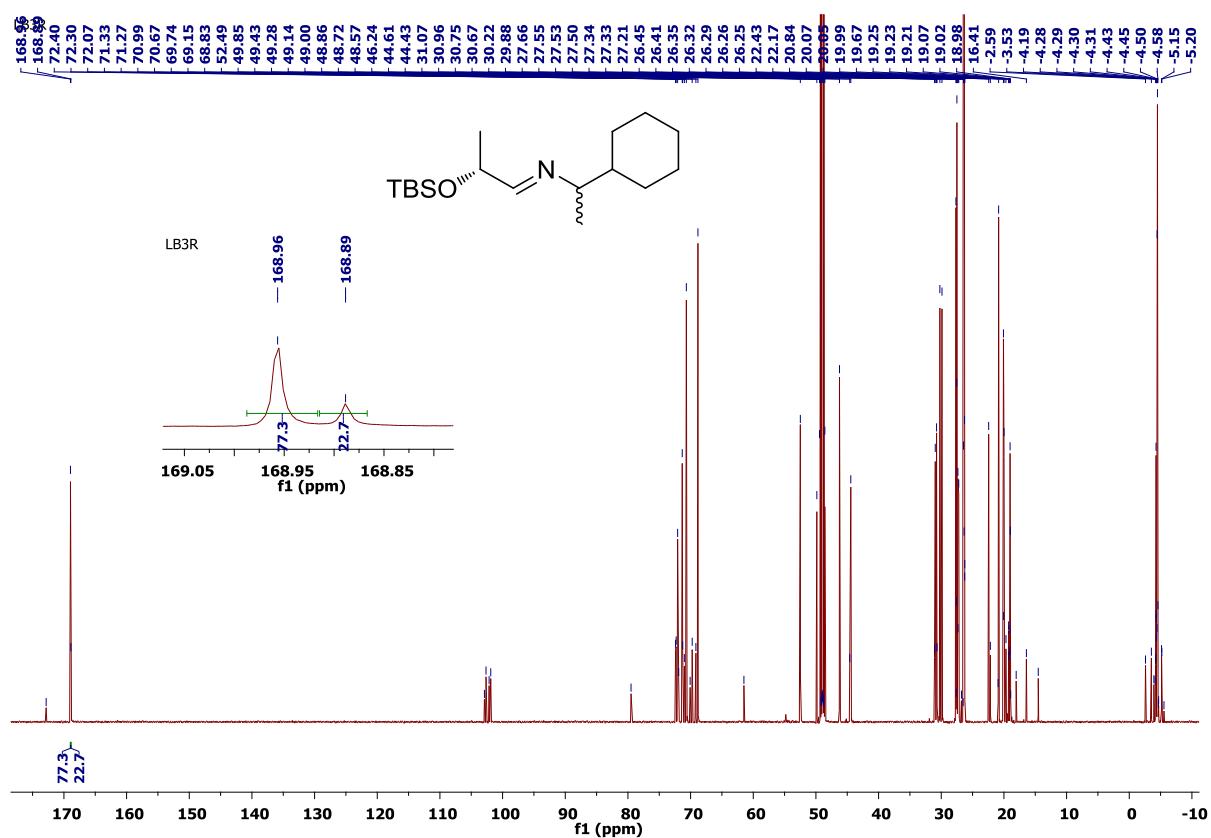
Sample 1 (LB)



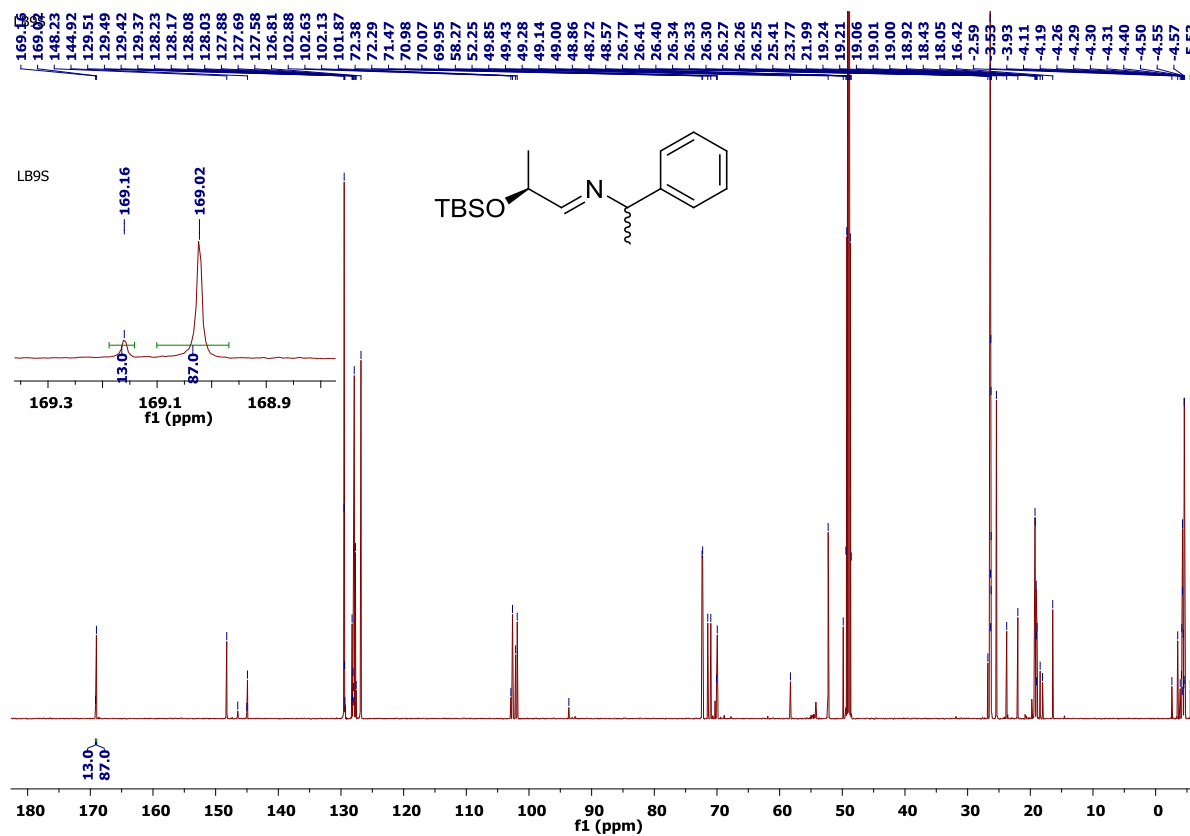
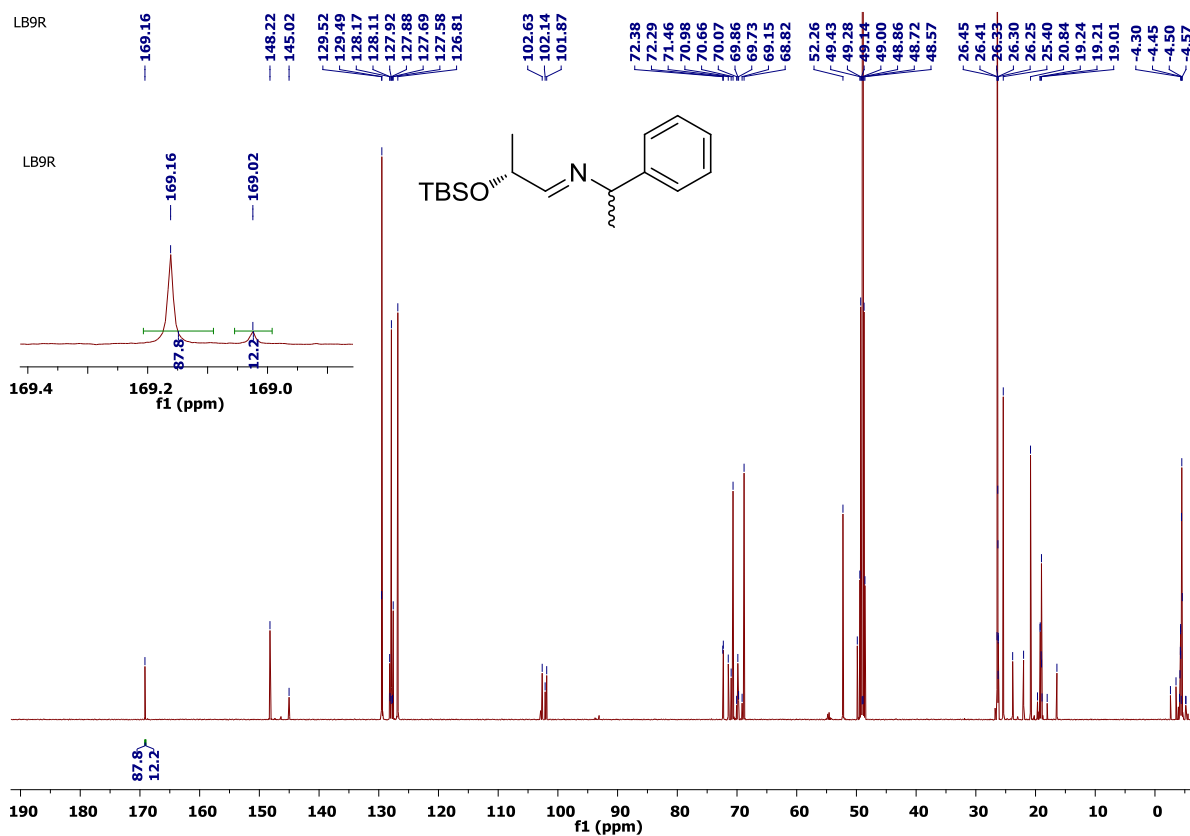
Sample 2 (LB)



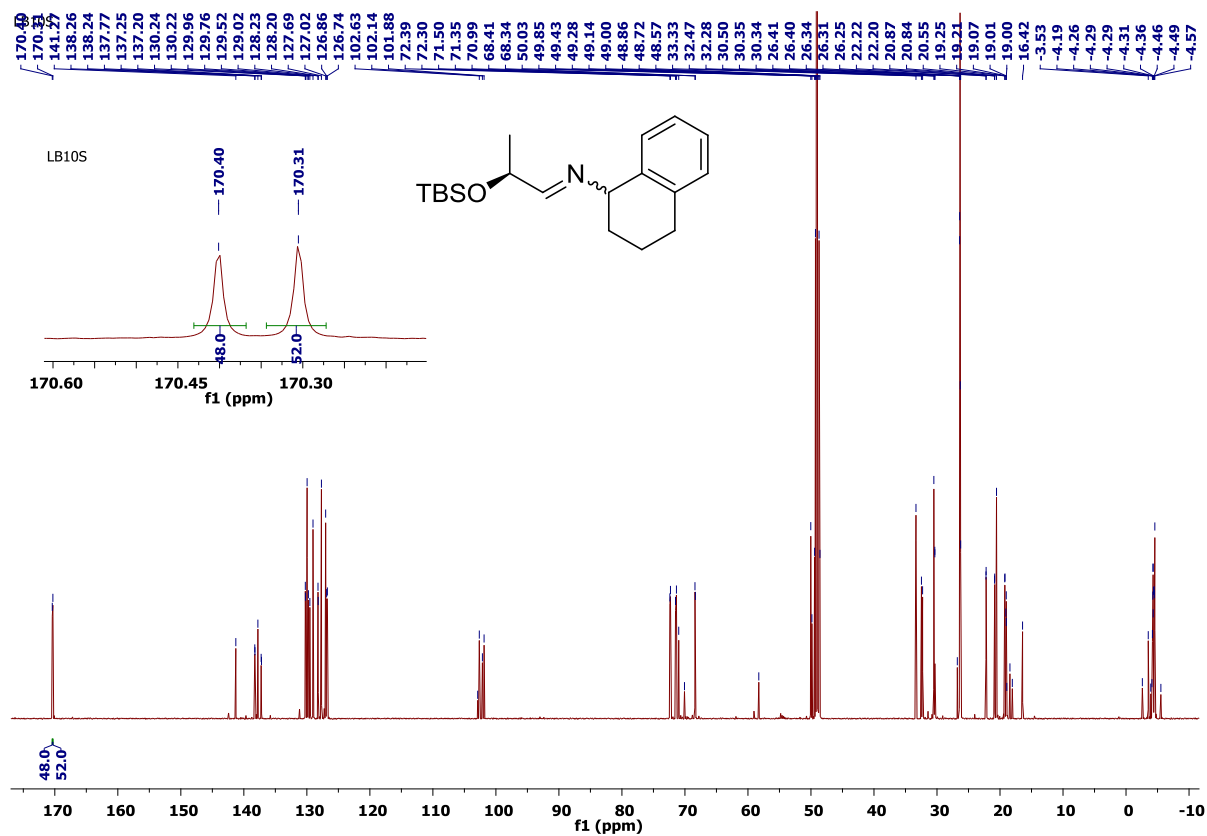
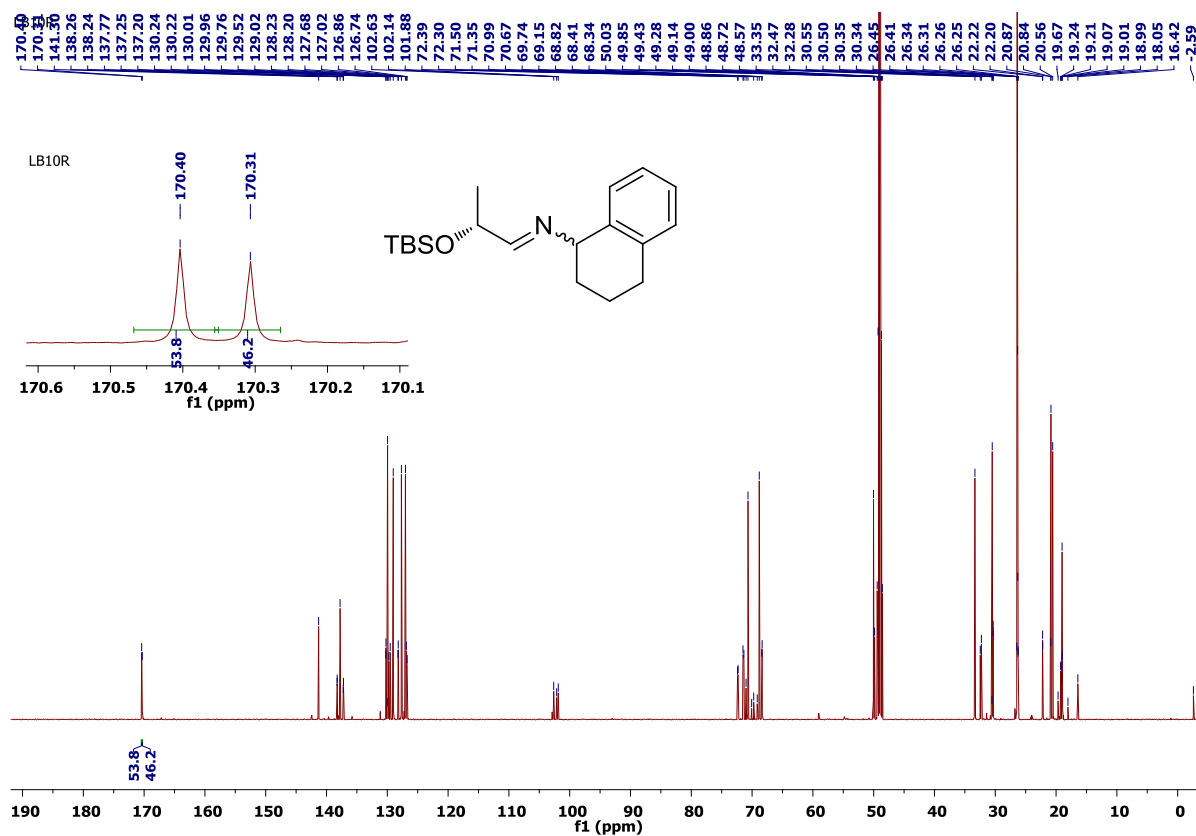
Sample 3 (LB)



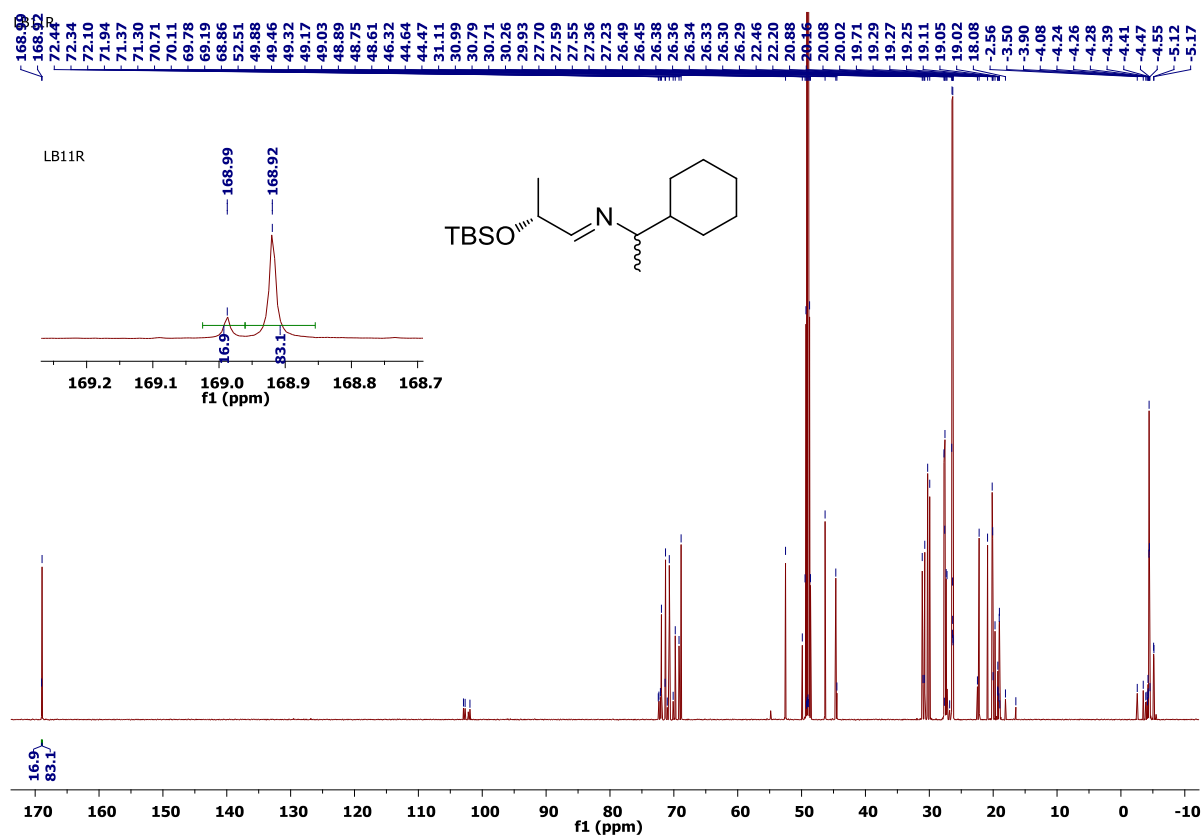
Sample 7 (LB)



Sample 8 (LB)



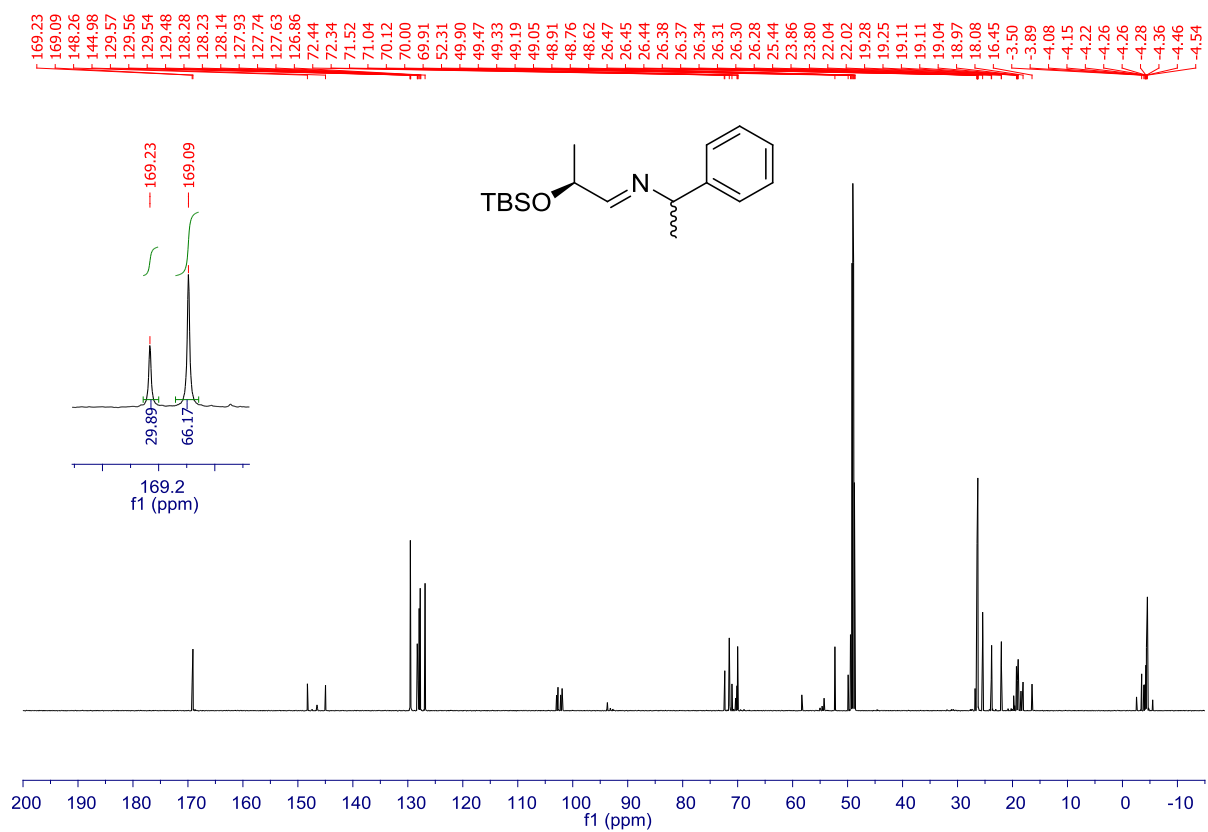
Sample 9 (LB)



