Supporting Information

Effects of structural modifications on the metal binding, anti-amyloid activity, and cholinesterase inhibitory activity of chalcones

Marina Y. Fosso,¹ Harry LeVine, 3rd,² Keith D. Green,¹ Oleg V. Tsodikov,¹ and Sylvie Garneau-Tsodikova^{1,*}

¹ Department of Pharmaceutical Sciences, University of Kentucky, Lexington, KY, 40536-0596, USA. ² Department of Molecular and Cellular Biochemistry and Sanders-Brown Center on Aging, University of Kentucky, Lexington, KY, 40536-0230, USA.

* Correspondence to: sylviegtsodikova@uky.edu

Materials and instrumentation. All chemicals were purchased from Sigma Aldrich (St. Louis, MO), Alfa Aesar (Ward Hill, MA), and AK scientific (Union City, CA) and used without further purification. Chemical reactions were monitored by thin layer chromatography (TLC) using Merck, Silica gel 60 F₂₅₀ plates. Visualization was achieved using UV light and KMnO₄ stain (1.5 g KMnO₄, 10 g K₂CO₃, 1.25 mL 10% NaOH, 200 mL H₂O). ¹H and ¹³C NMR spectra were recorded at 400 and 100 MHz, respectively, on a Varian 400 MHz spectrometer, using the indicated solvents. Chemical shift (δ) is given in parts per million (ppm). Coupling constants (J) are given in hertz (Hz), and conventional abbreviations used for signal shape are as follows: s =singlet; d = doublet; t = triplet; m = multiplet; dd = doublet of doublets; ddd = doublet of doublet of doublets; br s = broad singlet; dt = doublet of triplets. Liquid chromatography-mass spectrometry (LCMS) was carried out using an Agilent 1200 series Quaternary LC system equipped with a diode array detector, and Eclipse XDB- C_{18} column (250 mm x 4.6 mm, 5 µm), and an Agilent 6120 Quadrupole MSD mass spectrometer (Agilent Technologies, Santa Clara, CA). LCMS M + H signals were consistent with the expected molecular weights for all of the reported compounds. Cu²⁺ binding studies by UV-Vis were performed on a SpectraMax M5 multiplate reader (Molecular Devices, Sunnyvale, CA) using quartz cuvettes. N-biotinyl A β_{1-42} (bioA β_{42}) was purchased from Anaspec (Fremont, CA). ELISA plates (Costar 9018), NeutrAvidinTM (Promega), adhesive film (NUNC), polypropylene 96-well plates (Costar 3365), and polypropylene Eppendorf tubes (Fisher 02-681-248) used for $bioA\beta_{42}$ oligomer assembly and dissociation assays were all purchased from Fisher Scientific (Pittsburgh, PA). Plates were washed on a BioTek ELx50 plate washer (Biotek (Winooski, VT)) and absorbance was read on a Biotek HT Synergy plate reader. Elemental analysis of all final compounds were performed at Atlantic Microlab, Inc. (Norcross, GA).

General procedure A for the synthesis of 2-acetylpyridines. Methylmagnesium bromide (3.0 eq, 3 M in diethyl ether) was slowly added to a cooled (-20 °C) solution of 2-pyridinecarbonitrile (1.0 eq) in dry THF (5 mL), and the mixture was stirred between -20 °C and -10 °C until completion of the reaction (1-3 h). The reaction mixture was then cooled down to -40 °C, and aq. HCl was slowly added until the solution stopped fizzing. 5 min later, the solution was poured into sodium phosphate buffer (10 mL, 500 mM, pH 7) and extracted with Et₂O (5x10 mL). The combined organic layers were washed with H₂O (10 mL) and brine (10 mL), dried over MgSO₄, filtered, and concentrated under reduced pressure to give a crude product, which was purified by column chromatography (SiO₂, Hexanes:EtOAc/9:1).

Synthesis of 2-acetyl-6-methylpyridine (1b). Following general procedure A, the known compound $\mathbf{1b}^1$ (R_f 0.42, Hexanes:EtOAc/9:1) was obtained from 6-methyl-2pyridinecarbonitrile (400 mg, 3.39 mmol) and methylmagnesium bromide (3.4 mL, 10.16 mmol) as a light yellow oil (83% yield): ¹H NMR (400 MHz, CDCl₃, which matches the lit.¹) δ 7.80 (d, J = 7.2 Hz, 1H, aromatic), 7.66 (t, J = 7.2 Hz, 1H, aromatic), 7.28 (d, J = 7.2 1H, aromatic), 2.68 (s, 3H, CH₃), 2.58 (s, 3H, CH₃).



Synthesis of 2-acetyl-5-methylpyridine (1c). Following general procedure A, the known compound $1c^2$ (R_f 0.32, Hexanes:EtOAc/4:1) was obtained from 5methylpyridine-2-carbonitrile (400 mg, 3.39 mmol) and methylmagnesium bromide (3.4 mL, 10.16 mmol) as a light yellow oil (363 mg, 79% yield): ¹H NMR (400 MHz, CDCl₃, which matches the lit.²) δ 8.47 (m, 1H, aromatic), 7.92 (d, J = 8.0 Hz, 1H, aromatic), 7.61-7.58 (m, 1H, aromatic), 2.68 (s, 3H, CH₃), 2.39 (s, 3H, CH₃).



Synthesis of 2-acetyl-3-methylpyridine (1e). Following general procedure A, the known compound $1e^3$ (R_f 0.33, Hexanes:EtOAc/9:1) was obtained from 3methylpicolinonitrile (400 mg, 3.39 mmol) and methylmagnesium bromide (3.4 mL, 10.16 mmol) as a light yellow oil (310 mg, 68% yield): ¹H NMR (400 MHz, CDCl₃, which matches the lit.³) δ 8.47 (d, J = 4.4 Hz, 1H, aromatic), 7.54 (d, J = 7.6 Hz, 1H, aromatic), 7.28 $(dd, J_1 = 7.6 Hz, J_2 = 4.4 Hz, 1H, aromatic), 2.67 (s, 3H, CH_3), 2.54 (s, 3H, CH_3).$

Synthesis of 2-acetyl-5-bromopyridine (1g). Following general procedure A, the known compound $1g^4$ (R_f 0.49, Hexanes:EtOAc/9:1) was obtained from 5-bromo-2pyridinecarbonitrile (320 mg, 1.75 mmol) and methylmagnesium bromide (1.75 mL, 5.25 mmol) as a white solid (205 mg, 59% yield): ¹H NMR (400 MHz, CDCl₃, which matches the lit.⁴) δ 8.71 (d, J = 2.0 Hz, 1H, aromatic), 7.95 (dd, J₁ = 8.0 Hz, J₂ = 2.0 Hz, 1H, aromatic), 7.91 (d, J = 8.0 Hz, 1H, aromatic), 2.68 (s, 3H, CH₃).

Synthesis of 2-acetyl-4-bromopyridine (1h). Following general procedure A, 1h (R_f 0.40, Hexanes:EtOAc/9:1) was obtained from 4-bromopicolinonitrile (320 mg, 1.75 mmol) and methylmagnesium bromide (1.75 mL, 5.25 mmol) as an off-white solid (205 mg, 59% yield): ¹H NMR (400 MHz, CDCl₃, Fig. S1) δ 8.48 (d, J = 4.8 Hz, 1H, aromatic), 8.18 (d, J = 2.0 Hz, 1H, aromatic), 7.62 (dd, $J_1 = 4.8$ Hz, $J_2 = 2.0$ Hz, 1H, aromatic), 2.69 (s, 3H, CH₃); ¹³C NMR (100 MHz, CDCl₃, Fig. S2) δ 198.8, 154.4, 149.7, 134.0, 130.1, 125.2, 25.8; m/z calcd for C₇H₆BrNO 199.0; found 200.0 [M+H]⁺.

Synthesis of 2-acetyl-3-bromopyridine (1i). Following general procedure A, 1i (R_f 0.22, Hexanes:EtOAc/9:1) was obtained from 3-bromopyridine-2-carbonitrile (320 mg, 1.75 mmol) and methylmagnesium bromide (1.75 mL, 5.25 mmol) as a pale yellow liquid (231 mg, 66% yield): ¹H NMR (400 MHz, CDCl₃, Fig. S3) δ 8.55 (dd, J_1 = 4.8 Hz, $J_2 = 1.6$ Hz, 1H, aromatic), 7.97 (dd, $J_1 = 8.0$ Hz, $J_2 = 1.6$ Hz, 1H, aromatic), 7.25 (dd, $J_1 = 8.0$ Hz, $J_2 = 4.8$ Hz, 1H, aromatic), 2.67 (s, 3H, CH₃); ¹³C NMR (100 MHz, CDCl₃, Fig. S4) δ 199.5, 152.9, 147.2, 142.4, 126.5, 117.3, 28.0; *m/z* calcd for C₇H₆BrNO 199.0; found 200.0 [M+H]⁺.

General procedure B for the synthesis of chalcone derivatives 3a-i. A mixture of 2acetylpyridine (1a-i, 1.0 eq.) and 4-(dimethylamino)benzaldehyde (2, 1.0 eq.) in EtOH (5 mL) was treated with 20% aq. KOH (3 mL) and the resulting solution was stirred at rt for 3 h. Upon completion of the reaction, H₂O (5 mL) was added and the solid residues that formed were filtered out, rinsed with H₂O and ice-cold EtOH, and recrystallized from CH₂Cl₂/hexanes.



Synthesis of (*E*)-3-(4-(dimethylamino)phenyl)-1-(pyridin-2-yl)prop-2en-1-one (3a). Following general procedure B, the known compound $3a^5$ ($R_f 0.33$, Hexanes:EtOAc/3:1) was obtained from 1a (242 mg, 2.0 mmol) and 2 (298 mg, 2.0 mmol) as bright orange crystals (318 mg, 63% yield): ¹H

NMR (400 MHz, CDCl₃, which matches the lit.⁵) δ 8.72 (ddd, $J_1 = 4.8$ Hz, $J_2 = 1.6$ Hz, $J_3 = 0.8$ Hz, 1H, aromatic), 8.17 (dt, $J_1 = 8.0$ Hz, $J_2 = 1.2$ Hz, 1H, aromatic), 8.06 (d, J = 15.6 Hz, 1H, HC=CH-Ph), 7.92 (d, J = 15.6 Hz, 1H, HC=CH-Ph), 7.84 (td, $J_1 = 8.0$ Hz, $J_2 = 1.6$ Hz, 1H, aromatic), 7.63 (dt, $J_1 = 9.2$ Hz, $J_2 = 2.0$ Hz, 2H, aromatic), 7.44 (ddd, $J_1 = 7.6$ Hz, $J_2 = 4.8$ Hz, $J_3 = 1.2$ Hz, 1H, aromatic), 6.68 (dt, $J_1 = 9.2$ Hz, $J_2 = 2.0$ Hz, 2H, aromatic), 3.03 (s, 6H, N(CH₃)₂); Elemental analysis calcd. for C₁₆H₁₆N₂O: C, 76.16; H, 6.39; N, 11.10. Found: C, 75.88; H, 6.48; N, 11.02 (Fig. S5).



