

## Supporting Information for

### **Antimalarial diterpenoid dimers of a new carbon skeleton from *Aphanamixis grandifolia***

Hua Zhang,<sup>a,‡</sup> Jia Liu,<sup>a,‡</sup> Li-She Gan,<sup>b</sup> Seema Dalal,<sup>c</sup> Maria B. Cassera,<sup>c</sup> and Jian-Min Yue<sup>a,\*</sup>

<sup>a</sup>*State Key Laboratory of Drug Research, Shanghai Institute of Materia Medica, Chinese Academy of Sciences, 555 Zu Chong Zhi Road, Zhangjiang Hi-Tech Park, Shanghai 201203, P. R. China*

<sup>b</sup>*Institute of Modern Chinese Medicine, College of Pharmaceutical Sciences, Zhejiang University, Hangzhou 310058, P. R. China*

<sup>c</sup>*Department of Biochemistry and the Virginia Tech Center for Drug Discovery, MC 0308, Virginia Polytechnic Institute and State University, Blacksburg, Virginia 24061, United States*

<sup>‡</sup>*Equal contribution*

\*Corresponding author. E-mail: [jmyue@simm.ac.cn](mailto:jmyue@simm.ac.cn)

## 1. Experimental Section (p5 to p11)

**Figure S1.** Experimental ECD spectra of compounds **4** and **5**.

**Scheme S1.** Oxidative degradation of compounds **4/5**.

**Scheme S2.** Synthesis of (*S*)- (**6**) and (*R*)- (**7**) forms of 6-(hydroxymethyl)-4-methyl-5,6-dihydro-2H- pyran-2-one.

**Figure S2.** Chiral HPLC analysis of **4r/5r** from **4/5** and synthetic samples (**6 & 7**)

### ECD Calculations

**Figure S3.** Calculated ECD spectra of **2** and **3** versus their experimental ECD spectra.

**Figure S4.** Calculated ECD spectra of **4a** and **5a** versus their experimental ECD spectra.

## 2. Tabulated NMR data (p12 to p13)

**Table S1.**  $^1\text{H}$  and  $^{13}\text{C}$  NMR data for compounds **4** and **5** in  $\text{CD}_3\text{OD}$ .

**Table S2.**  $^1\text{H}$  and  $^{13}\text{C}$  NMR data for compounds **4a**, **4b**, **5a**, and **5b** in  $\text{CD}_3\text{OD}$ .

## 3. NMR spectra for all new Natural and synthetic compounds (p14 to p75)

**Figure S5.**  $^1\text{H}$  NMR spectrum for aphadilactone E (**1**) in  $\text{CD}_3\text{OD}$ .

**Figure S6.**  $^{13}\text{C}$  NMR spectrum for aphadilactone E (**1**) in  $\text{CD}_3\text{OD}$ .

**Figure S7.**  $^1\text{H}$ - $^1\text{H}$  COSY spectrum for aphadilactone E (**1**) in  $\text{CD}_3\text{OD}$ .

**Figure S8.** HSQC spectrum for aphadilactone E (**1**) in  $\text{CD}_3\text{OD}$ .

**Figure S9.** HMBC spectrum for aphadilactone E (**1**) in  $\text{CD}_3\text{OD}$ .

**Figure S10.** ROESY spectrum for aphadilactone E (**1**) in  $\text{CD}_3\text{OD}$ .

**Figure S11.** ESI(+) $\text{MS}$  spectrum for aphadilactone E (**1**).

**Figure S12.** HRESI(+) $\text{MS}$  spectrum for aphadilactone E (**1**).

**Figure S13.**  $^1\text{H}$  NMR spectrum for aphadilactone F (**2**) in  $\text{CD}_3\text{OD}$ .

**Figure S14.**  $^{13}\text{C}$  NMR spectrum for aphadilactone F (**2**) in  $\text{CD}_3\text{OD}$ .

**Figure S15.**  $^1\text{H}$ - $^1\text{H}$  COSY spectrum for aphadilactone F (**2**) in  $\text{CD}_3\text{OD}$ .

**Figure S16.** HSQC spectrum for aphadilactone F (**2**) in  $\text{CD}_3\text{OD}$ .

**Figure S17.** HMBC spectrum for aphadilactone F (**2**) in  $\text{CD}_3\text{OD}$ .

**Figure S18.** ROESY spectrum for aphadilactone F (**2**) in  $\text{CD}_3\text{OD}$ .

**Figure S19.** ESI(+) $\text{MS}$  spectrum for aphadilactone F (**2**).

**Figure S20.** HRESI(+) $\text{MS}$  spectrum for aphadilactone F (**2**).

**Figure S21.**  $^1\text{H}$  NMR spectrum for aphadilactone G (**3**) in  $\text{CD}_3\text{OD}$ .

**Figure S22.**  $^{13}\text{C}$  NMR spectrum for aphadilactone G (**3**) in  $\text{CD}_3\text{OD}$ .

**Figure S23.**  $^1\text{H}$ - $^1\text{H}$  COSY spectrum for aphadilactone G (**3**) in  $\text{CD}_3\text{OD}$ .

**Figure S24.** HSQC spectrum for aphadilactone G (**3**) in CD<sub>3</sub>OD.  
**Figure S25.** HMBC spectrum for aphadilactone G (**3**) in CD<sub>3</sub>OD.  
**Figure S26.** ROESY spectrum for aphadilactone G (**3**) in CD<sub>3</sub>OD.  
**Figure S27.** ESI(+)-MS spectrum for aphadilactone G (**3**).  
**Figure S28.** HRESI(+)-MS spectrum for aphadilactone G (**3**).

**Figure S29.** <sup>1</sup>H NMR spectrum for aphanamene H (**4**) in CD<sub>3</sub>OD.  
**Figure S30.** <sup>13</sup>C NMR spectrum for aphanamene H (**4**) in CD<sub>3</sub>OD.  
**Figure S31.** <sup>1</sup>H–<sup>1</sup>H COSY spectrum for aphanamene H (**4**) in CD<sub>3</sub>OD.  
**Figure S32.** HSQC spectrum for aphanamene H (**4**) in CD<sub>3</sub>OD.  
**Figure S33.** HMBC spectrum for aphanamene H (**4**) in CD<sub>3</sub>OD.  
**Figure S34.** ROESY spectrum for aphanamene H (**4**) in CD<sub>3</sub>OD.  
**Figure S35.** ESI(+)-MS spectrum for aphanamene H (**4**).  
**Figure S36.** ESI(–)-MS spectrum for aphanamene H (**4**).  
**Figure S37.** HRESI(+)-MS spectrum for aphanamene H (**4**).

**Figure S38.** <sup>1</sup>H NMR spectrum for aphanamene I (**5**) in CD<sub>3</sub>OD.  
**Figure S39.** <sup>13</sup>C NMR spectrum for aphanamene I (**5**) in CD<sub>3</sub>OD.  
**Figure S40.** <sup>1</sup>H–<sup>1</sup>H COSY spectrum for aphanamene I (**5**) in CD<sub>3</sub>OD.  
**Figure S41.** HSQC spectrum for aphanamene I (**5**) in CD<sub>3</sub>OD.  
**Figure S42.** HMBC spectrum for aphanamene I (**5**) in CD<sub>3</sub>OD.  
**Figure S43.** ROESY spectrum for aphanamene I (**5**) in CD<sub>3</sub>OD.  
**Figure S44.** ESI(+)-MS spectrum for aphanamene I (**5**).  
**Figure S45.** ESI(–)-MS spectrum for aphanamene I (**5**).  
**Figure S46.** HRESI(+)-MS spectrum for aphanamene I (**5**).

**Figure S47.** <sup>1</sup>H NMR spectrum for **4a** in CD<sub>3</sub>OD.  
**Figure S48.** <sup>13</sup>C NMR spectrum for **4a** in CD<sub>3</sub>OD.  
**Figure S49.** HSQC spectrum for **4a** in CD<sub>3</sub>OD.  
**Figure S50.** HMBC spectrum for **4a** in CD<sub>3</sub>OD.  
**Figure S51.** ESI(+)-MS spectrum for **4a**.  
**Figure S52.** HRESI(+)-MS spectrum for **4a**.

**Figure S53.** <sup>1</sup>H NMR spectrum for **4b** in CD<sub>3</sub>OD.  
**Figure S54.** <sup>13</sup>C NMR spectrum for **4b** in CD<sub>3</sub>OD.  
**Figure S55.** NOESY spectrum for **4b** in CD<sub>3</sub>OD.  
**Figure S56.** ESI(+)-MS spectrum for **4b**.  
**Figure S57.** HRESI(+)-MS spectrum for **4b**.

**Figure S58.** <sup>1</sup>H NMR spectrum for **5a** in CD<sub>3</sub>OD.

**Figure S59.**  $^{13}\text{C}$  NMR spectrum for **5a** in  $\text{CD}_3\text{OD}$ .

**Figure S60.** HSQC spectrum for **5a** in  $\text{CD}_3\text{OD}$ .

**Figure S61.** HMBC spectrum for **5a** in  $\text{CD}_3\text{OD}$ .

**Figure S62.** ESI(+)-MS spectrum for **5a**.

**Figure S63.** HRESI(+)-MS spectrum for **5a**.

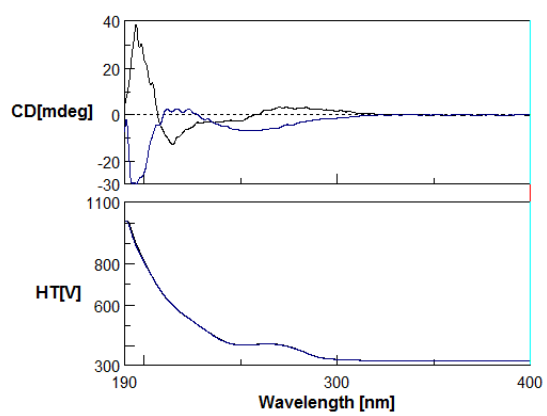
**Figure S64.**  $^1\text{H}$  NMR spectrum for **5b** in  $\text{CD}_3\text{OD}$ .

**Figure S65.**  $^{13}\text{C}$  NMR spectrum for **5b** in  $\text{CD}_3\text{OD}$ .

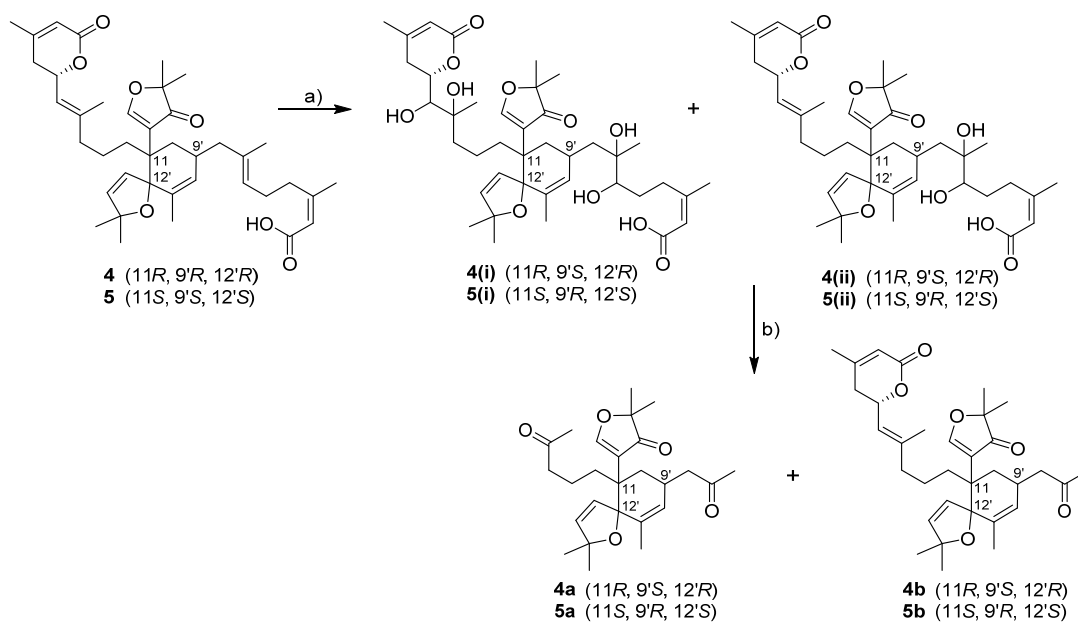
**Figure S66.** NOESY spectrum for **5b** in  $\text{CD}_3\text{OD}$ .

**Figure S67.** ESI(+)-MS spectrum for **5b**.

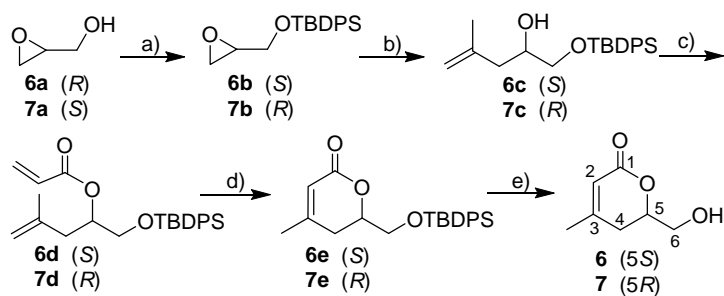
**Figure S68.** HRESI(+)-MS spectrum for **5b**.



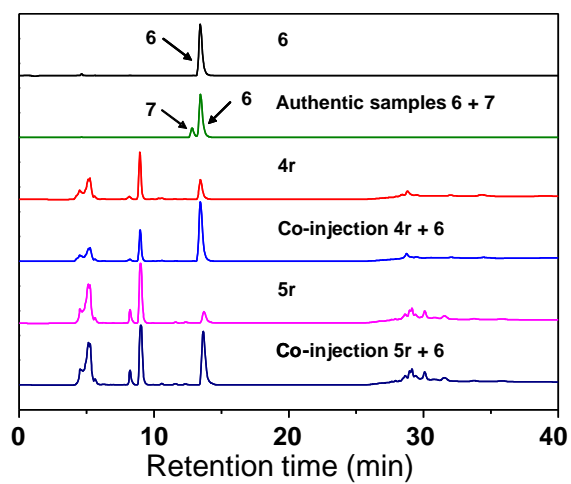
**Figure S1.** Experimental ECD spectra of **4** (black) and **5** (blue).



**Scheme S1.** Oxidative degradation of compounds **4/5** [Reaction conditions: a) 40 mol %  $\text{K}_2\text{OsO}_4 \cdot 2\text{H}_2\text{O}$ , 40 eq.  $\text{MeSO}_2\text{NH}_2$ , 120 eq.  $\text{K}_3\text{Fe}(\text{CN})_6$ , 120 eq.  $\text{K}_2\text{CO}_3$ ,  $^t\text{BuOH-H}_2\text{O}$  (1:1), r.t.; b)  $\text{Pb}(\text{OAc})_4$ , DCM, 0 °C].



**Scheme S2.** Synthesis of (*S*, **6**) and (*R*, **7**) forms of 6-(hydroxymethyl)-4-methyl-5,6-dihydro-2H-pyran-2-one [Reaction conditions: a) TBDPSiCl, imidazole, DMF; b) CuI, CH<sub>3</sub>CH(CH<sub>2</sub>)MgBr, THF, -30 to 0 °C; c) CH<sub>2</sub>CHCOCl, Et<sub>3</sub>N, DMAP, CH<sub>2</sub>Cl<sub>2</sub>, 0 °C; d) Grubbs' catalyst II, CH<sub>2</sub>Cl<sub>2</sub>, 50 °C; e) Bu<sub>4</sub>NF, THF].

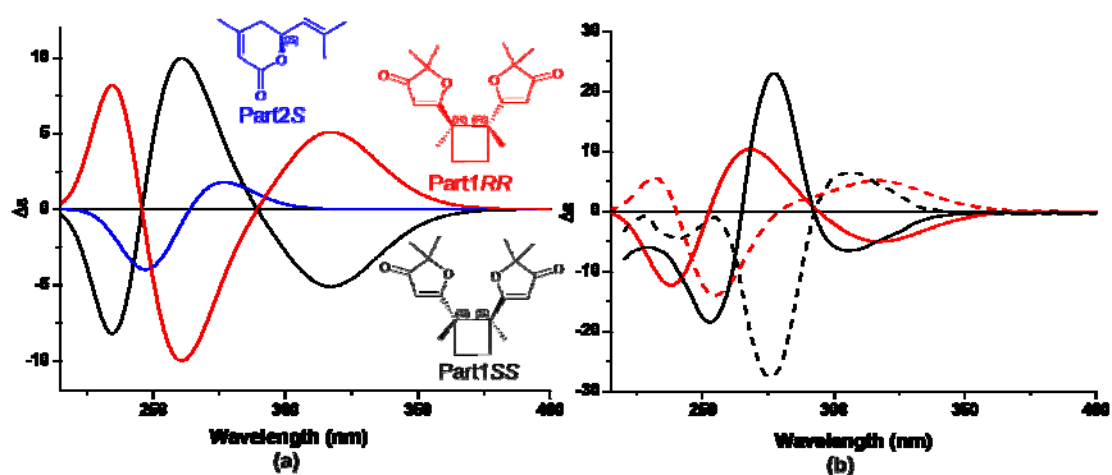


**Figure S2.** Chiral HPLC analysis of **4r/5r** from **4/5** and authentic synthetic samples (**6 & 7**)



## 1.1 ECD Calculations

**1.1.1 Calculated ECD spectra for compounds 2 and 3.** In order to further determine the structures of compounds **2** and **3**, their theoretical ECD spectra were calculated by TDDFT computational chemistry method and compared with the corresponding experimental ones. Firstly, in order to avoid the inaccurate large amounts of lowest energy conformers caused by the flexible chains, ECD spectra of three structural fragments from **2** and **3** (Part1SS, Part1RR, and Part2S, Figure S3(a)) were calculated separately. Linear combination of ECD spectra of Part1SS and two Part2S gave an ECD curve matching the experimental one of **2**, with first negative, second positive, and third negative Cotton effects. Similarly, the calculated ECD curve of Part1RR plus two Part2S could simulate the experimental data of **3**. The above studies allowed us to differentiate the absolute configurations of **2** and **3** as (5*S*,11*S*,11'*S*,5'*S*) and (5*S*,11*R*,11'*R*,5'*S*), respectively.

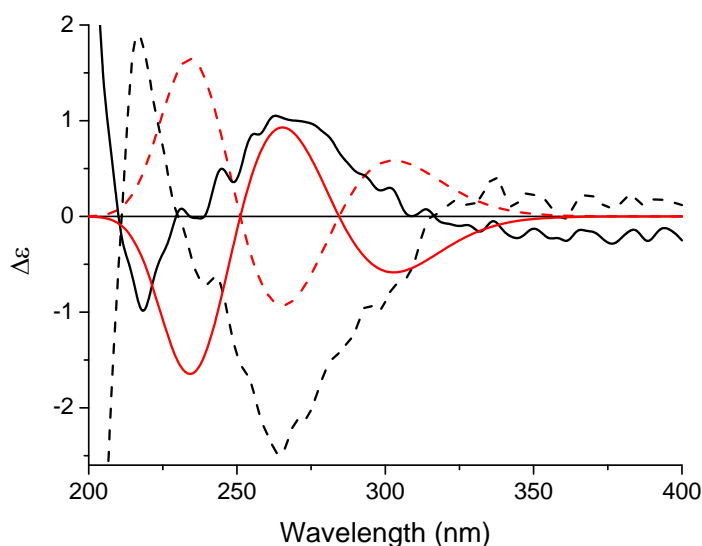


**Figure S3.** (a) B3LYP/6-311++G(2d,2p)//B3LYP/6-31+G(d) calculated ECD spectra for three structural fragments of **2** and **3**; (b) Experimental ECD spectra (220–400 nm) of **2** (black solid line) and **3** (black dashed line), and linear combination of (Part1SS + 2\*Part2S) (red solid line) and (Part1RR + 2\*Part2S) (red dashed line).

In general, conformational analyses were carried out via Monte Carlo searching using molecular mechanism with MMFF94 force field in the SPARTAN 08 software package.<sup>1</sup> The results showed three lowest energy conformers for Part1SS and only one for Part2S with relative energy below 2.0 kcal/mol. Subsequently, the conformers were re-optimized using DFT at the B3LYP/6-31+G(d) level in gas phase by the GAUSSIAN 09 program.<sup>2</sup> The B3LYP/6-31+G(d) harmonic vibrational frequencies were also calculated to confirm their stability. The energies, oscillator strengths, and rotational strengths (velocity) of the first 60 electronic excitations were calculated using the TDDFT methodology at the B3LYP/6-311++G(2d,2p) level in vacuum. The ECD spectra were simulated by the overlapping Gaussian function (half the bandwidth at 1/e peak height,  $\sigma = 0.3$  eV),<sup>3</sup> and the first seven electronic excitations for Part1SS and the first two electronic excitations for Part2S were adopted. To get the final spectra, the simulated spectra of the lowest energy conformers for each structure were averaged according to the Boltzmann distribution theory and their relative Gibbs free energy ( $\Delta G$ ). Theoretical ECD spectrum of Part1RR was obtained by directly inverting that of the

corresponding enantiomer Part1SS.

**1.1.2. Calculated ECD spectra for compounds 4a and 5a.** Theoretical ECD spectra of compounds **4a** and **5a** were also calculated using procedures same as those for **2** and **3**. In brief, conformational analyses of **5a** showed 10 lowest energy conformers with relative energy below 2.0 kcal/mol. The ECD spectra were simulated by the overlapping Gaussian function ( $\sigma = 0.3$  eV),<sup>3</sup> and the velocity rotatory strengths of the first four electronic excitations were adopted. In order to get the final ECD spectrum of **2a**, the simulated spectra of the 10 lowest energy conformers were averaged according to the Boltzmann distribution theory and their relative Gibbs free energy ( $\Delta G$ ). The theoretical ECD spectrum of **4a** was depicted by directly reversing that of **5a**. The results showed that the ECD spectrum of **4a** matched that of the enantiomer with (11*R*,9'*S*,12'*R*) configuration, and the ECD spectrum of **5a** matched that of the other enantiomer with (11*S*,9'*R*,12'*S*) configuration. Therefore, the absolute configurations of aphanamene H (**4**) and I (**5**) were identified to be (5*S*,11*R*,9'*R*,12'*R*) and (5*S*,11*S*,9'*S*,12'*S*), respectively.



**Figure S4.** Calculated (red color) ECD spectra of **4a** (solid line) and **5a** (dashed line) versus their experimental (black color) ECD spectra.

#### Notes and references

[1]. *Spartan 04*; Wavefunction Inc.:Irvine, CA.

[2] Gaussian 09, Rev. C 01. Frisch, M. J.; Trucks, G. W.; Schlegel, H. B.; Scuseria, G. E.; Robb, M. A.; Cheeseman, J. R.; Scalmani, G.; Barone, V.; Mennucci, B.; Petersson, G. A.; Nakatsuji, H.; Caricato, M.; Li, X.; Hratchian, H. P.; Izmaylov, A. F.; Bloino, J.; Zheng, G.; Sonnenberg, J. L.; Hada, M.; Ehara, M.; Toyota, K.; Fukuda, R.; Hasegawa, J.; Ishida, M.; Nakajima, T.; Honda, Y.; Kitao, O.; Nakai, H.; Vreven, T.; Montgomery, Jr., J. A.; Peralta, J. E.; Ogliaro, F.; Bearpark, M.; Heyd, J. J.; Brothers, E.; Kudin, K. N.; Staroverov, V. N.; Kobayashi, R.; Normand, J.; Raghavachari, K.; Rendell, A.; Burant, J. C.; Iyengar, S. S.; Tomasi, J.; Cossi, M.; Rega, N.; Millam, J. M.; Klene, M.; Knox, J. E.; Cross, J. B.; Bakken, V.; Adamo, C.; Jaramillo, J.; Gomperts, R.; Stratmann, R. E.; Yazyev, O.; Austin, A. J.; Cammi, R.; Pomelli, C.; Ochterski, J. W.; Martin, R. L.;

Morokuma, K.; Zakrzewski, V. G.; Voth, G. A.; Salvador, P.; Dannenberg, J. J.; Dapprich, S.; Daniels, A. D.; Farkas, Ö.; Foresman, J. B.; Ortiz, J. V.; Cioslowski, J.; Fox, D. J. Gaussian, Inc., Wallingford CT, 2009.

[3] Stephens, P. J.; Harada, N. ECD cotton effect approximated by the Gaussian curve and other methods. *Chirality* **2010**, 22, 229–233.

**Table S1.**  $^1\text{H}$  and  $^{13}\text{C}$  NMR data of compounds **4** and **5** in  $\text{CD}_3\text{OD}$ .

Position	<b>4</b>		<b>5</b>	
	$\delta_{\text{H}}$ (mult, $J_{\text{HH}}$ )	$\delta_{\text{C}}$	$\delta_{\text{H}}$ (mult, $J_{\text{HH}}$ )	$\delta_{\text{C}}$
1		168.0		168.0
2	5.79 (br s)	116.5	5.79 (br s)	116.5
3		161.0		161.1
4	2.32 (dd, 18.1, 4.5) 2.40 (br dd, 18.1, 10.5)	35.8	2.34 (dd, 18.1, 4.7) 2.40 (br dd, 18.1, 10.4)	35.8
5	5.21 (ddd, 10.5, 8.5, 4.5)	75.9	5.20 (ddd, 10.4, 8.5, 4.7)	75.9
6	5.34 (br d, 8.5)	124.0	5.33 (br d, 8.5)	123.8
7		143.3		143.5
8	2.06 (br t, 7.2, 2H)	40.7	2.06 (br t, 7.3, 2H)	40.7
9	1.24 (m) 1.38 (m)	24.4	1.22 (m) 1.41 (m)	24.6
10	1.62 (ddd, 13.1, 13.1, 4.5) 1.77 (m)	32.4	1.65 (ddd, 13.3, 13.3, 4.7)	32.6
11		51.7		51.8
12		198.3		198.4
13	5.52 (s)	105.9	5.52 (s)	105.9
14		210.2		210.1
15		89.7		89.7
16	1.34 (s, 3H)	23.6	1.34 (s, 3H)	23.6
17	1.34 (s, 3H)	23.2	1.34 (s, 3H)	23.2
18	1.79 (m) 1.89 (dd, 13.4, 11.0)	31.3	1.79 (m) 1.89 (dd, 13.2, 10.8)	31.3
19	1.71 (d, 1.4, 3H)	16.7	1.71 (d, 1.3, 3H)	16.8
20	2.01 (br s, 3H)	23.0	2.01 (br s, 3H)	23.0
1'		169.7		169.7
2'	5.68 (br s)	117.6	5.68 (br s)	117.6
3'		161.5		161.5
4'	2.70 (m, 2H)	34.1	2.70 (m, 2H)	34.1
5'	2.26 (br td, 7.5, 7.0, 2H)	27.7	2.25 (br td, 7.6, 6.9, 2H)	27.7
6'	5.28 (br t, 7.0)	127.2	5.28 (br t, 6.9, 2H)	127.2
7'		134.5		134.5
8'	2.01 (m) 2.16 (m)	46.9	2.01 (dd, 13.2, 8.9) 2.17 (dd, 13.2, 5.9)	46.9
9'	2.26 (m)	31.9	2.27 (m)	31.9
10'	5.50 (br s)	131.1	5.51 (br s)	131.2
11'		135.8		135.7
12'		96.1		96.1
13'	5.86 (d, 6.1)	128.1	5.84 (d, 6.1)	128.1
14'	5.88 (d, 6.1)	137.7	5.88 (d, 6.1)	137.6
15'		88.5		88.5
16'	1.14 (s, 3H)	29.7	1.15 (s, 3H)	29.7
17'	1.33 (s, 3H)	29.2	1.33 (s, 3H)	29.2
18'	1.68 (dd, 2.4, 1.3, 3H)	21.6	1.68 (dd, 2.5, 1.4, 3H)	21.5
19'	1.67 (br s, 3H)	16.1	1.67 (br s, 3H)	16.0
20'	1.92 (d, 1.4, 3H)	25.4	1.92 (d, 1.3, 3H)	25.4

**Table S2.**  $^1\text{H}$  and  $^{13}\text{C}$  NMR data for compounds **4a**, **4b**, **5a**, and **5b** in  $\text{CD}_3\text{OD}$ .

Position	4a/5a		4b		5b	
	$\delta_{\text{H}}$ (mult, $J$ in Hz)	$\delta_{\text{C}}$	$\delta_{\text{H}}$ (mult, $J$ in Hz)	$\delta_{\text{C}}$	$\delta_{\text{H}}$ (mult, $J$ in Hz)	$\delta_{\text{C}}^b$
1				168.1		168.0
2			5.79 (br s)	116.5	5.79 (br s)	116.5
3				161.2		161.1
4			2.37 (m)	35.8	2.34 (dd, 18.1, 4.5)	35.8
			2.42 (m)		2.43 (br dd, 18.1, 10.6)	
5			5.22 (m)	76.0	5.20 (m)	75.9
6			5.36 (br d, 8.6)	124.1	5.34 (br d, 8.5)	123.8
7		211.2		143.2		143.7
8	2.50 (m, 2H)	44.3	2.08 (br t, 7.2, 2H)	40.6	2.07 (br t, 7.2, 2H)	40.7
9	1.67 (m, 2H)	20.6	1.24 (m)	24.1	1.22 (m)	24.3
			1.53 (m)		1.57 (m)	
10	1.68 (m)	32.3	1.66 (m)	32.1	1.68 (m)	32.4
	1.76 (ddd, 13.2, 13.2, 3.8)		1.78 (ddd, 13.1, 13.1, 4.0)		1.78 (13.0, 13.0, 3.9)	
11		51.8		51.7		51.7
12		197.8		198.0		198.0
13	5.53 (s)	106.0	5.52 (s)	105.9	5.52 (s)	105.9
14		210.2		210.3		210.1
15		89.8		89.7		89.7
16	1.34 (s, 3H)	23.5	1.34 (s, 3H)	23.6	1.34 (s, 3H)	23.6
17	1.34 (s, 3H)	23.2	1.34 (s, 3H)	23.2	1.33 (s, 3H)	23.2
18	1.90 (dd, 13.4, 10.7)	30.9	1.88 (m)	30.9	1.88 (dd, 13.3, 9.9)	31.0
	1.95 (br dd, 13.4, 6.0)		1.93 (m)		1.94 (br dd, 13.3, 5.4)	
19	2.12 (s, 3H)	29.8	1.72 (d, 1.4, 3H)	16.7	1.74 (d, 1.3, 3H)	16.8
20			2.03 (br s, 3H)	23.0	2.02 (br s, 3H)	23.0
7'		210.5		210.3		210.3
8'	2.57 (dd, 17.3, 7.9)	49.8	2.58 (m)	49.3	2.57 (dd, 18.8, 9.8)	49.4
	2.64 (dd, 17.3, 6.1)		2.65 (m)		2.66 (dd, 18.8, 5.3)	
9'	2.73 (m)	29.8	2.62 (m)	29.7	2.65 (m)	29.8
10'	5.47 (m)	130.3	5.46 (br s)	130.3	5.47 (br s)	130.4
11'		136.4		136.5		136.5
12'		95.7		95.8		95.8
13'	5.85 (d, 6.1)	128.0	5.87 (d, 6.1)	128.0	5.85 (d, 6.1)	128.0
14'	5.88 (d, 6.1)	137.8	5.89 (d, 6.1)	137.8	5.89 (d, 6.1)	137.8
15'		88.5		88.5		88.5
16'	1.13 (s, 3H)	29.7	1.13 (s, 3H)	29.7	1.13 (s, 3H)	29.7
17'	1.31 (s, 3H)	29.2	1.32 (s, 3H)	29.2	1.32 (s, 3H)	29.2
18'	1.67 (dd, 2.4, 1.4)	21.6	1.68 (br s, 3H)	21.6	1.68 (br s, 3H)	21.6
19'	2.18 (s, 3H)	30.3	2.17 (s, 3H)	30.5	2.17 (s, 3H)	30.4

Figure S5.  $^1\text{H}$  NMR spectrum for aphadilactone E (1) in  $\text{CD}_3\text{OD}$ .

