SUPPLEMENTAL INFORMATION

Hybridation of Thiazolidinone with Hydroxamate Scaffold for

Developing Novel HDAC Inhibitors with Antitumor Activities

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Table of contents

Part A: Biology Experimental Procedures	
Part B: Chemistry Experimental Procedures	
Part C: Spectra of ¹ H and ¹³ C NMR and HR MS of target compounds	S16 – S63

Part A: Biology Experimental Procedures

A.1. HDAC1 activity assay.

HDAC1 activity assay was performed as described in the study report of Shanghai Chemparter CO., LTD. The inhibitory effect of compounds on HDAC1 function was determined in vitro using an optimized homogenous assay performed in 384-well plate (Perkin Elmer, Cat. No. 6007279) format. In this assay, enzyme solution and substrate solution (trypsin and Ac-peptide substrate) were prepared in 1x assay buffer and solid compounds for test were dissolved to 10 mM in 100% DMSO. Firstly, Compounds were transferred to 384-well plate by Echo® Liquid Handler (Labcyte, USA) with three times dilution in 100% DMSO. 15 μ L of enzyme solution or 1x assay buffer for low control was transferred to assay plate subsequently and incubated at room temperature for 15 min. And then 10 μ L of substrate solution was added to each well to start reaction and incubated at room temperature for 60 min. Fluorescence measurements were obtained using a multilabel plate reader Synergy MX with excitation at 355 nm and emission at 460nm. Fit the data in XL-fit to obtain IC50 values using equation $Y = Bottom + (Top-Bottom)/(1+10^{(LogIC_{50}-X)*Hill})$ Slope)) [Y is % inhibition and X is compound concentration].

A.2. Cell lines and culture conditions

Tumor cell lines used in this study were obtained from the American Type Culture Collection (ATCC). LNCaP, MCF-7 and Hela cell lines were cultured in RPMI 1640 medium, and A549 cell line was cultured in DMEM medium. Medium was supplemented with 10% FBS. All tumor cells were incubated at 37 °C and 5% CO₂

incubator.

A.3. Cell Viability assay

The cell viability of tumor cell lines in the presence of this series of compounds was determined by SRB (Sigma Aldrich) assay which was described previously.¹ In brief, cells were seeded into 96-well plates at the appropriate cell densities during the experiment. After incubation for 24 h, the cells were treated with various concentrations of the compound for 48 h. Control group were exposed to DMSO at a concentration equivalent to that of the compound-treated cells. After treatment for 48 h, 25 μ L of 50% TCA was added for cell fixation at 4 °C. At least 1 h later or more, the plates were washed by water for five times. The plates were allowed to dry using hair dryer followed by being dyed with 100 μ L 0.4% SRB for 10 min. After dying, the plates were washed by 1% acetic acid to remove the dye and allowed to dry using hair dryer. 100 μ L of 10 mM Tris-based solution was added to each well, and absorbance was measured using a 96-well plate reader at 515 nm. The IC₅₀ was calculated using GraphPad Software.

A.4. In vitro transwell migration assay

Transwell migration assay was performed as previously reported.² The inhibition of tumor cell migration was assessed by the Boyden chamber (Corning Falcon) migration assay in 24-well cell culture plate with 8.0- μ m pore. Briefly, A549 cells were collected, centrifuged, and re-suspended with serum-free medium. The top chambers were seeded with 5*10⁴ cells in 200 μ L of serum-free DMEM medium containing different concentrations of compound. The bottom chambers were filled

with 700 μ L of complete medium supplemented with different concentrations of compound. After 18 h incubation, non-migrated cells were removed with cotton swabs, and migrated cells were fixed with cold 4% paraformaldehyde and stained with 0.2% crystal violet for 1 min. Then the chambers were washed using water, and the membrane was left to dry. Images were taken with an inverted microscope (Olympus) and cells from three random areas per filter were counted.

A.5. Western blotting

Western blotting was performed as described previously.³ Cells were exposed to various concentrations of compounds for indicated time and lysed in RIPA buffer [(50 mM Tris-HCl (pH 7.4), 150 mM NaCl, 5 mM EDTA, 1 % Triton X-100, 1 % sodium deoxycholic acid, 0.1% SDS, 2 mM phenylmethanesulfonyl fluoride (PMSF), 30 mM Na₂HPO₄, 50 mM NaF, 1 mM Na₃VO₄] containing protease/phosphatase inhibitors (Roche). Lysates were combined with sample loading buffer and heated at 100 $^{\circ}$ C. Use a Bicinchoninic acid assay (Thermo Scientific) to quantify Protein concentration. Lysates were mixed with sample loading buffer and heated at 100 °C for 15 min. After separated by 8-15% SDS-PAGE, extracted protein were transferred to nitrocellulose membranes. Membranes were incubated in 5 % (w/v) bovine serum albumin (TBST/BSA) and stored overnight at $4 \,^{\circ}$ C on a shaker with specific primary antibodies (1/1,000 in TBST/BSA). Then membranes were washed with TBST and incubated for 45 min with secondary antibody (1/10,000 in TBST/BSA) at room temperature. . Immunoreactive proteins were visualized using the Odyssey Fluorescence Scanner (LI-COR Bioscience, Inc., Lincoln, NE, USA).

A.6 Cell cycle analysis

Cell cycle analysis was conducted by PI staining as described previously.⁴ LNCaP and A549 cells were plated in 6 cm dishes and were treated with compounds and the same amount of DMSO as control for 24 h. After ethanol fixation, cells were washed in PBS once and suspended in PBS with 200 µg/ml RNAase and 50 µg/ml propidium iodide (PI) in dark for 30 minutes. Then cells were analyzed by flow cytometry (FACS Calibur, BD Biosciences).

A.7 Apotosis assay

Apoptotic cells were monitored with Annexin V-FITC Apoptosis Detection Kit I (BD Biosciences) as described previously⁴. LNCaP and A549 cells were plated in 6 cm dishes and were treated with compounds and the same amount of DMSO as control for 48 h. Cells were washed with cold PBS, harvested and re-suspended in 100 μ L 1×binding buffer, and incubated with 5 μ l Annexin V fluorescein isothiocyanate and 5 μ l propidium iodide for 15 min in dark at room temperature. Then 400 μ l of 1×binding buffer was added and analyzed immediately with flow cytometry (FACS Calibur, BD Biosciences).