Synthesis of (*E*)-3-(4-(dimethylamino)phenyl)-1-(6-methylpyridin-2yl)prop-2-en-1-one (3b). Following general procedure B, 3b (R_f 0.17, Hexanes:EtOAc/9:1) was obtained from 1b (195 mg, 1.44 mmol) and 2 (215 mg, 1.44 mmol) as a dark yellow powder (212 mg, 55% yield): ¹H

NMR (400 MHz, CDCl₃, Fig. S6) δ 8.08 (d, J = 15.6 Hz, 1H, HC=CH-Ph), 7.96 (dd, $J_1 = 7.6$ Hz, $J_2 = 1.2$ Hz, 1H, aromatic), 7.89 (d, J = 16.4 Hz, 1H, HC=CH-Ph), 7.71 (t, J = 7.6 Hz, 1H, aromatic), 7.62 (dt, $J_1 = 9.2$ Hz, $J_2 = 2.0$ Hz, 2H, aromatic), 7.28 (dd, $J_1 = 7.6$ Hz, $J_2 = 1.2$ Hz, 1H, aromatic), 6.69 (dt, $J_1 = 9.2$ Hz, $J_2 = 2.0$ Hz, 2H, aromatic), 3.03 (s, 6H, N(CH₃)₂), 2.65 (s, 3H, CH₃); ¹³C NMR (100 MHz, CDCl₃, Fig. S7) δ 189.6, 157.6, 154.5, 152.0, 145.6, 136.9, 130.8 (2 carbons), 126.0, 123.2, 119.8, 115.9, 111.7 (2 carbons), 40.1 (2 carbons), 24.5; *m/z* calcd for C₁₇H₁₈N₂O 266.1; found 267.1 [M+H]⁺; Elemental analysis calcd. for C₁₇H₁₈N₂O: C, 76.66; H, 6.81; N, 10.52. Found: C, 75.18; H, 6.83; N, 10.36 (Fig. S8).



Synthesis of (*E*)-3-(4-(dimethylamino)phenyl)-1-(5-methylpyridin-2-yl)prop-2-en-1-one (3c). Following general procedure B, 3c (R_f 0.28, Hexanes:EtOAc/4:1) was obtained from 1c (195 mg, 1.44 mmol) and 2 (215 mg, 1.44 mmol) as an orange solid (312 mg, 81% yield): ¹H NMR

(400 MHz, CDCl₃, Fig. S9) δ 8.54 (m, 1H, aromatic), 8.09 (d, *J* = 8.0 Hz, 1H, aromatic), 8.07 (d, *J* = 15.6 Hz, HC=C<u>H</u>-Ph), 7.91 (d, *J* = 15.6 Hz, 1H, <u>H</u>C=CH-Ph), 7.66-7.62 (m, 3H, aromatic), 6.69 (dt, *J*₁ = 8.8 Hz, *J*₂ = 2.0 Hz, 2H, aromatic), 3.04 (s, 6H, N(C<u>H</u>₃)₂), 2.43 (s, 3H, Ph-C<u>H</u>₃); ¹³C NMR (100 MHz, CDCl₃, Fig. S10) δ 189.1, 152.7, 152.0, 149.1, 145.6, 137.3, 136.7, 130.8 (2 carbons), 123.1, 122.5, 115.7, 111.7 (2 carbons), 40.1 (2 carbons), 18.7; *m/z* calcd for C₁₇H₁₈N₂O 266.1; found 267.3 [M+H]⁺; Elemental analysis calcd. for C₁₇H₁₈N₂O: C, 76.66; H, 6.81; N, 10.52. Found: C, 76.37; H, 6.81; N, 10.43 (Fig. S11).



Synthesis of (*E*)-3-(4-(dimethylamino)phenyl)-1-(4-methylpyridin-2-yl)prop-2-en-1-one (3d). Following general procedure B, 3d (R_f 0.36, Hexanes:EtOAc/3:1) was obtained from 1d (270 mg, 2.0 mmol) and 2 (298

mg, 2.0 mmol) as a bright yellow solid (331 mg, 62% yield): ¹H NMR (400 MHz, CDCl₃, Fig. S12) δ 8.57 (d, J = 4.8 Hz, 1H, aromatic), 8.04 (d, J = 15.6 Hz, 1H, HC=CH-Ph), 7.99 (m, 1H, aromatic), 7.90 (d, J = 15.6 Hz, 1H, HC=CH-Ph), 7.62 (dt, $J_1 = 9.2$ Hz, $J_2 = 2.0$ Hz, 2H, aromatic), 7.26 (dd, $J_1 = 2.0$ Hz, $J_2 = 0.8$ Hz, 1H, aromatic), 6.67 (dt, $J_1 = 9.2$ Hz, $J_2 = 2.0$ Hz, 2H, aromatic), 3.03 (s, 6H, N(CH₃)₂), 2.43 (s, 3H, Ph-CH₃); ¹³C NMR (100 MHz, CDCl₃, Fig. S13) δ 189.5, 154.8, 152.0, 148.5, 148.2, 145.8, 130.9 (2 carbons), 127.3, 123.6, 123.1, 115.7, 111.7 (2 carbons), 40.1 (2 carbons), 21.1; *m/z* calcd for C₁₇H₁₈N₂O 266.1; found 267.1 [M+H]⁺; Elemental analysis calcd. for C₁₇H₁₈N₂O: C, 76.66; H, 6.81; N, 10.52. Found: C, 76.38; H, 6.79; N, 10.45 (Fig. S14).



Synthesis of (*E*)-3-(4-(dimethylamino)phenyl)-1-(3-methylpyridin-2yl)prop-2-en-1-one (3e). Following general procedure B, 3e (R_f 0.37, Hexanes:EtOAc/3:1) was obtained from 1e (195 mg, 1.44 mmol) and 2 (215 mg, 1.44 mmol) as a bright orange powder (153 mg, 57% yield): ¹H NMR

(400 MHz, CDCl₃, Fig. S15) δ 8.52 (ddd, $J_1 = 4.8$ Hz, $J_2 = 1.6$ Hz, $J_3 = 0.8$ Hz, 1H, aromatic), 7.68 (d, J = 16.0 Hz, 1H, HC=CH-Ph), 7.58 (ddd, $J_1 = 8.0$ Hz, $J_2 = 1.6$ Hz, $J_3 = 0.8$ Hz, 1H, aromatic), 7.57 (d, J = 16.0 Hz, 1H, HC=CH-Ph), 7.53 (dt, $J_1 = 9.2$ Hz, $J_2 = 2.0$ Hz, 2H, aromatic), 7.29 (dd, $J_1 = 8.0$ Hz, $J_2 = 4.8$ Hz, 1H, aromatic), 6.65 (dt, $J_1 = 9.2$ Hz, $J_2 = 2.0$ Hz, 2H, aromatic), 3.02 (s, 6H, N(CH₃)₂), 2.55 (s, 3H, CH₃); ¹³C NMR (100 MHz, CDCl₃, Fig. S16) δ 192.6, 154.4, 152.0, 146.0, 145.9, 139.6, 133.9, 130.6 (2 carbons), 125.0, 122.9, 119.2, 111.7 (2 carbons), 40.1 (2 carbons), 19.7; *m*/z calcd for C₁₇H₁₈N₂O 266.1; found 267.3 [M+H]⁺; Elemental analysis calcd. for C₁₇H₁₈N₂O: C, 76.66; H, 6.81; N, 10.52. Found: C, 76.44; H, 6.82; N, 10.47 (Fig. S17).



Synthesis of (*E*)-1-(6-bromopyridin-2-yl)-3-(4-(dimethylamino)phenyl)prop-2-en-1-one (3f). Following general procedure B, 3f (R_f 0.42, Hexanes:EtOAc/3:1) was obtained from 1f (201 mg, 1.0 mmol) and 2 (150 mg, 1.0 mmol) as orange crystals (193

mg, 58% yield): ¹H NMR (400 MHz, CDCl₃, Fig. S18) δ 8.12 (dd, $J_1 = 8.0$ Hz, $J_2 = 0.8$ Hz, 1H, aromatic), 7.97 (d, J = 15.6 Hz, 1H, HC=C<u>H</u>-Ph), 7.91 (d, J = 15.6 Hz, 1H, <u>H</u>C=CH-Ph), 7.69 (t, J = 7.2 Hz, 1H, aromatic), 7.624 (dt, $J_1 = 8.8$ Hz, $J_2 = 2.0$ Hz, 2H, aromatic), 7.617 (dd, $J_1 = 8.0$ Hz, $J_2 = 0.8$ Hz, 1H, aromatic), 6.68 (dt, $J_1 = 8.8$ Hz, $J_2 = 2.0$ Hz, 2H, aromatic), 3.04 (s, 6H, N(C<u>H</u>₃)₂); ¹³C NMR (100 MHz, CDCl₃, Fig. S19) δ 187.6, 156.2, 152.5, 147.0, 141.3, 139.4, 131.4 (2 carbons), 131.2, 123.1, 121.8, 115.1, 111.9 (2 carbons), 40.3 (2 carbons); *m/z* calcd for C₁₆H₁₅BrN₂O 330.0; found 331.1 [M+H]⁺; Elemental analysis calcd. for C₁₆H₁₅BrN₂O: C, 58.02; H, 4.57; Br, 24.12; N, 8.64. Found: C, 56.93; H, 4.62; Br, 23.43; N, 8.10 (Fig. S20).



Synthesis of (*E*)-1-(5-bromopyridin-2-yl)-3-(4-(dimethylamino)phenyl)prop-2-en-1-one (3g). Following general procedure B, 3g (R_f 0.47, Hexanes:EtOAc/5:1) was obtained from 1g (195 mg, 0.97 mmol) and 2 (145 mg, 0.97 mmol) as a bright orange

powder (192 mg, 59% yield): ¹H NMR (400 MHz, CDCl₃, Fig. S21) δ 8.75 (d, J = 2.0 Hz, 1H, aromatic), 8.06 (d, J = 8.0 Hz, 1H, aromatic), 7.99 (d, J = 16.0 Hz, 1H, HC=CH-Ph), 7.96 (dd, $J_1 = 8.0$ Hz, $J_2 = 2.0$ Hz, 1H, aromatic), 7.91 (d, J = 16.0 Hz, 1H, HC=CH-Ph), 7.61 (dt, $J_1 = 9.2$ Hz, $J_2 = 2.0$ Hz, 2H, aromatic), 6.68 (dt, $J_1 = 9.2$ Hz, $J_2 = 2.0$ Hz, 2H, aromatic), 3.04 (s, 6H, N(CH₃)₂); ¹³C NMR (100 MHz, CDCl₃, Fig. S22) δ 188.1, 153.3, 152.2, 149.8, 146.4, 139.6,

131.0 (2 carbons), 124.4, 124.1, 122.9, 114.9, 111.7 (2 carbons), 40.1 (2 carbons); m/z calcd for C₁₆H₁₅BrN₂O 330.0; found 331.1 [M+H]⁺; Elemental analysis calcd. for C₁₆H₁₅BrN₂O: C, 58.02; H, 4.57; Br, 24.12; N, 8.64. Found: C, 57.51; H, 4.53; Br, 23.73; N, 8.28 (Fig. S23).