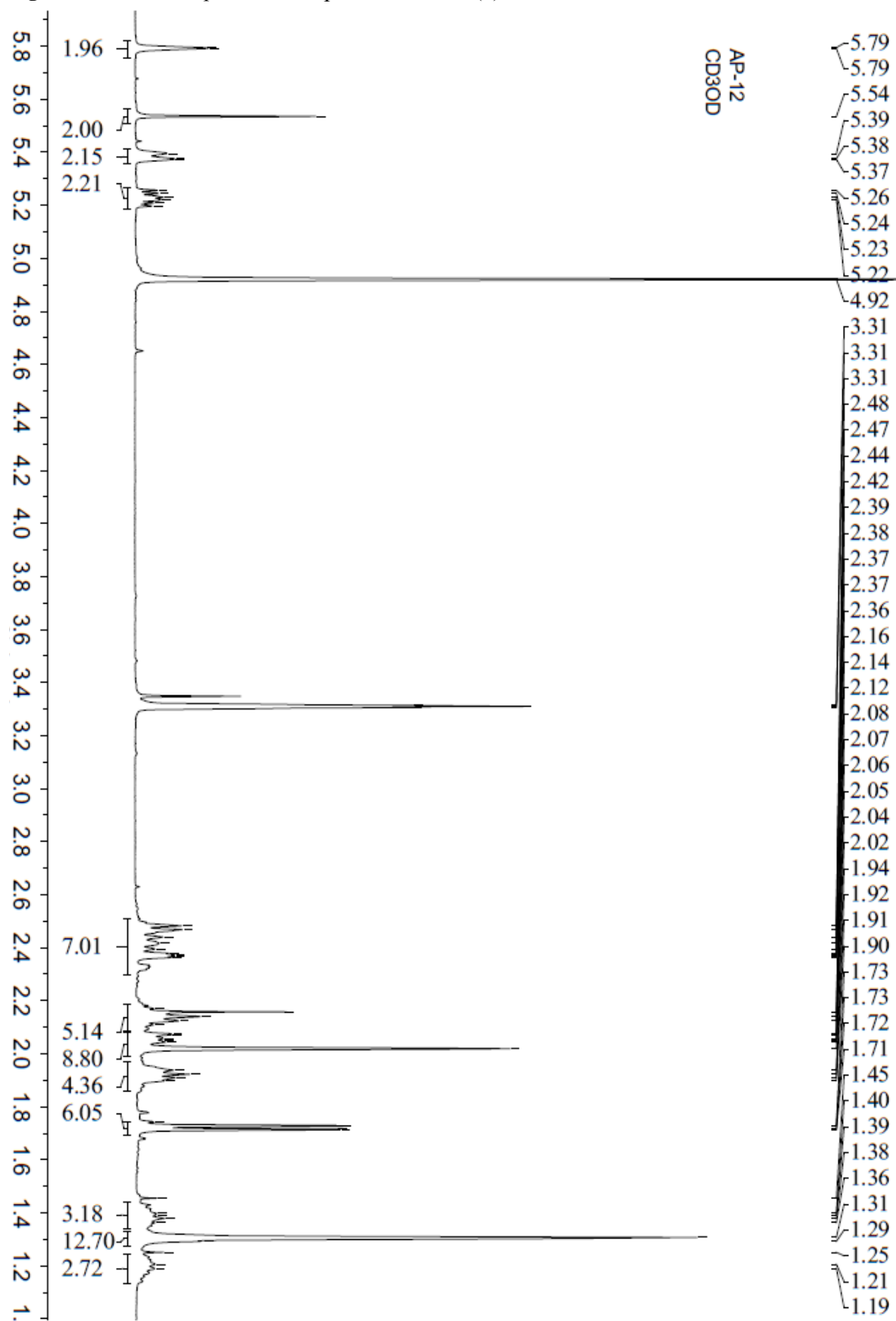
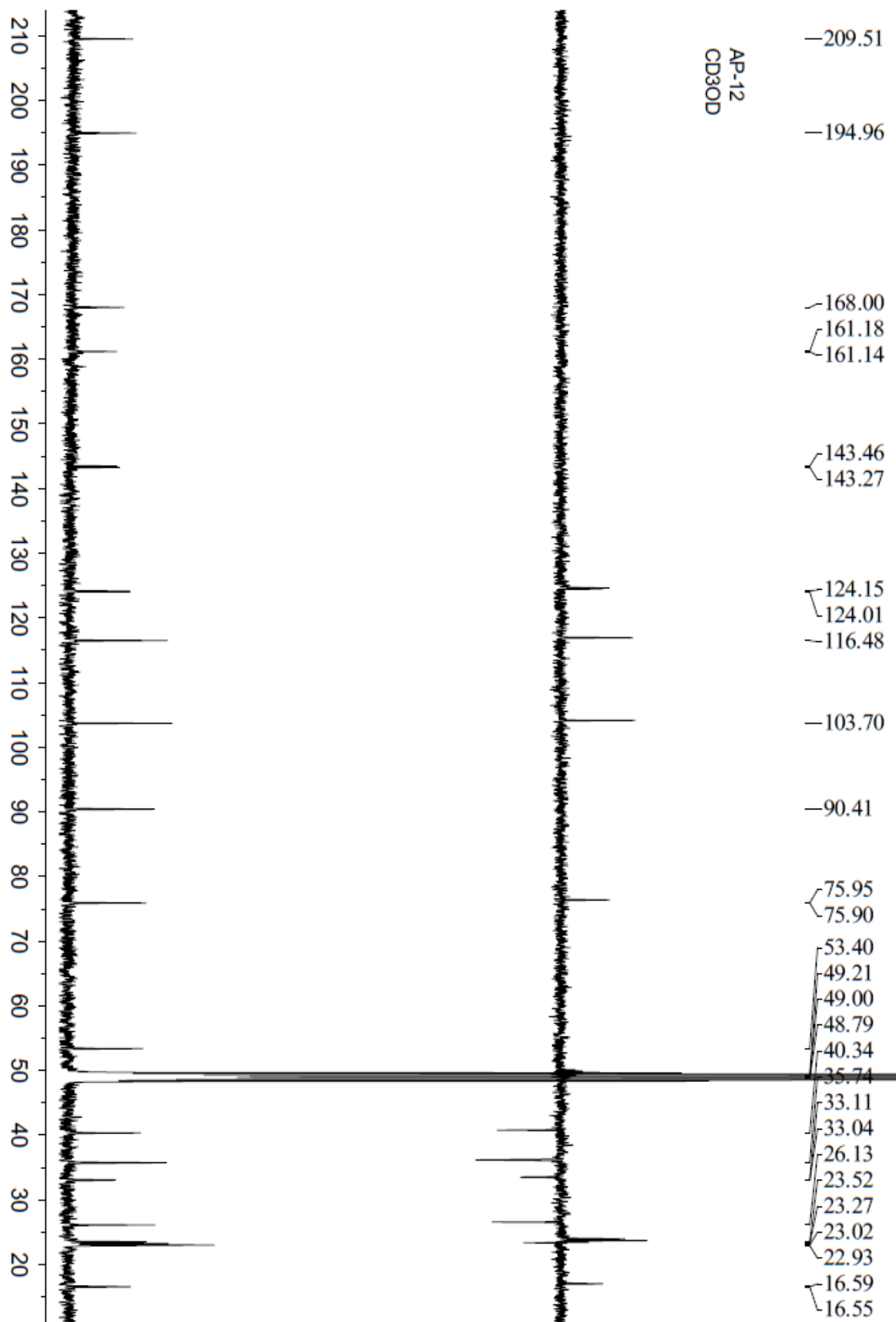
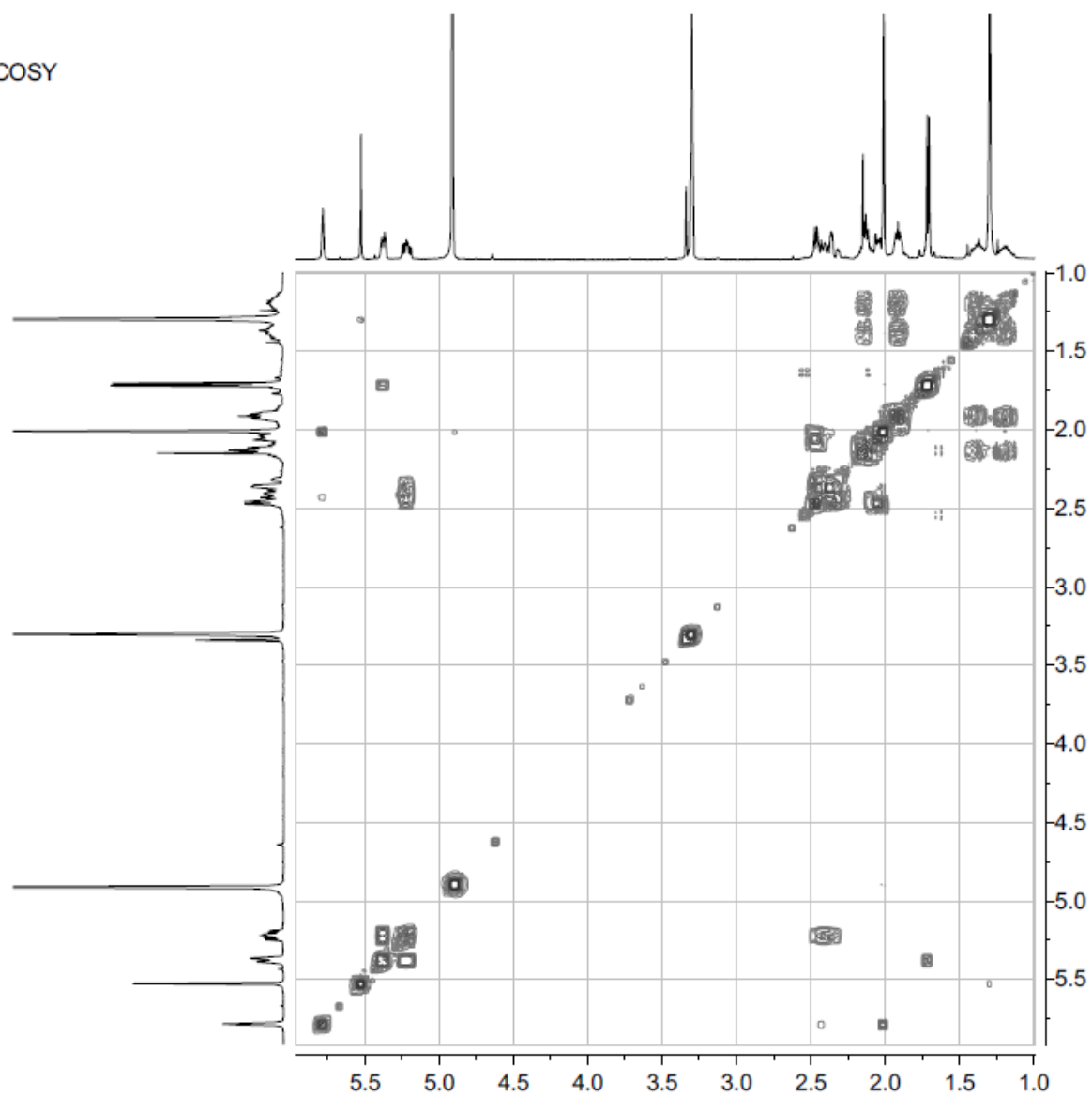


Figure S6.  $^{13}\text{C}$  NMR spectrum for aphadilactone E (1) in  $\text{CD}_3\text{OD}$ .



**Figure S7.**  $^1\text{H}$ - $^1\text{H}$  COSY spectrum for aphadilactone E (**1**) in  $\text{CD}_3\text{OD}$ .

AP-12 COSY  
CD3OD





**Figure S8.** HSQC spectrum for aphadilactone E (**1**) in CD<sub>3</sub>OD.

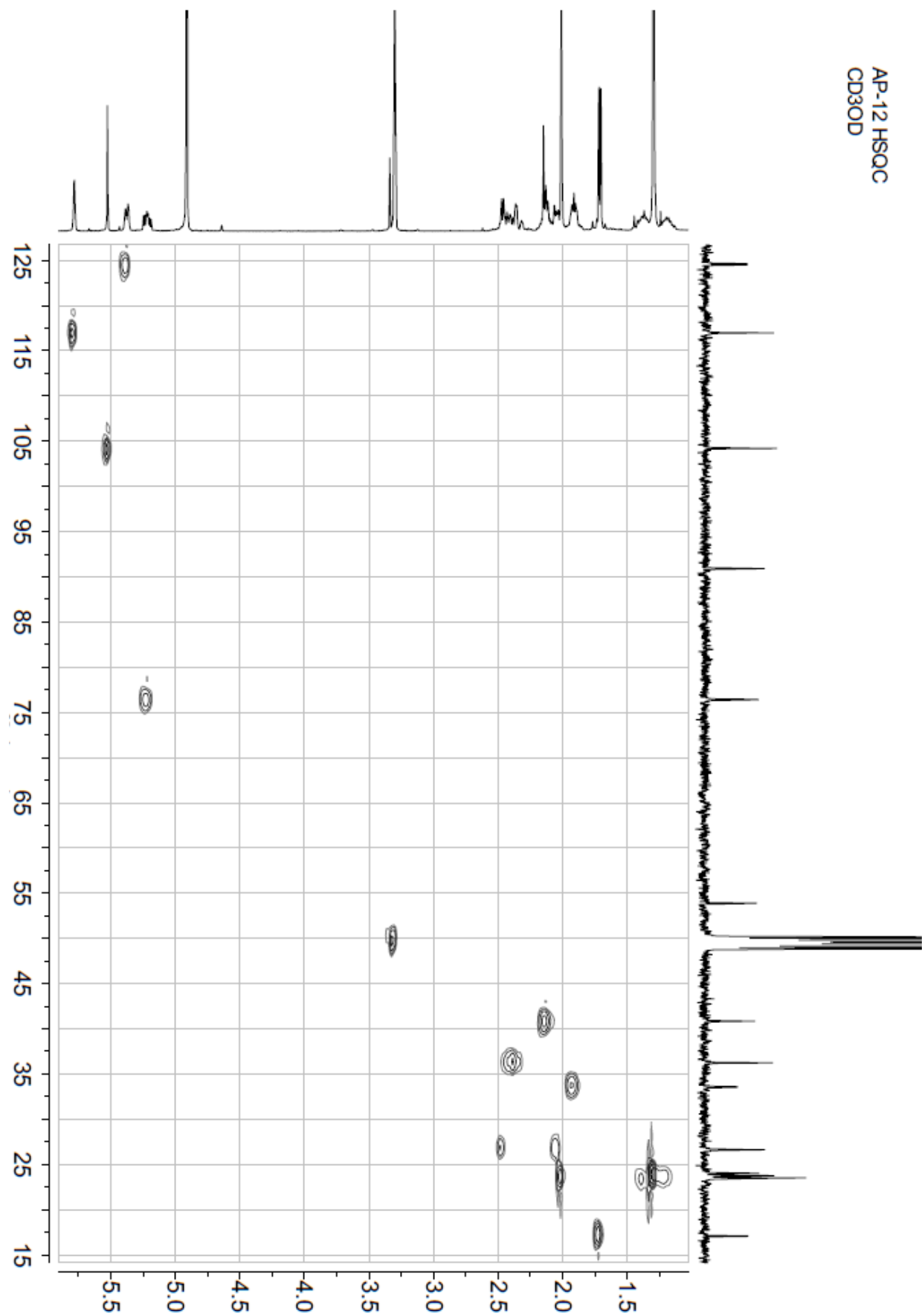
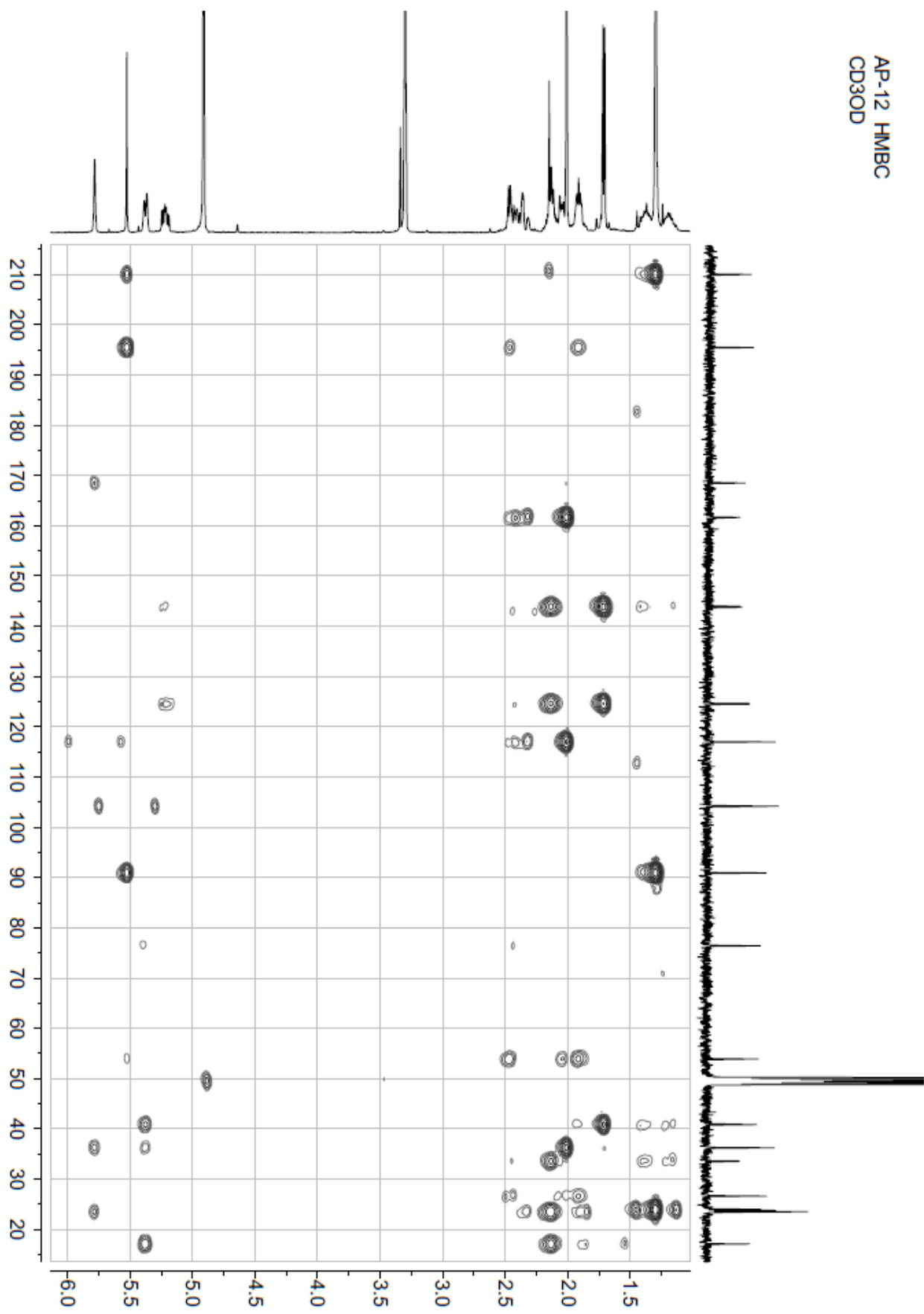


Figure S9. HMBC spectrum for aphadilactone E (1) in CD<sub>3</sub>OD.



**Figure S10.** ROESY spectrum for aphadilactone E (**1**) in CD<sub>3</sub>OD.

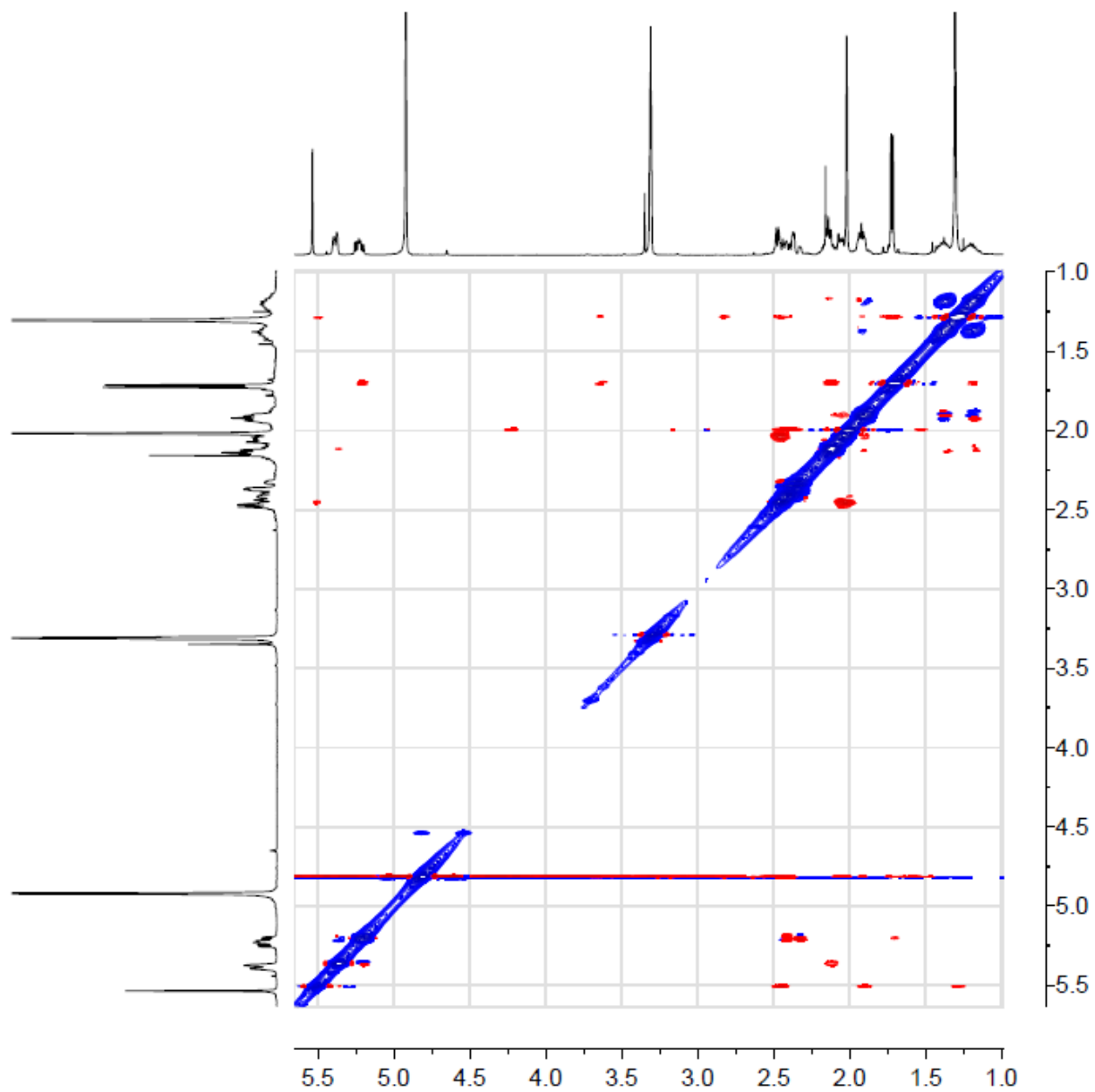


Figure S11. ESI(+)-MS spectrum for aphadilactone E (1).

### Display Report

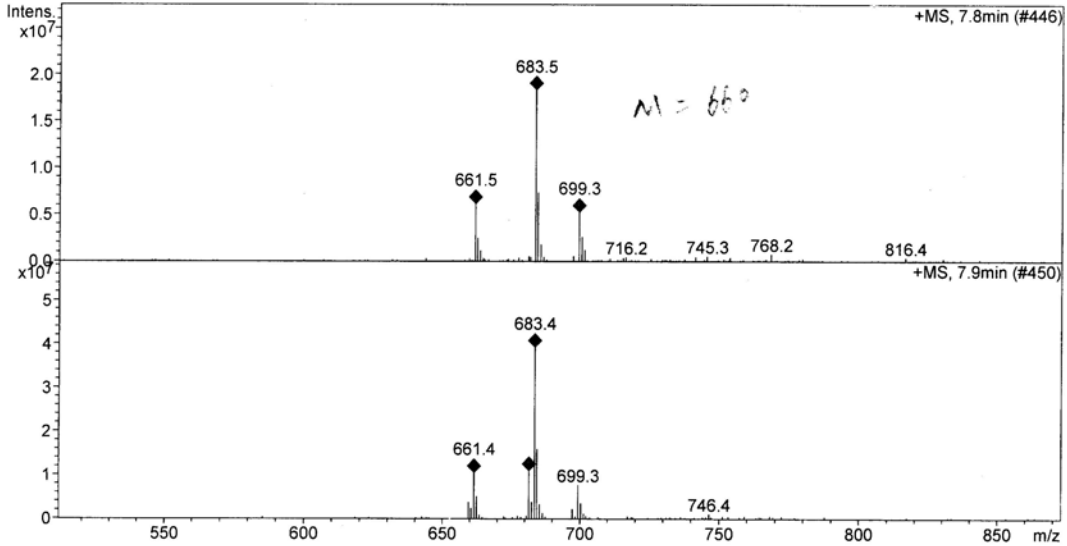
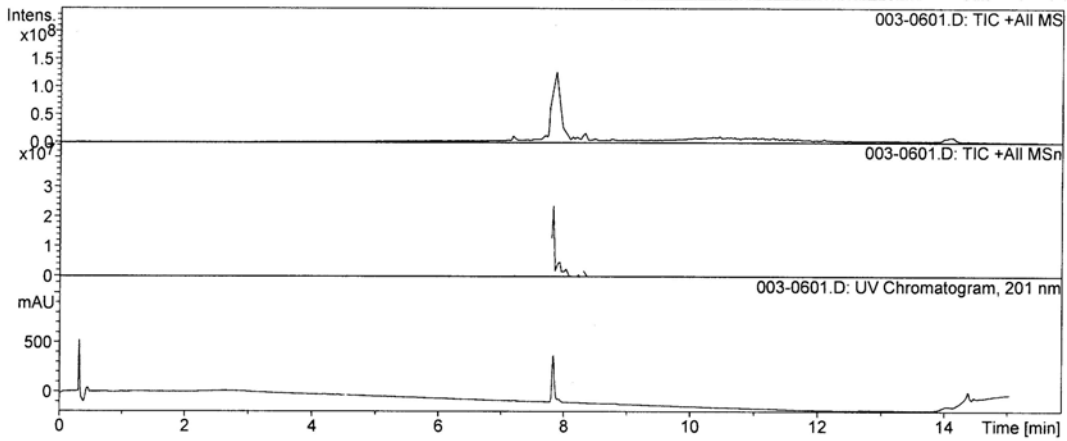
#### Analysis Info

Analysis Name 003-0601.D  
Method Copy of DSOPMS2P.M  
Sample Name yjm-AP-12  
Comment

Acquisition Date 03/17/11 10:10:56  
Operator Administrator  
Instrument esquire3000plus

#### Acquisition Parameter

Ion Source Type	ESI	Ion Polarity	Positive	Alternating Ion Polarity	off
Mass Range Mode	Std/Normal	Scan Begin	100 m/z	Scan End	1750 m/z
Capillary Exit	158.5 Volt	Skim 1	40.0 Volt	Trap Drive	85.4
Accumulation Time	12290 釐	Averages	3 Spectra	Auto MS/MS	on



**Figure S12.** HRESI(+)MS spectrum for aphadilactone E (1).

**Elemental Composition Report**

**Single Mass Analysis**

Tolerance = 3.0 PPM / DBE: min = -1.5, max = 50.0

Element prediction: Off

Number of isotope peaks used for i-FIT = 3

Monoisotopic Mass, Even Electron Ions

285 formula(e) evaluated with 1 results within limits (up to 50 closest results for each mass)

Elements Used:

C: 5-80 H: 2-120 O: 0-20 Na: 0-1

AP-12

LCT PXE KE324

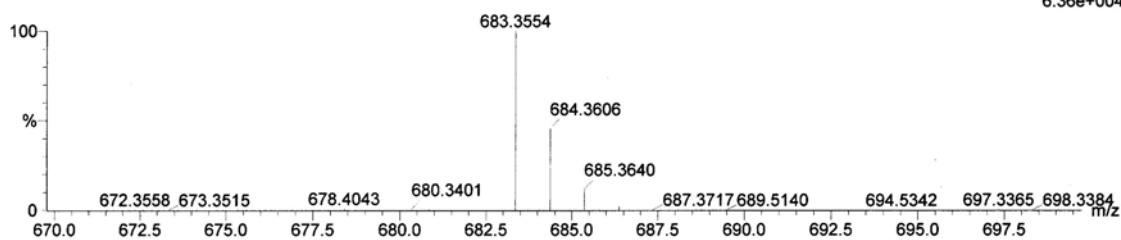
16-Oct-2014

09:44:15

AP-12\_1016 41 (0.865) AM2 (Ar,10000.0,0.00,1.00); ABS; Cm (39:50)

1: TOF MS ES+

6.36e+004



Minimum: -1.5  
Maximum: 5.0 3.0 50.0

Mass	Calc. Mass	mDa	PPM	DBE	i-FIT	i-FIT (Norm)	Formula
683.3554	683.3560	-0.6	-0.9	14.5	116.8	0.0	C40 H52 O8 Na