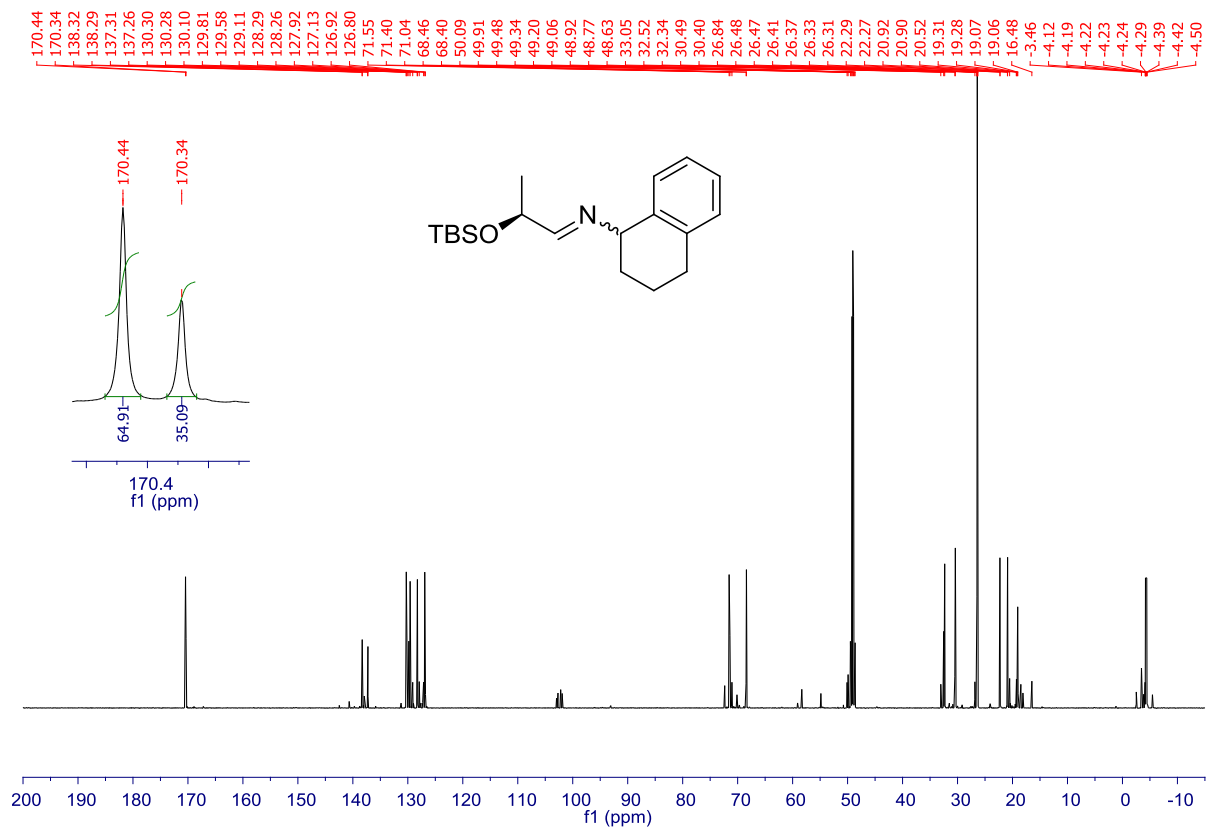
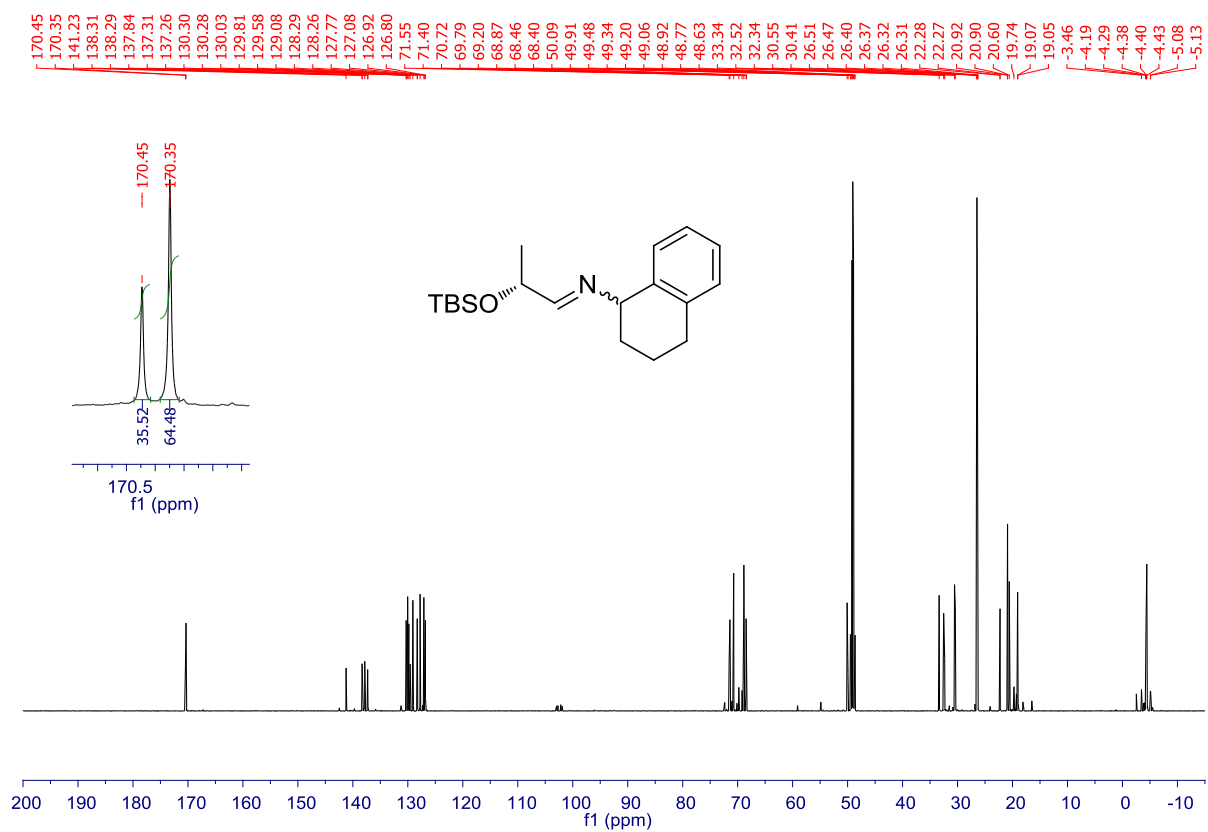
Chemical structure: CC(C)(C)Si(C)(C)OC1=CC=C(C=C1)C

¹³C NMR spectrum (ppm):

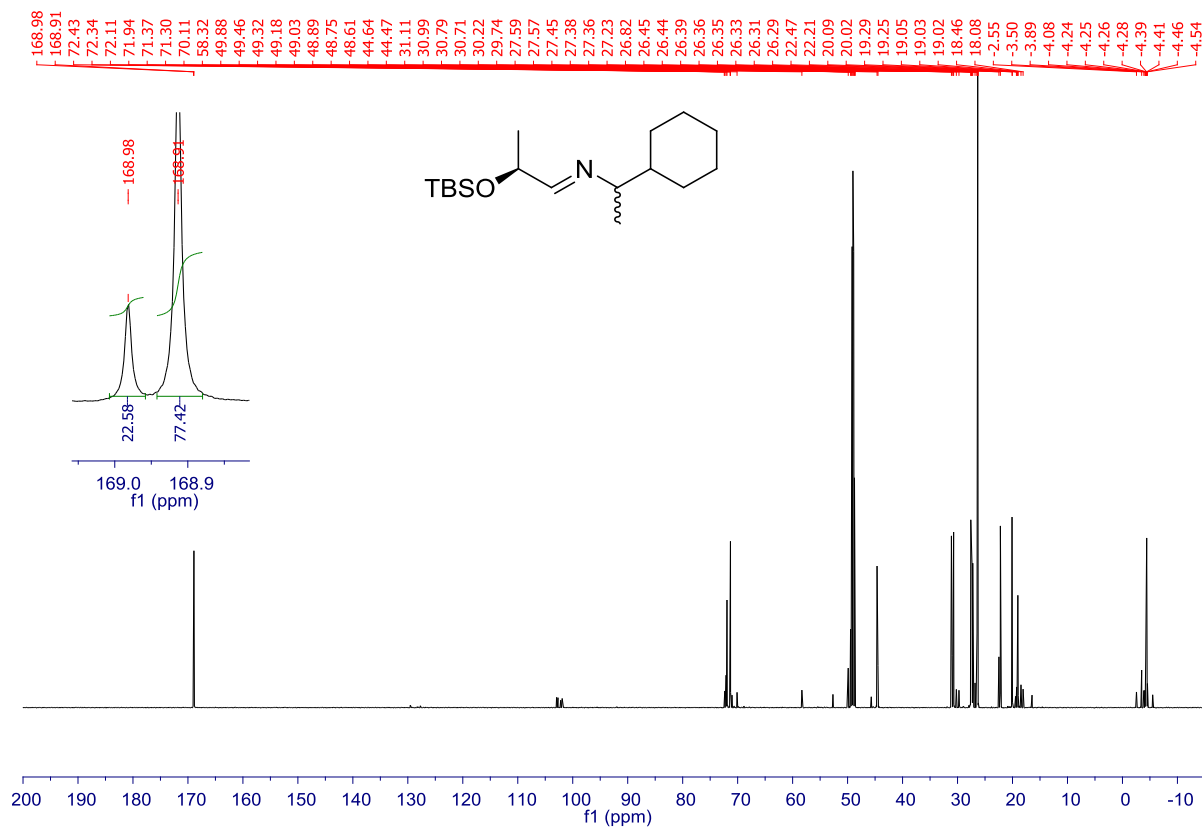
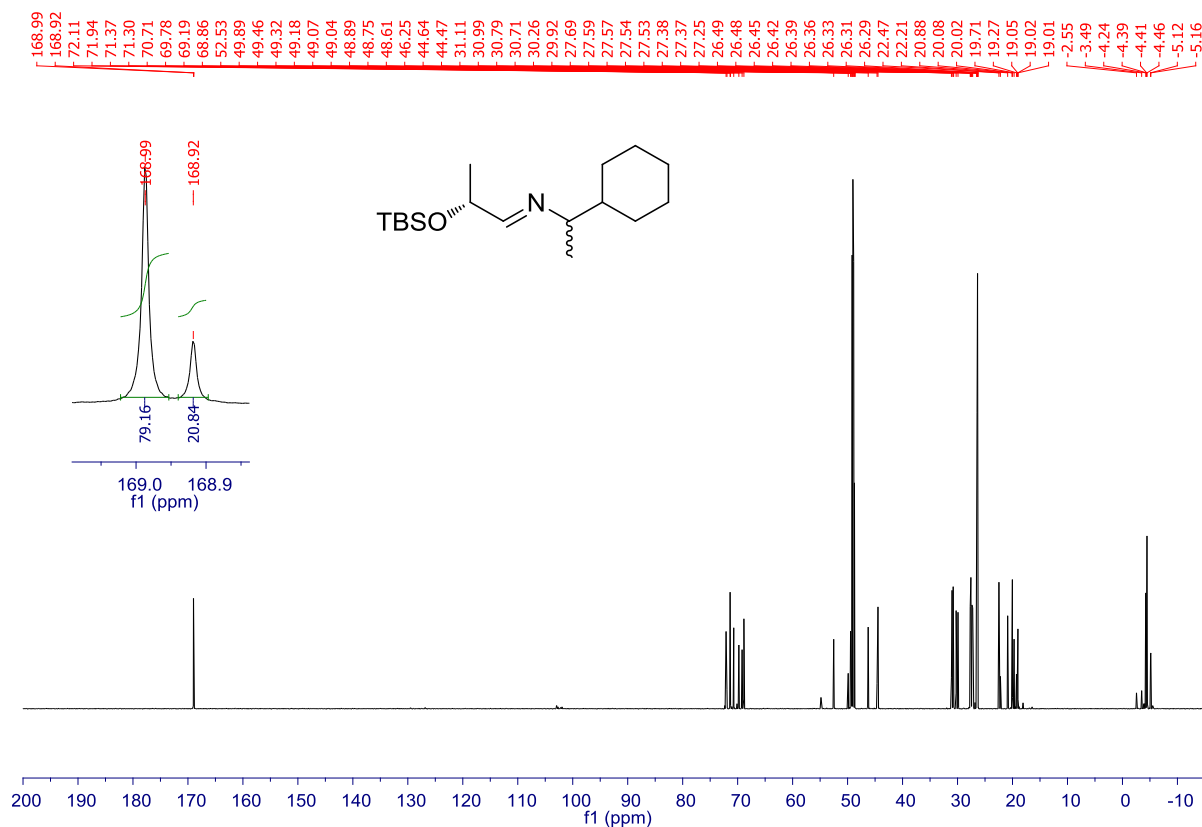
- 169.22
- 169.09
- 169.2
- 169.09
- 168.27
- 148.27
- 145.08
- 129.58
- 129.56
- 129.54
- 129.52
- 128.28
- 128.23
- 127.94
- 127.74
- 127.63
- 126.86
- 72.44
- 72.34
- 71.52
- 70.72
- 70.00
- 69.92
- 69.79
- 69.20
- 68.87
- 52.31
- 49.91
- 49.48
- 49.34
- 49.20
- 49.05
- 48.91
- 48.77
- 48.63
- 26.50
- 26.46
- 26.38
- 26.35
- 26.32
- 26.31
- 26.30
- 25.45
- 23.88
- 23.81
- 22.05
- 22.04
- 20.89
- 19.72
- 19.29
- 19.28
- 19.26
- 19.12
- 19.06
- 19.05
- 18.97
- 3.48
- 4.21
- 4.24
- 4.25
- 4.26
- 4.35
- 4.40
- 4.45
- 4.52
- 5.10
- 5.15



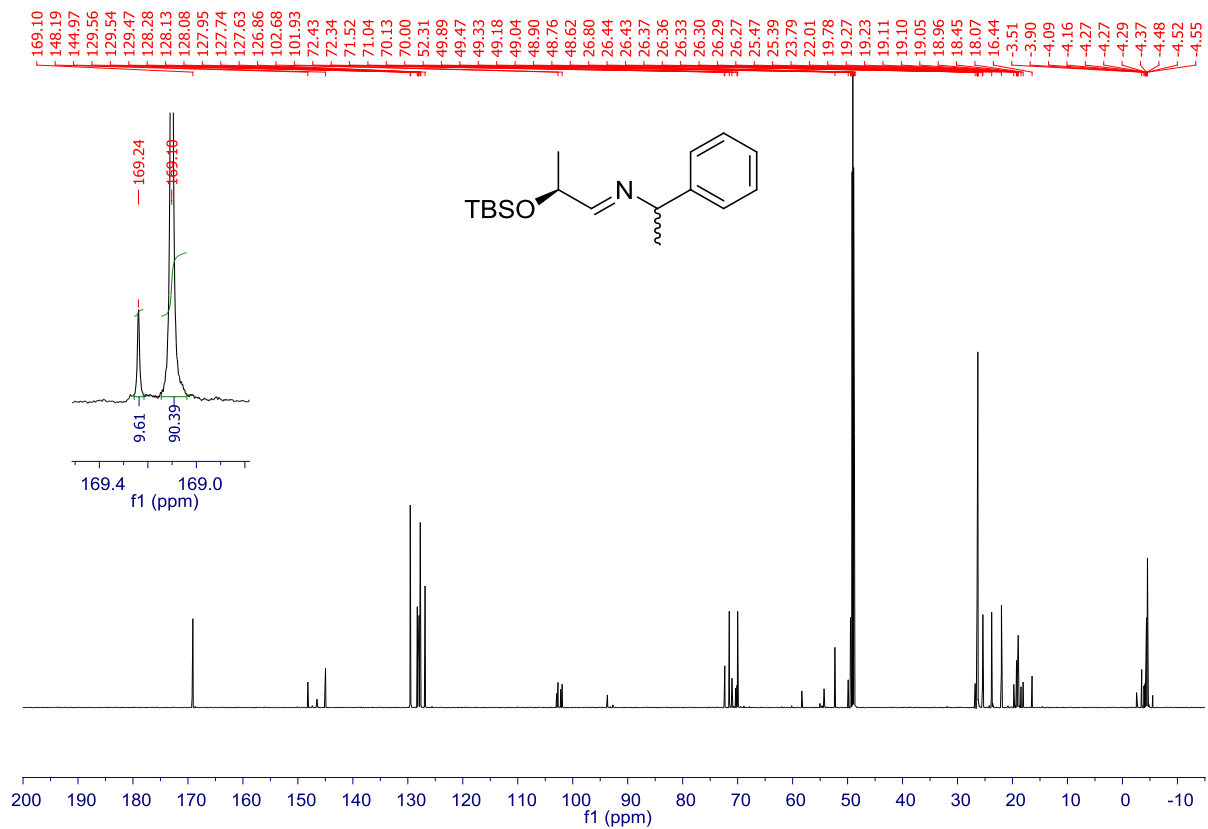
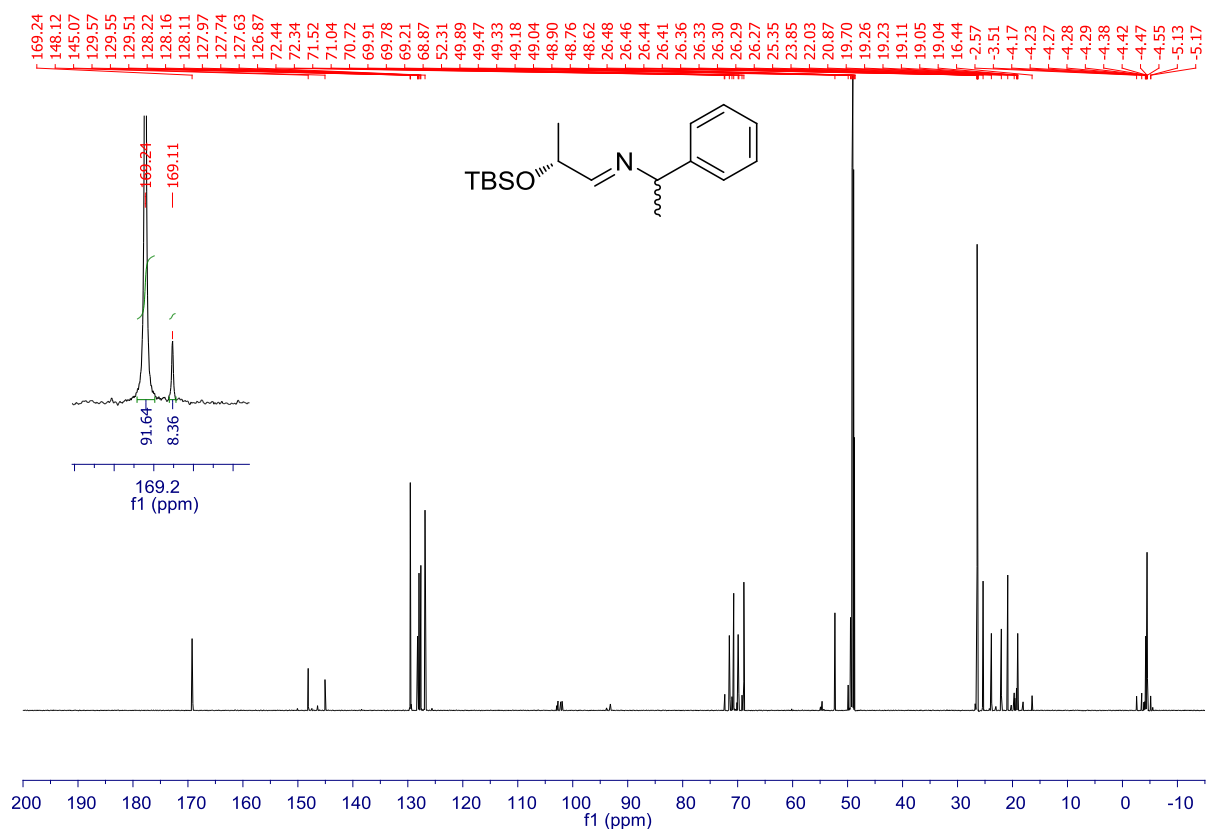
Sample 2 (SMG)



