

Electronic Supplementary Material (ESI) for Organic & Biomolecular Chemistry.
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Supporting Information:

One pot, oxidative N-S bond formation to access 2-Sulfenylimine Chromenes

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Content

Spectral data for the known compounds S1-S2.

¹H-NMR and ¹³C-NMR spectra for all compounds S3-S20.

Crystallographic data for 8c

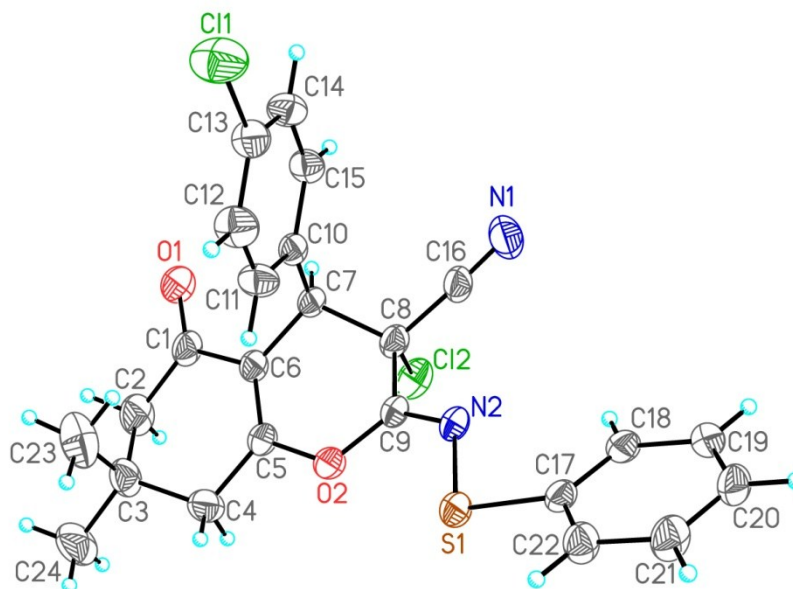


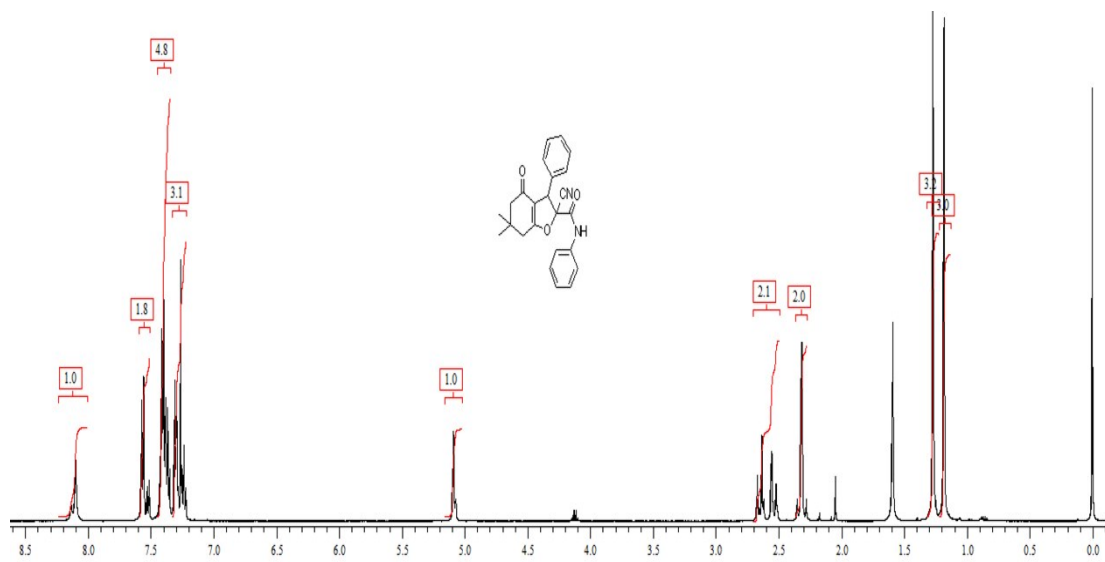
Figure caption: ORTEP diagram of **8c** with the atom-numbering. Displacement thermal ellipsoids are drawn at the 30% probability level and H atoms are shown as small spheres of arbitrary radius. There are two molecules of AY75 compound in the asymmetric unit ($Z'=2$), however only one is represented in the figure for clarity.

Crystal data for 8c: C₂₄H₂₀N₂O₂Cl₂S, $M = 471.39$, colorless diamond shaped crystal, 0.43 x 0.22 x 0.20 mm³, monoclinic, space group $P2_1/c$ (No. 14), $a = 16.6008(8)$, $b = 13.5612(7)$, $c = 21.8151(11)$ Å, $\alpha = 90$, $\beta = 106.926(1)$, $\gamma = 90^\circ$, $V = 4698.4(4)$ Å³, $Z = 8$, $D_c = 1.333$ g/cm³, $F_{000} = 1952$, CCD area detector, MoK α radiation, $\lambda = 0.71073$ Å, $T = 293(2)$ K, $2\theta_{\max} = 50.6^\circ$, 45900 reflections collected, 8595 unique ($R_{\text{int}} = 0.042$), Final $Goof = 1.13$, $RI = 0.0633$, $wR2 = 0.1452$, R indices based on 6645 reflections with $I > 2\sigma(I)$ (refinement on F^2), 563 parameters, $\mu = 0.388$ mm⁻¹, minimum and maximum residual density = -0.23 and 0.47 e/Å³, respectively. **CCDC 1415312** contains the supplementary crystallographic data for this paper. The data can be obtained free of charge from The Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/data_request/cif.

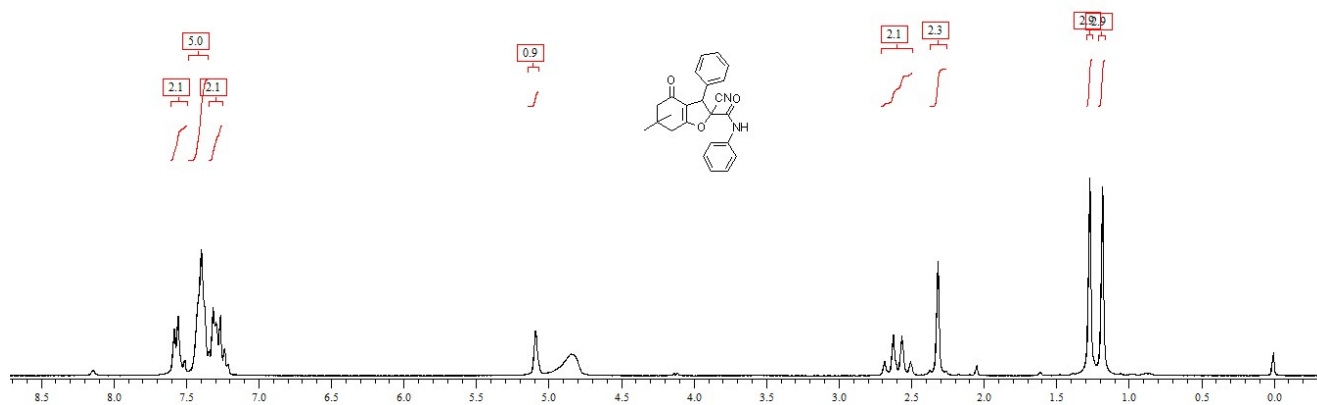
Data collection and Structure solution: X-ray data for **8c** compound were collected at room temperature using the Bruker Smart Apex CCD diffractometer with graphite monochromated MoK α radiation ($\lambda = 0.71073$ Å) with ω -scan method.¹ Preliminary lattice parameters and orientation matrices were obtained from four sets of frames. Unit cell dimensions were determined using 8332 reflections. Integration and scaling of intensity data were accomplished using SAINT program.¹ The structure was solved by Direct Methods using SHELXS97² and refinement was carried out by full-matrix least-squares technique using SHELXL97.² Anisotropic displacement parameters were included for all non-hydrogen atoms. All H atoms were positioned geometrically and treated as riding on their parent C atoms, with C-H distances of 0.93--0.97 Å, and with $U_{\text{iso}}(\text{H}) = 1.2U_{\text{eq}}(\text{C})$ or $1.5U_{\text{eq}}$ for methyl atoms.

1. SMART & SAINT. Software Reference manuals. Versions 6.28a & 5.625, Bruker Analytical X-ray Systems Inc., Madison, Wisconsin, U.S.A., 2001.
2. Sheldrick, G. M. SHELXS97 and SHELXL97, Programs for crystal structure solution and refinement; University of Gottingen: Germany, 1997.

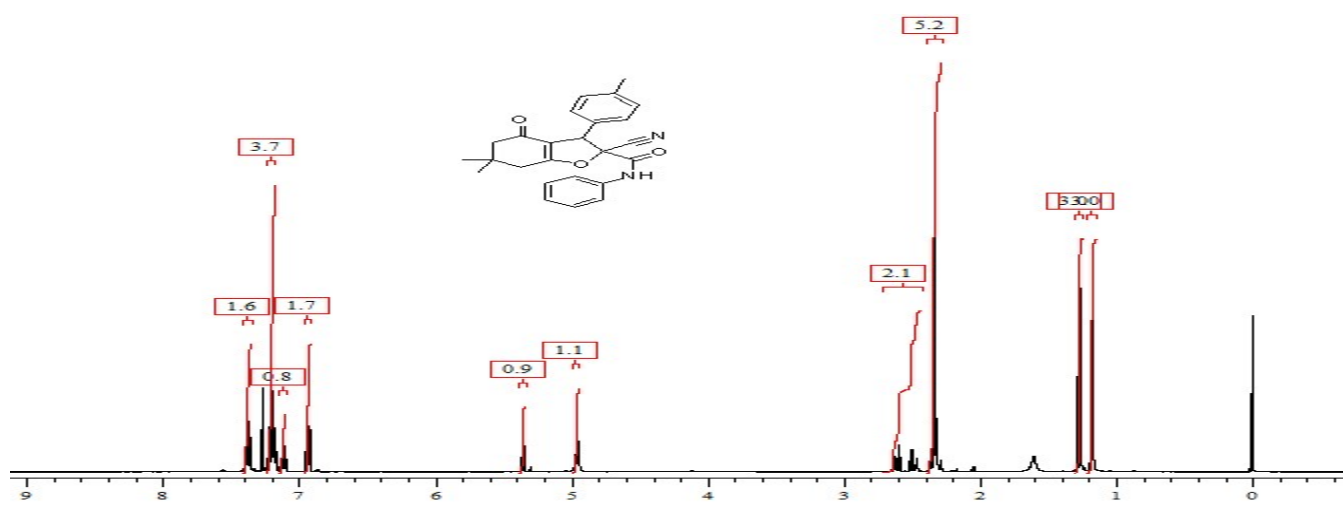
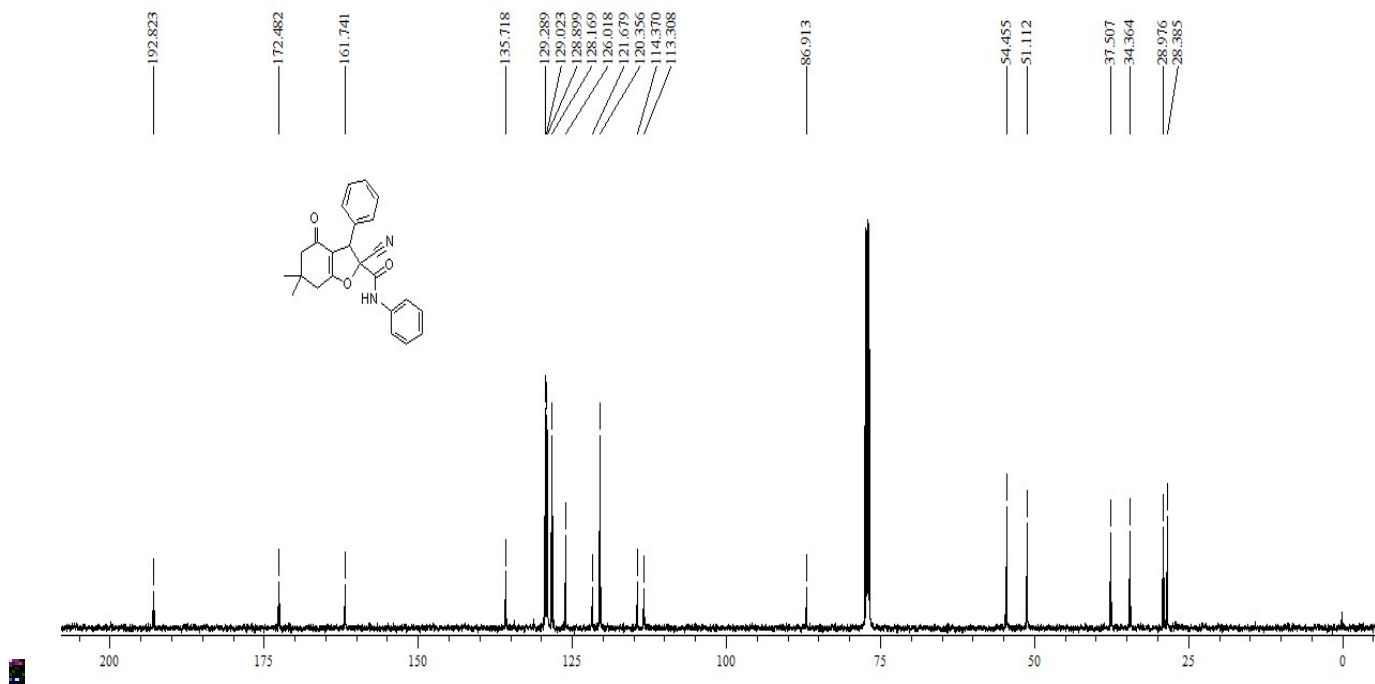
^1H and ^{13}C NMR spectra of 5 (a-l):