**Figure S13.**  $^1\text{H}$  NMR spectrum for aphadilactone F (**2**) in  $\text{CD}_3\text{OD}$ .

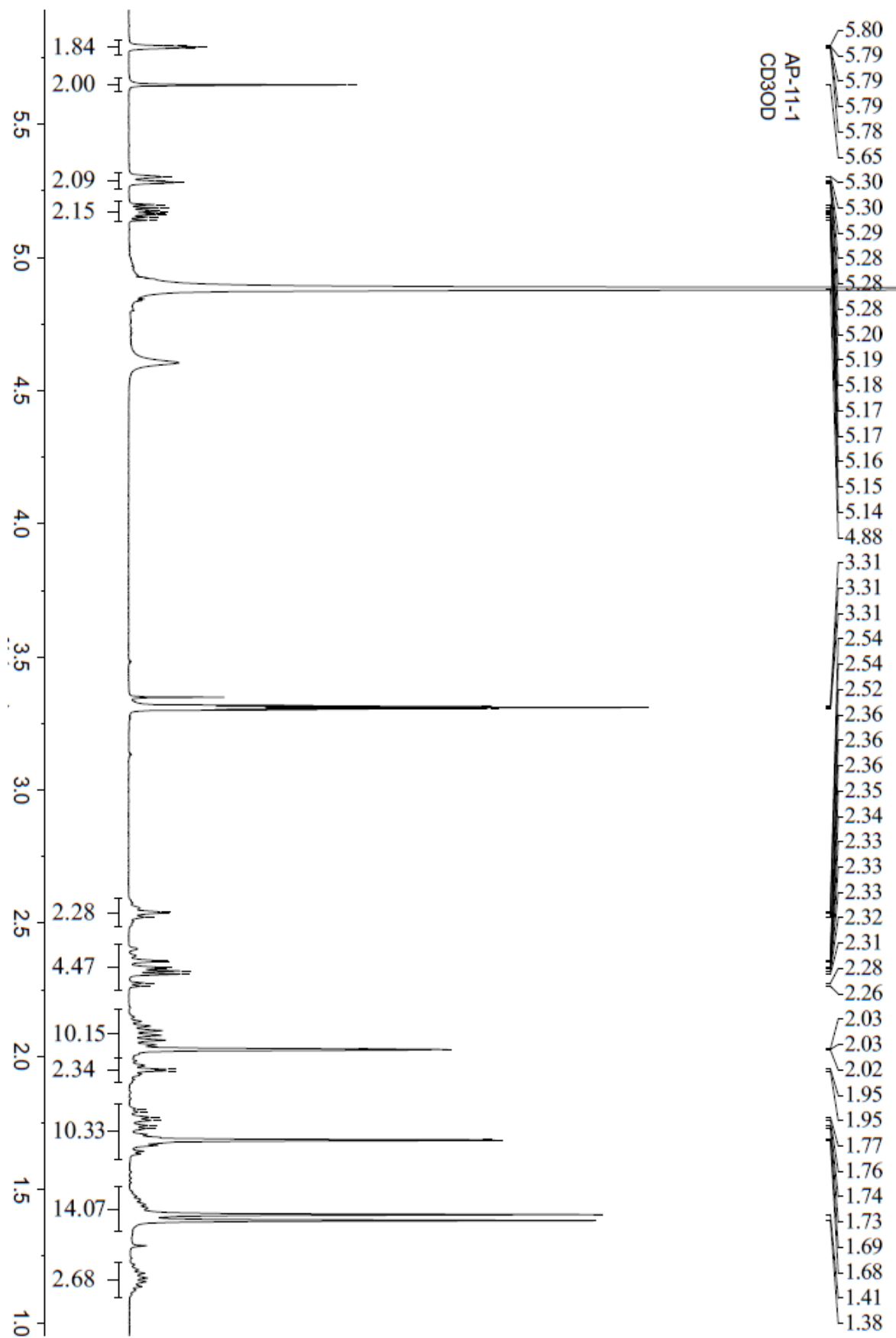
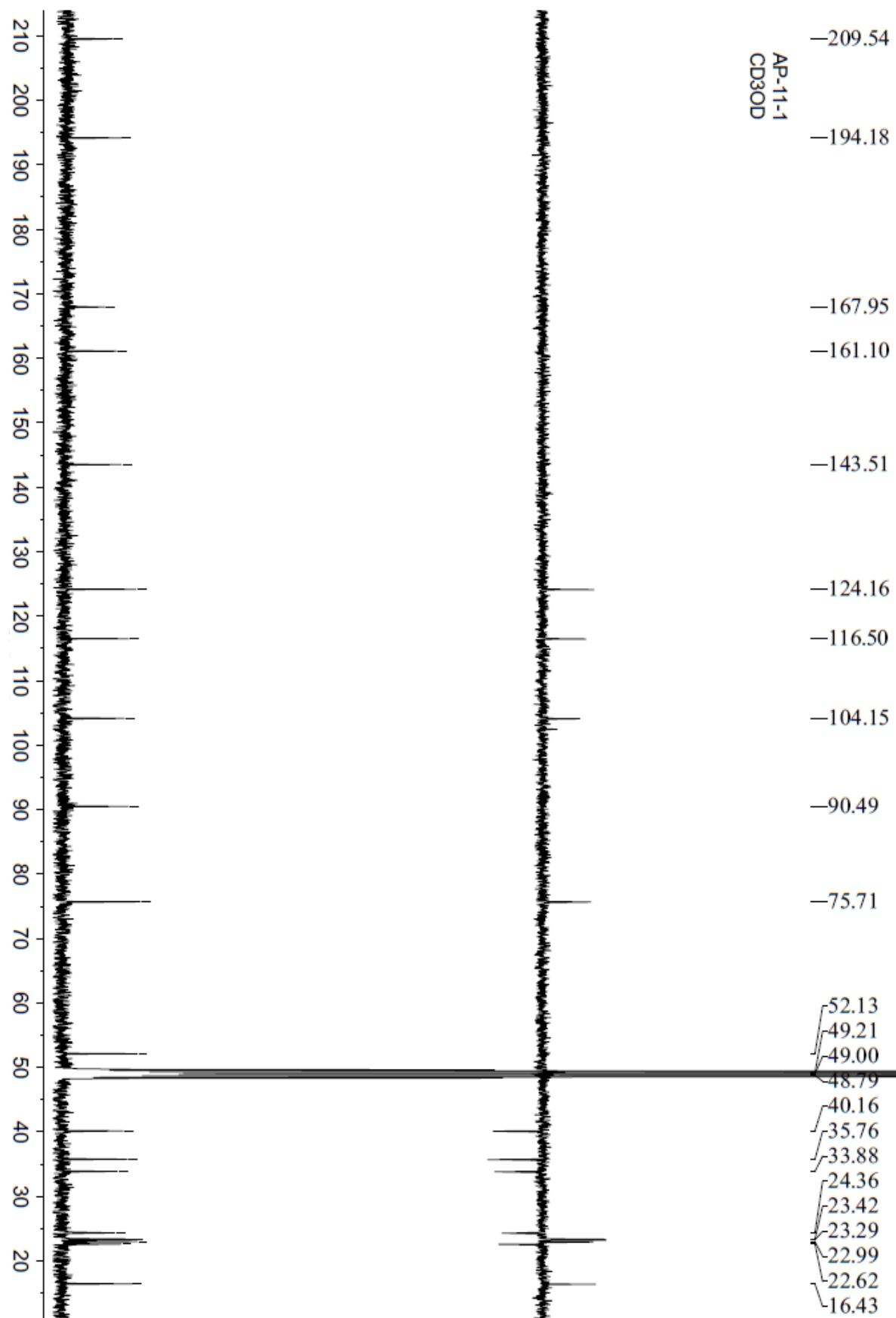
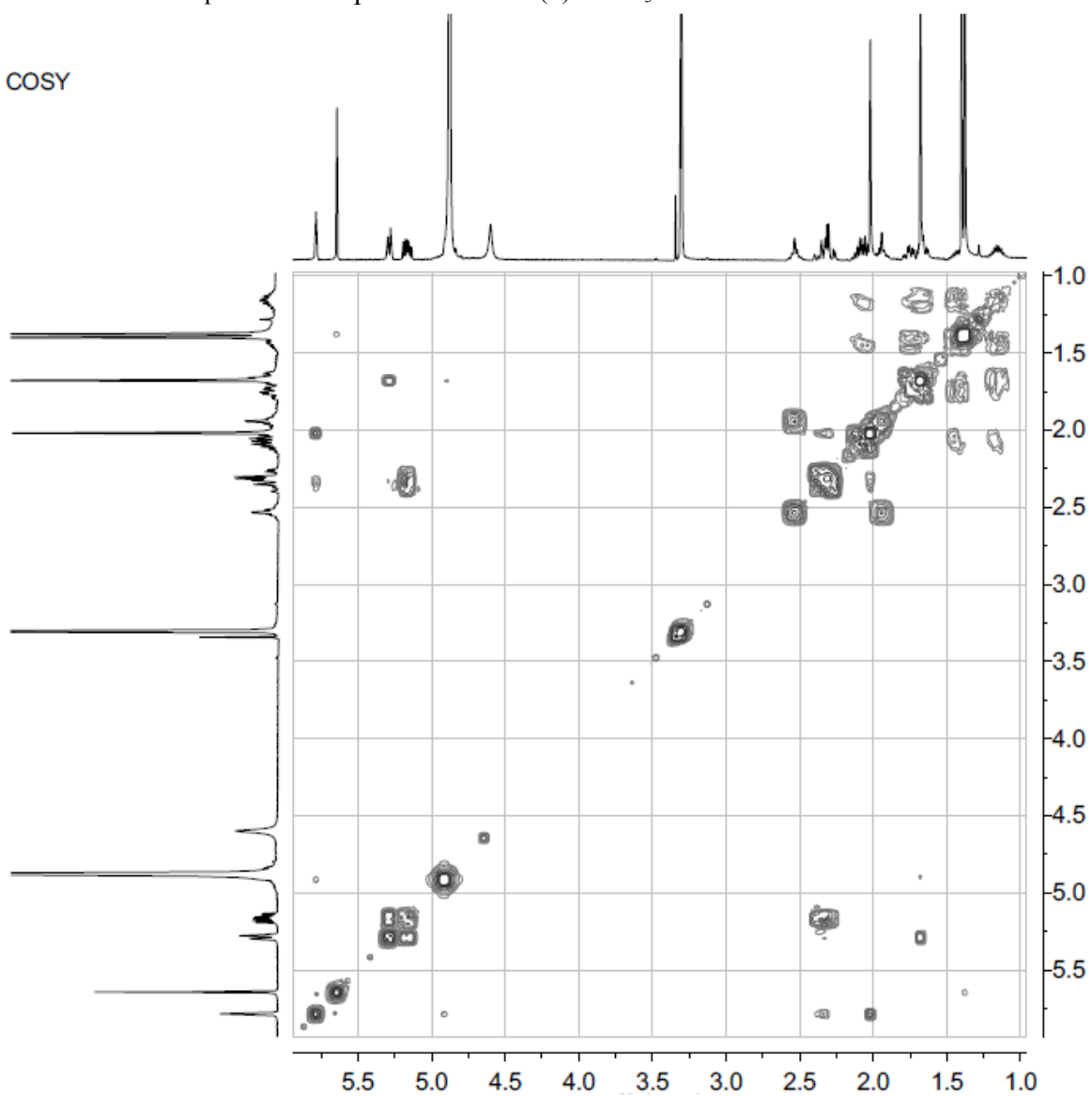


Figure S14.  $^{13}\text{C}$  NMR spectrum for aphadilactone F (2) in  $\text{CD}_3\text{OD}$ .



**Figure S15.**  $^1\text{H}$ - $^1\text{H}$  COSY spectrum for aphadilactone F (**2**) in  $\text{CD}_3\text{OD}$ .

AP-11-1 COSY  
CD3OD





**Figure S16.** HSQC spectrum for aphadilactone F (**2**) in CD<sub>3</sub>OD.

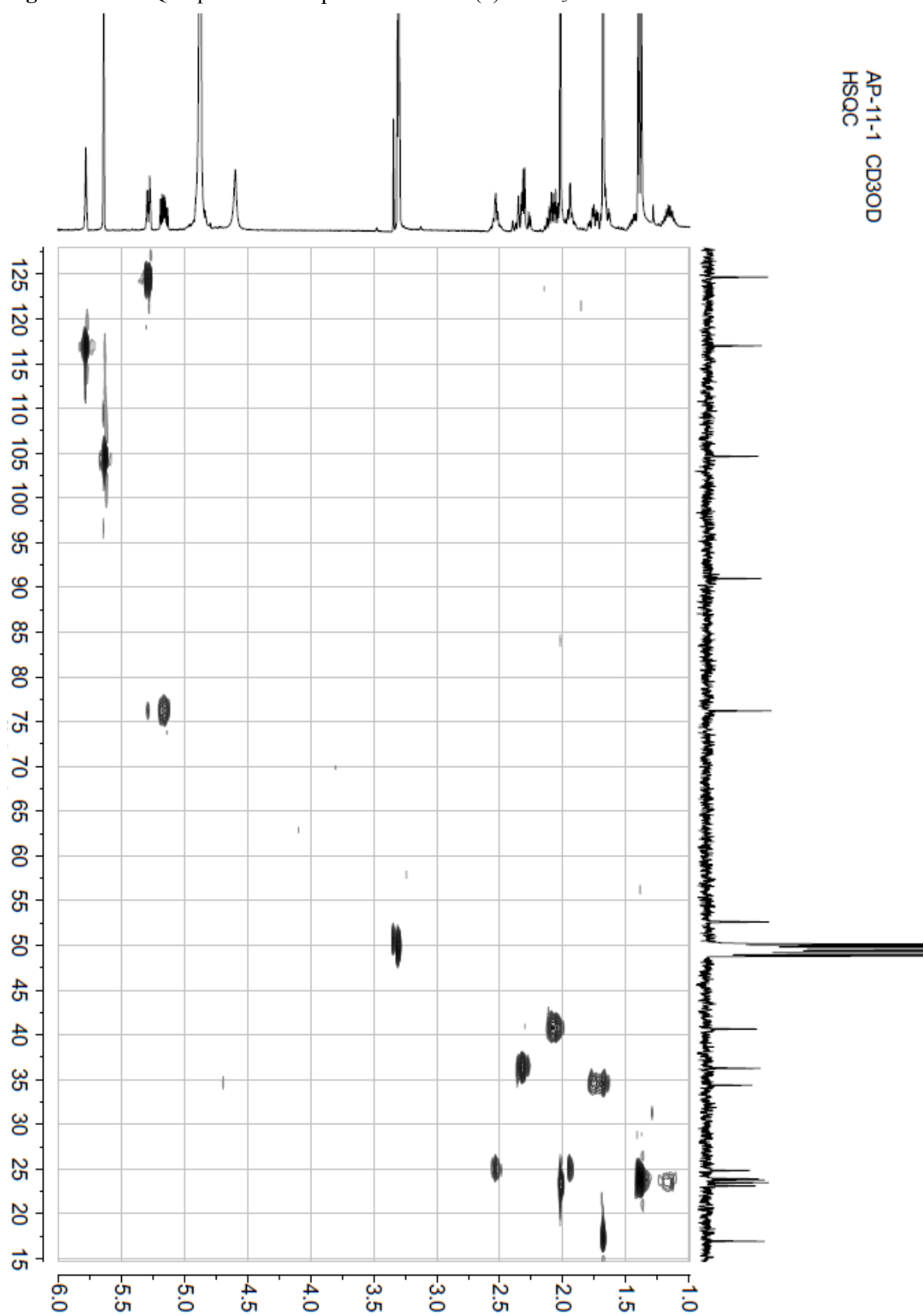
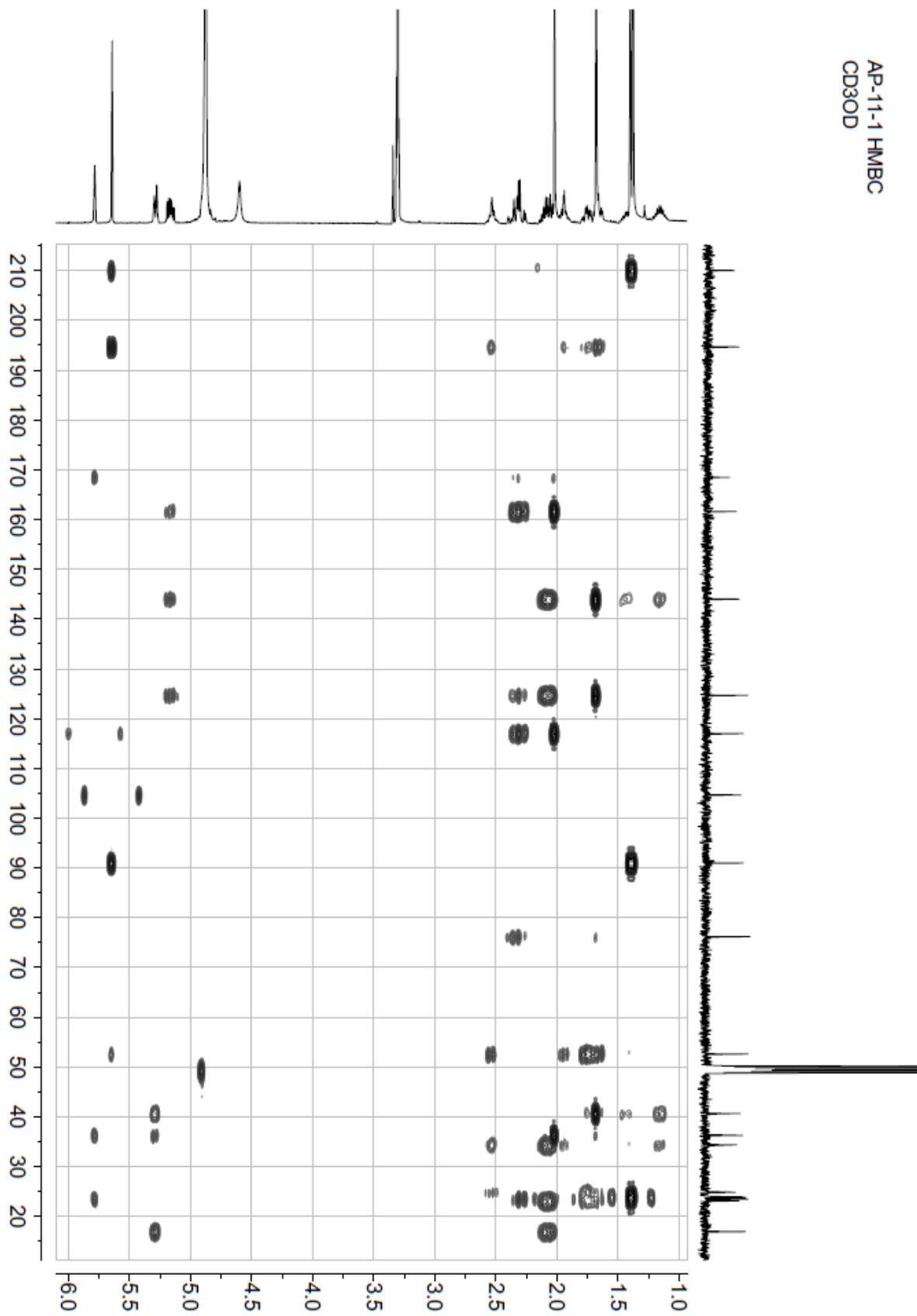


Figure S17. HMBC spectrum for aphadilactone F (2) in CD<sub>3</sub>OD.



**Figure S18.** ROESY spectrum for aphadilactone F (2) in CD<sub>3</sub>OD.

AP-11-1 ROESY

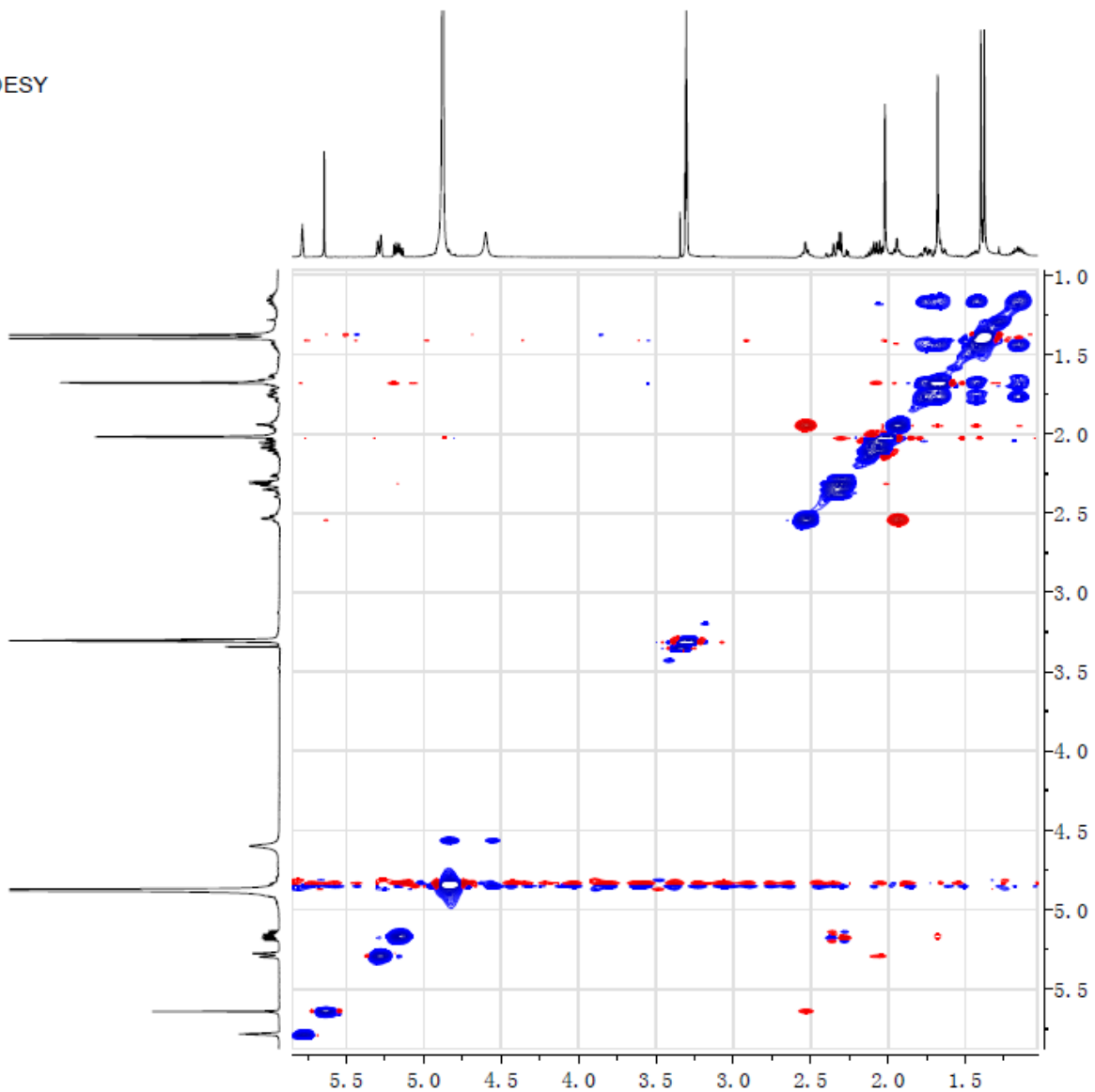


Figure S19. ESI(+)-MS spectrum for aphadilactone F (2).

### Display Report

#### Analysis Info

Analysis Name 011-1401.D  
Method Copy of DSOPMS2P.M  
Sample Name yjm-AP-11-1  
Comment

Acquisition Date 07/11/11 19:34:32  
Operator Administrator  
Instrument esquire3000plus

#### Acquisition Parameter

Ion Source Type	ESI	Ion Polarity	Positive	Alternating Ion Polarity	off
Mass Range Mode	Std/Normal	Scan Begin	100 m/z	Scan End	1750 m/z
Capillary Exit	158.5 Volt	Skim 1	40.0 Volt	Trap Drive	85.4
Accumulation Time	15000 纒	Averages	3 Spectra	Auto MS/MS	on

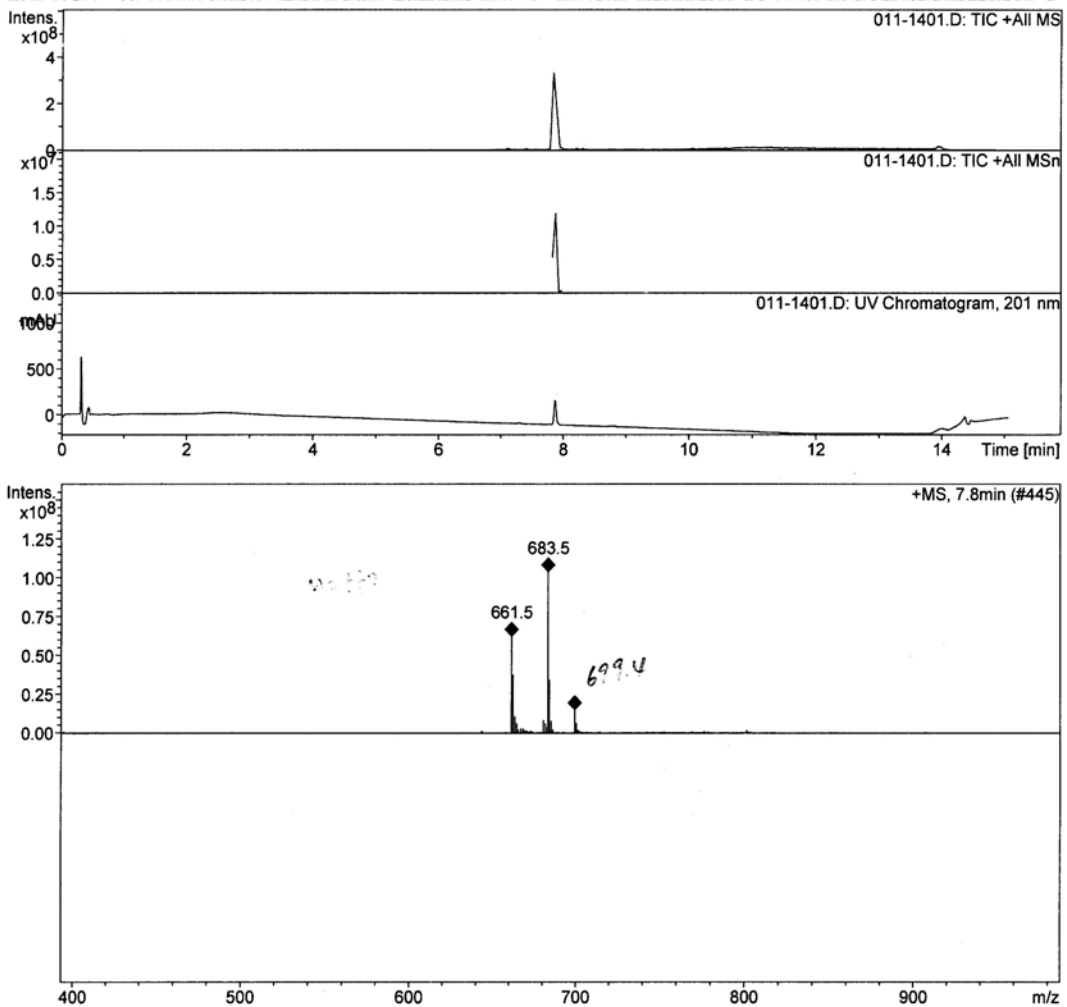


Figure S20. HRESI(+)MS spectrum for aphadilactone F (2).

**Elemental Composition Report**

**Single Mass Analysis**

Tolerance = 3.0 PPM / DBE: min = -1.5, max = 50.0  
 Element prediction: Off  
 Number of isotope peaks used for i-FIT = 3

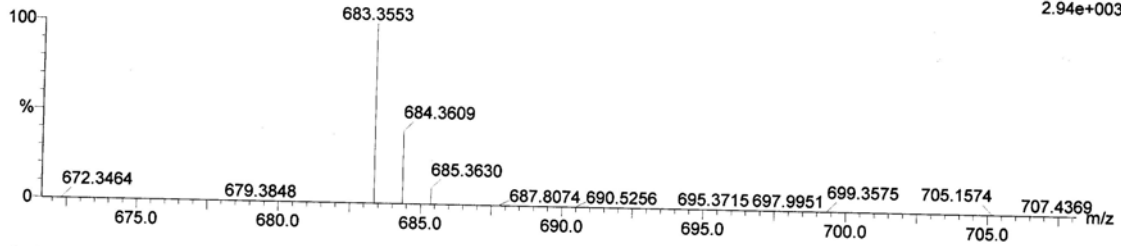
Monoisotopic Mass, Even Electron Ions  
 285 formula(e) evaluated with 1 results within limits (up to 50 closest results for each mass)  
 Elements Used:

C: 5-80 H: 2-120 O: 0-20 Na: 0-1  
 AP-11-1

LCT PXE KE324

16-Oct-2014  
 09:56:37  
 1: TOF MS ES+  
 2.94e+003

AP-11-1\_1016 52 (1.128) AM2 (Ar,10000.0,0.00,1.00); ABS; Cm (41:54)



Mass	Calc. Mass	mDa	PPM	DBE	i-FIT	i-FIT (Norm)	Formula
683.3553	683.3560	-0.7	-1.0	14.5	23.9	0.0	C40 H52 O8 Na

**Figure S21.**  $^1\text{H}$  NMR spectrum for aphadilactone G (**3**) in  $\text{CD}_3\text{OD}$ .

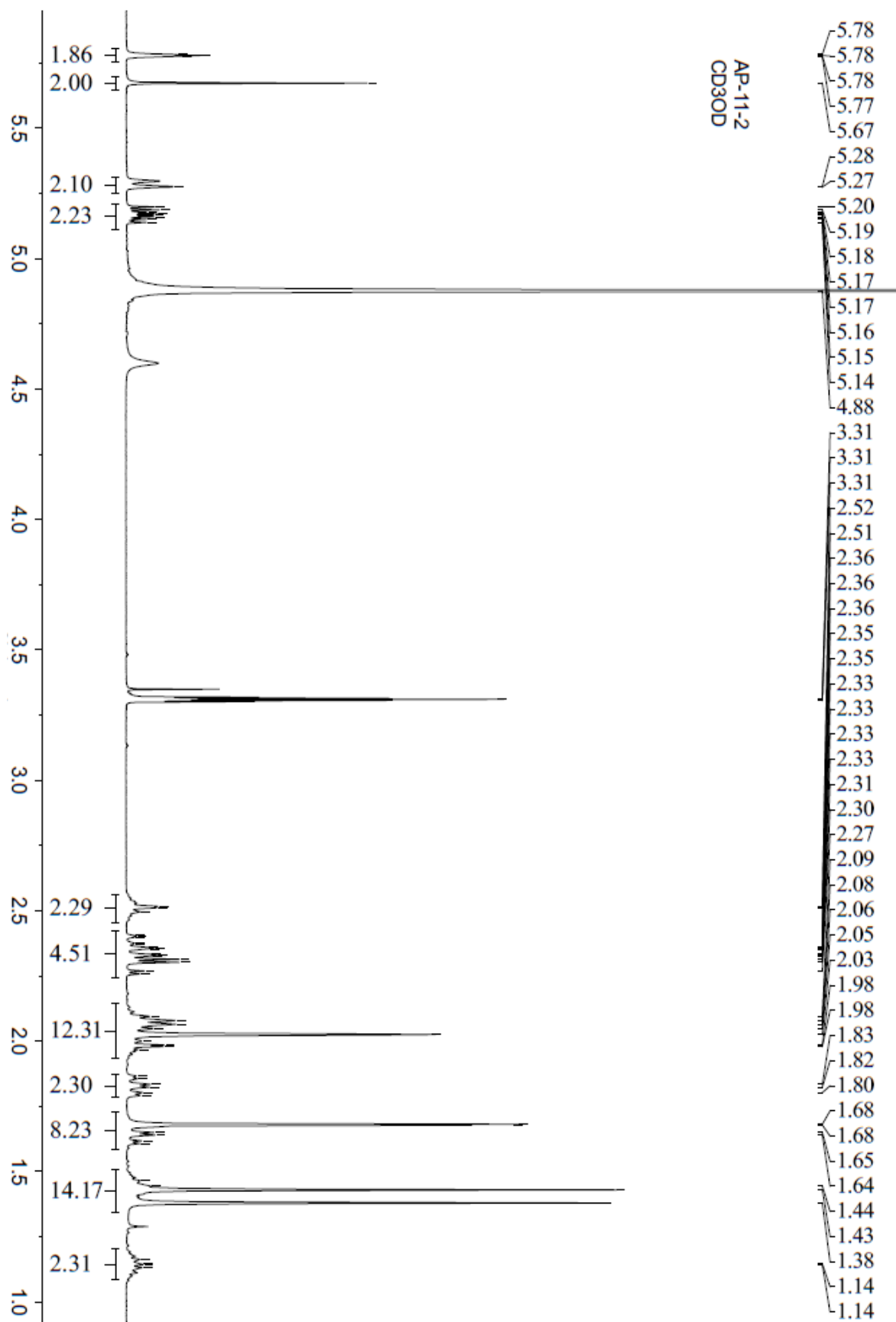
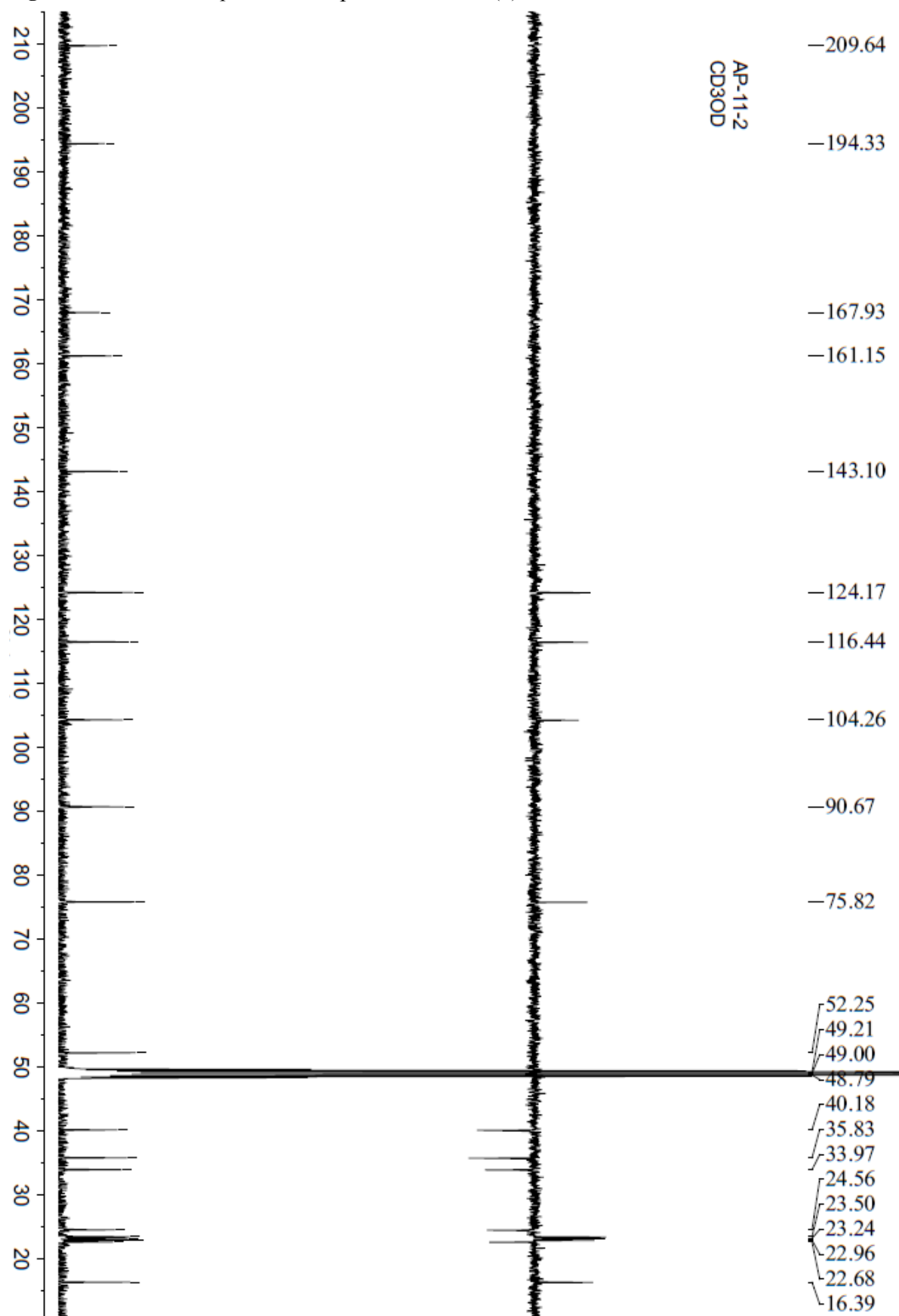


Figure S22.  $^{13}\text{C}$  NMR spectrum for aphadilactone G (3) in  $\text{CD}_3\text{OD}$ .



