

Supporting Information

A latent reactive handle for functionalising heparin-like and LMWH deca- and dodecasaccharides

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The following pages contains representative experimental protocols, data and spectra.

1. Synthetic procedures for compounds 4, 5, 10, 11, 12, 13, 14 and 15

All the chemicals used were purchased from commercial sources without further purification. All reactions were monitored by TLC on Merck silica gel plates ⁶⁰F₂₅₄. Silica gel 60 (particle size 0.035-0.070 mm) was used for column chromatography. ¹H NMR spectra were recorded at 400 MHz and ¹³C spectra at 100 MHz respectively on Bruker DPX spectrometers. Mass spectra (MS) were recorded using a Micromass Platform II spectrometer using an electro spray ionization source or via the EPSRC National Mass Spectrometry Service (Swansea). ESI spectra and isotope patterns for compounds with mass > 1000 are included with the NMR data. High resolution data are for the monoisotopic mass. Infrared spectra were obtained by using a Bruker Alpha instrument. Melting points were determined using Stuart Scientific SMP10 apparatus and are uncorrected. Optical rotations were obtained using an AA-1000 polarimeter. Elemental analyses were performed by Micro Analytical Laboratory, School of Chemistry, The University of Manchester. ESI MS conditions for final saccharide compounds: prior to MS, the sample for analysis was salt counterion switched from Na⁺ to NH₄⁺ using Amberlite IRC86-H⁺ resin (pretreated with 5% aqueous NH₄OH solution).

*N*¹-(7-nitrobenzo[*c*][1,2,5]oxadiazol-4-yl)ethane-1,2-diamine (4)

To a stirring solution of N-Boc ethylenediamine (180 μL, 1.14 mmol) and sodium hydrogen carbonate (239 mg, 2.08 mmol) in THF/EtOH (1:1, 2.0 mL) at 0 °C a solution of NBD-chloride (190mg, 0.95

mmol) in THF/EtOH (1:1, 2 mL) was added over 15 minutes. The reaction mixture was stirred at room temperature for 12 hours and then poured onto water. The aqueous layer was extracted with DCM (2 x 20 mL). The organic phases were combined, washed with brine, dried (MgSO₄), concentrated *in vacuo* and purified by silica gel flash chromatography eluting with DCM/MeOH, 2:1+5%NH₄OH to yield N-BOC protected **4** as a yellow/brown solid (266 mg, 0.82 mmol, 87%). R_f = 0.56 (DCM/MeOH, 9:1). Hydrochloric acid (240 μL, 3M) was added to a stirring solution of N-BOC protected **4** (226 mg, 0.70 mmol) in THF (3 mL) at 60 °C and the mixture was stirred for 4.5 hours, whereupon the reaction was shown to be complete by TLC analysis (DCM/MeOH, 9/1). The solution was concentrated *in vacuo* to yield **4** (150 mg, 0.67 mmol, 83%) as a yellow/brown solid. Mp: 236-238 °C; HRMS (TOF) *m/z* calcd for C₈H₁₀N₅O₃ [M+H]⁺ 224.0779, found 224.0789. Other analytical data matched those reported previously.¹

N-(6-aminohexyl)-4-(pyren-4-yl)butanamide hydrochloride (5)

Under an inert nitrogen atmosphere tert-butyl-N-(6-amino-hexyl)carbamate hydrochloride (228 mg, 0.90 mmol), 1-pyrenebutyric acid (216 mg, 0.75 mmol), TBTU (362 mg, 1.13 mmol) and DIPEA (392 μL, 2.25 mmol) were dissolved in anhydrous DMF (4 mL) and the mixture was stirred at 50 °C for 23 hours whereupon the reaction was shown to be complete by TLC analysis. The solution was diluted with diethyl ether (25 mL) and washed with water (5 x 10 mL). The organic phase was dried (MgSO₄) and concentrated *in vacuo*. The product was purified by silica gel flash chromatography eluting with EtOAc/hexane, 2/1) to yield N-BOC pyrene derivative **5** as an off- white solid (90 mg, 0.18 mmol, 25%). R_f = 0.61(EtOAc/hexane, 3:1). Hydrochloric acid (147 μL, 3M) was added to a stirring solution of N-BOC pyrene derivative **5** (85 mg, 0.18 mmol) in THF (2.0 mL) at 60 °C and the mixture was stirred for 4 hours whereupon the reaction was shown to be complete by TLC analysis (EtOAc/hexane, 2:1). The mixture was concentrated *in vacuo* to reveal **5** (70 mg, 0.18 mmol, 95%) as a white solid. Mp: 228-231 °C; LRMS (ESI+) *m/z* 387.0 [M+H]⁺. Other analytical data matched those reported previously.²

Phenyl-4-O-allyl-2-azido-2-deoxy-3,6-di-O-benzyl-1-thio-α-D-glucopyranoside (10)

Thioglycoside **1**¹⁸ (485 mg, 1.0 mmol) and sodium hydride (34 mg, 1.2 mmol, 60% dispersion in oil) were dissolved in dry DMF (10 mL) under nitrogen and the solution stirred at 0 °C for 30 minutes. Allyl bromide (132 μL, 1.5 mmol) was added to the stirring solution, the reaction temperature was then raised to 50 °C and stirred for 1 hour. Addition of water (5 mL) caused simultaneous quenching of remaining sodium hydride as well as precipitation of the allylated sugar from the reaction mixture. The product

was filtered and the filtrate extracted with DCM (30 mL) then washed with water (4 x 15 mL). The organic phase was dried (MgSO₄), filtered and solvents removed *in vacuo* to yield **10** as a white solid (41 mg). This was combined with the previously precipitated material (466 mg) to give an overall yield of **10** (507 mg, 0.98 mmol, 97%). mp 94-96 °C; [α]_D +154.2 (c = 10.0, DCM); ¹H NMR (CDCl₃, 400MHz) δ 7.52-7.26 (m, 15H, ArH), 5.80-5.70 (ddt, *J* = 17.2, 10.4, 5.6 Hz, 1H, H₈), 5.60 (d, *J* = 5.6 Hz, 1H, H₁), 5.21 (ddd, *J* = 17.2, 3.2, 1.6 Hz, 1H, H_{9a}), 5.14 (ddd, 1H, *J* = 10.4, 2.8, 1.2 Hz, H_{9b}), 4.90 (d, *J* = 10.4 Hz, 1H, CH₂Ar), 4.87 (d, *J* = 10.8 Hz, 1H, CH₂Ar), 4.63 (d, *J* = 12.0 Hz, 1H, CH₂Ar), 4.87 (d, *J* = 12.0 Hz, 1H, CH₂Ar), 4.32 (ddd, 1H, *J* = 10.0, 3.6, 2.0 Hz, 1H, H₅), 4.27 (ddt, 1H, *J* = 12.4, 5.6, 1.6 Hz, H_{7a}), 4.02 (ddt, *J* = 12.0, 5.6, 1.2 Hz, 1H, H_{7b}), 3.91 (dd, *J* = 10.4, 5.2 Hz, 1H, H₂), 3.80-3.74 (m, 2H, H₃, H_{6a}), 3.66-3.63 (m, 1H, H_{6b}), 3.62 (dd, *J* = 7.6, 6.4 Hz, 1H, H₄); ¹³C NMR (100 MHz; CDCl₃) δ 137.8, 137.7, 134.4, 132.0, 129.1, 128.5, 128.4, 128.2, 128.0, 127.9, 127.8, 127.7, 117.1, 87.2, 81.7, 78.1, 75.8, 73.9, 73.5, 71.8, 68.3, 63.9; MS ES [M+Na]⁺ *m/z* 540.0; HRMS (TOF⁺) *m/z* calcd for C₂₉H₃₁N₃O₄SNa 540.1927, found 540.1919; IR (neat) ν_{\max} 2923, 2853, 2105; Elemental analysis: calcd for C₂₉H₃₁N₃O₄S. C 67.3; H 6.0; N 8.1, found C 67.1; H 6.2; N 8.2.

Phenyl-2-azido-2-deoxy-3,6-di-O-benzyl-4-O-[(*S/R*)-2,3-dihydroxypropoxy]-1-thio- α -D-glucopyranoside (11**)**

The allylated glucosamine derivative **10** (500 mg, 0.97 mmol) was dissolved in an ice-cooled mixture of acetone:water (9:1, 5 mL). NMO (180 mg, 1.54 mmol) and osmium tetroxide (316 μ L, 2.5 mol% in *t*BuOH) was added to the solution and the mixture stirred for 8 hours at 0 °C warming to room temperature. The reaction was quenched with 10% aqueous sodium thiosulfate (5 mL) followed by extraction of the product into DCM (3 x 60 mL). The combined organics were dried (MgSO₄), filtered and evaporated *in vacuo* to yield **11** (521 mg, 0.95 mmol, 98 %) as a white crystalline solid. R_f (hexane:EtOAc, 1:1) 0.24; ¹H NMR (400 MHz, CDCl₃) δ 7.50-7.48 (m, 2H, ArH), 7.44-7.27 (m, 2H, ArH), 5.98 (d, *J* = 5.2 Hz, 1H, H₁), 4.94 (d, *J* = 10.4 Hz, 1H, CH₂Ar), 4.81 (d, *J* = 10.8 Hz, 1H, CH₂Ar), 4.63 (d, *J* = 12.0 Hz, 1H, CH₂Ar), 4.46 (d, *J* = 12.0 Hz, 1H, CH₂Ar), 4.30-4.24 (m, 1H, H₅), 3.92 (dd, *J* = 10.0, 5.2 Hz, 1H, H₂), 3.82-3.40 (m, 9H, H₃, H₄, H_{6ab}, H₇, H₈, H₉), 2.87 (d, *J* = 4.4 Hz, 1H, OH, C_{8major}), 2.69 (d, *J* = 5.2 Hz, 1H, OH, C_{8minor}), 1.86 (dd, *J* = 6.8, 5.2 Hz, 1H, OH, C_{9major}), 1.84 (dd, *J* = 7.2, 5.2 Hz, 1H, OH, C_{9minor}); ¹³C NMR (100 MHz; CDCl₃) δ 137.5, 137.4, 137.2, 133.3, 132.0, 132.0, 129.1, 128.6, 128.5, 128.4, 128.3, 128.2, 128.1, 128.1, 127.9, 127.7, 87.2, 81.6, 78.8, 78.7, 75.9, 75.8, 74.3, 73.6, 73.6, 71.7, 71.6, 71.2, 71.1, 68.3, 68.2, 64.3, 64.2, 63.6, 63.5; IR (neat) ν_{\max} 3422, 3061,

3031, 2922, 2869, 2104, 1045; MS ES [M+Na]⁺ *m/z* 574.0; HRMS (TOF⁺) *m/z* calcd for C₂₉H₃₃N₃O₆SNa 574.1982, found 574.1981.

Phenyl-2-azido-2-deoxy-3,6-di-*O*-benzyl-4-*O*-[(*S/R*)-2,3-bis(benzyloxy)propoxy]-1-thio- α -D-glucopyranoside (12)

To a solution of **11** (742 mg, 1.34 mmol) in anhydrous DMF (10 mL) was added sodium hydride (83.5 mg, 2.09 mmol, 60% dispersion in oil) and the mixture was stirred for 20 minutes at room temperature. Benzyl bromide (480 μ L, 4.0 mmol) was added and the reaction left to stir for 90 minutes at room temperature. The mixture was diluted with chloroform (80 mL), washed with water (4 x 20 mL) and then brine (20 mL). The organics were dried (MgSO₄), filtered and solvents removed *in vacuo* to give the crude product as a yellow gum. Purification by silica gel flash chromatography eluting with hexane/EtOAc, 5/1, 4/1 furnished **12** (757 mg, 1.04 mmol, 77%). ¹H NMR (400 MHz, CDCl₃) δ 7.60-7.58 (m, 2H, ArH), 7.47-7.44 (m, 2H, ArH), 7.40-7.30 (m, 21H, ArH), 5.65 (d, *J* = 5.2 Hz, 1H, H₁), 4.98-4.89 (m, 2H, CH₂Ar), 4.74-4.64 (m, 2H, CH₂Ar), 4.58-4.45 (m, 4H, CH₂Ar), 4.42-4.37 (m, 1H, H₅), 4.11-4.06 (m, 1H, H₈), 3.96 (dd, *J* = 10.4, 5.6 Hz, 1H, H₂), 3.86-3.70 (m, 5H, H₃, H₄, H_{6a}, H₇), 3.67-3.59 (m, 3H, H_{6a}, H₉); ¹³C NMR (100 MHz; CDCl₃) δ 138.5, 138.4, 138.1, 138.0, 137.9, 137.7, 137.6, 133.6, 133.5, 132.1, 132.0, 129.1, 128.5, 128.4, 128.3, 128.2, 128.1, 127.9, 127.8, 127.7, 127.6, 87.2, 81.62, 81.56, 78.8, 78.7, 77.5, 77.3, 77.2, 75.6, 73.4, 73.3, 73.2, 72.4, 72.3, 71.8, 71.8, 70.0, 69.8, 68.4, 64.0, 63.9; MS ES [M+Na]⁺ *m/z* 554.3; HRMS (FTMS-NSI⁺) *m/z* calcd for C₄₃H₄₉N₄O₆S₁ [M+NH₄]⁺ 749.3367, found 749.3368; IR (neat) ν_{\max} 3061, 3029, 2864, 2104, 1088.

Phenyl-2-azido-2-deoxy-3,6-di-*O*-benzyl-4-*O*-[(*S*)-2,3-bis(benzyloxy)propoxy]-1-thio- α -D-glucopyranoside (15)

To **1** (611 mg, 1.28 mmol) was added dry THF (10 mL) under N₂ and the solution cooled to 0 °C. NaH (60% in mineral oil) (76.0 mg, 1.9 mmol) was added in two portions over 30 min. while being kept under N₂. **13**²² (600 mg, 1.41 mmol) in dry THF (5 mL) was then added dropwise and the suspension allowed to warm to RT and heated at 50 °C overnight. TLC analysis (4/1, hexane/EtOAc) showed the reaction to be complete and quenching was effected with aqueous NaHCO₃ (1 mL). The solution was partitioned between EtOAc and H₂O. The layers were separated and the organic phase washed with 1M HCl, H₂O, saturated aqueous NaCl, dried (MgSO₄), filtered and evaporated. The crude product was purified twice by flash column chromatography (EtOAc/hexane gradient 1:10, 1:8, then DCM/Ether gradient 99:1) yielding **15** (115 mg, 0.16 mmol, 21%) as

a clear oil. R_f 0.40 (EtOAc/hexane, 1:4); $[\eta]_D$ -19.4 ($c = 0.7$, DCM); $^1\text{H NMR}$ (400 MHz; CDCl_3) δ 7.44-7.39 (m, 2H, ArH), 7.30-7.25 (m, 2H, ArH), 7.25-7.11 (m, 21H, ArH), 5.48 (d, $J = 5.3$ Hz, 1H, H_1), 4.80 (d, $J = 10.6$ Hz, 1H, CH_2Ar), 4.76 (d, $J = 10.6$ Hz, 1H, CH_2Ar), 4.55 (d, $J = 11.9$ Hz, 1H, CH_2Ar), 4.51 (d, $J = 11.9$ Hz, 1H, CH_2Ar), 4.39 (s, 2H, CH_2Ar), 4.35 (d, $J = 12.0$ Hz, 1H, CH_2Ar), 4.28 (d, $J = 12.0$ Hz, 1H, CH_2Ar), 4.23 (ddd, $J = 9.9, 3.8, 1.7$ Hz, 1H, H_5), 3.94-3.91 (m, 1H, $\text{CH}_2\text{CH}[\text{OBn}]\text{CH}_2\text{OBn}$), 3.79 (dd, $J = 10.3, 5.3$ Hz, 1H, H_2), 3.69-3.63 (m, 2H, H_3, H_4), 3.63-3.52 (m, 3H, H_{6A} , $\text{CH}_2\text{CH}[\text{OBn}]\text{CH}_2\text{OBn}$), 3.49-3.41 (m, 3H, H_{6B} , $\text{CH}_2\text{CH}[\text{OBn}]\text{CH}_2\text{OBn}$); $^{13}\text{C NMR}$ (100 MHz; CDCl_3) δ 138.6, 138.1, 138.0, 137.8, 133.7, 132.1, 129.1, 128.5, 128.4, 128.3, 128.2, 127.9, 127.8, 127.7, 127.6, 87.3, 81.6, 78.9, 77.4, 75.6, 73.5, 73.4, 73.3, 72.5, 71.9, 69.8, 68.5, 64.0; MS ES $[\text{M}+\text{Na}]^+$ m/z 754.0; HRMS (ES-TOF $^+$) m/z calcd for $\text{C}_{43}\text{H}_{45}\text{N}_3\text{O}_6\text{SNa}$ $[\text{M}+\text{Na}]^+$ 754.2921, found 754.2918.

1-*O*-*p*-toluene-sulfonyl-(*S*)-2,3-di-*O*-benzylglycerol (13)

(*S*)-2,3-Di-*O*-benzyloxypropanol (700 mg, 2.57 mmol) was dissolved in dry DCM (10 mL) under nitrogen. To this stirred pale yellow solution was added Et_3N (522 μL , 3.5 mmol), *p*-TsCl (540 mg, 2.83 mmol) and DMAP (16.0 mg, 0.13 mmol). Stirring was continued at room temperature for 24 h whereupon TLC analysis indicated no starting material remained. DCM (10 mL) was added and the organics washed with 1M HCl, H_2O , saturated aqueous NaCl, dried (MgSO_4), filtered and evaporated. The crude material was then purified by silica gel flash chromatography eluting with hexane/EtOAc, 8/1, 4/1 to yield **13** (680 mg, 1.60 mmol, 62%) as clear oil. $R_f = 0.75$ (hexane/EtOAc, 2/1); $^1\text{H NMR}$ (400 MHz; CDCl_3) δ 7.80-7.78 (m, 2H, ArH), 7.36-7.26 (m, 12H, ArH), 4.59 (s, 2H, CH_2Ar), 4.49 (s, 2H, CH_2Ar), 4.24 (dd, $J = 10.4, 4.2$ Hz, 1H, CH_2OTs), 4.14 (dd, $J = 10.4, 5.8$ Hz, 1H, CH_2OTs), 3.82-3.80 (m, 1H, CHOBn), 3.54 (dd, $J = 5.2, 0.7$ Hz, 2H, CH_2OBn), 2.44 (s, 3H, ArCH_3); MS ES $[\text{M}+\text{Na}]^+$ m/z 449.0; HRMS (ES-TOF $^+$) m/z calcd for $\text{C}_{24}\text{H}_{26}\text{O}_5\text{NaS}$ $[\text{M}+\text{Na}]^+$ 449.1393, found 449.1390. Other analytical data matched those previously reported.³

1-*O*-trifluoromethanesulfonyl-(*S*)-2,3-di-*O*-benzyl-glycerol (14)

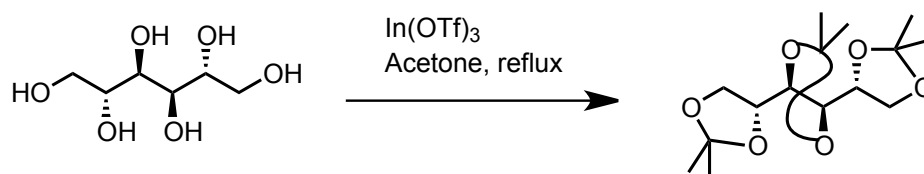
(*S*)-2,3-Di-*O*-benzyloxypropanol (3.0 g, 11.0 mmol) was dissolved in dry DCM (25 mL) under nitrogen. To this stirred pale yellow solution was added lutidine (2.10 mL, 17.6 mmol) and Tf_2O (2.80 mL, 16.5 mmol) at 0 °C. Stirring was continued for 1 h whereupon TLC analysis indicated no starting material remained. The volume of DCM was reduced to *ca* 5 mL *in vacuo*, the material loaded onto a short silica plug and eluted with hexane/EtOAc, 20/1 to yield **14** (4.40 g, 11.0 mmol, quant.) as clear oil. $R_f = 0.85$ (hexane/EtOAc, 3/1); $^1\text{H NMR}$ (400 MHz; CDCl_3) δ 7.41-7.32 (m, 10H, ArH), 4.71-4.60 (m, 4H, CH_2Ar , CH_2OTf), 4.59-4.52 (m, 2H, CH_2Ar), 3.93-3.88 (m, 1H, CH), 3.65-3.56 (m, 2H, CH_2OBn). The

material was stored under nitrogen in a freezer until use. Other analytical data matched those previously reported.⁴

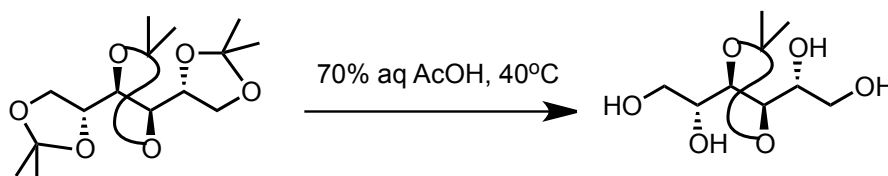
References:

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2. K. C. Etika, F. D. Jochum, M. A. Cox, P. Schattling, P. Theato and J. C. Grunlan, *Macromolecules*, 2010, **43**, 9447.
3. J. F. Tocanne, H. M. Verheij, J. A. F. Op Den Kamp and L. L. M. Van Deenen, *Chemistry and Physics of Lipids*, 1974, **13**, 389.
4. A. Esswein, H. Rembold and R. R. Schmidt, *Carbohydr. Res.*, 1990, **287**, 305.

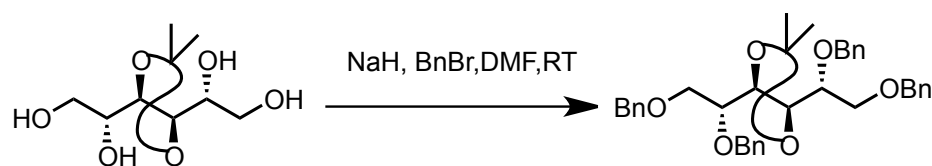
Synthesis of (S)-2,3-dibenzyloxy-1-propanol from D-mannitol



D-mannitol (5.0 g, 27.4 mmol) and $\text{In}(\text{OTf})_3$ (462 mg, 0.82 mmol) were suspended in acetone (50 mL) at RT under air. The suspension was heated to 65 °C for 3 h whereupon the suspension became clear. TLC analysis (DCM/MeOH, 4/1) showed the reaction to be complete and the solvent was removed in vacuo. The resulting white paste was dissolved in DCM (200 mL) and washed with water (50 mL). The organic phase was dried (MgSO_4), filtered and stripped to a clear oil which solidified upon high vacuum removal of final solvent traces to give the tris-dimethylacetal product (6.48 g, 21.5 mmol, 78%) as a white solid.

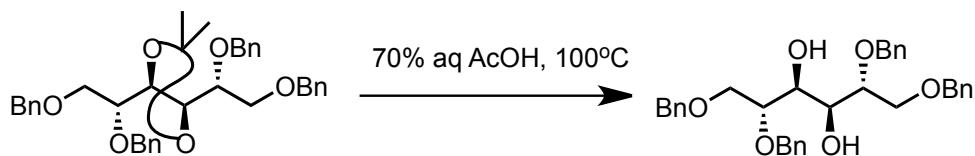


Tris-dimethylacetal-D-mannitol derivative (4.4 g, 14.6 mmol) was dissolved in MeOH (50 mL) at RT under air. 0.5M H_2SO_4 (5 mL) was added and the solution heated at 40 °C for 2 h. TLC analysis (hexane/EtOAc, 3/1) showed no starting material remained and the reaction was cooled to RT, neutralized to pH = 8 with 1M NaOH, the suspension filtered and solvent removed in vacuo. The crude oil was then purified by silica gel flash chromatography eluting with DCM/MeOH, 9/1 to furnish the product tetra-ol (2.18 g, 9.8 mmol, 67%) as a white solid.

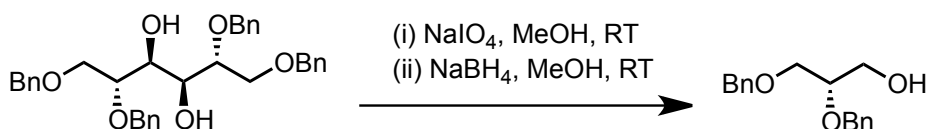


Tetra-ol derivative (2.20 g, 9.9 mmol) was dissolved in dry THF (50 mL) at RT under N_2 . The solution was cooled to 0 °C in an ice bath and NaH (1.98 g, 50.0 mmol, 60% in mineral oil) added portionwise over 15 minutes. The suspension was stirred at 0 °C for 30 minutes, BnBr (11.8 mL, 99.1 mmol) and TBAI (3.66 g, 9.91 mmol) were then added and the solution warmed to RT and stirred overnight. TLC analysis (DCM/MeOH, 4/1) showed no starting material and one product spot. The reaction solution was cooled once again to 0 °C and quenched with water (10 mL). This solution was then poured onto diethyl ether (100 mL) and the layers separated. The aqueous phase was extracted with diethyl ether (3 x 20

mL), the organics combined, dried (MgSO_4), filtered and stripped to a brown oil. This crude material was then purified by silica gel flash chromatography eluting firstly with hexane/EtOAc, 6/1 and then with hexane/diethyl ether, 4/1 to give the tetrabenzylated product (5.42 g, 9.30 mmol, 93%) as a pale pink oil.



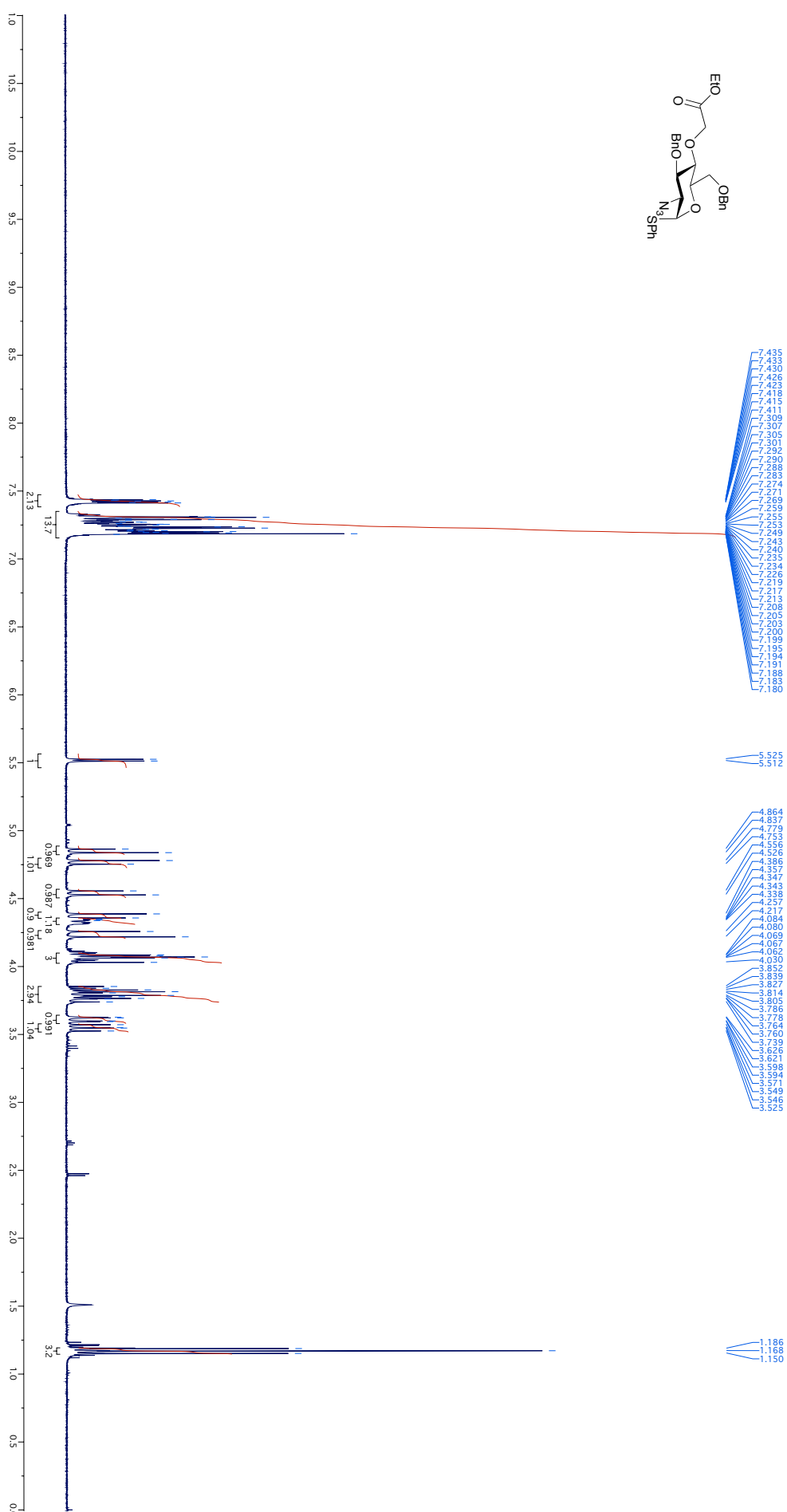
The tetrabenzylated-D-mannitol derivative (4.24 g, 7.3 mmol) was suspended in 60% aq. AcOH (40 mL) and heated to reflux overnight. TLC analysis (hexane/EtOAc, 4/1) showed no starting material and one product spot. The solvent was removed in vacuo azeotroping with toluene (2 x 100 mL) to give the crude material as a yellow oil which was purified by silica gel flash chromatography, eluting with hexane/EtOAc, 4/1, 2/1 to give the product 3,4-diol (2.85 g, 5.25 mmol, 72%) as a clear oil.



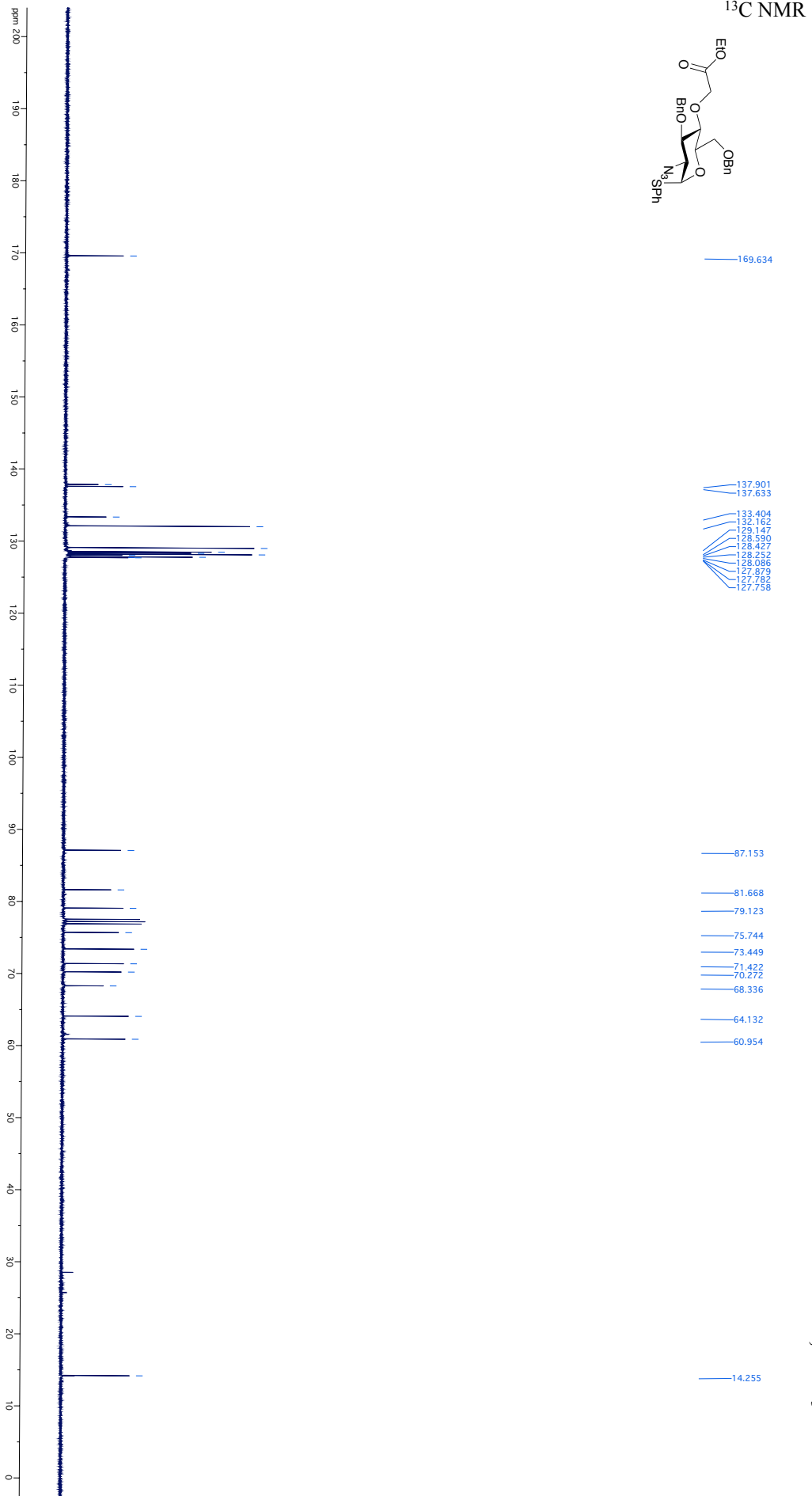
The D-mannitol-3,4-diol (2.70 g, 4.98 mmol) was dissolved in MeOH (50 mL) and cooled to 0 °C. NaIO_4 (1.20 g, 5.63 mmol) was added and the thick white suspension stirred vigorously for 30 minutes, filtered through Celite, washing with MeOH (25 mL) and then re-cooled to 0 °C whereupon NaBH_4 (259 mg, 4.48 mmol) was added and the suspension stirred for a further 2 h at 0 °C. TLC analysis (hexane/EtOAc, 3/1) showed a spot that matched the commercially available material and the reaction was quenched at 0 °C with glacial acetic acid. The suspension was filtered through Celite and the majority of the MeOH removed in vacuo. The residue was partitioned between EtOAc (100 mL) and brine (25 mL). The layers were separated and the organics extracted with EtOAc (3 a 25 mL), combined, dried (MgSO_4), filtered and solvent removed in vacuo to give the crude material as a yellow oil. This was purified by silica gel flash chromatography eluting with hexane/EtOAc, 4/1 to give (S)-2,3-dibenzyoxy-1-propanol (1.86 g, 6.8 mmol, 69%) as a clear oil. ^1H NMR and optical rotation data matched those available from a commercial supplier (Sigma Aldrich).

2. Spectral Data: ^1H , ^{13}C , COSY, HMQC/HSQC NMR and MS data for compounds 2-27

Phenyl 2-azido-3,6-di-O-benzyl-4-O-acetoxyethyl-2-deoxy-1-thio- α -D-glucopyranoside (α -2)

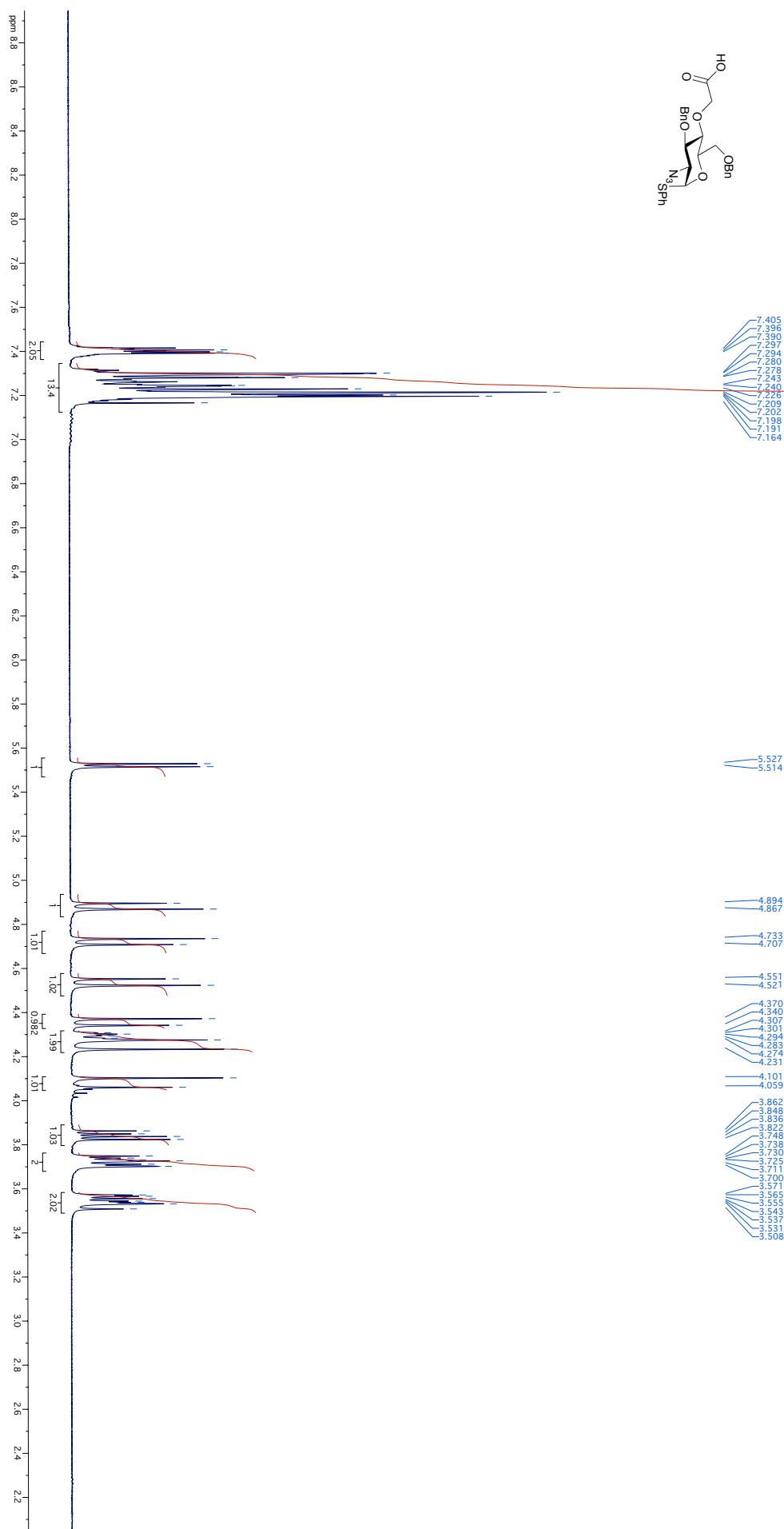


Phenyl 2-azido-3,6-di-O-benzyl-4-O-acetoxyethyl-2-deoxy-1-thio- α -D-glucopyranoside (α -2)



Phenyl 2-azido-3,6-di-O-benzyl-4-O-acetoxy-2-deoxy-1-thio- α -D-glucopyranoside (α -3)

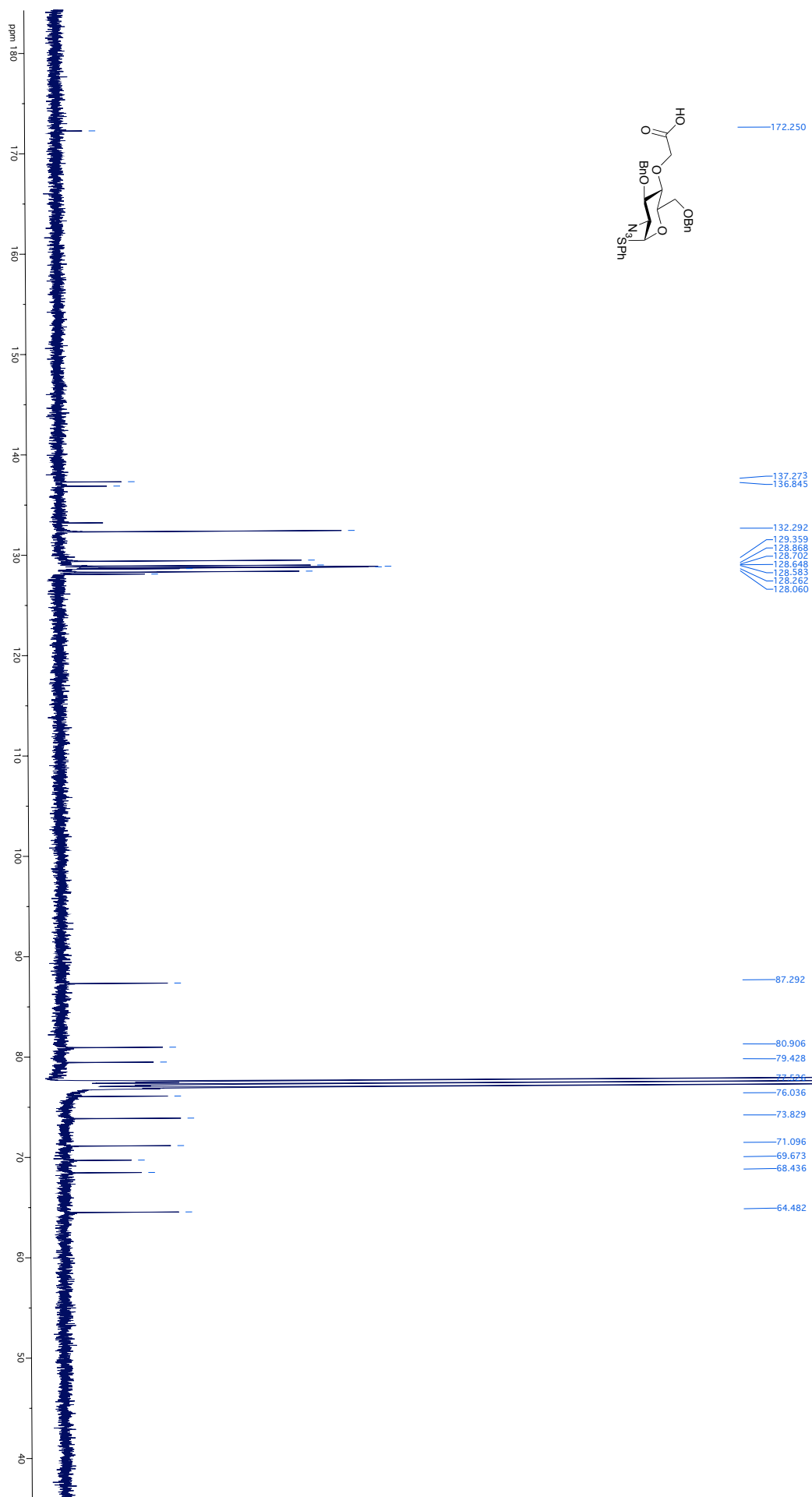
$^1\text{H NMR}$



400 MHz, CDCl_3

Phenyl 2-azido-3,6-di-O-benzyl-4-O-acetoxy-2-deoxy-1-thio- α -D-glucopyranoside (α -3)