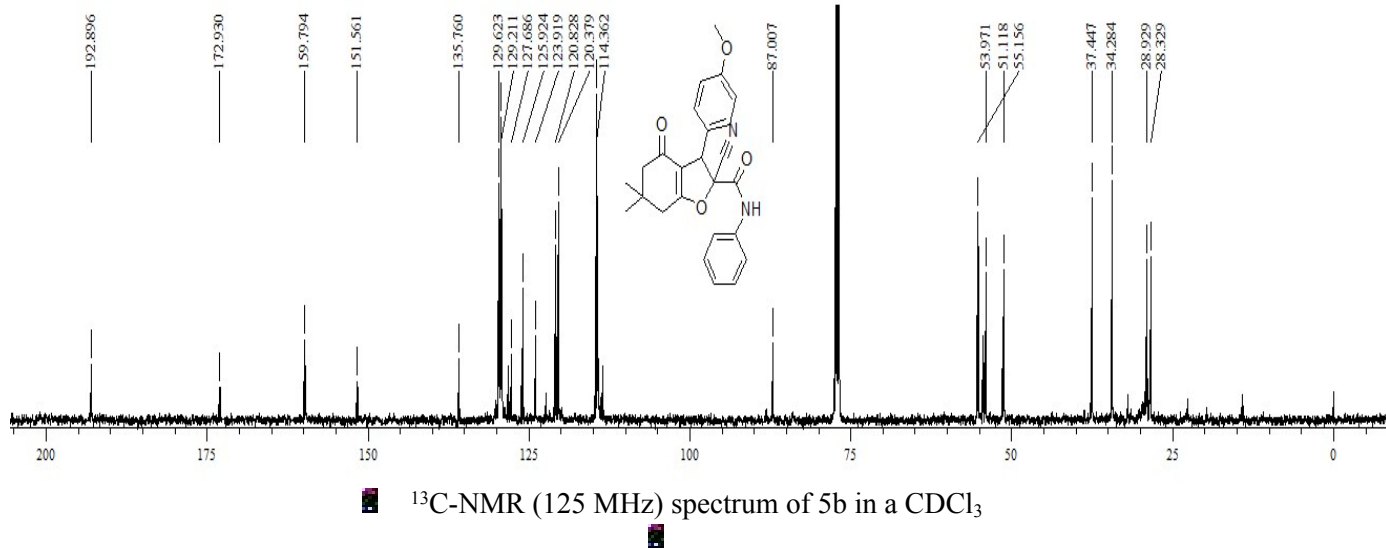
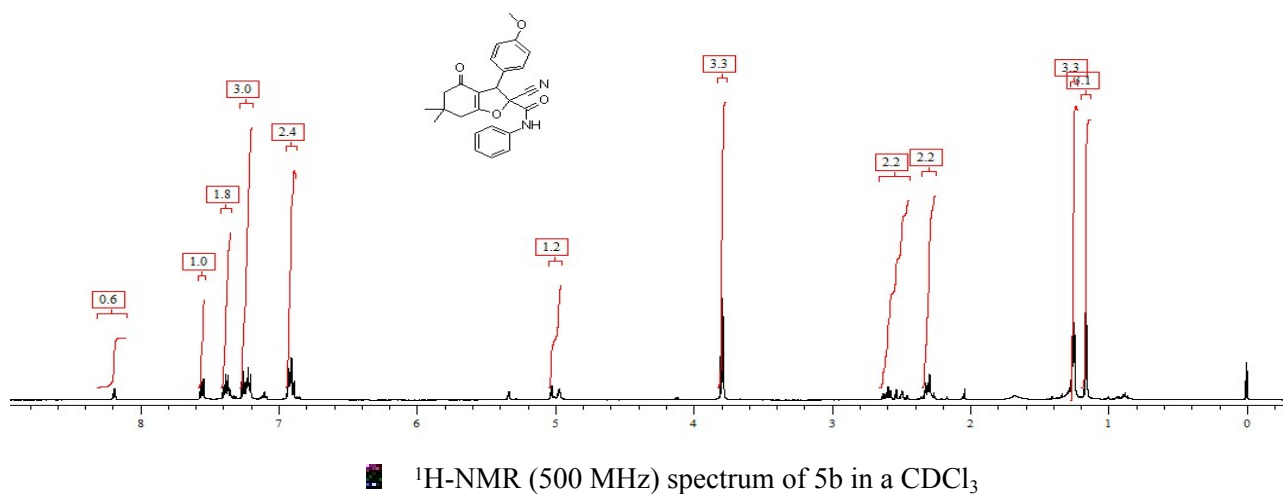
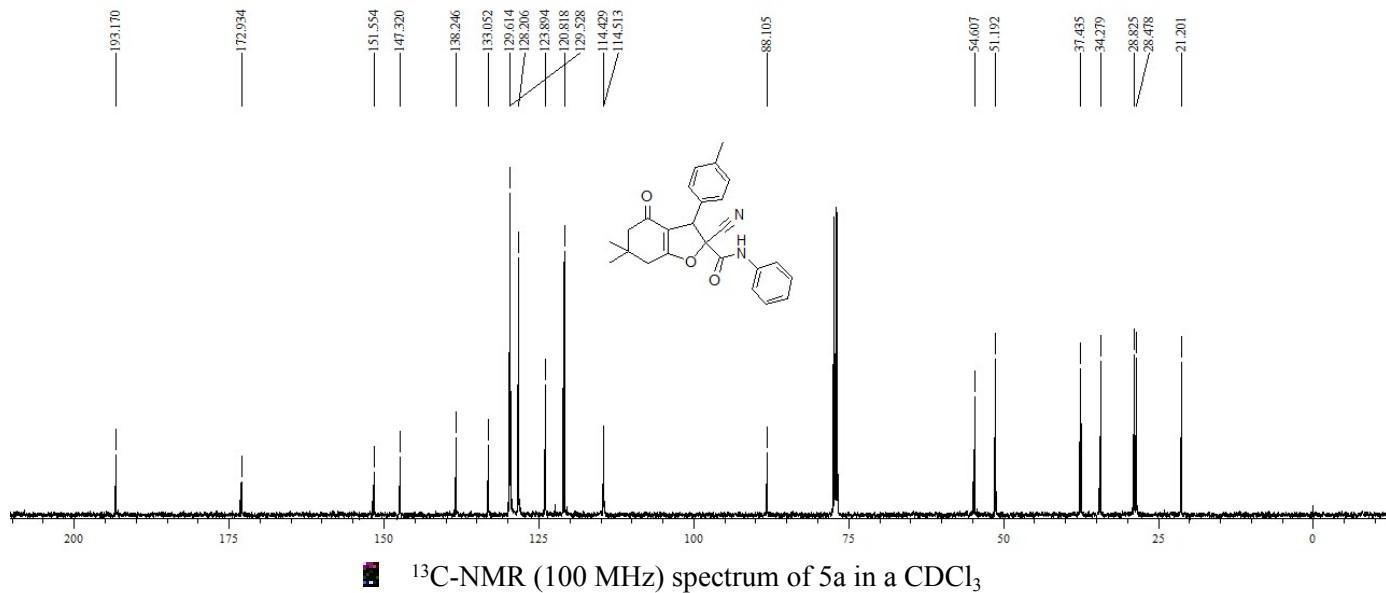


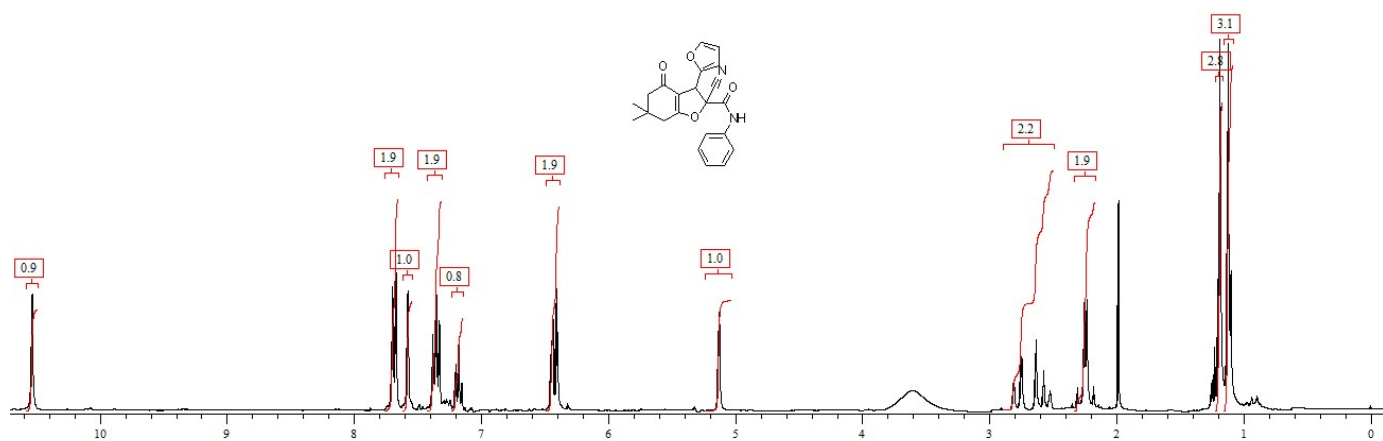
^1H -NMR (500 MHz) spectrum of 5 in a CDCl_3



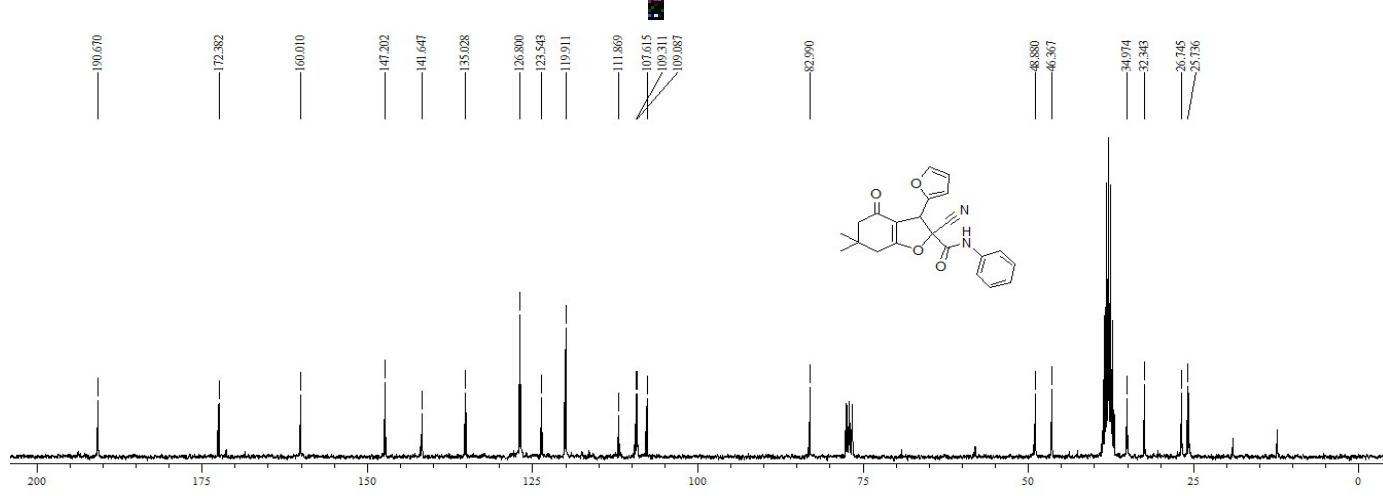
D_2O exchanged ^1H -NMR (300 MHz) spectrum of 5 in a CDCl_3



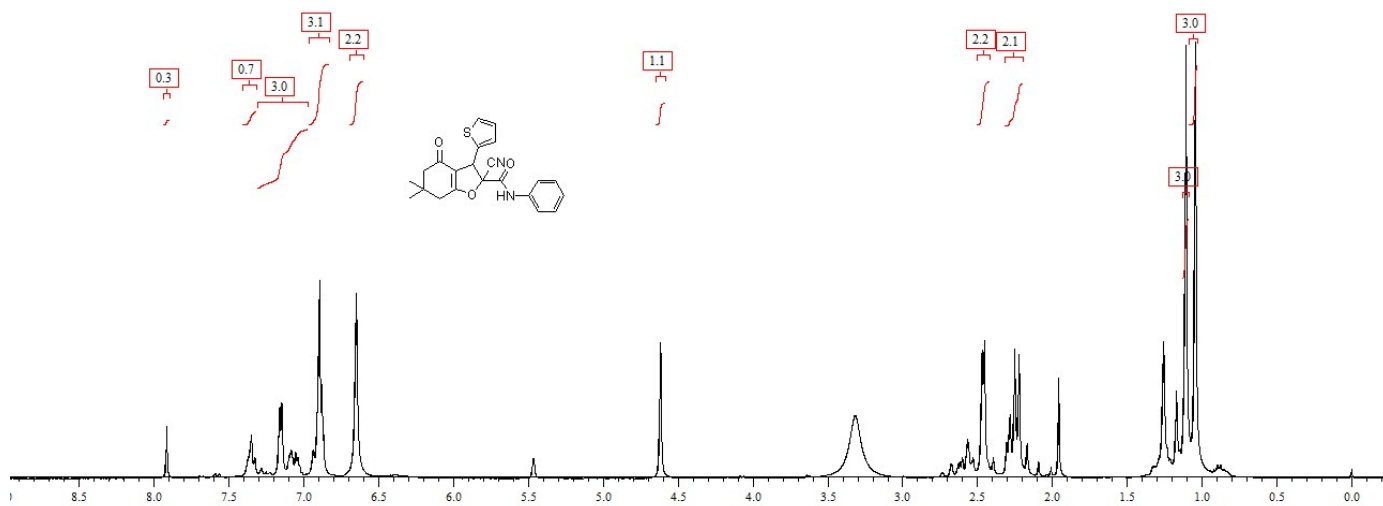




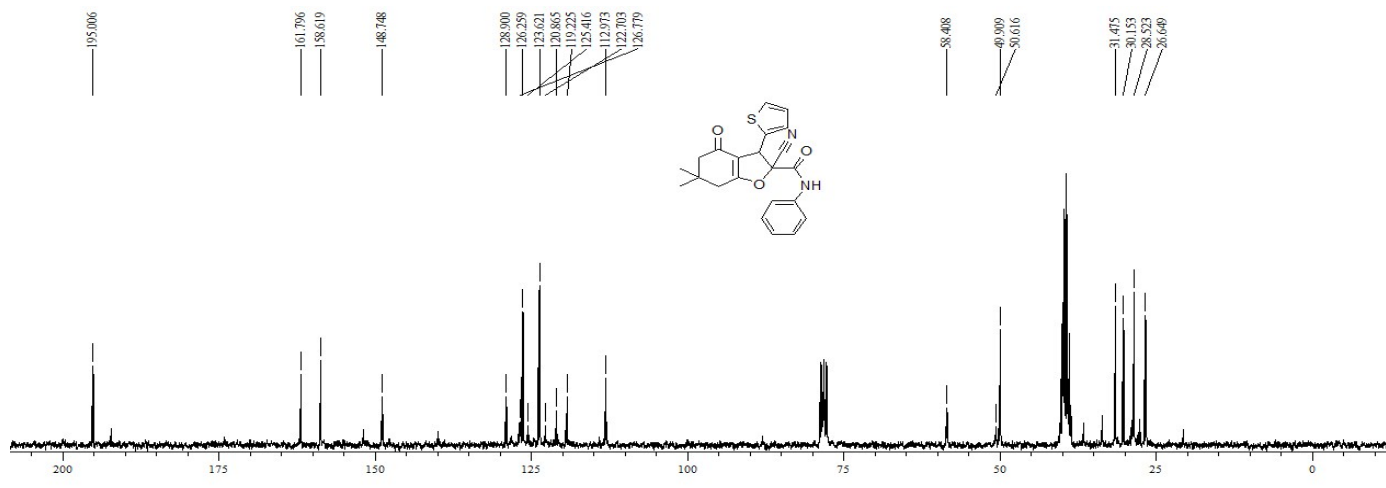
¹H-NMR (500 MHz) spectrum of 5c in a CDCl₃+DMSO-*d*₆