**Figure S23.**  $^1\text{H}$ - $^1\text{H}$  COSY spectrum for aphadilactone G (3) in  $\text{CD}_3\text{OD}$ .

AP-11-2 COSY  
CD3OD

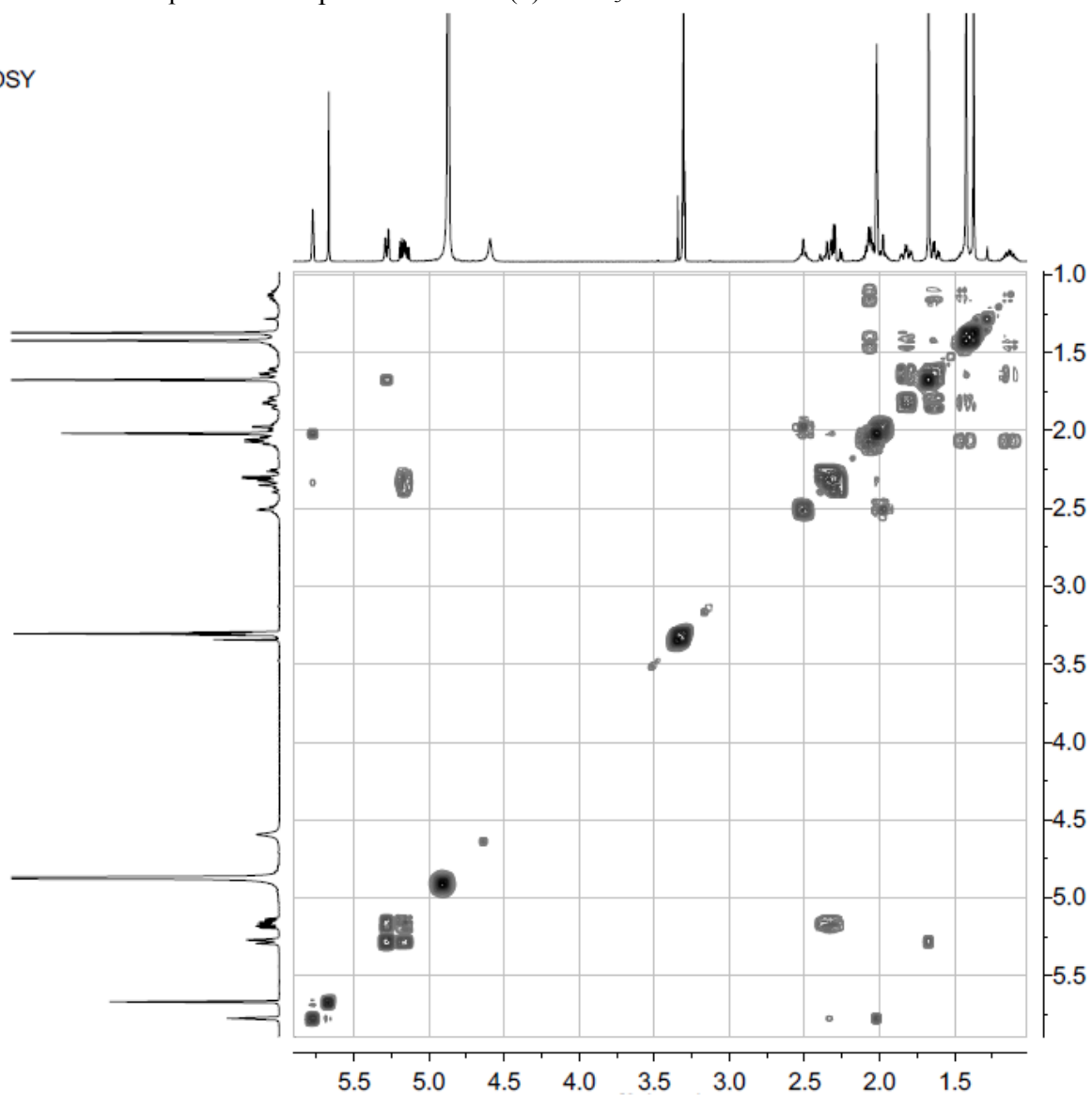




Figure S24. HSQC spectrum for aphadilactone G (3) in CD<sub>3</sub>OD.

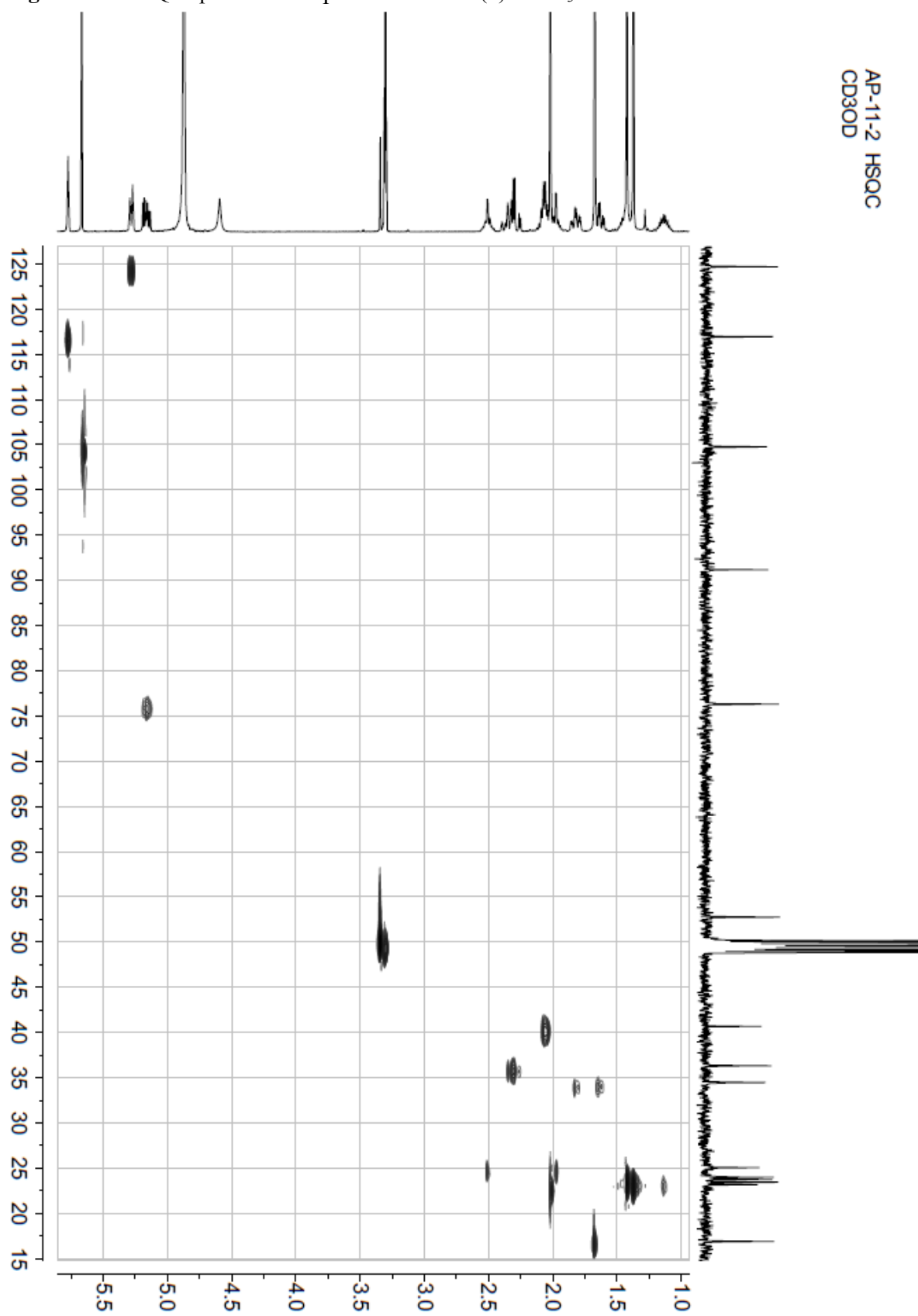
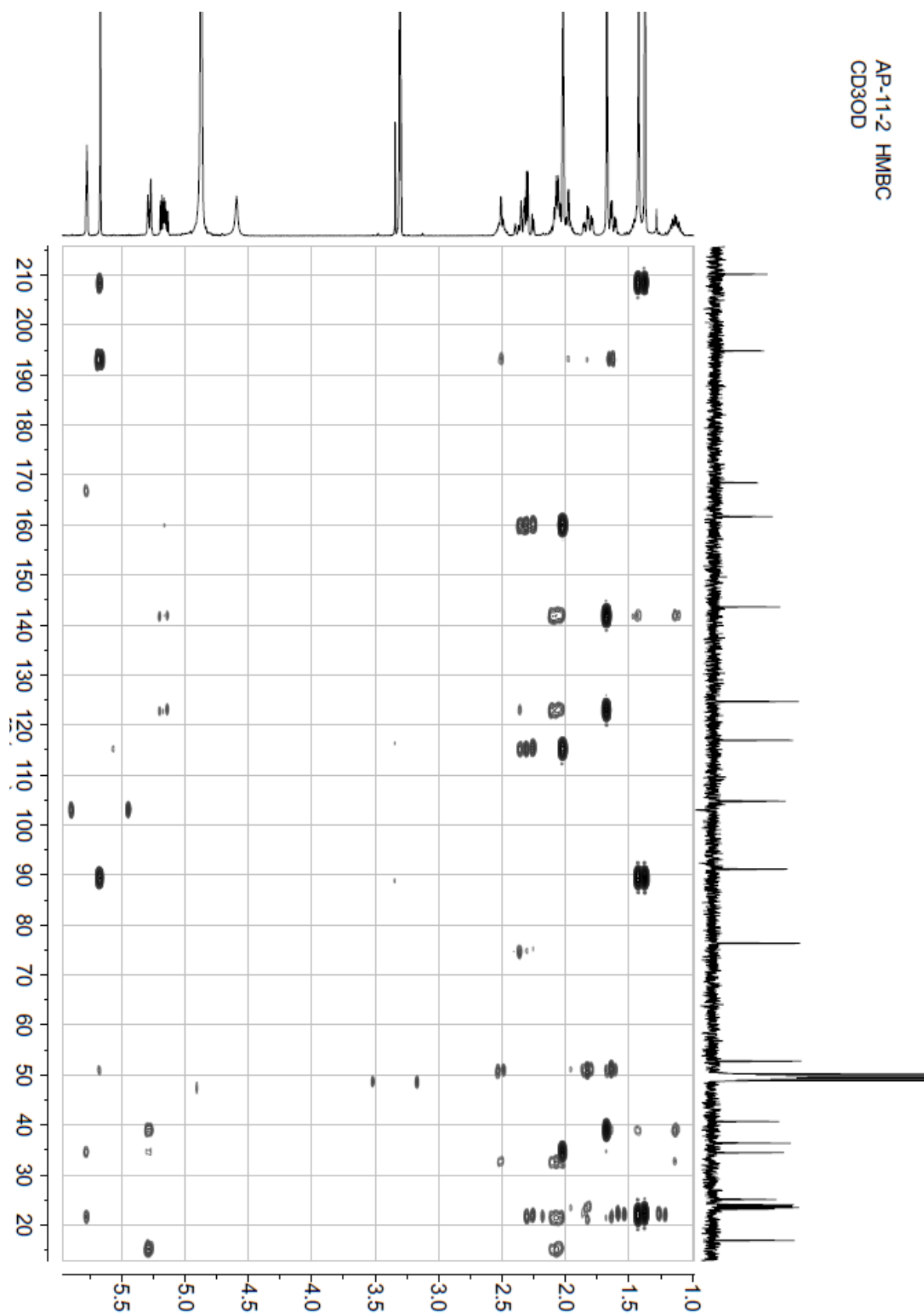


Figure S25. HMBC spectrum for aphadilactone G (3) in CD<sub>3</sub>OD.



**Figure S26.** ROESY spectrum for aphadilactone G (**3**) in CD<sub>3</sub>OD.

AP-11-2 ROESY  
CD3OD

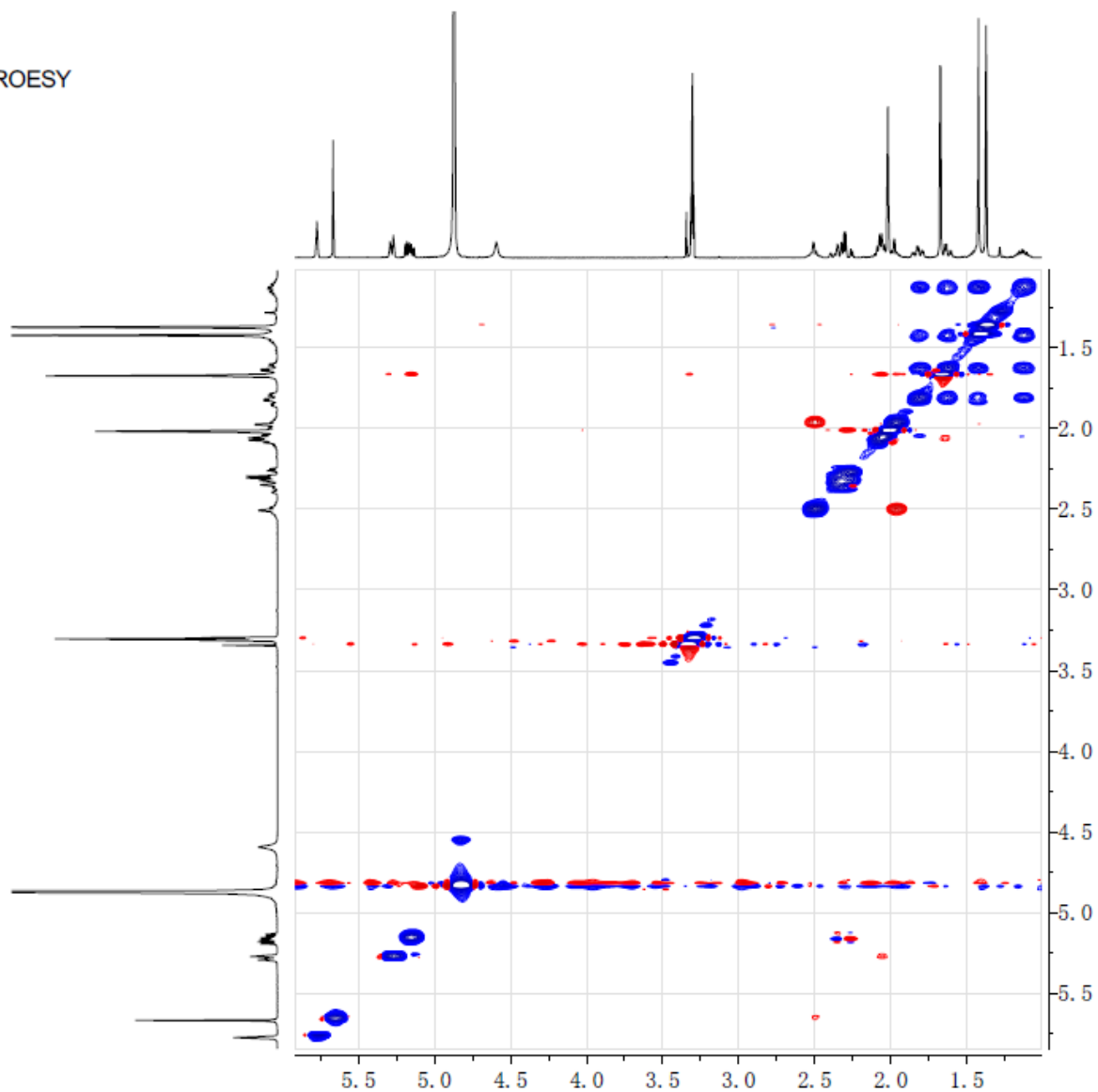
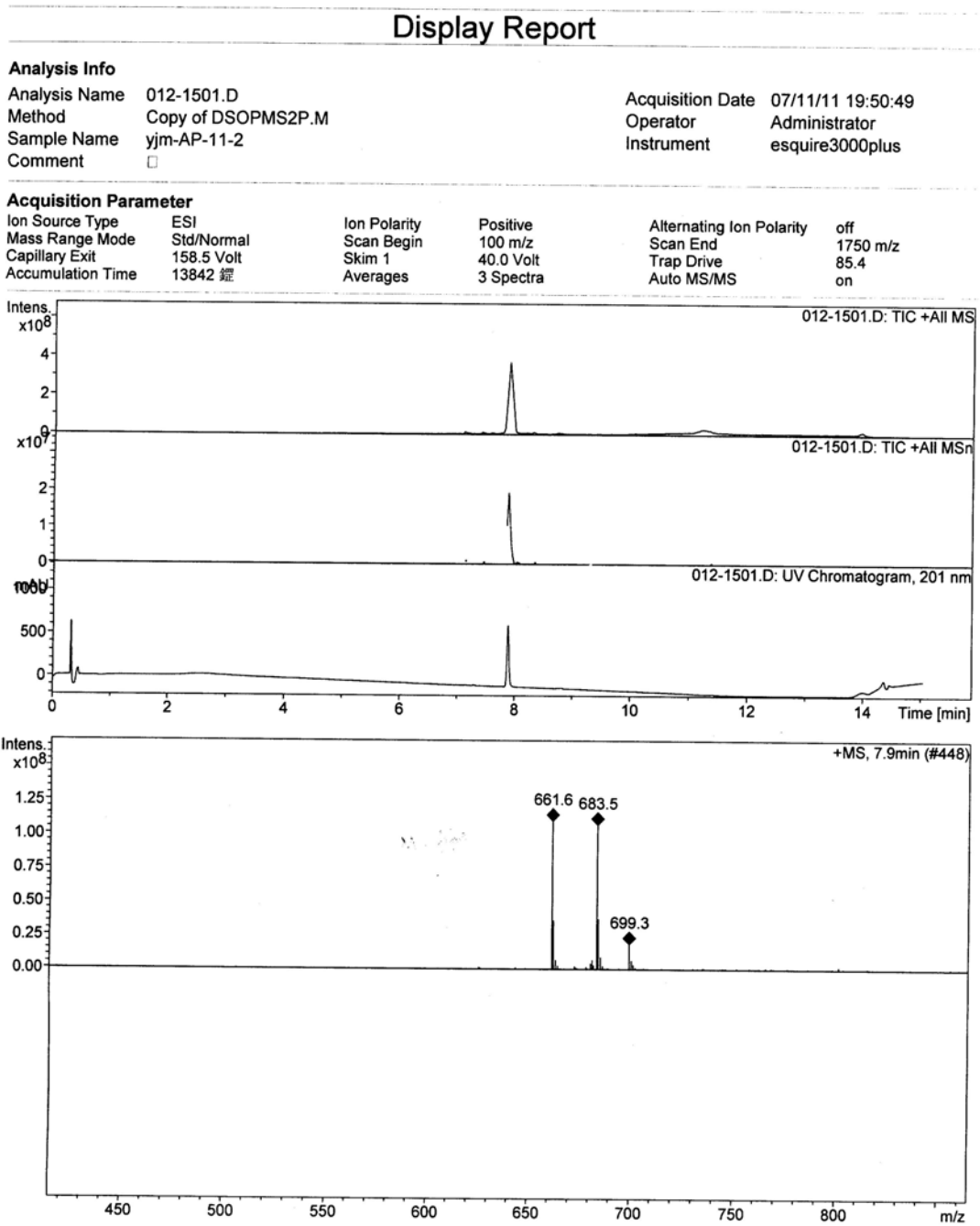


Figure S27. ESI(+)-MS spectrum for aphadilactone G (3).



**Figure S28.** HRESI(+)MS spectrum for aphadilactone G (3).

**Elemental Composition Report**

**Single Mass Analysis**

Tolerance = 3.0 PPM / DBE: min = -1.5, max = 50.0

Element prediction: Off

Number of isotope peaks used for i-FIT = 3

Monoisotopic Mass, Even Electron Ions

285 formula(e) evaluated with 2 results within limits (up to 50 closest results for each mass)

Elements Used:

C: 5-80 H: 2-120 O: 0-20 Na: 0-1

AP-11-2

LCT PXE KE324

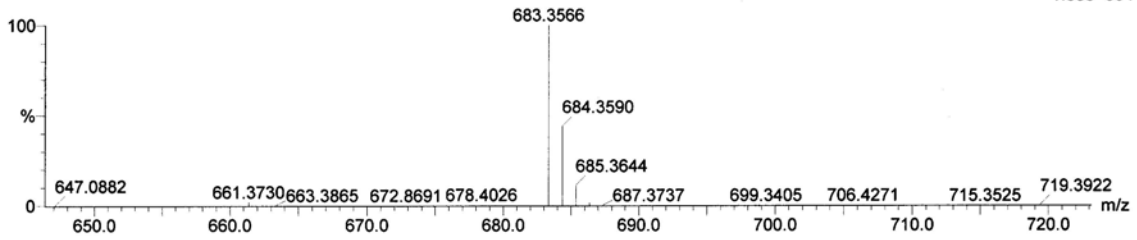
16-Oct-2014

09:51:18

AP-11-2\_1016 44 (0.953) AM2 (Ar,10000.0,0.00,1.00); ABS; Cm (33:48)

1: TOF MS ES+

1.88e+004



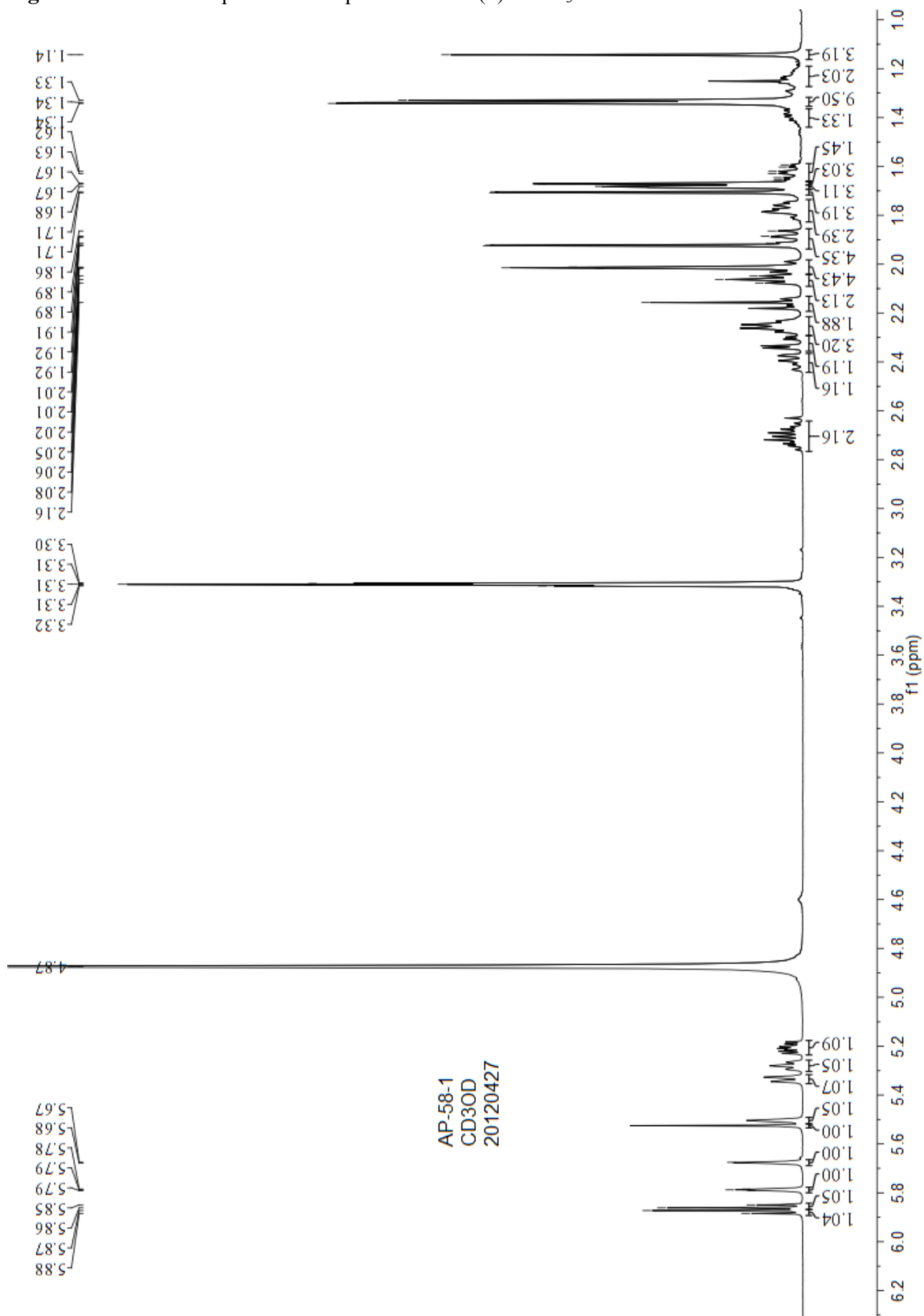
Minimum:

Maximum: 5.0 3.0 -1.5

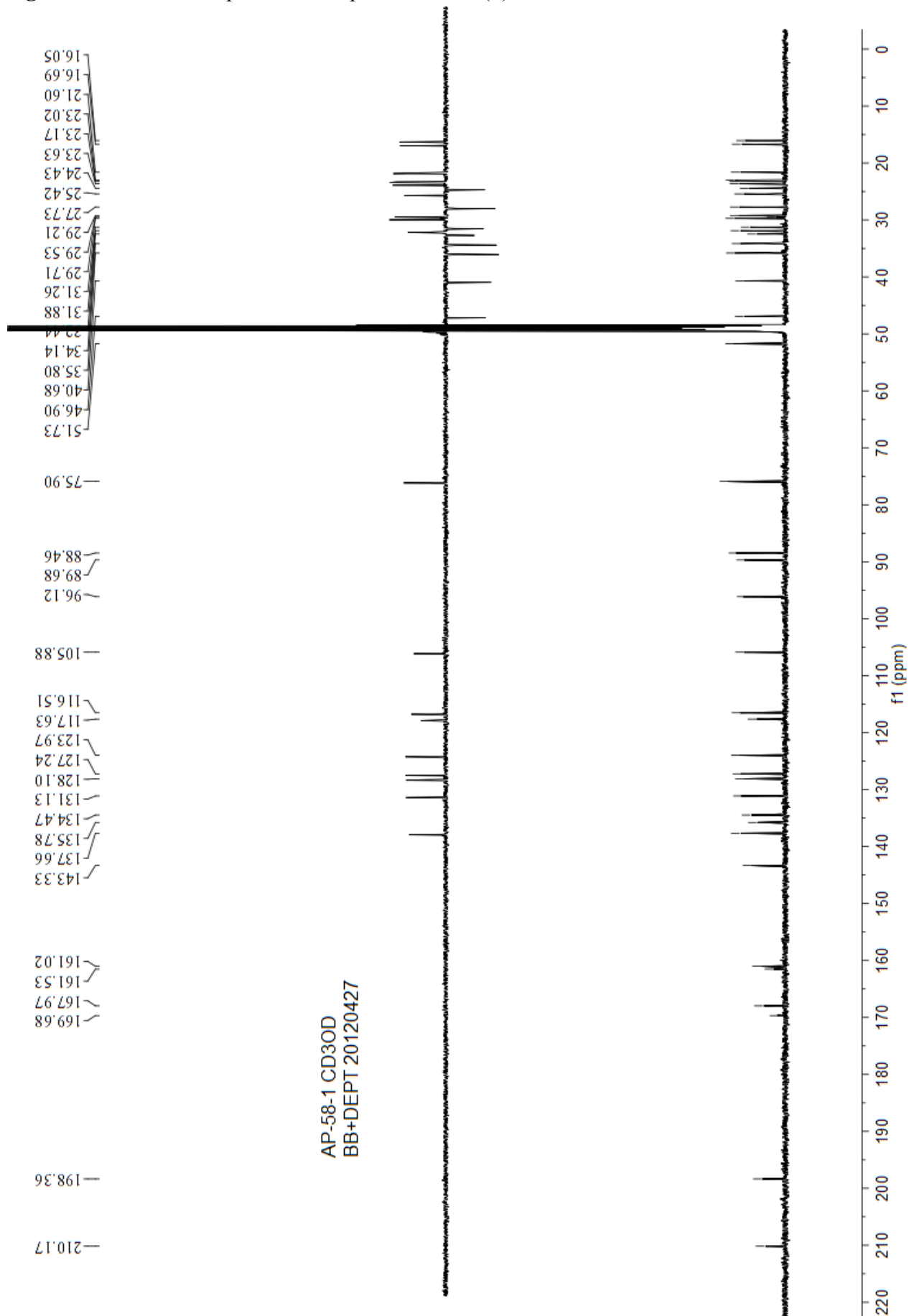
50.0

Mass	Calc. Mass	mDa	PPM	DBE	i-FIT	i-FIT (Norm)	Formula
683.3566	683.3560	0.6	0.9	14.5	51.1	0.0	C40 H52 O8 Na
	683.3584	-1.8	-2.6	17.5	56.2	5.2	C42 H51 O8

**Figure S29.**  $^1\text{H}$  NMR spectrum for aphanamene H (**4**) in  $\text{CD}_3\text{OD}$ .



**Figure S30.**  $^{13}\text{C}$  NMR spectrum for aphanamene H (**4**) in  $\text{CD}_3\text{OD}$ .



**Figure S31.**  $^1\text{H}$ - $^1\text{H}$  COSY spectrum for aphanamene H (**4**) in  $\text{CD}_3\text{OD}$ .

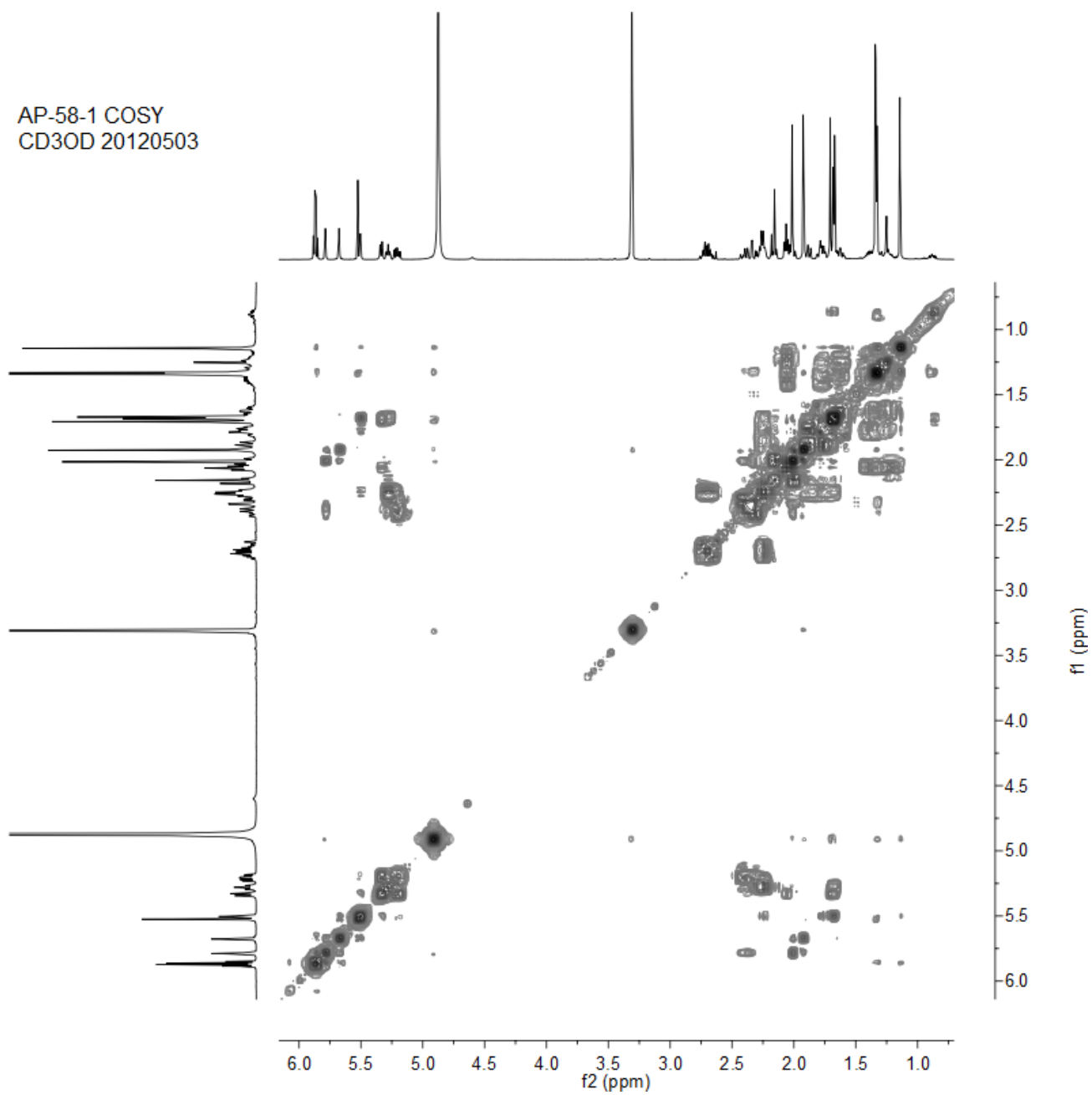




Figure S32. HSQC spectrum for aphanamene H (4) in CD<sub>3</sub>OD.