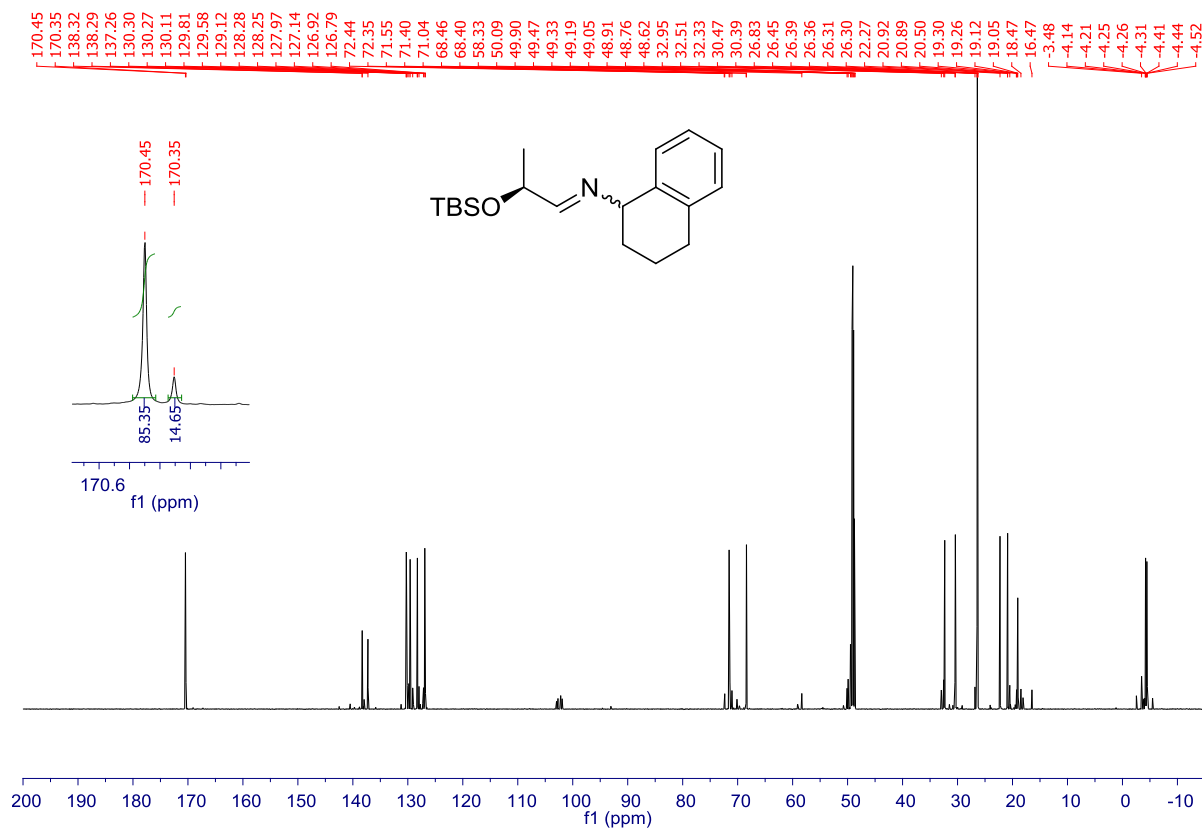
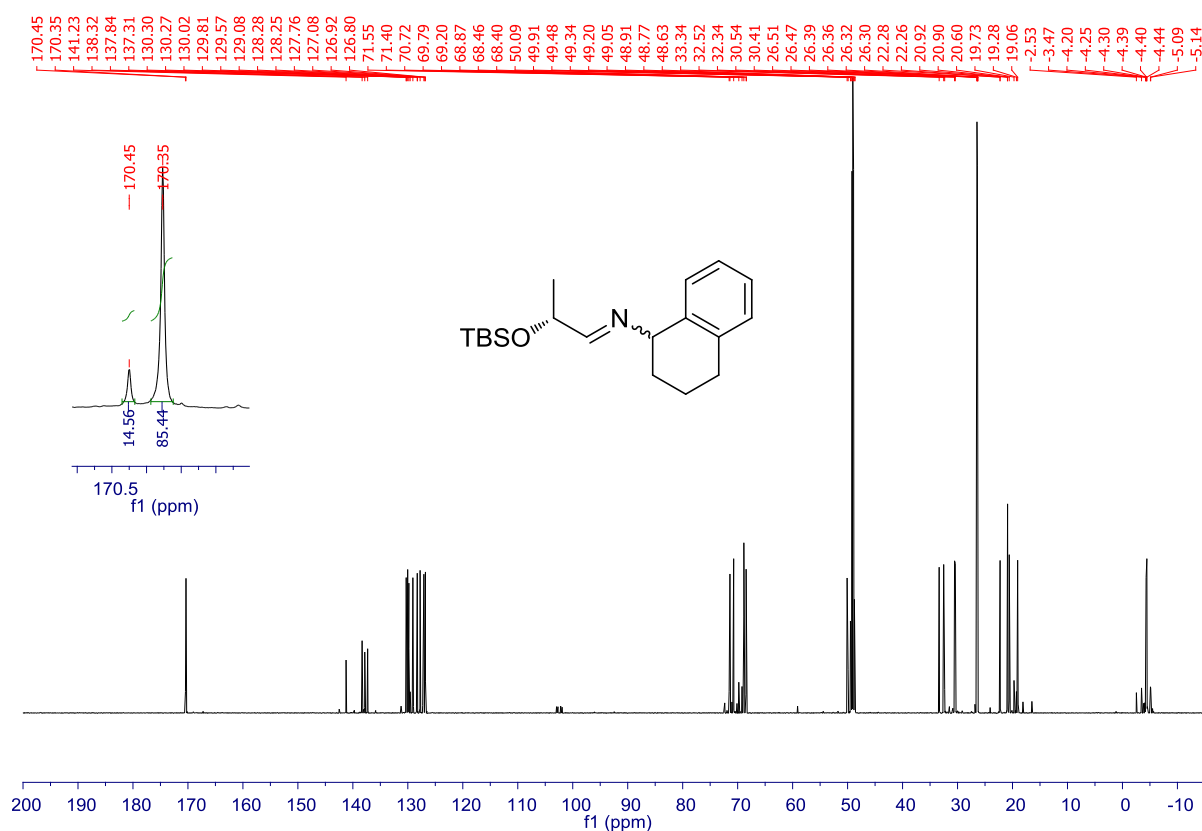
Sample 3 (SMG)



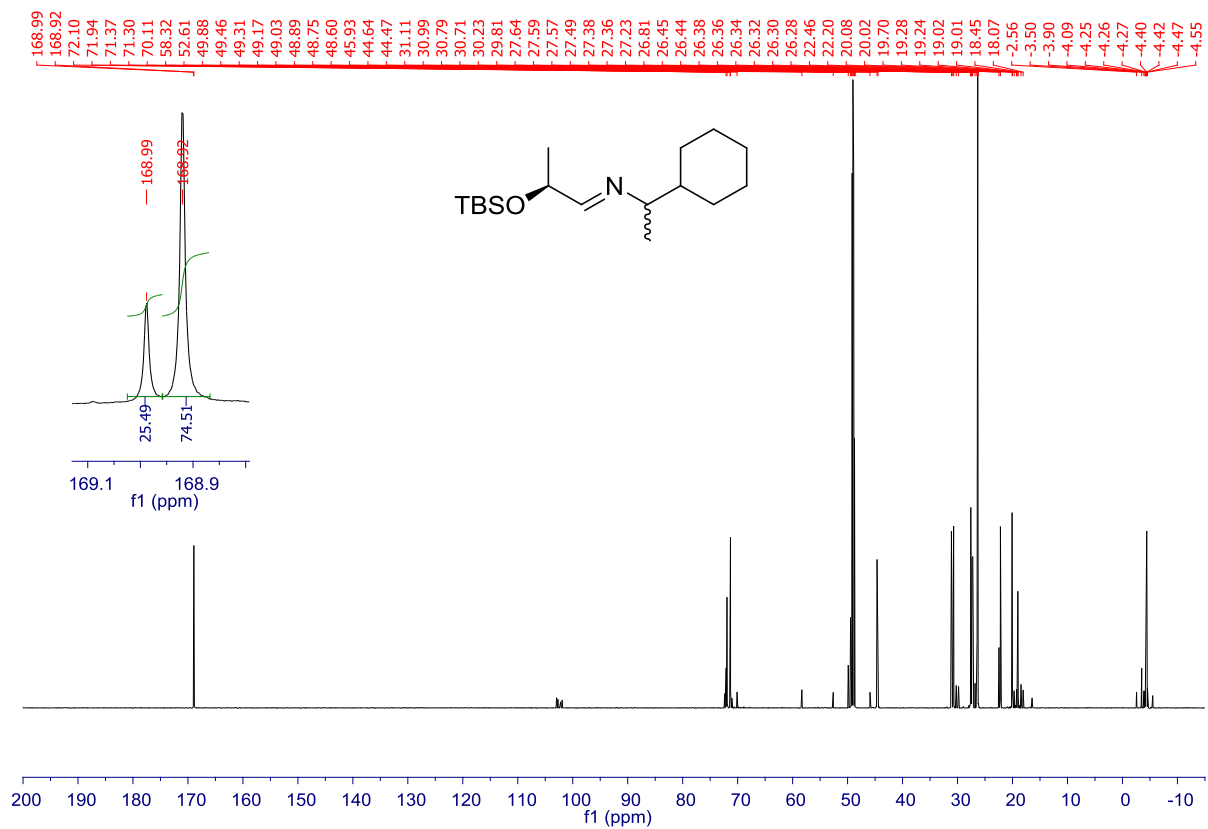
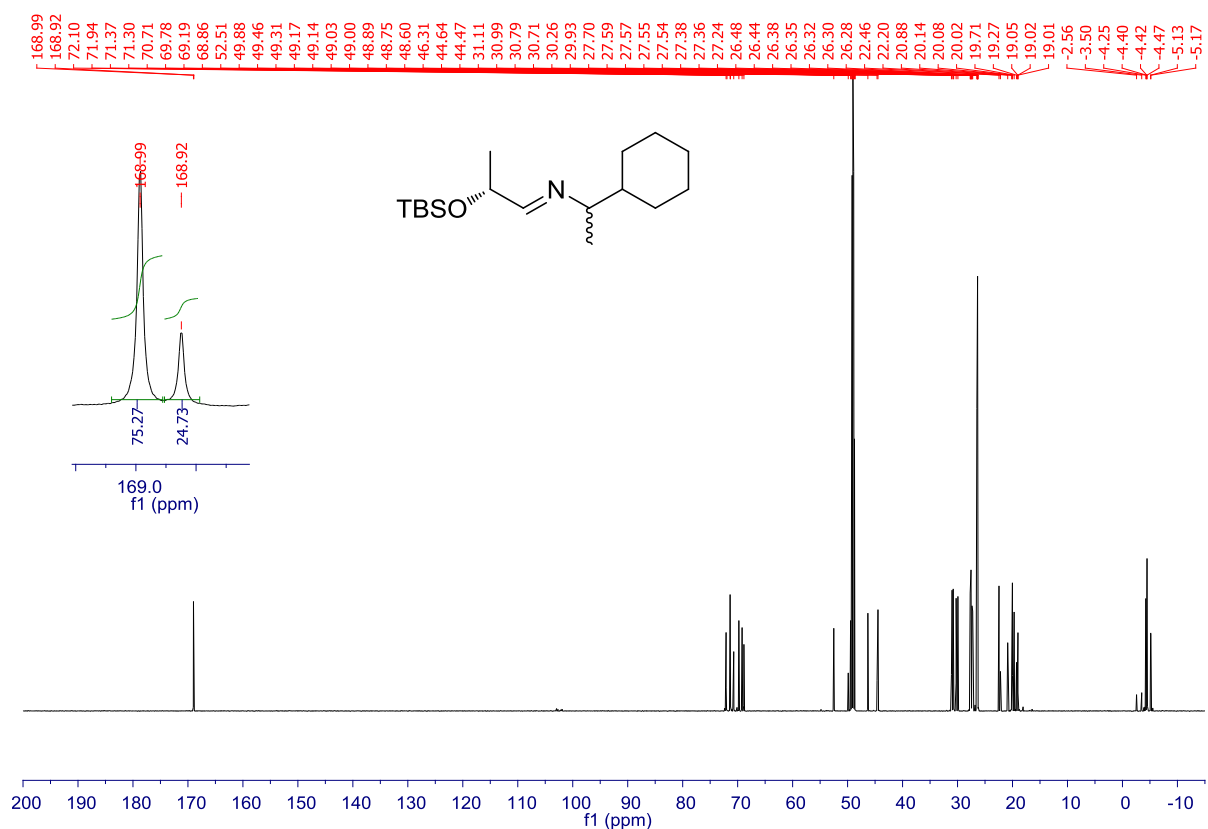
Sample 4 (SMG)



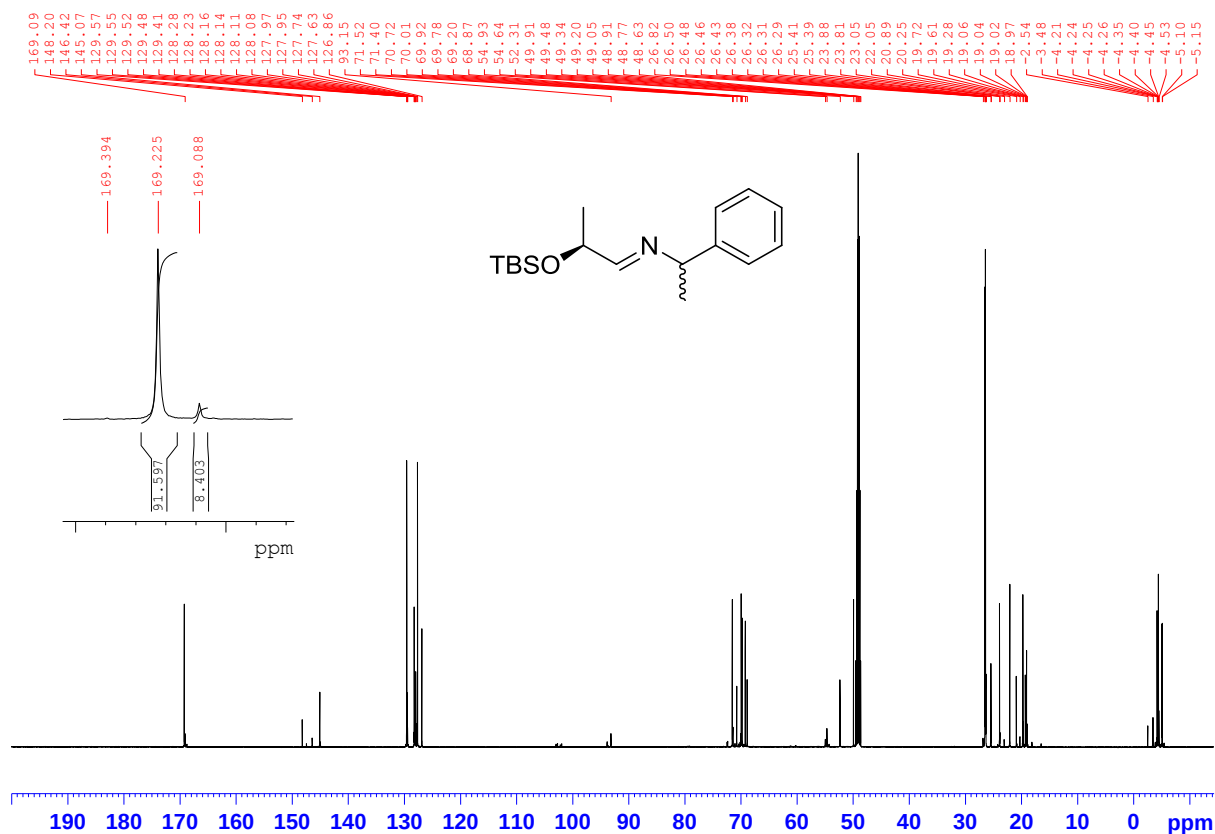
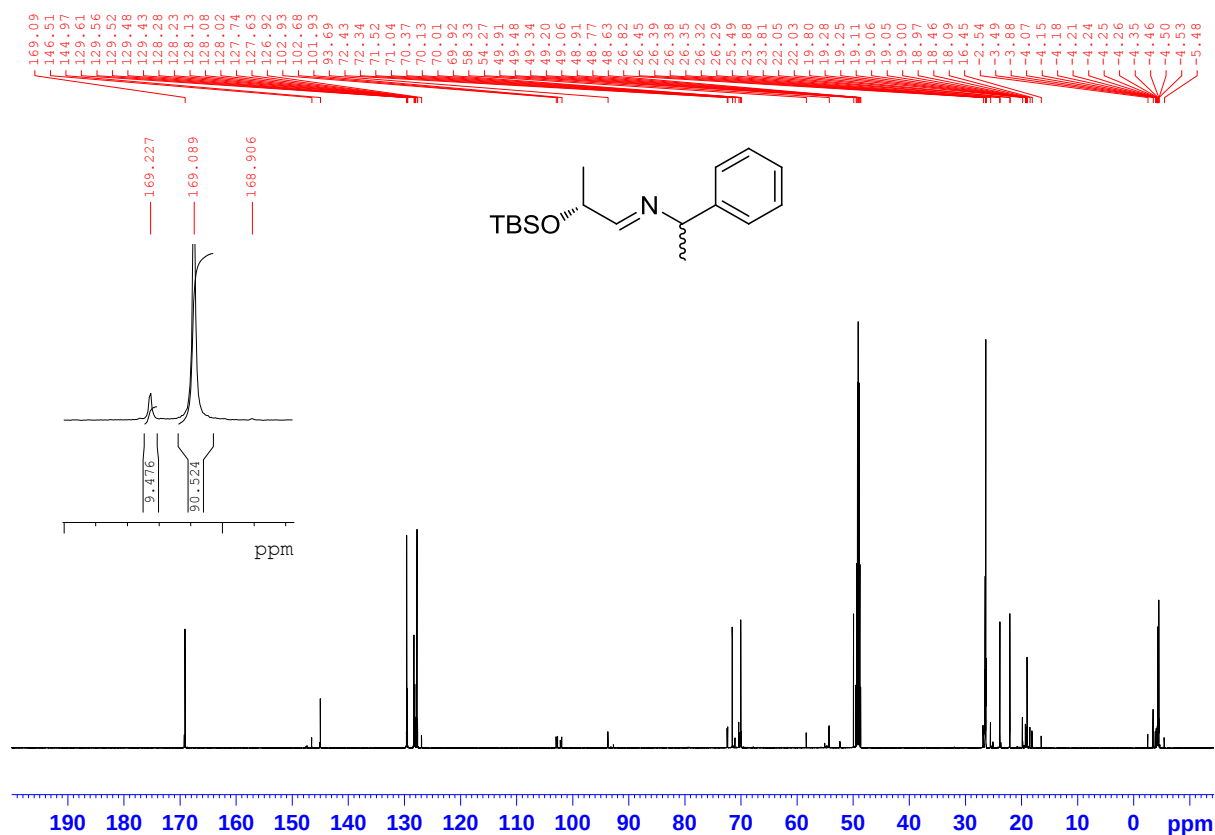
Sample 5 (SMG)