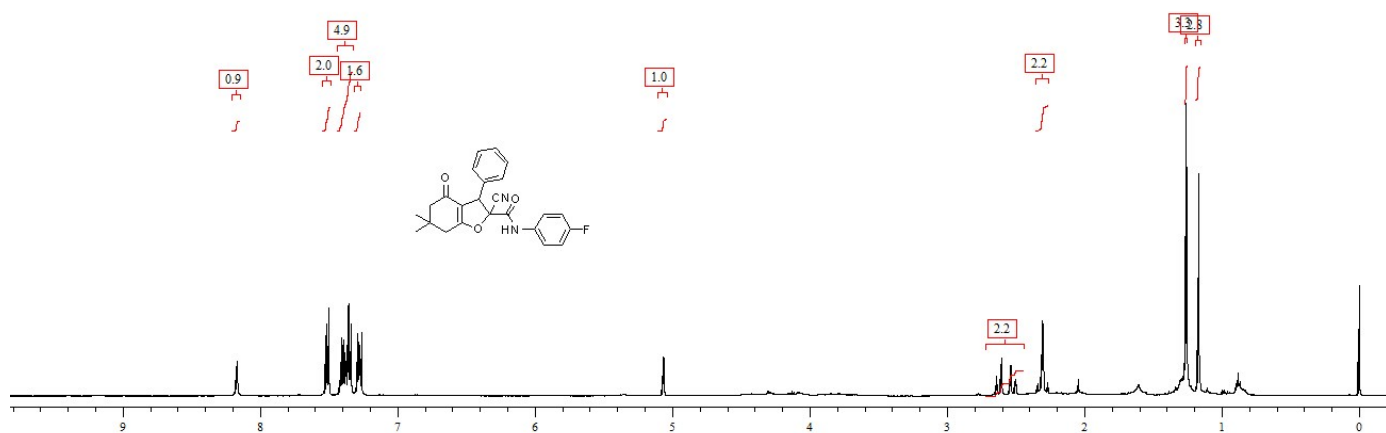
¹³C-NMR (75 MHz) spectrum of 5c in a CDCl₃+DMSO-*d*₆



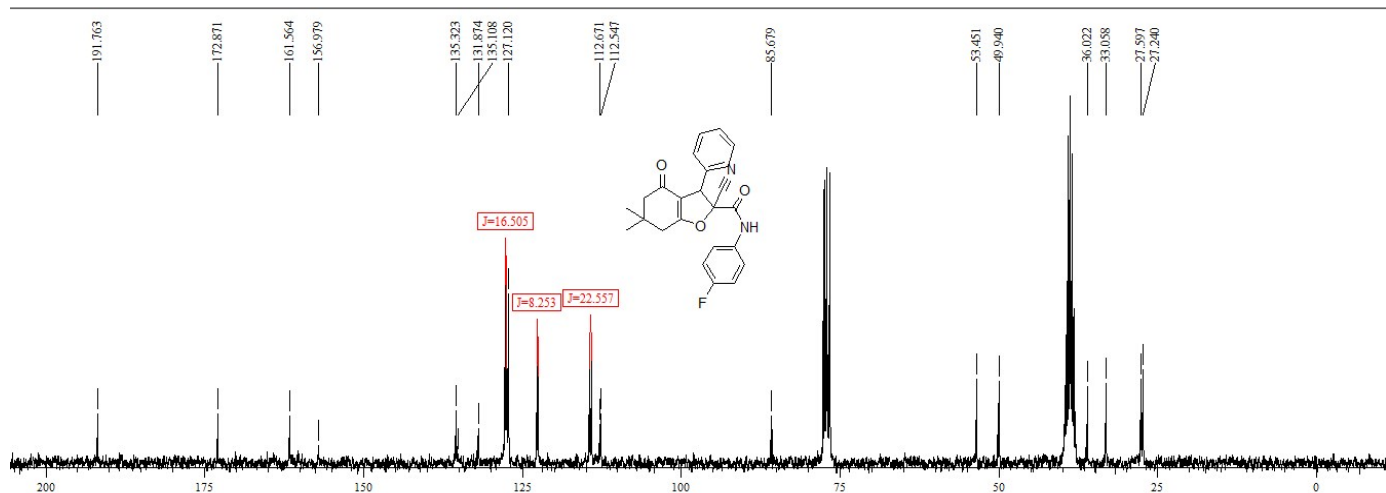
¹H-NMR (300 MHz) spectrum of 5d in a CDCl₃+DMSO-*d*₆



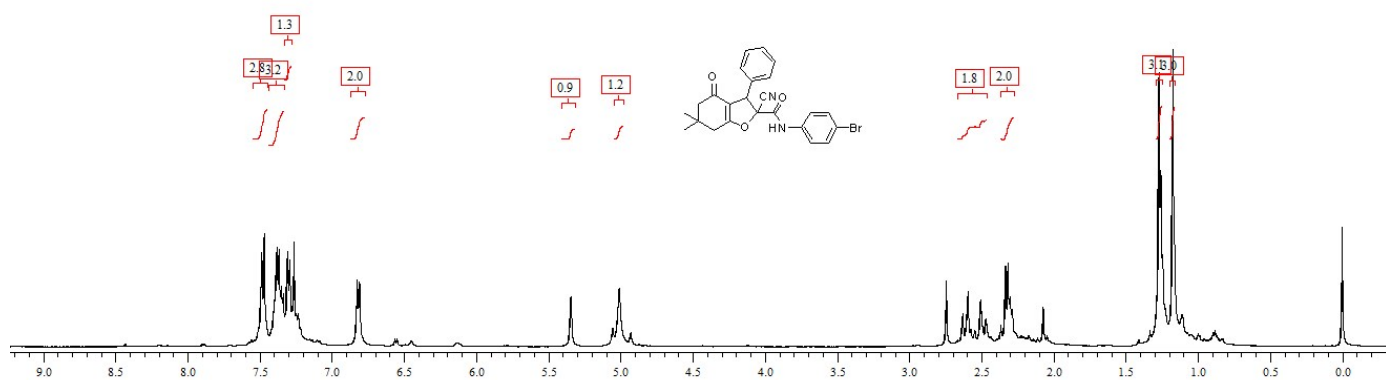
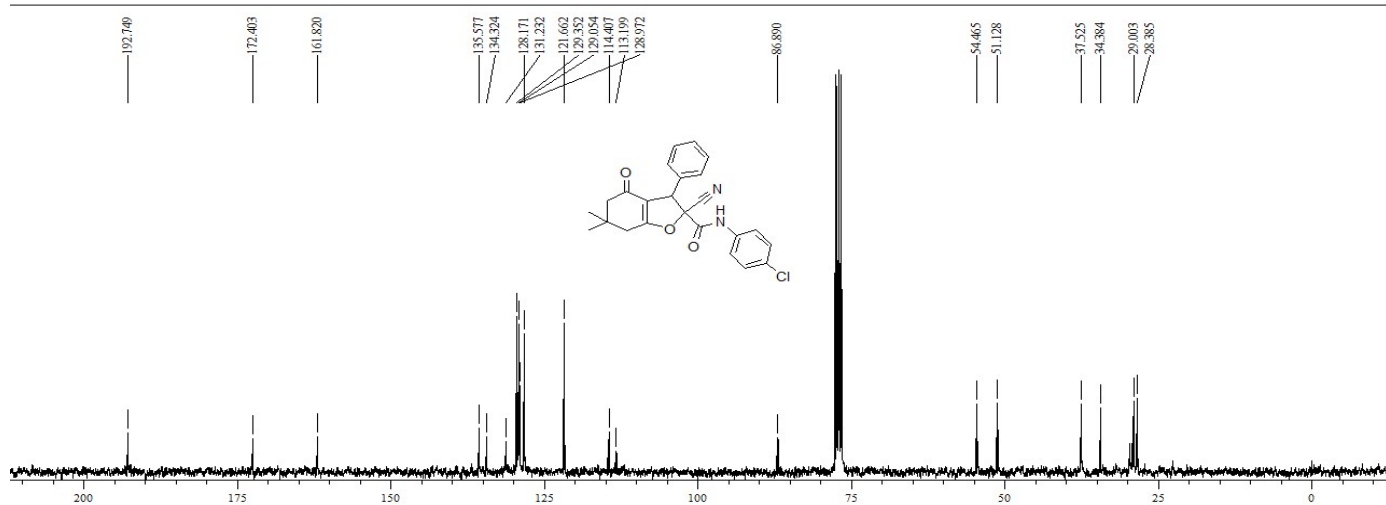
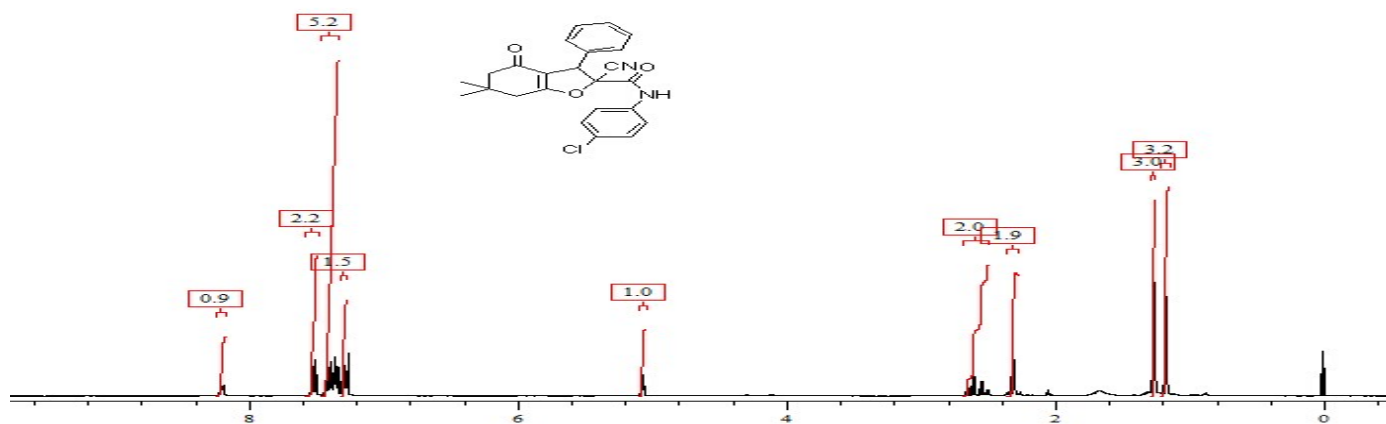
$^{13}\text{C-NMR}$ (75 MHz) spectrum of 5d in a $\text{CDCl}_3+\text{DMSO-}d_6$

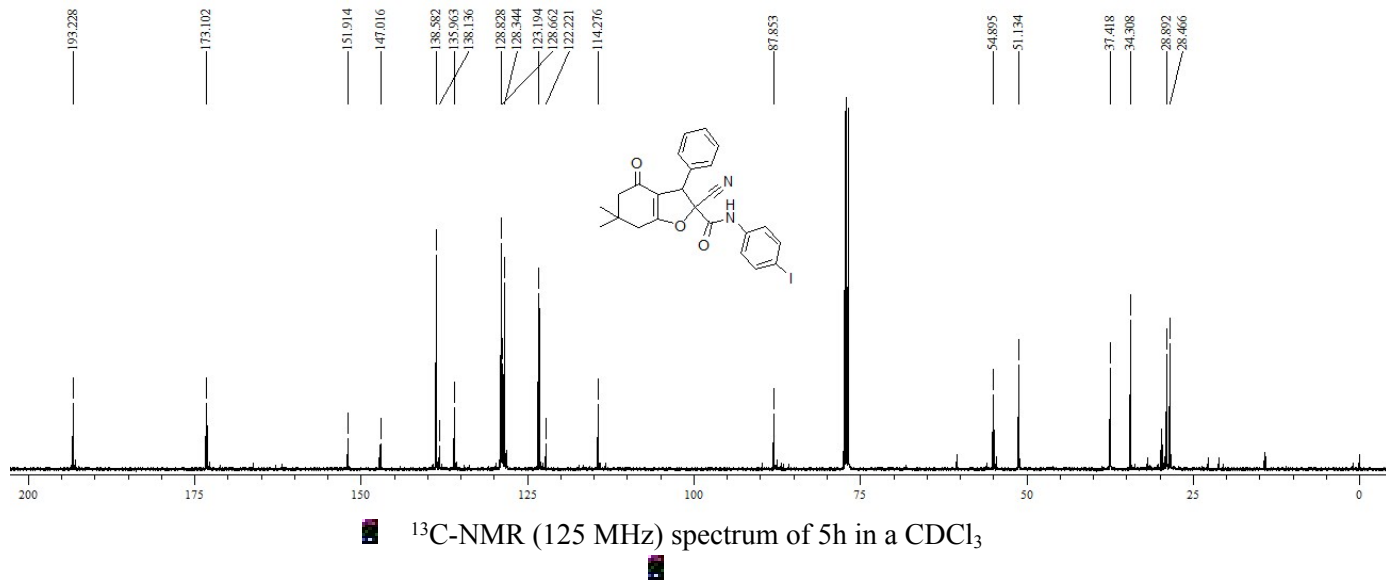
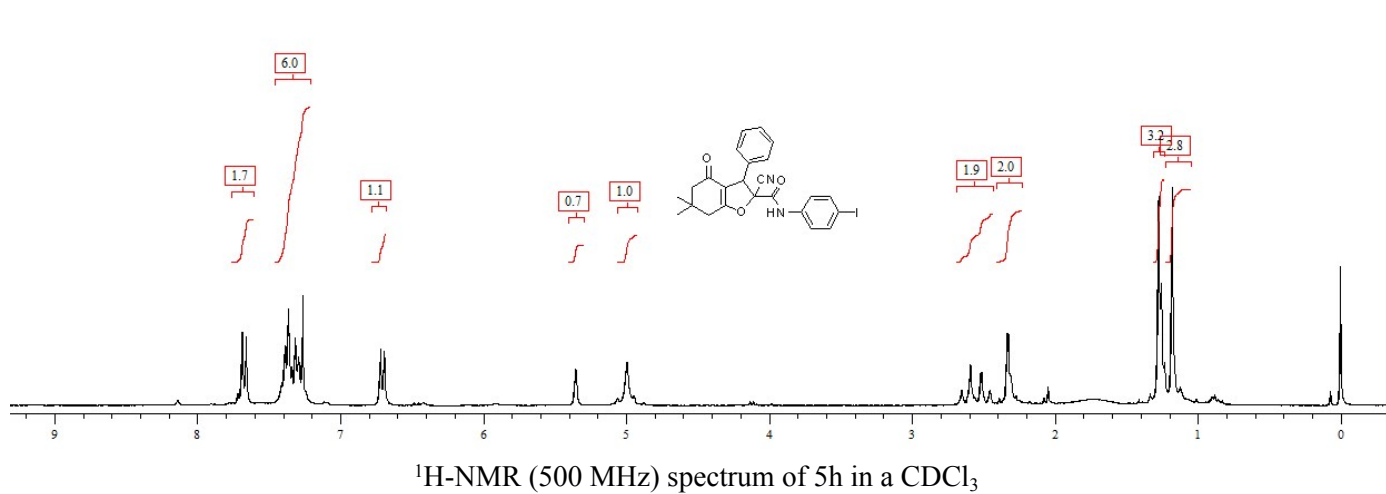
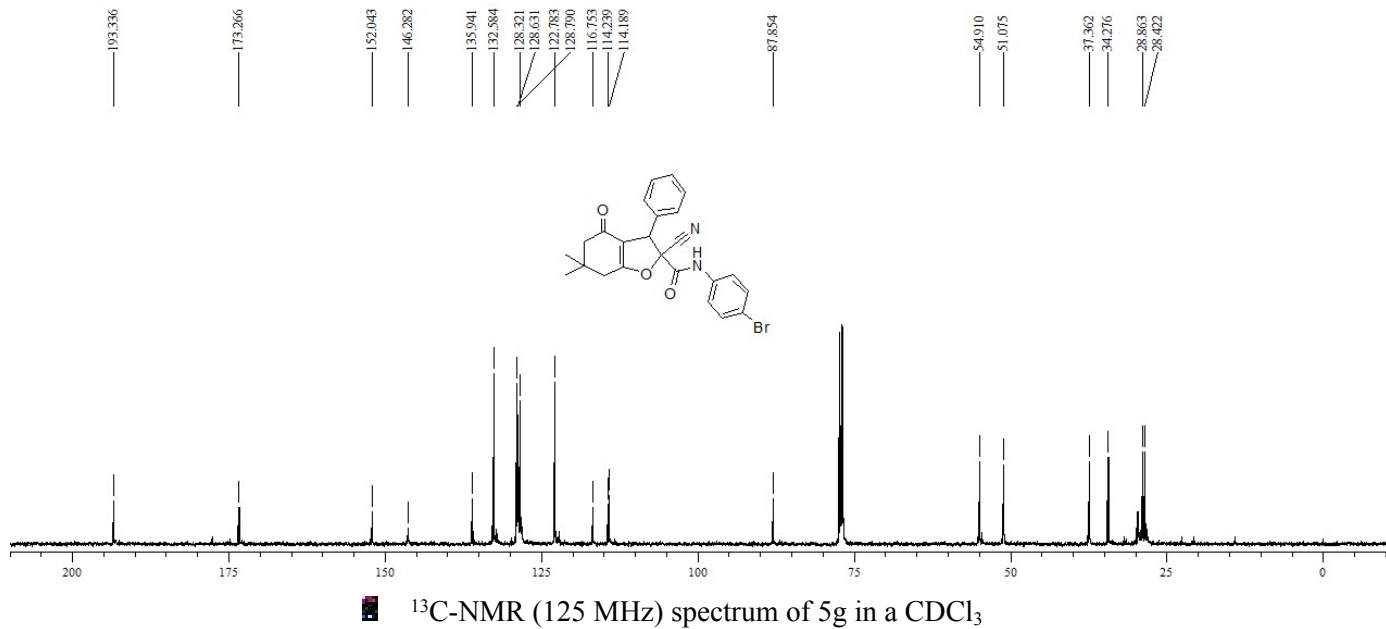