Synthesis of (*E*)-1-(4-bromopyridin-2-yl)-3-(4-(dimethylamino)phenyl)prop-2-en-1-one (3h). Following general procedure B, 3h (R_f 0.45, Hexanes:EtOAc/5:1) was obtained from 1h (195 mg, 0.97 mmol) and 2 (145 mg, 0.97 mmol) as a bright orange powder (230

mg, 71% yield): ¹H NMR (400 MHz, CDCl₃, Fig. S24) δ 8.52 (d, *J* = 4.8 Hz, 1H, aromatic), 8.33 (d, *J* = 2.0 Hz, 1H, aromatic), 7.99 (d, *J* = 15.6 Hz, 1H, HC=CH-Ph), 7.92 (d, *J* = 15.6 Hz, 1H, HC=CH-Ph), 7.61 (dt, *J*₁ = 8.8 Hz, *J*₂ = 2.0 Hz, 2H, aromatic), 7.60 (dd, *J*₁ = 4.8 Hz, *J*₂ = 2.0 Hz, 1H, aromatic), 6.67 (dt, *J*₁ = 8.8 Hz, *J*₂ = 2.0 Hz, 2H, aromatic), 3.04 (s, 6H, N(CH₃)₂); ¹³C NMR (100 MHz, CDCl₃, Fig. S25) δ 187.7, 156.0, 152.3, 149.4, 146.7, 134.0, 131.1 (2 carbons), 129.4, 126.2, 122.8, 114.9, 111.7 (2 carbons), 40.1 (2 carbons); *m*/z calcd for C₁₆H₁₅BrN₂O 330.0; found 331.1 [M+H]⁺; Elemental analysis calcd. for C₁₆H₁₅BrN₂O: C, 58.02; H, 4.57; Br, 24.12; N, 8.64. Found: C, 57.50; H, 4.64; Br, 23.69; N, 8.26 (Fig. S26).



Synthesis of (*E*)-1-(3-bromopyridin-2-yl)-3-(4-(dimethylamino)phenyl)prop-2-en-1-one (3i). Following general procedure B, 3i (R_f 0.21, Hexanes:EtOAc/3:1) was obtained from 1i (195 mg, 0.97 mmol) and 2 (145 mg, 0.97 mmol) as a bright orange foam (137

mg, 42% yield). *Note*: In this case, recrystallization was not successful. Therefore **3i** was purified by column chromatography (SiO₂, pure Hexanes to Hexanes:EtOAc/2:1, R_f 0.18, Hexanes:EtOAc/5:1): ¹H NMR (400 MHz, CDCl₃, Fig. S27) δ 8.60 (dd, J_1 = 4.8 Hz, J_2 = 1.6 Hz, 1H, aromatic), 7.97 (dd, J_1 = 8.4 Hz, J_2 = 1.6 Hz, 1H, aromatic), 7.53 (d, J = 16.0 Hz, 1H, HC=C<u>H</u>-Ph), 7.48 (dt, J_1 = 8.8 Hz, J_2 = 2.0 Hz, 2H, aromatic), 7.25 (dd, J_1 = 8.4 Hz, J_2 = 4.8 Hz, 1H, aromatic), 7.15 (d, J = 16.0 Hz, 1H, <u>H</u>C=CH-Ph), 6.64 (dt, J_1 = 8.8 Hz, J_2 = 2.0 Hz, 2H, aromatic), 3.02 (s, 6H, N(C<u>H</u>₃)₂); ¹³C NMR (100 MHz, CDCl₃, Fig. S28) δ 191.3, 155.8, 152.3, 148.1, 147.3, 141.6, 130.8 (2 carbons), 125.5, 122.3, 118.9, 117.9, 111.7 (2 carbons), 40.1 (2 carbons); *m/z* calcd for C₁₆H₁₅BrN₂O 330.0; found 331.1 [M+H]⁺; Elemental analysis calcd. for C₁₆H₁₅BrN₂O: C, 58.02; H, 4.57; Br, 24.12; N, 8.64. Found: C, 58.02; H, 4.63; Br, 24.28; N, 8.43 (Fig. S29).

Synthesis of 2'-hydroxy-3'-methylacetophenone (4b). Methyllithium (8.4 mL, 1.6 M in Et₂O) was slowly added to a cooled (0 °C) solution of 3-methylsalicylic acid (600 mg, 3.94 mmol) in dry THF (10 mL), and the mixture was stirred for 6 h at rt. A saturated aq. solution of NH₄Cl was added and the mixture was filtered through a bed of celite, eluting with EtOAc. The organic layer was separated and washed with brine, dried over MgSO₄, filtered, and concentrated under reduced pressure. Following purification by column chromatography (SiO₂, Hexanes:EtOAc/9:1), the known compound **4b**⁶ (R_f 0.74, Hexanes:EtOAc/9:1) was obtained as a light yellow oil (403 mg, 68% yield): ¹H NMR (400 MHz, CDCl₃, which matches the lit.⁶) δ 12.55 (s, 1H, O<u>H</u>), 7.58 (dd, J_1 = 8.0 Hz, J_2 = 1.2 Hz, 1H, aromatic), 7.33 (br dd, J_1 = 8.0 Hz, J_2 = 1.2 Hz, 1H, aromatic), 6.79 (t, J = 8.0 Hz, 1H, aromatic), 2.62 (s, 3H, C<u>H₃</u>), 2.25 (s, 3H, C<u>H₃</u>).

OH O Synthesis of 2'-hydroxy-6'-methylacetophenone (4e). A solution of

methylmagnesium bromide (5.5 mL, 3.0 M in Et₂O) and Et₃N (1.9 mL, 13.3 mmol) in anhydrous toluene (5 mL) was cooled down to 0 °C. A solution of ethyl 2-hydroxy-6-methylbenzoate (600 mg, 3.33 mmol) in anhydrous toluene (5 mL) was slowly added and the resulting mixture was stirred overnight till rt. A saturated aq. solution of NH₄Cl was added and the mixture was filtered through a bed of celite, eluting with EtOAc. The organic layer was separated and washed with brine, dried over MgSO₄, filtered, and concentrated under reduced pressure. Following purification by column chromatography (SiO₂, Hexanes:EtOAc/9:1), the known compound **4e**⁷ (R_f 0.32, Hexanes:EtOAc/9:1) was obtained as a yellow oil (379 mg, 76% yield): ¹H NMR (400 MHz, CDCl₃, which matches the lit.⁷) δ 12.30 (s, 1H, O<u>H</u>), 7.28 (t, *J* = 7.2 Hz, 1H, aromatic), 6.84 (d, *J* = 7.2 Hz, 1H, aromatic), 6.73 (d, *J* = 7.2 Hz, 1H, aromatic), 2.67 (s, 3H, C<u>H</u>₃), 2.60 (s, 3H, C<u>H</u>₃).

Br $\stackrel{OH}{\longrightarrow}$ Synthesis of 3'-bromo-2'-hydroxyacetophenone (4f). A solution of 2'hydroxyacetophenone (1.77 mL, 14.7 mmol) and *i*-Pr₂NH (0.21 mL, 1.47 mmol) in carbon disulfide (25 mL) was treated with NBS (2.61 g, 14.7 mmol) at 0 °C. The mixture was then stirred for 1.5 h at rt. H₂O was added and the mixture was extracted three times with EtOAc. The combined organic layers were washed with saturated aq. NaHCO₃, H₂O, and brine, dried over MgSO₄, filtered, and concentrated under reduced pressure. Following purification by column chromatography (SiO₂, Hexanes:EtOAc/9:1), the known compound **4f**⁸ (R_f 0.36, Hexanes:EtOAc/9:1) was obtained as a brown solid (292 mg, 9% yield): ¹H NMR (400 MHz, CDCl₃, which matches the lit.⁸) δ 12.97 (s, 1H, O<u>H</u>), 7.62 (d, *J* = 7.6 Hz, 1H, aromatic), 7.74 (dd, *J*₁ = 8.0 Hz, *J*₂ = 1.6 Hz, 1H, aromatic), 7.72 (dd, *J*₁ = 8.0 Hz, *J*₂ = 1.6 Hz, 1H, aromatic), 6.82 (t, *J* = 8.0 Hz, 1H, aromatic), 2.66 (s, 3H, C<u>H</u>₃).

Synthesis of 2'-amino-3'-methylacetophenone (5b). Following the procedure for the synthesis of 4b, the known compound 5b⁹ (R_f 0.66, Hexanes:EtOAc/3:1) was obtained from 2-amino-3-methylbenzoic acid (302 mg, 2.0 mmol) and methyllithium (3.7 mL, 6.0 mmol) as a yellow solid (139 mg, 47% yield): ¹H NMR (400 MHz, CDCl₃, which matches the lit.⁹) δ 7.62 (d, J = 7.6 Hz, 1H, aromatic), 7.18 (d, J = 7.6 Hz, 1H, aromatic), 6.58 (t, J = 7.6 Hz, 1H, aromatic), 2.57 (s, 3H, CH₃), 2.15 (s, 3H, CH₃).

General procedure C for the synthesis of chalcone derivatives 6a-h, 7a, 7b, and 7h. A mixture of 2'-hydroxyacetophenone (4a-h)/2'-aminoacetophenone (5a, 5b, 5h) (1.0 eq.), 4-(dimethylamino)benzaldehyde (2, 1.0 eq.), and NaOH (5.0 eq.) in EtOH (2.5 mL) was stirred at rt overnight. Upon completion of the reaction, EtOH was evaporated and the residue was neutralized with 1 N aq. HCl. The resulting mixture was stirred at rt for 10 min and the solid residues that formed were filtered out, rinsed with H₂O and ice-cold EtOH, and recrystallized from EtOH.



Synthesisof(E)-3-(4-(dimethylamino)phenyl)-1-(2-hydroxyphenyl)prop-2-en-1-one(6a). Following general procedure C, theknown compound $6a^{10}$ (R_f 0.56, Hexanes:EtOAc/2:1) was obtained from 4a(272 mg, 2.0 mmol) and 2 (298 mg, 2.0 mmol) as dark purple crystals (143

mg, 27% yield): ¹H NMR (400 MHz, CDCl₃, which matches the lit.¹⁰) δ 13.17 (s, 1H, O<u>H</u>), 7.93 (dd, $J_1 = 8.4$ Hz, $J_2 = 1.6$ Hz, 1H), 7.92 (d, J = 15.6 Hz, 1H, HC=C<u>H</u>-Ph), 7.58 (dt, $J_1 = 9.2$ Hz, $J_2 = 2.0$ Hz, 2H, aromatic), 7.46 (d, J = 15.6 Hz, 1H, <u>H</u>C=CH-Ph), 7.46 (ddd, $J_1 = 8.4$ Hz, $J_2 = 1.6$ Hz, 1H, $J_2 = 1.6$ Hz,

7.2 Hz, $J_3 = 1.2$ Hz, 1H, aromatic), 7.01 (dd, $J_1 = 8.4$ Hz, $J_2 = 1.2$ Hz, 1H, aromatic), 6.92 (ddd, $J_1 = 8.4$ Hz, $J_2 = 7.2$ Hz, $J_3 = 1.2$ Hz, 1H, aromatic), 6.71 (dt, $J_1 = 9.2$ Hz, $J_2 = 2.0$ Hz, 2H, aromatic), 3.07 (s, 6H, N(CH₃)₂); ¹³C NMR (CDCl₃, 100 MHz) δ 193.5, 163.5, 152.3, 146.5, 135.6, 130.8 (2 carbons), 129.3, 122.3, 120.4, 118.55, 118.46, 114.3, 111.8 (2 carbons), 40.1 (2 carbons); *m*/*z* calcd for C₁₇H₁₇NO₂ 267.1; found 268.3 [M+H]⁺; Elemental analysis calcd. for C₁₇H₁₇NO₂: C, 76.38; H, 6.41; N, 5.24. Found: C, 76.20; H, 6.42; N, 5.24 (Fig. S30).