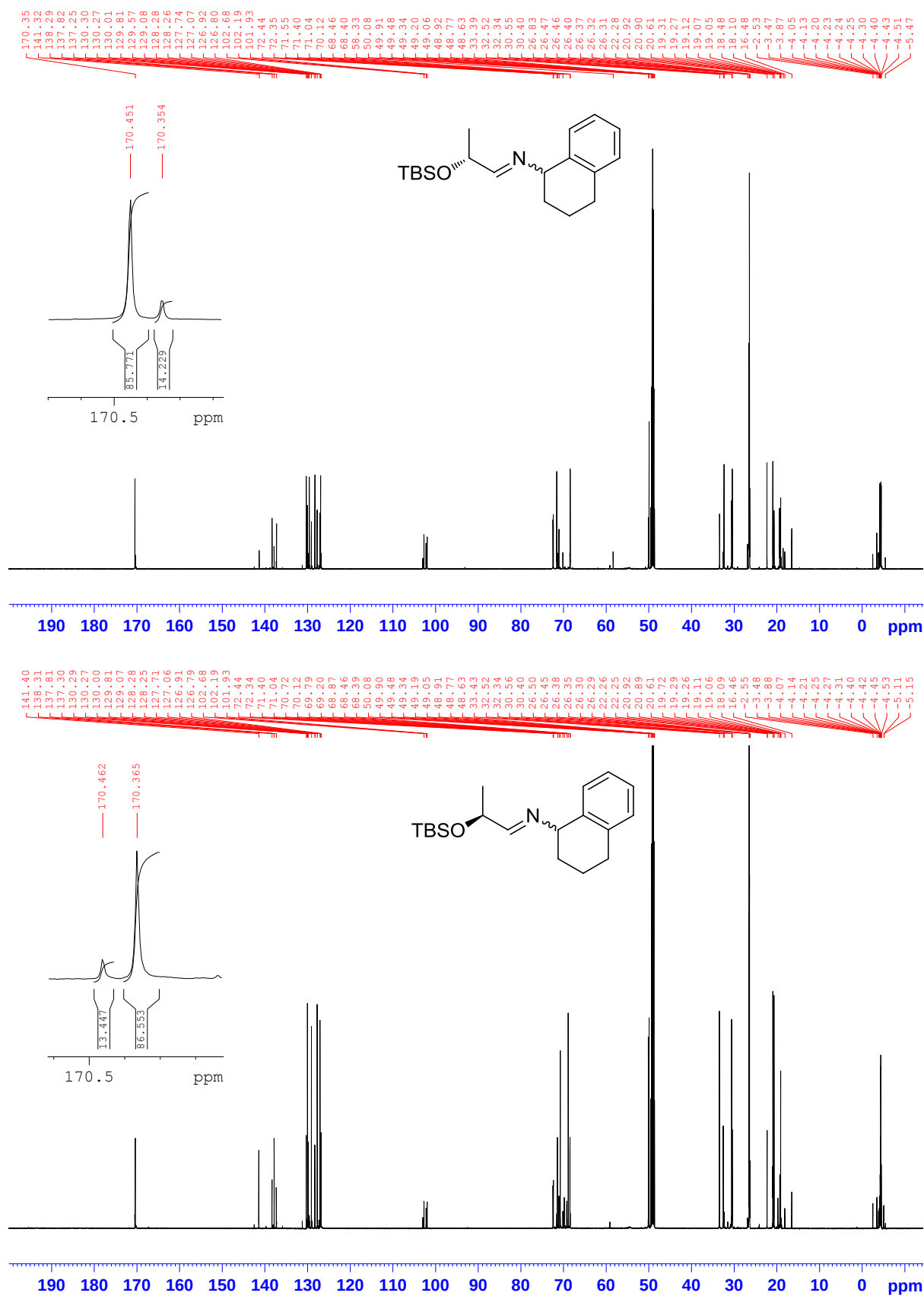
Sample 6 (SMG)



Sample 4 (RML)



Sample 5 (RML)

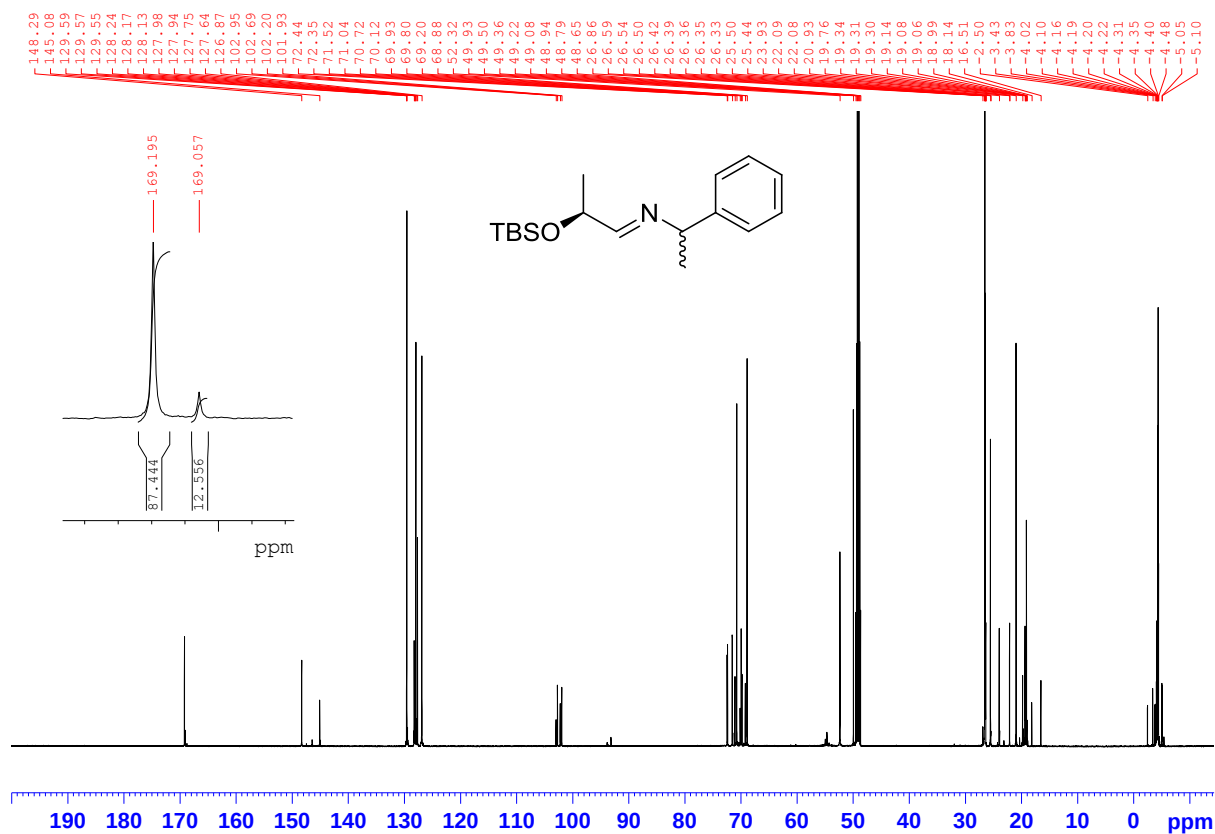
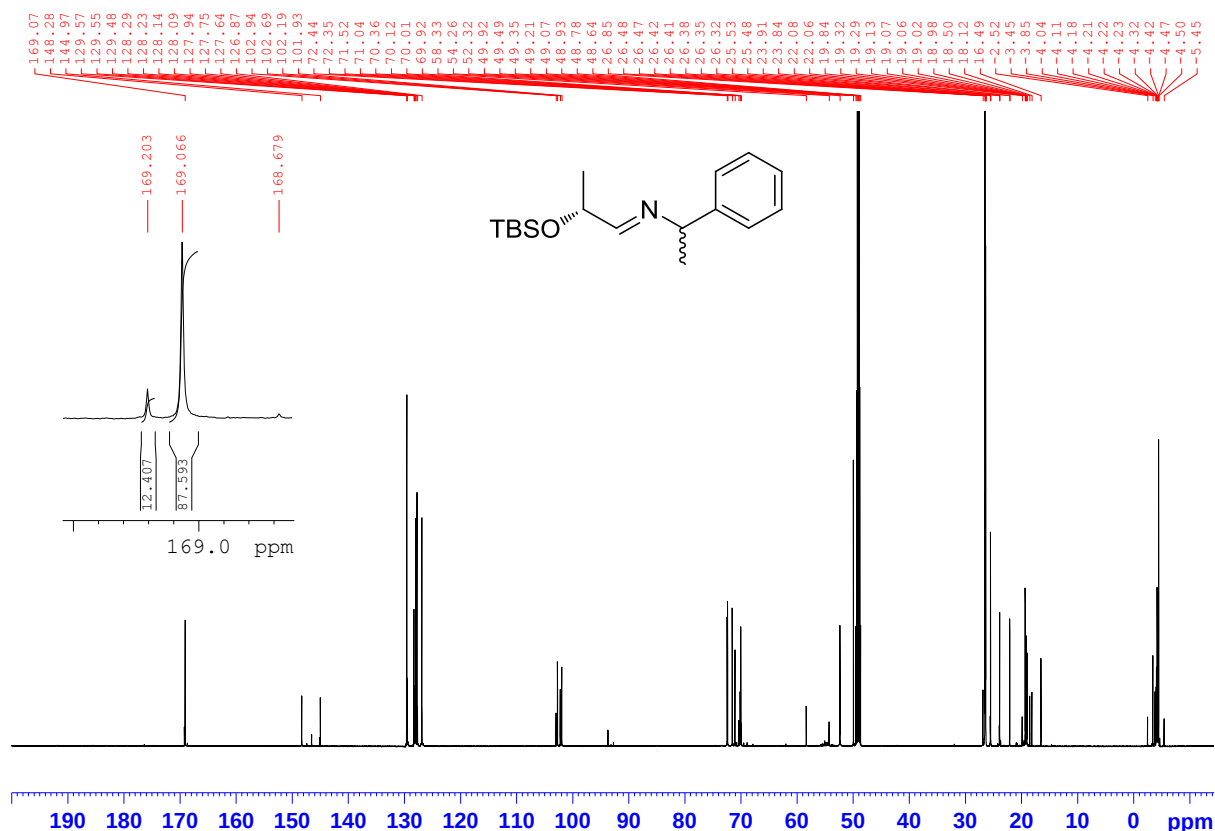


Top Spectrum: (S)-Enantiomer

Chemical structure: CC(C)[C@H](O)C(=N)C1CCCCC1 (with TBSO group on the chiral center)

Peak values (ppm): 168.94, 168.93, 168.92, 168.91, 168.90, 168.89, 168.88, 168.87, 168.86, 168.85, 168.84, 168.83, 168.82, 168.81, 168.80, 168.79, 168.78, 168.77, 168.76, 168.75, 168.74, 168.73, 168.72, 168.71, 168.70, 168.69, 168.68, 168.67, 168.66, 168.65, 168.64, 168.63, 168.62, 168.61, 168.60, 168.59, 168.58, 168.57, 168.56, 168.55, 168.54, 168.53, 168.52, 168.51, 168.50, 168.49, 168.48, 168.47, 168.46, 168.45, 168.44, 168.43, 168.42, 168.41, 168.40, 168.39, 168.38, 168.37, 168.36, 168.35, 168.34, 168.33, 168.32, 168.31, 168.30, 168.29, 168.28, 168.27, 168.26, 168.25, 168.24, 168.23, 168.22, 168.21, 168.20, 168.19, 168.18, 168.17, 168.16, 168.15, 168.14, 168.13, 168.12, 168.11, 168.10, 168.09, 168.08, 168.07, 168.06, 168.05, 168.04, 168.03, 168.02, 168.01, 168.00, 167.99, 167.98, 167.97, 167.96, 167.95, 167.94, 167.93, 167.92, 167.91, 167.90, 167.89, 167.88, 167.87, 167.86, 167.85, 167.84, 167.83, 167.82, 167.81, 167.80, 167.79, 167.78, 167.77, 167.76, 167.75, 167.74, 167.73, 167.72, 167.71, 167.70, 167.69, 167.68, 167.67, 167.66, 167.65, 167.64, 167.63, 167.62, 167.61, 167.60, 167.59, 167.58, 167.57, 167.56, 167.55, 167.54, 167.53, 167.52, 167.51, 167.50, 167.49, 167.48, 167.47, 167.46, 167.45, 167.44, 167.43, 167.42, 167.41, 167.40, 167.39, 167.38, 167.37, 167.36, 167.35, 167.34, 167.33, 167.32, 167.31, 167.30, 167.29, 167.28, 167.27, 167.26, 167.25, 167.24, 167.23, 167.22, 167.21, 167.20, 167.19, 167.18, 167.17, 167.16, 167.15, 167.14, 167.13, 167.12, 167.11, 167.10, 167.09, 167.08, 167.07, 167.06, 167.05, 167.04, 167.03, 167.02, 167.01, 167.00, 166.99, 166.98, 166.97, 166.96, 166.95, 166.94, 166.93, 166.92, 166.91, 166.90, 166.89, 166.88, 166.87, 166.86, 166.85, 166.84, 166.83, 166.82, 166.81, 166.80, 166.79, 166.78, 166.77, 166.76, 166.75, 166.74, 166.73, 166.72, 166.71, 166.70, 166.69, 166.68, 166.67, 166.66, 166.65, 166.64, 166.63, 166.62, 166.61, 166.60, 166.59, 166.58, 166.57, 166.56, 166.55, 166.54, 166.53, 166.52, 166.51, 166.50, 166.49, 166.48, 166.47, 166.46, 166.45, 166.44, 166.43, 166.42, 166.41, 166.40, 166.39, 166.38, 166.37, 166.36, 166.35, 166.34, 166.33, 166.32, 166.31, 166.30, 166.29, 166.28, 166.27, 166.26, 166.25, 166.24, 166.23, 166.22, 166.21, 166.20, 166.19, 166.18, 166.17, 166.16, 166.15, 166.14, 166.13, 166.12, 166.11, 166.10, 166.09, 166.08, 166.07, 166.06, 166.05, 166.04, 166.03, 166.02, 166.01, 166.00, 165.99, 165.98, 165.97, 165.96, 165.95, 165.94, 165.93, 165.92, 165.91, 165.90, 165.89, 165.88, 165.87, 165.86, 165.85, 165.84, 165.83, 165.82, 165.81, 165.80, 165.79, 165.78, 165.77, 165.76, 165.75, 165.74, 165.73, 165.72, 165.71, 165.70, 165.69, 165.68, 165.67, 165.66, 165.65, 165.64, 165.63, 165.62, 165.61, 165.60, 165.59, 165.58, 165.57, 165.56, 165.55, 165.54, 165.53, 165.52, 165.51, 165.50, 165.49, 165.48, 165.47, 165.46, 165.45, 165.44, 165.43, 165.42, 165.41, 165.40, 165.39, 165.38, 165.37, 165.36, 165.35, 165.34, 165.33, 165.32, 165.31, 165.30, 165.29, 165.28, 165.27, 165.26, 165.25, 165.24, 165.23, 165.22, 165.21, 165.20, 165.19, 165.18, 165.17, 165.16, 165.15, 165.14, 165.13, 165.12, 165.11, 165.10, 165.09, 165.08, 165.07, 165.06, 165.05, 165.04, 165.03, 165.02, 165.01, 165.00, 164.99, 164.98, 164.97, 164.96, 164.95, 164.94, 164.93, 164.92, 164.91, 164.90, 164.89, 164.88, 164.87, 164.86, 164.85, 164.84, 164.83, 164.82, 164.81, 164.80, 164.79, 164.78, 164.77, 164.76, 164.75, 164.74, 164.73, 164.72, 164.71, 164.70, 164.69, 164.68, 164.67, 164.66, 164.65, 164.64, 164.63, 164.62, 164.61, 164.60, 164.59, 164.58, 164.57, 164.56, 164.55, 164.54, 164.53, 164.52, 164.51, 164.50, 164.49, 164.48, 164.47, 164.46, 164.45, 164.44, 164.43, 164.42, 164.41, 164.40, 164.39, 164.38, 164.37, 164.36, 164.35, 164.34, 164.33, 164.32, 164.31, 164.30, 164.29, 164.28, 164.27, 164.26, 164.25, 164.24, 164.23, 164.22, 164.21, 164.20, 164.19, 164.18, 164.17, 164.16, 164.15, 164.14, 164.13, 164.12, 164.11, 164.10, 164.09, 164.08

Sample 7 (RML)



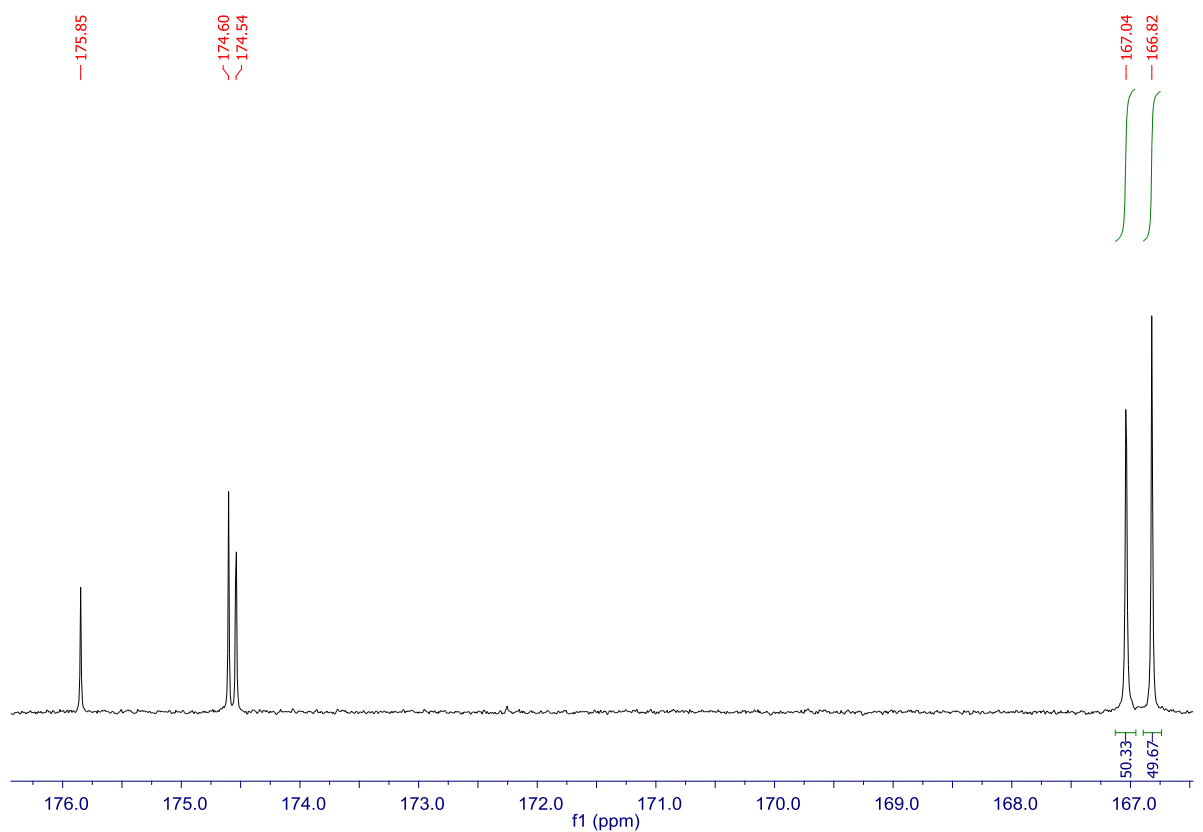
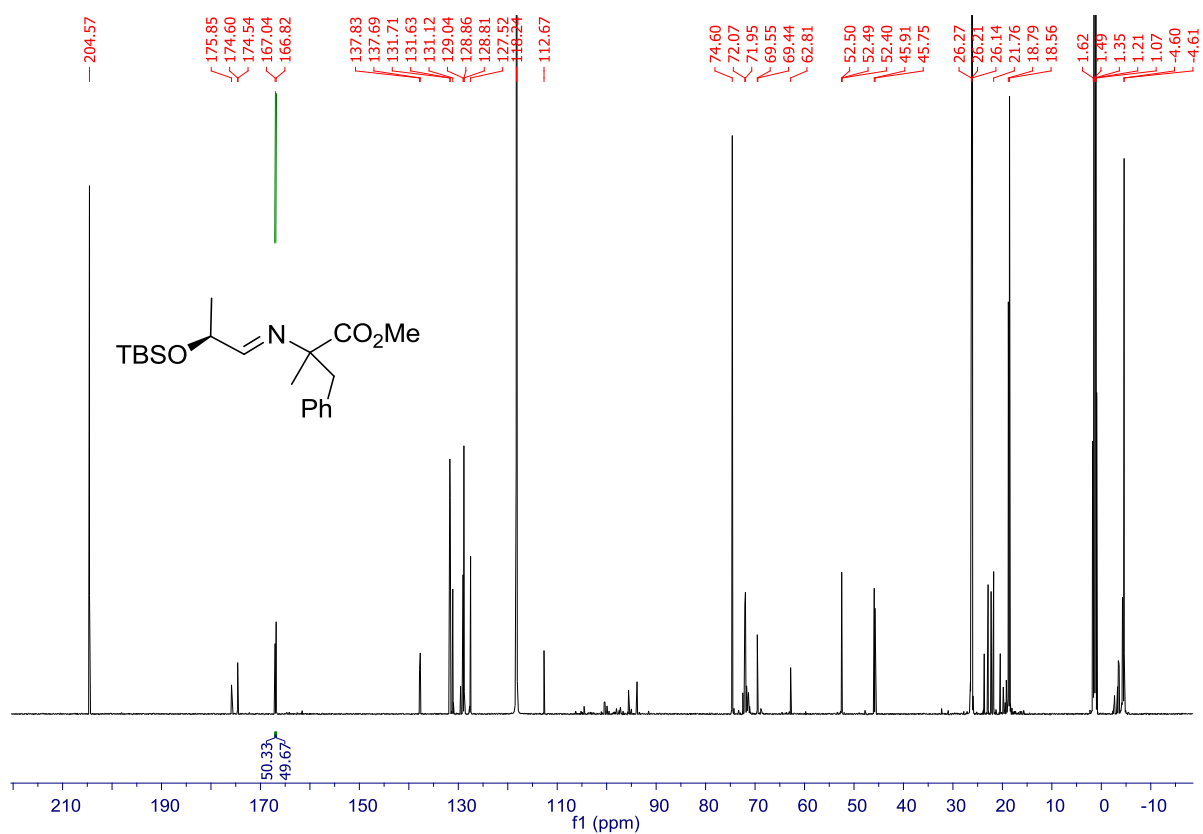
Chemical structure: CC(C)(C)Si(C)(C)C[C@H](C)/C=C/[N+]1CCCC1c2ccccc2