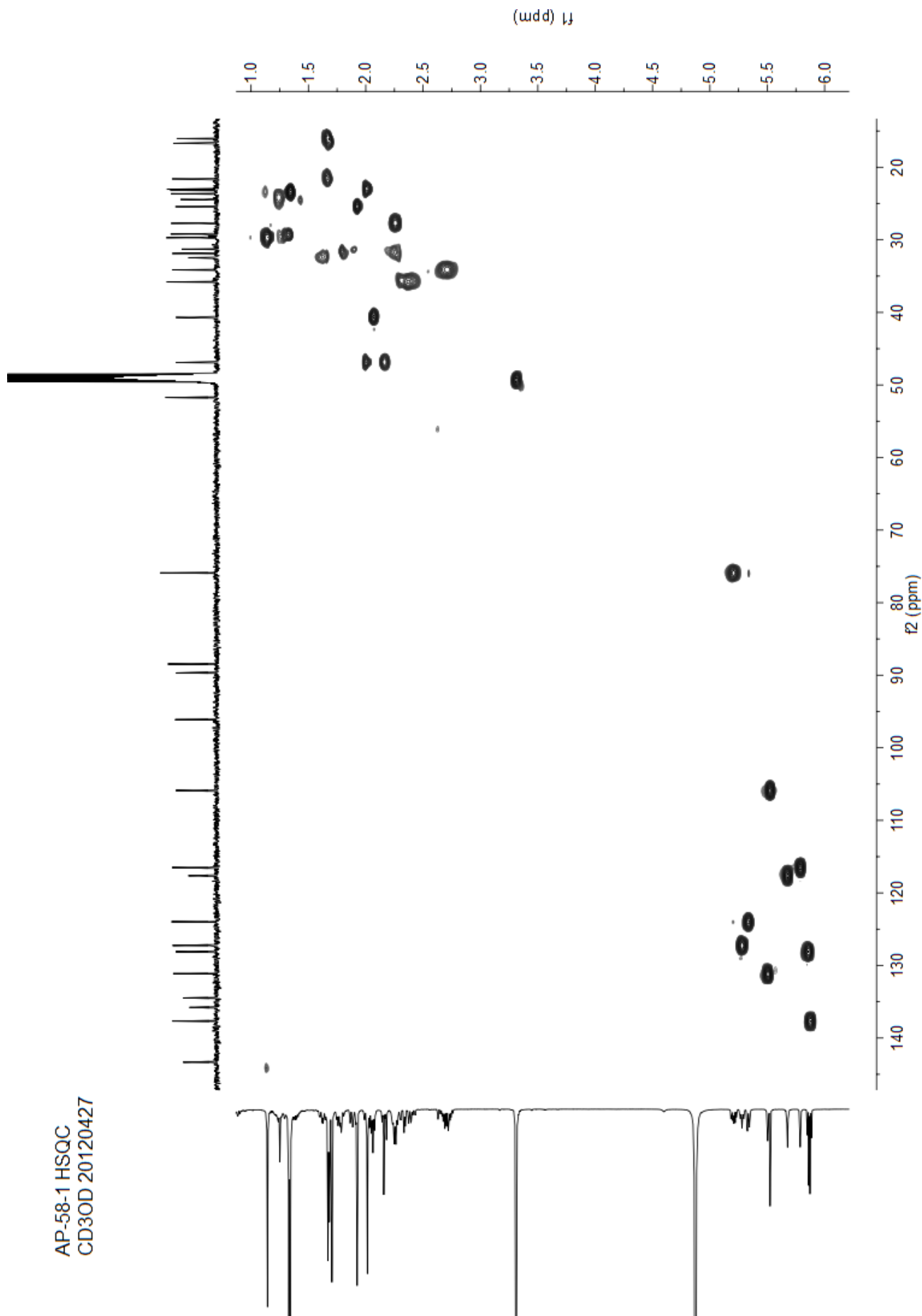
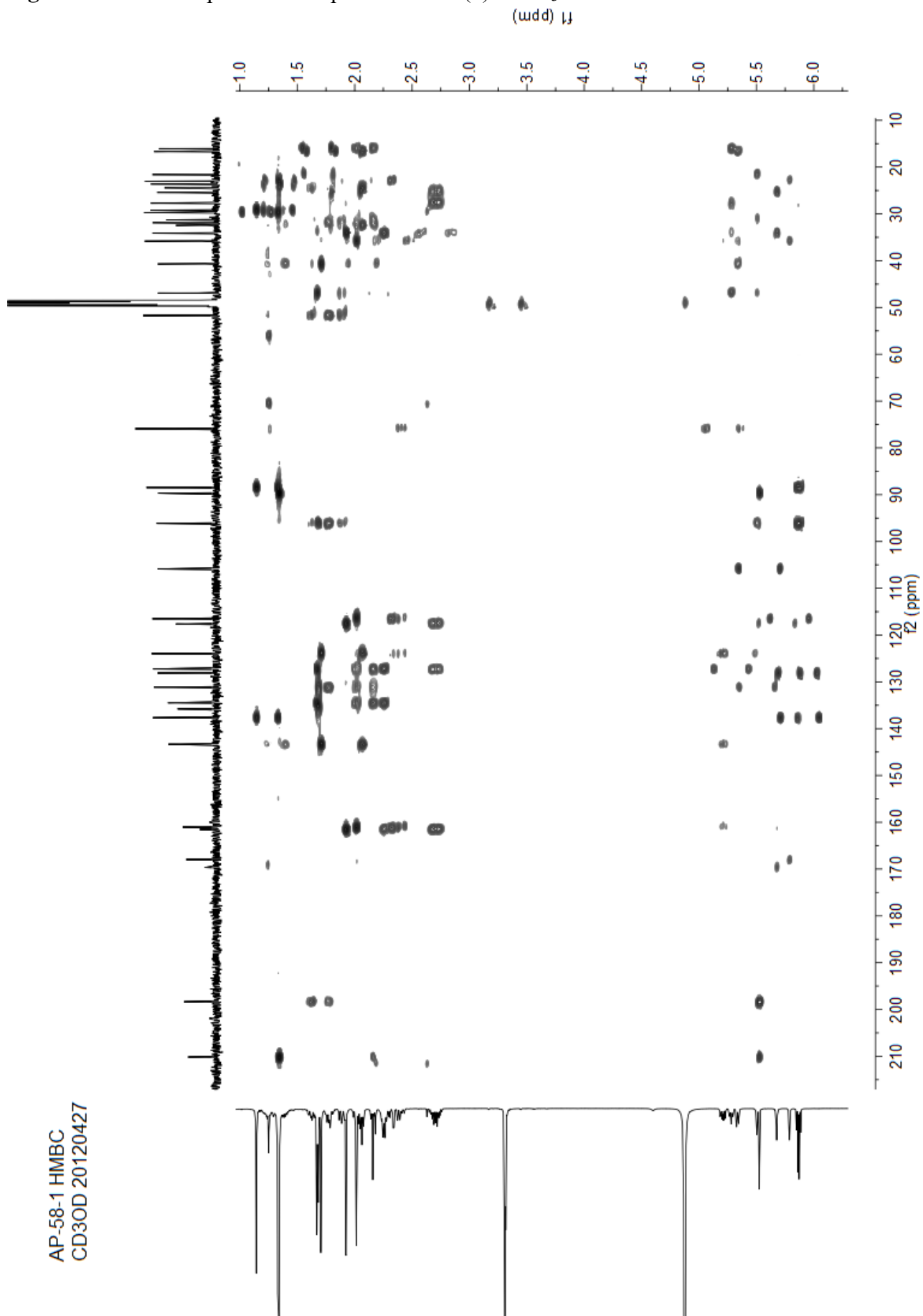


Figure S33. HMBC spectrum for aphanamene H (4) in CD<sub>3</sub>OD.



**Figure S34.** ROESY spectrum for aphanamene H (**4**) in CD<sub>3</sub>OD.

AP-58-1 ROESY  
CD3OD 20120502

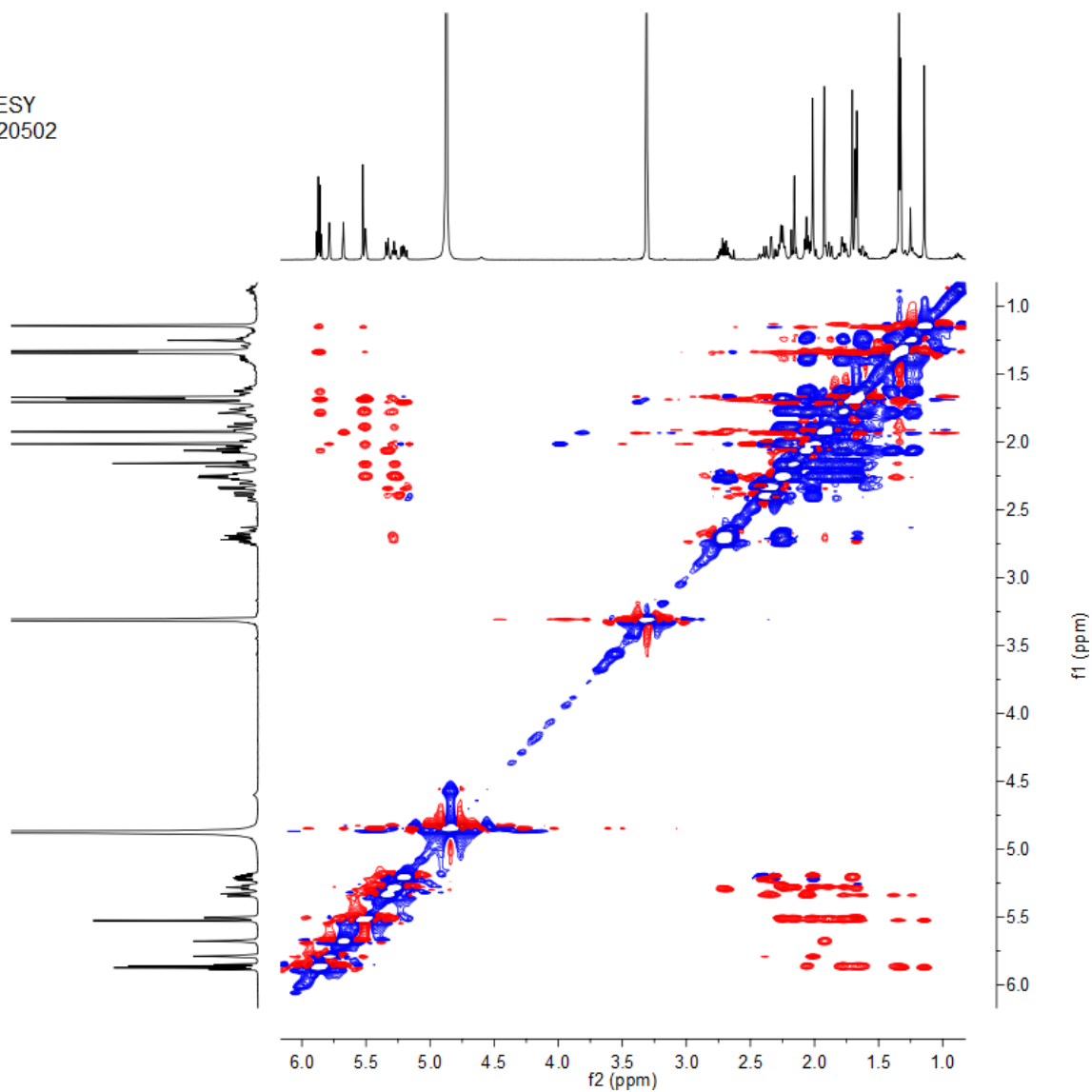


Figure S35. ESI(+)-MS spectrum for aphanamene H (4).

### Display Report

#### Analysis Info

Analysis Name 006-0901.D  
Method Copy of DSOPMS2P.M  
Sample Name yjm-AP-58-1  
Comment DA

Acquisition Date 04/27/12 17:52:12  
Operator Administrator  
Instrument esquire3000plus

#### Acquisition Parameter

Ion Source Type	ESI	Ion Polarity	Positive	Alternating Ion Polarity	off
Mass Range Mode	Std/Normal	Scan Begin	100 m/z	Scan End	1750 m/z
Capillary Exit	158.5 Volt	Skim 1	40.0 Volt	Trap Drive	85.4
Accumulation Time	15000 罫	Averages	3 Spectra	Auto MS/MS	on

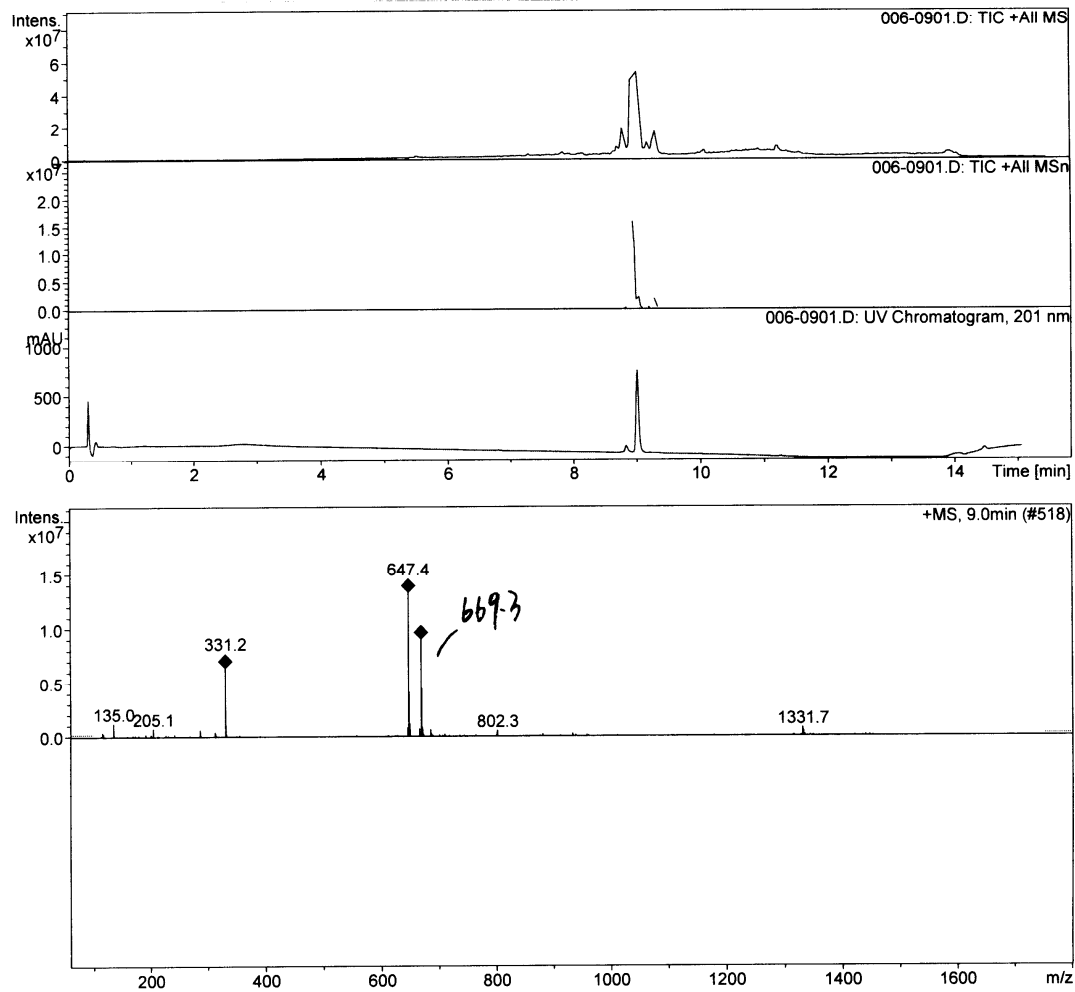


Figure S36. ESI(-)MS spectrum for aphanamene H (4).

### Display Report

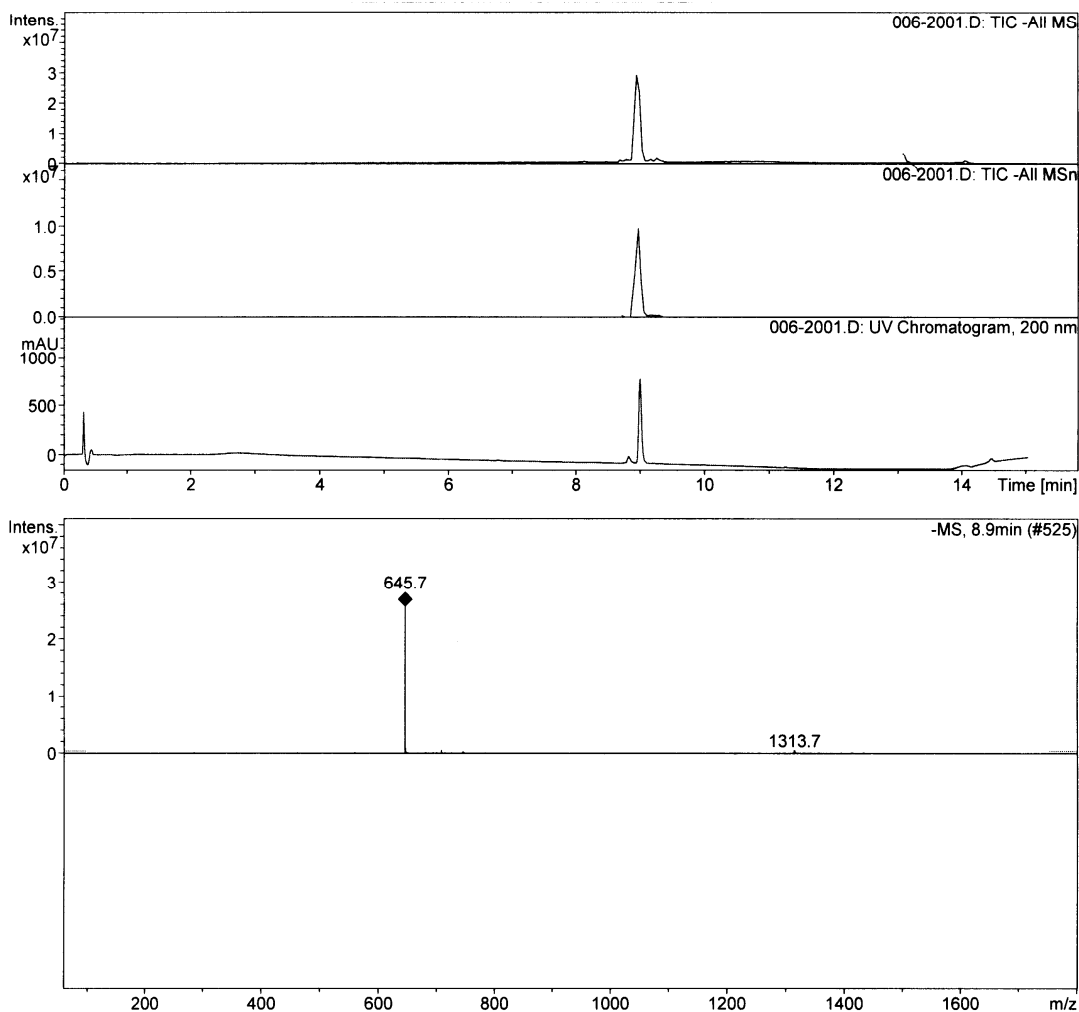
#### Analysis Info

Analysis Name 006-2001.D  
Method Copy of DSOPMS2N.M  
Sample Name yjm-AP-58-1  
Comment DAL

Acquisition Date 04/27/12 20:51:42  
Operator Administrator  
Instrument esquire3000plus

#### Acquisition Parameter

Ion Source Type	ESI	Ion Polarity	Negative	Alternating Ion Polarity	off
Mass Range Mode	Std/Normal	Scan Begin	100 m/z	Scan End	1750 m/z
Capillary Exit	-158.5 Volt	Skim 1	-40.0 Volt	Trap Drive	92.9
Accumulation Time	15000 鏃	Averages	3 Spectra	Auto MS/MS	on



**Figure S37.** HRESI(+)<sup>MS</sup> spectrum for aphanamene H (4).

**Elemental Composition Report**

**Single Mass Analysis**

Tolerance = 3.0 PPM / DBE: min = -1.5, max = 50.0  
 Element prediction: Off  
 Number of isotope peaks used for i-FIT = 3

Monoisotopic Mass, Even Electron Ions  
 173 formula(e) evaluated with 1 results within limits (up to 50 best isotopic matches for each mass)

Elements Used:

C: 10-80 H: 1-110 O: 0-30 Na: 1-1

liujia

LCT PXE KE324

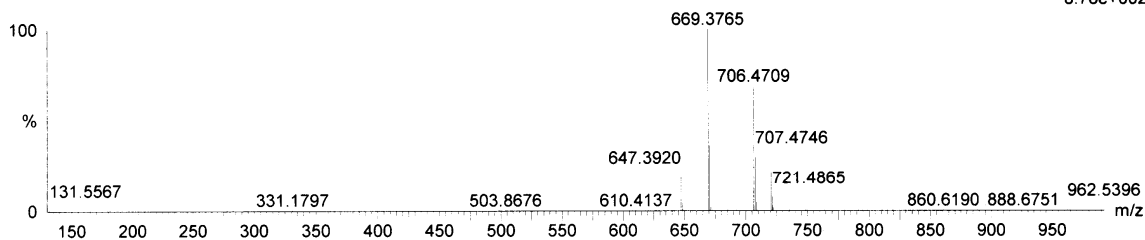
04-May-2012

13:21:50

1: TOF MS ES+

8.78e+002

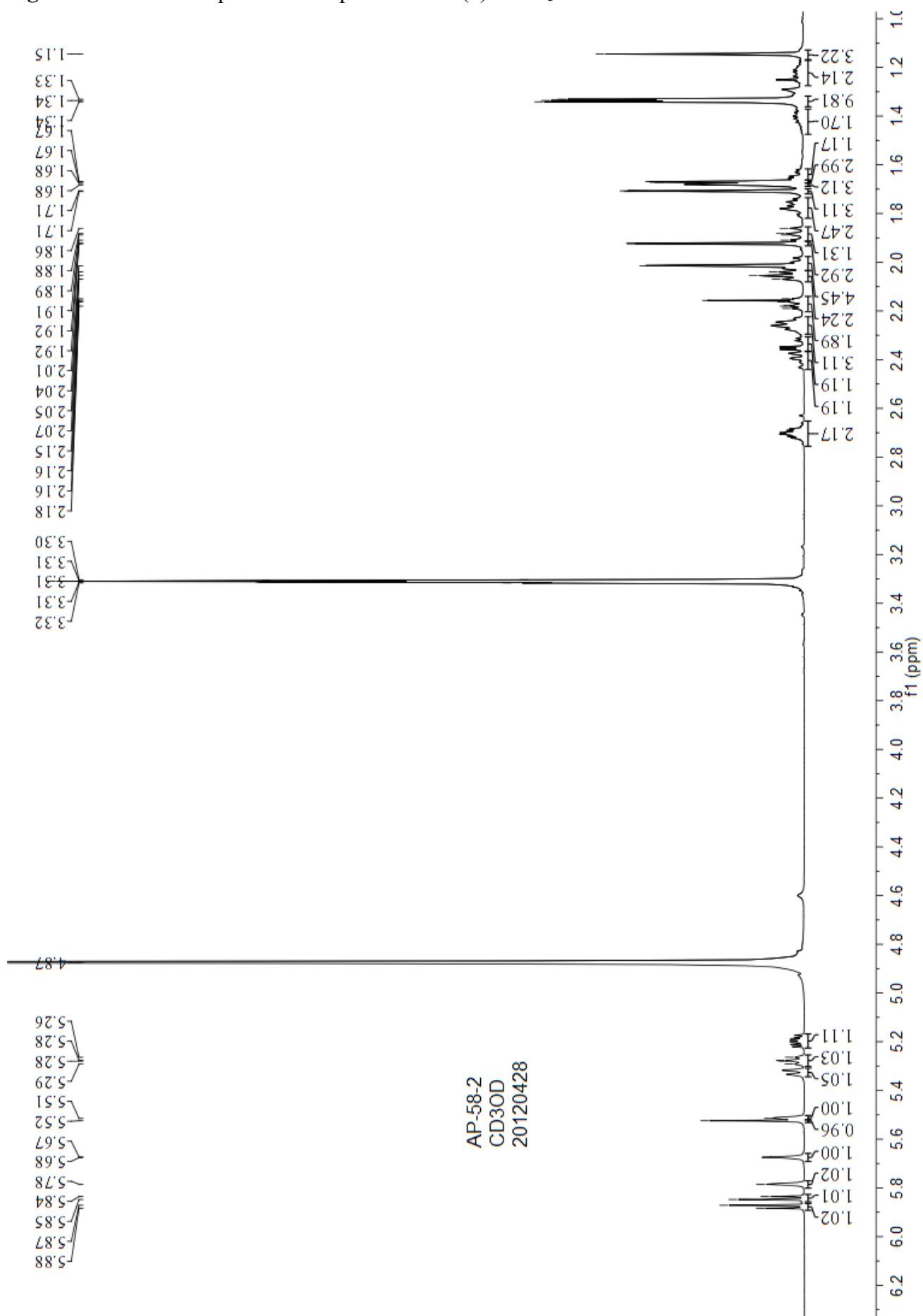
AP\_58\_1p 28 (0.600) AM2 (Ar,10000.0,0.00,1.00); ABS; Cm (25:39)



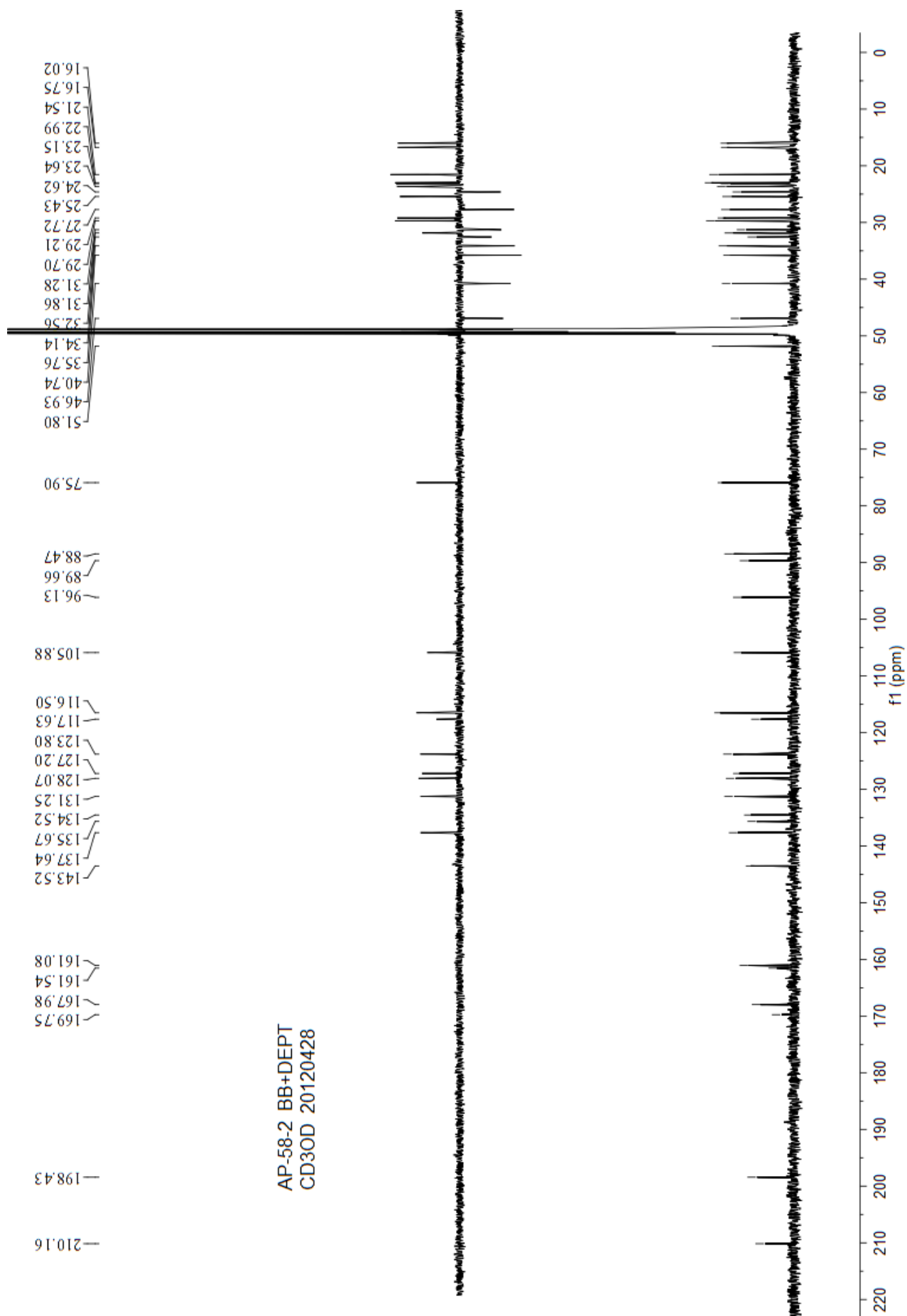
Minimum: -1.5  
 Maximum: 5.0 3.0 50.0

Mass	Calc. Mass	mDa	PPM	DBE	i-FIT	i-FIT (Norm)	Formula
669.3765	669.3767	-0.2	-0.3	13.5	13.8	0.0	C40 H54 O7 Na