Synthesis of (*E*)-3-(4-(dimethylamino)phenyl)-1-(2-hydroxy-3methylphenyl)prop-2-en-1-one (6b). Following general procedure C, 6b (R_f 0.50, Hexanes:EtOAc/4:1) was obtained from 4b (150 mg, 1.0 mmol) and 2 (149 mg, 1.0 mmol) as red crystals (239 mg, 85% yield): ¹H NMR

(400 MHz, CDCl₃, Fig. S31) δ 13.48 (s, 1H, O<u>H</u>), 7.90 (d, J = 15.6 Hz, 1H, HC=C<u>H</u>-Ph), 7.77 (d, J = 8.0 Hz, 1H, aromatic), 7.57 (dt, $J_1 = 9.2$ Hz, $J_2 = 2.8$ Hz, 2H, aromatic), 7.47 (d, J = 15.6 Hz, 1H, <u>HC</u>=CH-Ph), 7.32 (d, J = 8.0 Hz, 1H, aromatic), 6.81 (t, J = 8.0 Hz, 1H, aromatic), 6.72 (d, J = 9.2 Hz, 2H, aromatic), 3.05 (s, 6H, N(C<u>H₃)₂), 2.28 (s, 3H, C<u>H₃)</u>; ¹³C NMR (100 MHz, CDCl₃, Fig. S32) δ 193.7, 162.0, 152.1, 146.1, 136.4, 130.7 (2 carbons), 127.4, 127.0, 122.8, 119.6, 117.8 (2 carbons), 114.9, 112.0, 40.2 (2 carbons), 15.6; *m/z* calcd for C₁₈H₁₉NO₂ 281.1; found 282.1 [M+H]⁺; Elemental analysis calcd. for C₁₈H₁₉NO₂: C, 76.84; H, 6.81; N, 4.98. Found: C, 76.79; H, 6.76; N, 5.01 (Fig. S33).</u>



Synthesis of (*E*)-3-(4-(dimethylamino)phenyl)-1-(2-hydroxy-4methylphenyl)prop-2-en-1-one (6c). Following general procedure C, the known compound $6c^{11}$ (R_f 0.37, Hexanes:EtOAc/5:1) was obtained from 4c (150 mg, 1.0 mmol) and 2 (149 mg, 1.0 mmol) as a dark red powder

(168 mg, 60% yield): ¹H NMR (400 MHz, CDCl₃) δ 13.21 (s, 1H, O<u>H</u>), 7.88 (d, *J* = 15.2 Hz, 1H, HC=C<u>H</u>-Ph), 7.79 (d, *J* = 8.0 Hz, 1H, aromatic), 7.56 (dt, *J₁* = 9.2 Hz, *J₂* = 2.0 Hz, 2H, aromatic), 7.42 (d, *J* = 15.2 Hz, 1H, <u>HC</u>=CH-Ph), 6.80 (s, 1H, aromatic), 6.74-6.68 (m, 3H, aromatic), 6.69 (dt, *J₁* = 9.2 Hz, *J₂* = 2.0 Hz, 2H, aromatic), 3.05 (s, 6H, N(C<u>H</u>₃)₂), 2.35 (s, 3H, Ph-C<u>H</u>₃); ¹³C NMR (100 MHz, CDCl₃) δ 193.0, 163.6, 152.2, 147.2, 146.0, 130.7 (2 carbons), 129.2, 122.5, 119.8, 118.5, 118.1, 114.5, 111.8 (2 carbons), 40.1 (2 carbons), 21.9; *m/z* calcd for C₁₈H₁₉NO₂ 281.1; found 282.1 [M+H]⁺; Elemental analysis calcd. for C₁₈H₁₉NO₂: C, 76.84; H, 6.81; N, 4.98. Found: C, 76.33; H, 6.74; N, 4.94 (Fig. S34).



Synthesis of (*E*)-3-(4-(dimethylamino)phenyl)-1-(2-hydroxy-5methylphenyl)prop-2-en-1-one (6d). Following general procedure C, the known compound $6d^{12}$ (R_f 0.39, Hexanes:EtOAc/5:1) was obtained from 4d (150 mg, 1.0 mmol) and 2 (149 mg, 1.0 mmol) as dark purple crystals (218

mg, 76% yield): ¹H NMR (400 MHz, CDCl₃) δ 12.97 (s, 1H, O<u>H</u>), 7.89 (d, J = 15.2 Hz, 1H, HC=C<u>H</u>-Ph), 7.68 (d, J = 1.6 Hz, 1H, aromatic), 7.57 (dt, $J_1 = 9.2$ Hz, $J_2 = 2.0$ Hz, 2H, aromatic), 7.44 (d, J = 15.2 Hz, 1H, <u>HC</u>=CH-Ph), 7.26 (dd, $J_1 = 8.4$ Hz, $J_2 = 2.0$ Hz, 1H, aromatic), 6.90 (d, J = 8.4 Hz, 1H, aromatic), 6.69 (dt, $J_1 = 9.2$ Hz, $J_2 = 2.0$ Hz, 2H, aromatic), 3.05 (s, 6H, N(C<u>H</u>₃)₂), 2.34 (s, 3H, C<u>H</u>₃); ¹³C NMR (100 MHz, CDCl₃) δ 193.4, 161.4, 152.3, 146.3, 136.7, 130.8 (2 carbons), 129.1, 127.5, 122.4, 120.0, 118.2, 114.5, 111.8 (2 carbons), 40.1 (2 carbons), 20.6; *m*/z calcd for C₁₈H₁₉NO₂ 281.1; found 282.1 [M+H]⁺; Elemental analysis calcd. for C₁₈H₁₉NO₂: C, 76.84; H, 6.81; N, 4.98. Found: C, 76.83; H, 6.93; N, 5.00 (Fig. S35).



Synthesis of (*E*)-3-(4-(dimethylamino)phenyl)-1-(2-hydroxy-6methylphenyl)prop-2-en-1-one (6e). Following general procedure C, 6e (R_f 0.22, Hexanes:EtOAc/4:1) was obtained from 4e (150 mg, 1.0 mmol) and 2 (149 mg, 1.0 mmol) as a red powder (101 mg, 36% yield): ¹H NMR

(400 MHz, CDCl₃, Fig. S36) δ 10.72 (s, 1H, O<u>H</u>), 7.72 (d, J = 15.6 Hz, 1H, HC=C<u>H</u>-Ph), 7.50 (dt, $J_1 = 8.4$ Hz, $J_2 = 2.8$ Hz, 2H, aromatic), 7.24 (d, J = 8.4 Hz, 1H, aromatic), 7.02 (d, J = 15.6 Hz, 1H, <u>HC</u>=CH-Ph), 6.82 (d, J = 8.4 Hz, 1H, aromatic), 6.78-6.70 (m, 3H, aromatic), 3.05 (s, 6H, N(C<u>H</u>₃)₂), 2.54 (s, 3H, C<u>H</u>₃); ¹³C NMR (100 MHz, CDCl₃, Fig. S37) δ 196.3, 160.1, 152.2, 145.3, 137.9, 133.0, 130.7 (2 carbons), 124.0, 122.6, 122.4, 121.7, 115.2, 111.9 (2 carbons), 40.1 (2 carbons), 23.0; *m*/z calcd for C₁₈H₁₉NO₂ 281.1; found 282.2 [M+H]⁺; Elemental analysis calcd. for C₁₈H₁₉NO₂: C, 76.84; H, 6.81; N, 4.98. Found: C, 76.57; H, 6.79; N, 5.02 (Fig. S38).



Synthesis of (*E*)-1-(3-bromo-2-hydroxyphenyl)-3-(4-(dimethylamino)phenyl)prop-2-en-1-one (6f). Following general procedure C, 6f (R_f 0.34, Hexanes:EtOAc/4:1) was obtained from 4f (215 mg, 1.0 mmol) and 2 (149 mg, 1.0 mmol) as purple crystals (153

mg, 44% yield): ¹H NMR (400 MHz, CDCl₃, Fig. S39) δ 7.96 (d, *J* = 15.6 Hz, 1H, HC=C<u>H</u>-Ph), 7.90 (dd, *J*₁ = 7.6 Hz, *J*₂ = 1.6 Hz, 1H, aromatic), 7.73 (dd, *J*₁ = 7.6 Hz, *J*₂ = 1.6 Hz, 1H, aromatic), 7.58 (dt, *J*₁ = 9.2 Hz, *J*₂ = 2.0 Hz, 2H, aromatic), 7.43 (d, *J* = 15.6 Hz, 1H, <u>H</u>C=CH-Ph), 6.83 (t, *J* = 7.6 Hz, 1H, aromatic), 6.74 (br. dt, *J*₁ = 9.2 Hz, *J*₂ = 2.0 Hz, 2H, aromatic), 3.08 (s, 6H, N(C<u>H</u>₃)₂); ¹³C NMR (100 MHz, CDCl₃, Fig. S40) δ 192.9, 160.0, 152.3, 147.5, 138.7, 131.1 (2 carbons), 128.5, 122.6, 121.3, 119.2 (2 carbons), 113.7, 112.2, 112.1, 40.3 (2 carbons); *m/z* calcd for C₁₇H₁₆BrNO₂ 345.0; found 346.0 [M+H]⁺; Elemental analysis calcd. for C₁₇H₁₆BrNO₂: C, 58.98; H, 4.66; Br, 23.08; N, 4.05. Found: C, 58.90; H, 4.72; Br, 22.91; N, 3.95 (Fig. S41).



Synthesis of (*E*)-1-(4-bromo-2-hydroxyphenyl)-3-(4-(dimethylamino)phenyl)prop-2-en-1-one (6g). Following general procedure C, the known compound $6g^{11}$ (R_f 0.47, Hexanes:EtOAc/4:1) was obtained from 4g (215 mg, 1.0 mmol) and 2 (149 mg, 1.0 mmol) as

a red powder (108 mg, 31% yield): ¹H NMR (400 MHz, CDCl₃) δ 13.36 (s, 1H, O<u>H</u>), 7.92 (d, *J* = 15.2 Hz, 1H, HC=C<u>H</u>-Ph), 7.75 (d, *J* = 8.8 Hz, 1H, aromatic), 7.56 (d, *J* = 8.8 Hz, 2H, aromatic), 7.35 (d, *J* = 15.2 Hz, 1H, <u>H</u>C=CH-Ph), 7.18 (d, *J* = 2.0 Hz, 1H, aromatic), 7.03 (dd, *J_I* = 8.8 Hz, *J₂* = 2.0 Hz, 1H, aromatic), 6.68 (d, *J* = 8.8 Hz, 2H, aromatic), 3.06 (s, 6H, N(C<u>H₃)₂); ¹³C NMR (100 MHz, CDCl₃) δ 192.8, 164.0, 152.5, 147.2, 131.0 (2 carbons), 130.2, 129.8, 122.1, 122.0, 121.6, 119.3, 113.7, 111.8 (2 carbons), 40.1 (2 carbons); *m/z* calcd for C₁₇H₁₆BrNO₂ 345.0; found 346.0 [M+H]⁺; Elemental analysis calcd. for C₁₇H₁₆BrNO₂: C, 58.98; H, 4.66; Br, 23.08; N, 4.05. Found: C, 59.06; H, 4.73; Br, 22.96; N, 4.10 (Fig. S42).</u>



Synthesis of (*E*)-1-(5-bromo-2-hydroxyphenyl)-3-(4-(dimethylamino)phenyl)prop-2-en-1-one (6h). Following general procedure C, 6h (R_f 0.52, Hexanes:EtOAc/5:1) was obtained from 4h (215 mg, 1.0 mmol) and 2 (149 mg, 1.0 mmol) as a red powder (168 mg, 49%

yield): ¹H NMR (400 MHz, CDCl₃, Fig. S43) δ 13.14 (s, 1H, O<u>H</u>), 7.99 (d, J = 2.4 Hz, 1H, aromatic), 7.92 (d, J = 15.2 Hz, 1H, HC=C<u>H</u>-Ph), 7.58 (dt, $J_1 = 9.2$ Hz, $J_2 = 2.0$ Hz, 2H, aromatic), 7.51 (dd, $J_1 = 8.4$ Hz, $J_2 = 2.4$ Hz, 1H, aromatic), 7.33 (d, J = 15.2 Hz, 1H, <u>HC</u>=CH-

Ph), 6.89 (d, J = 8.4 Hz, 1H, aromatic), 6.69 (dt, $J_1 = 9.2$ Hz, $J_2 = 2.0$ Hz, 2H, aromatic), 3.06 (s, 6H, N(C<u>H</u>₃)₂); ¹³C NMR (100 MHz, CDCl₃, Fig. S44) δ 192.3, 162.4, 152.6, 147.6, 138.1, 131.5, 131.1 (2 carbons), 122.1, 122.7, 120.4, 113.4, 111.8 (2 carbons), 110.1, 40.1 (2 carbons); m/z calcd for C₁₇H₁₆BrNO₂ 345.0; found 346.0 [M+H]⁺; Elemental analysis calcd. for C₁₇H₁₆BrNO₂: C, 58.98; H, 4.66; Br, 23.08; N, 4.05. Found: C, 58.32; H, 4.68; Br, 23.15; N, 3.91 (Fig. S45).