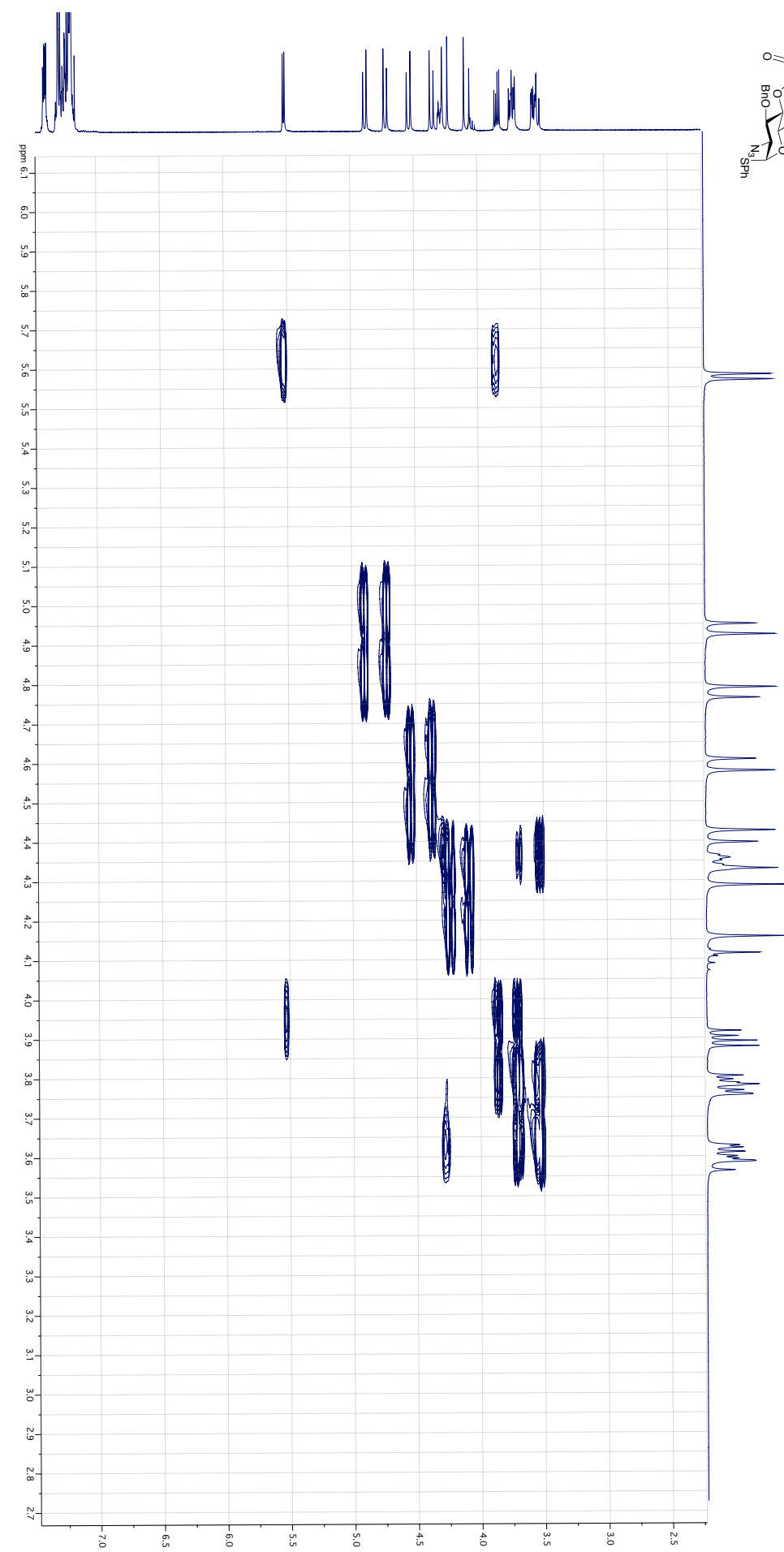
^{13}C NMR



100 MHz, CDCl_3

Phenyl 2-azido-3,6-di-O-benzyl-4-O-acetoxy-2-deoxy-1-thio- α -D-glucopyranoside (α -3)

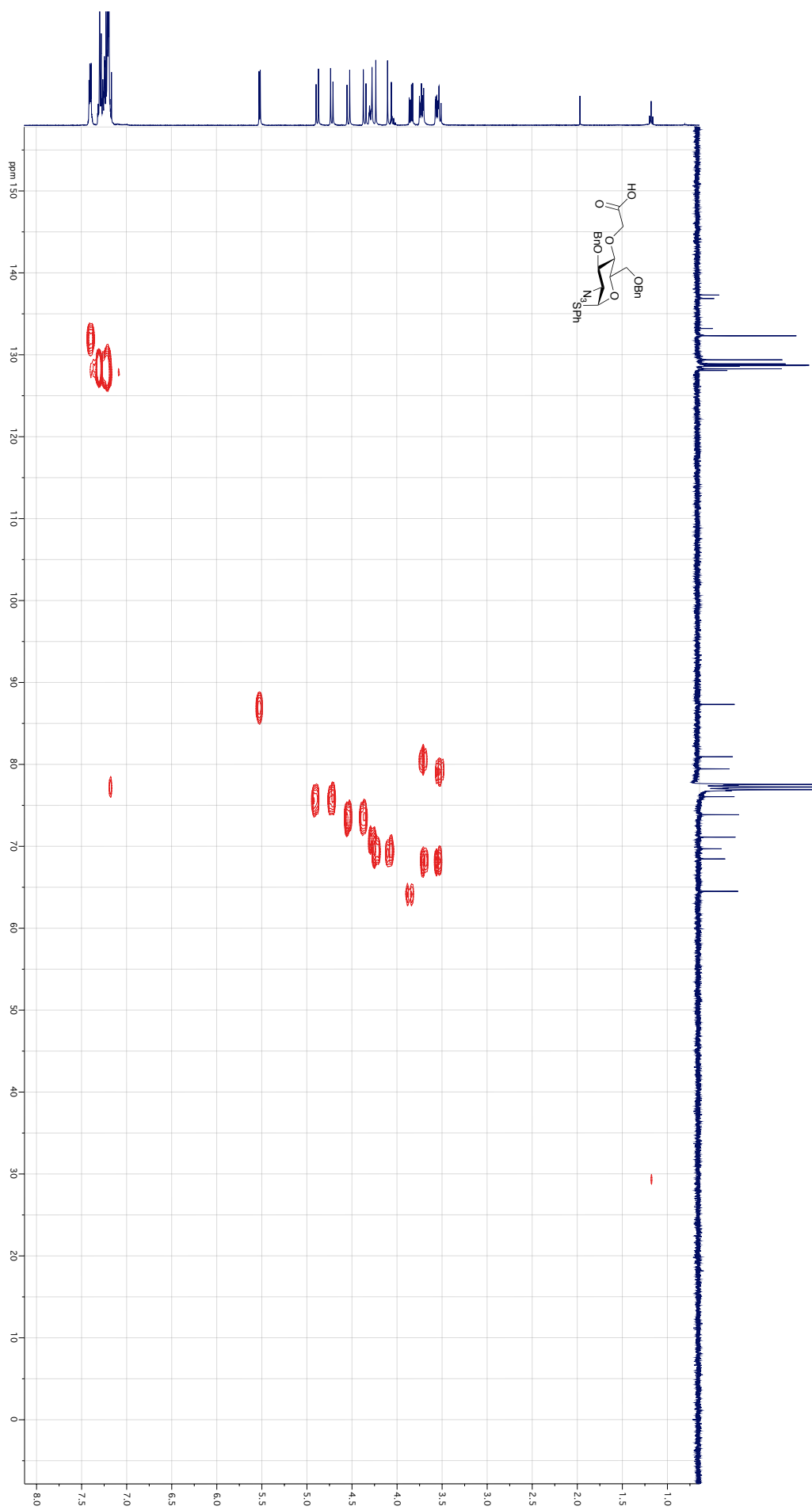
COSY



400 MHz, CDCl₃

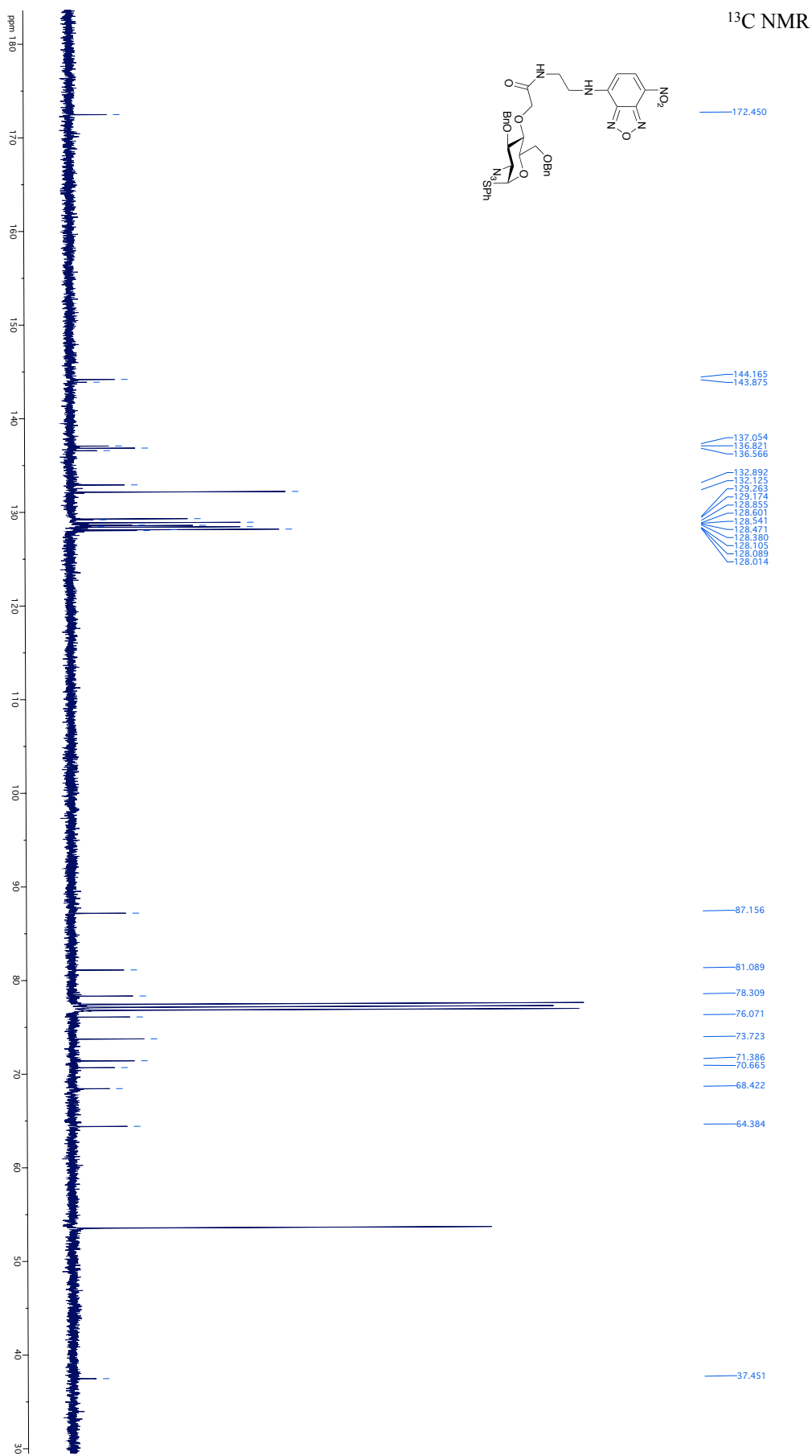
Phenyl 2-azido-3,6-di-O-benzyl-4-O-acetoxy-2-deoxy-1-thio- α -D-glucopyranoside (α -3)

HMQC



400 MHz, CDCl_3

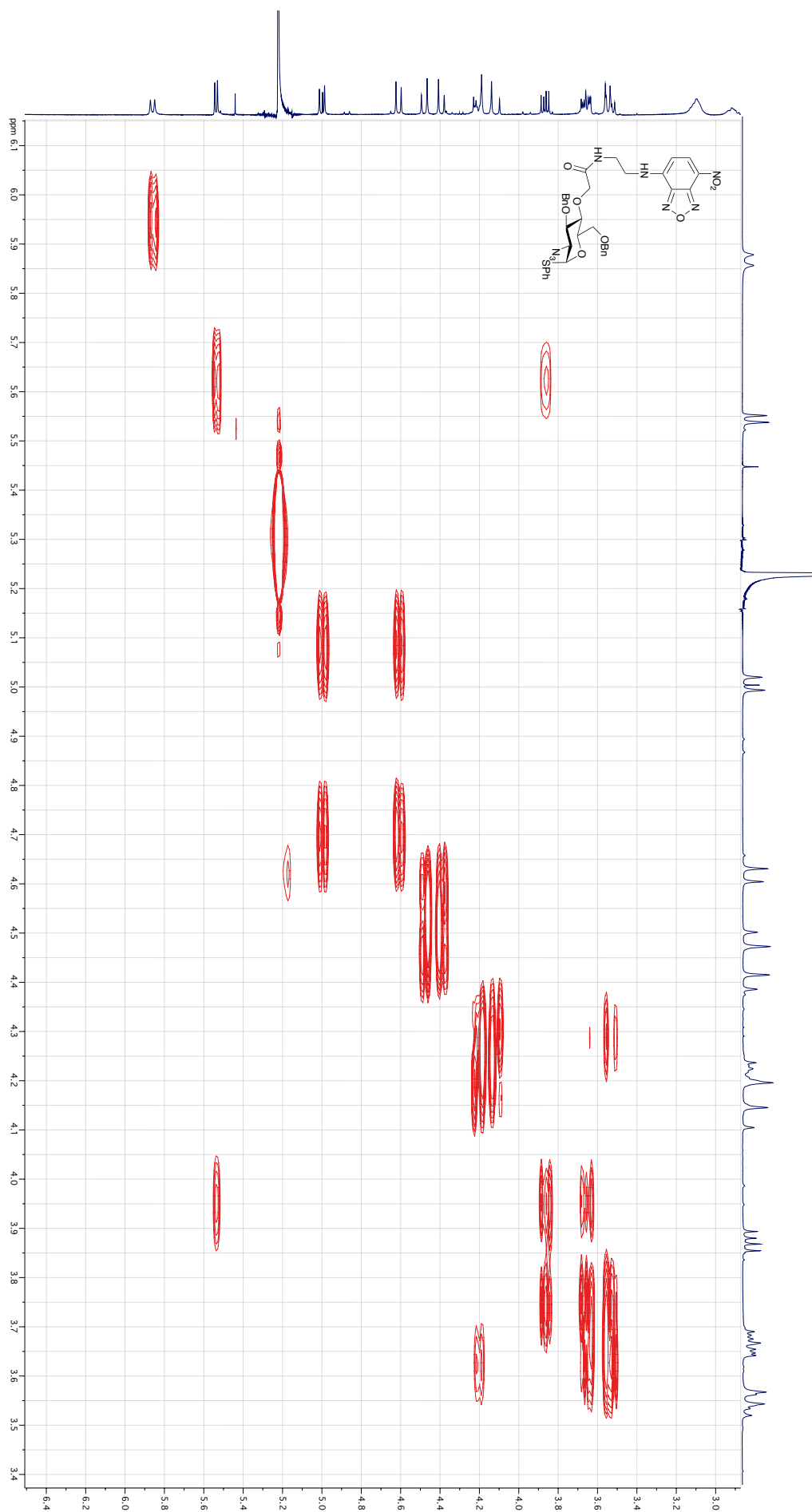
NBD-D-GlcN derivative (6)



100 MHz, CDCl₃

NBD-D-GlcN derivative (6)

COSY

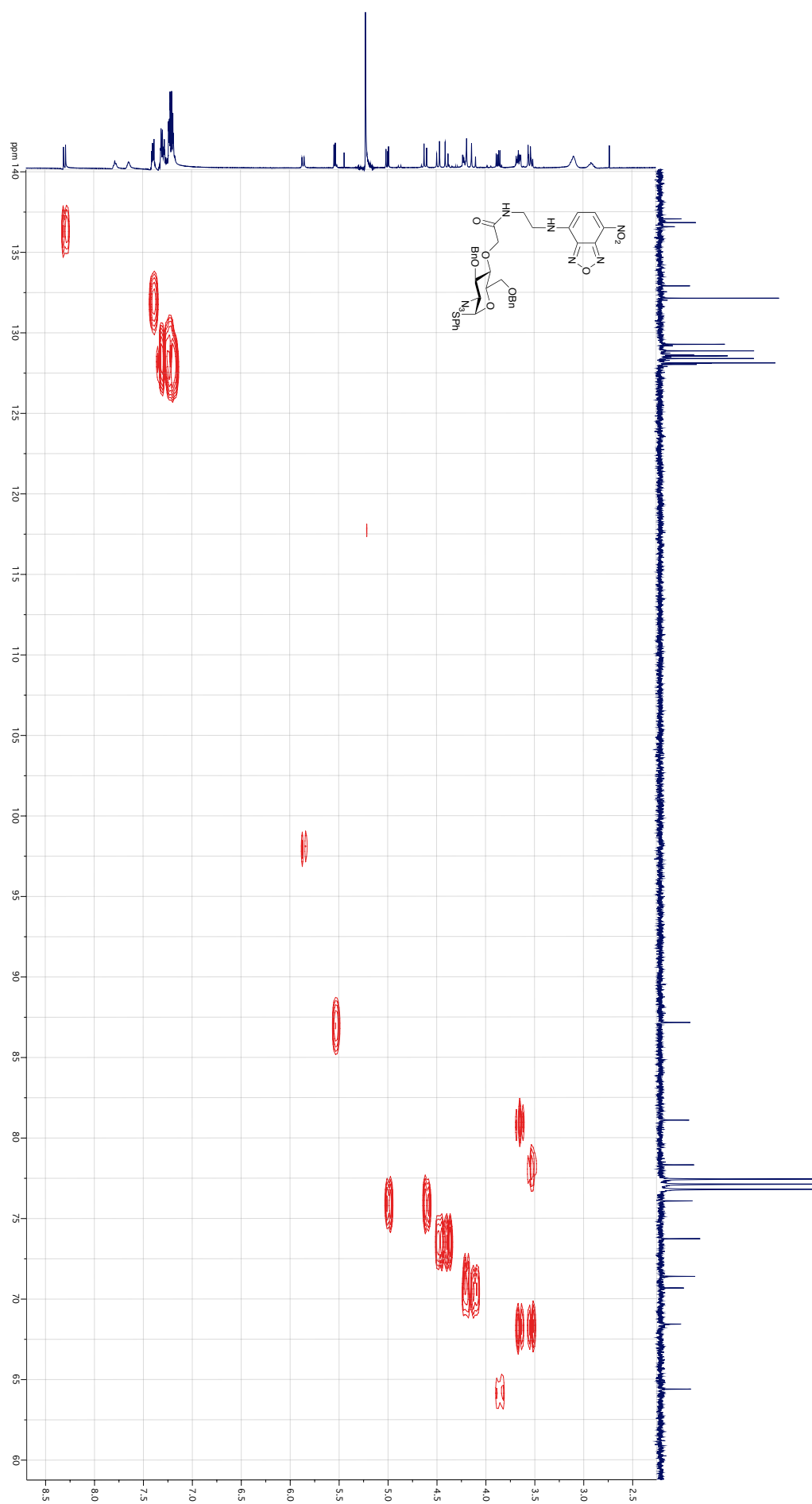


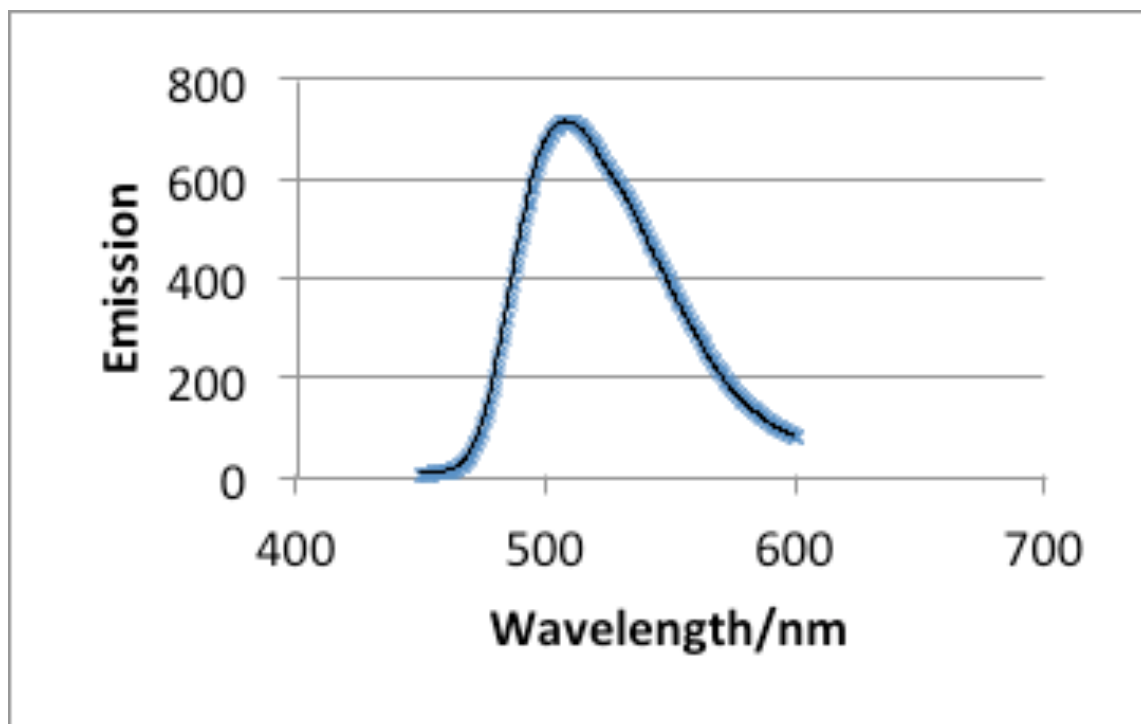
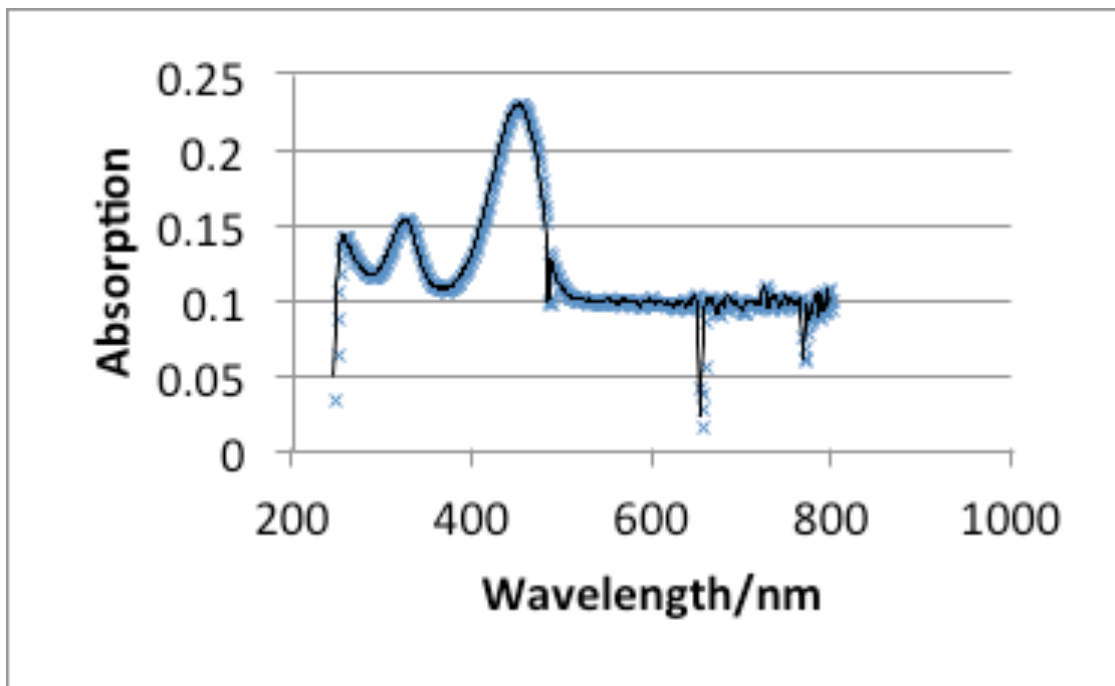
400 MHz, CDCl₃

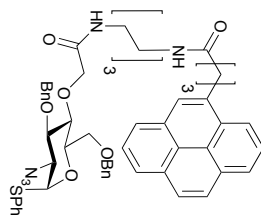
NBD-D-GlcN derivative (6)

HMQC

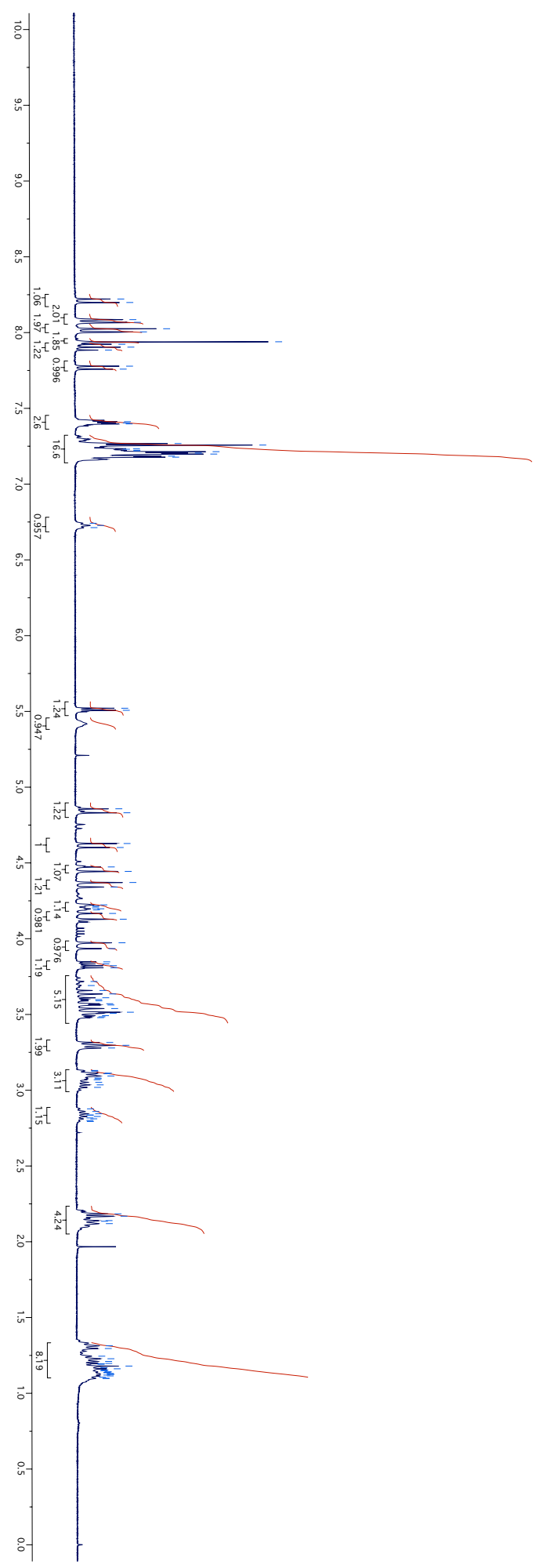
400 MHz, CDCl₃





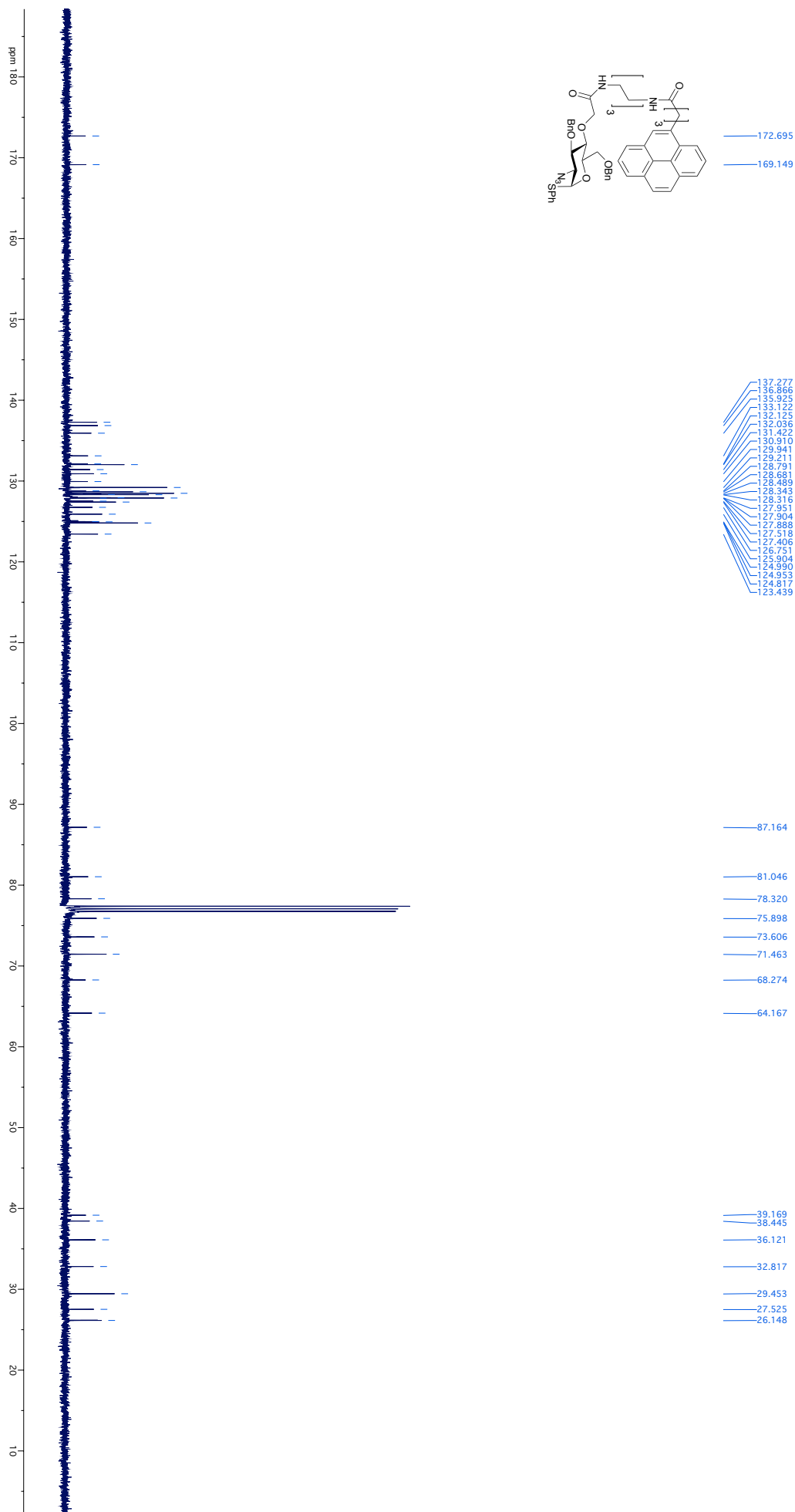


- 8.220
- 8.197
- 8.064
- 8.066
- 8.024
- 8.003
- 7.936
- 7.921
- 7.903
- 7.883
- 7.777
- 7.758
- 7.411
- 7.402
- 7.396
- 7.266
- 7.255
- 7.232
- 7.229
- 7.222
- 7.219
- 7.212
- 7.203
- 7.196
- 7.182
- 7.175
- 6.741
- 6.726
- 6.712
- 5.520
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- 4.856
- 4.830
- 4.627
- 4.601
- 4.472
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- 4.214
- 4.213
- 4.202
- 4.197
- 4.189
- 4.186
- 4.127
- 3.972
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- 3.717
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- 3.125
- 3.113
- 3.108
- 3.093
- 3.080
- 3.070
- 3.068
- 3.050
- 3.034
- 3.018
- 2.876
- 2.859
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- 2.836
- 2.826
- 2.816
- 2.811
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- 2.793
- 2.783
- 2.767
- 2.738
- 2.734
- 2.719
- 1.312
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- 1.284
- 1.227
- 1.209
- 1.196
- 1.193
- 1.193
- 1.178
- 1.168
- 1.161
- 1.153
- 1.150
- 1.140
- 1.140
- 1.137
- 1.134
- 1.129
- 1.127
- 1.125
- 1.124
- 1.119
- 1.114
- 1.105
- 1.098
- 1.097



Pyrene-D-GlcN derivative (7)

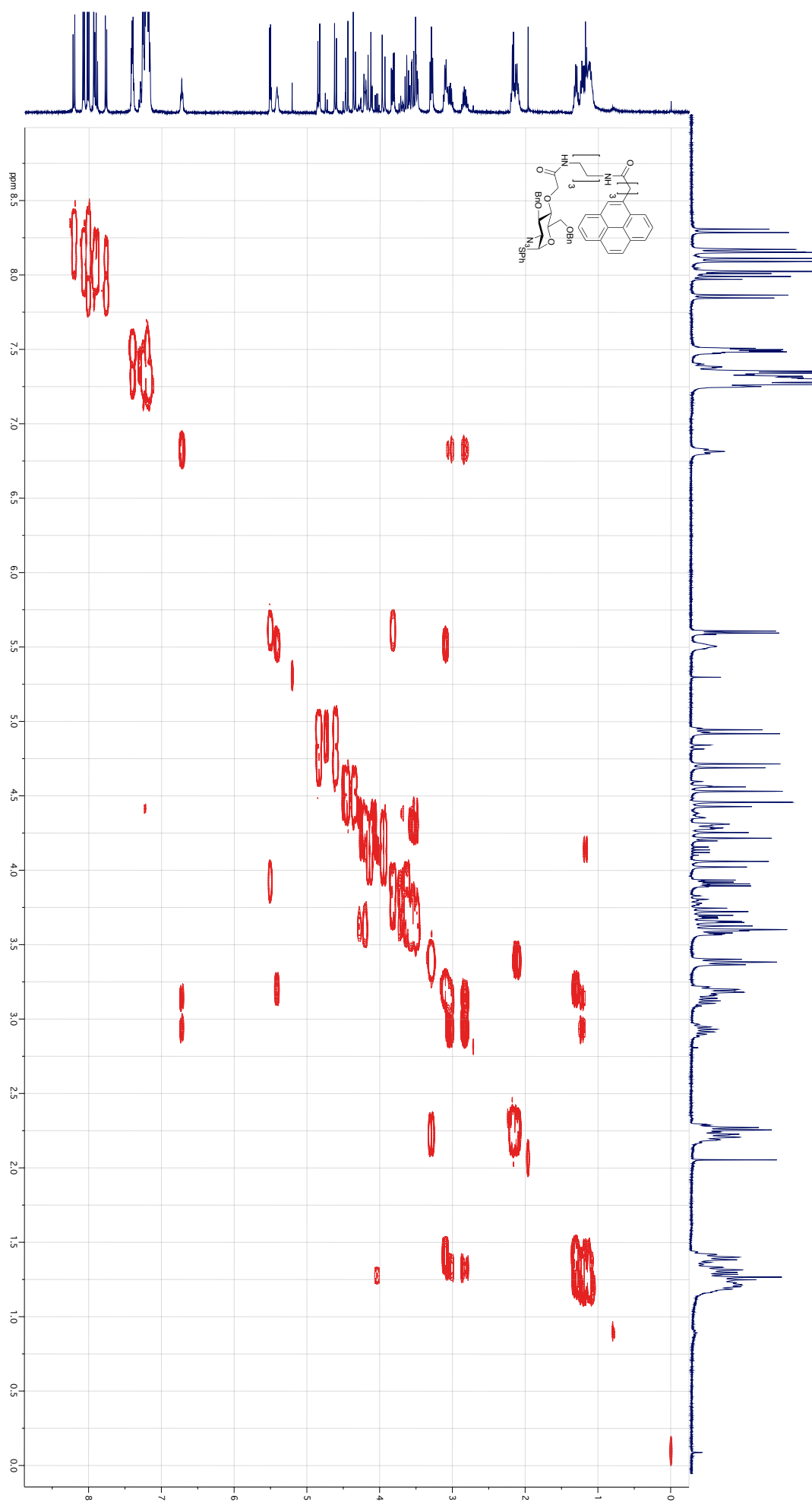
^{13}C NMR



100 MHz, CDCl₃

Pyrene-D-GlcN derivative (7)

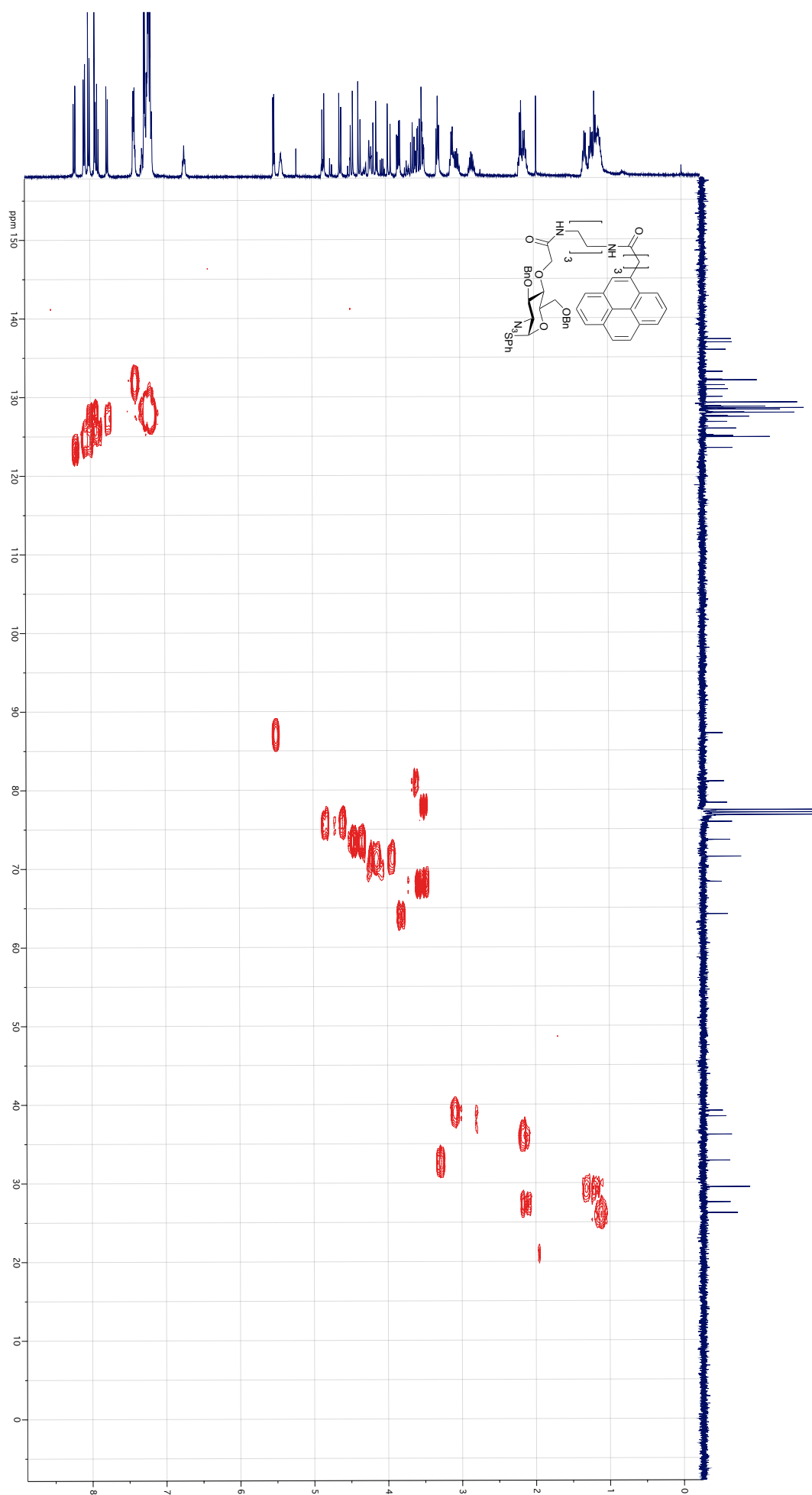
COSY



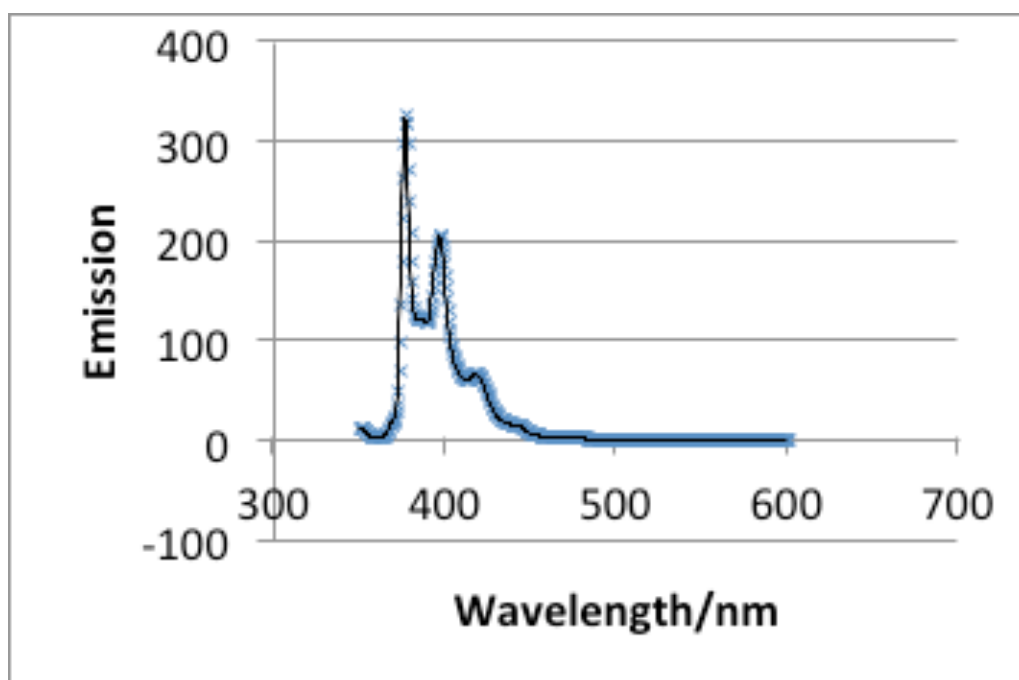
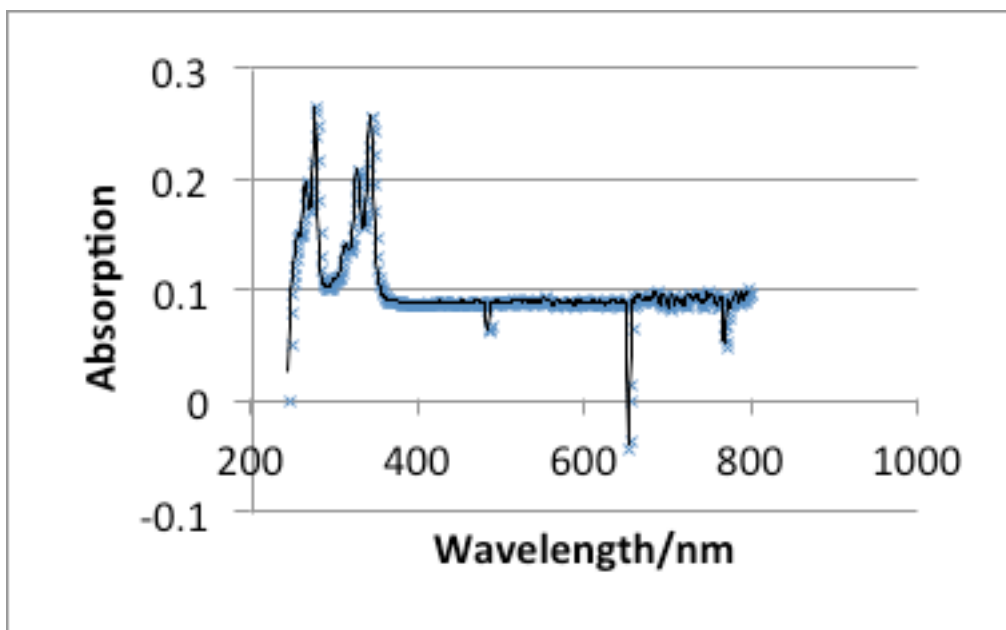
400 MHz, CDCl₃

Pyrene-D-GlcN derivative (7)

