

Concise asymmetric syntheses of novel phenanthroquinolizidines

Cintia Anton-Torrecillas,^a María Isabel Loza,^b José Brea^b and Jose C. Gonzalez-Gomez,^{*a}

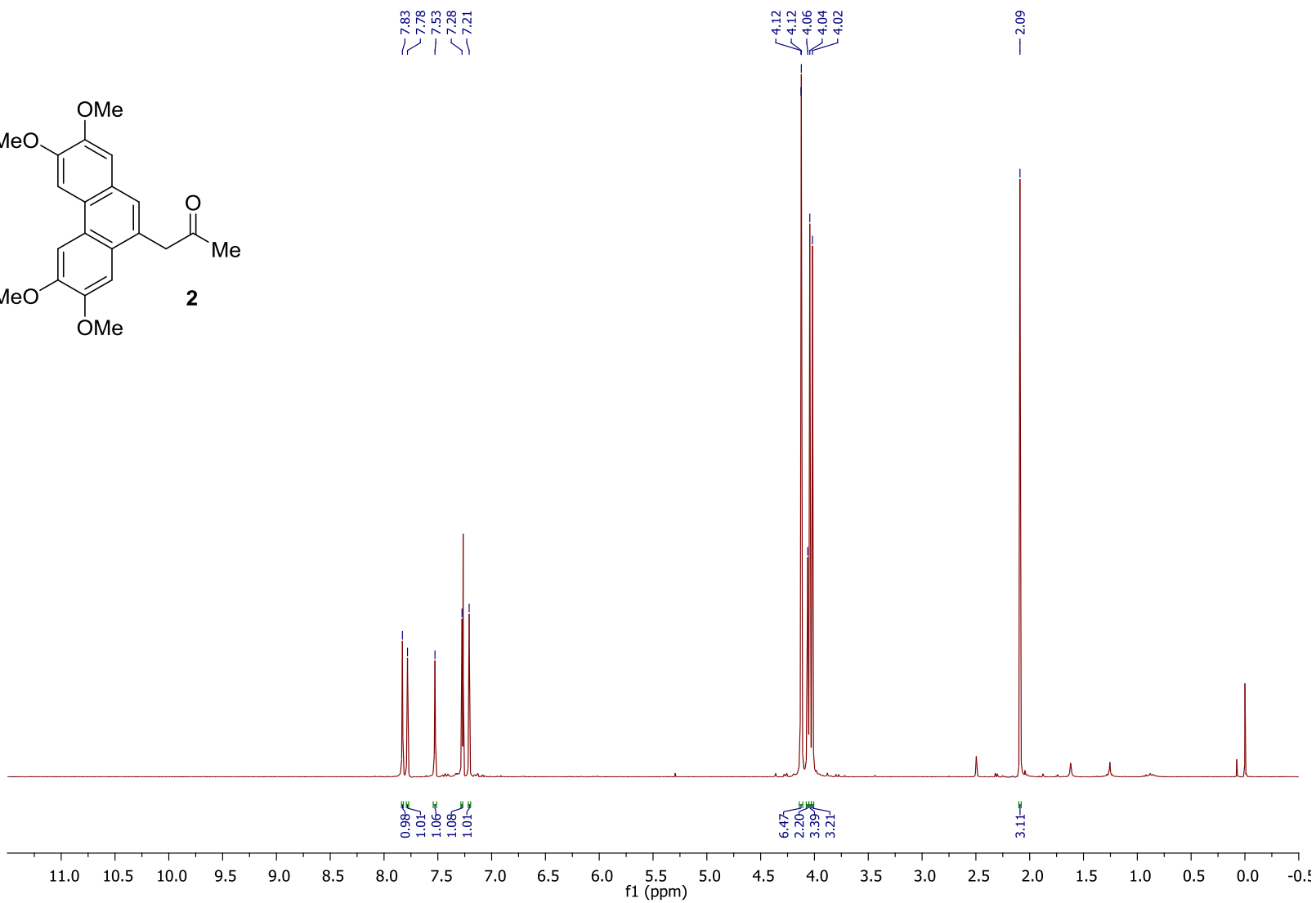
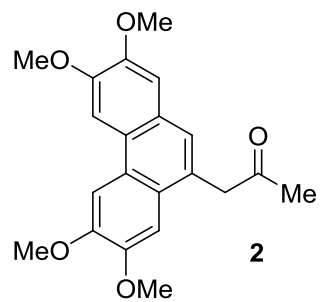
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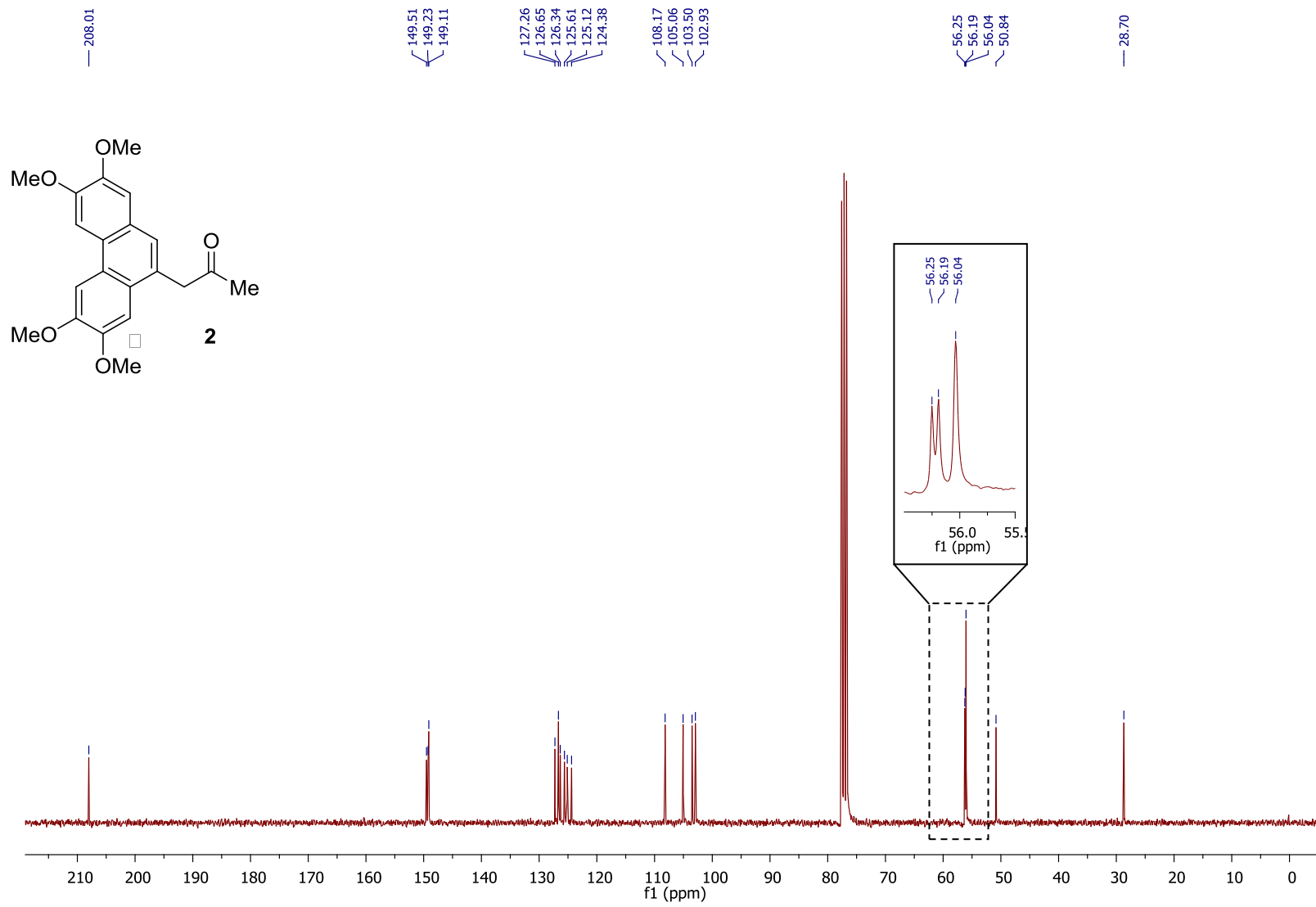
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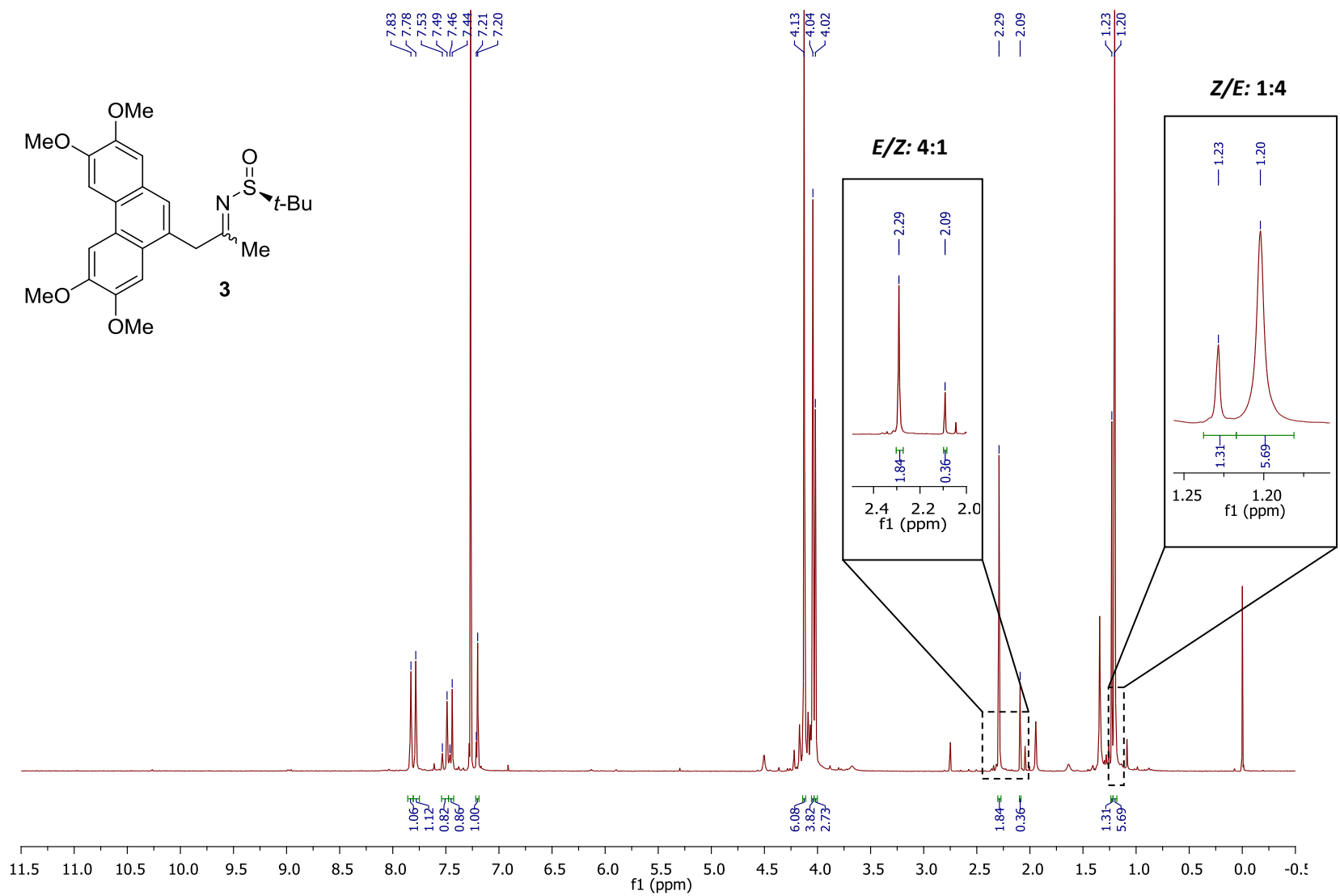
¹H NMR (300 MHz, CDCl₃)



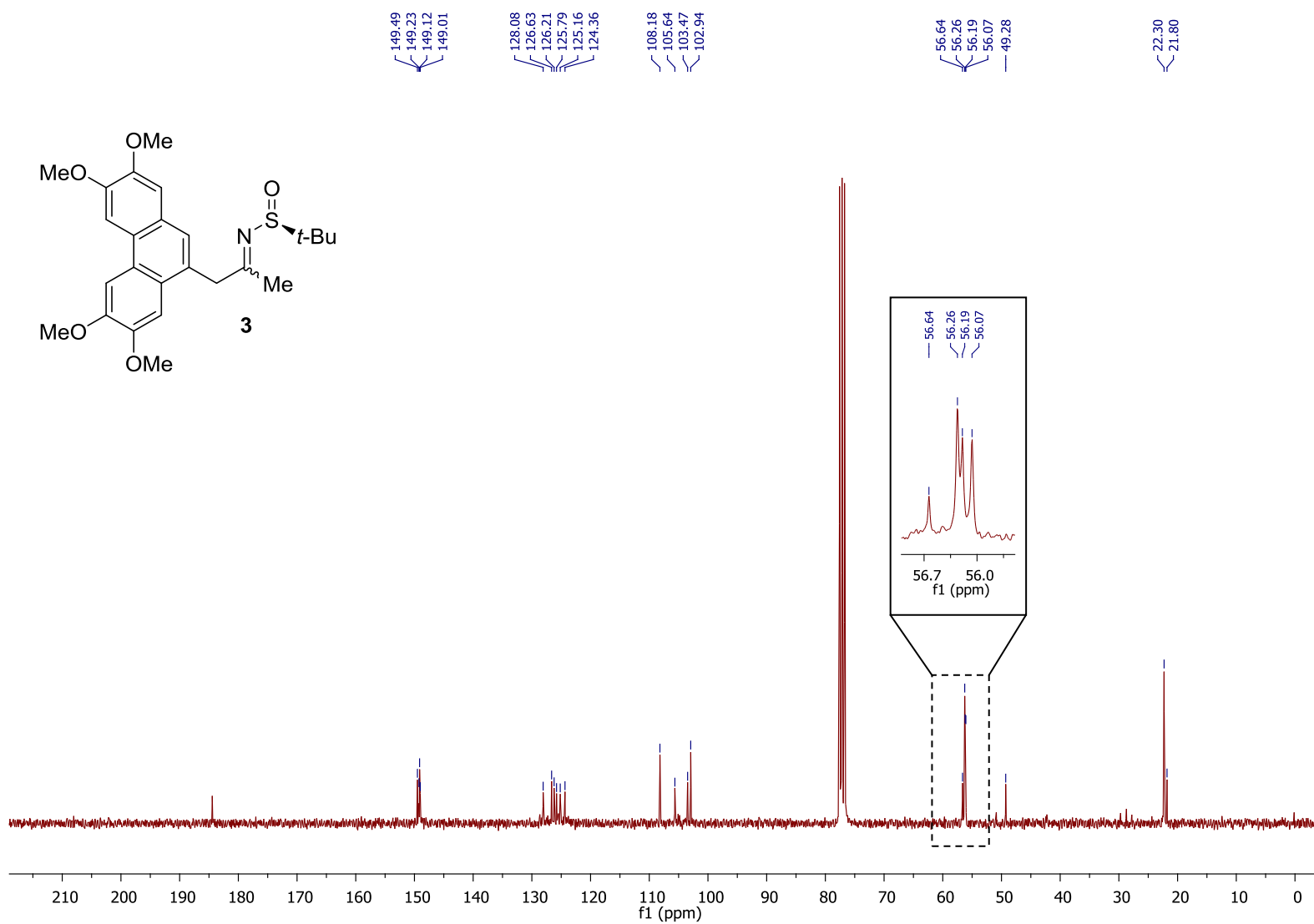
^{13}C NMR (75 MHz, CDCl_3)