Synthesis of (*E*)-1-(2-aminophenyl)-3-(4-(dimethylamino)phenyl)prop-2-en-1-one (7a). Following general procedure C with the exception of quenching the reaction with H₂O instead of 1 N aq. HCl, the known compound $7a^{13}$ (R_f 0.21, Hexanes:EtOAc/5:1) was obtained from 5a (135)

mg, 1.0 mmol) and **2** (149 mg, 1.0 mmol) as a bright orange powder (214 mg, 80% yield): ¹H NMR (400 MHz, CDCl₃) δ 7.85 (dd, $J_I = 8.8$ Hz, $J_2 = 1.6$ Hz, 1H, aromatic), 7.71 (d, J = 15.6 Hz, 1H, HC=C<u>H</u>-Ph), 7.52 (dt, $J_I = 9.2$ Hz, $J_2 = 2.0$ Hz, 2H, aromatic), 7.40 (d, J = 15.6 Hz, 1H, <u>HC</u>=CH-Ph), 7.25 (ddd, $J_I = 8.4$ Hz, $J_2 = 7.2$ Hz, $J_3 = 1.6$ Hz, 1H, aromatic), 6.70-6.66 (m, 4H, aromatic), 6.22 (br s, 2H, N<u>H</u>₂), 3.02 (s, 6H, N(C<u>H</u>₃)₂); ¹³C NMR (100 MHz, CDCl₃) δ 192.0, 151.8, 150.6, 143.9, 133.6, 130.8, 130.1 (2 carbons), 123.1, 119.9, 118.0, 117.2, 115.8, 111.8 (2 carbons), 40.1 (2 carbons); *m*/*z* calcd for C₁₇H₁₈N₂O 266.1; found 267.1 [M+H]⁺; Elemental analysis calcd. for C₁₇H₁₈N₂O: C, 76.66; H, 6.81; N, 10.52. Found: C, 76.51; H, 6.78; N, 10.47 (Fig. S46).



Synthesis of (*E*)-1-(2-amino-3-methylphenyl)-3-(4-(dimethylamino)phenyl)prop-2-en-1-one (7b). Following general procedure C, with the exception of quenching the reaction with H_2O instead of 1 N aq. HCl, 7b (R_f 0.34, Hexanes:EtOAc/4:1) was obtained

from **5b** (122 mg, 0.82 mmol) and **2** (122 mg, 0.82 mmol) as a brown powder (178 mg, 78% yield): ¹H NMR (400 MHz, CDCl₃, Fig. S47) δ 7.77 (d, *J* = 8.0 Hz, 1H, aromatic), 7.71 (d, *J* = 15.6 Hz, 1H, HC=C<u>H</u>-Ph), 7.52 (d, *J* = 9.2 Hz, 2H, aromatic), 7.43 (d, *J* = 15.6 Hz, 1H, <u>HC</u>=CH-Ph), 7.18 (d, *J* = 8.0 Hz, 1H, aromatic), 6.70 (d, *J* = 9.2 Hz, 2H, aromatic), 6.63 (t, *J* = 8.0 Hz, 1H, aromatic), 6.40 (br s, 2H, N<u>H</u>₂), 3.02 (s, 6H, N(C<u>H</u>₃)₂), 2.18 (s, 3H, C<u>H</u>₃); ¹³C NMR (100 MHz, CDCl₃, Fig. S48) δ 192.4, 151.6, 149.1, 143.7, 134.5, 130.1 (4 carbons), 128.8, 123.3, 119.3, 118.5, 115.1, 112.0, 40.2 (2 carbons), 17.4; *m/z* calcd for C₁₈H₂₀N₂O 280.2; found 281.2 [M+H]⁺; Elemental analysis calcd. for C₁₈H₂₀N₂O: C, 77.11; H, 7.19; N, 9.99. Found: C, 77.11; H, 7.17; N, 10.08 (Fig. S49).



Synthesis of (*E*)-1-(2-amino-5-bromophenyl)-3-(4-(dimethylamino)phenyl)prop-2-en-1-one (7h). Following general procedure C, with the exception of quenching the reaction with H_2O instead of 1 N aq. HCl, 7h (R_f 0.27, Hexanes:EtOAc/4:1) was obtained from 5h

(214 mg, 1.0 mmol) and **2** (149 mg, 1.0 mmol) as a dark yellow powder (181 mg, 52% yield): ¹H NMR (400 MHz, CDCl₃, Fig. S50) δ 7.92 (d, J = 2.4 Hz, 1H, aromatic), 7.72 (d, J = 15.2 Hz, 1H, HC=C<u>H</u>-Ph), 7.54 (dt, $J_1 = 9.2$ Hz, $J_2 = 2.0$ Hz, 2H, aromatic), 7.304 (dd, $J_1 = 8.4$ Hz, $J_2 = 2.4$ Hz, 1H, aromatic), 7.298 (d, J = 15.2 Hz, 1H, <u>HC</u>=CH-Ph), 6.69 (dt, $J_1 = 9.2$ Hz, $J_2 = 2.0$ Hz, 2H, aromatic), 6.57 (d, J = 8.4 Hz, 1H, aromatic), 6.24 (br s, 2H, N<u>H</u>₂), 3.03 (s, 6H, N(C<u>H</u>₃)₂); ¹³C NMR (100 MHz, CDCl₃, Fig. S51) δ 190.8, 152.0, 149.4, 145.0, 136.1, 132.8, 130.4 (2 carbons), 122.7, 121.2, 118.9, 117.0, 111.8 (2 carbons), 106.9, 40.1 (2 carbons); *m/z* calcd for

C₁₇H₁₇BrNO₂ 344.0; found 345.0 [M+H]⁺; Elemental analysis calcd. for C₁₇H₁₇BrN₂O: C, 59.14; H, 4.96; Br, 23.14; N, 8.11. Found: C, 59.03; H, 5.04; Br, 23.27; N, 7.96 (Fig. S52).

 Cu^{2+} binding studies of chalcones. A 100 mM master stock solution of CuCl₂ was first prepared by dissolving copper (II) chloride dihydrate powder in mQ ddH₂O. 20-fold dilution of the master stock solution yielded a 5 mM CuCl₂ stock solution, C₃. The latter was then subjected to a 2.5fold dilution to obtain a 2 mM CuCl₂ stock solution, C₂, which was further diluted to afford a 1 mM CuCl₂ stock solution, C₁. To a cuvette containing 980 µL of a 20.4 µM solution of chalcone in EtOH was added 20 µL of H₂O, C₁, C₂, or C₃, corresponding to treatment with 0, 1, 2, and 5 equivalents of CuCl₂, respectively. The resulting solution was gently mixed and allowed to stand for 2 min at rt before its absorbance was measured by UV-Vis from 250 to 750 nm.

 Zn^{2+} binding studies of chalcones by ¹H NMR spectroscopy. As previously described,⁵ a solution of chalcone (3a, 3b, 3d, 3f, 3h, 6a, or 7a) (4 mM) in acetonitrile- d_3 (CD₃CN) was treated with ZnCl₂ (3 equivalents) and the resulting mixture was analyzed by ¹H NMR spectroscopy.

Solution speciation studies. The pKa values for chalcones 3a, 3d, 3h, 6a, and 7a were determined by UV-visible variable-pH titrations as previously described.⁵ A mixture of chalcone $[20 \ \mu\text{L} \text{ of an ethanolic solution of } 3a/6a/7a (1 \text{ mM}) \text{ or } 50 \ \mu\text{L} \text{ of an ethanolic solution of } 3d/3h (1 \text{ mM})$ mM)], CuCl₂ (1 mM, 10 µL for 3a/6a/7a or 25 µL for 3d/3h) and buffer (10 mM NaOH, pH 12, 100 mM NaCl, 970 µL for 3a/6a/7a or 925 µL for 3d/3h) was transferred to a cuvette and titrated with small aliquots of HCl (0.1 N). The absorbance was measured upon each addition of HCl from 250 to 900 nm to obtain at least 30 spectra in the range of pH 12-2. In order to investigate the binding properties of ligands to Cu²⁺ at various pH values, a mixture of chalcone [20 µL of an ethanolic solution of 3a/6a/7a (1 mM) or 50 µL of an ethanolic solution of 3d/3h (1 mM)], CuCl₂ (1 mM, 10 µL for 3a/6a/7a or 25 µL for 3d/3h) and buffer (10 mM NaOH, pH 12, 100 mM NaCl, 970 µL for 3a/6a/7a or 925 µL for 3d/3h) was transferred to a cuvette and allowed to stand for 30 min at room temperature. It was then titrated with small aliquots of HCl (0.1 N). Upon each addition of HCl, the absorbance was measured from 250 to 900 nm to obtain at least 20 spectra in the range of pH 2-7 for 3a and 3d, and pH 2-10 for 3h, 6a, and 7a. Using the HypSpec program (Protonic Software, U.K.), the pKas and $\log \beta$ constants were determined. The speciation diagrams were obtained with Sigmaplot Software). UV-visible variable-pH titration spectra and solution speciation diagrams of 6a and 7a in the presence of CuCl₂ are presented in Fig. S53.

Assays for bioA β_{42} oligomer assembly and dissociation. The bioA β_{42} oligomer assembly and dissociation experiments were carried out as previously described.^{14,15}

NeutrAvidinTM-coated ELISA plates: Each well of an immunoassay ELISA plate was coated with 50 μ L of 1 μ g/mL NeutrAvidinTM in 10 mM sodium phosphate buffer, pH 7.5 (50 ng NeutrAvidinTM per well). The plate was sealed with adhesive film and stored at 4 °C overnight. The following day, 200 μ L of PBS containing 0.1% v/v Tween 20 was added to each well to block the uncoated well surface and the plate was stored at 4 °C at least overnight. These plates were used to measure both oligomer assembly and dissociation.

*bioAβ*₄₂ *oligomer assembly*: A 1 mg/mL stock solution of *N*-biotinyl Aβ₁₋₄₂ (bioAβ₄₂) in hexafluoroisopropanol (HFIP) was evaporated, treated with neat trifluoroacetic acid (TFA) for 10 min to allow complete disaggregation of the peptide, dried again, dissolved further in HFIP to remove residual TFA, and dried and re-dissolved to a concentration of 500 nM (50x) in DMSO. After standing with intermittent vortexing for 10 min, 2 µL of the 50x monomeric peptide solution was transferred into each well of a polypropylene 96-well plate. In a separate plate 100x DMSO-solubilized chalcone (5 mM-78 µM) was prepared and then diluted (50 µM-0.78 µM final concentration of chalcone) in PBS with or without 25 µM metal ion (ZnCl₂ or CuCl₂) (final 1% DMSO). 100 µL of the chalcone dilutions were added to each well containing the DMSO solubilized bioAβ₄₂ (final Aβ concentration 10 nM). 30 min later, 50 µL Tween 20 in ddH₂O (0.3% v/v) was added to each well to stop oligomer assembly. Once formed, these oligomers have a half-lifetime $t_{1/2} = 30$ h at 22 °C for dissociation into monomers.