HMQC

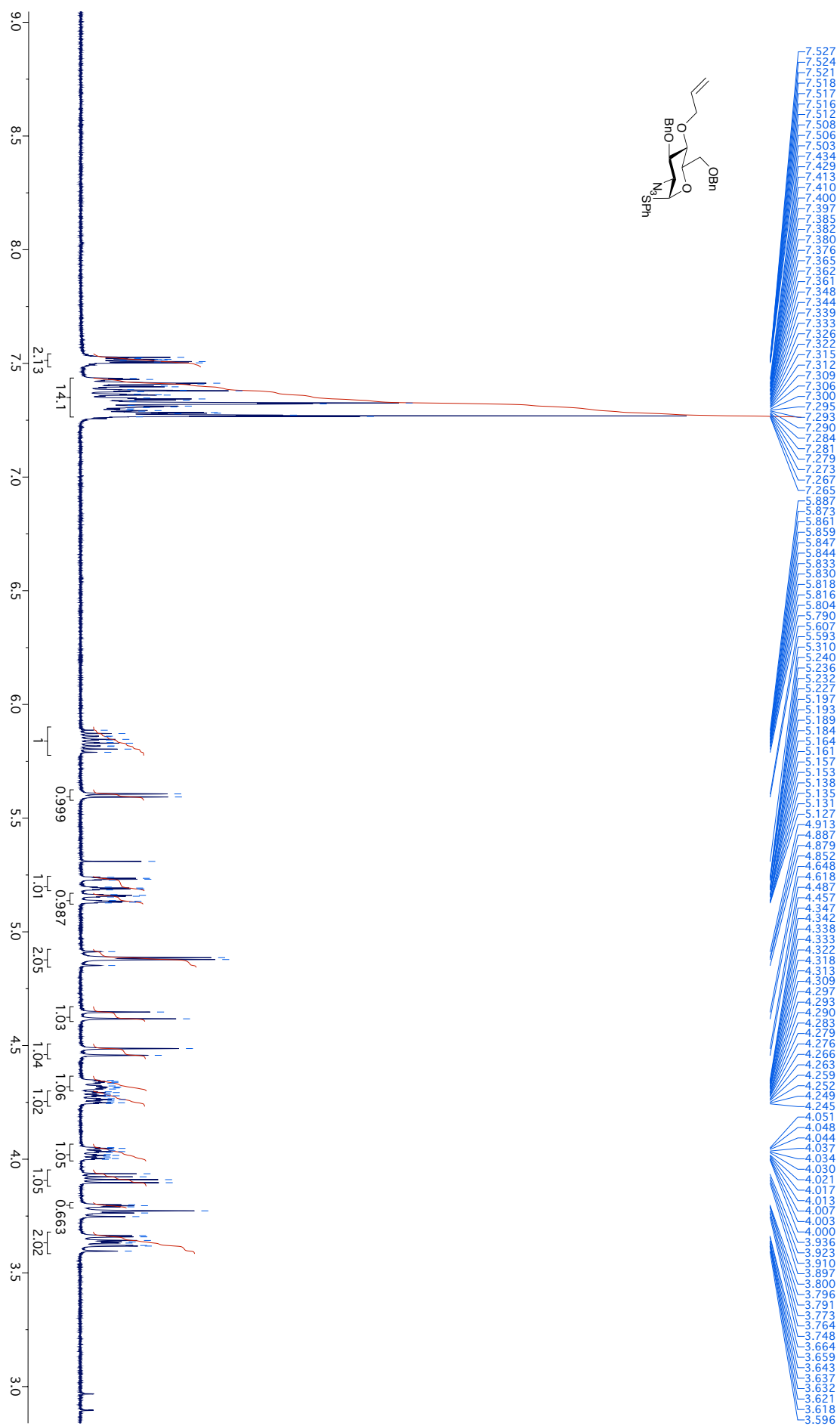


400 MHz, CDCl₃



Phenyl-4-O-allyl-2-azido-2-deoxy-3,6-di-O-benzyl-1-thio- α -D-glucopyranoside (10)

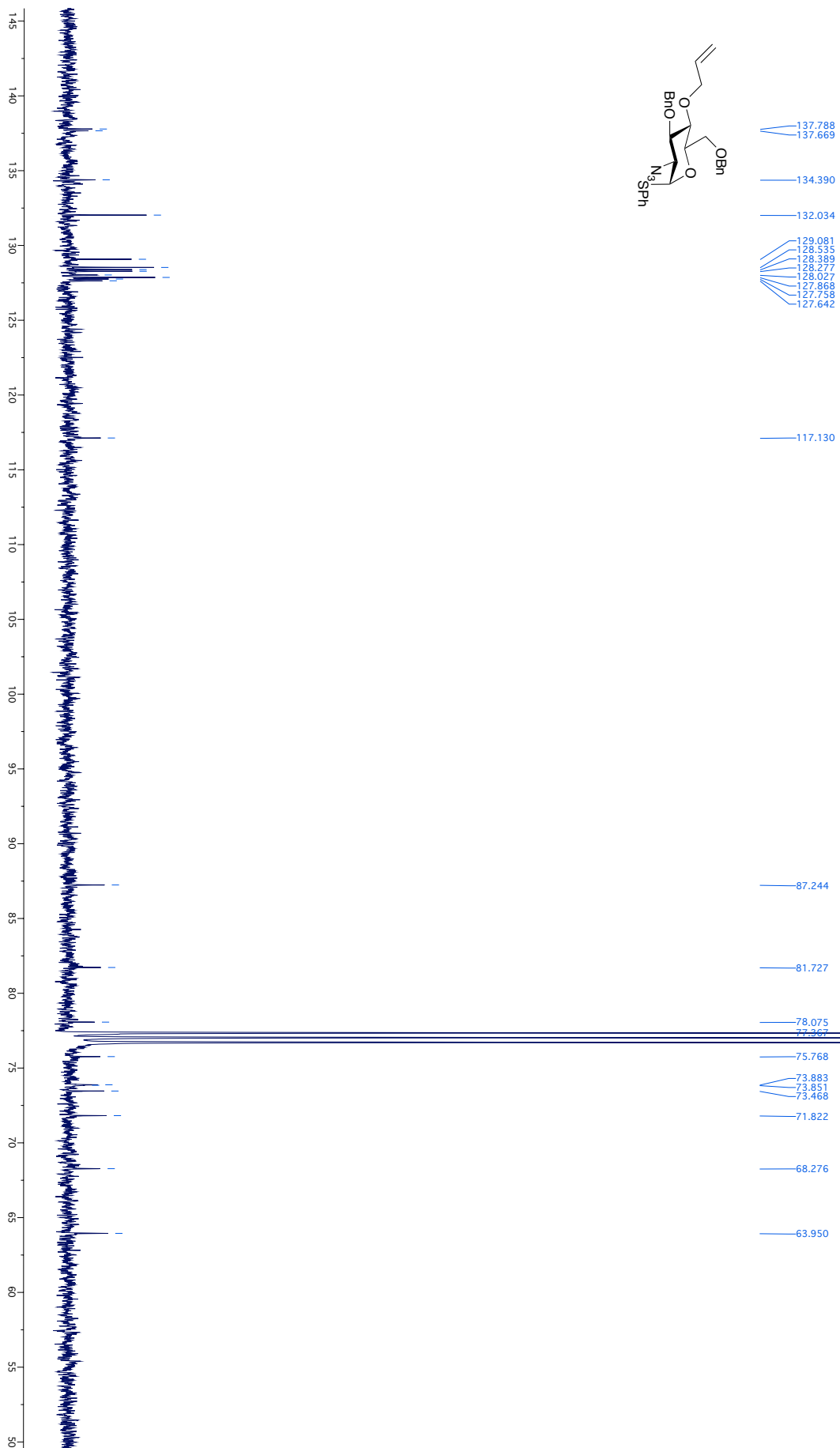
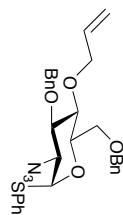
^1H NMR



400 MHz, CDCl_3

Phenyl-4-O-allyl-2-azido-2-deoxy-3,6-di-O-benzyl-1-thio- α -D-glucopyranoside (10)

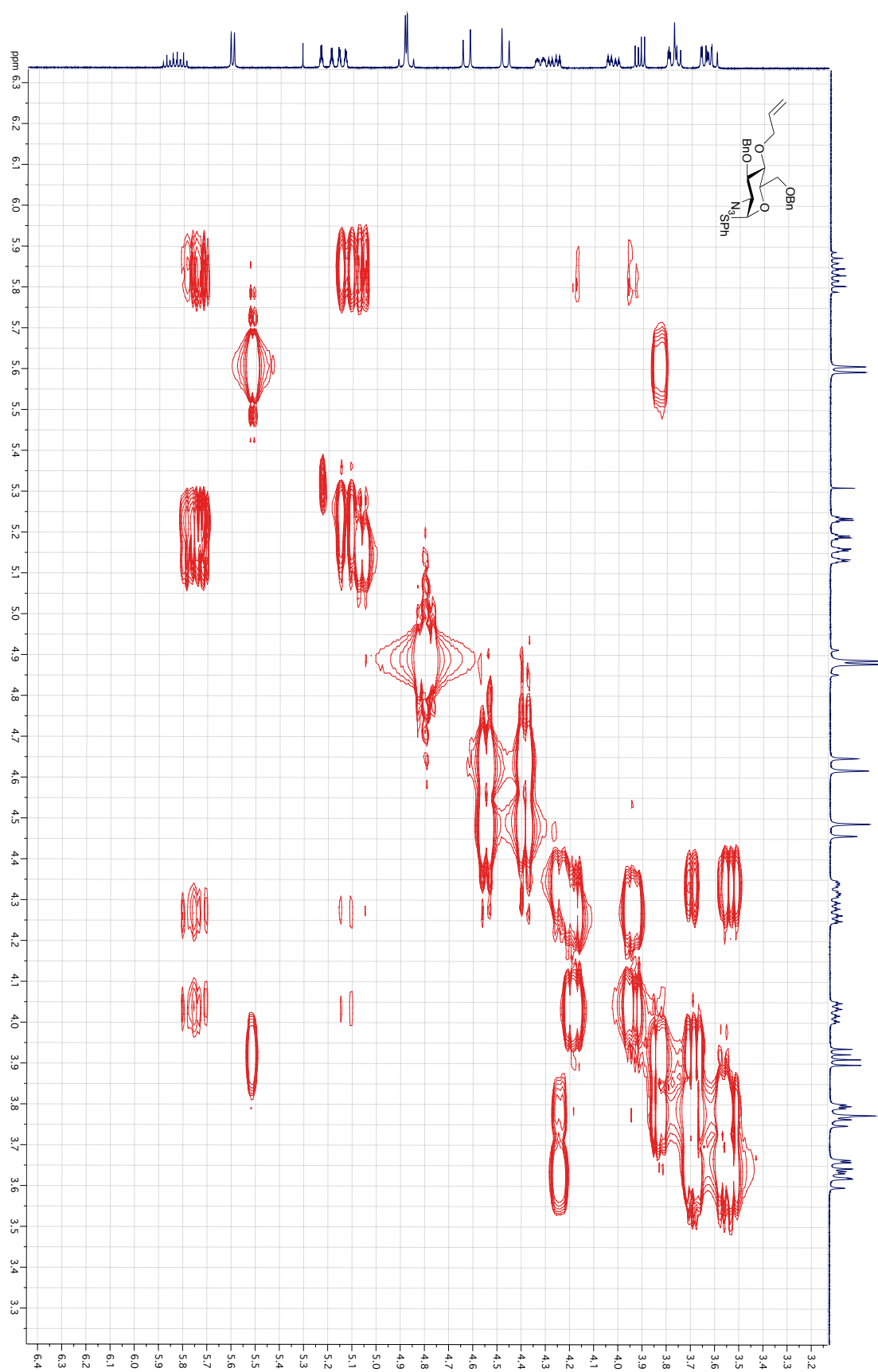
^{13}C NMR

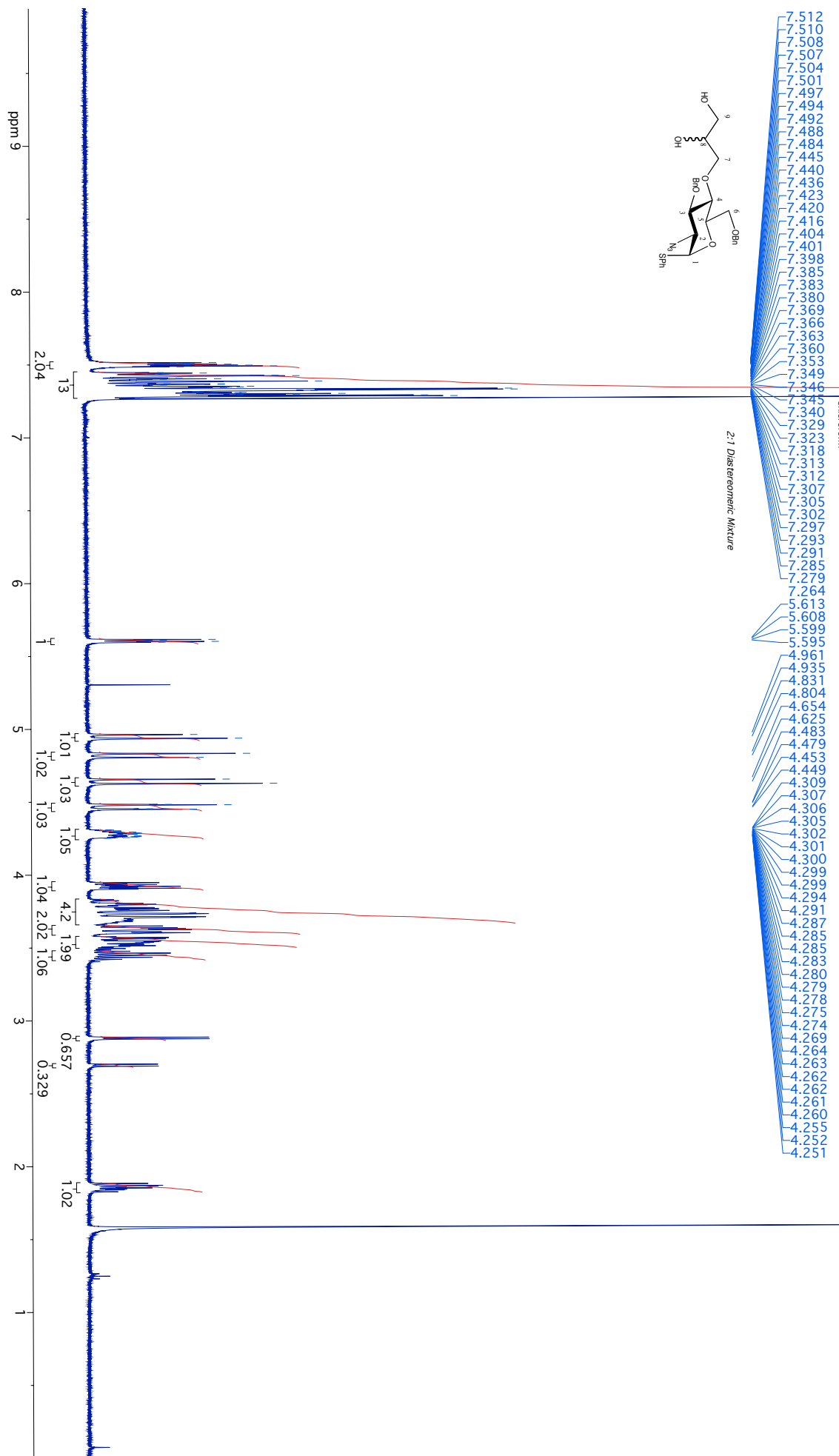


100 MHz, CDCl₃

Phenyl-4-O-allyl-2-azido-2-deoxy-3,6-di-O-benzyl-1-thio- α -D-glucopyranoside (10)

COSY

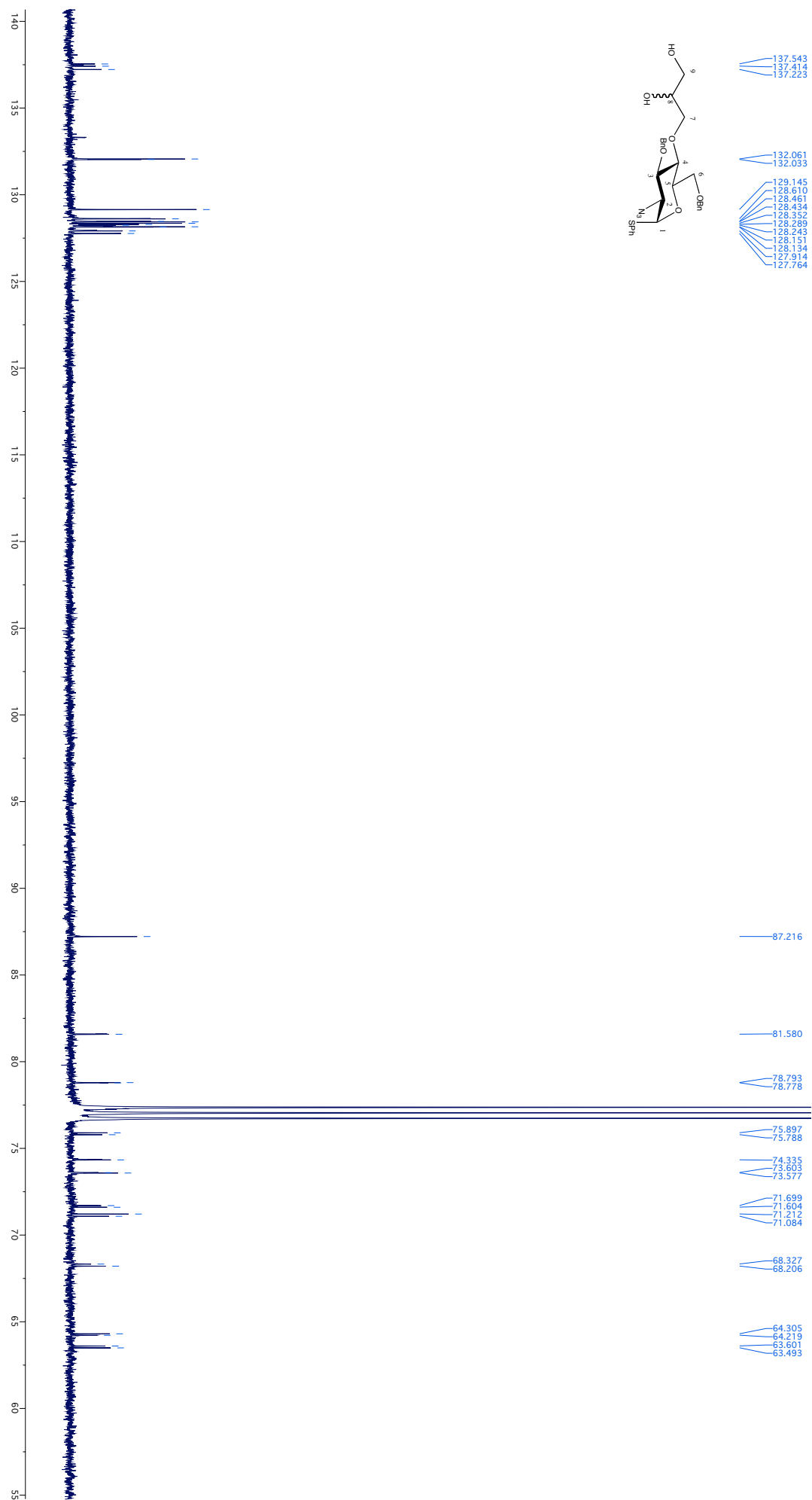




400 MHz, CDCl_3

Phenyl-2-azido-2-deoxy-3,6-di-O-benzyl-4-O-[(*S/R*)-2,3-dihydroxypropoxy]-1-thio- α -D-glucopyranoside (11)

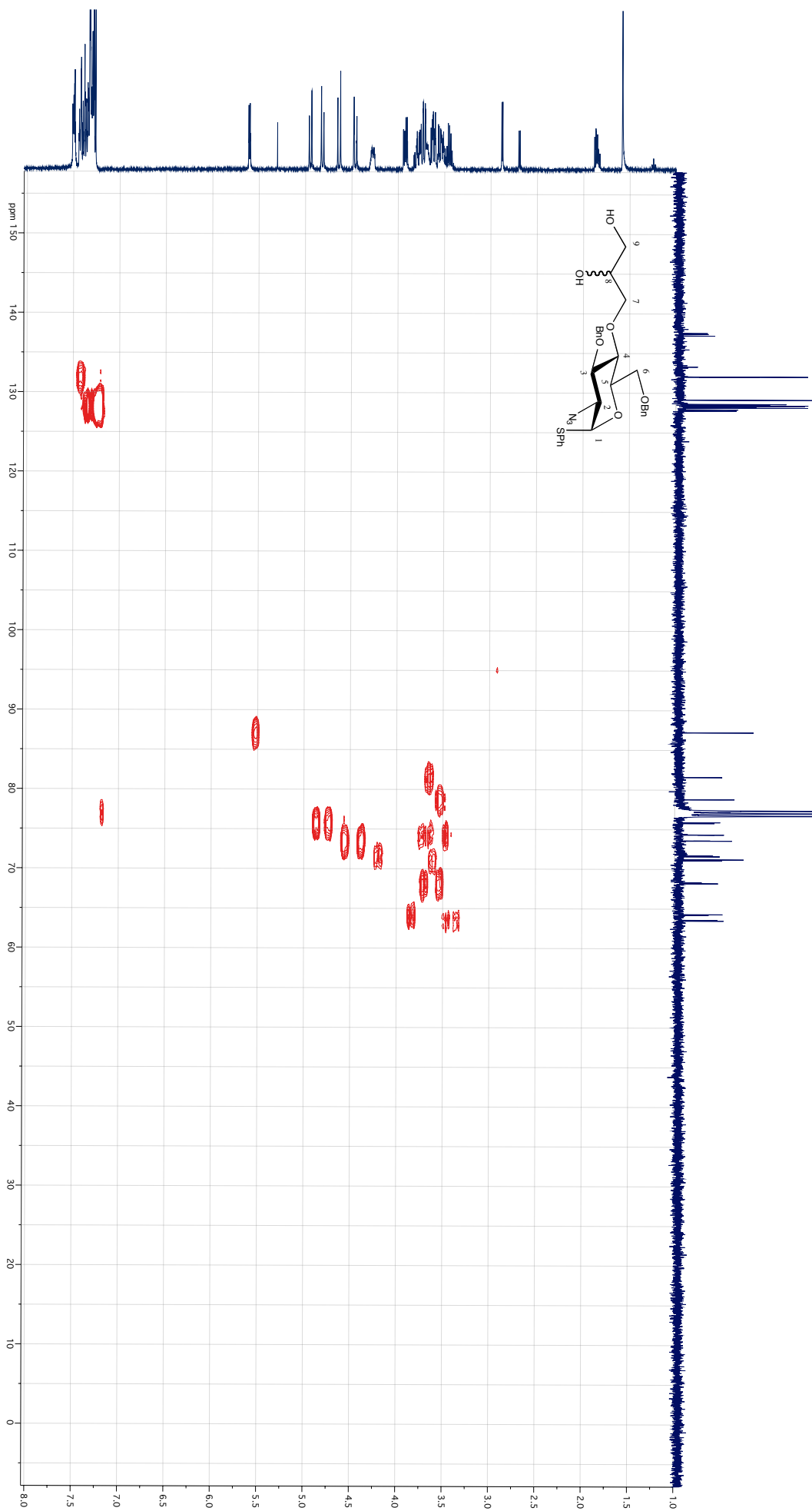
^{13}C NMR



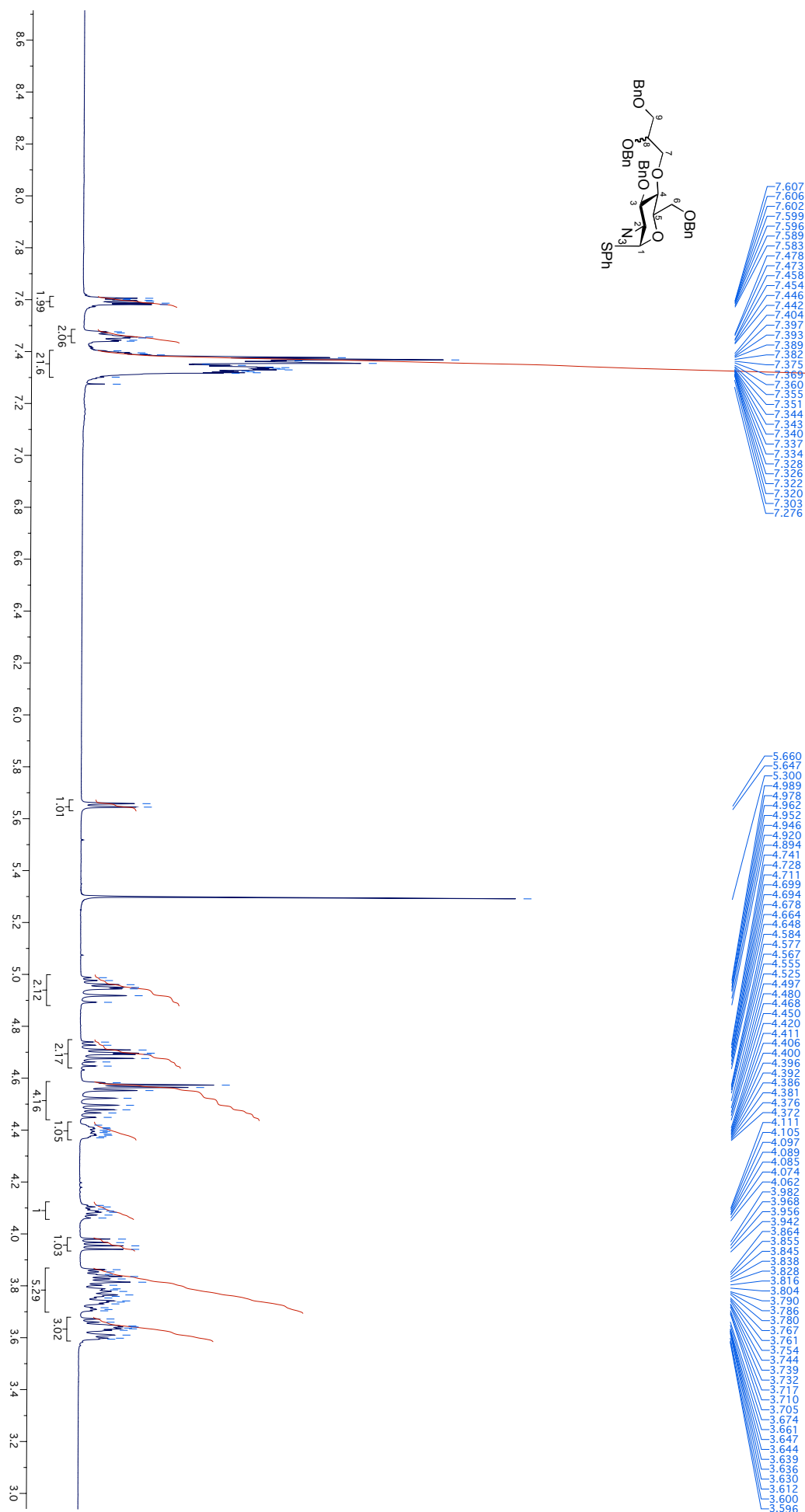
100 MHz, CDCl₃

Phenyl-2-azido-2-deoxy-3,6-di-O-benzyl-4-O-[(S/R)-2,3-dihydroxypropoxy]-1-thio- α -D-glucopyranoside (11)

HMQC



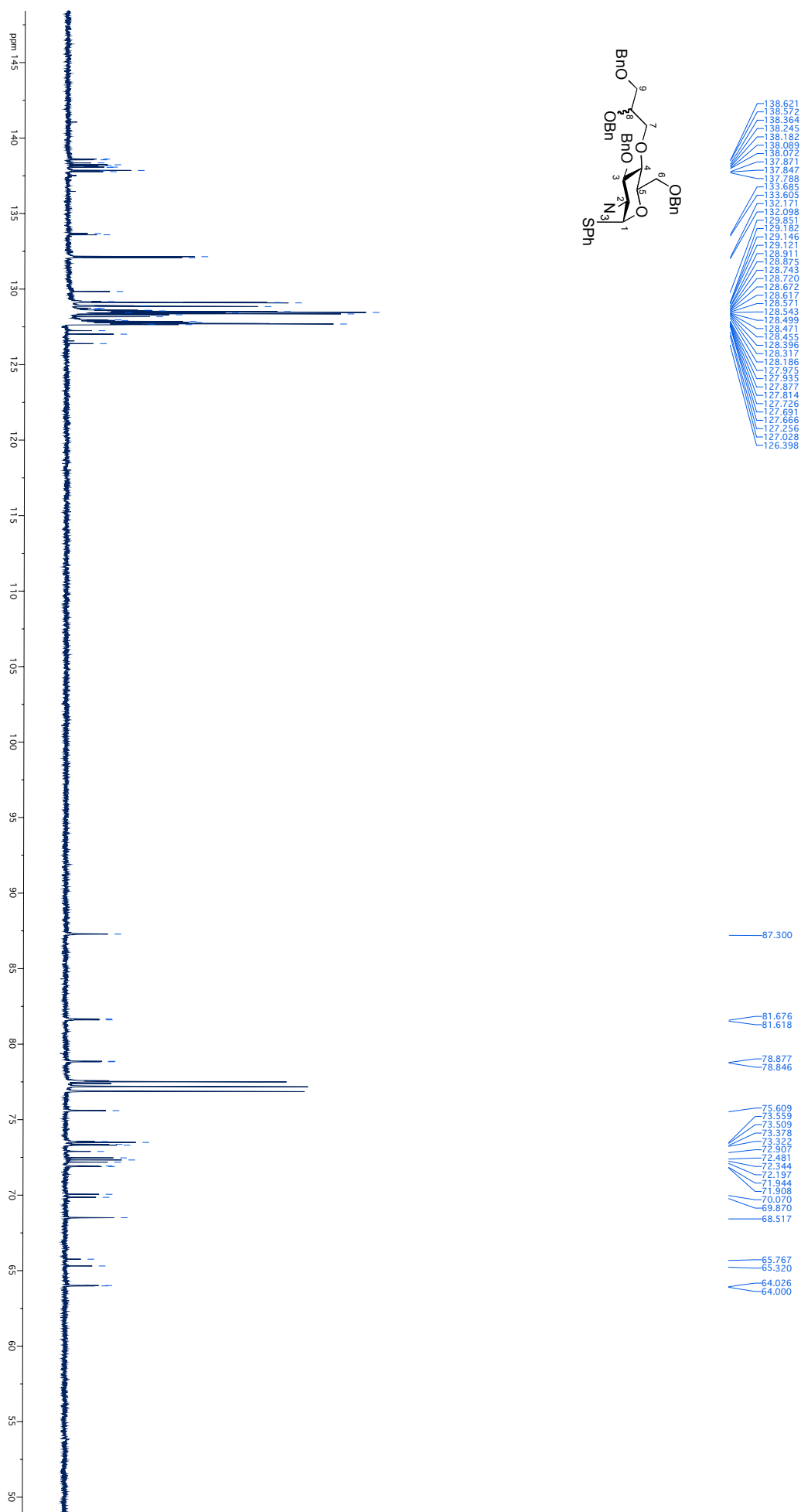
400 MHz, CDCl₃



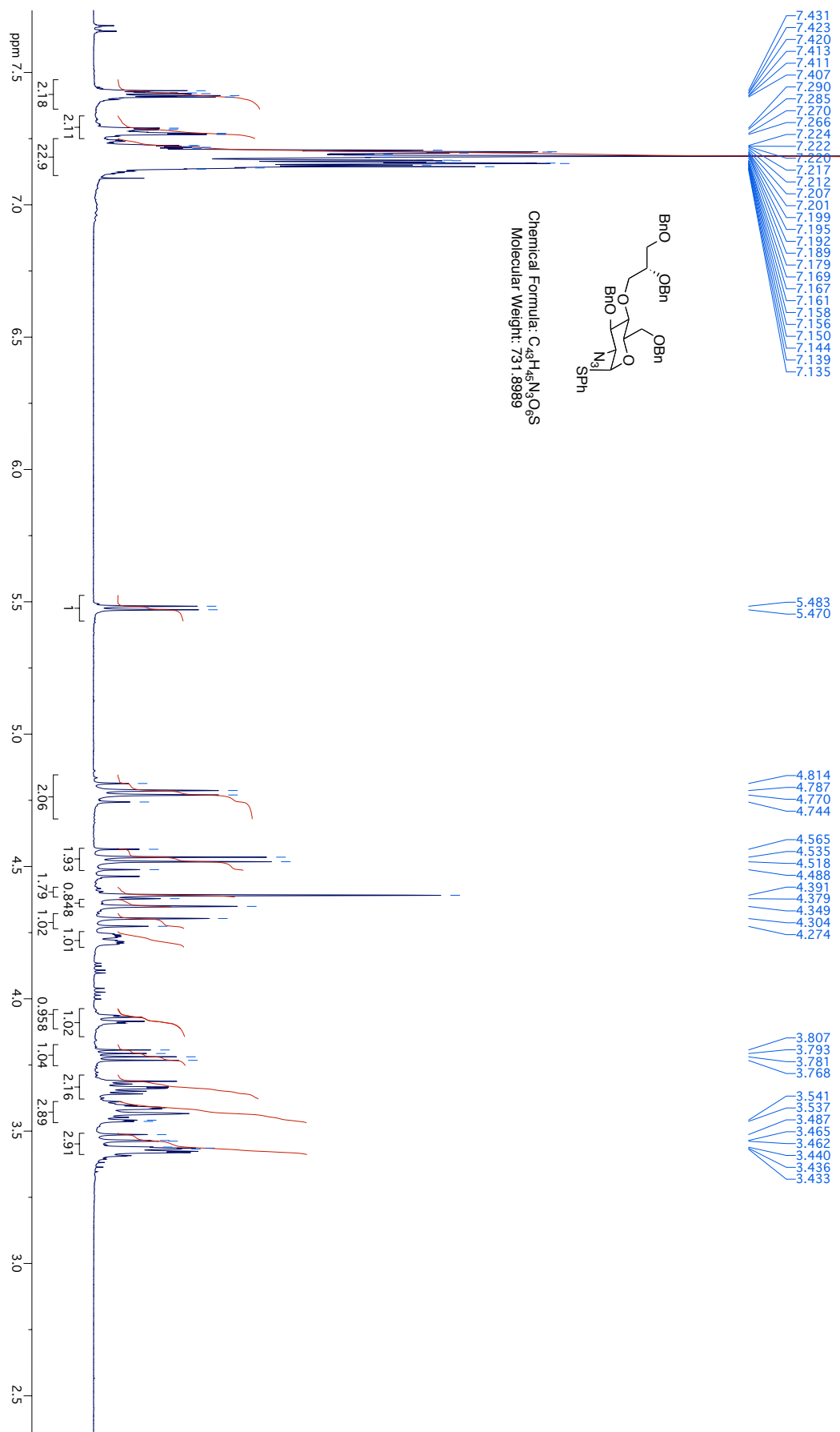
400 MHz, CDCl_3

Phenyl-2-azido-2-deoxy-3,6-di-O-benzyl-4-O-[(*S/R*)-2,3-bis(benzyloxy)propoxy]-1-thio- α -D-glucopyranoside (12)

^{13}C NMR

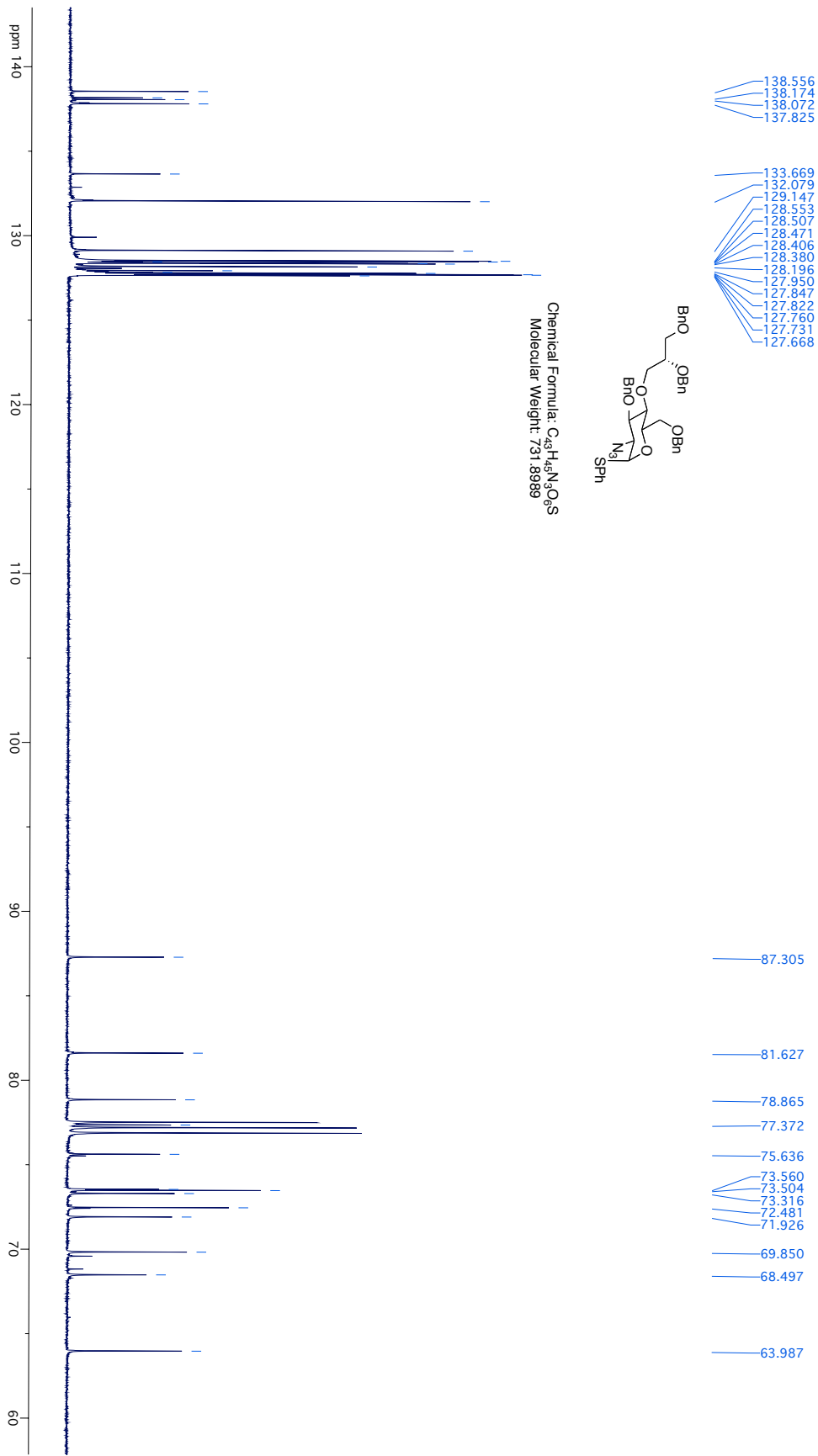


100 MHz, CDCl_3



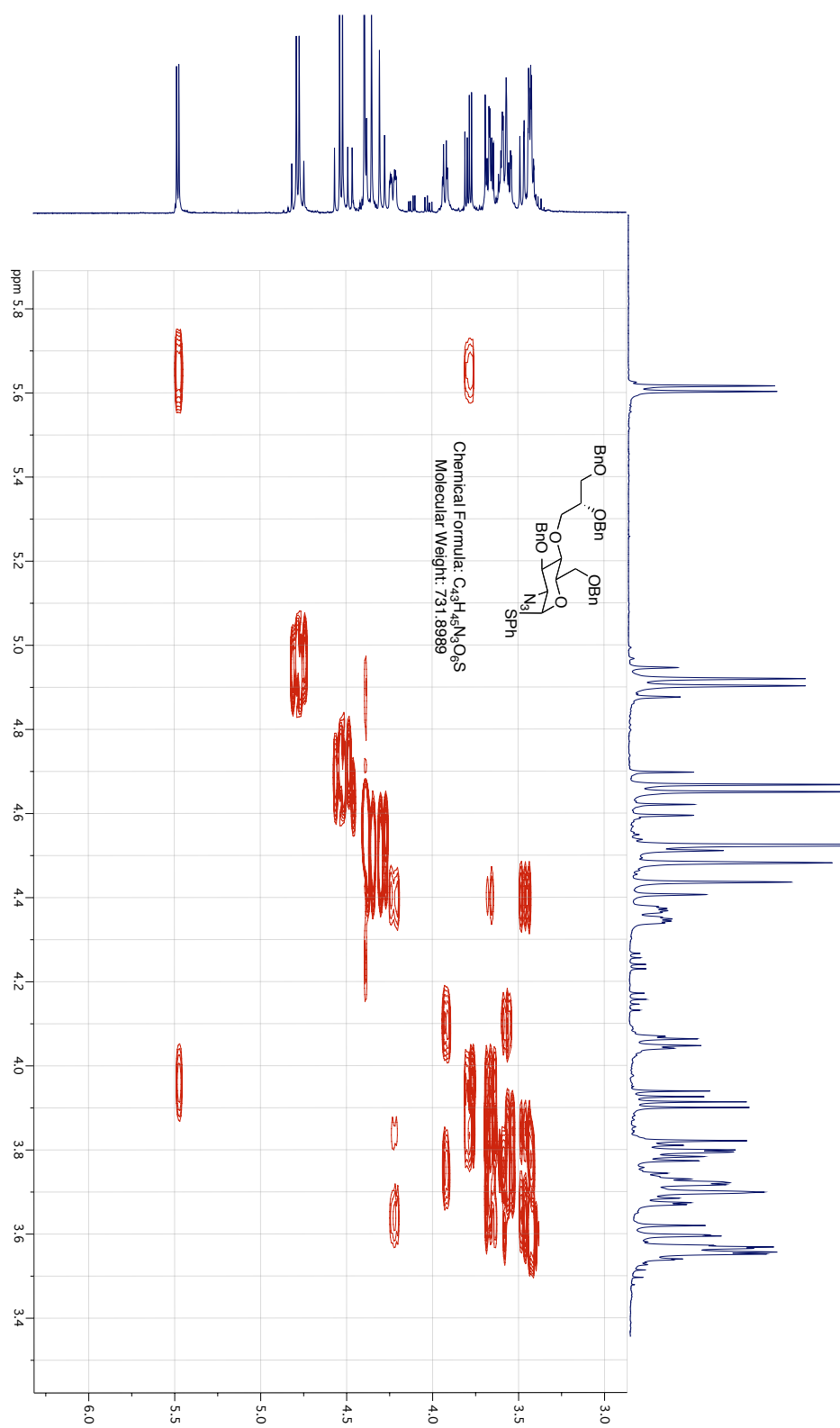
Phenyl-2-azido-2-deoxy-3,6-di-O-benzyl-4-O-[(S)-2,3-bis(benzyloxy)propoxy]-1-thio- α -D-glucopyranoside (15)