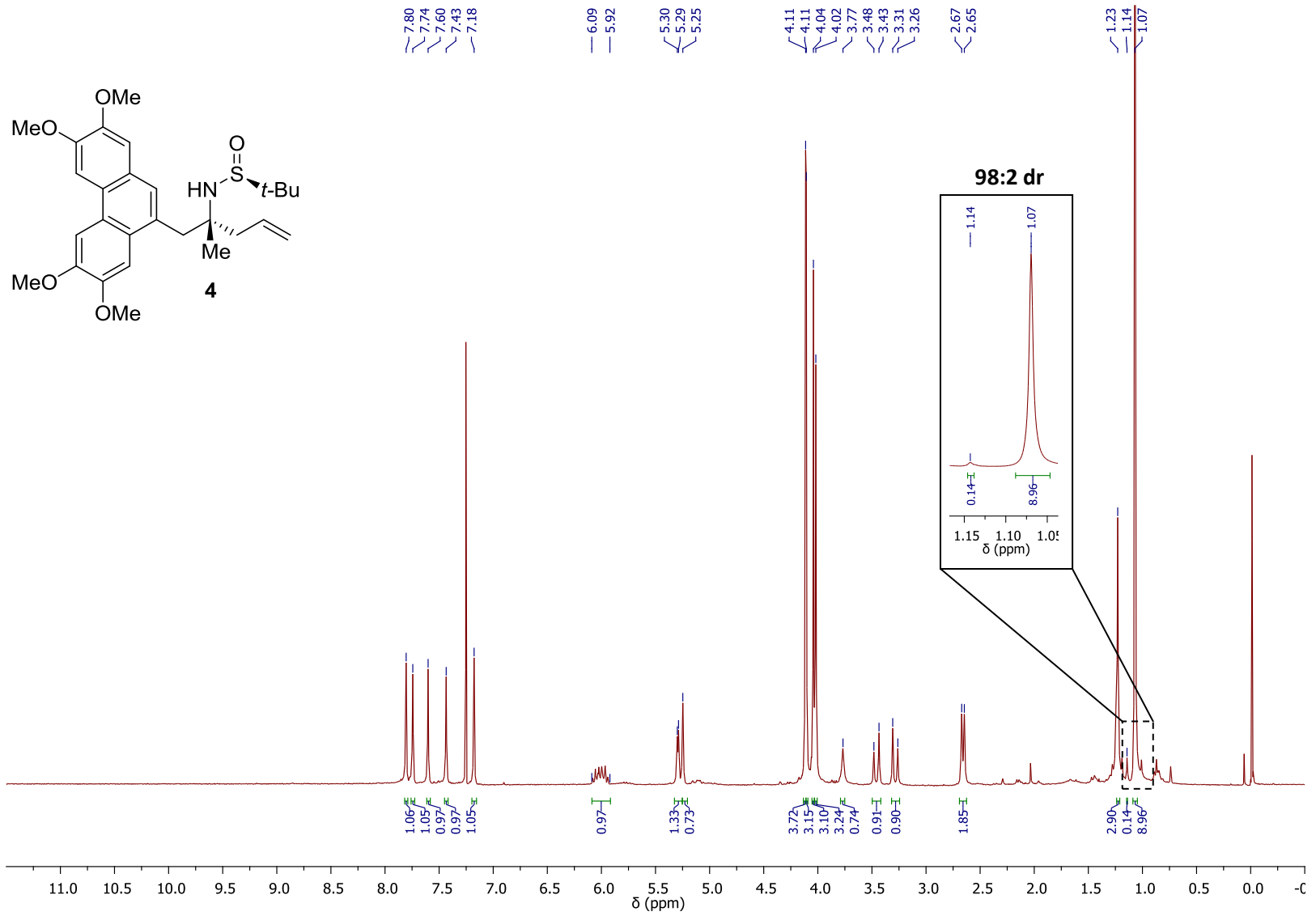
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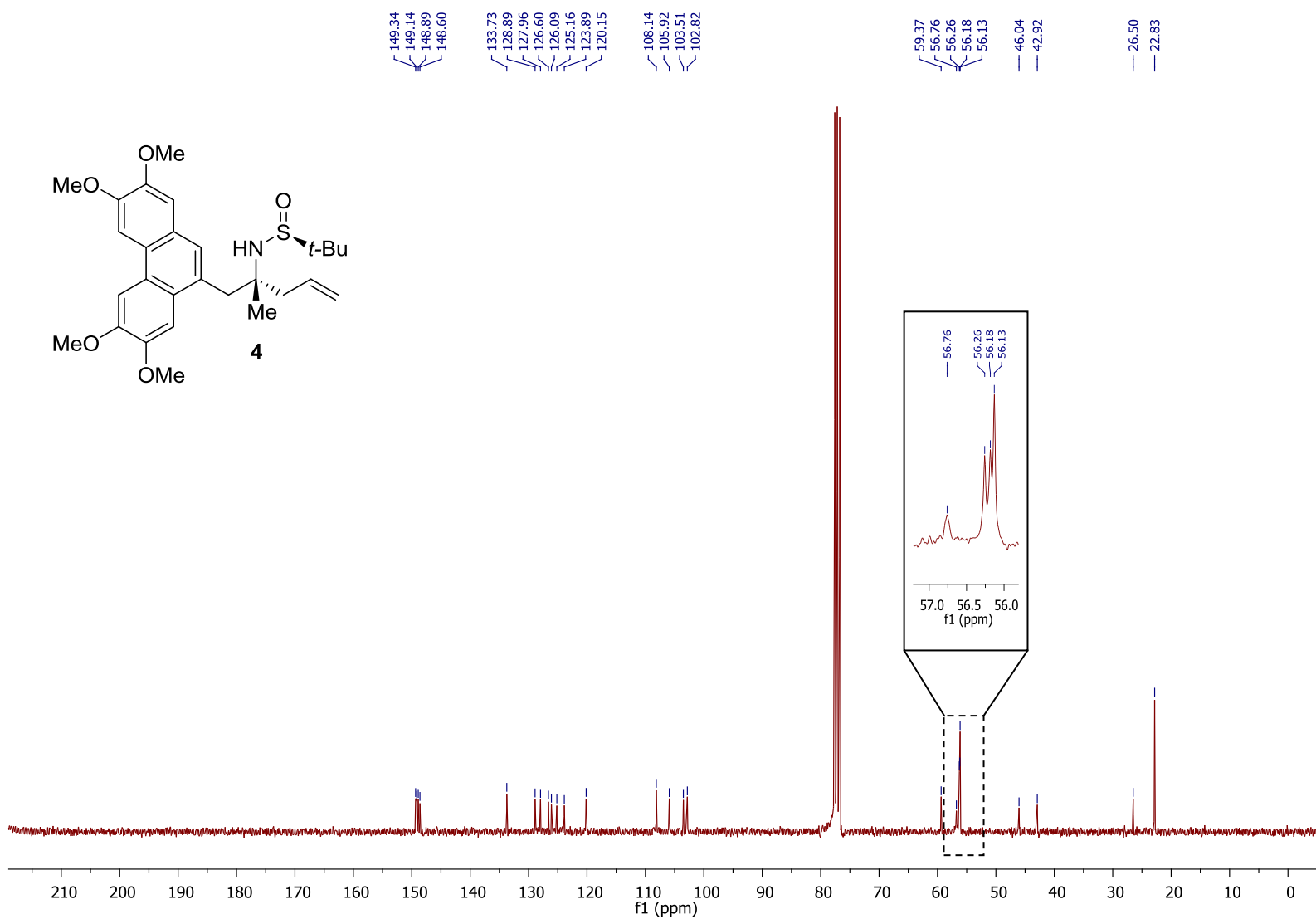
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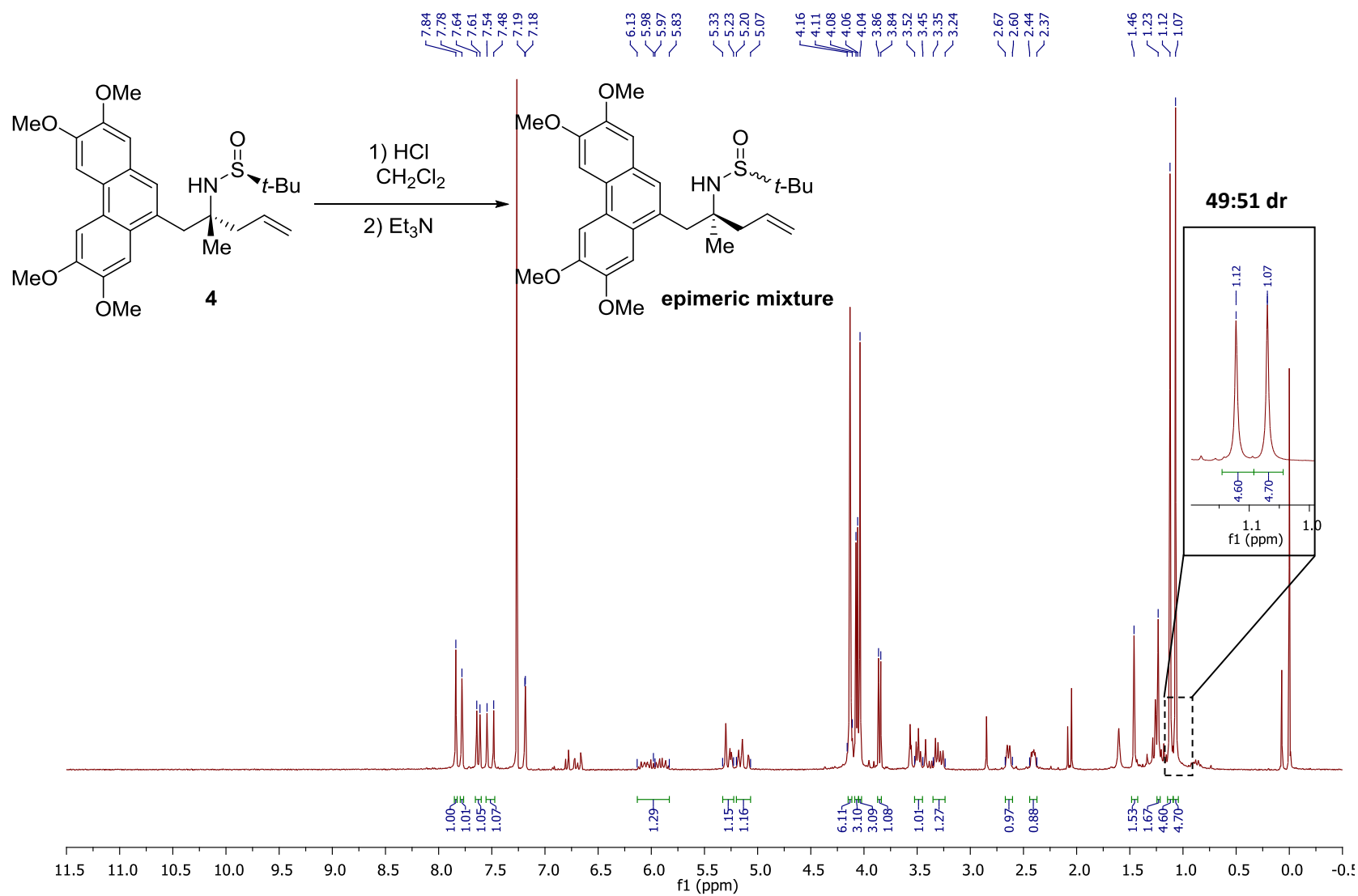
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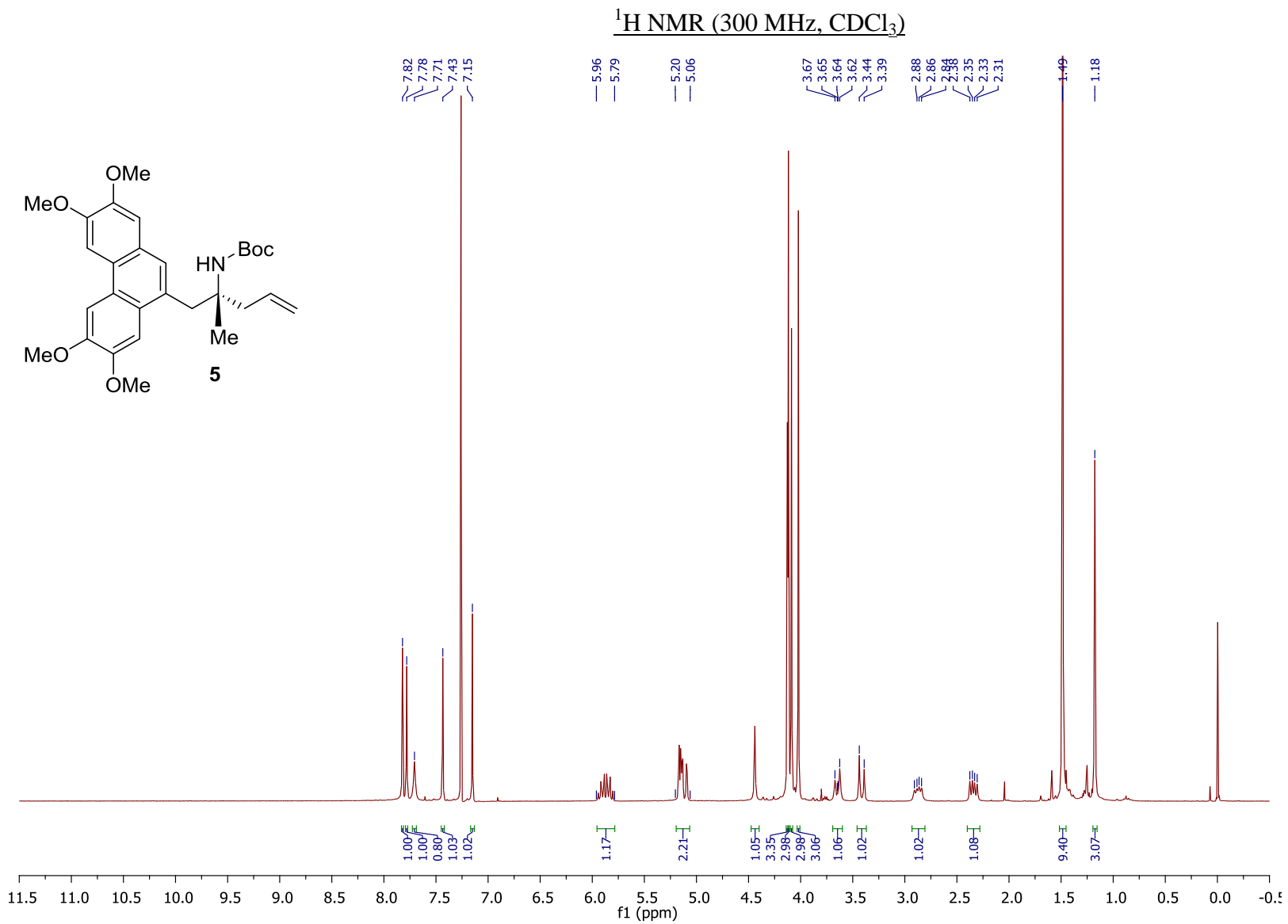