¹³C NMR spectrum (ppm):

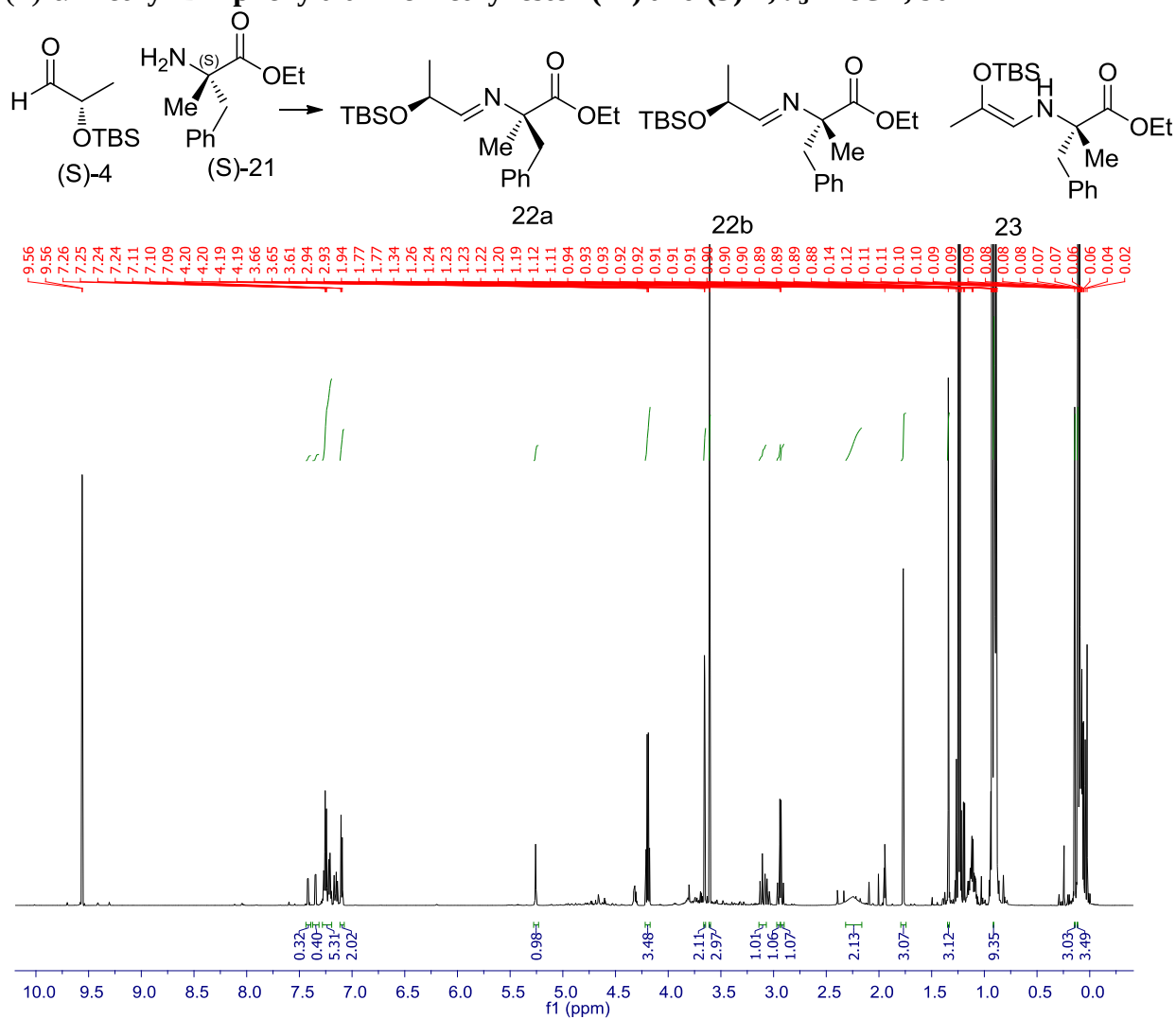
- 170.35, 170.31, 138.29, 138.29, 137.82, 137.30, 137.25, 130.50, 130.27, 129.82, 129.57, 129.08, 128.29, 128.26, 127.75, 127.07, 126.92, 126.80, 102.66, 102.11, 101.93, 72.44, 72.35, 71.55, 71.40, 71.04, 70.12, 68.46, 68.40, 50.09, 49.91, 49.48, 49.34, 49.20, 49.06, 48.92, 48.77, 48.63, 48.53, 48.39, 48.24, 48.10, 47.95, 47.81, 47.67, 47.53, 47.39, 47.25, 47.11, 46.97, 46.83, 46.69, 46.55, 46.41, 46.27, 46.13, 45.99, 45.85, 45.71, 45.57, 45.43, 45.29, 45.15, 45.01, 44.87, 44.73, 44.59, 44.45, 44.31, 44.17, 44.03, 43.89, 43.75, 43.61, 43.47, 43.33, 43.19, 43.05, 42.91, 42.77, 42.63, 42.49, 42.35, 42.21, 42.07, 41.93, 41.79, 41.65, 41.51, 41.37, 41.23, 41.09, 40.95, 40.81, 40.67, 40.53, 40.39, 40.25, 40.11, 39.97, 39.83, 39.69, 39.55, 39.41, 39.27, 39.13, 38.99, 38.85, 38.71, 38.57, 38.43, 38.29, 38.15, 38.01, 37.87, 37.73, 37.59, 37.45, 37.31, 37.17, 37.03, 36.89, 36.75, 36.61, 36.47, 36.33, 36.19, 36.05, 35.91, 35.77, 35.63, 35.49, 35.35, 35.21, 35.07, 34.93, 34.79, 34.65, 34.51, 34.37, 34.23, 34.09, 33.95, 33.81, 33.67, 33.53, 33.39, 33.25, 33.11, 32.97, 32.83, 32.69, 32.55, 32.41, 32.27, 32.13, 31.99, 31.85, 31.71, 31.57, 31.43, 31.29, 31.15, 31.01, 30.87, 30.73, 30.59, 30.45, 30.31, 30.17, 30.03, 29.89, 29.75, 29.61, 29.47, 29.33, 29.19, 29.05, 28.91, 28.77, 28.63, 28.49, 28.35, 28.21, 28.07, 27.93, 27.79, 27.65, 27.51, 27.37, 27.23, 27.09, 26.95, 26.81, 26.67, 26.53, 26.39, 26.25, 26.11, 25.97, 25.83, 25.69, 25.55, 25.41, 25.27, 25.13, 24.99, 24.85, 24.71, 24.57, 24.43, 24.29, 24.15, 24.01, 23.87, 23.73, 23.59, 23.45, 23.31, 23.17, 23.03, 22.89, 22.75, 22.61, 22.47, 22.33, 22.19, 22.05, 21.91, 21.77, 21.63, 21.49, 21.35, 21.21, 21.07, 20.93, 20.79, 20.65, 20.51, 20.37, 20.23, 20.09, 19.95, 19.81, 19.67, 19.53, 19.39, 19.25, 19.11, 18.97, 18.83, 18.69, 18.55, 18.41, 18.27, 18.13, 17.99, 17.85, 17.71, 17.57, 17.43, 17.29, 17.15, 17.01, 16.87, 16.73, 16.59, 16.45, 16.31, 16.17, 16.03, 15.89, 15.75, 15.61, 15.47, 15.33, 15.19, 15.05, 14.91, 14.77, 14.63, 14.49, 14.35, 14.21, 14.07, 13.93, 13.79, 13.65, 13.51, 13.37, 13.23, 13.09, 12.95, 12.81, 12.67, 12.53, 12.39, 12.25, 12.11, 11.97, 11.83, 11.69, 11.55, 11.41, 11.27, 11.13, 10.99, 10.85, 10.71, 10.57, 10.43, 10.29, 10.15, 10.01, 9.87, 9.73, 9.59, 9.45, 9.31, 9.17, 9.03, 8.89, 8.75, 8.61, 8.47, 8.33, 8.19, 8.05, 7.91, 7.77, 7.63, 7.49, 7.35, 7.21, 7.07, 6.93, 6.79, 6.65, 6.51, 6.37, 6.23, 6.09, 5.95, 5.81, 5.67, 5.53, 5.39, 5.25, 5.11, 4.97, 4.83, 4.69, 4.55, 4.41, 4.27, 4.13, 4.00, 3.86, 3.72, 3.58, 3.44, 3.30, 3.16, 3.02, 2.88, 2.74, 2.60, 2.46, 2.32, 2.18, 2.04, 1.90, 1.76, 1.62, 1.48, 1.34, 1.20, 1.06, 0.92, 0.78, 0.64, 0.50, 0.36, 0.22, 0.08, -0.06, -0.20, -0.34, -0.48, -0.62, -0.76, -0.90, -1.04, -1.18, -1.32, -1.46, -1.60, -1.74, -1.88, -2.02, -2.16, -2.30, -2.44, -2.58, -2.72, -2.86, -3.00, -3.14, -3.28, -3.42, -3.56, -3.70, -3.84, -3.98, -4.12, -4.26, -4.40, -4.54, -4.68, -4.82, -4.96, -5.10, -5.24, -5.38, -5.52, -5.66, -5.80, -5.94, -6.08, -6.22, -6.36, -6.50, -6.64, -6.78, -6.92, -7.06, -7.20, -7.34, -7.48, -7.62, -7.76, -7.90, -8.04, -8.18, -8.32, -8.46, -8.60, -8.74, -8.88, -9.02, -9.16, -9.30, -9.44, -9.58, -9.72, -9.86, -10.00, -10.14, -10.28, -10.42, -10.56, -10.70, -10.84, -10.98, -11.12, -11.26, -11.40, -11.54, -11.68, -11.82, -11.96, -12.10, -12.24, -12.38, -12.52, -12.66, -12.80, -12.94, -13.08, -13.22, -13.36, -13.50, -13.64, -13.78, -13.92, -14.06, -14.20, -14.34, -14.48, -14.62, -14.76, -14.90, -15.04, -15.18, -15.32, -15.46, -15.60, -15.74, -15.88, -16.02, -16.16, -16.30, -16.44, -16.58, -16.72, -16.86, -17.00, -17.14, -17.28, -17.42, -17.56, -17.70, -17.84, -17.98, -18.12, -18.26, -18.40, -18.54, -18.68, -18.82, -18.96, -19.10, -19.24, -19.38, -19.52, -19.66, -19.80, -19.94, -20.08, -20.22, -20.36, -20.50, -20.64, -20.78, -20.92, -21.06, -21.20, -21.34, -21.48, -21.62, -21.76, -21.90, -22.04, -22.18, -22.32, -22.46, -22.60, -22.74, -22.88, -23.02, -23.16, -23.30, -23.44, -23.58, -23.72, -23.86, -24.00, -24.14, -24.28, -24.42, -24.56, -24.70, -24.84, -24.98, -25.12, -25.26, -25.40, -25.54, -25.68, -25.82, -25.96, -26.10, -26.24, -26.38,

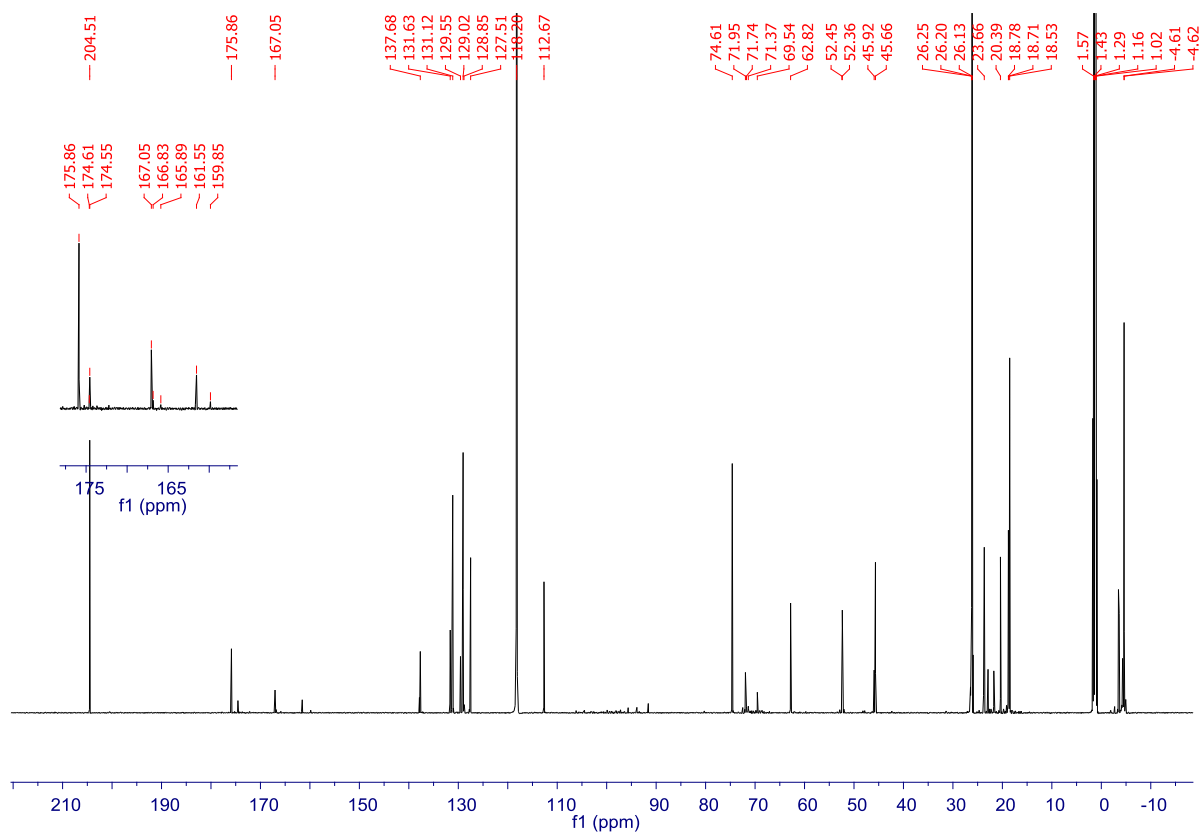


(±)- α -Methyl-DL-phenylalanine methyl ester-(21) and (S)-4, d_3 -MeCN

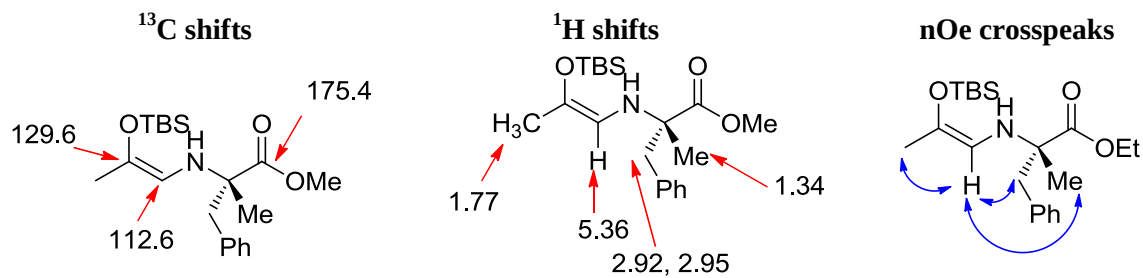


(±)- α -Methyl-DL-phenylalanine methyl ester-(21) and (S)-4, d_3 -MeCN, 90 min



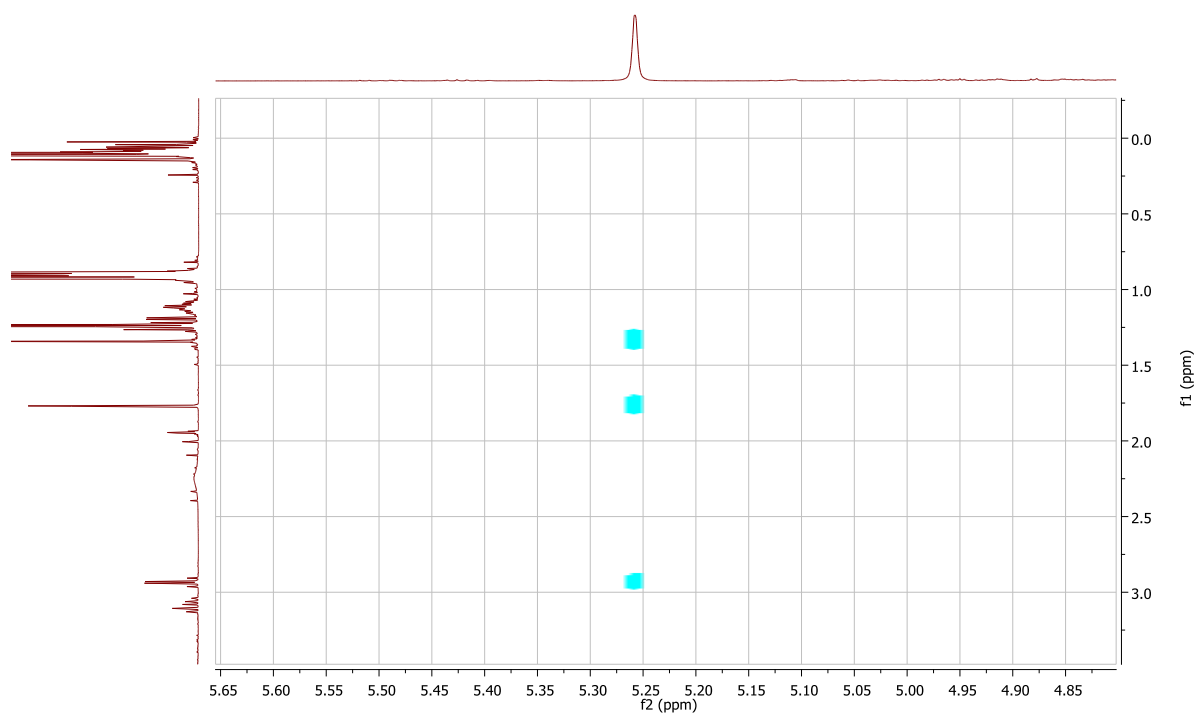
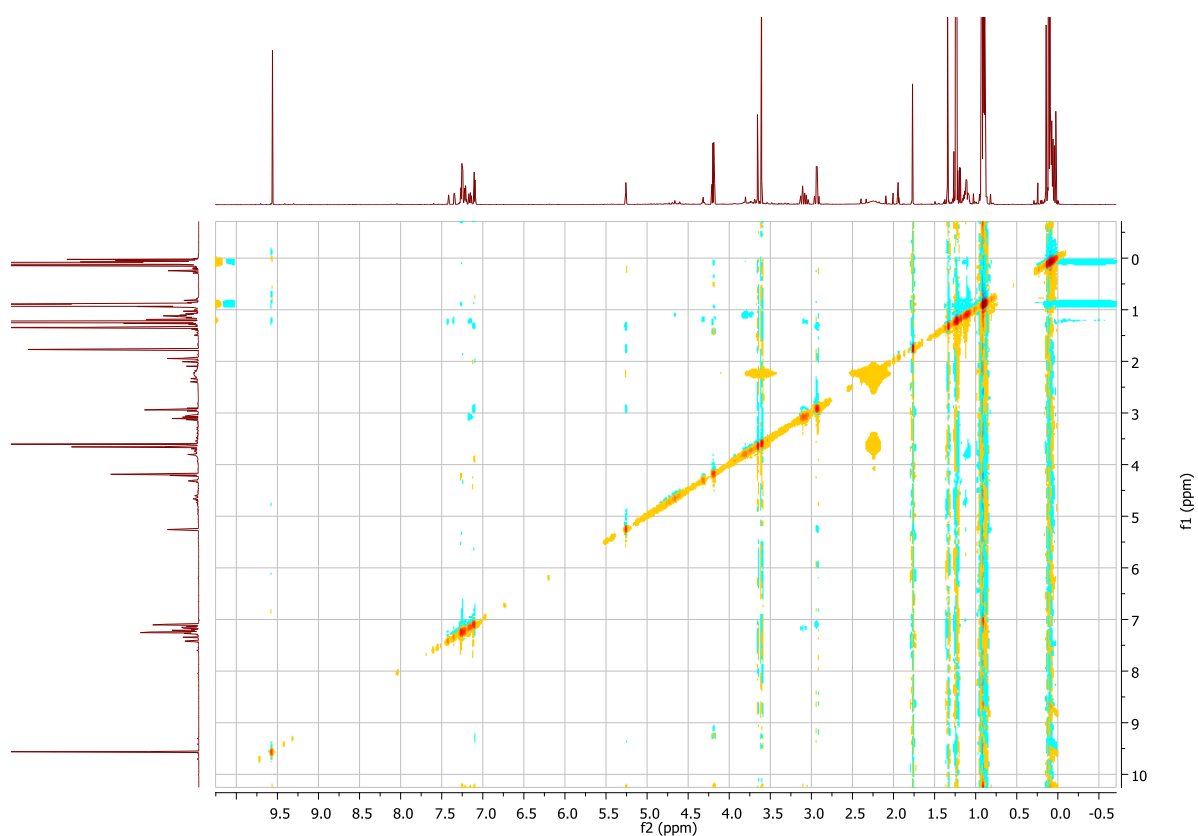