To measure the bioA β_{42} oligomers, the following steps were performed at rt (~22 °C). The blocking solution was flicked out of the wells of a NeutrAvidinTM-coated ELISA plate, 50 µL of the oligomer assembly reaction mixture was added, and the plate was sealed with adhesive film. The plate was shaken at 150 rpm for 2 h to allow monomeric and oligomeric bioA β_{42} to bind to the NeutrAvidinTM on the well surface. The plate was then washed three times with a low salt solution (34 mM aqueous NaCl, 20 mM Tris-HCl, pH 7.5, 0.1% v/v Tween 20 (TBST)) on a BioTek ELx50 plate washer. 50 µL of 50 ng/mL (1:20,000 diluted) Streptavidin-conjugated horseradish peroxidase (HRP) were added, the plate sealed with adhesive film, and the plate shaken at rt for 1 h. The plate was washed on the plate washer as previously, and 100 µL of 0.2 M citrate buffer, pH 4.0 containing 0.01% w/w H₂O₂ as HRP substrate and 1 mM tetramethylbenzidine were added. Since each captured bioA β_{42} peptide contains one biotin, NeutrAvidinTM-captured monomers will not react with the Streptavidin-HRP. Only oligomers will have biotin available for the Streptavidin-HRP to bind. The HRP reaction was stopped by the addition of 100 µL of 1% (v/v) H₂SO₄ to each well. The absorbance of each well was read at 450 nm with a Biotek HT Synergy plate reader.

bio $A\beta_{42}$ oligomer dissociation:

Pre-formation of bioA β_{42} *oligomers for dissociation*: A stock sufficient for four 96-well plates of pre-formed bioA β_{42} was prepared as follows. 1 µL of bioA β_{42} (1 mg/ml stock in HFIP) was pipetted into 20 µL of HFIP in a 0.65 mL polypropylene Eppendorf tube, dried down, and disaggregated as described for oligomer assembly. The dried peptide was re-dissolved in 250 µL of DMSO. After 10 min at rt with intermittent vortexing, the DMSO-solubilized bioA β_{42} was added to 12.5 mL of PBS in a 17x100 mm polypropylene tube, sealed with Parafilm[®], and mixed several times by inversion. The solution was then incubated at rt for 1 h with intermittent vortexing, then 375 µL of 100 mM Tween 20 (10% v/v in ddH₂O) was added for a final concentration of 0.3% v/v. The pre-formed stock oligomers (16.8 nM bioA β_{42}) were mixed by inversion and stored in aliquots at -75 °C.

Pre-formed oligomer dissociation: 25 μ L of the pre-formed bioA β_{42} oligomers adjusted to 0.6% v/v Tween 20 was pipetted into each well of a polypropylene 96-well plate, followed by the addition of 125 μ L PBS containing DMSO-solubilized chalcone (50 μ M-0.78 μ M final

concentration in two-fold serial dilutions in DMSO that is then added to PBS) with or without 25 μ M metal ion (ZnCl₂ or CuCl₂) (final 1% DMSO) to each well containing pre-formed oligomers. The total final concentration of bioA β_{42} peptide in each well was 2.8 nM. The plate was sealed and shaken at 150 rpm overnight at rt to allow the oligomers to dissociate. After 18 h of incubation, the plates were centrifuged at 1000xg for 10 min at rt to collect any condensate, and a 125 μ L aliquot was transferred from each well into a NeutrAvidinTM-coated ELISA plate to assay for oligomers as described for oligomer assembly.

In vitro cholinesterase (ChE) inhibition assays. Experiments were performed as previously described.^{16,17} Briefly, chalcones (102 pm to 200 μ M) were dissolved in sodium phosphate buffer ((100 μ L), 0.1 M, pH 8.0) (Buffer A) and subjected to a 5-fold serial dilution. ChE (either AChE or BChE) was added to the solution of inhibitors (50 μ L, containing 0.08 U/mL ChE (final concentration for both AChE and BChE) in Buffer A. The mixture of inhibitor and enzyme were incubated for 10 min before initiation with DTNB (50 μ L, 0.25 mM final concentration) and acylthiocholine (acetylthiocholine for AChE or butyrylthiocholine for BChE) (0.5 mM final concentration) in phosphate buffer. The reaction was monitored at 412 nm taking measurements every 30 s for 10 min using a SpectraMax M5 plate reader. Data was corrected with the negative control (no acylthicholine) and normalized to the positive control (no inhibitor) using the initial rates (first 5 min). All assays were performed in triplicate. The data was fitted to a sigmoidal curve and IC₅₀ values calculated using KaleidaGraph 4.1. Representive IC₅₀ curves are presented in Fig. S54.

Effect of metals on AChE inhibition. Chalcones were dissolved in Buffer A as above. To the solution, AChE mixed with metal ion (CuCl₂ or ZnCl₂, 200 μ M) in Buffer A was added and incubated for 10 min. The reactions were then initiated with the acetylthiocholine/DTNB mixture. Reactions were monitored and analyzed as above and previously described.¹⁸



Fig. S2: ¹³C NMR spectrum for compound 1h.



	At	lantic	Mic	rola	b, Inc.		
Sample No. MF	Y-2-84			Company	VSchool University	of Kentucky	
6180 Atlantic B	lvd. Suite M			Dept.	Pharmaceutical Sci	iences	
Norcross, GA 3	30071 Icrolab.com			Address	789 S. Limestone		
www.adandom	0.0.000		City	, State, Zip_	Lexington, KY 4053	36-0596	
Professor/Super	visor: Dr. Sylvie (Garneau-Tsodik	ova	Name_	Marina Fosso	Date	/2015
PO#) CC#_24	193S			Phone_	(859) 323-1945		
Element	Theory	Fo	ound		Single 🔀	Duplicate 🔲	
	70.40				Elements C, H, N Present	I, O	
0	76.16	75.88			Analyze C. H. N		
н	6.39	6.48			for:	Event of the D	
					M.P.	Explosive L	
N	11.10	11.02			To be dried: Yes		41
0	6.34				Rush Service	C	Mil be 5 PM EST
					Include Email Add	iress or FAX # Below	V 11.750.
L					sgt22	9@uky.edu	
		10 2015			81 0 1 0 0	A AAIF	
Date Received	JUN	19 2010		Date Corr	pleted JUN 2	2 2015	

Remarks: **Fig. S5:** Elemental analysis for compound **3a**.



Fig. S6: ¹H NMR spectrum for compound 3b.



Fig. S7: ¹³C NMR spectrum for compound **3b**.

Atlantic Microlab, Inc.

	V 0 161			2			
Sample No. MF 6180 Atlantic B Norcross, GA www.atlanticm	Ivd. Suite M 30071 icrolab.com		Company/School University of Kentucky Dept. Pharmaceutical Sciences Address 789 S. Limestone City State Zin Lexington, KY 40536-0596				
Professor/Super	rvisor: Dr. Sylvie ๆๆ35	Garneau-Tsodikov	<u>a Name</u> Name	Marina Fosso (859) 323-1945	Date ^{06/16/2015}		
Element	Theory	Fou	und	Single 🔀 🛛 Du	olicate 🗌		
С	76.66	75.18	75.03	Elements C, H, N, O Present:			
H	6.81	6.83	6.74	for:			
N	10.52	10.36	10.31	M.P.	sive B.P		
0	6.01	NO CHARGE FO	R DUPLICATES	Temp. 2.5 C Vac Rush ServiceRush service ompleted an nth degree for the day for 	gearantees analyses will be d results available by 5 PM EST 3 sample is received by 11 AM. r FAX # Below		
L				sgt229@uk	y.edu		
Date Received Remarks:	JUN	1 9 2015	Date Con	npleted JUN 2 2 20	115		

Fig. S8: Elemental analysis for compound 3b.



Sample No. MF	Y-3-42				t Iniu concit	a of Kontactus	
6180 Atlantic B Norcross, GA	lvd. Suite M 30071			Compa Dept.	Pharmaceutical Sciences 789 S. Limestone Lexington KY 40536-0596		
www.atlanticmi	crolab.com		01	Address			
Professor/Super	visor: Dr. Sylvie G	arneau-Tsodi	Ci kova	iy, State, Zip Name	Marina Fosso	Date ^{06/16/2015}	
PO# CC#_2	1935			Phone	(859) 323-1945		
Element	Theory	F	ound	deladeransen er annen ander	Single 🗙	Duplicate 🔲	
С	76.66	76.37			Elements C, H, Present:	N, O	
Н	6.81	6.81			Analyze C, H, N for:	N	
N	10.52	10.43			M.P.	Explosive [_] B.P	
0	6.01				Temp. 25°C Rush Service	Vac. <u>Justime 3-44</u> Rush service gubrantiess analyses will be completed and results available by 5 PM EST on the day the sample is creatived by 11 AM.	
					Include Email A sgt2	ddress or FAX # Below 229@uky.edu	
Date Received Remarks:	JUN	1 9 2015		_ Date Cor	npleted JUN	2 2 2015	

Fig. S11: Elemental analysis for compound 3c.



Fig. S12: ¹H NMR spectrum for compound **3d**.



230 220 210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 (ppm)

Fig. S13: ¹³C NMR spectrum for compound 3d.

Sample No.	-Y-2-87				Linh or rold	a f Kantur I	
6180 Atlantic E Norcross, GA	Bivd. Suite M 30071			Dept. Pharmaceutical Sciences			
www.atlanticmicrolab.com				Address	789 S. Limestone		
				City, State, Zip	Lexington, KY 40	536-0596	
Professor/Supe	rvisor: Dr. Sylvie C	Garneau-Tsodi	kova	Name	Marina Fosso	Date 06/16/2015	
(PO#)CC#	29935			Phone	(859) 323-1945		
Element	Theory	F	ound	d	Single 🗙	Duplicate	
С	76.66	76.38			Elements C, H, Present:	N, O	
Н	6.81	6.79			Hvgroscopic		
N	10.52	10.45			M.P To be dried: Yes	B.P	
0	6.01				Rush Service	Vac. VESTime 3-4% Rush service guarantees analyses will be completed and results available by 5 PM EST	
					Include Email A	ddress or FAX # Below	
					sgt2	29@uky.edu	
Date Received	JUN	1 9 2015		Date Cor	npleted JUN 2	2 2015	

Atlantic Microlab, Inc.

Remarks: **Fig. S14:** Elemental analysis for compound **3d**.



Fig. S16: ¹³C NMR spectrum for compound 3e.

Sample No. MF	Y-2-153				l Iniversity	of Kentucky	
6180 Atlantic Blvd. Suite M			Dept, Pharmaceutical Sciences				
www.atlanticm	icrolab.com			Address	789 S. Limestone		
			City	, State, Zip	Lexington, KY 405	536-0596	
Professor/Super	rvisor: Dr. Sylvie	Garneau-Tsodil	kova	Name	Marina Fosso	Date 06/16/2015	
(PO#) CC#2	29935			Phone	(859) 323-1945		
Element	Theory	F	ound	de Rommer yn Joan a'r	Single 🗙	Duplicate	
, C	76.66	76.44			Elements C, H, I Present:	N, O	
Н	6.81	6.82	_		Analyze C, H, N for:		
N	10.52	10.47			Hygroscopic	Explosive D B.P.	
0	6.01				To be dried: Yes Temp, <u>25°C. V</u> Rush Service	AL NO ac. <u>ALSTime 3-46</u> Rush service generates analyses will be completed and results available by 5 PM EST and the day the sample is received by 11 AM	
					Include Email Ac	dress or FAX # Below	
					sgt2	29@uky.edu	
<u></u>	l				<u> </u>		
Date Received	JUN	192015		Date Cor	npleted JUN 2	2 2015	
Remarks:							

Fig. S17: Elemental analysis for compound 3e.