¹³C NMR (75 MHz, CDCl₃)

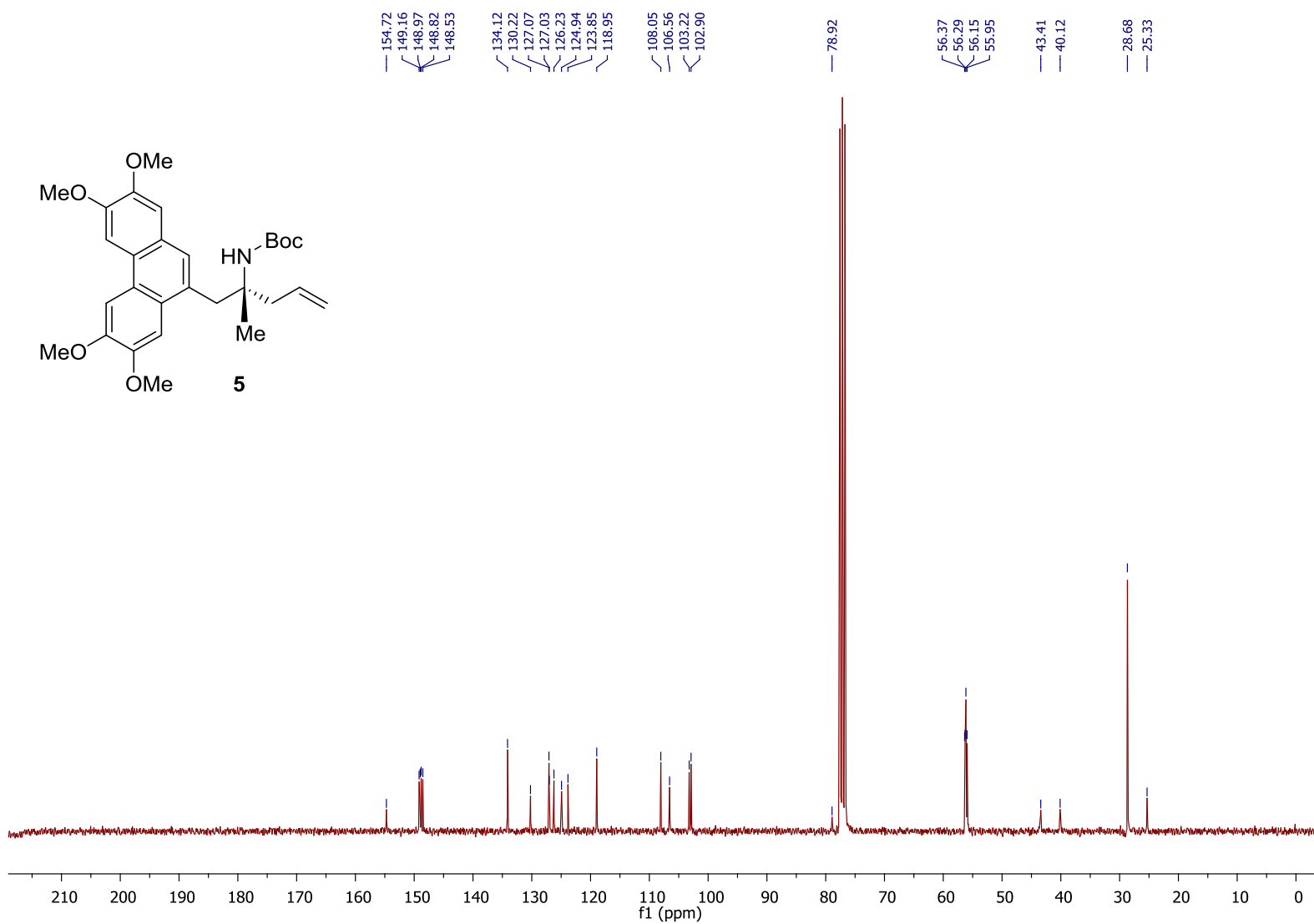


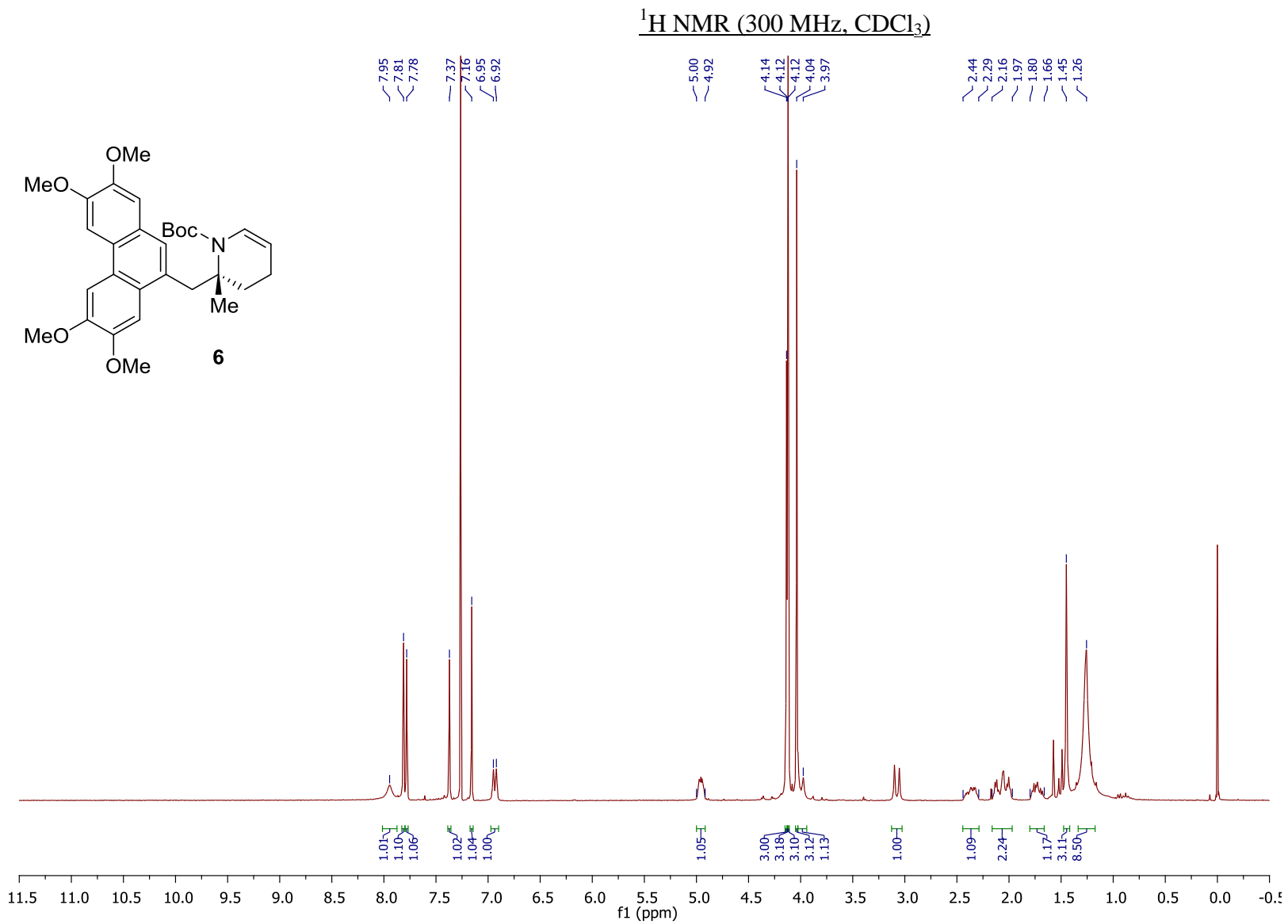
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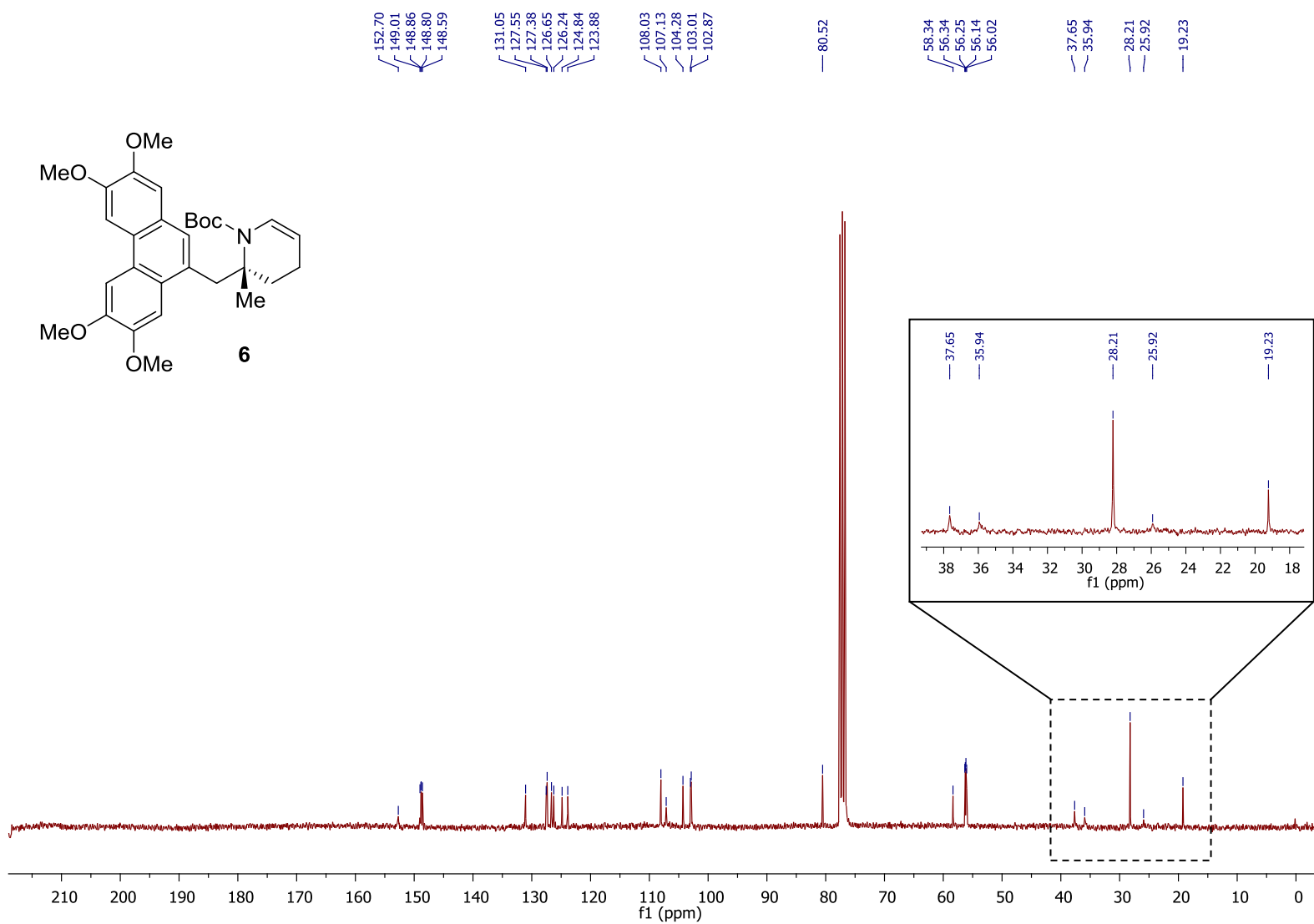


¹³C NMR (75 MHz, CDCl₃)

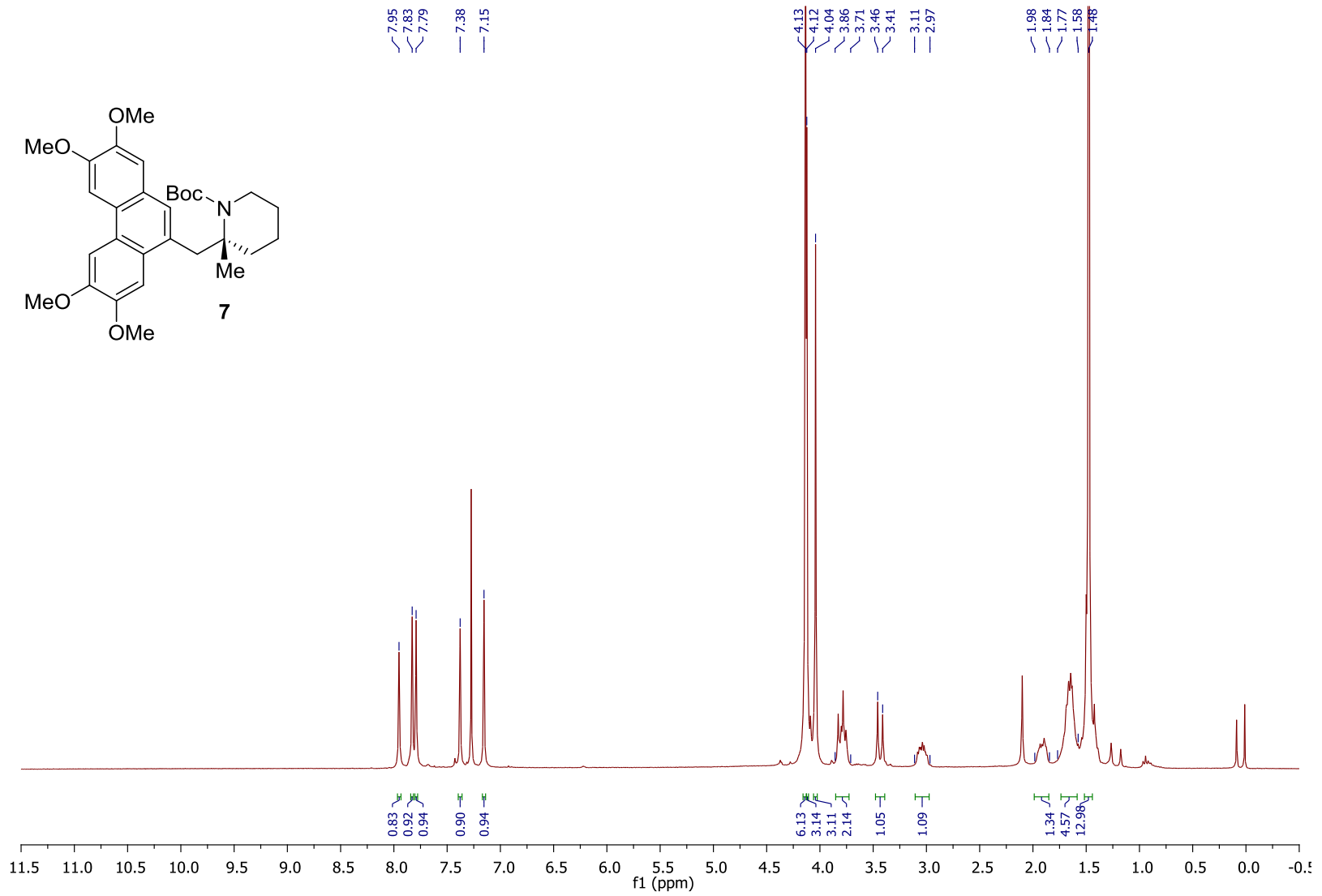
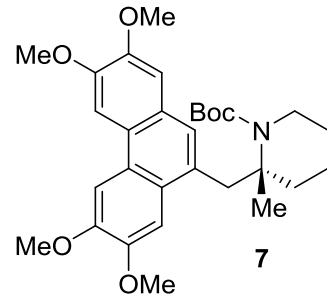