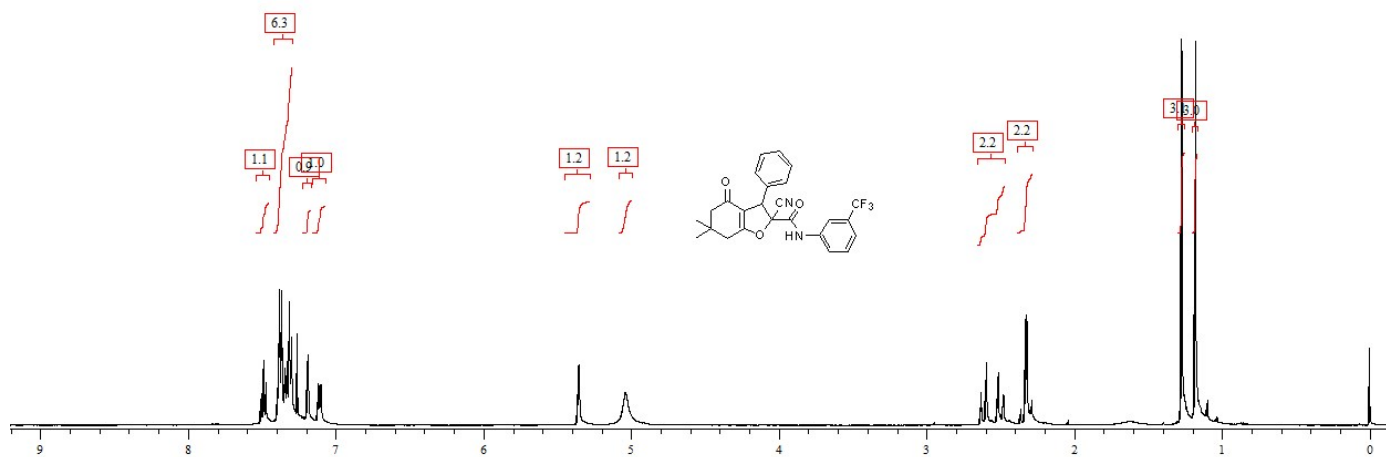
$^1\text{H-NMR}$ (500 MHz) spectrum of 5e in a CDCl_3



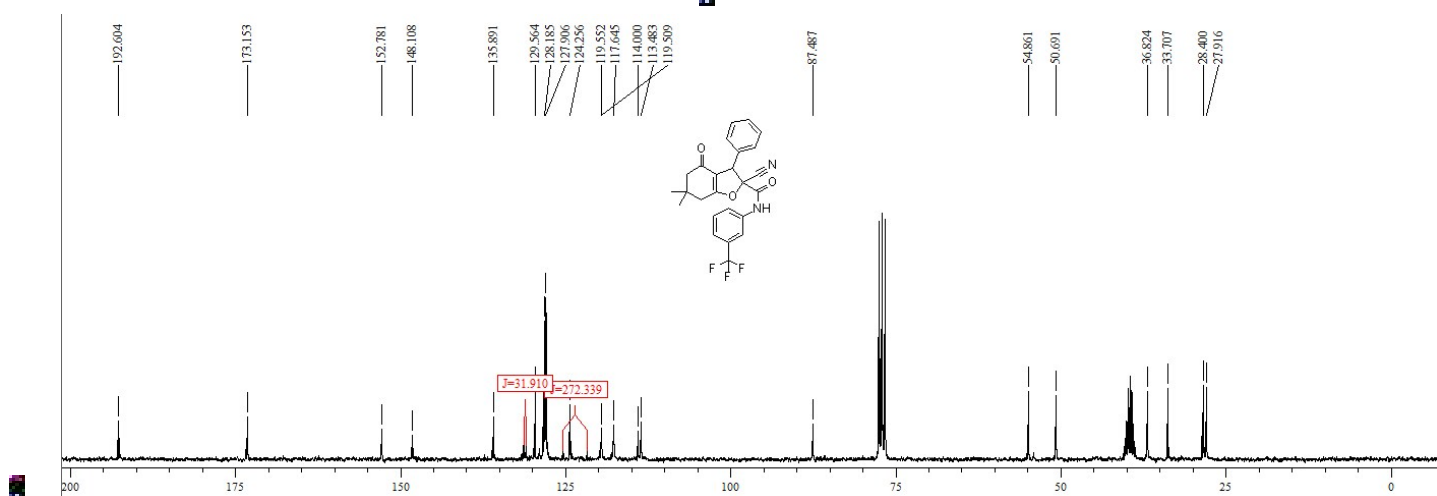
$^{13}\text{C-NMR}$ (75 MHz) spectrum of 5e in a $\text{CDCl}_3+\text{DMSO-}d_6$



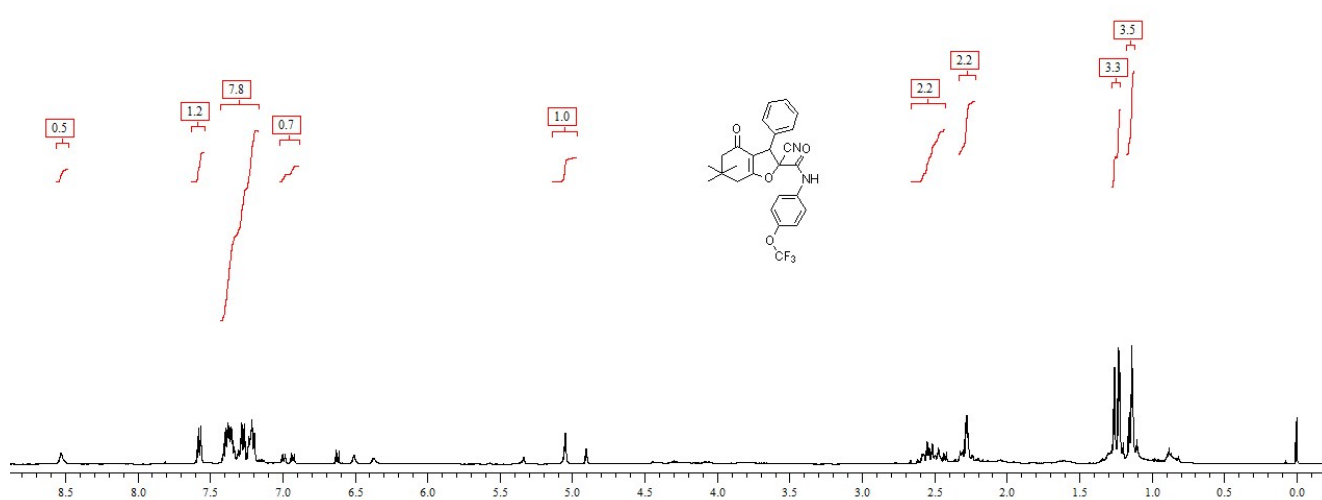




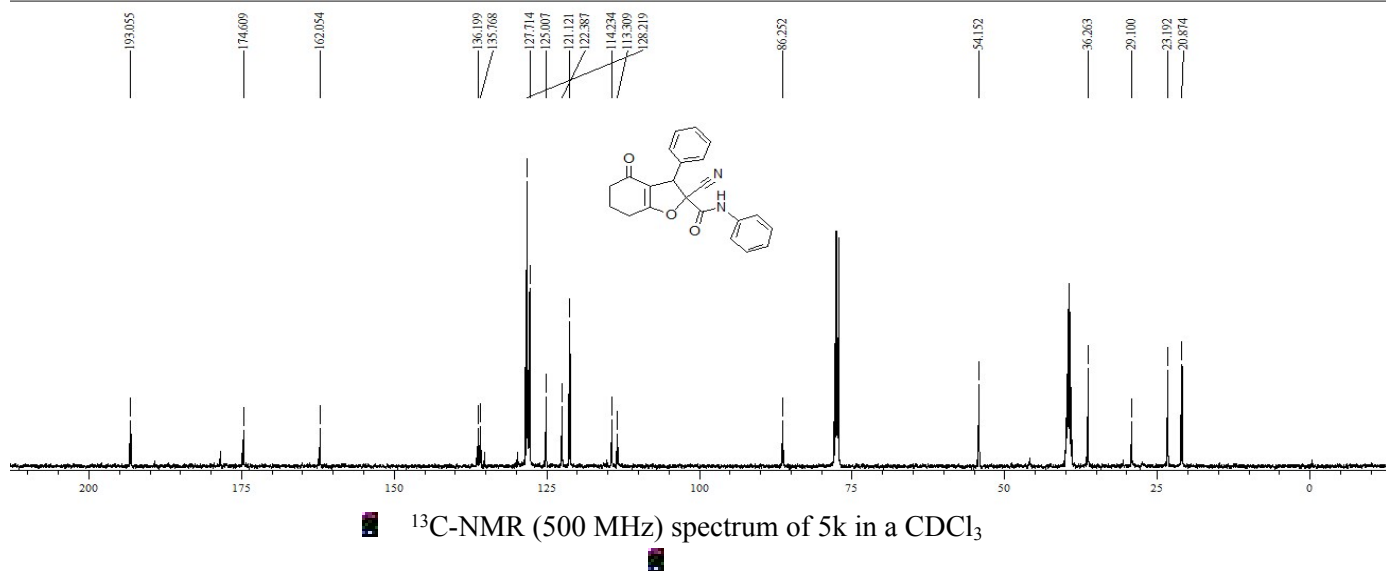
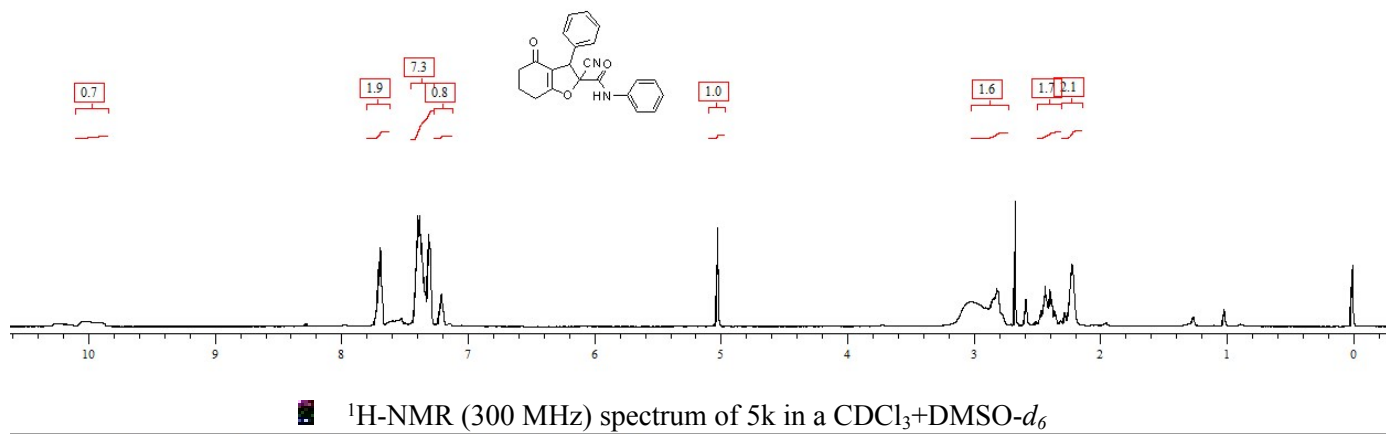
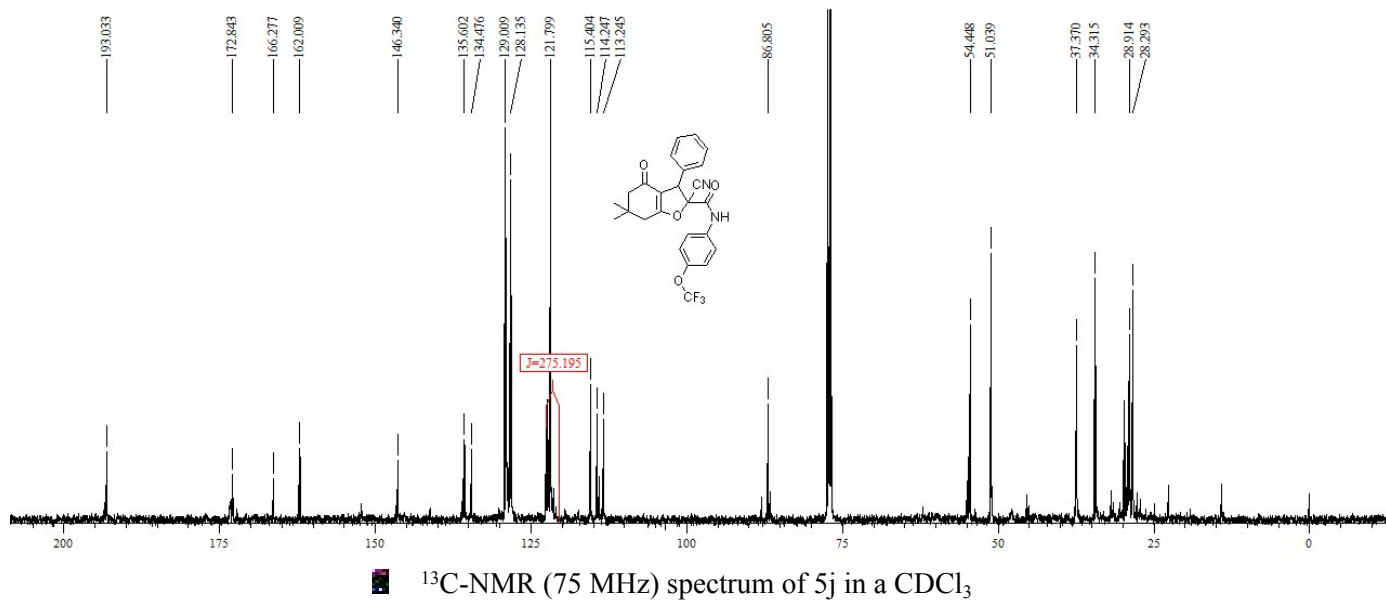
1 ¹H-NMR (500 MHz) spectrum of 5i in a CDCl₃