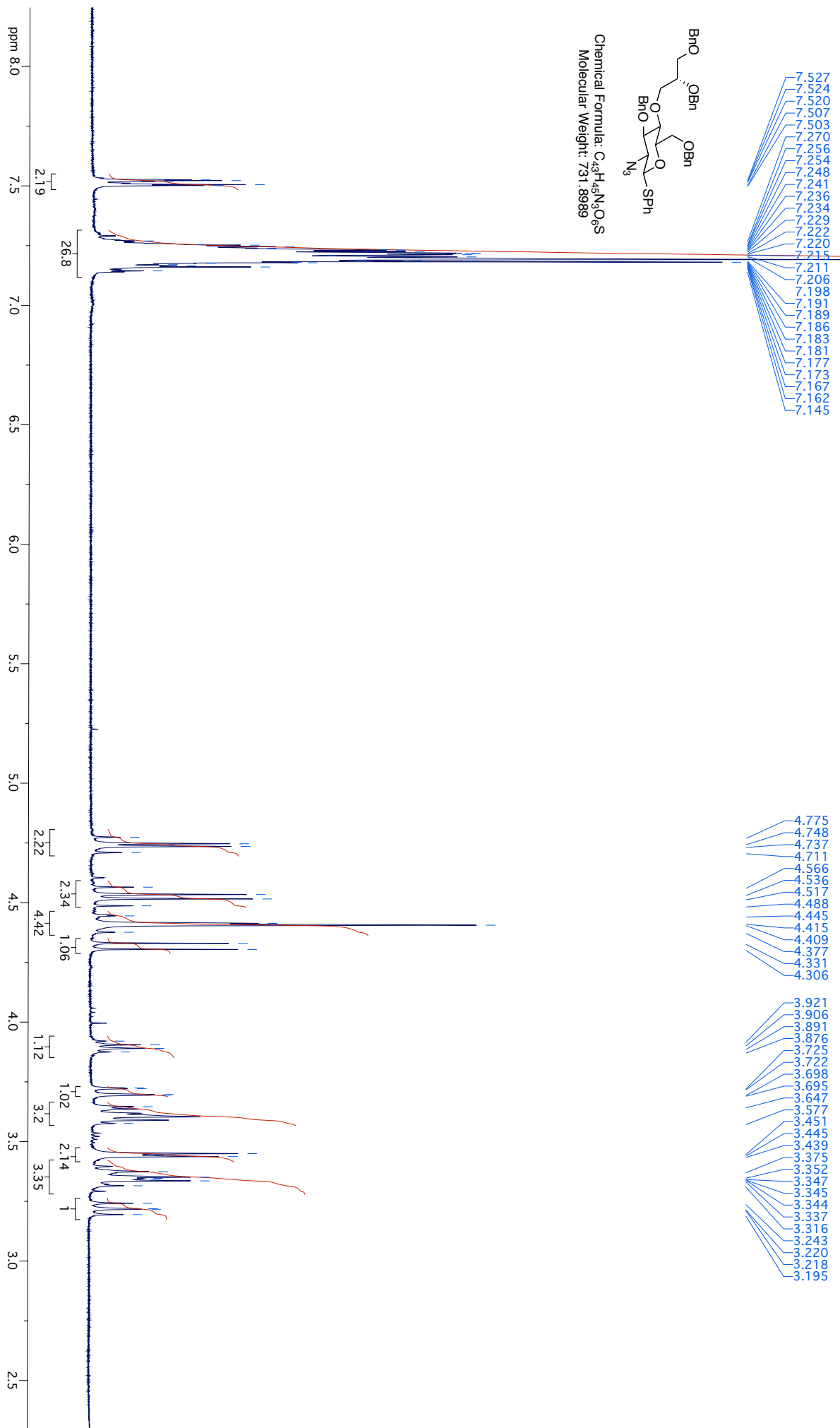
^{13}C NMR



Phenyl-2-azido-2-deoxy-3,6-di-O-benzyl-4-O-[(S)-2,3-bis(benzyloxy)propoxy]-1-thio- α -D-glucopyranoside (15)

COSY

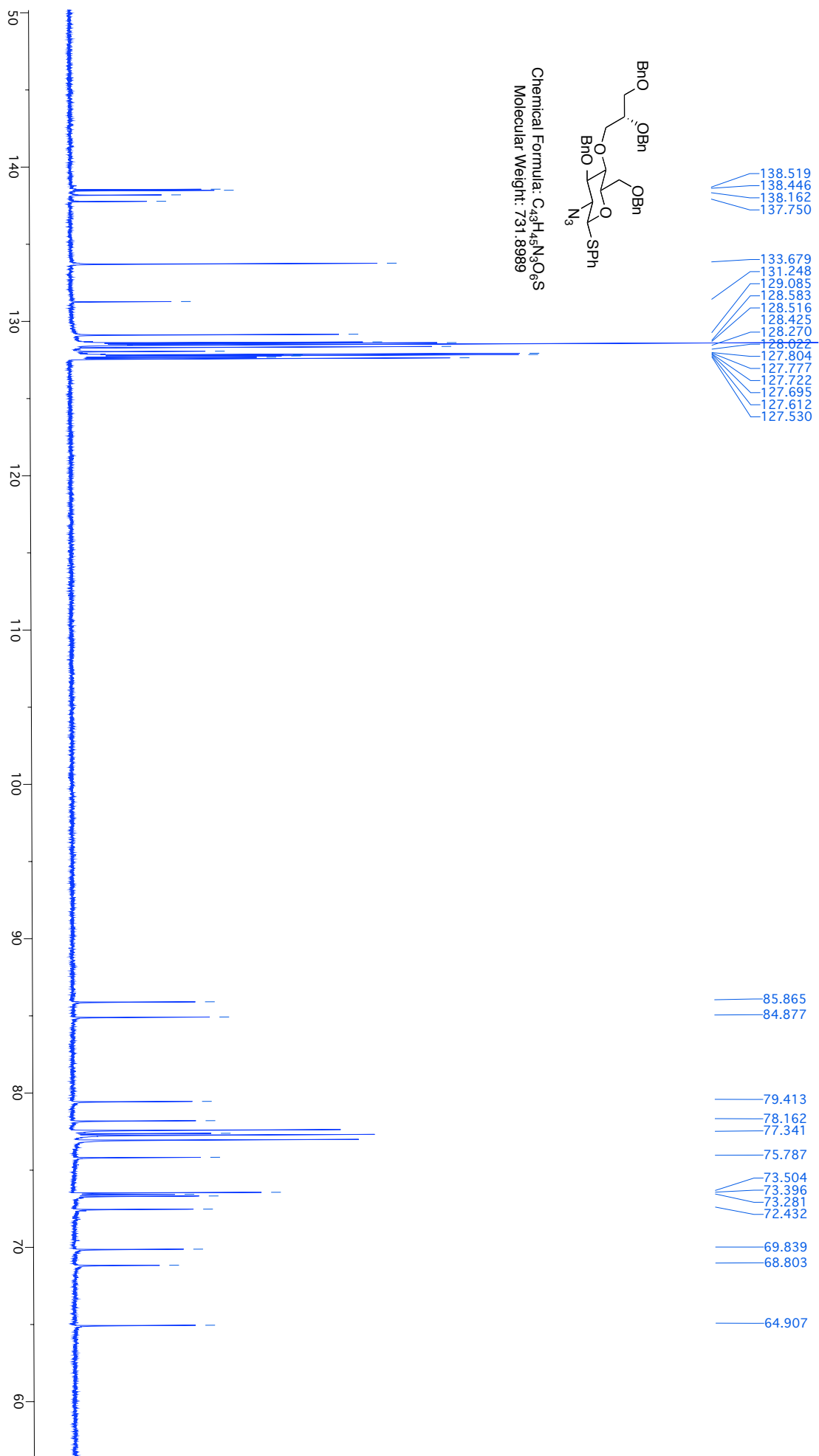




400 MHz, CDCl₃

Phenyl-2-azido-2-deoxy-3,6-di-O-benzyl-4-O-[(S)-2,3-bis(benzyloxy)propoxy]-1-thio-β-D-glucopyranoside (17)

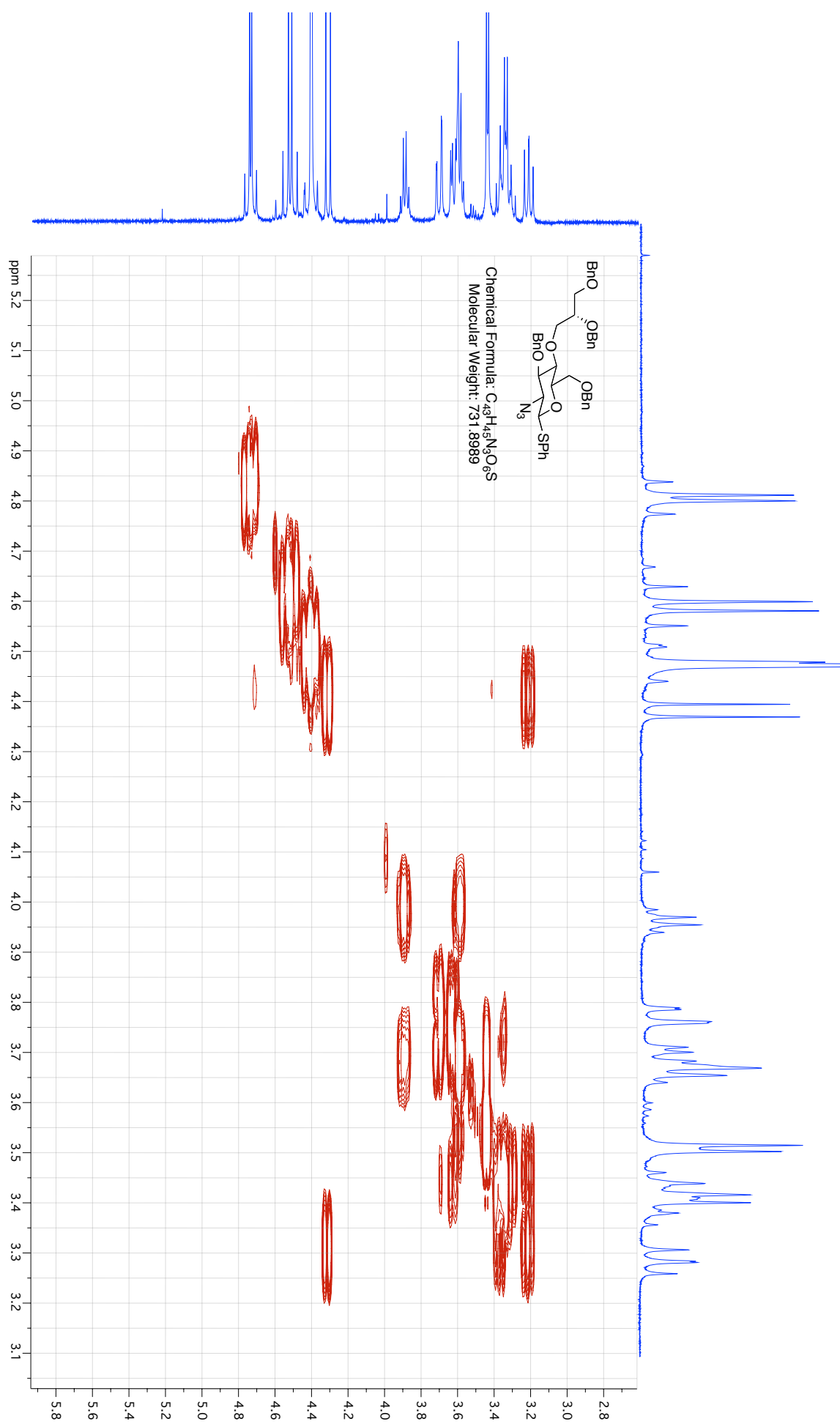
¹³C NMR



100 MHz, CDCl₃

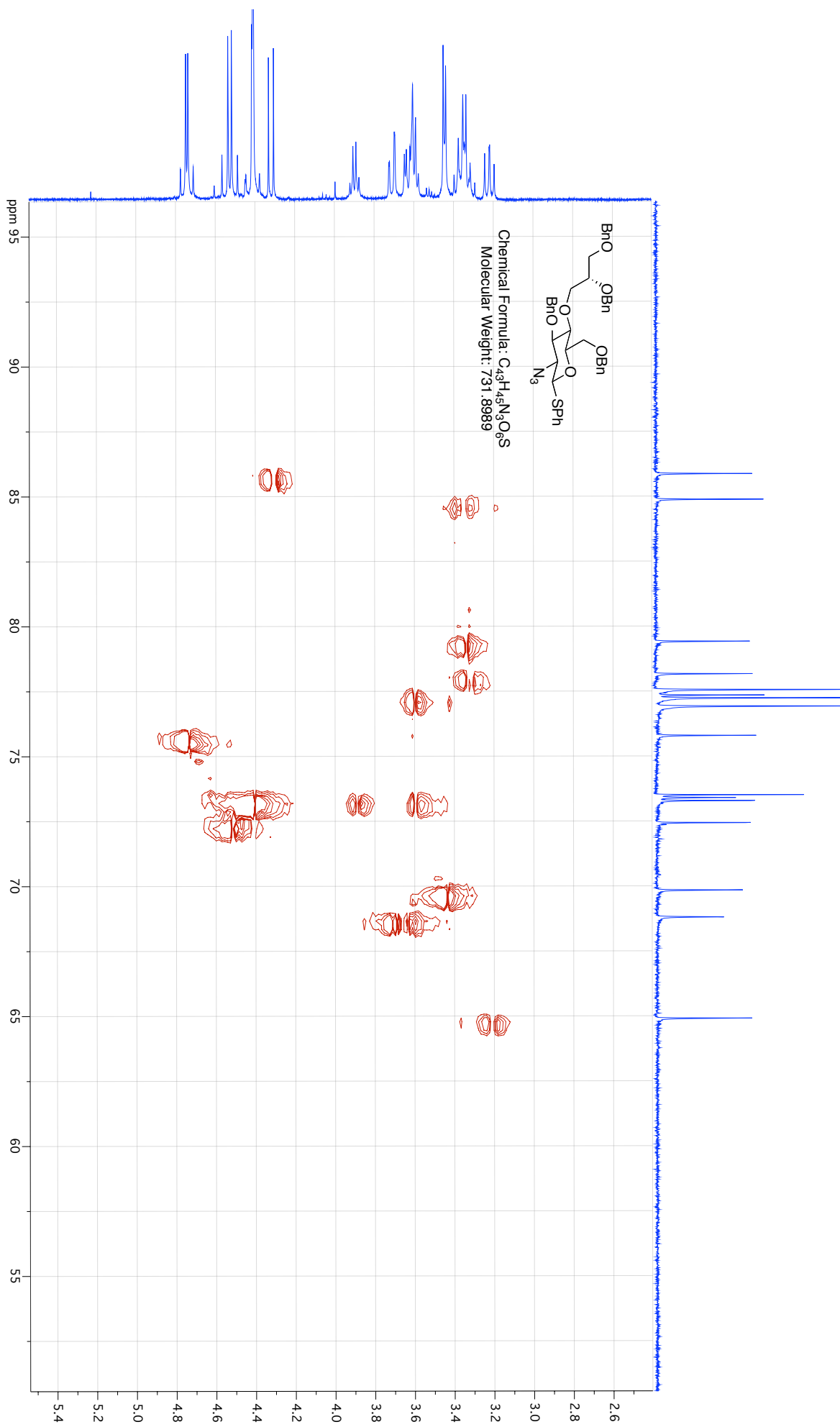
Phenyl-2-azido-2-deoxy-3,6-di-O-benzyl-4-O-[(S)-2,3-bis(benzyloxy)propoxy]-1-thio-β-D-glucopyranoside (17)

COSY

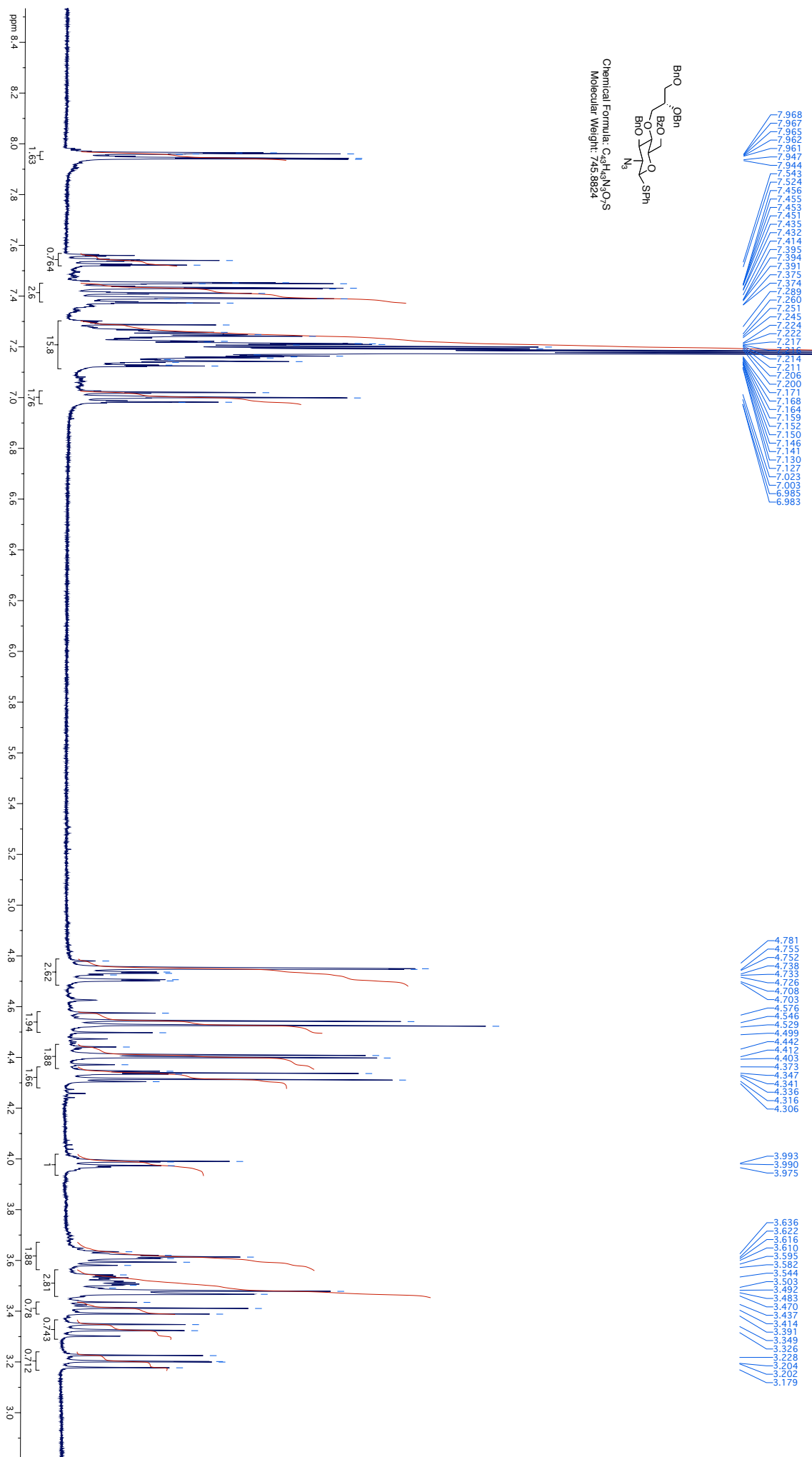


Phenyl-2-azido-2-deoxy-3,6-di-O-benzyl-4-O-[(S)-2,3-bis(benzyloxy)propoxy]-1-thio-β-D-glucopyranoside (17)

HMQC



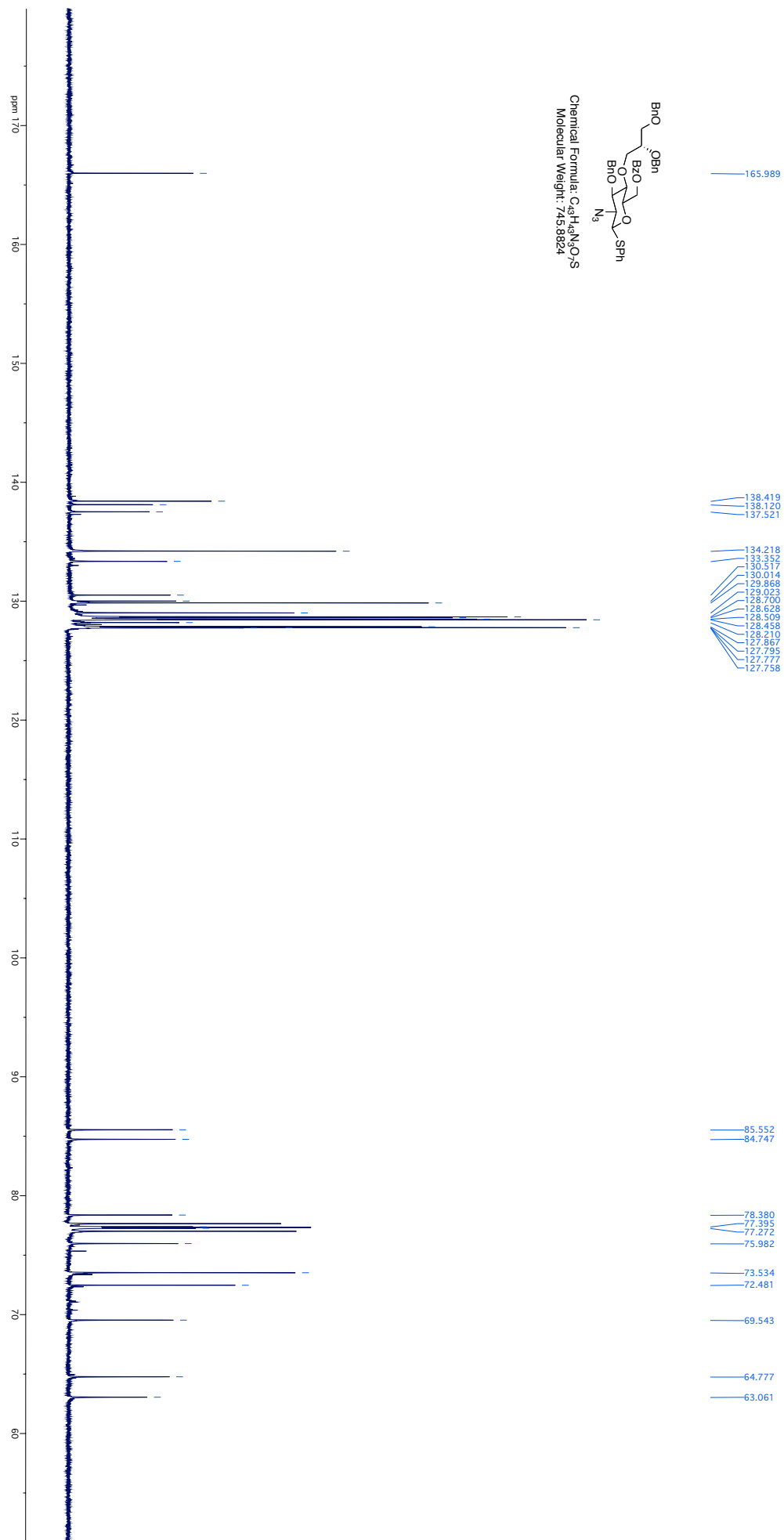
400 MHz, $CDCl_3$



400 MHz, CDCl₃

Phenyl-2-azido-2-deoxy-3-O-benzyl-4-O-[(S)-2,3-bis(benzyloxy)propoxy]-6-O-benzoyl-1-thio-β-D-glucopyranoside (18)

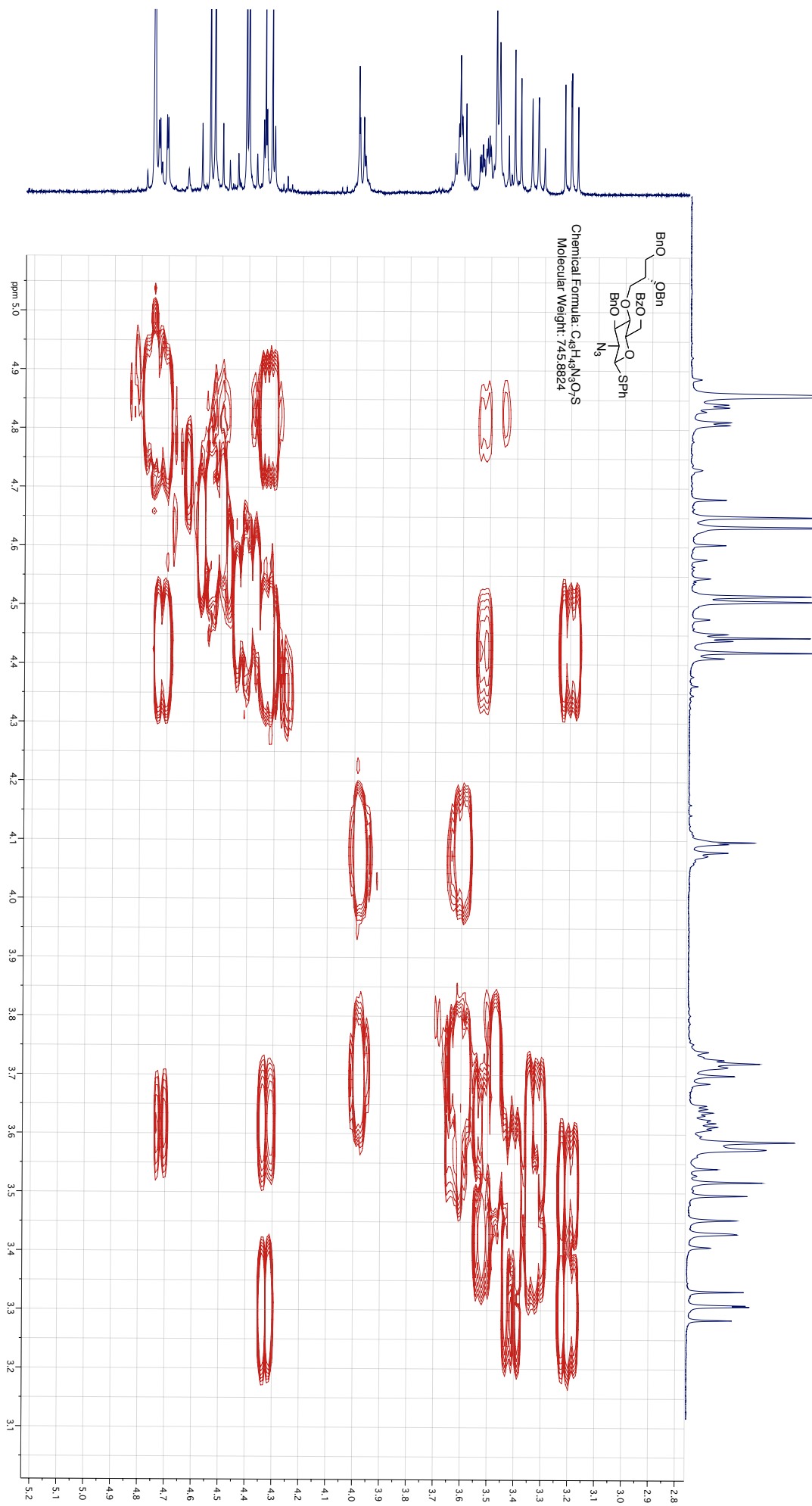
¹³C NMR



100 MHz, CDCl₃

Phenyl-2-azido-2-deoxy-3-O-benzyl-4-O-[(S)-2,3-bis(benzyloxy)propoxy]-6-O-benzoyl-1-thio-β-D-glucopyranoside (18)

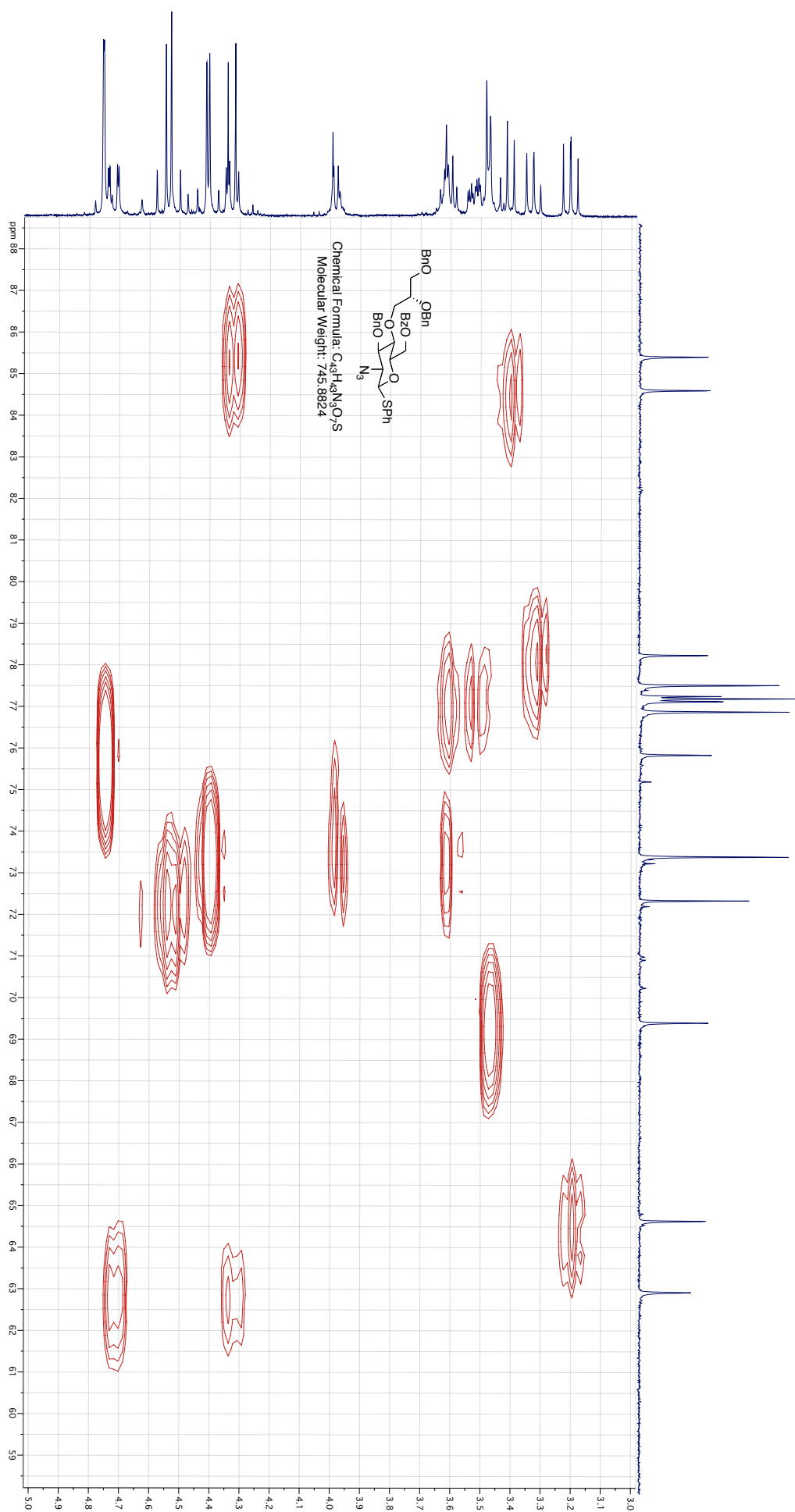
COSY



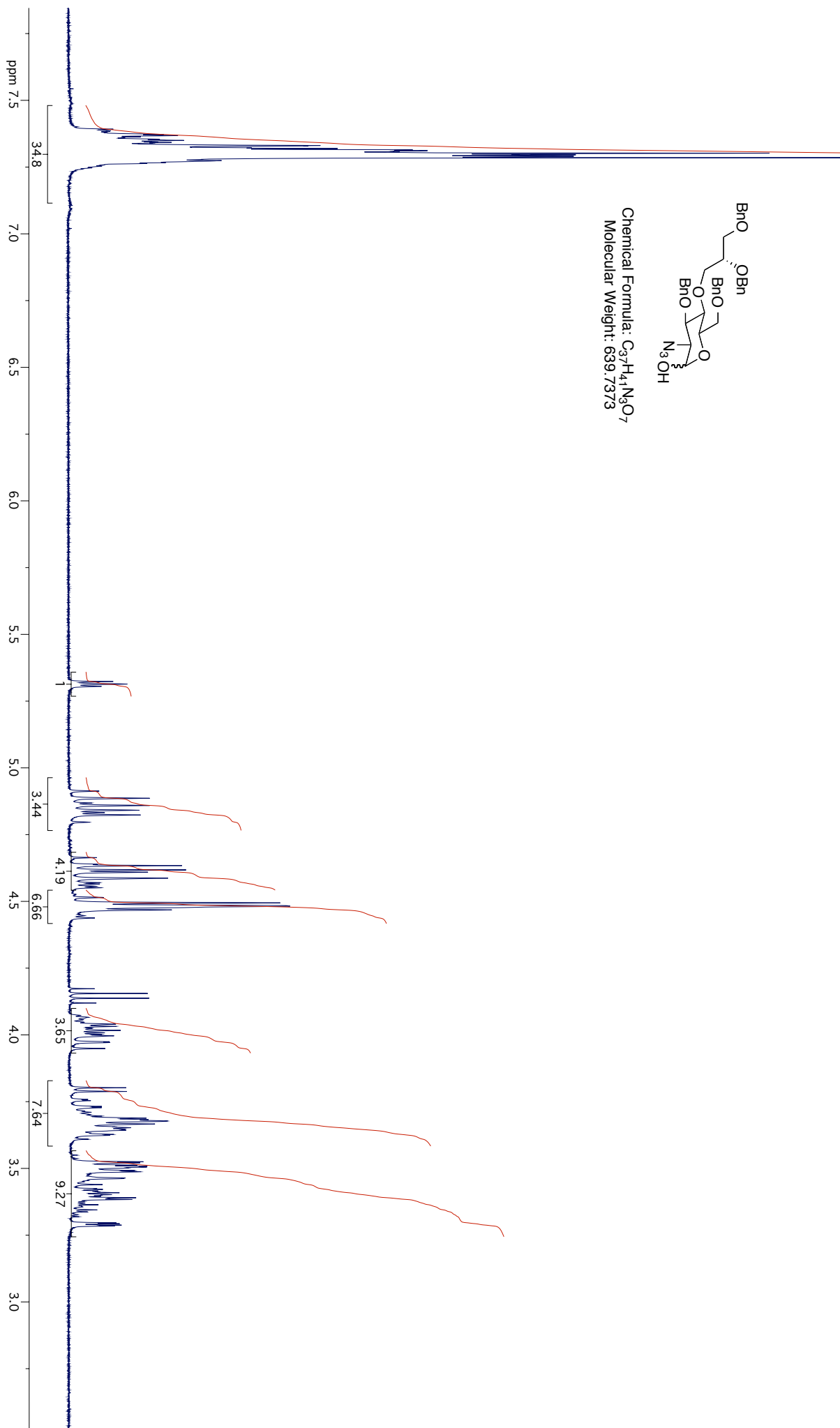
400 MHz, CDCl₃

Phenyl-2-azido-2-deoxy-3-O-benzyl-4-O-[(S)-2,3-bis(benzyloxy)propoxy]-6-O-benzoyl-1-thio-β-D-glucopyranoside (18)

HMQC



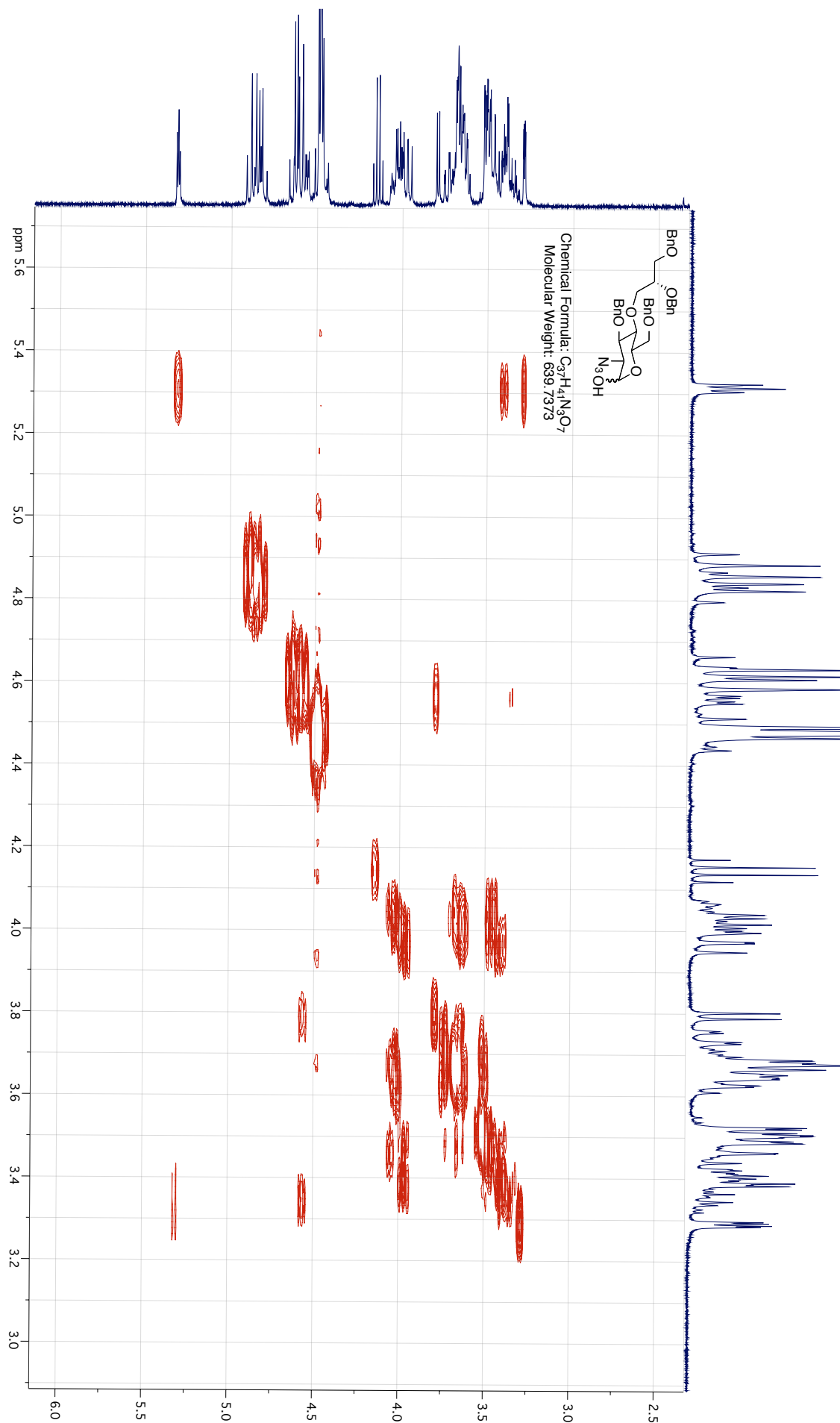
400 MHz, CDCl₃



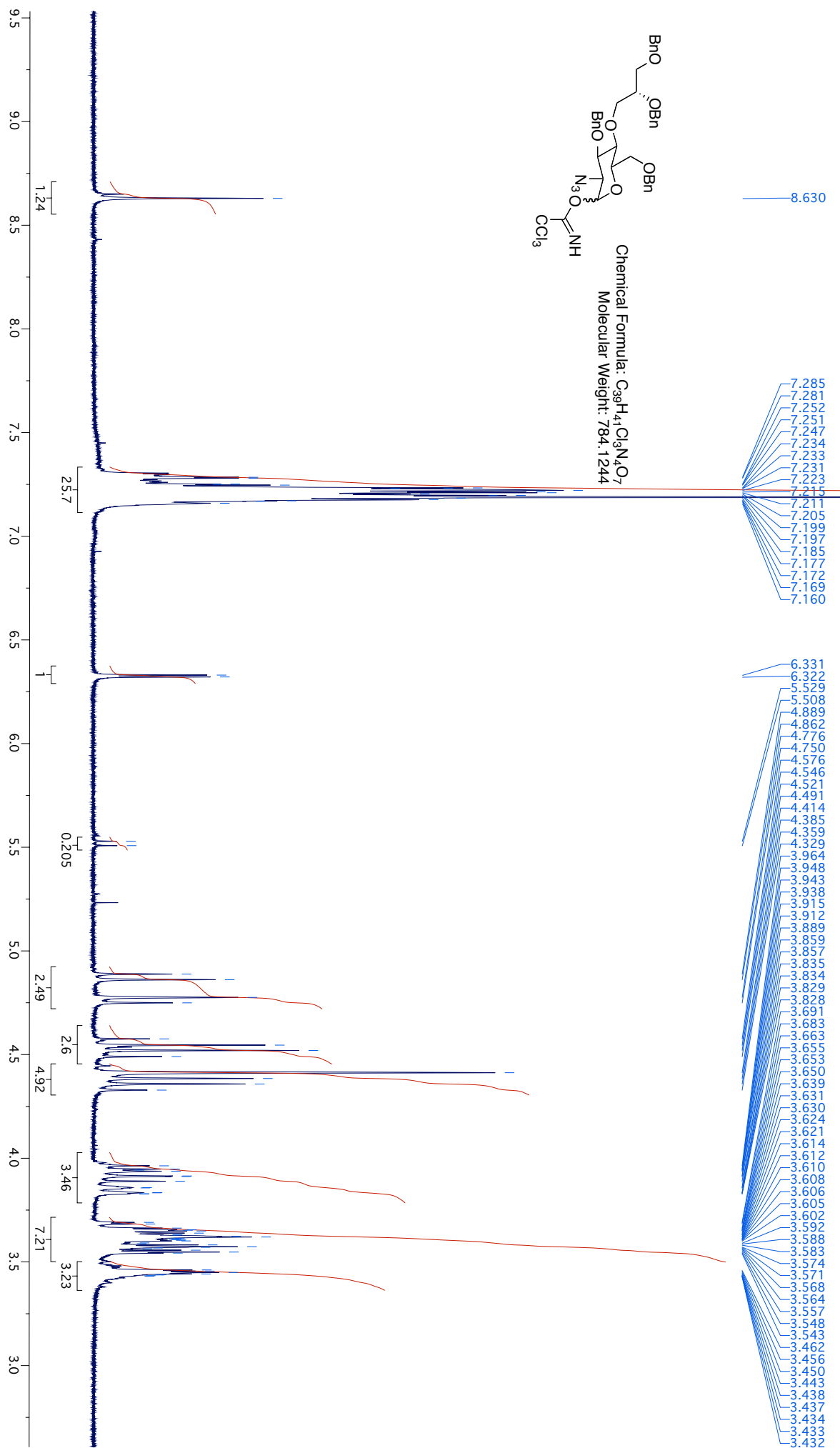
400 MHz, CDCl_3

2-Azido-3,6-di-O-benzyl-2-deoxy-4-O-[(S)-2,3-bis(benzyloxy)propoxy]- α -D-glucopyranose