**Figure S38.**  $^1\text{H}$  NMR spectrum for aphanamene I (**5**) in  $\text{CD}_3\text{OD}$ .

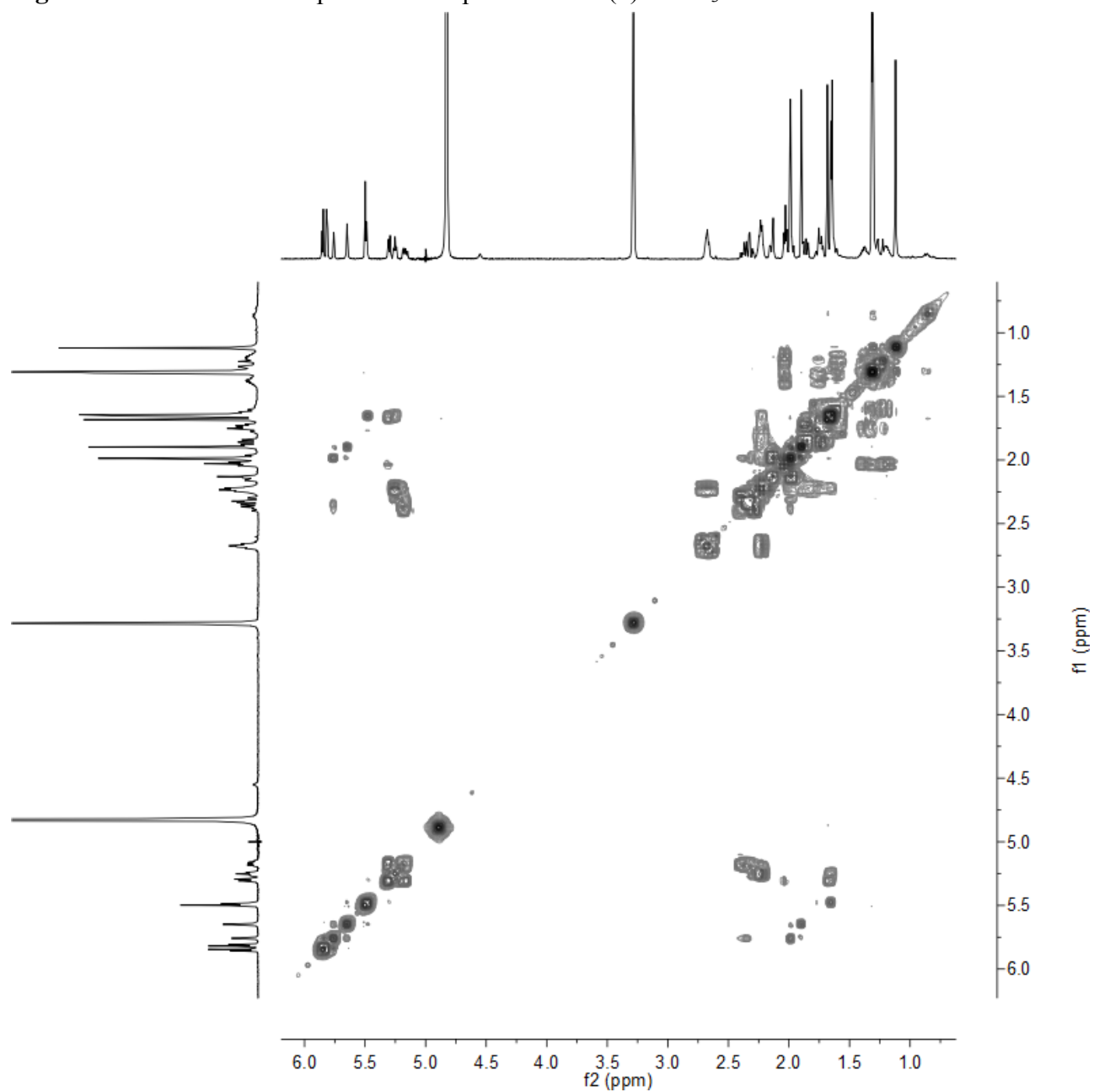


**Figure S39.**  $^{13}\text{C}$  NMR spectrum for aphanamene I (**5**) in  $\text{CD}_3\text{OD}$ .





**Figure S40.**  $^1\text{H}$ - $^1\text{H}$  COSY spectrum for aphanamene I (**5**) in  $\text{CD}_3\text{OD}$ .



**Figure S41.** HSQC spectrum for aphanamene I (**5**) in CD<sub>3</sub>OD.

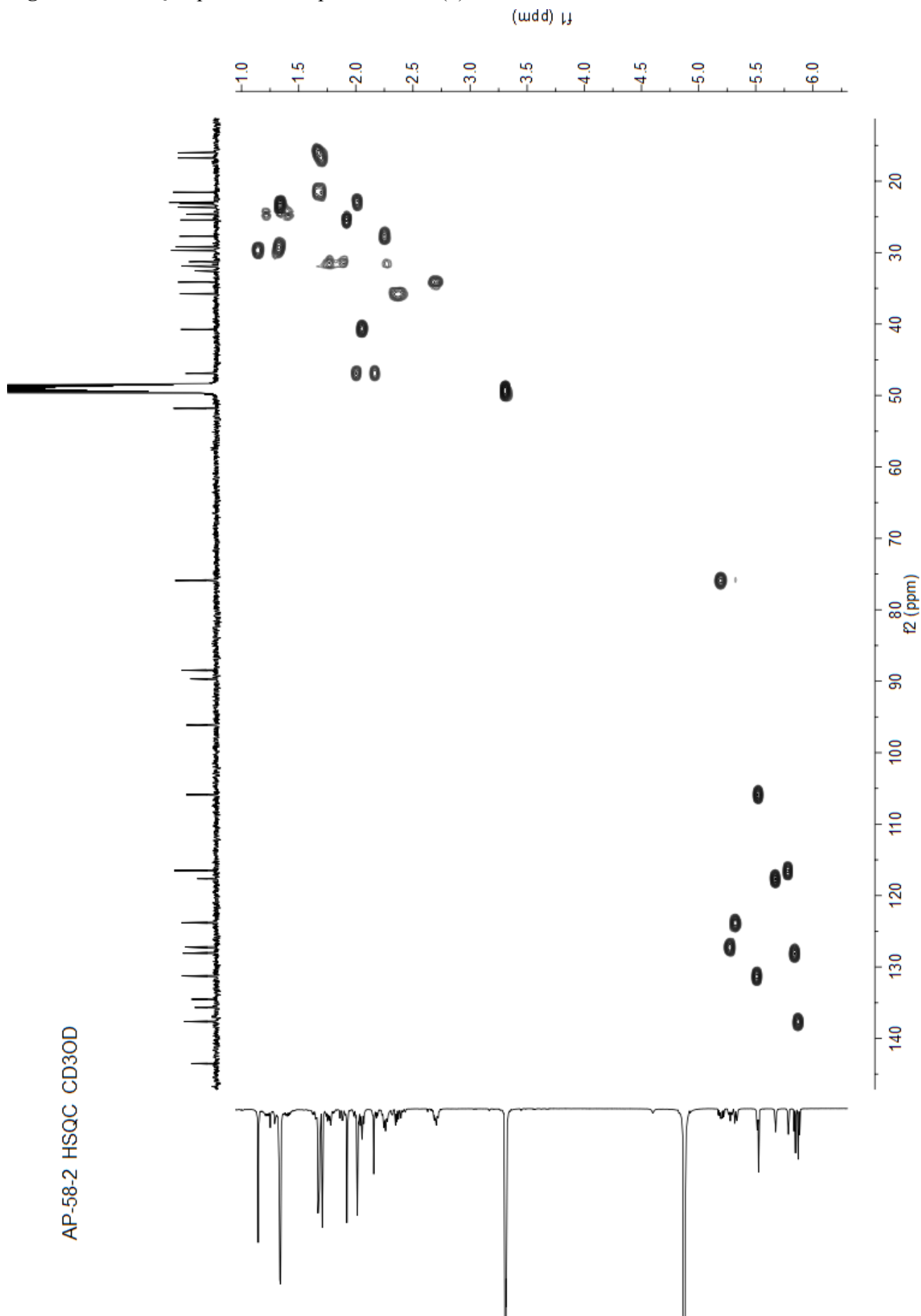
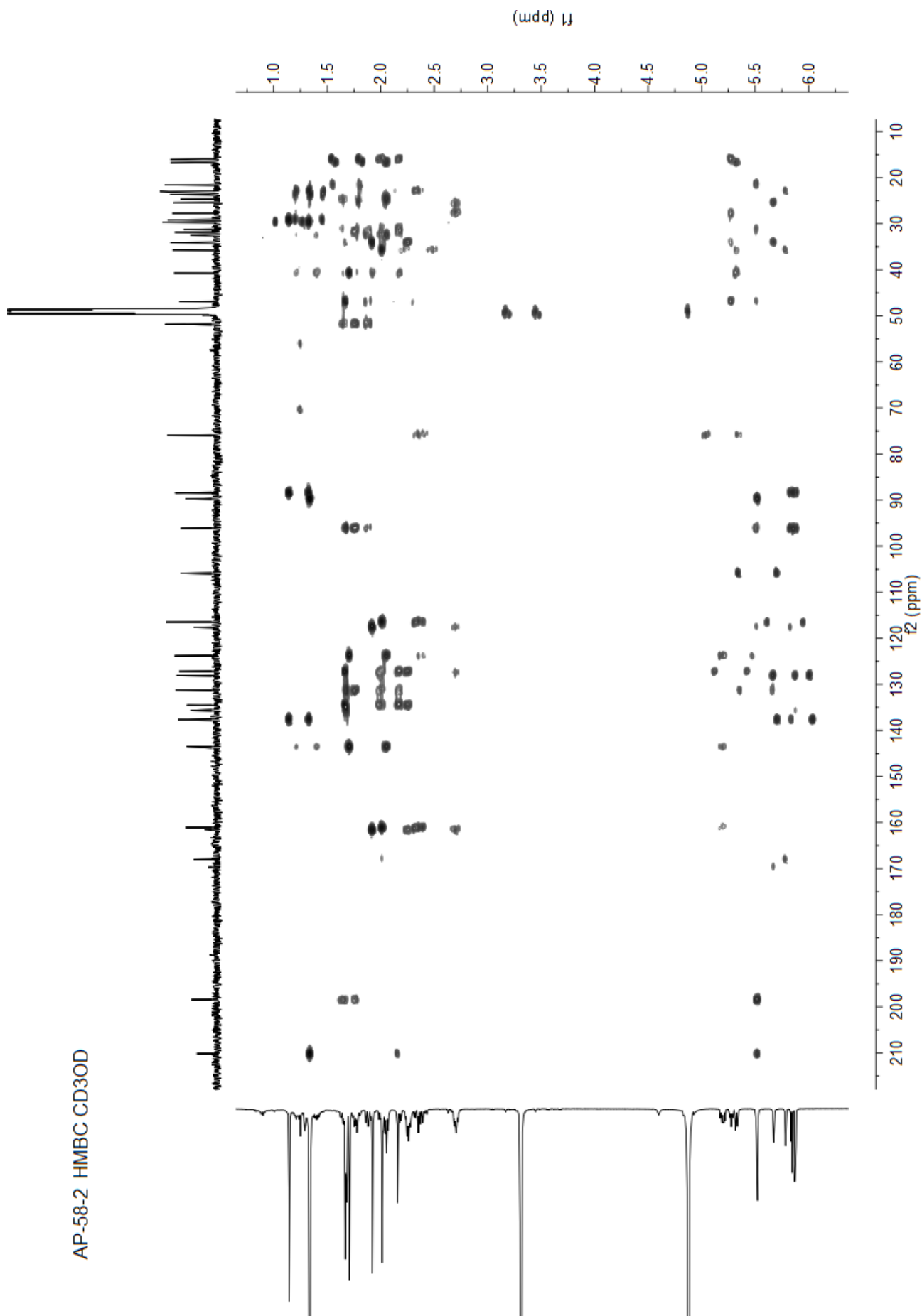
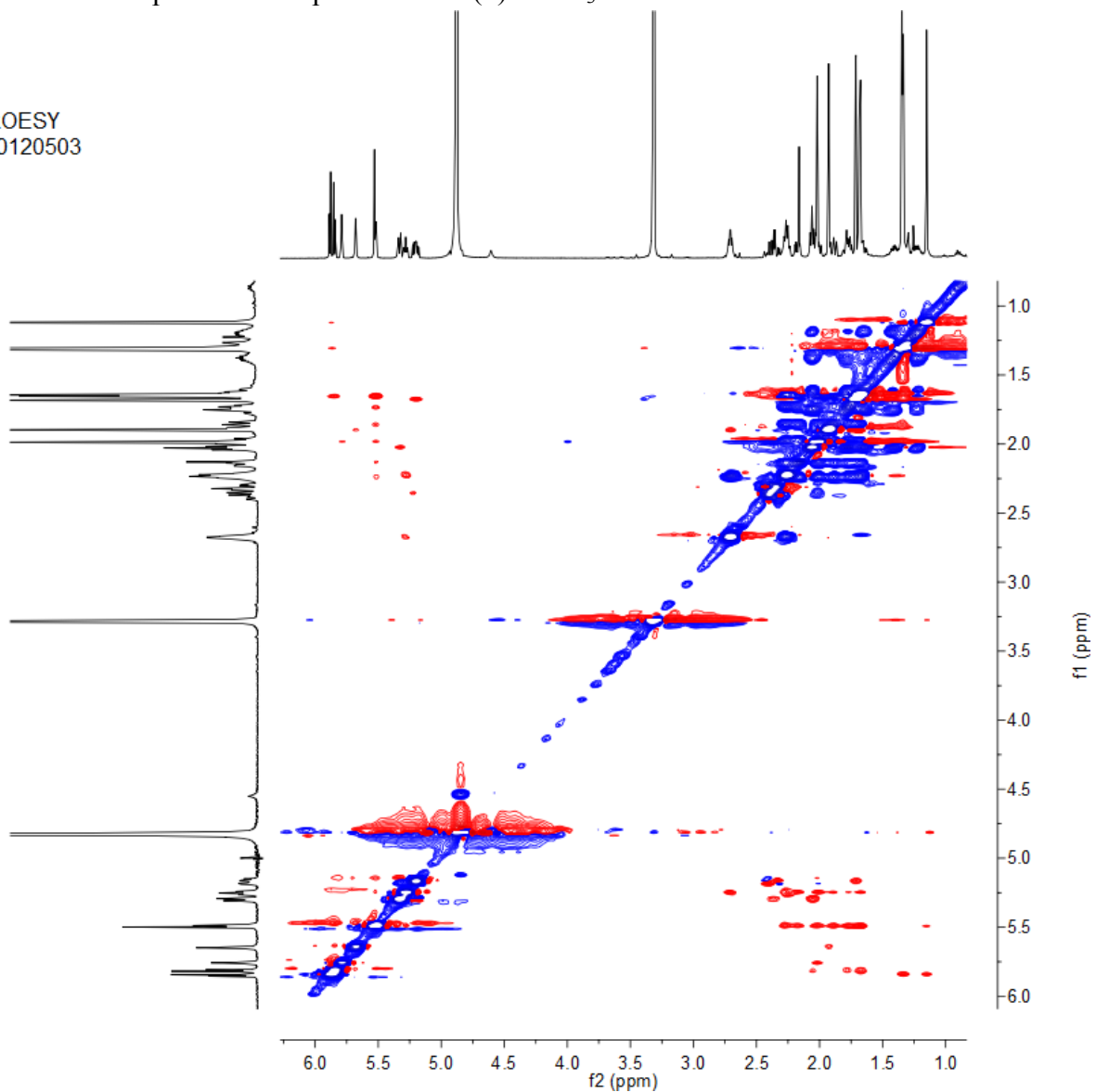


Figure S42. HMBC spectrum for aphanamene I (5) in CD<sub>3</sub>OD.



**Figure S43.** ROESY spectrum for aphanamene I (**5**) in CD<sub>3</sub>OD.

AP-58-2 ROESY  
CD3OD 20120503



**Figure S44.** ESI(+)<sup>MS</sup> spectrum for aphanamene I (5).

### Display Report

**Analysis Info**

Analysis Name 007-1001.D  
 Method Copy of DSOPMS2P.M  
 Sample Name yjm-AP-58-2  
 Comment DA1

Acquisition Date 04/27/12 18:08:31  
 Operator Administrator  
 Instrument esquire3000plus

**Acquisition Parameter**

Ion Source Type	ESI	Ion Polarity	Positive	Alternating Ion Polarity	off
Mass Range Mode	Std/Normal	Scan Begin	100 m/z	Scan End	1750 m/z
Capillary Exit	158.5 Volt	Skim 1	40.0 Volt	Trap Drive	85.4
Accumulation Time	15000 罫	Averages	3 Spectra	Auto MS/MS	on

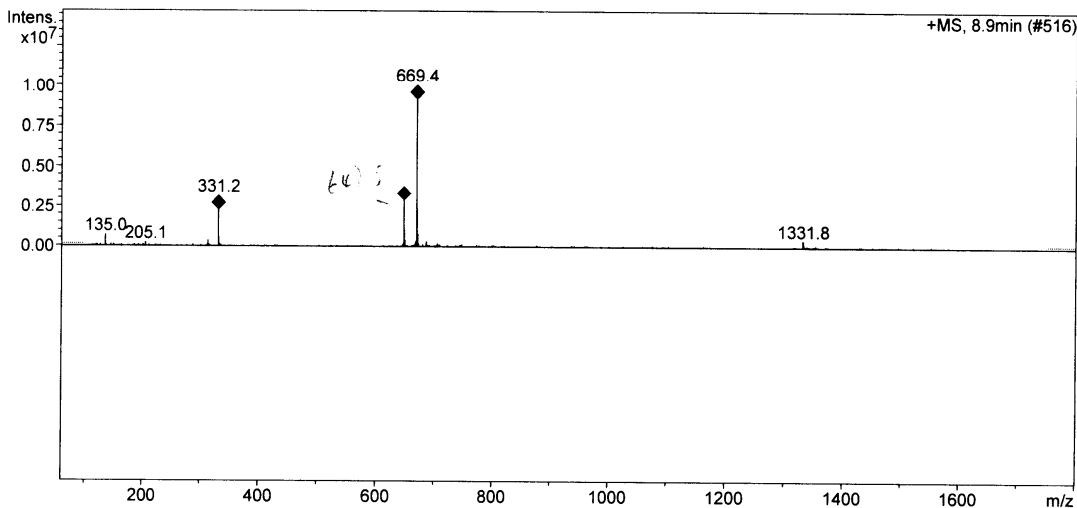
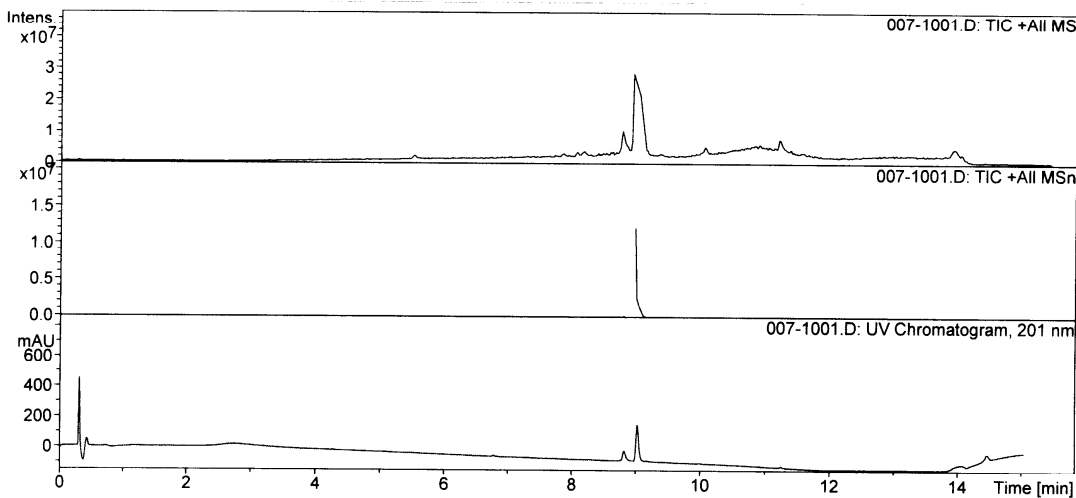


Figure S45. ESI(-)MS spectrum for aphanamene I (5).

### Display Report

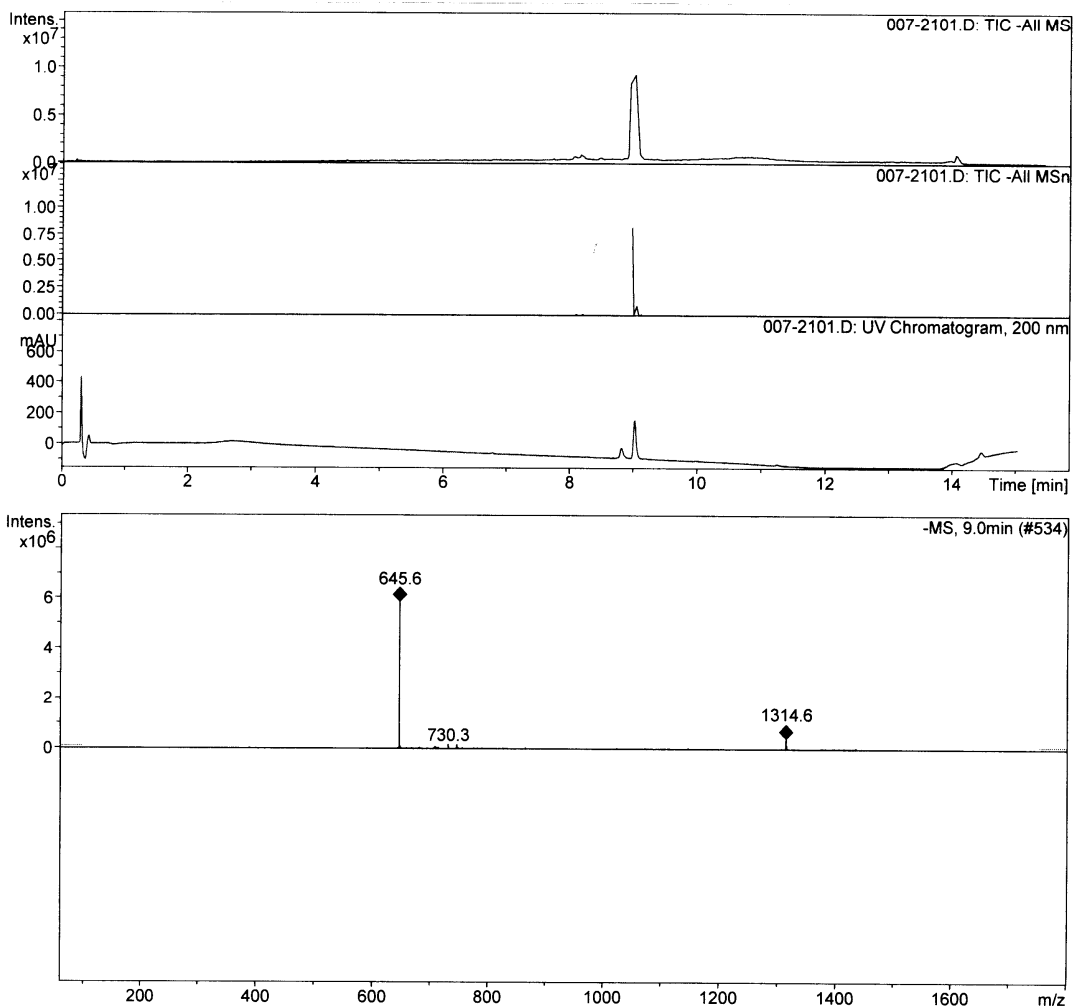
#### Analysis Info

Analysis Name 007-2101.D  
Method Copy of DSOPMS2N.M  
Sample Name yjm-AP-58-2  
Comment DAI

Acquisition Date 04/27/12 21:08:00  
Operator Administrator  
Instrument esquire3000plus

#### Acquisition Parameter

Ion Source Type	ESI	Ion Polarity	Negative	Alternating Ion Polarity	off
Mass Range Mode	Std/Normal	Scan Begin	100 m/z	Scan End	1750 m/z
Capillary Exit	-158.5 Volt	Skim 1	-40.0 Volt	Trap Drive	92.9
Accumulation Time	15000 罫	Averages	3 Spectra	Auto MS/MS	on



**Figure S46.** HRESI(+)-MS spectrum for aphanamene I (**5**).

**Elemental Composition Report**

**Single Mass Analysis**

Tolerance = 5.0 PPM / DBE: min = -1.5, max = 50.0  
 Element prediction: Off  
 Number of isotope peaks used for i-FIT = 3

Monoisotopic Mass, Even Electron Ions

173 formula(e) evaluated with 1 results within limits (up to 50 best isotopic matches for each mass)

Elements Used:

C: 10-80 H: 1-110 O: 0-30 Na: 1-1

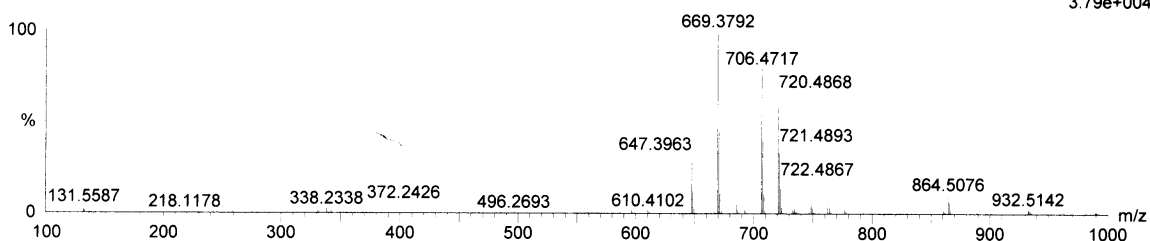
AP-58-2

LCT PXE KE324

04-May-2012

AP-58-2\_20120504 71 (1.555) AM2 (Ar,12000.0,0.00,0.70); ABS; Cm (53:84)

13:42:14  
 1: TOF MS ES+  
 3.79e+004



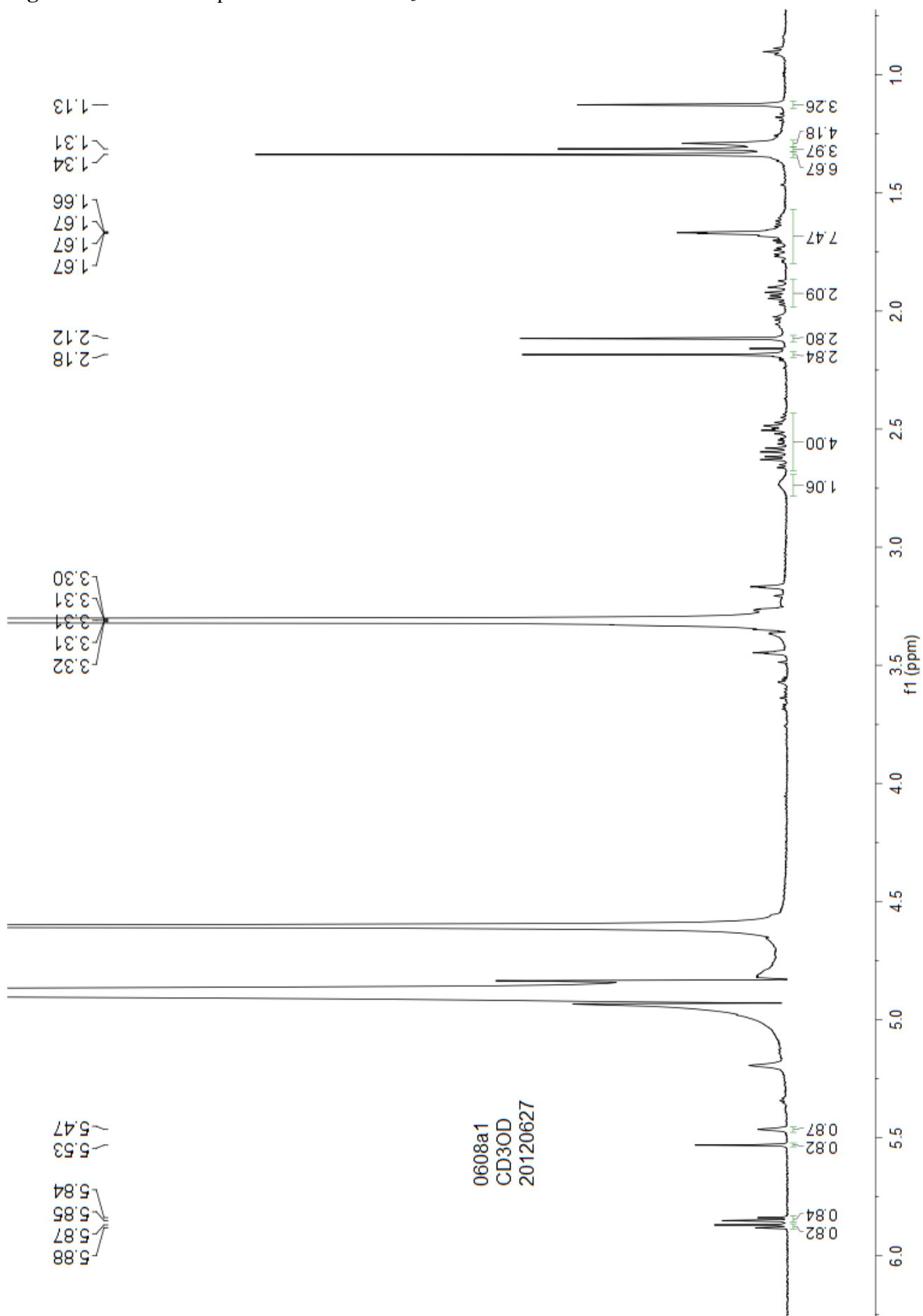
Minimum:

Maximum: 5.0 5.0 -1.5

50.0

Mass	Calc. Mass	mDa	PPM	DBE	i-FIT	i-FIT (Norm)	Formula
669.3792	669.3767	2.5	3.7	13.5	98.2	0.0	C40 H54 O7 Na

**Figure S47.**  $^1\text{H}$  NMR spectrum for **4a** in  $\text{CD}_3\text{OD}$ .





**Figure S48.**  $^{13}\text{C}$  NMR spectrum for **4a** in  $\text{CD}_3\text{OD}$ .

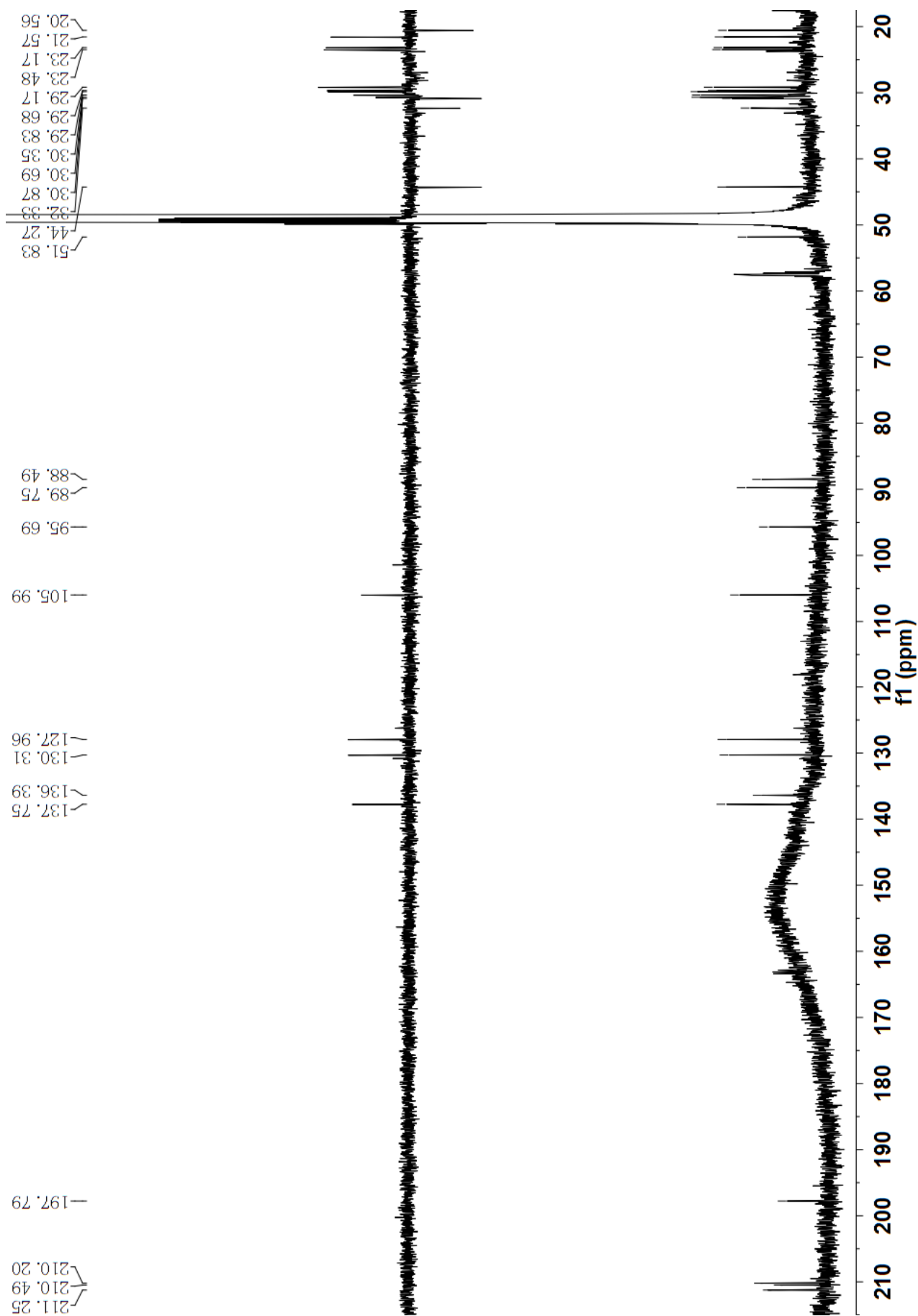


Figure S49. HSQC spectrum for **4a** in CD<sub>3</sub>OD.

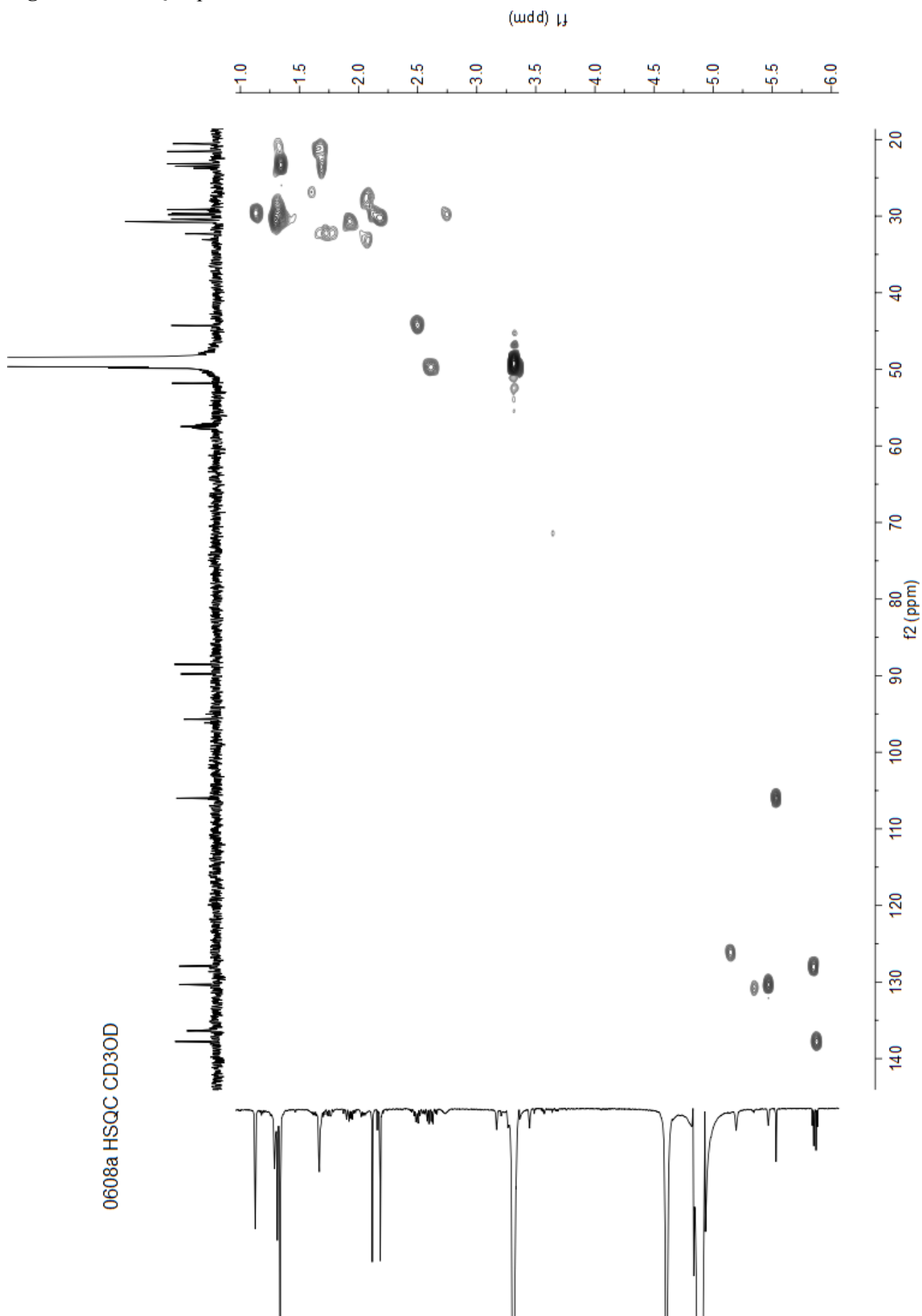


Figure S50. HMBC spectrum for **4a** in CD<sub>3</sub>OD.