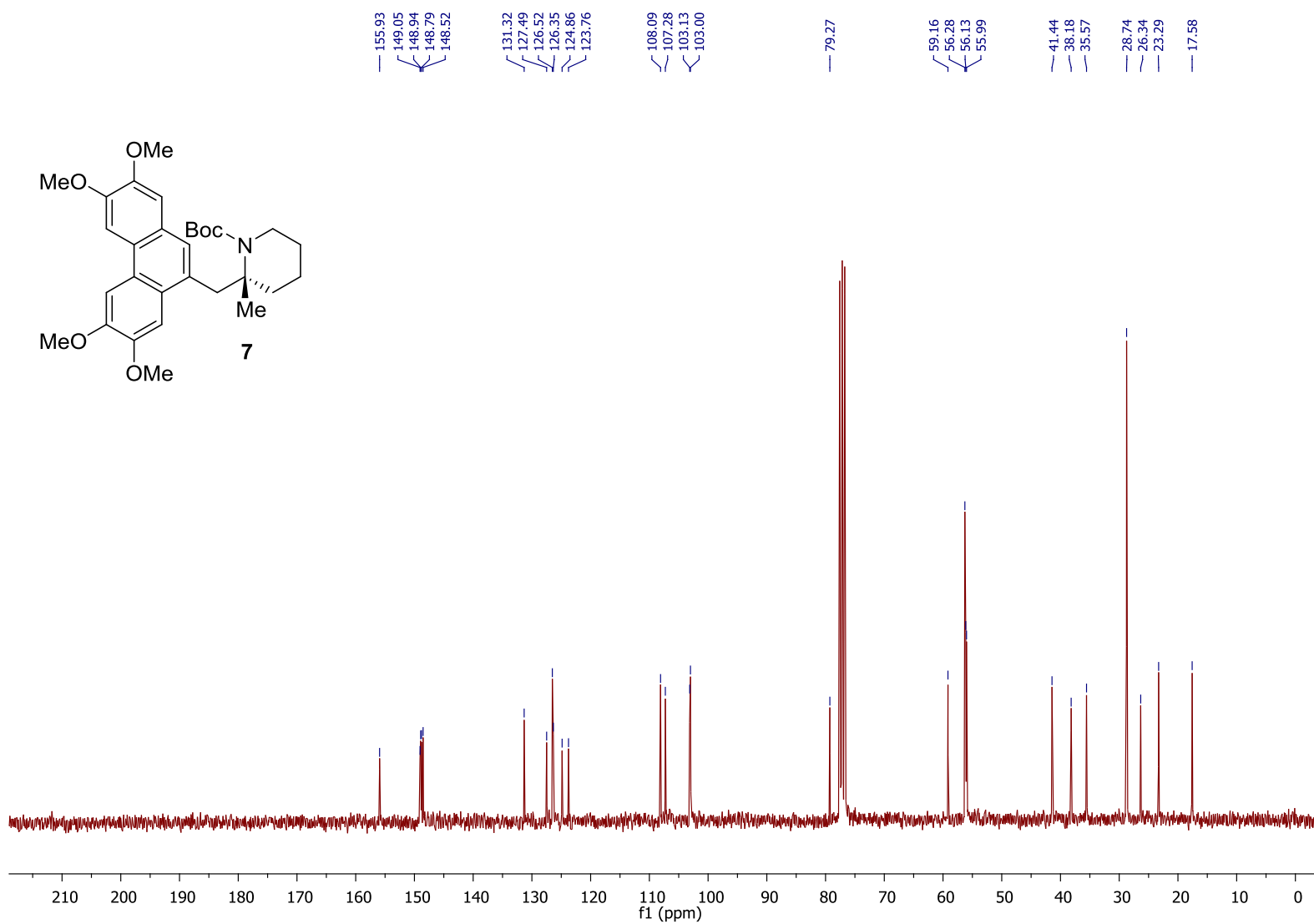
¹³C NMR (75 MHz, CDCl₃)



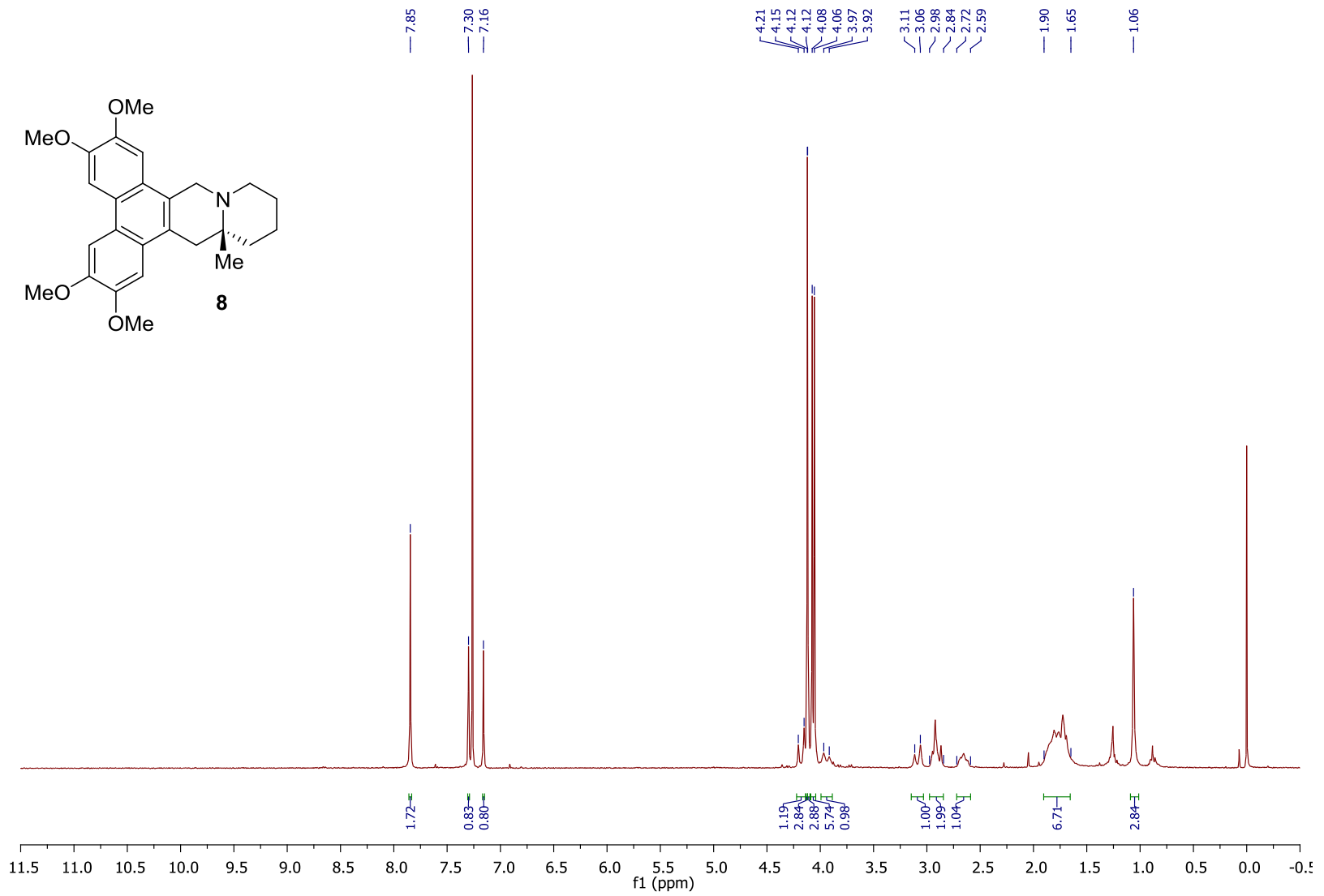
¹H NMR (300 MHz, CDCl₃)



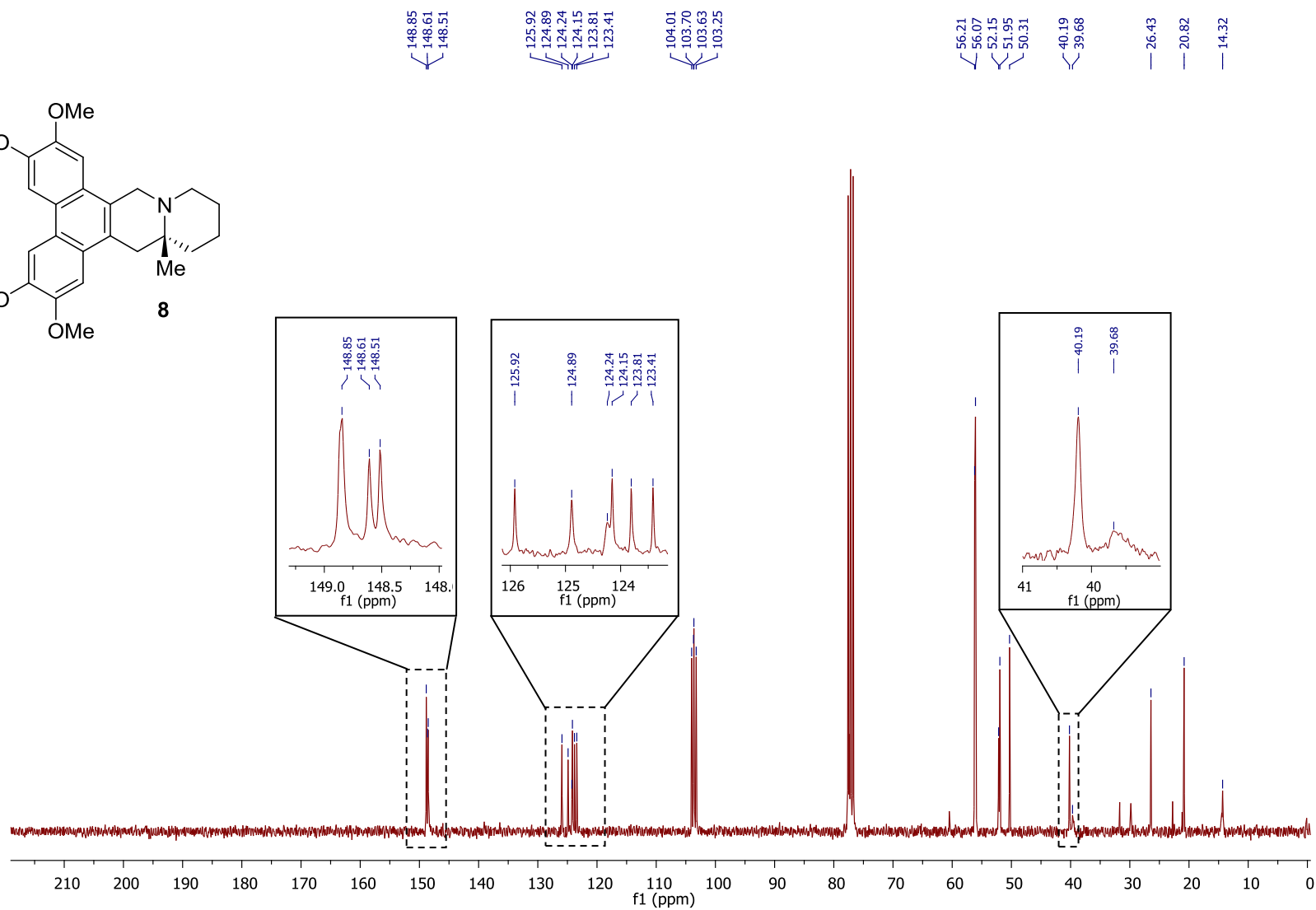
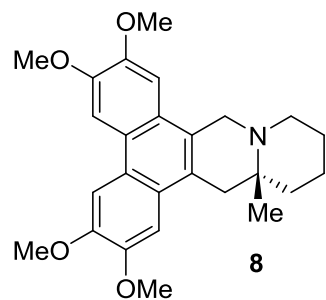
^{13}C NMR (75 MHz, CDCl_3)