The diagnostic signals for the enamine **23** were assigned as follows:

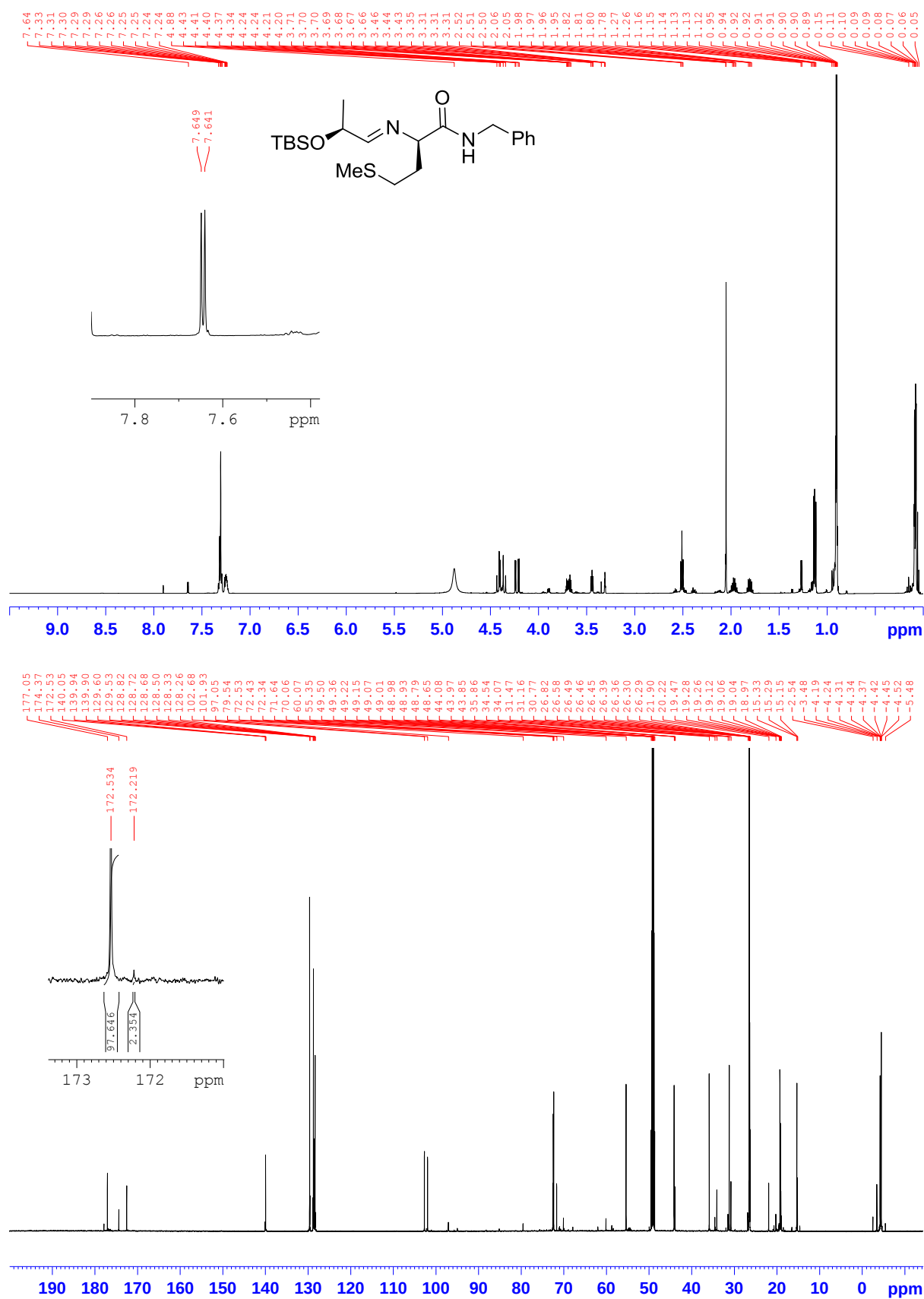


The geometry was determined from the nOe's shown. The NOESY spectrum is on the following page.

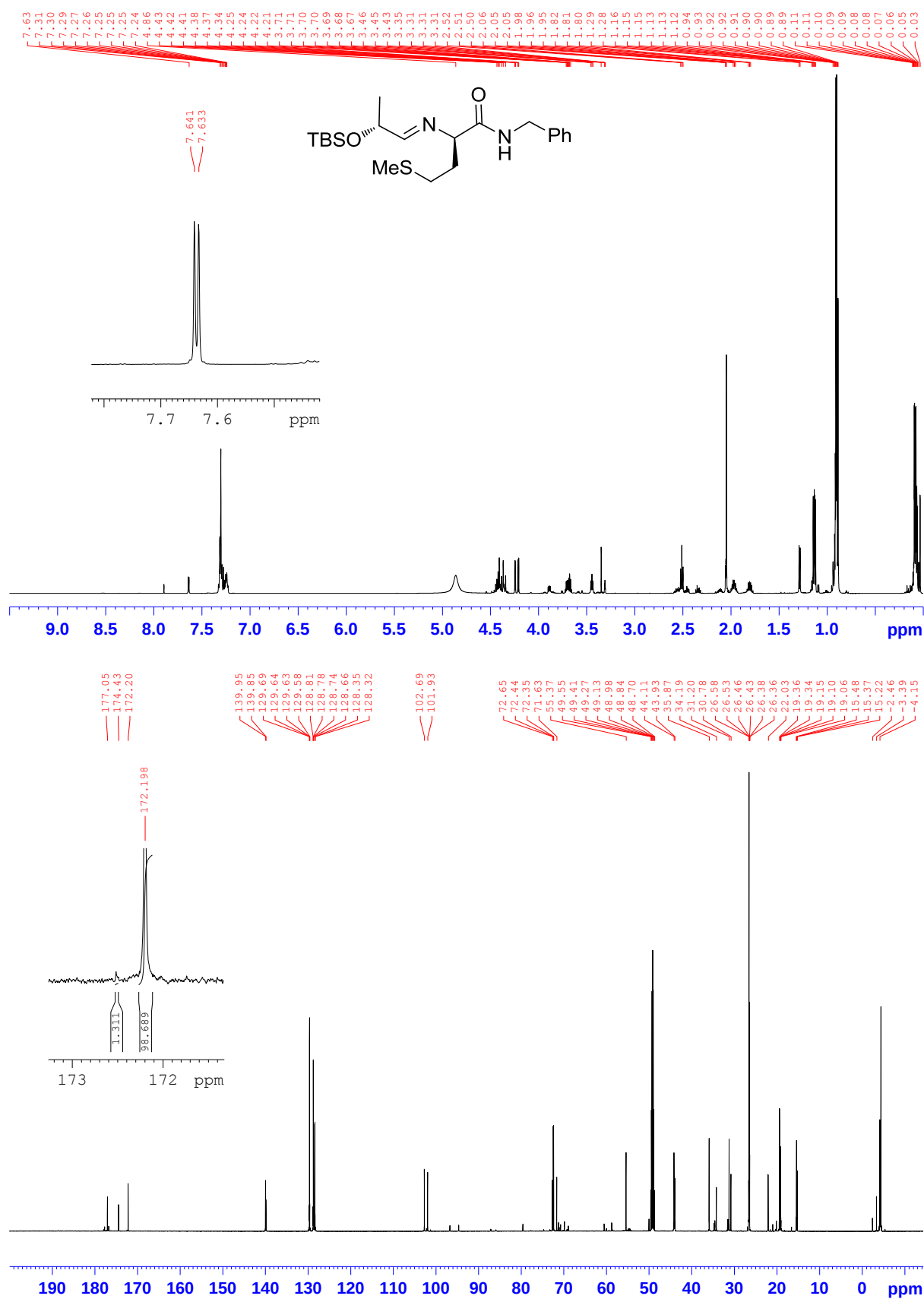
NOESY spectrum of mixture of imines 22 and enamine 23



Amide 26a and (S)-4



Amide 26a and (R)-4



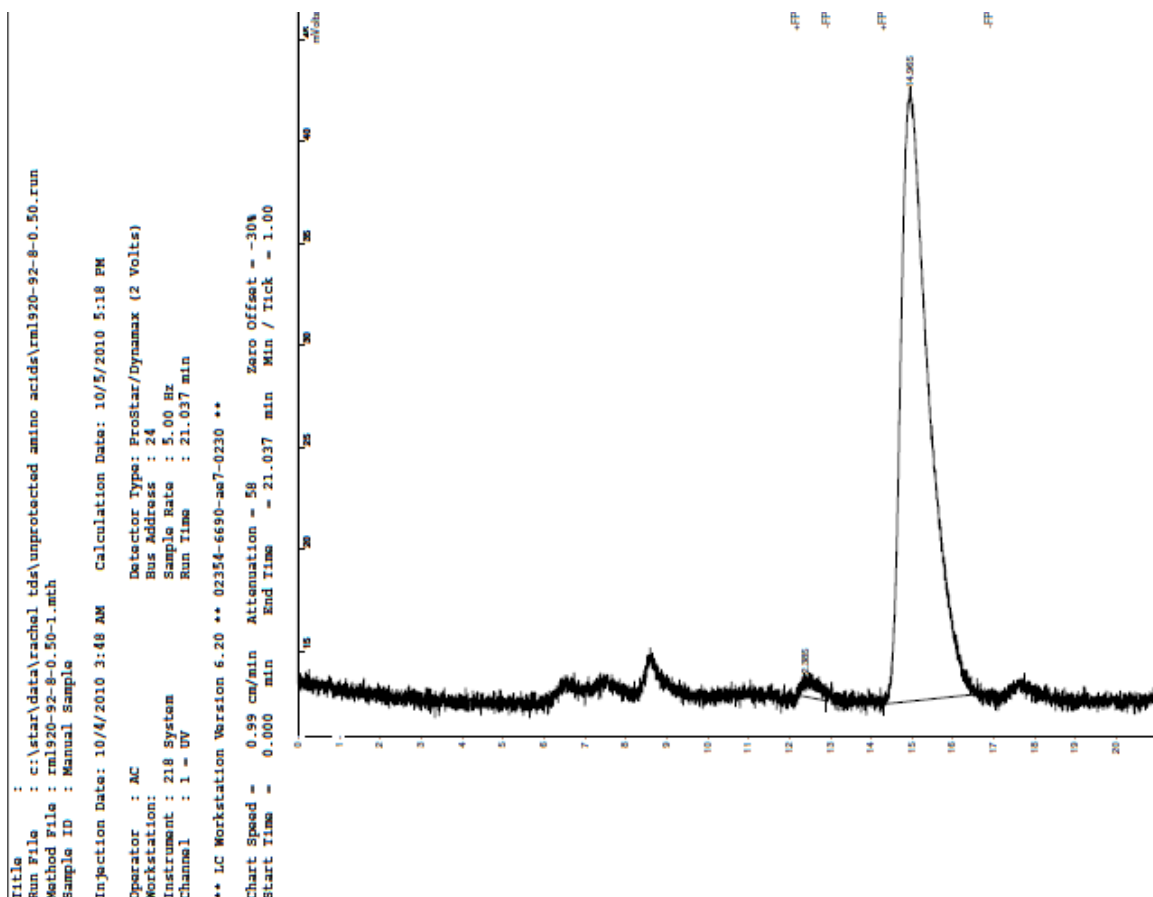
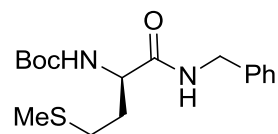
CC(C)[C@H](N=C[C@@H](CS)C(=O)NCC1=CC=CC=C1)OSi(C)(C)C(C)(C)C

RML920-D 11 1 C:\Bruker\TOPSPIN Rachel600

RML920-L 11 1 C:\Bruker\TOPSPIN Rachel600

173.0 172.5 172.0 171.5 [ppm]

Amide 25a chiral HPLC



Print Date: Tue Oct 05 17:13:50 2010

Page 1 of 1

Title :
 Run File : c:\star\data\rachel tds\unprotected amino acids\rm1920-92-8-0.50.run
 Method File : rm1920-92-8-0.50-1.mth
 Sample ID : Manual Sample

Injection Date: 10/4/2010 3:48 AM Calculation Date: 10/5/2010 5:18 PM

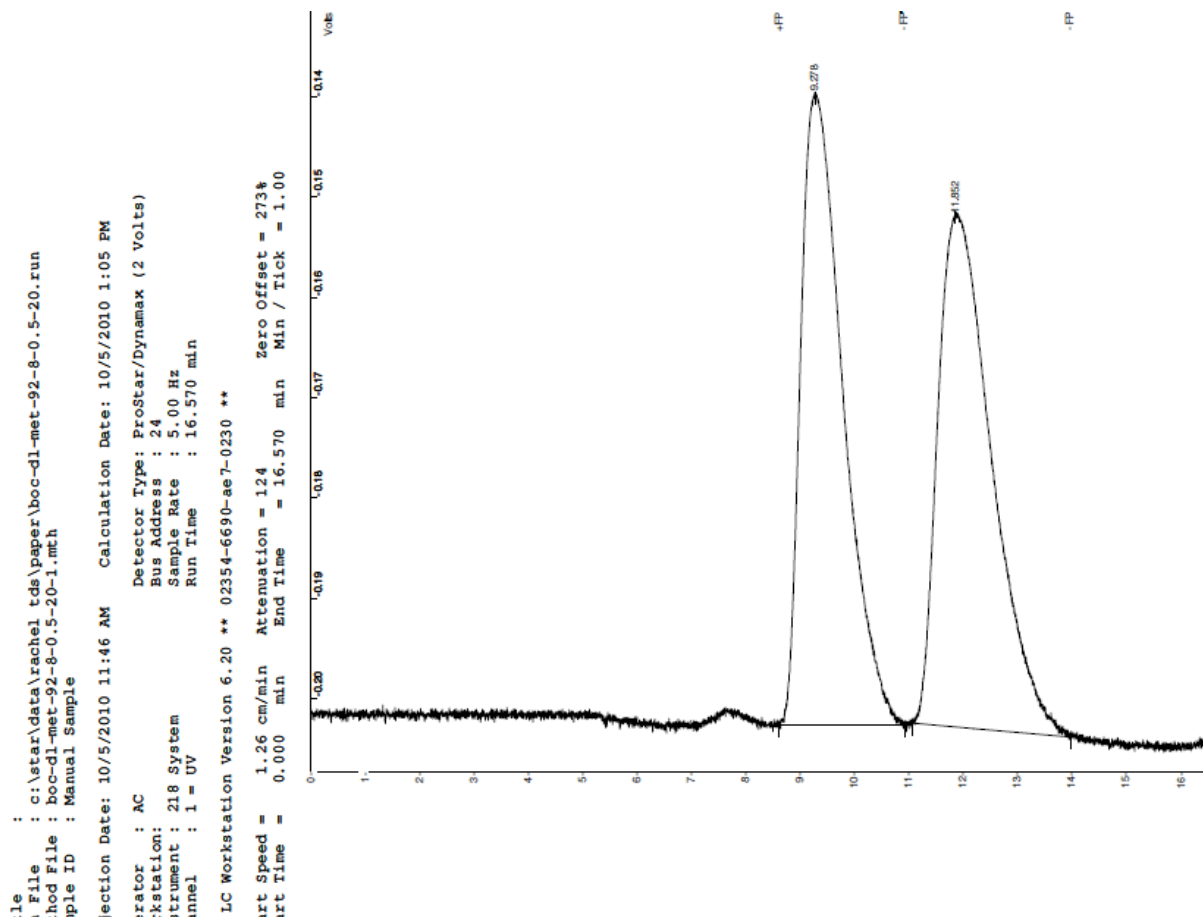
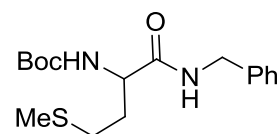
Operator : AC
 Workstation: Detector Type: ProStar/Dynamax (2 Volts)
 Instrument : 218 System Bus Address : 24
 Channel : 1 - UV Sample Rate : 5.00 Hz
 Run Time : 21.037 min

** LC Workstation Version 6.20 ** 02354-6690-as7-0230 **

Run Mode : Analysis
 Peak Measurement: Peak Area
 Calculation Type: Percent

Peak No.	Peak Name	Result (%)	Ret. Time (min)	Time Offset (min)	Area (counts)	Sep. Code	Width 1/2 (sec)	Status Codes
1		1.6603	12.385	0.000	24171	BB	0.5	
2		98.3397	14.965	0.000	1431657	BB	43.2	
Totals:		100.0000		0.000	1455828			

(±)-25a chiral HPLC



Print Date: Tue Oct 05 13:05:35 2010

Page 1 of 1

Title :
 Run File : c:\star\data\rachel tds\paper\boc-dl-met-92-8-0.5-20.run
 Method File : boc-dl-met-92-8-0.5-20-1.mth
 Sample ID : Manual Sample

Injection Date: 10/5/2010 11:46 AM Calculation Date: 10/5/2010 1:05 PM

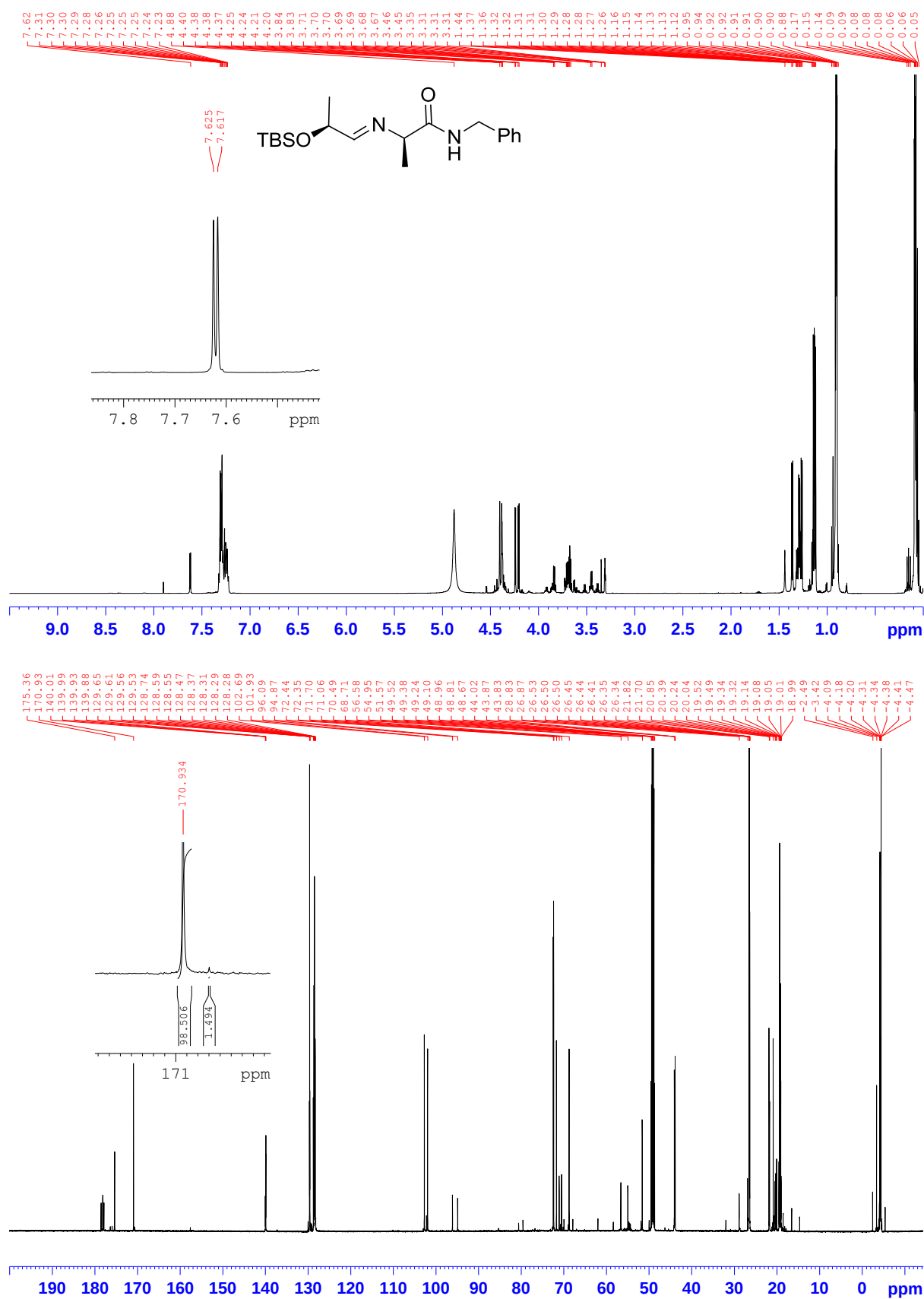
Operator : AC
 Workstation: ProStar/Dynamax (2 Volts)
 Instrument : 218 System
 Channel : 1 - UV
 Detector Type: ProStar/Dynamax (2 Volts)
 Bus Address : 24
 Sample Rate : 5.00 Hz
 Run Time : 16.570 min