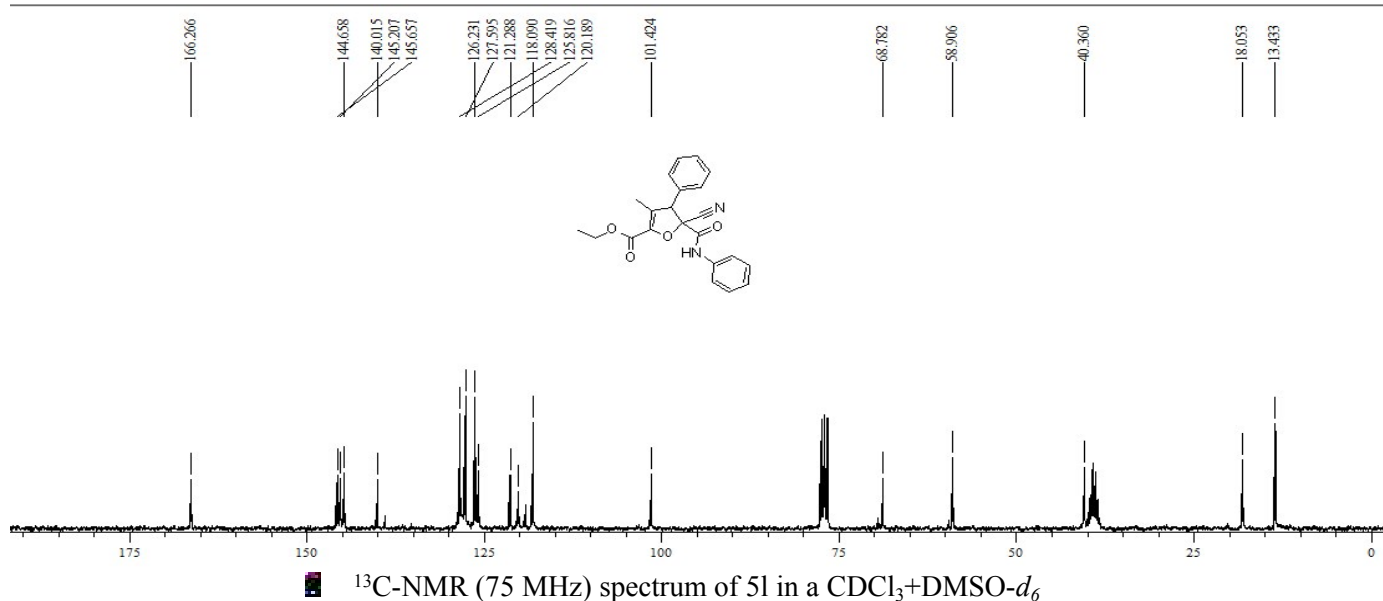
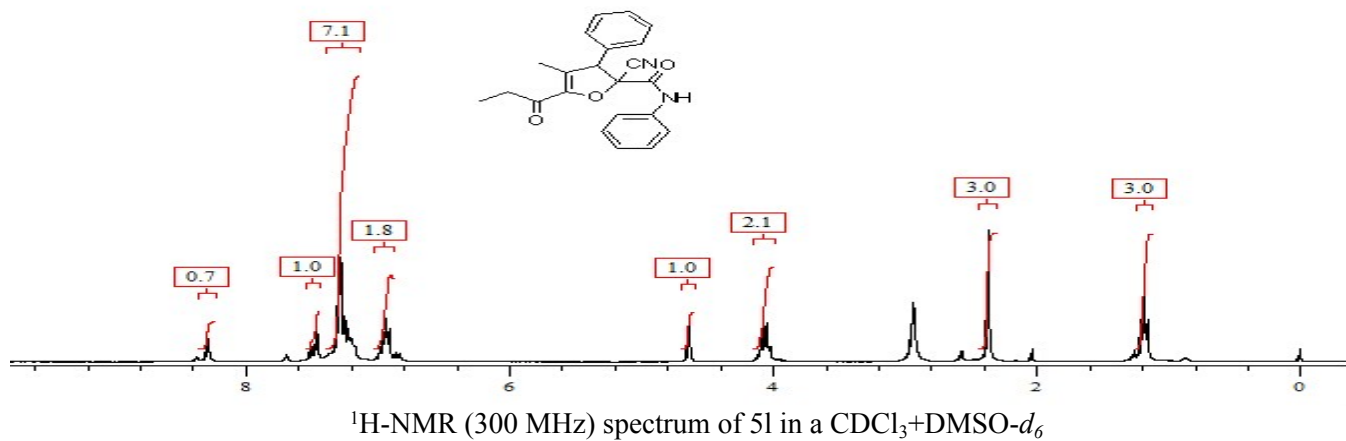


1 ¹³C-NMR (75 MHz) spectrum of 5i in a CDCl₃+DMSO-*d*₆

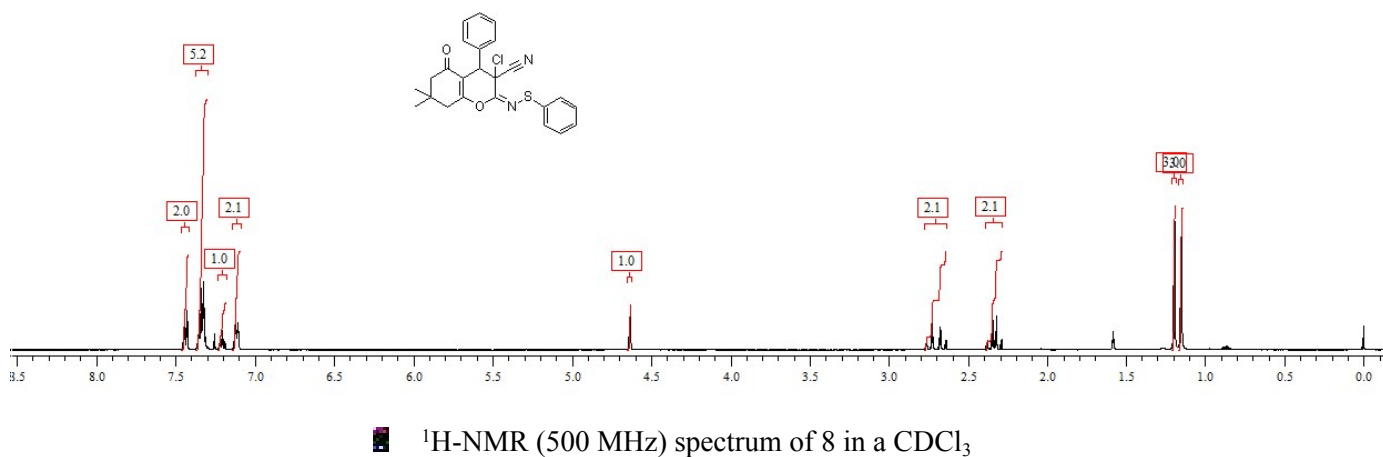


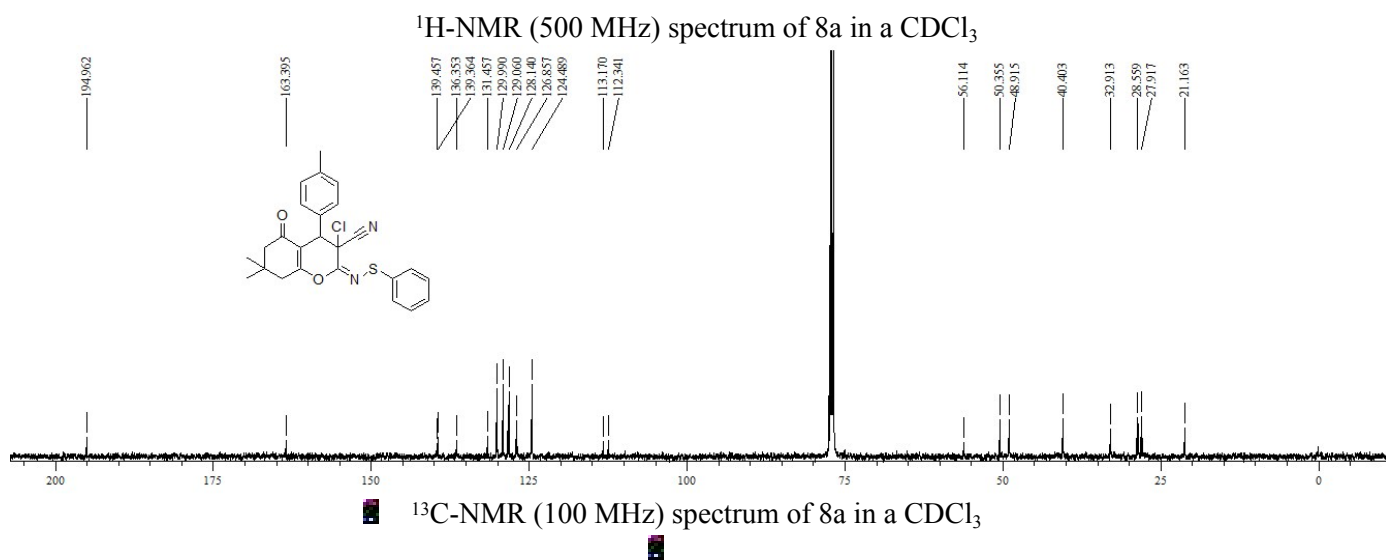
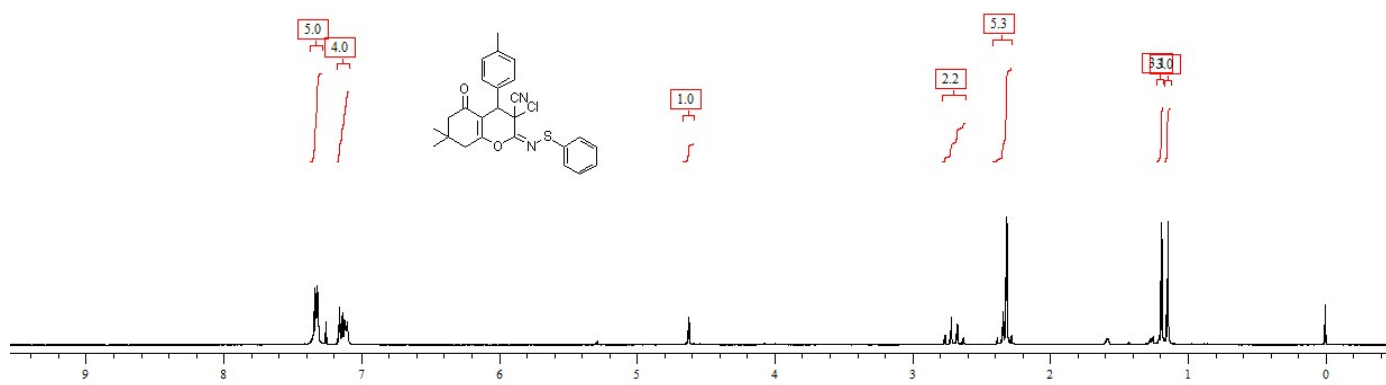
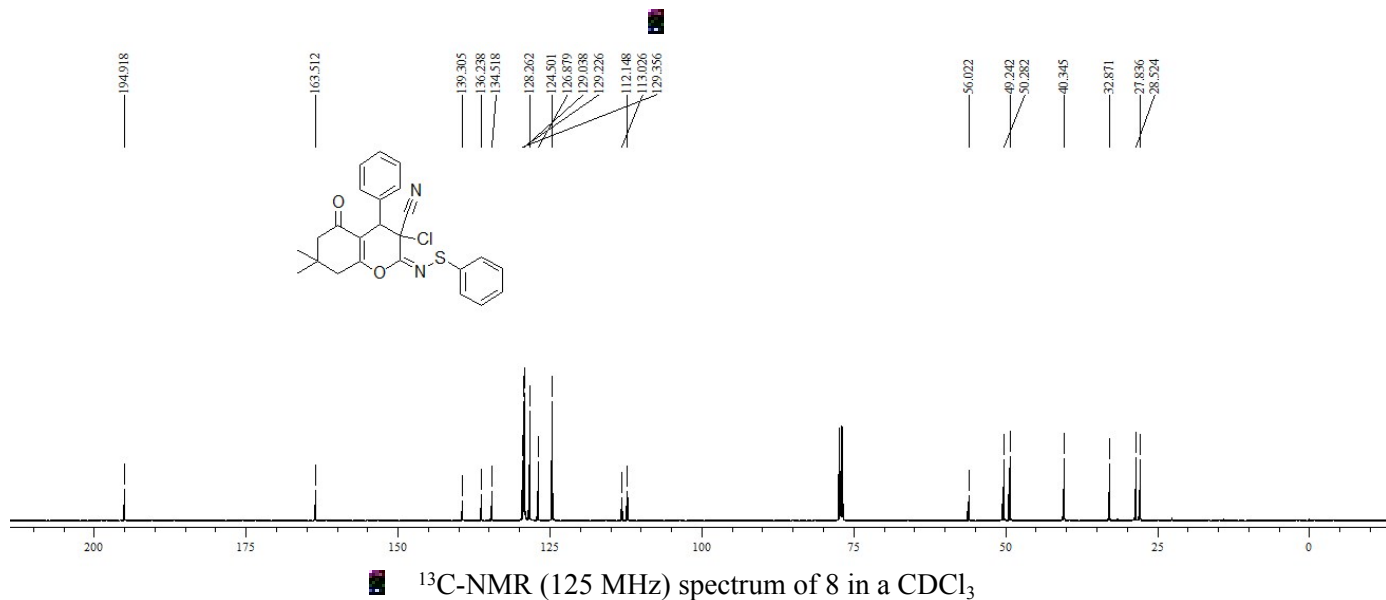
1 ¹H-NMR (500 MHz) spectrum of 5j in a CDCl₃