Part B: Chemistry Experimental Procedures

B.1. Chemistry: general methods

Reagents were purchased from Adamas-beta Ltd, Sigma-Aldrich Inc., J&K Inc., or Aladdin-reagents Inc., and used without further purification unless otherwise specified. All reactions were carried out with the use of standard techniques under an inert atmosphere (Ar or N₂). ¹H and ¹³C NMR spectra were generated on a Varian 300 MHz or Bruker 500 Hz instruments and obtained as CDCl₃ or DMSO- d_6 solutions (reported in ppm), using CDCl₃ as the reference standard (7.26 and 77.00 ppm) or DMSO- d_6 (2.50 and 39.51 ppm). Coupling constants (*J*) are reported in Hertz (Hz). Standard abbreviations indicating spin multiplicities are given as follow: s (singlet), d (doublet), t (triplet), q (quartet), br (broad) or m (multiplet). High resolution mass spectra were gathered on Bruker MicroTOF-Q II LCMS instrument operating in electrospray ionization (ESI). HPLC (Agilent Technologies 1200 Series) was employed for purity determination, using the following method: Eclipse XDB C18 column, 5 μ m, 4.6 mm×150 mm, column temperature 40 °C; solvent A: water; solvent B: methanol; gradient of 40–70% B (0–10 min), 70–90% B (10–15 min), 90–40% B (15–20 min); flow rate of 1.5 mL/min. Compound purity was determined by high pressure liquid chromatography (HPLC) with a confirming purity of ≥95% for all of the final biologically tested compounds.

B.2. General procedure for the preparation of target compounds

N^{1} -hydroxy- N^{4} -(3-(3-(2-methoxyphenyl)-4-oxo-2-thiazolidinyl)phenyl)succinamide

(10a) To a solution of hydroxyl amine hydrochloride (4.17 g, 60.0 mmol) in 10 mL MeOH and KOH (3.37 g, 60.0 mmol) was added to the mixture and stirred for 10 min at 40 °C, then the reaction mixture was cooled to 0 °C and filtered. Compound N-(3-(3-(2-methoxyphenyl)-4-oxo-2-thiazolidinyl)phenyl)succinamic acid methyl ester (16a) (500 mg, 1.2 mmol) was added to the filtrate followed by KOH (337 mg, 6.0 mmol) at room temperature for 30 min. The solvent was removed and extracted with EtOAc. The organic layer was washed with saturated NH₄Cl aqueous solution and brine, dried over Na₂SO₄, the residue was purified by column chromatography [eluting with EA followed by 10:1 CH₂Cl₂/MeOH] to give compound 10a as a

colorless oil (134 mg, 27.8% yield). ¹H NMR (DMSO-*d*₆, 300 MHz): δ 10.43 (br s, 1H), 9.97 (br s, 1H), 8.71 (br s, 1H), 7.68 (s, 1H), 7.43 (d, *J* = 8.1 Hz, 1H), 7.23–7.13 (m, 2H), 7.02 (d, *J* = 8.4 Hz, 2H), 6.99-6.97 (m, 1H), 6.81 (dd, *J* = 7.8, 7.8 Hz, 1H), 6.08 (s, 1H), 3.93 (d, *J* = 15.9 Hz, 1H), 3.83 (d, *J* = 15.9 Hz, 1H), 3.71 (s, 3H), 2.55–2.50 (m, 2H), 2.28–2.23 (m, 2H). ¹³C NMR (125 MHz, DMSO-*d*₆) δ 170.67, 170.28, 168.30, 154.84, 140.28, 139.42, 130.01, 129.24, 128.71, 125.46, 121.89, 120.26, 118.99, 117.53, 112.34, 63.27, 55.72, 32.07, 31.44, 27.33. HPLC purity: 96.2%, *t*_R = 3.291 min. HRMS (ESI): calcd for [C₂₀H₂₁N₃O₅S + Na]⁺ 438.1094, found 438.1093.

*N*¹-*hydroxy*-*N*⁷-(*3*-(*3*-(*2*-*methoxyphenyl*)-*4*-*oxo*-*2*-*thiazolidinyl*)*phenyl*)*heptanediamide* (**10b**) Compound **10b** (18.5% yield) was prepared according to the procedure described for the preparation of compound **10a.** ¹H NMR (DMSO-*d*₆, 300 MHz): δ 10.35 (br s, 1H), 9.88 (br s, 1H), 8.67 (br s, 1H), 7.66 (s, 1H), 7.45 (d, *J* = 8.1 Hz, 1H), 7.21–7.13 (m, 2H), 7.03 (d, *J* = 8.4 Hz, 2H), 7.00-6.98 (m, 1H), 6.81 (dd, *J* = 7.8, 7.8 Hz, 1H), 6.09 (s, 1H), 3.94 (d, *J* = 15.6 Hz, 1H), 3.83 (d, *J* = 15.6 Hz, 1H), 3.78 (s, 3H), 2.26 (t, *J* = 7.5 Hz, 2H), 1.95 (t, *J* = 7.5 Hz, 2H), 1.58–1.47 (m, 4H), 1.27–1.22 (m, 2H). ¹³C NMR (125 MHz, DMSO-*d*₆) δ 171.21, 170.67, 169.02, 154.85, 140.28, 139.42, 129.99, 129.25, 128.73, 125.48, 121.91, 120.26, 119.09, 117.62, 112.34, 63.27, 55.71, 36.22, 32.12, 32.08, 28.21, 24.90, 24.74. HPLC purity: 95.5%, *t*_R = 4.208 min. HRMS (ESI): calcd for [C₂₃H₂₇N₃O₅S + Na]⁺ 480.1564, found 480.1570. *N*-(*3*-(*hydroxycarbamoyl*)*propyl*)-*3*-(*3*-(*2*-*methoxyphenyl*)-*4*-*oxo*-*2*-*thiazolidinyl*)*benzamide* (**11a**) The synthesis of intermediate 19a

To a solution of **18** (300 mg, 0.90 mmol) in 8 mL DMF was added EDC·HCl (224 mg, 1.17 mmol) and HoBt (134 mg, 0.99 mmol), stirred for 10 min at 0 °C, then $NH_2(CH_2)_3COOCH_3$ (158 mg, 1.5 mmol) was added to the mixture. The solvent was removed and extracted with EtOAc and H₂O. The organic layer was washed with brine, dried over Na₂SO₄, the residue was purified by column chromatography [eluting with 4:1 PE/EA] to give compound **19a** as a colorless oil (290 mg, 75.4% yield).