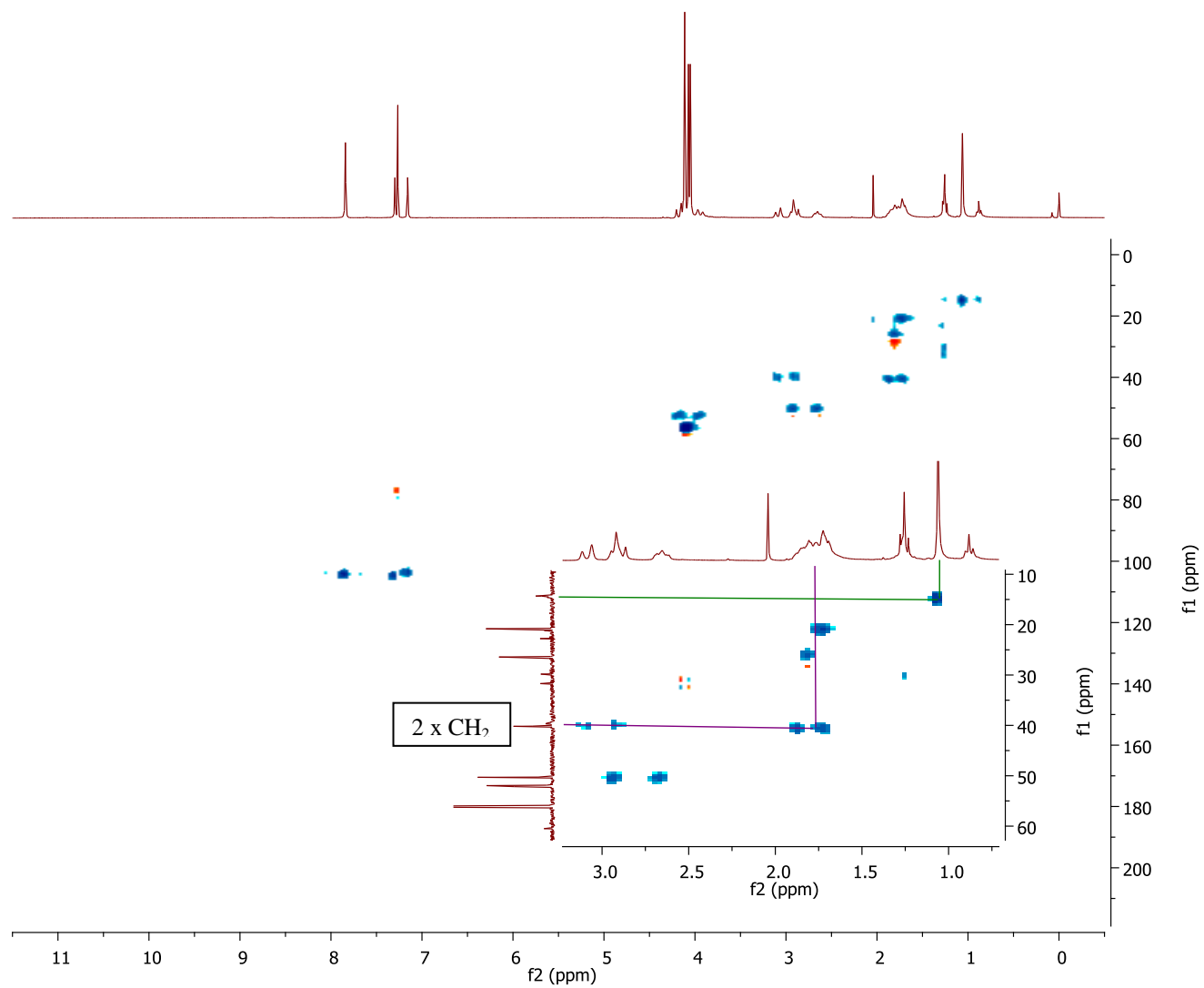
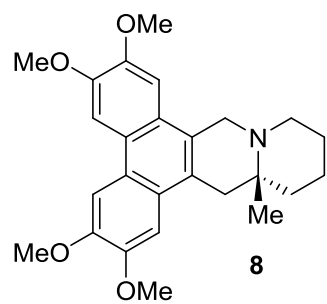
^1H NMR (300 MHz, CDCl_3)



^{13}C NMR (75 MHz, CDCl_3)

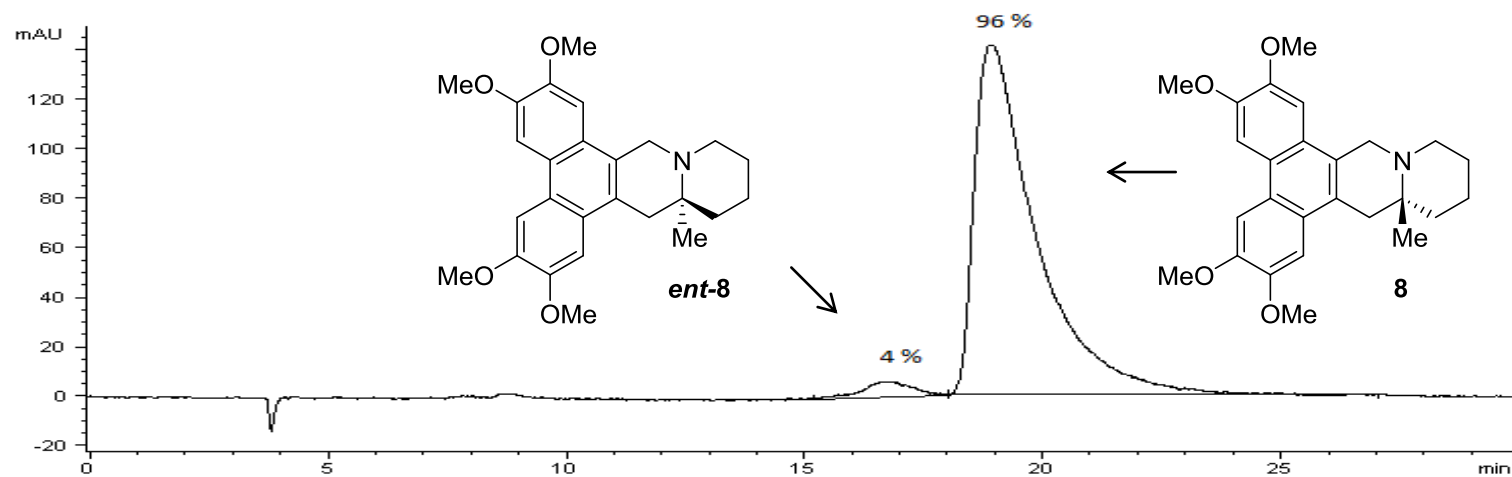
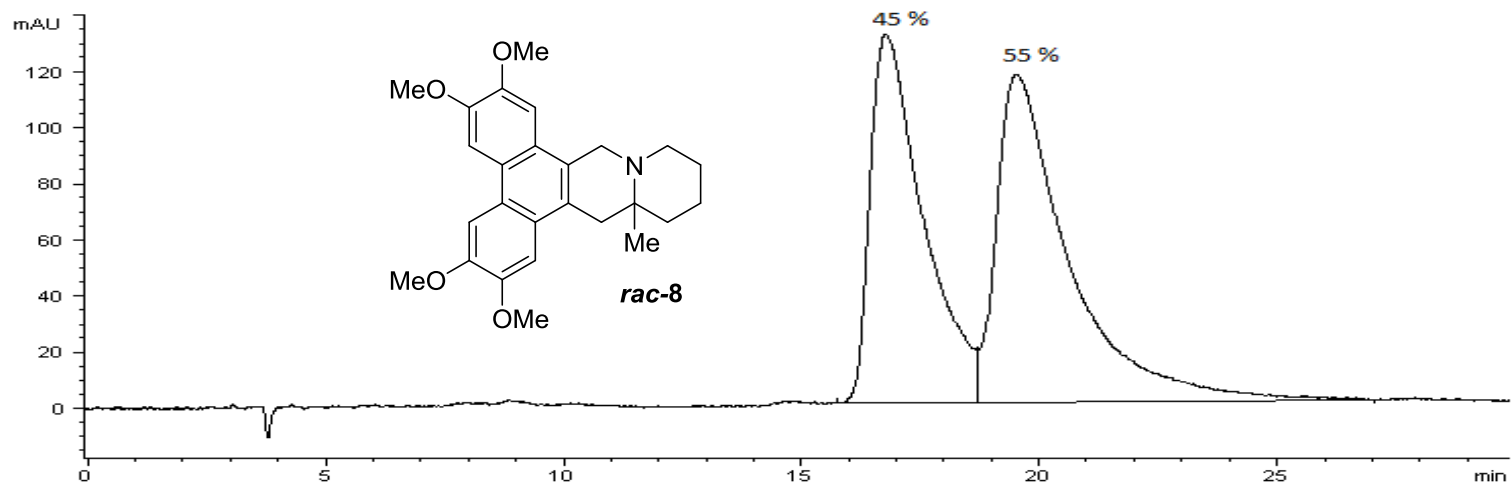


COSY (300 MHz, CDCl₃)

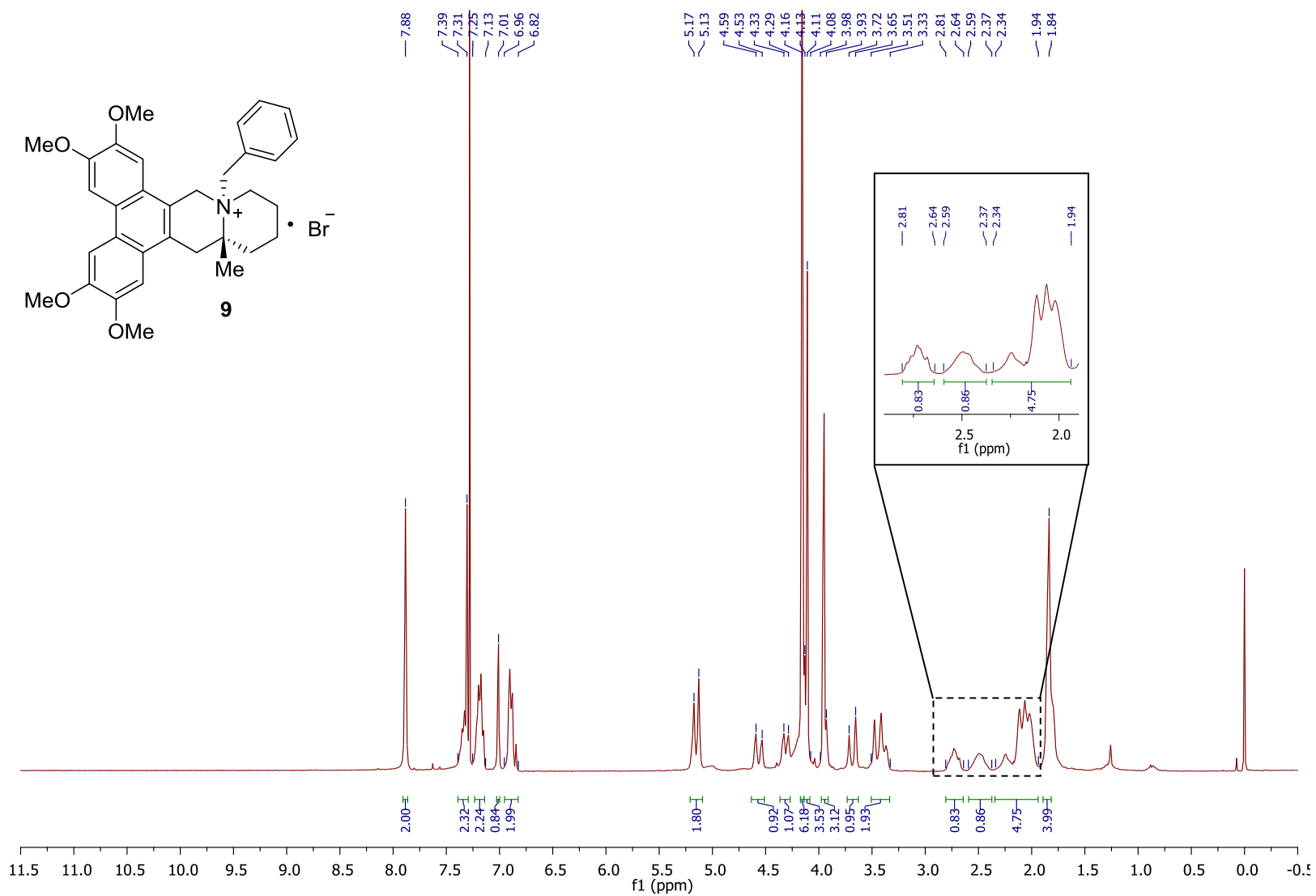


HPLC traces of compounds *rac-8* and *ent-8* / **8**

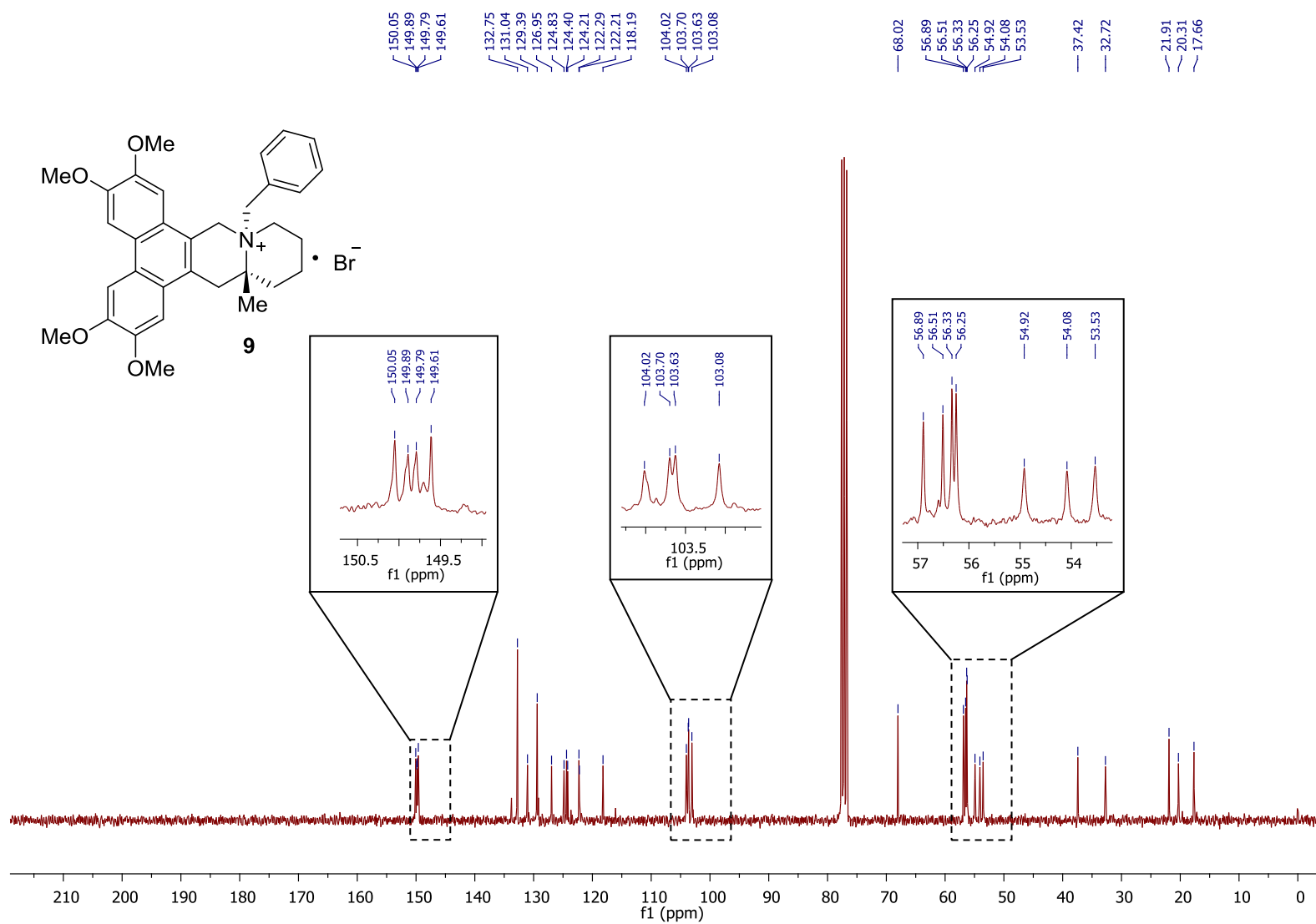
CHIRALPAK IB column 4.6 mm x 250 mL; isocratic elution with 75:25:0.3, *n*-hexane/*i*-PrOH/TEA, 1.0 mL·min⁻¹; UV detection at 254 nm.