COSY

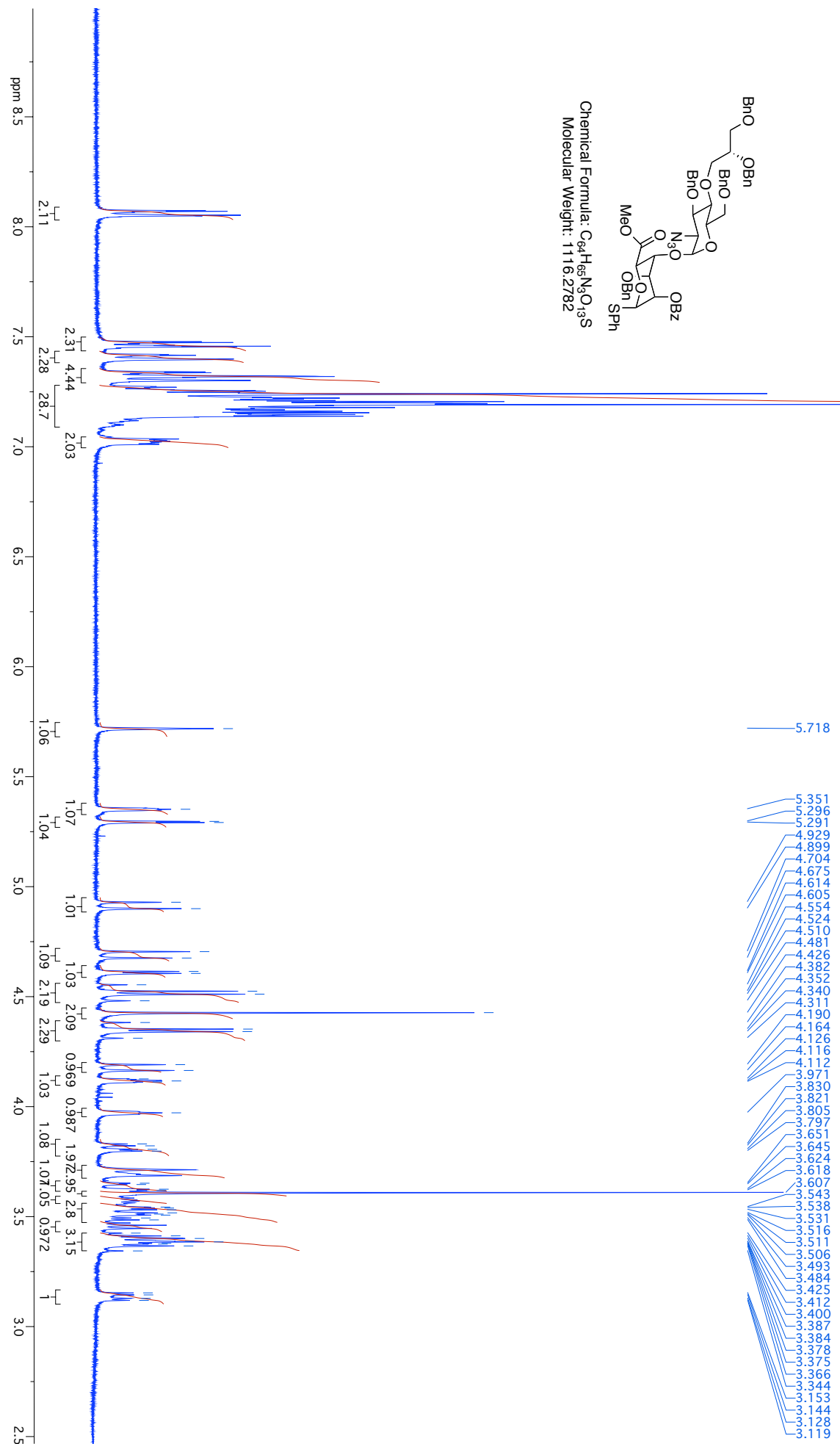


400 MHz, CDCl_3



Methyl (phenyl 4-O-(2-azido-3,6-di-O-benzyl-2-deoxy-4-O-[(S)-2,3-bis(benzyloxy)propoxy]- α -D-glucopyranosyl)-2-O-benzoyl-3-O-benzyl-1-thio- α -L-idopyranoside)-uronate (20)

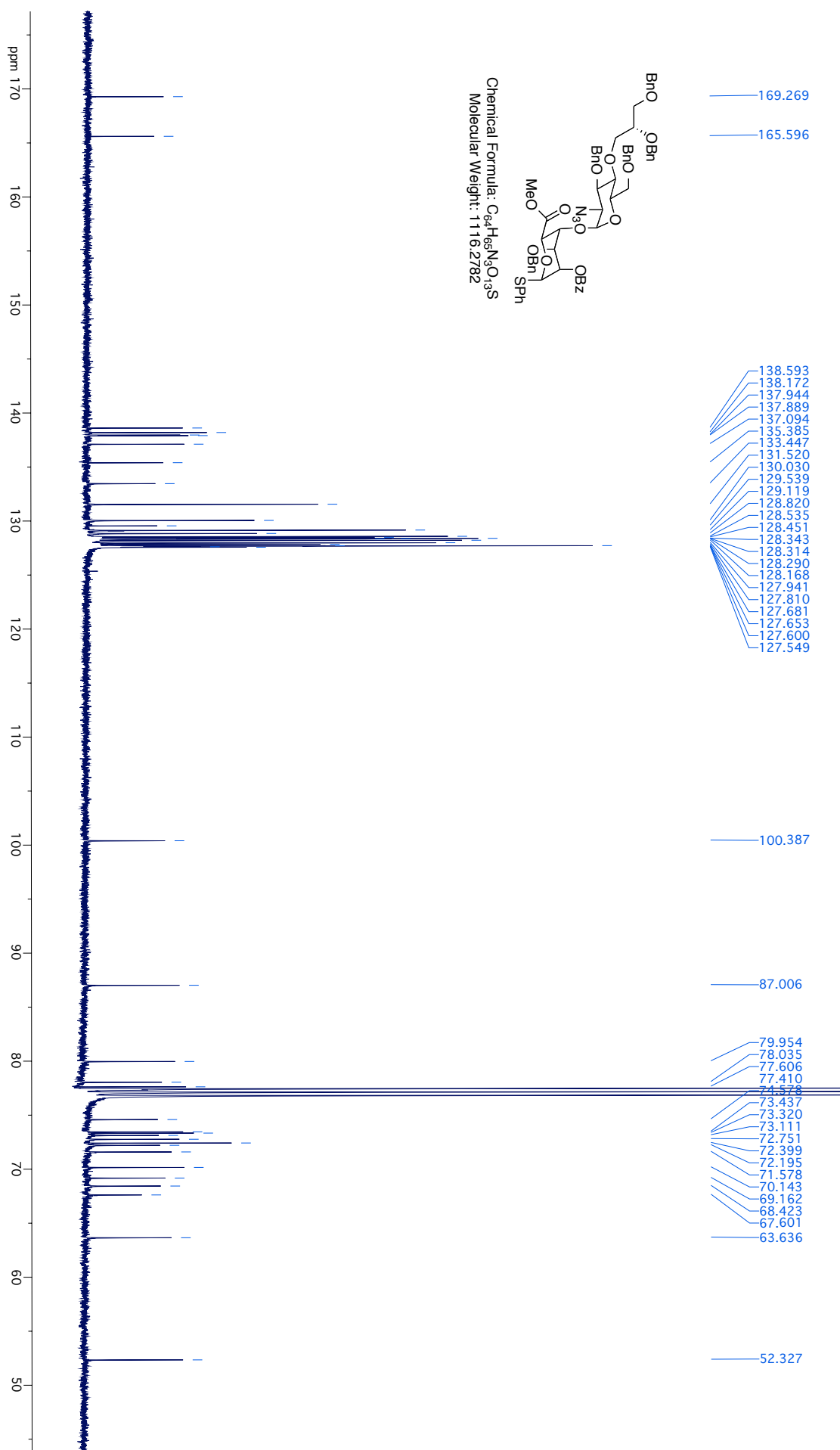
$^1\text{H NMR}$



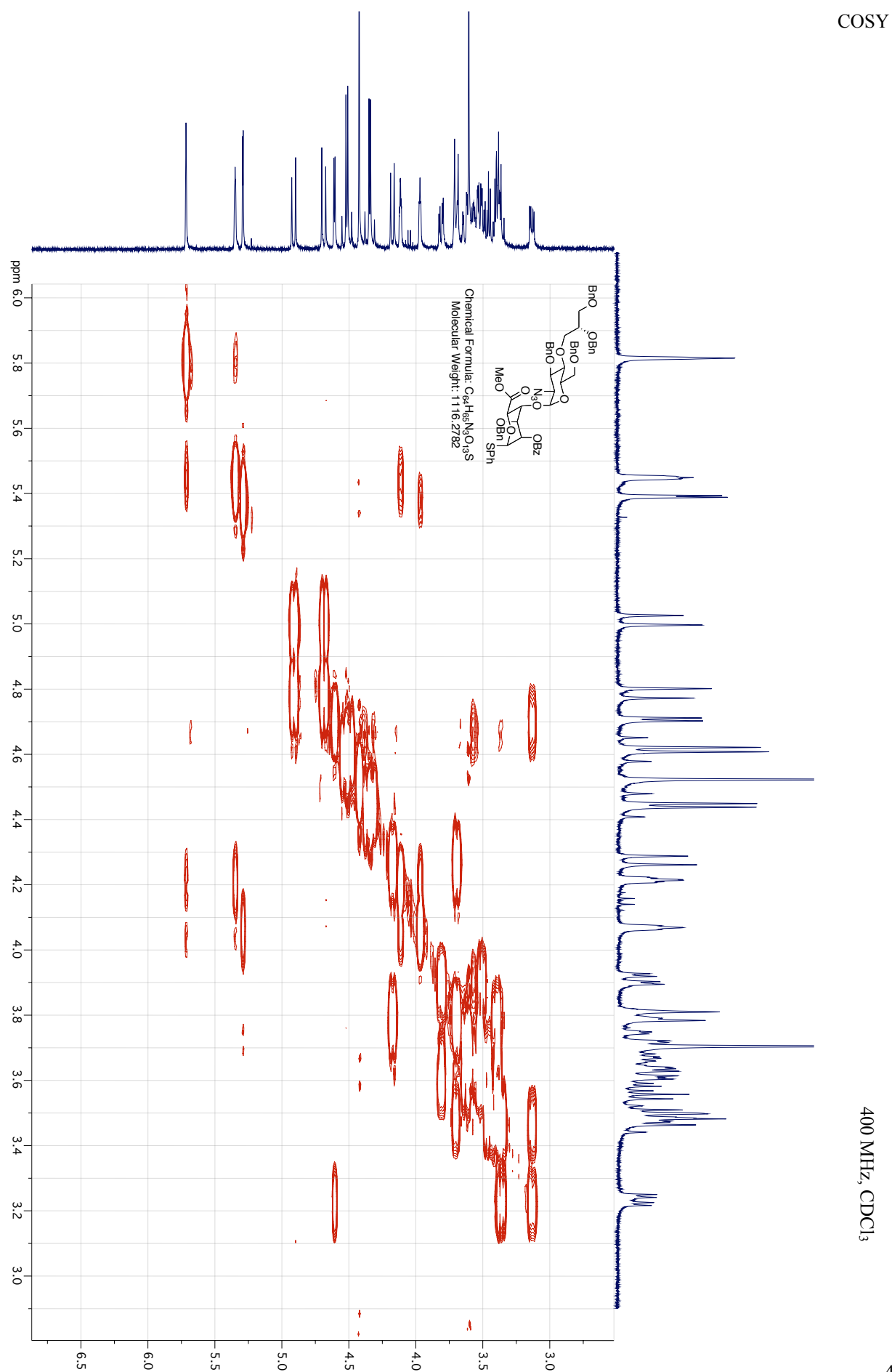
400 MHz, CDCl_3

Methyl (phenyl 4-O-(2-azido-3,6-di-O-benzyl-2-deoxy-4-O-[(S)-2,3-bis(benzyloxy)propoxy]- α -D-glucopyranosyl)-2-O-benzoyl-3-O-benzyl-1-thio- α -L-idopyranoside)-uronate (20)

^{13}C NMR

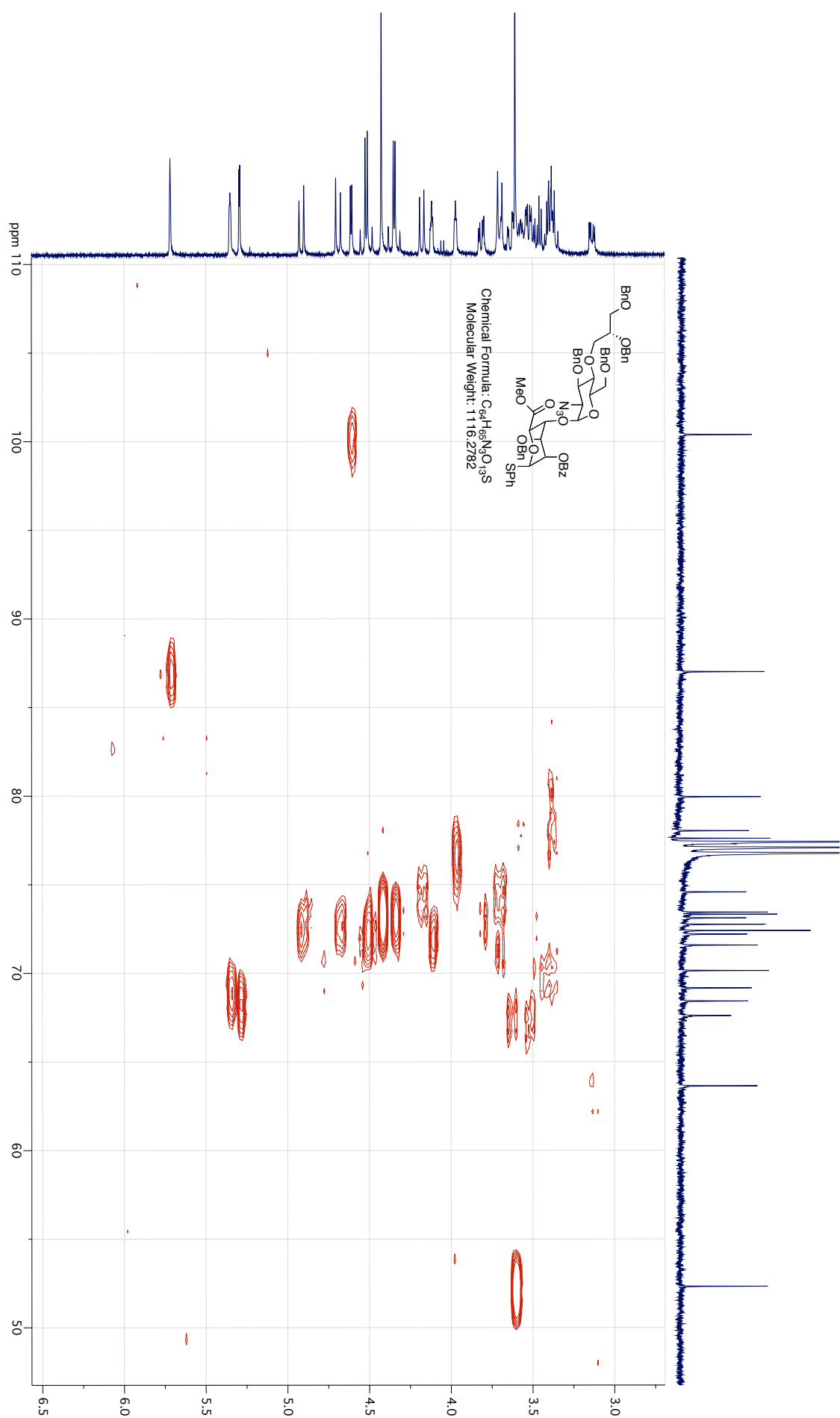


Methyl (phenyl 4-O-(2-azido-3,6-di-O-benzyl-2-deoxy-4-O-[(S)-2,3-bis(benzyloxy)propoxy]- α -D-glucopyranosyl)-2-O-benzoyl-3-O-benzyl-1-thio- α -L-idopyranoside)-uronate (20)



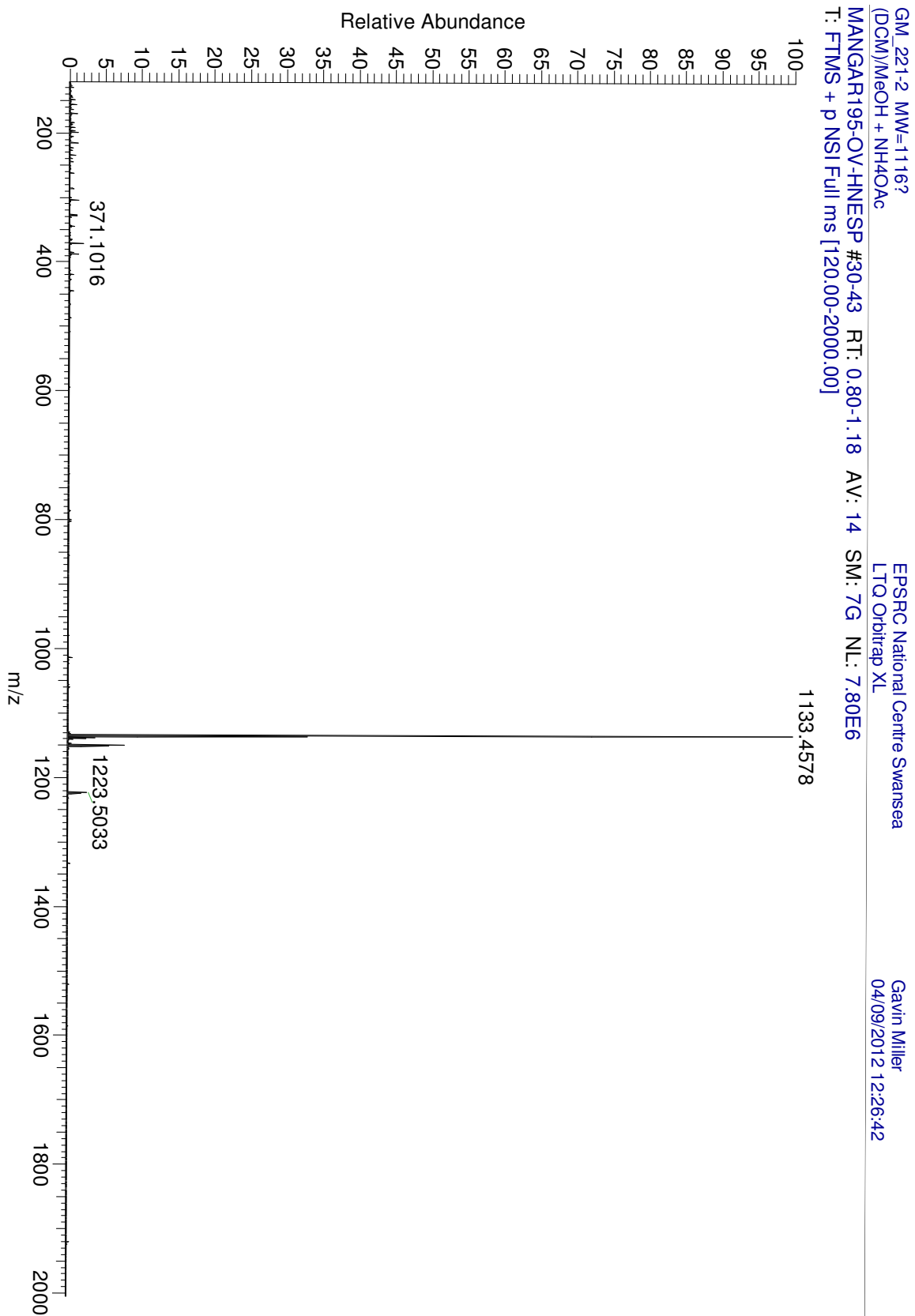
Methyl (phenyl 4-O-(2-azido-3,6-di-O-benzyl-2-deoxy-4-O-[(S)-2,3-bis(benzyloxy)propoxy]- α -D-glucopyranosyl)-2-O-benzoyl-3-O-benzyl-1-thio- α -L-idopyranoside)-uronate (20)

HMQC



Methyl (phenyl 4-O-(2-azido-3,6-di-O-benzyl-2-deoxy-4-O-[(S)-2,3-bis(benzyloxy)propoxy]- α -D-glucopyranosyl)-2-O-benzoyl-3-O-benzyl-1-thio- α -L-idopyranoside)-uronate (20)

ESI MS

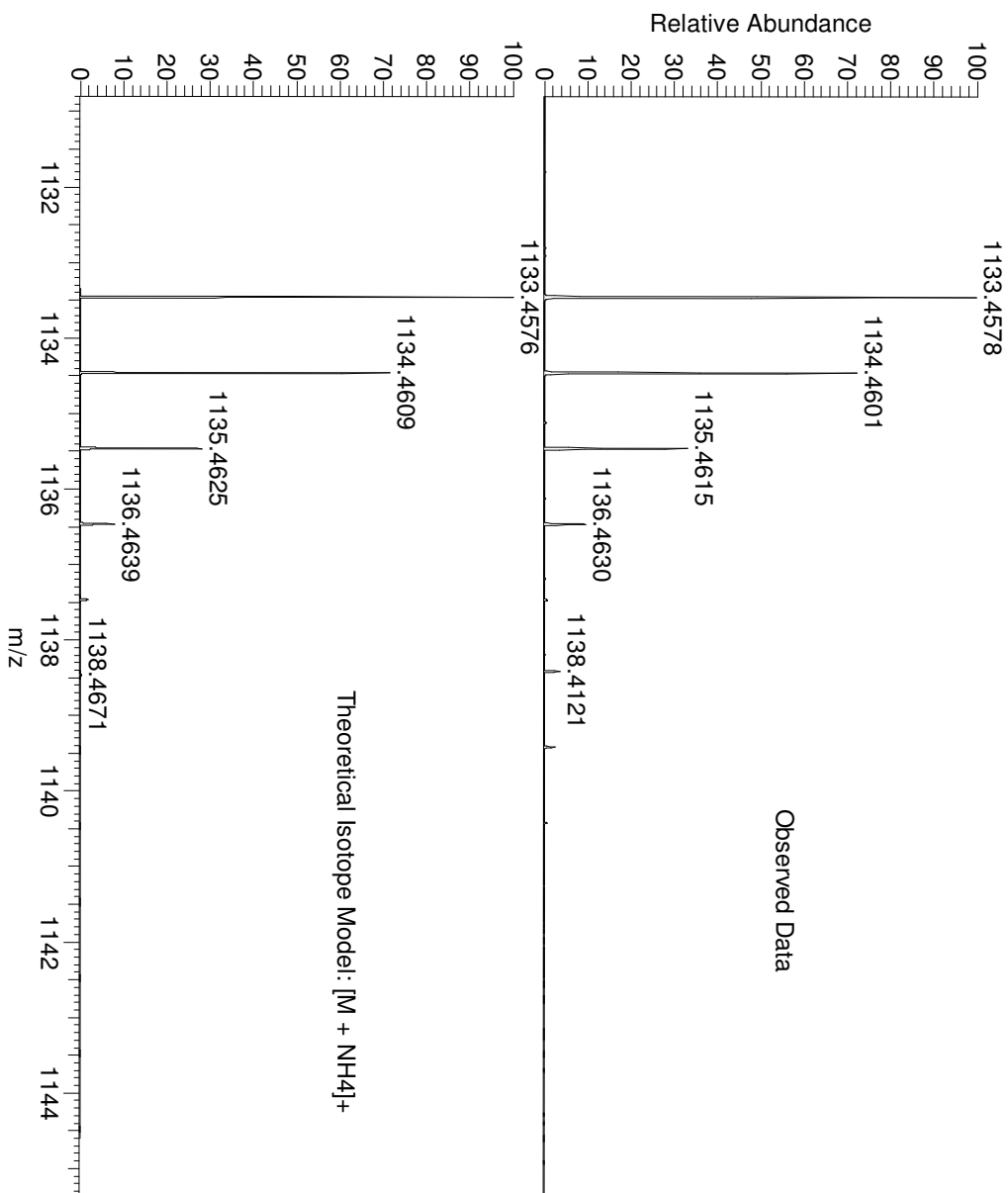


Methyl (phenyl 4-O-(2-azido-3,6-di-O-benzyl-2-deoxy-4-O-[(S)-2,3-bis(benzyloxy)propoxy]- α -D-glucopyranosyl)-2-O-benzoyl-3-O-benzyl-1-thio- α -L-idopyranoside)-uronate (20)

GM_221-2 MW=1116?
(DCM)/MeOH + NH4OAc

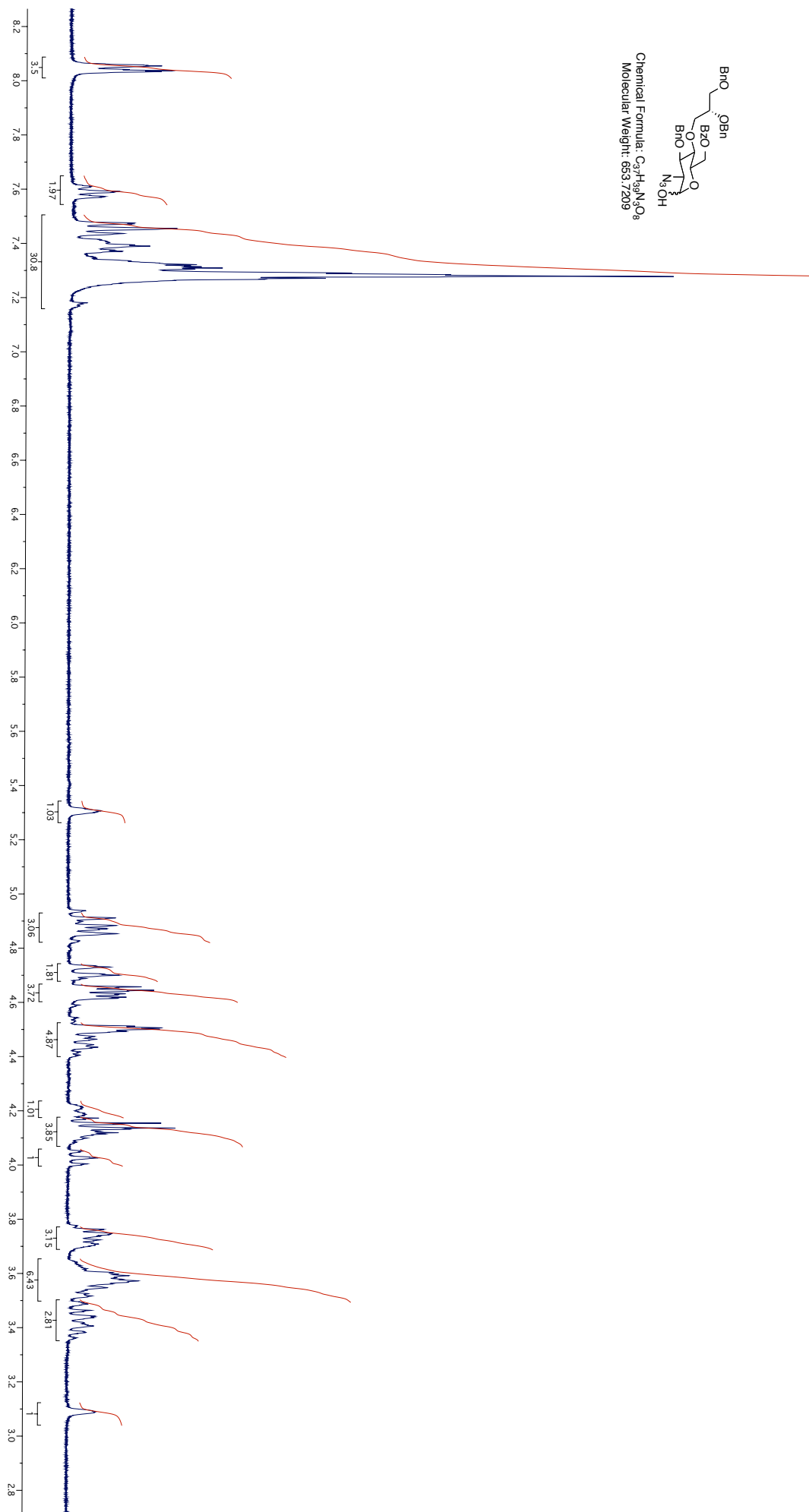
EPSRC National Centre Swansea
LTQ Orbitrap XL

Gavin Miller
04/09/2012 12:26:42



NL:
7.80E6
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43 RT: 0.80-1.18 AV: 14 T:
FTMS + p NSI Full ms
[120.00-2000.00]

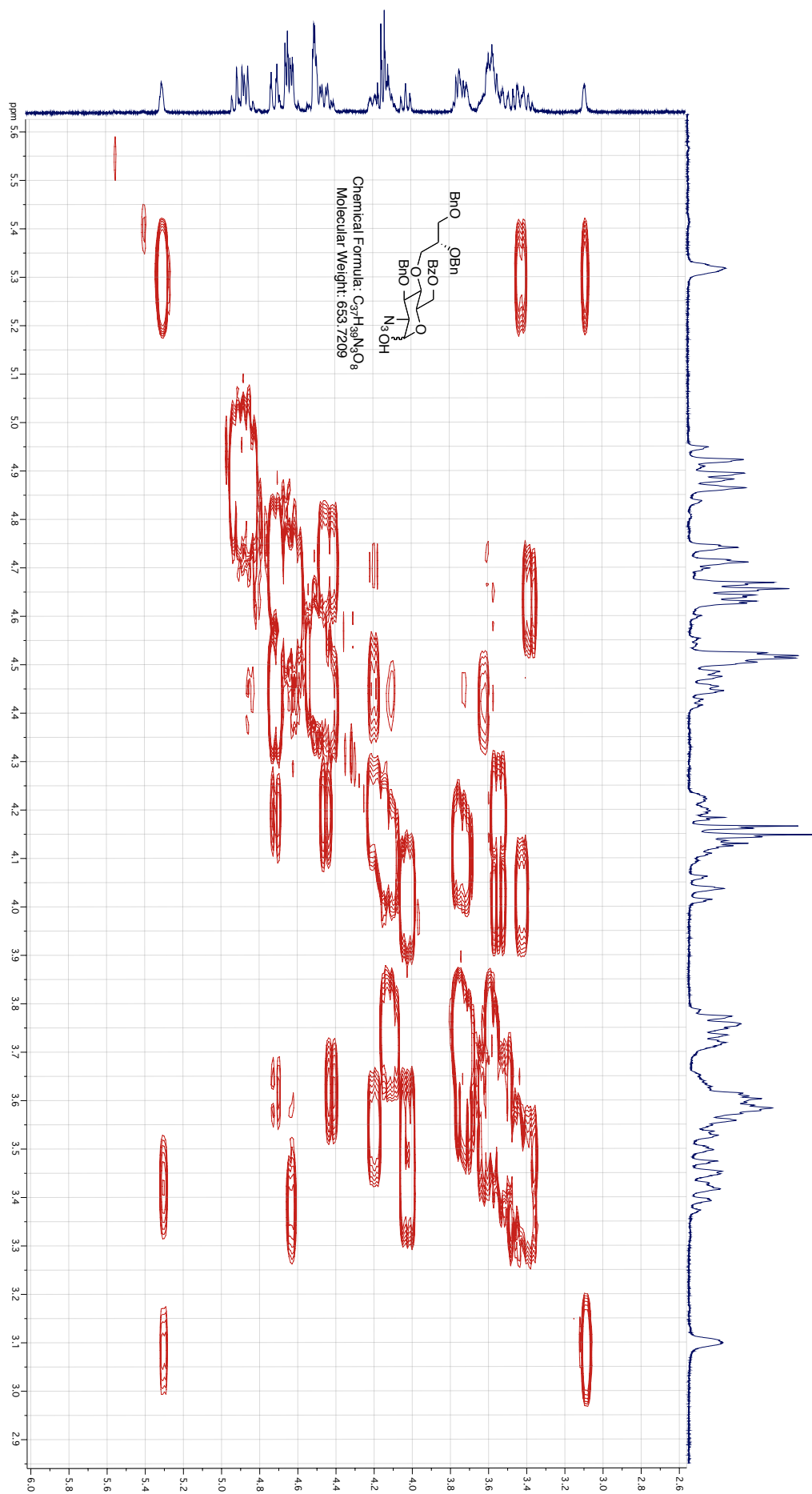
NL:
1.06E4
C₆₄H₆₅N₃O₁₃SNH₄:
C₆₄H₆₉N₄O₁₃S₁
P (gss, s/p:40) Chrg 1
R: 100000 Res .Pwr. @FWHM



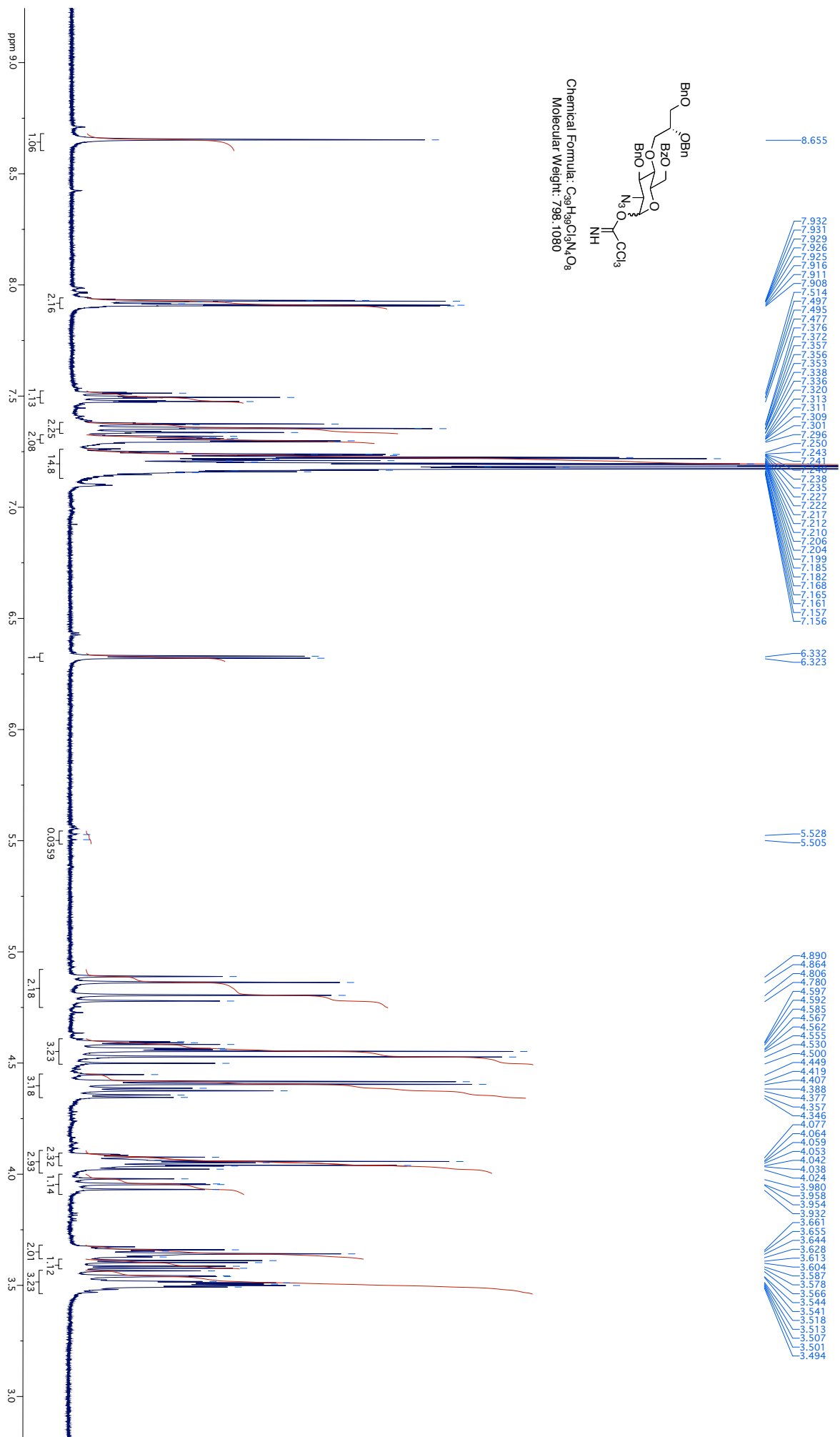
400 MHz, CDCl_3

2-Azido-3-O-benzyl-6-O-benzoyl-2-deoxy-4-O-[(S)-2,3-bis(benzyloxy)propoxy]- α -D-glucopyranose

COSY



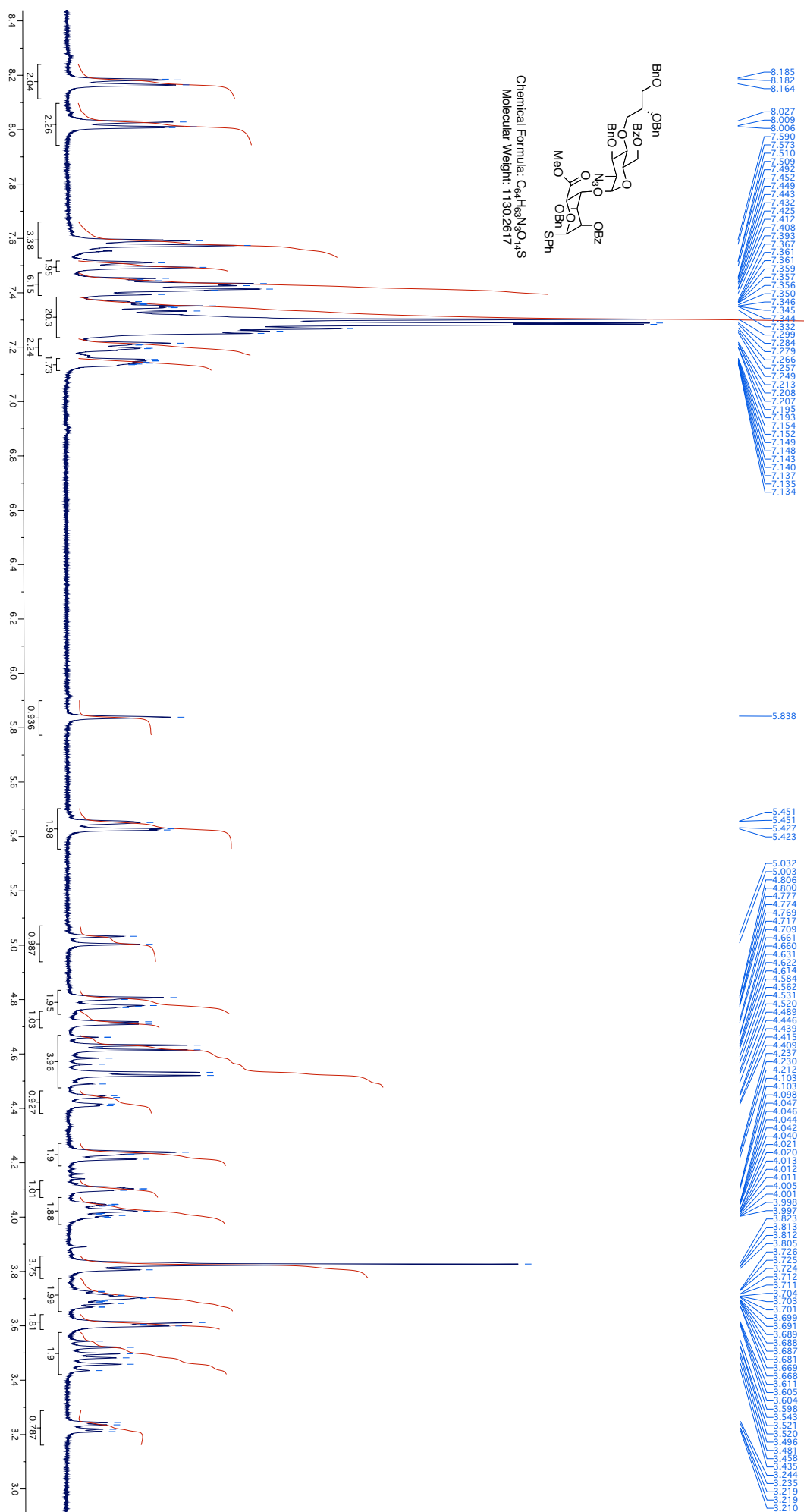
400 MHz, CDCl₃



400 MHz, CDCl_3

Methyl (phenyl 4-O-(2-azido-3-O-benzyl-6-O-benzoyl-2-deoxy-4-O-[(S)-2,3-bis(benzyloxy)propoxy]- α -D-glucopyranosyl)-2-O-benzoyl-3-O-benzyl-1-thio- α -L-idopyranoside)-uronate (21)

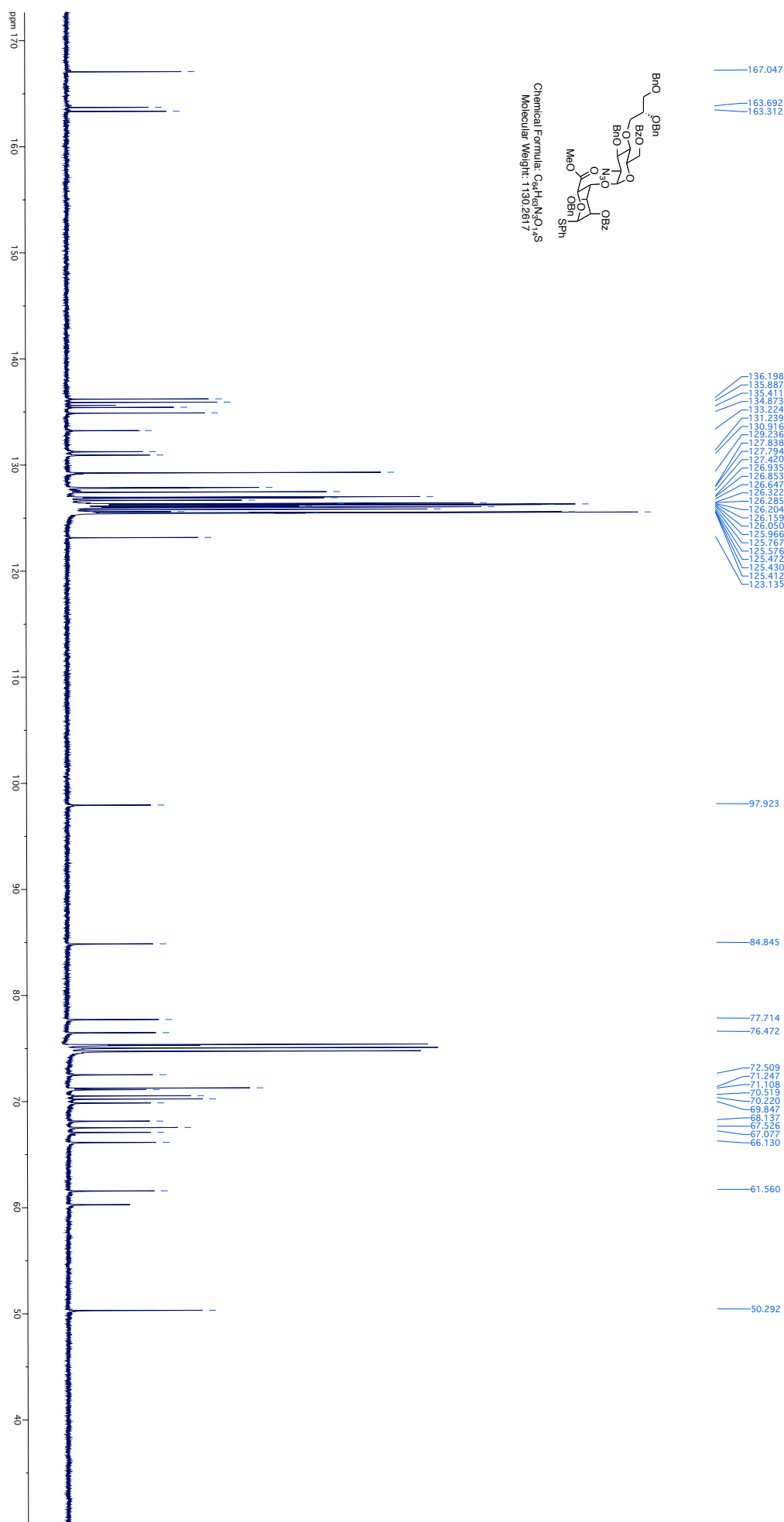
$^1\text{H NMR}$



400 MHz, CDCl_3

Methyl (phenyl 4-O-(2-azido-3-O-benzyl-6-O-benzoyl-2-deoxy-4-O-[(S)-2,3-bis(benzyloxy)propoxy]- α -D-glucopyranosyl)-2-O-benzoyl-3-O-benzyl-1-thio- α -L-idopyranoside)-uronate (21)