Compound **11a** (34.1% yield) was prepared according to the last step described for the preparation of compound **10a.** ¹H NMR (DMSO- d_6 , 300 MHz): δ 10.39 (br s, 1H), 8.72 (br s, 1H), 8.54–8.51 (m, 1H), 7.87 (s, 1H), 7.70 (d, J = 7.5 Hz, 1H), 7.52 (d, J =7.5 Hz, 1H), 7.34 (dd, J = 7.5 Hz, 7.5Hz, 1H), 7.23–7.177 (m, 1H), 7.04–6.99 (m, 2H), 6.80 (dd, J = 7.5, 7.5 Hz, 1H), 6.21 (s, 1H), 4.03 (d, J = 15.6 Hz, 1H), 3.86 (d, J =15.6 Hz, 1H), 3.77 (s, 3H), 3.27–3.19 (m, 2H), 2,01–1.97 (m, 2H), 1.74–1.69 (m, 2H). ¹³C NMR (125 MHz, DMSO- d_6) δ 170.63, 168.85, 165.52, 154.85, 140.07, 134.65, 130.15, 129.83, 129.32, 128.38, 127.41, 126.19, 125.43, 120.31, 112.37, 79.18, 63.09, 55.72, 32.20, 29.98, 25.29. HPLC purity: 95.4%, $t_R = 7.355$ min. HRMS (ESI): calcd for [C₂₁H₂₃N₃O₅S +Na]⁺ 452.1251, found 452.1255.

N-(5-(hydroxycarbamoyl)pentyl)-3-(3-(2-methoxyphenyl)-4-oxo-2-thiazolidinyl)benzamide (**11b**) Compound **11b** (31.1% yield) was prepared according to the procedure described for the preparation of compound **11a.** ¹H NMR (DMSO-*d*₆, 300 MHz): δ 10.37 (br s, 1H), 8.71 (br s, 1H), 8.49 (dd, *J* = 5.4, 5.4 Hz, 1H), 7.86 (s, 1H), 7.69 (d, *J* = 7.8 Hz, 1H), 7.52 (d, J = 8.1 Hz, 1H), 7.34 (dd, J = 7.8, 8.1 Hz, 1H), 7.20 (dd, J = 7.2, 7.2 Hz, 1H), 7.04–7.00 (m, 2H), 6.83–6.78 (m, 1H), 6.21 (s, 1H), 4.03 (d, J = 15.6 Hz, 1H), 3.85 (d, J = 15.6 Hz, 1H), 3.77 (s, 3H), 3.22–3.17 (m, 2H), 1.96–1.91 (m, 2H), 1.50–1.43 (m, 4H), 1.26–1.22 (m, 2H). ¹³C NMR (125 MHz, DMSO- d_6) δ 170.60, 169.04, 165.37, 154.84, 140.03, 134.73, 130.08, 129.81, 129.31, 128.34, 127.38, 126.16, 125.42, 120.29, 112.36, 63.07, 55.70, 48.59, 32.21, 32.19, 28.89, 26.10, 24.90. HPLC purity: 98.2%, $t_R = 3.495$ min. HRMS (ESI): calcd for [C₂₃H₂₇N₃O₅S + Na]⁺ 480.1564, found 480.1561.

4-{2-[2-(3-bromophenyl)-4-oxo-3-thiazolidinyl]phenoxy}-N-hydroxybutyramide (12a) The synthesis of intermediate 22a

To a solution of **21** (310 mg, 0.88 mmol) in 8 mL of DMF was added K₂CO₃ (243 mg, 1.76 mmol), then Br(CH₂)₃COOCH₃ (318 mg, 1.76 mmol) was added to the mixture. The solvent was removed and extracted with EtOAc and H₂O. The organic layer was washed with brine, dried over Na₂SO₄, the residue was purified by column chromatography [eluting with 4:1 PE/EA] to give compound **22a** as a colorless oil (340 mg, 86.1% yield).

Compound **12a** (65.2% yield) was prepared according to the last step described for the preparation of compound **10a.** ¹H NMR (DMSO- d_6 , 300 MHz): δ 10.44 (br s, 1H), 8.75 (br s, 1H), 7.63 (s, 1H), 7.44–7.40 (m, 2H), 7.25 (d, J = 7.8 Hz, 1H), 7.20 (d, J =7.8 Hz, 1H), 7.02–7.00 (m, 2H), 6.87–6.81(m, 1H), 6.18 (s, 1H), 4.06 (d, J = 15.6 Hz, 1H), 3.98–3.94 (m, 2H), 3.84 (d, J = 15.6 Hz, 1H), 2.21–2.16 (m, 2H), 2.04–1.98 (m, 2H). ¹³C NMR (125 MHz, DMSO- d_6) δ 170.54, 168.68, 154.00, 142.72, 131.45,

S9

130.59, 130.05, 129.35, 126.50, 125.53, 121.53, 120.34, 113.06, 67.30, 62.60, 32.02, 28.82, 24.92. HPLC purity: 97.3%, $t_{\rm R} = 6.745$ min. HRMS (ESI): calcd for $[C_{19}H_{19}BrN_2O_4S + Na]^+ 474.0141$, found 474.0135.