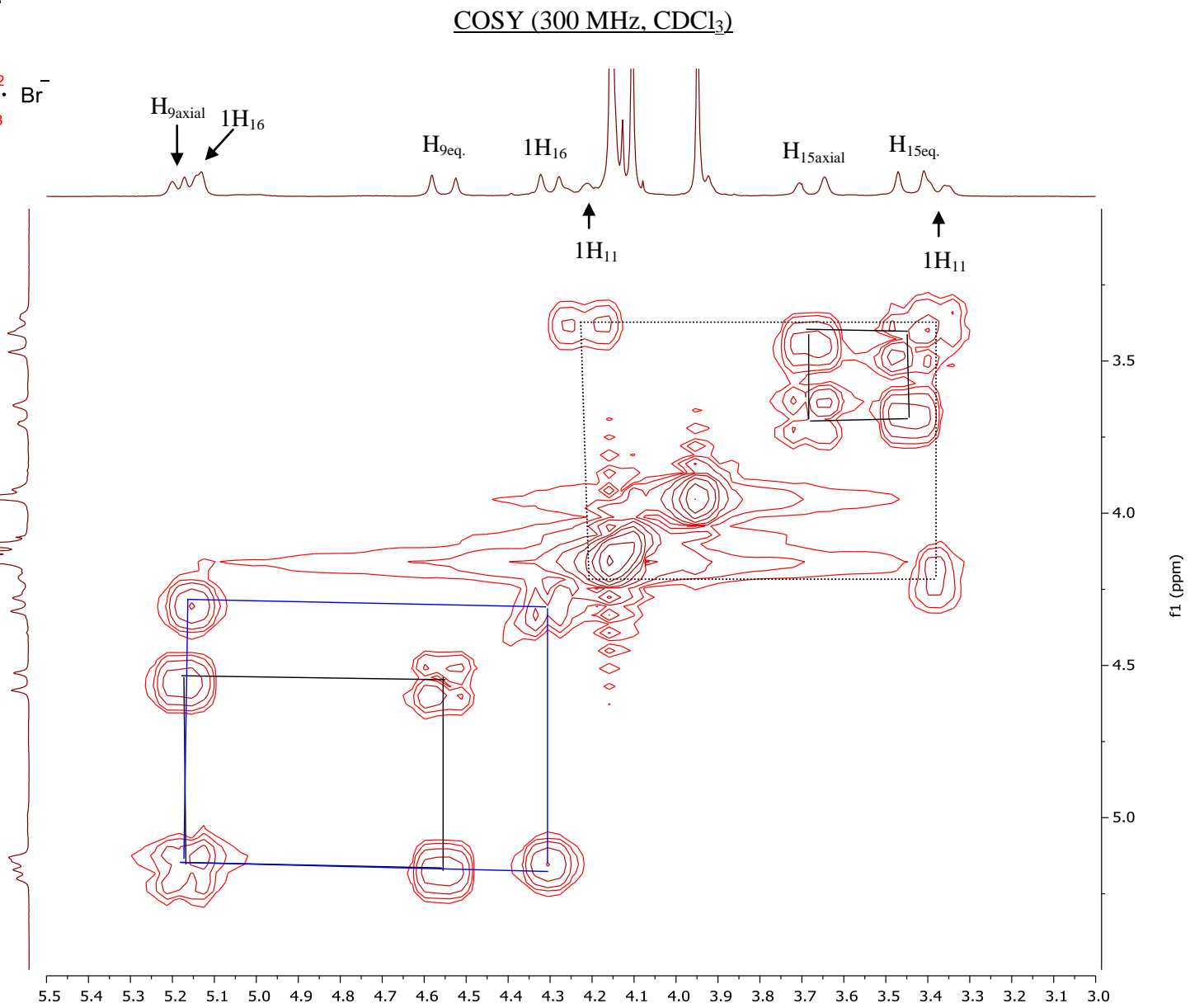
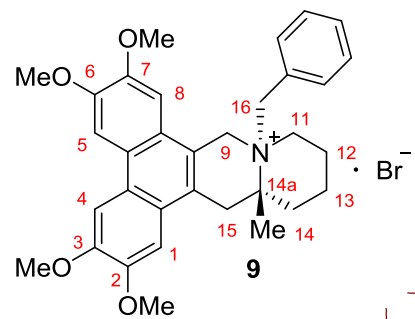


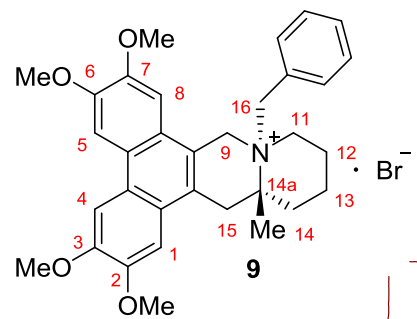
$^1\text{H NMR}$ (300 MHz, CDCl_3)



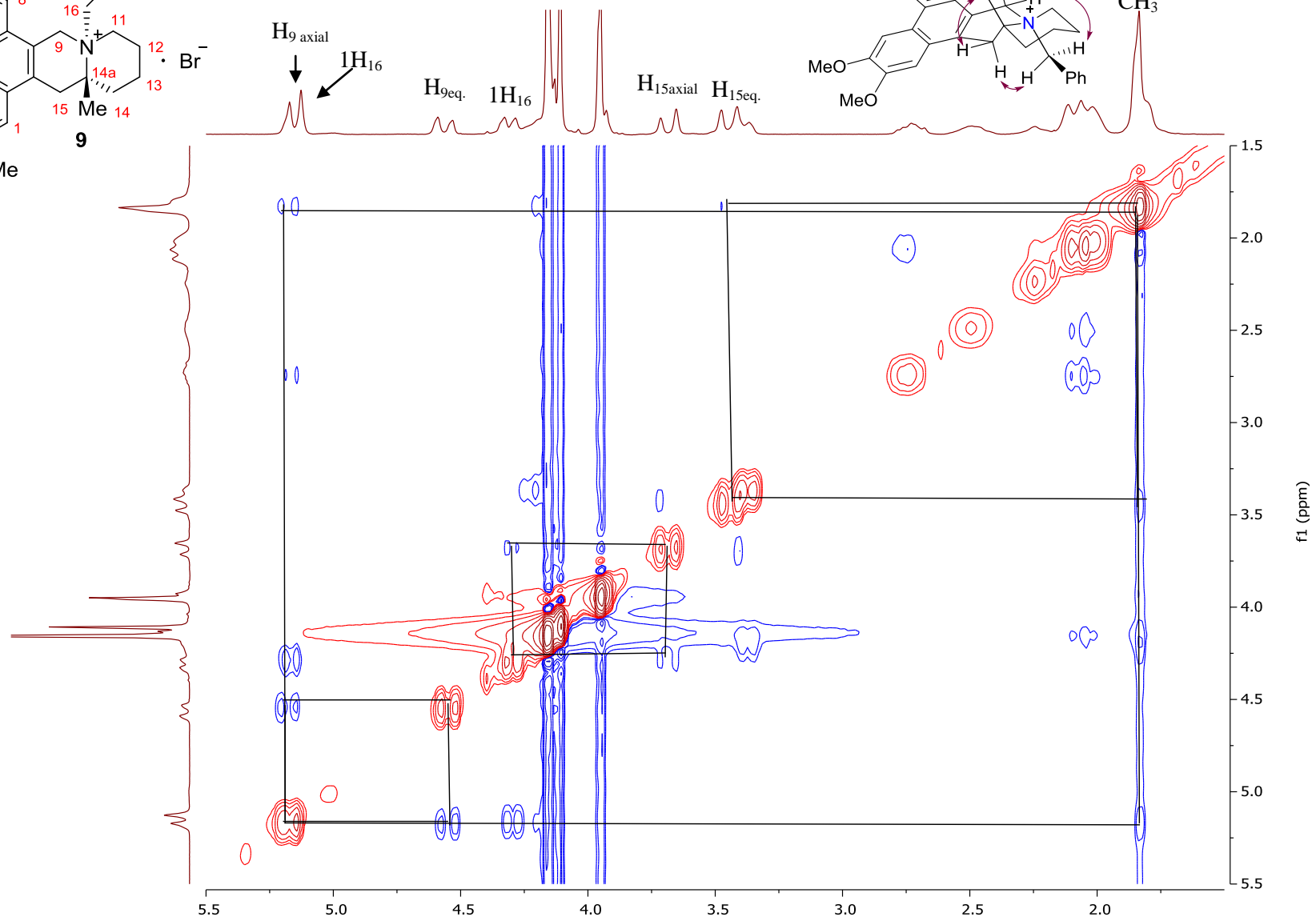
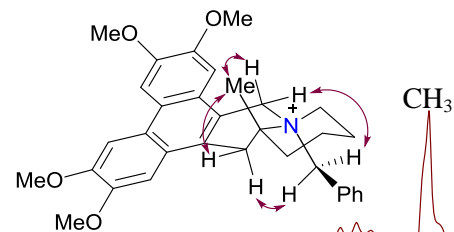
^{13}C NMR (75 MHz, CDCl_3)