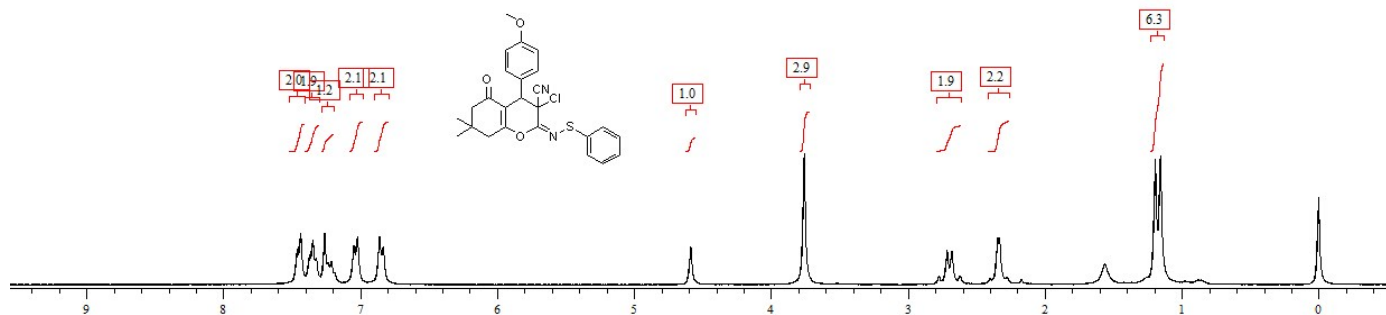




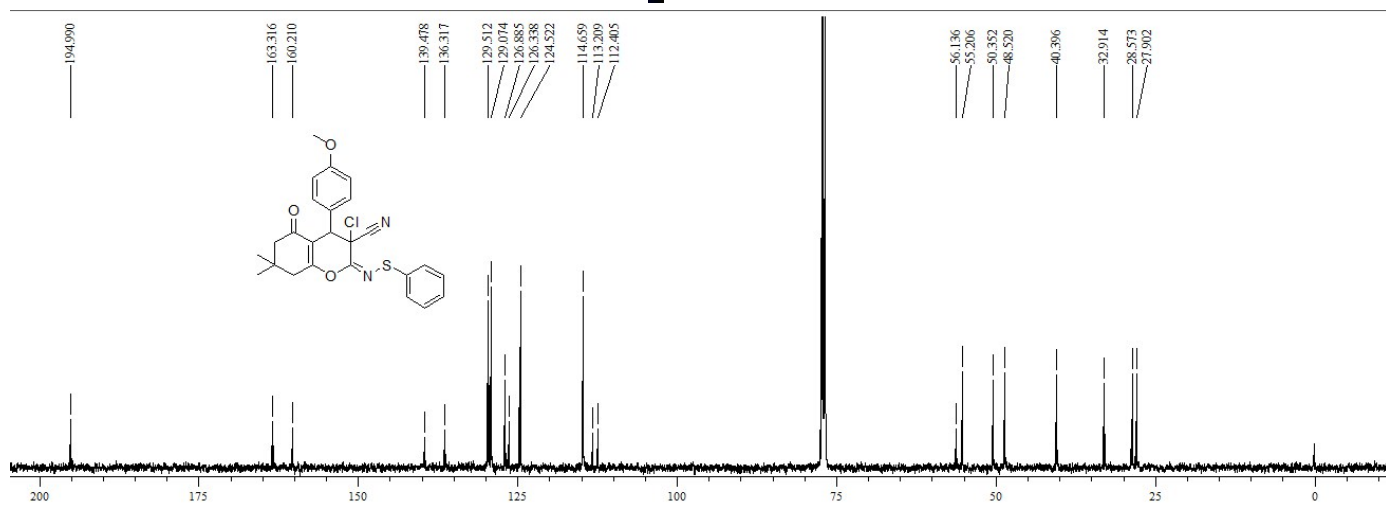
¹H and ¹³C NMR spectra of 8 (a-k) :



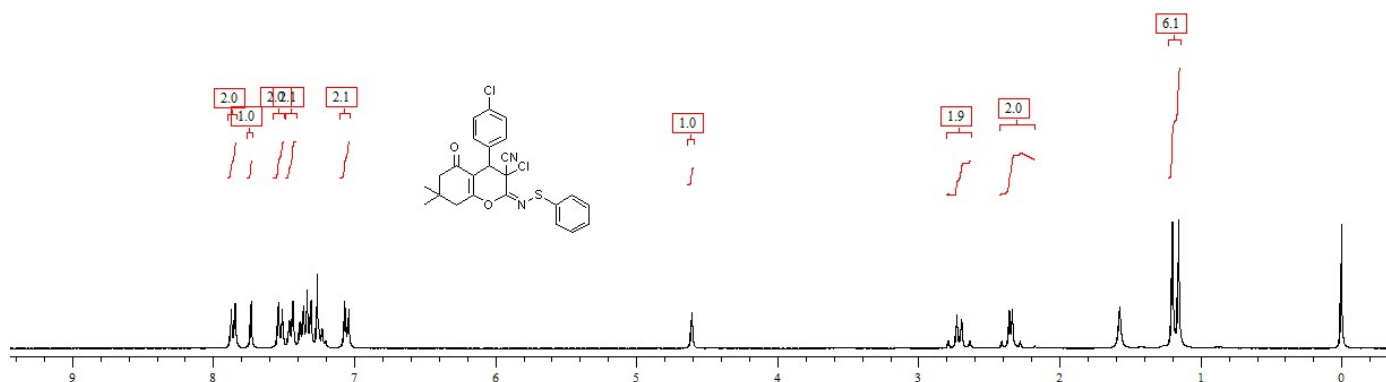




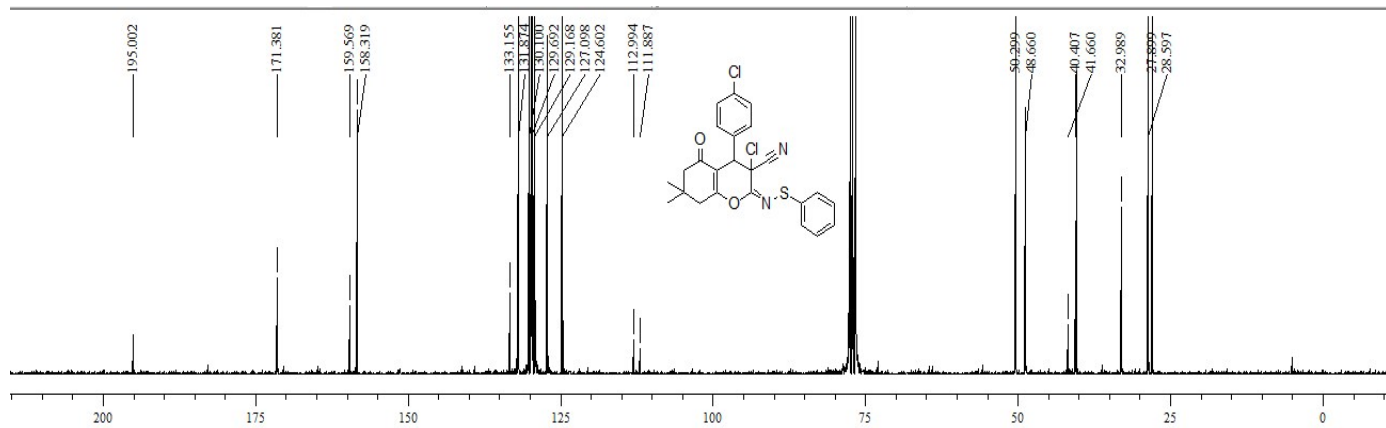
1 ¹H-NMR (300 MHz) spectrum of 8b in a CDCl₃



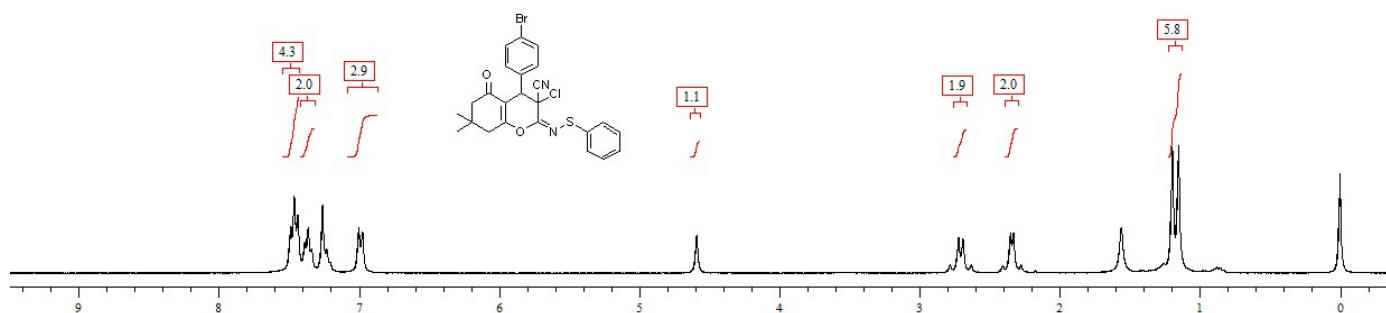
2 ¹³C-NMR (125 MHz) spectrum of 8b in a CDCl₃



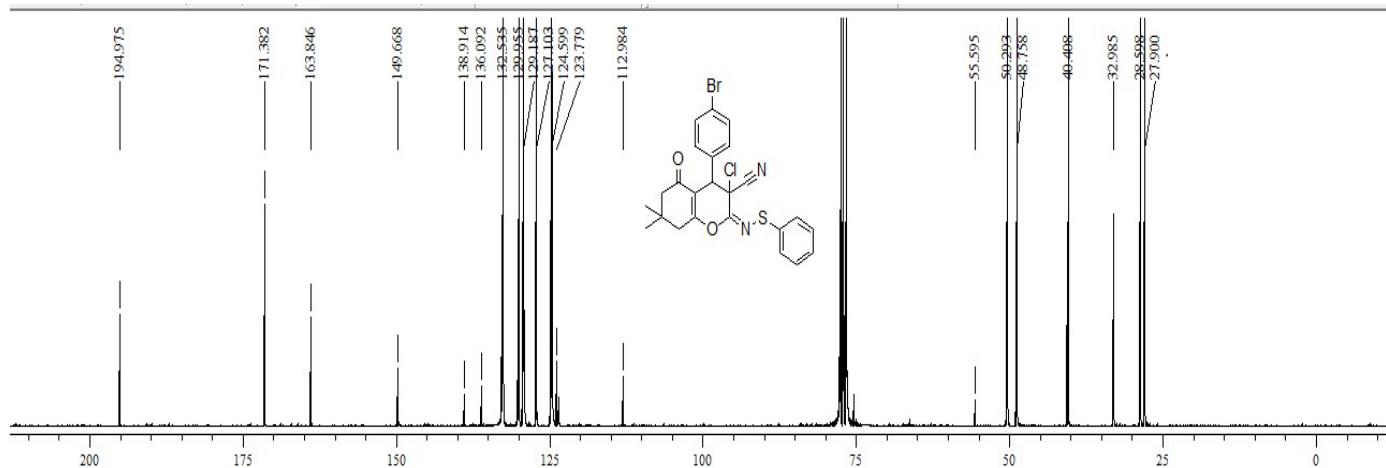
3 ¹H-NMR (300 MHz) spectrum of 8c in a CDCl₃