5-{2-[2-(3-bromophenyl)-4-oxo-3-thiazolidinyl]phenoxy}-N-hydroxypentaneamide

(12b) Compound 12b (29.5% yield) was prepared according to the procedure described for the preparation of compound 12a. ¹H NMR (DMSO-*d*₆, 300 MHz): δ 10.41 (br s, 1H), 8.73 (br s, 1H), 7.63 (s, 1H), 7.42 (dd, J = 7.5, 7.5 Hz, 2H), 7.25–7.17 (m, 2H), 7.01 (d, J = 7.8 Hz, 2H), 6.82 (dd, J = 7.5, 7.5 Hz, 1H), 6.16 (s, 1H), 4.02 (d, J = 15.6 Hz, 1H), 3.97–3.94 (m, 2H), 3.84 (d, J = 15.6 Hz, 1H), 2.06 (t, J = 6.9 Hz, 2H), 1.76-1.73 (m, 4H). ¹³C NMR (125 MHz, DMSO-*d*₆) δ 170.44, 168.95, 154.12, 142.68, 131.46, 130.59, 130.03, 129.32, 126.52, 125.59, 121.54, 120.25, 113.12, 67.42, 62.59, 32.00, 31.81, 28.17, 21.65. HPLC purity: 97.0%, *t*_R = 7.070 min. HRMS (ESI): calcd for [C₂₀H₂₁BrN₂O₄S + Na]⁺ 487.0298, found 487.0293.

6-{2-[2-(3-bromophenyl)-4-oxo-3-thiazolidinyl]phenoxy}-N-hydroxylhexanamide

(12c) Compound 12c (70.4% yield) was prepared according to the procedure described for the preparation of compound 12a. ¹H NMR (DMSO-*d*₆, 300 MHz): δ 10.39 (br s, 1H), 8.69 (br s, 1H), 7.63 (s, 1H), 7.41 (dd, *J* = 9.0, 9.0 Hz, 2H), 7.24 (d, *J* = 7.8 Hz, 1H), 7.19 (d, *J* = 7.8 Hz, 1H), 7.02 (d, *J* = 7.8 Hz, 2H), 6.82 (dd, *J* = 7.5, 7.5 Hz, 1H), 6.16 (s, 1H), 4.03 (d, *J* = 15.6 Hz, 1H), 3.96–3.91(m, 2H), 3.81 (d, *J* = 15.6 Hz, 1H), 2.03–1.98 (m, 2H), 1.79–1.75 (m, 2H), 1.64–1.57 (m, 2H), 1.50–1.42 (m, 2H). ¹³C NMR (125 MHz, DMSO-*d*₆) δ 170.40, 169.03, 154.17, 142.68, 131.46, 130.59, 130.01, 129.33, 126.51, 125.57, 121.54, 120.23, 113.15, 67.77, 62.57, 32.27,

32.03, 28.40, 25.13, 24.88. HPLC purity: 98.5%, $t_R = 7.562$ min. HRMS (ESI): calcd for $[C_{21}H_{23}BrN_2O_4S + Na]^+$ 501.0454, found 501.0439.

7-{2-[2-(3-bromophenyl)-4-oxo-3-thiazolidinyl]phenoxy}-N-hydroxyheptanamide

(12d) Compound 12d (24.1% yield) was prepared according to the procedure described for the preparation of compound 12a. ¹H NMR (DMSO-*d*₆, 300 MHz): δ 10.36 (br s, 1H), 8.67 (br s, 1H), 7.63 (s, 1H), 7.45–7.38 (m, 2H), 7.25–7.17 (m, 2H), 7.02 (d, *J* = 7.8 Hz, 2H), 6.83 (dd, *J* = 7.2, 7.2 Hz, 1H), 6.17 (s, 1H), 4.05 (d, *J* = 15.6 Hz, 1H), 4.02–3.97 (m, 2H), 3.80 (d, *J* = 15.6 Hz, 1H), 2.01–1.96 (m, 2H), 1.78 (t, *J* = 7.2 Hz, 2H), 1.55 (t, *J* = 7.2 Hz, 2H), 1.50–1.47 (m, 2H), 1.42–1.38 (m, 2H). ¹³C NMR (125 MHz, DMSO-*d*₆) δ 170.34, 169.11, 154.17, 142.65, 131.48, 130.58, 129.99, 129.36, 129.32, 126.49, 125.56, 121.56, 120.22, 113.12, 67.81, 62.57, 32.26, 32.06, 28.54, 28.34, 25.20, 25.15. HPLC purity: 95.0%, *t*_R = 9.573 min. HRMS (ESI): calcd for [C₂₂H₂₅BrN₂O₄S + Na]⁺ 515.0611, found 515.0597.

8-{2-[2-(3-bromophenyl)-4-oxo-3-thiazolidinyl]phenoxy}-N-hydroxyoctanamide (**12e**) Compound **12e** (65.2% yield) was prepared according to the procedure described for the preparation of compound **12a.** ¹H NMR (CDCl₃, 300 MHz): δ 7.52 (s, 1H), 7.37 (d, J = 7.5 Hz, 1H), 7.24–7.18 (m, 2H), 7.13 (dd, J = 7.5, 7.5 Hz, 1H), 6.87 (d, J = 7.8 Hz, 2H), 6.81 (dd, J = 7.5, 7.5 Hz, 1H), 5.97 (s, 1H), 4.04–3.92 (m, 4H), 2.15-2.11 (m, 2H), 1.85–1.81 (m, 2H), 1.66– 1.63 (m, 4H), 1.51–1.48 (m, 2H), 1.38–1.34 (m, 2H). ¹³C NMR (125 MHz, DMSO- d_6) δ 170.32, 169.09, 154.18, 142.64, 131.48, 130.58, 129.98, 129.37, 129.31, 126.48, 125.54, 121.55, 120.19, 113.10, 67.85, 62.55, 32.25, 32.07, 28.63, 28.59, 28.47, 25.37, 25.05. HPLC purity: 95.0%, $t_R = 8.642$ min. HRMS (ESI): calcd for $[C_{23}H_{27}BrN_2O_4S + Na]^+$ 529.0767, found 529.0743.