** LC Workstation Version 6.20 ** 02354-6690-ae7-0230 **

Run Mode : Analysis
 Peak Measurement: Peak Area
 Calculation Type: Percent

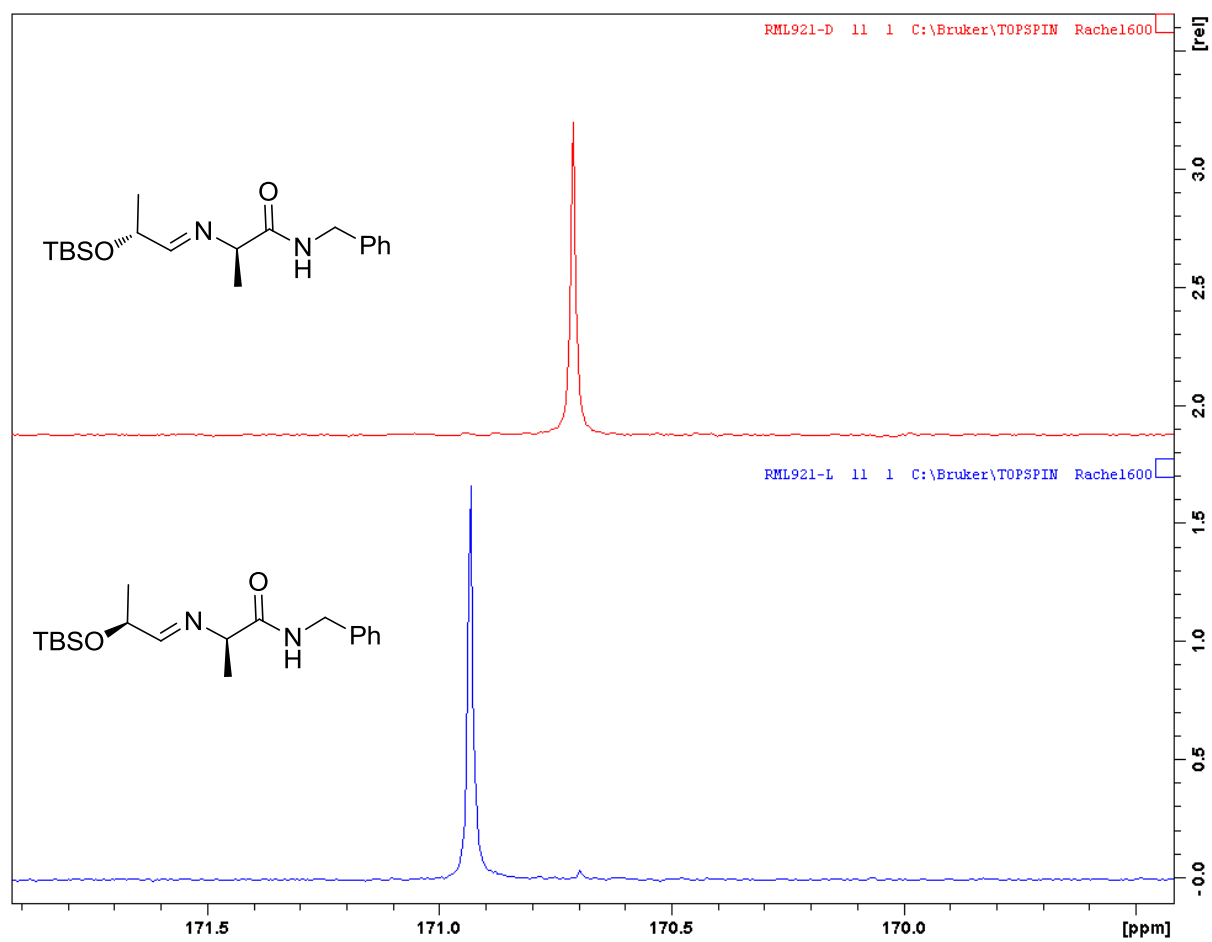
Peak No.	Peak Name	Result ()	Ret. Time (min)	Time Offset (min)	Area (counts)	Sep. Code	Width 1/2 (sec)	Status Codes
1		49.0595	9.278	0.000	3391570	BB	49.7	
2		50.9406	11.852	0.000	3521614	BB	62.9	
Totals:		100.0001		0.000	6913184			

Amide 26b and (S)-4

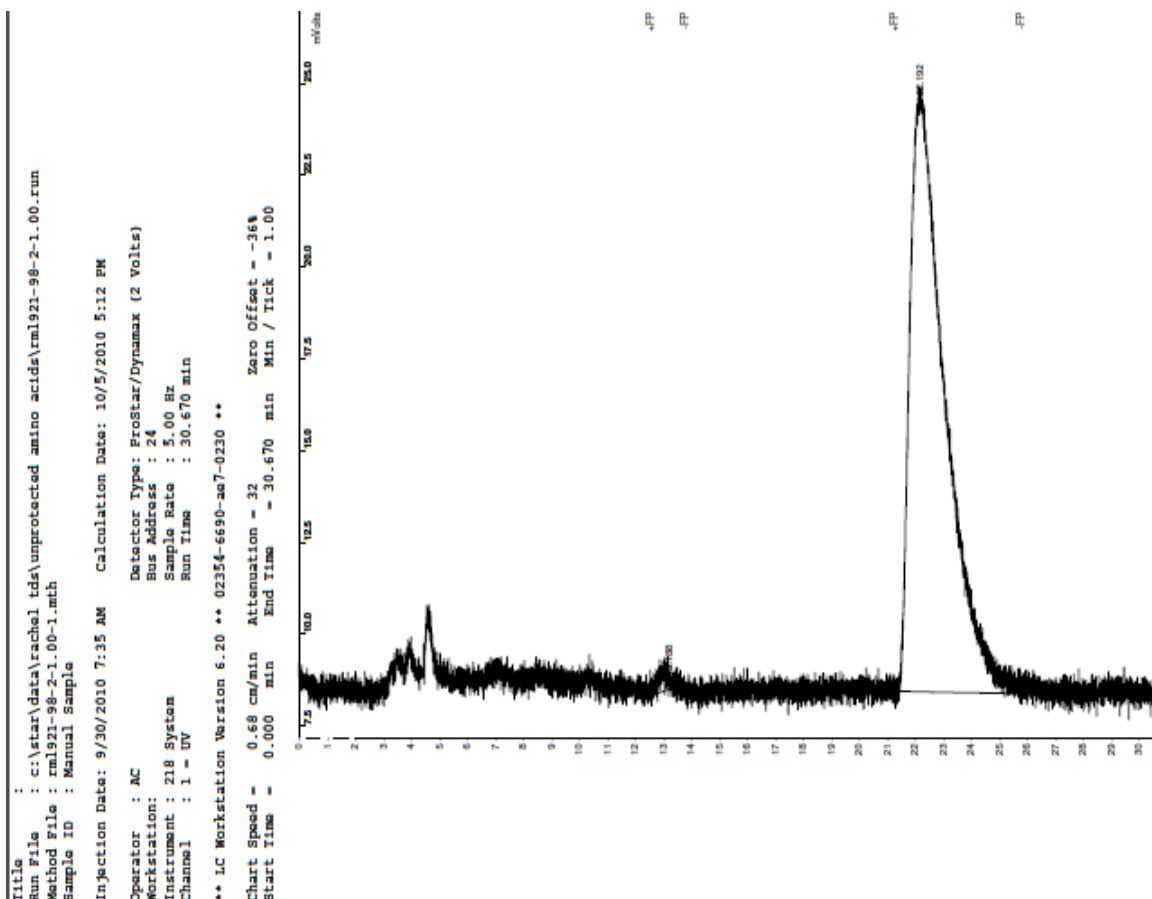
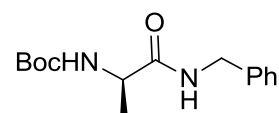


[illegible]

Amide 26b and (S)-4/Amide 26a and (R)-4 overlaid



Amide 25b Chiral HPLC



Print Date: Tue Oct 05 17:13:07 2010

Page 1 of 1

Title :
 Run File : c:\star\data\rachel tds\unprotected amino acids\rm1921-98-2-1.00.run
 Method File : rm1921-98-2-1.00-1.mth
 Sample ID : Manual Sample

Injection Date: 9/30/2010 7:35 AM Calculation Date: 10/5/2010 5:12 PM

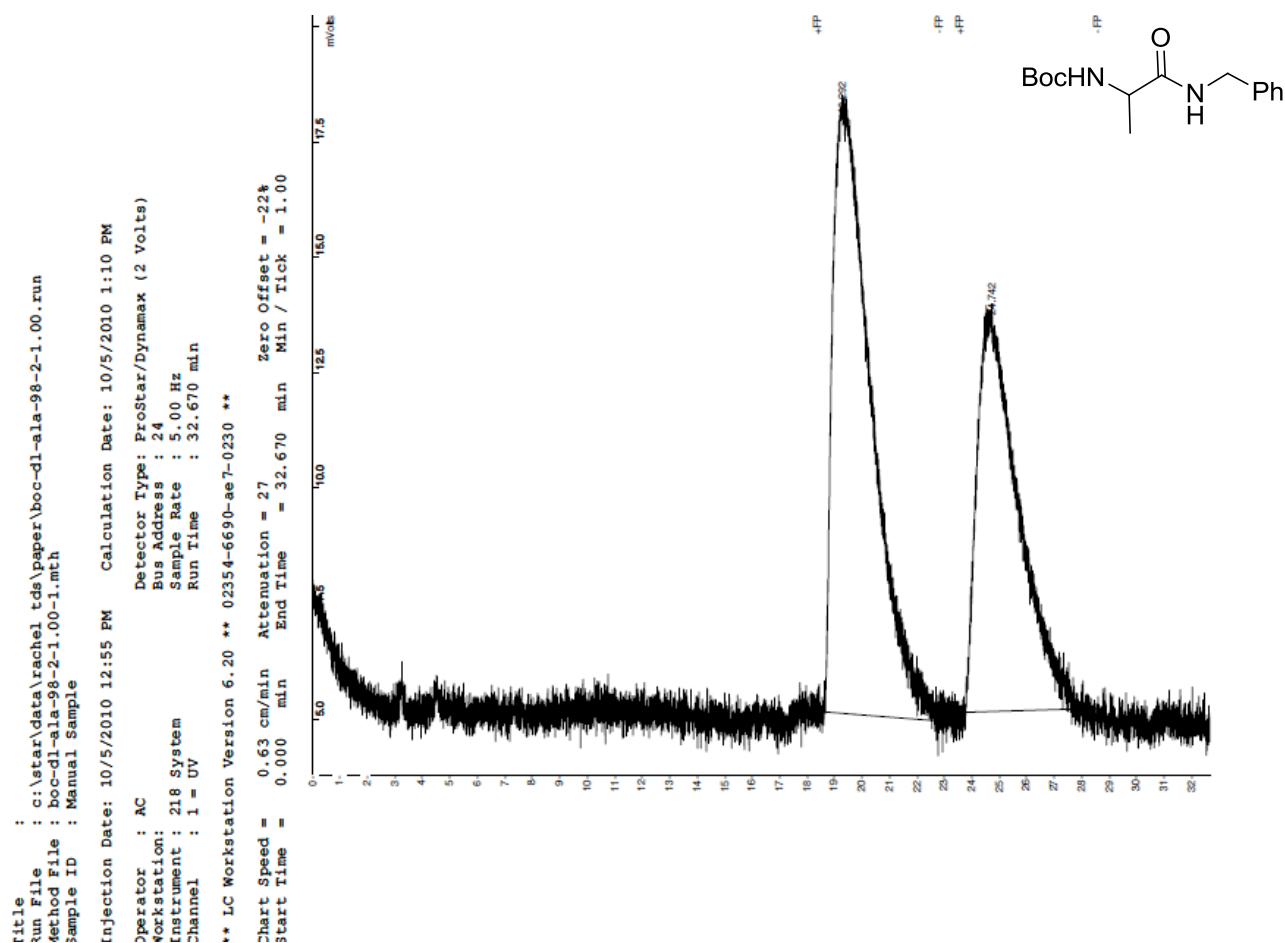
Operator : AC
 Workstation: 218 System
 Instrument : 218 System
 Channel : 1 = UV
 Detector Type: ProStar/Dynamax (2 Volts)
 Bus Address : 24
 Sample Rate : 5.00 Hz
 Run Time : 30.670 min

** LC Workstation Version 6.20 ** 02354-6690-ae7-0230 **

Run Mode : Analysis
 Peak Measurement: Peak Area
 Calculation Type: Percent

Peak No.	Peak Name	Result (%)	Ret. Time (min)	Time Offset (min)	Area (counts)	Sep. Code	Width 1/2 (sec)	Status Codes
1		0.9707	13.188	0.000	13881	BB	0.3	
2		99.0293	22.192	0.000	1416128	BB	76.2	
Totals:		100.0000		0.000	1430009			

(±)-25b chiral HPLC



Print Date: Tue Oct 05 13:00:47 2010

Page 1 of 1

Title :
 Run File : c:\star\data\rachel tds\paper\boc-dl-ala-98-2-1.00.run
 Method File : boc-dl-ala-98-2-1.00-1.mth
 Sample ID : Manual Sample

Injection Date: 10/5/2010 12:55 PM Calculation Date: 10/5/2010 1:10 PM

Operator : AC
 Workstation: 218 System
 Instrument : 218 System
 Channel : 1 - UV
 Detector Type: ProStar/Dynamax (2 Volts)
 Bus Address : 24
 Sample Rate : 5.00 Hz
 Run Time : 32.670 min

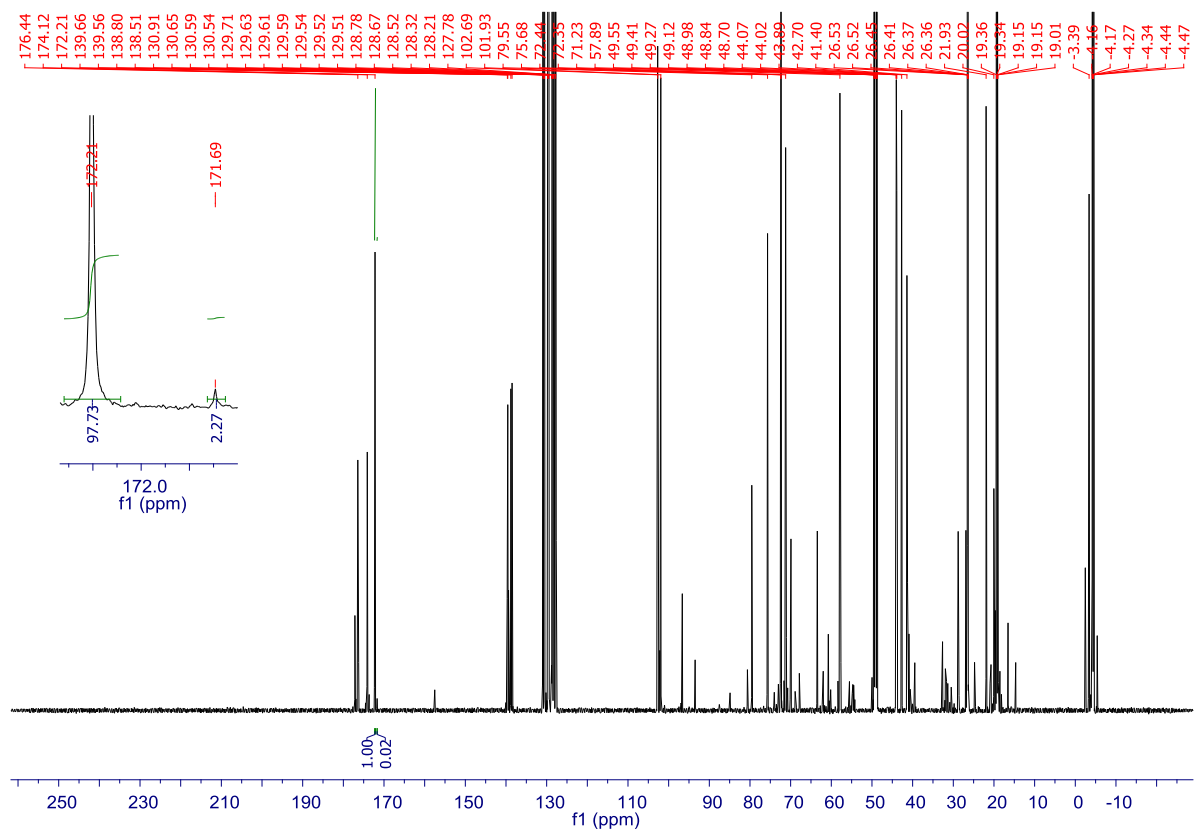
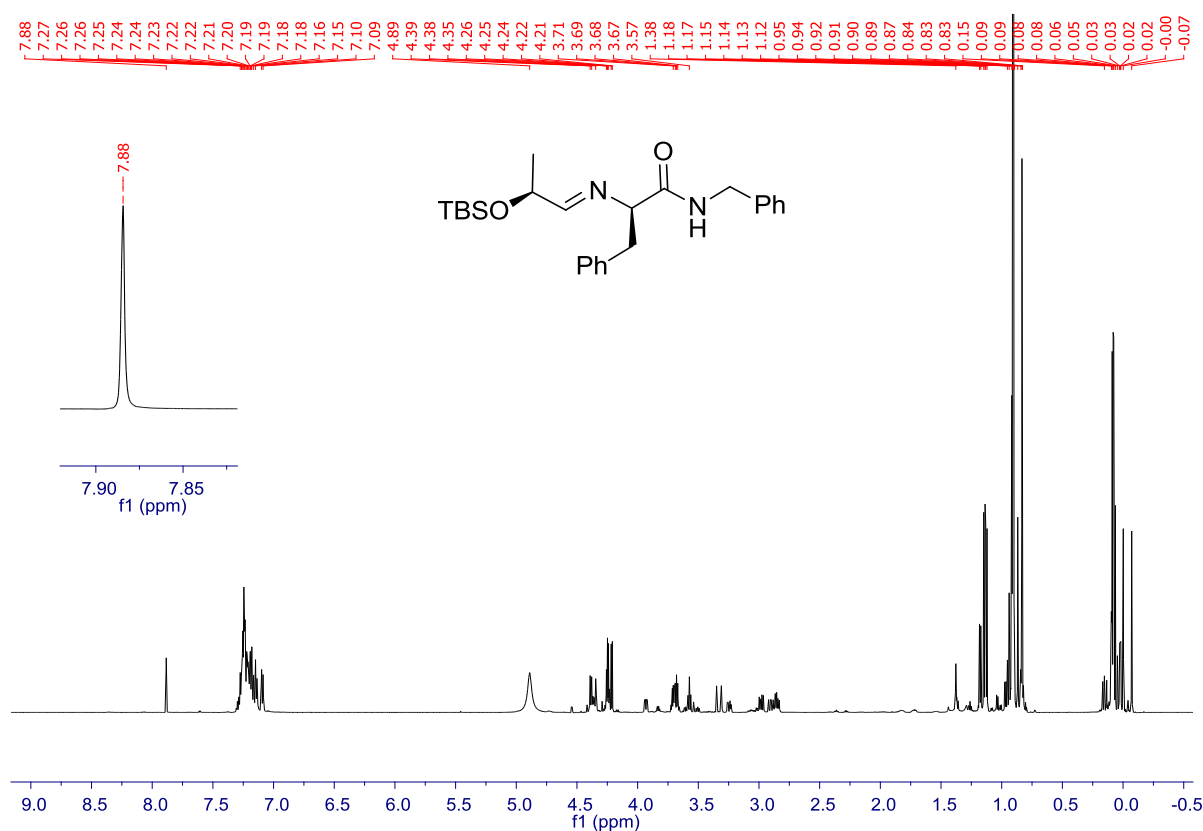
** LC Workstation Version 6.20 ** 02354-6690-ae7-0230 **