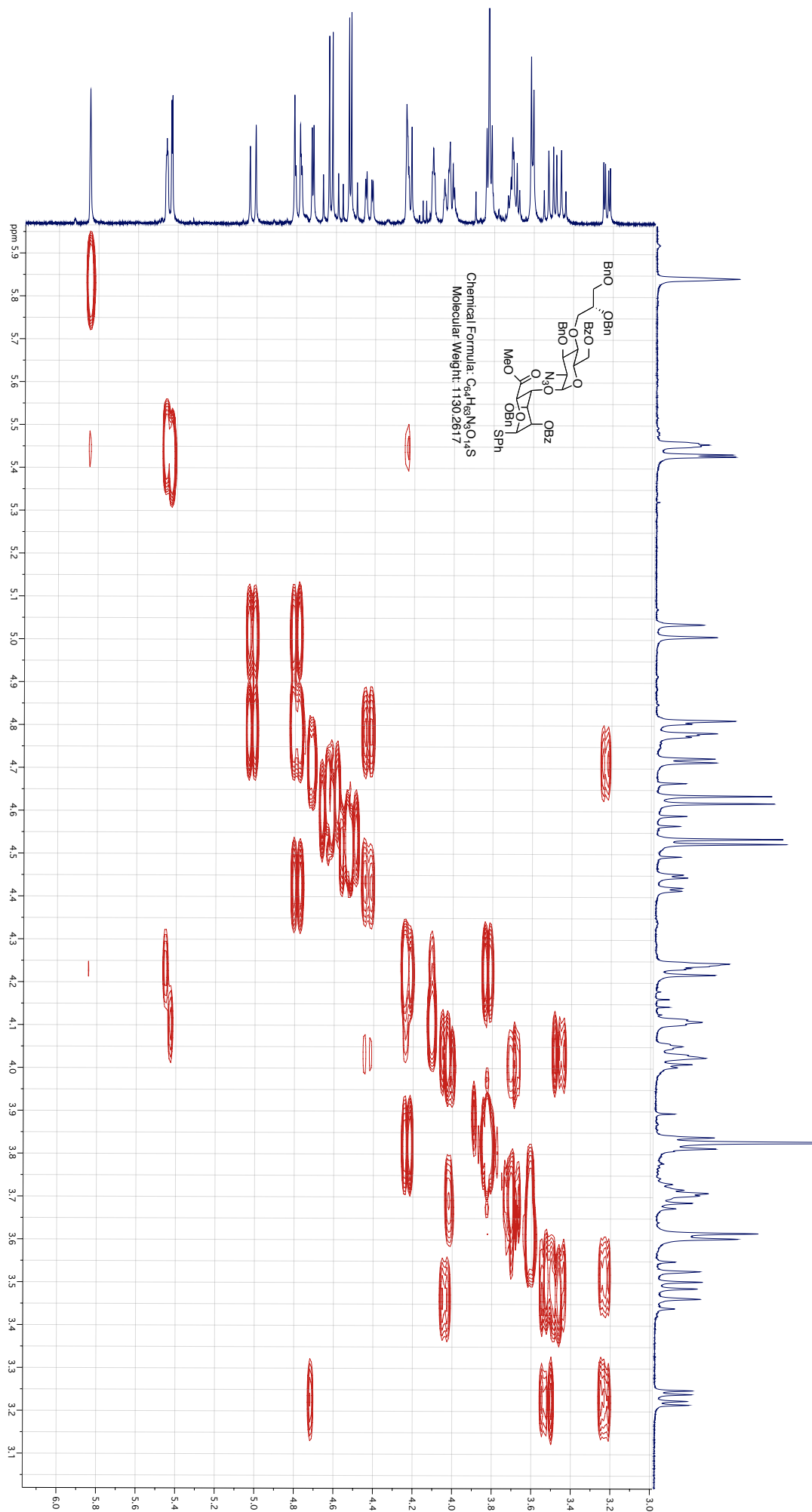
^{13}C NMR



100 MHz, CDCl₃

Methyl (phenyl 4-O-(2-azido-3-O-benzyl-6-O-benzoyl-2-deoxy-4-O-[(S)-2,3-bis(benzyloxy)propoxy]- α -D-glucopyranosyl)-2-O-benzoyl-3-O-benzyl-1-thio- α -L-idopyranoside)-uronate (21)

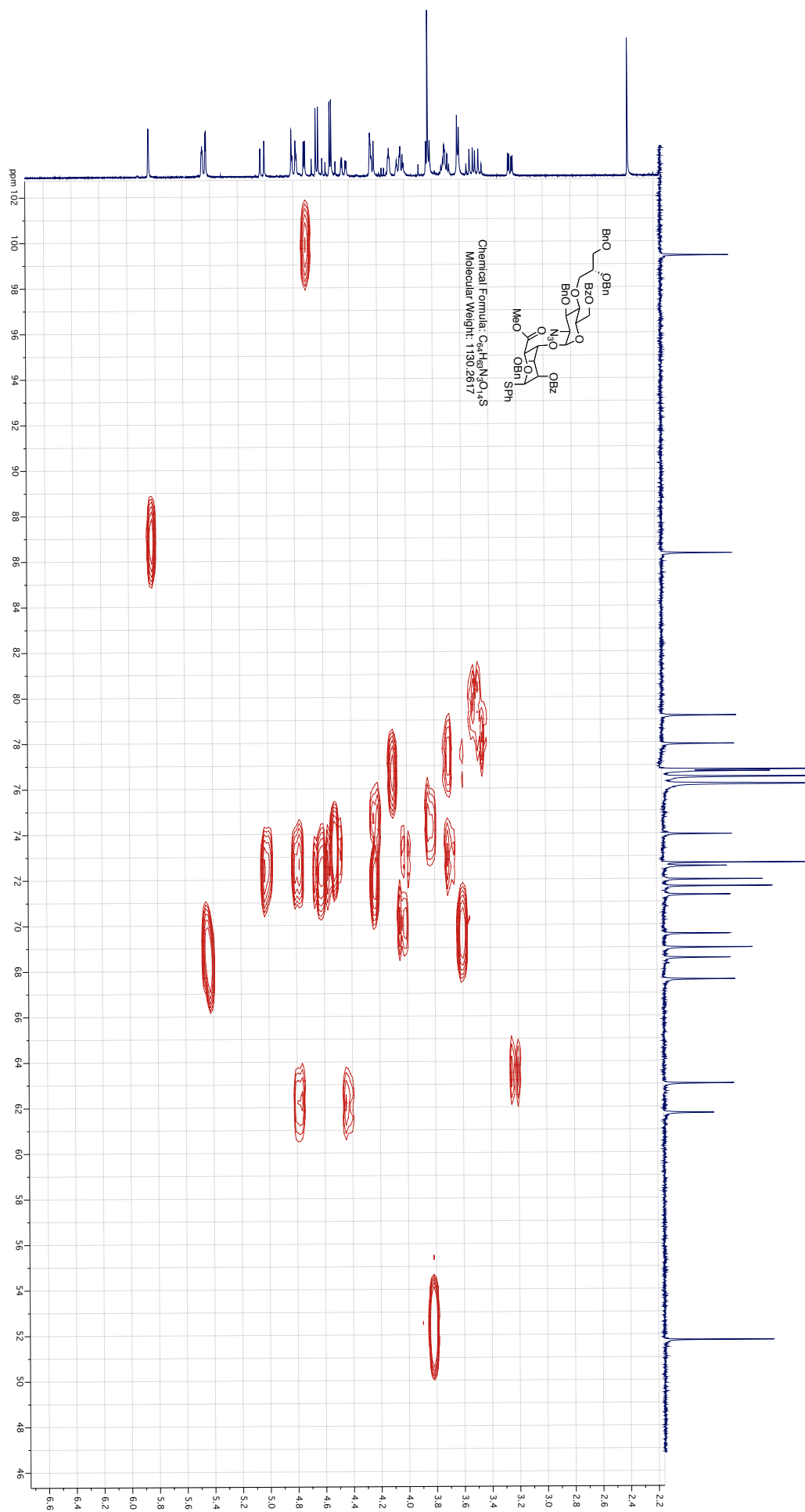
COSY



400 MHz, $CDCl_3$

Methyl (phenyl 4-O-(2-azido-3-O-benzyl-6-O-benzoyl-2-deoxy-4-O-[(S)-2,3-bis(benzyloxy)propoxy]- α -D-glucopyranosyl)-2-O-benzoyl-3-O-benzyl-1-thio- α -L-idopyranoside)-uronate (21)

HMQC



400 MHz, $CDCl_3$

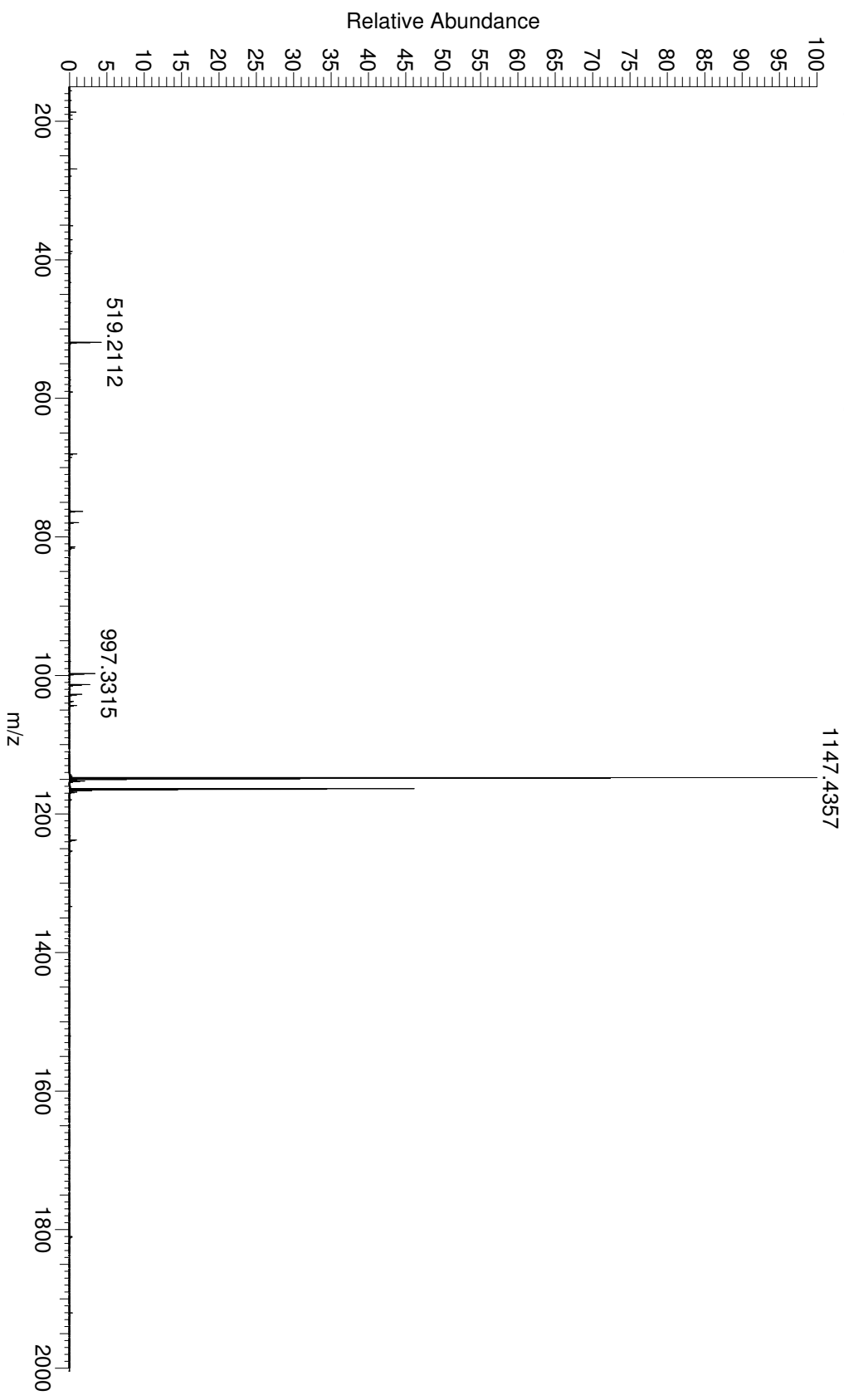
Methyl (phenyl 4-O-(2-azido-3-O-benzyl-6-O-benzoyl-2-deoxy-4-O-[(S)-2,3-bis(benzyloxy)propoxy]- α -D-glucopyranosyl)-2-O-benzoyl-3-O-benzyl-1-thio- α -L-idopyranoside)-uronate (21)

GM_229 MW=1130?
(DCM)/MeOH + NH₄OAc)

EPSRC National Centre Swansea
LTQ Orbitrap XL

Gavin Miller
16/11/2012 14:07:26

MANGAR221-OV-HNESP #21.44 RT: 0.79-1.44 AV: 24 NL: 1.50E8
T: FTMS + p NSI Full ms [150.00-2000.00]

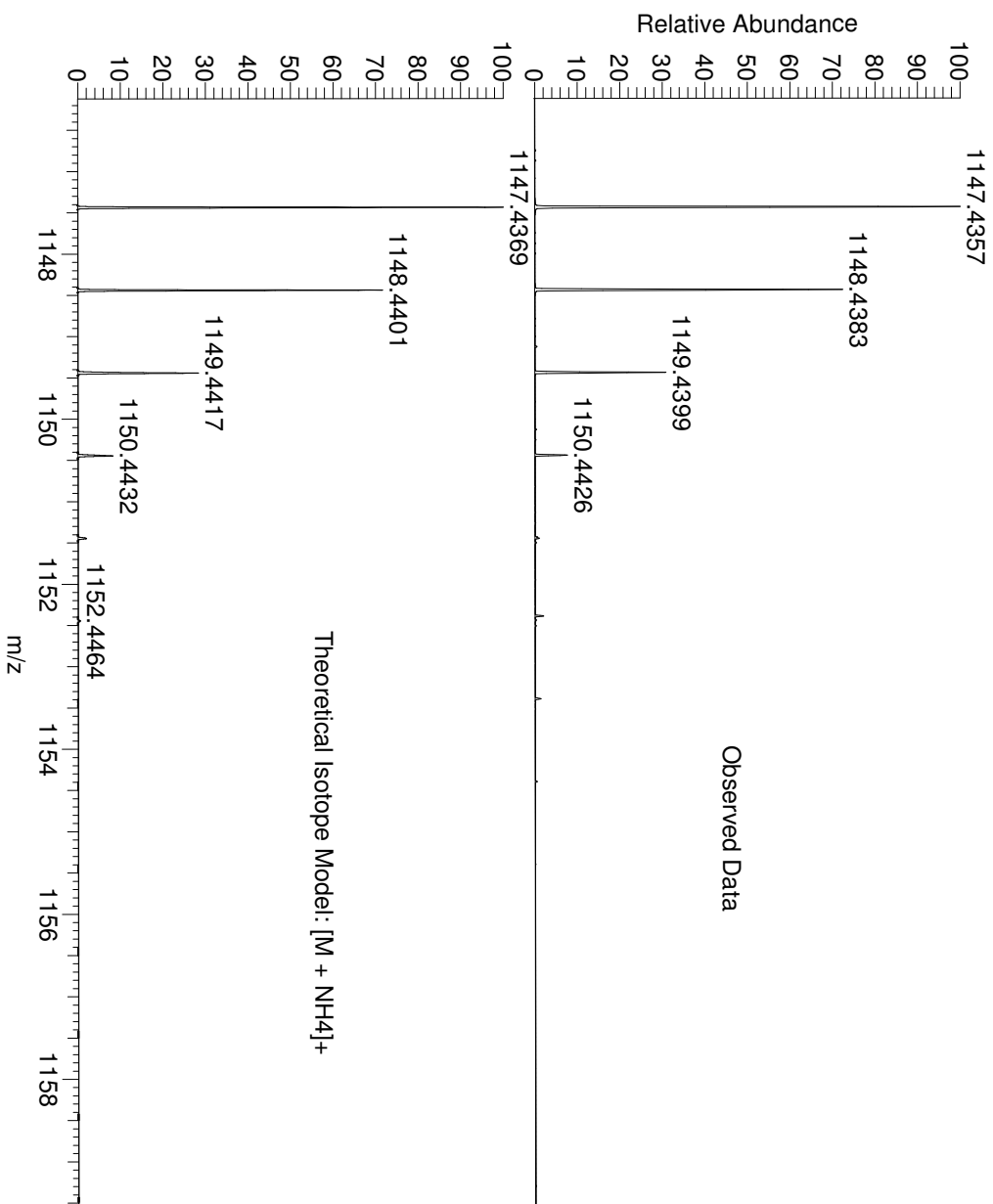


Methyl (phenyl 4-O-(2-azido-3-O-benzyl-6-O-benzoyl-2-deoxy-4-O-[(S)-2,3-bis(benzyloxy)propoxy]- α -D-glucopyranosyl)-2-O-benzoyl-3-O-benzyl-1-thio- α -L-idopyranoside)-uronate (21)

GM 229 MW=1130?
(DCM)/MeOH + NH4OAc)

EPSRC National Centre Swansea
LTQ Orbitrap XL

Gavin Miller
16/11/2012 14:07:26

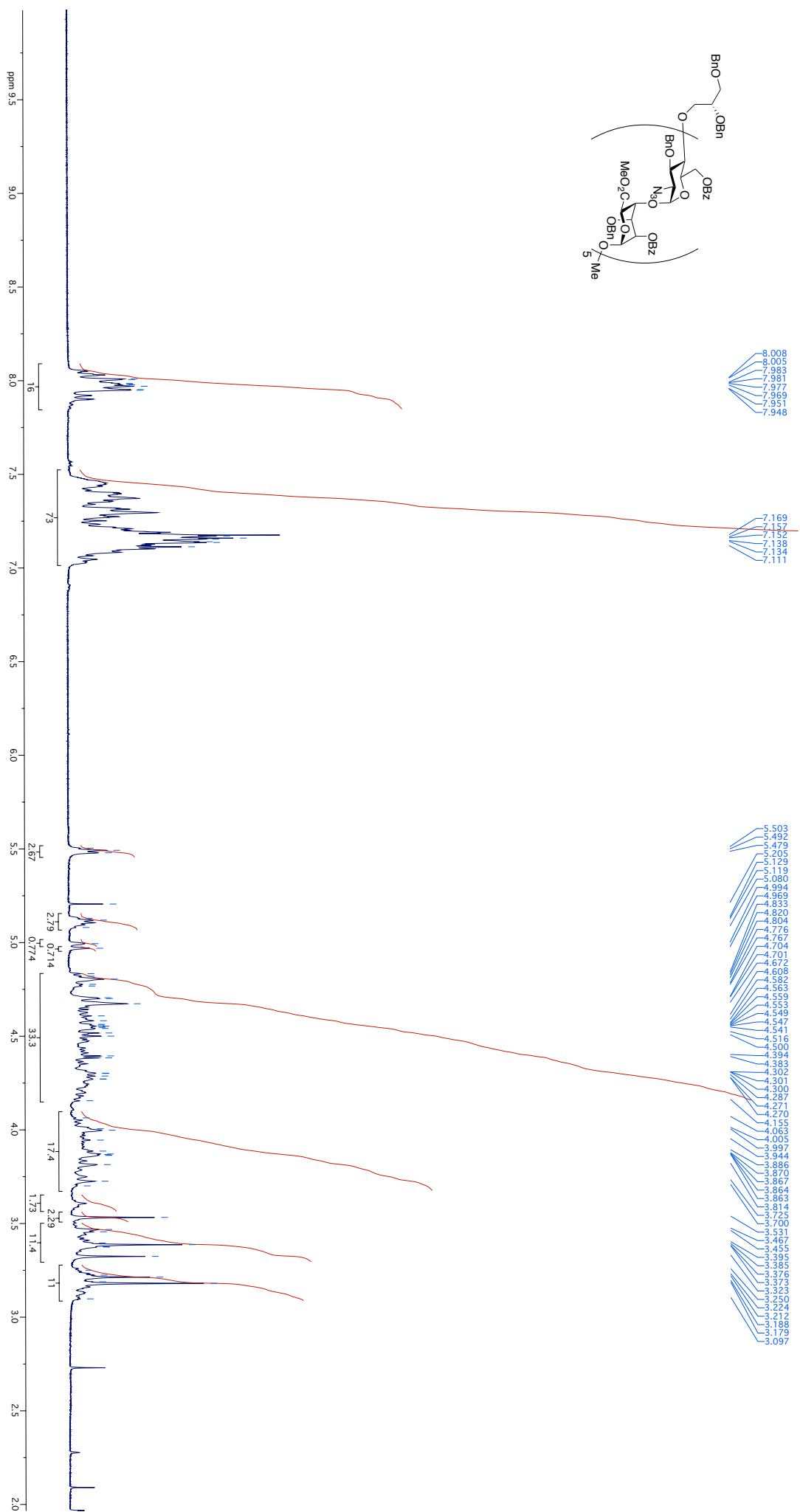


NL:
1.50E8
MANGAR221-OV-HNESPP#21-
44 RT: 0.79-1.44 AV: 24 T:
FTMS + p NSI Full ms
[150.00-2000.00]

NL:
1.06E4
C₆₄ H₆₃ N₃ O₁₄ SNH₄:
C₆₄ H₆₇ N₄ O₁₄ S₁
p (gss, s/p:40) Chrg 1
R: 100000 Res.: P.wr.: @FWHM

Decasaccharide 25

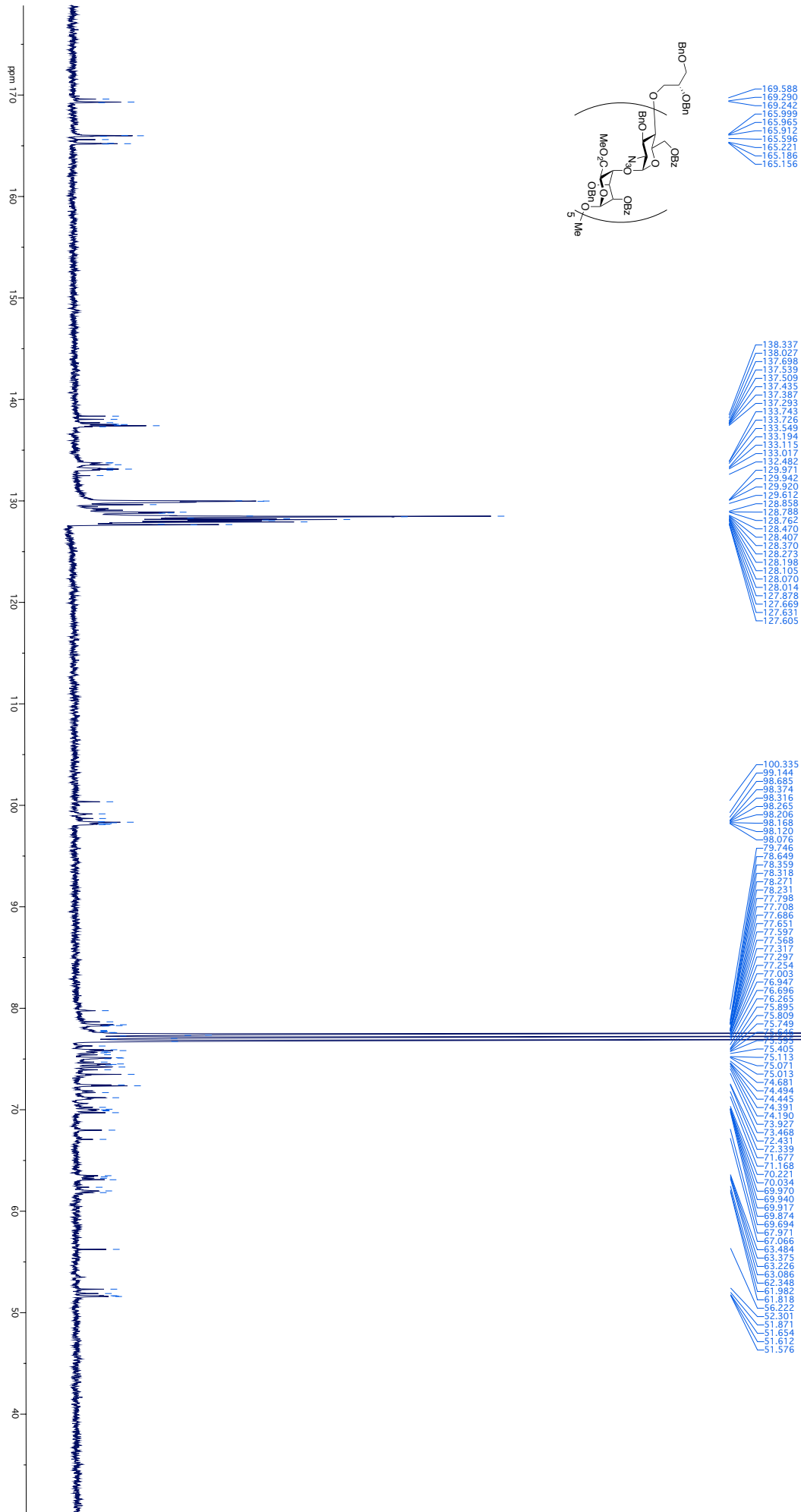
¹H NMR



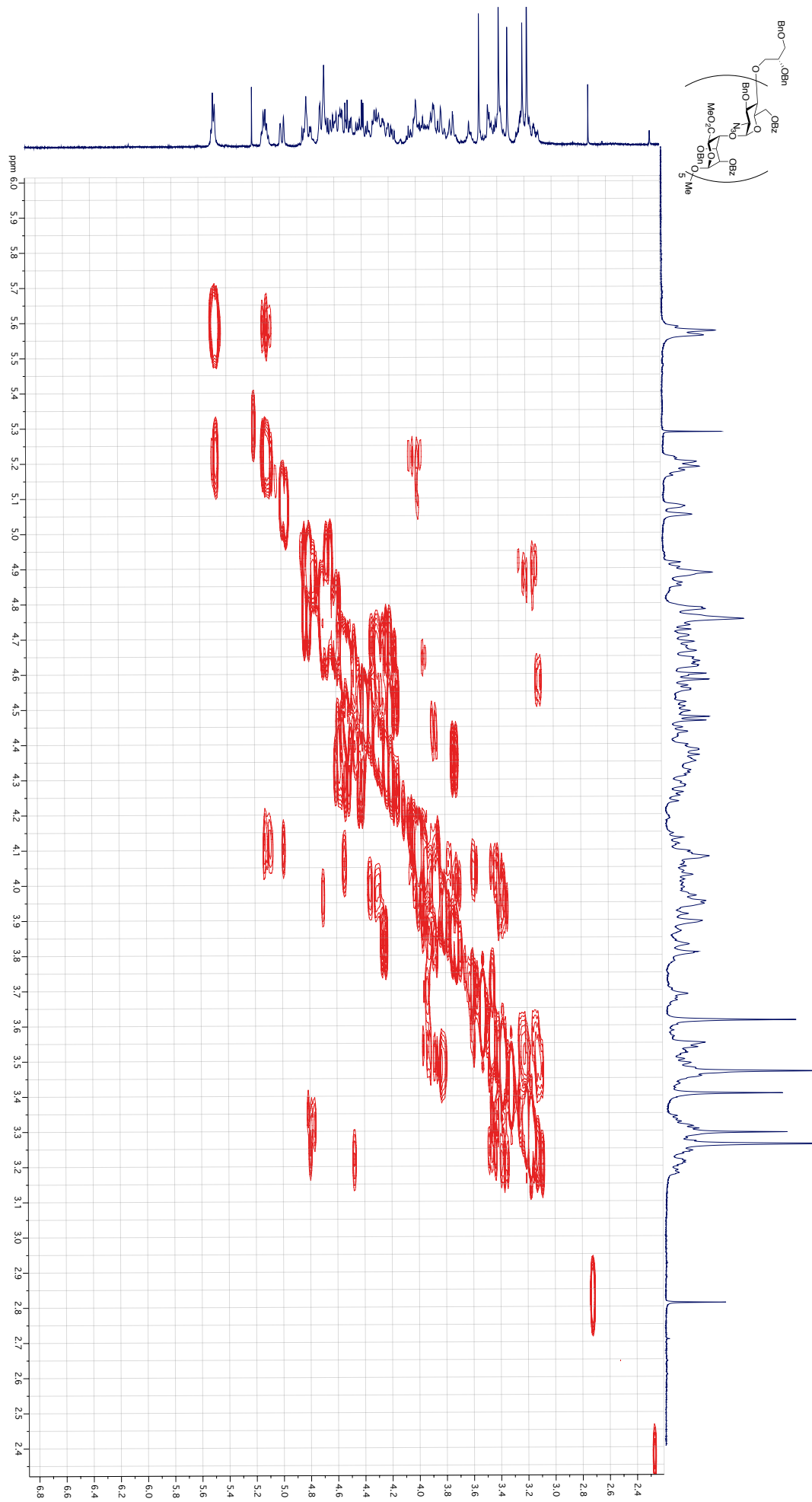
400 MHz, CDCl₃

Decasaccharide 25

¹³C NMR

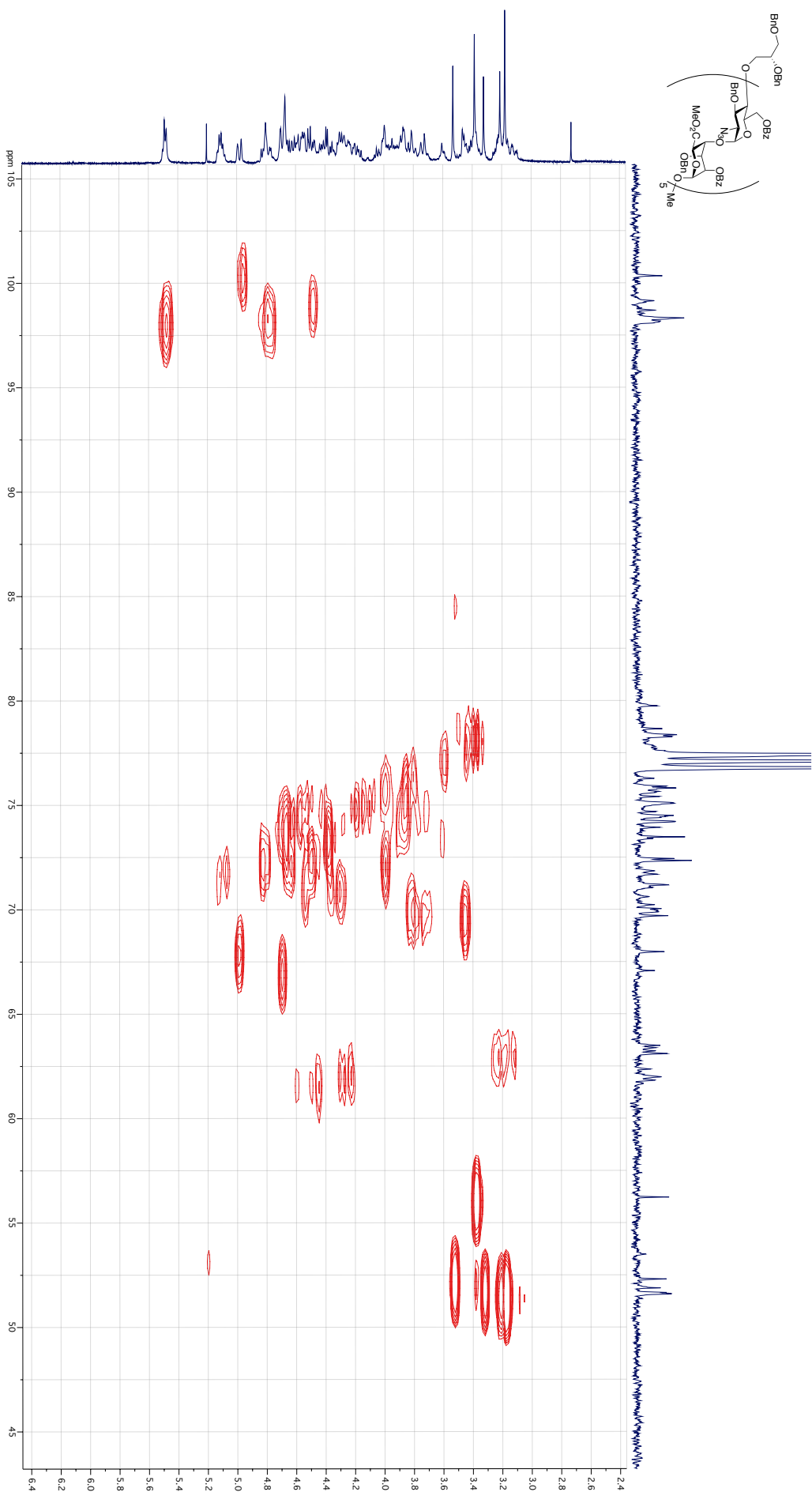


100 MHz, CDCl₃



400 MHz, CDCl₃

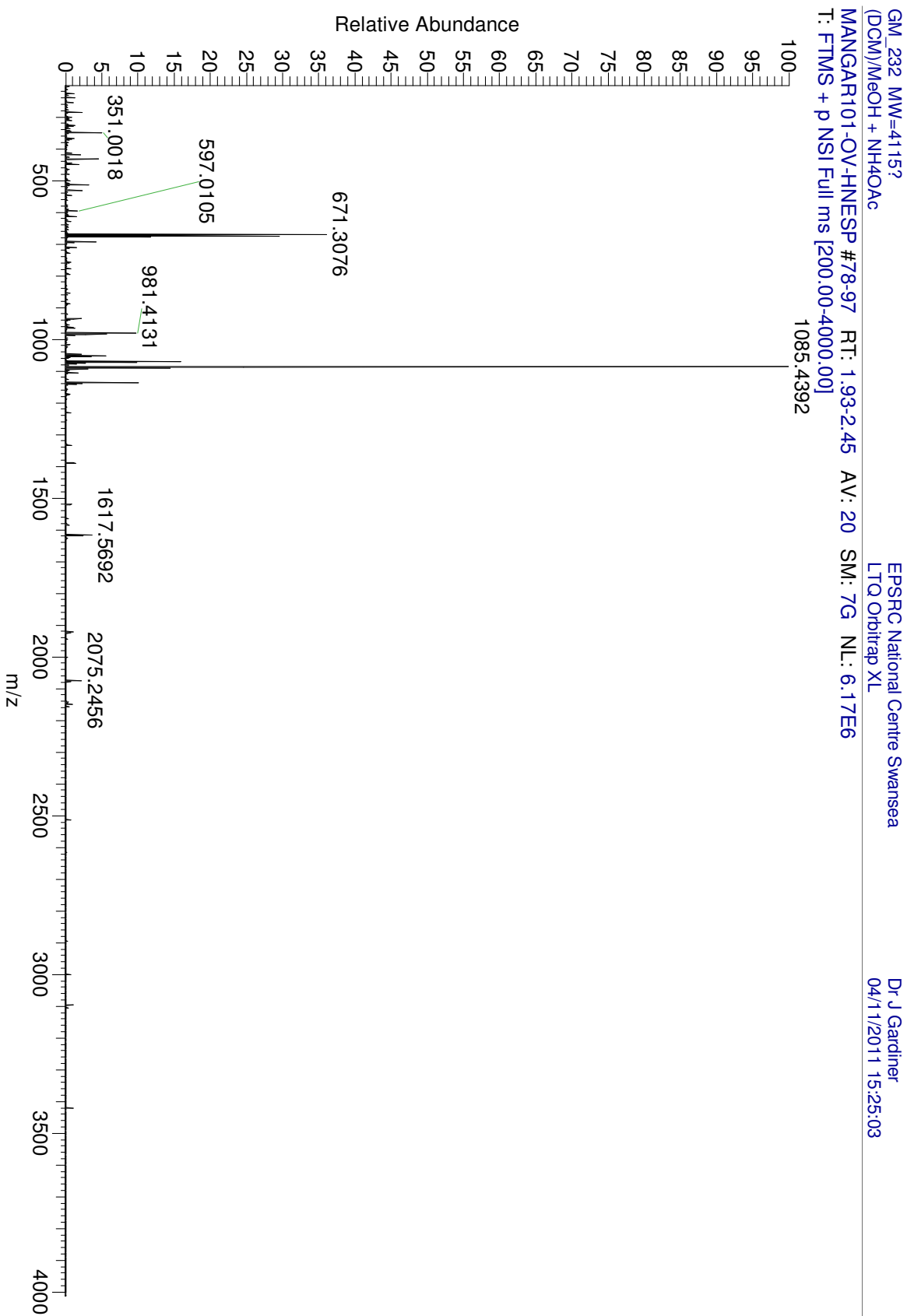
Decasaccharide 25



HMOC

400 MHz, CDCl₃

Decasaccharide 25- ESI MS

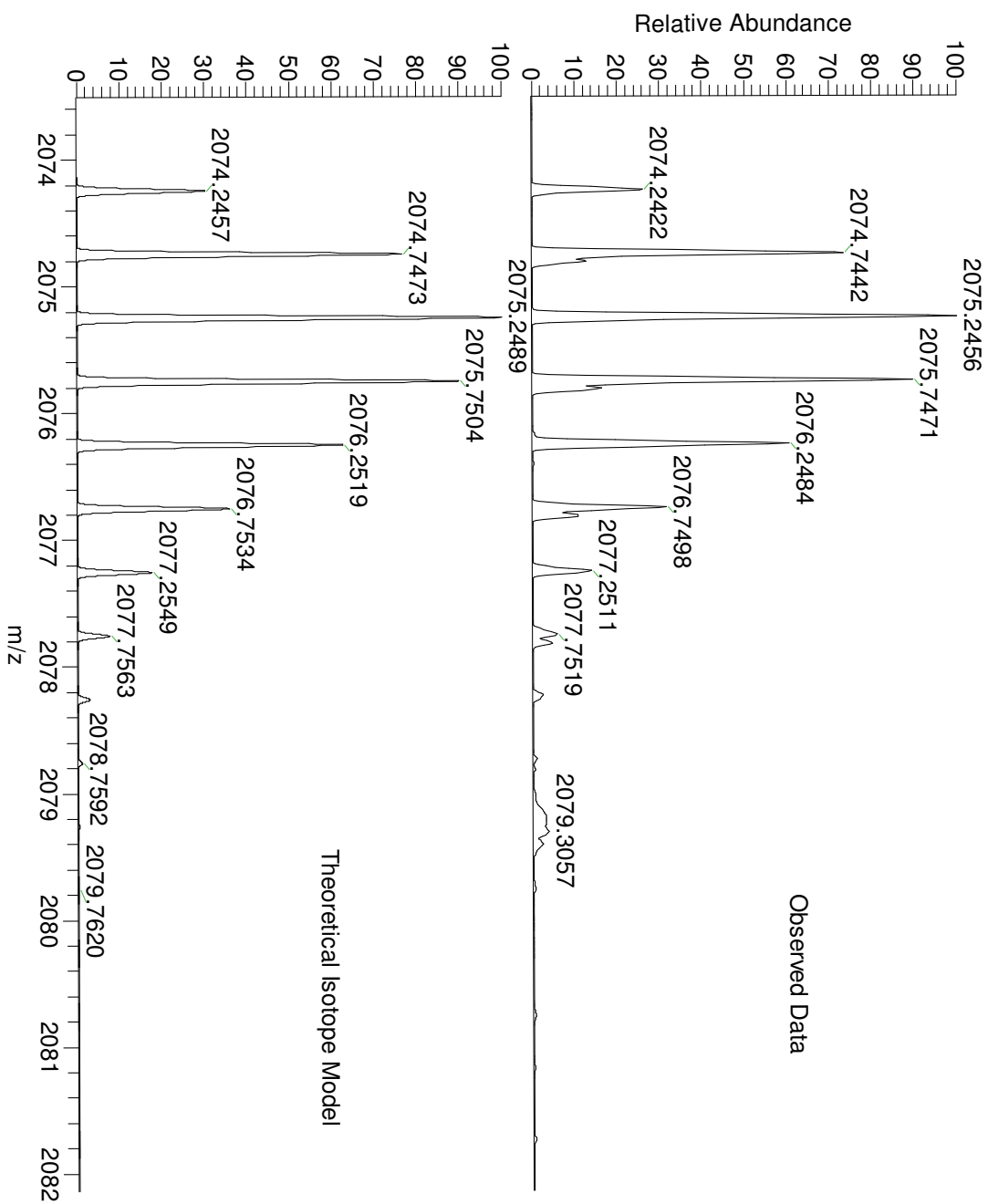


Decasaccharide 25-ESI MS Isotope Pattern

GM_232 MW=4115?
(DCM)/MeOH + NH4OAc

EPSRC National Centre Swansea
LTQ Orbitrap XL

Dr J Gardiner
04/11/2011 15:25:03

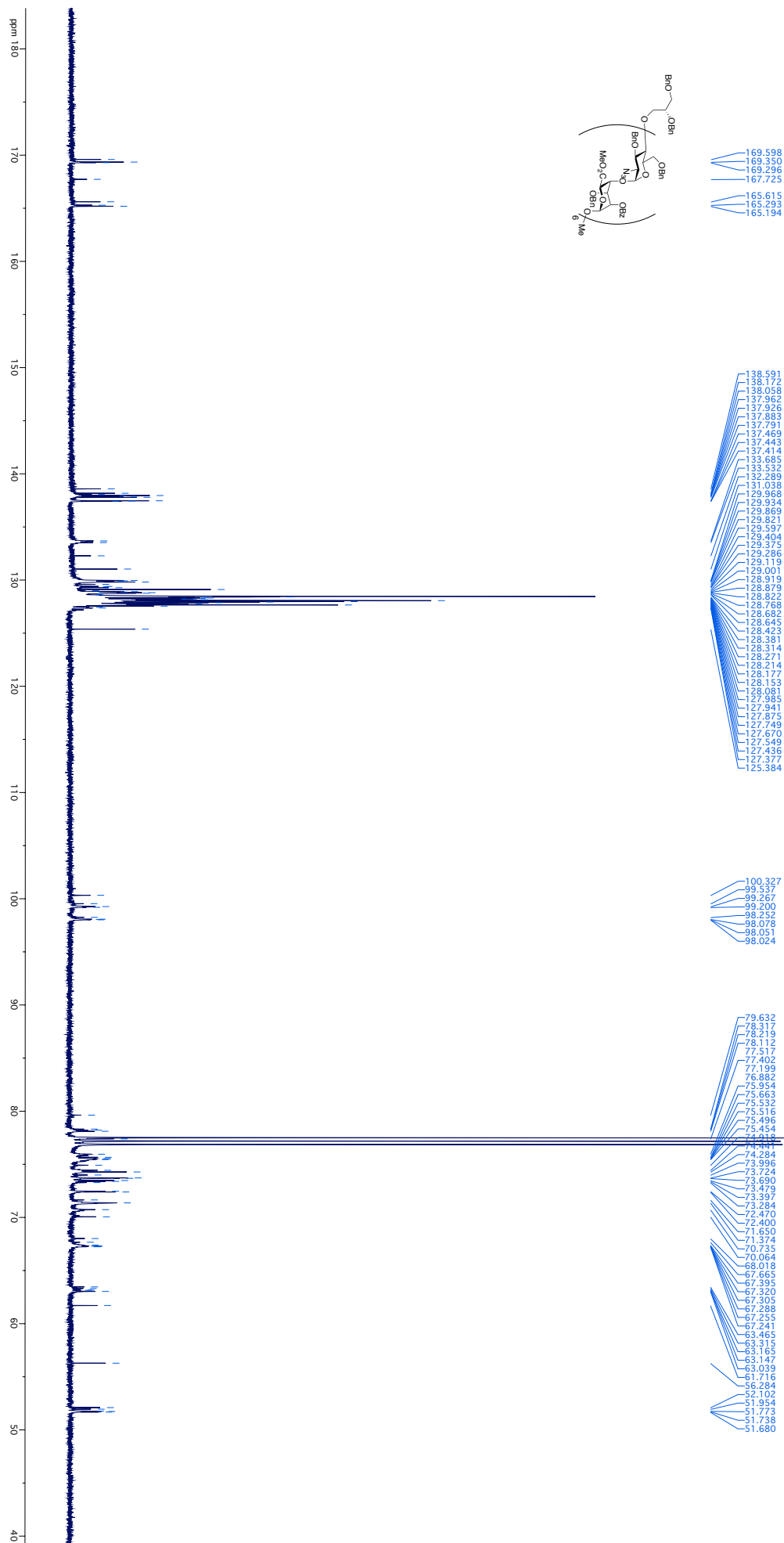


NL:
1.35E5
MANGAR101-OV-HNESP#78-
97 RT: 1.93-2.45 AV: 20 T:
FTMS + p NSI Full ms
[200.00-4000.00]