4-[2-[2-(3-(4-chlorophenylethyl amino carbonyl)phenyl)-4-oxo-3-thiazolidinyl] phenoxy]-N-hydroxybutyramide (**12f**) Compound **12f** (16.1% yield) was prepared according to the procedure described for the preparation of compound **12a.** ¹H NMR (DMSO-*d*₆, 300 MHz): δ 10.45 (br s, 1H), 8.75 (br s, 1H), 8.57 (t, *J* = 5.4 Hz, 1H), 7.85 (s, 1H), 7.67 (d, *J* = 7.8 Hz, 1H), 7.53 (d, *J* = 7.8 Hz, 1H), 7.38-7.35 (m, 1H), 7.33 (d, *J* = 8.1 Hz, 2H), 7.26-7.23 (m, 2H), 7.18 (dd, *J* = 7.8, 7.5 Hz, 1H), 6.96 (dd, *J* = 7.5, 7.5 Hz, 2H), 6.78 (dd, *J* = 7.5, 7.5 Hz, 1H), 6.19 (s, 1H), 4.05 (d, *J* = 15.6 Hz, 1H), 3.99–3.92 (m, 2H), 3.87 (d, *J* = 15.6 Hz, 1H), 3.46–3.42 (m, 2H), 2.82 (t, *J* = 7.5 Hz, 2H), 1.29–1.24 (m, 2H). ¹³C NMR (125 MHz, DMSO-*d*₆) δ 170.83, 168.73, 165.57, 154.06, 140.22, 138.54, 134.66, 130.73, 130.56, 130.23, 129.30, 128.43, 128.20, 127.39, 126.09, 125.62, 120.28, 113.05, 67.33, 63.14, 40.60, 34.27, 32.12, 30.67, 28.78, 24.87. HPLC purity: 95.9%, *t*_R = 9.445 min. HRMS (ESI): calcd for [C₂₈H₂₈ClN₃O₅S + Na]⁺ 576.1330, found 576.1346.

4-{2-[2-(5-bromo-2-thiopheneyl)-4-oxo-3-thiazolidinyl]phenoxy]-N-hydroxybutyramide (**12g**) Compound **12g** (22.5% yield) was prepared according to the procedure described for the preparation of compound **12a.** ¹H NMR (DMSO-*d*₆, 300 MHz): δ 10.42 (br s, 1H), 8.73 (br s, 1H), 7.26 (dd, *J* = 7.5, 7.5 Hz, 1H), 7.05 (d, *J* = 7.8 Hz, 1H), 6.99–6.96 (m, 2H), 6.89 (dd, *J* = 7.5, 7.5 Hz, 1H), 6.81–6.79 (m, 1H), 6.41 (s, 1H), 4.01 (d, *J* = 15.3 Hz, 1H), 3.96–3.93 (m, 2H), 3.88 (d, *J* = 15.3 Hz, 1H), 2.15 (t, *J* = 7.5 Hz, 2H), 1.94–1.90 (m, 2H). ¹³C NMR (125 MHz, DMSO-*d*₆) δ 169.96, 168.66, 154.08, 145.74, 130.06, 129.77, 129.59, 128.65, 125.09, 120.39, 113.14, 112.99, 67.34, 58.83, 32.02, 28.78, 24.82. HPLC purity: 96.2%, $t_{\rm R} = 6.843$ min. HRMS (ESI): calcd for $[C_{17}H_{17}BrN_2O_4S_2 + Na]^+ 478.9705$, found 478.9744.

4-[2-[2-(5-(5-pyrimidinyl)-2-thiopheneyl)-4-oxo-3-thiazolidinyl]phenoxy]-N-hydroxy -butyramide (**12h**) Compound **12h** (19.9% yield) was prepared according to the procedure described for the preparation of compound **12a.** ¹H NMR (DMSO-*d*₆, 300 MHz): δ 10.47 (br s, 1H), 9.10 (s, 1H), 9.06 (s, 2H), 8.73 (br s, 1H), 7.48 (d, J = 3.6Hz, 1H), 7.25 (dd, J = 7.2, 7.2 Hz, 1H), 7.07 (d, J = 3.6 Hz, 1H), 7.03 (d, J = 7.8 Hz, 1H), 6.85 (dd, J = 7.5, 7.5 Hz, 1H), 6.65–6.62 (m, 1H), 6.48 (s, 1H), 4.02 (d, J = 15.3Hz, 1H), 3.97–3.93 (m, 2H), 3.90 (d, J = 15.3 Hz, 1H), 2.20–2.13 (m, 2H), 1.98–1.93 (m, 2H). ¹³C NMR (125 MHz, DMSO-*d*₆) δ 170.16, 168.70, 157.20, 154.10, 153.01, 146.04, 136.83, 130.16, 129.59, 129.11, 127.65, 125.62, 125.22, 120.40, 113.02, 67.35, 58.86, 32.04, 28.78, 24.85. HPLC purity: 96.2%, *t*_R = 4.020 min. HRMS (ESI): calcd for [C₂₁H₂₀N₄O₄S₂ + Na]⁺ 479.0818, found 479.0831.

4-{4-[2-(5-bromo-2-thiopheneyl)-4-oxo-3-thiazolidinyl]phenoxy}-N-hydroxybutyr-

amide (**12i**) Compound **12i** (27.2 yield) was prepared according to the procedure described for the preparation of compound **12a.** ¹H NMR (DMSO-*d*₆, 300 MHz): δ 10.39 (br s, 1H), 8.68 (br s, 1H), 7.15 (d, *J* = 9.0 Hz, 2H), 6.95 (d, *J* = 3.6 Hz, 1H), 6.90-6.88 (m, 2H), 6.87 (d, *J* = 3.6 Hz, 1H), 6.65 (s, 1H), 3.97–3.92 (m, 3H), 3.85 (d, *J* = 15.9 Hz, 1H), 2.10 (t, *J* = 7.5 Hz, 2H), 1.92–1.87 (m, 2H). ¹³C NMR (125 MHz, DMSO-*d*₆) δ 153.37, 152.49, 142.54, 132.86, 118.59, 118.41, 117.54, 116.65, 105.17, 103.86, 63.49, 57.10, 33.35, 30.03, 26.58. HPLC purity: 96.4%, *t*_R = 7.964 min. HRMS (ESI): calcd for [C₁₇H₁₇BrN₂O₄S₂ + Na]⁺ 478.9705, found 478.9708.