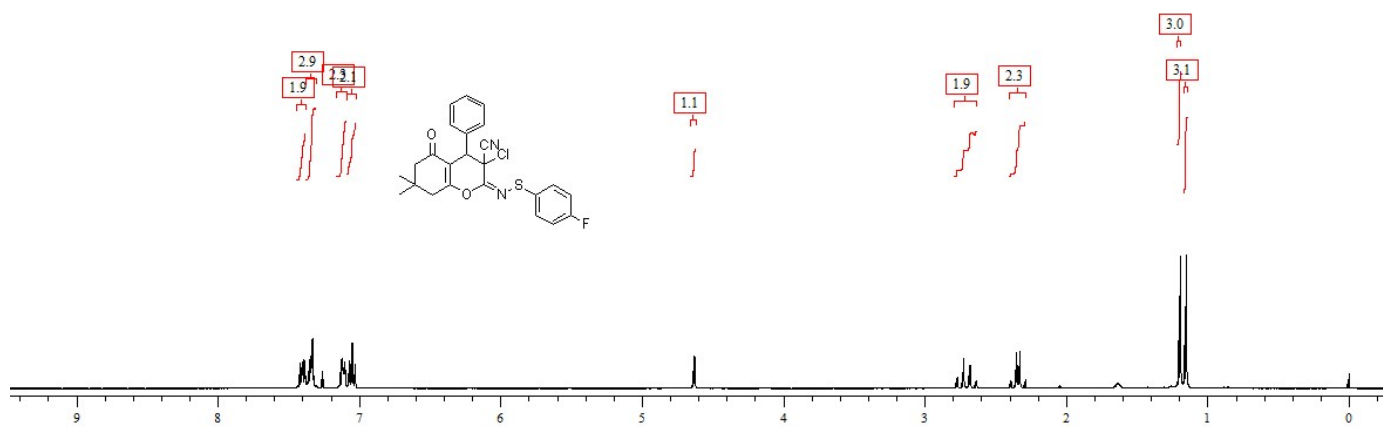
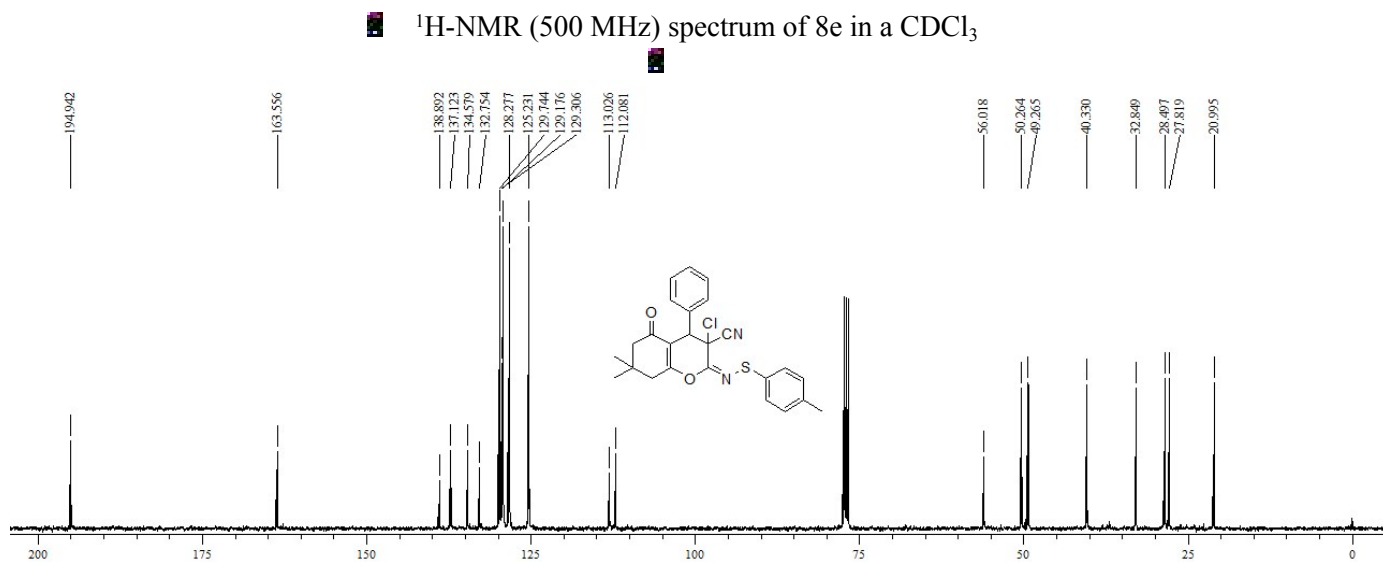
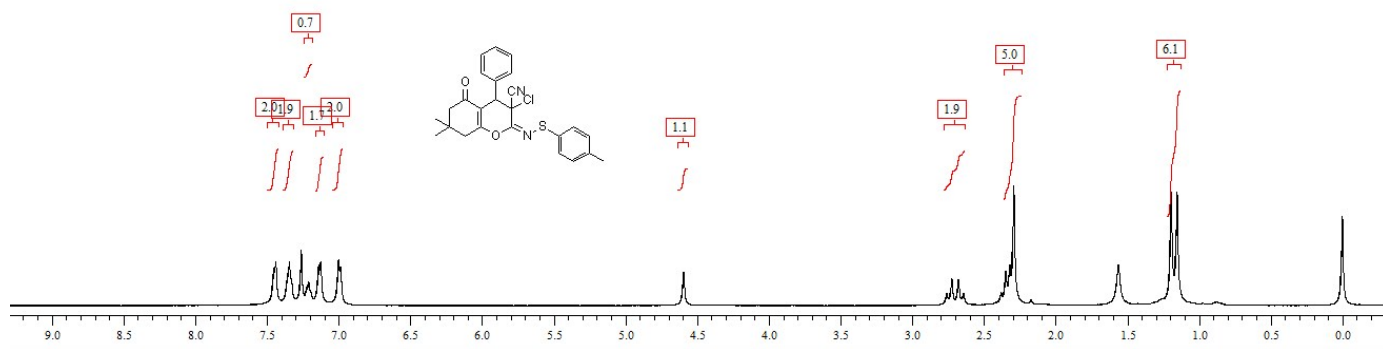
^{13}C -NMR (125 MHz) spectrum of 8c in a CDCl_3



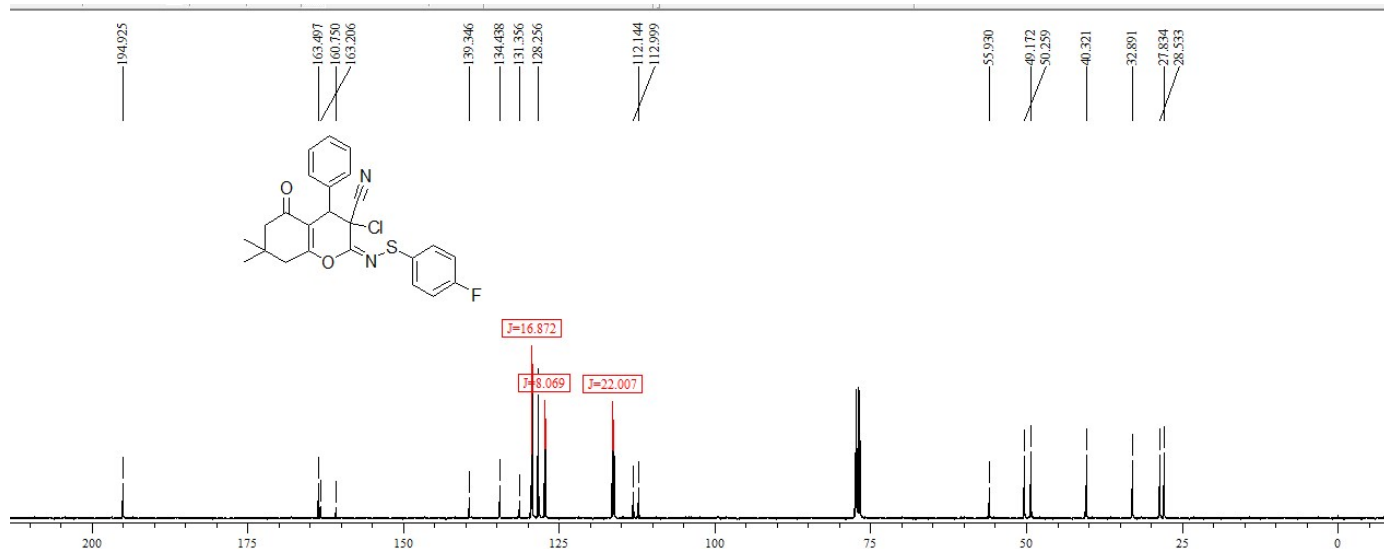
^1H -NMR (300 MHz) spectrum of 8d in a CDCl_3



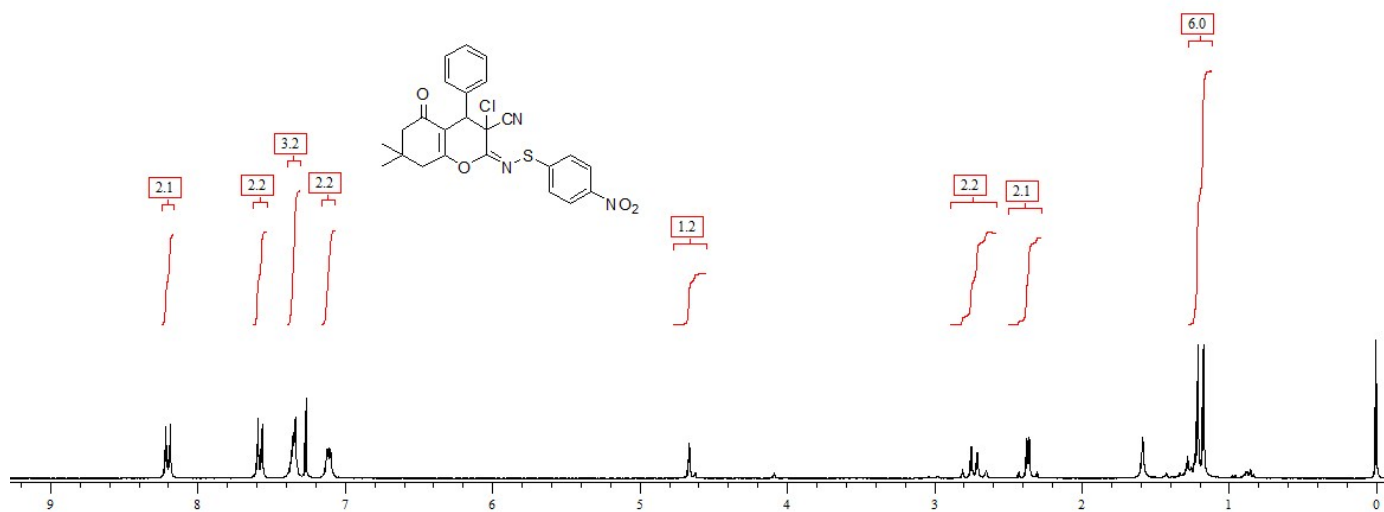
^{13}C -NMR (125 MHz) spectrum of 8d in a CDCl_3



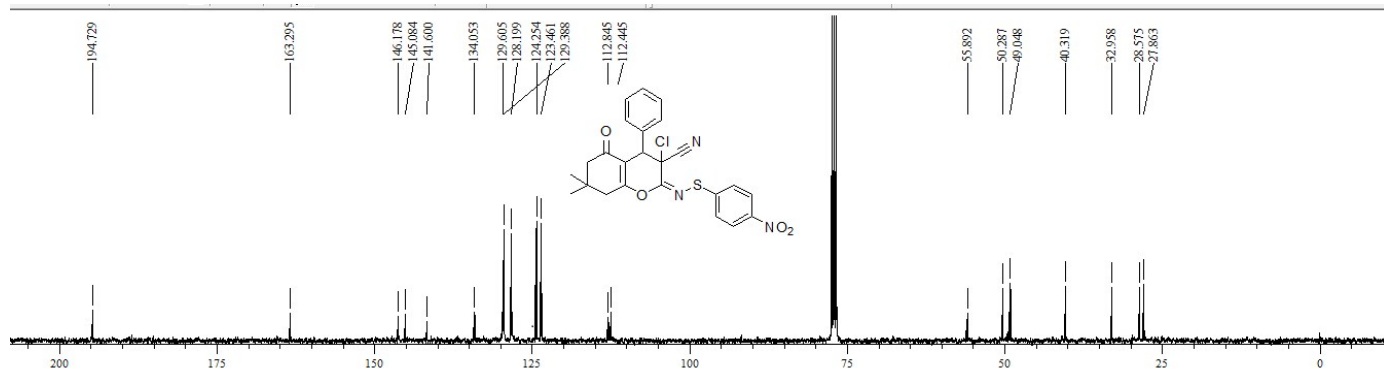
¹H-NMR (400 MHz) spectrum of 8f in a CDCl₃



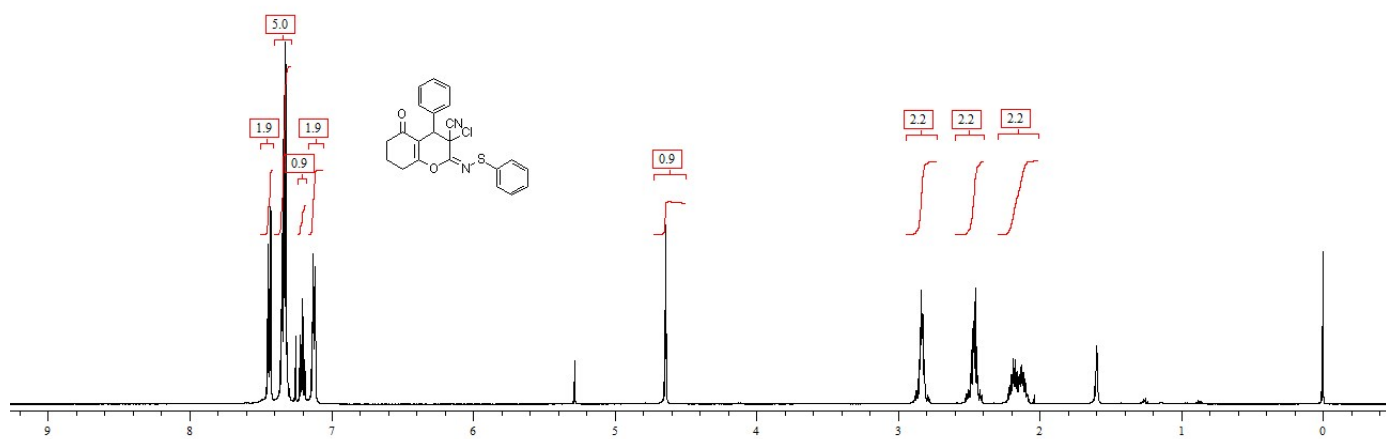
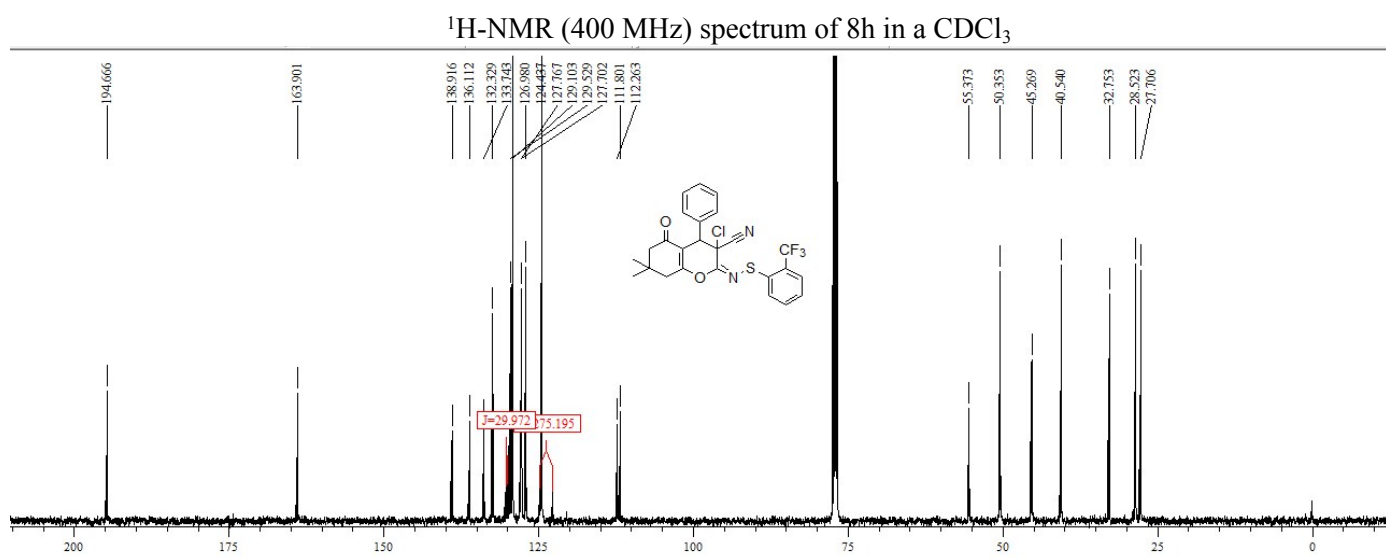
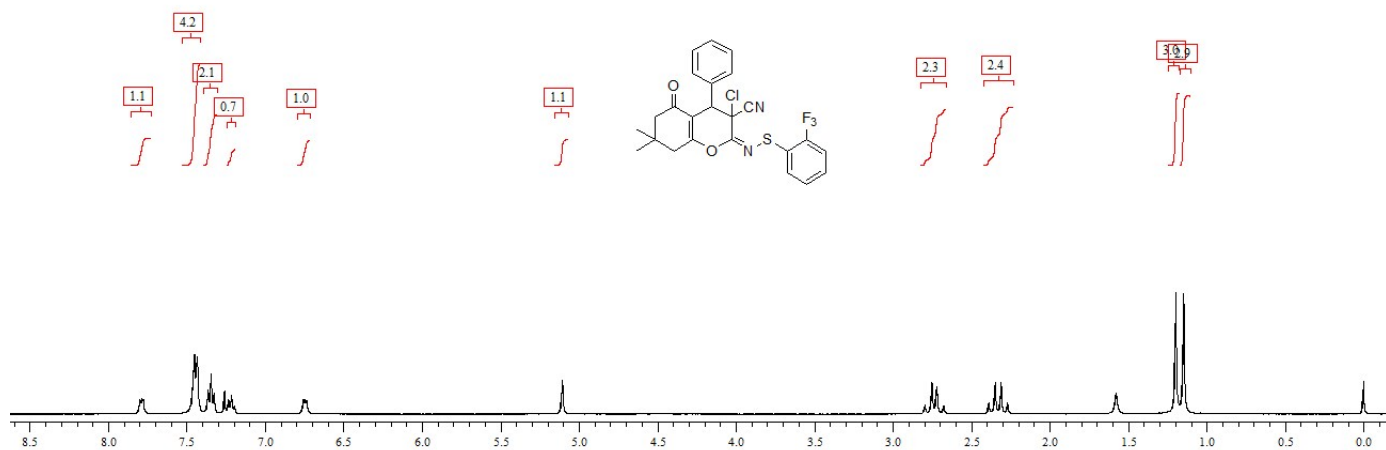
¹³C-NMR (100 MHz) spectrum of 8f in a CDCl₃