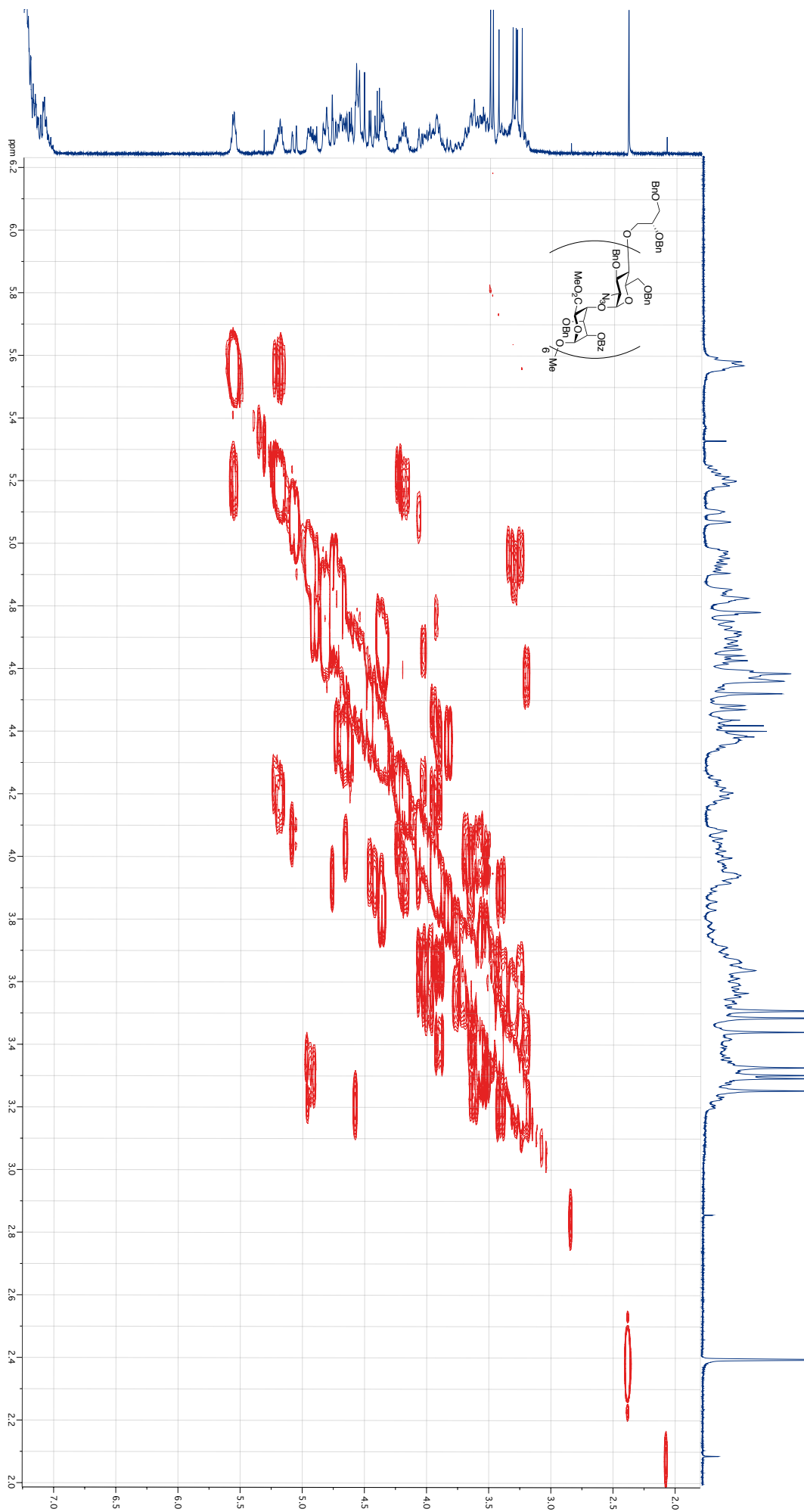
NL:
5.52E3
C₂₂₃ H₂₁₇ N₁₅ O₆₃ NH₄ NH₄:
C₂₂₃ H₂₂₅ N₁₇ O₆₃
p (gss, s /p:40) Chrg 2
R: 55000 Res .Pwr . @FWHM

Dodecasaccharide 24

¹³C NMR

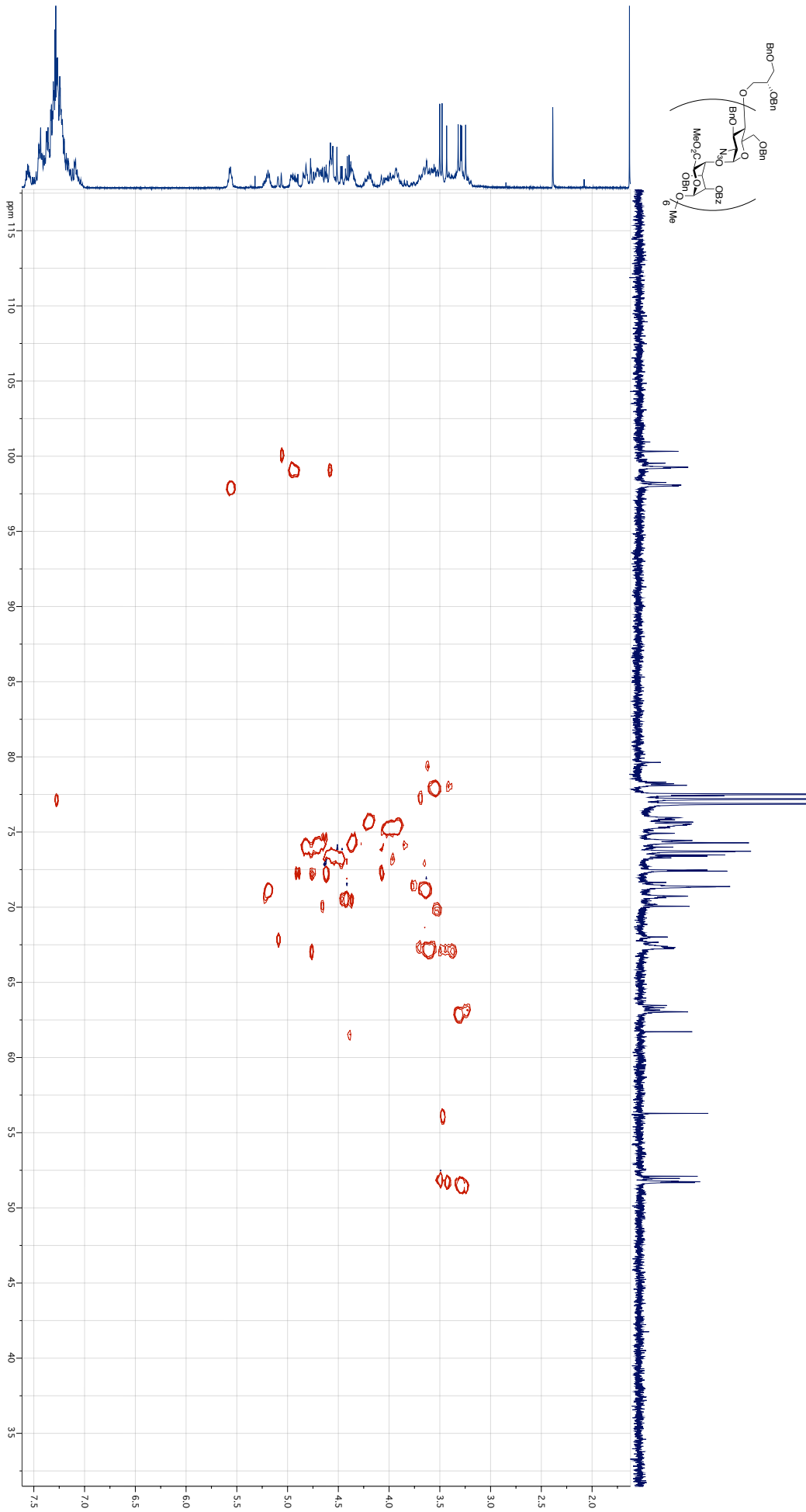


100 MHz, CDCl₃



400 MHz, CDCl₃

Dodecasaccharide 24

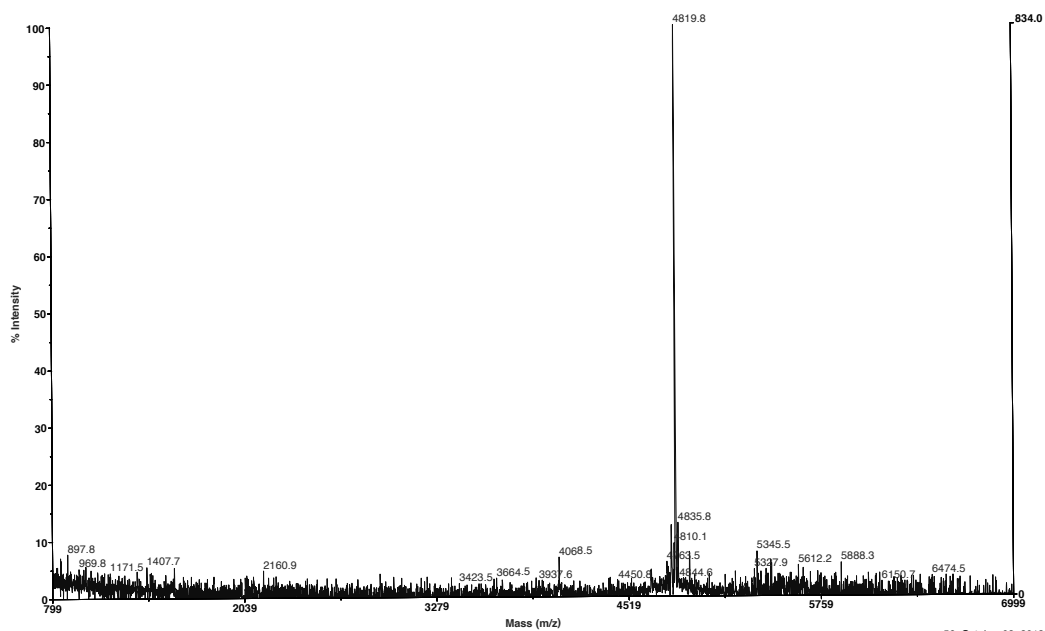


HSQC

400 MHz, CDCl₃

Dodecasaccharide 24 - MALDI MS and isotope pattern

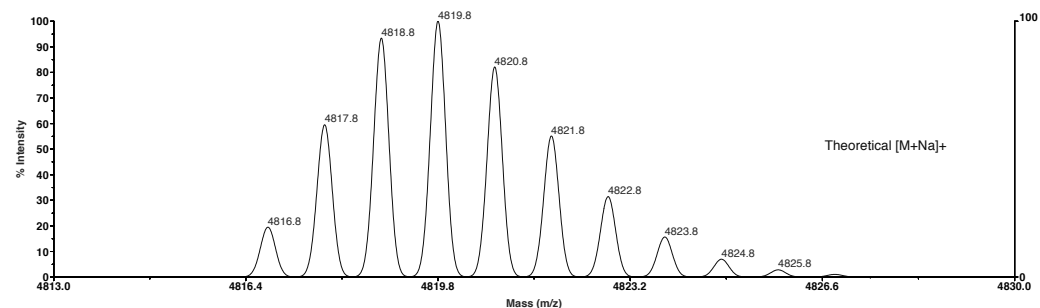
EPSRC National Mass Spectrometry Service Centre (NMSSC), Swansea
 <<MANGAR203-VA-MAP_0001>> Voyager Spec #1=>AdvBC(64,0.5,0.1)->NF0.7[BP = 4819.8, 834]



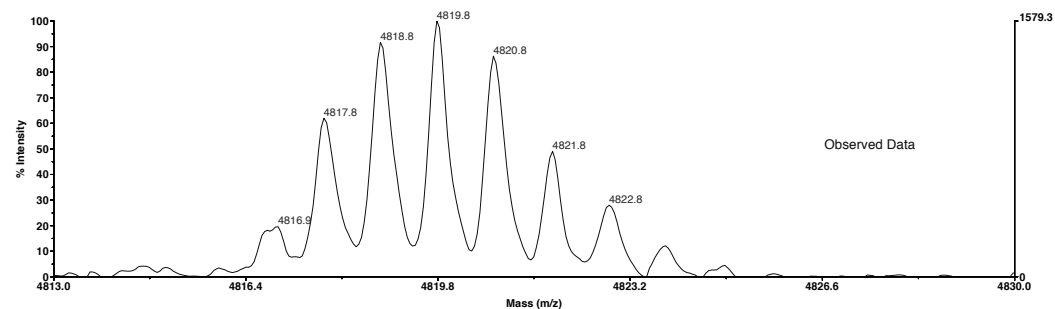
Acquired: 12:55:00, October 03, 2012
 G.Miller GM_302-2 MW=4797?? DCM PosRef [1:24] (DCTB,DCM) +NaOAc
 D:\2012\Oct12\MANGAR203-VA-MAP_0001.dat

Printed: 12:56, October 03, 2012

EPSRC National Mass Spectrometry Service Centre (NMSSC), Swansea
 ISO:C264H268N18O69 + (Na)1



<<MANGAR243-VM-MAP_0001>> Voyager Spec #1=>AdvBC(64,0.5,0.1)->SM5[BP = 594.1, 7804]

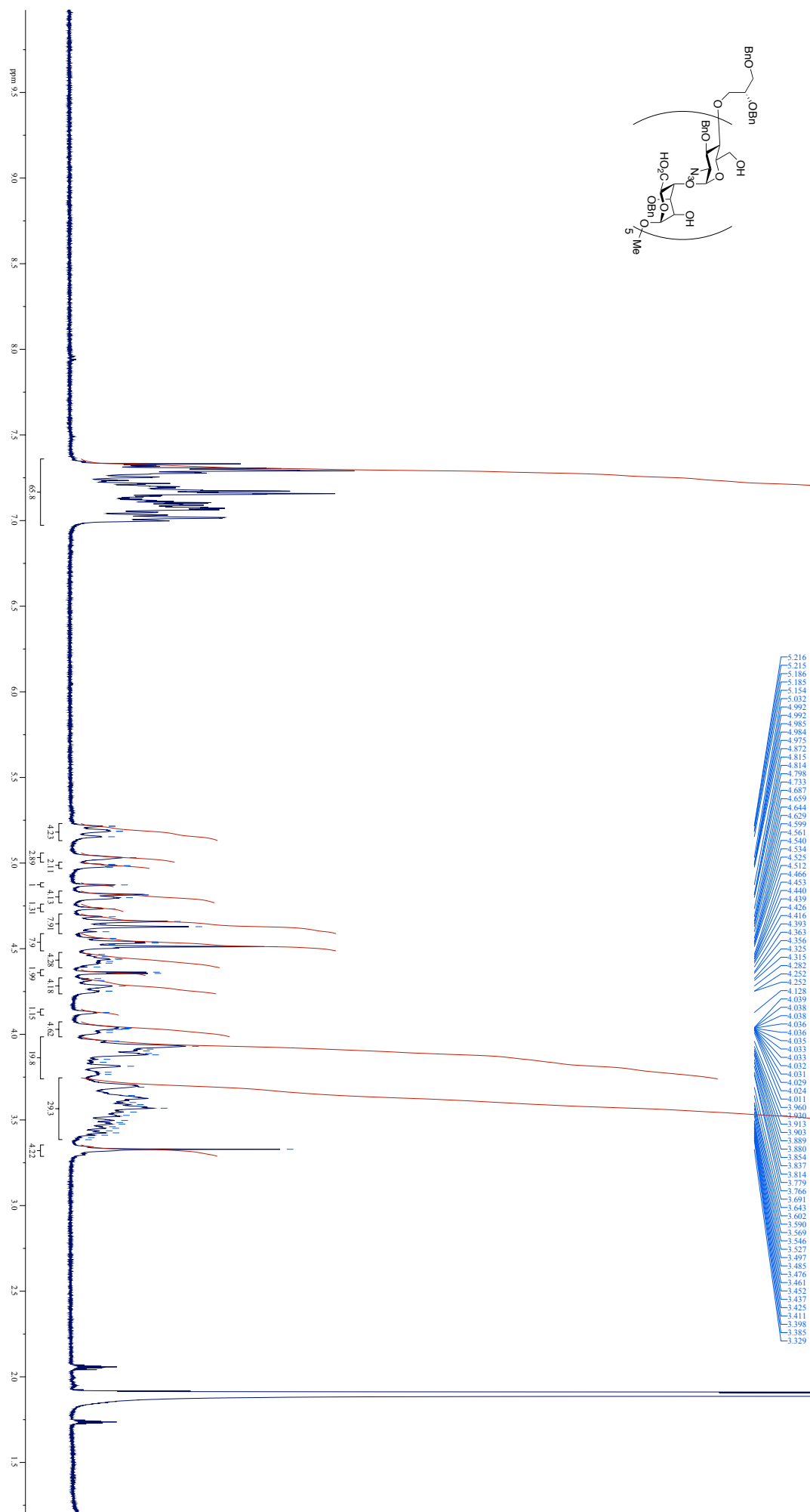


Acquired: 10:53:00, February 11, 2013
 Miller GM_302 MW=4797?? DCM PosRef [1:49] (Dith,DCM) +NaOAc
 D:\2013\Feb13\MANGAR243-VM-MAP_0001.dat

Printed: 12:00, February 11, 2013

Decasaccharide Saponification

¹H NMR



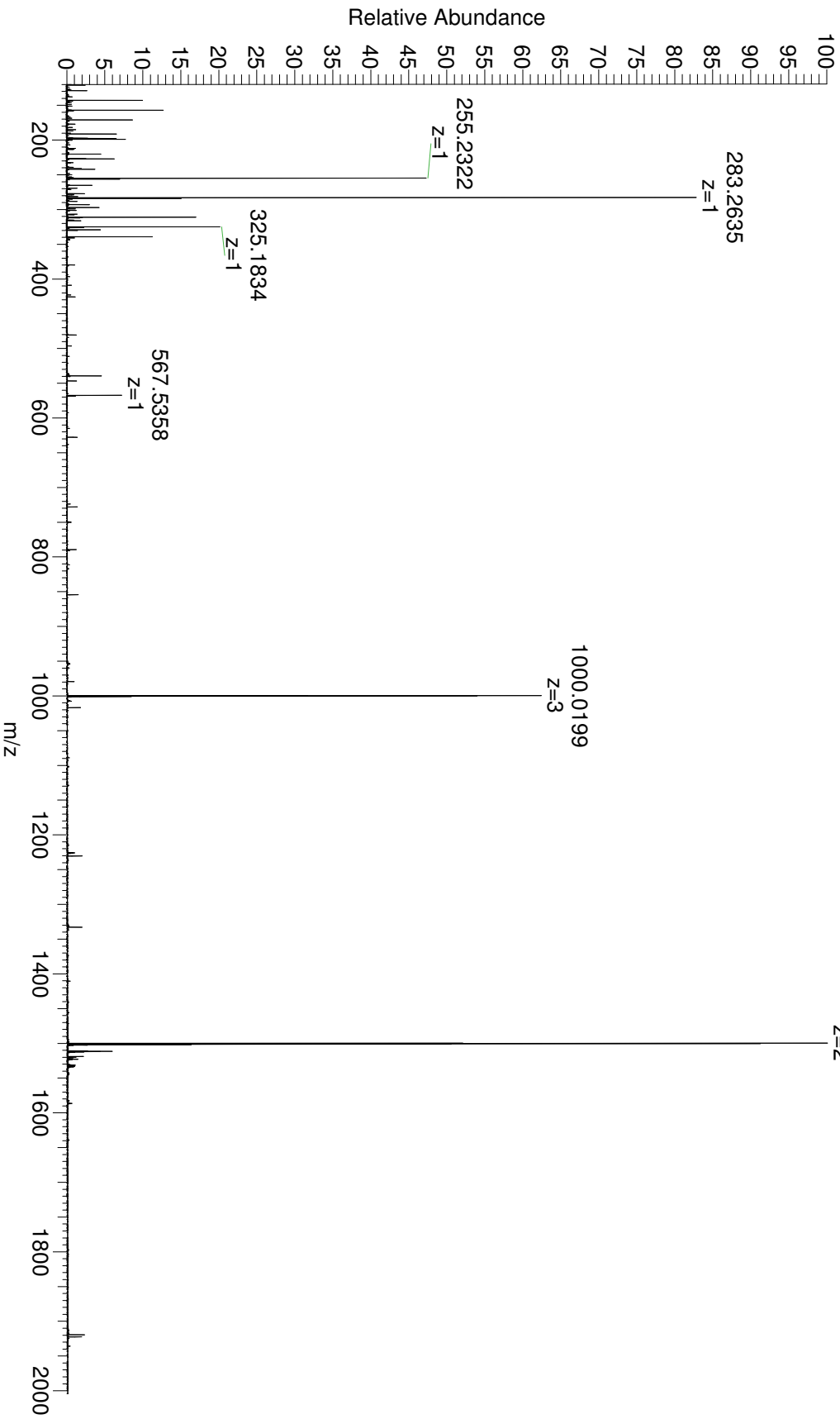
400 MHz, AcOH

Decasaccharide Saponification- ESI MS

GM_238 MW=3003?
(DCM)/MeOH
MANGAR104-OV-HNESN-3-#4-9 RT: 0.27-0.42 AV: 6 NL: 1.01E6
T: FTMS - p NSI Full ms [120.00-2000.00]

EPSPRC National Centre Swansea
LTO.Orbitrap XL

Gavin Miller
28/11/2011 11:22:42

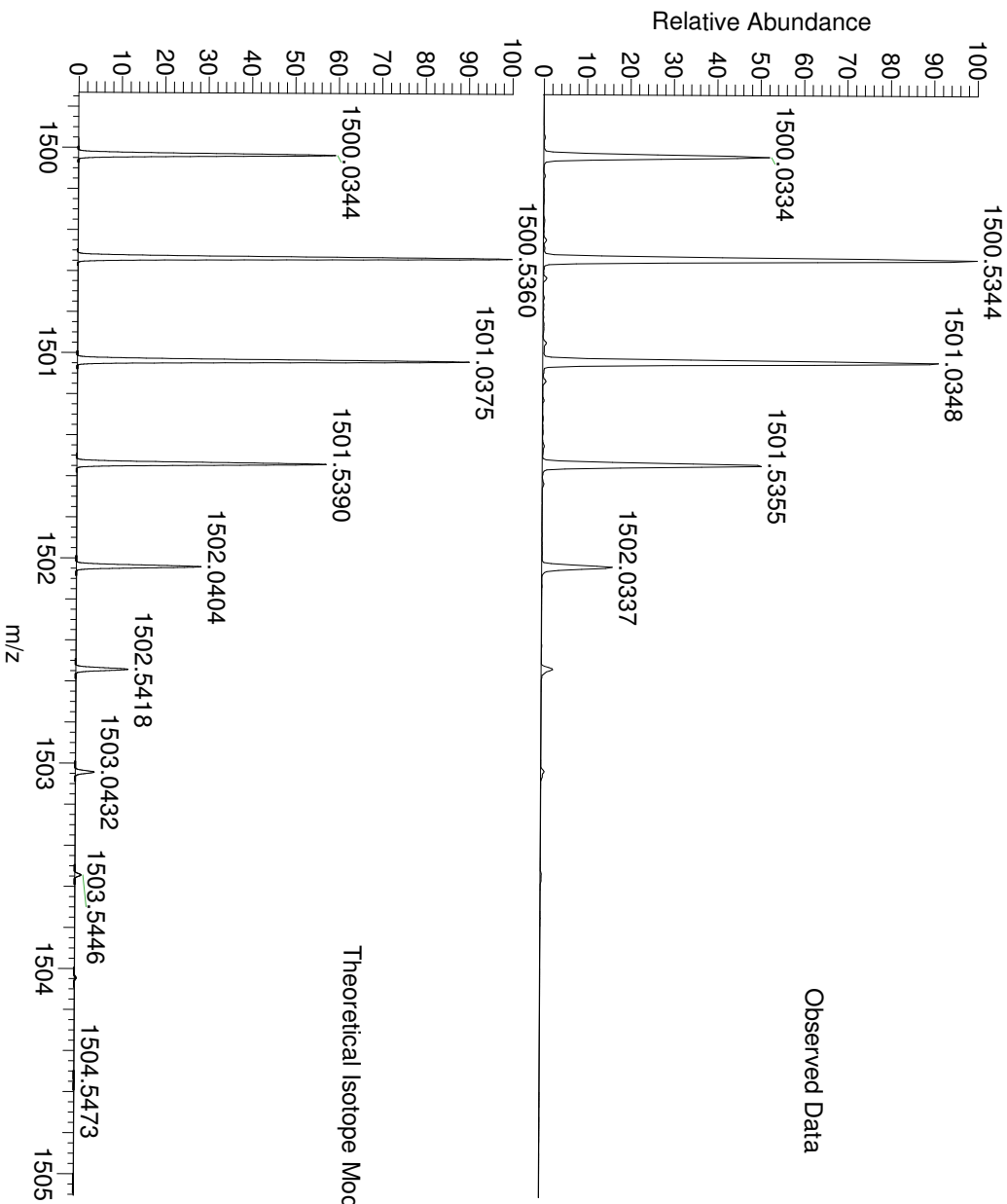


Decasaccharide Saponification-ESI MS Isotope Pattern

GM 238 MW=3003?
(DCM)/MeOH

EPSRC National Centre Swansea
LTQ Orbitrap XL

Gavin Miller
28/11/2011 11:22:42



Observed Data

Theoretical Isotope Model

NL:
1.01E6
MANGAR104-OV-HNESN-3#4-
9 RT: 0.27-0.42 AV: 6 T:
FTMS - p NSI Full ms
[120.00-2000.00]

NL:
6.58E3
C₁₄₈ H₁₆₅ N₁₅ O₅₃:
C₁₄₈ H₁₆₅ N₁₅ O₅₃
p (gss, s/p:40) Chrg -2
R: 100000 Res.Pwr. @FWHM

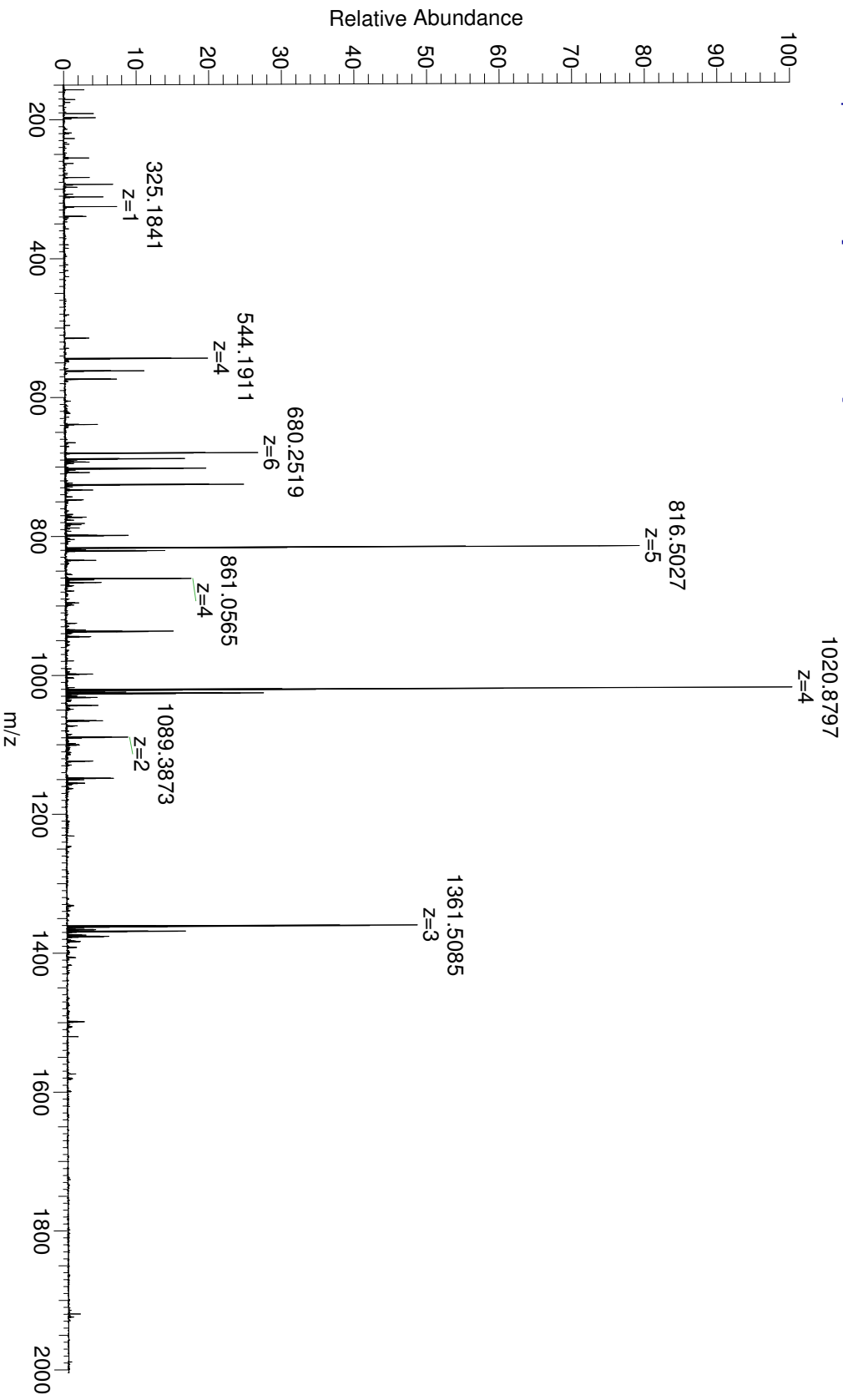
Dodecasaccharide Saponification- ESI MS

GM_308-1 MW=4088?
(MeOH)/(MeOH)

EPSRC National Centre Swansea
LTQ Orbitrap XL

Gavin Miller
19/11/2012 10:46:14

MANGAR222-OV-HNESN #16-18 RT: 1.39-1.57 AV: 3 NL: 2.28E6
T: FTMS - p NSI Full ms [150.00-2000.00]

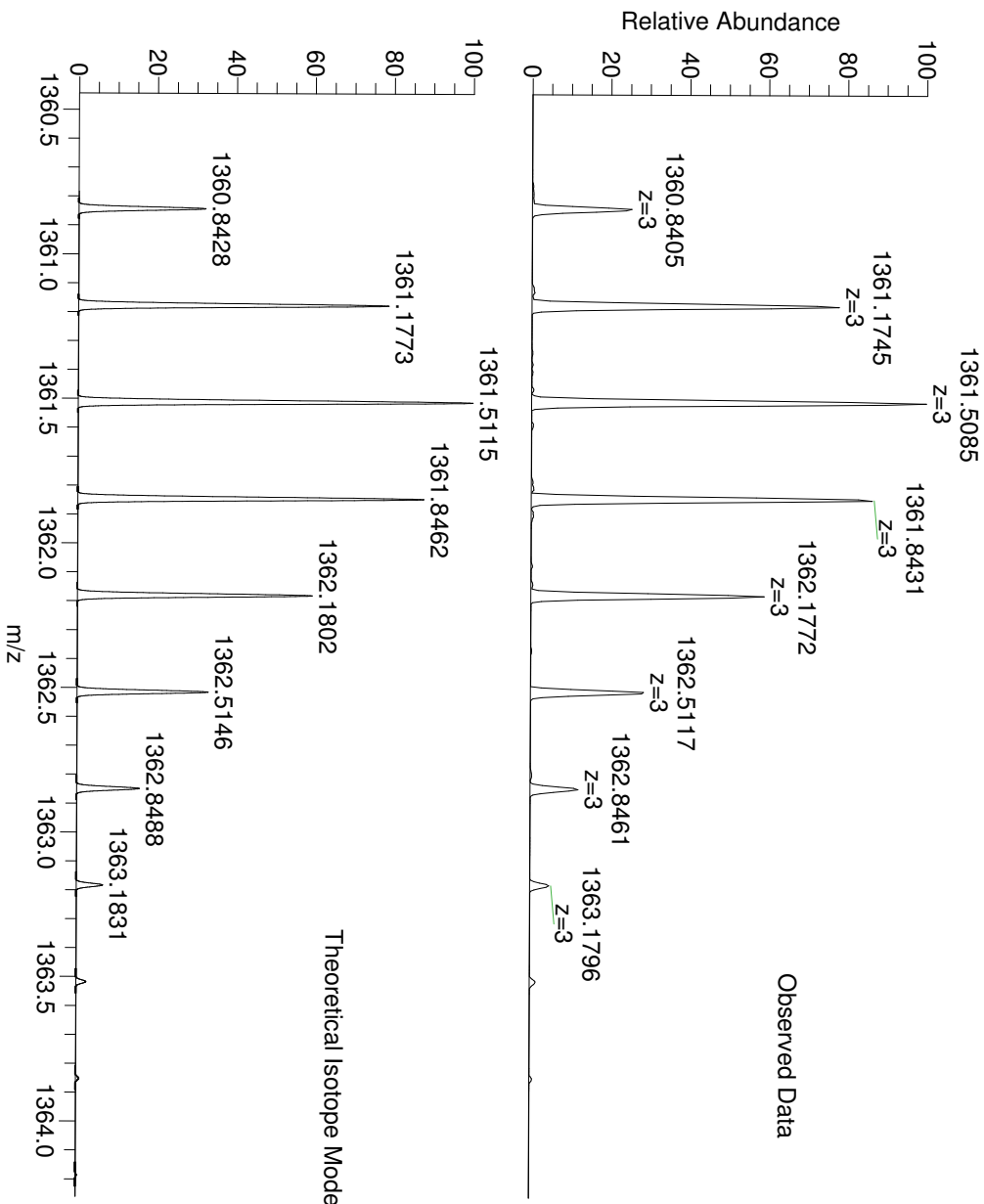


Dodecasaccharide Saponification-ESI MS Isotope Pattern

GM 308-1 MW=4088?
(MeOH)/(MeOH)

EPSRC National Centre Swansea
LTQ Orbitrap XL

Gavin Miller
19/11/2012 10:46:14

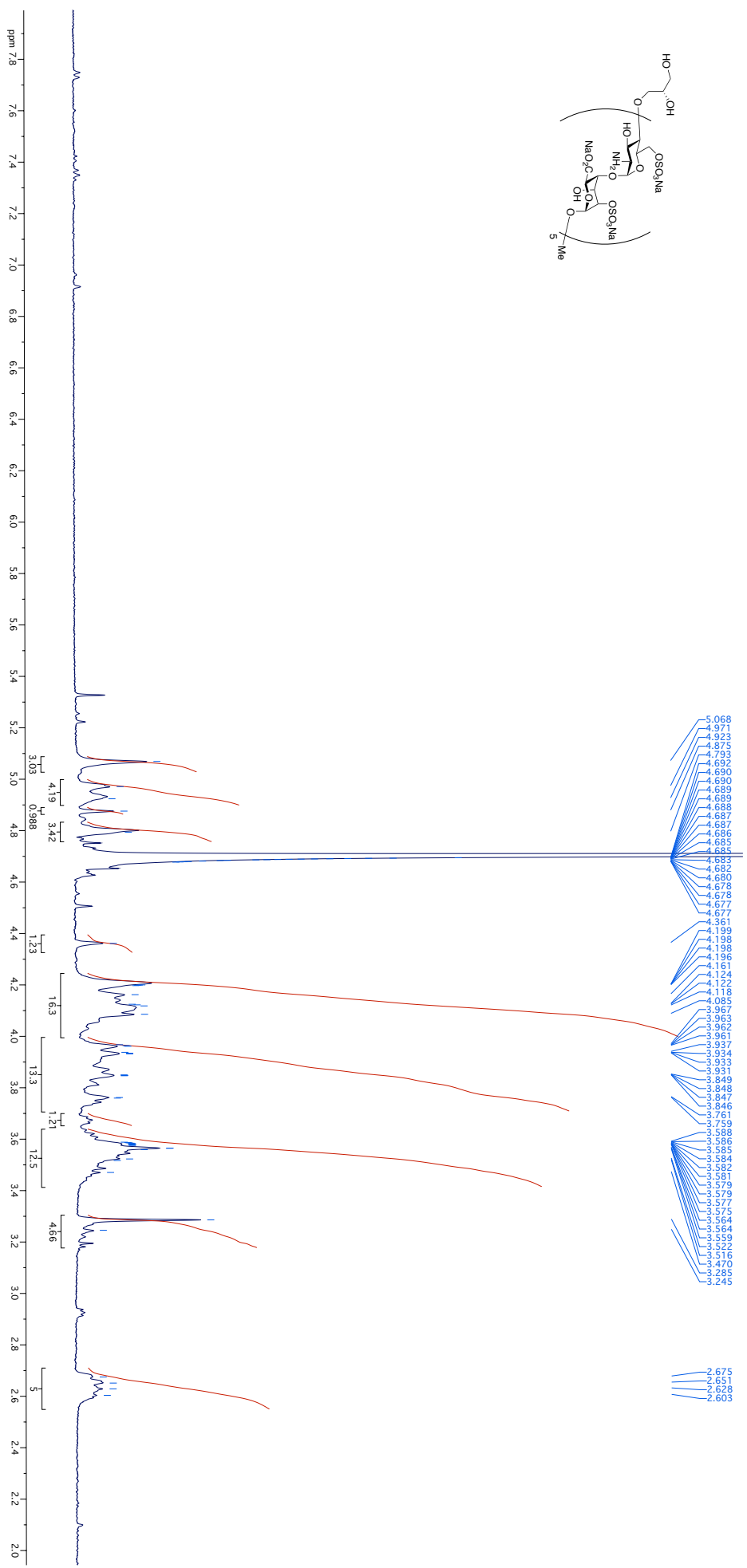


NL:
1.10E6
MANGARR222-OV-HNESN#16-
18 RT: 1.39-1.57 AV: 3 T:
FTMS - p NSI Full ms
[150.00-2000.00]

NL:
5.59E3
C₂₁₆H₂₂₉N₁₈O₆₃:
C₂₁₆H₂₂₉N₁₈O₆₃
p.(gss, s.(p:40) Chrg -3
R: 100000 Res.Pwr. @FWHM

Decasaccharide hydrogenation

¹H NMR



400 MHz, D₂O

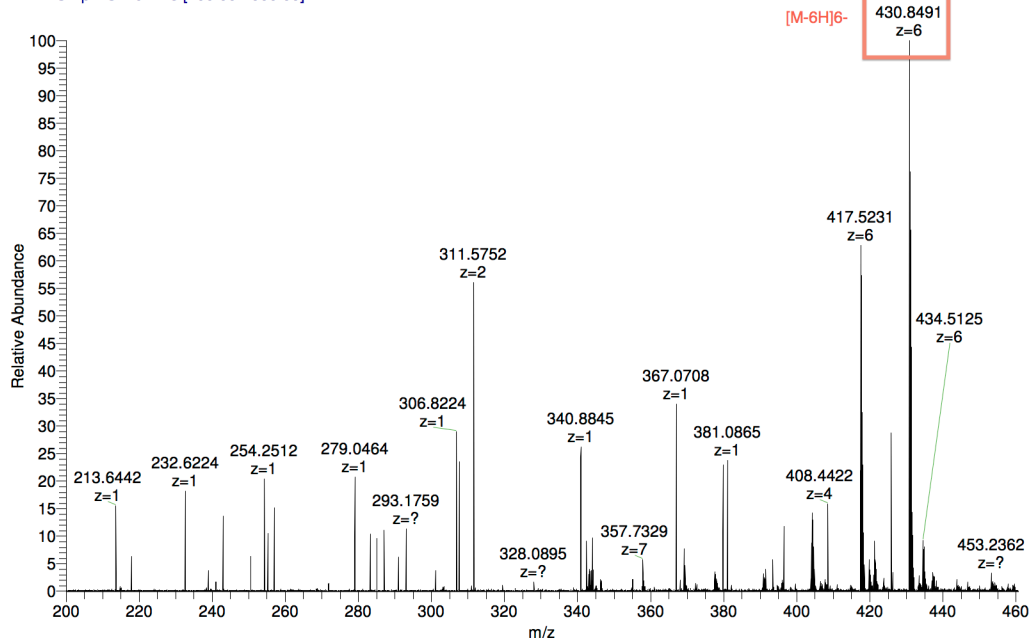
Decasaccharide hydrogenation- ESI MS

GM_247 MW=2848?
(H₂O)/MeOH + 10 μ l DEA

EPSRC National Centre Swansea
LTQ Orbitrap XL

Gavin Miller
19/01/2012 13:53:08

MANGAR119-OV-HNESN-2 #149-172 RT: 2.34-2.98 AV: 24 SM: 7G NL: 1.57E5
T: FTMS - p NSI Full ms [200.00-4000.00]

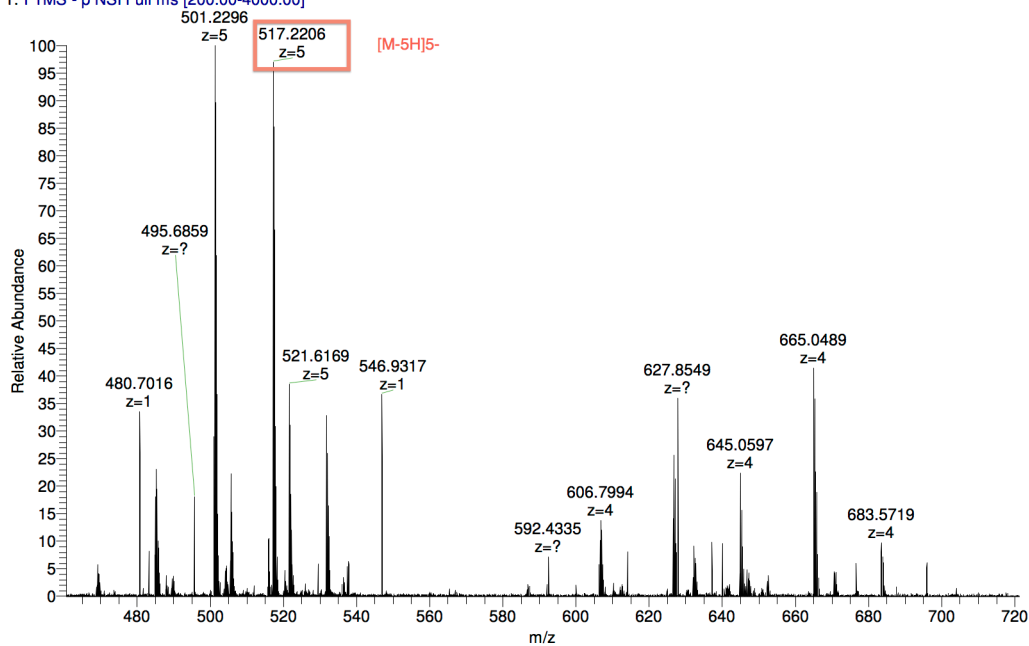


GM_247 MW=2848?
(H₂O)/MeOH + 10 μ l DEA

EPSRC National Centre Swansea
LTQ Orbitrap XL

Gavin Miller
19/01/2012 13:53:08

MANGAR119-OV-HNESN-2 #149-172 RT: 2.34-2.98 AV: 24 SM: 7G NL: 1.42E5
T: FTMS - p NSI Full ms [200.00-4000.00]