Run Mode : Analysis
 Peak Measurement: Peak Area
 Calculation Type: Percent

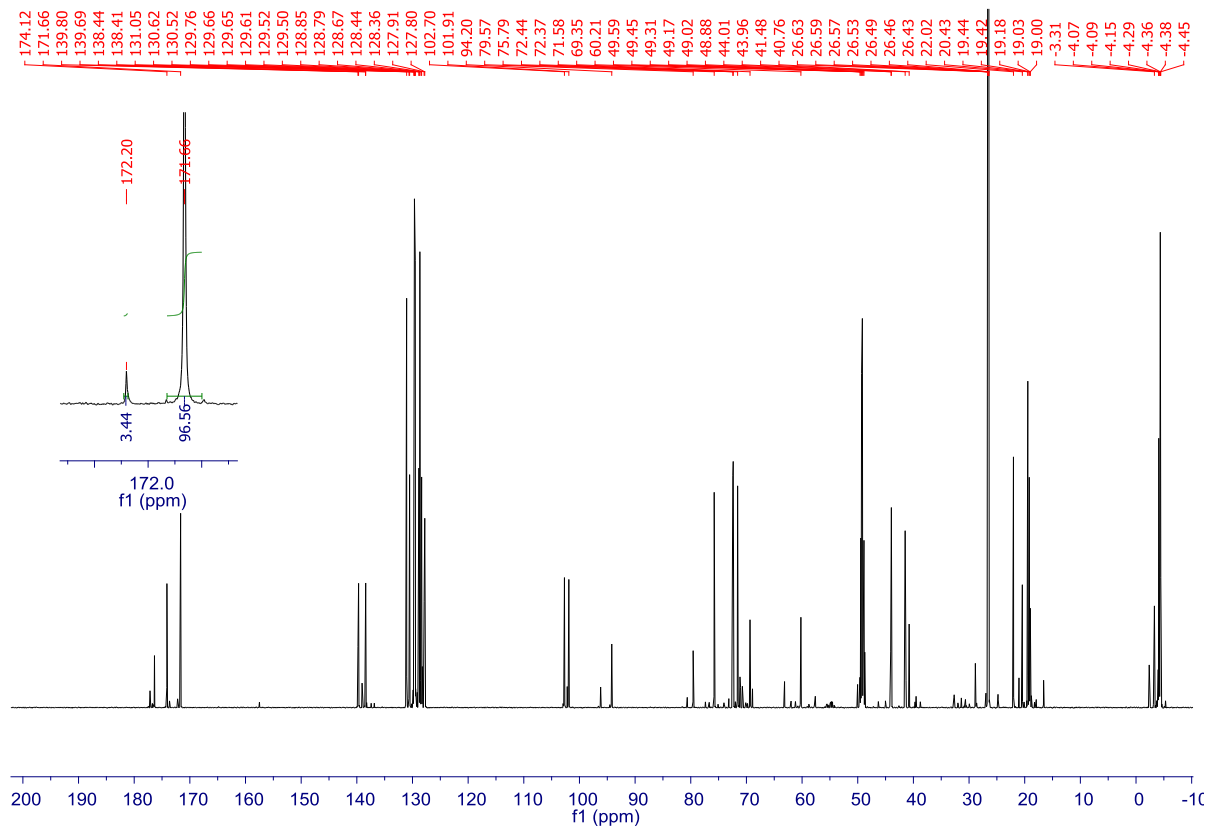
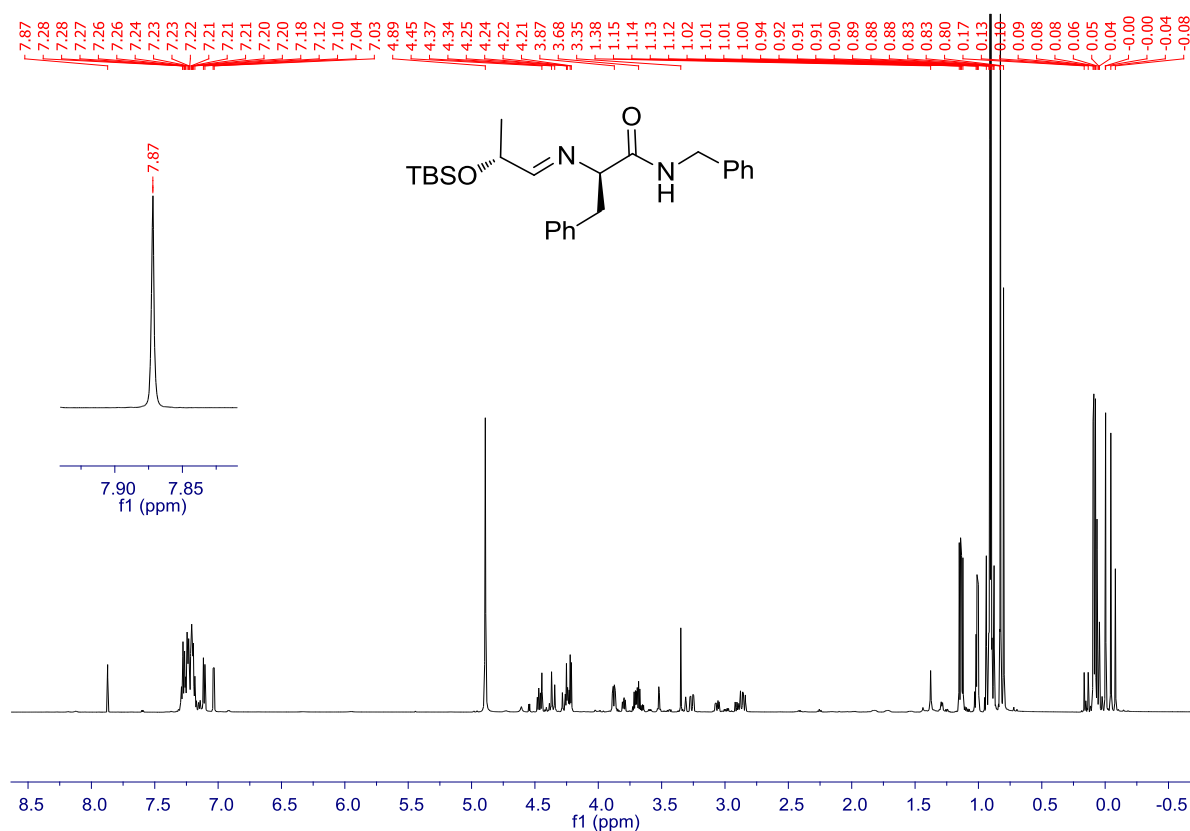
Peak No.	Peak Name	Result (%)	Ret. Time (min)	Time Offset (min)	Area (counts)	Sep. Code	Width 1/2 (sec)	Status Codes
1		59.1229	19.292	0.000	1295464	BB	85.5	
2		40.8771	24.742	0.000	895673	BB	89.4	
Totals:		100.0000		0.000	2191137			

Total Unidentified Counts : 2191137 counts

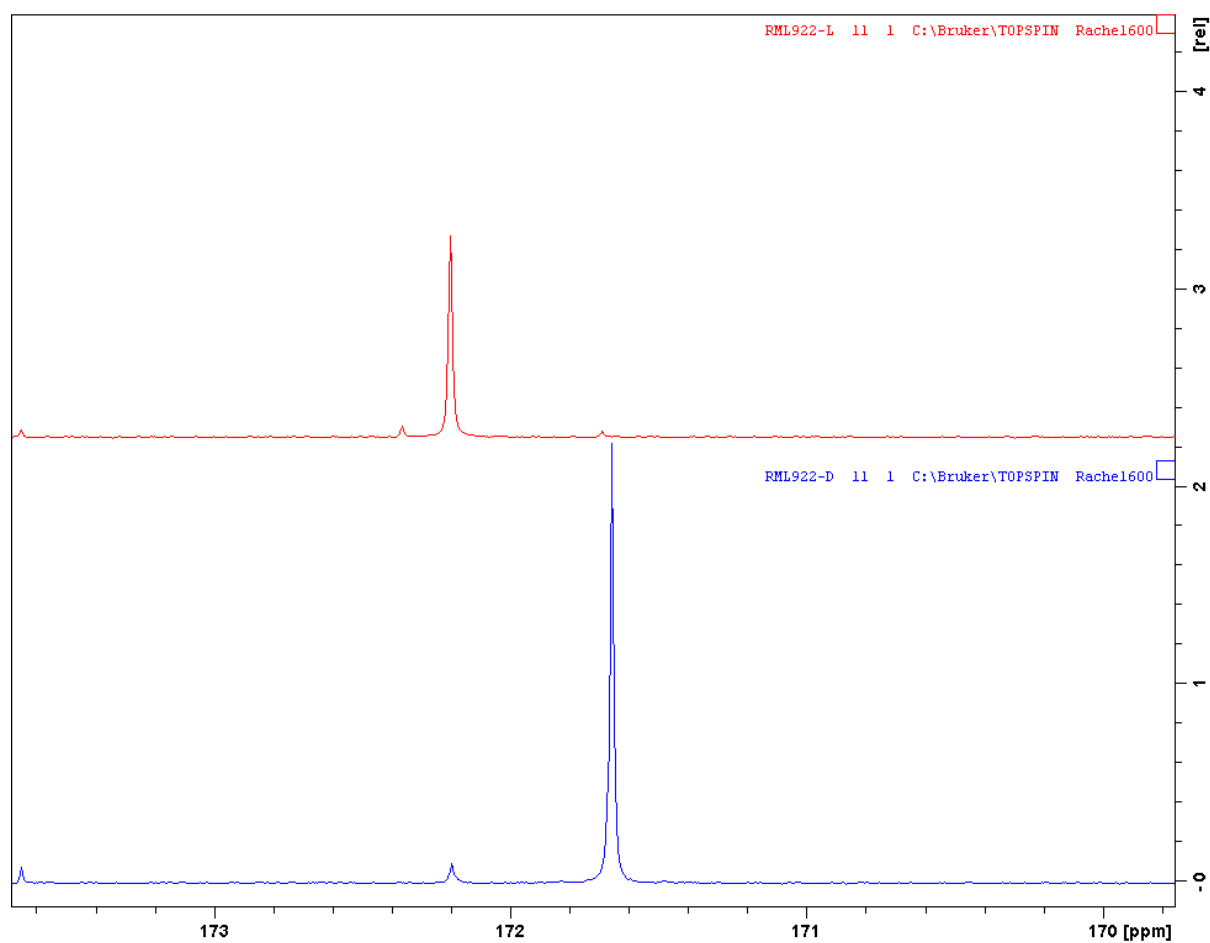
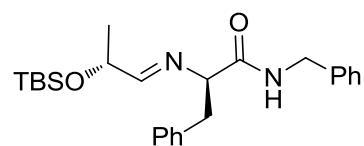
Amide 26c and (S)-4



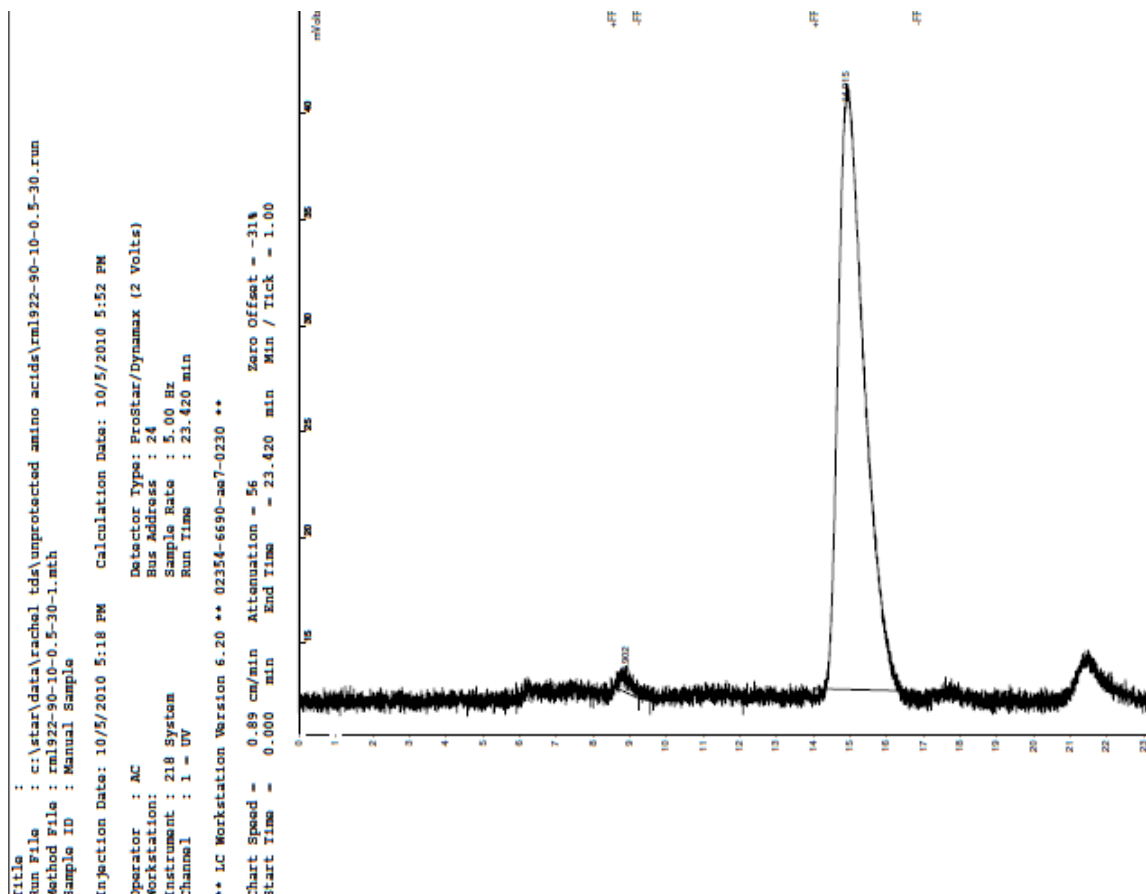
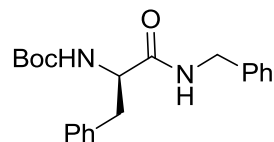
Amide 26c and (R)-4



Amide 26c and (S)-4/Amide 26a and (R)-4 overlaid



Amide 25c Chiral HPLC



Print Date: Tue Oct 05 17:14:32 2010

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Title :
 Run File : c:\star\data\rachel tds\unprotected amino acids\rm1922-90-10-0.5-30.run
 Method File : rm1922-90-10-0.5-30-1.mth
 Sample ID : Manual Sample

Injection Date: 10/5/2010 5:18 PM Calculation Date: 10/5/2010 5:52 PM

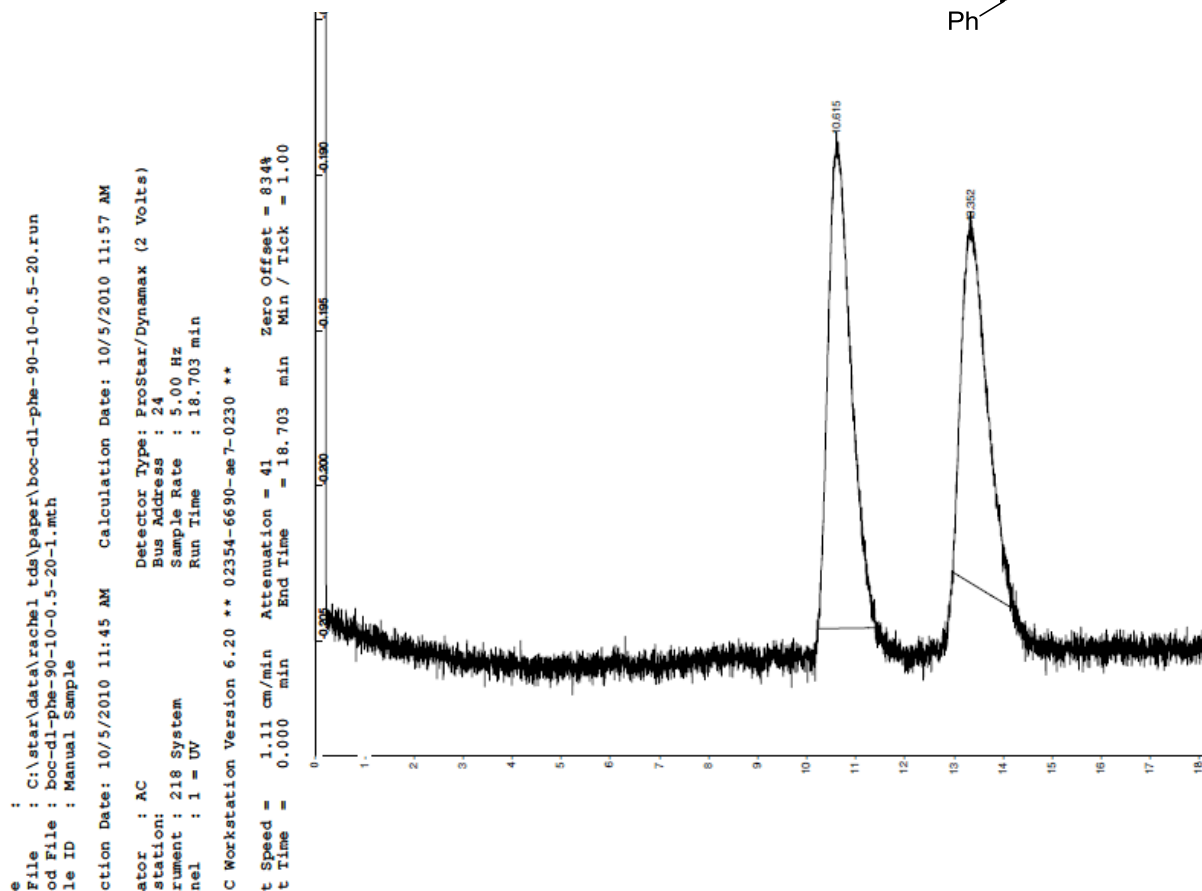
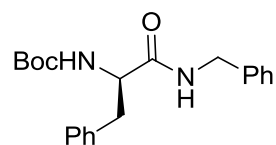
Operator : AC
 Workstation: ProStar/Dynamax (2 Volts)
 Instrument : 218 System
 Channel : 1 - UV
 Bus Address : 24
 Sample Rate : 5.00 Hz
 Run Time : 23.420 min

** LC Workstation Version 6.20 ** 02354-6690-ae7-0230 **

Run Mode : Analysis
 Peak Measurement: Peak Area
 Calculation Type: Percent

Peak No.	Peak Name	Result ()	Ret. Time (min)	Time Offset (min)	Area (counts)	Sep. Code	Width 1/2 (sec)	Status Codes
1		1.2955	8.902	0.000	18165	BB	0.5	
2		98.7045	14.915	0.000	1384045	BB	44.7	
Totals:		100.0000		0.000	1402210			

(±)-Amide 25c Chiral HPLC



Print Date: Tue Oct 05 13:08:07 2010

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Title :
 Run File : C:\star\data\rachel tds\paper\boc-dl-phe-90-10-0.5-20.run
 Method File : boc-dl-phe-90-10-0.5-20-1.mth
 Sample ID : Manual Sample

Injection Date: 10/5/2010 11:45 AM Calculation Date: 10/5/2010 11:57 AM

Operator : AC Detector Type: ProStar/Dynamax (2 Volts)
 Workstation: Bus Address : 24
 Instrument : 218 System Sample Rate : 5.00 Hz
 Channel : 1 = UV Run Time : 18.703 min

** LC Workstation Version 6.20 ** 02354-6690-ae7-0230 **

Run Mode : Analysis
 Peak Measurement: Peak Area
 Calculation Type: Percent

Peak No.	Peak Name	Result ()	Ret. Time (min)	Time Offset (min)	Area (counts)	Sep. Code	Width 1/2 (sec)	Status Codes
1		54.8020	10.615	0.000	505270	BB	29.9	
2		45.1980	13.352	0.000	416721	BB	31.0	
Totals:		100.0000		0.000	921991			

References

- 1 W. R. Li, W. R. Ewing, B. D. Harris, and M. M. Joullié, *J. Am. Chem. Soc.*, 1990, **112**, 7659–7672.
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- 4 R. M. Lanigan, P. Starkov, T. D. Sheppard, *J. Org. Chem.* 2013, **78**, 4512-4523.