Fig. S18: ¹H NMR spectrum for compound 3f.



Sample No. Mil 6180 Atlantic B Norcross, GA www.atlanticm Professor/Super (PO#)/ CC#_2	No. Suite M 30071 <i>icrolab.com</i> rvisor: <u>Dr. Sylvie</u> 9 9 3 5	Garneau-Tsodikov	Company/School University of Kentucky Dept. Pharmaceutical Sciences Address 789 S. Limestone City, State, Zip Lexington, KY 40536-0596 va Name Marina Fosso Date 06/16 Phone (859) 323-1945		
Element	Theory	Foi	und	Single X	Duplicate
С	58.02	56.76	56.93	Present:	N, O, Br
Н	4.57	4.60	4.62	for:	, Br
Br	24.12	23.43		M.P	
N	8.46	8.20	8.10	Temp. 25°C V Rush Service	Acc. <u>VISTime 3-4 k</u> Rush service guerantees analyses will be
0	4.83	NO CHARGE FO	R DUPLICATES	Include Email Ac	on the day the sample is received by 11 AM Idress or FAX # Below
					9 a uky. edu
Date Received	JUN	1 9 2015	Date Con	npleted JUN 2	2 2 2015

Fig. S20: Elemental analysis for compound 3f.



Fig. S22: ¹³C NMR spectrum for compound **3**g.

Sample No. MF	Y-2-147			
6180 Atlantic Blvd. Suite M Norcross, GA 30071 www.atlanticmicrolab.com			Compar Dept. Address	ny/School Unievrsity of Kentucky Pharmaceutical Sciences 789 S. Limestone
Professor/Super	rvisor: ^{Dr.} Sylvie { <u>935</u>	Garneau-Tsodikov	City, State, Zip. /a Name.	Marina Fosso Date 06/16/2015 (859) 323-1945
Element	Theory	Fou	und	Single X Duplicate
С	58.02	57.51	57.40	Present:
Н	4.57	4.53	4.49	for:
Br	24.12	23.73		M.P B.P
N	8.46	8.28	8.34	Temp. 25°C Vac. VugsTime 3-4% Rush Service Rush service goarantees analyses will be completed and results available to 50 PM FST
0	4.83	NO CHARGE FO	R DU PLICAT ES	on the day the sample is received by 11 AM. Include Email Address or FAX # Below
				sgt229@uky.edu
Date Received	JU	1 9 2015	Date Com	npleted JUN 2 2 2015

Remarks: **Fig. S23:** Elemental analysis for compound **3g**.



Fig. S24: ¹H NMR spectrum for compound **3h**.



230 220 210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 (ppm)

Fig. S25: ¹³C NMR spectrum for compound 3h.

Atlantic Microlab, Inc.

Sample No. MF	Y-2-149			-	
6180 Atlantic B Norcross, GA 3 www.atlanticmi	Ivd. Suite M 30071 crolab.com		Company/School University of Kentucky Dept. Pharmaceutical Sciences Address 789 S. Limestone		
Professor/Super	visor: <u>Dr. Sylvie</u> 9935	Garneau-Tsodikov	City, State, Zip /a Name Phone	Marina Fosso (859) 323-1945	Date ^{06/16/2015}
Element	Theory	Fou	Ind	Single 🗙	Duplicate
С	58.02	57.50	57.37	Present:	N, O, Br
H	4.57	464	4.55	for:	, Br
Br	24.12	23.69		M.P To be dried: Yes	Explosive [_] B.P [X] No [7]
N	8.46	8.26	8.27	Temp. 25°C N Rush Service	Ac. Ve Time 3-46 Rush service guarantees analyses will be completed and results available by 5 PM FST
0	4.83	NO CHARGE FO	R DUPLICATES	Include Email Ad	on the day the sample is received by 11 AM. ddress or FAX # Below
					229 Quky. ed
Date Received	JUL	1 9 2015	Date Con	npleted JUN	2 2 2015

Fig. S26: Elemental analysis for compound 3h.



Sample No. MF	At Y-2-148	lantic	Mic	rola	b, Inc.
6180 Atlantic B Norcross, GA www.atlanticm Prefessor/Super PO#/ CC#2	Ivd. Suite M 30071 icrolab.com rvisor: <u>Dr. Sylvie</u> 29935	Garneau-Tsodiko	Ci ova	Compar Dept. Address ty, State, Zip Name Phone	N/School Onlevisity of Kentucky Pharmaceutical Sciences 789 S. Limestone Lexington, KY 40536-0596 Marina Fosso (859) 323-1945 Date 06/16/2015
Element	Theory	Fo	ound	Sauce	Single X Duplicate
С	58.02	58.02			Elements C, H, N, O, Br Present:
Н	4.57	4.63			Analyze C, H, N, Br
Br	24.12	24.28	1		M.P B.P
N	8.46	8.43			Temp. 25°C. Vac. V 425 Time 3-4h Rush Service Rush service guarantees analyses will be
0	4.83		1		on the day the sample is received by 11 AM. Include Email Address or FAX # Below
					sgt229@uky.edu
Date Received	JUN	1 9 2015		Date Con	JUN 2 2 2015

Remarks:

Fig. S29: Elemental analysis for compound 3i.

Atlantic Microlab, Inc.

Sample No. MFY-2-85 6180 Atlantic Blvd. Suite M Norcross, GA 30071 www.atlanticmicrolab.com Professor/Supervisor: Dr. Sylvie Garneau-Tsodikova PO#/ CC#_29935			Compa Dept Address City, State, Zip va Name Phone	any/School University of Kentucky Pharmaceutical Sciences 789 S. Limestone Lexington, KY 40536-0596 Marina Fosso SST 323 1945
Element	Theory	Fo	und	Single Duplicate
С	76.38	76.20		Present:
Н	6.41	6.42		for:
N	5.24	5.24		M.P B.P To be dried: Yes X No
0	11.97			Temp. 25°C Vac. V using Time 3-4 K Rush Service Rush service guarantees analyses will be completed and results available by 5 PM EST on the day the sample is received by 11 AM.
				Include Email Address or FAX # Below sqt229@uky.edu
Date Received	JUN 1	g 2015	Date Co	

Fig. S30: Elemental analysis for compound 6a.





Sample No MF	Y-3-47					
6180 Atlantic B	lvd. Suite M		Compa	ny/School Universit	y of Kentucky	
Norcross, GA 3	30071		Dept. Pharmaceutical Sciences			
www.atlanticmi	crolab.com		Address	789 S. Limestone)	
			City, State, Zip	Lexington, KY 40	536-0596	
Professor/Super	visor: Dr. Sylvie	Garneau-Tsodiko	va Name	Marina Fosso	Date 06/16/2015	
(<u>PO#/CC#_2</u>	9935		Phone	(859) 323-1945	5410	
Element	Theory	Foi	und	Single 🗙	Duplicate	
С	76.84	76.79		Elements C, H, Present:	N, O	
Н	6.81	6.76		Analyze C, H, N for:		
N	4.98	5.01		M.P.	Explosive	
0	11.37			Temp, <u>25°C</u> v	AC. V 425 Time 3-46	
				Include Email Ad	completed and results available by 5 PM EST on the day the sample is received by 11 AM. Idress or FAX # Below	
				sgt22	29@uky.edu	
Date Received _ Remarks:	JUN	1 9 2015	Date Com	pleted JUN 2	2 2015	

Fig. S33: Elemental analysis for compound 6b.

Atlantic Microlab, Inc.

Sample No. MH 6180 Atlantic B Norcross, GA www.atlanticm	IV-2-92 IV. Suite M 30071 <i>icrolab.com</i>	Garneau-Tsodikov	Compai Dept. Address City, State, Zip	ny/School University Pharmaceutical Sc 789 S. Limestone Lexington, KY 405 Marina Fosso	of Kentucky iences 36-0596 Date ^{06/16/2015}
PO#) CC#2	993.5		Phone	(859) 323-1945	
Element	Theory	Fou	und	Single 🔀	Duplicate
С	76.84	76.33	76.18	Present: Analyze C H N	N, O
H	6.81	6.74	6.86	for:	
N	4.98	4.94	493	M.P To be dried: Yes	
0	11.37	NO CHARGE FOF	DUPLICATES	Rush Service	AC. V 4 4 I IME <u>2.74 h</u> Rush service guarantees analyses will be completed and results available by 5 PM EST on the day the sample is received by 11 AM.
				Include Email Ad	dress or FAX # Below
1				sgt22	29@uky.edu
Date Received Remarks:	JUN	1 9 2015	Date Co	mpleted JUN 2	2 2015

Fig. S34: Elemental analysis for compound 6c.

Sample No. MF 6180 Atlantic Bl Norcross, GA 3 www.atlanticmi	Y-2-93 Ivd. Suite M 80071 crolab.com visor: Dr. Sylvie (Garneau-Tsodikov	Compar Dept. Address City, State, Zip a Name.	ny/School University Pharmaceutical Sc 789 S. Limestone Lexington, KY 405 Marina Fosso	of Kentucky iences 36-0596 Date 06/16/2015
PO#) CC#_2	1935		Phone	(859) 323-1945	
Element	Theory	Fou	Ind	Single X	Duplicate
С	76.84	76.83		Present:	N, O
Н	6.81	6.93		for:	
	4.98	5.00		M.P.	
0	11.37			Temp. <u>25°C</u> V Rush Service	ac. VyesTime 3-46, tush service-guarantiaes analyses will be completed and results available by 5 PM EST on the day the sample is received by 11 AM.
				Include Email Ad sgt22	dress or FAX # Below 29@uky.edu
					-
Date Received	JUN	192015	Date Cor	mpleted JUN 2	2 2015

Remarks:

Fig. S35: Elemental analysis for compound 6d.



Fig. S36: ¹H NMR spectrum for compound 6e.



Comple No. MF	Y-3-38						
6180 Atlantic B Norcross, GA 3 www.atlanticmi Professor/Super	Ivd. Suite M 30071 Scrolab.com	Garneau-Tsodikov	Compai Dept. Address City, State, Zip a Name	Company/School University of Kentucky Dept. Pharmaceutical Sciences Address 789 S. Limestone City, State, Zip Lexington, KY 40536-0596 Name Marina Fosso Date06/16/2			
(PO#// CC#	9935		Phone	(859) 323-1945			
Element	Theory	Fou	Ind	Single 🗙	Duplicate		
C	76.84	76.57		Elements C, H, Present:	N, O		
Н	6.81	6.79		Analyze C, H, N for:	1		
N	4.09	502		Hygroscopic X M.P.	Explosive		
	4.90			To be dried: Yes	No D 210		
0	11.37			Rush Service	Vac. V 425 ime 3-446 Rush service guarantees analyses will be completed and results available by 5 PM EST		
				Include Email A	on the day the sample is received by 11 AM. ddress or FAX # Below		
				sgt2	29@uky.edu		
Date Received	JUN 1	9 2015	Date Con	npleted JUN 2	2 2015		

Remarks: **Fig. S38:** Elemental analysis for compound **6e**.