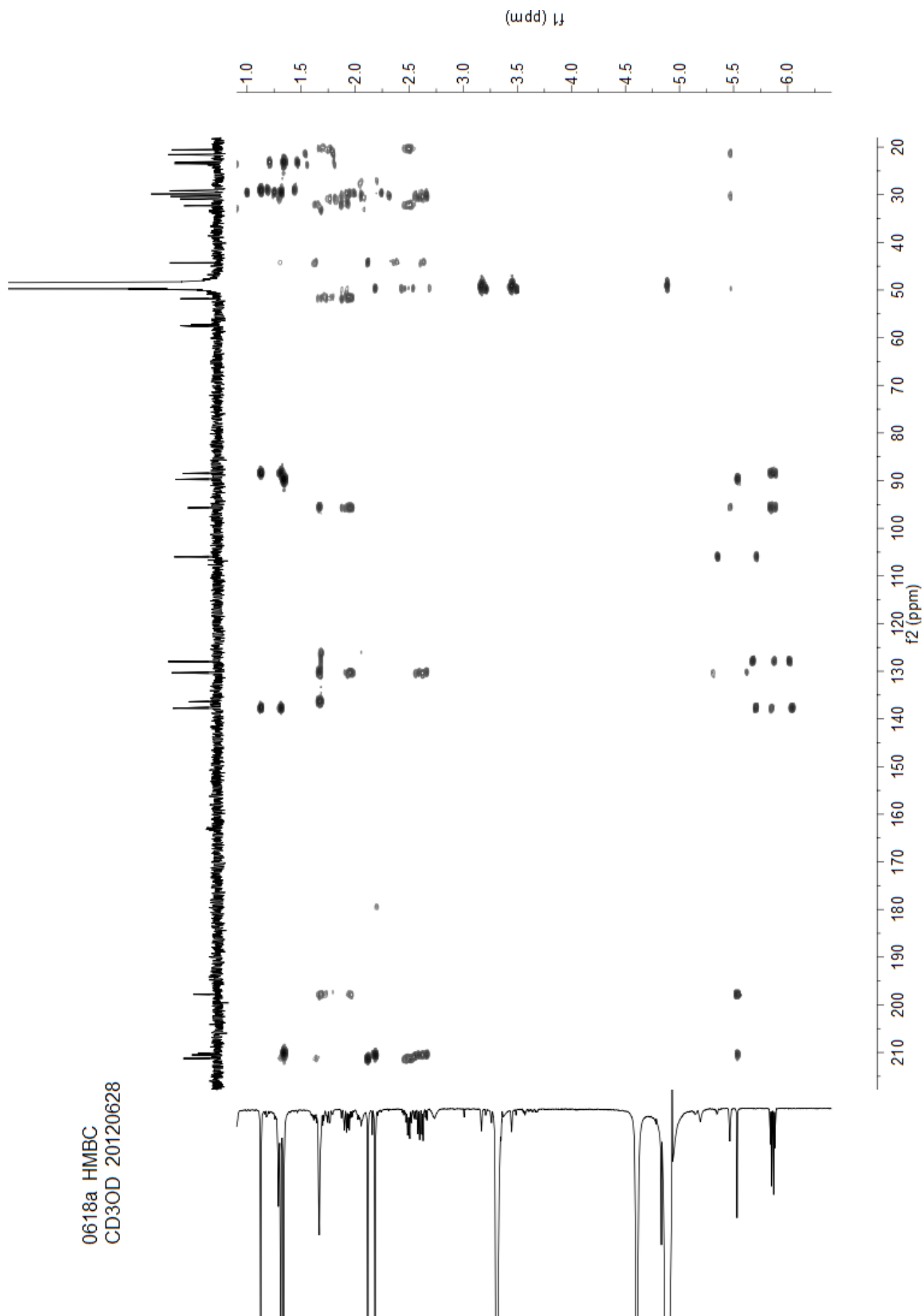


Figure S51. ESI(+)MS spectrum for 4a.

### Display Report

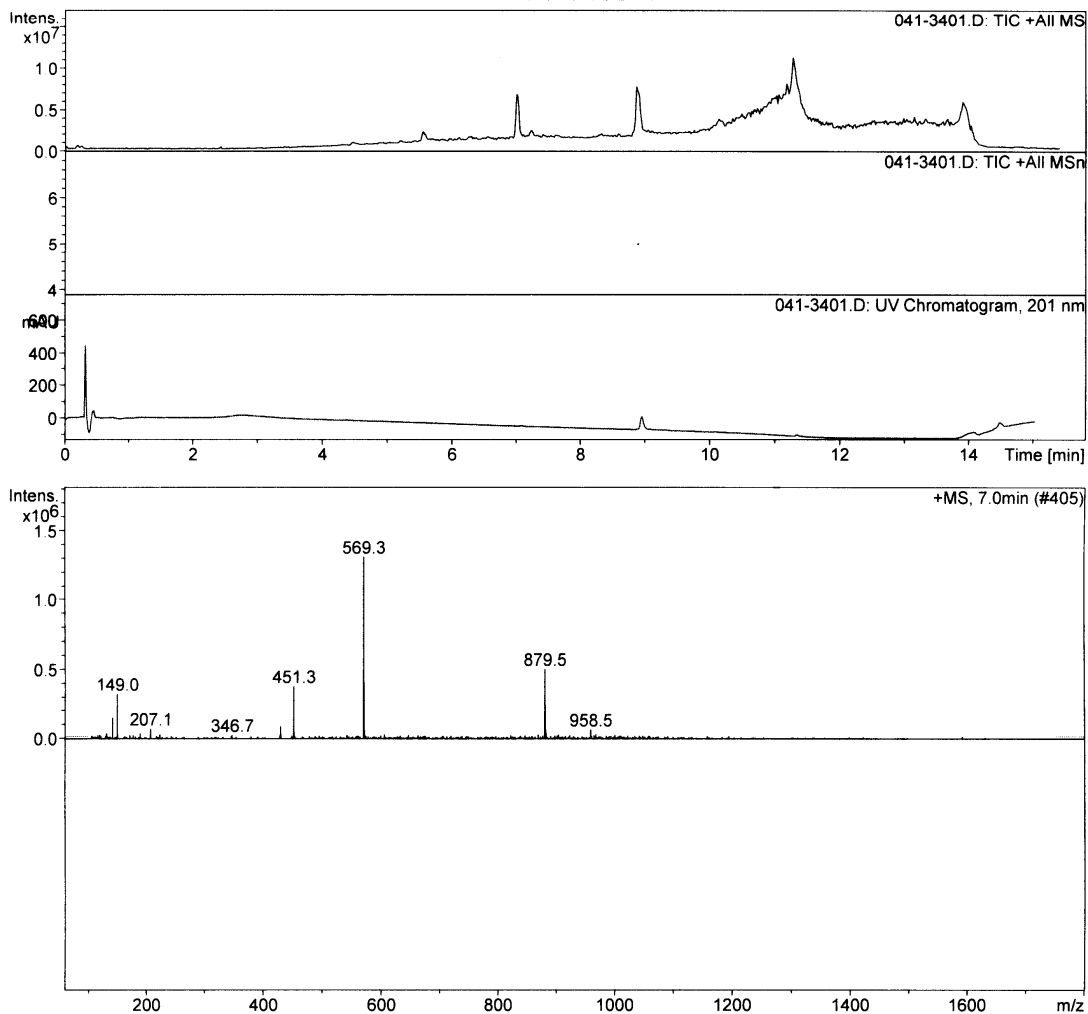
**Analysis Info**

Analysis Name 041-3401.D  
Method Copy of DSOPMS2P.M  
Sample Name yjm-0608a  
Comment

Acquisition Date 06/18/12 19:49:10  
Operator Administrator  
Instrument esquire3000plus

**Acquisition Parameter**

Ion Source Type	ESI	Ion Polarity	Positive	Alternating Ion Polarity	off
Mass Range Mode	Std/Normal	Scan Begin	100 m/z	Scan End	1750 m/z
Capillary Exit	158.5 Volt	Skim 1	40.0 Volt	Trap Drive	85.4
Accumulation Time	15000 罫	Averages	3 Spectra	Auto MS/MS	on



**Figure S52. HRESI(+)-MS spectrum for 4a.**

**Elemental Composition Report**

**Single Mass Analysis**

Tolerance = 3.0 PPM / DBE: min = -1.5, max = 50.0  
 Element prediction: Off  
 Number of isotope peaks used for i-FIT = 3

Monoisotopic Mass, Even Electron Ions

179 formula(e) evaluated with 1 results within limits (up to 50 closest results for each mass)

Elements Used:

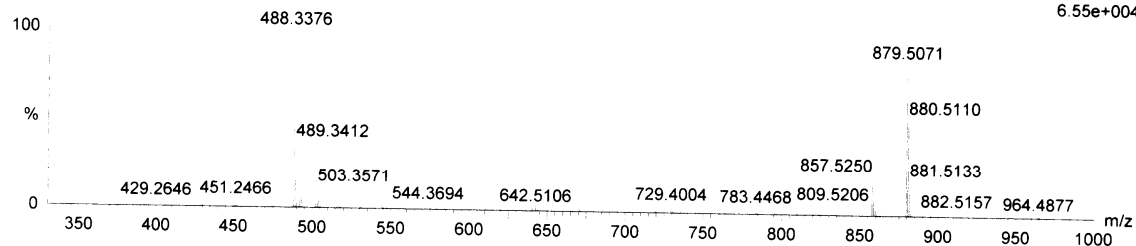
C: 10-60 H: 1-110 O: 0-30 Na: 0-1

LJ

LCT PXE KE324

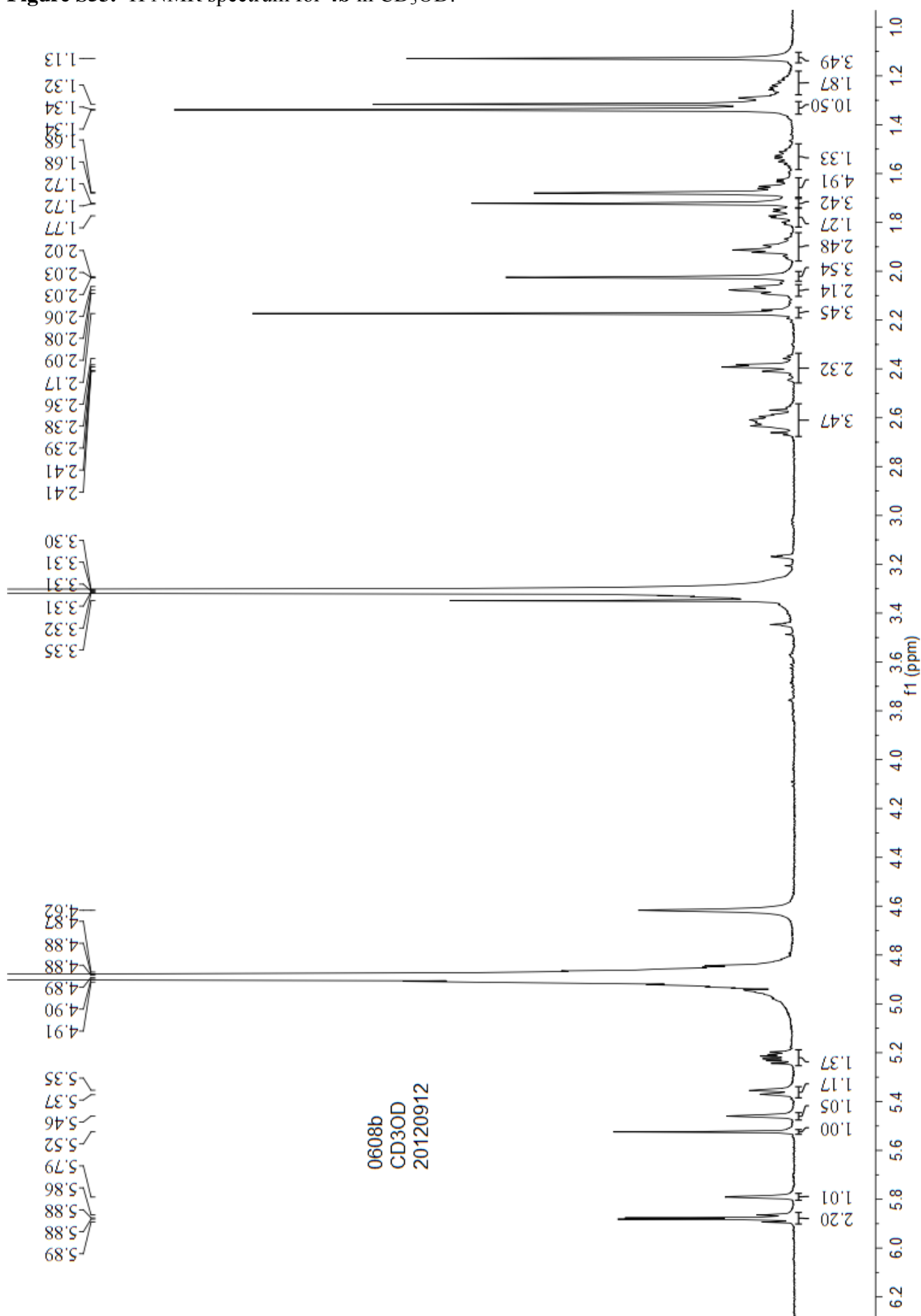
0608a 69 (1.481) AM2 (Ar,12000.0,0.00,0.70); ABS; Cm (50:86)

11-Jun-2012  
 09:09:34  
 1: TOF MS ES+  
 6.55e+004

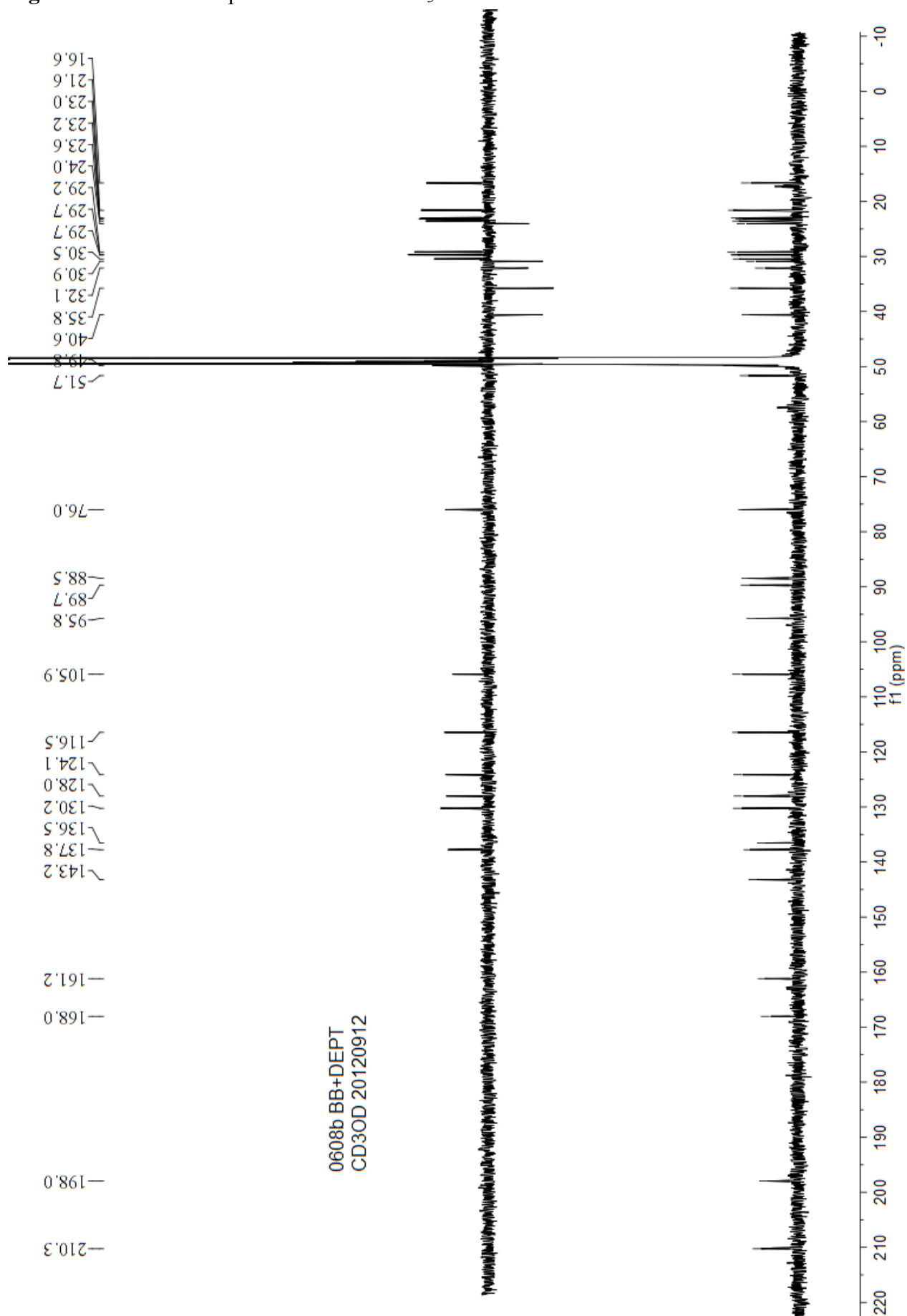


Mass	Calc. Mass	mDa	PPM	DBE	i-FIT	i-FIT (Norm)	Formula
451.2466	451.2460	0.6	1.3	8.5	27.9	0.0	C26 H36 O5 Na
Minimum:				-1.5			
Maximum:			3.0	50.0			

**Figure S53.**  $^1\text{H}$  NMR spectrum for **4b** in  $\text{CD}_3\text{OD}$ .



**Figure S54.**  $^{13}\text{C}$  NMR spectrum for **4b** in  $\text{CD}_3\text{OD}$ .



**Figure S55.** NOESY spectrum for **4b** in CD<sub>3</sub>OD.

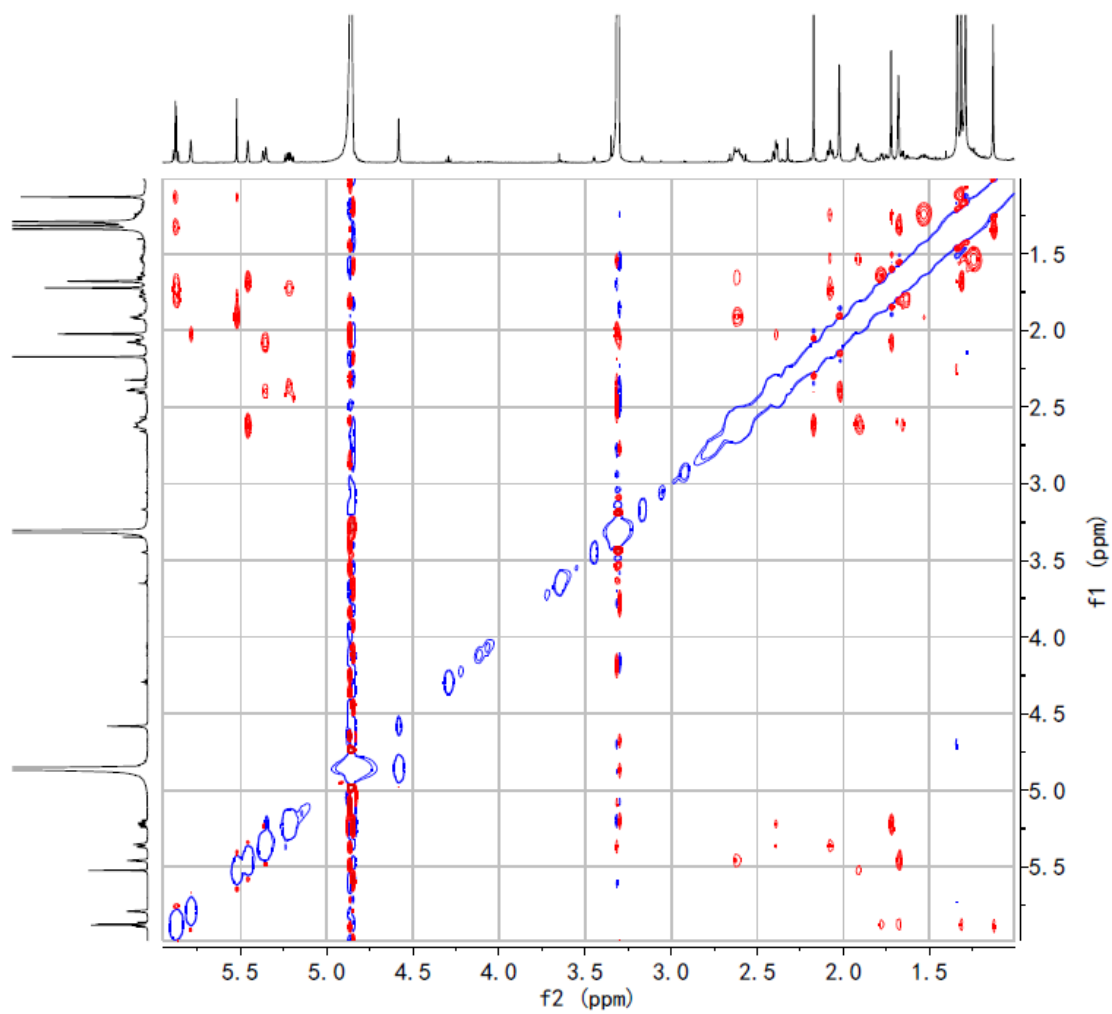




Figure S56. ESI(+)MS spectrum for 4b.

### Display Report

#### Analysis Info

Analysis Name 006-0801.D  
Method Copy of DSOPMS2P.M  
Sample Name yjm-0614b  
Comment 2\

Acquisition Date 09/12/12 11:55:01  
Operator Administrator  
Instrument esquire3000plus

#### Acquisition Parameter

Ion Source Type	ESI	Ion Polarity	Positive	Alternating Ion Polarity	off
Mass Range Mode	Std/Normal	Scan Begin	100 m/z	Scan End	1750 m/z
Capillary Exit	158.5 Volt	Skim 1	40.0 Volt	Trap Drive	85.4
Accumulation Time	15000 經	Averages	3 Spectra	Auto MS/MS	on

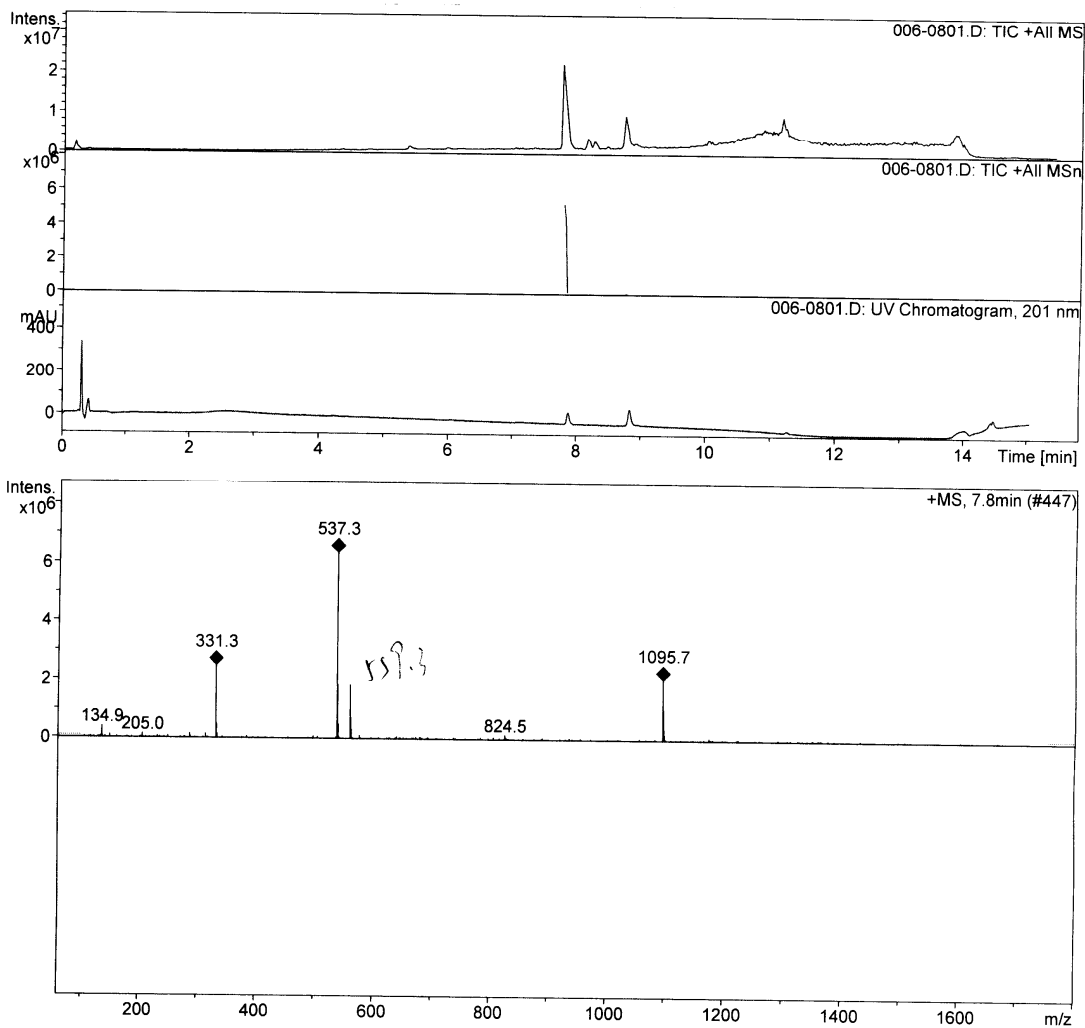


Figure S57. HRESI(+)MS spectrum for 4b.

Elemental Composition Report

Single Mass Analysis

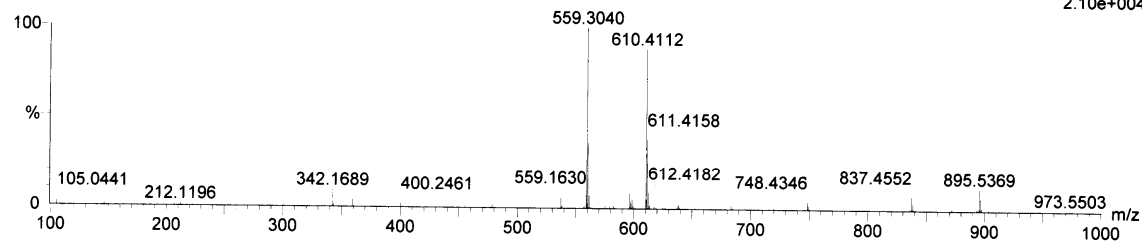
Tolerance = 2.0 PPM / DBE: min = -1.5, max = 50.0  
 Element prediction: Off  
 Number of isotope peaks used for i-FIT = 3

Monoisotopic Mass, Even Electron Ions  
 268 formula(e) evaluated with 1 results within limits (up to 50 closest results for each mass)

Elements Used:  
 C: 6-60 H: 2-110 O: 0-30 Na: 0-1

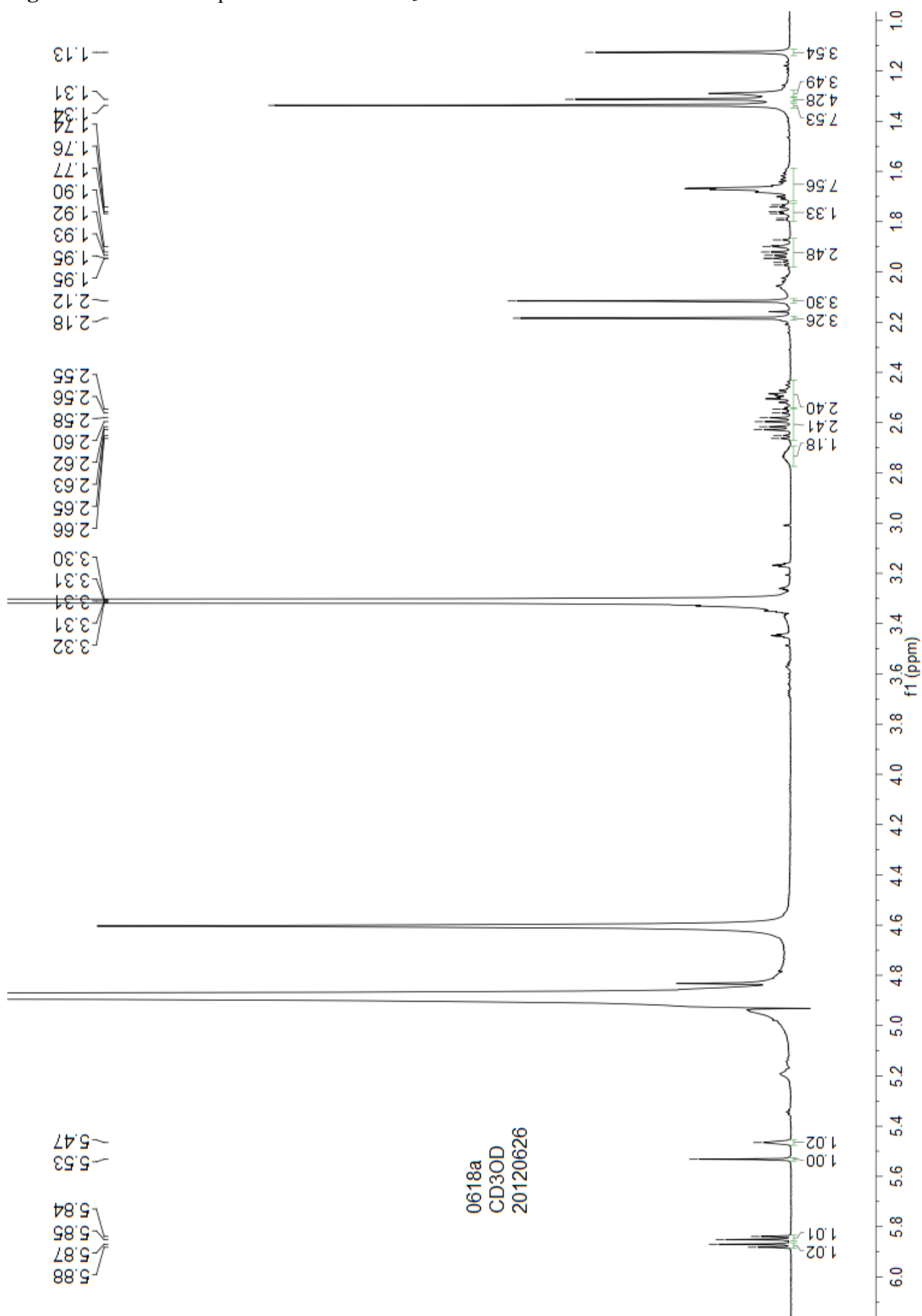
0614b  
 LCT PXE KE324  
 0614b\_20120914 14 (0.283) AM2 (Ar,10000.0,0.00,1.00); ABS; Cm (5:23)

14-Sep-2012  
 10:51:02  
 1: TOF MS ES+  
 2.10e+004



Mass	Calc. Mass	mDa	PPM	DBE	i-FIT	i-FIT (Norm)	Formula
559.3040	559.3036	0.4	0.7	11.5	88.9	0.0	C33 H44 O6 Na

**Figure S58.**  $^1\text{H}$  NMR spectrum for **5a** in  $\text{CD}_3\text{OD}$ .



**Figure S59.**  $^{13}\text{C}$  NMR spectrum for **5a** in  $\text{CD}_3\text{OD}$ .

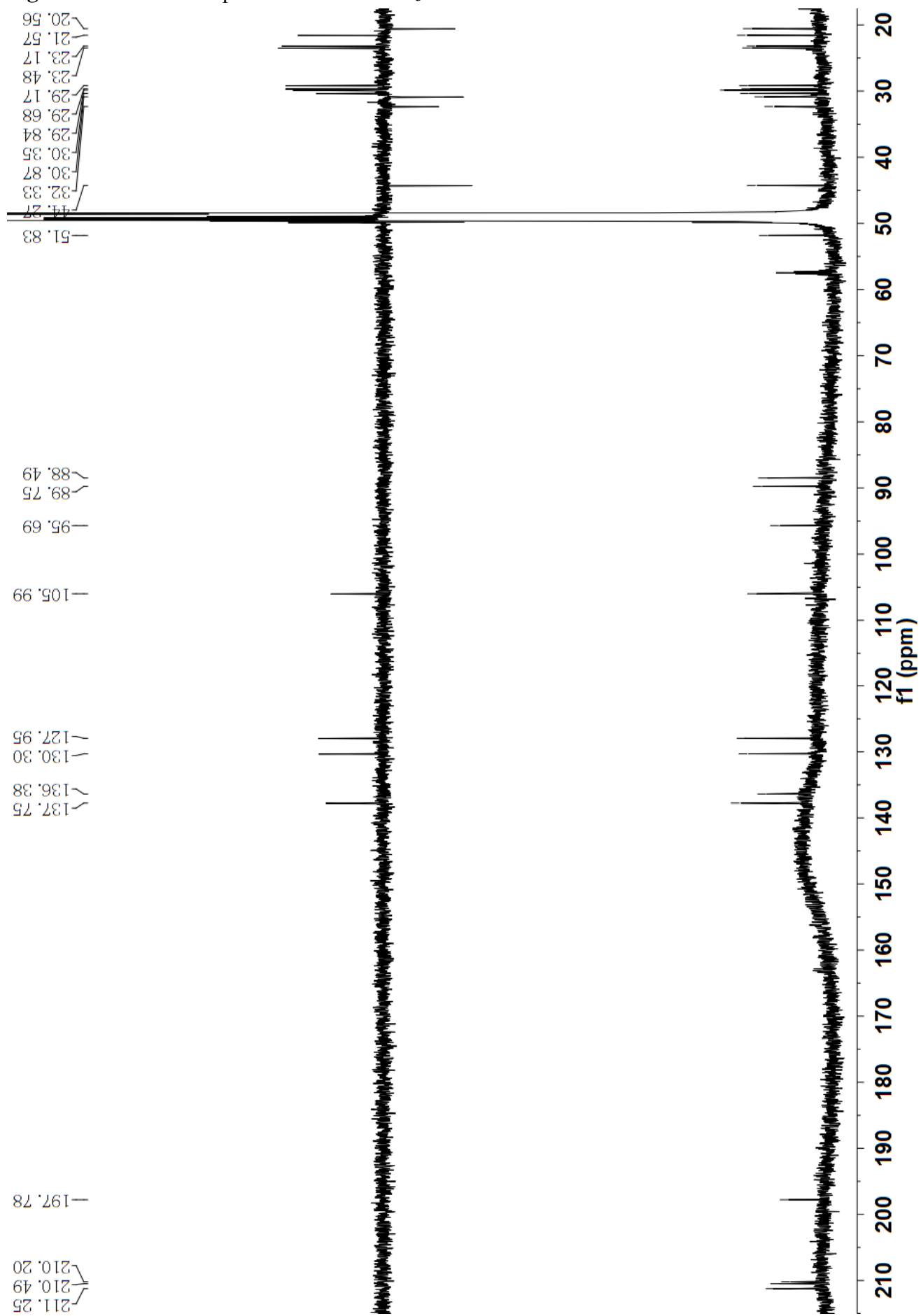


Figure S60. HSQC spectrum for **5a** in CD<sub>3</sub>OD.