4-{4-[2-(5-(5-pyrimidinyl)-2-thiopheneyl)-4-oxo-3-thiazolidinyl]phenoxy}-N-hydroxy-

butyramide (**12j**) Compound **12j** (35.1% yield) was prepared according to the procedure described for the preparation of compound **12a.** 1H NMR (DMSO-*d*₆, 300 MHz): 10.39 (br s, 1H), 9.10 (s, 1H), 9.06 (s, 2H), 8.68 (br s, 1H), 7.48 (d, J = 3.6 Hz, 1H), 7.25 (d, J = 7.2 Hz, 2H), 7.07 (d, J = 3.6 Hz, 1H), 7.03 (d, J = 7.8 Hz, 2H), 6.75 (s, 1H), 4.01 (d, J = 15.3 Hz, 1H), 3.96–3.93 (m, 2H), 3.90 (d, J = 15.3 Hz, 1H), 2.21–2.08 (m, 2H), 1.90–1.88 (m, 2H). ¹³C NMR (125 MHz, DMSO-*d*₆) δ 169.70, 168.56, 157.19, 153.02, 146.42, 136.72, 129.78, 129.07, 127.59, 125.70, 114.53, 66.90, 59.66, 32.53, 28.65, 24.73. HPLC purity: 98.7%, *t*_R = 3.788 min. HRMS (ESI): calcd for [C₂₁H₂₀N₄O₄S₂ + Na]⁺ 479.0818, found 479.0828.

 N^{1} -(2-(2-(3-bromophenyl)-4-oxo-3-thiazolidinyl)phenyl)- N^{7} -hydroxyheptanediamide

(13a) Compound 13a (25.1% yield) was prepared according to the procedure described for the preparation of compound 10a. ¹H NMR (CDCl₃, 300 MHz): δ 7.74 (br s, 1H) 7.49 (s, 1H), 7,36–7.35 (m, 1H), 7.23–7.22 (m, 2H), 7.16–7.01 (m, 3H), 6.00 (s, 1H), 4.06 (d, *J* = 15.0 Hz, 1H), 3.93 (d, *J* = 15.0 Hz, 1H), 2.46–2.33 (m, 2H), 2.19–2.06 (m, 2H), 1.69–1.68 (m, 4H), 1.48–1.37 (m, 2H). ¹³C NMR (125 MHz, DMSO-*d*₆) δ 171.35, 171.26, 169.02, 141.94, 135.09, 131.69, 130.45, 126.89, 124.50, 121.56, 35.84, 32.79, 32.17, 28.32, 25.01, 24.83. HPLC purity: 96.6%, *t*_R = 5.787 min. HRMS (ESI): calcd for [C₂₂H₂₄BrN₃O₄S + Na]⁺ 528.0563, found 528.0575.

 N^{1} -(2-(2-(3-bromophenyl)-4-oxo-3-thiazolidinyl)phenyl)- N^{8} -hydroxyoctanediamide

(13b) Compound 13b (35.1% yield) was prepared according to the procedure described for the preparation of compound 10a. ¹H NMR (CDCl₃, 300 MHz): δ 9.33 (br s, 1H), 7.76–7.70 (m, 2H), 7.47 (s, 1H), 7.41–7.7.34 (m, 1H), 7.30–7.23 (m, 2H),

7.16–7.05 (m, 3H), 6.04 (s, 1H), 4.06 (d, J = 15.0 Hz, 1H), 3.92 (d, J = 15.0 Hz, 1H), 2.32 (t, J = 7.2 Hz, 2H), 2.17 (t, J = 7.2 Hz, 2H), 1.80–1.60 (m, 4H), 1.48–1.30 (m, 4H). ¹³C NMR (125 MHz, DMSO- d_6) δ 171.39, 171.27, 169.08, 141.94, 135.08, 131.70, 130.52, 130.44, 128.03, 126.88, 124.54, 121.57, 79.16, 62.48, 35.94, 32.81, 32.26, 28.49, 25.07, 25.01. HPLC purity: 97.2%, $t_R = 6.285$ min. HR MS (ESI): calcd for [C₂₃H₂₆BrN₃O₄S + Na]⁺ 542.0720, found 542.0732.

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Analysis Info

Acquisition Date 4/29/2015 10:32:35 AM D:\Data\waixi\chenyihua\20150429LIYUNQI\YF-415_P1-D-1_01_1513.d Analysis Name Tune_pos_low_LC with calibration_2min.m ECNU-Chem Method Operator YF-415 Sample Name Instrument maXis impact 282001.00122 Comment

Acquisition Parameter



Page 1 of 1





Analysis Info Acquisition Date 4/29/2015 10:35:43 AM Analysis Name D:\Data\waixi\chenyihua\20150429LIYUNQI\YF-457_P1-D-2_01_1514.d Method Tune_pos_low_LC with calibration_2min.m Operator ECNU-Chem Sample Name YF-457 Instrument maXis impact 282001.00122 Comment Acquisition Parameter







Analysis InfoAcquisition Date4/29/2015 10:38:50 AMAnalysis NameD:\Data\waixi\chenyihua\20150429LIYUNQI\WJ-429_P1-D-3_01_1515.dTune_pos_low_LC with calibration_2min.mOperatorECNU-ChemSample NameWJ-429InstrumentmaXis impact 282001.00122CommentCommentCommentComment

Acquisition Parameter



Page 1 of 1





Analysis Info

Acquisition Date 4/29/2015 10:41:56 AM D:\Data\waixi\chenyihua\20150429LIYUNQI\YF-457A_P1-D-4_01_1516.d Analysis Name Tune_pos_low_LC with calibration_2min.m ECNU-Chem Method Operator YF-457A Sample Name Instrument maXis impact 282001.00122 Comment