Sample No. M	FY-3-39					v of Kentucky	
6180 Atlantic I	Blvd. Suite M			Dept. Pharmaceutical Sciences			
www.atlanticmicrolab.com				Address	789 S. Limestone		
			(City, State, Zip	536-0596		
Professor/Sune	ervisor: Dr. Sylvie (Garneau-Tsodiko	ova	Name	Marina Fosso	Date 06/16/2015	
PO#/ CC#	29935			Phone	(859) 323-1945		
Element	Theory	Fo	und		Single 🗙	Duplicate 🔲	
С	58.98	58.90	Τ		Elements C, H, Present:	N, O, Br	
н	4.66	4.72	1		Analyze C, H, M	l, Br	
Br	23.08	22.91			Hygroscopic 🔀	Explosive	
			-		To be dried: Yes	Vac. Vyestime <u>3-4 h</u>	
N	4.05	3.95	ļ		Rush Service 🔲	Rush service guerantees analyses will be completed and results available by 5 PM EST on the deaths available by 5 PM EST	
0	9.24				Include Email A	ddress or FAX # Below	
					sgt2	29@uky.edu	
Date Receive	JUN	1 9 2015		Date Cor	npleted_JUN 2	2 2015	

Remarks:

Fig. S41: Elemental analysis for compound 6f.

Atlantic Microlab, Inc.

Sample No. MF	Y-2-125					
6180 Atlantic E Norcross, GA	Bivd. Suite M 30071			Compar Dept.	ny/School University of Kentucky Pharmaceutical Sciences	
www.atlanticm	icrolab.com rvisor: <u>Dr</u> . Sylvie (こそそろろ	Garneau-Tsodi	City, S kova	Address State, Zip Name Phone	789 S. Limestone Lexington, KY 405 Marina Fosso (858) 323-1945	36-0596 Date ^{06/16/2015}
Element	Theory	F	ound		Single 🗙	Duplicate
С	58.98	59.06			Elements C, H, N Present:	I, O, Br
Н	4.66	473			for:	Br
Br	23.08	22.96			M.P	
N	4.05	4 10			Temp. <u>25°C.</u> Va Rush Service	ac. VyzS Time 3-46
0	9.24				Include Email Add	the day the sample is received by 11 AM.
					sgt22	9@uky.edu
Date Received Remarks:	JUN	1 9 2015	E	ate Corr	pleted JUN 2	2 2015

Fig. S42: Elemental analysis for compound 6g.



Sample No. MF	Y-2-91						
6180 Atlantic Blvd. Suite M Norcross, GA 30071			Company/School Oniversity of Kentucky Dept. Pharmaceutical Sciences				
www.atlanticm	icrolab.com		Address	Address 789 S. Limestone			
Professor/Super	visor: Dr. Sylvie	Garneau-Tsodiko	City, State, Zip va Name	Marina Fosso	Date ^{06/16/2015}		
(PU#/CC#	192	Contraction and the second	Phone	(859) 323-1935			
Element	Theory	Fo	und	Single 🔀	Duplicate		
С	58.98	58.32	58.37	Elements C, H, Present:	N, O, Br		
Н	4.66	4.68	4.58	for:	, Br		
Br	23.08	23.15		M.P.	Explosive		
N	4.05	3.91	3.91	Temp. 25°C	A NO L ac. Lus Time 3-4 6-		
0	9.24	NO CHARGE FO	R DUPLICATES	Include Email Ad	completed and results available by 5 PM EST on the day the sample is received by 11 AM. Idress or FAX # Below		
				sgt22	29@uky.edu		
Date Received . Remarks:	JUN 1	9 2015	Date Corr	pleted JUN 2	2 2015		

Fig. S45: Elemental analysis for compound **6h**.

Atlantic Microlab, Inc.

Sample No. N	1FY-2-89			-			
6180 Atlantic	Blvd. Suite M		Compa Dept.	ny/School University of Kentucky Pharmaceutical Sciences			
www.atlanticr	nicrolab.com		Address	Address 789 S. Limestone City, State, Zip Lexington, KY 40536-0596			
			City, State, Zip				
Professor/Sup	ervisor: Dr. Sylvie	Garneau-Tsodiko [,]	va Name	Marina Fosso	Date 06/16/2015		
PO#/cc#	29935		Phone	Phone (859) 323-1945			
Element	Theory	Fo	und	Single 🗙	Duplicate		
C	76.66	76.51		Elements C, H, Present:	N, O		
Н	6.81	6.78		Analyze C, H, M	۷ 		
N	10.52	10.47		Hygroscopic			
0	6.01			Temp. 25°C	Vac. V425 Time 3-4.2		
				Include Email A	on the day the sample is received by 11 AM		
		_		sgt2	29@uky.edu		
Date Receive Remarks:	ed JUN	1 9 2015	Date Cor	mpleted JUN	2 2 2015		

Fig. S46: Elemental analysis for compound 7a.



Sample No. MF	Y-3-1				-	
6180 Atlantic B Norcross, GA 3 www.atlanticmi	lvd. Suite M 30071 crolab.com		Cit	Company/School University of Kentucky Dept, Pharmaceutical Sciences Address 789 S. Limestone City, State, Zip Lexington, KY 40536-0596		
Professor/Super	visor: Dr. Sylvie (9935	3arneau-Tsodiko	va	Name Phone	Marina Fosso Date 06/16/201 (859) 323-1945 0	
Element	Theory	Fo	und	54-14-1-1-1-1-1-1-1-1-1-1-1-1-1-1-1-1-1-	Single 🛛 Duplicate 🗌	
С	77.11	77.11			Elements C, H, N, O Present:	
Н	7.19	7.17			Hydroscopic X Explosive	
N	9.99	10.08			M.P B.P To be dried: Yes 🕅 No	
0	5.71				Temp. 2.5°C. Vac. Jus Time 3-42 Rush Service Converted starting snapper with by 5 Miss on the day the snappe is necessed by 11 AM Include Email Address or FAX # Below sg1229@uky.edu	
Date Received	JU	IN 192015	I	Date Com	npleted JUN 2 2 2015	

Remarks: **Fig. S49:** Elemental analysis for compound **7b**.



Fig. S50: ¹H NMR spectrum for compound **7h**.



Fig. S51: ¹³C NMR spectrum for compound **7h**.

Atlantic Microlab, Inc.

ME	V 0 106			,		
Sample No. MF 6180 Atlantic B Norcross, GA 3 www.atlanticm Professor/Super (O#) CC#2	17-2-126 Ivd. Suite M 30071 icrolab.com visor: <u>Dr. Sylvie (</u> 	Gameau-Tsodiko	Compa Dept. Address City, State, Zip Va Name Phone	Company/School University of Kentucky Dept. Pharmaceutical Sciences Address 789 S. Limestone City, State, Zip Lexington, KY 40536-0596 Name Marina Fosso Phone (859) 323-1945		
Element	Theory	Fo	und	Single 🛛 Duplicate 🗌		
С	59.14	59.03		Elements C, H, N, O, Br Present:		
Н	4.96	5.04		for:		
Br	23.14	23.27		M.P B.P		
N	8.11	7.96		Temp. 25°C. Vac. Ves Time <u>3</u> ~4.2 Rush Service Rush service guarantees analyses will be	<u>+</u>	
0	4.63			on the day the sample is received by 11 AM Include Email Address or FAX # Below	51 [
				sgt229@uky.edu		
Date Received Remarks:	JUN	1 9 2015	Date Con	npleted JUN 2 2 2015		

Fig. S52: Elemental analysis for compound 7h.



Fig. S53: UV-visible variable-pH spectra (left) and solution speciation diagrams (right) of **6a** and **7a** in the presence of CuCl₂. 20 μ M of the chalcone (S: **6a** or **7a**) was incubated for 30 minutes with CuCl₂ ([Cu²⁺]/[S] = 1/2). Titrations were then performed at room temperature from pH 12-2. There were no differences between the curves obtained in the absence and presence of CuCl₂, suggesting that these chalcones (**6a** and **7a**) do not bind Cu²⁺.



Fig. S54: Representative IC_{50} curves for compounds **3c** and **7b** with AChE and BChE in the absence of presence of $CuCl_2$ or $ZnCl_2$.

References

- (1) Nayad, S.; Lee, H.; Jeong, J. H. Polyhedron 2012, 43, 55.
- (2) Alhamadsheh, M. M.; Gupta, S.; Hudson, R. A.; Perera, L.; Tillekeratne, L. M. V. Chem. *Eur. J.* **2008**, *12*, 570.
- (3) Crabb, T. A.; Heywood, G. C. Org. Magn. Resonance 1982, 20, 242.
- (4) Reck, F.; Zhou, F.; Eyermann, C. J.; Kern, G.; Carcanague, D.; Ioannidis, G.; Illingworth, R.; Poon, G.; Gravestock, M. B. J. Med. Chem. 2007, 50, 4868.
- (5) Liu, Y.; Kochi, A.; Pithadia, A. S.; Lee, S.; Nam, Y.; Beck, M. W.; He, X.; Lee, D.; Lim, M. H. *Inorg. Chem.* 2013, *52*, 8121.
- (6) Seidel, J. L.; Epstein, W. W.; Davidson, D. W. J. Chem. Ecol. 1990, 16, 1791.
- (7) Uto, Y.; Ueno, Y.; Kiyotsuka, Y.; Miyazawa, Y.; Kurata, H.; Ogata, T.; Yamada, M.; Deguchi, T.; Konishi, M.; Takagi, T.; Wakimoto, S.; Ohsumi, J. Eur. J. Med. Chem. 2010, 45, 4788.
- (8) Setoh, M.; Ishii, N.; Kono, M.; Miyanohana, Y.; Shiraishi, E.; Harasawa, T.; Ota, H.; Odani, T.; Kanzaki, N.; Aoyama, K.; Hamada, T.; Kori, M. J. Med. Chem. 2014, 57, 5226.
- (9) Mangas-Sanchez, J.; Busto, E.; Gotor-Fernandez, V.; Gotor, V. Org. Lett. 2012, 14, 1444.

- (10) Liu, M.; Wilairat, P.; Croft, S. L.; Tan, A. L.; Go, M. L. *Bioorg. Med. Chem.* **2003**, *11*, 2729.
- (11) Dang, S.; Liu, J.-G.; Wang, G.-H. Hecheng Huaxue 2008, 16, 460.
- (12) Mohamed, H. A. S.; Sivakumar, T. Int J Drug Des Disc 2011, 2, 497.
- (13) Ullah, A.; Ansari, F. L.; Ihsan ul, H.; Nazir, S.; Mirza, B. Chem. Biodivers. 2007, 4, 203.
- (14) LeVine, H., 3rd Analytical biochemistry 2006, 356, 265.
- (15) LeVine, H., 3rd; Ding, Q.; Walker, J. A.; Voss, R. S.; Augelli-Szafran, C. E. *Neurosci. Lett.* **2009**, *465*, 99.
- (16) Bornstein, J. J.; Eckroat, T. J.; Houghton, J. L.; Jones, C. K.; Green, K. D.; Garneau-Tsodikova, S. *MedChemComm* **2011**, *2*, 406.
- (17) Eckroat, T. J.; Green, K. D.; Reed, R. A.; Bornstein, J. J.; Garneau-Tsodikova, S. *Bioorg. Med. Chem.* **2013**, *21*, 3614.
- (18) Kochi, A.; Eckroat, T. J.; Green, K. D.; Mayhoub, A. S.; Lim, M. H.; Garneau-Tsodikova, S. *Chem. Sci.* **2013**, *4*, 4137.