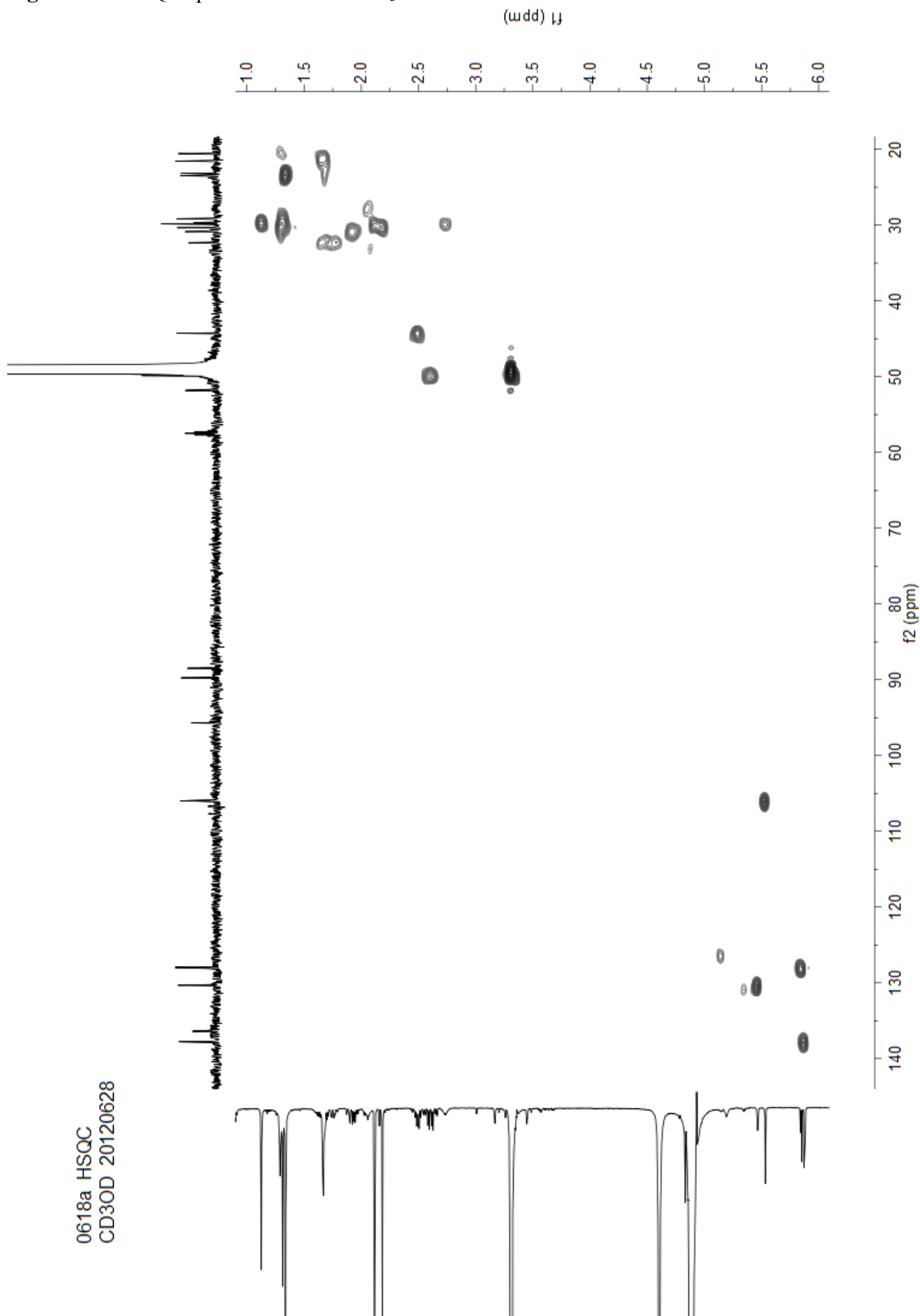


Figure S61. HMBC spectrum for **5a** in CD<sub>3</sub>OD.

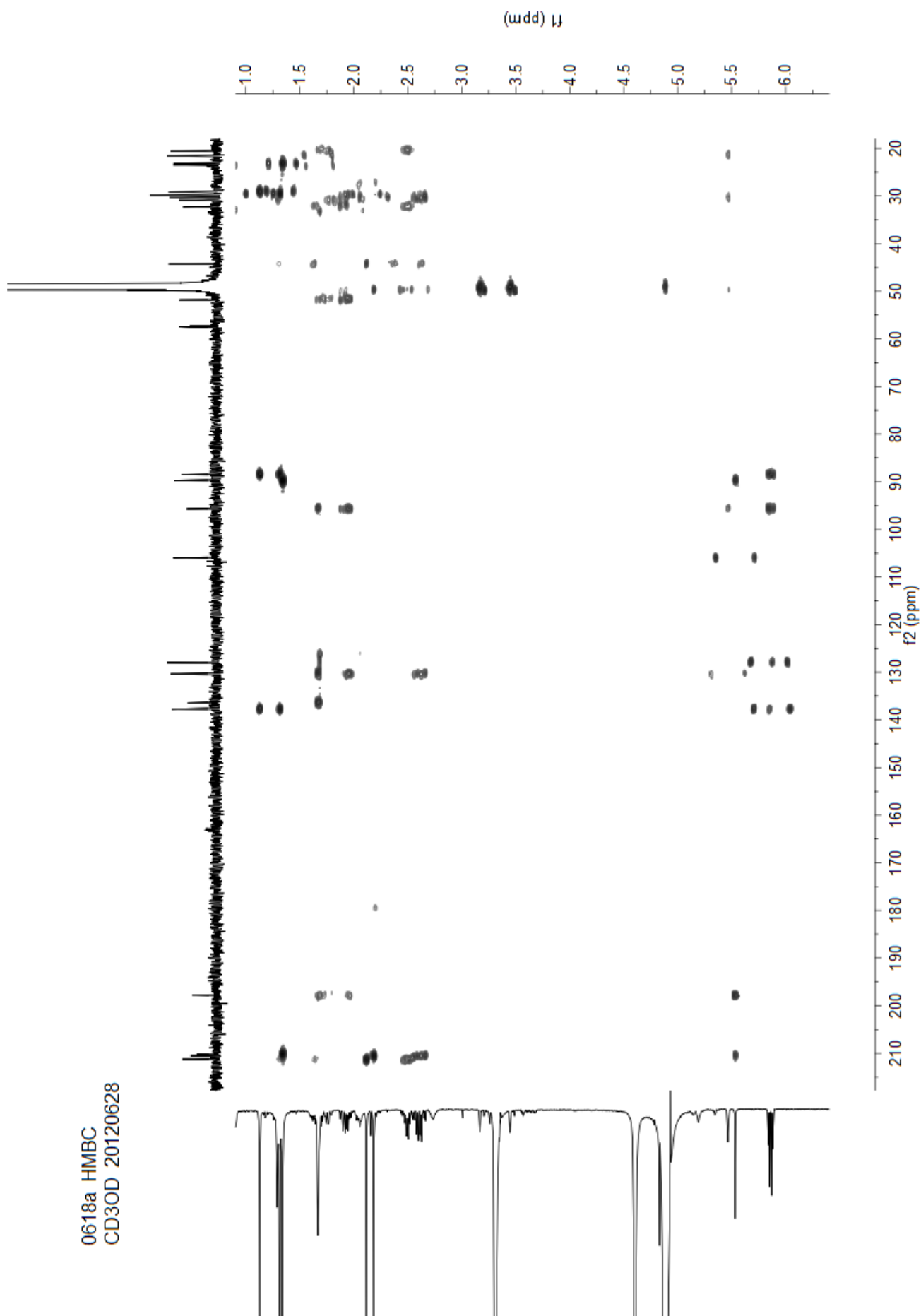


Figure S62. ESI(+)MS spectrum for 5a.

### Display Report

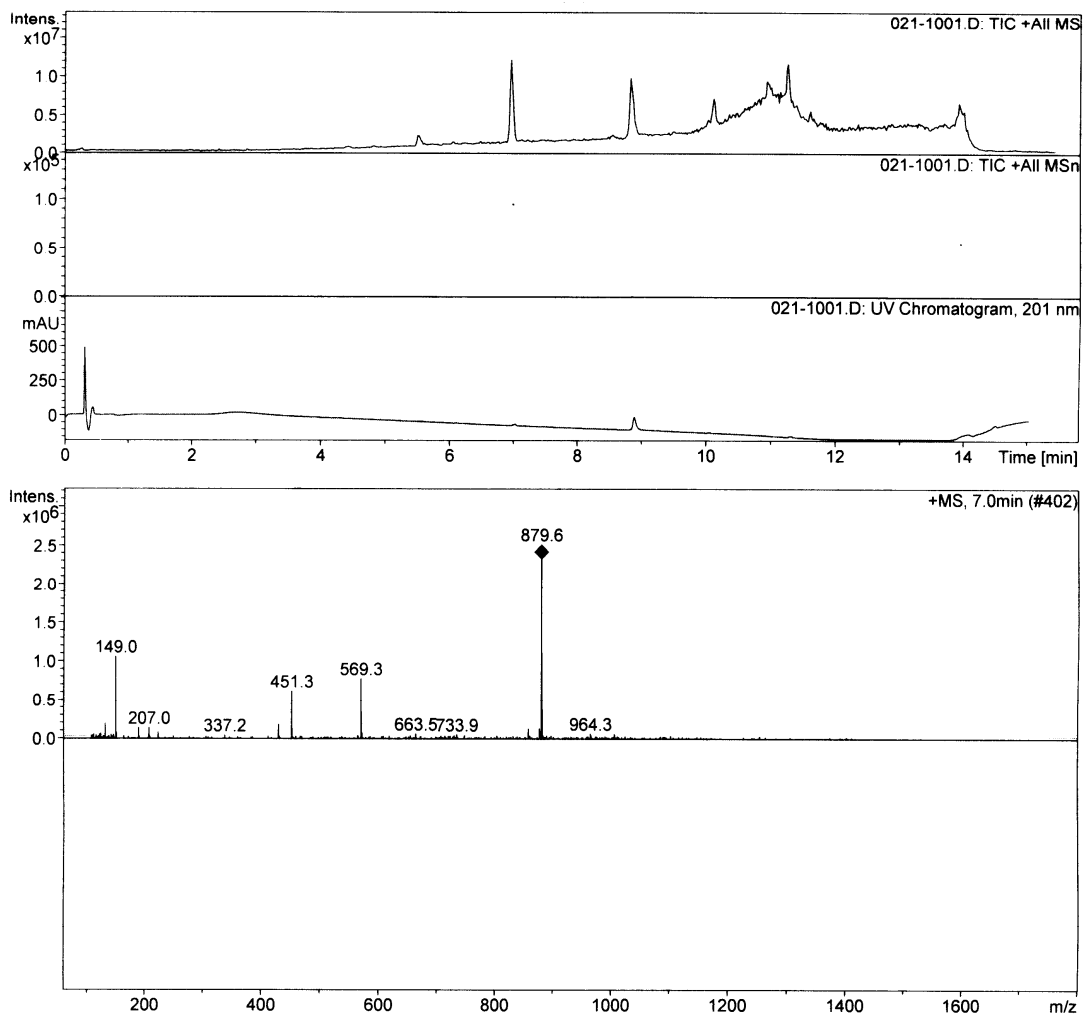
#### Analysis Info

Analysis Name 021-1001.D  
Method Copy of DSOPMS2P.M  
Sample Name yjm-0618a  
Comment v

Acquisition Date 06/28/12 18:11:28  
Operator Administrator  
Instrument esquire3000plus

#### Acquisition Parameter

Ion Source Type	ESI	Ion Polarity	Positive	Alternating Ion Polarity	off
Mass Range Mode	Std/Normal	Scan Begin	100 m/z	Scan End	1750 m/z
Capillary Exit	158.5 Volt	Skim 1	40.0 Volt	Trap Drive	85.4
Accumulation Time	15000 罫	Averages	3 Spectra	Auto MS/MS	on



**Figure S63.** HRESI(+)MS spectrum for **5a**.

**Elemental Composition Report**

**Single Mass Analysis**

Tolerance = 3.0 PPM / DBE: min = -1.5, max = 50.0  
 Element prediction: Off  
 Number of isotope peaks used for i-FIT = 3

Monoisotopic Mass, Even Electron Ions

185 formula(e) evaluated with 1 results within limits (up to 50 closest results for each mass)

Elements Used:

C: 6-60 H: 2-110 O: 0-30 Na: 0-1

LJ

LCT PXE KE324

19-Jul-2012

13:59:20

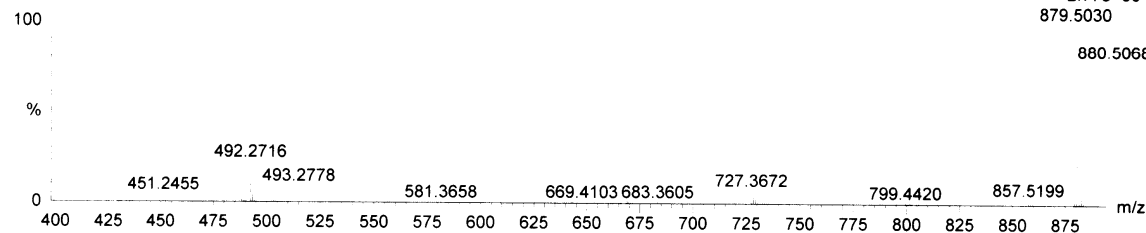
0618a\_20120719 28 (0.600) AM2 (Ar,12500.0,0.00,0.70); ABS; Cm (21:44)

1: TOF MS ES+

2.77e+004

879.5030

880.5068

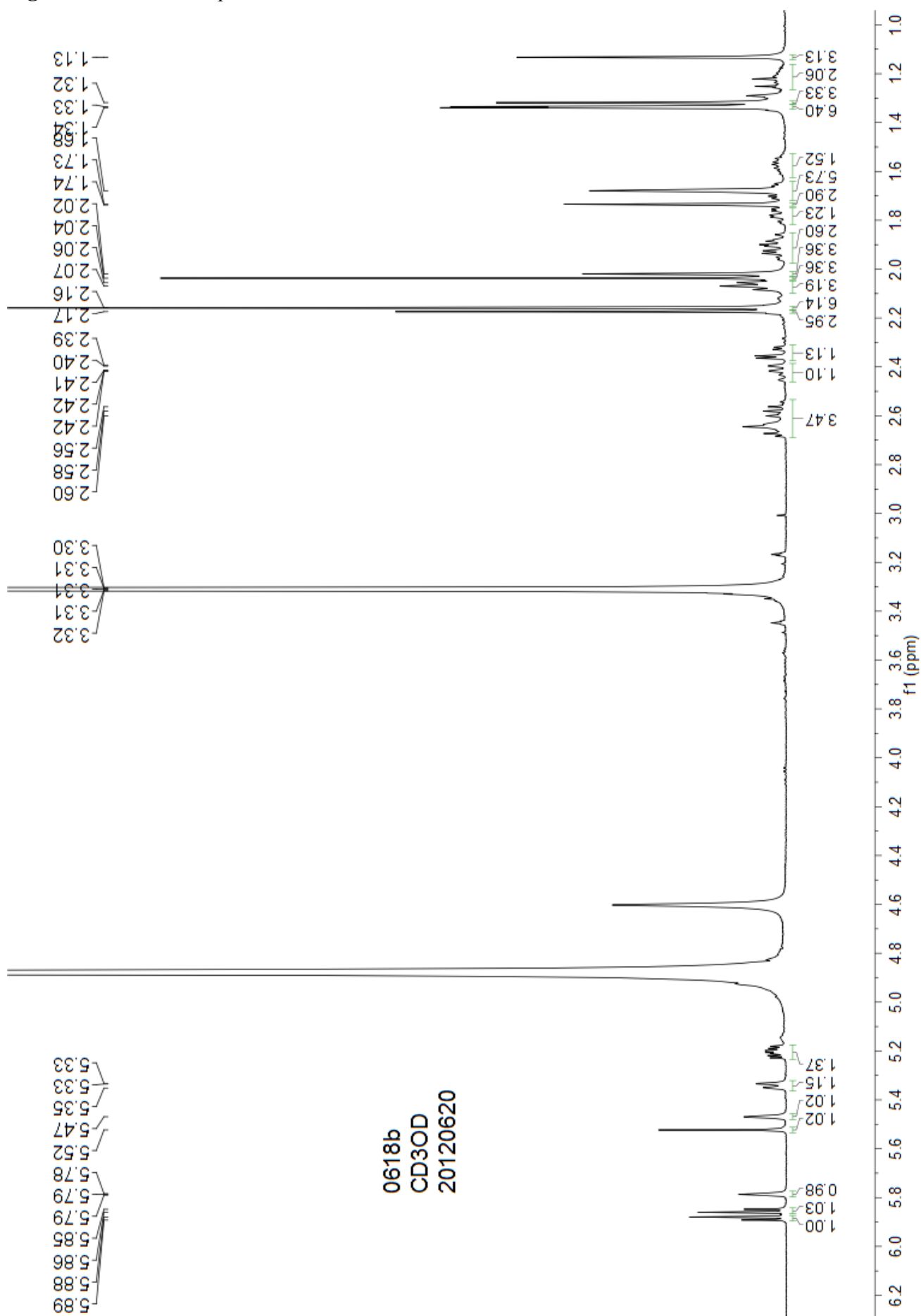


Minimum: -1.5  
 Maximum: 3.0 3.0 50.0

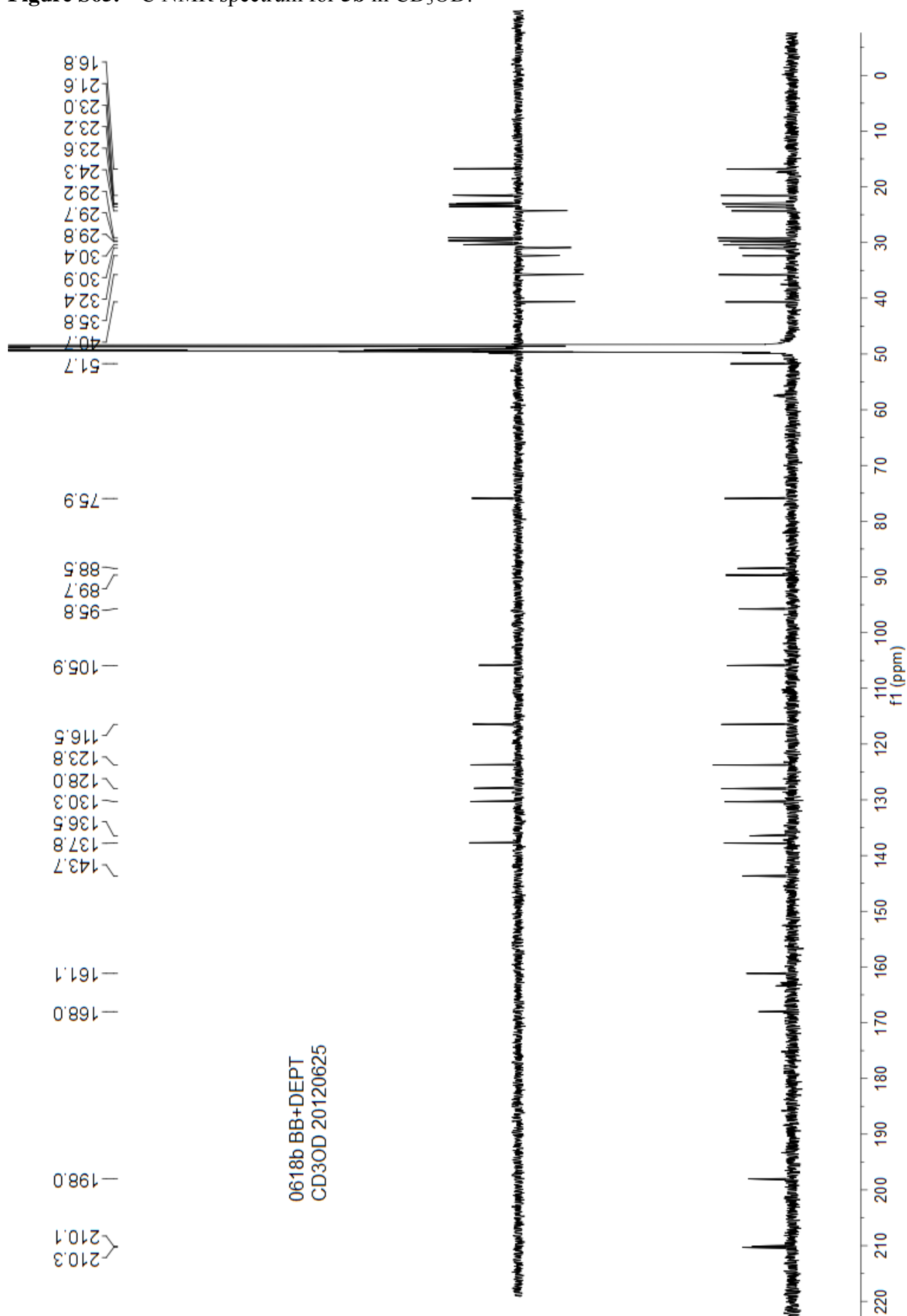
Mass	Calc. Mass	mDa	PPM	DBE	i-FIT	i-FIT (Norm)	Formula
451.2455	451.2460	-0.5	-1.1	8.5	16.1	0.0	C26 H36 O5 Na



**Figure S64.**  $^1\text{H}$  NMR spectrum for **5b** in  $\text{CD}_3\text{OD}$ .



**Figure S65.**  $^{13}\text{C}$  NMR spectrum for **5b** in  $\text{CD}_3\text{OD}$ .



**Figure S66.** NOESY spectrum for **5b** in CD<sub>3</sub>OD.

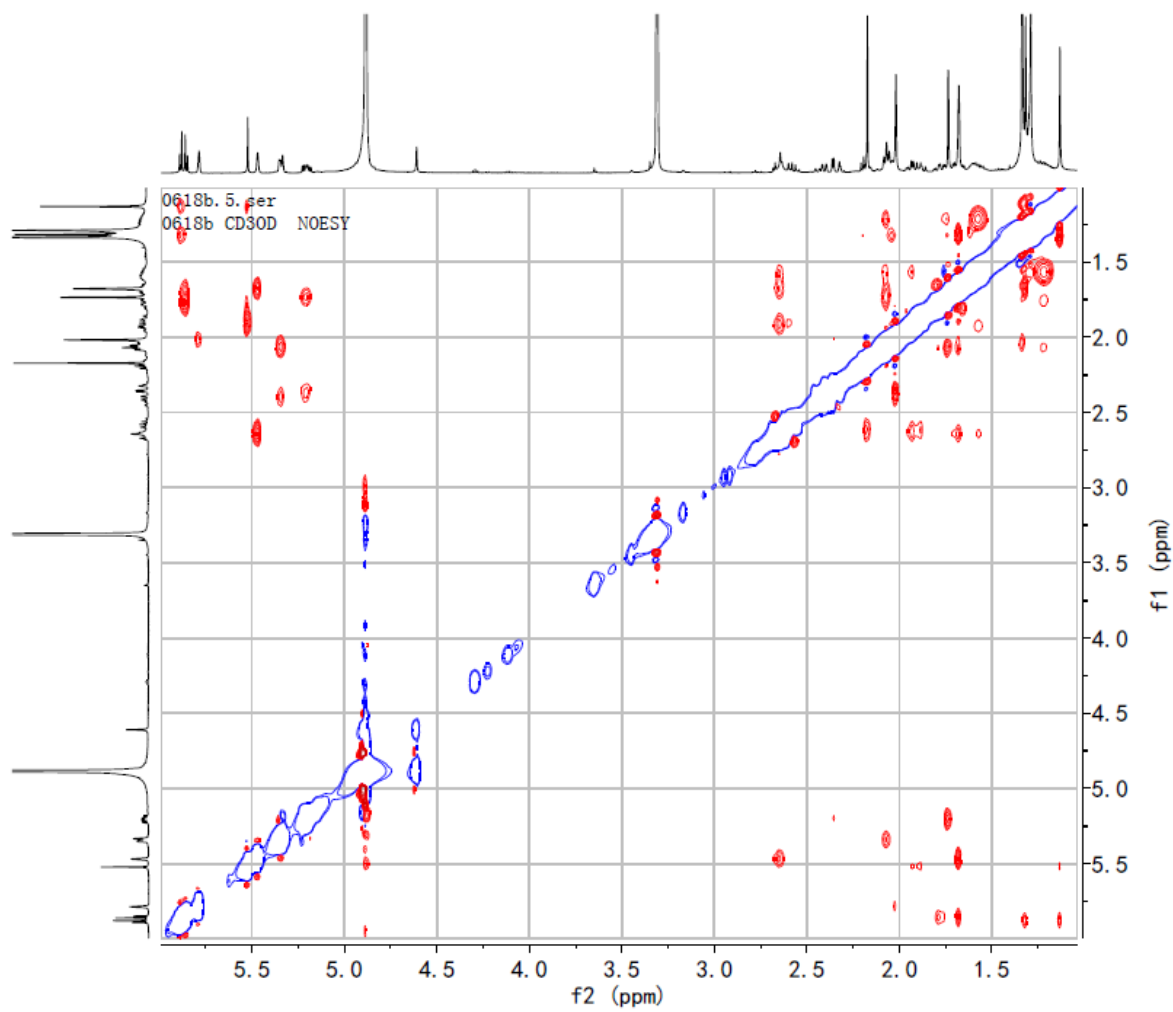


Figure S67. ESI(+)MS spectrum for 5b.

### Display Report

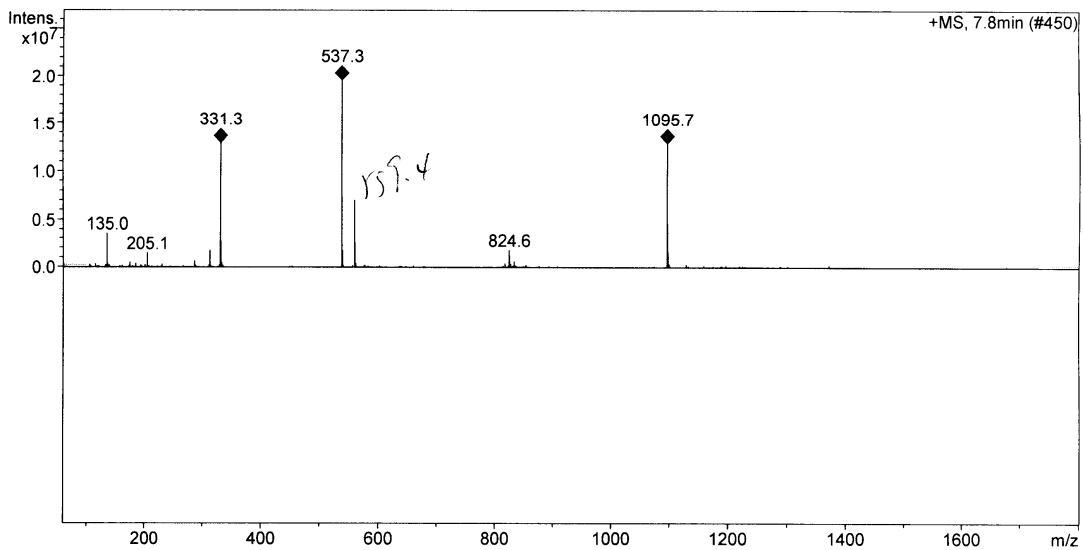
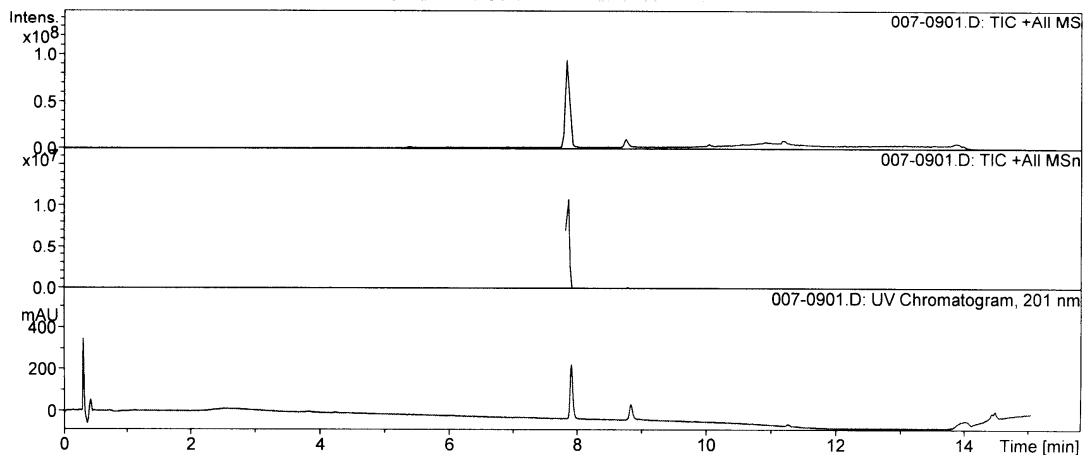
#### Analysis Info

Analysis Name 007-0901.D  
Method Copy of DSOPMS2P.M  
Sample Name yjm-0618b  
Comment 2\

Acquisition Date 09/12/12 12:11:21  
Operator Administrator  
Instrument esquire3000plus

#### Acquisition Parameter

Ion Source Type	ESI	Ion Polarity	Positive	Alternating Ion Polarity	off
Mass Range Mode	Std/Normal	Scan Begin	100 m/z	Scan End	1750 m/z
Capillary Exit	158.5 Volt	Skim 1	40.0 Volt	Trap Drive	85.4
Accumulation Time	15000 扫描	Averages	3 Spectra	Auto MS/MS	on



**Figure S68.** HRESI(+)**MS** spectrum for **5b**.

**Elemental Composition Report**

**Single Mass Analysis**

Tolerance = 2.0 PPM / DBE: min = -1.5, max = 50.0

Element prediction: Off

Number of isotope peaks used for i-FIT = 3

Monoisotopic Mass, Even Electron Ions

268 formula(e) evaluated with 1 results within limits (up to 50 closest results for each mass)

Elements Used:

C: 6-60 H: 2-110 O: 0-30 Na: 0-1

0618b

LCT PXE KE324

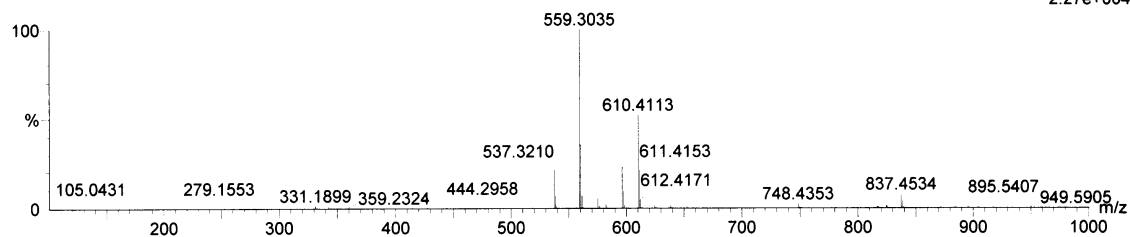
14-Sep-2012

10:56:07

1: TOF MS ES+

2.27e+004

0618b\_20120914 28 (0.583) AM2 (Ar,10000.0,0.00,1.00); ABS; Cm (6:29)



Minimum:

Maximum: 3.0 2.0 -1.5

Maximum: 50.0

Mass	Calc. Mass	mDa	PPM	DBE	i-FIT	i-FIT (Norm)	Formula
559.3035	559.3036	-0.1	-0.2	11.5	75.6	0.0	C33 H44 O6 Na