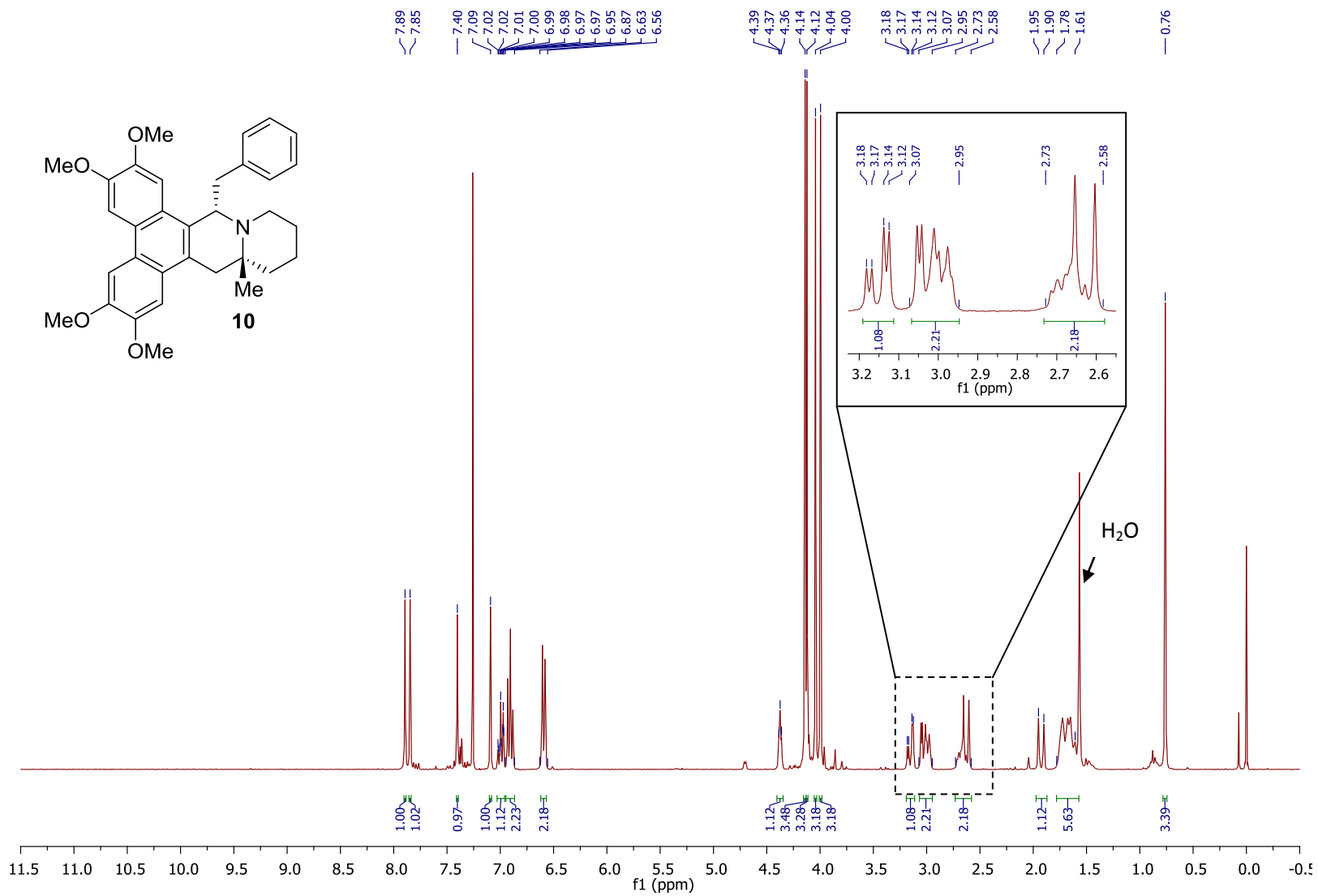




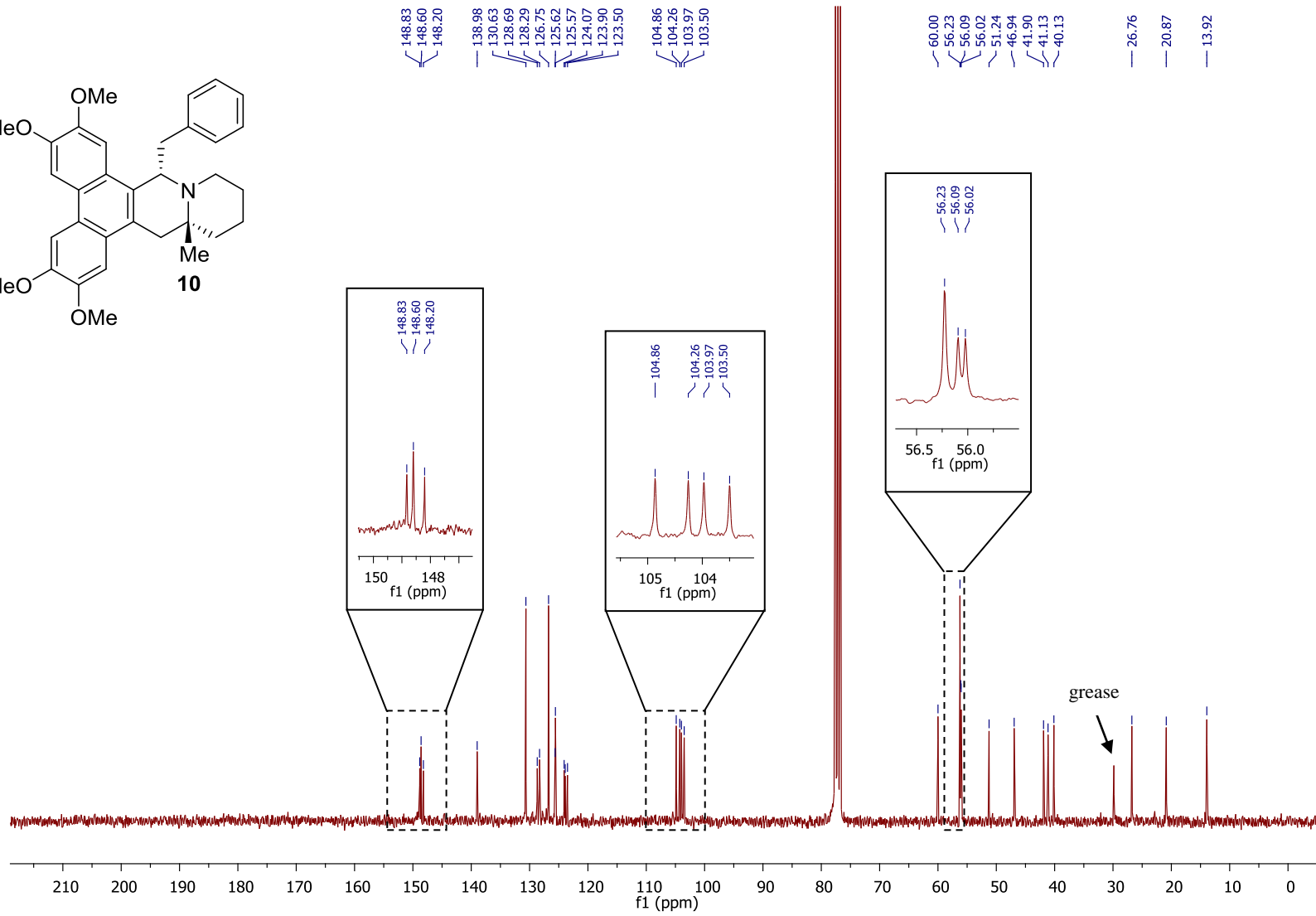
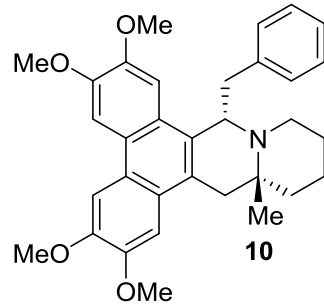
NOESY (300 MHz, CDCl₃)



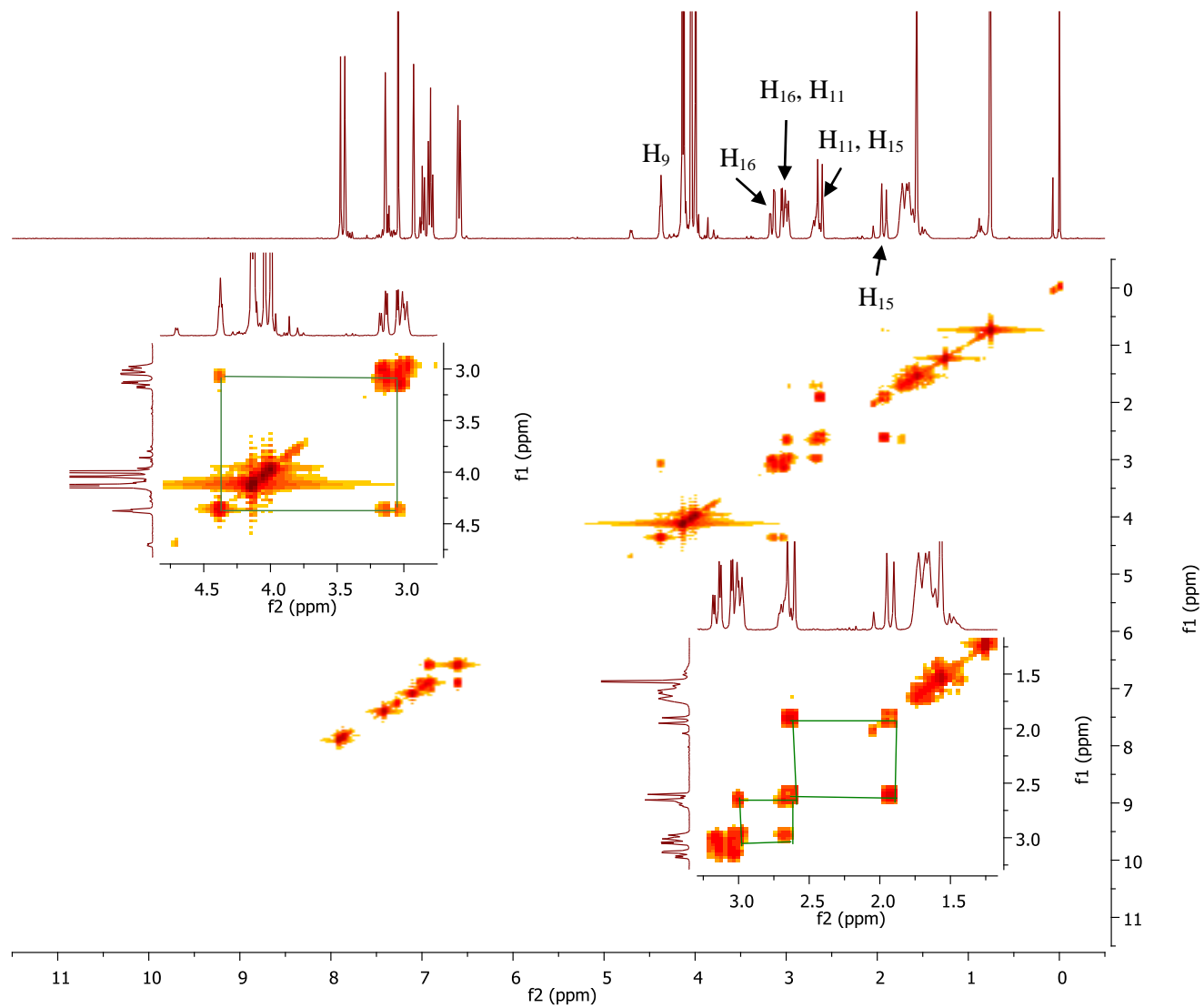
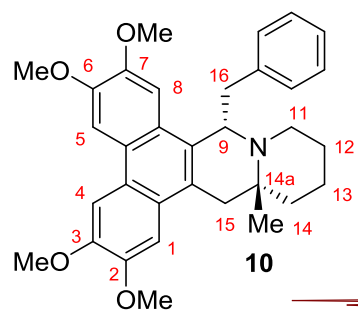
$^1\text{H NMR}$ (300 MHz, CDCl_3)



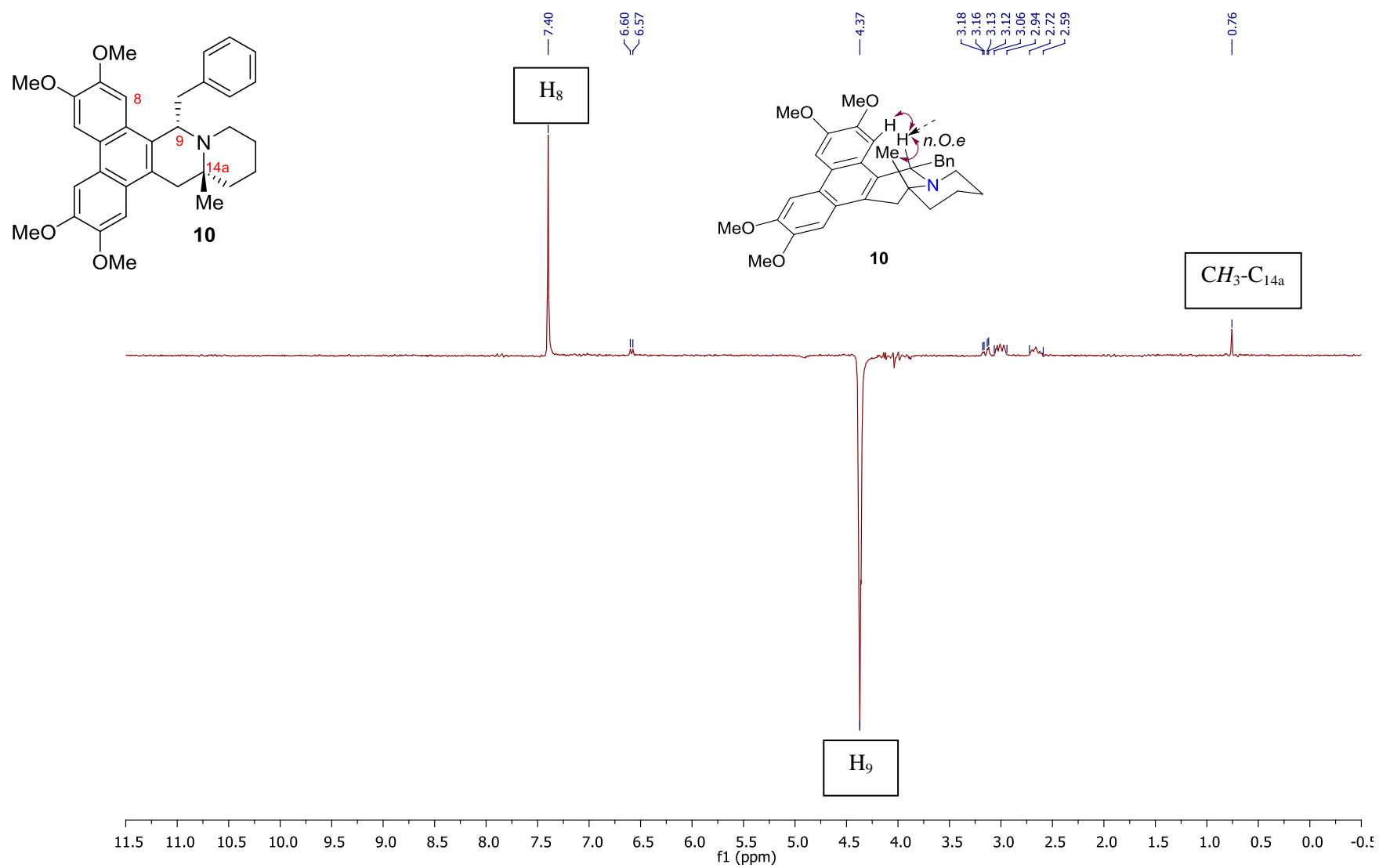
^{13}C NMR (75 MHz, CDCl_3)



COSY (300 MHz, CDCl₃)

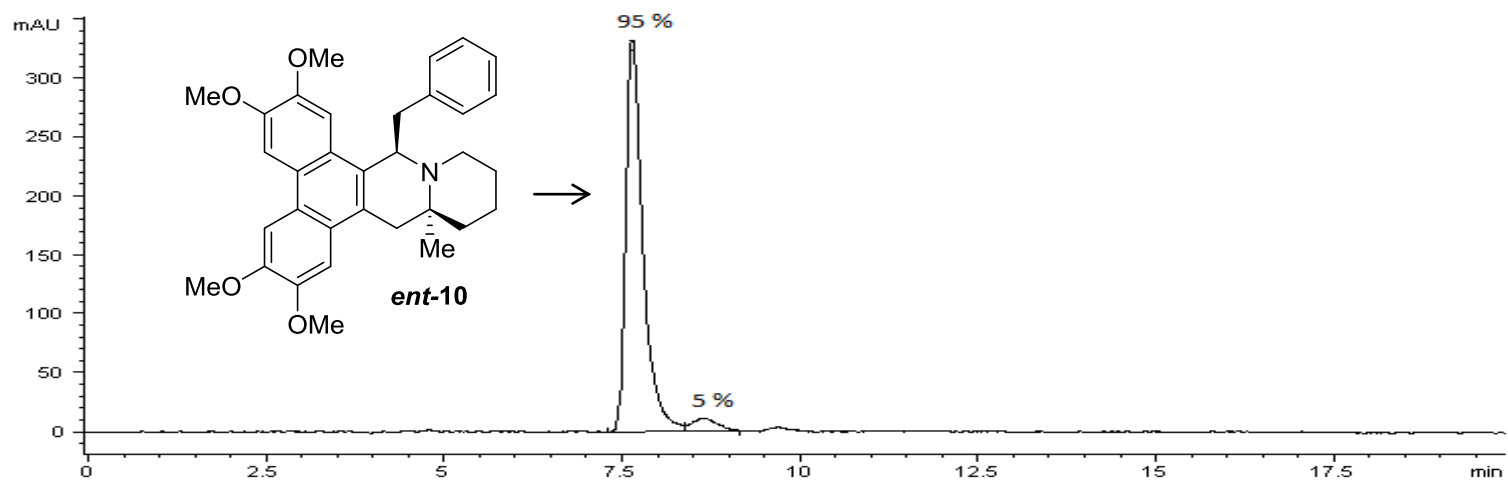
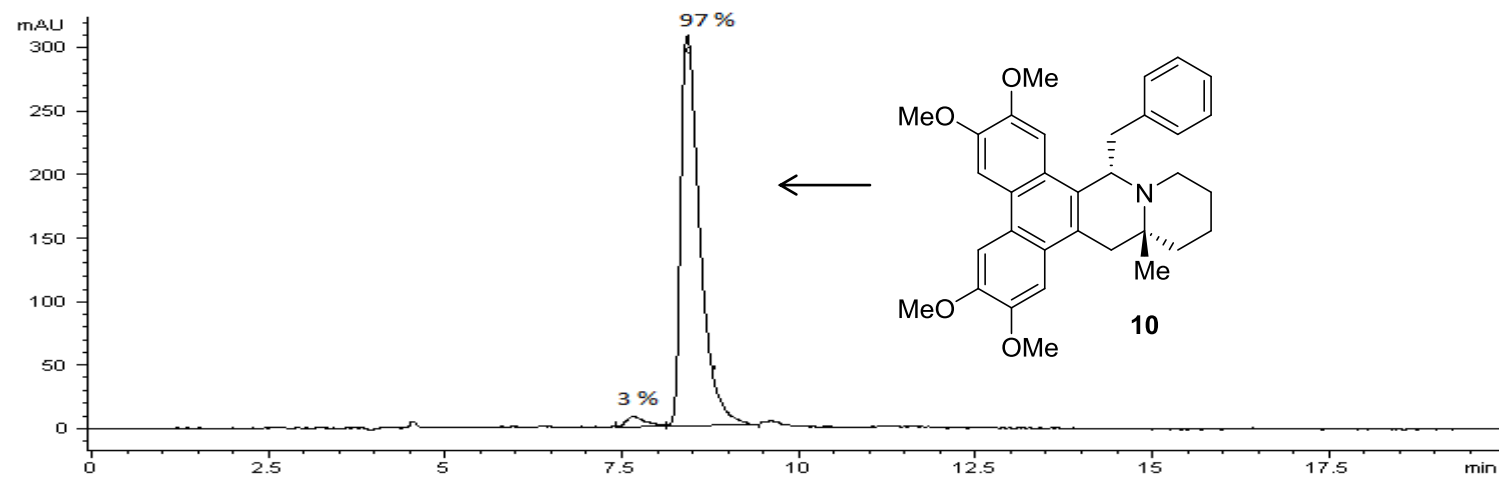


DPFGSE-NOE when H₉ is irradiated (300 MHz, CDCl₃)



HPLC traces of compounds **10**/ *ent-10*

CHIRALPAK IB column 4.6 mm x 250 mL; isocratic elution with 75:25:0.3, *n*-hexane/*i*-PrOH/TEA 1.0 mL·min⁻¹; UV detection at 254 nm.