Analysis Info

Acquisition Date 4/29/2015 10:45:04 AM D:\Data\waixi\chenyihua\20150429LIYUNQI\LW-451_P1-D-5_01_1517.d Analysis Name Tune_pos_low_LC with calibration_2min.m Method Operator **ECNU-Chem** Sample Name LW-451 Instrument maXis impact 282001.00122 Comment







Analysis Info Acquisition Analysis Name D:\Data\waixi\chenyihua\20150429LIYUNQI\YF-465A_P1-D-6_01_1518.d Method Tune_pos_low_LC with calibration_2min.m Operator

YF-465A

Acquisition Date 4/29/2015 10:48:13 AM

Operator Instrument

ECNU-Chem maXis impact 282001.00122

Method Sample Name Comment

Source Type Focus Scan Begin Scan End		ESI Active 50 m/z 1200 m/z			lon Polarity Set Capillary Set End Plate Offset Set Collision Cell RF			Positive 3700 V -500 V 500.0 Vpp			Set Nebulizer Set Dry Heater Set Dry Gas Set Divert Valve			1.5 Bar 180 °C 6.0 l/min Waste	
Intens x10 1.2	s. .6 .5						- 487.0293	489.027							
0.7	0						0319	0298 63							
0.2 0.0	0 465	470) 47	5 48	30	485	488	490	1	495		00	505	510 m/z	
#	m/z	Res.	S/N		1%	FWHN	Л							010 11#2	
1	487.0293	34765	16144.2	1046488	90.3	0.014	0								
2	488.0319	23964	3073.2	198943	17.2	0.020	4								
3	489.0274	35934	17915.0	1158641	100.0	0.013	6								
4	490.0298	23353	3031.7	195895	16.9	0.021	0								
5	491.0263	16142	788.6	50895	4.4	0.030	4								
Meas. 487.0	m/z # 0293 1	Ion Forn C20H21E	nula BrN2NaO4S	m/: 487.029	z err 8	[ppm] 0.9	mSig	gma 40.4	2	Score 63.24	rdb 10.5	e⁻ Con even	if N-Rul o	e k	





Analysis Info Acquisition Date 4/29/2015 10:51:22 AM D:\Data\waixi\chenyihua\20150429LIYUNQI\LW-479_P1-D-7_01_1519.d Analysis Name Tune_pos_low_LC with calibration_2min.m Method Operator **ECNU-Chem** Sample Name LW-479 Instrument maXis impact 282001.00122 Comment

Acquisition Parameter Source Type ESI Ion Polarity Positive Set Nebulizer 1.5 Bar Focus Active Set Capillary 3700 V Set Dry Heater 180 °C -500 V Scan Begin Set End Plate Offset 6.0 l/min 50 m/z Set Dry Gas Scan End 1200 m/z Set Collision Cell RF 500.0 Vpp Set Divert Valve Waste Intens. 503.0420 <u>501.0439</u> x10⁵ 1.25 1.00 0.75 <u>502.0465</u> 504.0445 0.50 0.25 0.00 480 485 490 495 500 505 510 515 m/z FWHM # m/z Res. S/N I 1% 501.0439 24334 977.5 96047 94.1 0.0206 1 2 502.0465 17668 179.7 17713 17.4 0.0284 503.0420 24536 102064 0.0205 3 1033.1 100.0 17506 4 504.0445 176.6 17515 17.2 0.0288 Meas. m/z # Ion Formula err [ppm] mSigma Score rdb e⁻ Conf N-Rule m/z C21H23BrN2NaO4S 501.0439 1 501.0454 3.0 40.3 3 38.62 10.5 even ok

S36





Analysis Info Acquisition Date 4/29/2015 10:54:30 AM D:\Data\waixi\chenyihua\20150429LIYUNQI\YF-493_P1-D-8_01_1520.d Analysis Name Method Tune_pos_low_LC with calibration_2min.m Operator ECNU-Chem YF-493 Instrument Sample Name maXis impact 282001.00122 Comment

Positive

Set Nebulizer

1.5 Bar

Acquisition Parameter Source Type ESI Ion Polarity Focus Scan Begin Active Set Capillary Scan

us n Begi n End	n	- - 5 1	Active 50 m/z 1200 m/z		Set Ca Set Er Set Co	apillary nd Plate ollision C	Offset Cell RF	3700 V -500 V 500.0 Vp	þ	Set Dry Heater Set Dry Gas Set Divert Valve	180 °C 6.0 I/min Waste	
I	nten x10	s.)6						5.0597				
	0.	.8-										
	0.	.6-										
	0.	.4-						6.0623	3.0603 374			
	0.	.2-						- 51				
	0.	.0 490	495	500	505		510	515	520	525 530	, † , , , , , , 535 m/z	z
	#	m/z	Res.	S/N	I	۱%	FWHM					
-	1	515.0597	34654	8666.0	763020	96.4	0.0149					
	2	516.0623	23747	1881.7	165565	20.9	0.0217					
	3	517.0578	34510	9007.7	791772	100.0	0.0150					
	4	518.0603	22676	1710.8	150258	19.0	0.0228					
	5	519.0574	15492	438.8	38520	4.9	0.0335					

Meas. m/z	#	Ion Formula	m/z	err [ppm]	mSigma		Score	rdb	e [−] Conf	N-Rule
515.0597	1	C22H25BrN2NaO4S	515.0611	2.6	32.6	1	58.76	10.5	even	ok





Analysis Info Acquisition Date 4/29/2015 10:57:38 AM Analysis Name D:\Data\waixi\chenyihua\20150429LIYUNQI\LW-507_P1-D-9_01_1521.d Method Tune_pos_low_LC with calibration_2min.m Operator ECNU-Chem Sample Name LW-507 Instrument maXis impact 282001.00122