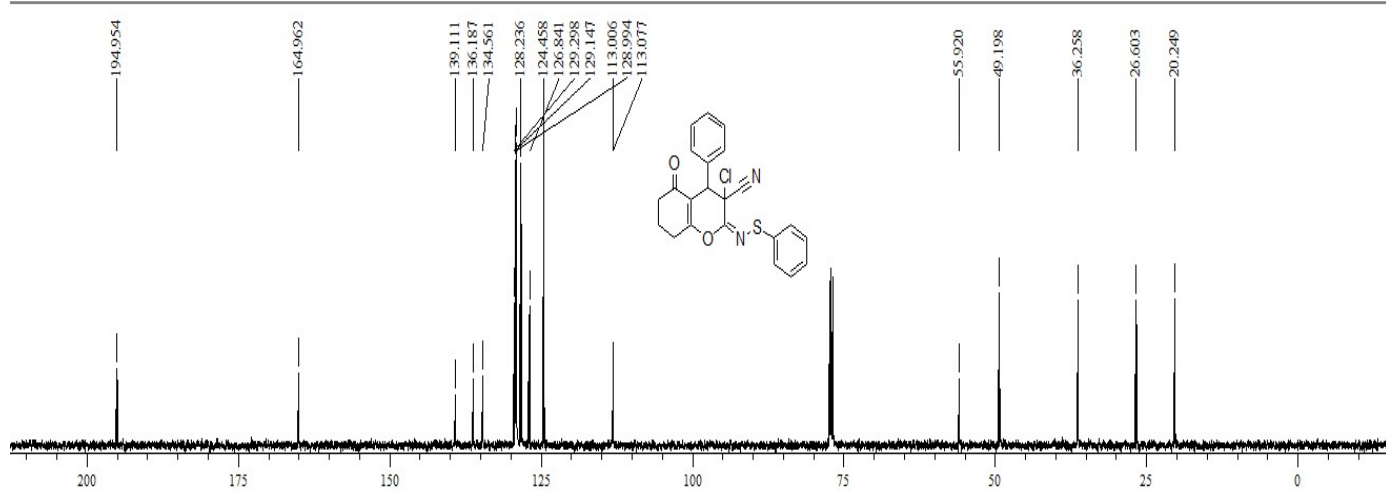


¹H-NMR (500 MHz) spectrum of 8g in a CDCl₃

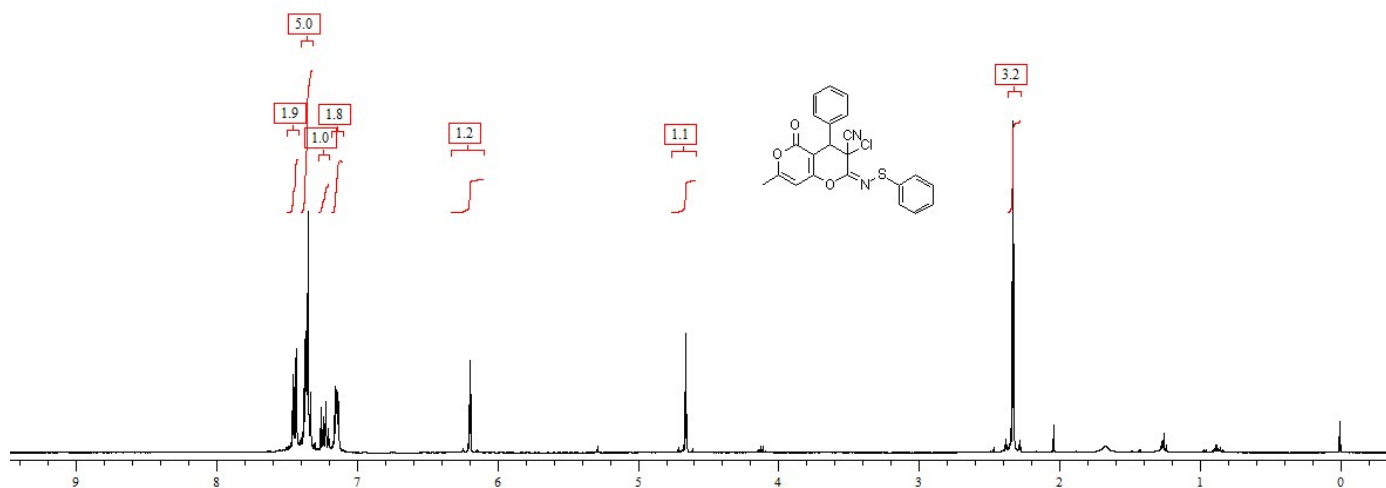


¹³C-NMR (100 MHz) spectrum of 8g in a CDCl₃

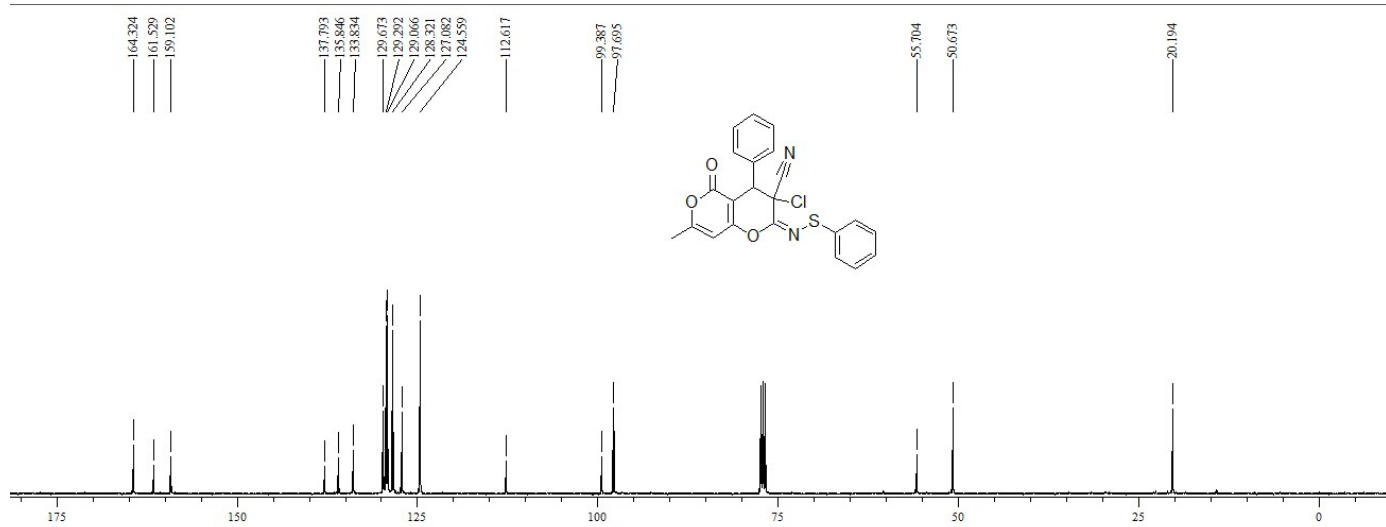




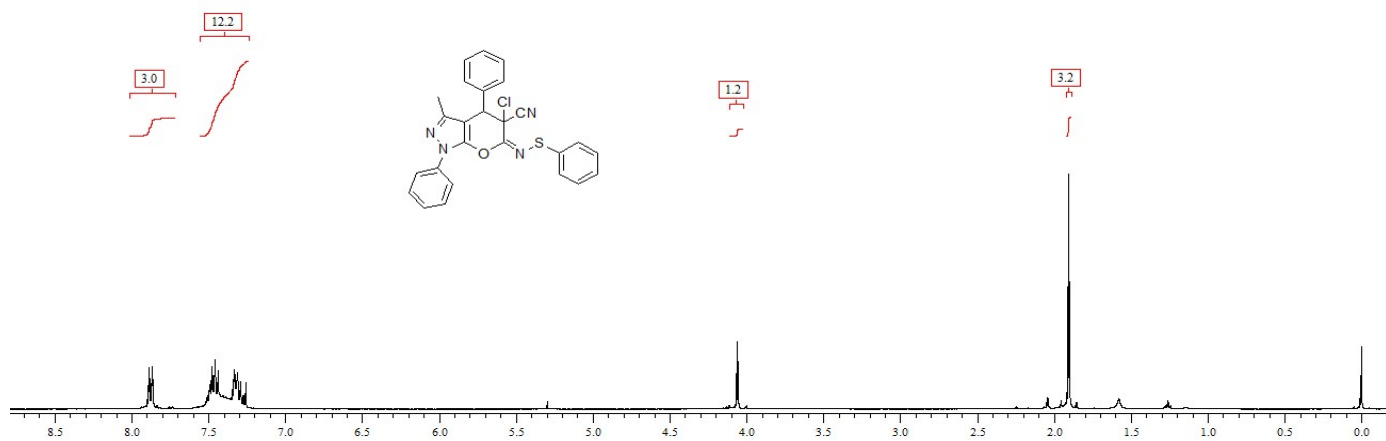
$^{13}\text{C-NMR}$ (125 MHz) spectrum of 8i in a CDCl_3



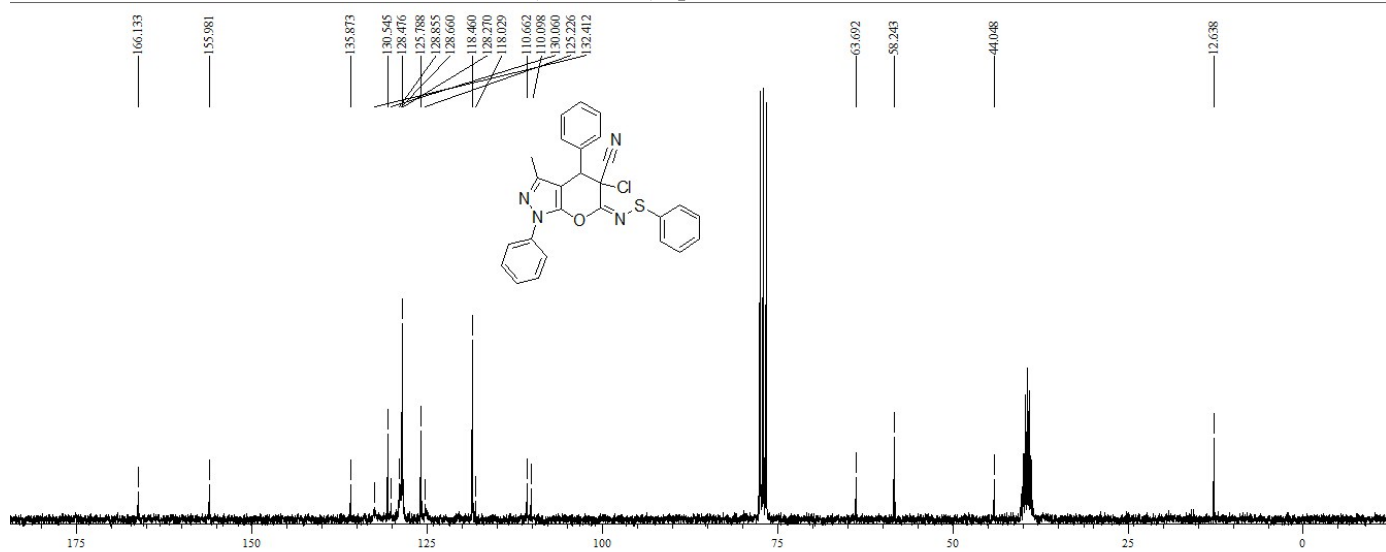
$^1\text{H-NMR}$ (500 MHz) spectrum of 8j in a CDCl_3



$^{13}\text{C-NMR}$ (125 MHz) spectrum of 8j in a CDCl_3



¹H-NMR (500 MHz) spectrum of 8k in a CDCl₃



¹³C-NMR (125 MHz) spectrum of 8k in a CDCl₃+DMSO-*d*₆