Cytotoxic Studies

All the cell lines employed were purchased to the American Tissue Culture Collection (ATCC) and grown following the instructions and media from the provider.

Cells were seeded in 96-well culture microplates and kept at 37°C in a 5% CO₂ atmosphere for 24 h. Solutions of test compounds in MeOH were then added to each microplate, keeping 1% of MeOH, and incubated for different times depending on the cell line studied. After the incubation time cell viability was measured by using the tetrazolium dye assay (MTT method). Experiments were repeated three times.

Data was expressed as a percentage of inhibition in basis of the cell viability observed in vehicle-treated well following the formula:

%inhibition=100-((AO*100)/AT); where AO is the absorbance observed in the test well and AT is the absorbance observed in the vehicle-treated wells (with 1% of MeOH to consider the effect of the solvent). Concentration-% inhibition were fitted by using GraphPad Prism (v2.1) following the equation:

$$y = \frac{E_{\max}}{1 + \frac{IC_{50}^n}{x^n}}$$

where E_{max} is the inhibition observed at the maximum concentration examined for each compound (100 μM), IC₅₀ is the concentration that inhibits cell viability in a 50% and n is the Hill slope.

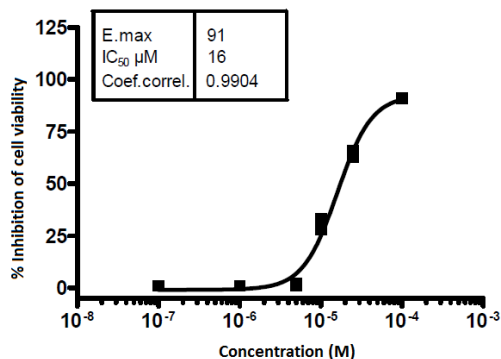
The seeding numbers and incubation times for each cell lines, as well as the concentration-response curves obtained for the compounds examined (including CDDP as a control experiment) were the following:

MCF-7

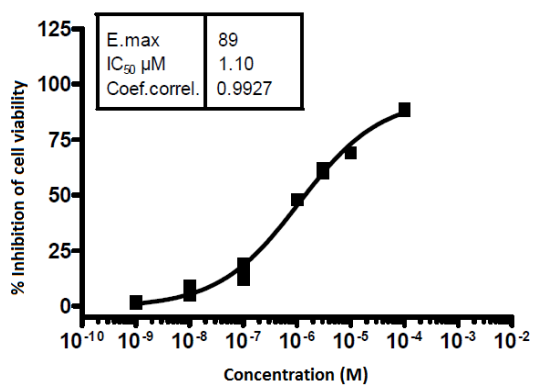
Seeding number: 10000

Incubation time with test compound: 96 h

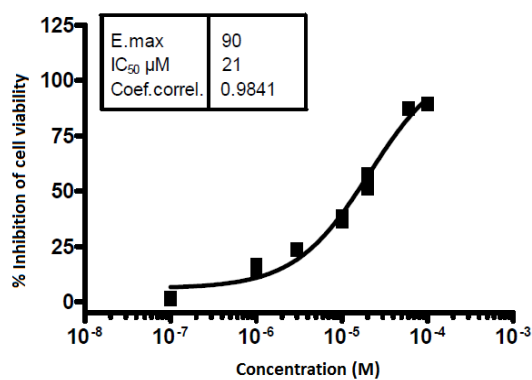
CDDP



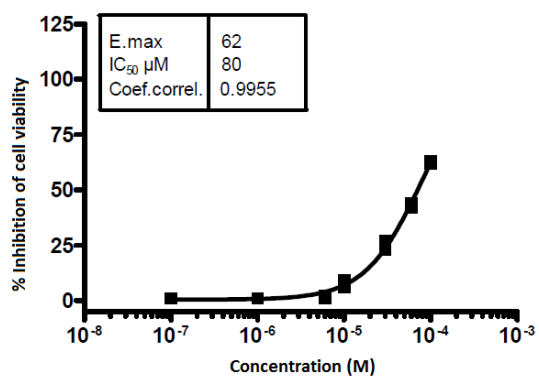
8·HCl



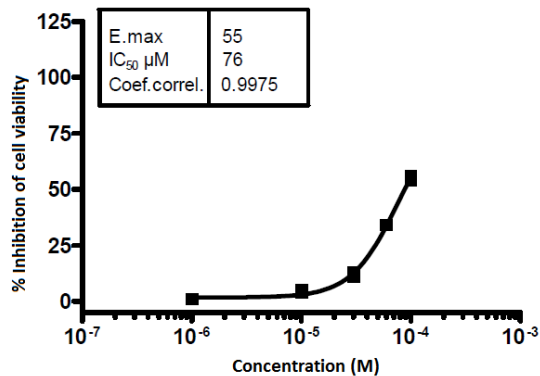
ent-8·HCl



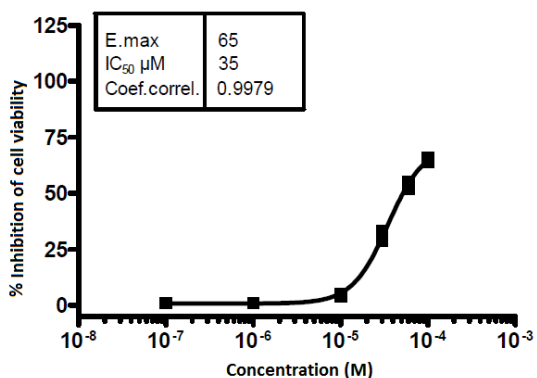
9



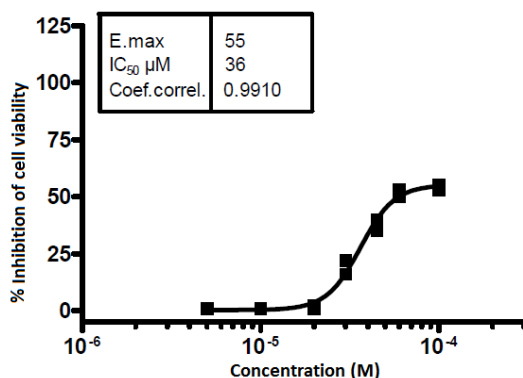
ent-9



10



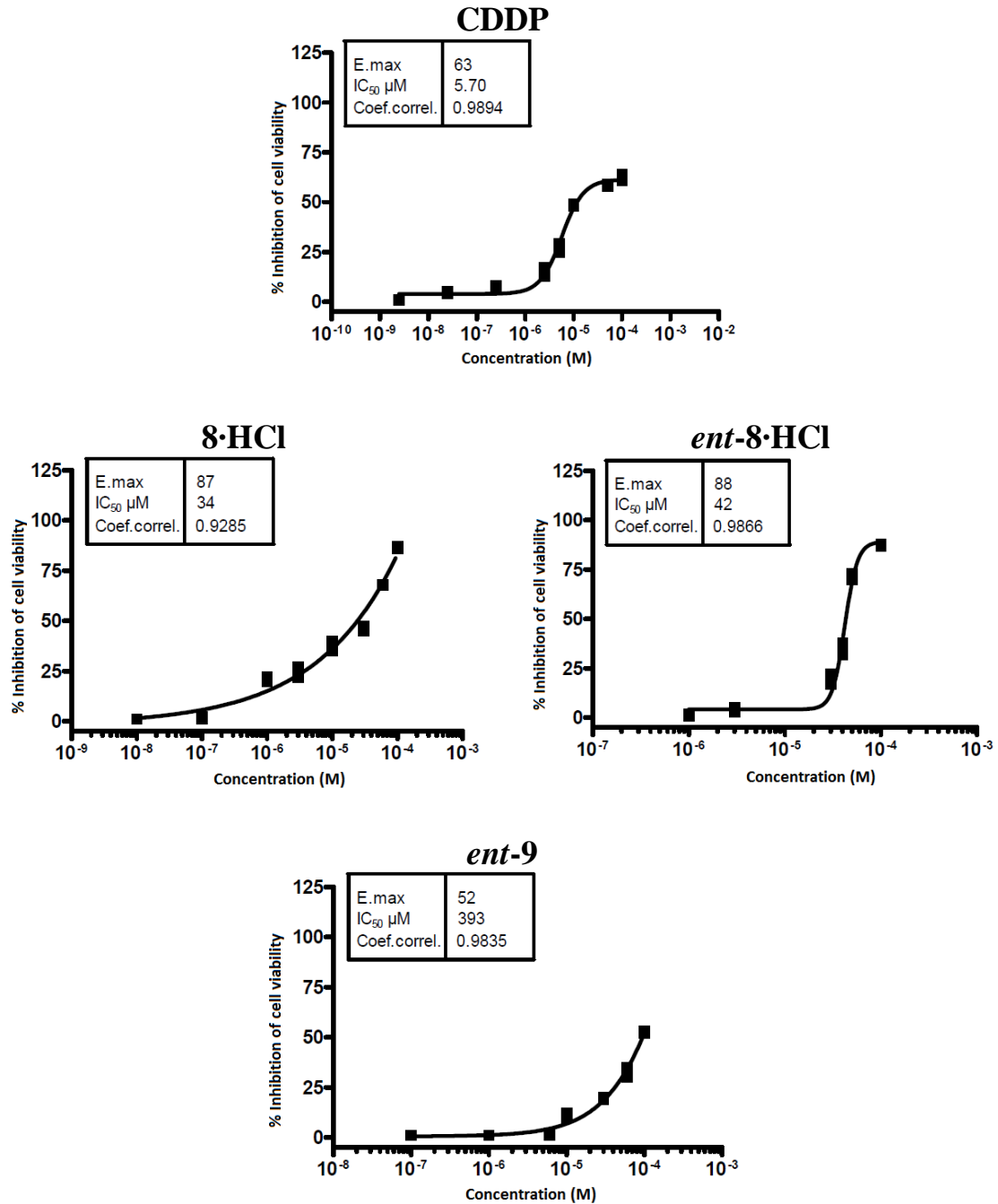
ent-10



NCI-H460

Seeding number: 15000

Incubation time with test compound: 48 h

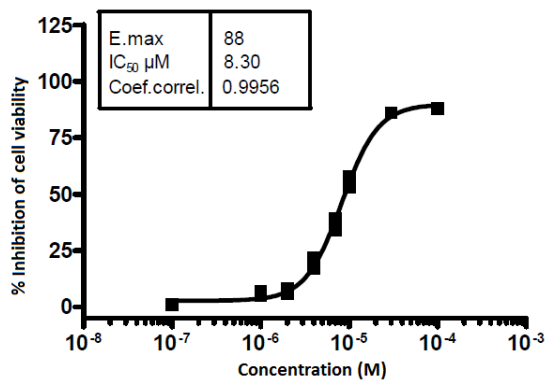


HL-60

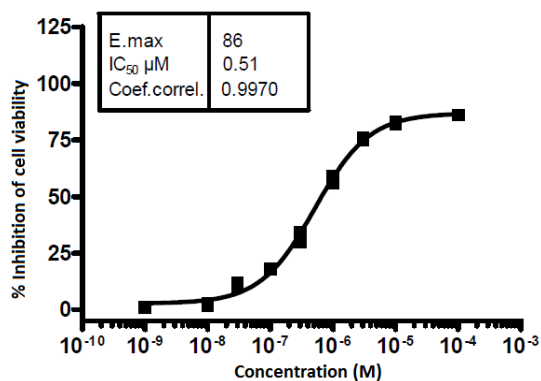
Seeding number: 10000

Incubation time with test compound: 24 h

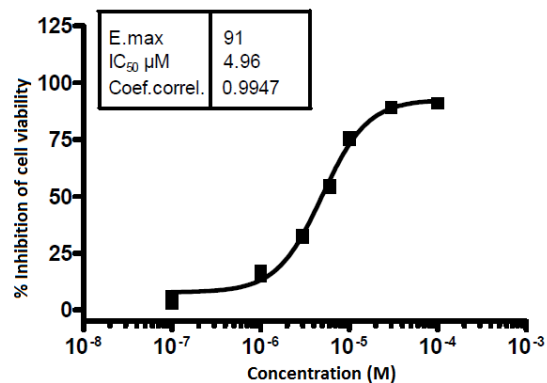
CDDP



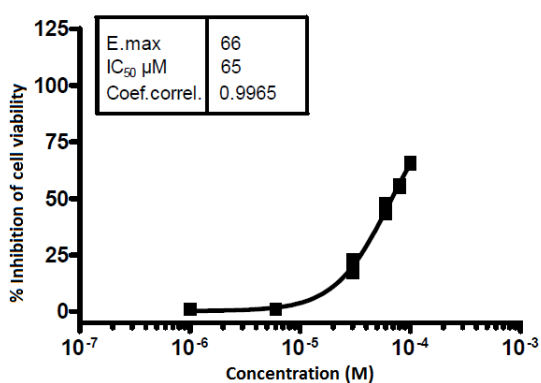
8·HCl



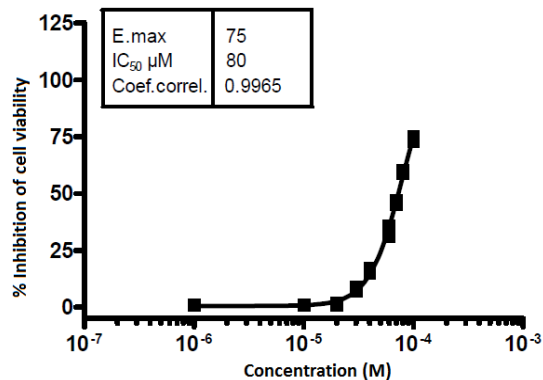
ent-8·HCl



9



10



NCI/ADR-RES

Seeding number: 15000

Incubation time with test compound: 48 h