Acquisition Parameter Source Type ESI Ion Polarity Positive Set Nebulizer 1.5 Bar Focus Active Set Capillary 3700 V Set Dry Heater 180 °C Set End Plate Offset -500 V 6.0 l/min Scan Begin 50 m/z Set Dry Gas Scan End 1200 m/z Set Collision Cell RF 500.0 Vpp Set Divert Valve Waste Intens. 529.0743 531.072 x10⁶ 1.0 0.8 0.6 530.0769 0.4 0.2 0.0 505 510 515 520 525 530 535 540 545 550 m/z FWHM # m/z Res. S/N I 1% 529.0743 34567 11298.0 884953 95.5 0.0153 1 2 530.0769 23614 2358.8 184629 19.9 0.0224 531.0725 34565 926460 0.0154 3 11836.9 100.0 4 532.0750 23353 2317.4 181253 19.6 0.0228 5 533.0723 15903 580.7 45419 4.9 0.0335 Meas. m/z # Ion Formula rdb e⁻ Conf N-Rule m/z err [ppm] mSigma Score C23H27BrN2NaO4S 529.0743 1 529.0767 4.6 38.0 1 100.00 10.5 even ok





Analysis InfoAcquisition Date4/29/2015 1:57:28 PMAnalysis NameD:\Data\waixi\chenyihua\20150429LIYUNQI\YF-554_P1-E-2_01_1525.dMethodTune_pos_low_LC with calibration_2min.mOperatorECNU-ChemSample NameYF-554InstrumentmaXis impact 282001.00122CommentCommentCommentComment







Analysis Info

Acquisition Date 4/29/2015 2:09:58 PM D:\Data\waixi\chenyihua\20150429LIYUNQI\YF-457B_P1-E-6_01_1529.d Analysis Name Tune_pos_low_LC with calibration_2min.m Method Operator **ECNU-Chem** YF-457B Sample Name Instrument maXis impact 282001.00122 Comment







Analysis InfoAcquisition Date4/29/2015 2:58:15 PMAnalysis NameD:\Data\waixi\chenyihua\20150429LIYUNQI\YF-456A-2_P1-E-7_01_1535.dTune_pos_low_LC with calibration_2min.mOperatorECNU-ChemSample NameYF-456A-2InstrumentmaXis impact 282001.00122CommentCommentCommentCommentComment

Acquisition Parameter



Page 1 of 1





Analysis Info

Acquisition Date 4/29/2015 3:01:23 PM Analysis Name D:\Data\waixi\chenyihua\20150429LIYUNQI\YF-457C-2_P1-E-8_01_1536.d Tune_pos_low_LC with calibration_2min.m Method Operator **ECNU-Chem** YF-457C-2 Sample Name Instrument maXis impact 282001.00122 Comment

Analysis Info Acquisition Date 4/29/2015 3:04:30 PM Analysis Name D:\Data\waixi\chenyihua\20150429LIYUNQI\YF-456B-2_P1-E-9_01_1537.d Method Tune_pos_low_LC with calibration_2min.m Operator ECNU-Chem Sample Name YF-456B-2 Instrument maXis impact 282001.00122

Acquisition Parameter Source Type ESI Ion Polarity Positive Set Nebulizer 1.5 Bar Focus Active Set Capillary 3700 V Set Dry Heater 180 °C Scan Begin Set End Plate Offset -500 V 6.0 l/min 50 m/z Set Dry Gas Scan End 1200 m/z Set Collision Cell RF 500.0 Vpp Set Divert Valve Waste Intens. 479.082 x10⁶ 1.25 1.00 0.75 0.50 0.25 0.00] 455 485 460 465 470 475 490 495 480 FWHM # m/z Res. S/N I ۱% 479.0828 34779 10985.6 1167102 100.0 0.0138 1 2 480.0850 23255 2208.7 234785 20.1 0.0206 3 481.0806 17680 967.2 102877 8.8 0.0272 mSigma N-Rule Meas. m/z # Ion Formula m/z err [ppm] Score rdb e⁻ Conf C21H20N4NaO4S2 33.2 479.0828 1 479.0818 -2.0 1 100.00 13.5 even ok

m/z

Analysis Info

Acquisition Date 4/29/2015 3:07:37 PM D:\Data\waixi\chenyihua\20150429LIYUNQI\WJ-506-2_P1-F-1_01_1538.d Analysis Name Tune_pos_low_LC with calibration_2min.m Method Operator **ECNU-Chem** Sample Name WJ-506-2 Instrument maXis impact 282001.00122 Comment

Analysis Info Acquisition Date 4/29/2015 3:10:45 PM Analysis Name D:\Data\waixi\chenyihua\20150429LIYUNQI\WJ-520-2_P1-F-2_01_1539.d Tune_pos_low_LC with calibration_2min.m Operator ECNU-Chem Method WJ-520-2 Instrument maXis impact 282001.00122 Comment VI Sample Name D:\Data\waixi\chenyihua\20150429LIYUNQI\WJ-520-2_P1-F-2_01_1539.d

Acquisition Parameter Source Type ESI Ion Polarity Positive Set Nebulizer 1.5 Bar Focus Active Set Capillary 3700 V Set Dry Heater 180 °C Set End Plate Offset -500 V Scan Begin 6.0 l/min 50 m/z Set Dry Gas Scan End 1200 m/z Set Collision Cell RF 500.0 Vpp Set Divert Valve Waste Intens. 542.073 544.071 x10⁶ 2.0 1.5 543.0755 1.0 0.5 0.0 520 525 530 535 540 545 550 555 560 m/z FWHM # m/z Res. S/N ۱% I 542.0732 38580 19697.1 1996708 98.3 0.0141 1 2 543.0755 28686 4126.3 417403 20.6 0.0189 2030477 0.0144 3 544.0715 37734 20123.7 100.0 4 545.0736 27734 3956.0 398235 19.6 0.0197 5 546.0711 18836 943.3 94787 4.7 0.0290 Meas. m/z # Ion Formula rdb e⁻ Conf N-Rule m/z err [ppm] mSigma Score C23H26BrN3NaO4S 542.0732 1 542.0720 -2.4 41.9 2 100.00 11.5 even ok