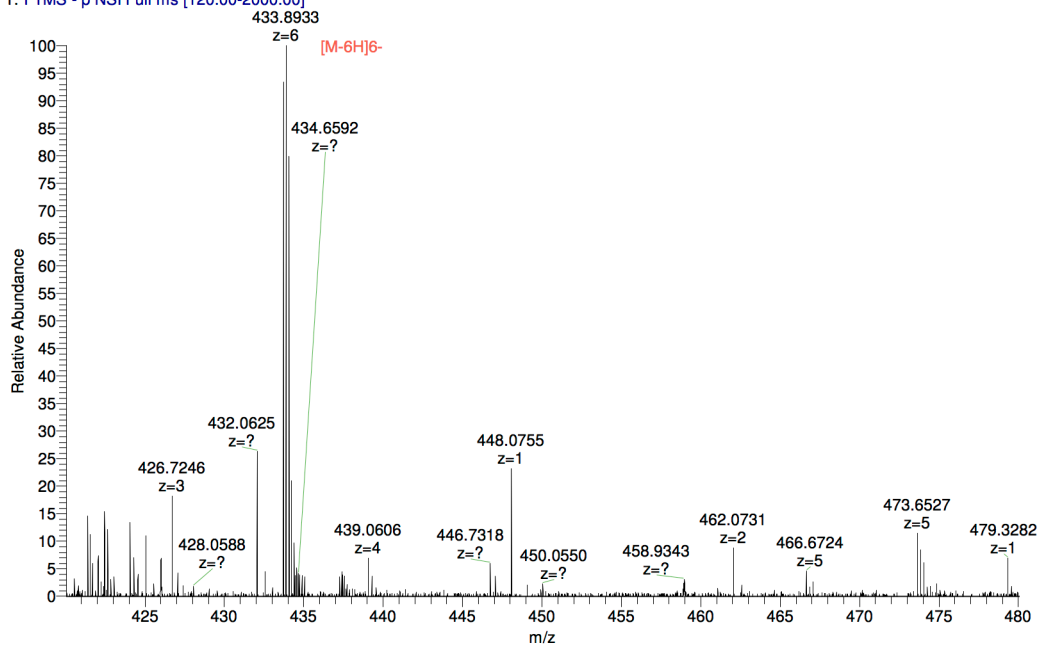
Dodecasaccharide hydrogenation- ESI MS

GM_311-1 MW=2610?
(H₂O)/MeOH + DEA

EPSRC National Centre Swansea
LTQ Orbitrap XL

Gravin Miller
29/01/2013 11:19:47

MANGAR245-OV-HNESN #6-10 RT: 0.33-0.44 AV: 5 SM: 7G NL: 7.22E5
T: FTMS - p NSI Full ms [120.00-2000.00]

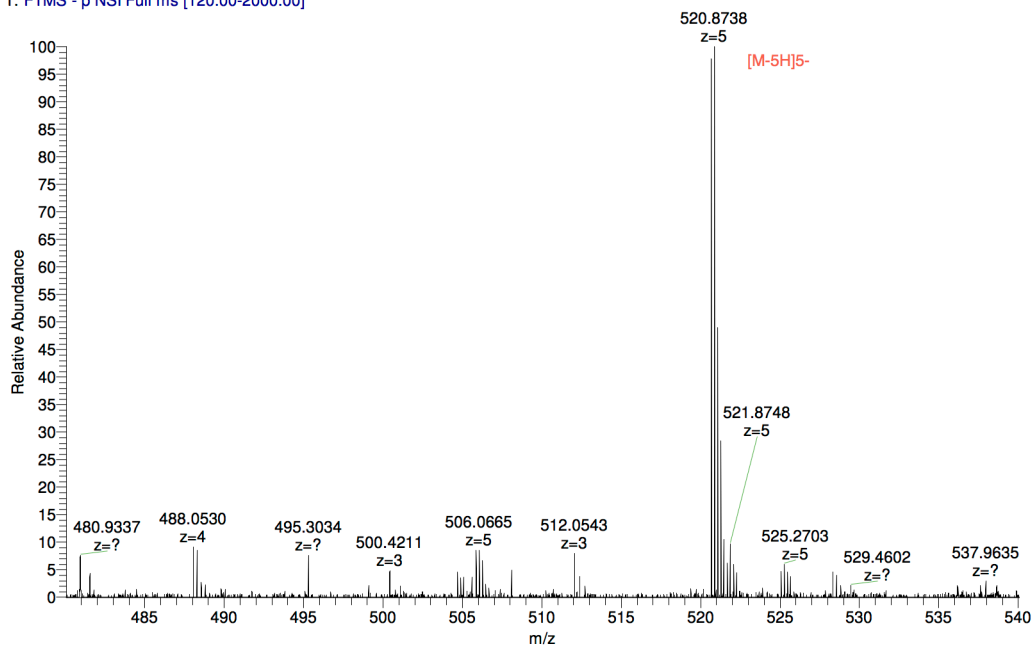


GM_311-1 MW=2610?
(H₂O)/MeOH + DEA

EPSRC National Centre Swansea
LTQ Orbitrap XL

Gravin Miller
29/01/2013 11:19:47

MANGAR245-OV-HNESN #6-10 RT: 0.33-0.44 AV: 5 SM: 7G NL: 6.91E5
T: FTMS - p NSI Full ms [120.00-2000.00]

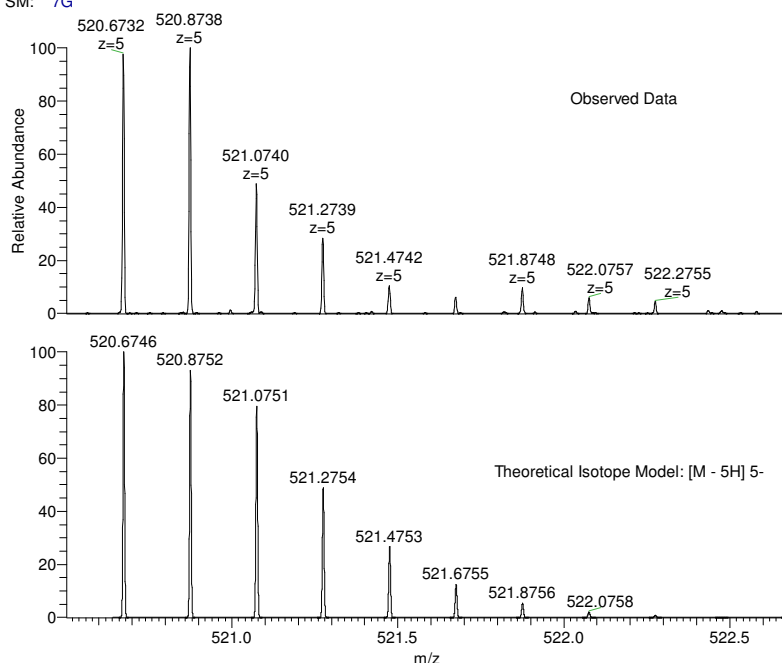


Dodecasaccharide hydrogenation- ESI MS isotope patterns

GM_311-1 MW=2610?
(H₂O)/MeOH + DEA
SM: 7G

EPSRC National Centre Swansea
LTQ Orbitrap XL

Gavin Miller
29/01/2013 11:19:47



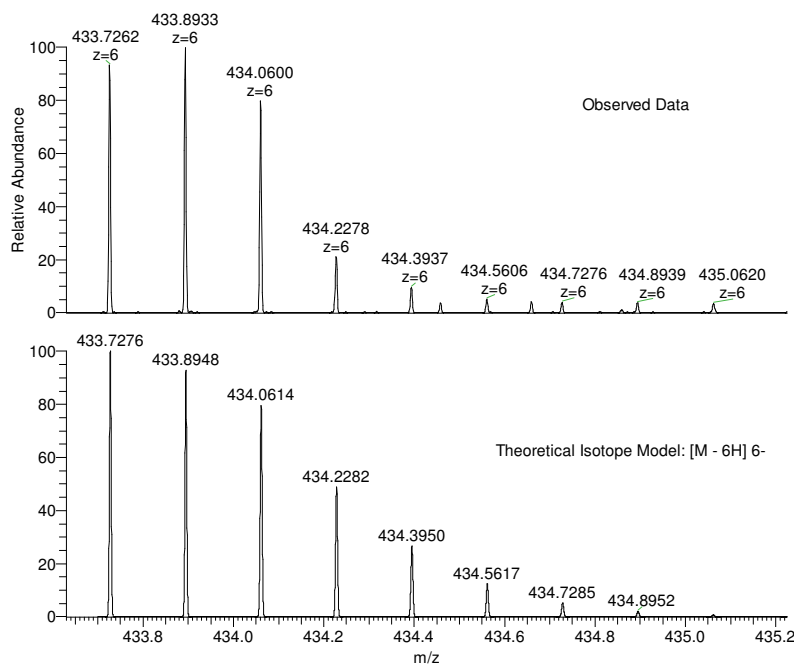
NL:
6.91E5
MANGAR245-OV-HNESN#6-
10 RT: 0.33-0.44 AV: 5 T:
FTMS - p NSI Full ms
[120.00-2000.00]

NL:
6.01E3
C₇₆ H₁₁₉ N₆ O₈₁ S₆:
C₇₆ H₁₁₉ N₆ O₈₁ S₆:
p (gss, s /p:40) Chrg -5
R: 100000 Res .Pwr . @FWHM

GM_311-1 MW=2610?
(H₂O)/MeOH + DEA

EPSRC National Centre Swansea
LTQ Orbitrap XL

Gavin Miller
29/01/2013 11:19:47



NL:
7.22E5
MANGAR245-OV-HNESN#6-
10 RT: 0.33-0.44 AV: 5 T:
FTMS - p NSI Full ms
[120.00-2000.00]

NL:
6.01E3
C₇₆ H₁₁₈ N₆ O₈₁ S₆:
C₇₆ H₁₁₈ N₆ O₈₁ S₆:
p (gss, s /p:40) Chrg -6
R: 100000 Res .Pwr . @FWHM

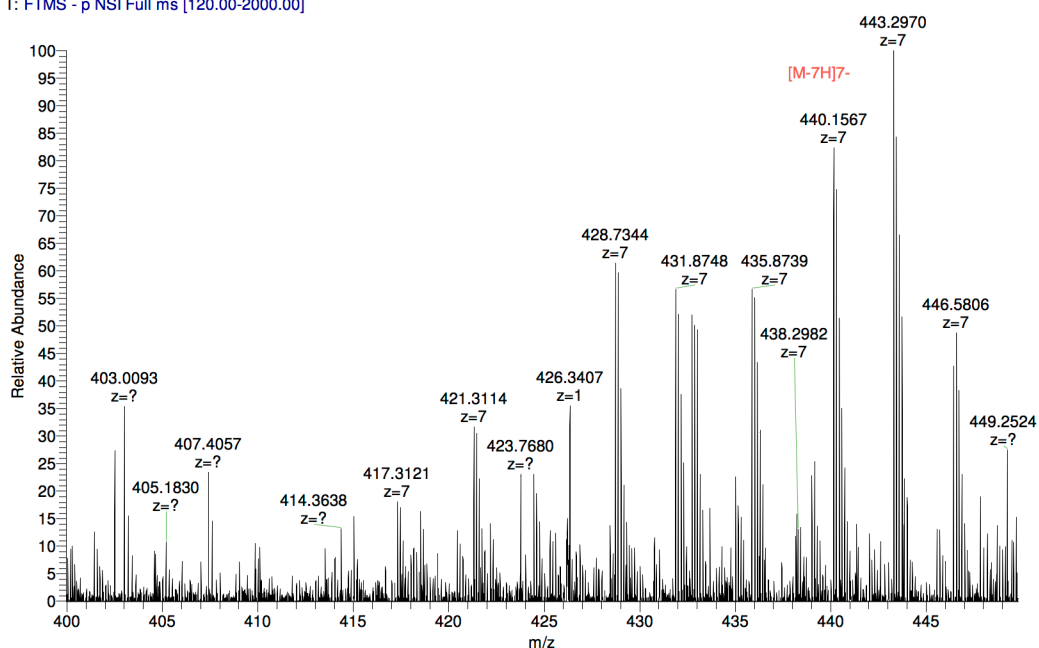
Dodecasaccharide N-sulfation- ESI MS

GM_329-3 MW=3090?
(H₂O)/MeOH + DEA

EPSRC National Facility Swansea
LTQ Orbitrap XL

Gavin Miller
16/05/2013 10:19:11

MANGAR283-OV-HNESN #1-7 RT: 0.20-0.68 AV: 7 SM: 7G NL: 1.65E5
T: FTMS - p NSI Full ms [120.00-2000.00]

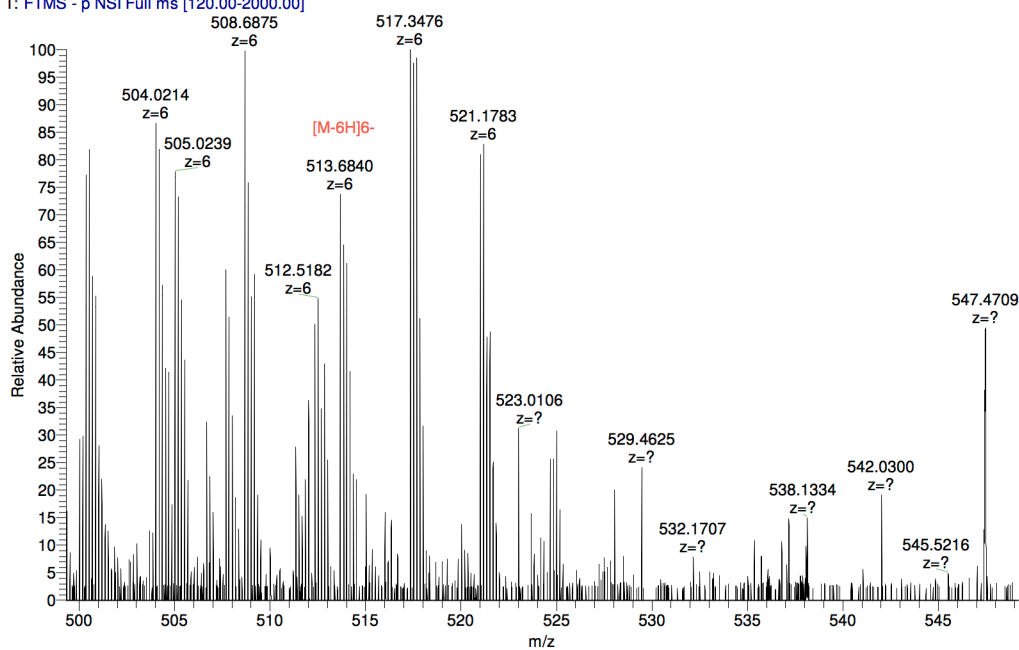


GM_329-3 MW=3090?
(H₂O)/MeOH + DEA

EPSRC National Facility Swansea
LTQ Orbitrap XL

Gavin Miller
16/05/2013 10:19:11

MANGAR283-OV-HNESN #1-7 RT: 0.20-0.68 AV: 7 SM: 7G NL: 4.41E4
T: FTMS - p NSI Full ms [120.00-2000.00]



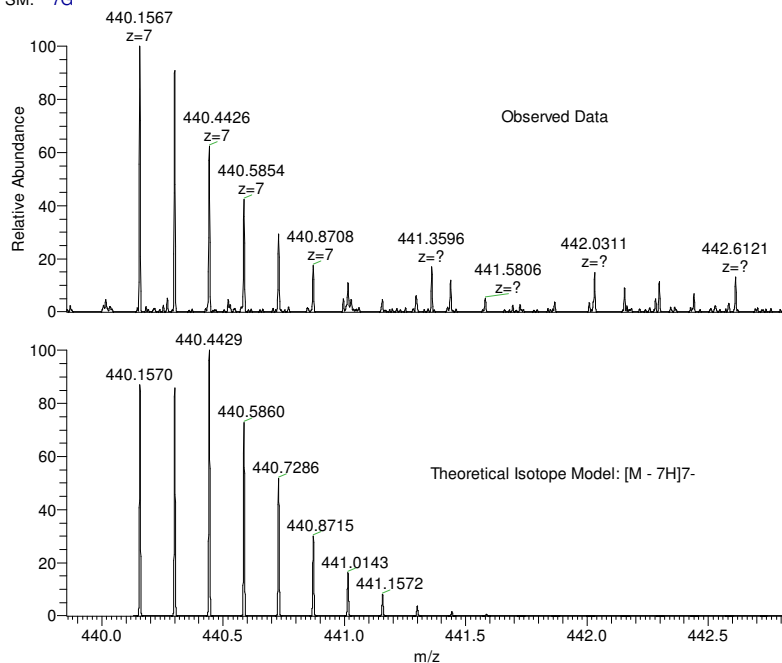
Dodecasaccharide N-sulfation- ESI MS isotope patterns

GM_329-3 MW=3090?
(H₂O)/MeOH + DEA

EPSRC National Facility Swansea
LTQ Orbitrap XL

Gavin Miller
16/05/2013 10:19:11

SM: 7G



NL:
1.36E5
MANGAR283-OV-HNESN#1-7
RT: 0.20-0.68 AV: 7 T: FTMS -
p NSI Full ms [120.00-2000.00]

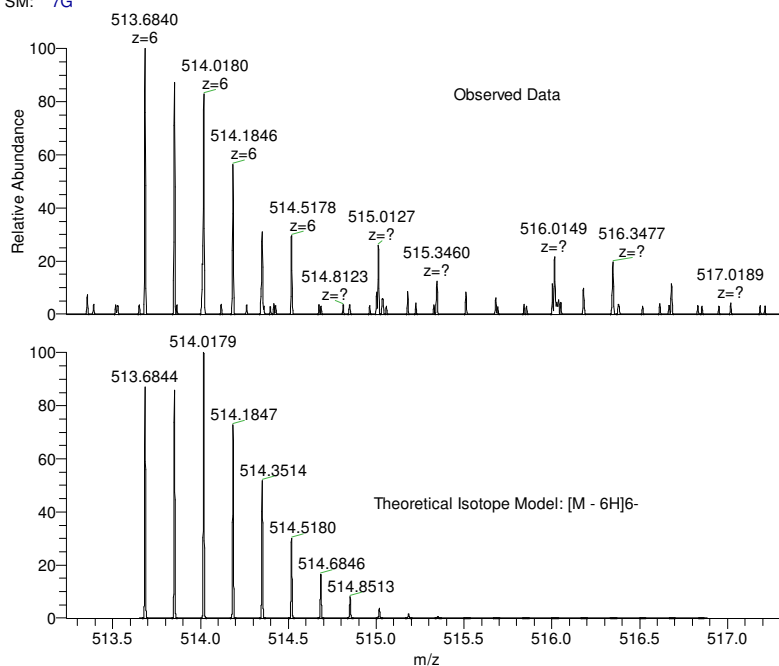
NL:
4.83E3
C₇₆ H₁₁₇ N₆ O₉₉ S₁₂:
C₇₆ H₁₁₇ N₆ O₉₉ S₁₂:
p (gss, s/p:40) Chrg -7
R: 100000 Res .Pwr . @FWHM

GM_329-3 MW=3090?
(H₂O)/MeOH + DEA

EPSRC National Facility Swansea
LTQ Orbitrap XL

Gavin Miller
16/05/2013 10:19:11

SM: 7G

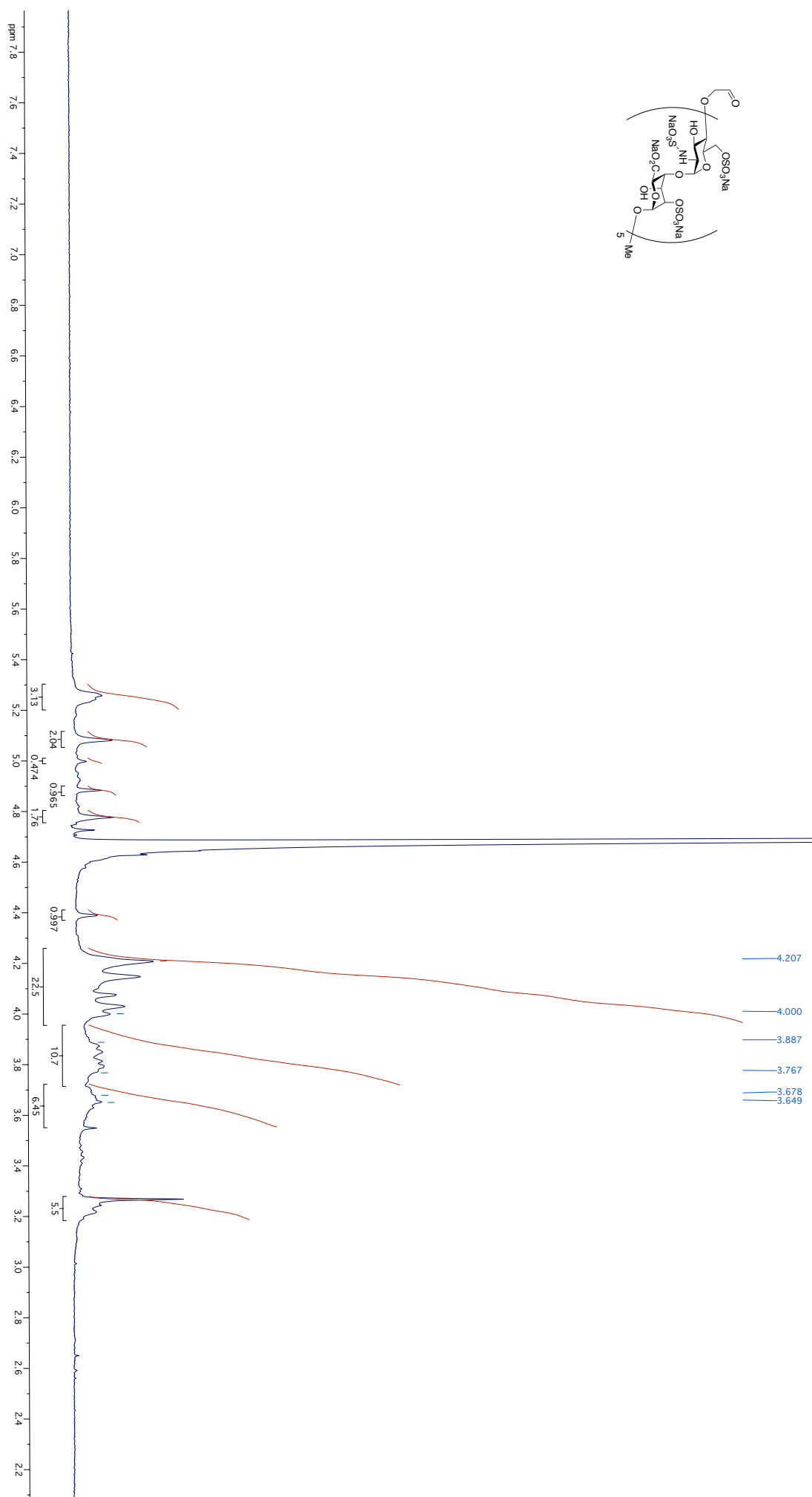


NL:
3.25E4
MANGAR283-OV-HNESN#1-7
RT: 0.20-0.68 AV: 7 T: FTMS -
p NSI Full ms [120.00-2000.00]

NL:
4.83E3
C₇₆ H₁₁₈ N₆ O₉₉ S₁₂:
C₇₆ H₁₁₈ N₆ O₉₉ S₁₂:
p (gss, s/p:40) Chrg -6
R: 100000 Res .Pwr . @FWHM

Decasaccharide periodate cleavage product 27

¹H NMR



400 MHz, D₂O

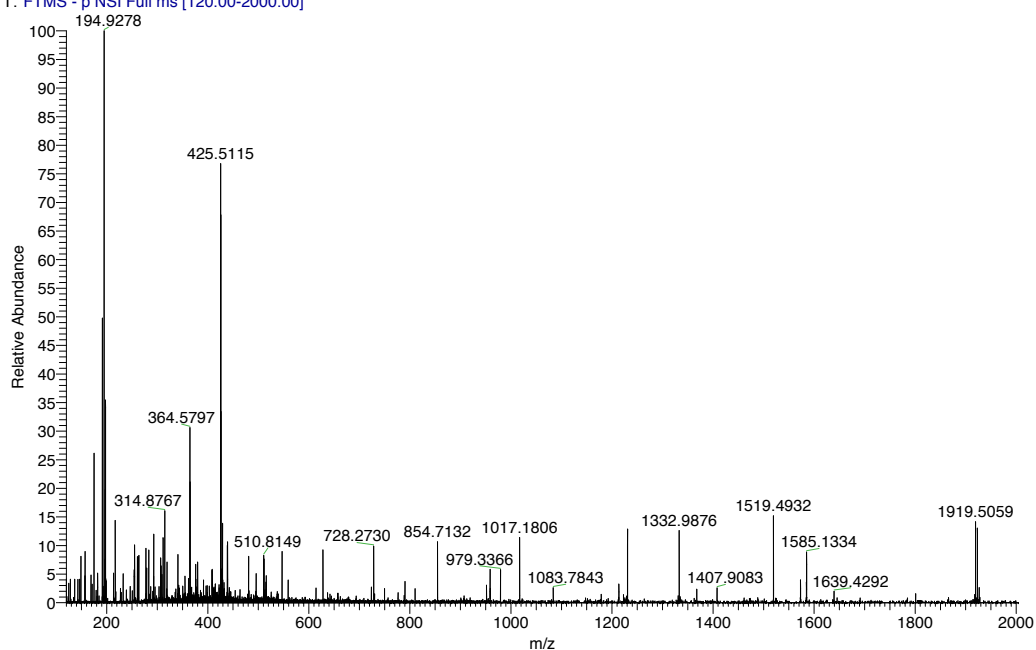
Decasaccharide periodate cleavage product 27- ESI MS

GM_260 MW=3302?
(H₂O/MeCN)/H₂O/MeCN +DEA

EPSRC National Centre Swansea
LTQ Orbitrap XL

Gravin Miller
10/02/2012 15:59:21

MANGAR131-OM-HNESN #5-10 RT: 0.30-0.45 AV: 6 SM: 7G NL: 3.37E5
T: FTMS - p NSI Full ms [120.00-2000.00]

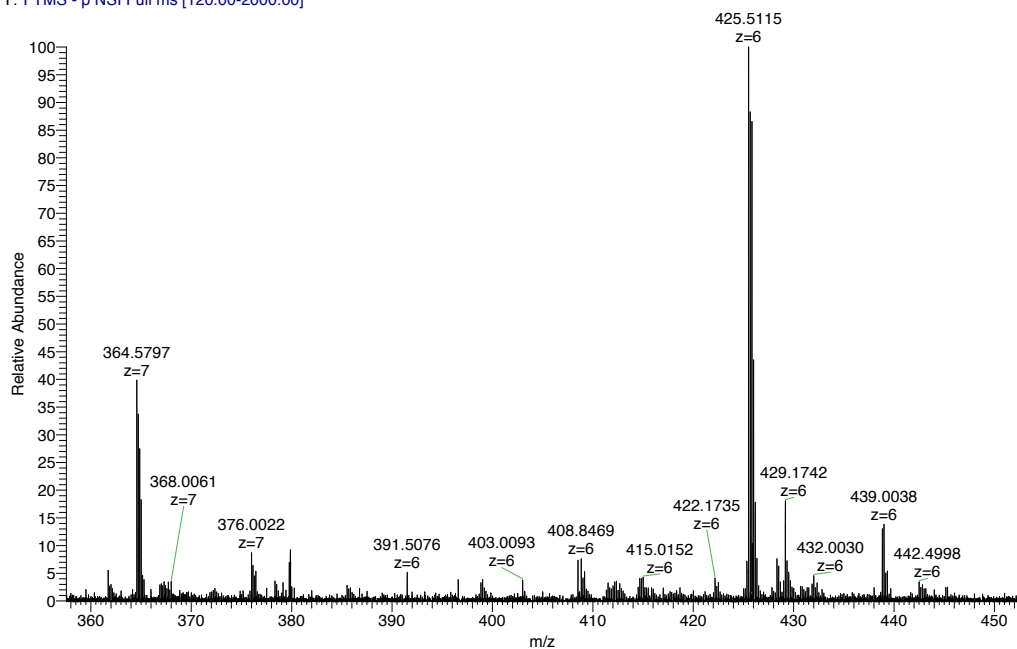


GM_260 MW=3302?
(H₂O/MeCN)/H₂O/MeCN +DEA

EPSRC National Centre Swansea
LTQ Orbitrap XL

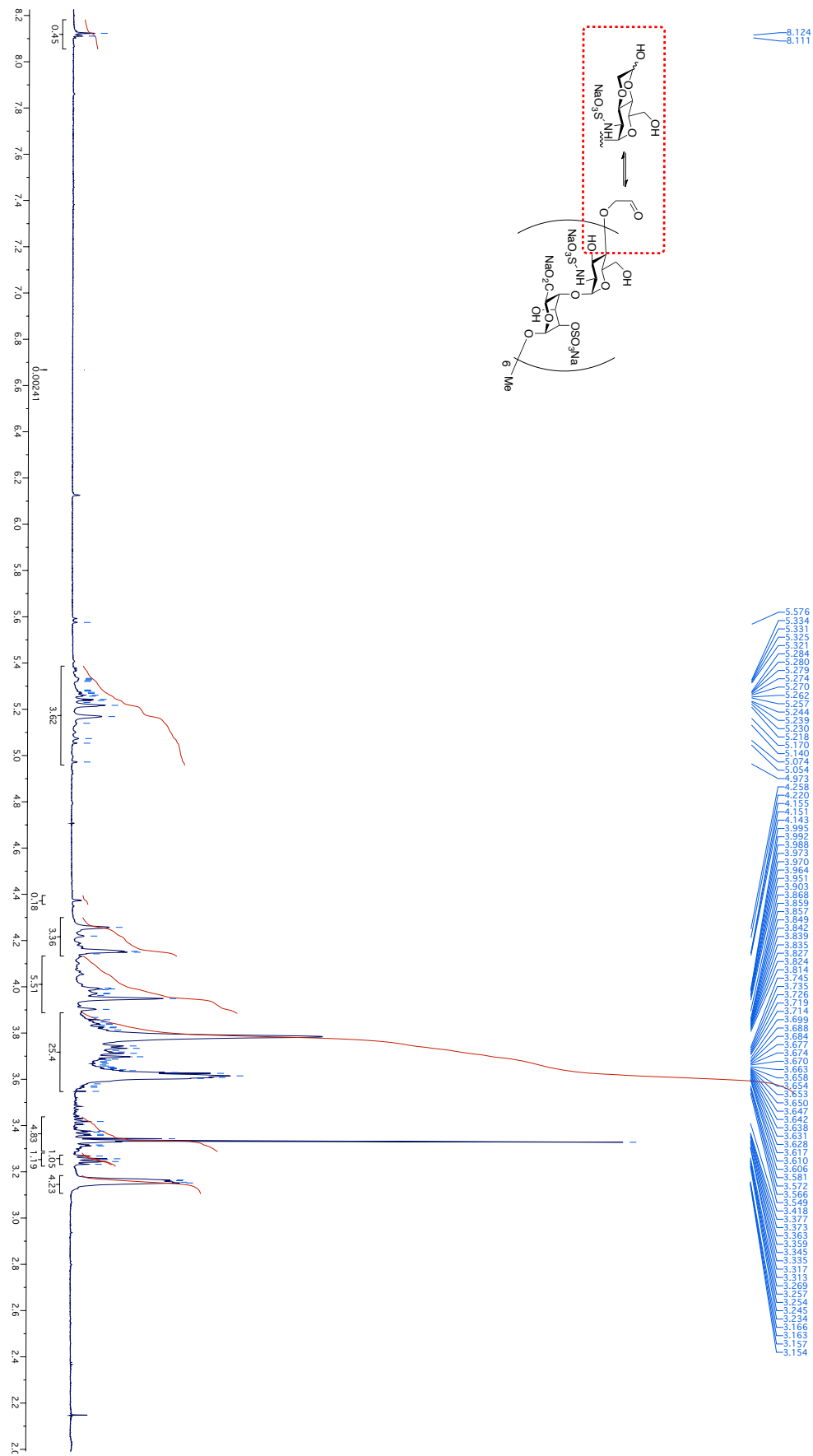
Gravin Miller
10/02/2012 15:59:21

MANGAR131-OM-HNESN #5-10 RT: 0.30-0.45 AV: 6 SM: 7G NL: 2.59E5
T: FTMS - p NSI Full ms [120.00-2000.00]



Dodecasaccharide periodate cleavage product 26

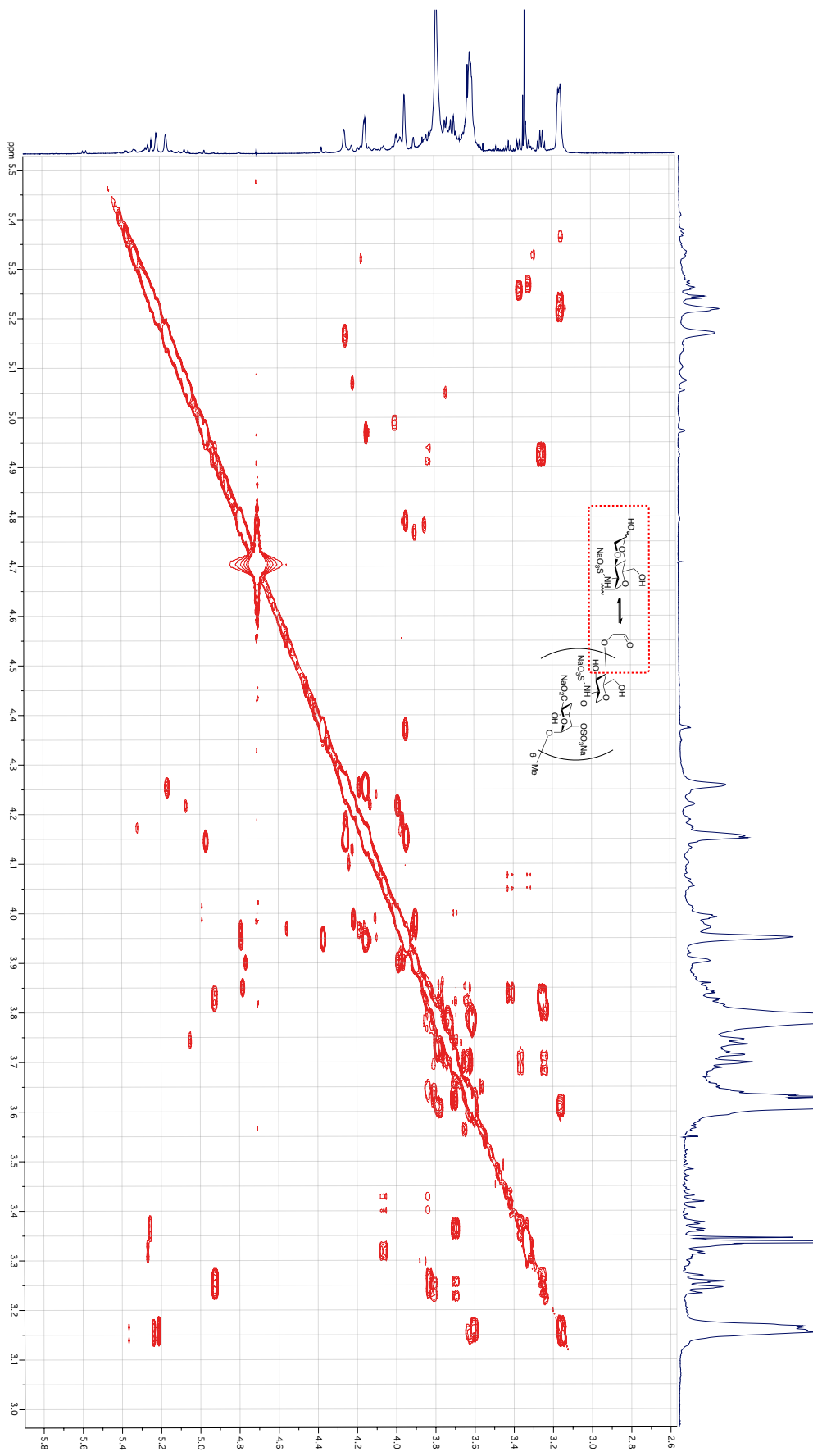
¹H NMR



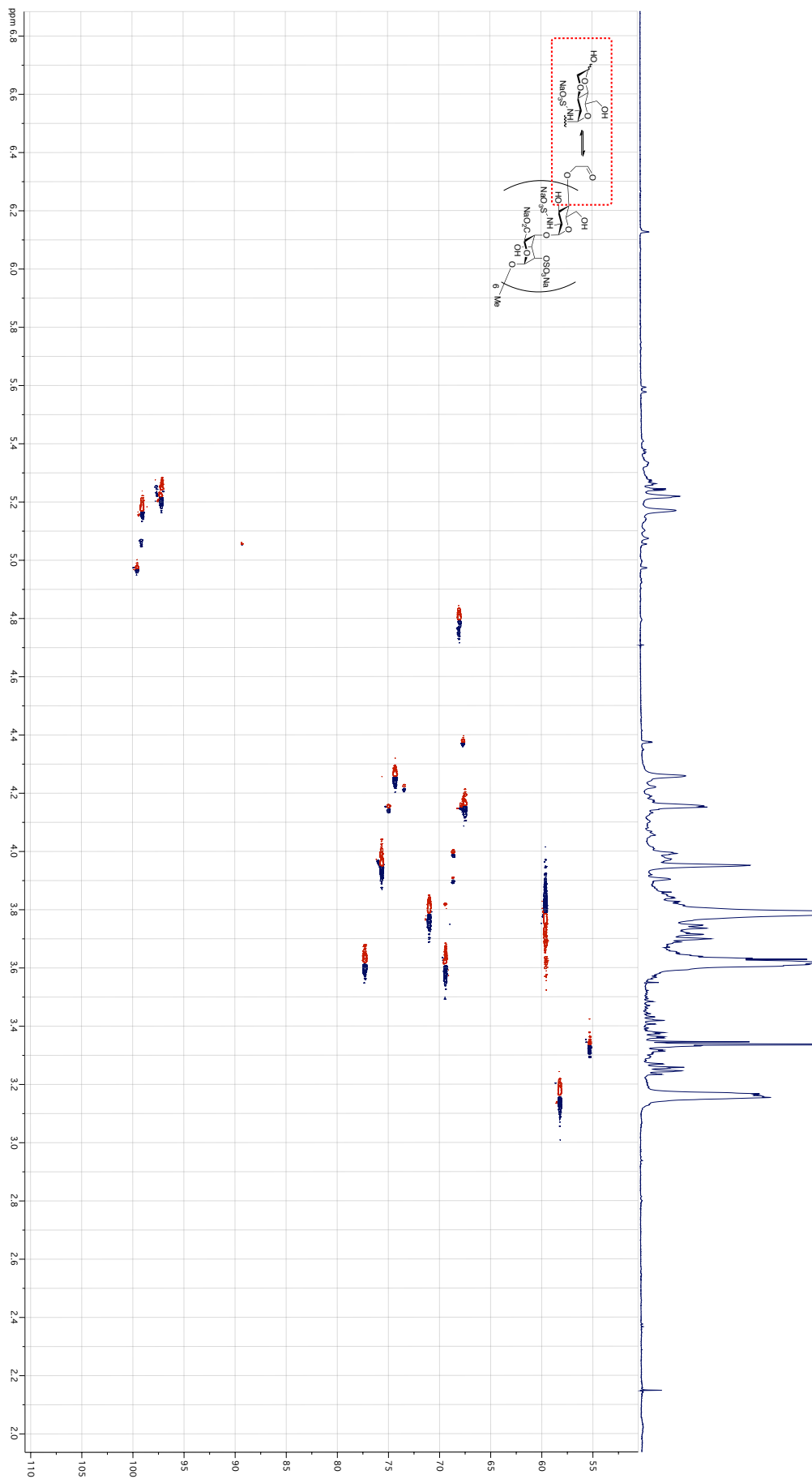
800 MHz, CDCl₃

Dodecasaccharide periodate cleavage product 26

COSY



800 MHz, D₂O



800 MHz, D₂O

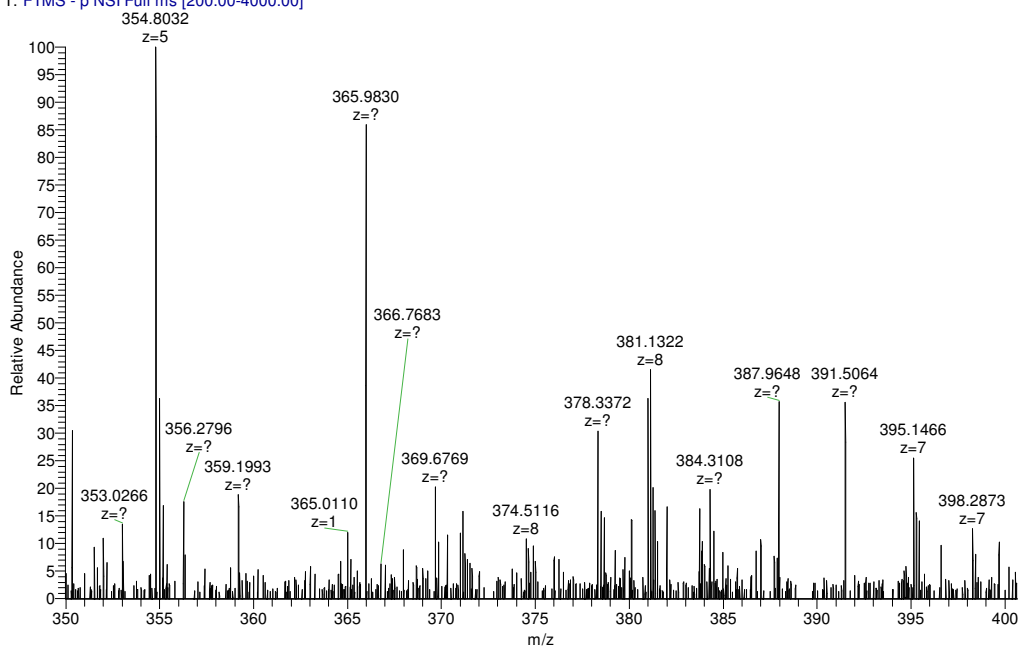
Dodecasaccharide periodate cleavage product 26- ESI MS

GM_340-1.1 MW=3058?
(H₂O)/MeOH + DEA

EPSRC National Centre Swansea
LTQ Orbitrap XL

Gravin Miller
04/04/2013 10:43:31

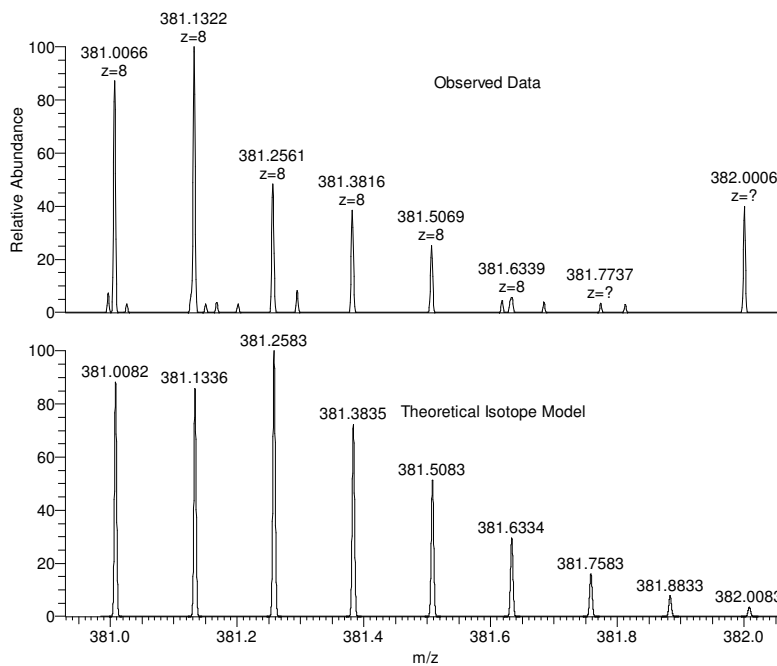
MANGAR278-OV-HNESN #1-6 RT: 0.21-0.35 AV: 3 SM: 7G NL: 8.03E4
T: FTMS - p NSI Full ms [200.00-4000.00]



GM_340-1.1 MW=3058?
(H₂O)/MeOH + DEA

EPSRC National Centre Swansea
LTQ Orbitrap XL

Gravin Miller
04/04/2013 10:43:31



NL:
3.34E4
MANGAR278-OV-HNESN#1-6
RT: 0.21-0.35 AV: 3 T: FTMS -
p NSI Full ms [200.00-4000.00]

NL:
4.84E3
C₇₅ H₁₁₂ N₆ O₉₈ S₁₂:
C₇₅ H₁₁₂ N₆ O₉₈ S₁₂
p (gss, s/p:40) Chrg -8
R: 100000 Res .Pwr . @FWHM